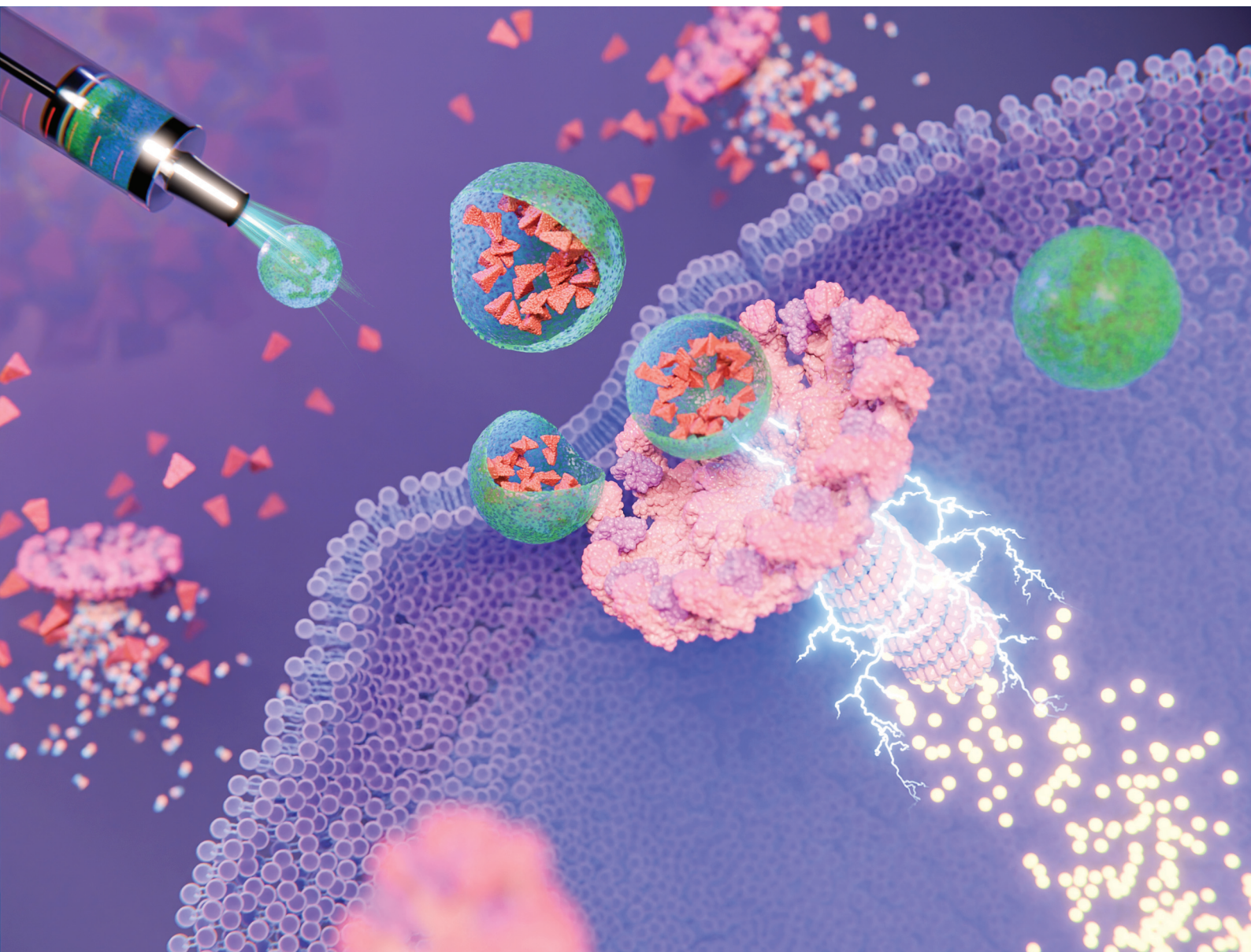


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REVIEW

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Role of NLRP3 inflammasome in nanoparticle adjuvant-mediated immune response

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The nucleotide-binding oligomerization domain (NOD)-like receptor (NLR) family pyrin domain-containing 3 (NLRP3) inflammasome is pivotal in orchestrating the immune response induced by nanoparticle adjuvants. Understanding the intricate mechanisms underlying the activation of NLRP3 inflammasome by these adjuvants is crucial for deciphering their immunomodulatory properties. This review explores the involvement of the NLRP3 inflammasome in mediating immune responses triggered by nanoparticle adjuvants. It delves into the signaling pathways and cellular mechanisms involved in NLRP3 activation, highlighting its significance in modulating the efficacy and safety of nanoparticle-based adjuvants. A comprehensive grasp of the interplay between NLRP3 inflammasome and nanoparticle adjuvants holds promise for optimizing vaccine design and advancing immunotherapeutic strategies.

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Introduction

Vaccines are essential public health interventions that protect against serious illness and complications. The outbreak of COVID-19 [caused by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2)] has raised a massive demand for the development of vaccines.^{1,2} The successful cooperation between multi-disciplinary research, such as virology, biology, chemistry, medicine, bioengineering, computational sciences, and nanotechnology, revolutionized vaccine development in record time.^{3,4} Vaccination is the process of delivering antigenic substances (*i.e.*, live attenuated, inactivated, viral vector, nucleic acid (DNA/RNA), or recombinant protein) to develop the immune response towards specific pathogens and infectious diseases (Fig. 1).^{5–8} However, in several instances, antigens or antigenic determinants are combined with substances that enhance the antigen-specific immunity regarding the breadth of the immune response and are called “vaccine adju-

vants”.⁹ Despite immunogenicity, the addition of an adjuvant to the vaccine brings several advantages, including potency, reduction in the booster doses, cost, stability, and others.^{7,10} Adjuvants can provide a swift and efficient immune response and will be effective towards a wide range of pathogens, even with antigenic drift (“The genetic diversity in viruses occurs due to the accumulation of mutations in the genes responsible for coding virus-surface proteins that are recognized by host antibodies.”).¹¹ Adjuvants are classified broadly as immunomodulatory molecules (including mineral salts, microbial products, cytokines, glycolipids [saponins], polymers, and others.) and non-immunostimulatory delivery systems (such as lipid nanoparticles [LNPs], emulsions, and others.) or combinations of both.^{12,13}

Since 1920, various forms of aluminum salts (alum) containing adjuvants have been used as successful adjuvants in many human vaccines, such as tetanus, diphtheria toxoids, and others.^{15,16} In general, antigens are bound to the adjuvants *via* non-covalent binding including electrostatic, hydrophobic interactions, hydrogen bonding, and van der Waals forces.¹⁷ Later in the 1990s, new adjuvants, *i.e.*, the oil-in-water emulsion system (MF59), were developed to deliver influenza vaccine for the elderly.¹⁸ Emulsion adjuvants are based on either oil-in-water or water-in-oil and a three-phase water-oil-water system, where purified mineral oils and biodegradable oils (*e.g.*, squalene, squalane, and others) are used as oils in such emulsions.¹⁷ Further, liposome-based adjuvant (AS01) was used in the herpes zoster vaccine. AS01 is composed of two lipids, 3-O-desacyl-4'-monophosphoryl lipid A (MPL), a toll-like receptor 4 (TLR 4) agonist, and QS-21 (from *Quillaja*

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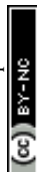
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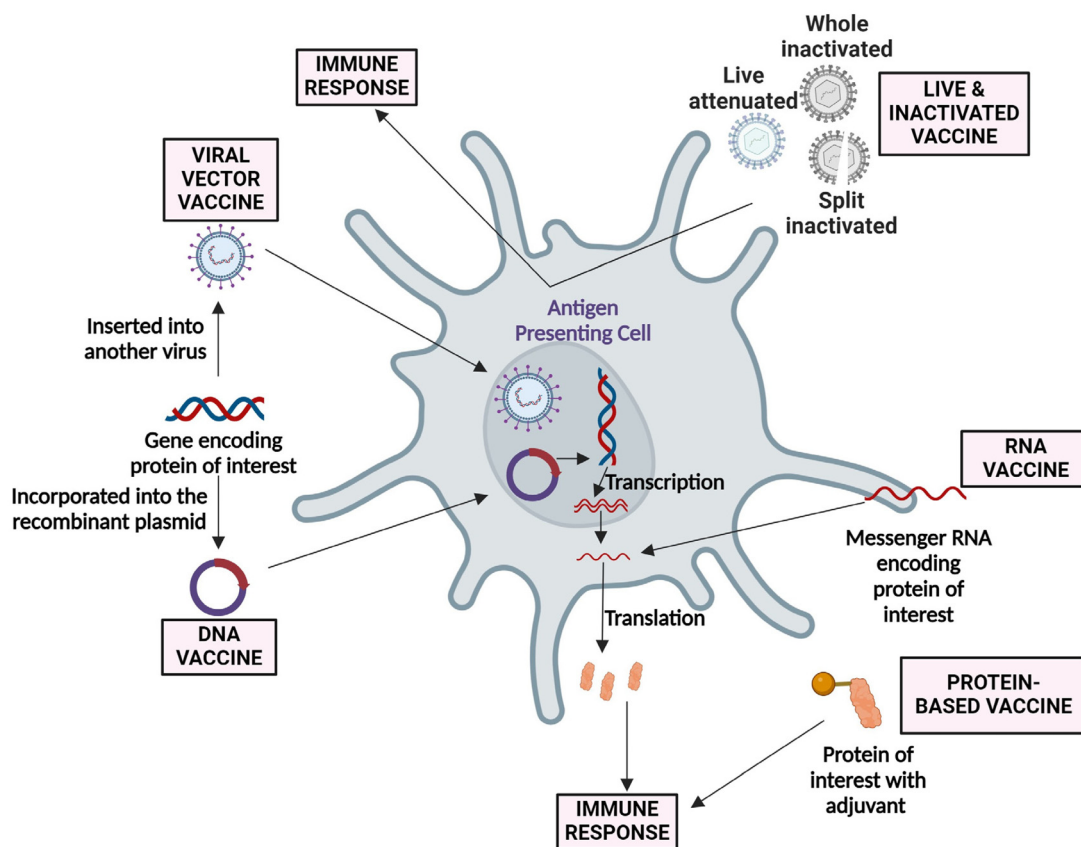


Fig. 1 Approaches followed in developing live, viral vector, DNA, RNA, and protein-based vaccine for SARS-CoV-2. Currently, most of the vaccines are in the clinical phase. An adenovirus-based vaccine (Ad5-nCoV) has been approved for military usage, and an RNA-based vaccine (mRNA-1273) is gearing up for phase III clinical trial.¹⁴

saponaria), which trigger the activity of NLRP3 inflammasome pathway.¹⁹ Later, the hybrid adjuvant AS04 (combination of MPL lipid and aluminum hydroxide) was developed for human hepatitis B and human papillomavirus vaccines.^{9,20} Further, in 1994, virosomes or virus-like particles (VLPs) purified from the influenza virus, which are spherical vesicles (~150 nm size) consisting of the lipid membrane and envelope proteins successfully approved as adjuvants for human vaccines.²¹ The sudden and unexpected COVID-19 outbreak prompted scientists and pharmaceutical firms to expedite the search for new adjuvants and innovative vaccine approaches to swiftly prevent SARS-CoV-2 infection. In this regard, the protein- or nucleic acid (mRNA or DNA)-based vaccines were seriously studied for the developing vaccines against COVID-19 due to their simplicity and facile scale-up possibility compared to inactivated vaccines. Undoubtedly, the knowledge gained from the development of COVID-19 vaccines will have a profound influence on the future development of vaccines, particularly those utilizing mRNA technology and LNPs.^{1,2} The use of mRNA technology reduced the cell culture experiments and lowered the chances of contamination compared to other conventional complicated vaccine methods.^{1,22} LNPs, used to deliver the nucleic acids, are known to act as vaccine adjuvants. A handful of studies have been published on the innate

immune response of empty LNPs.^{3,23,24} In addition, some novel adjuvants have been approved for COVID-19 vaccines, and several have been in clinical trials. A saponin-based adjuvant (*i.e.*, Matrix-M) was used to develop the recombinant protein vaccine (NVX-CoV2373) under clinical evaluation in Phase II.²⁵ A recent study in 2020 reported the usage of alum as an adjuvant in an inactivated vaccine against SARS-CoV-2 developed by Beijing-based Sinovac Biotech, which is in clinical trials.²⁶ This review encompasses the studies conducted by multiple scientists investigating a diverse range of adjuvants, mainly focusing on nanomaterials. These adjuvant nanomaterials play a crucial role in activating the NLRP3 inflammasome and stimulating the immune response in the host, which is advantageous in the field of vaccinology.

In our day-to-day life, people come in contact with various microorganisms, leading to multiple diseases/infections, some of which are life-threatening. To protect and keep ourselves healthy, we need efficacious vaccines that can help us fight against these infections. Earlier vaccines were not so potent, and to address the same, scientists have investigated ways to produce more potent vaccines that are also cost-effective and provide a rapid and effective immune response. The idea was to add adjuvants as they fulfil all the criteria to develop potent and effective vaccines.



vants, such as chitosan, liposomes, biodegradable microparticles, and nanoparticles, are showing promising results.^{30,31} These alternative adjuvants can improve the immune response, leading to the development of efficient vaccines.

Inflammasomes are crucial in the immune system and are essential for activating inflammatory caspases.³⁵ These protein complexes contain specialized pattern recognition receptors (PRRs) that bind to pro-caspase-1 with the help of an adaptor molecule called apoptosis speck-like protein that contains a caspase activation and recruitment domain (ASC).^{36,37} Understanding the functioning of inflammasomes can help us develop new approaches to combat inflammatory diseases and promote overall health.³⁸ There are two types of inflammasomes: the 'canonical inflammasomes' and the 'non-canonical inflammasomes'. The 'canonical inflammasomes', which

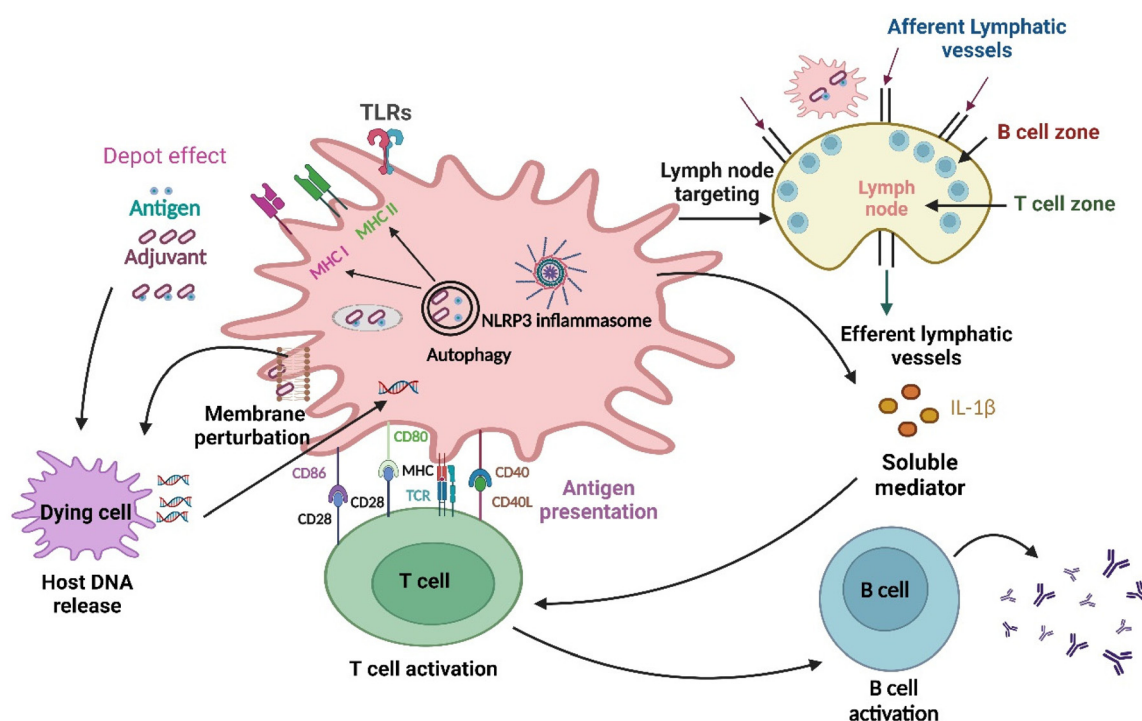


Fig. 2 Different pathways through which nanomaterials activate the immune system: at first, the antigen is released into the bloodstream at a rate regulated by adjuvants, which is the depot effect, and subsequently, the NLRP3 inflammasome gets activated. Changes in the lipid membrane structure also occur in the dendritic cell membrane, leading to the activation of an abortive phagocytic response. The delivery of extracellular and intracellular antigens to major histocompatibility complex (MHC) class II and I molecules for presentation to CD4⁺ T and CD8⁺ T lymphocytes, respectively, is facilitated by autophagic regulation.³² Further nanoparticles as adjuvants are delivered to the draining lymph nodes (primary site for immune activation), which target the immune cells residing in them and activate those cells.³³ Furthermore, innate and adaptive immune responses are activated by the TLR4 with help from the nanoparticles. The development of humoral immunity by repetitive antigen display on the B cell receptor leads to co-aggregation, triggering, and activation of B cells. The cell-based immunity differentiation of T cells occurs in Th1 or Th2 effector cells. Dendritic cells (highly specialized antigen-presenting cells) can take up foreign material in the extracellular milieu alongside MHC molecules, such as MHC I and MHC II, and also leads to cross-presentation, which is the shuttling of antigen to MHC class I processing pathway for the activation of CD8⁺ T cell. DNA released following cell damage can also trigger an innate immune response. Lastly, soluble mediators, such as cytokines, chemokines, and immunomodulatory substances, may impact innate and adaptive immune activity, including T cell polarization.³⁴

include NLRP1, NLRP3, NLRC4, AIM2, and pyrin, are well-known. The 'non-canonical inflammasomes' comprise caspase-11 in mice and caspase-4 or -5 in humans.^{35,39} In general, inflammasomes recognize various pathogen-associated molecular patterns (PAMPs), damage-associated molecular patterns (DAMPs), or any other disturbances in cellular homeostasis.⁴⁰ The activation of pro-caspase-1 by all canonical inflammasomes leads to the cleavage of pro-inflammatory cytokines, such as interleukin (IL)-1 β and IL-18, resulting in the secretion of their active forms. Activated caspase-1 also cleaves a pore-forming protein called Gasdermin D (GSDMD), which creates a large plasma membrane pore in the cell where the inflammasome was activated.^{41,42} This pore allows for the fast release of IL-1 β and IL-18, which leads to cell swelling, resulting in a pro-inflammatory form of cell death called "Pyroptosis". Furthermore, pyroptosis extends the inflammatory response by releasing more DAMPs and alarmins, including IL-1 α .⁴³ The downstream response triggered by all these canonical inflammasomes is a pivotal process in the immune system's response, and its understanding can lead to the development of more effective treatments for inflammatory diseases.³⁸

The mechanism of action of particulate adjuvants is not entirely understood yet. Still, recent studies have highlighted the potential of biodegradable poly(lactic-co-glycolic acid) (PLGA), polystyrene microparticles, and nanoparticles, when combined with TLR agonists, in significantly boosting NLRP3-dependent IL-1 β production by murine dendritic cells (DCs).⁴⁴ These discoveries are a significant step, as they are a progressive step towards understanding the mechanisms underlying inflammasome activation. Overall, these findings could lead to the development of new and effective methods for enhancing immune responses.⁴⁵ It has been noted that the inclusion of a TLR agonist is necessary for adjuvant-induced IL-1 β secretion *in vitro*.⁴⁶ However, the adjuvants alone can also stimulate IL-1 β secretion *in vivo*, indicating that endogenous factors can also play a role in the process.⁴⁷

Recent research has demonstrated that particulate adjuvants can activate the NLRP3 inflammasome, a significant breakthrough. While the mechanisms leading to this activation have been partially resolved, the scientific community is presently involved in understanding the role of NLRP3 in inducing adaptive immunity through particulate adjuvants.⁴⁸ NLRP3, a protein-coding gene, encodes a pyrin-like protein containing a pyrin domain, a nucleotide-binding site (NBS) domain, and a leucine-rich repeat (LRR) motif.⁴⁹ This protein interacts with the apoptosis-associated speck-like protein PYCARD/ASC, which contains a caspase recruitment domain and is a component of the NLRP3 inflammasome complex. This complex acts as an upstream activator of NF- κ B signaling and is crucial in regulating inflammation, immune response, and apoptosis.⁵⁰ The NLRP3 inflammasome is essential to the innate immune system and is crucial in protecting humans against microbial infections and cellular damage.⁵¹ However, the improper activation of this inflammasome has been linked to various inflammatory disorders, such as Alzheimer's disease, diabetes, atherosclerosis, cancer, cryopyrin-associated

periodic syndromes, and others.^{51,52} To better understand the mechanism behind NLRP3 activation, researchers have identified multiple molecular and cellular events that trigger its activation, including ionic flux, mitochondrial dysfunction, reactive oxygen species production, and lysosomal damage.^{53,54} The NLRP3 is an important intracellular protein encoded for NOD-, LRR-, and pyrin domain-containing protein 3, which works as a sensor protein for detecting various microbial protein motifs.⁴⁵ In particular, NLRP3 is responsible for the formation and activation of the NLRP3 inflammasome, thereby resulting in activating the pro-inflammatory response and secreting the IL-1 β and IL-18 cytokines *via* the caspase 1-dependent mechanism and inducing the inflammatory and pyroptotic cell death.^{45,55} Pyroptosis eliminates infected cells, which stops viruses from sustaining further reproduction in the event of a viral infection. It also liberates the cell's contents, amplifying the innate immune response even more.⁵⁶ GSDMD plays an essential role in the execution of pyroptosis. GSDMD activation depends on cleavage by caspases 1, 4, 5, 8, and 11, which release the N-terminal fragment, oligomerization, translocation, and ultimately perforation of the plasma membrane.⁵⁷ NLRP3 activation leads to GSDMD cleavage and allows caspases to cause inflammation and cell death. Despite being protective and essential to host defense, inappropriate activation of the NLRP3 inflammasome results in excessive release of cytokines and a hyperinflammatory response, both locally and systemically.⁵⁶ For NLRP3 to be an effective therapeutic target in the clinical treatment of inflammatory conditions, further research will be needed focusing on the physiological role of NLRP3 in healthy persons, as its role is not entirely understood.⁵⁸ However, the function of NLRP3 in physiology is unequivocally proven by preclinical models. They have a role in intestinal homeostasis regulation⁵⁹ and aging⁶⁰ and function as a crucial transcription factor in type II helper T cell (Th2) differentiation.⁶¹ They also have a role in metabolism, such as regulation of β -cell proliferation, insulin release, and triglyceride levels.⁵⁸ In the liver, it has been found to negatively regulate the development of non-alcoholic fatty liver disease (NAFLD).⁶² In cancer, NLRP3 inflammasome is critical in regulating tumor growth by directly triggering pyroptotic cell death or secreting cytokines that cause cell death.⁶³ Apart from its role in physiology, NLRP3 inflammasomes have been involved in many hepatic diseases, such as alcoholic liver disease, non-alcoholic steatohepatitis, viral hepatitis, liver fibrosis, and, most importantly, nanoparticle-induced liver injury.⁶⁴ Not only in liver diseases, the NLRP3 inflammasome also holds a pivotal position in managing the pathological processes in conditions such as acute myocardial infarction (AMI), systemic lupus erythematosus (SLE), inflammatory bowel disease (IBD), Crohn's disease, bacterial infections, ocular illnesses, and others.⁶⁵ Further research is needed to fully comprehend how NLRP3 responds to these signaling events and initiates the assembly of the NLRP3 inflammasome. NLRP3 was also found to be involved in autoinflammatory diseases and affects various infections in mice models. The pathogenesis of many acquired inflammatory diseases was



During the initial stages of vaccine development, various aluminum adjuvants, notably aluminum hydroxide (Alhydrogel) and aluminum phosphate (Adju-Phos), were successfully used as primary adjuvants in vaccines against tetanus, pertussis, and diphtheria.⁹³ Additionally, aluminum potassium sulfate (Potash Alum), a mixture of aluminum and magnesium hydroxides (Inject alum), and aluminum hydroxy phosphate sulfate (AHSA) are also part of aluminum-based

adjuvants.¹⁶ Indeed, the applied alum-based adjuvants are in the form of nanoscale materials against various infectious diseases.⁹⁴ Although alum-based adjuvants are poor simulators of cell-mediated immunity, their humoral immunity, cost-effectiveness, and high safety profile make them successful in vaccine development.⁹³

A study by Shi *et al.* compared the immunological effect of a rabies vaccine containing nano-sized aluminum particles (nano alum) and micron-sized aluminum (alum) along with five other adjuvants. Mice immunized with nano-alum showed protection and increased anti-rabies immunoglobulin (IgG) levels over other adjuvants.⁹⁵ Aluminum hydroxide is one of the best aluminum adjuvants widely used in vaccine development owing to its high safety profile. However, aqueous aluminum hydroxide particles form aggregates, resulting in micro-particles (increased size, 1–20 μm). Li *et al.* synthesized aluminum hydroxide nanoparticles (~112 nm) conjugated with model proteins (ovalbumin [OVA] and *B. anthracis* PA) to overcome this limitation. These conjugated nanoparticles induced higher antigen-specific antibody response than micro aluminum hydroxide adjuvants.¹⁶ Sun *et al.* developed γ -aluminum oxyhydroxide (AIOOH) nanorods functionalized with ovalbumin as a model protein to study the immune response. To understand the NLRP3 trigger mechanism in THP-1 cells and

murine bone marrow-derived dendritic cells (BMDCs), vivid AIOOH nanorods were synthesized with different shapes, crystallinity, and hydroxyl group contents. The results confirmed that nanorods with higher crystallinity showed better cellular uptake by forming agglomerates, wherein the shape, crystallinity, and hydroxyl content of AIOOH nanoparticles also played an essential role in the NLRP3 trigger mechanism and increased the OVA-specific immune response as shown in Fig. 3A.⁹⁶

Similarly, aluminum phosphate is also considered a commonly used aluminum salt-based adjuvant. Vrieling *et al.* synthesized aluminum phosphate nanoparticles by sonicating the commercially available aluminum phosphate.⁹⁸ Nanoparticles are stabilized with amino acids, such as threonine, asparagine, aspartic acid, and L-alanyl-L-1-aminoethylphosphonic acid (LAPA), with size distribution in the 400–600 nm range. The adsorption capacity of the particles was studied using lysozyme as a model protein, and it became slightly reduced after the functionalization with asparagine. The immune response induced by the aluminum phosphate particles was analyzed by immunizing mice with diphtheria toxoid adjuvanted with the particles. Only the arginine functionalization increased the specific IgG levels but didn't increase the diphtheria-neutralizing antibodies.⁹⁸ Lebre *et al.* developed chitosan-aluminum

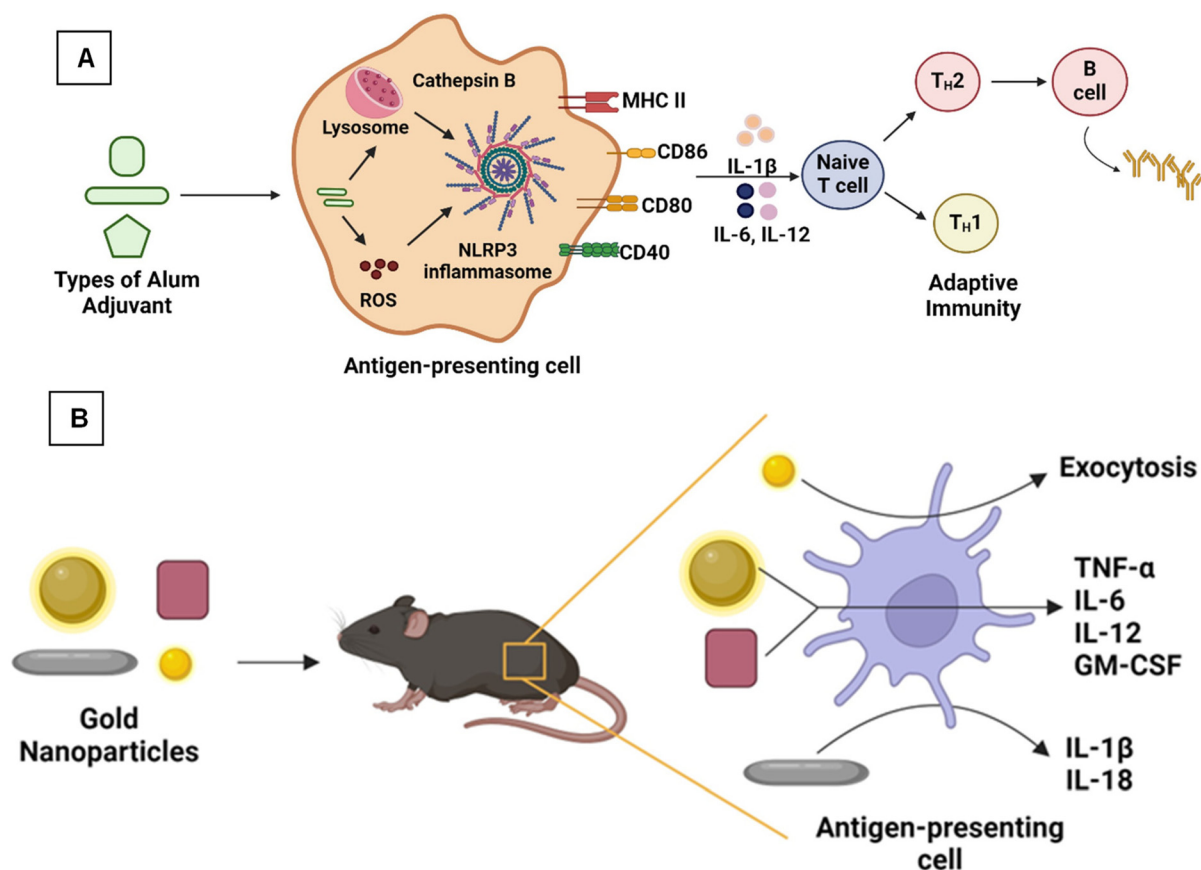


Fig. 3 (A) Aluminum oxyhydroxide nanoparticles showing an adaptive immune response⁹⁶ and (B) gold nanoparticles of different morphology producing antibodies specific to West Nile virus.⁹⁷



nanoparticles (CH-Al NPs) with a mean diameter of 280 nm, a positive surface charge, and better stability. Chitosan (CH) is a natural polysaccharide used for various bio-medical applications because of its low toxicity and biocompatibility.⁹⁹ The *in vitro* studies showed the low toxicity of the particles. The *in vivo* studies showed that mice immunized with hepatitis-B antigen (HBsAg) adsorbed CH-Al NPs showed a higher level of anti-HBsAg IgG.⁹⁹ Later, the same group tried to find out the mechanism of action of CH-Al NPs by undertaking similar *in vitro* and *in vivo* studies and found that CH-Al NPs showed NLRP3 activation mechanism (Fig. 3A), which induced cytokine secretion. It is concluded that CH-Al NPs induce innate and adaptive immune responses and can act as an efficient vaccine adjuvant, compared with alum and CH NPs.

Nanoparticle adjuvants activate NLRP3 inflammasome

Using nanoscale materials as an antigen delivery system increases their targeting ability and bioavailability, leading to increased uptake by the APCs, thereby presenting a better release profile of antigenic material.¹² Further, nanoscale materials are garnering attraction as nano-immunomodulators owing to their unique properties, such as immune suppression or simulating activity.⁴⁷ As shown in Fig. 4, various nanoscale

adjuvants, including metal and metal oxide-based vaccine adjuvants (*e.g.*, aluminum, gold, silver, SiO₂),^{95,100,101} polymeric (*e.g.*, chitosan, polystyrene, *etc.*) nanoparticles,^{102–104} and carbon-based materials, such as nanodiamonds and carbon nanotubes, have been developed.^{101,105}

Gold NPs (AuNPs) are widely explored materials in the biomedical field because of their unique physicochemical properties. Niikura *et al.* studied the effect of the size and shape of AuNPs in producing immunological responses.⁹⁷ The spherical, rod, and cubic-shaped AuNPs were studied *in vivo* and *in vitro*. The AuNPs were coated with West Nile Virus Envelope (WNVE) to understand the immune response. The 40 nm spherical AuNPs showed comparatively higher amounts of WNVE-specific antibodies in the mice. Whereas rods showed the highest level of internalization by APCs and induced the production of inflammatory cytokines (IL-1 β and IL-18), as shown in Fig. 3B.⁹⁷ In a similar study, Vandebriel *et al.* compared the effect of the shape of AuNPs in NLRP3 inflammasome activation. PEGylated gold nanorods, nanostars, and nanospheres were exposed to differentiated THP-1 cells (wildtype, ASC, or NLRP3-deficient), and then the cells were studied for gene expression analysis. NLRP3 inflammasome activation by gold nanorods decreased sterol/cholesterol biosynthesis, oxidative phosphorylation, and purinergic receptor signaling. A notable finding at the individual gene level was the low production of the protein paraoxonase-2 (PON2),

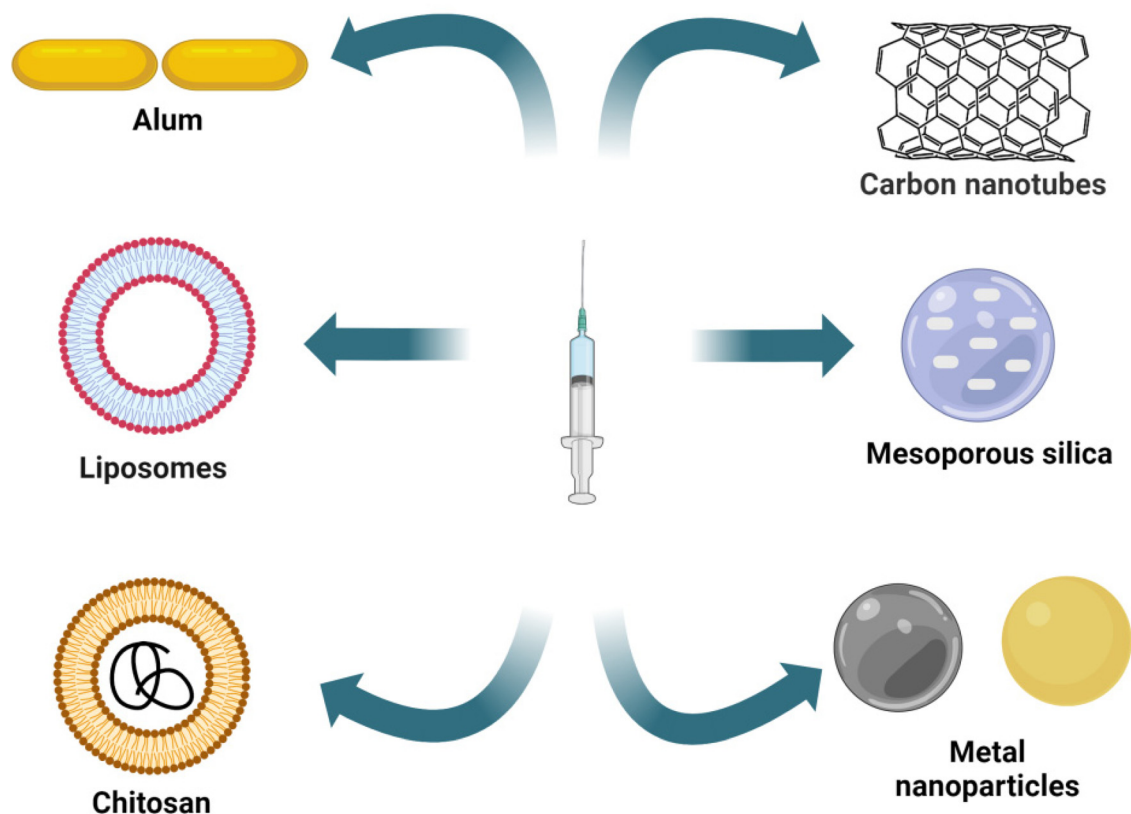


Fig. 4 Representation of different nanomaterials/nanoparticles used as vaccine adjuvants.



which is responsible for reducing oxidative stress and possibly related to increased ROS production.¹⁰⁰ In conclusion, these two studies proved that the surface area of these differently shaped and sized particles plays a prominent role in the immune response and inflammatory cytokine production.^{97,100}

Like AuNPs, silver nanoparticles (AgNPs) are also used as vaccine adjuvants. Xu *et al.* investigated the immunological effect of pristine AgNPs as vaccine adjuvants in both *in vivo* and *in vitro* conditions by using ovalbumin (OVA) and bovine serum albumin as model antigens. The mice immunized intraperitoneally (i.p.) with AgNPs showed increased levels of antigen-specific IgG, which depends upon the level of NPs used. Also, the NPs displayed Th2-biased immune responses. Finally, they have concluded that the immune response exhibited by the AgNPs is due to the recruitment and activation of local leukocytes, mainly macrophages.¹⁰¹

Polymeric nanomaterials are another class of nanomaterials being explored in the biomedical field. Polymers are considered one of the best biomaterials, mainly because of their flexible physical and mechanical properties, achieved by varying the chemistry.¹⁰⁶ Among them, PLGA is a widely used biomedical copolymer. Desai *et al.* synthesized PLGA-based nanospheres and used *Staphylococcal Enterotoxin B* (SEB) toxoid as a model vaccine. Both *in vitro* and *in vivo* (rabbit) studies of nanospheres containing SEB toxoid have been proven to exhibit enhanced immune response, almost similar to that of alum.¹⁰⁷ Gu *et al.* synthesized PLGA nanoparticles encapsulated with immunopotentiator *Angelica sinensis* polysaccharide (ASP). The nanoparticle's surface was coated with cationic polymer polyethyleneimine (ASP-PLGA-PEI) so that the positive charge on the nanoparticles would increase the

targeting and activation ability towards the APCs, thereby increasing the immune response. The immune response was further studied *in vivo* using porcine circovirus type 2 (PCV2) as a model protein known to cause porcine circovirus-associated disease (PCAVD).^{104,108} ASP-PLGA-PEI NPs effectively activated the macrophages and also enhanced antigen uptake by APCs. The *in vivo* studies showed a higher antigen-specific IgG immune response, indicating that ASP-PLGA-PEI NPs can act as vaccine adjuvants, as shown in the schematic representation of Fig. 5.¹⁰⁴ In a work by Wang *et al.*, biodegradable poly (g-glutamic acid) (g-PGA) nanoparticles were used for antigen delivery and activation of the immune system both *in vitro* and *in vivo*. FITC-labelled nanoparticles encapsulating Texas Red-labelled ovalbumin (TR-OVA/FITC-NPs) were used to study the cellular intake of OVA. An increased concentration of OVA in the cells was observed compared to control samples. To examine the adjuvant effect of nanoparticles, mice were immunized with human immunodeficiency virus type 1 (HIV-1) p24 encapsulating NPs, which induced HIV-1-specific humoral and cellular immune responses.¹⁰⁹

CNTs are one of the allotropic forms of carbon, with a rolled-up graphene layer structure, sometimes with fullerene caps. CNTs are used extensively in different fields, including the biomedical field, because of their unique physicochemical, mechanical, optical, and electronic properties.¹¹⁰ Zhu *et al.* studied the immune-responsive and adjuvant properties of multi-walled carbon nanotubes (MWCNTs) both *in vitro* and *in vivo*.¹⁰⁵ The MWCNTs were covalently functionalized with model antigen ovalbumin (MWCNT-OVA). Increased uptake of MWCNT-OVA by mouse dendritic cells (DC2.4 cell line) was observed along with the upregulation of co-stimulators (CD40/

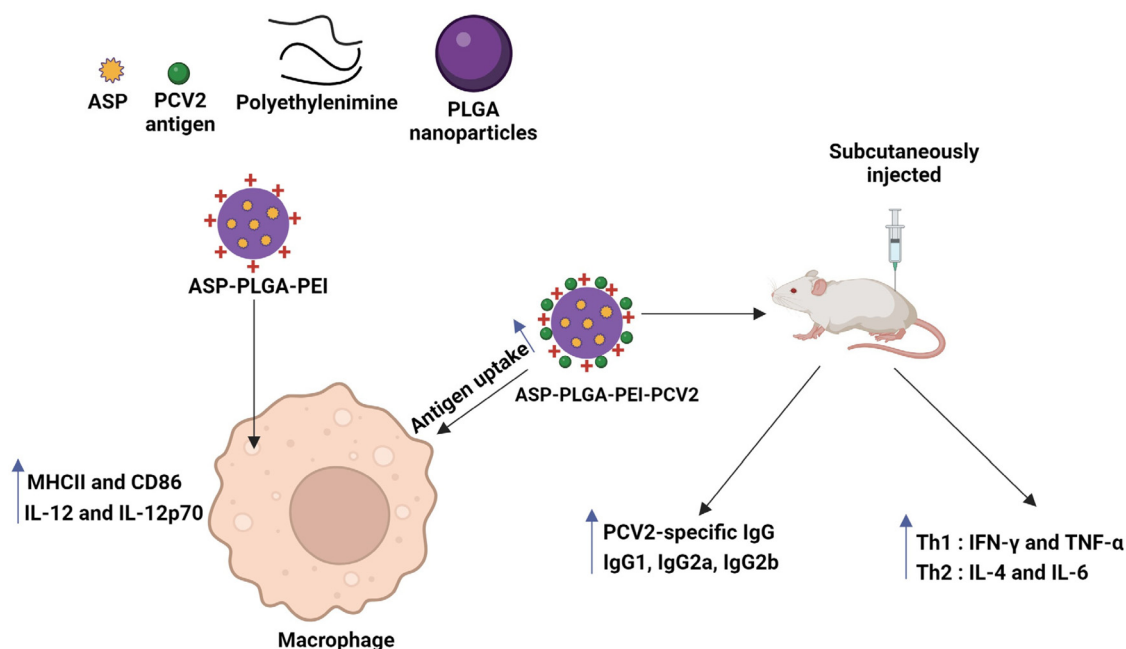


Fig. 5 ASP-PLGA-PEI nanoparticles inducing macrophage activation and antigen uptake by macrophage and also the mice immunized with PCV2 antigen adsorbed ASP-PLGA-PEI showed significant enhancement of PCV2-specific IgG immune response.¹⁰⁴



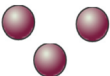






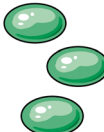


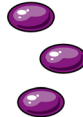



production in BMDCs *via* NLRP3- and ASC-dependent mechanisms.¹²¹

Despite benefits, health risks are associated with health when using adjuvants. The most potential side effect of adjuvants is the autoimmune/inflammatory syndrome induced by adjuvants (ASIA) first described by Shoenfeld *et al.* in the year 2011.¹²² This syndrome comprises various immune-mediated illnesses that are more prone to arise in genetically predisposed individuals following exposure to adjuvants. The key characteristic of this illness is the development of autoantibodies.¹²³ Aluminum adjuvants have been linked to a disease known as macrophagic myofasciitis (MMF) in people who have received hepatitis A and B and tetanus toxoid vaccinations. MMF patients experience arthromyalgias, persistent tiredness, muscular weakness, and even multiple sclerosis.¹²⁴ There are various types of aluminum adjuvants that differ in shape and size and might have a toxicologic effect on the body, and evaluation of the toxicity is critical. Badran *et al.* conducted a study to check the safety (toxicological evaluation) of highly used aluminum adjuvants such as Al oxyhydroxide (AlOOH) and Al hydroxyphosphate (AlOHPO₄) having different physicochemical properties.¹²⁵ The toxicology results confirmed that the toxicity of both these Al-based adjuvants depends on the size, charge and shape.¹²⁵ Though gold nanoparticles exhibited high cell uptake and caused immune stimulation *via* proinflammatory cytokine generation, their long-term cytotoxicity and *in vivo* biodistribution, excretion, and possible *in vivo* biodegradation are not completely understood.^{126–128} Adjuvants in veterinary vaccines show systemic, nonspecific side effects, including fever, arthritis, uveitis, anorexia, discomfort, and tiredness.¹²⁹ In human studies, alum adjuvants showed side effects, such as local site response and flu-like symptoms, while CpG/DNA adjuvants headache is the commonly reported side effect. Systemic reactogenicity is also a concern for commercial adjuvants such as oil emulsions and saponins that promote local tissue damage. The various systemic reactogenicity include headache, fever, malaise, diarrhea, arthralgia, myalgia, and lethargy.¹³⁰ From these multiple studies, it is clear that adjuvants have some side effects, so they must be tested preclinically to check their immunological and toxicological safety.

Future perspective

Adjuvants in vaccines play a huge role in providing immunity. However, due to various restrictions and limitations, a small number of adjuvants compatible with humans pass the test (clinical trials) and get approved. Generally, as seen from numerous studies in the scientific community, the use of

Nanomaterial adjuvants	Shape	Model used	Comments/summary	Ref.
Nanoalum (190 nm)	—	A titer of inactivated rabies virus of aG strain (108 TCID ₅₀ per ml). It is ultrafiltrated and inactivated <i>Ovalbumin</i> Bacillus anthracis Mouse BMDCs.	Nanoalum showed better effectiveness over other adjuvants due to complex formation with antigen.	95
Aluminum hydroxide (112 nm)	Rod shape 	THP-1 & BMDC	The aluminum hydroxide NPs adjuvant activity is stronger than that of the conventional aluminum hydroxide microparticles	16
γ -Aluminum oxyhydroxide (~20 nm and lengths of 150–200 nm)	Rod Shape 	Model antigen (lysozyme)	AlOOH nanorods are the most redox-active materials with low crystallinity and the highest hydroxyl content, which activate the NLRP3 inflammasome and cause BMDCs and THP-1 cells to produce IL-1 β .	96
Aluminum phosphate NPs (stabilized) {200–300 nm}	—	Hepatitis B surface antigen (HBsAg)	Aluminum phosphate was stabilized by threonine, asparagine, and L-alanyl-L-1-aminoethylphosphonic acid	98
Chitosan aluminum (CH-Al) nanomaterials {280 nm}	Round shaped 	West Nile virus (WNV) envelope protein	CH-NP formulations modulated dendritic cell cytokine production. CH-Al NPs activated the NLRP3 inflammasome, increasing IL-1 β release, suppressing type 1 T helper (Th1) and Th17 cell-polarizing cytokines, IL-12p70 or IL-23. They did not enhance pro-inflammatory cytokine production but increased dendritic cell maturation.	99
Gold NPs (AuNP) {20 and 40 nm spheres in diameter, 40 \times 10 nm rods 40 \times 40 \times 40 nm cube}	  	Differentiated THP-1 cells (wildtype, ASC, or NLRP3-deficient)	The 40 nm spherical West Nile virus E-coated AuNP (Sphere40-Es) produced the most significant level of WNV-specific antibodies.	97
Gold nanorods (40 \times 16) nm and (60 \times 14) nm	Rod-shaped  	AgNPs increased antigen-specific IgG production in a nanoparticle concentration-dependent fashion.	Nanorods activated NLRP3 inflammasome.	100
Silver nanoparticles (141 nm) {spherical}	Spherical shape 	Ovalbumin and bovine serum albumin.	AgNPs increased antigen-specific IgG production in a nanoparticle concentration-dependent fashion.	101
Poly(lactic polyglycolic acid) copolymer (PLGA) (100–150 nm)	Spherical shape 	Staphylococcal Enterotoxin B (SEB) toxoid	This study suggested the use of biodegradable nanosphere as a vaccine adjuvant.	107

Nanomaterial adjuvants	Shape	Model used	Comments/summary	Ref.
<i>Angelic acid</i> polysaccharide-Polylactic polyglycolic acid-Polyethyleneimine (ASP-PLGA-PEI) {201.7–278.6 nm}		Fluorescein isothiocyanate-labelled <i>ovalbumin</i> (FITC-OVA)	The ASP-PLGA-PEI nanoparticles notably stimulated macrophages. This nanoparticle promotes the expression of MHCII, and CD86, producing IL-1 β and IL-12p70	104
Poly (gamma-glutamic acid) nanoparticles {200 nm}	—	Ovalbumin	Poly (g-glutamic Acid) nanoparticles have a good potential for AIDS Vaccine	109
Multi-walled carbon nanotubes {200–400 nm}		Ovalbumin	MWCNTs improve the immune response and have great qualities for use as an adjuvant	105
Double-walled carbon nanotubes (DWCNTs) {0.5–2.5 nm, 1.2–3.2 nm}		LPS-primed human monocytes	In LPS-primed human monocytes, DWCNTs promote IL-1 β and IL-18 production but not TNF α or IL-6	111
Mesoporous Silica SBA-15 {2 μ m long, 10 to 12 nm internal diameter}		Bovine serum albumin as a model antigen	SBA-15 induced co-production of IgG2a and IgG1 isotypes in mice model, which is higher than aluminum hydroxide salts in comparison	112

extensively discussed. Aluminum hydroxide NPs conjugated with model proteins induced higher antigen-specific antibody response than micro aluminum hydroxide adjuvants. It is also concluded that the chitosan-aluminum nanoparticles are more efficient vaccine adjuvants than the chitosan nanoparticles. Gold nanoparticles play a significant role in immune response and inflammatory cytokine production, and studies have proved that the shape and size of these particles determine the efficiency of their response. Alum is considered one of the best vaccine adjuvants and both *in vitro* and *in vivo* studies have shown that the polymeric PLGA nanospheres have similar efficiency in immune response. In like ways, carbon-based nanomaterials have also been attributed to the activation of cytokine secretion and increased cellular uptake. Therefore, this review gives a wholesome summary of different nanomaterials and their efficiency as vaccine adjuvants.

There are no conflicts to declare.

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