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Design, synthesis and *in silico* evaluation of newer 1,4-dihydropyridine based amlodipine bioisosteres as promising antihypertensive agents†

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Hypertension remains a major global health concern, prompting ongoing research into innovative therapeutic approaches. This research encompasses the strategic design, synthesis, and computational assessment of a novel series of 1,4-dihydropyridine based scaffolds with the objective of developing promising antihypertensive agents as viable alternatives to the well-established dihydropyridine based drugs such as amlodipine, felodipine, nicardipine, etc. The crystal structure of the lead compound determined using X-ray crystallography offers crucial insights into its 3D-conformation and intermolecular interactions. *In silico* molecular docking experiments conducted against the calcium channel responsible for blood pressure regulation revealed superior docking scores for all the bioisosteres P1–P14 than the standard amlodipine, indicating their potential for enhanced therapeutic efficacy. Extensive ADMET profiling and structure–activity relationship (SAR) elucidated favourable pharmacokinetic properties and essential structural modifications influencing antihypertensive effectiveness. Specifically, P6–P10, P12 and P14 hybrids were found in accordance with Lipinski rules and exhibited druglikeness attributes, involving high GI absorption and no BBB permeance. In particular, P7 was found to be crystalline in nature having the highest binding affinity with the concerned calcium channels with excellent ADMET profile. The findings highlight the significance of the presence of triazole tethered aryl/heteroaryl ring in the synthesized hybrids, providing a foundation for further preclinical and clinical translation as antihypertensive medications.

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1. Introduction

Hypertension, characterized by elevated blood pressure levels, is an enduring and pervasive global health concern, impacting a substantial proportion of the world's population.¹ It represents a key risk factor for cardiovascular diseases, strokes, and renal complications, contributing significantly to global morbidity and mortality rates.^{2,3} Consequently, the management of hypertension constitutes a paramount concern in contemporary medical treatment, necessitating the ongoing quest for innovative and effective antihypertensive drugs.

Among the arsenal of medications employed to manage hypertension, calcium channel blockers (CCBs) have proven to be effective and are widely prescribed by healthcare professionals.⁴ Amlodipine, a 1,4-dihydropyridine based calcium

channel blocker, has established itself as a frontline antihypertensive agent by efficiently lowering the blood pressure levels by exerting vasodilatory effects on smooth muscle cells.^{5,6} Nevertheless, even though amlodipine demonstrates impressive antihypertensive capabilities, it is not devoid of certain limitations, including side effects and varied therapeutic responses among patients.⁷ Therefore, there is an unmet demand for innovative novel analogs of amlodipine that can surmount these challenges enhancing its overall pharmacological characteristics.

In the contemporary era, the concept of bioisosterism has gained prominence in the field of pharmaceutical industry as a strategic approach to drug design. Bioisosteres are the structurally different compounds that mimic the essential pharmacophoric characteristics of a reference compound to display similar biological actions.⁸ This concept has been extremely valuable in facilitating the development of safer and more effective pharmaceutical agents.⁹ Exploration of bioisosteres in the context of well-established antihypertensive drug like amlodipine offers an intriguing opportunity to fine-tune its structural and pharmacological characteristics.

1,4-Dihydropyridine based drugs such as amlodipine are the Ca²⁺ antagonists¹⁰ that are widely used in the treatment of cardiovascular disorders. These drugs allosterically interact

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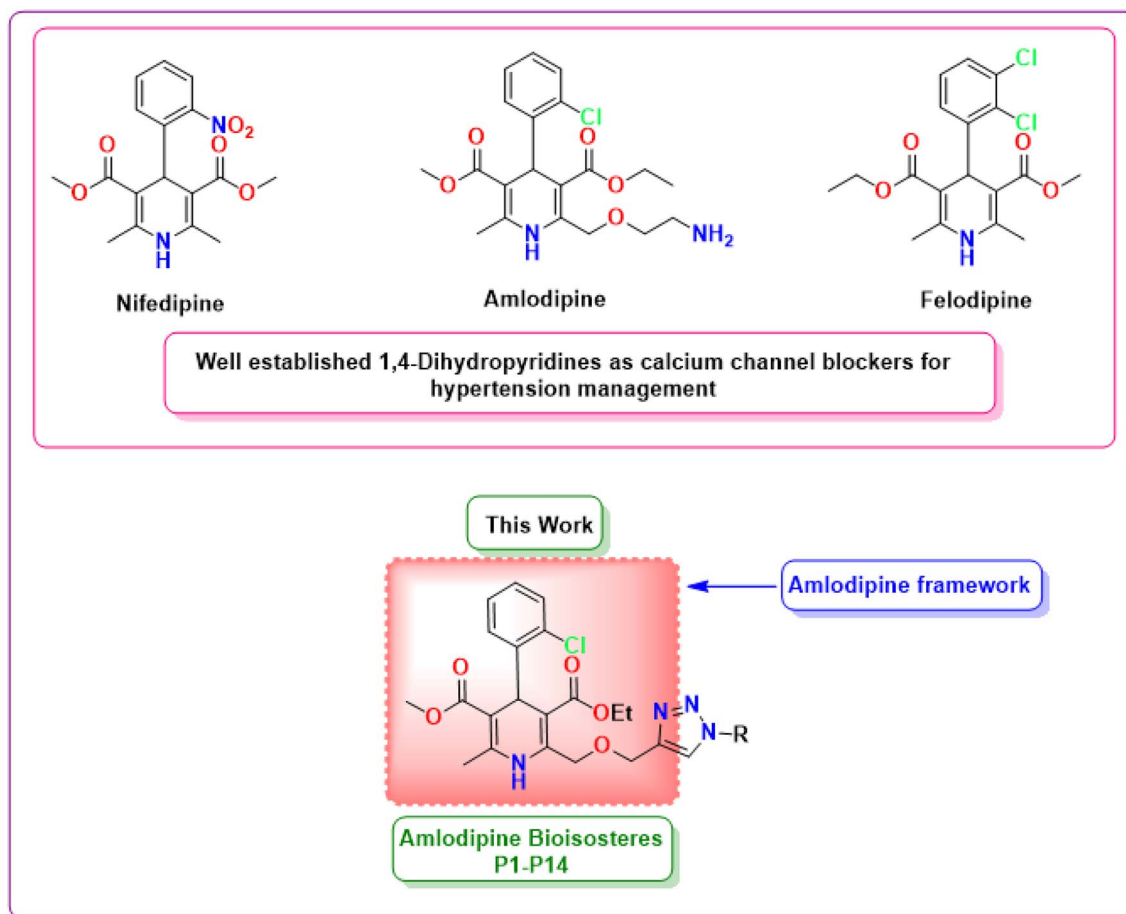


Fig. 1 Chemical structures of well-established 1,4-dihydropyridines as antihypertensive drugs and the framework of amlodipine bioisosteres synthesized in this work.

with the receptor sites on CaV1.2 channels, the most predominant voltage gated Ca^{2+} (CaV) channel type in cardiac and vascular smooth muscle myocytes. Specifically, 1,4-dihydropyridines are primarily used to treat hypertension and are considered to act as allosteric modulators that influence L-type voltage-dependent Ca^{2+} channels activation.¹¹ Binding of 1,4-dihydropyridines to receptors in L-type voltage dependent Ca^{2+} channel inhibits the entry of Ca^{2+} ions through voltage-gated Ca^{2+} channels into both the cardiac and vascular smooth muscles.¹² The target proteins CavAb complex and calmodulin were procured from the X-ray crystallographic data as reported by Tang *et al.*¹³ and Johnson C. N. *et al.*,¹⁴ respectively to successfully establish the *in silico* efficiency of our candidates as promising anti-hypertensive agents.

The present study undertakes a comprehensive exploration of newer 1,4-dihydropyridine based amlodipine bioisosteres incorporating molecular design, synthetic methodologies, and advanced computational analysis (Fig. 1). In particular, the *in silico* evaluation, the fundamental aspect of this study plays a pivotal role in analyzing the critical features of the synthesized hybrids, including their binding interactions with target proteins, pharmacokinetic characteristics and potential therapeutic efficacy.¹⁵ This approach not only accelerates the drug

discovery process but also reduces the risks and resource expenditures involved with traditional trial-and-error methods. This study holds the potential to significantly advance the field of cardiovascular medicine by introducing a newer category of antihypertensive drugs that could redefine the standard therapy for hypertension, thereby advancing the frontiers of medicinal chemistry.

2. Methodology

2.1 Materials and methods

All the reagents and solvents (analytical grade) were procured from the commercial providers and employed directly for the synthesis without any further purifications. Thin layer chromatographic (TLC) plates precoated with silica gel were used to monitor the reaction optimizations. Melting points were determined using an uncorrected Buchi instrument (M-560). KBr disks were utilized to record infrared spectra on a SHIMADZU IR Affinity 1S spectrophotometer. The JEOL, ECX-400P Spectrometer USA was used to record ^1H -NMR and ^{13}C NMR spectra in deuterated $\text{DMSO}-d_6$ at 400 MHz and 100 MHz, respectively with TMS (tetramethylsilane) as internal reference standard. The mass spectrometry measurements of all the



compounds were acquired using a 6530 Accurate-Mass Q-TOF LC/MS spectrometer.

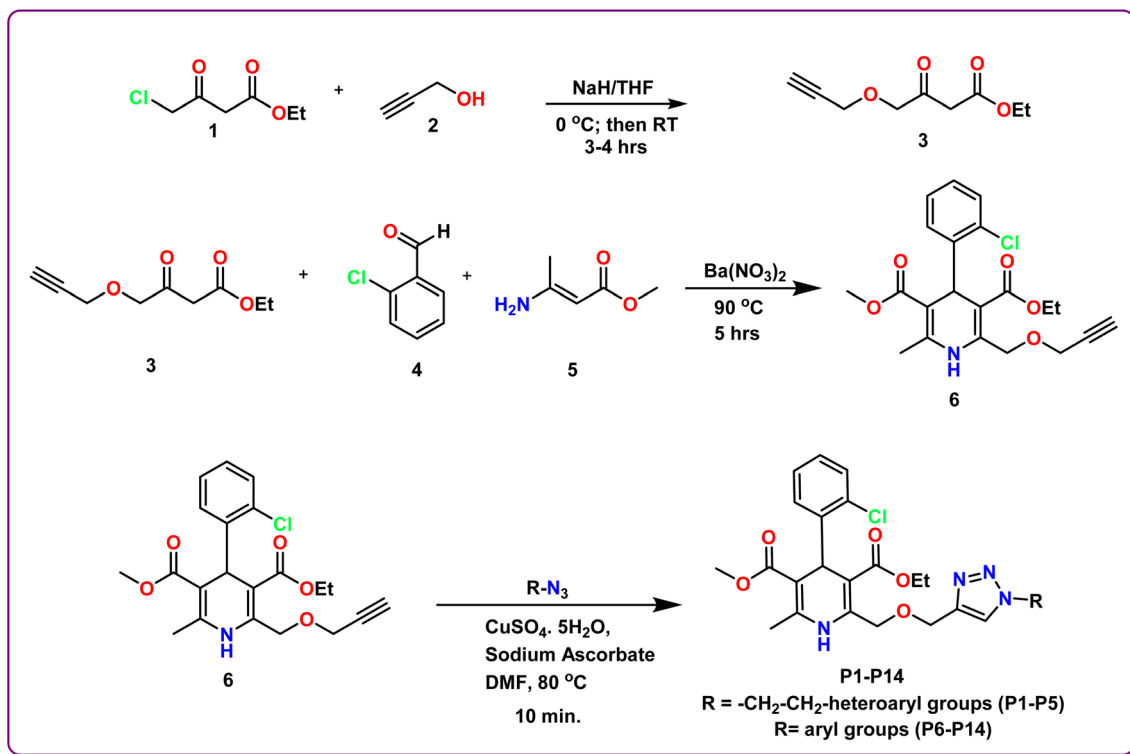
2.1.1 General procedure for the synthesis of ethyl 3-oxo-4-(prop-2-yn-1-yloxy)butanoate (3). Sodium hydride (2 mmol) was firstly added to THF at ice cold condition and was stirred vigorously for 15 min. After this, ethyl 4-chloroacetoacetate (1; 1 mmol) and propargyl alcohol (2; 1 mmol) were added to it dropwise. The reaction mixture was then stirred for around 3–4 h at room temperature and the completion of reaction was monitored *via* thin layer chromatography. After the completion of reaction, distilled water was added to the reaction mixture and was extracted using ethyl acetate. Anhydrous Na₂SO₄ was further added to remove any water content and the separated ethyl acetate layer was then evaporated under reduced pressure. The obtained product was purified *via* column chromatography to obtain pure dark yellow colored liquid 3 in 72% yield.

2.1.2 General procedure for the synthesis of 3-ethyl 5-methyl 4-(2-chlorophenyl)-6-methyl-2-((prop-2-yn-1-yloxy)methyl)-1,4-dihydropyridine-3,5-dicarboxylate (6). In a round bottom flask, a mixture of ethyl 3-oxo-4-(prop-2-yn-1-yloxy)butanoate (3; 1 mmol), 2-chlorobenzaldehyde (4; 1 mmol), methyl 3-aminocrotonate (5; 1 mmol), and barium nitrate (0.1 mmol) was heated at 90 °C for around 5 hours (Scheme 1). The reaction's progress was monitored by TLC at various time intervals. After the reaction was completed, the mixture was allowed to settle at room temperature before being extracted with ethyl acetate and any water content was removed using anhydrous Na₂SO₄. The solution was evaporated at low pressure, and the crude product was purified using column chromatography to procure 6 as a yellow solid in 65% yield.

2.1.3 General procedure for the synthesis of 1,4-dihydropyridine based amlodipine bioisosteres (P1–P14). 3-Ethyl 5-methyl 4-(2-chlorophenyl)-6-methyl-2-((prop-2-yn-1-yloxy)methyl)-1,4-dihydropyridine-3,5-dicarboxylate (6; 1 mmol) and different aryl and heteroaryl azides (1 mmol) were dissolved in DMF (10 ml) in a round bottom flask. Following this, a solution of sodium ascorbate (0.4 mmol) and copper sulphate (0.2 mmol) in H₂O was introduced to the reaction mixture and was refluxed for 10 min (Scheme 1). TLC was employed to track the progress of the reaction. Following the completion of the reaction, water was added to the reaction mixture and the product was extracted using ethyl acetate. Subsequently, anhydrous Na₂SO₄ was used to eliminate any water content in the organic layer and the solvent was evaporated under reduced pressure. Crude products obtained were then purified using column chromatography to obtain pure hybrids P1–P14 in good yields (Fig. 2).

2.2 Single crystal X-ray structure

Fine crystals of ligand P7 were grown by slow evaporation of saturated mother solvent methanol & chloroform which was layered by hexane and diffused by ether at room temperature for SC-XRD studies. Good quality rectangular shaped yellowish crystals were taken and exposed to X-rays on a Bruker diffractometer employing a graphite monochromatized Mo/K α radiation ($\lambda = 0.71073$ Å) at temperature 107 K. The crystal data was reduced using CrysAlis pro software available with the diffractometer. The structure was solved by direct methods using SHELXL-2016/4 and refined by the full-matrix least-squares method on Olex2.refine 1.5.¹⁶ All calculations were carried out using the OLEX2 package of



Scheme 1 Schematic route for the synthesis of 1,4-dihydropyridine based amlodipine bioisosteres P1–P14.

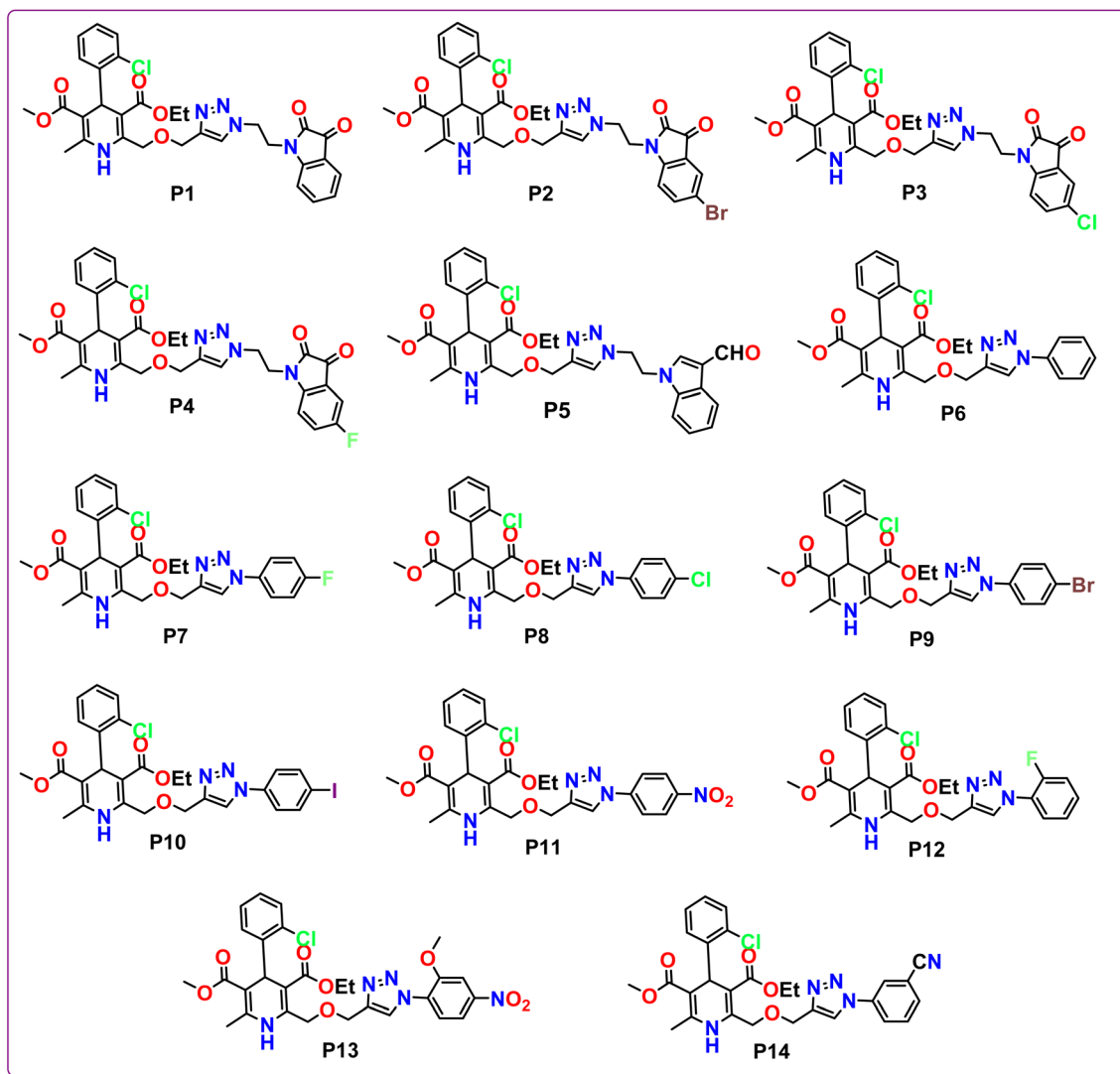


Fig. 2 Chemical structures of the synthesized 1,4-dihydropyridine based amlodipine bioisosteres **P1–P14**.

the crystallographic programs.¹⁷ For the molecular graphics, the program Mercury (2022.3.0) was used.¹⁸

2.3 Molecular docking studies

Molecular docking examinations were executed using AutoDock Vina to evaluate the binding potential and molecular interactions of the synthesized hybrids **P1–P14** with the proteins (PDB ID: 5KMD and 6M7H).¹⁹ The 3-D structures of 5KMD and 6M7H were procured from the protein data bank. The protein preparation process comprised of multiple steps, including the removal of co-factors, water molecules, and co-crystallized ligands from the protein structure. Subsequently, polar hydrogen atoms and kollmann charges were introduced to the protein prepared using AutoDock Tools 1.5.6. Following this, docking was executed separately for each ligand with different proteins by setting the grid size according to the active binding sites of the receptors. Best docked pose with lowest binding affinity value was recorded for each compound. The protein–ligand interactions were further visualized using BIOVIA Discovery Studio 2021.

2.4 *In silico* ADMET analysis

Effective drug discovery requires superior pharmacokinetics and minimal toxicity, making them more drug-like. *In silico* techniques for screening potential candidates for Absorption, Distribution, Metabolism, Excretion and Toxicity (ADMET) play a crucial role in mitigating the risk of late-stage mortality and refining the selection of most promising candidates. Consequently, in this study, all the synthesized hybrids **P1–P14** underwent screening for their druglikeness, oral bioavailability and pharmacokinetic characteristics using the SwissADME^{20,21} and preADMET tools.

3. Result and discussion

3.1 Chemistry

The methodology for the development of novel 1,4-dihydropyridine based amlodipine bioisosteres **P1–P14** was established in three steps. The precursor ethyl 3-oxo-4-(prop-2-yn-1-yloxy)butanoate **3** was prepared according to the reported



method in literature using ethyl 4-chloroacetoacetate and propargyl bromide in the presence of NaH in dry THF solvent. Further, ethyl 3-oxo-4-(prop-2-yn-1-yloxy)butanoate **3**, 2-chlorobenzaldehyde **4** and methyl 3-aminocrotonate **5** were employed to obtain novel 3-ethyl 5-methyl 4-(2-chlorophenyl)-6-methyl-2-((prop-2-yn-1-yloxy)methyl)-1,4-dihydropyridine-3,5-dicarboxylate **6** by modified Hantzsch synthesis as reported procedure in literature. These synthesized 1,4-dihydropyridine scaffolds were then tethered with various aryl and heteroaryl azides *via* Cu-catalyzed 1,3-dipolar cycloaddition click reaction under different reaction conditions to procure the desired amlodipine bioisosteres **P1–P14** in excellent yields. The structures of the synthesized scaffolds were substantiated by various spectroscopic techniques, including Nuclear Magnetic Resonance (^1H NMR and ^{13}C NMR), Fourier-transform infrared (FT-IR), High Resolution Mass Spectrometry (HRMS), and Single Crystal XRD.

Evaluation of ^1H NMR of **P1** revealed the characteristic peaks at δ 8.57 and δ 8.23 as singlets for one proton each signifying the

existence of $-\text{NH}$ of 1,4-dihydropyridine ring and $-\text{CH}$ of the triazole ring, respectively. A prominent singlet peak of 1 proton at δ 5.29 ppm corresponds to the $-\text{CH}$ of the chiral centre of 1,4-dihydropyridine at 4-position. The ^{13}C NMR spectrum of this compound depicted the presence of characteristic peak for both the carbonyl groups of the isatin ring at δ 183.37 ppm and δ 158.59 ppm. Further, the presence of two ester groups at 1,4-dihydropyridine ring is confirmed by the prominent characteristic peaks at δ 167.60 and δ 166.72 ppm. Additionally, the HRMS data of **P1** demonstrated the calculated mass ion $(\text{M} + \text{H})^+$ as 620.1834, satisfying the observed mass of 620.1922.

The crystal structure of ligand **P7** was determined by Single Crystal X-ray diffraction method which revealed that the molecular structure of compound consists of a fluorophenyl substituted triazole ring. It further revealed the presence of vertical appended chlorobenzene on the top of the dihydropyridine ring at 124.9° angle. Ethyl ester and methyl ester groups are attached at 3rd and 5th position of 1,4-dihydropyridine ring at angles 117.8° and 118.91° , respectively, confirming

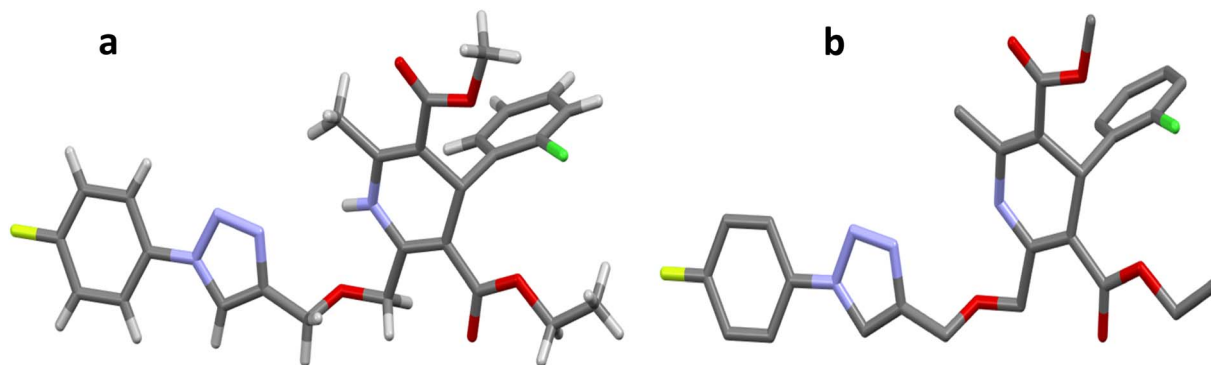


Fig. 3 Molecular structures (a and b) of compound **P7**. H atoms are omitted in (b) for clarity. C = gray, N = blue, F = yellow, Cl = green, O = red, H = gray.

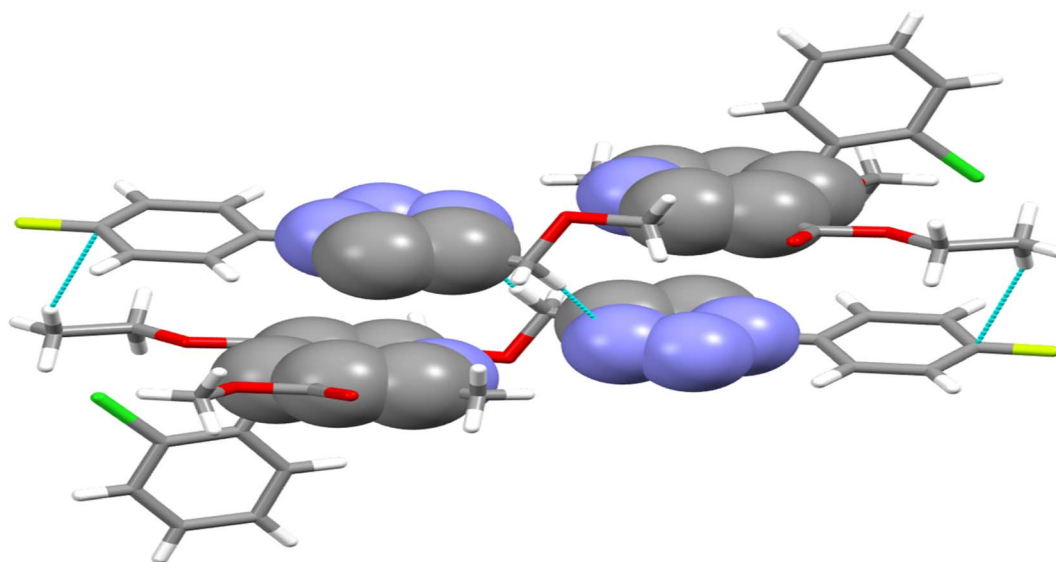


Fig. 4 Double-decker π -stacked arrangement of triazole and dihydropyridine ring and hydrogen bonding between the adjacent **P7** molecules.

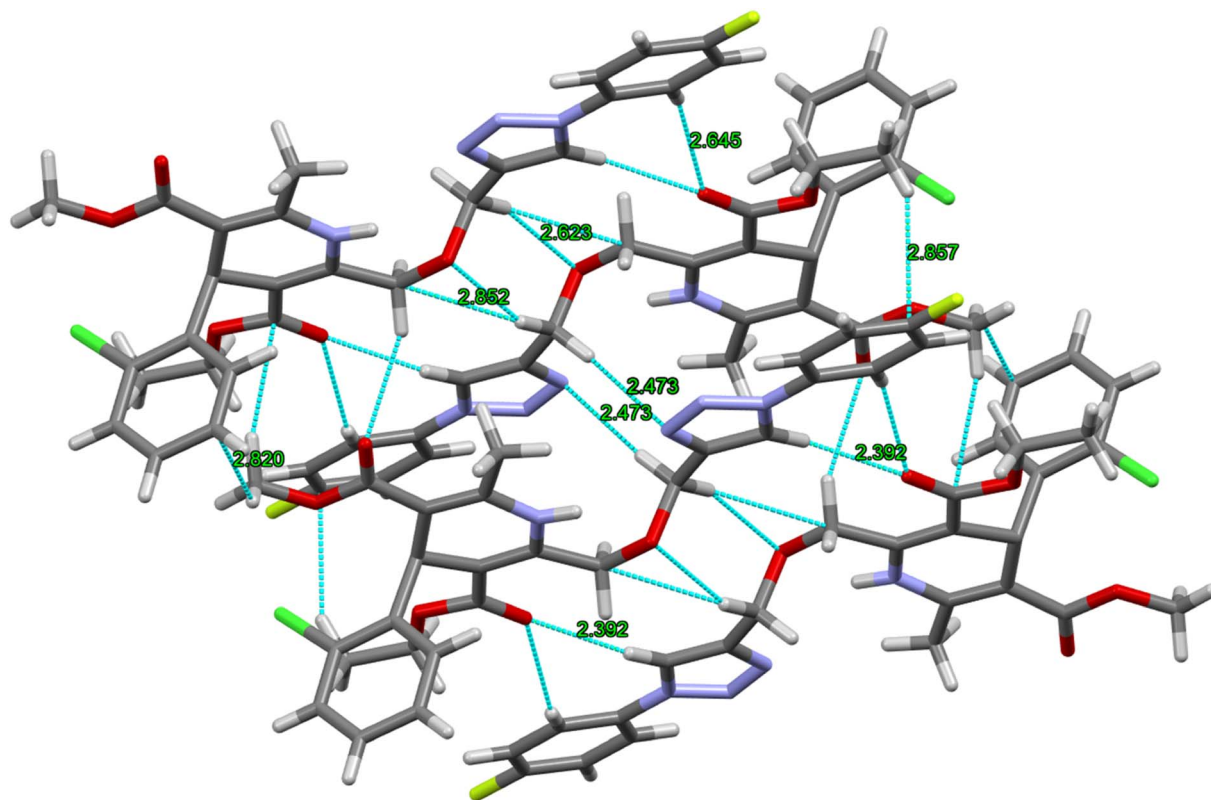


Fig. 5 A network of P7 *via* intra- and intermolecular H-bonding between the adjacent molecules.

the streamlined shape of the compound which is clearly shown in Fig. 3. The chair form geometry when the two adjacent molecules encapsulate the hexane solvent *via* $-\text{CH}-\text{Pi}$ interactions in the compound from chlorobenzene to the fluoro-benzene ring *via* triazole ring is confirmed by the bond angles analysis as depicted in Fig. 4. The bond distances of adjacent atoms from chlorine to fluorine lie in 1.355(2)–1.749(2) Å whereas inter and intra molecular hydrogen bond lengths lie close to (2.471–2.853 Å). These weaker coordinating interactions

form the core of crystal engineering and are therefore essential. Interestingly, crystal structure of P7 shown in the present study display a variety of non-covalent interactions such as a network of inter-molecular H-bonding and $\pi-\pi$ stacking arrangements in the crystal structures (Fig. 5).

3.2 Molecular docking evaluation

Exploration of interactions between proteins and ligands is critical in the process of structure-based drug design.

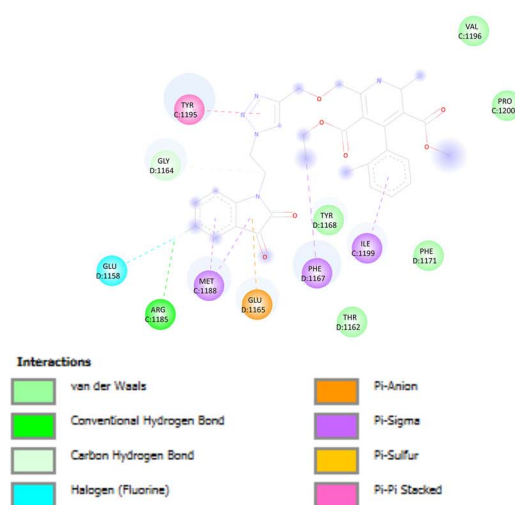
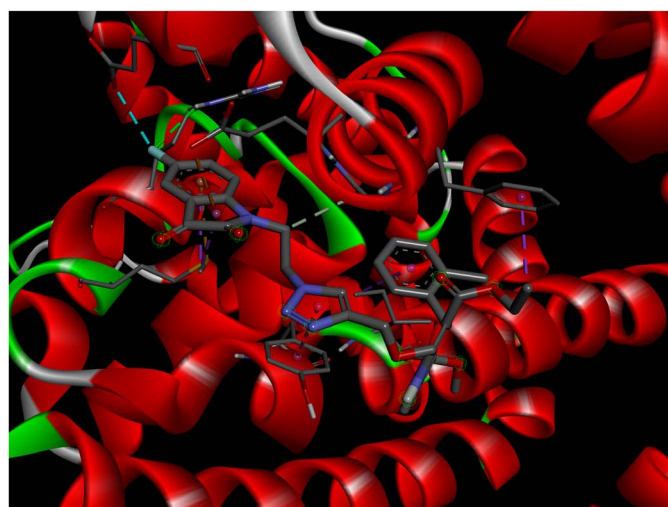


Fig. 6 3D and 2D representation of binding interactions of P4 with 5KMD.



Molecular docking studies were conducted to elucidate the key pharmacophores responsible for calcium channel blocker activity or anti-hypertensive activity. This investigation examined the interactions between a set of molecule ligands (amlodipine bioisosteres) and their respective protein targets 5KMD and 6M7H. The molecular docking analysis of all the synthesized hybrids **P1–P14** revealed excellent docking scores and remarkable interactions with the essential amino acid residues located within the receptor's binding pocket of 5KMD and 6M7H, as compared to the standard amlodipine (Tables S3 and S4 in ESI† file). A stronger binding affinity with the receptor is indicated by the more negative binding affinity value in the docking results.

Evaluation of the molecular docking results with unique binding sites on calcium voltage gated (CaV) channels 5KMD demonstrated a remarkable binding affinity of -8.6 kcal \cdot mol $^{-1}$ for **P4** containing fluorinated isatin group in case of

heteroaryl rings that is considerably higher than that of the standard amlodipine (-5.6 kcal \cdot mol $^{-1}$). However, the bioisoster **P7** containing the triazole tethered fluorobenzene ring displayed the highest binding affinity of -8.8 kcal \cdot mol $^{-1}$ for 5KMD. As displayed in Fig. 6, **P4** exhibits impressive interactions with a number of amino acids. Significant Pi-Sigma interactions with MET1188, PHE1167, and ILE1199, are exhibited by the isatin ring, $-\text{CH}_3$ of $-\text{COOEt}$ group and chlorobenzene ring, respectively. Triazole ring is involved in Pi-Pi stacking with TYR1195. Further, $-\text{F}$ atom of isatin ring is involved in hydrogen bond interactions with GLU1158 and ARG1185. Pi-Anion interaction is also displayed by the indole ring of isatin with GLU1165. In case of **P7**, Pi-Sigma interactions are displayed by the chlorobenzene ring with the amino acid residue ILE1199. Amide-Pi Stacked interactions are exhibited by the triazole ring and fluorobenzene ring with TYR1195 and GLY1164. Furthermore, $-\text{CH}_3$ of $-\text{COOEt}$ group

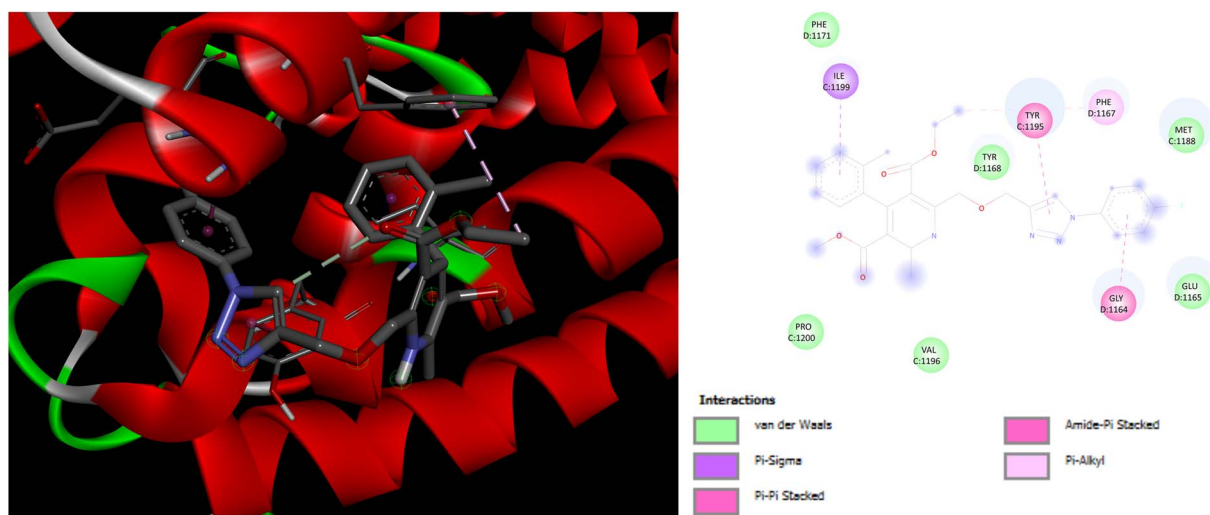


Fig. 7 3D and 2D representation of binding interactions of **P7** with 5KMD.

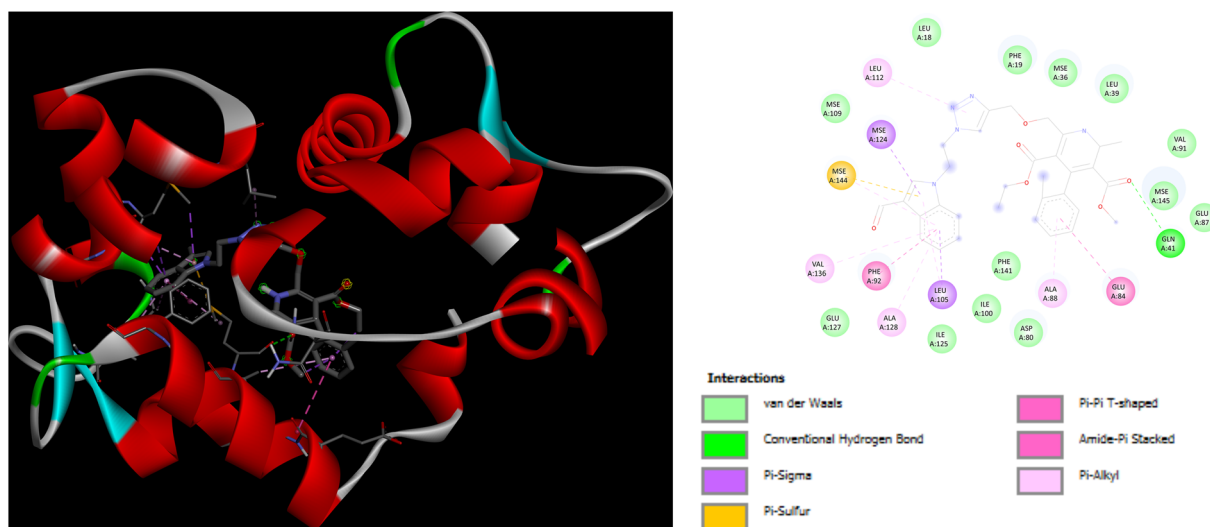


Fig. 8 3D and 2D representation of binding interactions of **P5** with 6M7H.

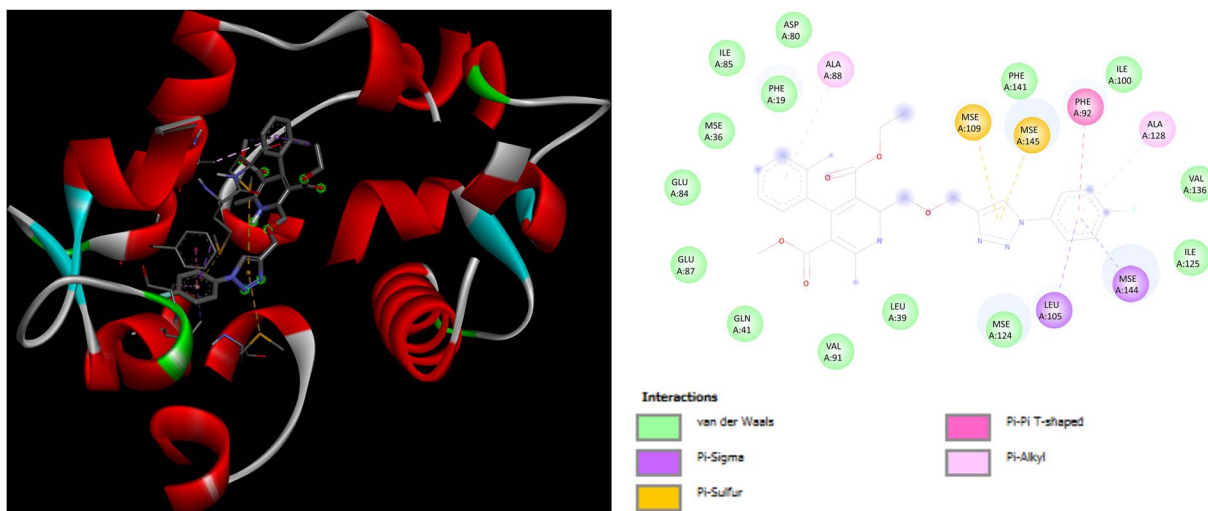


Fig. 9 3D and 2D representation of binding interactions of P7 with 6M7H.

is involved in the Pi-Alkyl interactions with the amino acid residue PHE1167. It is also involved in a number of van der Waals interactions with several amino acid residues (Fig. 7).

In addition to this, docking studies of these compounds were assessed on the calmodulin (CaM)-dependent protein kinase II (PDB ID: 6M7H). P5 and P7 demonstrated the highest binding

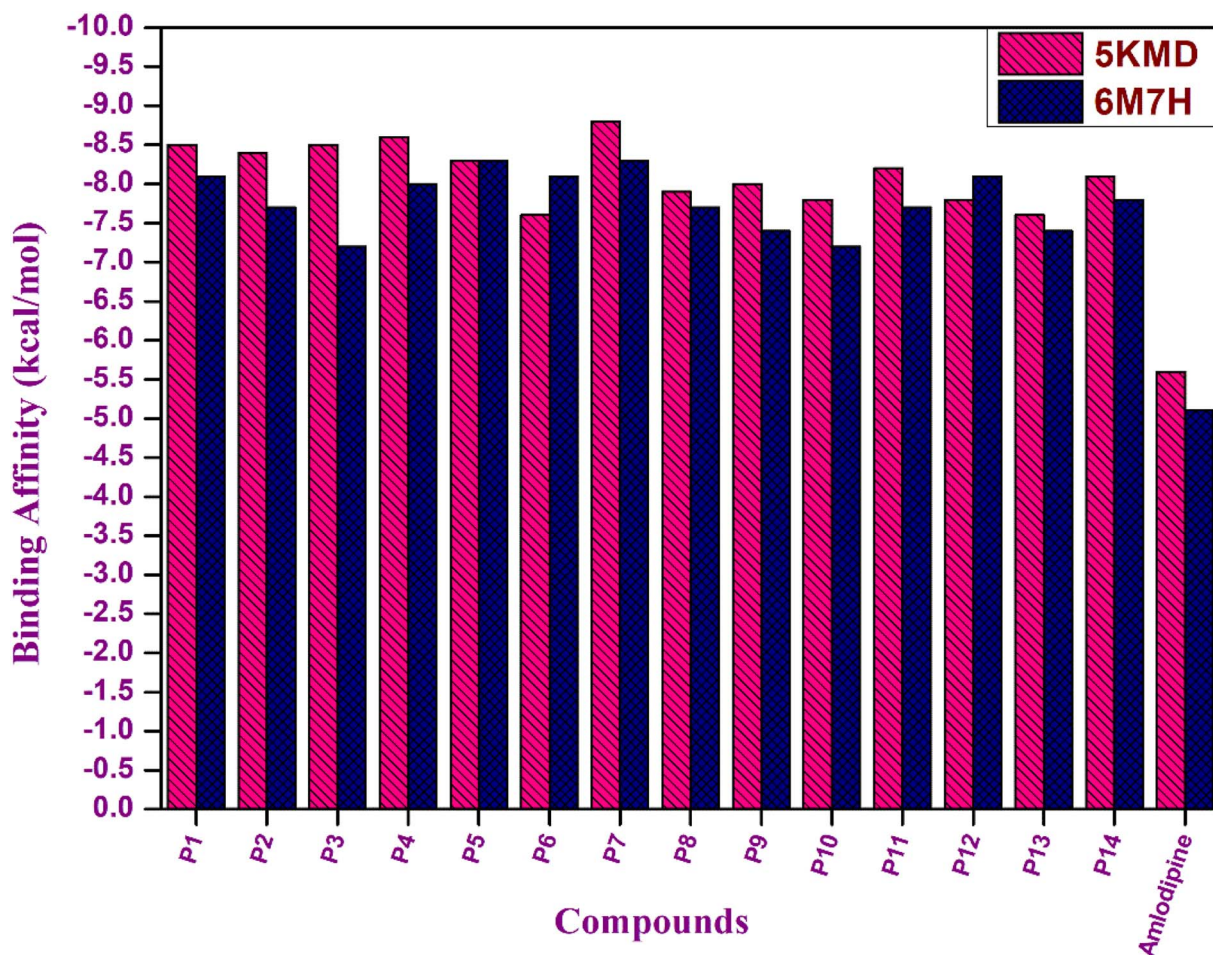


Fig. 10 Graphical representation of binding affinity of hybrids P1–P14 with target proteins 5KMD and 6M7H.



affinity of $-8.3 \text{ kcal mol}^{-1}$ as compared to amlodipine ($-5.1 \text{ kcal mol}^{-1}$). In case of docking interactions of **P5** with 6M7H, the indole ring is involved in a number of interactions, namely; Pi-Sigma, Pi-sulfur, Pi-Pi T shaped, and Pi-alkyl interactions with amino acid residues MSE124, LEU105, MSE144, PHE92, VAL136, and ALA128, respectively as depicted in Fig. 8. Further, Pi-Pi T shaped and Pi-alkyl interactions with GLU84 and ALA88, respectively are also exhibited by the chlorobenzene ring of **P5** along with the Pi-alkyl interactions of LEU112 with the triazole ring. Fig. 9 demonstrates the binding interactions of **P7** with 6M7H, showing the Pi-alkyl interactions of fluorobenzene and chlorobenzene ring with ALA128, and ALA88, respectively. Fluorobenzene ring is also associated with the Pi-Pi T-shaped interactions with PHE92, along with Pi-Sigma interactions with MSE144 and LEU105. Residues MSE109 and MSE145 are involved in Pi-sulfur interactions with the triazole ring of **P7**.

The docking investigations for all the synthesized compounds **P1–P14** demonstrated superior binding affinities

when compared to the standard amlodipine in case of both the proteins, particularly, **P7** exhibited the highest binding affinities with 5KMD and 6M7H, underscoring the significance of these bioisosteres as potential leads for effective anti-hypertensive activity (Fig. 10).

3.3 *In silico* structure activity relationship (SAR) assessment

The structure activity relationship is a fundamental concept in drug design that is used to comprehend the relationship between the chemical structure of a compound and its pharmacological activity. Its ultimate goal is to optimize the chemical structure of a compound in order to maximize its intended pharmacological activity while minimizing undesired side effects.²² As evident from the computational evaluation of the synthesized amlodipine bioisosteres **P1–P14**, hybrids containing heteroaryl rings tethered *via* triazole linkage (**P1–P5**) exhibited excellent binding affinity and remarkable interactions with the amino acid residues in case of both the proteins 5KMD and 6M7H as compared to the hybrids possessing the aryl

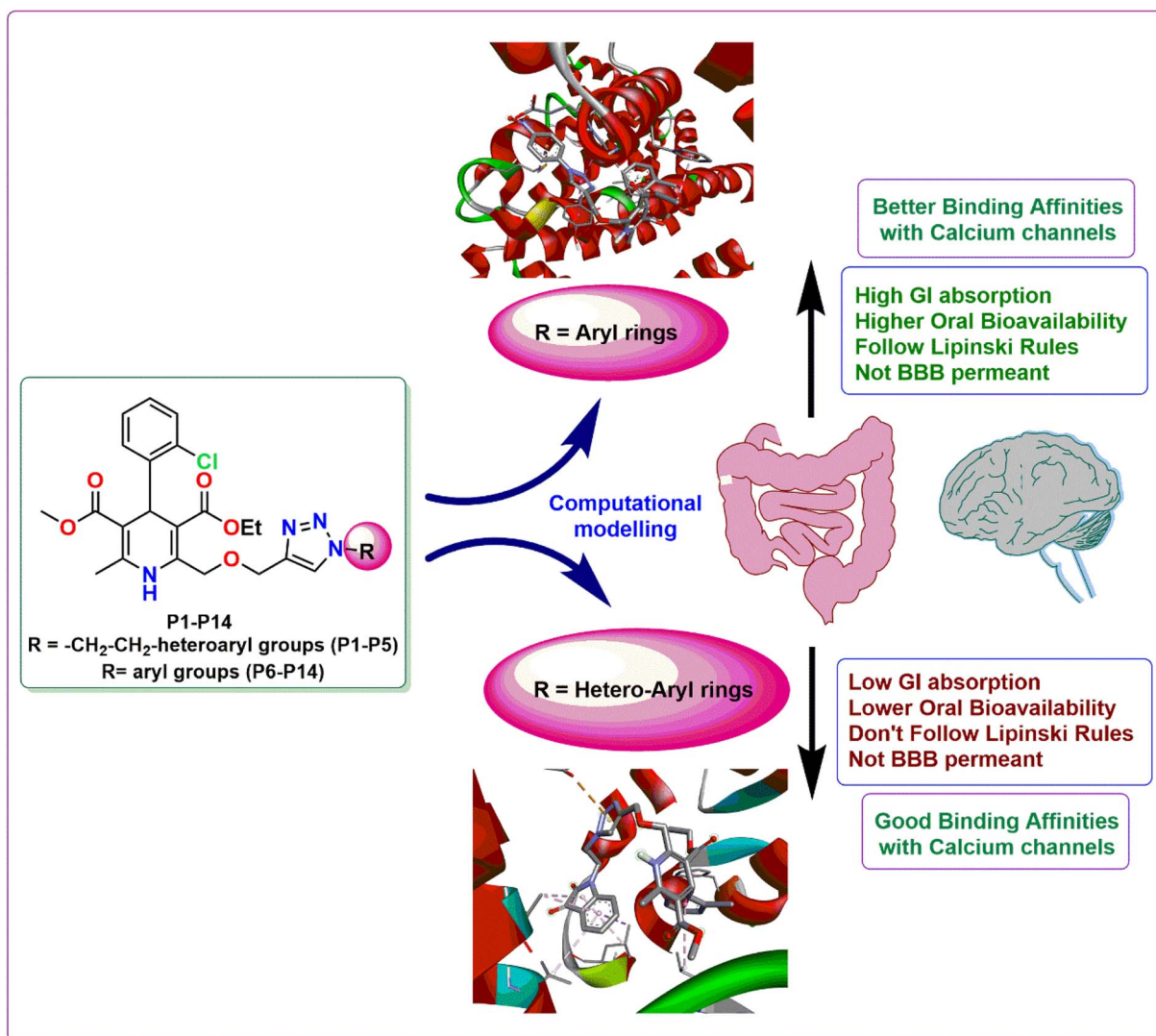


Fig. 11 Demonstration of the structure activity relationship of the synthesized 1,4-dihydropyridine based amlodipine bioisosteres **P1–P14**.

linkage (P6–P14). However, aryl linked hybrids were found to be in accordance with the Lipinski rule and have shown high GI absorption. In contrast, P1–P5 heteroaryl ring containing hybrids displayed two violations in Lipinski rule with low GI absorption (Fig. 11). Hence, it can be inferred that despite of better binding affinity of P1–P5, hybrids P6–P14 possess drug-like attributes and can serve as lead for future investigations. Better computational results than the standard drug amlodipine suggest that the existence of triazole tethered aryl/heteroaryl ring is essential for the manifestation of pharmacological attributes associated with anti-hypertensive activity.

4. Conclusion

In conclusion, this study represents a significant advancement in the quest for innovative antihypertensive medications. A promising series of 1,4-dihydropyridine based scaffolds were developed as prospective replacements to amlodipine through a meticulous approach encompassing strategic design, synthesis and extensive *in silico* evaluation. This comprehensive study involved the use of crystallography, molecular docking, ADMET profiling, and analysis of Structure Activity Relationships (SAR). The examination of crystal structure provided valuable structural insights, laying the foundation for subsequent computational evaluations. The existence of triazole appended aryl/heteroaryl linkage was substantiated by the better molecular docking results with the concerned calcium channels as compared to amlodipine. ADMET and SAR evaluation uncovered pivotal insights for the future refinement of antihypertensive activity. Furthermore, excellent binding affinity, high GI absorption, no BBB permeance, and the drug-likeness nature of the synthesized bioisosteres, especially P6–P10, P12 and P14 as established from the *in silico* ADMET assessments, suggest their suitability for further development. In particular, P7 showcased extraordinary *in silico* profiling and its crystal structure manifests the molecular interactions with biological systems. This collective effort establishes a robust foundation for the progression and potential clinical applications of these candidates, signalling a promising avenue in antihypertensive drug discovery. In essence, this research significantly contributes to the growing panorama of innovative therapeutics by offering prospective solutions for hypertension management and underscoring the importance of exploring novel bioisosteres in drug design and development.

Accession codes

CCDC 2294665 contains the supplementary crystallographic data for ligand S1 for this paper.

Conflicts of interest

The authors declare no conflicts of interests.

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References

- 1 K. T. Mills, A. Stefanescu and J. He, *Nat. Rev. Nephrol.*, 2020, **16**, 223–237.
- 2 M. Burnier and B. M. Egan, *Circ. Res.*, 2019, **124**, 1124–1140.
- 3 W. B. Kannel, *J. Cardiovasc. Pharmacol.*, 1989, **13**, S4–S10.
- 4 H. Scholz, *Cardiovasc. Drugs Ther.*, 1997, **10**, 869–872.
- 5 K. G. Bulsara and M. Cassagnol, in *StatPearls [Internet]*, StatPearls Publishing, 2023.
- 6 U. Hostalek-Gottwald and Z. Gaciong, *Curr. Med. Res. Opin.*, 2022, **38**, 1047–1053.
- 7 S. Mostafa, H. Shabana, F. Khalil, I. M. El Mancy, H. A.-M. Zedan, A. Elmoursi, I. G. Ramadan, S. E. D. Mohamed, A. Kassem and I. S. Kamel, *High Blood Pressure Cardiovasc. Prev.*, 2022, **29**, 565–576.
- 8 T. Iida, J. Kanazawa, T. Matsunaga, K. Miyamoto, K. Hirano and M. Uchiyama, *J. Am. Chem. Soc.*, 2022, **144**, 21848–21852.
- 9 B. S. Jayashree, P. S. Nikhil and S. Paul, *Med. Chem.*, 2022, **18**, 915–925.
- 10 J. Striessnig and N. J. Ortner, in *Encyclopedia of Molecular Pharmacology*, Springer, 2022, pp. 375–383.
- 11 R. Bansal, P. Jain, G. Narang, A. Kaur, C. Calle and R. Carron, *Lett. Drug Des. Discovery*, 2023, **20**, 1632–1644.
- 12 H. J. Little, *Pharmacol. Rev.*, 2021, **73**, 1298–1325.
- 13 L. Tang, T. M. Gamal El-Din, T. M. Swanson, D. C. Pryde, T. Scheuer, N. Zheng and W. A. Catterall, *Nature*, 2016, **537**, 117–121.
- 14 C. N. Johnson, R. Pattanayek, F. Potet, R. T. Rebbeck, D. J. Blackwell, R. Nikolaienko, V. Sequeira, R. Le Meur, P. B. Radwański, J. P. Davis, A. V. Zima, R. L. Cornea, S. M. Damo, S. Györke, A. L. George and B. C. Knollmann, *Cell Calcium*, 2019, **82**, 102063.
- 15 B. N. Sağlık, B. K. Çavuşoğlu, D. Osmaniye, S. Levent, U. A. Çevik, S. İlgin, Y. Özkay, Z. A. Kaplancıklı and Y. Öztürk, *Bioorg. Chem.*, 2019, **85**, 97–108.
- 16 G. M. Sheldrick, *Acta Crystallogr., Sect. C: Struct. Chem.*, 2015, **71**, 3–8.
- 17 O. V. Dolomanov, *J. Appl. Crystallogr.*, 2009, **42**, 339–341.
- 18 C. F. Macrae, P. R. Edgington, P. McCabe, E. Pidcock, G. P. Shields, R. Taylor, M. Towler and J. V. D. Streek, *J. Appl. Crystallogr.*, 2006, **39**, 453–457.
- 19 O. Trott and A. J. Olson, *J. Comput. Chem.*, 2010, **31**, 455–461.
- 20 A. Daina, O. Michielin and V. Zoete, *Sci. Rep.*, 2017, **7**, 42717.
- 21 A. Daina, O. Michielin and V. Zoete, *J. Chem. Inf. Model.*, 2014, **54**, 3284–3301.
- 22 R. Guha, *J. Comput.-Aided Mol. Des.*, 2008, **22**, 857–871.

