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An efficient alternative to DBU in the oxathiaphospholane (OTP) method for the solid phase synthesis of P-stereodefined phosphorothioate analogs†

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This study presents a modified (extended) 1,3,2-oxathiaphospholane (OTP) method for the synthesis of P-stereodefined phosphorothioate analogs in the presence of previously unused organic bases. TBD (5,7-triazabicyclo[4.4.0]dec-5-ene) and Verkade's proazaphosphatane (2,8,9-trimethyl-2,5,8,9-tetraaza-1-phosphabicyclo[3.3.3]undecane) are herein used for the first time as efficient organic bases compared to DBU, which is commonly used in the OTP approach.

Introduction

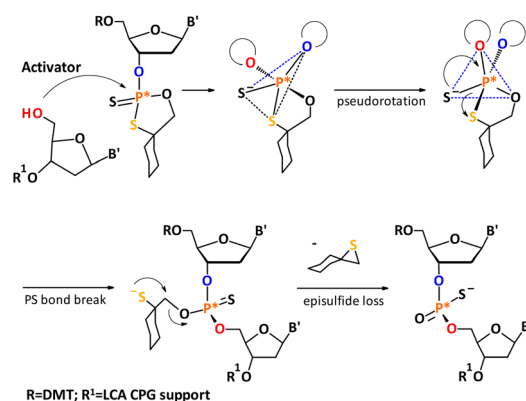
Only a few methods have been developed for stereocontrolled synthesis of phosphorothioate oligonucleotides, including methods that utilize *N*-acyl oxazaphospholidine¹ (Beaucage *et al.*), oxazaphosphorinane² (Just *et al.*) and bicyclic oxazaphospholidine³ (Agrawal *et al.*) derivatives. Wada and co-workers have developed a methodology to synthesize stereochemically regular PS-ODNs using diastereopure nucleoside 3'-*O*-(1,3,2-oxazaphospholidine) monomers (originally proposed by Agrawal *et al.*).⁴ The first method for stereocontrolled chemical synthesis of phosphorothioate oligonucleotides, which was developed by Stec *et al.*, is based on a chemistry employing P-diastereomerically pure monomers possessing the 2-thio-1,3,2-oxathiaphospholane moiety.⁵ Their applicability and limitations have been summarized in several reports.⁶ Recently, Baran *et al.* described a method using a citrus-derived PSI reagent.⁷

General mechanistic principle of the OTP method

In the OTP method, the P-chiral nucleoside 3'-*O*-(2-thio-1,3,2-oxathiaphospholane) monomers are prepared as a mixture of both P-isomers, which can be separated by silica gel column chromatography. Separated P-diastereomers may be used for the synthesis of diastereomerically pure fragments of phosphorothioate oligonucleotides. In this synthesis, the

oxathiaphospholane reacts with the 5'-OH group of a nucleoside (or a growing oligonucleotide attached to the solid support) in the presence of 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU). This leads to an intermediate, which undergoes pseudorotation (permutational isomerization), followed by the elimination of the episulfide to form an internucleotide phosphorothioate linkage (Scheme 1).⁸ In the synthesis of PS-DNA oligomers, repetitive yields in the range of 92–94% are usually observed, making the synthesis of oligomers longer than 12–14 nucleotides problematic; although successful but very low yields of 23-mers have also been reported.⁹

The OTP approach promoted by DBU has been successfully used for the stereocontrolled synthesis of phosphorothioate oligonucleotides and their analogues. For instance, in our laboratory, the OTP method was also used for the stereocontrolled synthesis of PS-LNA analogs (Locked Nucleic Acids)¹⁰



Scheme 1 Synthesis of P-stereodefined phosphorothioate analogs developed by Stec *et al.* A mechanism of DBU-promoted condensation step.

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and PS-GNA analogs (Glycol Nucleic Acids).¹¹ Recently, there has been successful contribution to research in the field of P-diastereomerically pure PS-RNA and PS-(2'-OMe) RNA derivatives.¹²

The constantly increasing number of approved nucleic acid therapeutics shows their great potential.¹³ It is necessary to provide more detailed information on the influence of the absolute configuration of the phosphorus atoms in P-stereodefined oligonucleotides. The studies carried out so far have shown their potential. It has been demonstrated that the configuration of the phosphorus atom has an influence on biological activity when interacting with biomolecules. R_P -PS linkages are preferred for duplex formation with complementary ORN strands,¹⁴ while S_P -PS linkages are chosen to increase stability towards the 3'-exonuclease present in the human plasma.¹⁵ PS DNA in which both the R_P and S_P -PS linkages are properly arranged would therefore be better drug candidates than those synthesized in a non-stereocontrolled manner. Therefore, the stereoselective synthesis of such compounds is an important goal to achieve. In view of the growing interest of molecular biologists, biochemists, as well as pharmaceutical companies in the synthesis of modified oligonucleotides and their therapeutic application,¹⁶ more efficient organic bases must be considered.

Results and discussion

Novel and efficient organic bases in OTP method

In the search for new organic bases, we focused on strong, yet non-nucleophilic bases. We found bases consisting of nitrogenated bases and proazaphosphatranes (Fig. 1).

Here we discovered that TBD (1,5,7-triazabicyclo[4.4.0]dec-5-ene) and one of the Verkade bases, 2,8,9-trimethyl-2,5,8,9-tetraaza-1-phosphabicyclo[3.3.3]undecane, can be used as alternatives and efficient bases for the 1,3,2-oxathiaphospholane ring-opening condensation (Scheme 2).

TBD is a strong bicyclic guanidine base (pK_a in acetonitrile: 26.0)¹⁸ with a steric hindrance and the ability, when protonated, to delocalize the positive charge between amino groups and the

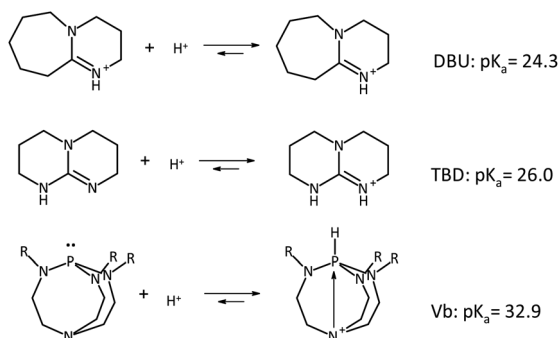
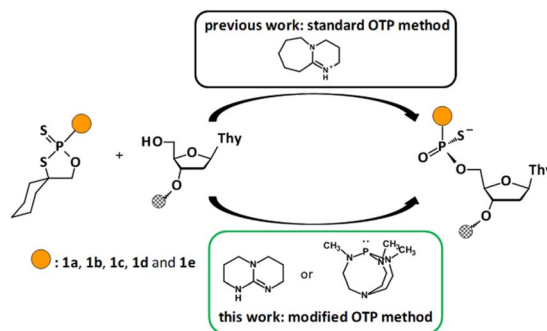


Fig. 1 Non-nucleophilic bases used in this study. DBU (1,8-diazabicyclo[5.4.0]undec-7-ene), TBD (1,5,7-triazabicyclo[4.4.0]dec-5-ene), Verkade base (Vb, in this study: R = methyl group, 2,8,9-trimethyl-2,5,8,9-tetraaza-1-phosphabicyclo[3.3.3]undecane), their protonated molecules and pK_a s in acetonitrile.¹⁷



Scheme 2 Synthesis of dinucleoside phosphorothioate 1p5T by the oxathiaphospholane approach.

double bond. Because of their strong basicity and relatively weak nucleophilicity, proazaphosphatranes have also attracted attention. Proazaphosphatranes have been used in multiple types of catalytic processes for organic synthesis.¹⁹ Pioneered by Verkade *et al.*,²⁰ the 'molecular football' shaped compound is commonly known as Verkade's superbase (in this paper labeled as Vb).

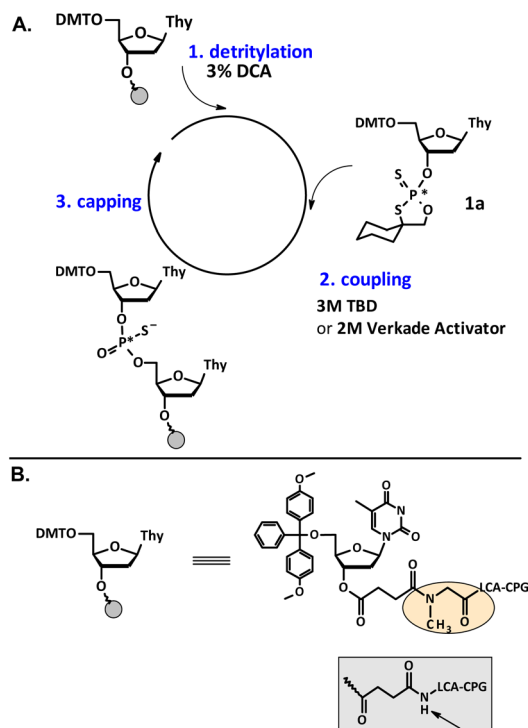
The term 'superbase' is given to organic compounds whose basicity is greater than that of 1,8-bis(dimethylamino)naphthalene (proton sponge),²¹ with pK_a of 18.6 (conjugate acid form) in acetonitrile. The pK_a of the protonated form of proazaphosphatranes ranges from 31.6 to 33.6 in acetonitrile, depending on the substituents attached to the equatorial nitrogen atoms R (in this study R = Me). The conjugate acid $[HP(MeNCH_2CH_2)_3N]^+$ has a pK_a of 32.9 in acetonitrile;²² thus, this base is by eight orders of magnitude stronger than DBU.²³

General protocol of the OTP method

Synthesis of P-stereodefined oligonucleotides by oxathiaphospholane method was carried out manually at a 1 μ mol scale, according to the previously published protocols. This method uses a CPG-support (controlled pore glass, LCAA CPG 500 Å, ChemGenes, USA), to which a 5'-O-DMT-nucleoside unit (in this study: loading 35 μ mol g^{-1}) is attached with a succinyl-sarcosinyl type linker (Scheme 3). To prevent the DBU-promoted intramolecular cleavage of the standard LCAA (long chain alkylamino) linker, a modified sarcosine linker $[-COCH_2CH_2CON(CH_3)CH_2CO-LCAA-CPG]$ is used.²⁴ Coupling steps are carried out using 20 mg of 3'-O-OTP-derivatives of the standard 5'-O-DMT-protected nucleosides in the presence of 1 M DBU in acetonitrile. Manual preparation (without the synthesizer) is considered demanding in terms of the preparation of reagents and technical skills during the oligonucleotide synthesis. Our previous observations indicate that some parameters, such as amount of monomer, coupling time, and base concentration play a major role in determining the coupling yield during synthesis.

UV monitoring of the solid-phase synthesis efficacy

The efficiency of solid phase synthesis was studied by condensation of 1a-e with thymidine (attached to the solid support).



Scheme 3 (A) Synthetic cycle for the preparation of oligonucleotide fragment from **1a**; (B) structures of sarcosine linker and linker with no sarcosine are highlighted in light orange and grey, respectively.

We analyzed the coupling efficiency of the set of oxathiaphospholane monomers **1** (as a mixture of P-diastereomers) in the presence of novel organic non-nucleophilic bases. The experimental protocol involves use of previously tested 5'-O-DMT-protected 3'-O-(2-thio-"spiro"-4,4-pentamethylene-1,3,2-oxathiaphospholane) derivatives, *i.e.* 2'-deoxynucleoside (**1a**), locked nucleic acid (**1b**), 2'-O-methyl ribonucleoside (**1c**), and 2'-O-TBDMS-ribonucleoside (**1d**) monomers. It should be noted that previously unknown oxathiaphospholane derivatives of

morpholino nucleosides (**1e**)²⁵ were also synthesized and used in this modified OTP protocol (Fig. 2).

Detailed procedure for synthesis of mU-OTP is presented in the ESI.† For all synthesized products the same support (which possessed the thymidine 3'-O-attached to LCA CPG linker with loading 35 $\mu\text{mol g}^{-1}$) was used to eliminate any possible effect of support variability. The coupling efficacy is controlled by measurement of absorption at 504 nm coming from the released DMT⁺ cation.

The efficiency of monomer DNA condensation in the standard OTP procedure using 1 M DBU (50-fold molar excess) is up to 95%. Attempts to use the same concentrations of novel bases did not lead to satisfactory results. When using 3 M TBD (150-fold molar excess) and 2 M Verkade base (25-fold molar excess), the efficacy was 93% (Table 1 and Fig. S4–18, ESI†). Even with double coupling, we did not observe a significantly increased yield of condensation. This result suggests that TBD and Verkade base can be successfully used alternatively.

Working earlier with LNA-derived OTP monomers using the standard DBU-assisted OTP method, we observed that in the cycles where LNA-OTP monomers were incorporated, double coupling was necessary (20 mg + 20 mg, ~25-fold molar excess) and in both steps the coupling time was extended to 20 min (Table 1 and Fig. S1, ESI†). Although the repetitive average yield of the coupling (88%, based on the DMT⁺ cation assay) was relatively low as compared with DNA OTP (**1a**), it was considered to be satisfactory enough to attempt synthesis of chimeric P-stereodefined PS-(DNA/LNA). In the case of 3 M TBD solution in acetonitrile, the yield of a single coupling step of LNA monomer was approximately 65%. Thus, it was necessary to use double coupling (20 mg + 20 mg of OTP monomer **1b**) and the coupling time was extended to 20 minutes. Repetitive yield, calculated from the DMT⁺ cation assay, was 86%. It is noted that the repetitive yield of condensation reduced when using 3 M TBD and 2 M Vb (86%, assessed by the DMT⁺ cation assay) despite the use of double coupling. In the case of synthesizing LNA oligonucleotides, 1 M DBU is still the best option. In the next step, the 2'-OMe monomer (**1c**) was investigated. In standard protocol with 1 M DBU, only *ca.* 84% repetitive yield was achieved (based on a DMT⁺ cation assay), even with double coupling. Additional experiments using 3 M TBD revealed

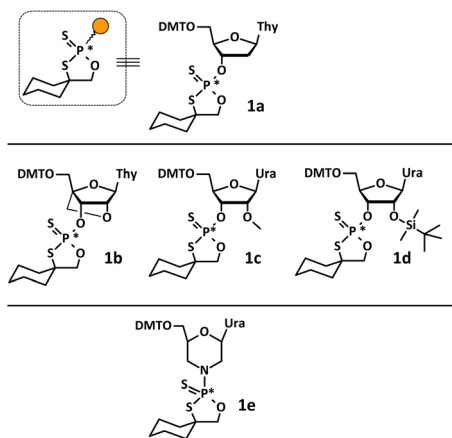


Fig. 2 All OTP monomers: DNA (**1a**), LNA (**1b**), 2'-OMe (**1c**), 2'-OTBDMS (**1d**), morpholino (**1e**) as a mixture of P-diastereomers used in this study.

Table 1 Coupling yields with OTP monomers in solid phase synthesis of the dimers calculated from the DMT⁺ decay assay

OTP monomer	Standard OTP method		Modified OTP method	
	DBU ^a		TBD ^b	Verkade ^c
DNA (1a)	95		93	93
LNA (1b)	88 ^d		86 ^d	86 ^d
2'-OMe (1c)	84 ^d		86 ^d	83 ^d
2'-OTBDMS (1d)	91 ^d		92 ^d	92
mU (1e)	61 ^d		93 ^d	95

^a 1 M DBU solution in anhydrous acetonitrile. ^b 2 M TBD solution in anhydrous acetonitrile. ^c 3 M Verkade solution in anhydrous acetonitrile. ^d Double coupling.



Next, longer analogs were synthesized. The yield of the first coupling step of 5T_T and 5T_V (as a mixture of P-diastereomers) with the 5'-OH group of a deoxyribonucleoside attached to a solid support was acceptable (93%) and the repetitive yields associated with the subsequent coupling steps were equally high (Table 2, Fig. S19 and 20, ESI†).

There are no conflicts of interest to declare.

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