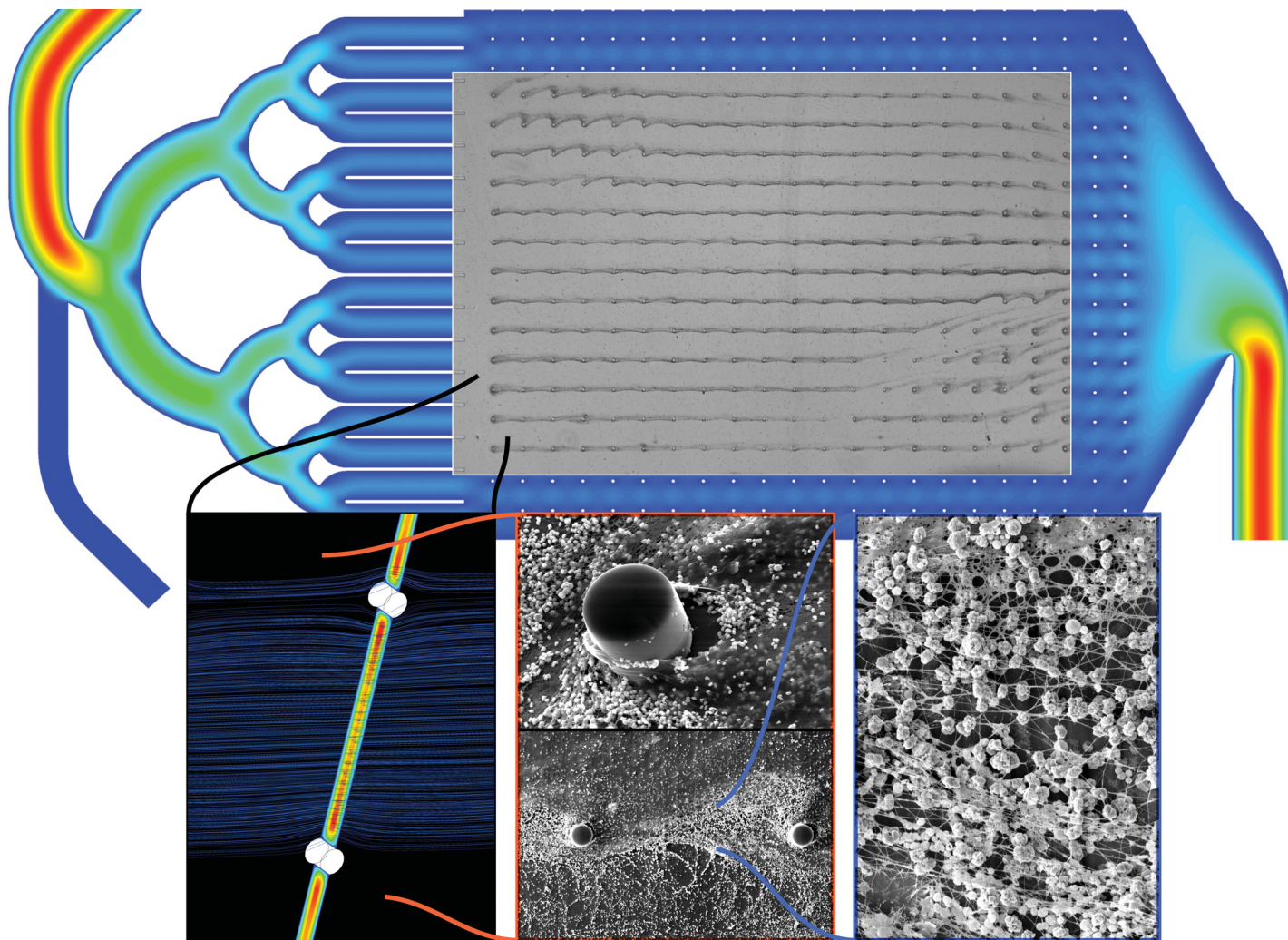




Showcasing research from Daniel Van Opdenbosch's group, Chair for Biogenic Polymers, Technical University of Munich

Grow with the flow – observing the formation of rheotactically patterned bacterial cellulose networks

In this work, the flow-dependent growth dynamics of microbial exopolysaccharide networks in obstacle-studded microfluidic cells are tracked by shadowgraphy. Our setup allows quantifying the material density as a function of time and location with almost single microbial cell resolution. In this manner, an overall bi-logistic growth and local wall shear stress-dependent material formation around obstacles were determined.

As featured in:



See Daniel Van Opdenbosch *et al.*, *Mater. Adv.*, 2022, **3**, 6191.

PAPER

[View Article Online](#)
[View Journal](#) | [View Issue](#)Cite this: *Mater. Adv.*, 2022,
3, 6191Received 2nd February 2022,
Accepted 17th May 2022

DOI: 10.1039/d2ma00115b

rsc.li/materials-advancesGrow with the flow – observing the formation of
rheotactically patterned bacterial cellulose
networks†Moritz Klotz,  Dardan Bajrami  and Daniel Van Opdenbosch*

The use of matrix-forming organisms is a promising way to obtain hierarchically structured materials. In this work, we investigate the rheotactic bio-mediated alignment and network formation of bacterial cellulose streamers, resulting in hierarchically patterned textiles. For this purpose, we devised a panoramic-view observation setup for a microfluidic cell. We observed a split growth, with first, a flow-aligned streamer formation and second, a streamer connection phase. The growth phase parameters showed dependence on flow rate. After aligned growth, a mineralisation step was conducted to the grown textiles with calcium carbonate and -sulfate to obtain nature-inspired organic/inorganic composite materials. Hereby, an anisotropic deposition of the gypsum phase due to the prealigned bacterial cellulose substrate was observed.

1 Introduction

The growth of natural materials differs fundamentally from the production of engineering materials: while engineering materials' requirements are typically fulfilled by choice of substance and shaping techniques, nature forms its materials from readily available elements into hierarchical structures.¹ Hence, biological materials derive their widely varying properties mainly from the adaptation of internal structuring to meet functional requirements.^{2,3} The most prominent aspects hereof are hierarchical structure and structural anisotropy. In addition, dispersed polymer phases enable stress relaxation and structural regression of the material due to their viscoelasticity, sacrificial bonds and 'hidden lengths'.⁴

In order to obtain bio-mimetic mechanical properties in engineering materials, different approaches can be pursued: applying biomorphism and biotemplating, it is possible to convert a naturally grown template into an inorganic replica material retaining most, or all, of the hierarchical levels of structuring, leading to exciting mechanical properties.^{5–8} With some restrictions, it is also possible to obtain hierarchical structures by the self-organizing material synthesis in an artificial process.⁹ However, up-scaling these processes, which were reviewed by Deuerling *et al.* as 'Level 1 – Transfer of

structure',¹⁰ to produce competitive engineering materials remains a challenge.

Nature, on the other hand, readily produces large-scale composite materials *via* hierarchical structuring. Therefore, bio-mediated material structuring, reviewed by Deuerling *et al.* as 'Level 2 – Usage of biological systems',¹⁰ offers the potential to control material formation from the micro to the macro level. Here, biological systems are influenced by stimuli to which the living cells and microbes react by movement, 'taxis'. The growth into a certain culture medium as an internal stimulus,¹⁰ as well as the directed growth after external stimuli, *e.g.* light,^{11–13} mechanical influences^{14–17} and a variety of other attractants,^{18–20} can serve to create a biological, hierarchically structured template to be used for different purposes than intended by nature. The advantage of this approach is that biological materials are typically already hierarchically structured, starting at the molecular level: Biofilms, for example, are micro-organisms embedded in their extracellular polymeric matrix (EPS).²¹ While they are undesirable in most living and technical systems, they feature a hierarchical structure.^{22,23} Specifically, bacterial cellulose biofilms (BCB) consisting mainly of bacterial cellulose (BC), exhibit several structural levels: starting from the cellulose molecules, the bacteria produce microfibrils at the nanometer level which consist of amorphous and crystalline regions. The microfibrils arrange into ribbons, building bundles, which then cluster into bacterial cellulose networks.^{24,25}

Several works have considered bio-mediated structuring of BCB, respectively the BC fibers: Kondo *et al.* and Uraki *et al.* similarly used already-deposited cellulose strands as templates for further deposition.^{16,26} Anisotropic deposition of BC is

Technical University of Munich, Campus Straubing for Biotechnology and Sustainability, Chair for Biogenic Polymers, Schulgasse 16, D-94315 Straubing, Germany. E-mail: moritz.klotz@tum.de

† Electronic supplementary information (ESI) available. See DOI: <https://doi.org/10.1039/d2ma00115b>



possible by using nematic ordered cellulose (NOC) or stretched cellulose as templates.^{27,28} Also of interest is the alignment *via* growth on polydimethylsiloxane (PDMS) with a ridged morphology.^{29–31} By varying ridge sizes, Putra *et al.* archived a 2.3 times higher fracture stress and a 3.8 times lower elastic modulus, compared to air-grown BCB. His group was also able to produce mechanically anisotropic behavior with BCB grown in thin silicon tubes, as well as by influencing the culture interface *via* silicon oil.^{32,33} Also promising are the works of Sano *et al.* and Liu *et al.* who successfully used an electromagnetic field to align BCB.^{34,35}

Wan *et al.* and Luo *et al.* carried out structuring by a flow approach: Wan *et al.* were able to align the BCB with the flow and increase the tensile strength in the BC fiber direction from 0.9 MPa to 1.2 MPa, while the fracture strain decreased by 8%. Luo *et al.* even achieved a tensile strength of 1.5 MPa with a flow velocity of 150 mm^{−1}, with a fracture strain decrease of 12%.^{36,37} Using flow-producing rising bubbles, Chae *et al.* also attained a unidirectional deposition of BCB.³⁸

Since structuring of BCB by flow has already yielded interesting results, it is worth considering the mechanism: if a biofilm grows in a flowing medium, its viscoelastic morphology responds to the mechanical stimulus of the flow.³⁹ The form thereby differs if the biofilm is fully surface attached or if it adheres at least at one point, while the rest of the biofilm moves with the flow as a so-called streamer. The streamer formation and its physics have been studied by several research groups.^{40–43} As intuitively expected, the geometry and the flow velocities of the system have a great influence on the streamers' shapes.^{15,44–47} Interestingly, whether streamers start to form is largely independent of the Reynolds number, as summarized by Karimi *et al.*⁴¹ The dynamics of the flow can also influence the growth system, as can be seen in experiments with pulsative or split flow.^{48,49} Condensed, the streamers morphology but also the strength of attachment are strongly influenced by their viscoelastic behavior⁵⁰ which is a function of its structure. The structure formation in turn is influenced by the flow: in contrast to fish, copepods and some protists, which possess a receptor for flow forces and exhibit active rheotaxis, bacteria react in a passive way to the shear stress factor, comparable with other passive physical movement phenomena like gyro- and magnetotaxis.⁵¹ Bacterial rheotaxis is triggered by bacterial motility and morphology.⁵² While the swimming direction of motile bacteria is changed by torque, non-motile bacteria expand along their biofilm, due to the predominant shear forces under flow conditions.^{47,53} For BC producing bacteria, it is likely that extrusion of BC and cell division based fiber-anchoring²⁴ provide this movement. Together with the morphology induced alignment, which occurs for ellipsoidal bacteria like *Komagataeibacter xylinus* (*K. xylinus*),^{37,50} we consider the flow based forming of the BCB as a type of rheotaxis. So far, rheotaxis and streamer formation has been studied mainly in single, isolated samples. The formation of streamer networks has been investigated rarely, and mainly with the aim of being able to avoid them, and the associated clogging of fluidic systems. Worth mentioning is the work by

Hassanpourfard *et al.* who investigated fast-formed *Pseudomonas fluorescens* streamer-networks and found that they still moved by a viscoelastic 'stick-slip' dynamic, which allows the formation of channels, resulting in regions of varying density.^{54,55}

In summary, the *in situ* structuring and alignment of biofilm streamers, in particular of BCB, can be realised by a variety of approaches. However, in order to produce engineering materials, processes have to be adapted to include larger-scale hierarchical structuring and outer shapes, such as honeycomb structures.^{16,26} At this point, rheotactic structuring has advantages: flow bodies can have any shape and size and there is a high degree of flexibility due to the large number of parameters such as flow, species and medium composition. Another advantage is the direct introduction of additives into the growth system, which allows the formation of organic/inorganic composite *via* an *in situ* or quasi *in situ* mineralisation route.

Generally, composites with BCB, respectively, BC as the substrate are of interest, especially for medical purposes, as the reviews from Shan *et al.* and Liu *et al.* illustrate. Mainly for the investigation of new tissue materials, dozens of BC combinations with polymers or nanomaterials were tested.^{56,57} Excellent mechanical properties were obtained by impregnation of the grown BC *ex situ*.⁵⁸ However, *in situ* mineralisation by which the composites are produced during BCB growth, or quasi *in situ* mineralisation, where the BCB is treated in the same vessel directly after growth, are promising. Literature thereof, in the combination organic/inorganic, so far is limited to (nano)particle,^{59–61} fibers^{35,62} and minerals *e.g.* hydroxyapatite or clay.^{63–66} Inspired by biological materials with tough or self-healing properties like nacre,⁶⁷ we assume that rather the latter combination is suitable to obtain a scalable, hierarchically and nature oriented engineering material. Materials which can be processed by a wet mineralisation route in a quasi *in situ* setup.

CaCO₃/cellulose composites, serving as a model for quasi *in situ* mineralisation, can be achieved by different approaches, such as the already mentioned direct deposition of particles,^{59,60} or *via* the precipitation from precursors. The latter works *via* diffusive processes⁶⁸ or faster, *via* direct mixing. The intermediate which occurs during the precipitation of CaCO₃ is amorphous calcium carbonate (ACC), which can serve as the inorganic phase after stabilization^{69–71} or after crystallisation.^{72–75} ACC can be used in a one step process, without the need for a second reagent. Also a one step process is the deposition of gypsum from super-saturated solutions. Despite gypsum's low cost and the possibility to overcome drawbacks such as brittleness, gypsum/cellulose composites have not been the focus of many publications so far. However, due to the possibility of a one-step mineralisation and its typical aceros morphology, which lends itself to textured mineralisation, it is well suited for use with prealigned BCB. Gypsum mineralisation on a cellulose substrate was already achieved by Nissinen *et al.*, who used supersaturated CaCl₂–Na₂SO₄ solutions.⁷⁶

Bio-mediated materials with hierarchical structures can be made by rheotactic approaches as demonstrated by Wan *et al.*



and Luo *et al.*^{36,37} Building on these results, we aim to extend the fundamental knowledge of flow based material patterning for future upscaling and material production. Investigating the growth dynamics leads to an improved understanding of the rheotactic biofilm formation and enables prospective reproducible material production. We hypothesized that an initial homogeneous distribution of EPS-forming microbes on obstacles, positioned with a long-range order and subjected to volume-driven flow, will lead to their uniform growth, resulting in a network of streamers. The growth parameters thereby are a key aspect to tailor a well defined material with specific structural features on several hierarchical levels. Such a material can serve as a matrix for organic/inorganic composites whose hierarchical characteristics span its different phases. Therefore, we evaluated in a panoramic-view observation setup the obstacle-induced streamer and streamer-network formation of *K. xylinus* at three different flow rates. After growth, we assessed two potential mineralisation routes by a quasi *in situ* process: in a one step process either the amorphous precursor ACC or a supersaturated solution of CaSO_4 was added directly into the growth chamber.

2 Experimental

2.1 Setup

Fig. 1 shows the experimental setup for the growth experiments. A batch culture (1) of *K. xylinus* was pumped by a syringe pump (2) through a pressure sensor (3) and a double bubble trap (4, 5) into a microfluidic cell (8) in a circular flow. Growth in the microfluidic cell was observed using an optical setup (6–10).

Komagataeibacter xylinus. The *K. xylinus* strain DSM 2004 was obtained from the Deutsche Sammlung von Mikroorganismen und Zellkulturen (DSMZ, Braunschweig, Germany). For each of the 12 experiments, a batch culture ((1) in Fig. 1) of 150 ml DSMZ medium 105 containing 100 g l^{-1} glucose (VWR International, Darmstadt, Germany), 10 g l^{-1} yeast extract (VWR International, Darmstadt, Germany) and 20 g l^{-1} CaCO_3 (Carl Roth, Karlsruhe, Germany) at pH 6,8 was prepared in a 250 ml cone-shaped flask with a $0.2 \mu\text{m}$ PTFE membrane lid (Duran, Schott, Mitterteich, Germany), and inoculated from *K. xylinus* grown on an agar plate (15 g l^{-1} agar, VWR International, Darmstadt, Germany). BC producing bacteria like *K. xylinus* are located almost exclusively in the BCB pellicle. To maintain a sufficient concentrated inoculum from below the pellicle, before use, inoculated cultures were stored at ambient temperature until the growth of the pellicle stagnated at a thickness of approximately 5 mm. The stagnation could be attributed to the conical shape of the used flasks.⁷⁷ Preliminary tests showed that glucose was sufficiently present and BCB production was unaffected by this storage time. Purity was checked before and after each experiment by streaking the *K. xylinus* culture on agar plates.

Microfluidic cell. For each experiment, a new microfluidic cell ((8) in Fig. 1) was cast from PDMS (SYLGARD 184 Silicone Elastomer Kit, Mixture Silicone/Initiator 1/10, Dow Chemical, Midland, United States) via the MicCell Tool Kit (GeSiM,

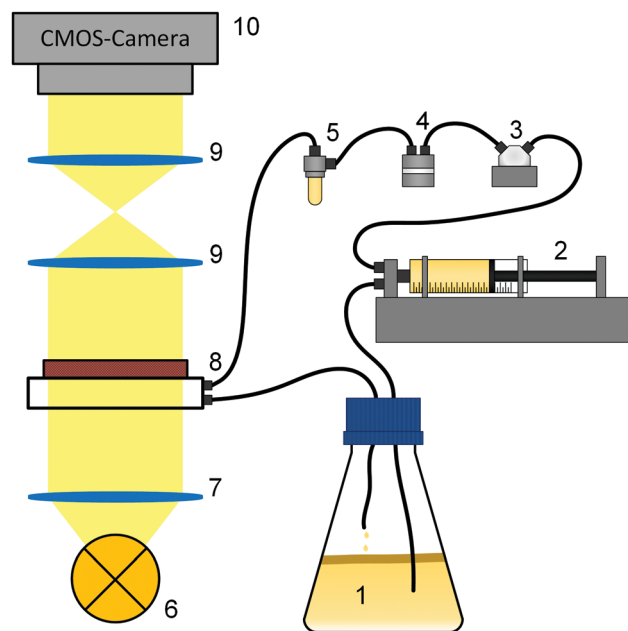


Fig. 1 Sketch of the experimental setup. The inoculum (1) of *K. xylinus* was taken from below the BCB pellicle by a syringe pump (2), injected via a pressure sensor (3) and a double bubble trap (4, 5) into the microfluidic cell (8), and recycled into the batch culture (1). The microfluidic cell was illuminated in transmission with nearly parallel light (7) emitted by a white LED (6). After passing an optional optical filter which allows schlieren techniques (9, not used in the current experiments), the observation took place using a camera (10). For clarity, the beam path is simplified.

Radeberg, Germany). After casting and cross-linking for 24 h at 45°C , the sealing glass was permanently connected to the PDMS by a 45 s plasma treatment. The microfluidic cell, shown in Fig. 2, with its polycarbonate carrier and sealing glass on the PDMS was transparent, allowing observation of the $50 \mu\text{m}$ channel in transmission.

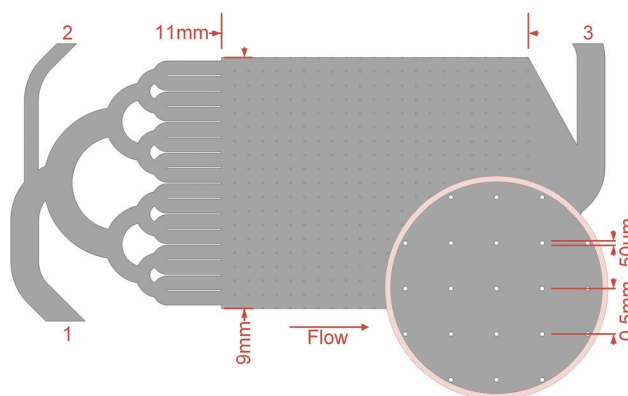


Fig. 2 Sketch of the microfluidic cell. The inoculum was injected into the larger inlet (1) from where it was evenly distributed into the inlet channels. This regularly allowed an even distribution of the fluid into the growth chamber, which was filled with evenly arranged obstacles with diameters of $50 \mu\text{m}$ in 0.5 mm distance. After the growth, the smaller inlet (2) was used to add mineralisation medium. The inset shows a magnified section of the growth chamber.



Optical setup. In the optical setup ((6–10) in Fig. 1) the microfluidic cell was illuminated (6) by a white LED without diffusor (QAUR1361L030, Signal-Construct, Niefern-Oeschelbronn, Germany). After passing the convex lens (7), the slightly divergent light passed the microfluidic cell (8). The optical filter (9) with $f = 100$ mm, while part of the setup, was not used in this experiment. The microfluidic cell was imaged on the $13.1 \text{ mm} \times 8.8 \text{ mm}$ camera chip (10) slightly magnified ($1.28 \times$) with $5472 \text{ px} \times 3648 \text{ px}$ (acA5472-17um, Basler, Ahrensburg, Germany), resulting in a viewing window of $10.2 \text{ mm} \times 6.8 \text{ mm}$ and a resolution of $7 \mu\text{m}$ (144 line pairs per millimeter, measured with USAF 1951 Target).

2.2 Experimental procedure

For each of the three different flow rates, four repeating experiments were conducted at ambient temperature $21 \pm 1^\circ\text{C}$. The necessary volume flow for the mean flow rates of 1.0 mm s^{-1} , 12.5 mm s^{-1} and 25.0 mm s^{-1} were calculated for the growth chamber of the microfluidic cell.

Before each of the 12 experiments, all tubes (PTFE, inner diameter $1/32''$), fittings and the second bubble trap were autoclaved. After assembly of the components, the setup was flushed with 20% H_2O_2 (Carl Roth, Karlsruhe, Germany) for one hour to sterilize all unautoclavable parts including the microfluidic cell, both bubble traps, the valve and the pressure sensor (CPS 2184 Z, 5 bar, CETONI, Korbussen, Germany).

At the start of an experiment, a syringe pump (neMESYS 290N with QmixElements, CETONI, Korbussen, Germany) equipped with a 3-2-way valve ((2) in Fig. 1), pumped inoculum from below the BCB pellicle in cycles through the microfluidic cell and back into the flask. During the experiments, which lasted up to 70 h, the syringes were automatically refilled. All pump parameters are listed in Table 1.

Since the long-lasting experiments were susceptible to air bubbles which degrade the grown biofilm, a double bubble trap ((4,5) Fig. 1) was used. The first (Bubble Trap for Microfluidics, ELVEFLOW, Paris, France) for large bubbles appearing in the starting phase and the second (Microfluidic Reservoir XS, ELVEFLOW, Paris, France), for small bubbles appearing during the experiments, which were not removable by the first trap. Additionally, the second bubble trap buffered pressure impulses and allowed a more even flow. With the pressure sensor, a servo loop was adjusted. The experiments were conducted until the slowly increasing pressure reached 1 bar and the servo loop started to minimize the flow rate from Table 1. For the evaluation, only the data before the start of the

servo loop with the constant adjusted flow (below 1 bar) were used. To document the growth, 8-bit black/white images were recorded *via* Micro-Manager⁷⁸ every 10 minutes.

After each experiment, the setup was flushed with 5–10 ml deionised water (DI) to remove all water-soluble substances. For this, the maximum pressure was step-wise increased, up to a maximum of 1.5 bar. After the following mineralisation steps which were conducted on some samples, the microfluidic cells were dried for 24 h at 45°C and opened mechanically to obtain the biofilm.

2.3 Growth analysis

BCB concentration calibration. To relate the BCB transmitted light intensities imaged during growth to BCB concentrations, the Lambert–Beer law $A = \epsilon l_{\text{cell}} c$ was used. A accounts for all attenuating phenomena, including absorbance and scattering. ϵ is the molar attenuation coefficient, l_{cell} the path length of $50 \mu\text{m}$ (chamber thickness) and c the BCB concentration.

For determining ϵ , the biofilm thickness l_{BCB} of one reference sample was obtained by breaking the sample after freezing in liquid nitrogen. l_{BCB} then was determined by SEM at 19 measuring points at the fracture plane. We assume that the vacuum-dried BCB was contracted compact and dense with marginal inter-spaces. This is supported by the SEM images (see Fig. S3(d), ESI†). Since only BCB influenced light absorbance due to the use of the reference images at t_0 , we defined the reference mass concentration c_{ref} with the measured BCB density ρ_{BCB} by eqn (1). To measure ρ_{BCB} , in total 48 BCB pieces of six separate batch-grown samples were washed, dried at 65°C and weighed. Afterwards, they were soaked for 24 h in DI water at <0.1 bar to remove embedded air. Their densities were determined by Archimedes' principle (224-1S, Sotoris Entris, Göttingen, Germany) with a mean result of $\rho_{\text{BCB}} = 1.12 \pm 0.03 \text{ g cm}^{-3}$.

$$c_{\text{ref}} = \frac{\rho_{\text{BCB}} l_{\text{BCB}}}{l_{\text{cell}}} \quad (1)$$

With c_{ref} , l_{cell} and the reference absorbances A_{ref} , ϵ was determined as $\epsilon = (1.30 \pm 0.27) \times 10^5 \text{ mm}^2 \text{ g}^{-1}$, see Fig. S1 (ESI†). Thereby A_{ref} was obtained by eqn (2), with t_0 as the first image without biofilm and t as the last taken image during the growth experiment.

$$A(i, j, t) = \log_{10} \frac{I(i, j, t_0)}{I(i, j, t)} \quad (2)$$

BCB concentration. With the determined ϵ , all c_{BCB} values were calculated for all the measurement data *via* the Lambert–Beer law, eqn (3). Pixel values i, j stayed within the 8-bit range of the images. This showed that no saturation was reached during the experiments and all calculated concentrations were within the measuring range.

$$c_{\text{BCB}}(i, j, t) = \frac{A(i, j, t)}{\epsilon l_{\text{cell}}} \quad (3)$$

Statistical analysis. For the evaluation of biofilm growth in the function of the three flow rates 1.0 mm s^{-1} , 12.5 mm s^{-1} and 25.0 mm s^{-1} , the borders of the viewing window were cut

Table 1 Pump parameters with the calculated mean flow rates in the growth chamber of the microfluidic cell. Syringe volumes differed to maintain a precise flow; the refill rate differed to avoid formation of bubbles in the syringes

Flow rate/ (mm s^{-1})	Syringe inject rate/($\mu\text{l s}^{-1}$)	Syringe volume/ml	Syringe refill rate/($\mu\text{l s}^{-1}$)
1.0	0.450	1	50
12.5	5.625	2	75
25.0	11.25	10	100



off to a section of $9 \times 6 \text{ mm}^2$, so that the identical sector inside of the growth chamber was analyzed in every experiment. Normalised cumulative histograms $\gamma(c_{\text{BCB},b}, t)$ of the obtained BCB concentrations were calculated to determine proportions of the material with concentrations of c_{BCB} or higher. Hence, values of γ represent the area fractions where the minimum concentrations are c_{BCB} . All 210 bins $c_{\text{BCB},b}$ in their logarithmic concentration range, ranging from -0.010 up to 0.347 g cm^{-3} , were summarised starting by the highest occurring bin $c_{\text{BCB},b_{\text{max}}}$.

After conversion, the isolines of $\gamma(c_{\text{BCB},b}, t)$ were fitted between $0 < \gamma \leq 1$ via a bi-logistic fit, which was performed by the least-squares algorithm of the Python Imfit package. With the maximum curve values S , the inflexion points Δt of the sigmoid curves and the logistic growth constants λ , fits were performed after eqn (4). The plots shown in this work ended when the fitted parameter of at least two measurements reached zero or unrealistic values.

$$c_{\text{BCB}}(\gamma, t) = \frac{S_1}{1 + e^{-\lambda_1(t-\Delta t_1)}} + \frac{S_2}{1 + e^{-\lambda_2(t-\Delta t_2)}} \quad 0 < \gamma \leq 1 \quad (4)$$

Cumulative analysis. From the $c_{\text{BCB}}(i,j,t)$ data, the average BCB concentrations $\bar{c}_{\text{BCB}}(i,j,t)$ were calculated. For determining the concentration increase rates, the resulting mean data of the three flow rates were fitted by linear regression via the Python SciPy theilslopes Theil-Sen estimator.

Azimuthal analysis. To obtain the average development of the concentration around the obstacles for every experiment, the azimuthal BCB concentrations $c_{\text{BCB}}(\alpha, t)$ were determined by calculating the radial average for any azimuthal angle α per image and timestep t , in a radius between $28 \mu\text{m} \leq r \leq 250 \mu\text{m}$ from the obstacles' centers. Per image, the data from 187 obstacles (see Fig. 2) were averaged.

Obstacle analysis. The spatial concentrations near the obstacles were determined as two profiles, averaged over 187 obstacles per timestep: one profile parallel and one perpendicular to the flow. The profile midpoints are the obstacle centers and with an x -axis length of $\pm 225 \mu\text{m}$, the total profile lengths cover approximately half the distance between two obstacles. Vertically, the y -axes of the profiles cover distances of $\pm 25 \mu\text{m}$, equating to the obstacles' diameters.

Robustness. To enhance the robustness of the obtained concentrations, all calculated averages are medians. All uncertainties are median absolute deviations.

2.4 Flow simulation

Two-dimensional finite volume element simulation of the fluid flow (CFD) in the microfluidic cell without biofilm was performed with ANSYS FLUENT (Ansys, Pennsylvania, USA) for the three different flow rates. The defined working fluid was the culturing medium before inoculation (dynamic viscosity = 1.34 mPa s (Ubbelohde viscometer with a type 50103/0c capillary, SI Analytics, Mainz, Germany), density = $0.99 \pm 0.01 \text{ g ml}^{-1}$). As the flow rates and Reynolds numbers were low, the flow was considered incompressible, laminar and steady state. The system was set as isothermal, without any gravity effects and the boundary conditions at the walls were considered as non-slip

and the pressure at the outlet as atmospheric. For the pressure-velocity linking, the coupled method was used. For discretisation, "second order upwind" was selected for pressure and momentum. The convergence criteria were set as 10^{-10} for all equations. For evaluation of the method, a grid independence study was performed, Fig. S6 (ESI†).

2.5 Mineralisation

Mineralisation of biofilms was conducted by crystallisation of CaCO_3 from ACC or CaSO_4 from a supersaturated solution. For the CaCO_3 mineralisation, the microfluidic cells, after they were flushed with water, were rinsed with $3 \text{ M CaCl}_2 \cdot 2\text{H}_2\text{O}$ (Th. Geyer, Renningen, Germany) solutions for at least one hour. Under a continuous flow of $1 \mu\text{l s}^{-1}$, the second inlet ((2) in Fig. 2) was first flushed with DI-water and then with a $3 \text{ M Na}_2\text{CO}_3$ (Carl Roth, Karlsruhe, Germany) solution manually injected by a syringe until the microfluidic cells clogged. The ACC precipitated directly in the growth chambers, where the ensuing mineralisation could be observed. After a rest period of about 17 h, the cells were flushed with at least 5 ml DI-water at a pressure up to 1.5 bar.

For the CaSO_4 mineralisation, solutions of $0.4 \text{ M CaCl}_2 \cdot 2\text{H}_2\text{O}$ and Na_2SO_4 (Carl Roth, Karlsruhe, Germany) were mixed and injected via a $0.2 \mu\text{m}$ syringe-filter into the cell inlets until the filter clogged after 1–2 ml solution. After a rest period of about 17 h, when no further mineralisation was observable, the cell was dried at 45°C , without flushing. Preliminary tests showed that a supersaturated solution of this concentration stays stable for approximately four minutes before crystallisation initiates.

2.6 Metrology

Imaging. Grown and optionally mineralised samples were analyzed via optical microscopy (OM, Dialux 20, Leitz, Wetzlar, Germany with camera a7 RII, Sony, Tokyo, Japan and adapter DSLRCC, Micro Tech Lab, Graz, Austria) and scanning electron microscopy (SEM, DSM 940A, Zeiss, Oberkochen, Germany).

X-Ray diffractometry. To trace ACC mineralisation, X-ray diffractometry in Bragg Brentano geometry (XRD, Miniflex, Rigaku, Tokyo, Japan with D/teX Ultra, Copper Ka, Divergence Slits 0.625° , Soller Slits 2.5°) was used. The ACC formation was repetitively measured at a speed of $10^\circ \text{ min}^{-1}$ between $28^\circ < 2\theta < 31^\circ$ in a rotating liquid sample holder (Item no: 1D0807AA with Mylar film, Rigaku, Tokyo, Japan) until no changes to the diffractograms of the mixed $3 \text{ M CaCl}_2 \cdot 2\text{H}_2\text{O}$ and $0.25 \text{ M Na}_2\text{CO}_3$ solutions were recorded.

For the assessment of biofilm mineralisation, the mineralised biofilms were measured for $10^\circ < 2\theta < 60^\circ$ and $0.5^\circ \text{ min}^{-1}$ while being spun to equalize any in-plane texture. Additionally, they were measured in transmission mode (Item no: 2101G102, Rigaku, Tokyo, Japan) with the flow direction parallel to the scattering vector, at $2\theta/2$. The measuring speed was $0.1^\circ \text{ min}^{-1}$, with divergence Slit = 0.1 mm . Since in transmission mode the sample height equals the XRD beam diameter, sample-dependent peak broadening and thus



crystallite sizes could only be obtained for the rotating in-plane measurements.

Evaluation of the data was performed by Rietveld refinement (BGMN,⁷⁹ Profex interface⁸⁰) with a verified machine line function (Standards NIST 640e and 660c). The lattice parameters of the refined phases calcite, vaterite, gypsum and halite were allowed to be in a range of $\pm 2\%$, the thermal displacement parameter were in the range of $0 \text{ nm} < B < 0.1 \text{ nm}$ and the sample offset was considered in the refinement. The crystallite sizes of the minerals were refined anisotropically and the preferred orientations by spherical harmonic functions ($1 + Y_{40}$ after Järvinen⁸¹) with directional weighting factors w . Not all experiments could be evaluated, due to a lack of sufficient sample volume and thus lack of anisotropical refinement. The comparatively few, weak and broad cellulose reflexes were not considered for refinement.

Cellulose orientation. Evaluation of cellulose orientation was performed by visualisation of the cellulose birefringence *via* polarized light microscopy (RMA 5, Mikroskop Technik Rathenow, Rathenow, Germany, with a λ (525 nm phase shift) retardation plate and Eos 550D, Canon, Tokyo, Japan). Between two crossed polarizers, the dried samples were oriented in the flow direction at 0° , at -45° and $+45^\circ$. The images at the different angles were taken at fixed exposure times and gain for comparability.

3 Results

3.1 Biofilm growth

At all flow rates, the biofilm growth started evenly at the obstacles and spread in the direction of the flow, connecting the obstacles, Fig. 3(b)–(i). From there, the development of a film, which connected the rows perpendicularly was visible and can be seen in Fig. 3(e)–(g) or as a video, see the ESI.[†] The growth speed at 1.0 mm s^{-1} was visibly slower than at higher flow rates. Independent of the flow rates, when the cells were increasingly filled by biofilm, the images darkened, indicating an increased concentration of BCB. When the flow was reduced by clogging of the setup, the darkening reduced and stagnated.

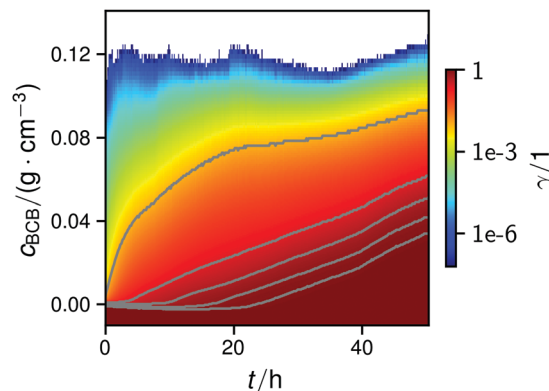


Fig. 4 Map of the normalized cumulative histogram $\gamma(c_{\text{BCB}}, t)$ at a flow rate of 12.5 mm s^{-1} . The gray plotted lines are isolines of $\gamma = 0.01, 0.25, 0.50, 0.75$, and 0.9 . They were fitted with a bi-logistic function $c_{\text{BCB}}(\gamma, t)$, eqn (4).

Scanning electron micrographs showed that the BCB was built from several paper-like layers parallel to the flow, which were continuously connected by BC fibers. Between the obstacles and in the flow direction, the BCB appears to be thicker with a higher fraction of bacteria, which are roughly estimated at $0.01\text{--}0.1 \text{ ml ml}^{-1}$. At the higher concentrated spots, the thickness of the dried film only reached a few microns. For brevity, these images are shown as the ESI,[†] Fig. S2 and Fig. S3.

Statistic growth. The distribution of the BCB concentration could be illustrated over time. Fig. 4 shows exemplary which fraction γ of the total area of the microfluidic chamber reached at least a particular concentration c_{BCB} . A closer look at the plotted area fractions shows a bi-logistic growth with the second logistic function starting after the first reached its saturation.

In all conducted experiments, the c_{BCB} of the recorded data did not exceed 0.2 g cm^{-3} which was below the setup saturation. It should be noted, however that data collection always ended before the second logistic function could reach saturation due to clogging of the setup.

By fitting the histogram data with a bi-logistic function (eqn (4)), parameters from Fig. 5 were obtained. The fits'

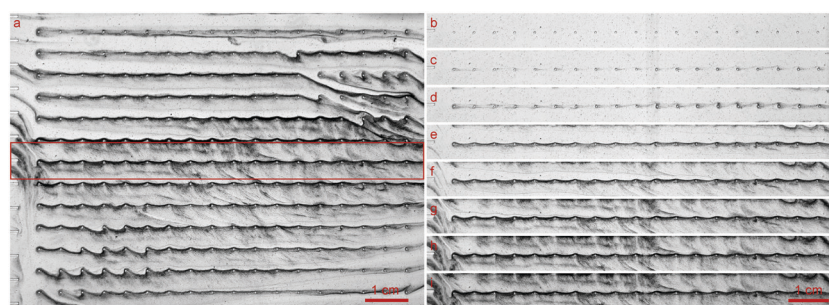
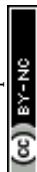


Fig. 3 Grown biofilm of *K. xylinus* in the microfluidic cell growth chamber (see Fig. 2) at a flow rate of 1.0 mm s^{-1} running from left to right. (a) Due to the panoramic viewing window of the optical setup, the biofilm growth around over 230 obstacles could be determined. The inlet channels, whose exits can be seen on the left, regularly allowed an even distribution of the fluid into the growth chamber. (b)–(i) is the cutout highlighted in (a) by a red frame. (b)–(h) Growth at the middle row in time steps of 6 h. Image (h) corresponds to the large image (a) and captures the moment after 36 h of growth when the pressure reached 1 bar and the flow reduces due to the servo loop. Image (i), unchanged from (h), shows the moment 94 h after the flow rate stagnated due to clogging of the setup.



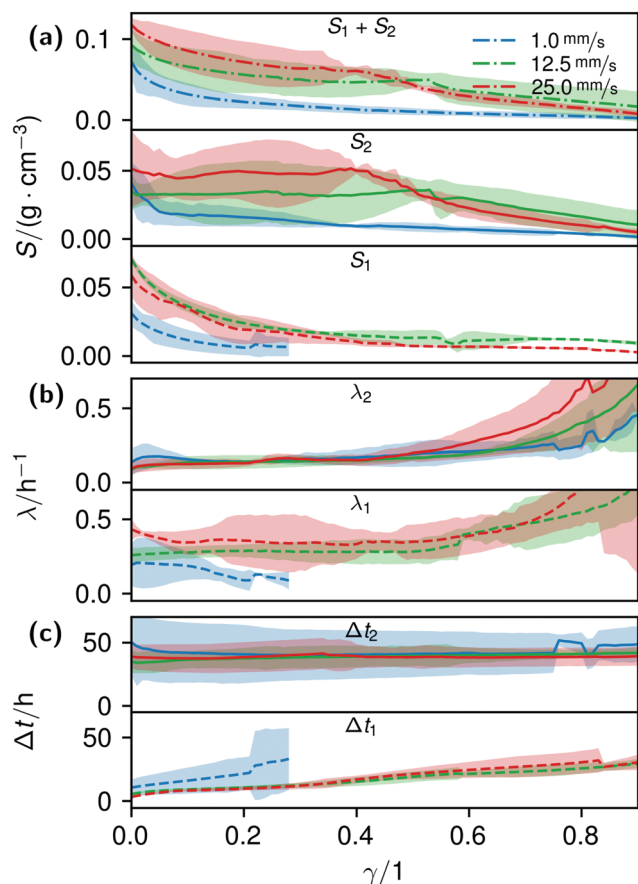


Fig. 5 Plots of the histogram parameters (e.g. Fig. 4) from eqn (4). The plots end when the fitted parameter of at least two of the four conducted measurements reached zero, or unrealistic values. Dashed lines (Index 1) are the parameters of the first term, solid lines (Index 2) of the second term of eqn (4). Dash-dotted lines in (a) represent the cumulative values S of the first and second terms.

uncertainties increase when the data increasingly shows only sections of entire logistic growth curves at high γ .

Cumulative growth. At higher flow rates, the average BCB concentrations of the biofilms rose initially linearly with a slope of $8.7 \pm 2.7 \times 10^{-4} g cm^{-3} h^{-1}$ for $12.5 mm s^{-1}$ and $9 \pm 4 \times 10^{-4} g cm^{-3} h^{-1}$ for $25.0 mm s^{-1}$. At $1.0 mm s^{-1}$, it started slower and slightly accelerated over time, see Fig. 6. A linear regression fit yielded a slope of $24 \pm 9 \times 10^{-5} g cm^{-3} h^{-1}$.

Azimuthal growth. Azimuthal analyses of the obstacles' surroundings are shown in Fig. 7. BCB concentrations initially increased along the flow, with a higher increase rate at higher flow rates, Fig. 7(a). In (b) it can be seen that the concentrations become more and more isotropic with time.

Obstacle growth. In Fig. 8, the BCB spatial concentration profiles around the obstacles, parallel (a) and perpendicular (b) to the flow, are displayed. While the concentration spreads evenly perpendicular to the flow (b), a maximum appears at the flow-facing sides. This effect increases with the flow rate, in particular between $1.0 mm s^{-1}$ and $12.5 mm s^{-1}$.

Cellulose orientation. The orientation of the BC was investigated from the dried BCB textiles by polarized light

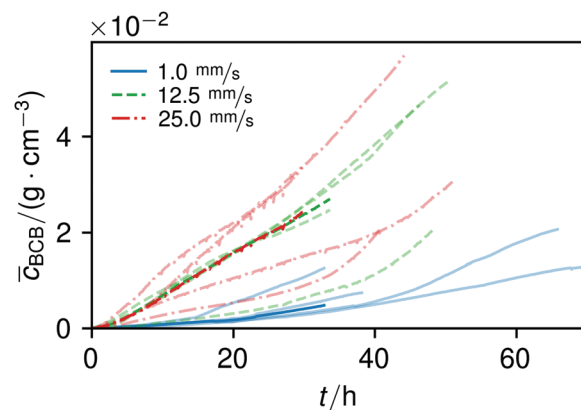


Fig. 6 Increase of the average BCB concentration over time, together with the median of the different flow series. To show the variance of the measurements, the data from the single experiments are overlaid in lighter tones.

microscopy. Fig. 9 shows the birefringence of the cellulose under different angles. According to the Michel-Lévy interference colour chart, the bright regions of the dried analysed sample had a film thickness of $2.5 \pm 1 \mu m$ which is in agreement with the measured thickness at the BCB concentration calibration.

3.2 Flow simulation

The local Reynolds number Re was determined in the CFD simulation for all flow rates. With the results of $Re_{max} = 24$ in the inlet, $Re_{max} = 20$ on the chamber and $Re_{max} = 3.7$ around the obstacles the flow was considered laminar, since all values were below $Re_{Kar} \sim 40$, at which Karman vortices start to appear after cylindrical obstacles.⁸²

Even though the flow was laminar all the time, at higher flow the field at t_0 was no longer perfectly distributed in the growth chamber, see Fig. S7 (ESI†). Nevertheless, the wall shear stress τ around parallel obstacles located in the middle of the growth chamber showed a high uniformity at every flow rate (Fig. 11(a)). This also illustrates the spatially resolved flow field, Fig. 10.

3.3 Mineralisation

ACC. To assess the ACC mineralisation method, the formation of a crystalline phase from ACC was analysed *in situ* by mixing 3 M $CaCl_2 \cdot 2H_2O$ and 0.25 M Na_2CO_3 solutions. The development of the main calcite reflex (104) from ACC can be seen in Fig. S8 (ESI†) after 1–2 minutes. In parallel, the formation of crystals from the ACC slurry was observed after approximately two minutes. Beneficially for the conducted mineralisations, it emerged that ACC from mixtures where both reagents had a concentration of 3 M required more time to form and crystallise.

Crystallisation. ACC was distributed over the BCB in the growth chamber until no further change in brightness was visible. Then the formation of $CaCO_3$ crystals could be observed. The formation appeared stronger where the ACC was previously more concentrated. However, areas with higher BCB concentration also tend to show increased crystal

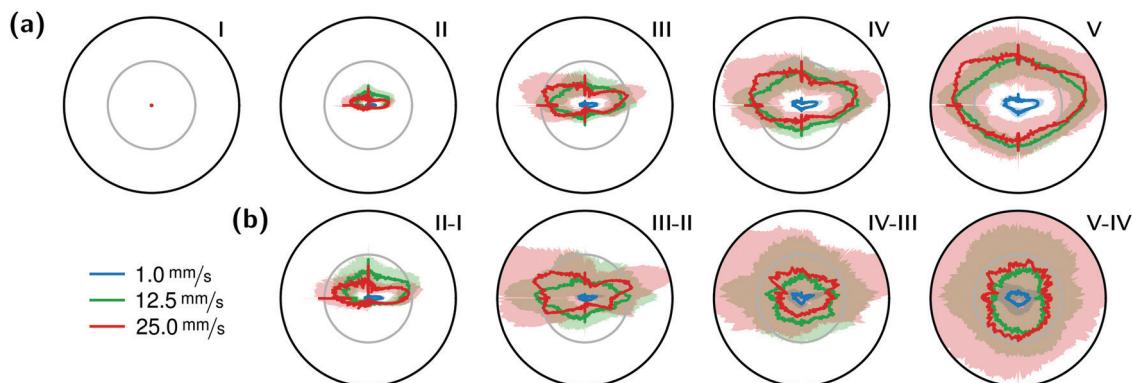


Fig. 7 Azimuthal plots of the obstacles' surroundings with the flow running from left to right in timesteps I–V of 7.5 h, whereby timestep I was 10 minutes after the start. Inner circles in row (a) represent a BCB concentration of 0.02 g cm^{-3} , outer circles 0.04 g cm^{-3} . Row (b) shows the differences between two timesteps from the upper row. Here, the inner circles represent 0.01 g cm^{-3} and the outer circles 0.02 g cm^{-3} .

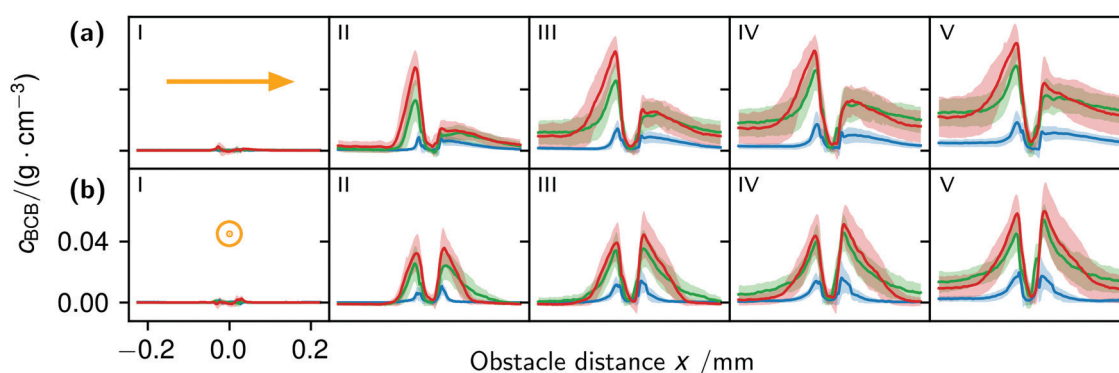


Fig. 8 Profile plots parallel (a) and perpendicular (b) to the flow in timesteps I–V of 7.5 h, where timestep I was 10 minutes after the start. The obstacle center is the x-axis center and fluid flow is indicated by the orange arrows. The blue line represents flow at 1.0 mm s^{-1} , the green line at 12.5 mm s^{-1} and the red line at 25.0 mm s^{-1} .

formation. In areas without BCB, no crystal growth was visible. Micrographs of the dried BCB showed that the crystals were distributed over the entire BCB surface with sizes from $0.1 \mu\text{m}$ to $100 \mu\text{m}$. No formation of aragonite was observed by microscope or XRD. A dependence of the ratio between the two CaCO_3 phases calcite and vaterite due to the different flow rates could not be detected. Nonetheless, the fraction of vaterite and thus the ratio between the phases varied considerably between the experiments. For future evaluation of the variation, the total mass balance between the organic fractions (BC amorphous, BC crystalline; hemicelluloses, bacteria, ...) and the mineral phases would need to be determined, which was not feasible with the current samples.

Gypsum formation, identified by XRD as $\text{CaSO}_4 \cdot 2\text{H}_2\text{O}$, was observed to be more localised: approximately two minutes after injection, several hundred observable crystals formed. Growth was visibly concentrated on these aggregates which increased their sizes up to half a millimeter. These aggregates consisted of differently sized needles which appeared to have a slightly preferred orientation in the direction of the flow. During drying, the observed number of aggregates and single needle-like crystals smaller than a tenth of a millimeter increased.

Small amounts of halite, residual from the mineralisation reagents were observed, but not further considered.

Micrographs of the dried samples of both mineralisation paths showed that the crystals grew on both the BCB surfaces and between the layers, pressing them apart by being embedded in the BC network. An overview of the SEM images can be seen in the Fig. S4 and S5 (ESI†).

Oriented crystallisation. Further evaluation of the inorganic phases from the Rietveld refined data are compiled in Table 2. Notable is the smaller crystallite size L of calcite grown on BCB, grown at 25.0 mm s^{-1} . Gypsum shows a significant difference in the ratio of L and w between the a - and c -axes to the b -axis at all flow rates. In contrast, the w values of calcite and vaterite indicate no significant orientation.

4 Discussion

4.1 Biofilm growth

4.1.1 Dependence on flow rate. A correlation between the flow rate and the increase of the final BCB concentration was demonstrated: by increasing the flow from 1 mm s^{-1} to



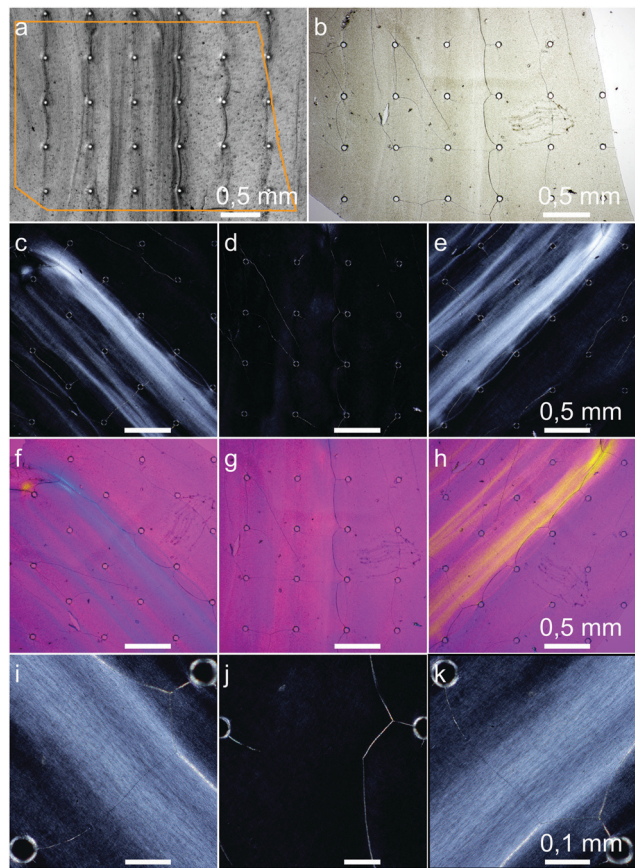


Fig. 9 Micrographs of cellulose structured at a flow velocity of 1.0 mm s^{-1} . (a) shows a clipping of the last image during the growth experiment near the inlet channels. The orange contour marked the piece of the dried cellulose film (b) which was investigated by polarized light microscopy. Images (c), (f), and (i) shown the sample oriented at -45° , (e), (h), and (k) by $+45^\circ$ and (d), (g), and (j) by 0° in relation to the polarizers. For images (f), (g), and (h), a $n \lambda$ retardation plate was inserted.

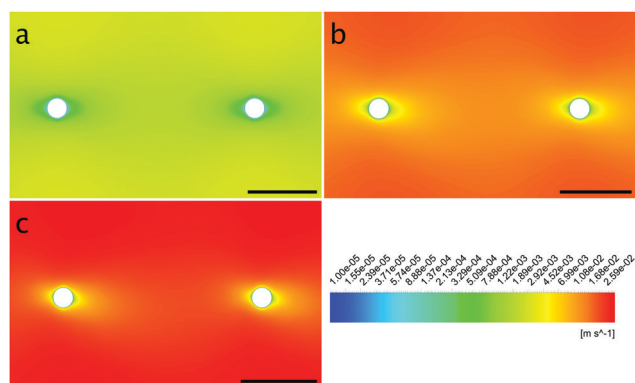


Fig. 10 Contour plots of the obstacles' surroundings at the flow rates 1.0 mm s^{-1} (a), 12.5 mm s^{-1} (b) and 25.0 mm s^{-1} (c) with flow from left to right.

12.5 mm s^{-1} , it was raised by a factor of 3.6. Since no difference in the rate of increase between 12.5 mm s^{-1} and 25 mm s^{-1} was

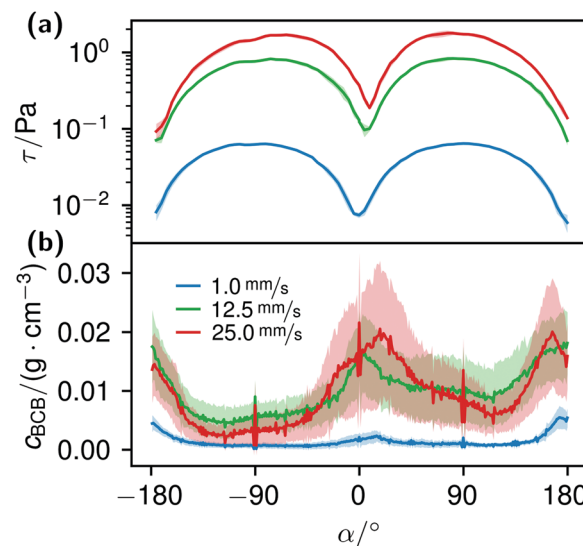


Fig. 11 Image (a) shows the calculated wall shear stress around nine parallel obstacles located in the middle of the microfluidic cell at different flow rates. In image (b) the azimuthal concentration $c_{\text{CCB}}(\alpha)$ around the obstacles, 15 h after the start of the experiments, is pictured. The flow direction was from 0° to $\pm 180^\circ$.

Table 2 Results of the Rietveld refinements. Crystallite size L and $w_{\perp}$ values are from the measurements, measured with Bragg–Brentano geometry. $w_{\parallel}$ values are from the measurements in transmission mode. With the former reflexes from lattice planes perpendicular to the flow, and with the latter, lattice planes in the direction of the flow were measurable. # denotes how many samples per flow rate were available and used to calculate the values. The gypsum crystal size value $L_b \gg 208$ indicates the method's detection limit

Mineral	Flow rate/ (mm s^{-1})	#	(hkl)	L/nm	$w_{\perp}/1$	$w_{\parallel}/1$
Calcite	1.0	1	(100)	102 ± 3	0.28 ± 0.01	0.36 ± 0.01
			(001)	99 ± 4	0.44 ± 0.02	0.29 ± 0.01
		2	(100)	104 ± 5	0.33 ± 0.01	0.38 ± 0.04
	12.5	2	(100)	103 ± 7	0.35 ± 0.02	0.24 ± 0.04
			(001)	54 ± 1	0.33 ± 0.01	0.33 ± 0.01
		3	(001)	50 ± 1	0.33 ± 0.01	0.33 ± 0.01
Vaterite	12.5	1	(100)	28 ± 0	0.30 ± 0.00	0.31 ± 0.01
			(001)	27 ± 0	0.40 ± 0.01	0.37 ± 0.02
		3	(100)	23 ± 0	0.31 ± 0.01	0.29 ± 0.01
	25.0	3	(100)	28 ± 2	0.38 ± 0.03	0.41 ± 0.03
			(001)	28 ± 2	0.38 ± 0.03	0.41 ± 0.03
		1	(001)	28 ± 2	0.38 ± 0.03	0.41 ± 0.03
Gypsum	1.0	1	(100)	120 ± 7	0.04 ± 0.00	0.57 ± 0.02
			(010)	$\gg 208 \pm 25$	0.87 ± 0.01	0.09 ± 0.00
		2	(001)	123 ± 6	0.09 ± 0.00	0.33 ± 0.01
	12.5	2	(100)	127 ± 14	0.09 ± 0.01	0.47 ± 0.04
			(010)	141 ± 5	0.79 ± 0.03	0.13 ± 0.01
		1	(001)	122 ± 20	0.13 ± 0.01	0.40 ± 0.04
	25.0	1	(100)	87 ± 5	0.07 ± 0.00	0.53 ± 0.02
			(010)	208 ± 7	0.77 ± 0.03	0.05 ± 0.00
		1	(001)	116 ± 8	0.15 ± 0.01	0.41 ± 0.01
			(010)	116 ± 8	0.15 ± 0.01	0.41 ± 0.01
			(001)	116 ± 8	0.15 ± 0.01	0.41 ± 0.01

observed, it can be assumed that it saturates between 1 mm s^{-1} and 12.5 mm s^{-1} , Fig. 6.

To assess the system's productivity, the total volume of the pumped inoculum should be considered. Hence in the observed section, the productivity was $(1.8 \pm 0.7) \times 10^{-5} \text{ g (l h)}^{-1}$ for 1 mm s^{-1} , $(6 \pm 2) \times 10^{-6} \text{ g (l h)}^{-1}$ for 12.5 mm s^{-1} and $(3 \pm 1) \times 10^{-6} \text{ g (l h)}^{-1}$

for 25.0 mm s^{-1} . These productivities may be low, compared to the values Campano *et al.* reviewed in their Table 3.⁸³ However, our productivity only represents biomass from the viewing window and not from the whole setup, including the reservoir.

The interesting information from these values, which decrease with higher flow rates is another: they show that the concentration increase rate at higher flow rates is not explicable by a better nutrient or oxygen supply due to the higher agitation. Literature reports confirm that agitated bacteria cultures in general do not necessarily show a better performance than static ones.^{84,85} A higher biomass production follows from lower velocities, as Krsmanovic *et al.* reviewed.⁸⁶ This effect can also be observed in microfluidic systems.⁴⁵ Considering the production of BCB, Liu *et al.* found that the cell viability of *K. xylinus* was clearly increased by agitating. But the data also shows that even low agitation reduced the BCB production. Thus, referring to the BCB yield per cell, the agitation had a strong disadvantageous effect.⁸⁷ For aligned BCB, Luo *et al.* found similar results, with an observed biomass reduction from around 38 mg ml^{-1} to 24 mg ml^{-1} at flow rates of 30 mm s^{-1} and 150 mm s^{-1} (and around 55 mg ml^{-1} for the static reference).³⁷ The reason for this observation is rather not a genetic mechanosensing, but a prolonged lag-phase due to washout of biofilm substances.^{44,49}

4.1.2 Network formation. Interestingly, despite the significantly higher volume-dependent productivity of the 1 mm s^{-1} in our experiments, we partly observed a shorter lag phase, a higher growth constant and a higher concentration for the higher velocities as shown in Fig. 5. How does this fit together? An explanation for the observed flow dependence is that with an increasing flow rate, more bacteria attached to the obstacles' sides facing the incoming flow, resulting in an overall higher BCB production. Fig. 8 confirms this hypothesis, since the flow-facing side shows a significantly higher concentration which increases with the flow rate. We propose a mechanism as follows.

K. xylinus, which can be considered non-motile when floating freely, in principle, is not influenced by flow, except for a rotary type of stroke.^{51,52} Hence, it 'goes with the flow'. When the flow hits an obstacle, bacteria adhere to the obstacle. Once adhered, the bacterial BC then facilitates the attachment of further bacteria, leading to their rapid accumulation. The calculated shear stress at the obstacles, Fig. 11(a), supports this idea: with a maximum wall shear stress around 2 Pa for 25.0 mm s^{-1} perpendicular to the flow direction and a ten times lower minimum at the flow facing direction, we assume that the bacteria withstand the shear force at the flow-facing side. Although we could not find data about the specific adhesion force of *K. xylinus* to PDMS, it is likely that the critical shear stress, at which attachment and detachment are in balance, is not smaller by orders of magnitudes to *P. aeruginosa* with $1.1 \pm 0.2 \text{ Pa}$, due to their similar shapes and sizes.⁸⁸ The work of Chaen *et al.* supports this by demonstrating BCB structuring, formed at an estimated 1 Pa with *G. hansanii*. In our experiments, it can be assumed that the adhesion is even higher: *K. xylinus* seems to be more difficult to wash off polar surfaces such as PDMS due to microbial cellulose

hydrogen bond attachment.^{89,90} And additionally, if *K. xylinus* already extruded fibers, we assume that they further enhance adhesion, making it more difficult to detach.

The correlation of the minima and maxima in Fig. 11(a) and (b) indicates that *K. xylinus* started to form its biofilm with its origin in the areas that were protected from shear forces.⁸⁵ Hence *K. xylinus* initially attached to the obstacles' flow-facing sides, from where it excreted BC, whereby the amount of initial bacteria depended on the flow speed, see sketch Fig. 12, left hand side. Extruded BC then elongated with the flow, embracing the obstacles (Fig. 8(b) (II)) and forming an obstacle-to-obstacle network in the direction of the flow, Fig. 3(b)–(e) and sketch Fig. 12 center image. While for higher flow rates, the median BCB concentration is equal in front of and behind the obstacles (Fig. 7(a)), the spatial distribution has a maximum on the flow-facing side as already illustrated for all flow rates, Fig. 8(a). After the BCB streamers connected after about 10 h, this maximum value does not increase further. Rather, the obstacle-to-obstacle concentration rose, building a highly concentrated network in the direction of the flow, with an increasingly harmonised concentration distribution. All tested flow rates resulted in a distinct obstacle-to-obstacle network. At higher flow, the network was more highly concentrated. This concentration rise can be attributed to a better nutrient supply of the consisting network due to the higher dynamic pressure.

The initial bilateral flow-dependent concentration distributions became weaker with time, as Fig. 7(b) indicates. It changed from being aligned with the flow, to more uniform with and lateral to the flow. The lateral spreading of the BCB can also be observed with its spatial concentration profile in Fig. 8(b). Looking at the single strands, the spreading initially occurs unilaterally and becomes uniform after the strands connected. Fig. 8(b) indicates that when facing the flow, statistically the concentration distribution is slightly shifted to the right. But sometimes, it also occurred left-sided, as it can be seen with the spreading of the BCB in Fig. 3(e)–(g). We speculate that with time, the biofilm also grew in the microfluidic inlet. This caused minimal pressure differences between the grown strands due to different inlet flows. Under perfect initial conditions, the resulting cross flow in principle occurs randomly. But in CFD simulations, the used microfluidic cell already shows a very slight initial cross flow at 12.5 mm s^{-1} and 25 mm s^{-1} due to a short inlet channel (Fig. S7, ESI†). It is likely that the observed slight right-handed concentration distribution (Fig. 8(b)) can be contributed to a unilateral biofilm growth in the microfluidic inlet, caused by the initial cross flow. However, since the network concentration equalized with time, this probably had no effect on the final network. Its total concentration distribution becomes more and more uniform and a connected network with higher concentrated regions along the obstacles forms, see sketch Fig. 12, right image.

The angular changes of the total concentration from directional to uniform were visible after a similar growth duration at all tested flow rates with a minimal delay at 1 mm s^{-1} . This becomes visible in Fig. 7(b) IV–III and V–IV and is called the second growth phase in the following, being distinguishable from the first logistic growth phase, Fig. 4. In summary, the growth can be described in two sequential processes: first, the



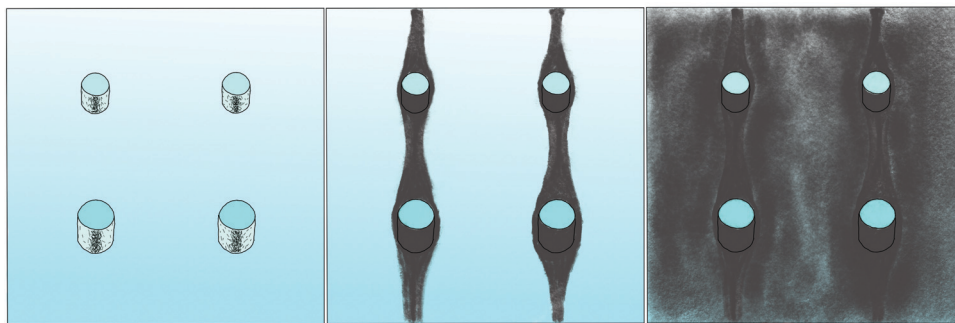


Fig. 12 Sketch of the BCB network formation around the PDMS obstacles with the initial bacteria accumulation (left), the first growth phase (middle) and the second growth phase (right). In this sketch, the flow comes from below.

interconnection of the obstacles in the direction of the flow and second, their connection with a general increase of the concentration over the whole biofilm.

The formation of the flow-aligned obstacle-to-obstacle network in the first phase can also be observed in the statistic growth data, as dashed lines in Fig. 5. Since the main concentration is located in the direction of the flow between the obstacles (Fig. 3) and the area of these interspaces equals $\gamma \approx 0.1$, it further can be assumed that the region of the first growth phase below $\gamma = 0.2$ in Fig. 5 mainly represents the obstacle-to-obstacle growth. That the relevant values of S_1 are mostly in this region fits this assumption. On the basis of the fitted parameters (Fig. 5), the forming of this network becomes traceable: a higher flow rate enhanced the growth constant, the final concentration and shortens the time-point of the inflection. We assume that the former change is due to an ongoing and flow dependent catching of floating bacteria similar to the “fishing-line” observation of Marty *et al.*⁹¹ The change of the latter two started to saturate with a higher flow. On the one hand, in contrast to the second growth phase, nutrients could reach easily the inner of the relatively small strands, while a higher dynamic pressure did not enhance the final concentration by the increased fluent exchange. On the other hand, the inflection time-point stops decreasing. It appears that the lag-phase became the speed-determining step, indicating that influences such as the nutrient resorption of the bacteria and other influences dominated.

The overall increase in concentration at the flow rates 12.5 mm s^{-1} and 25 mm s^{-1} in both phases was quite similar in their main grow directions (first phase with the flow; second phase uniformly) and mostly in a range between 0.005 and 0.01 g cm^{-3} in 7.5 h , see Fig. 7(b). At 1.0 mm s^{-1} it was around 0.001 g cm^{-3} in the same time-step. Interestingly, the growth constant of the second growth phase is widely the same for all flow rates, Fig. 5. Different from the “fishing-line” effect in the first phase, in the second growth phase the catching of bacteria seemed no longer to be the dominant mechanism. Rather, an equilibrium was reached at every flow rate, resulting in a growth constant which stayed stable and can be seen as the real growth constant for biofilm with the given culturing parameters. Because on the one hand, by a higher shear stress, more bacteria were embedded in the existing biofilm. On the

other hand, there were more high-shear regions, where bacteria and biofilm substances could no longer adhere or become detached. We assume that these high-shear regions arose from a smaller available volume, due to the increasing filling of the microfluidic cell. This is different from the observation by Drescher *et al.*, who consistently saw a more rapid clogging with higher flow rates.⁴⁵ By contrast to the present work, their setup was pressure-, and not flow-controlled, resulting in a defined maximum flow rate and therefore in a maximum shear stress the biofilms have to endure.

4.1.3 Statistic growth. As already mentioned, the BCB growth in the microfluidic chamber is characterised by two logistic growth phases. Fig. 5 shows the fitted parameters of the bi-logistic function, eqn (4). As already stated, the first phase can be related to the growth of the obstacle-to-obstacle BCB strands, whereas the second phase is related to the strands' connection. However, that they are clearly distinguishable in the statistic growth (Fig. 5), is due to their saturation. This saturation, in turn, was mainly triggered by a lack of space on which *K. xylinus* biofilm can spread. Presumably because of the comparatively thin microfluidic cell ($50 \mu\text{m}$), the BCB concentration reached levels where the nutrient supply decreased and thus the BCB concentrations leveled out. With a higher flow rate and the resulting higher dynamic pressure, this supply could be longer preserved by an elevated fluent exchange, leading to a higher total concentration $S_1 + S_2$, Fig. 5(a). In total, $S_1 + S_2$ can be interpreted as the concentration distribution over the observed area. It illustrates that the obtained BCB gel showed regions of different densities, similar to the biofilm of Hassanpourfard *et al.*^{54,55} Their observed ‘stick-slip’ behavior was not observable in our experiment, since the viscoelasticity of their EPS strongly differs from a web of bacterial cellulose. After drying, the different densities resulted in inhomogeneous film thickness. The thickness shrank by a factor of 10 to 30.

The average BCB concentration increase $\bar{c}$ is nearly the same for 12.5 mm s^{-1} and 25 mm s^{-1} , Fig. 6. Since a higher flow rate does increase the total concentration over a broad range as in Fig. 5(a), the final average BCB concentrations will differ: Fig. 6 only shows the increase until the setup clogged. The end of this BCB gel redensification was only observable for two of the 1 mm s^{-1} flow rate experiments after approximately 60–70 h. Since the S_2 values in Fig. 5 are extrapolated by obtaining the



best fit, it is to be expected that all $\bar{c}$ saturate. The appearing linear increase in Fig. 6 thus was the superposition of both growth phases. In the end, the saturated $\bar{c}$ of 25 mm s^{-1} then will be higher than that of 12.5 mm s^{-1} . Moreover, the time-point of the saturations can be estimated. Since Δt_2 is very similar for all flow rates, and in the region of approximately 40 h, the observed 60–70 h are likely for all flow rates.

The median of the second phases' inflection points Δt_2 and growth constants λ_2 changed only insignificantly. It shows, as already stated, that the growth in the second phase was mainly independent of effects like an increased and continuous aggregation of bacteria. It should be noted that due to the increasingly shorter isolines at higher γ , in particular the uncertainty of λ strongly rose, making its interpretation difficult.

4.1.4 Hierarchical structure. That BC fibers can be aligned by growing in flow was observed in several experiments.^{36,37} The principal mechanism behind the alignment was the subject of several studies: Stoodley *et al.* observed that ellipsoidal bacteria, which *K. xylinus* belongs to, align with the flow within their streamers.⁵⁰ Furthermore, Holm *et al.* confirmed the intuitive assumption that fibers also align with the flow as largely correct. Although short fibers near the wall tend to orient perpendicular to the flow direction, the longer they become, the more they swim with the flow.^{92,93} Additional to this orientation of the BC fibers, for *K. xylinus* there is also the fact that already deposited cellulose strands serve as templates for further depositions.^{16,26}

Our experiments showed rheotactically aligned BC already at the lowest flow rate of 1 mm s^{-1} , Fig. 9. In images (c)–(k), the birefringence of the long optical axis in the direction of the (001) planes of the highly crystalline bacterial cellulose can be seen. Since the birefringence is strongest visible at $\pm 45^\circ$ between the flow direction and the crossed microscope polarizers, the fibers which are elongated along the (001) planes were aligned in the direction of the flow. The fact that also higher concentrated regions are outside of the highly concentrated obstacle-to-obstacle strands shows that the majority of the bacterial cellulose was flow-aligned. On closer inspection, the crystalline birefringent regions of the aligned fibers can even be seen in Fig. 9(i), (k) with an approximated mean length of about $10 \mu\text{m}$.

Besides the alignment, Chae *et al.* found, in their flow-driven alignment experiment with the BCB producing *G. hansenii*, an additional structural characteristic: in contrast to other alignment methods like BC stretching, the BC chains showed parallel packing. This results in macroscopic net dipoles *via* the alignment from the fibers' reducing end to their non-reducing end.³⁸ We assume it is possible that the current flow driven alignment approach results in the same packing and that the hierarchical structure features persist even more distinct from the molecular to the macroscopic level by rheotactic structuring compared to other structuring approaches.

4.2 Mineralisation

To obtain a composite material with structured BCB as the substrate, we performed two quasi *in situ* mineralisations.

In the gypsum mineralisation, flower-like gypsum aggregates occurred. Nissinen *et al.* and Nindiyasari *et al.*^{76,94} demonstrated gypsum/cellulose composites with mineralisation from hemihydrate or supersaturated $\text{CaCl}_2\text{--Na}_2\text{SO}_4$ solutions. The reason for the more uniform crystal distribution and size by Nissinen *et al.* can be seen in the used cellulose substrate. While Nissinen *et al.* used a cast cellulosic film, bacterial cellulose consists of ribbon-shaped cellulosic nanofibers with a width of 40–60 nm.²⁵ Cellulose in general can be seen as supportive for gypsum precipitation,⁹⁴ but the anhydride formed from the supersaturated solution requires water for the hydration steps over bassanite to the final $\text{CaSO}_4 \cdot 2\text{H}_2\text{O}$.⁹⁵ Water-soaked cellulose can supply an anhydrid nuclei. If the water release decreases, due to a decreasing size of the fibers, the formation of nuclei becomes more unlikely,⁹⁴ resulting in a degraded and more heterogeneous crystal distribution.

To determine whether the anisotropic structure of the substrate affects the orientation of the crystals, we performed two different XRD measurements: measurements with Bragg–Brentano geometry and applied spin, to discriminate texture within or out of the sample plane ($w_\perp$). With measurements in transmission mode, textures along the sample plane in the direction of the flow are determinable ($w_\parallel$). Gypsum refined XRD data show that L and $w_\perp$ in the direction of the (010) planes are increased against (100) and (001). The (001) planes show the same effect against the (100) planes to a lesser extent, Table 2. Thus the data indicate the texture in two directions: first, the (010) plane of the gypsum crystals, laid parallel to the BC film. This can also be seen in Fig. S5(b)–(d), ESI† where the acerous morphology has a distinct and visible tabular fraction largely facing the observer. We assume that this favored orientation normal to the BCB surface is due to the strong BCB shrinkage, since XRD measurements are obtained after drying. The gypsum crystals were probably twisted by the shrinking in a way that the biggest surface ((010) plane) finally lay between the BC layers, reducing the fibers' tensions. A slight decrease of this anisotropy with a higher flow rate and thus a higher concentration is explainable by better embedding, reducing the twisting of the crystals. Second, since $w_\parallel$ of the (100) plane is slightly more distinctive than the (001) plane, slight orientation of the crystals in the direction of the flow is indicated. This orientation is probably caused by heterogeneous nucleation. Some of these crystals then formed large aggregates, which overall showed a slight statistic orientation with the flow, Fig. S5(a), (ESI†). The observed aggregates do not correspond to the determined L values. Since the XRD data represent the whole samples and SEM images are limited to the samples' surfaces, further evaluation of the interlayer BC structure, which is out of the scope of this work, might resolve the missing correspondence.

The stable ACC suspension was well distributed over the samples, resulting in a good final distribution of CaCO_3 with small crystallites embedded in the BCB. Since the directional w does not differ significantly from 0.33, the growth of the obtained CaCO_3 phases calcite and vaterite seemed to be isotropic and not affected by the aligned BCB substrate, Table 2. However, the mean crystallite size L of calcite was probably influenced, reducing to



half at a flow rate of 25 mm s^{-1} . Due to the lack of repeating measurements for the lower flow rates, this value has some scope for interpretation. But, an influence on L caused by the higher BCB concentration in areas $\gamma < 0.5$ is quite possible, since with a larger surface, more heterogeneous nucleation can take place.

We processed a one precursor mineralisation of the BCB samples, immediately after growth in the same setting. This quasi *in situ* mineralisation route, based on liquid impregnation,⁷² provides – besides its simplicity – advantages by comparison to other *in situ* or *in vitro* routes. Regarding the former, the growth process and thus BC properties such as crystallinity are not affected by mineralisation ingredients and can be conducted at optimal process parameters.⁵⁹ The latter on the other hand brings along some difficulties by transferring fragile samples into a new vessel where mineralisation can be performed, *e.g.* by a diffusion based⁶⁸ or time-consuming gas diffusion based mineralisation.^{68,96,97} The one-precursor method further was beneficial, since in our setup already minimal rinsing between the reagents prohibited the precipitation and a double diffusion method applied by Grassmann *et al.*⁹⁸ was rather inapplicable.

In conclusion, the slight effect of the anisotropic substrate on gypsum crystallisation seems promising for the production of a phase overlapping hierarchical engineering material. Even though gypsum had some limitations, other acerous minerals such as montmorillonite could overcome the insufficient distribution. However, due to its good distribution and embedment, the CaCO_3 route seems promising to create a biomimetic organic/inorganic composite. An interesting work was conducted by Yu *et al.* where a BC composite with tunable toughness was produced.⁹⁹ It is quite imaginable to apply the presented process with pre-aligned BCB as a substrate in a quasi *in situ* route. Indeed, fabrication steps like purification by boiling with sodium hydroxide, or blending of the BCB must be adapted. But even if bacteria remain after adaption in the composite similar to our experiments, they can induce interesting properties such as improved absorption capacity or higher maximum tensile/compressive strength.¹⁰⁰

4.3 Experimental setup

In principle, three types of biofilm reactors are suited for the targeted rheotactical cultivation of biofilm streamers.¹⁰¹ Rotational reactors, where either the inoculum or the sample is in rotation to produce a flow,^{36,37} or loop type reactor, where the reactor is fixed and constantly flushed by medium or inoculum. While the former two are mostly used to produce biofilms in large quantities, the latter method is suitable to evaluate biofilms during growth.^{15,44–46} Besides a better possibility to maintain axenic conditions, the loop type setup enables the use of flow- or pressure driven material synthesis. Since in open or rotational reactors, the inoculum primarily flows on top of the grown BCB, tracing increases in concentration to flow rates, as in this work, is not feasible.

The main limitation of the loop setup is the limited space of the reactor (in this work the microfluidic cell). In our experiments, the saturation occurred due to this limitation. While in other reactors, it can be expected that different growth

parameters will be obtained, since especially the way of nutrient supply differs by the higher degree of freedom of the medium/inoculum flow, an eventual saturation is inevitable. And, if the structuring element where the BCB grows is similar to the obstacle pattern used in this work, a two phase growth will probably also arise. Hence, for obstacle-induced growth, where a defined microbial streamer shall form a cohesive streamer network, our results give an appropriate basis that can be adapted to produce structured material with loop type reactors, but also with rotational reactors.^{36,37} To summarise, with our loop type reactor with a panorama view observation setup, we were able to observe the formation of the BCB network and spatial streamers simultaneously. To our best knowledge, investigation about the formation of larger bacterial streamer networks, in particular BCB streamer networks are not available. Thus, the flow-influenced formation of two hierarchical levels could be observed directly, knowing that also the subjacent levels like the direction of the cellulose fibers and ribbons are aligned by the flow, as apparent in Fig. 9.

Our method is suitable to measure the BCB concentration profile over the grown network *via* the Lambert–Beer equation in a dynamic range of 1 : 932. Even after several days of growth, the optical examination did not reach its saturation. Besides the attenuation of the light by BCB, a slight (focused) shadowgraph effect was visible at the thin edges between BCB and the surrounding inoculum (*e.g.* visible in Fig. 3(f) at the spreading BCB edge). This effect was caused by different refractive indexes of the inoculum and the BCB and the resulting refractive gradient.¹⁰² On the one hand, this effect resulted in a minimal and spatial finite overestimation of low measured BCB concentrations, but on the other hand the advantage is that the boundary layers become more visible. By including negative attenuation, our evaluation of the growth data was not affected by this focused shadowgraphy effect. Furthermore, at high refractive gradient edges caused by a higher BCB concentration, the effect largely disappeared, since the light attenuation of the BCB reduced the amount of deflected light.

Since the attenuation coefficient of BCB is relatively high compared to other EPS, the manner observation was sufficient. But, when growing biofilm streamers consisting of other bacteria and EPS, the sensitivity of the EPS detection and thus the determination of the concentration could become an issue, when the attenuation coefficient is lower. Here, the focused shadowgraphy or even better the schlieren techniques could play a major role in the sensitivity enhancement. Simply inserting a knife edge in the optional optical filter ((9) in Fig. 1) with a cutoff of 80% would intensify the refractive gradient edge by a factor of 5. Changing the lens to $f = 500 \text{ mm}$ would enhance the factor with the same cutoff to 25.¹⁰³

5 Outlook

We investigated the rheotactically formation of a bacterial cellulose network consisting of several single bacterial streamers. The formation could be separated into two logistic growth phases. The first was the strain formation between



the streamers in the direction of the flow. In the second phase, the grown strains connected, building a hierarchically structured BCB network with flow-aligned BC fibers. The first phase was dependent on the flow rate, since a higher flow provided the streamers with more bacteria. In the second phase, a higher flow with its higher dynamic pressure resulted in a higher final BCB concentration. These findings may form the basis for an upscaled material production of anisotropic cellulose substrates. With the flow rate and design of the flow reactor the concentration and distribution of the aligned BCB in the strands and in the whole material can be adjusted. A BCB-material interwoven with tailored BCB-strands can then be adapted to meet specific mechanical requirements.

We also conducted a quasi *in situ* mineralisation route to trace crystal growth in the hierarchically organised BCB. With ACC and a supersaturated CaSO_4 solution, mineral deposition was obtained. CaCO_3 thereby showed a more uniform distribution than gypsum. However, with the deposition of the needle shaped gypsum crystals, the anisotropic substrate features could be continued from the organic to the inorganic phase. It is quite conceivable that other minerals *e.g.* from the group of montmorillonite, or a biomineralisation parth are sufficient for anisotropic hierarchical structured composites. These results illustrate that an evenly distributed mineral phase is attainable. This, in turn, can be utilised to tune the mechanical properties of bio-mimetic hierarchically structured BCB-mineral composite materials. Future applications of the obtained materials thereby could be manifold: in addition to obtaining tough materials, tailor-made surfaces for use as catalysis carriers, for defined adsorption tasks, or as a stationary phase in separating columns are also conceivable.

Author contributions

Moritz Klotz: data curation, formal analysis, investigation, methodology, validation, visualization, writing – original draft. Dardan Bajrami: software (CFD), formal analysis (CFD). Daniel Van Opdenbosch: conceptualization, funding acquisition, project administration, supervision, writing – review & editing.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

We thank the German Science Foundation (DFG) for funding our research *via* grant VA 1349/5.

References

- 1 P. Fratzl and R. Weinkamer, Nature's hierarchical materials, *Prog. Mater. Sci.*, 2007, **52**(8), 1263–1334.
- 2 C. Mattheck and I. Tesari, The mechanical self-optimisation of trees, *WIT Trans. Ecol. Environ.*, 2004, **73**, 10.

- 3 A. Öchsner, W. Ahmed, J. Fish and T. Belytschko, *Biomechanics of Hard Tissues*, Wiley Online Library, 2011.
- 4 G. E. Fantner, E. Oroudjev, G. Schitter, L. S. Golde, P. Thurner, M. M. Finch, P. Turner, T. Gutsman, D. E. Morse and H. Hansma, *et al.*, Sacrificial bonds and hidden length: Unraveling molecular mesostructures in tough materials, *Biophys. J.*, 2006, **90**(4), 1411–1418.
- 5 P. Greil, T. Lifka and A. Kaindl, Biomimetic cellular silicon carbide ceramics from wood: I. processing and microstructure, *J. Eur. Ceram. Soc.*, 1998, **18**(14), 1961–1973.
- 6 D. Van Opdenbosch, M. Thielen, R. Seidel, G. Fritz-Popovski, T. Fey, O. Paris, T. Speck and C. Zollfrank, The pomelo peel and derived nanoscale-precision gradient silica foams, *Bioinspired, Biomimetic Nanobiomater.*, 2012, **1**(2), 117–122.
- 7 G. Fritz-Popovski, R. Morak, T. Schöberl, D. Van Opdenbosch, C. Zollfrank and O. Paris, Pore characteristics and mechanical properties of silica templated by wood, *Bioinspired, Biomimetic Nanobiomater.*, 2014, **3**(3), 160–168.
- 8 D. Van Opdenbosch, G. Fritz-Popovski, W. Wagermaier, O. Paris and C. Zollfrank, Moisture-driven ceramic bilayer actuators from a biotemplating approach, *Adv. Mater.*, 2016, **28**(26), 5235–5240.
- 9 H. Imai, Self-organized formation of hierarchical structures, in *Biomineralization I*, Springer, 2006, pp. 43–72.
- 10 S. Deuerling, S. Kugler, M. Klotz, C. Zollfrank and D. Van Opdenbosch, A perspective on bio-mediated material structuring, *Adv. Mater.*, 2018, **30**(19), 1703656.
- 11 D.-P. Häder, Wie orientieren sich Cyanobakterien im Licht, *Biol. Unserer Zeit*, 1984, **14**(3), 78–83.
- 12 J. Arlt, V. A. Martinez, A. Dawson, T. Pilizota and W. C. Poon, Painting with light-powered bacteria, *Nat. Commun.*, 2018, **9**, 1–7.
- 13 M. Klotz, S. Deuerling, S. Kugler, C. Zollfrank and D. Van Opdenbosch, light-diffractive patterning of porphyrinium purpureum, *Photochem. Photobiol. Sci.*, 2020, 22–34.
- 14 X.-X. Wang, L. Xie and R. Wang, Biological fabrication of nacreous coating on titanium dental implant, *Biomaterials*, 2005, **26**(31), 6229–6232.
- 15 R. Rusconi, S. Lecuyer, N. Autrusson, L. Guglielmini and H. A. Stone, Secondary flow as a mechanism for the formation of biofilm streamers, *Biophys. J.*, 2011, **100**(6), 1392–1399.
- 16 Y. Uraki, J. Nemoto, H. Otsuka, Y. Tamai, J. Sugiyama, T. Kishimoto, M. Ubukata, H. Yabu, M. Tanaka and M. Shimomura, Honeycomb-like architecture produced by living bacteria, *Gluconacetobacter xylinus*, *Carbohydr. Polym.*, 2007, **69**(1), 1–6.
- 17 N. Oxman, J. Laucks, M. Kayser, J. Duro-Royo and C. Gonzales-Urbe, Silk pavilion: a case study in fiber-based digital fabrication, in *FABRICATE Conference Proceedings*, ta Verla, 2014, 248–255.
- 18 J. Loehr, D. Pfeiffer, D. Schöler and T. M. Fischer, Magnetic guidance of the magnetotactic bacterium *Magnetospirillum gryphiswaldense*, *Soft Matter*, 2016, **12**(15), 3631–3635.
- 19 C. Pierce, E. Mumper, E. Brown, J. Brangham, B. Lower, S. Lower, F. Yang and R. Sooryakumar, Tuning bacterial



- hydrodynamics with magnetic fields, *Phys. Rev. E*, 2017, **95**(6), 062612.
- 20 E. Bonabeau, G. Theraulaz, J.-L. Deneubourg, S. Aron and S. Camazine, Selforganization in social insects, *Trends Ecol. Evol.*, 1997, **12**(5), 188–193.
 - 21 Z. Lewandowski and H. Beyenal, *Fundamentals of Biofilm Research*, CRC press, 2013.
 - 22 I. Dogsa, M. Kriechbaum, D. Stopar and P. Laggnier, Structure of bacterial extracellular polymeric substances at different pH values as determined by SAXS, *Biophys. J.*, 2005, **89**(4), 2711–2720.
 - 23 T. Coviello, W. Burchard, E. Geissler and D. Maier, Static and dynamic light scattering by a thermoreversible gel from *Rhizobium leguminosarum* 8002 exopolysaccharide, *Macromolecules*, 1997, **30**(7), 2008–2015.
 - 24 S. Koizumi, Y. Tomita, T. Kondo and T. Hashimoto, What factors determine hierarchical structure of microbial cellulose -interplay among physics, chemistry and biology, in *Macromolecular Symposia*, Wiley Online Library, 2009, vol. 279, 1, pp. 110–118.
 - 25 P. Zugenmaier, *Crystalline cellulose and derivatives: characterization and structures*, Berlin Heidelberg, Springer, 2008.
 - 26 T. Kondo and W. Kasai, Autonomous bottom-up fabrication of three-dimensional nano/microcellulose honeycomb structures, directed by bacterial nanobuilder, *J. Biosci. Bioeng.*, 2014, **118**(4), 482–487.
 - 27 T. Kondo, M. Nojiri, Y. Hishikawa, E. Togawa, D. Romanovicz and R. M. Brown, Biodirected epitaxial nanodeposition of polymers on oriented macromolecular templates, *Proc. Natl. Acad. Sci. U. S. A.*, 2002, **99**(22), 14008–14013.
 - 28 A. Nagashima, T. Tsuji and T. Kondo, A uniaxially oriented nanofibrous cellulose scaffold from pellicles produced by *Gluconacetobacter xylinus* in dissolved oxygen culture, *Carbohydr. Polym.*, 2016, **135**, 215–224.
 - 29 A. Putra, A. Kakugo, H. Furukawa, J. P. Gong, Y. Osada, T. Uemura and M. Yamamoto, Production of bacterial cellulose with well oriented fibril on PDMS substrate, *Polym. J.*, 2008, **40**(2), 137–142.
 - 30 S. Zang, Z. Sun, K. Liu, G. Wang, R. Zhang, B. Liu and G. Yang, Ordered manufactured bacterial cellulose as biomaterial of tissue engineering, *Mater. Lett.*, 2014, **128**, 314–318.
 - 31 J. Yang, L. Wang, W. Zhang, Z. Sun, Y. Li, M. Yang, D. Zeng, B. Peng, W. Zheng and X. Jiang, *et al.*, Reverse reconstruction and bioprinting of bacterial cellulose-based functional total intervertebral disc for therapeutic implantation, *Small*, 2018, **14**(7), 1702582.
 - 32 A. Putra, A. Kakugo, H. Furukawa, J. P. Gong and Y. Osada, Tubular bacterial cellulose gel with oriented fibrils on the curved surface, *Polymer*, 2008, **49**(7), 1885–1891.
 - 33 A. Putra, A. Kakugo, H. Furukawa and J. P. Gong, Oriented bacterial cellulose culture controlled by liquid substrate of silicone oil with different viscosity and thickness, *Polym. J.*, 2009, **41**(9), 764–770.
 - 34 M. B. Sano, A. D. Rojas, P. Gatenholm and R. V. Davalos, Electromagnetically controlled biological assembly of aligned bacterial cellulose nanofibers, *Ann. Biomed. Eng.*, 2010, **38**(8), 2475–2484.
 - 35 M. Liu, C. Zhong, X. Zheng, L. Ye, T. Wan and S. R. Jia, Oriented bacterial cellulose/glass fiber nanocomposites with enhanced tensile strength through electric field, *Fibers Polym.*, 2017, **18**(7), 1408–1412.
 - 36 Y. Wan, D. Hu, G. Xiong, D. Li, R. Guo and H. Luo, Directional fluid induced self-assembly of oriented bacterial cellulose nanofibers for potential biomimetic tissue engineering scaffolds, *Mater. Chem. Phys.*, 2015, **149**, 7–11.
 - 37 H. Luo, W. Li, Z. Yang, H. Ao, G. Xiong, Y. Zhu, J. Tu and Y. Wan, Preparation of oriented bacterial cellulose nanofibers by flowing medium-assisted biosynthesis and influence of flowing velocity, *J. Polym. Eng.*, 2018, **38**(3), 299–305.
 - 38 I. Chae, S. M. Bokhari, X. Chen, R. Zu, K. Liu, A. Borhan, V. Gopalan, J. M. Catchmark and S. H. Kim, Shear-induced unidirectional deposition of bacterial cellulose microfibrils using rising bubble stream cultivation, *Carbohydr. Polym.*, 2021, **255**, 117328.
 - 39 S. Das and A. Kumar, Formation and post-formation dynamics of bacterial biofilm streamers as highly viscous liquid jets, *Sci. Rep.*, 2014, **4**(1), 1–6.
 - 40 A. Persat, C. D. Nadell, M. K. Kim, F. Ingremeau, A. Sryaporn, K. Drescher, N. S. Wingreen, B. L. Bassler, Z. Gitai and H. A. Stone, The mechanical world of bacteria, *Cell*, 2015, **161**(5), 988–997.
 - 41 A. Karimi, D. Karig, A. Kumar and A. Ardekani, Interplay of physical mechanisms and biofilm processes: Review of microfluidic methods, *Lab Chip*, 2015, **15**(1), 23–42.
 - 42 Y. Yawata, J. Nguyen, R. Stocker and R. Rusconi, Microfluidic studies of biofilm formation in dynamic environments, *J. Bacteriol.*, 2016, **198**(19), 2589–2595.
 - 43 M. G. Mazza, The physics of biofilms: An introduction, *J. Phys. D: Appl. Phys.*, 2016, **49**(20), 203001.
 - 44 P. Thomen, J. Robert, A. Monmeyran, A.-F. Bitbol, C. Douarche and N. Henry, Bacterial biofilm under flow: First a physical struggle to stay, then a matter of breathing, *PLoS One*, 2017, **12**(4), 1–24.
 - 45 K. Drescher, Y. Shen, B. L. Bassler and H. A. Stone, Biofilm streamers cause catastrophic disruption of flow with consequences for environmental and medical systems, *Proc. Natl. Acad. Sci. U. S. A.*, 2013, **110**(11), 4345–4350.
 - 46 A. Marty, C. Roques, C. Causserand and P. Bacchin, Formation of bacterial streamers during filtration in microfluidic systems, *Biofouling*, 2012, **28**(6), 551–562.
 - 47 R. Rusconi, S. Lecuyer, L. Guglielmini and H. A. Stone, Laminar flow around corners triggers the formation of biofilm streamers, *J. R. Soc., Interface*, 2010, **7**(50), 1293–1299.
 - 48 D. R. Espeso and A. M.-C. Rodriguez, Modeling and simulation of bacterial biofilms, *PhD dissertation*, Universidad Carlos III de Madrid, 2013.
 - 49 N. B. Aznavah, M. Safdar, G. Wolfaardt and J. Greener, Micropatterned biofilm formations by laminar flow-templating, *Lab Chip*, 2014, **14**(15), 2666–2672.
 - 50 P. Stoodley, R. Cargo, C. J. Rupp, S. Wilson and I. Klapper, Biofilm material properties as related to shear-induced



- deformation and detachment phenomena, *J. Ind. Microbiol. Biotechnol.*, 2002, **29**(6), 361–367.
- 51 M. Marcos, H. Fu, T. Powers and R. Stocker, Bacterial rheotaxis, *Bull. Am. Phys. Soc.*, 2011, **109**, 4780–4785.
 - 52 J. D. Wheeler, E. Secchi, R. Rusconi and R. Stocker, Not just going with the flow: The effects of fluid flow on bacteria and plankton, *Annu. Rev. Cell Dev. Biol.*, 2019, **35**, 213–237.
 - 53 J. A. Aufrecht, J. D. Fowlkes, A. N. Bible, J. Morrell-Falvey, M. J. Doktycz and S. T. Retterer, Pore-scale hydrodynamics influence the spatial evolution of bacterial biofilms in a microfluidic porous network, *PLoS One*, 2019, **14**(6), e0218316.
 - 54 M. Hassanpourfard, Z. Nikakhtari, R. Ghosh, S. Das, T. Thundat, Y. Liu and A. Kumar, Bacterial floc mediated rapid streamer formation in creeping flows, *Sci. Rep.*, 2015, **5**(1), 1–12.
 - 55 M. Hassanpourfard, R. Ghosh, T. Thundat and A. Kumar, Dynamics of bacterial streamers induced clogging in microfluidic devices, *Lab Chip*, 2016, **16**(21), 4091–4096.
 - 56 N. Shah, M. Ul-Islam, W. A. Khattak and J. K. Park, Overview of bacterial cellulose composites: A multipurpose advanced material, *Carbohydr. Polym.*, 2013, **98**(2), 1585–1598.
 - 57 W. Liu, H. Du, M. Zhang, K. Liu, H. Liu, H. Xie, X. Zhang and C. Si, Bacterial cellulosebased composite scaffolds for biomedical applications: A review, *ACS Sustainable Chem. Eng.*, 2020, **8**(20), 7536–7562.
 - 58 Z. Cheng, Z. Ye, A. Natan, Y. Ma, H. Li, Y. Chen, L. Wan, C. Aparicio and H. Zhu, Boneinspired mineralization with highly aligned cellulose nanofibers as template, *ACS Appl. Mater. Interfaces*, 2019, **11**(45), 42486–42495.
 - 59 F. Mohammadkazemi, M. Faria and N. Cordeiro, *In situ* biosynthesis of bacterial nanocellulose-CaCO₃ hybrid bio-nanocomposite: One-step process, *Mater. Sci. Eng., C*, 2016, **65**, 393–399.
 - 60 G. Serafica, R. Mormino and H. Bungay, Inclusion of solid particles in bacterial cellulose, *Appl. Microbiol. Biotechnol.*, 2002, **58**(6), 756–760.
 - 61 S. Yano, H. Maeda, M. Nakajima, T. Hagiwara and T. Sawaguchi, Preparation and mechanical properties of bacterial cellulose nanocomposites loaded with silica nanoparticles, *Cellulose*, 2008, **15**(1), 111–120.
 - 62 M. Pomet, J. Juntaro, J. Y. Heng, A. Mantalaris, A. F. Lee, K. Wilson, G. Kalinka, M. S. Shaffer and A. Bismarck, Surface modification of natural fibers using bacteria: Depositing bacterial cellulose onto natural fibers to create hierarchical fiber reinforced nanocomposites, *Biomacromolecules*, 2008, **9**(6), 1643–1651.
 - 63 S. Shi, S. Chen, X. Zhang, W. Shen, X. Li, W. Hu and H. Wang, Biomimetic mineralization synthesis of calcium-deficient carbonate-containing hydroxyapatite in a three-dimensional network of bacterial cellulose, *J. Chem. Technol. Biotechnol.*, 2009, **84**(2), 285–290.
 - 64 Y. Wan, Y. Huang, C. Yuan, S. Raman, Y. Zhu, H. Jiang, F. He and C. Gao, Biomimetic synthesis of hydroxyapatite/bacterial cellulose nanocomposites for biomedical applications, *Mater. Sci. Eng., C*, 2007, **27**(4), 855–864.
 - 65 M. Ul-Islam, T. Khan and J. K. Park, Nanoreinforced bacterial cellulose-montmorillonite composites for biomedical applications, *Carbohydr. Polym.*, 2012, **89**(4), 1189–1197.
 - 66 N. Khodamoradi, V. Babaeipour and M. Sirousazar, Bacterial cellulose/montmorillonite bionanocomposites prepared by immersion and in-situ methods: Structural, mechanical, thermal, swelling and dehydration properties, *Cellulose*, 2019, **26**(13), 7847–7861.
 - 67 J. Sun and B. Bhushan, Hierarchical structure and mechanical properties of nacre: A review, *RSC Adv.*, 2012, **2**(20), 7617–7632.
 - 68 M. W. Rauch, M. Dressler, H. Scheel, D. Van Opdenbosch and C. Zollfrank, Mineralization of calcium carbonates in cellulose gel membranes, *Eur. J. Inorg. Chem.*, 2012, (32), 5192–5198.
 - 69 D. Kuo, T. Nishimura, S. Kajiyama and T. Kato, Bioinspired environmentally friendly amorphous CaCO₃-based transparent composites comprising cellulose nanofibers, *ACS Omega*, 2018, **3**(10), 12722–12729.
 - 70 D. Gebauer, V. Olynyk, M. Salajkova, J. Sort, Q. Zhou, L. Bergström and G. Salazar-Alvarez, A transparent hybrid of nanocrystalline cellulose and amorphous calcium carbonate nanoparticles, *Nanoscale*, 2011, **3**(9), 3563–3566.
 - 71 T. Saito, Y. Oaki, T. Nishimura, A. Isogai and T. Kato, Bioinspired stiff and flexible composites of nanocellulose-reinforced amorphous CaCO₃, *Mater. Horiz.*, 2014, **1**(3), 321–325.
 - 72 M. Stroescu, A. Stoica-Guzun, S. I. Jinga, T. Dobre, I. M. Jipa and L. M. Dobre, Influence of sodium dodecyl sulfate and cetyl trimethylammonium bromide upon calcium carbonate precipitation on bacterial cellulose, *Korean J. Chem. Eng.*, 2012, **29**(9), 1216–1223.
 - 73 Q. Zhu, J. Wang, J. Sun and Q. Wang, Preparation and characterization of regenerated cellulose biocomposite film filled with calcium carbonate by *in situ* precipitation, *BioResources*, 2020, **15**(4), 7893.
 - 74 M. Ciobanu, E. Bobu and F. Ciolacu, *In situ* cellulose fibres loading with calcium carbonate precipitated by different methods, *Cellul. Chem. Technol.*, 2010, **44**(9), 379.
 - 75 A. Stoica-Guzun, M. Stroescu, S. Jinga, I. Jipa, T. Dobre and L. Dobre, Ultrasound influence upon calcium carbonate precipitation on bacterial cellulose membranes, *Ultrason. Sonochem.*, 2012, **19**(4), 909–915.
 - 76 T. Nissinen, M. Li, N. Brielles and S. Mann, Calcium sulfate hemihydrate-mediated crystallization of gypsum on Ca²⁺-activated cellulose thin films, *CrystEngComm*, 2013, **15**(19), 3793–3798.
 - 77 M. Iguchi, S. Yamanaka and A. Budhiono, Bacterial cellulose: A masterpiece of nature's arts, *J. Mater. Sci.*, 2000, **35**(2), 261–270.
 - 78 A. D. Edelstein, M. A. Tsuchida, N. Amodaj, H. Pinkard, R. D. Vale and N. Stuurman, Advanced methods of microscope control using µManager software, *J. Biol. Methods*, 2014, **1**(2), 1–18.
 - 79 J. Bergmann, P. Friedel and R. Kleeberg, BGMN – a new fundamental parameters based Rietveld program for



- laboratory X-ray sources, its use in quantitative analysis and structure investigations, *CPD Newsletter*, 1998, **20**, 5–8.
- 80 N. Doeblin and R. Kleeberg, Profex: A graphical user interface for the Rietveld refinement program BGMN, *J. Appl. Crystallogr.*, 2015, **48**(5), 1573–1580.
 - 81 M. Järvinen, Texture models in powder diffraction analysis, in *Materials Science Forum*, Trans Tech Publ, 1998, vol. 278, pp. 184–199.
 - 82 J. H. Lienhard, *et al.*, Synopsis of lift, drag, and vortex frequency data for rigid circular cylinders, *Technical Extension Service*, Washington State University Pullman, WA, 1966, vol. 300.
 - 83 C. Campano, A. Balea, A. Blanco and C. Negro, Enhancement of the fermentation process and properties of bacterial cellulose: A review, *Cellulose*, 2016, **23**(1), 57–91.
 - 84 W. Czaja, D. Romanovicz and R. Malcolm Brown, Structural investigations of microbial cellulose produced in stationary and agitated culture, *Cellulose*, 2004, **11**(3), 403–411.
 - 85 P. Singhsa, R. Narain and H. Manuspiya, Physical structure variations of bacterial cellulose produced by different *Komagataeibacter xylinus* strains and carbon sources in static and agitated conditions, *Cellulose*, 2018, **25**(3), 1571–1581.
 - 86 M. Krsmanovic, D. Biswas, H. Ali, A. Kumar, R. Ghosh and A. K. Dickerson, Hydrodynamics and surface properties influence biofilm proliferation, *Adv. Colloid Interface Sci.*, 2020, 102336.
 - 87 M. Liu, C. Zhong, X.-Y. Wu, Y.-Q. Wei, T. Bo, P.-P. Han and S.-R. Jia, Metabolomic profiling coupled with metabolic network reveals differences in *Gluconacetobacter xylinus* from static and agitated cultures, *Biochem. Eng. J.*, 2015, **101**, 85–98.
 - 88 M. R. Nejadnik, H. C. Van Der Mei, H. J. Busscher and W. Norde, Determination of the shear force at the balance between bacterial attachment and detachment in weakadherence systems, using a flow displacement chamber, *Appl. Environ. Microbiol.*, 2008, **74**(3), 916–919.
 - 89 J. Kuncova-Kallio and P. J. Kallio, PDMS and its suitability for analytical microfluidic devices, in 2006 International Conference of the IEEE Engineering in Medicine and Biology Society. IEEE, 2006, 2486–2489.
 - 90 D. J. Gardner, G. S. Oporto, R. Mills and M. A.-S. A. Samir, Adhesion and surface issues in cellulose and nanocellulose, *J. Adhes. Sci. Technol.*, 2008, **22**(5–6), 545–567.
 - 91 A. Marty, C. Causserand, C. Roques and P. Bacchin, Impact of tortuous flow on bacteria streamer development in microfluidic system during filtration, *Biomicrofluidics*, 2014, **8**(1), 014105.
 - 92 R. Holm and D. Söderberg, Shear influence on fibre orientation, *Rheol. Acta*, 2007, **46**(5), 721–729.
 - 93 S. B. Lindström and T. Uesaka, Simulation of semidilute suspensions of non-brownian fibers in shear flow, *J. Chem. Phys.*, 2008, **128**(2), 024901.
 - 94 F. Nindiyasari, E. Griesshaber, T. Zimmermann, A. P. Manian, C. Randow, R. Zehbe, L. Fernandez-Diaz, A. Ziegler, C. Fleck and W. W. Schmahl, Characterization and mechanical properties investigation of the cellulose/gypsum composite, *J. Compos. Mater.*, 2016, **50**(5), 657–672.
 - 95 A. Van Driessche, L. G. Benning, J. Rodriguez-Blanco, M. Ossorio, P. Bots and J. García-Ruiz, The role and implications of bassanite as a stable precursor phase to gypsum precipitation, *Science*, 2012, **336**(6077), 69–72.
 - 96 H. K. Raut, A. F. Schwartzman, R. Das, F. Liu, L. Wang, C. A. Ross and J. G. Fernandez, Tough and strong: Cross-lamella design imparts multifunctionality to biomimetic nacre, *ACS Nano*, 2020, **14**(8), 9771–9779.
 - 97 A.-W. Xu, M. Antonietti, H. Cölfen and Y.-P. Fang, Uniform hexagonal plates of vaterite CaCO₃ mesocrystals formed by biomimetic mineralization, *Adv. Funct. Mater.*, 2006, **16**(7), 903–908.
 - 98 O. Grassmann and P. Löbmann, Morphogenetic control of calcite crystal growth in sulfonic acid based hydrogels, *Chem. – Eur. J.*, 2003, **9**(6), 1310–1316.
 - 99 K. Yu, E. M. Spiesz, S. Balasubramanian, D. T. Schmieden, A. S. Meyer and M.-E. Aubin-Tam, Scalable bacterial production of moldable and recyclable biomineralized cellulose with tunable mechanical properties, *Cell Rep. Phys. Sci.*, 2021, 100464.
 - 100 Y. Wan, J. Wang, M. Gama, R. Guo, Q. Zhang, P. Zhang, F. Yao and H. Luo, Biofabrication of a novel bacteria/bacterial cellulose composite for improved adsorption capacity, *Composites, Part A*, 2019, **125**, 105560.
 - 101 H. Kanematsu and D. M. Barry, *Formation and control of biofilm in various environments*, Singapore, Springer, 2020.
 - 102 G. S. Settles and E. Covert, Schlieren and shadowgraph techniques: Visualizing phenomena in transport media, *Appl. Mech. Rev.*, 2002, **55**(4), B76–B77.
 - 103 G. S. Settles, *Schlieren and Shadowgraph Techniques*, Berlin Heidelberg New York, Springer, 2006.

