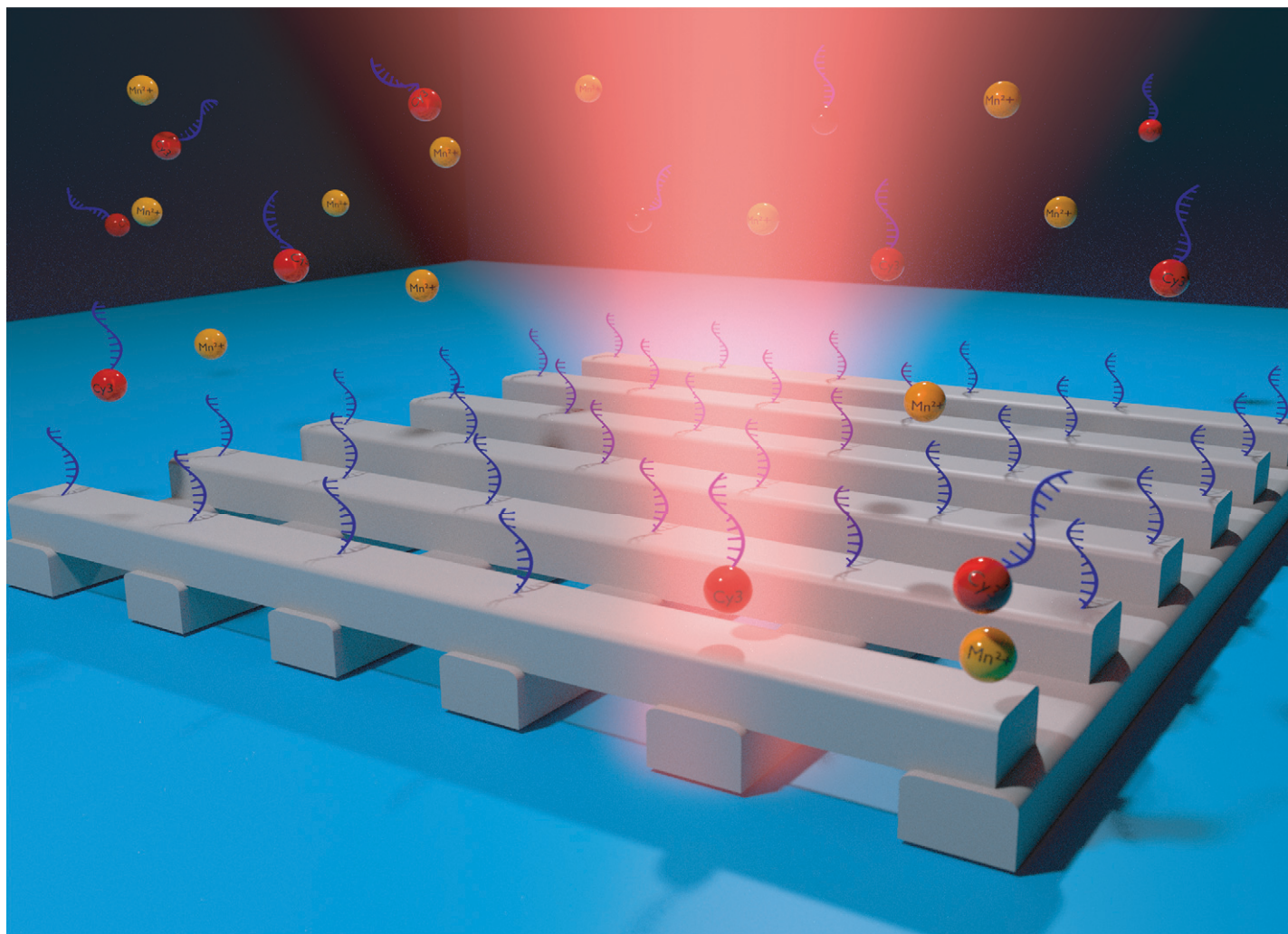




**Showcasing research from Professor Jacak's laboratory,
School of Medical Engineering, University of Applied
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New buffer systems for photopainting of single biomolecules

Advanced buffer systems are introduced to enhance the efficiency of photochemically induced surface modification at the single-molecule level. Utilization of buffers with paramagnetic cations and radical oxygen-promoting agents enables Laser-Assisted Protein Adsorption by Photobleaching (LAPAP) of fluorescently labeled oligonucleotides or biotin onto 2D and 3D acrylate scaffolds structured by multi-photon lithography. Specific cation interaction sites for cyanine, coumarin, and rhodamine fluorophores are identified through quantum mechanical calculations. A remarkable up to three-fold increase in LAPAP efficiency for cyanine fluorophores is demonstrated, while constant excitation parameters are maintained.

As featured in:



See Jaroslaw Jacak *et al.*,
RSC Appl. Interfaces, 2024, 1, 110.

Cite this: *RSC Appl. Interfaces*, 2024,
1, 110Received 4th August 2023,
Accepted 3rd October 2023

DOI: 10.1039/d3lf00125c

rsc.li/RSCApplInter

New buffer systems for photopainting of single biomolecules†

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We present newly developed buffer systems that significantly improve the efficiency of a photochemically induced surface modification at the single molecule level. Buffers with paramagnetic cations and radical oxygen promoting species facilitate laser-assisted protein adsorption by photobleaching (LAPAP) of single fluorescently labelled oligonucleotides or biotin onto multi-photon-lithography-structured 2D and 3D acrylate scaffolds. Single molecule fluorescence microscopy has been used to quantify photopainting efficiency. We identify specific cation interaction sites for members of the cyanine, coumarin and rhodamine classes of fluorophores using quantum mechanical calculations. We show that our buffer systems provide an up to three-fold LAPAP-efficiency increase for the cyanine fluorophore, while keeping excitation parameters constant.

1 Introduction

Photopatterning techniques are attracting increasing interest due to their widespread applications in lithography, biointerfaces, and microfluidics.^{1–3} Manipulation of surface functionalities is important for applications involving adaptive surface properties.^{4–14} Surfaces that can regulate biological functions by stimuli-responsive compounds allow the control of molecular uptake, manipulation of cell adhesion and sensing,^{12,15–26} or the induction of protein denaturation.^{27,28} Physical and chemical properties can be (ir-)reversibly tuned under a stimulus such as light, temperature, chemicals, pH, or an electric field.^{29–40} Light provides high spatio-temporal resolution to control the functions of such surfaces without a direct contact. This is achieved by photoisomerization, -coupling or -cleavage through chemical bond breakage/formation,^{20,41–48} or addition of photo-switchable molecules such as azobenzene,^{49–55} spiropyran,^{56–58} dithienylethene,^{59,60} and molecular shuttles.^{61–63} Such photo-switchable modifications can be switched between multiple molecular states *via* light irradiation.^{64–69}

There is a continuing need for biologically active molecular concentration gradients or local, precise surface modifications to decipher cellular functions *in vitro*.^{70–72} One possibility to improve the modification quality of the surface is irreversible patterning *via* laser-assisted protein adsorption by photobleaching (LAPAP).⁷³ Here, photopainting is introduced by intentional bleaching of fluorophores, where the formed radicals subsequently bind to the surface. LAPAP has been introduced using lamps and photomasks to covalently bind biotin-4-fluorescein (B4F) to bovine serum albumin (BSA) adsorbed on a glass surface for surface functionalization.⁷⁴ These experiments yielded resolution-limited molecular patterns. Recently, the resolution has been extended to the nanoscale by combining LAPAP and nanostructured polymeric anchors fabricated *via* stimulated emission depletion (STED) nanolithography.^{75,76}

Our motivation to improve the LAPAP efficiency was driven by spectroscopic studies by Stennett *et al.* and Ciuba *et al.*^{77,78} It has been demonstrated that for TAMRA, Cy3/Cy3B and Cy5 in aqueous solvent, Mn²⁺ in the solution increases the intersystem crossing (ISC) efficiency. Related studies on the applicability of cyanine dyes show that they reversibly switch between a fluorescent state and a dark state when illuminated at different wavelengths.^{79,80}

In the context of LAPAP, photobleaching and radical formation can be induced using paramagnetic transition metal- (*e.g.* Zn²⁺) or heavy metal ions (*e.g.* iodine) as well as ROS-promoters.^{81–83} The role of ROS in the photobleaching process of several classes of fluorophores fostered the idea of using photoactive molecules as ROS promoters for increased

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† Electronic supplementary information (ESI) available. See DOI: <https://doi.org/10.1039/d3lf00125c>

photopainting efficiency.⁸⁴ Water soluble porphyrin is a well-known ROS promoter, due to singlet oxygen formation.^{85–88}

In this work, we present new buffer systems for improved LAPAP patterning efficiency. We characterise the effect of Mn^{2+} as well as ruthenium porphyrin (RuPo) on three fluorophore class representatives by optimizing buffer systems for single-molecule LAPAP.

Our buffer systems enhance the efficiency for single molecule LAPAP (smLAPAP) on Multi-Photon Lithography (MPL) structured scaffolds. Experiments with representatives of cyanine- (Cy3), coumarin- (ATTO 390) and rhodamine- (ATTO 514) fluorophores have been performed. The interaction of Mn^{2+} has been corroborated by quantum mechanical calculations yielding optimal fluorophore/ Mn^{2+} complexes with specific ion binding sites. Using our optimized buffers, single oligonucleotides and biotins have been photopainted efficiently on 2D/3D acrylate scaffolds with low unspecific surface adsorption. Photopainting efficiency has been confirmed using single molecule fluorescence microscopy (SMFM).

2 Experimental section

2.1 Materials and methods

2.1.1 Structuring and photoresist. For multi-photon lithography (MPL), a mixture of the monomers Ormocomp^{89,90} (Micro Resist Technology, Germany) and poly(ethylene glycol) diacrylate (PEGDA, $M_n = 575$, Sigma Aldrich, USA) was used in a 4:1 mass ratio. 1 wt% of Irgacure 819 (Sigma Aldrich, USA) was added as photoinitiator.

2.1.2 Chemicals. Bovine serum albumin (BSA, Sigma Aldrich) was used for surface passivation. LAPAP was performed with Cy3 conjugated to the model oligonucleotide NP77 (ref. 91) (sequence: CGC TAG ACT AGT ACA ATT GGC AGA CTG CCT TGA TCG GTA CGA ACA GAT CGC GTC AGG TAT, 3' modification: C6-spacer amino modification – Cy3 carboxylic acid) (Cy3-NP77, Microsynth AG, Switzerland) (“Cy-LAPAP”), ATTO 390 conjugated to biotin (ATTO390-biotin, ATTO-TEC GmbH, Germany) (“Cou-LAPAP”) and ATTO 514 conjugated to biotin (ATTO514-biotin, ATTO-TEC GmbH, Germany) (“Rho-LAPAP”). For visualization of the fluorophore–biomolecule conjugates bound *via* LAPAP, ATTO 647N conjugated to the complementary oligonucleotide cNP77 (sequence: ATA CCT GAC GCG ATC TGT TCG TAC CGA TCA AGG CAG TCT GCC AAT TGT ACT AGT CTA GCG, 5' modification: C6-spacer amino modification – ATTO 647N) (ATTO647N-cNP77, Microsynth AG, Switzerland) was used for Cy-LAPAP and ATTO 655 conjugated to streptavidin (ATTO655-SA, ATTO-TEC GmbH, Germany) was used for Cou-LAPAP and Rho-LAPAP. Ethylenediamine tetra acetic acid disodium salt (EDTA) (Roth, Germany) dissolved in double-distilled water and phosphate buffered saline (PBS) were used for washing the sample.

2.1.3 Buffer systems. Buffer I consisted of double-distilled water (Labochem, Germany) (Cy-LAPAP) or PBS (Cou-LAPAP, Rho-LAPAP). Buffer II consisted of buffer I with 2.5 mM

manganese(II)-chloride tetrahydrate ($\text{MnCl}_2 \cdot 4\text{H}_2\text{O}$, “ Mn^{2+} ”) (Sigma Aldrich, USA). Buffer III consisted of buffer II with 1 μM tris-(2,2'-bipyridyl)-ruthenium(II)-chloride hexahydrate ($\text{C}_{30}\text{H}_{24}\text{Cl}_2\text{N}_6 \text{Ru} \cdot 6\text{H}_2\text{O}$, “RuPo”) (Sigma Aldrich, USA).

2.2 Lithography setup

MPL was performed with a customized lithography system (Workshop Of Photonics (WOP), Lithuania) using an ultra-short pulsed laser at 515 nm (CARBIDE, 1 MHz repetition rate, >290 fs pulse duration, Light Conversion) and a 3-axis stage (AEROTECH Nanopositioner, USA) for sample movement, as described previously.⁹² The same instrument was used for imaging in a confocal configuration. The scattered light was collected using an Avalanche PhotoDiode (APD) (PDM, Micro Photon Devices, Italy) operated with a custom-made software ($\lambda_{\text{exc}} = 515 \text{ nm}$ fs pulsed, $I = 5 \mu\text{W}$). MPL, imaging, and LAPAP were performed with an oil objective lens (63 $\times$, 1.4 NA, Zeiss, Germany).

2.3 Single molecule fluorescence microscopy (SMFM)

Fluorescence images were acquired using a modified Olympus IX81. A nanometre precision XYZ piezo stage (Physik Instrumente GmbH, Germany) on top of a mechanical stage with $1 \times 1 \text{ cm}$ range (JPK Instruments, Germany) was used for sample positioning. The sample was illuminated through a 60 $\times$ magnification objective lens (60 $\times$, 1.42 NA, Olympus, Austria) and an additional 1.6 $\times$ magnification tube lens with a 642 nm diode laser (Omicron, Germany). A dichroic filter (ZT405/488/561/640rpc, Chroma, Germany), an emission filter (446/523/600/677 nm BrightLine quad-band band-pass filter, Semrock, USA), and an additional emission filter (HQ 700/75 M, NC209774, Chroma, Germany) were used for imaging of the probes of the immobilized fluorophore–biomolecule conjugates (ATTO647N-cNP77 for Cy-LAPAP and ATTO655-SA for Cou-LAPAP and Rho-LAPAP). The fluorescence signal was detected using an Andor iXonEM+ 897 (back illuminated) EMCCD camera (16 μm pixel size) (Andor Technology, United Kingdom).⁹³ 1500 images were recorded with 20 ms illumination time and 4.15 kW cm^{-2} excitation intensity at $\lambda_{\text{exc}} = 640 \text{ nm}$.

2.4 Sample preparation

Two- and three-dimensional MPL acrylate scaffolds (photoresist described above) were prepared with an excitation intensity of 42 GW cm^{-2} at a 1 mm s^{-1} writing speed and developed by rinsing the sample with acetone (Roth, Germany) (air-dried). Surface passivation was achieved *via* incubation with 2% BSA solution in PBS for 1 h at 4 $^\circ\text{C}$. The remaining BSA was washed out in 3 washing steps with PBS.

We prepared solutions of: 1 μM Cy3-NP77 oligonucleotide (Cy-LAPAP), 50 μM ATTO390-biotin (Cou-LAPAP) and 20 μM ATTO514-biotin (Rho-LAPAP) for LAPAP. Positioning of the laser's focal spot during LAPAP was achieved by detecting the



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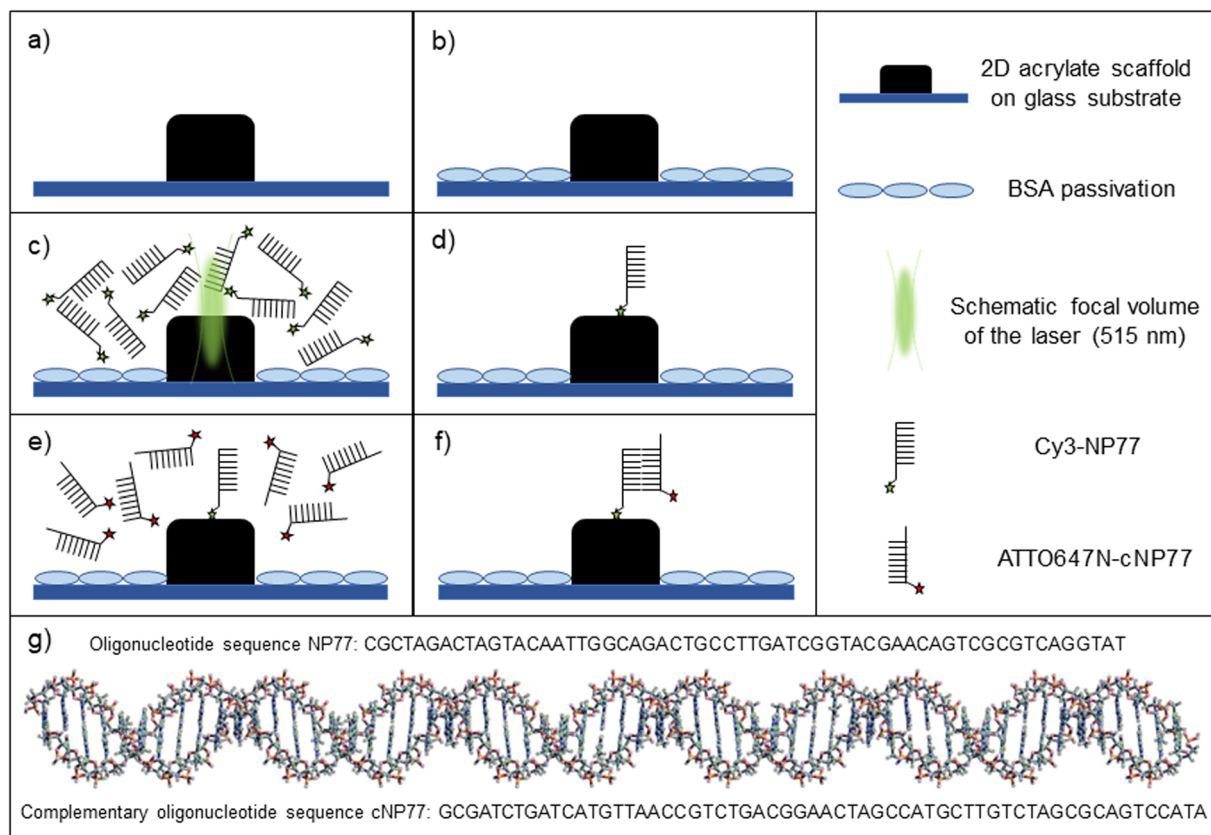


Fig. 1 Schematic representation of the 2D acrylate scaffold functionalization via LAPAP at a single molecule level (smLAPAP). (a) A 2D acrylate scaffold (4 : 1 weight ratio Ormocomp : PEGDA, 1 wt% Irgacure 819) was prepared via MPL on glass substrate. (b) Passivation of the surrounding glass substrate with BSA. (c) Incubation with Cy3 conjugated to NP77- (Cy3-NP77) model oligonucleotide and photobleaching of the fluorophores in the focal spot of the laser (515 nm) (LAPAP). (d) After washing, only bleached fluorophores bound to the polymer-substrate remain on the scaffold. (e) Incubation with ATTO 647N labelled cNP77 (ATTO647N-cNP77) complementary oligonucleotide. (f) Immobilized oligonucleotide helix on the polymer surface. (g) Oligonucleotide helix composed of NP77 and cNP77. Sketches are not to drawn scale.

ATTO647N-cNP77 and ATTO655-SA, which were used as probes to reveal the LAPAP immobilized biomolecules (see Materials and methods).

More specifically, for Cy-LAPAP, we immobilized a cyanine derivative (Cy3) conjugated to an NP77-model oligonucleotide (Cy3-NP77) on the acrylate scaffold. The immobilized oligonucleotides were detected by incubation with the fluorescently labelled complementary cNP77 oligonucleotide (ATTO647N-cNP77). For Cou-LAPAP we used a coumarin derivative (ATTO 390) conjugated to a biotin (ATTO390-biotin). For Rho-LAPAP we used a rhodamine derivative (ATTO 514) conjugated to a biotin (ATTO514-biotin). The immobilized biotins (Cou-LAPAP, Rho-LAPAP) were detected with ATTO 655 conjugated to a streptavidin (ATTO655-SA) and imaged by SMFM.

Ciuba *et al.*⁷⁸ have previously shown that buffers containing paramagnetic transition metal ions (*e.g.* Mn^{2+}) have a pronounced effect on photobleaching driven by enhanced ISC. In addition to Mn^{2+} , we have investigated the effect of a co-initiator that promotes radical formation, *i.e.*, bleaching, of the fluorophore *via* enhanced singlet state oxygen formation. As a co-initiator, tris-(2,2'-bipyridyl)-ruthenium(II)-chloride hexahydrate (RuPo) has been chosen,

which is an efficient singlet oxygen generator^{111–113} with an absorption spectrum matching the LAPAP excitation wavelength¹¹⁴ (see ESI† Table S5 for presumed 1-photon or 2-photon absorption).

Therefore, we have developed two new buffer systems containing Mn^{2+} and Mn^{2+} + RuPo, respectively. Buffer I consists either of double-distilled water (Cy-LAPAP) or PBS (Cou-LAPAP, Rho-LAPAP). Buffer II consists of Mn^{2+} in buffer I. Buffer III consists of Mn^{2+} and RuPo in buffer I. All in all, we have tested three fluorophore classes with three buffers (see Materials and methods).

In a first step, the patterning capabilities were tested on 2D acrylate scaffolds. The scaffolds ($10 \times 100 \times 2 \mu\text{m}$ bars with a periodicity of $10 \mu\text{m}$) consisted of Ormocomp⁸⁹ and poly(ethylene glycol)diacrylate (PEGDA, weight ratio 4 : 1) (see ESI† Fig. S1).

For precise positioning of the photobleaching laser beam on the polymer surface, the interface was determined by detecting the light scattered from the polymer/buffer interface. For that, our MPL lithography setup has been used in confocal imaging mode.⁹² Positioning in axial-direction (sub-100 nm) of the excitation point spread function (PSF) on the polymer surface, laser intensity and

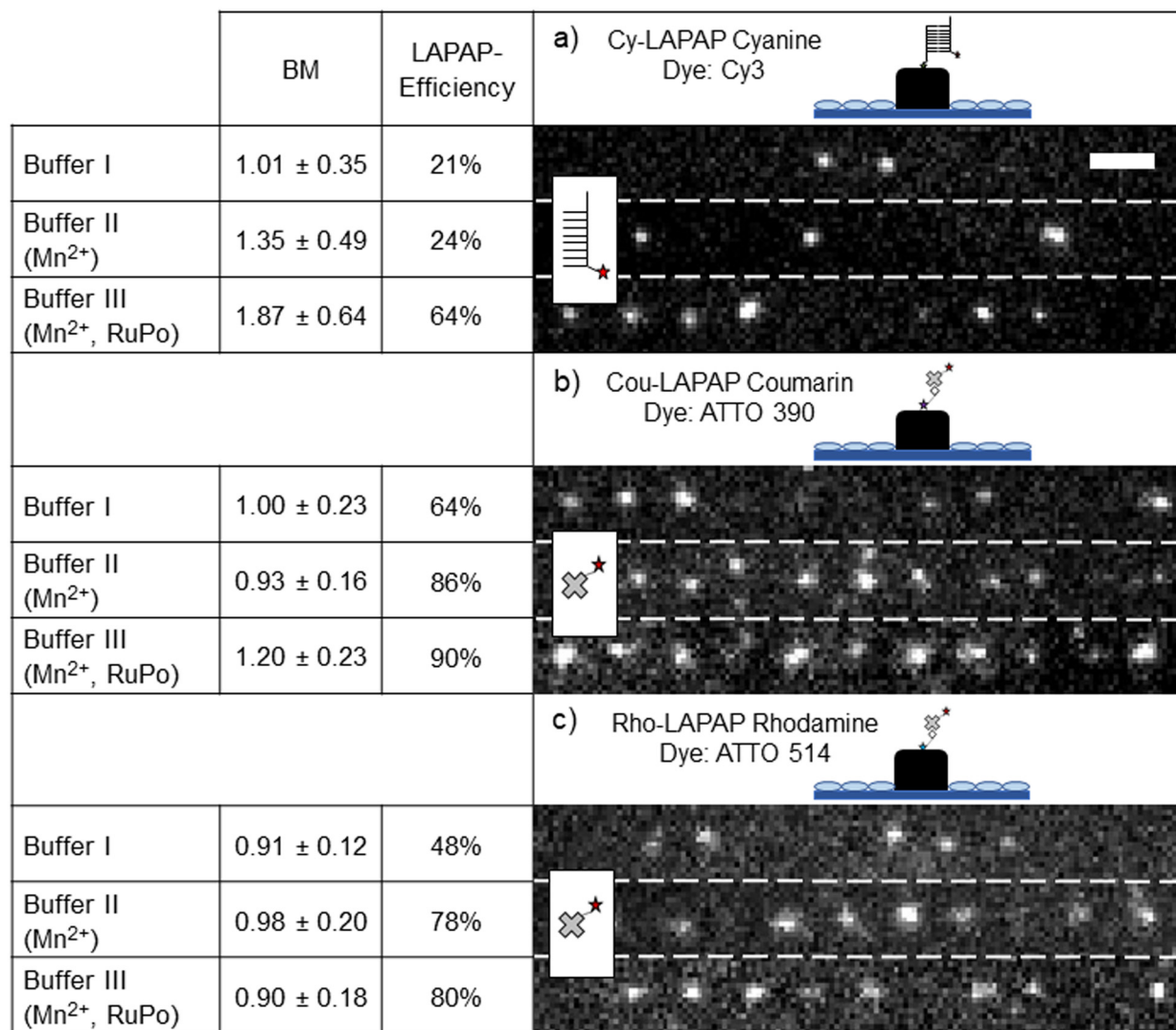


Fig. 2 LAPAP on 2D scaffolds. 3 buffer systems (buffer I: neat water for Cy-LAPAP and PBS for Cou-LAPAP and Rho-LAPAP; buffer II: buffer I + Mn^{2+} ; buffer III: buffer I + Mn^{2+} + RuPo) with different fluorophore families were tested for LAPAP. Immobilized molecules were confirmed with ATTO647N-cNP77 and ATTO655-SA, respectively. (a) Cy-LAPAP cyanine dye: Cy3-NP77. With buffer I, 1.01 ± 0.35 BM at LAPAP-efficiency of 21% was observed. With buffer II, BM increases to 1.35 ± 0.49 with a LAPAP-efficiency of 24%. Buffer III yielded 1.87 ± 0.64 BM, and an increase of LAPAP-efficiency by three-fold (64%). (b) Cou-LAPAP coumarin dye: ATTO390-biotin. With buffer I, 1.00 ± 0.23 BM at LAPAP-efficiency of 64% was estimated. With buffer II, LAPAP-efficiency increased to 86% and 0.93 ± 0.16 BM. Buffer III yields 1.20 ± 0.23 BM with a LAPAP-efficiency of 90%. (c) Rho-LAPAP rhodamine dye: ATTO514-biotin. With buffer I, 0.91 ± 0.12 BM at LAPAP-efficiency of 48% was determined. With buffer II, LAPAP-efficiency increased to 78% and 0.98 ± 0.20 BM. Buffer III yielded similar results as buffer II. Scale bar for all fluorescence images 2 μm .

pulse duration were adjusted. For LAPAP optimization, parameters such as BM and LAPAP-efficiency were evaluated for each experiment (see Fig. 2).

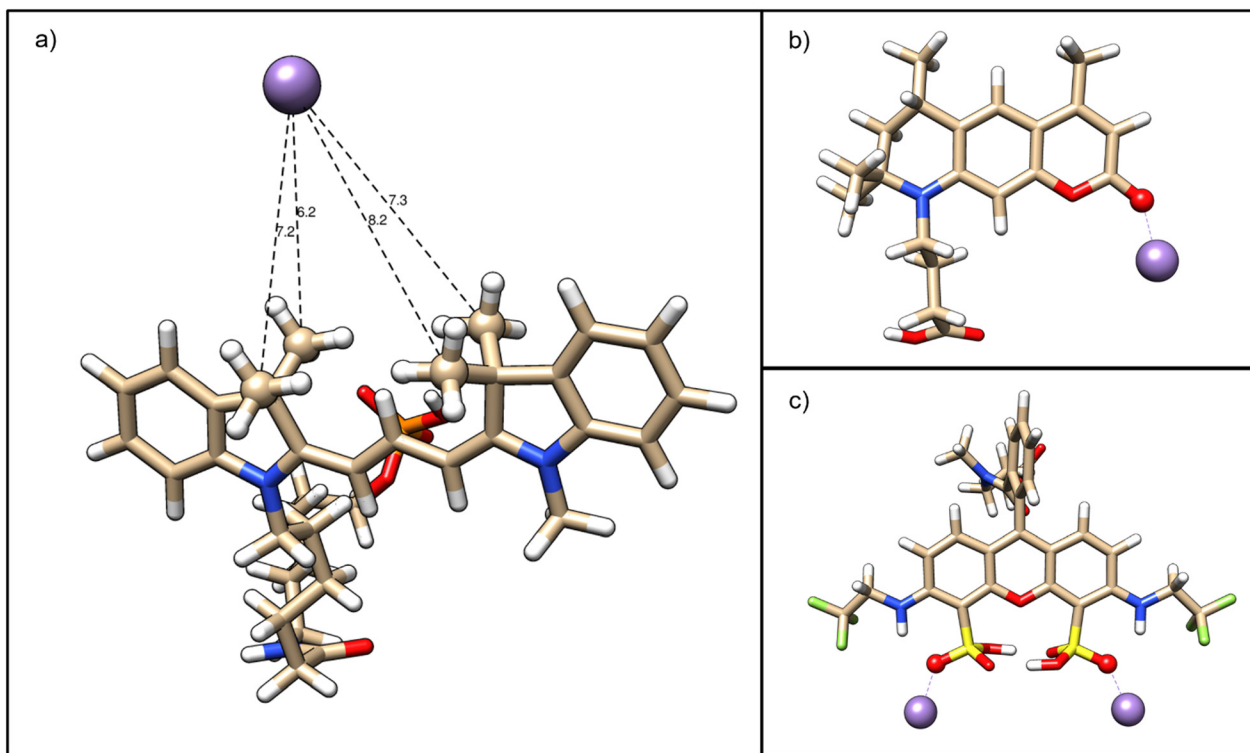
For BM estimation, average SMIs of individual ATTO647N-cNP77 and ATTO655-SA probe molecules were determined prior to LAPAP experiments. The SMI of both probe molecules, sparsely distributed on a glass surface, was 357 ± 29 arb.u. and 292 ± 28 arb.u., respectively (see Materials and methods).

Fig. 2 shows the number of bound biomolecules (BM) and the LAPAP efficiencies for all dyes and buffer systems investigated. Hereby, LAPAP-efficiency refers to the percentage of fluorescent spots on the scaffold while BM refers to the average number of bound fluorophores on the

scaffold, which do show fluorescence, estimated from the signals in a diffraction-limited LAPAP area (FWHM ~ 330 nm, see ESI† Fig. S2). As shown in Fig. 2a, Cy-LAPAP with buffer I yielded 1.01 ± 0.35 BM and LAPAP-efficiency of 21% ($n_{\text{images}} = 5$, $n_{\text{scaffold}} = 3/\text{image}$ histogram mode = 1). This means, that one out of five LAPAP-modified spots on the scaffold showed some fluorescence and, in those cases, the fluorescence intensity corresponded to that of one fluorophore, on average. Buffer II increased the BM to 1.35 ± 0.49 with 24% LAPAP efficiency ($n_{\text{images}} = 4$, $n_{\text{scaffold}} = 3/\text{image}$, histogram mode = 1). Buffer III yields 1.87 ± 0.64 BM, and LAPAP-efficiency is increased to 64% ($n_{\text{images}} = 6$, $n_{\text{scaffold}} = 3/\text{image}$, histogram mode = 2) ($t_{\text{pulse}} = 1$ ps, 3000 pulses per spot). For

The highest LAPAP-efficiency was achieved for buffer III with Cou-LAPAP yielding up to 90% of functionalized diffraction limited LAPAP illumination areas (see Fig. 2b). The biggest increase in LAPAP-efficiency was observed for Cy-

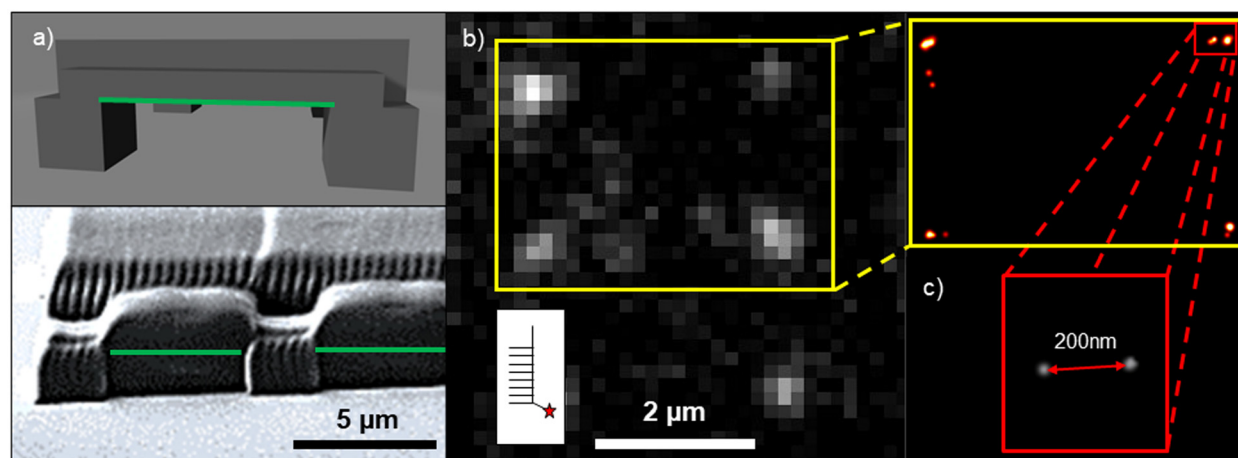
For ATTO 390 (see Fig. 3b), our approach identified a negatively charged furan ring oxygen ($q = -0.57e$) and an even more strongly negatively charged ketone oxygen ($q = -0.68e$) attached to the furan ring, which is directly connected to the



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Our buffers allow efficient smLAPAP with 1.4 times reduced excitation intensity compared to standard buffers for tested fluorophore classes (for *e.g.*, Cy3-LAPAP: buffer I: 85 GW cm⁻², with buffer III: 60 GW cm⁻²). At LAPAP excitation intensity settings greater than 90 GW cm⁻² ($t_{\text{pulse}} = 1$ ps, 250 pulses per spot) the damage threshold is exceeded, and upon photo damage, the illuminated areas exhibit an unbleachable autofluorescence. In comparison, the “true” LAPAP spots were only visible after incubation and showed a stepwise photobleaching behaviour (see ESI† Fig. S3). The combined decrease of photodamage likelihood and excitation intensity for enhanced LAPAP-efficiency in buffer III will

Fig. 4b shows fluorescence signals of ATTO647N-cNP77 complementing Cy3-NP77 immobilized *via* LAPAP on a 3D MPL scaffold Fig. 4c illustrates the localized emitters of the 4 spots in the yellow insert in Fig. 4b. In the red insert two ATTO647N-cNP77 are presented. The emitters are separated by ~ 200 nm within the LAPAP spot. Each emitter has been localized with an average position accuracy (PA) of 24 ± 5 nm.



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Single molecule intensity analysis of the same spot confirms this assumption (intensity of spot = 702 arb.u. divided by SMI = 357 ± 29 arb.u. = 1.97 BM).

Conclusions

In summary, we demonstrated that our newly developed buffer systems significantly improve the LAPAP-efficiency and allow for reduced painting power, enabling smLAPAP on MPL structured 2D and 3D acrylate scaffolds. We showed that our buffer systems improve LAPAP efficiency for Cy3, ATTO 514 and ATTO 390.

Increased LAPAP efficiency can be attributed to specific electrostatic Mn^{2+} -fluorophore interactions as previously argued by Ciuba *et al.*,⁷⁸ which we characterized in quantum mechanical DFT calculations. More precisely, the calculations predict a rather weak binding pocket for Mn^{2+} in Cy3, where LAPAP efficiency increases only slightly. For Cou-LAPAP and Rho-LAPAP, where the Mn^{2+} binding is stronger, the LAPAP efficiency increased from 64% to 86% and from 48% to 78%, respectively. The oligonucleotide-, biotin-modifications have not been considered in the calculations. Additional enhancement of the LAPAP efficiency has always been observed for RuPo-containing buffers. The presented experimental results clearly show the positive influence of paramagnetic transition metal ions and ROS on LAPAP efficiency and could expand its application range as a surface modification tool. Further investigations, in particular spectroscopic and computational studies, will be needed to thoroughly characterize the effects of ligand binding on the photobleaching reaction as well as ISC rates and to further refine the LAPAP-mechanism.

Our results pave the way for future LAPAP applications with low power light sources, including but not limited to surface modifications for biotechnological application for *e.g.*, 3D cell cultures or *in vitro* assays as well as molecular sensors which require local modifications.

Author contributions

J. J. and C. N. designed the experiments. C. N. performed the experiments and analysed the data. H. K. performed the quantum mechanical calculations. G. G. distributed 2D and 3D scaffolds. D. S. and T. K. helped interpret and analyse the data. C. N. H. K., D. S. and J. J. wrote the manuscript. All authors reviewed and approved the final manuscript.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

We thank Heidi Piglmayer-Brezina for taking SEM images of the 2D and 3D acrylate scaffolds. We would also like to thank Univ.-Prof. Dr. Thomas Renger for fruitful discussions. We thank Dr. Markus Axmann for proofreading the manuscript.

The work was supported by Austrian Science Fund *via* grant P 31827-B.21.

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