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## Binding of exogenous cyanide reveals new active-site states in [FeFe] hydrogenases†

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[FeFe] hydrogenases are highly efficient metalloenzymes for hydrogen conversion. Their active site cofactor (the H-cluster) is composed of a canonical [4Fe-4S] cluster ( $[4Fe-4S]_H$ ) linked to a unique organometallic di-iron subcluster ( $[2Fe]_H$ ). In  $[2Fe]_H$  the two Fe ions are coordinated by a bridging 2-azapropane-1,3-dithiolate (ADT) ligand, three CO and two  $CN^-$  ligands, leaving an open coordination site on one Fe where substrates ( $H_2$  and  $H^+$ ) as well as inhibitors (e.g.  $O_2$ , CO,  $H_2S$ ) may bind. Here, we investigate two new active site states that accumulate in [FeFe] hydrogenase variants where the cysteine (Cys) in the proton transfer pathway is mutated to alanine (Ala). Our experimental data, including atomic resolution crystal structures and supported by calculations, suggest that in these two states a third  $CN^-$  ligand is bound to the apical position of  $[2Fe]_H$ . These states can be generated both by "cannibalization" of  $CN^-$  from damaged  $[2Fe]_H$  subclusters as well as by addition of exogenous  $CN^-$ . This is the first detailed spectroscopic and computational characterisation of the interaction of exogenous  $CN^-$  with [FeFe] hydrogenases. Similar  $CN^-$ -bound states can also be generated in wild-type hydrogenases, but do not form as readily as with the Cys to Ala variants. These results highlight how the interaction between the first amino acid in the proton transfer pathway and the active site tunes ligand binding to the open coordination site and affects the electronic structure of the H-cluster.

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## Introduction

Hydrogenases are the most powerful natural catalyst for the production and utilization of molecular hydrogen.<sup>4,5</sup> Depending on the metal content of the cofactor at their active site, hydrogenases are classified as [FeFe], [NiFe] or [Fe]

hydrogenases.<sup>7</sup> For the [FeFe] type, the cofactor at the active site is called the H-cluster and consists of a canonical [4Fe-4S] cluster ( $[4Fe-4S]_H$ ) linked through a cysteine thiolate to a unique organometallic diiron cluster ( $[2Fe]_H$ ) (Fig. 1A).<sup>8,9</sup> In  $[2Fe]_H$ , the two irons (distinguished as proximal,  $Fe_p$ , and distal,  $Fe_d$ , depending on their relative distance from  $[4Fe-4S]_H$ ) are bridged by a 2-azapropane-1,3-dithiolate (ADT) ligand and a CO ligand, while additional CO and  $CN^-$  ligands are terminally bound to each Fe. The open coordination site at the apical position on  $Fe_d$  is where activation/formation of  $H_2$  occurs, but also where inhibitors including CO,<sup>10</sup>  $H_2S^{2,11}$  and  $O_2$ <sup>12</sup> bind. Binding of  $O_2$  generally leads to the destruction of the H-cluster, making [FeFe] hydrogenases highly oxygen-sensitive.<sup>12-15</sup>

Several states of the H-cluster differing in electron and proton distribution at the two subclusters and ligand binding to the open coordination site have been identified. However, the precise structure and the involvement of some of these states in the catalytic cycle (Fig. 1B) of [FeFe] hydrogenase is still a matter of debate.<sup>6,16</sup> In  $[2Fe]_H$  the strong-field CO and  $CN^-$  ligands stabilize low-spin and low-oxidation states for the two Fe ions, which cycle between Fe(II) and Fe(I) during catalysis. For instance, the active oxidized state  $H_{ox}$  has mixed valence  $Fe_p$ (II)  $Fe_d$ (I) in  $[2Fe]_H$ .<sup>17</sup> The one-electron reduced state  $H_{red}$  retains the  $Fe_p$ (II)  $Fe_d$ (I) valence in  $[2Fe]_H$  but has a reduced  $[4Fe-4S]_H$ . The

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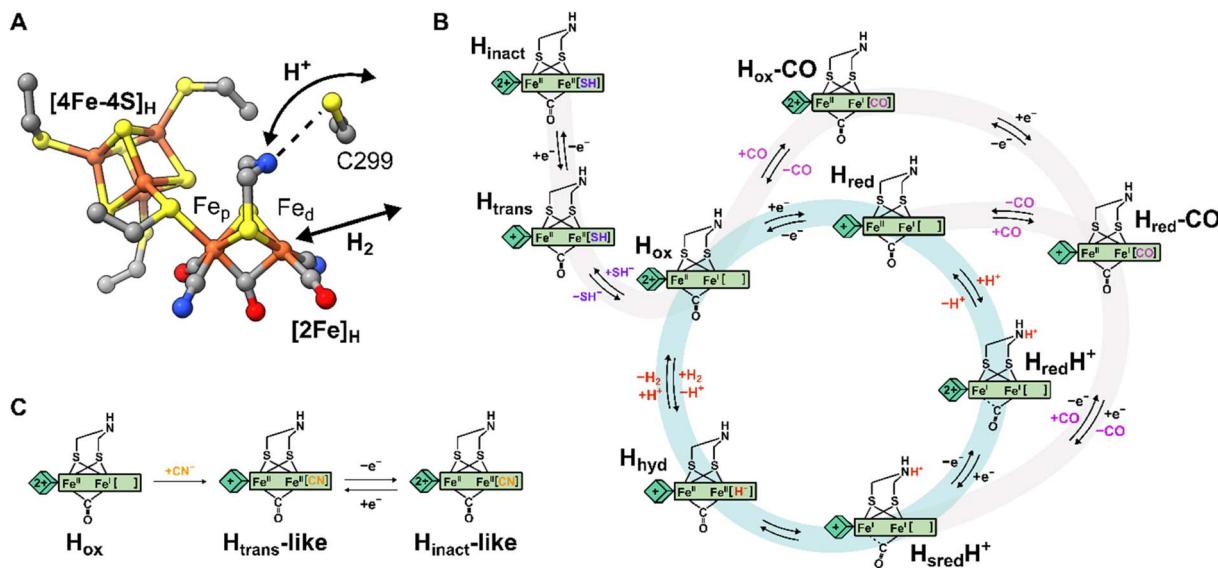
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**Fig. 1** Structure of the H-cluster and proposed catalytic cycle. (A) Structure of the [2Fe]<sub>H</sub> and [4Fe-4S]<sub>H</sub> subclusters from *Clostridium pasteurianum* HydA1 (CpHydA1, PDB: 4XDC)<sup>1</sup> with the Cys in the proton transfer pathway also shown (C299 in CpHydA1, C178 in DdHydAB, C169 in CrHydA1). (B) Proposed catalytic cycle also showing the pathways of reversible inactivation by H<sub>2</sub>S and CO, respectively. The pathway for H<sub>inact</sub> formation is only applicable to DdHydAB as the H<sub>trans</sub> state has not been identified for other [FeFe] hydrogenases. A different catalytic cycle has been proposed by some authors.<sup>6</sup> (C) Schematic showing the proposed chemical structure of the H<sub>trans</sub>-like and H<sub>inact</sub>-like states identified in this study.

reduced states H<sub>red</sub>H<sup>+</sup> and H<sub>sred</sub>H<sup>+</sup> are thought to have an Fe<sub>p</sub>(i) Fe<sub>d</sub>(i) configuration at [2Fe]<sub>H</sub> that is favored by concomitant protonation of the ADT ligand.<sup>18–20</sup> The crucial two-electron reduced H<sub>hyd</sub> state contains a terminal hydride on Fe<sub>d</sub> and an overoxidized [2Fe]<sub>H</sub> with a formal Fe<sub>p</sub>(ii)Fe<sub>d</sub>(ii) configuration.<sup>21–25</sup> Recently, states with a terminal hydride on Fe<sub>d</sub> differing in the redox state of [4Fe-4S]<sub>H</sub> have been identified.<sup>26</sup>

A similar overoxidized [2Fe]<sub>H</sub> can be found in two inactive states called H<sub>trans</sub> and H<sub>inact</sub>.<sup>11,27–30</sup> These states form upon reversible inactivation of [FeFe] hydrogenases by sulfide, which binds to the H-cluster under oxidizing conditions in some enzymes,<sup>2,11</sup> or by binding of a nearby cysteine thiolate in others.<sup>31–33</sup> In H<sub>inact</sub>, an RS<sup>−</sup> ligand (where R can be H or the rest of the cysteine amino acid) is thus bound in the apical position to Fe<sub>d</sub>, [2Fe]<sub>H</sub> is in the overoxidized state and the [4Fe-4S]<sub>H</sub> subcluster is oxidized ([4Fe-4S]<sub>H</sub><sup>2+</sup>–[Fe<sub>p</sub>(ii)Fe<sub>d</sub>(ii)–SR]<sub>H</sub>). Notably, the H<sub>inact</sub> state is stable under air as the RS<sup>−</sup> ligand prevents O<sub>2</sub> from binding to Fe<sub>d</sub>. Reversible one-electron reduction of H<sub>inact</sub>, in which SH<sup>−</sup> is bound, yields the H<sub>trans</sub> state, which has a reduced [4Fe-4S]<sub>H</sub><sup>+</sup> ([4Fe-4S]<sub>H</sub><sup>+</sup>–[Fe<sub>p</sub>(ii)Fe<sub>d</sub>(ii)–SH]<sub>H</sub>). Conversion of H<sub>trans</sub> to the active hydrogenase appears to require an additional reduction step.<sup>29</sup> However, the exact mechanism of conversion is not clear, but several theories have been proposed.<sup>2,11,34</sup> So far, H<sub>trans</sub> has not been identified in enzymes that bind a cysteine thiolate in H<sub>inact</sub>.<sup>33</sup>

Efficient exchange of protons between the solvent and the active site is crucial during H<sub>2</sub> conversion and is facilitated by a proton channel (also called the proton transfer pathway, PTP). This pathway is formed by a series of largely conserved (at least in prototypical hydrogenases)<sup>5</sup> amino acids and water

molecules that form a network of hydrogen bonds connecting the protein surface with the H-cluster.<sup>35–37</sup> Site-directed mutagenesis of amino acids along the proton transfer pathway can impair or even completely abolish catalytic activity, as a consequence of the slower proton exchange with the H-cluster.<sup>36,38–40</sup> Additionally, it was observed that some H-cluster states accumulate differently in wild-type enzymes and in variants with deficient proton transfer. Closest to the H-cluster is a cysteine residue (C299 in *Clostridium pasteurianum* HydA1, C178 in *Desulfovibrio desulfuricans* HydAB, DdHydAB, and C169 in *Chlamydomonas reinhardtii* HydA1) whose thiol is within hydrogen-bond distance of the bridgehead amine of the ADT ligand (Fig. 1A). When this Cys was mutated to alanine (Ala) or serine (Ser) in *Chlamydomonas reinhardtii* HydA1 (CrHydA1 C169A and C169S) and to Ser in *Clostridium pasteurianum* HydA1 (CpHydA1 C299S), these hydrogenases formed readily the H<sub>hyd</sub> state.<sup>21–23,39,41,42</sup> In addition, the CrHydA1 C169S variant was shown, using electron paramagnetic resonance (EPR) spectroscopy, to accumulate a state similar to H<sub>trans</sub>, but the precise nature of this state remains unknown.<sup>21,22,43</sup> Finally, the C169A variant of CrHydA1 has been reported to react with oxygen to form an H<sub>ox</sub>–O<sub>2</sub> state (so far observed only in this particular mutant), which has been suggested to have a superoxide bound to Fe<sub>d</sub> and an oxidized [4Fe-4S] cluster, yielding a [4Fe-4S]<sub>H</sub><sup>2+</sup>–[Fe<sub>p</sub>(i)Fe<sub>d</sub>(iii)–O<sub>2</sub><sup>−</sup>]<sub>H</sub> electronic configuration.<sup>44</sup> However, the infrared (IR) spectrum of this state is very similar to the H<sub>inact</sub> state and so a formal Fe<sub>p</sub>(ii)Fe<sub>d</sub>(ii) valence would seem more likely.

In this study, we investigated the effects of replacing the Cys in the proton transfer pathway with alanine in the hydrogenases DdHydAB and CrHydA1. DdHydAB is an exceptionally active

bidirectional hydrogenase that contains two additional [4Fe-4S] clusters (F-clusters) for electron transfer between the H-cluster and the protein surface. The mutation of amino acids along the proton transfer pathway of *DdHydAB* has not been investigated before. The C169A variant of *CrHydA1* has already been studied in particular in relation to the  $H_{\text{hyd}}$  and  $H_{\text{ox}}-\text{O}_2$  states,<sup>23,41,44</sup> but here we report two new active site states in *CrHydA1* C169A never identified before. In both *DdHydAB* C178A and *CrHydA1* C169A, we observed formation of unprecedented H-cluster states similar to  $H_{\text{trans}}$  and  $H_{\text{inact}}$ . By combining their spectroscopic and structural characterization, we demonstrated that these  $H_{\text{trans}}$ -like and  $H_{\text{inact}}$ -like states form upon binding of  $\text{CN}^-$  to the H-cluster (Fig. 1C). These  $\text{CN}^-$ -bound states form also in wild-type (WT) hydrogenases, but are stabilized in the Cys to Ala mutants. This study highlights how the interaction between the Cys in the proton transfer pathway and the H-cluster (specifically the bridgehead amine in [2Fe]<sub>H</sub>) tunes the electronic structure of the H-cluster and regulates ligand binding to the apical position of  $\text{Fe}_d$ .

## Results

### *DdHydAB* C178A is isolated in an $H_{\text{trans}}$ -like state

The *DdHydAB* C178A mutant was recombinantly expressed in *E. coli* as an “apo”-hydrogenase (*i.e.* containing the [4Fe-4S]<sub>H</sub> subcluster and all the accessory F-clusters but lacking [2Fe]<sub>H</sub>) and artificially matured *in vitro*, as routinely performed with the WT enzyme.<sup>45</sup> The WT *DdHydAB* is commonly isolated after maturation (under 2%  $\text{H}_2$  and 98%  $\text{N}_2$ ) as a mixture of states,

mainly  $\text{H}_{\text{ox}}$ ,  $\text{H}_{\text{ox}}-\text{CO}$ , and  $\text{H}_{\text{red}}\text{H}^+$ .<sup>45</sup> As shown in the IR spectra in Fig. 2A, after artificial maturation the C178A mutant was, surprisingly, isolated in an almost pure unprecedented state that greatly differs from the states normally observed in freshly matured WT *DdHydAB* (Table S1†). In particular, the spectrum of the as isolated C178A mutant exhibits a broad absorption at  $1853\text{ cm}^{-1}$  attributed to the bridging CO ligand, a single broad band at  $1989\text{ cm}^{-1}$  attributed to the terminal CO ligands (potentially resulting from the two overlapping CO bands, exhibiting a shoulder at  $\sim 2002\text{ cm}^{-1}$ ), and three bands at  $2116$ ,  $2100$  and  $2087\text{ cm}^{-1}$  attributed to  $\text{CN}^-$  ligands (instead of the expected two bands). Relative to the IR bands observed for the  $\text{H}_{\text{ox}}$  state in WT *DdHydAB*, the IR bands of the C178A variant are shifted to higher energies (blue-shifted) suggesting decreased electron density on [2Fe]<sub>H</sub>. The frequencies of the IR bands are very similar to those of the  $H_{\text{trans}}$  state in WT *DdHydAB* (Table S1†),<sup>2,29</sup> therefore, we hypothesized that this new state could be an  $H_{\text{trans}}$ -like state (*i.e.* with the same electronic configuration  $[4\text{Fe}-4\text{S}]_{\text{H}}^+ - [\text{Fe}_p(\text{II})\text{Fe}_d(\text{II})]_{\text{H}}$ ). An  $H_{\text{inact}}$ -like state ( $[4\text{Fe}-4\text{S}]_{\text{H}}^{2+} - [\text{Fe}_p(\text{II})\text{Fe}_d(\text{II})]_{\text{H}}$ ) was formed both by oxidation of the as-isolated C178A variant under anaerobic conditions (using hexaammineruthenium(III) chloride, HAR) or by exposure to atmospheric oxygen (Fig. 2A, green bands), yielding identical IR spectra in each case. Exposure of the as-isolated sample to air did not result in any significant decrease in IR signal intensity, suggesting that all active site species present in the  $H_{\text{trans}}$ -like state transform into an  $H_{\text{inact}}$ -like state. Notably, this  $H_{\text{inact}}$ -like state is another example of an air-stable state in [FeFe] hydrogenases. Therefore, in addition to having the same

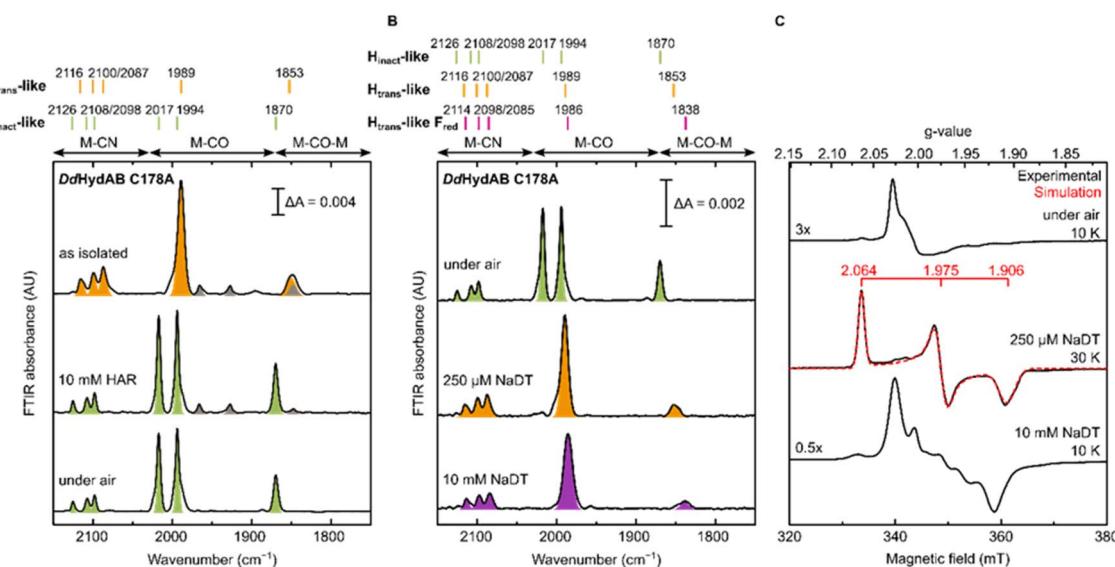


Fig. 2 IR and EPR spectra of *DdHydAB* C178A exhibit new  $H_{\text{trans}}$ -like and  $H_{\text{inact}}$ -like states. (A) IR spectra of freshly matured *DdHydAB* C178A (in 25 mM Tris pH 8.0, 25 mM KCl): as isolated; oxidized under anaerobic conditions (with 10 mM HAR); oxidized under air. Bands from the  $H_{\text{trans}}$ -like and  $H_{\text{inact}}$ -like states are colored in orange and green, respectively. (B and C) Samples of *DdHydAB* C178A were prepared under different conditions. One aliquot was used for room temperature IR measurements (B), while the remaining sample was used for CW X-band EPR measurements (C). In (A and B), bands are color coded as follows: green for  $H_{\text{inact}}$ -like state, orange for  $H_{\text{trans}}$ -like state, purple for  $H_{\text{trans}}$ -like  $\text{F}_{\text{red}}$  state. The gray bands correspond to traces of an unidentified state that potentially lacks the third  $\text{CN}^-$  ligand (which protects the H-cluster from  $\text{O}_2$  attack), as this state disappears after exposure to air. In (C), experimental spectra are in black, overlaid in one case with spectral simulations (dashed red line). EPR experimental conditions: microwave frequency = 9.64 GHz; microwave power = 1 mW for the first two conditions (under air, 250  $\mu\text{M}$  NaDT), 0.1 mW for the bottom one (10 mM NaDT); temperature is specified in the figure.



electronic configuration of  $H_{trans}$  and  $H_{inact}$ , respectively, we hypothesized that the new states might have an additional ligand (likely different from  $SH^-$ ) bound at the open coordination site.

To confirm our assignment of  $H_{trans}$ -like and  $H_{inact}$ -like states, we measured EPR spectra of *DdHydAB* C178A poised in different states as confirmed by IR spectroscopy (Fig. 2C). The  $H_{inact}$ -like state is EPR silent, like the  $H_{inact}$  state in WT *DdHydAB*.<sup>30</sup> Only a signal from a [3Fe-4S] cluster could be detected, probably due to oxidative damage to the F-cluster located in vicinity of the protein surface. Addition of one equivalent of reducing agent (sodium dithionite, NaDT) to the *DdHydAB* C178A sample, which had partially converted to the  $H_{inact}$ -like state during storage (Fig. S1A†) reverts it to the  $H_{trans}$ -like state (Fig. 2B). The  $H_{trans}$ -like state exhibits a rhombic EPR signal ( $g = 2.06, 1.98, 1.91$ ) similar to the one of  $H_{trans}$  in WT *DdHydAB* (Fig. S2†),<sup>30</sup> confirming our initial assignment of *DdHydAB* C178A being isolated in an  $H_{trans}$ -like state. Addition of excess NaDT results in only minor shifts to lower energies (red-shift) of all IR bands and gives rise to a complex EPR interaction spectrum. We interpreted this behaviour with the H-cluster remaining in an EPR-active  $H_{trans}$ -like state while the accessory F-clusters are being reduced to EPR-active states by the excess of NaDT, resulting in strong dipolar spin-coupling ( $H_{trans}$ -like  $F_{red}$  state) as observed previously for the WT enzyme in the  $H_{ox}$  or  $H_{ox}$ -CO state with reduced F-clusters.<sup>46</sup> Samples in the as-isolated state and purged with CO showed no change to the IR spectrum (Fig. S1B†) indicating that this state was unable to bind CO, most likely due to an already occupied coordination site.

### Evidence for an additional $CN^-$ ligand at the H-cluster in *DdHydAB* C178A

For all the states we could observe in *DdHydAB* C178A ( $H_{trans}$ -like,  $H_{inact}$ -like and  $H_{trans}$ -like  $F_{red}$ ), the IR spectra always exhibit three bands in the  $CN^-$  region. To test if all the bands derive from  $CN^-$  vibrations associated with the H-cluster, we performed the artificial maturation of the C178A mutant with a precursor of the [2Fe]<sub>H</sub> cluster with both  $CN^-$  ligands labelled with <sup>13</sup>C. As shown in Fig. 3A, we observed an isotope shift (46–44  $cm^{-1}$ ) of all three  $CN^-$  absorptions in the IR spectrum of the  $H_{inact}$ -like state. We could interpret these results in three ways: (i) three  $CN^-$  ligands are present at the H-cluster; (ii) two  $CN^-$  vibrations couple in an unusual way giving rise to three IR bands; (iii) spectra represent two very similar states with one strongly overlapping  $CN^-$  band, while the other  $CN^-$  band in each state is distinct. The second hypothesis is not likely as the  $CN^-$  ligands are on different Fe ions and such a structure is unlikely to give significant quadratic coupling.<sup>29,47</sup> Previous isotope editing experiments on the CO ligands in WT enzyme showed very little perturbation in the vibrational frequency of the pCO ligand (the terminal CO on  $Fe_p$ ), when dCO (the terminal CO on  $Fe_d$ ) or  $\mu$ CO (the bridging CO) were exchanged with <sup>13</sup>CO.<sup>29,47</sup>

The isotope shift could be reproduced by quantum mechanics/molecular mechanics (QM/MM) calculations of an

H-cluster model ( $Fe(p)Fe(d)$  redox state) of the C178A variant of *DdHydAB*, with  $CN^-$  as the exogenous ligand on  $Fe_d$ , as shown in Fig. 3B. Two protonation states of the amine in the ADT ligand were calculated: singly protonated (ADT) and doubly protonated (ADTH). The experimental <sup>13</sup>CN isotope shifts of 43–46  $cm^{-1}$  could be satisfactorily reproduced with both models: 43–46  $cm^{-1}$  (ADT) and 43–45  $cm^{-1}$  (ADTH). The absolute experimental frequencies are reasonably well reproduced by scaled harmonic frequencies, though with some differences between ADT and ADTH models. The terminal CO modes were somewhat better predicted by the ADTH model while the  $CN^-$  modes and bridging CO modes were better predicted with the ADT model. We note that the CO frequencies are quite dependent on the quality of the model, density functional and scaling factor while the  $CN^-$  frequencies are less so (Fig. S3 and Materials and methods in the ESI†). The calculated relative intensities of the three  $CN^-$  modes, however, differ more strongly between models. The ADT model predicts an increase in  $CN^-$  mode intensity with decreasing energy, consistent with the experimental intensities, while the ADTH model does not. The reason is that the order of the assigned  $CN^-$  modes differs between ADT/ADTH models; the exogenous  $CN^-$  mode is the highest-energy  $CN^-$  mode for the ADT model but it is the lowest for the ADTH. These differences can be explained by a stronger exogenous  $CN^-$ -binding in the ADTH model (aided by stronger H-bonding to the doubly protonated amine). Other conformers of the ADT and ADTH models were explored (Fig. S4†) but were found to be energetically unfavorable. Overall the calculations suggest the  $H_{inact}$ -like state as best described by an  $[Fe_p(p)Fe_d(d)]_H$  model featuring an exogenous  $CN^-$  ligand in the apical position with a singly protonated bridgehead amine of the ADT ligand.

To further investigate the properties of the H-cluster in the C178A variant of *DdHydAB*, we solved crystal structures of the enzyme in the  $H_{inact}$ -like and the  $H_{trans}$ -like states using X-ray crystallography (Fig. 4 and S5–S9†). After exposure to air to form the  $H_{inact}$ -like state, the protein was crystallized under aerobic conditions (Fig. 4A and S7†) as performed previously for the  $SH$ -bound  $H_{inact}$  state in the WT enzyme.<sup>11</sup> IR and resonance Raman (RR) measurements on crystals<sup>48</sup> prepared under the same conditions confirmed that the enzyme was indeed in the  $H_{inact}$ -like state (Fig. S5†). However, small shifts in the band positions compared with solution measurements were observed and are likely related to temperature-dependent changes or crystal packing effects. Such effects have been observed previously for [FeFe]<sup>11,49</sup> and [NiFe] hydrogenases.<sup>48,50,51</sup> We also solved a structure of the  $H_{trans}$ -like state of the enzyme from crystals grown under anaerobic conditions (2%  $H_2$ , 98%  $N_2$ ) (Fig. 4B). *DdHydAB* C178A crystallized in an orthorhombic space group  $P2_12_12_1$  as observed for the previously reported WT *DdHydAB* in the  $H_{inact}$  state.<sup>11</sup> The crystals diffracted up to 1.0  $\text{\AA}$  and the structures were solved at high resolution, 1.04  $\text{\AA}$  for the  $H_{inact}$ -like state and 1.01  $\text{\AA}$  for the  $H_{trans}$ -like state. The overall structure of the C178A variant in the  $H_{inact}$ -like state and the WT enzyme in the  $H_{inact}$  state is virtually identical with an RMSD of 0.237  $\text{\AA}$  (calculated for all  $C\alpha$  atoms of all residues, Table S2) (Fig. S7 and S8†). The structure at atomic resolution clearly



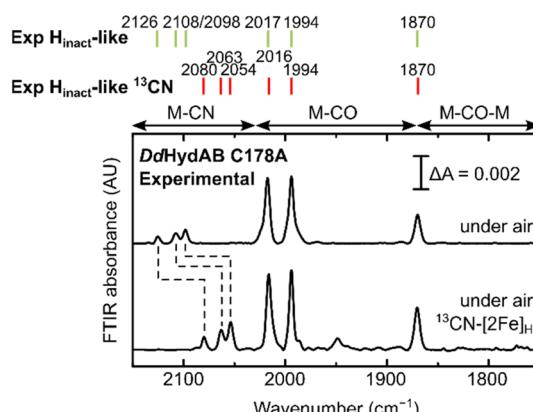
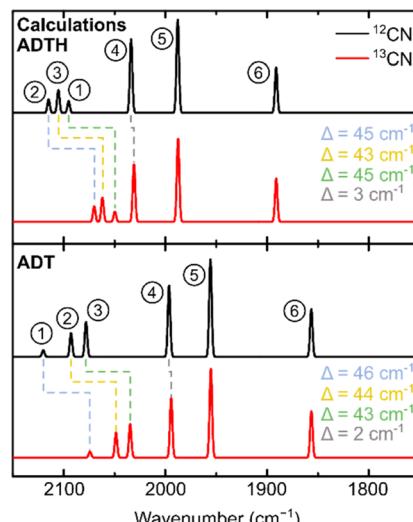
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Fig. 3 (A) Experimental IR spectra of *DdHydAB* C178A matured with natural abundance (top) and  $^{13}\text{CN}^-$ -labeled  $[2\text{Fe}]_H$  precursor (bottom), both exposed to air to form the  $\text{H}_{\text{inact}}$ -like state. The small additional feature in the  $^{13}\text{CN}$  spectrum may represent a small amount of an unknown degradation product. (B) Calculated IR spectra of the H-cluster model with either a singly protonated (ADT) or doubly protonated (ADTH) ADT ligand. Insets are  $[2\text{Fe}]_H$  structures with chemical groups associated with modes labelled. A scaling factor of 0.964 was used.

shows a diatomic ligand in the apical position of  $\text{Fe}_d$  and the Ala residue that replaced the Cys at position 178. Moreover, we identified two additional well-defined water molecules appearing near the Ala178. Interestingly, in the crystal structure of the C299A variant from *CpHydA1* reported by Duan *et al.*,<sup>36</sup> the

space of the missing thiol group was replaced by an additional  $\text{H}_2\text{O}$  molecule (Wat962), which was located at 3.4–3.7 Å from the NH group of the ADT ligand and at 3.6–3.7 Å from the Wat826 molecule of the PTP.<sup>36</sup> This new water molecule was hypothesised to rescue proton transfer activity in the absence of the

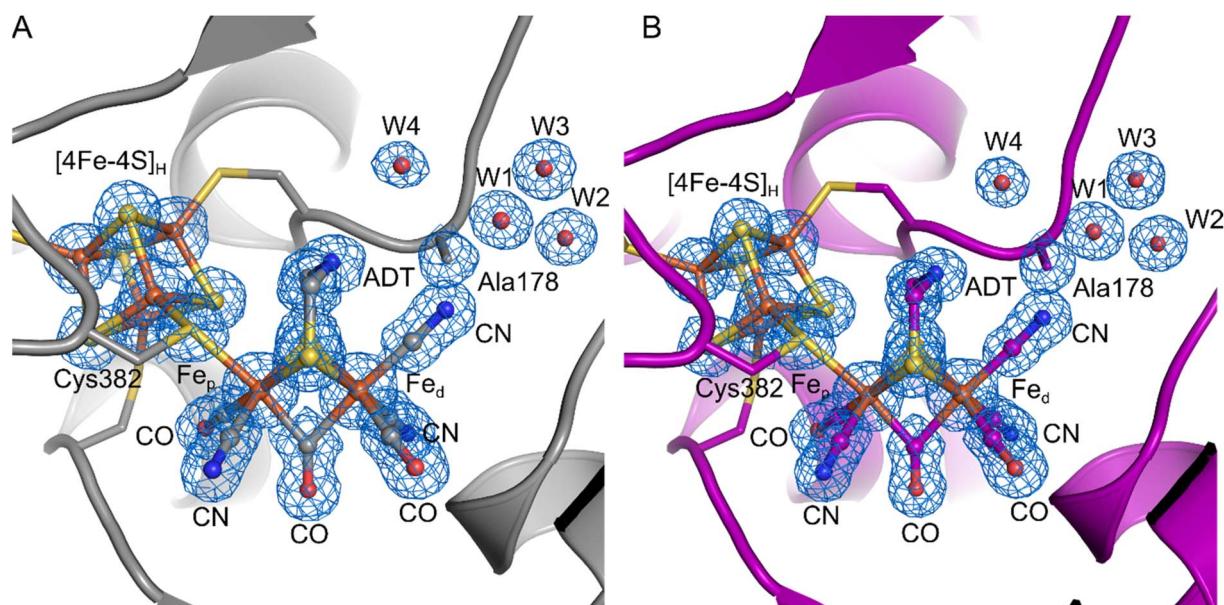


Fig. 4 Crystal structures of the *DdHydAB* C178A mutant in two different states. (A) *DdHydAB* in the  $\text{H}_{\text{inact}}$ -like state (PDB ID 8BJ7) is shown as cartoon and colored in gray. (B) *DdHydAB* in the  $\text{H}_{\text{trans}}$ -like state (PDB ID 8BJ8) is shown as cartoon and colored in magenta. Close-up view of the active site showing the H-cluster, the Cys ligating the cofactor, the side chain of Ala178 and the well-defined water molecules with a distance  $<4.0$  Å from Ala178. The protein backbone is shown as cartoon, the amino acid side chains and the H-cluster including the bound  $\text{CN}^-$  ligand are shown as stick model, and water molecules are shown as spheres. The cofactors and amino acid side chains are colored according to the element-specific color code. A 2Fo–Fc electron density map (blue mesh, contoured at  $1.0\sigma$ ) is shown for the H-cluster including the  $\text{CN}^-$  ligand, the side chain of Ala178, and the water molecules.



thiol; however, the authors could not measure any significant catalytic activity.

The overall architecture of the  $H_{inact}$ -like and the  $H_{trans}$ -like states are also virtually identical with an RMSD of 0.053 Å (calculated for all  $C\alpha$  atoms of all residues, Table S2) (Fig. S8†). While for the structure of WT *DdHydAB* in the  $H_{inact}$  state a reduced occupancy of the  $[2Fe]_H$  subcluster led to better agreement between modelled and experimental data,<sup>11</sup> here we observed no negative difference density when refining the structural models with an occupancy of 100% for the  $[2Fe]_H$  subcluster. This could be evidence for a better incorporation of the  $[2Fe]_H$  subcluster during artificial maturation or higher stability of the H-cluster during crystallization in the C178A mutant compared to the WT protein.

The crystal structure clearly shows a well-defined, relatively symmetric bridging CO (Fig. S9†), with roughly equal  $Fe_p-C_b$  and  $Fe_d-C_b$  bond distances. A similar observation was made for the  $H_{inact}$  state in wild type *DdHydAB*,<sup>11</sup> whereas other  $[FeFe]$  hydrogenase structures show slight lengthening of the  $Fe_p-C_b$  bond and shortening of the  $Fe_d-C_b$  bond.<sup>8,9,52</sup> The differences here are attributed to the oxidation states and coordination environment of  $Fe_p$  and  $Fe_d$ . In our structure with cyanide bound and the previously published  $H_{inact}$  state<sup>11</sup> both Fe ions were  $Fe^{(II)}$  and hexacoordinate. Meanwhile for structures obtained of the active enzyme,<sup>8,9,52</sup> the Fe ions are more reduced (for  $H_{ox}$   $Fe_d$  is reduced to  $Fe^{(I)}$  and for  $H_{red}H^+$  both Fe ions are reduced to  $Fe^{(I)}$ ) and  $Fe_d$  is pentacoordinate. These effects lead to shortening of the  $Fe_d-C_b$  bond giving a semi-bridging CO.

On the basis of these results and the fact that the IR spectra of all  $H_{inact}$ -like and  $H_{trans}$ -like states in *DdHydAB* C178A exhibit three  $CN^-$  bands, we suggest that in these states a third  $CN^-$  is present at the H-cluster, bound at the apical position on  $Fe_d$ . Therefore, in the structure we modelled a  $CN^-$  ligand coordinated to the distal iron ion through its carbon atom, with a  $Fe-C$  distance of 1.90 Å. To confirm the assignment of the ligands, we calculated an omit map in the absence of the  $[2Fe]_H$  subsite and the additional ligand at the apical position on  $Fe_d$ . The omit map (Fig. S9†) supports the positioning of the atoms in the electron density. In addition, we are able to distinguish between the N and O atoms of the terminal and bridging ligands when increasing the contouring level to  $\sigma = 2.8$  Å (Fig. S9C†). This result suggests that the exogenous  $CN^-$  ligand remains in the apical position, which agrees with the QM/MM calculations as well as the illumination experiments on *CrHydA1* C169A (see below).

### A similar $H_{trans}$ -like state in *CrHydA1* C169A

To gain further understanding of these  $CN^-$  bound states and on the role of the Cys in the proton transfer pathway in their formation, we decided to re-investigate the C169A variant of *CrHydA1*. As mentioned in the introduction, this particular mutation in *CrHydA1* has already been studied and showed accumulation of the  $H_{hyd}$  and  $H_{ox}-O_2$  states.<sup>23,41,44</sup> In contrast with what we observed with *DdHydAB* C178A, and consistent with the previous reports on *CrHydA1* C169A, this mutant is isolated after artificial maturation without an additional  $CN^-$

ligand on the H-cluster. Indeed, *CrHydA1* C169A is initially isolated under 2%  $H_2$  in a mixture of the  $H_{hyd}$  state (which can be enriched upon addition of NaDT) and another state that has been previously assigned as  $H_{ox}$  on the basis of the position of the IR bands (Fig. 5A).<sup>23,53</sup> However, the EPR spectrum of the as-isolated enzyme lacks the characteristic rhombic signal for the  $H_{ox}$  state and shows only the  $H_{hyd}$  state as major component (75%) (Fig. 5B). The EPR spectrum of the  $H_{hyd}$  state in *CrHydA1* C169A ( $g = 2.075, 1.942, 1.884$ ) is nearly identical to the one reported for the same state in the C169S mutant ( $g = 2.07, 1.94, 1.88$ ).<sup>22</sup> Therefore, we suggest that the state originally labelled as  $H_{ox}$  may instead be an EPR silent state with a similar electronic structure to the  $H_{red}$  state ( $H_{red}$ -like, light blue IR bands in Fig. 5A), which has a reduced  $[4Fe-4S]_H$  ( $[4Fe-4S]_H^{+/-}[Fe_{p(H)}Fe_{d(I)}]_H$ ). Oxidation of this state with one equivalent of oxidizing agent (HAR) under anaerobic conditions formed a new state with its most intense IR band at 1948 cm<sup>-1</sup>, with an EPR spectrum ( $g = 2.104, 2.046, 2.000$ ) similar to that observed for the  $H_{ox}$  state in WT *CrHydA1* (dark blue bands in Fig. 5A). Interestingly, these  $H_{red}$ -like and  $H_{ox}$ -like states have similar FTIR spectra to those observed for the recently characterised  $H_{ox}H$  and  $H'_{red}H$  states,<sup>53-55</sup> with bands shifted to higher energy compared with the  $H_{red}$  and  $H_{ox}$  states in WT *CrHydA1*.

Despite being initially isolated in states lacking an additional  $CN^-$  ligand ( $H_{red}$ -like,  $H_{hyd}$ ), incubation of *CrHydA1* C169A (pH 8) in the glovebox (2%  $H_2$ , 98%  $N_2$ ) for 24 h at room temperature led to the appearance in the IR spectrum of an  $H_{trans}$ -like state similar to the one observed in *DdHydAB* C178A, including a third  $CN^-$  band appearing at high energy (2114 cm<sup>-1</sup>, Fig. 5A). This  $H_{trans}$ -like state could also be enriched upon addition of exogenous  $CN^-$  to freshly maturated enzyme (Fig. 5A), which is initially isolated as a mixture of  $H_{red}$ -like and  $H_{hyd}$  (Fig. 5A).  $CN^-$  binding to the  $H_{hyd}$  state seems less favored, consistent with the presence of a terminal hydride bound to  $Fe_d$  in  $H_{hyd}$  (Fig. 5A). Therefore, complete conversion of the as-isolated enzyme to the  $H_{trans}$ -like state required addition of half an equivalent of oxidizing agent (HAR) to first oxidize the  $H_{hyd}$  state (Fig. 5A). Addition of excess  $CN^-$  induces partial degradation of the H-cluster, as demonstrated by the appearance in the IR spectra of a broad band around 2037 cm<sup>-1</sup>, which suggests formation of  $[Fe(CN)_6]^{4-}$  (Fig. 5A),<sup>3</sup> as already reported for other Fe-containing metalloenzymes like CODH upon treatment with  $CN^-$ .<sup>56</sup> Therefore, after the formation of the  $H_{trans}$ -like state, samples were buffer exchanged to remove degradation products as well as the excess of free  $CN^-$  to give cleaner IR spectra as in Fig. 5 and 6.

The EPR spectrum of the  $H_{trans}$ -like state in *CrHydA1* C169A could be simulated with two components having similar rhombic signals (Fig. 5B). The first component ( $g = 2.068, 1.977, 1.916$ ), accounting for ca. 87% of the signal, resembles the EPR spectrum of the  $H_{trans}$ -like state in *DdHydAB* C178A as well as a similar  $H_{trans}$ -like state previously observed in *CrHydA1* C169S ( $g = 2.065, 1.969, 1.906$ ), which was never assigned to a particular structure of the H-cluster.<sup>21,22,43</sup> The second rhombic component ( $g = 2.066, 2.008, 1.935$ ) could potentially relate to a different protein or H-cluster conformation. As observed for *DdHydAB* C178A, oxidation of *CrHydA1* C169A in the  $H_{trans}$ -like state yields



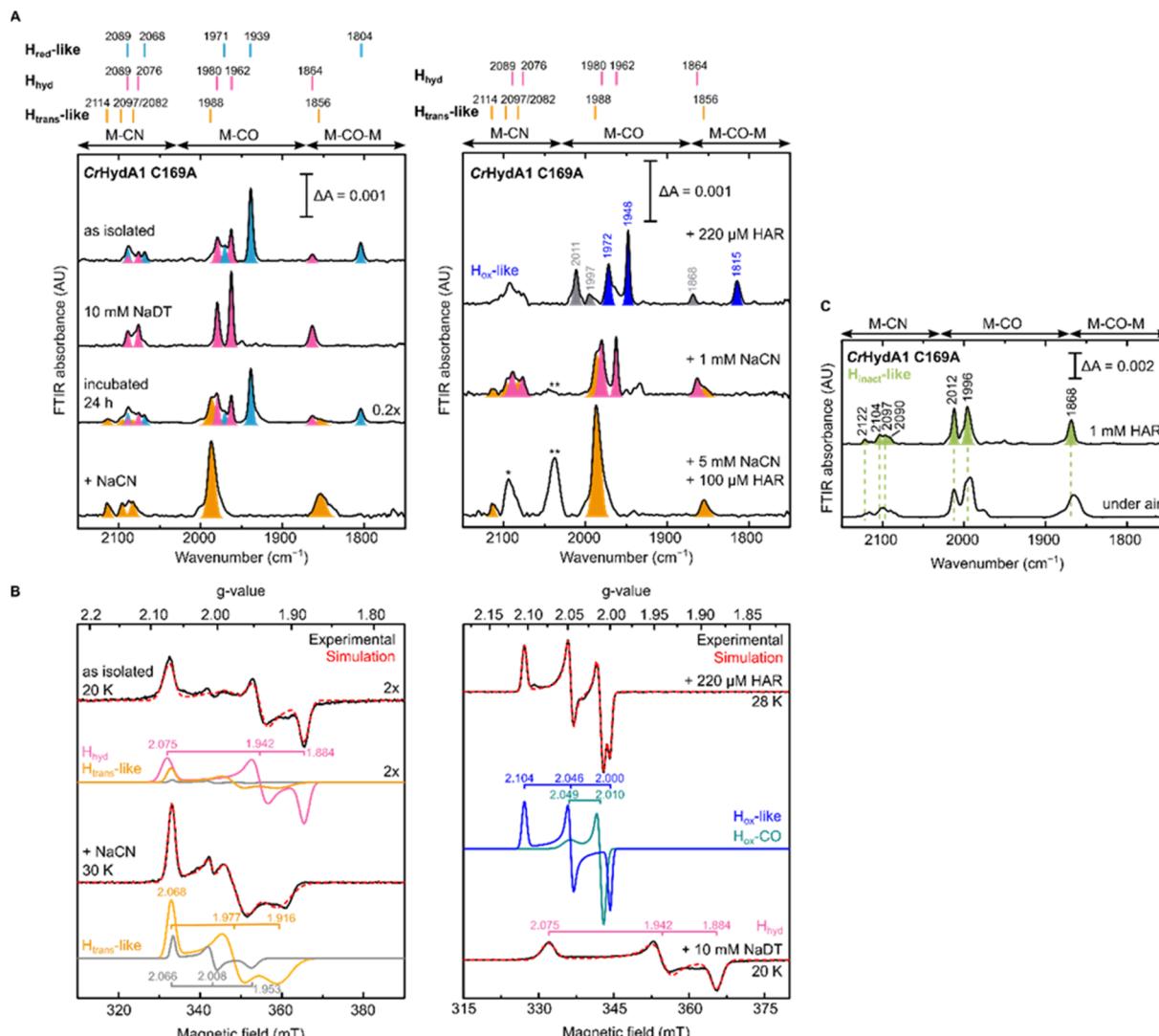


Fig. 5 IR and EPR spectra suggest formation of a CN<sup>−</sup>-dependent H<sub>trans</sub>-like state also in CrHydA1 C169A. (A) Room temperature IR spectra of CrHydA1 C169A under different conditions: as isolated; with NaDT; after 24 h incubation under 2% H<sub>2</sub>; after addition of 5 mM NaCN, 100 μM HAR and then buffer exchanged ("+NaCN"); oxidized with 220 μM (1.1 eq.) HAR; after addition of 1 mM NaCN; after addition of 5 mM NaCN and 100 μM (0.5 eq.) HAR. Bands are color-coded as follows: light blue for H<sub>red</sub>-like, pink for H<sub>hyd</sub>, orange for the H<sub>trans</sub>-like, and blue for the H<sub>ox</sub>-like state. Bands in gray correspond to traces of the H<sub>inact</sub>-like and H<sub>ox</sub>-CO states. For the H<sub>ox</sub>-like state the bands in the complex CN<sup>−</sup> region could not be assigned. In the H<sub>hyd</sub> state, CN<sup>−</sup> binding is likely disfavored as a hydride is bound to Fe<sub>d</sub>. The single asterisk marks the band of HCN (2093 cm<sup>-1</sup>), while the double asterisk marks the band of [Fe(CN)<sub>6</sub>]<sup>4−</sup> (2037 cm<sup>-1</sup>),<sup>3</sup> which suggests partial cofactor degradation upon CN<sup>−</sup> addition. Clean spectra for the H<sub>trans</sub>-like state where obtained after buffer-exchanging the protein to eliminate HCN, free CN<sup>−</sup> and degradation products. (B) CW X-band EPR spectra for some of the conditions shown in (A). Experimental spectra are shown in black and are overlaid with spectral simulations (dashed red line) with component spectra underneath. The pink component corresponds to the H<sub>hyd</sub> state. The orange component is likely the H<sub>trans</sub>-like state, while the gray component may represent an alternative, as yet unidentified, state. Presence of the H<sub>trans</sub>-like components in the EPR spectrum of as isolated enzyme suggests that a small amount of H<sub>trans</sub>-like state has already formed in freshly matured CrHydA1 C169A (the small shoulder at 1988 cm<sup>-1</sup> in the IR spectrum of the same sample is also consistent with the presence of traces of the H<sub>trans</sub>-like state). The blue trace corresponds to the H<sub>ox</sub>-like state, while the dark cyan trace corresponds to the H<sub>ox</sub>-CO state. EPR experimental conditions: microwave frequency = 9.64 GHz; microwave power = 1 mW; temperature is specified in the figure. (C) IR spectra of the H<sub>inact</sub>-like states in CrHydA1 C169A. From the H<sub>trans</sub>-like state, two similar but slightly different H<sub>inact</sub>-like states form by oxidation of the enzyme with HAR under anaerobic (top) or by oxidation by atmospheric oxygen (bottom).

an H<sub>inact</sub>-like state. However, two similar H<sub>inact</sub>-like states were formed depending on whether the oxidation was performed under anaerobic conditions by HAR or by atmospheric oxygen (Fig. 5B). A reason for this difference could be damage to the [4Fe-4S]<sub>H</sub> during the different oxidative treatments. Notably,

illumination of the air-oxidized CrHydA1 C169A at cryogenic temperature (Fig. S10†) did not reveal any photosensitivity, confirming again the assignment of a terminally bound CN<sup>−</sup> for the H<sub>trans</sub>-like and H<sub>inact</sub>-like states, since a CO species in the apical position would likely be photolyzed.<sup>29,30</sup>

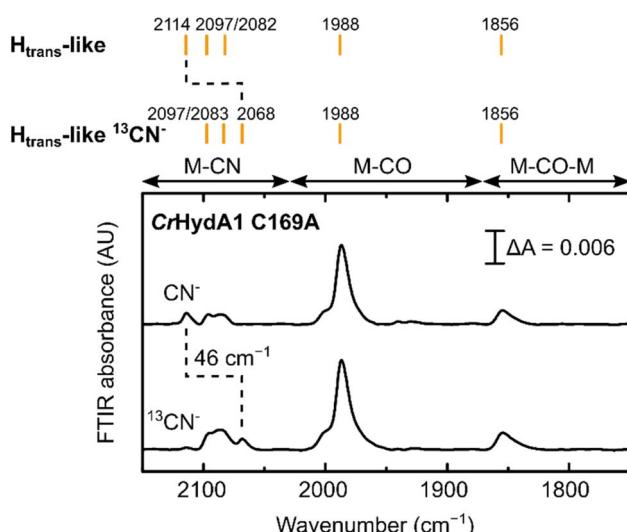


Fig. 6 IR spectra of *CrHydA1* C169A prepared in the  $H_{trans}$ -like state using natural abundance and  $^{13}C$  labelled  $CN^-$  exhibit an expected isotope shift of one of the  $CN^-$  bands. Small peaks in the region between 1930 and 1940  $cm^{-1}$  are likely due to small contributions from states without  $CN^-$  bound.

Treatment of freshly-maturated *CrHydA1* C169A with  $^{13}CN^-$  yielded an  $H_{trans}$ -like state which exhibited an isotope shift ( $46\text{ cm}^{-1}$ ) of one of the  $CN^-$  band in the IR spectrum (Fig. 6). This result unambiguously confirms that a third  $CN^-$  ligand is bound at the H-cluster in the  $H_{trans}$ -like and  $H_{inact}$ -like states. Additionally, this result allows us to assign the exogenous  $CN^-$  ligand on  $Fe_d$  to the band with the highest vibrational frequency (in samples prepared with  $^{12}CN^-$ ) among the three  $CN^-$  bands. In line with this observation, resonance Raman (RR) measurements on *CrHydA1* C169A in the  $H_{inact}$ -like state (Fig. S11A†) revealed 1–3  $cm^{-1}$  downshifts in the region characteristic for metal-ligand vibrations with contributions from the cyanide ligands ( $390$ – $600\text{ cm}^{-1}$ ), when comparing samples prepared with  $^{12}CN^-$  and  $^{13}CN^-$ . This is consistent with an expected lower vibrational frequency in the presence of a heavier atom. Calculated Raman spectra of the apical  $CN^-$ -bound H-cluster models in a  $[Fe_p(n)Fe_d(n)]_H$  redox state (note: using the *DdHydAB* QM/MM model) reproduce the experimental RR spectra and  $^{13}C$  isotope shifts fairly well (Fig. S11†). An observed mode at  $603\text{ cm}^{-1}$  (experimental), assigned as a bridging CO bending mode, could only be reproduced (calculated  $605\text{ cm}^{-1}$ ) with the ADT model, suggesting the ADT ligand to be singly protonated.

### Addition of $CN^-$ to WT hydrogenases

Can these  $CN^-$ -dependent states only be formed in  $[FeFe]$  hydrogenases with a disrupted proton transfer pathway? IR spectra of artificially matured WT *DdHydAB* quite often show a band at  $1987\text{ cm}^{-1}$ ,<sup>45</sup> which has previously been difficult to assign to any known state of the H-cluster (Fig. 7A). However, we note that the vibrational frequency of this band is similar to the one of the terminal CO ligands in the  $CN^-$ -dependent  $H_{trans}$ -like state in *DdHydAB* C178A ( $1989\text{ cm}^{-1}$ ) (Fig. 2). Indeed, addition of exogenous  $CN^-$  to WT *DdHydAB* caused an increase in

intensity of this band, together with the appearance of other bands characteristic of the  $CN^-$ -dependent  $H_{trans}$ -like state (Fig. 7B).

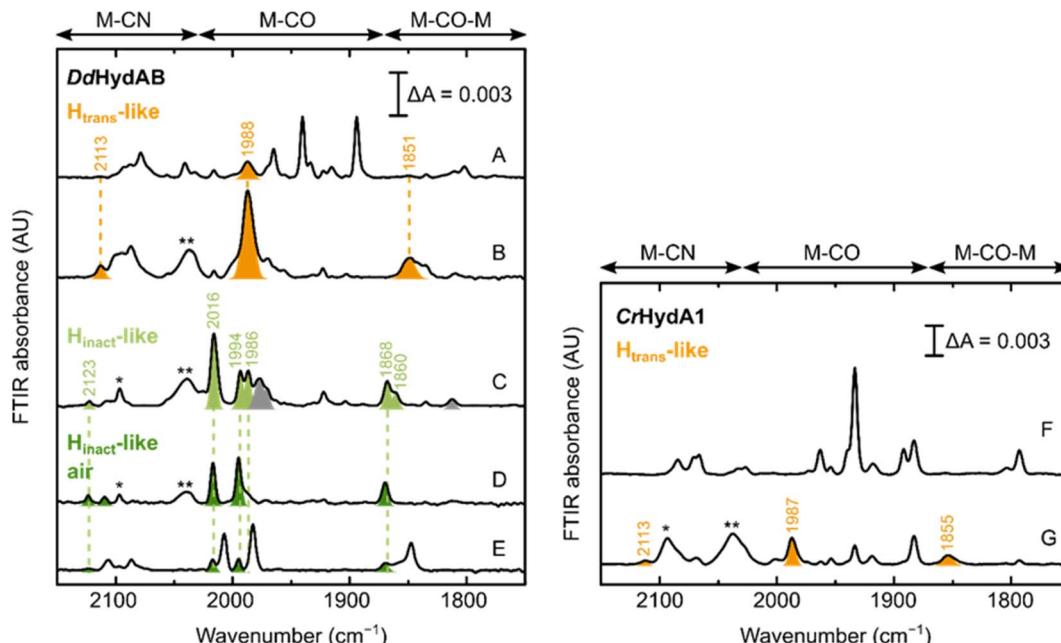
Oxidation with HAR under anaerobic conditions gives rise to multiple bands in the IR spectrum, some belonging to the  $H_{ox}$ –CO state and others similar to the  $H_{inact}$ -like state of the C178A mutant. Subsequent exposure of the enzyme to oxygen, yields a simpler IR spectrum with only an  $H_{inact}$ -like state present (slightly blue-shifted compared to the analogous state observed under anaerobic conditions, potentially reflecting a difference in the oxidation state of F-clusters, fully oxidized under aerobic conditions) (Fig. 7D). This same  $CN^-$ -dependent  $H_{inact}$ -like state is present in small amounts when WT *DdHydAB* is prepared in the  $H_{inact}$  state by addition of sulfide,<sup>2</sup> and probably derives from the small amount of the  $CN^-$ -dependent  $H_{trans}$ -like state that forms during artificial maturation of this enzyme. Addition of exogenous  $CN^-$  to WT *CrHydA1* also induces formation of a  $CN^-$ -dependent  $H_{trans}$ -like state (Fig. 7F and G). However, we noted that in this case addition of excess  $CN^-$  caused substantial degradation of the H-cluster (more than with the *CrHydA1* C169A mutant or WT *DdHydAB*). Therefore, we did not investigate the formation of the  $H_{inact}$ -like state in WT *CrHydA1* in more detail.

## Discussion

In this work, we identified two new H-cluster redox states with electronic structures similar to those of the  $H_{trans}$  and  $H_{inact}$  states.<sup>2,11,29</sup> These states have been characterized in detail *via* a combination of spectroscopic, crystallographic and computational techniques. We revealed that the  $H_{trans}$ -like and  $H_{inact}$ -like states form upon reaction of  $[FeFe]$  hydrogenases with  $CN^-$ . Isotope labelling experiments and X-ray crystallography, supported by computational calculations, suggest that  $CN^-$  binds at the open coordination site of the H-cluster and, therefore, protects it from  $O_2$  binding, as it blocks the vacant site. Unlike typical  $[FeFe]$  hydrogenase inhibitors such as CO and  $H_2S$ , which bind to the H-cluster reversibly,<sup>2,57</sup>  $CN^-$  binding appears to be irreversible, at least under the conditions studied here (pH 8). As noted earlier, purging the *DdHydAB* C178A variant with CO had no effect (Fig. S11†) and all samples are initially prepared under an atmosphere of 2%  $H_2$ , suggesting that neither CO nor  $H_2$  can effectively compete off  $CN^-$ . Although  $CN^-$  binding to the H-cluster confers air-stability to  $[FeFe]$  hydrogenases, the irreversible nature of  $CN^-$  binding does not make the formation of the  $H_{inact}$ -like state a suitable strategy to protect  $[FeFe]$  hydrogenases during aerobic handling, in contrast to the reversible formation of the  $H_2S$ -dependent  $H_{inact}$  state.<sup>2,58</sup>

For the Cys-to-Ala variants and, to a smaller extent, also for the artificially matured WT *DdHydAB*, the  $H_{trans}$ -like state could form even in the absence of exogenous  $CN^-$ . We suggest that the source of  $CN^-$  in this case derives from the degradation of the  $[2Fe]_H$  synthetic cofactor during artificial maturation. Degradation of the  $[2Fe]_H$  cofactor leads to the dissociation of the CO and  $CN^-$  ligands, which can in turn bind to the ‘intact’ H-clusters. This process of cofactor ‘cannibalization’ is a well-known source of the  $H_{ox}$ –CO state in WT enzymes,<sup>29</sup> but this is the





**Fig. 7** Formation of the  $H_{trans}$ -like and  $H_{inact}$ -like states in WT [FeFe] hydrogenase. IR spectra of WT *DdHydAB* and WT *CrHydA1* under several conditions. Left panel – *DdHydAB*: as isolated (A); with 10 mM NaCN (B); with 10 mM NaCN and 10 mM HAR (C); with 10 mM NaCN, 10 mM HAR and after exposure to air (D); after preparation of the  $H_{inact}$  state with  $Na_2S$  as described in ref. 2. Right panel – *CrHydA1*: as isolated (F) and with 10 mM NaCN. Bands from the  $H_{trans}$ -like state are colored in orange, those from the  $H_{inact}$ -like state obtained under anaerobic conditions in light green and those from the  $H_{inact}$ -like state under air in dark green. The single and double asterisk indicate HCN and  $[Fe(CN)_6]^{4-}$  respectively. In (A), the additional bands (not colored) correspond to the  $H_{ox}$  and  $H_{red}H^+$  states, with small contributions from the  $H_{ox}$ -CO and  $H_{red}$  states. A small proportion of the  $H_{trans}$ -like state is often present even in freshly-matured *DdHydAB*. In (C), the gray bands belong to the  $H_{ox}$ -CO state. In (D), the small blue-shift observed when the enzyme is exposed to oxygen after addition of NaCN and HAR ( $H_{inact}$ -like air) might be caused by oxidation of the F-clusters not already oxidized by HAR. As shown in (E), the same “ $H_{inact}$ -like air” state is routinely present as a minor component when the  $H_{inact}$  state (not colored) is prepared by addition of  $Na_2S$  and HAR,<sup>2</sup> as a result of the oxidation of the  $H_{trans}$ -like state formed during artificial maturation (shown in A). In (F), the additional bands belong to the  $H_{red}$  state, with smaller contributions from the  $H_{red}H^+$  and  $H_{sred}H^+$  states. The  $H_{red}$  and  $H_{sred}H^+$  states are present also in (G), after addition of  $CN^-$  to *CrHydA1*. Addition of  $CN^-$  to *CrHydA1* seems to induce a lot more H-cluster degradation, as demonstrated by the loss in intensity in the IR spectra (while the protein concentration was similar, 420  $\mu M$  in (F) and 500  $\mu M$  in (G)) and the presence of an intense band for  $[Fe(CN)_6]^{4-}$ .

first time that  $CN^-$  binding is also observed. The reason why *DdHydAB* enzymes (mutant and WT) form more of the  $H_{trans}$ -like state compared with *CrHydA1* is because this enzyme requires longer maturation times with a large excess of cofactor and higher temperature (see Materials and methods in the ESI†), therefore, promoting degradation of the  $[2Fe]_H$  cofactor and allowing the accumulation of more free  $CN^-$  in solution. In contrast, artificial maturation of *CrHydA1* can be achieved in one hour with only a small excess of the  $[2Fe]_H$  cofactor. Nevertheless, *DdHydAB* shows an unusually high affinity towards strong-field ligands, *i.e.* CO and  $CN^-$ , with a tendency to stabilize them in the apical position of the distal Fe ion, as is well known for the CO-inhibited state  $H_{ox}$ -CO in native *DdHydAB*. For example, Goldet *et al.* showed that *DdHydAB* had a 25-fold higher  $K_I$  (inhibition constant for CO during  $H_2$  oxidation) than *CrHydA1*, and a 330-fold higher  $K_I$  than *HydA1* [FeFe] hydrogenase from *Clostridium acetobutylicum* (*CaHydA*).<sup>57</sup> In addition, the C169A variant of *CrHydA1*,<sup>23,41,44</sup> the C299A variant of *CpHydA1*<sup>36</sup> and the C298A variant of *CaHydA*<sup>59</sup> have been produced and studied before. In none of these cases was spontaneous formation of  $CN^-$ -bound states observed during artificial maturation, further highlighting that the C178A variant of *DdHydAB* had especially

high affinity for  $CN^-$ . The  $H_{inact}$ -like crystal structure presented in this work (Fig. 4) at atomic resolution, shows a diatomic ligand at the apical position on  $Fe_d$ , which we have modelled as  $CN^-$ . The reason for this high affinity of *DdHydAB* towards strong-field ligands is not well understood and needs to be investigated further. Very recently, Duan *et al.* showed binding of  $CN^-$  to WT *CpHydA1* and *CrHydA1* using X-ray crystallography and IR spectroscopy.<sup>60</sup> Their structures show an identical binding mode for the  $CN^-$  ligand, but the authors propose additional hydrogen bonding interactions from the ADT ligand and nearby cysteine to the nitrogen of the  $CN^-$  ligand. While their IR spectra of  $CN^-$ -bound WT *CpHydA1* appear to be analogous to those from the  $H_{inact}$ -like states of *DdHydAB* C178A and *CrHydA1* C169A reported here, their IR spectrum of  $CN^-$ -bound *CrHydA1* appears to be analogous to our spectrum of *CrHydA1* C169A in the  $H_{trans}$ -like state.

Another example that illustrates the exceptional ligand binding properties of *DdHydAB* is the formation of the  $H_2S$ -dependent  $H_{inact}$  state. Previous work on WT *DdHydAB* showed that anaerobic oxidation in the presence of sulfide results in binding of  $SH_2$  to the open coordination site, forming the  $H_{inact}$  state, for which the crystal structure revealed a  $SH^-$

ligand on the apical position of  $\text{Fe}_\text{d}$ . Sulfide binding to the active site also requires an overoxidized  $[\text{2Fe}]_\text{H}$  subcluster (*i.e.*  $[\text{Fe}_\text{p}(\text{II})\text{Fe}_\text{d}(\text{II})]$ ).<sup>2,11</sup> This has been shown to occur in both *DdHydAB* and *CrHydA1*,<sup>2</sup> but much less effectively in *CpHydA1*, *CaHydA* and *MeHydA* from *Megasphaera elsdenii*.<sup>34</sup> Interestingly, during electrochemistry *DdHydAB* inactivates with a slightly more negative potential ( $E_\text{switch}$ ) than *CrHydA1* suggesting that  $\text{H}_2\text{S}$  binding is faster and  $\text{H}_2\text{S}$  release is slower for *DdHydAB* compared with *CrHydA1*.<sup>2</sup>

In both enzymes, *DdHydAB* and *CrHydA1*, the Cys-to-Ala mutation in the proton transfer pathway favors the formation of the  $\text{CN}^-$  bound states. We hypothesize that this may be related to the fact that the H-cluster in the Cys-to-Ala variants seems to be electron-deficient compared to the WT enzymes, as suggested by the blue-shifted bands observed in the IR spectra. This is also in line with previous reports on the altered  $\text{H}_\text{ox}/\text{H}_\text{red}$  thermodynamics in *CrHydA1 C169S*.<sup>22</sup> In contrast, in the *HydA1* [FeFe] hydrogenase from *Clostridium acetobutylicum*, the Cys mutation in the PTP to the ionizable residue aspartic acid seems to favor formation of  $\text{H}_\text{ox}$  over  $\text{H}_\text{red}$ .<sup>40</sup>

Why does  $\text{CN}^-$  binding favor  $\text{H}_\text{trans}$ -/ $\text{H}_\text{inact}$ -like states? While  $\text{CN}^-$  is generally considered a good  $\pi$ -acceptor, recent experimental and theoretical studies have shown that it is dominated by  $\sigma$ -donating properties with only weak  $\pi$ -accepting properties.<sup>61,62</sup> Therefore,  $\text{CN}^-$  might stabilize higher oxidation states (*e.g.* the overoxidized  $[\text{2Fe}]_\text{H}$  subcluster in the  $\text{H}_\text{trans}$ -like and  $\text{H}_\text{inact}$ -like states  $[\text{Fe}_\text{p}(\text{II})\text{Fe}_\text{d}(\text{II})]_\text{H}$ ) relative for example to CO, which is a better  $\pi$ -acceptor than  $\text{CN}^-$ . According to the IR spectra, the Cys-to-Ala variants appear more electron-deficient and, therefore, they have higher affinity for  $\text{CN}^-$  compared to CO. Although we note that the interaction between the cysteine in the PTP and the H-cluster is crucial to modulate the electronic structure at the active site, the detailed understanding of how the Cys-to-Ala mutation affects the electronic distribution at the H-cluster is beyond the scope of this work. However, we suggest that this mutation affects the hydrogen-bonding network surrounding the H-cluster, in particular concerning the ADT amine which has been shown to be involved in electronic delocalization from the two Fe ions in  $[\text{2Fe}]_\text{H}$  model compounds.<sup>63</sup> The reduced steric hindrance in the Cys-to-Ala mutants might also provide easier access to the open coordination site for the additional  $\text{CN}^-$  ligand.

Interestingly, during the synthesis of the  $[\text{2Fe}]_\text{H}$  precursor, only two of the CO ligands in  $\text{Fe}_2[(\text{SCH}_2)_2\text{NH}](\text{CO})_6$  can be substituted with  $\text{CN}^-$ .<sup>64</sup> This shows how the protein scaffold plays a crucial role in stabilizing an additional  $\text{CN}^-$  bound to  $[\text{2Fe}]_\text{H}$ . One reason could be that within the H-cluster, electron transfer from  $[\text{2Fe}]_\text{H}$  to  $[\text{4Fe-4S}]_\text{H}$  allows formation of an overoxidized binuclear site,  $[\text{Fe}_\text{p}(\text{II})\text{Fe}_\text{d}(\text{II})]_\text{H}$ , which is necessary for  $\text{CN}^-$  binding.

Our report of  $\text{CN}^-$  binding to [FeFe] hydrogenases also sheds light on the nature of previously uncharacterized active-site states. We have demonstrated that the unknown states present as impurities in artificially matured samples of WT *DdHydAB* are indeed  $\text{CN}^-$  bound states caused by the long artificial maturation of this enzyme. Previous EPR studies on *CrHydA1 C169S* have shown the formation of an unidentified

$\text{H}_\text{trans}$ -like state, exhibiting almost identical  $g$ -values to the one studied here.<sup>21,22,43</sup> Thus, it is also plausible that the previously observed  $\text{H}_\text{trans}$ -like state is caused by binding of  $\text{CN}^-$  to the H-cluster, favored by the Cys-to-Ser mutation. A recent study reported the accumulation over a long time-scale (24 h) of this very similar  $\text{H}_\text{trans}$ -like state for both the C169S variant and WT *CrHydA1* artificially matured inside *E. coli* cells, leading to the inhibition of  $\text{H}_2$  production by the culture.<sup>43</sup> We hypothesize that this  $\text{H}_\text{trans}$ -like state is also  $\text{CN}^-$ -dependent as the one described here.

Cyanide binding to metals in biology is well known, with the most classic example being cytochrome c oxidase of the mitochondrial respiratory chain, where  $\text{CN}^-$  binds between heme  $\text{a}_3$  and the  $\text{Cu}_\text{B}$  site.<sup>65,66</sup> Interestingly, in the reduced structure, with  $\text{Fe}(\text{II})$  the Fe-C distance is 2.4 Å (ref. 65) and shortens to 2.0 Å in the oxidized structure, with  $\text{Fe}(\text{III})$ ,<sup>66</sup> suggesting a shorter stronger bond. Cyanide has also been reported to bind ferric heme-proteins with a very high affinity, *e.g.* myoglobin<sup>67-69</sup> and hemoglobin.<sup>70,71</sup> In [NiFe] hydrogenase,  $\text{CN}^-$  is thought to bind transiently to the  $\text{Ni}(\text{II})$  in the  $\text{Ni-SI}_\text{a}$  state, promoting oxidation to  $\text{Ni}(\text{III})$  and formation of the  $\text{Ni-B}$  state.<sup>72</sup> In CODH,  $\text{CN}^-$  binds again to a  $\text{Ni}(\text{II})$  ion with a 1.8 Å Ni-C bond,<sup>73</sup> and inhibits CO oxidation rather than  $\text{CO}_2$  reduction suggesting that it also binds favorably to a more oxidized active site. Overall, our results are consistent with literature observations that  $\text{CN}^-$  binds preferentially to more oxidized active sites, or alternatively that binding of  $\text{CN}^-$  favors metal oxidation.

## Conclusions

Here, we have reported for the first time, a detailed spectroscopic and computational characterisation of the binding of  $\text{CN}^-$  to the active site of [FeFe] hydrogenases.  $\text{CN}^-$  binding is clearly favored in the Cys-to-Ala mutants, exemplifying the crucial role of the second coordination sphere of the H-cluster in preventing  $\text{CN}^-$  binding, and reflecting the electronic structure adaptations of the H-cluster environment to facilitate stabilization of a terminal  $\text{Fe}(\text{II})$ -hydride species during catalysis. Overall, our studies showed how the interaction between the Cys in the PTP and the ADT in  $[\text{2Fe}]_\text{H}$  tunes the electronic structure of the active site, controlling ligand binding at the open coordination site.

## Data availability

Data supporting the findings of this study are available in the article and the associated ESI files. Structural data for *DdHydAB C178A* have been deposited into the Protein Data Bank (PDB) under the following accession codes: 8BJ7 for *DdHydAB C178A* in the  $\text{H}_\text{inact}$ -like state and 8BJ8 for *DdHydAB C178A* in the  $\text{H}_\text{trans}$ -like state.†

## Author contributions

Conceptualization: M. A. M., J. A. B. and P. R.-M.; methodology: M. A. M., K. B., Y. P., C. L., N. B., I. S., R. B.; investigation: M. A. M., K. B., Y. P., C. L., C. W.; writing – original



draft: M. A. M., J. A. B and P. R.-M; writing & editing: all authors; supervision: M. A. M., I. S., R. B., I. Z., S. D., J. A. B and P. R.-M.; funding acquisition: J. A. B, P. R.-M., S. D., I. S., I. Z.

## Conflicts of interest

There are no conflicts to declare.

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## References

- 1 J. Esselborn, N. Muraki, K. Klein, V. Engelbrecht, N. Metzler-Nolte, U. P. Apfel, E. Hofmann, G. Kurisu and T. Happe, *Chem. Sci.*, 2016, **7**, 959–968.
- 2 P. Rodriguez-Macia, E. J. Reijerse, M. van Gastel, S. DeBeer, W. Lubitz, O. Rüdiger and J. A. Birrell, *J. Am. Chem. Soc.*, 2018, **140**, 9346–9350.
- 3 S. Yoshikawa, D. H. O'Keeffe and W. S. Caughey, *J. Biol. Chem.*, 1985, **260**, 3518–3528.
- 4 W. Lubitz, H. Ogata, O. Rüdiger and E. Reijerse, *Chem. Rev.*, 2014, **114**, 4081–4148.
- 5 H. Land, M. Senger, G. Berggren and S. T. Stripp, *ACS Catal.*, 2020, **10**, 7069–7086.
- 6 M. Haumann and S. T. Stripp, *Acc. Chem. Res.*, 2018, **51**, 1755–1763.
- 7 P. M. Vignais, B. Billoud and J. Meyer, *FEMS Microbiol. Rev.*, 2001, **25**, 455–501.
- 8 J. W. Peters, W. N. Lanzilotta, B. J. Lemon and L. C. Seefeldt, *Science*, 1998, **282**, 1853–1858.
- 9 Y. Nicolet, C. Piras, P. Legrand, C. E. Hatchikian and J. C. Fontecilla-Camps, *Structure*, 1999, **7**, 13–23.
- 10 B. J. Lemon and J. W. Peters, *Biochemistry*, 1999, **38**, 12969–12973.
- 11 P. Rodriguez-Macia, L. M. Galle, R. Bjornsson, C. Lorent, I. Zebger, Y. Yoda, S. P. Cramer, S. DeBeer, I. Span and J. A. Birrell, *Angew. Chem., Int. Ed.*, 2020, **59**, 16786–16794.
- 12 S. T. Stripp, G. Goldet, C. Brandmayr, O. Sanganas, K. A. Vincent, M. Haumann, F. A. Armstrong and T. Happe, *Proc. Natl. Acad. Sci. U.S.A.*, 2009, **106**, 17331–17336.
- 13 C. Lambertz, N. Leidel, K. G. V. Havelius, J. Noth, P. Chernev, M. Winkler, T. Happe and M. Haumann, *J. Biol. Chem.*, 2011, **286**, 40614–40623.
- 14 K. D. Swanson, M. W. Ratzloff, D. W. Mulder, J. H. Artz, S. Ghose, A. Hoffman, S. White, O. A. Zadvornyy, J. B. Broderick, B. Bothner, P. W. King and J. W. Peters, *J. Am. Chem. Soc.*, 2015, **137**, 1809–1816.
- 15 J. Esselborn, L. Kertess, U.-P. Apfel, E. Hofmann and T. Happe, *J. Am. Chem. Soc.*, 2019, **141**, 17721–17728.
- 16 J. A. Birrell, P. Rodriguez-Macia, E. J. Reijerse, M. A. Martini and W. Lubitz, *Coord. Chem. Rev.*, 2021, **449**, 214191.
- 17 E. J. Reijerse, V. Pelmenschikov, J. A. Birrell, C. P. Richers, M. Kaupp, T. B. Rauchfuss, S. P. Cramer and W. Lubitz, *J. Phys. Chem. Lett.*, 2019, **10**, 6794–6799.
- 18 A. Adamska, A. Silakov, C. Lambertz, O. Rüdiger, T. Happe, E. Reijerse and W. Lubitz, *Angew. Chem., Int. Ed.*, 2012, **51**, 11458–11462.
- 19 S. Katz, J. Noth, M. Horch, H. S. Shafaat, T. Happe, P. Hildebrandt and I. Zebger, *Chem. Sci.*, 2016, **7**, 6746–6752.
- 20 C. Sommer, A. Adamska-Venkatesh, K. Pawlak, J. A. Birrell, O. Rüdiger, E. J. Reijerse and W. Lubitz, *J. Am. Chem. Soc.*, 2017, **139**, 1440–1443.
- 21 D. W. Mulder, M. W. Ratzloff, M. Bruschi, C. Greco, E. Koonce, J. W. Peters and P. W. King, *J. Am. Chem. Soc.*, 2014, **136**, 15394–15402.
- 22 D. W. Mulder, Y. Guo, M. W. Ratzloff and P. W. King, *J. Am. Chem. Soc.*, 2017, **139**, 83–86.
- 23 M. Winkler, M. Senger, J. Duan, J. Esselborn, F. Wittkamp, E. Hofmann, U.-P. Apfel, S. T. Stripp and T. Happe, *Nat. Commun.*, 2017, **8**, 16115.
- 24 V. Pelmenschikov, J. A. Birrell, C. C. Pham, N. Mishra, H. Wang, C. Sommer, E. Reijerse, C. P. Richers, K. Tamasaku, Y. Yoda, T. B. Rauchfuss, W. Lubitz and S. P. Cramer, *J. Am. Chem. Soc.*, 2017, **139**, 16894–16902.
- 25 E. J. Reijerse, C. C. Pham, V. Pelmenschikov, R. Gilbert-Wilson, A. Adamska-Venkatesh, J. F. Siebel, L. B. Gee, Y. Yoda, K. Tamasaku, W. Lubitz, T. B. Rauchfuss and S. P. Cramer, *J. Am. Chem. Soc.*, 2017, **139**, 4306–4309.
- 26 C. Lorent, S. Katz, J. Duan, C. J. Kulka, G. Caserta, C. Teutloff, S. Yadav, U.-P. Apfel, M. Winkler, T. Happe, M. Horch and I. Zebger, *J. Am. Chem. Soc.*, 2020, **142**, 5493–5497.
- 27 A. J. Pierik, W. R. Hagen, J. S. Redeker, R. B. Wolbert, M. Boersma, M. F. Verhagen, H. J. Grande, C. Veeger, P. H. Mutsaers, R. H. Sands, *et al.*, *Eur. J. Biochem.*, 1992, **209**, 63–72.
- 28 D. S. Patil, J. J. Moura, S. H. He, M. Teixeira, B. C. Prickril, D. V. DerVartanian, H. D. Peck Jr, J. LeGall and B. H. Huynh, *J. Biol. Chem.*, 1988, **263**, 18732–18738.
- 29 W. Roseboom, A. L. De Lacey, V. M. Fernandez, E. C. Hatchikian and S. P. J. Albracht, *JBIC, J. Biol. Inorg. Chem.*, 2006, **11**, 102–118.
- 30 S. P. Albracht, W. Roseboom and E. C. Hatchikian, *J. Biol. Inorg. Chem.*, 2006, **11**, 88–101.



31 S. Morra, M. Arizzi, F. Valetti and G. Gilardi, *Biochemistry*, 2016, **55**, 5897–5900.

32 P. S. Corrigan, J. L. Tirsch and A. Silakov, *J. Am. Chem. Soc.*, 2020, **142**, 12409–12419.

33 M. Winkler, J. Duan, A. Rutz, C. Felbek, L. Scholtysek, O. Lampret, J. Jaenecke, U.-P. Apfel, G. Gilardi, F. Valetti, V. Fourmond, E. Hofmann, C. Léger and T. Happe, *Nat. Commun.*, 2021, **12**, 756.

34 C. Felbek, F. Arrigoni, D. de Sancho, A. Jacq-Bailly, R. B. Best, V. Fourmond, L. Bertini and C. Léger, *ACS Catal.*, 2021, **11**, 15162–15176.

35 G. Hong, A. J. Cornish, E. L. Hegg and R. Pachter, *Biochim. Biophys. Acta Bioenerg.*, 2011, **1807**, 510–517.

36 J. Duan, M. Senger, J. Esselborn, V. Engelbrecht, F. Wittkamp, U.-P. Apfel, E. Hofmann, S. T. Stripp, T. Happe and M. Winkler, *Nat. Commun.*, 2018, **9**, 4726.

37 M. Senger, V. Eichmann, K. Laun, J. Duan, F. Wittkamp, G. Knör, U.-P. Apfel, T. Happe, M. Winkler, J. Heberle and S. T. Stripp, *J. Am. Chem. Soc.*, 2019, **141**, 17394–17403.

38 A. J. Cornish, K. Gärtner, H. Yang, J. W. Peters and E. L. Hegg, *J. Biol. Chem.*, 2011, **286**, 38341–38347.

39 P. Knorzer, A. Silakov, C. E. Foster, F. A. Armstrong, W. Lubitz and T. Happe, *J. Biol. Chem.*, 2012, **287**, 1489–1499.

40 S. Morra, A. Giraudo, G. Di Nardo, P. W. King, G. Gilardi and F. Valetti, *PLoS One*, 2012, **7**, e48400.

41 S. Rumpel, C. Sommer, E. Reijerse, C. Fares and W. Lubitz, *J. Am. Chem. Soc.*, 2018, **140**, 3863–3866.

42 C. C. Pham, D. W. Mulder, V. Pelmenschikov, P. W. King, M. W. Ratzloff, H. Wang, N. Mishra, E. E. Alp, J. Zhao, M. Y. Hu, K. Tamasaku, Y. Yoda and S. P. Cramer, *Angew. Chem., Int. Ed.*, 2018, **57**, 10605–10609.

43 M. Lorenzi, P. Ceccaldi, P. Rodriguez-Macia, H. J. Redman, A. Zamader, J. A. Birrell, L. S. Meszaros and G. Berggren, *J. Biol. Inorg. Chem.*, 2022, **27**, 345–355.

44 S. Mebs, R. Kositzki, J. Duan, L. Kertess, M. Senger, F. Wittkamp, U. P. Apfel, T. Happe, S. T. Stripp, M. Winkler and M. Haumann, *Biochim. Biophys. Acta Bioenerg.*, 2018, **1859**, 28–41.

45 J. A. Birrell, K. Wrede, K. Pawlak, P. Rodriguez-Macia, O. Rüdiger, E. J. Reijerse and W. Lubitz, *Isr. J. Chem.*, 2016, **56**, 852–863.

46 P. Rodriguez-Macia, K. Pawlak, O. Rüdiger, E. J. Reijerse, W. Lubitz and J. A. Birrell, *J. Am. Chem. Soc.*, 2017, **139**, 15122–15134.

47 M. Senger, S. Mebs, J. Duan, F. Wittkamp, U.-P. Apfel, J. Heberle, M. Haumann and T. Stripp Sven, *Proc. Natl. Acad. Sci. U.S.A.*, 2016, **113**, 8454–8459.

48 C. Lorent, V. Pelmenschikov, S. Frielingsdorf, J. Schoknecht, G. Caserta, Y. Yoda, H. Wang, K. Tamasaku, O. Lenz, S. P. Cramer, M. Horch, L. Lauterbach and I. Zebger, *Angew. Chem., Int. Ed.*, 2021, **60**, 15854–15862.

49 J. A. Birrell, V. Pelmenschikov, N. Mishra, H. Wang, Y. Yoda, K. Tamasaku, T. B. Rauchfuss, S. P. Cramer, W. Lubitz and S. DeBeer, *J. Am. Chem. Soc.*, 2020, **142**, 222–232.

50 M.-E. Pandelia, H. Ogata, L. J. Currell, M. Flores and W. Lubitz, *JBIC, J. Biol. Inorg. Chem.*, 2009, **14**, 1227–1241.

51 G. Caserta, V. Pelmenschikov, C. Lorent, A. F. Tadjoung Waffo, S. Katz, L. Lauterbach, J. Schoknecht, H. Wang, Y. Yoda, K. Tamasaku, M. Kaupp, P. Hildebrandt, O. Lenz, S. P. Cramer and I. Zebger, *Chem. Sci.*, 2021, **12**, 2189–2197.

52 Y. Nicolet, A. L. de Lacey, X. Vernède, V. M. Fernandez, E. C. Hatchikian and J. C. Fontecilla-Camps, *J. Am. Chem. Soc.*, 2001, **123**, 1596–1601.

53 M. Senger, S. Mebs, J. Duan, O. Shulenina, K. Laun, L. Kertess, F. Wittkamp, U.-P. Apfel, T. Happe, M. Winkler, M. Haumann and S. T. Stripp, *Phys. Chem. Chem. Phys.*, 2018, **20**, 3128–3140.

54 M. Senger, K. Laun, F. Wittkamp, J. Duan, M. Haumann, T. Happe, M. Winkler, U.-P. Apfel and S. T. Stripp, *Angew. Chem., Int. Ed.*, 2017, **56**, 16503–16506.

55 M. A. Martini, O. Rüdiger, N. Breuer, B. Nöring, S. DeBeer, P. Rodriguez-Macia and J. A. Birrell, *J. Am. Chem. Soc.*, 2021, **143**, 18159–18171.

56 A. Ciaccafava, D. Tombolini, L. Domnik, J. Fesseler, J.-H. Jeoung, H. Dobbek, M. A. Mroginski, I. Zebger and P. Hildebrandt, *Chem. Sci.*, 2016, **7**, 3162–3171.

57 G. Goldet, C. Brandmayr, S. T. Stripp, T. Happe, C. Cavazza, J. C. Fontecilla-Camps and F. A. Armstrong, *J. Am. Chem. Soc.*, 2009, **131**, 14979–14989.

58 A. A. Oughli, S. Hardt, O. Rüdiger, J. A. Birrell and N. Plumeré, *Chem. Commun.*, 2020, **56**, 9958–9961.

59 T. Lautier, P. Ezanno, C. Baffert, V. Fourmond, L. Cournac, J. C. Fontecilla-Camps, P. Soucaille, P. Bertrand, I. Meynial-Salles and C. Léger, *Faraday Discuss.*, 2011, **148**, 385–407.

60 J. Duan, A. Hemschemeier, D. J. Burr, S. T. Stripp, E. Hofmann and T. Happe, *Angew. Chem., Int. Ed.*, 2023, **62**(7), e202216903.

61 N. Levin, S. Peredkov, T. Weyhermüller, O. Rüdiger, N. B. Pereira, D. Grötzsch, A. Kalinko and S. DeBeer, *Inorg. Chem.*, 2020, **59**, 8272–8283.

62 S. K. Singh, J. Eng, M. Atanasov and F. Neese, *Coord. Chem. Rev.*, 2017, **344**, 2–25.

63 Ö. F. Erdem, M. Stein, S. Kaur-Ghumaan, E. J. Reijerse, S. Ott and W. Lubitz, *Chem.-Eur. J.*, 2013, **19**, 14566–14572.

64 M. Schmidt, S. M. Contakes and T. B. Rauchfuss, *J. Am. Chem. Soc.*, 1999, **121**, 9736–9737.

65 K. Muramoto, K. Ohta, K. Shinzawa-Itoh, K. Kanda, M. Taniguchi, H. Nabekura, E. Yamashita, T. Tsukihara and S. Yoshikawa, *Proc. Natl. Acad. Sci. U. S. A.*, 2010, **107**, 7740–7745.

66 N. Yano, K. Muramoto, M. Mochizuki, K. Shinzawa-Itoh, E. Yamashita, S. Yoshikawa and T. Tsukihara, *Acta Crystallogr., Sect. F: Struct. Biol. Cryst. Commun.*, 2015, **71**, 726–730.

67 Y. Dou, J. S. Olson, A. J. Wilkinson and M. Ikeda-Saito, *Biochemistry*, 1996, **35**, 7107–7113.

68 M. Bolognesi, C. Rosano, R. Losso, A. Borassi, M. Rizzi, J. B. Wittenberg, A. Boffi and P. Ascenzi, *Biophys. J.*, 1999, **77**, 1093–1099.

69 P. Ascenzi, A. di Masi, F. Gullotta, M. Mattu, C. Ciaccio and M. Coletta, *Biochem. Biophys. Res. Commun.*, 2010, **393**, 196–200.



70 M. Milani, Y. Ouellet, H. Ouellet, M. Guertin, A. Boffi, G. Antonini, A. Bocedi, M. Mattu, M. Bolognesi and P. Ascenzi, *Biochemistry*, 2004, **43**, 5213–5221.

71 A. Boffi, E. Chiancone, E. S. Peterson, D. L. Rousseau and J. M. Friedman, *Biochemistry*, 1997, **36**, 4510–4514.

72 S. V. Hexter, M.-W. Chung, K. A. Vincent and F. A. Armstrong, *J. Am. Chem. Soc.*, 2014, **136**, 10470–10477.

73 J.-H. Jeoung and H. Dobbek, *J. Am. Chem. Soc.*, 2009, **131**, 9922–9923.

