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**Microbial Vesicle-Mediated Communication: Convergence to understand interactions within and between domains of life**

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3 **Microbial Vesicle-Mediated Communication: Convergence to understand**  
4 **interactions within and between domains of life**  
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43 Keywords: Extracellular Vesicles, Health and Safety of Environmental Nanomaterials,  
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45 soil science, convergence  
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49 ***Environmental Significance Statement***  
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51 Cells secrete extracellular vesicles (EVs), nanoscale biological packages that contain  
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53 complex mixtures of molecular cargo. The multiple roles of microbial EVs include their  
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3 function as carriers for molecular messengers that facilitate interspecies communication  
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5 and have been studied extensively in mammalian systems. For environmental systems,  
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7 however, the prevalence, characteristics, and functions of these biological particles are  
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9 only now being revealed. Here, we argue that the study of microbial EVs in the  
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11 environment requires biochemical insights from studies of donor and receiving  
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13 organisms as well as knowledge of soft colloid mobility and interactions with other  
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15 components of the environment. Such questions of EV function, transport, and  
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17 environmental impact can be addressed best by harnessing theories and methodologies  
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19 developed by the biological, colloid, and geochemical sciences.  
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## 26 ***Abstract***

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28 All cells produce extracellular vesicles (EVs). These biological packages contain  
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30 complex mixtures of molecular cargo and have a variety of functions, including  
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32 interkingdom communication. Recent discoveries highlight the roles microbial EVs may  
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34 play in the environment with respect to interactions with plants as well as nutrient  
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36 cycling. These studies have also identified molecules present within EVs and  
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38 associated with EV surfaces that contribute to these functions. In parallel, studies of  
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40 engineered nanomaterials have developed methods to track and model small particle  
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42 behavior in complex systems and measure the relative importance of various surface  
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44 features on transport and function. While studies of EV behavior in complex  
45  
46 environmental conditions have not yet employed transdisciplinary approaches, it is  
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48 increasingly clear that expertise from disparate fields will be critical to understand the  
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50 role of EVs in these systems. Here, we outline how the convergence of biology, soil  
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3 geochemistry, and colloid science can both develop and address questions surrounding  
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5 the basic principles governing EV-mediated interkingdom interactions.  
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### 10 ***Introduction***

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12 In every ecosystem, at all scales, life forms communicate constantly with each other  
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14 and their environment, often using secreted components to react to, affect, and  
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16 exchange information about their surroundings. These interactions ultimately shape  
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18 higher order organismal and ecosystem function. Combinations of chemical, physical,  
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20 and biological processes govern this interkingdom communication and, as such, these  
21  
22 processes can be understood only through a synergistic combination of methods and  
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24 insight from multiple disciplinary perspectives. Indeed, with theory and methodology  
25  
26 from just one field, it would be impossible to develop or investigate critical questions  
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28 about these complex interactions in the context of environmental systems. Despite the  
29  
30 obvious need for collaboration, thus far studies of interkingdom communication have  
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32 remained relatively constrained to singular or similar disciplines. This predicament is  
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34 exemplified in studies of extracellular vesicles (EVs).  
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42 EVs are nanoscale proteoliposomes that are secreted by all forms of life and are a  
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44 ubiquitous part of every environment (1, 2). Compared to other cellular secretions, EVs  
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46 uniquely enable the simultaneous interaction of a broad mixture of molecular cargo  
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48 embedded and enclosed within the EV with the surroundings (2-8). These biologically  
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50 complex nanostructures present a convergence challenge since a complete  
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52 understanding of their function requires expertise from an array of disciplines. Here, we  
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3 discuss how a multidisciplinary approach can address this challenge in bacterial EV  
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5 studies.  
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10 To date, most bacterial EV studies focus on their activities in mammalian systems (9,  
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12 10) whereas EV research in environmental systems is still emerging, with only a few  
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14 studies considering EV's roles in microbial communities and plant systems (11-16).  
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17 Despite the recent expansion of EV research into plant systems, interactions between  
18  
19 EVs and surrounding soil systems remain under-investigated. Our understanding of EVs  
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21 in complex conditions can be expanded by investigating the soil rhizosphere  
22  
23 environment. Rhizosphere ecosystem functions rely on interkingdom interactions, soil  
24  
25 composition, primary production, and major element cycling in natural and managed  
26  
27 terrestrial landscapes. Thus, understanding EV roles in this complex environment  
28  
29 requires a rich area for interdisciplinary collaboration (Figure 1). Substantial  
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31 advancement in understanding the role of EVs in the environment will require answers  
32  
33 to fundamental questions: What are the relevant characteristics of EVs to relate their  
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35 biogenesis, transport, and function? How do these change in response to environmental  
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37 conditions? How do these changes impact ecosystem function?  
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44 Having diameters as small as 20 nm, many EVs may be considered naturally occurring  
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46 biotic nanoparticles as nanomaterials are defined as any material that possesses at  
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48 least one dimension between 1-100 nm. Questions tackled under the umbrella of the  
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50 environmental health and safety of nanomaterials (nanoEHS) exemplify the advantages  
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52 of a convergence approach to science. Here, findings rooted in areas such as colloid  
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3 science, environmental biogeochemistry, ecology and toxicology provided theoretical  
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5 frameworks and methods to inform first principles investigations (17). NanoEHS  
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7 research has highlighted the fact that environmental conditions surrounding nanoscale  
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9 and microscopic materials control their surface chemical properties and subsequently  
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11 can govern their transport and attachment to other surfaces (18). Intriguingly, linking EV  
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13 properties to findings in NanoEHS research, recent work on biological EVs has revealed  
14  
15 how function relies on properties of the EV exterior for transport and attachment to other  
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17 surfaces, and that the environmental context of the cells governs what is exposed on  
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19 the EV surface (14, 19-22). We anticipate that an intersection of disciplinary domains  
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21 will generate clarity and novel concepts in the understanding of the roles and activities  
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23 of EVs.  
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31 Significant new understanding of EV transport, fate, and function could be gained by  
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33 studying biological phenomena of EVs in the context of nanomaterial and  
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35 biogeochemical processes in the environment. Here we introduce some foundational  
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37 and emerging insights that will contribute to our transdisciplinary approach to  
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39 understanding, predicting, and ultimately harnessing EV-mediated interkingdom  
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41 communication in soils.  
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### 47 ***EV Surfaces, Cargo, and Biological Function***

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49 While EVs require considerable energy to produce, they also play important roles in  
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51 inter-kingdom communication (10, 23-26). EVs include membrane vesicles, outer  
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53 membrane vesicles, and exosomes, among other similar structures (Figure 2). In  
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3 microbial systems, EVs bud from the outermost cell membrane and are packaged with  
4 specific cargo (9, 27-30). In mammalian and potentially plant systems, EVs can be  
5 produced by budding from the cell membrane or through pathways originating at  
6 intracellular, multivesicular bodies (31-34). While there are innumerable differences in  
7 the generation, composition, and biological function amongst these different types of  
8 vesicles, many overarching similarities exist, enabling high-level comparisons of  
9 disparate EV populations.  
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21 The universal concepts of EV functional features that are governed by their surface  
22 composition and cargo can be seen using Gram-negative bacterial EVs as a case  
23 study. Termed outer membrane vesicles (OMVs) based on their bacterial outer  
24 membrane origin, these EVs are typically much smaller (20-200 nm in diameter) than  
25 eukaryotic EVs (20-1000 nm in diameter) (Figure 2). Their cargo of enriched, depleted,  
26 and “bulk flow” amounts of cellular envelope components relative to the producing cell  
27 reveal that they are products of a regulated secretory process (9, 35-37). OMVs are  
28 secreted for a variety of reasons, including secretion of misfolded proteins, membrane  
29 remodeling, nutrient acquisition, communication, and as decoys for phage and antibiotic  
30 molecules (9, 38, 39). Additionally, host-pathogen studies show that bacterial EVs can  
31 deliver cocktails of virulence factors into host cells and activate host immune responses  
32 (5, 28, 40-42). Importantly, bacterial EV-elicited immunogenicity has been exploited for  
33 use in vaccine development (38, 43).  
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3 A few studies have begun to elucidate bacterial EV function in plant systems or the  
4 natural environment (13, 44). Just like all other bacteria, phyto-bacteria produce EVs  
5 that have the potential to interact with plants in the phyllosphere and rhizosphere.  
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7 Several phyto-bacterial EV proteomes have been characterized, revealing that these  
8 EVs contain numerous metabolites as well as virulence factors (16, 45-48).  
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10 Additionally, some of this cargo, such as cellulases and xylanases, retains its ability to  
11 digest plant cell wall components (45, 49). As in mammalian systems, bacterial EVs  
12 also activate plant immune responses. EVs from several different bacterial plant  
13 pathogens activate general plant innate immune responses including a reactive oxygen  
14 species burst and transcription of pattern recognition receptors (11). This work also  
15 showed some dependence on the co-receptors BAK1 and SOBIR1 for EV-mediated  
16 immune activation, though a complete mechanism has not yet been identified (11).  
17  
18 Recent studies extended these initial findings to demonstrate that EVs from the model  
19 plant pathogen *Pseudomonas syringae* activate plant immune responses including *ICS1*  
20 transcription and salicylic acid production that result in protection from future pathogen  
21 attack (15, 16). Furthermore, it was shown for the first time that EVs from plant  
22 beneficial bacteria such as *Pseudomonas fluorescens* also lead to protection from  
23 pathogen attack, though they do so through plant immune pathways different from those  
24 activated by plant pathogen EVs (15). Together, these findings add a new layer of  
25 complexity to plant-microbe interactions and reveal a previously unstudied role for EVs  
26 in the environment, likely highlighting only a very small part of EV's overall functional  
27 contributions.  
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3 Surface properties mediate the physical interactions between bacterial EVs and the  
4 plant host and consequently can substantially impact the host-pathogen relationship.  
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6 OMV surfaces consist of a subset of outer membrane components, lipopolysaccharide,  
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8 phospholipids, membrane proteins, and associated small molecules and metal ions (9,  
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10 35). This composition mediates the ability of bacterial EVs to adhere to plant surfaces,  
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12 which contributes to increased bacterial spread and virulence (14, 21, 22, 50). For  
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14 example, OMVs from the pathogenic bacterium *Xylella fastidiosa* adhere to xylem cell  
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16 walls, limiting the ability of the *X. fastidiosa* cells to attach and form bacterial  
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18 communities (biofilms), and facilitating spread throughout the xylem, which leads to  
19  
20 increased infection of plant tissue (14). Additionally, secreted hydrophobic molecules  
21  
22 are found in association with OMVs along with lipases, esterases, and other cargo that  
23  
24 could aid in plant cell wall degradation and virulence (21, 22). OMV surface components  
25  
26 may also confer a particular charge, which could result in attraction to or repulsion from  
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28 plant cell wall components as well as components of the phyllosphere and rhizosphere  
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30 environment. While early studies reveal the potential of bacterial EVs to adhere to  
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32 surfaces, future work incorporating EV charge and surface composition should be useful  
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34 to predict transport and fate in biological systems.  
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44 In the natural environment, microbes must adapt to chemical and physical changes. A  
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46 notable role for EV production in microbial adaptation to changing environments is  
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48 highlighted from studies of *X. fastidiosa* OMVs (14). *X. fastidiosa* is transmitted via  
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50 sharpshooter insect vectors and, therefore, survives by interacting differently with  
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52 surfaces in two very different environments: the insect gut and the plant xylem (51, 52).  
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3 In the insect, the bacterium must adhere tightly to the host to withstand significant sheer  
4 force as the insect feeds and sap flows through its mouth parts (14, 53, 54). In contrast,  
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6 if the bacteria attached firmly to the xylem cell wall, it would be unable to spread  
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8 throughout the host plant (14). Thus, EV production presents an opportunity through  
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10 which *X. fastidiosa* modulates its attachment in response to its environment (14, 22).  
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12 Several other reports further document the use of EVs as microbial tools for responding  
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14 to environment change (55-58).  
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21 These studies have revealed several governing properties of EV function in complex  
22 systems within the phyllosphere – namely, that cargo within the EV and associated with  
23 the EV surface influences interactions with hosts and attachment to biological surfaces,  
24 and that EVs play a role in survival of the producing cell during environmental  
25 transitions. The same factors likely play a role in the context of the soil rhizosphere,  
26 influencing how EVs interact with and are changed by plant roots, soil particles,  
27 nutrients, and groundwater. It remains largely unknown what factors govern EV  
28 packaging and release into the environment, and how EV cargo may affect the abiotic  
29 or living constituents in the environment. These questions pertain not only to the  
30 rhizosphere, but also to other habitats that harbor active microbial communities.  
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### 47 ***EVs in the Context of Element Biogeochemical Cycles***

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49 Within the emerging understanding of EV characteristics and biological function, the  
50 roles of EVs in environmental processes remain at an early stage of exploration.  
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52 Nevertheless, evidence is mounting that EVs could influence the soil rhizosphere, for  
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3 example by participating in the biogeochemical cycling of major and trace elements (59-  
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5 64). Microbes play critical roles in nutrient cycling by inducing changes to various soil  
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7 elements through processes like nitrogen fixation. More specifically, it is the membranes  
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9 of the microbes that are critical in performing a variety of these critical functions,  
10  
11 including sequestering limited nutrients, such as iron and trace metals, from the  
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13 environment for delivery into the cell, and contributing to the role microorganisms play  
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15 by regulating the concentrations and availability of soil constituents (63, 65-68).  
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19 Membrane properties of bacteria and their secreted EVs are similar, as many of the  
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21 same molecules found in the outermost microbial membrane are also found in EVs (19,  
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23 35, 69, 70). Therefore, it is important to consider whether EVs may play supplementary,  
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25 complementary, or redundant roles in biogeochemical regulation and cycling. As with  
26  
27 other biological functions, the production of EVs with such functionalities entails great  
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29 energetic costs for microorganisms in terms of expelling macromolecular complexes,  
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31 especially when compared to similar cellular processes enabled by membrane-bound  
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33 proteins at the cell envelope (61, 63). Thus, the concept of EVs with scavenging and  
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35 element cycling functionalities needs to be investigated in the context of competitive  
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37 advantages compared to parent cell capabilities.  
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45 EV involvement in element biogeochemical processes has recently been revealed in  
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47 several microbial species. For example, *Geobacter sulfurreducens* produces EVs with  
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49 membrane proteins capable of extracellular electron transfer (62), a process known to  
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51 occur at the cell envelope of the parent organism. Likewise, the pathogen  
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53 *Mycobacterium tuberculosis* produces EVs packaged with membrane-bound  
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3 siderophores that have high affinity to bind iron, similar to those associated with the cell  
4 envelope (61). These EVs are produced under iron-limited conditions as a mechanism  
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6 to sequester the metal and deliver it to the EV-producing cell or a nearby cell of the  
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8 same species (61). Similar phenomena for acquisition of sparingly soluble elements  
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10 also occur in soil-relevant microorganisms such as *Pseudomonas aeruginosa*, which  
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12 uses EVs as an intermediate step in iron acquisition (63).  
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19 EVs also contribute to biogeochemical cycling of carbon and major substrate nutrients.  
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21 For example *Prochlorococcus* species, a ubiquitous marine cyanobacterium that is  
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23 responsible for a notable proportion of Earth's photosynthetic activity, produce EVs in  
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25 abundances that are 1 to 10 times the number of whole cells in sea water (64). These  
26  
27 EVs are able to sustain and promote the growth of heterotrophic bacteria in culture,  
28  
29 indicating the potential of EVs to function as an intermediary of ecological carbon  
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31 transfer in surface oceans. Additionally, EVs play a critical role in the valorization of  
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33 lignin, a particularly recalcitrant sink of the terrestrial carbon cycle (71).  
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40 In addition to EV roles in nutrient cycling, EVs may also play a role in physically altering  
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42 the mineral phases surrounding the parent organisms as a means for promoting  
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44 microbial survival. For example, heterotrophic bacteria isolated from a black shale  
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46 deposit in southwestern Poland produce EVs that are the primary component of biofilms  
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48 and sites of phosphate, carbonate and sulphate mineral precipitation (60). Mineral  
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50 particles containing Cu, P, Mg, Si, Al and Ca were observed to form within vesicles from  
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52 these organisms, suggesting an active and targeted role for vesicles in element cycling  
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3 in geological deposits (60). Similarly, *Shewanella oneidensis* reduces soluble uranium  
4 species, which leads to formation of sparingly soluble uraninite mineral precipitates (59).  
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7 EV production in this species functions as a protective response for the cells by  
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9 enabling them to shed these uraninite crusts that would otherwise inhibit overall cellular  
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12 function (59).  
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17 Whereas such studies highlight the roles EVs play in soil environments with respect to  
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19 element cycling, mineral interactions, extracellular electron transfer, and nutrient  
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21 sequestration, EVs are generally considered the alternative to better-known processes  
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23 that occur directly at the cell envelope. Thus, the relative importance of EVs in these  
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25 functions remains unknown. Evaluation of EV contributions to interactions with the  
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27 organism's surroundings therefore should include comparisons to the cell-bound  
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29 processes that accomplish the same function for the organism. As-yet-unappreciated  
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31 advantages to functionalizing EVs include prolonged environmental persistence and  
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33 increased distribution of these nanoscale particles compared to EV-producing cells.  
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37 Uncovering principles governing EV mobility within the environment as well as how  
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39 environmental conditions affect that mobility will be essential to develop a  
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41 comprehensive understanding of how EVs impact biogeochemical processes. In the  
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43 future, our understanding of the roles EVs play in these processes and where they act  
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45 may provide opportunities to engineer or manage the soil environment for enhanced  
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47 plant productivity among other desirable outcomes.  
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### 53 ***EVs as Colloids: Transport and Attachment***

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3 From a biophysical perspective, EVs can be considered colloids: small particles  
4 (typically less than 1  $\mu\text{m}$ ) dispersed in liquid medium. While their complex and  
5 sometimes multi-tasking functionalities have begun to be revealed, the colloidal  
6 properties of EVs have thus far received relatively little attention. EVs likely have similar  
7 properties to those of more simplistic phospholipid vesicles; however, given their  
8 complex composition, EVs may exhibit unexpected colloidal properties (72). Some of  
9 this work has been conducted in the context of soft matter particles, using for example  
10 quartz crystal microbalance measurements to understand lipid particle attachment (73,  
11 74). Although not a direct comparison to EVs, these reports do suggest that both the  
12 surfaces of lipid particles and the surrounding environmental conditions influence the  
13 fate and persistence of these particles, specifically relating to attachment and possibly  
14 transformation or uptake as well. Studying EVs holistically in the context of their  
15 surrounding environment (i.e. as a collection of particles in a complex system) will  
16 provide fresh insights into vesicle fate and function (27, 29, 75). To do this, phenomena  
17 understood for colloid behavior may be helpful in predicting what governs the fate of  
18 EVs on their journey from parent cell through their surrounding environments.

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42 Particle transport depends on size, i.e. smaller particles will be transported differently  
43 than larger ones. In particular, nanoparticles tend to be more sensitive to thermal forces,  
44 and thus Brownian motion, while larger particles tend to be more sensitive to shear or  
45 gravitational forces (76). Analogously, we predict that EV transport will be distinct from  
46 that of cellular colloids. Furthermore, as described above, EVs and EV-producing cells  
47 have distinct compositional and surface characteristics, implying that the chemical as  
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3 well as the physical factors controlling particle transport will be different for EVs  
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5 compared to their parent cells. While the physical transport of particles based on size is  
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7 well-studied (77), knowledge of the biochemical composition of EV surfaces has rarely  
8  
9 been directly connected to their mobilization and deposition potential. Hence, the  
10  
11 determination of a metric to characterize surface chemistry is necessary to properly  
12  
13 evaluate EVs as colloidal particles.  
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19 An additional consideration is that all types of particles, including biological particles  
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21 (78-103), may undergo aggregation or deposition in environmental and physiological  
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23 systems. Both aggregation and deposition can be considered two-step processes where  
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25 largely physical phenomena transport particles to the vicinity of a surface (including  
26  
27 another particle) and near-field chemical factors (i.e. chemical factors that are only  
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29 experienced by a particle when in close proximity of the surface in question) determine  
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31 whether particle attachment occurs. For example, if ten collisions of a particle with a  
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33 surface result on average in only one particle attaching, the attachment efficiency of that  
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35 particle would be 0.1. Therefore, the attachment (or sticking) efficiency of a particle ( $\alpha$ )  
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37 reflects its relative affinity for the surfaces it encounters. The  $\alpha$  value of small particles  
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39 such as colloidal EVs may predict their tendency to disperse in environmental systems.  
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41 This concept has already been applied to bacterial systems, where simple models for  
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43 colloid deposition have been applied to describe bacterial deposition in porous media  
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45 (104-106).  
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3 Beyond particle attachment and transport into organisms, the attachment efficiency  
4 potentially also determines particle absorption, distribution, metabolism, and excretion  
5 behavior in organisms. This suggests that attachment efficiency could also determine  
6 these behaviors in EVs, as has been observed in the case of engineered nanoparticles  
7 (ENPs) When examined from a colloid chemistry perspective, this may lead to a wide  
8 range of environmental and biological interactions (107) that include bio-uptake (ENPs  
9 entering into cells or organisms) (108), biomagnification (ENPs observed at greater  
10 concentrations farther up the food chain) (109) trophic transfer (ENPs transferred up the  
11 food chain) (110) and maternal transfer (ENPs taken up by one generation of organism  
12 and passed on to the next generation) (111, 112). It is also possible that EVs could  
13 exhibit prolonged persistence in the environment compared to the EV-producing cell,  
14 which could result in accumulation at high levels and have unexpected implications for  
15 ecosystem function. Although we have yet to measure EV persistence in environmental  
16 conditions, EVs are remarkably stable when exposed to extreme conditions and  
17 physiological stress (15, 113-116), lending to possible biouptake or trophic transfer.  
18 Furthermore, EVs commonly possess genetic material which can propagate throughout  
19 environmental systems or food chains (117-123). Although this is not a direct colloid  
20 interaction, attachment could be the first step to allow for the genetic disruption that  
21 would then influence ecosystems indirectly through biomagnification or trophic transfer.  
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49 As noted above, complex and variable surface properties mediate EV attachment to  
50 biological surfaces, which is critical for bacterial EV interactions with plant hosts. While  
51 theoretical models for the attachment of particles to surfaces (124-126) provide useful  
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3 guidelines for overall trends, quantitative predictions for the deposition of both mineral  
4 and biological particles are hampered by complexities such as surface heterogeneity  
5 and interactions with solutes and adsorbing molecules. As a result, evaluation of the  
6 attachment efficiency,  $\alpha$ , in these systems has been largely empirical, relying on either  
7 column deposition studies (127-129) or, more recently, batch aggregation studies (130-  
8 132). The latter method is particularly well-suited to determining attachment efficiency in  
9 complex systems, which allows for direct parameterization of models to predict particle  
10 fate and serves as a “functional assay” to describe a wide range of behaviors in  
11 complex systems (133). For example, one study used laboratory determinations of  
12 values for  $\alpha$  to predict the fate of ENPs in simulated wetlands using a simple transport  
13 model (134). The approach has also been applied to systems ranging from activated  
14 sludge (131) and soils (130) to river basins (135). As a functional assay, the parameter  
15  $\alpha$  encompasses a large number of underlying variables ranging from temperature, pH,  
16 and van der Waals interactions, to steric repulsion, ion strength and surface  
17 composition. Hence,  $\alpha$  allows us to account for multiple variables in a single parameter,  
18 making this sort of assay amenable to predicting outcomes in complex systems.  
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42 Applying measurements of particle attachment behavior to EVs in the rhizosphere  
43 demonstrates the potential strength of combining theory, expertise, and methodology  
44 from colloid science, biology, and soil geochemistry to predict the fate of EVs in the  
45 environment. The resulting insights may even help us to retroactively relate functional  
46 assay-derived transport properties of EVs to their biological properties and impacts by  
47 connecting biological function with resultant surface properties and thus delivery. This  
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3 systems-level understanding can only be achieved by thorough integration of our  
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5 otherwise discrete fields.  
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### 10 ***Discussion***

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12 Understanding how cells communicate and impact one another across different  
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14 kingdoms and across physical space within complex natural environments such as the  
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16 rhizosphere presents significant research challenges. Scientific approaches in the  
17  
18 seemingly disparate areas of biology, geochemistry and colloid theory have each  
19  
20 independently developed a deep understanding of fundamental concepts pertinent to  
21  
22 this undertaking. For example, soil biogeochemistry studies have characterized  
23  
24 fundamental aspects of nutrient cycling, colloid science research has developed models  
25  
26 to predict transport of nanomaterials, and the fields of molecular biology and  
27  
28 biochemistry have revealed EV functionalities in simplified systems. Harnessed  
29  
30 together, these initial findings and understanding will allow us to describe and make  
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32 predictions about physiologically relevant EV properties, transport, and function in the  
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34 rhizosphere. This focus on a shared thread of inquiry is a proven core to successful  
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36 convergence research (136).  
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44 Convergence enables the ability to take advantage of available experimental strategies  
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46 across our represented disciplines. However, multi- and transdisciplinary dialogues  
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48 thus far have already revealed new challenges, highlighting the need to develop novel  
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50 methods and approaches.  
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3 For instance, in transitioning colloid chemistry methodologies from NP applications to  
4 the characterization of EVs, methodological challenges arise. First, EVs are comprised  
5 primarily of organic biomolecular ‘soft’ materials. Therefore, in many environmental  
6 systems, EVs are difficult to differentiate from other material in the soil matrix,  
7 presenting a significant barrier to many traditional methods for colloid study. Second,  
8 the ability to validate proposed models of EV transport is severely limited by a lack of  
9 existing data. Notably, these two categories of challenges – the obfuscating nature of  
10 the environment and a paucity of data - were also among the first hurdles within the  
11 convergent field of nanoEHS. We anticipate that solutions adapting approaches across  
12 fields may prove useful both for detection and validation of EV behavior in complex  
13 systems. For example, using biochemical techniques to fluorescently label lipids or  
14 proteins in EVs, or using genetic techniques to tag particular EV cargo may benefit our  
15 colloidal studies of EVs by facilitating transport, aggregation, and deposition  
16 measurements.

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37 Similarly, in biological studies of EVs there is a notable need to track these nanoscale  
38 packages and predict their movement and function in various environments. Using  
39 modeling principles from colloid science and a detailed understanding of soil  
40 composition from soil geochemistry, we may be able to predict which conditions will be  
41 most interesting and environmentally relevant to test experimentally and which cargo  
42 play critical roles in transport and function. Specifically, in the context of the  
43 rhizosphere, soil biogeochemistry has developed some understanding of how microbes  
44 and, perhaps, EVs play active roles in element cycling and biogeochemical processes.

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3 EVs may function differently than microbes in these processes due to their smaller size,  
4 relative increase in surface area, and potential to travel further and persist longer in soil.  
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6 Modeling techniques from colloid science and molecular characterization from biological  
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8 studies will significantly advance understanding based on biogeochemical approaches  
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10 alone by providing the necessary variables to define functional mechanisms.  
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14 Transdisciplinary approaches to the design stage as well as in driving analyses will be  
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16 important so that models may be informed appropriately and can be validated  
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18 experimentally. This convergent approach could solve a common problem in biological  
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20 experimentation where the number of conditions, controls and variables needed to  
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22 address increasingly complex questions exceeds that which is possible or practical in a  
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24 wet-lab environment.  
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31 Another major challenge in the EV field arises from technical limitations of purification  
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33 methods. Current approaches for and limitations of EV purification from mammalian,  
34  
35 bacterial, and plant cells have been discussed recently (43, 137-143). The most  
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37 common purification methods involve sequential ultracentrifugation and filtration steps  
38  
39 to pellet EVs followed by density purification to remove contaminants. One notable  
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41 limitation of this approach is the inability to separate EVs based on their molecular  
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43 cargo. Although some techniques like affinity purification can isolate groups of EVs that  
44  
45 all contain one similar cargo protein, currently there are no methods to isolate EVs with  
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47 the same complex mixture of cargo. A convergent approach to EV research could  
48  
49 reveal new ways to isolate EVs based on cargo or surface properties. Alternatively,  
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51 using the techniques from multiple disciplines may reveal new ways to track and  
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3 measure EVs either individually or as collective populations that bypass the need to  
4 isolate specific populations of EVs. For example, perhaps discrete populations of EVs  
5 could deposit in sediment and percolate to deeper layers, while others stay more on the  
6 surface of sediment, stay suspended in a water body, or are even taken up by various  
7 organisms preferentially (108, 144, 145). EV research could be extended to current  
8 studies in mesocosms (simulated and controlled environmental systems) to evaluate  
9 whether these differences exist and the extent to which they could be used to isolate  
10 various EV populations.  
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24 Considering the challenges of studying EV-mediated interkingdom communication from  
25 three different perspectives reveals many questions that are now possible to explore  
26 through shared and co-developed methodologies (Figure 1). For example, which vesicle  
27 properties affect surface chemistry and thus the fate and persistence of EVs in relevant  
28 environments? The transport and attachment portion of the EV journey during  
29 interkingdom communication requires an understanding of how vesicle surface  
30 composition (and variations in surface composition) impact surface charge, steric  
31 interactions, and other colloidal properties of EV suspensions. Biology, chemical  
32 biology, and analytical chemistry, among other related disciplines, can provide the tools  
33 needed to identify and characterize these surface-associated molecules and inform  
34 transport models. Further, we wonder how variations in physical properties of a larger  
35 scale, like size or shape, influence the mechanisms by which EVs are transported.  
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3 predictions experimentally presents an excellent opportunity to move all three fields  
4 forward.  
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10 An improved understanding of EV-mediated interkingdom communication in the  
11 rhizosphere would inform a myriad of future applications. Depending on EV persistence  
12 and transport profiles, potential agricultural applications might include naturally derived  
13 protective sprays, irrigation additives, or seed treatments to reduce disease incidence  
14 either by boosting plant immune responses or by interacting directly with the pathogens.  
15 Naturally produced or engineered EVs could potentially also be used to improve nutrient  
16 uptake in plants by packaging macro- and micro-nutrients in more bioavailable forms.  
17 Importantly, results from studies of naturally produced EVs and their functions will likely  
18 reveal specific cargo that results in desirable interactions and properties. This cargo  
19 could then be added to artificial lipid vesicles to improve their functionality. EVs may  
20 also prove useful as biomarkers for a variety of conditions, for example in crop disease  
21 and soil quality, as has been shown analogously in mammalian systems (31). For  
22 environmental management, a thorough understanding of EV composition, biogenesis,  
23 fate, and transport may enable optimal compositions of ENPs that deposit specific  
24 nutrients or that sequester heavy metals and other contaminants. While these  
25 applications have direct implications for agriculture and environmental management,  
26 studies of the properties governing EV function and fate will also impact our  
27 understanding of EVs in mammalian systems as well as microbial processes and  
28 complex microbial communities. The combination of theory and methods at the  
29 intersection of biology, colloid science, and geochemistry shows significant potential to  
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3 advance our understanding of EV composition and function as well as potential to  
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5 jumpstart a variety of environmentally, agriculturally, and economically important  
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7 applications, thus providing an example of impactful convergent research.  
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### 28 29 ***Declaration of Interests*** 30

31 The authors declare no conflicting interests.  
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### 35 36 ***Figure Legends*** 37

38 **Figure 1. Convergence to address EV-mediated inter-kingdom communication.** A  
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40 schematic describing the different compartments of the natural environment in which  
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42 EVs may be found, and the questions associated with the roles of EVs in these  
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44 compartments. Top Left. How do EVs released by microorganisms influence the  
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46 function of both plants and other microorganisms in the natural environment? Bottom  
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48 left. How do EVs interact with leaf and root cells? Top right. In what ways do time and  
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50 environmental stressors affect EV viability? Middle right. What roles do EVs have in  
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3 inter-microbial communication? Bottom right. How can EVs impact nutrient cycling in the  
4 environment?  
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10 **Figure 2. “Extracellular vesicle” encompasses a variety of secreted cellular**  
11 **structures.** Although we focus on microbial EVs throughout this perspective, the  
12 general term “EV” can refer to a wide array of vesicle structures. In general, terminology  
13 for EVs is determined according to their size and mechanism of biogenesis. This  
14 schematic depicts the various routes of EV biogenesis from mammalian and microbial  
15 cells. A detailed depiction of mammalian EVs, including specific cargo, can be found in  
16 recent reviews (2, 146-149). Depictions of cargo in EVs from bacterial and fungal  
17 species are also available (9, 38, 40, 140, 150-154). A) Top: Gram-negative bacteria  
18 produce outer membrane vesicles (OMVs) and outer inner membrane vesicles (OIMVs).  
19 Bottom: Explosive outer membrane vesicles can also result from explosive cell lysis.  
20 OM: Outer Membrane. PG: Peptidoglycan. IM: Inner Membrane. B) Gram-positive  
21 bacteria produce cytoplasmic membrane vesicles (CMVs). Fungal cells produce EVs  
22 presumably via a similar biogenesis pathway. C) Left: Mammalian cells produce a  
23 variety of EVs including exosomes, exomeres, microvesicles, migrosomes, and  
24 oncosomes. EE: Early Endosome. Lys: Lysosome. MVB: Multi-vesicular Body. ER:  
25 Endoplasmic Reticulum. Right: Apoptotic bodies are formed when mammalian cells  
26 undergo apoptotic cell death. Size in nm indicates diameter.  
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## 51 **References**

52  
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54 1. Deatherage BL, Cookson BT. Membrane vesicle release in bacteria, eukaryotes, and archaea: a  
55 conserved yet underappreciated aspect of microbial life. *Infection and immunity*. 2012;80(6):1948-57.  
56  
57



- 1
- 2
- 3
- 4 2. van Niel G, D'Angelo G, Raposo G. Shedding light on the cell biology of extracellular vesicles. *Nature Reviews Molecular Cell Biology*. 2018;19(4):213-28.
- 5
- 6 3. Beveridge TJ. Structures of gram-negative cell walls and their derived membrane vesicles. *J Bacteriol*. 1999;181(16):4725-33.
- 7
- 8 4. Schooling SR, Beveridge TJ. Membrane vesicles: an overlooked component of the matrices of biofilms. *Journal of bacteriology*. 2006;188(16):5945-57.
- 9
- 10 5. Kaparakis-Liaskos M, Ferrero RL. Immune modulation by bacterial outer membrane vesicles. *Nat Rev Immunol*. 2015;15(6):375-87.
- 11
- 12 6. McCaig WD, Koller A, Thanassi DG. Production of outer membrane vesicles and outer membrane tubes by *Francisella novicida*. *Journal of bacteriology*. 2013;195(6):1120-32.
- 13
- 14 7. Margolis L, Sadovsky Y. The biology of extracellular vesicles: The known unknowns. *PLOS Biology*. 2019;17(7):e3000363.
- 15
- 16 8. Avila-Calderón ED, Ruiz-Palma MdS, Aguilera-Arreola MG, Velázquez-Guadarrama N, Ruiz EA, Gomez-Lunar Z, et al. Outer Membrane Vesicles of Gram-Negative Bacteria: An Outlook on Biogenesis. *Frontiers in Microbiology*. 2021;12:345.
- 17
- 18 9. Schwechheimer C, Kuehn MJ. Outer-membrane vesicles from Gram-negative bacteria: biogenesis and functions. *Nat Rev Microbiol*. 2015;13(10):605-19.
- 19
- 20 10. Kaparakis-Liaskos M, Kufer TA. *Bacterial Membrane Vesicles: Biogenesis, Functions and Applications*: Springer International Publishing; 2020.
- 21
- 22 11. Bahar O, Mordukhovich G, Luu DD, Schwessinger B, Daudi A, Jehle AK, et al. Bacterial Outer Membrane Vesicles Induce Plant Immune Responses. *Mol Plant Microbe Interact*. 2016;29(5):374-84.
- 23
- 24 12. Cai Q, Qiao L, Wang M, He B, Lin F-M, Palmquist J, et al. Plants send small RNAs in extracellular vesicles to fungal pathogen to silence virulence genes. *Science*. 2018;360(6393):1126.
- 25
- 26 13. Rybak K, Robatzek S. Functions of Extracellular Vesicles in Immunity and Virulence. *Plant Physiology*. 2019;179(4):1236.
- 27
- 28 14. Ionescu M, Zaini PA, Baccari C, Tran S, da Silva AM, Lindow SE. *Xylella fastidiosa* outer membrane vesicles modulate plant colonization by blocking attachment to surfaces. *Proc Natl Acad Sci U S A*. 2014;111(37):E3910-8.
- 29
- 30 15. McMillan HM, Zebell SG, Ristaino JB, Dong X, Kuehn MJ. Protective plant immune responses are elicited by bacterial outer membrane vesicles. *Cell Reports*. 2021;34(3).
- 31
- 32 16. Janda M, Ludwig C, Rybak K, Meng C, Stigliano E, Botzenhardt L, et al. Biophysical and proteomic analyses suggest functions of *Pseudomonas syringae* pv tomato DC3000 extracellular vesicles in bacterial growth during plant infection. *bioRxiv*. 2021:2021.02.08.430144.
- 33
- 34 17. Avery D, Hendren CO, Wiesner MR, Nel AE, Lowry GV, Hull MS, et al. The NSF-EPA Centers for the Environmental Implications of Nanotechnology: What They Taught Us. *Nanotechnology Environmental Health and Safety: Risks, Regulation and Management*. 3rd ed2018. p. 151 - 68.
- 35
- 36 18. Lowry GV, Gregory KB, Apte SC, Lead JR. Transformations of Nanomaterials in the Environment. *Environmental Science & Technology*. 2012;46(13):6893-9.
- 37
- 38 19. Orench-Rivera N, Kuehn MJ. Environmentally controlled bacterial vesicle-mediated export. *Cell Microbiol*. 2016;18(11):1525-36.
- 39
- 40 20. Bonnington KE, Kuehn MJ. Outer Membrane Vesicle Production Facilitates LPS Remodeling and Outer Membrane Maintenance in *Salmonella* during Environmental Transitions. *MBio*. 2016;7(5).
- 41
- 42 21. Ionescu M, Yokota K, Antonova E, Garcia A, Beaulieu E, Hayes T, et al. Promiscuous Diffusible Signal Factor Production and Responsiveness of the *Xylella fastidiosa* Rpf System. *mBio*. 2016;7(4):e01054-16.
- 43
- 44 22. Feitosa-Junior OR, Stefanello E, Zaini PA, Nascimento R, Pierry PM, Dandekar AM, et al. Proteomic and Metabolomic Analyses of *Xylella fastidiosa* OMV-Enriched Fractions Reveal Association with Virulence Factors and Signaling Molecules of the DSF Family. *Phytopathology*. 2019;109(8):1344-53.
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23. Samuel M, Bleackley M, Anderson M, Mathivanan S. Extracellular vesicles including exosomes in cross kingdom regulation: a viewpoint from plant-fungal interactions. *Frontiers in plant science*. 2015;6:766-.
24. Woith E, Fuhrmann G, Melzig MF. Extracellular Vesicles-Connecting Kingdoms. *Int J Mol Sci*. 2019;20(22):5695.
25. Cai Q, He B, Weiberg A, Buck AH, Jin H. Small RNAs and extracellular vesicles: New mechanisms of cross-species communication and innovative tools for disease control. *PLOS Pathogens*. 2020;15(12):e1008090.
26. Soares RP, Xander P, Costa AO, Marcilla A, Menezes-Neto A, Del Portillo H, et al. Highlights of the São Paulo ISEV workshop on extracellular vesicles in cross-kingdom communication. *J Extracell Vesicles*. 2017;6(1):1407213.
27. Kim JH, Lee J, Park J, Gho YS, editors. Gram-negative and Gram-positive bacterial extracellular vesicles. *Seminars in cell & developmental biology*; 2015: Elsevier.
28. Pathirana RD, Kaparakis-Liaskos M. Bacterial membrane vesicles: Biogenesis, immune regulation and pathogenesis. *Cellular Microbiology*. 2016;18(11):1518-24.
29. de Toledo Martins S, Szwarc P, Goldenberg S, Alves LR. Extracellular vesicles in fungi: composition and functions. *Fungal Physiology and Immunopathogenesis*. 2019:45-59.
30. Oliveira DL, Nakayasu ES, Joffe LS, Guimarães AJ, Sobreira TJP, Nosanchuk JD, et al. Characterization of yeast extracellular vesicles: evidence for the participation of different pathways of cellular traffic in vesicle biogenesis. *PloS one*. 2010;5(6):e11113-e.
31. Nawaz M, Camussi G, Valadi H, Nazarenko I, Ekström K, Wang X, et al. The emerging role of extracellular vesicles as biomarkers for urogenital cancers. *Nature Reviews Urology*. 2014;11(12):688-701.
32. Cui Y, Gao J, He Y, Jiang L. Plant extracellular vesicles. *Protoplasma*. 2020;257(1):3-12.
33. Abels ER, Breakefield XO. Introduction to Extracellular Vesicles: Biogenesis, RNA Cargo Selection, Content, Release, and Uptake. *Cellular and Molecular Neurobiology*. 2016;36(3):301-12.
34. Colombo M, Raposo G, Théry C. Biogenesis, Secretion, and Intercellular Interactions of Exosomes and Other Extracellular Vesicles. *Annual Review of Cell and Developmental Biology*. 2014;30(1):255-89.
35. Bonnington KE, Kuehn MJ. Protein selection and export via outer membrane vesicles. *Biochimica et Biophysica Acta (BBA) - Molecular Cell Research*. 2014;1843(8):1612-9.
36. Toyofuku M, Nomura N, Eberl L. Types and origins of bacterial membrane vesicles. *Nature Reviews Microbiology*. 2019;17(1):13-24.
37. Guerrero-Mandujano A, Hernández-Cortez C, Ibarra JA, Castro-Escarpulli G. The outer membrane vesicles: Secretion system type zero. *Traffic*. 2017;18(7):425-32.
38. Jan AT. Outer Membrane Vesicles (OMVs) of Gram-negative Bacteria: A Perspective Update. *Front Microbiol*. 2017;8:1053.
39. Manning AJ, Kuehn MJ. Contribution of bacterial outer membrane vesicles to innate bacterial defense. *BMC Microbiology*. 2011;11(1):258.
40. Kuehn MJ, Kesty NC. Bacterial outer membrane vesicles and the host-pathogen interaction. *Genes Dev*. 2005;19(22):2645-55.
41. Ellis TN, Kuehn MJ. Virulence and Immunomodulatory Roles of Bacterial Outer Membrane Vesicles. *Microbiology and Molecular Biology Reviews*. 2010;74(1):81.
42. Gilmore WJ, Johnston EL, Zavan L, Bitto NJ, Kaparakis-Liaskos M. Immunomodulatory roles and novel applications of bacterial membrane vesicles. *Molecular Immunology*. 2021;134:72-85.
43. Klimentová J, Stulík J. Methods of isolation and purification of outer membrane vesicles from gram-negative bacteria. *Microbiological Research*. 2015;170:1-9.

- 1
- 2
- 3
- 4 44. Bielska E, Birch PRJ, Buck AH, Abreu-Goodger C, Innes RW, Jin H, et al. Highlights of the mini-
- 5 symposium on extracellular vesicles in inter-organismal communication, held in Munich, Germany,
- 6 August 2018. *J Extracell Vesicles*. 2019;8(1):1590116.
- 7 45. Solé M, Scheibner F, Hoffmeister AK, Hartmann N, Hause G, Rother A, et al. *Xanthomonas*
- 8 *campestris* pv. *vesicatoria* Secretes Proteases and Xylanases via the Xps Type II Secretion System and
- 9 Outer Membrane Vesicles. *J Bacteriol*. 2015;197(17):2879-93.
- 10 46. Kulkarni HM, Swamy CV, Jagannadham MV. The proteome of the outer membrane vesicles of an
- 11 Antarctic bacterium *Pseudomonas syringae* Lz4W. *Data Brief*. 2015;4:406-9.
- 12 47. Sidhu VK, Vorhölter FJ, Niehaus K, Watt SA. Analysis of outer membrane vesicle associated
- 13 proteins isolated from the plant pathogenic bacterium *Xanthomonas campestris* pv. *campestris*. *BMC*
- 14 *Microbiol*. 2008;8:87.
- 15 48. Chowdhury C, Jagannadham MV. Virulence factors are released in association with outer
- 16 membrane vesicles of *Pseudomonas syringae* pv. *tomato* T1 during normal growth. *Biochim Biophys*
- 17 *Acta*. 2013;1834(1):231-9.
- 18 49. Tayi L, Maku R, Patel HK, Sonti RV. Action of Multiple Cell Wall-Degrading Enzymes Is Required
- 19 for Elicitation of Innate Immune Responses During *Xanthomonas oryzae* pv. *oryzae* Infection in Rice. *Mol*
- 20 *Plant Microbe Interact*. 2016;29(8):599-608.
- 21 50. Gouran H, Gillespie H, Nascimento R, Chakraborty S, Zaini PA, Jacobson A, et al. The Secreted
- 22 Protease PrtA Controls Cell Growth, Biofilm Formation and Pathogenicity in *Xylella fastidiosa*. *Scientific*
- 23 *Reports*. 2016;6(1):31098.
- 24 51. Redak RA, Purcell AH, Lopes JRS, Blua MJ, Mizell Iii RF, Andersen PC. The Biology of Xylem Fluid-
- 25 Feeding Insect Vectors of *Xylella fastidiosa* and their Relation to Disease Epidemiology. *Annual Review of*
- 26 *Entomology*. 2003;49(1):243-70.
- 27 52. Chatterjee S, Wistrom C, Lindow SE. A cell-cell signaling sensor is required for virulence and
- 28 insect transmission of *Xylella fastidiosa*. *Proceedings of the National Academy of Sciences*.
- 29 2008;105(7):2670.
- 30 53. Almeida RPP, Purcell AH. Patterns of *Xylella fastidiosa* Colonization on the Precibarium of
- 31 Sharpshooter Vectors Relative to Transmission to Plants. *Annals of the Entomological Society of*
- 32 *America*. 2006;99(5):884-90.
- 33 54. Killiny N, Almeida RPP. *Xylella fastidiosa* Afimbrial Adhesins Mediate Cell Transmission to Plants
- 34 by Leafhopper Vectors. *Applied and Environmental Microbiology*. 2009;75(2):521.
- 35 55. Bauwens A, Kunsmann L, Marejková M, Zhang W, Karch H, Bielaszewska M, et al. Intrahost
- 36 milieu modulates production of outer membrane vesicles, vesicle-associated Shiga toxin 2a and
- 37 cytotoxicity in *Escherichia coli* O157:H7 and O104:H4. *Environ Microbiol Rep*. 2017;9(5):626-34.
- 38 56. Kulkarni HM, Swamy CV, Jagannadham MV. Molecular characterization and functional analysis
- 39 of outer membrane vesicles from the antarctic bacterium *Pseudomonas syringae* suggest a possible
- 40 response to environmental conditions. *J Proteome Res*. 2014;13(3):1345-58.
- 41 57. Eberlein C, Baumgarten T, Starke S, Heipieper HJ. Immediate response mechanisms of Gram-
- 42 negative solvent-tolerant bacteria to cope with environmental stress: cis-trans isomerization of
- 43 unsaturated fatty acids and outer membrane vesicle secretion. *Appl Microbiol Biotechnol*.
- 44 2018;102(6):2583-93.
- 45 58. Volgers C, Savelkoul PHM, Stassen FRM. Gram-negative bacterial membrane vesicle release in
- 46 response to the host-environment: different threats, same trick? *Crit Rev Microbiol*. 2018;44(3):258-73.
- 47 59. Shao PP, Comolli LR, Bernier-Latmani R. Membrane Vesicles as a Novel Strategy for Shedding
- 48 Encrusted Cell Surfaces. *Minerals*. 2014;4(1).
- 49 60. Matlakowska R, Skłodowska A, Nejbort K. Bioweathering of Kupferschiefer black shale (Fore-
- 50 Sudetic Monocline, SW Poland) by indigenous bacteria: implication for dissolution and precipitation of
- 51 minerals in deep underground mine. *FEMS Microbiology Ecology*. 2012;81(1):99-110.
- 52
- 53
- 54
- 55
- 56
- 57
- 58
- 59
- 60

61. Prados-Rosales R, Weinrick BC, Piqué DG, Jacobs WR, Jr., Casadevall A, Rodriguez GM. Role for Mycobacterium tuberculosis membrane vesicles in iron acquisition. *Journal of bacteriology*. 2014;196(6):1250-6.
62. Liu X, Jing X, Ye Y, Zhan J, Ye J, Zhou S. Bacterial Vesicles Mediate Extracellular Electron Transfer. *Environmental Science & Technology Letters*. 2020;7(1):27-34.
63. Lin J, Zhang W, Cheng J, Yang X, Zhu K, Wang Y, et al. A Pseudomonas T6SS effector recruits PQS-containing outer membrane vesicles for iron acquisition. *Nature Communications*. 2017;8(1):14888.
64. Biller SJ, Schubotz F, Roggensack SE, Thompson AW, Summons RE, Chisholm SW. Bacterial vesicles in marine ecosystems. *Science*. 2014;343(6167):183-6.
65. Ahmed E, Holmström SJM. Siderophores in environmental research: roles and applications. *Microb Biotechnol*. 2014;7(3):196-208.
66. Johnstone TC, Nolan EM. Beyond iron: non-classical biological functions of bacterial siderophores. *Dalton Trans*. 2015;44(14):6320-39.
67. Kramer J, Özkaya Ö, Kümmerli R. Bacterial siderophores in community and host interactions. *Nature Reviews Microbiology*. 2020;18(3):152-63.
68. Fein JB, Yu Q, Nam J, Yee N. Bacterial cell envelope and extracellular sulfhydryl binding sites: Their roles in metal binding and bioavailability. *Chemical Geology*. 2019;521:28-38.
69. Choi C-W, Park EC, Yun SH, Lee S-Y, Lee YG, Hong Y, et al. Proteomic Characterization of the Outer Membrane Vesicle of Pseudomonas putida KT2440. *Journal of Proteome Research*. 2014;13(10):4298-309.
70. Bhar S, Edelmann MJ, Jones MK. Characterization and proteomic analysis of outer membrane vesicles from a commensal microbe, Enterobacter cloacae. *Journal of Proteomics*. 2021;231:103994.
71. Salvachúa D, Werner AZ, Pardo I, Michalska M, Black BA, Donohoe BS, et al. Outer membrane vesicles catabolize lignin-derived aromatic compounds in Pseudomonas putida KT2440. *Proceedings of the National Academy of Sciences*. 2020;117(17):9302.
72. Yousefi N, Tufenkji N. Probing the Interaction between Nanoparticles and Lipid Membranes by Quartz Crystal Microbalance with Dissipation Monitoring. *Frontiers in Chemistry*. 2016;4:46.
73. Tellechea E, Johannsmann D, Steinmetz NF, Richter RP, Reviakine I. Model-Independent Analysis of QCM Data on Colloidal Particle Adsorption. *Langmuir*. 2009;25(9):5177-84.
74. Richter R, Mukhopadhyay A, Brisson A. Pathways of lipid vesicle deposition on solid surfaces: a combined QCM-D and AFM study. *Biophysical journal*. 2003;85(5):3035-47.
75. Buzás EI, Tóth EÁ, Sódar BW, Szabó-Taylor KÉ. Molecular interactions at the surface of extracellular vesicles. *Seminars in Immunopathology*. 2018;40(5):453-64.
76. Thill A, Moustier S, Aziz J, Wiesner MR, Bottero JY. Flocc restructuring during aggregation: Experimental evidence and numerical simulation. *J Colloid Interf Sci*. 2001;243(1):171-82.
77. Elimelech M. Effect of Particle Size on the Kinetics of Particle Deposition under Attractive Double Layer Interactions. *Journal of Colloid and Interface Science*. 1994;164(1):190-9.
78. Yuan B, Pham M, Nguyen TH. Deposition Kinetics of Bacteriophage MS2 on a Silica Surface Coated with Natural Organic Matter in a Radial Stagnation Point Flow Cell. *Environmental Science & Technology*. 2008;42(20):7628-33.
79. Gutierrez L, Li X, Wang J, Nangmenyi G, Economy J, Kuhlenschmidt TB, et al. Adsorption of rotavirus and bacteriophage MS2 using glass fiber coated with hematite nanoparticles. *Water Research*. 2009;43(20):5198-208.
80. Gutierrez L, Mylon SE, Nash B, Nguyen TH. Deposition and Aggregation Kinetics of Rotavirus in Divalent Cation Solutions. *Environmental Science & Technology*. 2010;44(12):4552-7.
81. Liu Y, Janjaroen D, Kuhlenschmidt MS, Kuhlenschmidt TB, Nguyen TH. Deposition of Cryptosporidium parvum Oocysts on Natural Organic Matter Surfaces: Microscopic Evidence for Secondary Minimum Deposition in a Radial Stagnation Point Flow Cell. *Langmuir*. 2009;25(3):1594-605.

- 1  
2  
3 82. Janjaroen D, Ling F, Monroy G, Derlon N, Mogenroth E, Boppart SA, et al. Roles of ionic strength  
4 and biofilm roughness on adhesion kinetics of *Escherichia coli* onto groundwater biofilm grown on PVC  
5 surfaces. *Water Research*. 2013;47(7):2531-42.
- 6 83. Shen Y, Monroy GL, Derlon N, Janjaroen D, Huang C, Morgenroth E, et al. Role of Biofilm  
7 Roughness and Hydrodynamic Conditions in *Legionella pneumophila* Adhesion to and Detachment from  
8 Simulated Drinking Water Biofilms. *Environmental Science & Technology*. 2015;49(7):4274-82.
- 9 84. Gutierrez L, Nguyen TH. Interactions between Rotavirus and Suwannee River Organic Matter:  
10 Aggregation, Deposition, and Adhesion Force Measurement. *Environmental Science & Technology*.  
11 2012;46(16):8705-13.
- 12 85. Armanious A, Aeppli M, Jacak R, Refardt D, Sigstam T, Kohn T, et al. Viruses at Solid–Water  
13 Interfaces: A Systematic Assessment of Interactions Driving Adsorption. *Environmental Science &*  
14 *Technology*. 2016;50(2):732-43.
- 15 86. Tomaszewski JE, Schwarzenbach RP, Sander M. Protein Encapsulation by Humic Substances.  
16 *Environmental Science & Technology*. 2011;45(14):6003-10.
- 17 87. Sander M, Madliger M, Schwarzenbach RP. Adsorption of Transgenic Insecticidal Cry1Ab Protein  
18 to SiO<sub>2</sub>. 1. Forces Driving Adsorption. *Environmental Science & Technology*. 2010;44(23):8870-6.
- 19 88. Madliger M, Gasser CA, Schwarzenbach RP, Sander M. Adsorption of Transgenic Insecticidal  
20 Cry1Ab Protein to Silica Particles. Effects on Transport and Bioactivity. *Environmental Science &*  
21 *Technology*. 2011;45(10):4377-84.
- 22 89. Servagent-Noinville S, Revault M, Quiquampoix H, Baron MH. Conformational Changes of Bovine  
23 Serum Albumin Induced by Adsorption on Different Clay Surfaces: FTIR Analysis. *Journal of Colloid and*  
24 *Interface Science*. 2000;221(2):273-83.
- 25 90. Baron MH, Revault M, Servagent-Noinville S, Abadie J, Quiquampoix H. Chymotrypsin  
26 Adsorption on Montmorillonite: Enzymatic Activity and Kinetic FTIR Structural Analysis. *Journal of*  
27 *Colloid and Interface Science*. 1999;214(2):319-32.
- 28 91. Leprince F, Quiquampoix H. Extracellular enzyme activity in soil: effect of pH and ionic strength  
29 on the interaction with montmorillonite of two acid phosphatases secreted by the ectomycorrhizal  
30 fungus *Hebeloma cylindrosporum*. *European Journal of Soil Science*. 1996;47(4):511-22.
- 31 92. Quiquampoix H, Staunton S, Baron MH, Ratcliffe RG. Interpretation of the pH dependence of  
32 protein adsorption on clay mineral surfaces and its relevance to the understanding of extracellular  
33 enzyme activity in soil. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*. 1993;75:85-  
34 93.
- 35 93. Quiquampoix H, Burns RG. Interactions between Proteins and Soil Mineral Surfaces:  
36 Environmental and Health Consequences. *Elements*. 2007;3(6):401-6.
- 37 94. Daly CA, Allen C, Rozanov N, Chong G, Melby ES, Kuech TR, et al. Surface Coating Structure and  
38 Its Interaction with Cytochrome c in EG6-Coated Nanoparticles Varies with Surface Curvature. *Langmuir*.  
39 2020;36(18):5030-9.
- 40 95. Liang D, Dahal U, Zhang Y, Lochbaum C, Ray D, Hamers RJ, et al. Interfacial water and ion  
41 distribution determine  $\zeta$  potential and binding affinity of nanoparticles to biomolecules. *Nanoscale*.  
42 2020;12(35):18106-23.
- 43 96. Caudill ER, Hernandez RT, Johnson KP, O'Rourke JT, Zhu L, Haynes CL, et al. Wall teichoic acids  
44 govern cationic gold nanoparticle interaction with Gram-positive bacterial cell walls. *Chemical Science*.  
45 2020;11(16):4106-18.
- 46 97. Sheavly JK, Pedersen JA, Van Lehn RC. Curvature-driven adsorption of cationic nanoparticles to  
47 phase boundaries in multicomponent lipid bilayers. *Nanoscale*. 2019;11(6):2767-78.
- 48 98. Melby ES, Allen C, Foreman-Ortiz IU, Caudill ER, Kuech TR, Vartanian AM, et al. Peripheral  
49 Membrane Proteins Facilitate Nanoparticle Binding at Lipid Bilayer Interfaces. *Langmuir*.  
50 2018;34(36):10793-805.
- 51  
52  
53  
54  
55  
56  
57  
58  
59  
60

- 1  
2  
3 99. Grant SB, List EJ, Lidstrom ME. Kinetic analysis of virus adsorption and inactivation in batch  
4 experiments. *Water Resources Research*. 1993;29(7):2067-85.
- 5 100. Redman JA, B. Grant S, Olson TM, Adkins JM, Jackson JL, Castillo MS, et al. Physicochemical  
6 mechanisms responsible for the filtration and mobilization of a filamentous bacteriophage in quartz  
7 sand. *Water Research*. 1999;33(1):43-52.
- 8 101. Redman JA, Grant SB, Olson TM, Hardy ME, Estes MK. Filtration of Recombinant Norwalk Virus  
9 Particles and Bacteriophage MS2 in Quartz Sand: Importance of Electrostatic Interactions.  
10 *Environmental Science & Technology*. 1997;31(12):3378-83.
- 11 102. Grant SB. Virus Coagulation in Aqueous Environments. *Environmental Science & Technology*.  
12 1994;28(5):928-33.
- 13 103. Penrod SL, Olson TM, Grant SB. Deposition Kinetics of Two Viruses in Packed Beds of Quartz  
14 Granular Media. *Langmuir*. 1996;12(23):5576-87.
- 15 104. Bouwer EJ, Rittman BE. Comment on "Use of Colloid Filtration Theory in Modelling Movement of  
16 Bacteria through a Contaminated Sandy Aquifer. *Environ Sci Technol*. 1992;26(2):400-1.
- 17 105. Duston KL, Wiesner MR, Ward CH, Battelle Mem I. Bacterial attachment and transport through  
18 porous media: The effects of bacterial cell characteristics. Columbus: Battelle Press; 1997. 565- p.
- 19 106. Redman JA, Walker SL, Elimelech M. Bacterial adhesion and transport in porous media: role of  
20 the secondary minimum. *Environmental Science & Technology*. 2004;38(6):1777-85.
- 21 107. Wiesner MR, Lowry GV, Jones KL, Hochella MF, Di Giulio RT, Casman E, et al. Decreasing  
22 Uncertainties in Assessing Environmental Exposure, Risk, and Ecological Implications of Nanomaterials.  
23 *Environ Sci Technol*. 2009;43(17):6458-62.
- 24 108. Geitner NK, Cooper JL, Avellan A, Castellon BT, Perrotta BG, Bossa N, et al. Size-Based  
25 Differential Transport, Uptake, and Mass Distribution of Ceria (CeO<sub>2</sub>) Nanoparticles in Wetland  
26 Mesocosms. *Environmental Science & Technology*. 2018;52(17):9768-76.
- 27 109. Judy JD, Unrine JM, Bertsch PM. Evidence for Biomagnification of Gold Nanoparticles within a  
28 Terrestrial Food Chain. *Environmental Science & Technology*. 2011;45(2):776-81.
- 29 110. Geitner NK, Marinakos SM, Guo C, O'Brien N, Wiesner MR. Nanoparticle Surface Affinity as a  
30 Predictor of Trophic Transfer. *Environ Sci Technol*. 2016;50(13):6663-9.
- 31 111. Blickley T, Matson C, Vreeland W, Rittschoff D, Di Giulio R, McClellan-Green P. Effects of dietary  
32 CdSe/ZnS quantum dot exposure in estuarine fish: bioavailability, oxidative stress responses,  
33 reproduction, and maternal transfer. *Aquat Toxicol*. 2014;148:27-49.
- 34 112. Kim SW, Kwak JI, An Y-J. Multigenerational Study of Gold Nanoparticles in *Caenorhabditis*  
35 *elegans*: Transgenerational Effect of Maternal Exposure. *Environmental Science & Technology*.  
36 2013;47(10):5393-9.
- 37 113. Ott E, Kawaguchi Y, Kölbl D, Rabbow E, Rettberg P, Mora M, et al. Molecular repertoire of  
38 *Deinococcus radiodurans* after 1 year of exposure outside the International Space Station within the  
39 Tanpopo mission. *Microbiome*. 2020;8(1):150.
- 40 114. Podolich O, Kukharenko O, Haidak A, Zaets I, Zaika L, Storozhuk O, et al. Multimicrobial  
41 Kombucha Culture Tolerates Mars-like Conditions Simulated on Low Earth Orbit. *Astrobiology*.  
42 2018;19(2):183-96.
- 43 115. Podolich O, Kukharenko O, Zaets I, Orlovskaya I, Palchykovska L, Zaika L, et al. Fitness of Outer  
44 Membrane Vesicles From *Komagataeibacter intermedius* Is Altered Under the Impact of Simulated  
45 Mars-like Stressors Outside the International Space Station. *Frontiers in Microbiology*. 2020;11:1268.
- 46 116. Lossouarn J, Dupont S, Gorlas A, Mercier C, Bienvenu N, Marguet E, et al. An abyssal mobilome:  
47 viruses, plasmids and vesicles from deep-sea hydrothermal vents. *Research in Microbiology*.  
48 2015;166(10):742-52.
- 49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

117. Fulsundar S, Harms K, Flaten GE, Johnsen PJ, Chopade BA, Nielsen KM. Gene transfer potential of outer membrane vesicles of *Acinetobacter baylyi* and effects of stress on vesiculation. *Applied and environmental microbiology*. 2014;80(11):3469-83.
118. Domingues S, Nielsen KM. Membrane vesicles and horizontal gene transfer in prokaryotes. *Current Opinion in Microbiology*. 2017;38:16-21.
119. Tran F, Boedicker JQ. Genetic cargo and bacterial species set the rate of vesicle-mediated horizontal gene transfer. *Scientific Reports*. 2017;7(1):8813.
120. Velimirov B, Ranftler C. Unexpected aspects in the dynamics of horizontal gene transfer of prokaryotes: the impact of outer membrane vesicles. *Wien Med Wochenschr*. 2018;168(11-12):307-13.
121. Grüll MP, Mulligan ME, Lang AS. Small extracellular particles with big potential for horizontal gene transfer: membrane vesicles and gene transfer agents. *FEMS Microbiology Letters*. 2018;365(19).
122. Abe K, Nomura N, Suzuki S. Biofilms: hot spots of horizontal gene transfer (HGT) in aquatic environments, with a focus on a new HGT mechanism. *FEMS Microbiology Ecology*. 2020;96(5).
123. Emamalipour M, Seidi K, Zununi Vahed S, Jahanban-Esfahlan A, Jaymand M, Majdi H, et al. Horizontal Gene Transfer: From Evolutionary Flexibility to Disease Progression. *Frontiers in Cell and Developmental Biology*. 2020;8:229.
124. Hunter RJ. *Zeta Potential in Colloid Science*. New York: Academic Press Inc.; 1981. 386 p.
125. Tufenkji N, Elimelech M. Deviation from the classical colloid filtration theory in the presence of repulsive DLVO interactions. *Langmuir*. 2004;20(25):10818–28.
126. Elimelech M, Gregory J, Jia X, Williams RA. *Particle Deposition and Aggregation: Measurement, Modeling, and Simulations*. Oxford: Butterworth-Heinemann; 1995.
127. Elimelech M, O'Melia CR. Effect of particle size on collision efficiency in the deposition of brownian particles with electrostatic energy barriers. *Langmuir*. 1990;6:1153-63.
128. Elimelech M, O'Melia CR. Effect of electrolyte type on the electrophoretic mobility of polystyrene latex colloids. *Colloids and Surfaces*. 1990;44:165-78.
129. Song L, Elimelech M. Calculation of Particle Deposition Rate under Unfavourable Particle-Surface Interactions. *J Chem Soc Faraday Trans*. 1993b;89(18):3443-52.
130. Turner AA, Rogers NMK, Geitner NK, Wiesner MR. Nanoparticle affinity for natural soils: a functional assay for determining particle attachment efficiency in complex systems. *Environmental Science: Nano*. 2020;7(6):1719-29.
131. Barton LE, Therezien M, Auffan M, Bottero JY, Wiesner MR. Theory and Methodology for Determining Nanoparticle Affinity for Heteroaggregation in Environmental Matrices Using Batch Measurements. *Environmental Engineering Science*. 2014;31(7):421-7.
132. Geitner NK, O'Brien NJ, Turner AA, Cummins EJ, Wiesner MR. Measuring Nanoparticle Attachment Efficiency in Complex Systems. *Environ Sci Technol*. 2017;51(22):13288-94.
133. Hendren CO, Lowry GV, Unrine JM, Wiesner MR. A functional assay-based strategy for nanomaterial risk forecasting. *Science of The Total Environment*. 2015;536:1029-37.
134. Geitner NK, Bossa N, Wiesner MR. Formulation and Validation of a Functional Assay-Driven Model of Nanoparticle Aquatic Transport. *Environ Sci Technol*. 2019;53(6):3104-9.
135. Praetorius A, Tufenkji N, Goss K-U, Scheringer M, von der Kammer F, Elimelech M. The road to nowhere: equilibrium partition coefficients for nanoparticles. *Environmental Science: Nano*. 2014;1(4):317-23.
136. Roco MC. Principles and Methods That Facilitate Convergence. In: Bainbridge WS, Roco MC, editors. *Handbook of Science and Technology Convergence*. Cham: Springer International Publishing; 2014. p. 1-20.
137. Gandham S, Su X, Wood J, Nocera AL, Alli SC, Milane L, et al. Technologies and Standardization in Research on Extracellular Vesicles. *Trends in Biotechnology*. 2020;38(10):1066-98.

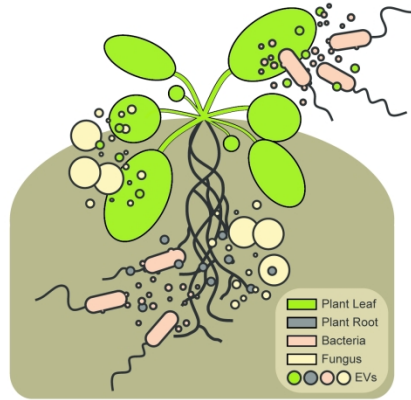
- 1  
2  
3 138. Chutkan H, Macdonald I, Manning A, Kuehn MJ. Quantitative and qualitative preparations of  
4 bacterial outer membrane vesicles. *Methods Mol Biol.* 2013;966:259-72.  
5 139. Dauros Singorenko P, Chang V, Whitcombe A, Simonov D, Hong J, Phillips A, et al. Isolation of  
6 membrane vesicles from prokaryotes: a technical and biological comparison reveals heterogeneity. *J*  
7 *Extracell Vesicles.* 2017;6(1):1324731.  
8 140. Qing G, Gong N, Chen X, Chen J, Zhang H, Wang Y, et al. Natural and engineered bacterial outer  
9 membrane vesicles. *Biophysics Reports.* 2019;5(4):184-98.  
10 141. Rutter BD, Innes RW. Extracellular Vesicles Isolated from the Leaf Apoplast Carry Stress-  
11 Response Proteins. *Plant physiology.* 2017;173(1):728-41.  
12 142. Rutter BD, Innes RW. Growing pains: addressing the pitfalls of plant extracellular vesicle  
13 research. *New Phytologist.* 2020;228(5):1505-10.  
14 143. Rutter BD, Rutter KL, Innes RW. Isolation and Quantification of Plant Extracellular Vesicles. *Bio-*  
15 *protocol.* 2017;7(17):e2533.  
16 144. Matson C, King RS, Castellon BT, Perrotta BG, Baker L, Simonin M, et al. Mesocosm approaches  
17 to study the influence of environmental complexity on nanoparticle fate and transport, transformations,  
18 bioavailability, and toxicity. December 01, 2019. p. B11M-2235.  
19 145. Espinasse BP, Geitner NK, Schierz A, Therezien M, Richardson CJ, Lowry GV, et al. Comparative  
20 Persistence of Engineered Nanoparticles in a Complex Aquatic Ecosystem. *Environmental Science &*  
21 *Technology.* 2018;52(7):4072-8.  
22 146. Yáñez-Mó M, Siljander PRM, Andreu Z, Bedina Zavec A, Borràs FE, Buzas EI, et al. Biological  
23 properties of extracellular vesicles and their physiological functions. *J Extracell Vesicles.*  
24 2015;4(1):27066.  
25 147. Chuo ST-Y, Chien JC-Y, Lai CP-K. Imaging extracellular vesicles: current and emerging methods.  
26 *Journal of Biomedical Science.* 2018;25(1):91.  
27 148. Holm MM, Kaiser J, Schwab ME. Extracellular Vesicles: Multimodal Envoys in Neural  
28 Maintenance and Repair. *Trends in Neurosciences.* 2018;41(6):360-72.  
29 149. Srivastava A, Amreddy N, Pareek V, Chinnappan M, Ahmed R, Mehta M, et al. Progress in  
30 extracellular vesicle biology and their application in cancer medicine. *WIREs Nanomedicine and*  
31 *Nanobiotechnology.* 2020;12(4):e1621.  
32 150. Brown L, Wolf JM, Prados-Rosales R, Casadevall A. Through the wall: extracellular vesicles in  
33 Gram-positive bacteria, mycobacteria and fungi. *Nature Reviews Microbiology.* 2015;13(10):620-30.  
34 151. Zamith-Miranda D, Nimrichter L, Rodrigues ML, Nosanchuk JD. Fungal extracellular vesicles:  
35 modulating host-pathogen interactions by both the fungus and the host. *Microbes and Infection.*  
36 2018;20(9):501-4.  
37 152. Turnbull L, Toyofuku M, Hynen AL, Kurosawa M, Pessi G, Petty NK, et al. Explosive cell lysis as a  
38 mechanism for the biogenesis of bacterial membrane vesicles and biofilms. *Nature communications.*  
39 2016;7:11220-.  
40 153. Toyofuku M, Cárcamo-Oyarce G, Yamamoto T, Eisenstein F, Hsiao C-C, Kurosawa M, et al.  
41 Prophage-triggered membrane vesicle formation through peptidoglycan damage in *Bacillus subtilis*.  
42 *Nature communications.* 2017;8(1):481-.  
43 154. Giesbrecht P, Labischinski H, Wecke J. A special morphogenetic wall defect and the subsequent  
44 activity of "murosomes" as the very reason for penicillin-induced bacteriolysis in staphylococci. *Arch*  
45 *Microbiol.* 1985;141(4):315-24.  
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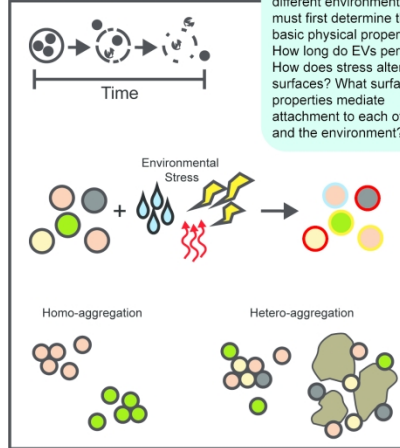
Figure 1

## EV-mediated Inter-kingdom Communication

Many factors determine EV function in complex environments. By integrating information and techniques from multiple fields, we can begin to form a complete picture of what roles EVs play and in which conditions they are produced.

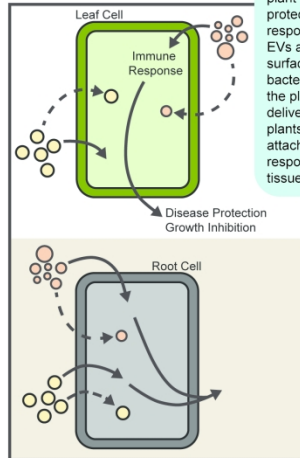


### Governing Themes



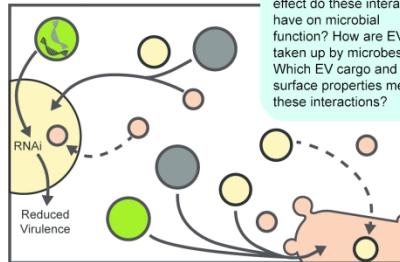
To understand the functions EVs play in different environments, we must first determine their basic physical properties. How long do EVs persist? How does stress alter their surfaces? What surface properties mediate attachment to each other and the environment?

### EV-host



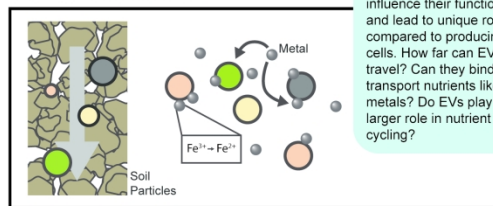
EV interactions with plant leaves induce protective plant immune responses. In contrast, EVs also attach to plant surfaces to facilitate bacterial spread through the plant. Can EVs deliver nutrients to plants? What mediates attachment? Do these responses occur in root tissue?

### EV-microbe



EVs can interact with microbes in the environment including bacteria and fungi. What effect do these interactions have on microbial function? How are EVs taken up by microbes? Which EV cargo and surface properties mediate these interactions?

### EV-environment



As they move through the environment, EV transport and nutrient sorption could influence their function and lead to unique roles compared to producing cells. How far can EVs travel? Can they bind and transport nutrients like metals? Do EVs play a larger role in nutrient cycling?

Caption : Figure 1. Convergence to address EV-mediated inter-kingdom communication. A schematic describing the different compartments of the natural environment in which EVs may be found, and the microorganisms associated with the roles of EVs in these compartments. Top Left. How do EVs released by microorganisms influence the function of both plants and other microorganisms in the natural environment? Bottom left. How do EVs interact with leaf and root cells? Top right. In what ways do time and environmental stressors affect EV viability? Middle right. What roles do EVs have in inter-microbial communication? Bottom right. How can EVs impact nutrient cycling in the environment?

Figure 2

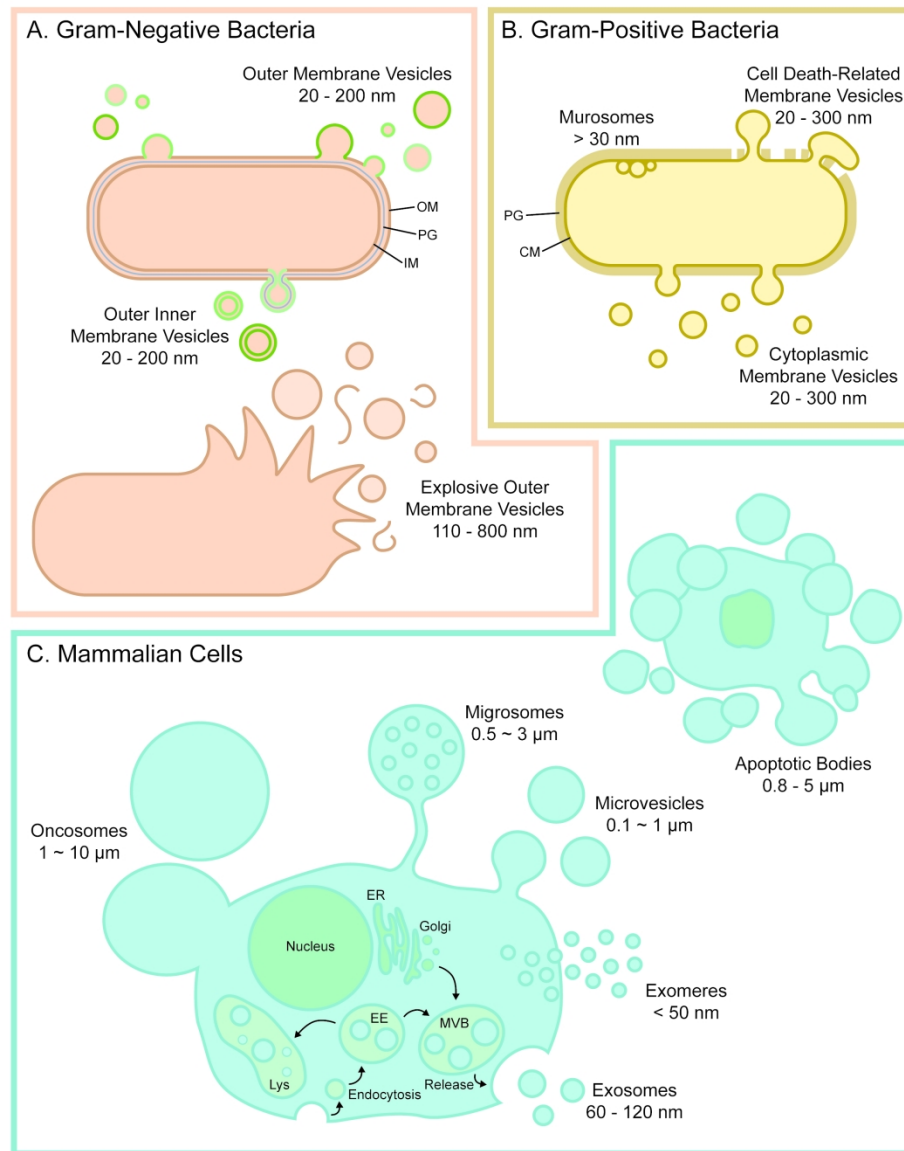


Figure 2. "Extracellular vesicle" encompasses a variety of secreted cellular structures. Although we focus on microbial EVs throughout this perspective, the general term "EV" can refer to a wide array of vesicle structures. In general, terminology for EVs is determined according to their size and mechanism of biogenesis. This schematic depicts the various routes of EV biogenesis from mammalian and microbial cells. A detailed depiction of mammalian EVs, including specific cargo, can be found in recent reviews (2, 146-149). Depictions of cargo in EVs from bacterial and fungal species are also available (9, 38, 40, 140, 150-154). A) Top: Gram-negative bacteria produce outer membrane vesicles (OMVs) and outer inner membrane vesicles (OIMVs). Bottom: Explosive outer membrane vesicles can also result from explosive cell lysis. OM: Outer Membrane. PG: Peptidoglycan. IM: Inner Membrane. B) Gram-positive bacteria produce cytoplasmic membrane vesicles (CMVs). Fungal cells produce EVs presumably via a similar biogenesis pathway. C) Left: Mammalian cells produce a variety of EVs including exosomes, exomeres, microvesicles, migrosomes, and oncosomes. EE: Early Endosome. Lys: Lysosome. MVB: Multi-vesicular Body. ER: Endoplasmic Reticulum. Right: Apoptotic bodies are formed when mammalian cells undergo apoptotic cell death. Size in nm indicates diameter.

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