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Photophysics of fluorescent nanoparticles based on organic dyes – challenges and design principles

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Fluorescent nanoparticles have become attractive for bioanalysis and imaging, due to their high brightness and photostability. Many different optical materials have been applied in fluorescent nanoparticles with a broad range of properties and characteristics. One appealing approach is the incorporation of molecular organic fluorophores in nanoparticles with the intention of transferring their known attractive solution-state properties directly to the nanoparticles. However, as molecular dyes are packed closely together in the nanoparticles their interactions most often result in fluorescence quenching and change in spectral properties making this approach challenging. In this perspective we will first discuss the origins of quenching and spectral shifts observed in dye based nanoparticles. On this background, we will then describe various designs of dye based NPs and how they address the challenges of dye–dye interactions and quenching. Our aim is to provide a general framework for understanding the supramolecular mechanisms that determine the photophysics of dye based nanoparticles. This framework of molecular photophysics and its relation to the internal structure of dye based nanoparticles can hopefully serve to assist rational design and optimization of new and improved dye based nanoparticles.

Introduction

Fluorescent nanoparticles (NPs) have proven attractive for use in bioanalysis and bioimaging.^{1,2} Compared to traditional molecular fluorescent labels the NPs can offer a number of advantages including highly increased brightness and increased photostability. Another key advantage of NPs is that the emissive materials are often protected from the environment making the optical properties insensitive to complex biological environments. In general, NPs also display low propensity for unspecific binding or localization in biological systems. Motivated by these potential advantages many different types of nanoparticles have been reported. In a very general manner, they can be divided into inorganic based or organic based, where inorganic NPs were more dominating in the early years. Among the inorganic nanoparticles there are many variations^{3,4} including the well-known quantum dots (QDs)^{5–8} and upconversion NPs.^{9,10} Fluorescent NPs where the absorption and emission stem from organic molecules and materials include NPs based on molecular dyes (neat, or embedded in a matrix material),¹¹ conjugated polymers,^{12,13} and amorphous carbon materials (carbon dots).^{14,15}

In the case of QDs their spectral properties mainly result from electronic states delocalized over the entire QD and thus intimately linked to the size. By varying the size of the particle, it

is possible to create emission colours spanning all wavelengths from the UV to IR.⁶ While this provides a direct way of engineering the optical properties of a single material, it can also be a limitation since it requires very strict preparation conditions and size control. In addition, QDs do usually not have discrete absorption bands, which limits selective excitation.^{6,16} Contrary to this, the optical properties of NPs based on organic dyes and conjugated polymers are largely defined by the covalent building blocks and thus independent of the size and size distribution of the NPs, while also in general being brighter than their inorganic counterparts.¹¹ It is exactly the well-defined correlation between the covalent structure of organic molecules and their photophysical properties that is the basis for their large potential as fluorescent labels and probes.^{17,18} 150 years of organic synthesis and study of dyes have provided a very diverse range of structures and photophysical properties, and a detailed understanding of the structure–property relations. Therefore, it is also attractive to design dyes and polymers optimized for use in fluorescent NPs, or simply use this vast back catalogue of already existing dyes and polymers and try to transfer desired photophysical properties to NPs.

Packing a large number of these organic units in one NP can bring about advantages such as very high brightness and protection from the environment. However, as molecular dyes are packed closely together in the NPs their interactions most often result in fluorescence quenching, known as aggregation caused quenching (ACQ), and change in spectral properties making this approach less straightforward and more

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is instead delocalized over two or more units. Depending on the systems and the details of the new states formed, such phenomena are described by different models like exciton coupling and excimers. We will denote all such phenomena as “strong interactions”. Strong interactions can lead to large spectral shifts in both absorption and emission spectra, just like the new delocalized excited states can display very different radiative and non-radiative rates and thus fluorescence quantum yields and lifetimes.

In the context of molecular materials and dye based NPs the most common model for strong interactions is based on the molecular exciton coupling model by Kasha, which describes the new states and transitions as a result of Coulomb coupling between the transition dipole moments in neighbouring chromophores.^{29,30} From this model it is evident that the energy and transition probabilities of the aggregate exciton transitions strongly depend on the detailed relative arrangement of the chromophores. For a dimer, this can be illustrated by the famous cases of head-to-tail (J) and side-by-side (H) dimers, both showing new delocalized exciton states (Fig. 1). The Coulomb coupling J_C depends on the distance (R) and magnitude of the monomer transition dipole moment (μ), the angle between the transition moments (θ), and the local dielectric constant (ϵ):

$$J_C = \mu^2(1 - 3 \cos^2 \theta)/4\pi\epsilon R^3 \quad (3)$$

The Coulomb coupling of the monomer transitions leads to two new excitonic transitions. The in-phase combination of the monomer transition dipole moments leads to an allowed transition, shifted by J_C relative to the monomer. For the out-of-phase combination, the transition dipoles cancel out and lead to a forbidden transition, also shifted by J_C relative to the monomer. In J-aggregates with head-to-tail arrangement of the transition dipoles ($0^\circ \leq \theta < 54.7^\circ$) the in-phase combination has the lowest energy, and absorption and emission spectra are found to red-shift according to the exciton coupling strength by

J_C relative to the monomer transition. For the side-by-side arrangement in H-aggregates ($54.7^\circ < \theta \leq 90^\circ$) the allowed in-phase combination has higher energy than the forbidden out-of-phase combination. Since the absorption is dominated by the most allowed transitions, blue-shifts are observed for H-aggregates (absorption to the S_2 exciton state). On the other hand, fluorescence is originating from the lowest excited state (according to Kasha's rule) and will come from the forbidden $S_1 \rightarrow S_0$ transition and appear with a large Stokes shift relative to the dominating exciton absorption band ($S_0 \rightarrow S_2$). Fluorescence from H-aggregates is often weak or completely absent due to the forbidden nature of the transition, which results in low rates of emission compared to the competing non-radiative deactivation pathways. In contrast, fluorescence from J-aggregates can be very efficient due to the high radiative rate and has been actively pursued by many research groups.³¹ However, it is important to remember that the quantum yield is a result of the competition between the radiative rate and the non-radiative rates and cannot be predicted from the aggregate structure alone. From eqn (3) it is clear that dyes with large transition moments (μ), and thus large molar absorption coefficients, are most likely to display strong exciton coupling. As seen from eqn (3) and Fig. 1 any change in the angle between the chromophores will have large impact on the energy and transition probabilities in the aggregate. For crystals, examples of any angle can be found and in intermediate cases, both transitions can be observed (Davydov splitting).^{27,32} In NPs with less ordered structures a range of coexisting conformations can be expected and thus also inhomogeneous photophysical properties of the NPs.

The term excimer is usually used to describe a dimer between an excited and a ground state molecule formed dynamically in solution.³³ Excimers are mainly formed by planar aromatic hydrocarbons, pyrene being the textbook example, and are characterized by a broad redshifted emission from their excited state delocalized over the π -stack dimer. In molecular solids similar emission features can be observed from some π -stack materials that either are already in a geometry favourable for excimer formation or due to structural relaxation in more dynamic systems favouring the excimer state.^{27,34} Since the excimer states are completely different from the monomer excited state with lower energy and different radiative and non-radiative rates, the formation of excimer states will lead to large redshifts in emission and often significant quenching.

The perspective by Bialas *et al.* provides a thorough discussion of different approaches for direct dye assembly to achieve specific aggregate structures with predictable properties such as excimers or J-aggregates.³⁵ Their argument is that better understanding of the photophysical processes occurring in the solid-state will help in the development of new fluorescent solid-state materials. However, even the simplest systems require thorough analysis of aggregate photophysics and the control of aggregate structure might prove even more difficult for NPs that also need to be stable in water and biological systems. In an example by Pivonski *et al.*³⁶ they carefully

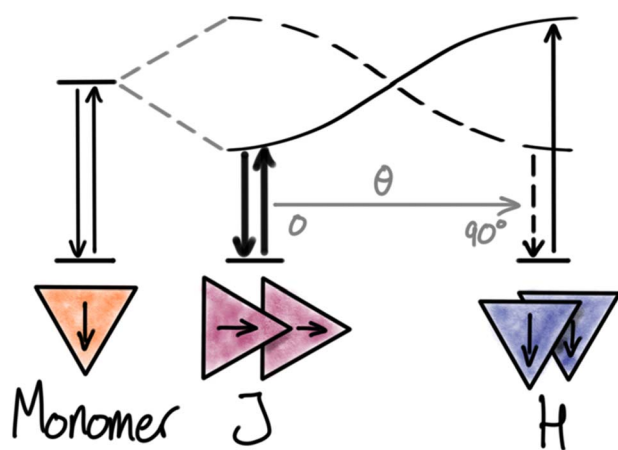


Fig. 1 Kasha's exciton model for dimers. Relative orientation as a function of the angle (θ) between the transition dipole moments of dyes generating J- and H-aggregates with new electronic transitions.



designed a D- π -A- π -D structure, which forms a semi-crystalline J-aggregate structure with a size of ~ 3.5 nm named J-dots.

Another example by Chen *et al.*³⁷ shows the encapsulation of the cyanine dye IR-140 in a hollow silica nanoparticle; the close packing results in strong interaction and J-aggregated emission in the infrared region. For the above-mentioned examples, each of the particles have their advantages. However, the formation of new emitter states by strong interactions in general leads to unpredictable photophysical properties and therefore can require a lot of trial and error before favourable photophysical properties may be achieved.

Energy migration and quenching

Energy migration is a key feature of dye based NPs. Even when the chromophores are sufficiently separated to avoid strong interactions, long range weak Coulomb interactions still mediate efficient energy transfer. This non-radiative Coulomb energy transfer is also known as resonance energy transfer (RET) or Förster resonance energy transfer (FRET).^{27,38,39} While energy migration between identical dyes does not itself change the photophysical properties, it often facilitates fluorescence quenching by bringing the excitation to a site where quenching occurs. In this way energy migration contributes to the phenomena of ACQ to a large extent. In a NP containing only one type of dye, homo energy transfer can occur as long as the dyes are close enough and there is a spectral overlap between absorption and emission of the chromophore. With efficient energy transfer, the excited state (the exciton) can migrate in the NP visiting many dyes during its lifetime (τ), until it eventually deactivates radiatively or non-radiatively. However, if the exciton reaches a site with a structural defect or impurity of lower energy it may get trapped there and deactivate (non-radiatively) on the trap (Fig. 2). If energy migration is efficient even a very low density of such dark trap-states may quench a large fraction or all of the NP emission.⁴⁰ The most clear indication of such energy transfer mediated quenching is shortened multi-exponential fluorescence decays as the density of dye or traps in NPs is increased.⁴¹ In the absence of traps, mobile excitons (high loading of dye) will have the same mono-

exponential decay and lifetime (τ) as localized excitons in a diluted system. However, in systems with energy migration and traps the lifetime of the excitons will depend on their proximity to a trap/quencher site leading to multi-exponential decays and shortened lifetimes. The efficiency of the energy transfer process itself can, in disordered/isotropic systems, be assessed by measuring the rate of anisotropy loss.⁴²

The origin of dark trap states in dye based NPs is in most cases unknown and difficult to assess due to their low concentration and non-emissive nature. However, some likely origins can be considered based on structure and properties of dyes and NPs, and from knowledge of trap states in other optical materials. Traps can originate from impurities in the dye stock. Most commercial dyes have a purity $>95\%$, the impurities could originate from synthetic by-products or degradation (*e.g.* photo degradation) of the dye. Such impurities can either have a low excited state energy and thus act as the trap state, or they can quench the exciton by electron transfer, in which case the trap state is a charge separated photoinduced electron transfer (PET) state.^{43–45} The trap state can also be a “structural defect” in the NP, like a dark H-dimer or excimer occurring in a system where the majority of dyes are organized in an emissive structure such as J-aggregates or separated by a matrix. For small NPs where the surface to volume ratio is large, surface traps may be an important contribution to quenching. Inorganic QDs illustrate this well, as they are highly crystalline materials and it is well known that the dominating sites of quenching are on the surface, and it is therefore essential to passivate these surface defects and protect the QDs from the surrounding environment.^{5,46} In relation to organic dye based nanoparticles it is much less clear how important surface quenching is, and it likely varies between different optical materials. However, Chen *et al.* found that introduction of an amphiphilic ligand, DSPE-PEG, to dye based SMILES NPs not only increased stability in water but also slightly increased the QY.⁴⁷ For some dyes contact with water or dissolved oxygen is known to reduce the quantum yield and it is likely that such quenching processes occur at the NP surface and become amplified by the energy migration in NPs.⁴⁸

The relative efficiency of energy migration for dyes can be estimated from their Förster distance (R_0) for homo-FRET. Here R_0 reports the distance between two identical dyes at which the likelihood of energy transfer is 50%. Since energy migration is a Coulomb interaction, R_0 scales with the transition moments and the spectral overlap between fluorescence and absorption. This allows us to predict that dyes with strong transitions (large molar absorption coefficient, ϵ) and small Stokes shifts will also make up materials and NPs with the most efficient energy migration and thus larger susceptibility to quenching by trap states. For good chromophores the R_0 for homo FRET can reach up to 60 Å.^{18,41}

Efficient energy migration in NPs can however also be utilized as an advantage. Examples of this includes NP blinking by reversible formation of dark states quenching the entire NP allowing their use in super resolution microscopy⁴⁰ and NPs acting as antenna systems for acceptor dyes as discussed below.

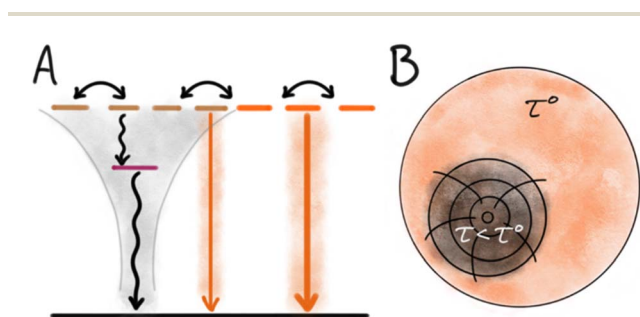


Fig. 2 Quenching by trap states. (A) Schematic state diagram with indication of energy migration between multiple similar dyes in the NPs. Presence of a dark trap state leads to full or partial quenching of many nearby dyes. (B) Illustration of the impact of a trap state able to quench a large volume of the NPs and result in shortened multi-exponential lifetimes.



Table 1 Relation between fluorophore loading and concentration^a

wt%	C (mol L ⁻¹)	V_{dye} (nm ³)	r dye-dye (Å)
100	2.00	0.8	12
30.0	0.60	2.8	17
10.0	0.20	8.3	25
1.00	0.02	83	54
0.10	0.002	830	117

^a Calculated assuming formula weight 500 g mol⁻¹ and mass density 1 g cm⁻³.

Inner filter effects

Both primary (excitation attenuation) and secondary (self-absorption) inner filter effects are common issues in optical spectroscopy of high optical density materials.^{49–51} In the case of NPs with high dye concentration, these effects should be kept in mind. A large effect will most likely only be an issue when working with highly absorbing dyes (high ϵ) in larger NPs.⁵² However, primary inner filter effects that can lead to deviation in quantitative measurements such as quantum yield and excitation spectra may appear even in smaller particles. For a NP with 0.6 mol L⁻¹ concentration of a dye (Table 1) with $\epsilon \approx 10^5$ M⁻¹ cm⁻¹ this could be relevant already when sizes exceed 20 nm. In the presence of secondary inner filter effects, the blue side of the emission spectrum will be truncated due to reabsorption and as a consequence the spectrum appears red shifted.^{41,49} Reabsorption will also lead to stretching of the fluorescence decay in time resolved measurements resulting in apparently longer fluorescence lifetimes.^{53,54}

Parameters describing NP performance

Many approaches and materials are applied for dye based NPs by different research groups who also report different parameters. This can make it difficult to compare how different approaches deliver the best possible photophysical properties. Therefore, it is desirable to use some key parameters that allow for comparison between the many different NP types. Klymchenko and co-workers recently reviewed this in detail.^{11,52} Here we will only briefly discuss the parameters that we find most important.

Brightness per volume

A commonly used parameter to evaluate the performance of an emitter is the brightness (B) defined as the product of the fluorescence quantum yield and the molar absorption coefficient ($B = \Phi \times \epsilon$). When comparing different emitters this number usually gives a good idea of the relative emission output that can be achieved at a given excitation power. For dye based NPs the brightness can be given as $B = \Phi \times \epsilon \times n$, where n is the number of dyes in the particle. This makes sense when comparing specific NPs for imaging applications.⁴ However, n , and thus B , depends strongly on the NP size making the

brightness values misleading in the comparison of different materials and NP constructs. Reisch and Klymchenko therefore proposed the use of brightness per volume (B_V) instead, normalised by using the nanoparticle volume V_{NP} :^{11,52}

$$B_{V,\text{NP}} = \Phi \epsilon n / V_{\text{NP}} \quad (4)$$

Another way to calculate B_V is to normalise the brightness to the average volume occupied by each dye.

$$B_V = \Phi \epsilon / V_{\text{dye}} \quad (5)$$

Here V_{dye} is the average volume in nm³ containing one dye in the NP. By using brightness per volume, it is possible to compare the emission efficiency of all types of dye based NPs irrespective of the NP size and size distribution. For chromophores of similar size, V_{dye} also provides an interesting measure of how efficiently the chromophoric units are packed in the NP. V_{dye} in the NP can *e.g.* be compared to the volume of the dye obtained from calculations or crystal structures.⁵⁵

Another key parameter that can differ between research groups is the description of “dye loading”, with the use of weight percentage dye compared to the total mass of the NP. Sometimes the weight used includes bulky side groups or the bulky anion instead of the actual mass of the chromophore unit. This gives inconsistent values of dye loading. Using the volume per chromophore (V_{dye}) will give a more fair perspective on how much of the NP actually contributes to the absorption and emission. Table 1 relates the fluorophore weight percent to the concentration in NPs and how much volume is used per fluorophore (assuming a fluorophore with formula weight 500 g mol⁻¹, and a mass density of 1 g cm⁻³). At 100 wt% (the ultimate limit) the volume used per dye is below 1 nm³. Some of the brightest NP approaches reach a dye loading of around 10–30 wt% where the volume per fluorophore is increased 3–10 times compared to this ideal scenario.

Fluorescence quantum yield and lifetime

The brightness per volume gives a good indication of the overall performance of the NP. The quantum yield of fluorescence from the NP will often vary depending on the loading due to the ACQ effects discussed above. When evaluating the quantum yield of the dyes in the NP it is tempting to use the solution values for reference. This may be reasonable for rigid dyes that are not sensitive to the solvent viscosity and polarity. However, in general, a better reference will be a highly diluted NP where the dye is immobilized in the same matrix, or a dilute solid solution of the dye in a polymer thin film.⁴¹

Another tool to evaluate NPs is their fluorescent lifetime. Although rarely used in this context it can prove to be a very strong tool to diagnose possible defects. If there is only one type of emitter (and no quenchers) present in the NPs a mono-exponential decay should be observed as it is the case for molecular fluorophores in dilute solid solution. If there are multiple configurations or quenchers present, the lifetime will be shortened and multi-exponential. In the paper by Kacenauskaitė *et al.* the fluorescence decays were used as



a diagnostic tool to elucidate exactly the role of quenching by energy migration to trap sites.⁴¹ In some cases the fluorescence lifetime increases when the fluorophore is encapsulated in NPs *e.g.* stilbenes and cyanines due to the restriction of torsional freedom, which decreases non-radiative deactivation. In particular, in these cases, the use of a solid solution (*e.g.* polymer thin film) as the reference for the intrinsic molecular properties becomes important. The bottom-line is the fluorescence lifetime provides important insights for understanding NP properties.

Nanoparticle design principles

In this section, we will discuss selected NP design strategies. There are many aspects to consider when working with fluorescent NPs; here we will however narrow the discussion to address only the relation between NP structure/composition and photophysical properties. Over the years many different approaches have been reported to create efficient organic based fluorescent NPs. These approaches can be divided into several classes of NP designs listed below.

Nanoparticles with low loading of chromophores

A simple solution to overcome the above-mentioned challenges with ACQ and altered photophysical properties is to encapsulate low concentration of dyes in a matrix that makes up most of the NP. In such diluted dye NPs (Fig. 3A) dyes are on average far apart, randomly distributed in the matrix and thus less likely to aggregate. Energy migration is also disabled when r (dye–dye) $>$ R_0 (homo FRET). There are various types of matrices used to encapsulate dyes in this way. The matrix can be polymer based,^{56–60} and several examples of this type are commercially

available like the polystyrene based FluoSpheres.⁶¹ Another type that was used early on with low dye loading is silica-based nanoparticles.^{62–65} NPs with low loading of chromophores is a reliable strategy for generating fluorescent dye based NPs. However, as soon as the loading is increased to obtain higher brightness, ACQ will become dominating. This is due to the formation of quenched aggregates or dimers as well as energy migration to these trap sites (Fig. 3B) as the average dye–dye distance becomes comparable to or smaller than R_0 (Table 1). This proves to be a severe limitation for these types of NPs as the dye loading has to be kept very low and essentially most of the NP structure is the non-emissive matrix surrounding the dyes.

In the same category, we find emulsion based nanoparticles, using a water immiscible core and surfactant shell or specially designed amphiphilic dyes, which have been reported as fluorescent nanoemulsions (NEs).^{66,67} To ensure high efficiency of these types of emulsions the dyes have to be specifically designed to reduce both ACQ and leakage. In the work by Collot *et al.* different fluorophore families were used in combination with various oils to form NEs.⁶⁸ The viscous oil also reduces deactivation by conformational relaxation of some of the fluorophores, thereby increasing the QY drastically compared to solution properties. Still this approach requires low concentration of the dye between 0.25 wt% to 2.5 wt% where the QY is approximately halved. Later a bulky anion tetraphenylborate (TPB) was introduced to increase the loading of cationic dyes in the oily core up to 8 wt% dye while still maintaining separation of the chromophores and keeping ACQ effects at a minimum.⁶⁹

Another approach to fluorescent NEs uses the so-called fluorofluorophores that are dyes equipped with fluorinated side chains making them soluble in perfluorocarbons.^{70,71} In work by Sletten *et al.* highly fluorinated rhodamine and coumarin chromophores were used to form fluorescent perfluorocarbon

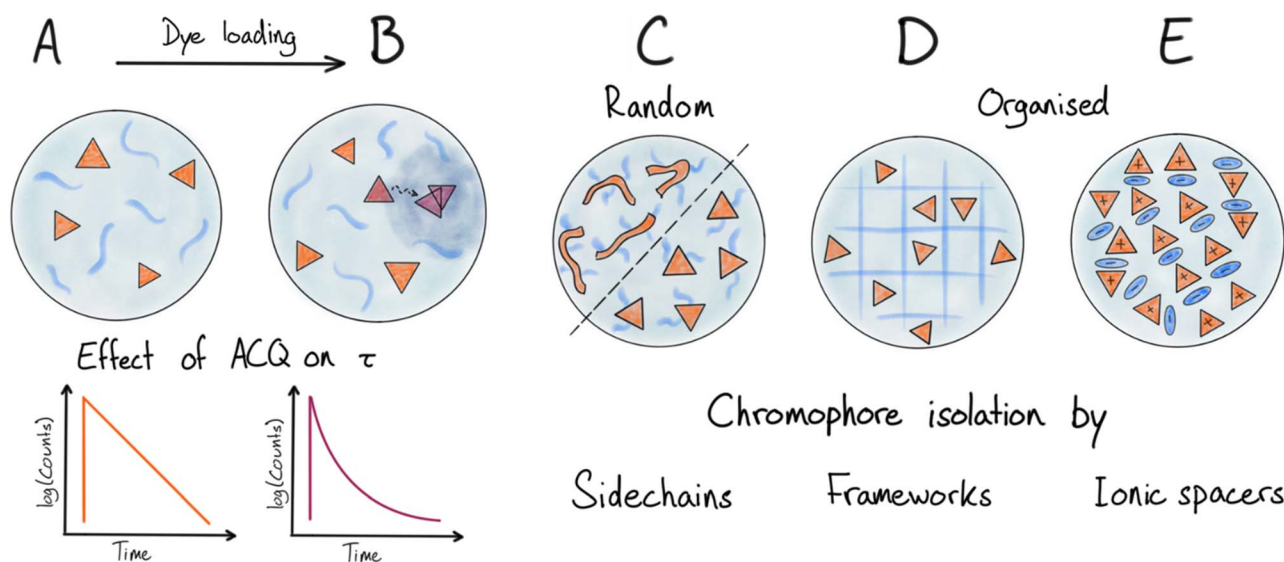


Fig. 3 Illustrations of nanoparticle design principles. (A and B) Low to medium concentration of chromophores in matrix, and effect on fluorescence lifetime upon beginning aggregation and energy transfer mediated quenching as dye loading increases. (C), (D) and (E) Illustrate various strategies to limit chromophore interactions and mitigate ACQ at high dye densities: (C) covalently attached bulky side chains on dyes and semiconducting polymers. (D) Organization of dyes in an ordered framework/lattice. (E) Organization and separation of ionic dyes by large counterions.



emulsions with sizes of around 120 nm.⁷⁰ Recently fluorinated cyanine dyes extending to the far red region were also introduced.⁷² NEs of these fluoro-cyanine dyes ranged in size from 190–250 nm and were found to be more stable to leaching compared to the previously reported fluorinated rhodamine. Still in all these NEs the dye loading is very low (<0.1 wt%).

Syntheses of specific amphiphiles containing chromophores to form fluorescent micelles and vesicles have also been reported,^{73–75} which can also be obtained by post functionalizing with fluorophores after micelle formation.⁷⁶

In all of the above examples, the dyes have been randomly embedded (dissolved) in a matrix material. As the dye loading is increased formation of dimers or aggregates will take place (Fig. 3B). Even with a matrix that dissolves the dyes well ACQ is observed at relative low loadings of a few wt% or lower. This can be rationalized as a result of formation of aggregates/dimers with low emission quantum yield (*e.g.* H-aggregates) that not only are self-quenching but also quench nearby monomer dyes by acting as energy acceptors (traps), thereby amplifying the ACQ. As mentioned above this will be particularly true for dyes with large R_0 values for homo-energy transfer where energy migration will extend the range of quenching (Fig. 2 and 3B).

Special organic materials where ACQ is limited by chromophore design

Some materials and chromophore types are more resistant to ACQ than general organic dyes, giving these materials a special advantage for the preparation of very bright NPs. Here we will discuss two such cases: polydots and the so-called AIEs.

Polydots. Nanoparticles based on semiconducting fluorescent polymers, like those used in OLEDs, used as the emissive material are known as polydots or Pdots.^{12,13,77–80} Semiconducting polymers consist of a π -conjugated backbone with bulky side groups and can be used to produce fluorescent nanoparticles without the use of an additional matrix to dilute the active component. Due to the extended π -conjugation in the polymer backbone, exciton migration through the polymer is highly efficient.^{81,82}

In polydot systems one can consider the covalently attached side chains as a matrix separating the dye components (the π -conjugated polymer backbone). However, contrary to systems with dyes randomly dissolved in polymer matrices, much higher effective loading is achieved since the backbones are kept from strong interaction by a much smaller fraction of the matrix (side chains) fixed in specific positions around the π -system (Fig. 3C).^{83,84}

Because of their use in the production of OLEDs, there are already many types of semiconducting polymers available with different spectral properties and optimized for high quantum yields in the solid state. However, the development of new conjugated polymers can be time consuming and difficult. Designing new polydots with entirely new properties can therefore be a demanding process. The optical properties also slightly change during the formation of NPs compared to its solution properties. This is likely due to changes in the conjugation length and conformations of the polymer backbone,

which may broaden and shift both absorption and emission.^{12,85,86} This can also induce quenching trap states in the structure which will influence QY and fluorescence lifetime. Piwonski *et al.* introduced non-planar conjugated polymers PCzBT and PCzDTBT for producing ultra-small polydots with sizes of around 3 nm.⁸⁷ The quantum yield of the PCzDTBT polydots were lower than that of the corresponding polymer in THF, 0.2 and 0.5, respectively. Later they expanded the polydot spectroscopic range toward NIR emission with reasonable quantum yields.⁸⁸ Further information on polydots can be found in the review by Chiu and Wu.⁷⁷

AIE nanoparticles. Nanoparticles based on so called aggregation induced emission (AIE) chromophores have attracted a lot of attention in recent years.^{89–91} Aggregation induced emission is, as Würthner points out, in itself not a new phenomenon as similar structures with emission turn-on in the solid state have been known for a long time.⁹² Yet, since 2011 Bin Liu, Ben Zhong Tang and others have expanded the research area into several branches including AIE nanoparticles.^{93–96} The chromophores are commonly designed with flexible π -systems that in solution efficiently relax non-radiatively by twisting. In the solid state, however, these motions are locked, which enables fluorescence while the contorted non-planar structure also limits quenching by π - π -stacking and strong interactions. Other features of most AIE dyes are moderate absorption coefficients and radiative rates, which stem from the twisted π -systems, and large relaxation of the excited state causing a large Stokes shift. These properties will reduce both exciton coupling and energy migration, and are likely important aspects of their resistance to ACQ.

General methods for suppressing ACQ

Several different strategies have been introduced to increase the loading of fluorescent dyes in general. A higher loading of dyes can be achieved by actively preventing close contact like in π - π -stacking. This is possible by introducing large or bulky groups either on the chromophore itself or on the counter ions associated with charged dyes.

Dyes with bulky side groups. By introducing large bulky side groups on the chromophores ACQ is limited as the π -stacking is hindered (Fig. 3C). This is a strategy very similar to that used for semiconductor polymers. This allows for a higher loading in polymer particles. An example is the modification of perylene diimides (PDIs) with bulky side groups at both bay and imide positions for improving fluorescence efficiency in high dye loaded polymer films⁹⁷ and NPs with sizes of 40–50 nm.⁹⁸ This allows for an increase in the NP dye loading up to 5 wt%, beyond which QY decreases substantially. The Würthner group has introduced even bulkier substituents to PDIs and achieved highly improved molecular fluorescence from neat solid state materials.^{99,100} In this case the PDI chromophore accounts for ~23% of the mass of the material while the bulky side groups account for the rest. These sidechain protected PDIs are yet to be applied in fluorescent NPs.

Framework isolation. Chromophore separation can also be achieved by encapsulation in well-defined lattices such as



metal-organic frameworks (MOFs)^{101,102} or zeolites¹⁰³(Fig. 3D). For MOFs, where the lattice consists of metal ions linked by organic bridging ligands, the chromophore can either be encapsulated in the pores or used as the bridging ligand. This allows for precise control of content and packing and thereby avoiding ACQ and formation of dimer traps. Zeolite lattices are limited to only containing chromophores in their porous network but there are still some examples displaying their potential as fluorescent NPs.^{104,105}

Ionic organization. Another strategy for organizing dyes makes use of electrostatically driven ion pair formation between charged dyes and various counterions. A class of materials with similar features to ionic liquids called GUMBOs¹⁰⁶ (group of uniform materials based on organic salts) has been used to produce a type of fluorescent NP. The GUMBOs and nano-GUMBOs utilize large counterions to achieve liquid crystalline like packing. The large counterions also partly function as spacers between the cationic organic dyes and thereby reduce ACQ, yet the optical properties are still distorted.^{107–110} With the nanoGUMBO approach, control of the internal structure of the particle is still a challenge and a more directed approach seems to be needed to counteract ACQ and achieve more efficient fluorescence.

Counterion separation. Ionic organization is also used in polymer NPs loaded with cationic dyes paired with the large counterion tetraphenylborate (TPB) or its larger fluorinated derivatives (Fig. 3E). The use of a large anion increases the separation between the dyes and thereby limits ACQ. Early studies reported by the Yao group showed improved QY of dyes encapsulated in poly(vinylpyrrolidone) polymer using tetraphenylborate (TBP) or tetrakis(4-fluorophenyl)borate (TFBP) anions.^{111–113} This concept has been further developed and improved by the Klymchenko group showing large improvement of the fluorescent properties with even larger anions. In 2015 Shulov *et al.*¹¹⁴ screened TBP and several fluorinated derivatives with rhodamine B without a polymer matrix. Here it was shown that it was possible to produce NPs with sizes ranging from 25 nm to 45 nm with the different counterion derivatives. The QY of the particles depend significantly on the counterion, and the non-fluorinated TBP yielded particles with low QY while the most fluorinated derivative yielded a QY of around 0.6. In most cases, however, the cationic dyes paired with large bulky anions are loaded into polymers like PMMA and PLGA.¹¹⁵ With this approach, the Klymchenko group has developed a range of very bright NPs and shown how they are able to modulate properties such as size, brightness, and blinking.^{40,42,116} These NPs have successfully been applied for sensing of nucleic acids^{117–119} including microRNA,¹²⁰ by exploiting energy transfer within the NP core and to chromophores attached to the surface. This is discussed extensively in the reviews by Klymchenko and coworkers.^{11,52} Recently the Klymchenko group also showed that the fluorinated TPB anions can be exchanged for a less cytotoxic alkylated barbiturate anion.¹²¹ This strategy of loading polymer NPs with cationic dyes paired with large counterions has in cases yielded dye loadings up to 0.22 mol L⁻¹; however, these still show reduced QY compared to NPs with lower dye content.¹¹⁵

Supramolecular counterions. Supramolecular organization of charged π -systems in the solid state and nanostructures through ion-pairing assembly has been used by many groups and has recently been explored and discussed by Maeda and co-workers.^{122,123} In this context, Maeda and co-workers showed how a neutral anion receptor can be used to convert small spherical anions such as Cl⁻ and BF₄⁻ into larger planar anion-receptor complexes, favouring alternating stacking with planar cations.¹²⁴ Recently, the Flood group in collaboration with the Laursen group used a similar approach to organize fluorescent dyes in solids while counteracting ACQ. This new supramolecular concept for dye based fluorescent materials was named small-molecule ionic isolation lattices (SMILES).⁵⁵ Addition of the macrocyclic anion receptor cyanostar to cationic fluorescent dyes converts anions like ClO₄⁻, PF₆⁻ or BF₄⁻ to large disc-shaped anion complexes that direct alternating charge-by-charge packing and ensure efficient isolation of cationic fluorophores (Fig. 3E). The anion complex produced with cyanostar displays a large electronic bandgap, which ensures that the cationic dyes are isolated both structurally and electronically. The SMILES approach allows for easy use of commercially available cationic dyes and has been demonstrated for a range of cationic dye families such as cyanines, rhodamines, oxazines, styryls and trianguleniums, which all are organized in similar isolation lattices with 13–18 Å separation of the dyes in all directions and a volume per dye (V_{dye}) \approx 3.5 nm³, corresponding to 0.5 mol L⁻¹. Chen *et al.* reported a simple nanoprecipitation method to produce bright \sim 20 nm SMILES NPs in water stabilized by a shell of DSPE-PEG detergent.⁴⁷ For a rhodamine derivative, a QY of up to 0.3 was reported. Furthermore, UV excitation of the strongly absorbing cyanostar anion complex can result in energy transfer to the embedded fluorophores further increasing the brightness, showing that the anion complexes not only function as spacers but also as an antenna system.¹²⁵

Doping and mix of strategies

Most of the strategies discussed above reduce ACQ by separating the chromophores (dyes) to avoid strong interactions. This is achieved using various matrices, spanning from polymers and silica acting as simple solvents with random distribution of the dyes, to highly organized structures like MOFs and SMILES. To achieve high brightness per volume a high density of chromophores is needed, which in turn requires use of an effective strategy to keep the chromophores separated. However, even when effective separation of dyes is obtained in NPs, fluorescence QYs do not reach the values expected from the chromophores in solid solution. Much of the remaining quenching that limits the performance of NPs with high chromophore density stems from the highly efficient energy migration making them sensitive to even single trap states.¹¹⁵ Compromises are often made where the number of traps or the energy transfer efficiency is reduced by further increasing the distance between chromophores *e.g.* by introducing non absorbing spacers either as an additional matrix or as “blank salts” in the case of ionic organization.^{41,126,127} For dye loaded



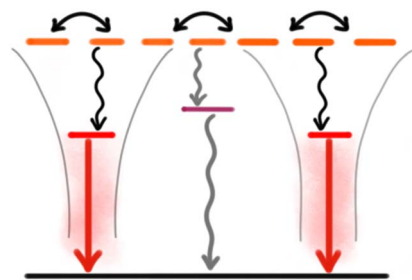


Fig. 4 Schematic state diagram of a doped NP system with indication of energy migration between multiple similar (donor) dyes in the NP. Efficient energy transfer to a low number of isolated dopant (acceptor) dyes outcompetes quenching by the trap state.

polymer NPs the local density of dyes can also be modulated by the polarity of the polymer matrix to either favour segregation or isotropic distribution of dyes.¹¹⁵

A promising approach is to use the efficient energy migration as an advantage by introducing a red shifted dopant emitter into the NP. A small fraction of dopant emitters can efficiently harvest most excitons and thus outcompete energy migration to the dark trap states (Fig. 4). This doping strategy was demonstrated for several SMILES materials showing large improvement of quantum yields and fluorescence lifetimes approaching the intrinsic molecular values and was since translated to SMILES NPs.^{41,128} Introduction of dopant dyes in the NP core has been used in several NP strategies to boost the fluorescence brightness,^{42,64,129–131} while introduction of acceptor dyes to the NP surface also has been used to develop sensing properties *e.g.* for DNA/RNA hybridization assays.^{119,120} Dopant dyes have also been used to achieve more narrow and redshifted emission of AIE NPs and polydots.^{44,131} In a recent example by Zhou *et al.* they introduced acceptor fluorophores into a AIEdot and utilize effective energy transfer while isolating the molecular chromophores to achieve more narrow emission compared to that of the AIEdot alone.¹³⁰

Nevertheless, doping with acceptor dyes is not a universal solution to ACQ and the brightness problem. Some dye combinations have negative impact or no effect at all on the optical properties. An example by Wu *et al.* shows polydots doped with 3 different molecular dyes where only 2 combinations show improved fluorescence intensity.⁴⁴ This decrease could be due to quenching of the dopant dye by PET. Such PET quenching of dopant dyes can be avoided in well-organized systems like SMILES.⁴¹

Future outlook

A lot of progress in the development of organic dye based NPs has been achieved in recent years, and several materials and strategies are now available that effectively avoid strong interactions and thus allow for NPs with high chromophore density and high brightness per volume. Doping strategies additionally offer a means to further improve brightness and tuning of spectral properties. In the race for high brightness, polydots

and counterion based methods are now approaching the limit set by the separating matrix and the intrinsic properties of the dyes. Still we believe that dye based NPs have much more to offer in terms of advanced photophysical properties.

NPs with NIR emission are particularly interesting for deep tissue imaging.^{132–134} NPs based on NIR dyes are however still challenging and often show low quantum yields.^{135,136} Typical NIR dyes like cyanines present challenges stemming from their large π -surfaces and tendency for aggregation, large oscillator strength that also favours strong coupling, their sensitivity to quenching by water and generally very fast non-radiative deactivation rates as dictated by the energy gap law.^{48,137} Strategies that will address these challenges and provide bright tuneable NIR-I and NIR-II NPs will be highly valuable.

New and advanced fluorescence based sensing and imaging modalities are often based on photophysical properties beyond just emission wavelength and brightness. This is the case for *e.g.* time-resolved techniques,^{58,128,138} super resolution,⁴⁰ and circular polarized emission.¹³⁹ Development of dye based NPs specifically for such readouts is a promising direction.

Inclusion of multiple different chromophores in NPs may provide new and advanced photophysical properties *via* supramolecular photophysics – examples include engineering of *e.g.* Stokes shifts and emission lifetimes *via* doping (discussed above). However, a limitation for all of the present NP strategies is the stochastic nature of these composites, and more organized structures with control of relative distances, orientations and locations of the dyes within the NP could greatly widen and improve the scope of dye based NPs.

Conclusions

This perspective discusses key aspects that must be considered when working with fluorescent organic dye based NPs. We discuss the various photophysical effects that limit brightness of fluorescent NPs; strong dye–dye interactions, and the presence of trap sites, and energy migration to these. On this background the properties and performance of various NP designs can be understood and hopefully even improved. We believe that dye based NPs have great potential stemming from the possible transfer of advanced properties from the molecular dyes into NPs when the organization and supramolecular interactions are well understood and controlled.

Author contributions

SGS and BWL conceived the idea and co-wrote the perspective.

Conflicts of interest

There are no conflicts to declare.

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References

- W. R. Algar, Heroes or Villains? How Nontraditional Luminescent Materials Do and Do Not Enhance Bioanalysis and Imaging, *Chem. Mater.*, 2020, **32**(12), 4863–4883.
- O. S. Wolfbeis, An overview of nanoparticles commonly used in fluorescent bioimaging, *Chem. Soc. Rev.*, 2015, **44**(14), 4743–4768.
- Z. Li, Q. Sun, Y. Zhu, B. Tan, Z. P. Xu and S. X. Dou, Ultra-small fluorescent inorganic nanoparticles for bioimaging, *J. Mater. Chem. B*, 2014, **2**(19), 2793–2818.
- W. R. Algar, M. Massey, K. Rees, R. Higgins, K. D. Krause, G. H. Darwish, W. J. Peveler, Z. Xiao, H.-Y. Tsai, R. Gupta, K. Lix, M. V. Tran and H. Kim, Photoluminescent Nanoparticles for Chemical and Biological Analysis and Imaging, *Chem. Rev.*, 2021, **121**(15), 9243–9358.
- A. P. Alivisatos, Semiconductor Clusters, Nanocrystals, and Quantum Dots, *Science*, 1996, **271**(5251), 933.
- U. Resch-Genger, M. Grabolle, S. Cavaliere-Jaricot, R. Nitschke and T. Nann, Quantum dots versus organic dyes as fluorescent labels, *Nat. Methods*, 2008, **5**(9), 763–775.
- F. P. García de Arquer, D. V. Talapin, V. I. Klimov, Y. Arakawa, M. Bayer and E. H. Sargent, Semiconductor quantum dots: Technological progress and future challenges, *Science*, 2021, **373**(6555), eaaz8541.
- K. D. Wegner and N. Hildebrandt, Quantum dots: bright and versatile *in vitro* and *in vivo* fluorescence imaging biosensors, *Chem. Soc. Rev.*, 2015, **44**(14), 4792–4834.
- A. Gnach and A. Bednarkiewicz, Lanthanide-doped up-converting nanoparticles: Merits and challenges, *Nano Today*, 2012, **7**(6), 532–563.
- U. Resch-Genger and H. H. Gorris, Perspectives and challenges of photon-upconversion nanoparticles – Part I: routes to brighter particles and quantitative spectroscopic studies, *Anal. Bioanal. Chem.*, 2017, **409**(25), 5855–5874.
- A. Reisch and A. S. Klymchenko, Fluorescent Polymer Nanoparticles Based on Dyes: Seeking Brighter Tools for Bioimaging, *Small*, 2016, **12**(15), 1968–1992.
- J. Pecher and S. Mecking, Nanoparticles of Conjugated Polymers, *Chem. Rev.*, 2010, **110**(10), 6260–6279.
- L. R. MacFarlane, H. Shaikh, J. D. Garcia-Hernandez, M. Vespa, T. Fukui and I. Manners, Functional nanoparticles through π -conjugated polymer self-assembly, *Nat. Rev. Mater.*, 2021, **6**(1), 7–26.
- M. O. Alas, F. B. Alkas, A. Aktas Sukuroglu, R. Genc Alturk and D. Battal, Fluorescent carbon dots are the new quantum dots: an overview of their potential in emerging technologies and nanosafety, *J. Mater. Sci.*, 2020, **55**(31), 15074–15105.
- S. K. Bhunia, A. Saha, A. R. Maity, S. C. Ray and N. R. Jana, Carbon Nanoparticle-based Fluorescent Bioimaging Probes, *Sci. Rep.*, 2013, **3**(1), 1473.
- B. N. G. Giepmans, T. J. Deerinck, B. L. Smarr, Y. Z. Jones and M. H. Ellisman, Correlated light and electron microscopic imaging of multiple endogenous proteins using Quantum dots, *Nat. Methods*, 2005, **2**(10), 743–749.
- L. D. Lavis and R. T. Raines, Bright Ideas for Chemical Biology, *ACS Chem. Biol.*, 2008, **3**(3), 142–155.
- J. R. Lakowicz, *Principles of fluorescence spectroscopy*, Springer, 3rd edn, 2006.
- H.-S. Peng and D. T. Chiu, Soft fluorescent nanomaterials for biological and biomedical imaging, *Chem. Soc. Rev.*, 2015, **44**(14), 4699–4722.
- S. Y. Lim, W. Shen and Z. Gao, Carbon quantum dots and their applications, *Chem. Soc. Rev.*, 2015, **44**(1), 362–381.
- K. Jiang, S. Sun, L. Zhang, Y. Lu, A. Wu, C. Cai and H. Lin, Red, Green, and Blue Luminescence by Carbon Dots: Full-Color Emission Tuning and Multicolor Cellular Imaging, *Angew. Chem., Int. Ed.*, 2015, **54**(18), 5360–5363.
- Y. Song, S. Zhu and B. Yang, Bioimaging based on fluorescent carbon dots, *RSC Adv.*, 2014, **4**(52), 27184–27200.
- J. Liu, R. Li and B. Yang, Carbon Dots: A New Type of Carbon-Based Nanomaterial with Wide Applications, *ACS Cent. Sci.*, 2020, **6**(12), 2179–2195.
- V. Strauss, J. T. Margraf, C. Dolle, B. Butz, T. J. Nacken, J. Walter, W. Bauer, W. Peukert, E. Spiecker, T. Clark and D. M. Guldi, Carbon Nanodots: Toward a Comprehensive Understanding of Their Photoluminescence, *J. Am. Chem. Soc.*, 2014, **136**(49), 17308–17316.
- S. Tao, T. Feng, C. Zheng, S. Zhu and B. Yang, Carbonized Polymer Dots: A Brand New Perspective to Recognize Luminescent Carbon-Based Nanomaterials, *J. Phys. Chem. Lett.*, 2019, **10**(17), 5182–5188.
- M. Pope and C. E. Swenberg, *Electronic processes in organic crystals and polymers*, Oxford University Press, Oxford, 2nd edn, 1999.
- J. Gierschner, J. Shi, B. Milián-Medina, D. Roca-Sanjuán, S. Varghese and S. Park, Luminescence in Crystalline Organic Materials: From Molecules to Molecular Solids, *Adv. Opt. Mater.*, 2021, **9**(13), 2002251.
- S. J. Strickler and R. A. Berg, Relationship between Absorption Intensity and Fluorescence Lifetime of Molecules, *J. Chem. Phys.*, 1962, **37**(4), 814–822.
- M. Kasha, H. R. Rawls and M. A. El-Bayoumi, The exciton model in molecular spectroscopy, *Pure Appl. Chem.*, 1965, **11**(3–4), 371–392.
- N. J. Hestand and F. C. Spano, Expanded Theory of H- and J-Molecular Aggregates: The Effects of Vibronic Coupling and Intermolecular Charge Transfer, *Chem. Rev.*, 2018, **118**(15), 7069–7163.
- F. Würthner, T. E. Kaiser and C. R. Saha-Möller, J-Aggregates: From Serendipitous Discovery to Supramolecular Engineering of Functional Dye Materials, *Angew. Chem., Int. Ed.*, 2011, **50**(15), 3376–3410.
- A. Austin, N. J. Hestand, I. G. McKendry, C. Zhong, X. Zhu, M. J. Zdilla, F. C. Spano and J. M. Szarko, Enhanced Davydov Splitting in Crystals of a Perylene Diimide Derivative, *J. Phys. Chem. Lett.*, 2017, **8**(6), 1118–1123.



- 33 B. Stevens and E. Hutton, Radiative Life-time of the Pyrene Dimer and the Possible Role of Excited Dimers in Energy Transfer Processes, *Nature*, 1960, **186**(4730), 1045–1046.
- 34 R. D. Pensack, R. J. Ashmore, A. L. Paoletta and G. D. Scholes, The Nature of Excimer Formation in Crystalline Pyrene Nanoparticles, *J. Phys. Chem. C*, 2018, **122**(36), 21004–21017.
- 35 D. Bialas, E. Kirchner, M. I. S. Röhr and F. Würthner, Perspectives in Dye Chemistry: A Rational Approach toward Functional Materials by Understanding the Aggregate State, *J. Am. Chem. Soc.*, 2021, **143**(12), 4500–4518.
- 36 H. Piwoński, S. Nozue, H. Fujita, T. Michinobu and S. Habuchi, Organic J-Aggregate Nanodots with Enhanced Light Absorption and Near-Unity Fluorescence Quantum Yield, *Nano Lett.*, 2021, **21**(7), 2840–2847.
- 37 W. Chen, C.-A. Cheng, E. D. Cosco, S. Ramakrishnan, J. G. P. Lingg, O. T. Bruns, J. I. Zink and E. M. Sletten, Shortwave Infrared Imaging with J-Aggregates Stabilized in Hollow Mesoporous Silica Nanoparticles, *J. Am. Chem. Soc.*, 2019, **141**(32), 12475–12480.
- 38 T. Förster, Energy migration and fluorescence, *J. Biomed. Opt.*, 2012, **17**(1), 011002.
- 39 W. R. Algar, N. Hildebrandt, S. S. Vogel and I. L. Medintz, FRET as a biomolecular research tool — understanding its potential while avoiding pitfalls, *Nat. Methods*, 2019, **16**(9), 815–829.
- 40 A. Reisch, P. Didier, L. Richert, S. Oncul, Y. Arntz, Y. Mély and A. S. Klymchenko, Collective fluorescence switching of counterion-assembled dyes in polymer nanoparticles, *Nat. Commun.*, 2014, **5**(1), 4089.
- 41 L. Kacenauskaite, S. G. Stenspil, A. H. Olsson, A. H. Flood and B. W. Laursen, Universal Concept for Bright, Organic, Solid-State Emitters—Doping of Small-Molecule Ionic Isolation Lattices with FRET Acceptors, *J. Am. Chem. Soc.*, 2022, **144**(43), 19981–19989.
- 42 K. Trofymchuk, A. Reisch, P. Didier, F. Fras, P. Gilliot, Y. Mely and A. Klymchenko, Giant light-harvesting nanoantenna for single-molecule detection in ambient light, *Nat. Photonics*, 2017, **11**(10), 657–663.
- 43 J.-S. Yang and T. M. Swager, Porous Shape Persistent Fluorescent Polymer Films: An Approach to TNT Sensory Materials, *J. Am. Chem. Soc.*, 1998, **120**(21), 5321–5322.
- 44 C. Wu, Y. Zheng, C. Szymanski and J. McNeill, Energy Transfer in a Nanoscale Multichromophoric System: Fluorescent Dye-Doped Conjugated Polymer Nanoparticles, *J. Phys. Chem. C*, 2008, **112**(6), 1772–1781.
- 45 H. J. Egelhaaf, J. Gierschner and D. Oelkrug, Polarizability effects and energy transfer in quinquethiophene doped bithiophene and OPV films, *Synth. Met.*, 2002, **127**(1), 221–227.
- 46 B. O. Dabbousi, J. Rodriguez-Viejo, F. V. Mikulec, J. R. Heine, H. Mattoussi, R. Ober, K. F. Jensen and M. G. Bawendi, (CdSe)ZnS Core–Shell Quantum Dots: Synthesis and Characterization of a Size Series of Highly Luminescent Nanocrystallites, *J. Phys. Chem. B*, 1997, **101**(46), 9463–9475.
- 47 J. Chen, S. M. A. Fatemina, L. Kacenauskaite, N. Bærentsen, S. G. Stenspil, J. Bredehoeft, K. L. Martinez, A. H. Flood and B. W. Laursen, Ultra-Bright Fluorescent Organic Nanoparticles Based on Small-Molecule Ionic Isolation Lattices, *Angew. Chem., Int. Ed.*, 2021, **60**(17), 9450–9458.
- 48 J. Maillard, K. Klehs, C. Rumble, E. Vauthey, M. Heilemann and A. Fürstenberg, Universal quenching of common fluorescent probes by water and alcohols, *Chem. Sci.*, 2021, **12**(4), 1352–1362.
- 49 J. B. Birks and G. T. Wright, Fluorescence Spectra of Organic Crystals, *Proc. Phys. Soc., London, Sect. B*, 1954, **67**(9), 657.
- 50 R. Katoh, K. Suzuki, A. Furube, M. Kotani and K. Tokumaru, Fluorescence Quantum Yield of Aromatic Hydrocarbon Crystals, *J. Phys. Chem. C*, 2009, **113**(7), 2961–2965.
- 51 S. Kumar Panigrahi and A. Kumar Mishra, Inner filter effect in fluorescence spectroscopy: As a problem and as a solution, *J. Photochem. Photobiol., C*, 2019, **41**, 100318.
- 52 A. H. Ashoka, I. O. Aparin, A. Reisch and A. S. Klymchenko, Brightness of fluorescent organic nanomaterials, *Chem. Soc. Rev.*, 2023, **52**(14), 4525–4548.
- 53 Y. Sakai, M. Kawahigashi, T. Minami, T. Inoue and S. Hirayama, Deconvolution of non-exponential emission decays arising from reabsorption of emitted light, *J. Lumin.*, 1989, **42**(6), 317–324.
- 54 J. S. Connolly, A. F. Janzen and E. B. Samuel, Fluorescence lifetimes of chlorophyll a: solvent, concentration and oxygen dependence, *Photochem. Photobiol.*, 1982, **36**(5), 559–563.
- 55 C. R. Benson, L. Kacenauskaite, K. L. VanDenburgh, W. Zhao, B. Qiao, T. Sadhukhan, M. Pink, J. Chen, S. Borgi, C.-H. Chen, B. J. Davis, Y. C. Simon, K. Raghavachari, B. W. Laursen and A. H. Flood, Plug-and-Play Optical Materials from Fluorescent Dyes and Macrocycles, *Chem*, 2020, **6**(8), 1978–1997.
- 56 T. Behnke, C. Würth, E.-M. Laux, K. Hoffmann and U. Resch-Genger, Simple strategies towards bright polymer particles *via* one-step staining procedures, *Dyes Pigm.*, 2012, **94**(2), 247–257.
- 57 T. Behnke, J. E. Mathejczyk, R. Brehm, C. Würth, F. Ramos Gomes, C. Dullin, J. Napp, F. Alves and U. Resch-Genger, Target-specific nanoparticles containing a broad band emissive NIR dye for the sensitive detection and characterization of tumor development, *Biomaterials*, 2013, **34**(1), 160–170.
- 58 K. Hoffmann, T. Behnke, D. Drescher, J. Kneipp and U. Resch-Genger, Near-Infrared-Emitting Nanoparticles for Lifetime-Based Multiplexed Analysis and Imaging of Living Cells, *ACS Nano*, 2013, **7**(8), 6674–6684.
- 59 K. Hoffmann, T. Behnke, M. Grabolle and U. Resch-Genger, Nanoparticle-encapsulated vis- and NIR-emissive fluorophores with different fluorescence decay kinetics for lifetime multiplexing, *Anal. Bioanal. Chem.*, 2014, **406**(14), 3315–3322.
- 60 D. Kage, L. Fischer, K. Hoffmann, T. Thiele, U. Schedler and U. Resch-Genger, Close Spectroscopic Look at Dye-Stained



- Polymer Microbeads, *J. Phys. Chem. C*, 2018, **122**(24), 12782–12791.
- 61 I. Johnson and M. T. Z. Spence, *The Molecular Probes Handbook*, Life Technologies, 11th edn, 2010.
- 62 S. Liang; C. L. John; S. Xu; J. Chen; Y. Jin; Q. Yuan; W. Tan and J. X. Zhao, Silica-Based Nanoparticles: Design and Properties, in *Advanced Fluorescence Reporters in Chemistry and Biology II: Molecular Constructions, Polymers and Nanoparticles*, ed. A. P. Demchenko, Springer Berlin Heidelberg, Berlin, Heidelberg, 2010, pp 229–251.
- 63 E.-B. Cho, D. O. Volkov and I. Sokolov, Ultrabright Fluorescent Silica Mesoporous Silica Nanoparticles: Control of Particle Size and Dye Loading, *Adv. Funct. Mater.*, 2011, **21**(16), 3129–3135.
- 64 D. Genovese, S. Bonacchi, R. Juris, M. Montalti, L. Prodi, E. Rampazzo and N. Zaccheroni, Prevention of Self-Quenching in Fluorescent Silica Nanoparticles by Efficient Energy Transfer, *Angew. Chem., Int. Ed.*, 2013, **52**(23), 5965–5968.
- 65 S. Bonacchi, D. Genovese, R. Juris, M. Montalti, L. Prodi, E. Rampazzo and N. Zaccheroni, Luminescent Silica Nanoparticles: Extending the Frontiers of Brightness, *Angew. Chem., Int. Ed.*, 2011, **50**(18), 4056–4066.
- 66 A. S. Klymchenko, F. Liu, M. Collot and N. Anton, Dye-Loaded Nanoemulsions: Biomimetic Fluorescent Nanocarriers for Bioimaging and Nanomedicine, *Adv. Healthcare Mater.*, 2021, **10**(1), 2001289.
- 67 A. S. Klymchenko, E. Roger, N. Anton, H. Anton, I. Shulov, J. Vermot, Y. Mely and T. F. Vandamme, Highly lipophilic fluorescent dyes in nano-emulsions: towards bright non-leaking nano-droplets, *RSC Adv.*, 2012, **2**(31), 11876–11886.
- 68 X. Wang, N. Anton, P. Ashokkumar, H. Anton, T. K. Fam, T. Vandamme, A. S. Klymchenko and M. Collot, Optimizing the Fluorescence Properties of Nanoemulsions for Single Particle Tracking in Live Cells, *ACS Appl. Mater. Interfaces*, 2019, **11**(14), 13079–13090.
- 69 V. N. Kilin, H. Anton, N. Anton, E. Steed, J. Vermot, T. F. Vandamme, Y. Mely and A. S. Klymchenko, Counterion-enhanced cyanine dye loading into lipid nano-droplets for single-particle tracking in zebrafish, *Biomaterials*, 2014, **35**(18), 4950–4957.
- 70 E. M. Sletten and T. M. Swager, Fluorofluorophores: Fluorescent Fluorous Chemical Tools Spanning the Visible Spectrum, *J. Am. Chem. Soc.*, 2014, **136**(39), 13574–13577.
- 71 E. M. Sletten and T. M. Swager, Readily accessible multifunctional fluoruous emulsions, *Chem. Sci.*, 2016, **7**(8), 5091–5097.
- 72 I. Lim, A. Vian, H. L. van de Wouw, R. A. Day, C. Gomez, Y. Liu, A. L. Rheingold, O. Campàs and E. M. Sletten, Fluorous Soluble Cyanine Dyes for Visualizing Perfluorocarbons in Living Systems, *J. Am. Chem. Soc.*, 2020, **142**(37), 16072–16081.
- 73 M. Collot, J. Schild, K. T. Fam, R. Bouchaala and A. S. Klymchenko, Stealth and Bright Monomolecular Fluorescent Organic Nanoparticles Based on Folded Amphiphilic Polymer, *ACS Nano*, 2020, **14**(10), 13924–13937.
- 74 I. Shulov, Y. Arntz, Y. Mély, V. G. Pivovarenko and A. S. Klymchenko, Non-coordinating anions assemble cyanine amphiphiles into ultra-small fluorescent nanoparticles, *Chem. Commun.*, 2016, **52**(51), 7962–7965.
- 75 X. Zhang, Z. Chen and F. Würthner, Morphology Control of Fluorescent Nanoaggregates by Co-Self-Assembly of Wedge- and Dumbbell-Shaped Amphiphilic Perylene Bisimides, *J. Am. Chem. Soc.*, 2007, **129**(16), 4886–4887.
- 76 G. Sun, M. Y. Berezin, J. Fan, H. Lee, J. Ma, K. Zhang, K. L. Wooley and S. Achilefu, Bright fluorescent nanoparticles for developing potential optical imaging contrast agents, *Nanoscale*, 2010, **2**(4), 548–558.
- 77 C. Wu and D. T. Chiu, Highly Fluorescent Semiconducting Polymer Dots for Biology and Medicine, *Angew. Chem., Int. Ed.*, 2013, **52**(11), 3086–3109.
- 78 K. Li and B. Liu, Polymer-encapsulated organic nanoparticles for fluorescence and photoacoustic imaging, *Chem. Soc. Rev.*, 2014, **43**(18), 6570–6597.
- 79 D. Tuncel and H. V. Demir, Conjugated polymer nanoparticles, *Nanoscale*, 2010, **2**(4), 484–494.
- 80 C. Wu, B. Bull, C. Szymanski, K. Christensen and J. McNeill, Multicolor Conjugated Polymer Dots for Biological Fluorescence Imaging, *ACS Nano*, 2008, **2**(11), 2415–2423.
- 81 J. Yu, C. Wu, Z. Tian and J. McNeill, Tracking of Single Charge Carriers in a Conjugated Polymer Nanoparticle, *Nano Lett.*, 2012, **12**(3), 1300–1306.
- 82 L. C. Groff, X. L. Wang and J. D. McNeill, Measurement of Exciton Transport in Conjugated Polymer Nanoparticles, *J. Phys. Chem. C*, 2013, **117**(48), 25748–25755.
- 83 C. Wu, T. Schneider, M. Zeigler, J. Yu, P. G. Schiro, D. R. Burnham, J. D. McNeill and D. T. Chiu, Bioconjugation of Ultrabright Semiconducting Polymer Dots for Specific Cellular Targeting, *J. Am. Chem. Soc.*, 2010, **132**(43), 15410–15417.
- 84 C. Wu, S. J. Hansen, Q. Hou, J. Yu, M. Zeigler, Y. Jin, D. R. Burnham, J. D. McNeill, J. M. Olson and D. T. Chiu, Design of Highly Emissive Polymer Dot Bioconjugates for *In Vivo* Tumor Targeting, *Angew. Chem., Int. Ed.*, 2011, **50**(15), 3430–3434.
- 85 C. Szymanski, C. Wu, J. Hooper, M. A. Salazar, A. Perdomo, A. Dukes and J. McNeill, Single Molecule Nanoparticles of the Conjugated Polymer MEH–PPV, Preparation and Characterization by Near-Field Scanning Optical Microscopy, *J. Phys. Chem. B*, 2005, **109**(18), 8543–8546.
- 86 J. Pecher and S. Mecking, Nanoparticles from Step-Growth Coordination Polymerization, *Macromolecules*, 2007, **40**(22), 7733–7735.
- 87 H. Piwoński, T. Michinobu and S. Habuchi, Controlling photophysical properties of ultrasmall conjugated polymer nanoparticles through polymer chain packing, *Nat. Commun.*, 2017, **8**(1), 15256.
- 88 H. Piwoński, W. Li, Y. Wang, T. Michinobu and S. Habuchi, Improved Fluorescence and Brightness of Near-Infrared and Shortwave Infrared Emitting Polymer Dots for



- Bioimaging Applications, *ACS Appl. Polym. Mater.*, 2020, **2**(2), 569–577.
- 89 J. Luo, Z. Xie, J. W. Y. Lam, L. Cheng, H. Chen, C. Qiu, H. S. Kwok, X. Zhan, Y. Liu, D. Zhu and B. Z. Tang, Aggregation-induced emission of 1-methyl-1,2,3,4,5-pentaphenylsilole, *Chem. Commun.*, 2001, (18), 1740–1741.
- 90 Y. Hong, J. W. Y. Lam and B. Z. Tang, Aggregation-induced emission: phenomenon, mechanism and applications, *Chem. Commun.*, 2009, (29), 4332–4353.
- 91 Z. Zhao, H. Zhang, J. W. Y. Lam and B. Z. Tang, Aggregation-Induced Emission: New Vistas at the Aggregate Level, *Angew. Chem., Int. Ed.*, 2020, **59**(25), 9888–9907.
- 92 F. Würthner, Aggregation-Induced Emission (AIE): A Historical Perspective, *Angew. Chem., Int. Ed.*, 2020, **59**(34), 14192–14196.
- 93 Y. Ren, J. W. Y. Lam, Y. Dong, B. Z. Tang and K. S. Wong, Enhanced Emission Efficiency and Excited State Lifetime Due to Restricted Intramolecular Motion in Silole Aggregates, *J. Phys. Chem. B*, 2005, **109**(3), 1135–1140.
- 94 J. Mei, N. L. C. Leung, R. T. K. Kwok, J. W. Y. Lam and B. Z. Tang, Aggregation-Induced Emission: Together We Shine, United We Soar!, *Chem. Rev.*, 2015, **115**(21), 11718–11940.
- 95 S. Chen, H. Wang, Y. Hong and B. Z. Tang, Fabrication of fluorescent nanoparticles based on AIE luminogens (AIE dots) and their applications in bioimaging, *Mater. Horiz.*, 2016, **3**(4), 283–293.
- 96 K. Li, Y. Jiang, D. Ding, X. Zhang, Y. Liu, J. Hua, S.-S. Feng and B. Liu, Folic acid-functionalized two-photon absorbing nanoparticles for targeted MCF-7 cancer cell imaging, *Chem. Commun.*, 2011, **47**(26), 7323–7325.
- 97 B. Zhang, H. Soleimaninejad, D. J. Jones, J. M. White, K. P. Ghiggino, T. A. Smith and W. W. H. Wong, Highly Fluorescent Molecularly Insulated Perylene Diimides: Effect of Concentration on Photophysical Properties, *Chem. Mater.*, 2017, **29**(19), 8395–8403.
- 98 K. Trofymchuk, A. Reisch, I. Shulov, Y. Mély and A. S. Klymchenko, Tuning the color and photostability of perylene diimides inside polymer nanoparticles: towards biodegradable substitutes of quantum dots, *Nanoscale*, 2014, **6**(21), 12934–12942.
- 99 D. Schmidt, M. Stolte, J. Süß, A. Liess, V. Stepanenko and F. Würthner, Protein-like Enwrapped Perylene Bisimide Chromophore as a Bright Microcrystalline Emitter Material, *Angew. Chem., Int. Ed.*, 2019, **58**(38), 13385–13389.
- 100 M. Stolte, T. Schembri, J. Süß, D. Schmidt, A.-M. Krause, M. O. Vysotsky and F. Würthner, 1-Mono- and 1,7-Disubstituted Perylene Bisimide Dyes with Voluminous Groups at Bay Positions: In Search for Highly Effective Solid-State Fluorescence Materials, *Chem. Mater.*, 2020, **32**(14), 6222–6236.
- 101 J. Della Rocca, D. Liu and W. Lin, Nanoscale Metal–Organic Frameworks for Biomedical Imaging and Drug Delivery, *Acc. Chem. Res.*, 2011, **44**(10), 957–968.
- 102 A. Monguzzi, M. Ballabio, N. Yanai, N. Kimizuka, D. Fazzi, M. Campione and F. Meinardi, Highly Fluorescent Metal–Organic-Framework Nanocomposites for Photonic Applications, *Nano Lett.*, 2018, **18**(1), 528–534.
- 103 G. Calzaferrri, S. Huber, H. Maas and C. Minkowski, Host-Guest Antenna Materials, *Angew. Chem., Int. Ed.*, 2003, **42**(32), 3732–3758.
- 104 T. Doussineau, M. Smaïhi, S. Balme and J.-M. Janot, Fluorescent Hydroxyflavone–Zeolite Nanoparticles: Ship-in-a-Bottle Synthesis and Photophysical Properties, *ChemPhysChem*, 2006, **7**(3), 583–589.
- 105 L. M. Grimm, S. Sinn, M. Krstić, E. D'Este, I. Sonntag, E. A. Prasetyanto, T. Kuner, W. Wenzel, L. De Cola and F. Biedermann, Fluorescent Nanozeolite Receptors for the Highly Selective and Sensitive Detection of Neurotransmitters in Water and Biofluids, *Adv. Mater.*, 2021, **33**(49), 2104614.
- 106 A. M. O. Azevedo, J. L. M. Santos, I. M. Warner and M. L. M. F. S. Saraiva, GUMBOS and nanoGUMBOS in chemical and biological analysis: A review, *Anal. Chim. Acta*, 2020, **1133**, 180–198.
- 107 D. K. Bwambok, B. El-Zahab, S. K. Challa, M. Li, L. Chandler, G. A. Baker and I. M. Warner, Near-Infrared Fluorescent NanoGUMBOS for Biomedical Imaging, *ACS Nano*, 2009, **3**(12), 3854–3860.
- 108 S. Das, D. Bwambok, B. El-Zahab, J. Monk, S. L. de Rooy, S. Challa, M. Li, F. R. Hung, G. A. Baker and I. M. Warner, Nontemplated Approach to Tuning the Spectral Properties of Cyanine-Based Fluorescent NanoGUMBOS, *Langmuir*, 2010, **26**(15), 12867–12876.
- 109 A. N. Jordan, S. Das, N. Siraj, S. L. de Rooy, M. Li, B. El-Zahab, L. Chandler, G. A. Baker and I. M. Warner, Anion-controlled morphologies and spectral features of cyanine-based nanoGUMBOS – an improved photosensitizer, *Nanoscale*, 2012, **4**(16), 5031–5038.
- 110 P. K. S. Magut, S. Das, V. E. Fernand, J. Losso, K. McDonough, B. M. Naylor, S. Aggarwal and I. M. Warner, Tunable Cytotoxicity of Rhodamine 6G via Anion Variations, *J. Am. Chem. Soc.*, 2013, **135**(42), 15873–15879.
- 111 H. Yao, M. Yamashita and K. Kimura, Organic Styryl Dye Nanoparticles: Synthesis and Unique Spectroscopic Properties, *Langmuir*, 2009, **25**(2), 1131–1137.
- 112 H. Yao and K. Ashiba, Highly fluorescent organic nanoparticles of thiacyanine dye: A synergetic effect of intermolecular H-aggregation and restricted intramolecular rotation, *RSC Adv.*, 2011, **1**(5), 834–838.
- 113 T. Enseki and H. Yao, Controlled Formation of Fluorescent Organic Nanoparticles of Carbocyanine Dye via Ion-association Approach, *Chem. Lett.*, 2012, **41**(10), 1119–1121.
- 114 I. Shulov, S. Oncul, A. Reisch, Y. Arntz, M. Collot, Y. Mely and A. S. Klymchenko, Fluorinated counterion-enhanced emission of rhodamine aggregates: ultrabright nanoparticles for bioimaging and light-harvesting, *Nanoscale*, 2015, **7**(43), 18198–18210.
- 115 A. Reisch, K. Trofymchuk, A. Runser, G. Fleith, M. Rawiso and A. S. Klymchenko, Tailoring Fluorescence Brightness and Switching of Nanoparticles through Dye Organization



- in the Polymer Matrix, *ACS Appl. Mater. Interfaces*, 2017, **9**(49), 43030–43042.
- 116 A. Reisch, A. Runser, Y. Arntz, Y. Mély and A. S. Klymchenko, Charge-Controlled Nanoprecipitation as a Modular Approach to Ultrasmall Polymer Nanocarriers: Making Bright and Stable Nanoparticles, *ACS Nano*, 2015, **9**(5), 5104–5116.
- 117 N. Melnychuk and A. S. Klymchenko, DNA-Functionalized Dye-Loaded Polymeric Nanoparticles: Ultrabright FRET Platform for Amplified Detection of Nucleic Acids, *J. Am. Chem. Soc.*, 2018, **140**(34), 10856–10865.
- 118 C. Severi, N. Melnychuk and A. S. Klymchenko, Smartphone-assisted detection of nucleic acids by light-harvesting FRET-based nanoprobe, *Biosens. Bioelectron.*, 2020, **168**, 112515.
- 119 N. Melnychuk, S. Egloff, A. Runser, A. Reisch and A. S. Klymchenko, Light-Harvesting Nanoparticle Probes for FRET-Based Detection of Oligonucleotides with Single-Molecule Sensitivity, *Angew. Chem., Int. Ed.*, 2020, **59**(17), 6811–6818.
- 120 S. Egloff, N. Melnychuk, A. Reisch, S. Martin and A. S. Klymchenko, Enzyme-free amplified detection of cellular microRNA by light-harvesting fluorescent nanoparticle probes, *Biosens. Bioelectron.*, 2021, **179**, 113084.
- 121 B. Andreiuk, I. O. Aparin, A. Reisch and A. S. Klymchenko, Bulky Barbiturates as Non-Toxic Ionic Dye Insulators for Enhanced Emission in Polymeric Nanoparticles, *Chem.–Eur. J.*, 2021, **27**(50), 12877–12883.
- 122 Y. Haketa, K. Yamasumi and H. Maeda, π -Electronic ion pairs: building blocks for supramolecular nanoarchitectonics via $i\pi$ – $i\pi$ interactions, *Chem. Soc. Rev.*, 2023, **52**(20), 7170–7196.
- 123 Y. Haketa and H. Maeda, Dimension-controlled ion-pairing assemblies based on π -electronic charged species, *Chem. Commun.*, 2017, **53**(20), 2894–2909.
- 124 Y. Haketa, S. Sasaki, N. Ohta, H. Masunaga, H. Ogawa, N. Mizuno, F. Araoka, H. Takezoe and H. Maeda, Oriented Salts: Dimension-Controlled Charge-by-Charge Assemblies from Planar Receptor–Anion Complexes, *Angew. Chem., Int. Ed.*, 2010, **49**(52), 10079–10083.
- 125 J. Chen, S. G. Stenspil, S. Kaziannis, L. Kacenauskaite, N. Lenngren, M. Kloz, A. H. Flood and B. W. Laursen, Quantitative Energy Transfer in Organic Nanoparticles Based on Small-Molecule Ionic Isolation Lattices for UV Light Harvesting, *ACS Appl. Nano Mater.*, 2022, **5**(10), 13887–13893.
- 126 D. S. Biswas, N. Melnychuk, C. Severi, P. Didier and A. S. Klymchenko, Giant Light-Harvesting in Dye-Loaded Nanoparticles Enhanced by Blank Hydrophobic Salts, *Adv. Opt. Mater.*, 2024, **12**(4), 2301671.
- 127 N. Adarsh and A. S. Klymchenko, Ionic aggregation-induced emission dye with bulky counterions for preparation of bright near-infrared polymeric nanoparticles, *Nanoscale*, 2019, **11**(29), 13977–13987.
- 128 S. G. Stenspil, J. Chen, M. B. Liisberg, A. H. Flood and B. W. Laursen, Control of the fluorescence lifetime in dye based nanoparticles, *Chem. Sci.*, 2024, **15**, 5531–5538.
- 129 Z. Zhang, Y. Yuan, Z. Liu, H. Chen, D. Chen, X. Fang, J. Zheng, W. Qin and C. Wu, Brightness Enhancement of Near-Infrared Semiconducting Polymer Dots for *in Vivo* Whole-Body Cell Tracking in Deep Organs, *ACS Appl. Mater. Interfaces*, 2018, **10**(32), 26928–26935.
- 130 X. Zhou, L. Zhao, K. Zhang, C. Yang, S. Li, X. Kang, G. Li, Q. Wang, H. Ji, M. Wu, J. Liu, Y. Qin and L. Wu, Ultrabright AIEdots with tunable narrow emission for multiplexed fluorescence imaging, *Chem. Sci.*, 2023, **14**, 113–120.
- 131 X. Zhu, J.-X. Wang, L.-Y. Niu and Q.-Z. Yang, Aggregation-Induced Emission Materials with Narrowed Emission Band by Light-Harvesting Strategy: Fluorescence and Chemiluminescence Imaging, *Chem. Mater.*, 2019, **31**(9), 3573–3581.
- 132 S. Wang, B. Li and F. Zhang, Molecular Fluorophores for Deep-Tissue Bioimaging, *ACS Cent. Sci.*, 2020, **6**(8), 1302–1316.
- 133 E. D. Cosco, A. L. Spearman, S. Ramakrishnan, J. G. P. Lingg, M. Saccomano, M. Pengshung, B. A. Arús, K. C. Y. Wong, S. Glasl, V. Ntziachristos, M. Warmer, R. R. McLaughlin, O. T. Bruns and E. M. Sletten, Shortwave infrared polymethine fluorophores matched to excitation lasers enable non-invasive, multicolour *in vivo* imaging in real time, *Nat. Chem.*, 2020, **12**(12), 1123–1130.
- 134 E. Thimsen, B. Sadtler and M. Y. Berezin, Shortwave-infrared (SWIR) emitters for biological imaging: a review of challenges and opportunities, *Nanophotonics*, 2017, **6**(5), 1043–1054.
- 135 Y. Yang, C. Sun, S. Wang, K. Yan, M. Zhao, B. Wu and F. Zhang, Counterion-Paired Bright Heptamethine Fluorophores with NIR-II Excitation and Emission Enable Multiplexed Biomedical Imaging, *Angew. Chem., Int. Ed.*, 2022, e202117436.
- 136 C. Sun, M. Zhao, X. Zhu, P. Pei and F. Zhang, One-Pot Preparation of Highly Dispersed Second Near-Infrared J-Aggregate Nanoparticles Based on FD-1080 Cyanine Dye for Bioimaging and Biosensing, *CCS Chem.*, 2022, **4**(2), 476–486.
- 137 R. Englman and J. Jortner, The energy gap law for radiationless transitions in large molecules, *Mol. Phys.*, 1970, **18**(2), 145–164.
- 138 H. S. Afsari, M. Cardoso Dos Santos, S. Lindén, T. Chen, X. Qiu, P. M. P. van Bergen en Henegouwen, T. L. Jennings, K. Susumu, I. L. Medintz, N. Hildebrandt and L. W. Miller, Time-gated FRET nanoassemblies for rapid and sensitive intra- and extracellular fluorescence imaging, *Sci. Adv.*, 2016, **2**(6), e1600265.
- 139 P. Stachelek, L. MacKenzie, D. Parker and R. Pal, Circularly polarised luminescence laser scanning confocal microscopy to study live cell chiral molecular interactions, *Nat. Commun.*, 2022, **13**(1), 553.

