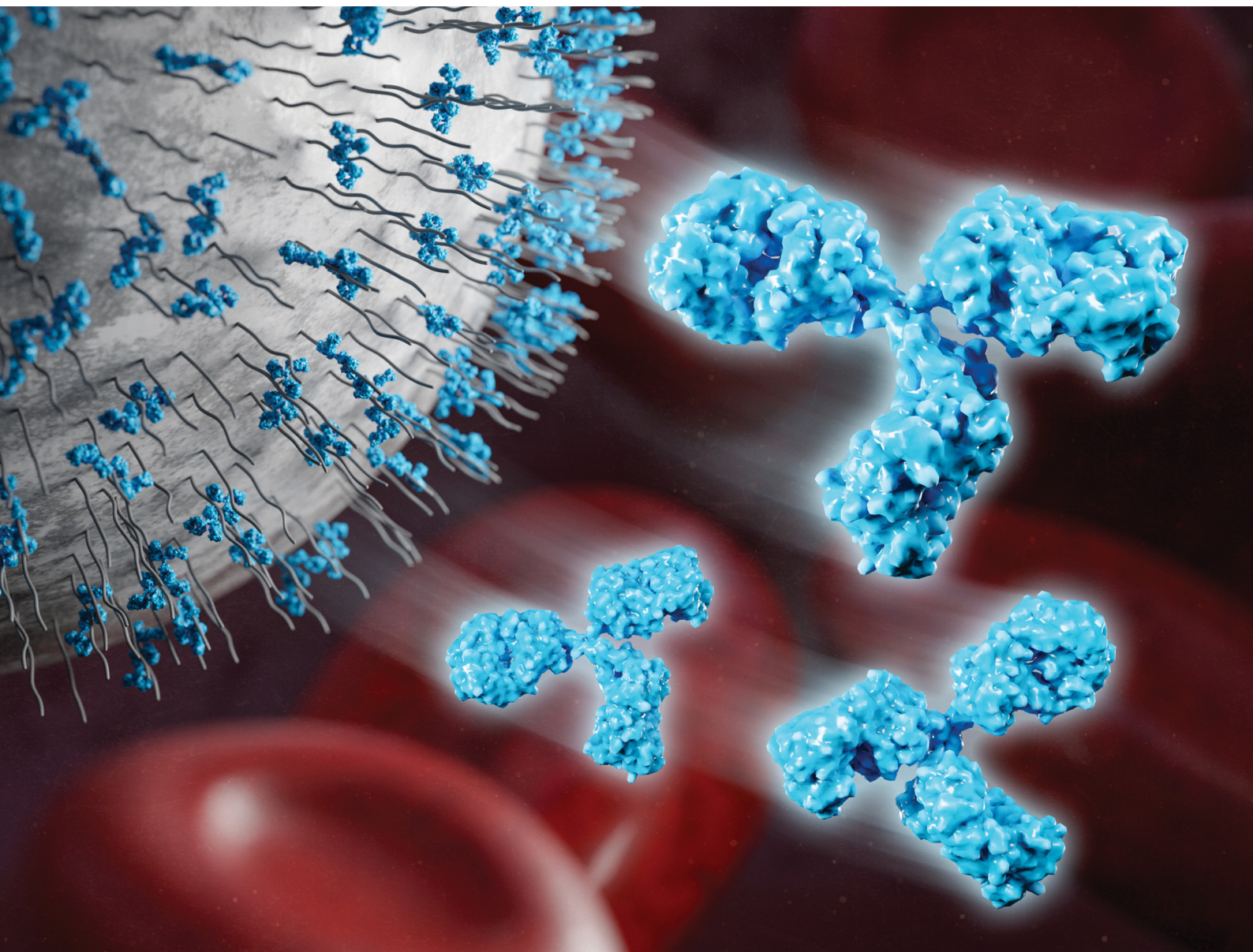


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Anti-PEG antibodies enriched in the protein corona of PEGylated nanocarriers impact the cell uptake†

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Poly(ethylene glycol) (PEG) is the gold standard used to reduce unspecific protein adsorption and prolong nanocarrier circulation time. However, this stealth effect could be counteracted by the increasing prevalence of anti-PEG antibodies in the bloodstream. Up to now, the presence of anti-PEG antibodies in the protein corona and their effect on cell uptake has not been investigated yet. Our results showed a high concentration and prevalence of anti-PEG antibodies in the German population. PEGylated nanocarriers exhibited a higher level of anti-PEG antibodies in the protein corona compared to non-PEGylated, which lead to higher uptake in macrophages. Consequently, the anti-PEG antibodies in the protein corona could mitigate the stealth effect of PEG, leading to accelerated blood clearance and unwanted side effects.

Introduction

Many materials in biomedical applications use poly(ethylene glycol) (PEG).¹ Attaching PEG chains to a biopharmaceutical agent – so-called “PEGylation” – is an important approach to improve the agent’s pharmacokinetics and pharmacodynamics.² PEGylation can increase water solubility and stability, decrease enzymatic degradation, reduce immunogenicity, and extend the blood circulation half-life.³ Additionally, PEG reduces unspecific protein adsorption on nanocarriers (NCs) and prolongs their circulation time by adsorption of specific apolipoproteins like apolipoprotein A1 or clusterin.⁴ This is known as the “stealth effect”.^{4,5} However, in some cases it was observed that PEGylation resulted in an enrichment of opsonins such as immunoglobulins, promoting

New concepts

In our study, we report for the first time on anti-PEG antibody quantification in the protein corona of nanocarriers. The prevalence of antibodies directed against PEG among the general population has been reported before, as well as the accelerated blood clearance of PEGylated nanomedicines. However, it was not yet clear whether these antibodies are indeed enriched on nanocarrier surfaces and how their presence correlates with cellular uptake. With our findings, we verify the assumption that nanocarrier-associated anti-PEG antibodies contribute to enhanced uptake in macrophage cell lines and, thus, induce effects opposite to the original intention of PEGylation.

unspecific cell uptake.⁶ This could be related to PEG-binding antibodies.⁷ These anti-PEG antibodies could bind to PEGylated NCs, leading to increased uptake in macrophages and therefore counteract the stealth effect. Thus, it is important to determine the prevalence of anti-PEG antibodies in the blood plasma, investigate the interaction of the antibodies with NCs, and monitor the influence on cell uptake.

In 1983, Richter *et al.* first reported the potential immunogenicity of PEG itself.⁸ They observed anti-PEG antibodies (mainly IgM isotype) in approximately 0.2% of healthy blood donors, which at that time was considered to be of no clinical significance and therefore probably did not interfere with the clinical use of PEGylated therapeutics.⁹ Later on, various research groups observed that administering repeated doses of PEGylated NCs led to accelerated blood clearance and weakened efficacy of PEGylated therapeutics.¹⁰ In contrast to most antidrug antibodies, anti-PEG IgG and IgM antibodies were observed in both PEGylated therapeutics-treated patients and healthy (treatment-naïve) individuals.

In 2016, Chen *et al.* reported pre-existing anti-PEG antibodies in healthy Han Chinese and found that 44.3% of participants tested positive for anti-PEG IgG and IgM antibodies.¹¹ Additionally, a recent study by Huckaby *et al.* characterized the anti-PEG antibody structure in complexes with PEG chains by X-ray crystallography.¹² They demonstrated how antibodies could bind

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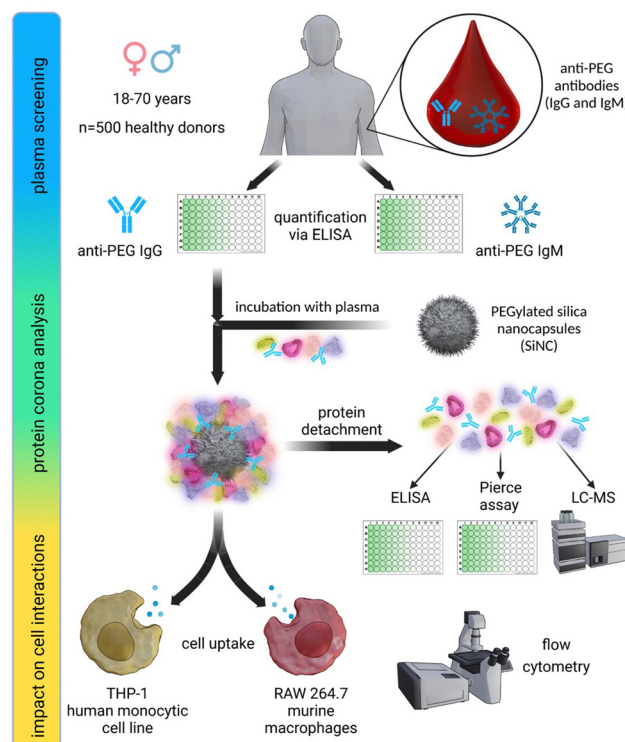
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† Electronic supplementary information (ESI) available: Experimental Section, additional plasma screening data, MST parameters and data, LC–MS data, additional cell uptake data, F_c blocking experiments. See DOI: <https://doi.org/10.1039/d3nh00198a>



Regardless of the potentially serious consequences of circulating anti-PEG antibodies, their impact on the effect of NC therapeutics and related side effects remains uncertain up to now. Based on the currently available literature, it can be assumed that observed effects originated in the association of anti-PEG antibodies with PEGylated nanocarriers, but their presence in the protein corona was not investigated so far. Therefore, we analyzed the prevalence of anti-PEG related antibodies in healthy individuals, determined their concentration in the protein corona of PEGylated NCs, and studied their influence on cellular uptake. Finally, we show that anti-PEG antibody levels should definitely be considered for nanocarrier-based drug delivery.

As mentioned, an increasing number of studies report pre-existing anti-PEG antibodies in Chinese and North American



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populations.^{7,11,23} Against this background, we performed a plasma screening to evaluate the prevalence of anti-PEG antibodies among the German population. To this end, we received 500 plasma samples from healthy blood donors, representing the plasma source we typically use for protein corona studies. The samples were chosen randomly and collected together with information about the donor's age (year of birth) and gender. It is important to note that the plasma samples were collected in 2019 before PEGylated Covid-19 vaccines were approved and applied. We analyzed the anti-PEG antibody prevalence and concentration in plasma samples using a modified ELISA test.¹¹ In the assay, anti-PEG antibodies in the plasma samples bound to poly(ethylene glycol) diamine immobilized on the plates.

An enzyme-linked secondary antibody then specifically detected anti-human IgG or IgM antibodies. Additionally, we performed a competition assay to confirm that anti-PEG antibodies specifically bind to PEG and did not randomly adsorb on the plates. For the competition assay, free PEG was added to the plasma samples to compete with the immobilized PEG chains. This reduced the number of detectable antibodies. Consequently, the sample was considered positive for anti-PEG antibodies if the color reaction was reduced by at least 35% compared to the reading without the addition of free PEG. The relative concentrations of anti-PEG IgG or IgM antibodies in the plasma samples were determined by comparison to standard curves obtained from a serial dilution of chimeric anti-PEG antibodies c3.3-IgG or cAGP4-IgM.²⁴ Fig. 2 shows the schematic setup of the ELISA protocol and the corresponding results of the plasma

screening. As indicated in Fig. 2(b), anti-PEG antibodies were present in the majority of the samples with minor differences between male and female donors. 81.8% of the female donors were positive for anti-PEG IgG, and 55.1% were positive for anti-PEG IgM. The male donors showed a slightly lower prevalence of 74.4% for anti-PEG IgG and 54.1% for anti-PEG IgM. Overall, 48.8% of all donors were positive for both IgG and IgM, and in only 16.6% no anti-PEG antibodies could be detected. This means that in ~83% of all donor samples anti-PEG antibodies were found. In general, anti-PEG IgG was more prevalent than IgM, while both followed a similar trend, as shown in Fig. 2(c). The prevalence of anti-PEG IgG and IgM is shown depending on donor age. Samples were grouped into 10 years time intervals and groups of <20 and >60 years of age. Here, it can be seen that anti-PEG prevalence roughly followed a linear trend and decreased with increasing age for both immunoglobulin isotypes.

As already shown, anti-PEG IgG occurred in a higher concentration than IgM in all age groups. Fig. 2(d) and (e) display the anti-PEG IgG and IgM concentrations. Both immunoglobulin isotypes varied the most and showed the highest absolute concentrations in the age group between 21–30 years. The outliers and mean values of antibody concentration slightly decreased with increasing age, more prominently for anti-PEG IgG than for IgM. More details on the plasma screening including the anti-PEG IgG and IgM concentration of all samples and the age distribution can be found in the (Fig. S1, ESI†).

These results agree with findings from previous studies. Chen *et al.* reported anti-PEG IgG and IgM antibodies in

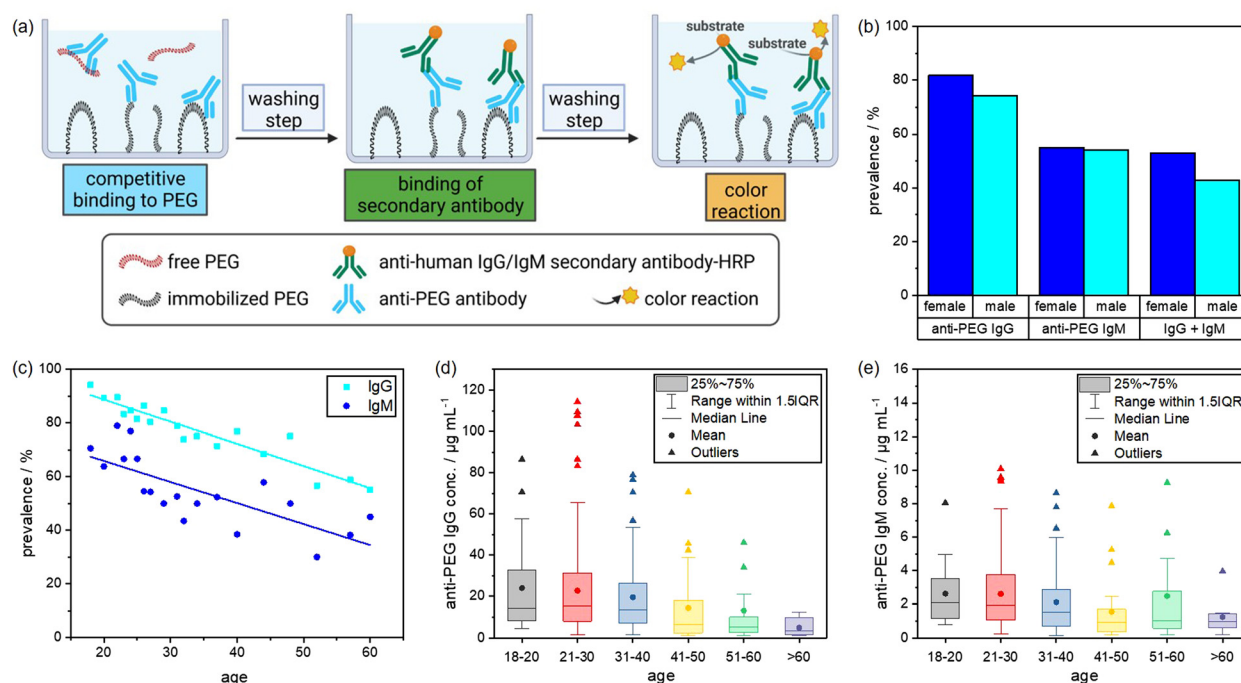


Fig. 2 Plasma screening to analyze anti-PEG antibody prevalence and concentration in a sample of the German population ($n = 500$). (a) Schematic setup of the ELISA test. (b) Prevalence of anti-PEG IgG and IgM antibodies (blue: female, cyan: male samples). (c) Prevalence distribution depending on age (cyan: IgG, blue: IgM), the line indicates the linear regression. (d) Concentration of anti-PEG IgG antibodies in age groups. (e) Concentration of anti-PEG IgM antibodies in age groups. Differences in average concentrations shown in (d) and (e) are not statistically significant due to outliers in the high concentration range.

The obtained results are listed in Table 1 and the corresponding binding curves are shown in Fig. S2 and S3 (ESI[†]). The binding of anti-PEG IgG to mPEG with a molecular weight of 10 000 or 20 000 g mol⁻¹ yielded the lowest K_d values (24–44 nM), which indicates the highest binding affinity. The difference between the two K_d values is not significant, but for lower PEG molecular weights the obtained K_d values were considerably higher. This means that binding strength decreased with

^a K_d values are mean $\pm$ S.D. from several measurements (except for mPEG_{2k} where $n = 1$ and error of fit is given instead). Details see ESI.

The NC size (hydrodynamic diameter) was determined from dynamic light scattering (DLS) and ranges between 148 and 212 nm for all NCs. The zeta potential is related to the net surface charge of the NCs. As expected, the zeta potential for all PEGylated SiNCs was slightly negative, while SiNCs with CTMA-Cl exhibited a positive zeta potential. We prepared the protein corona using undiluted pooled human plasma and determined the anti-PEG antibody level in this plasma batch by ELISA. The detected anti-PEG IgG concentration was $9.6 \pm 0.8 \mu\text{g mL}^{-1}$.

Table 2 Characterization of nanocarrier systems regarding physicochemical properties

	SiNC-CTMA-Cl	SiNC PEG $n = 25$ medium	SiNC PEG $n = 50$ low	SiNC PEG $n = 50$ medium	SiNC PEG $n = 50$ high	SiNC PEG $n = 80$ medium
Surfactant	Cetyltrimethylammonium-chloride (CTMA-Cl)	Lutensol [®] AT25	Lutensol [®] AT50	Lutensol [®] AT50	Lutensol [®] AT50	Lutensol [®] AT80
$n(\text{Lutensol}^{\text{®}})/\text{mol}$	-	2.85×10^{-5}	1.42×10^{-5}	2.85×10^{-5}	5.70×10^{-5}	2.85×10^{-5}
$M_w \text{ PEG}/\text{g mol}^{-1}$	-	1230	2460	2460	2460	3940
D_h (PDI)/nm	262 ± 65 (0.13)	231 ± 39 (0.14)	153 ± 3 (0.11)	212 ± 47 (0.25)	164 ± 8 (0.23)	207 ± 34 (0.35)
Zeta potential/mV	$+13 \pm 1$	-2 ± 1	-5 ± 1	-7 ± 1	-9 ± 1	-7 ± 1

which corresponds to 0.06% of the total immunoglobulins found in this plasma batch (Table S2, ESI[†]). We analyzed the protein corona using different methods to establish the overall protein concentration by Pierce assay and the detailed protein composition in the protein corona by LC-MS. The percentage of total immunoglobulins was then converted into immunoglobulin concentration based on the Pierce assay. As before, ELISA was used to measure the anti-PEG IgG concentration in the protein corona samples. Fig. 3 shows the corresponding results of the protein corona analysis. The list of TOP25 identified proteins in the protein corona is shown in Fig. S5 (ESI[†]). For a more detailed comparison, the fraction of anti-PEG IgG antibodies of the total immunoglobulin concentration is displayed in Fig. 3(b). The overall determined protein amount was similar for all PEGylated SiNCs, as expected from the general effect of PEGylation, while SiNC-CTMA-Cl showed a significantly higher amount of adsorbed proteins.

Additionally, we detected a slightly higher concentration of immunoglobulins in the protein corona of CTMA-Cl stabilized NCs. In contrast, PEGylated NCs exhibited the highest concentration and fraction of anti-PEG IgG in the protein corona.

We observed no significant difference in anti-PEG IgG fraction in the protein corona derived from NCs with a PEG chain length of $n = 50$ or $n = 80$ PEG repeating units and a medium or high surface density. In contrast, a lower surface density of PEG led to a slight decrease of the anti-PEG IgG fraction in the protein corona. This was even more noticeable when a shorter PEG chain length ($n = 25$ repeating units) was present on the NC surface. This trend agrees with the determined binding affinities as mentioned before, which increased with longer PEG chains. In general, anti-PEG IgG is enriched in the protein corona of PEGylated NCs compared to non-PEGylated NCs or human plasma. Our results indicate that a certain density and chain length was necessary for a “saturation” of the protein corona with anti-PEG antibodies. Afterwards, higher density or longer chain length did not lead to increased binding of anti-PEG antibodies. Interestingly, even the CTMA-Cl NCs showed a slight enrichment of anti-PEG IgG compared to plasma levels. It is not clear what the reason for this enrichment is, but might be related to the charge of the NCs. It is known that especially charged surfaces induce interactions with immunoglobulins, but so far this effect was not investigated on the level of specific antibodies.^{35,36}

PEGylation in general often leads to a decrease of unspecific protein adsorption and enrichment of stealth proteins like clusterin, which in combination helps to reduce unspecific

cellular uptake.⁴ The results of the protein corona analysis revealed that anti-PEG IgG became enriched in the protein

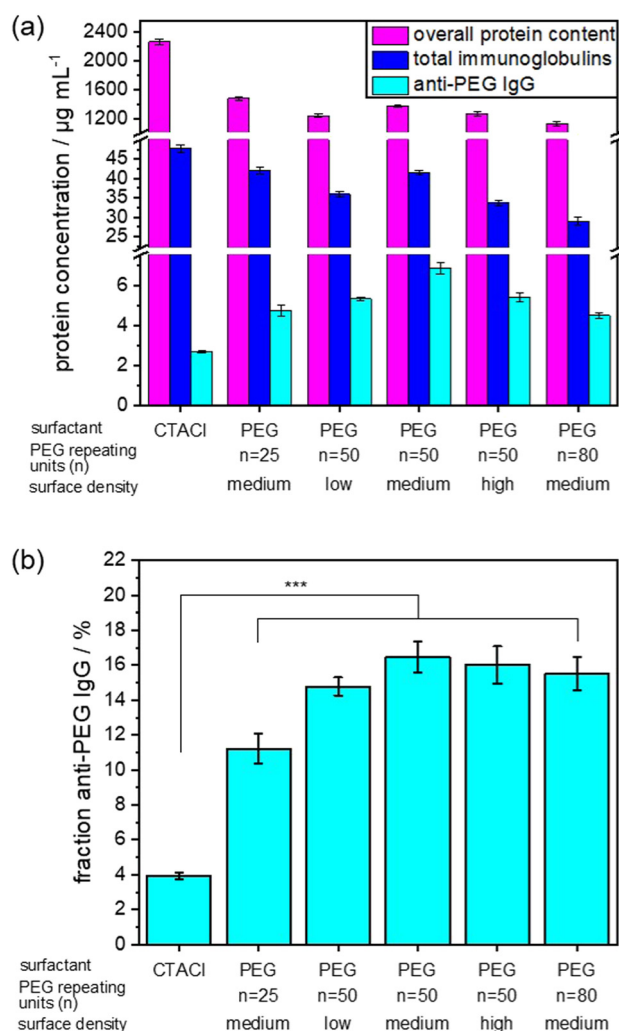
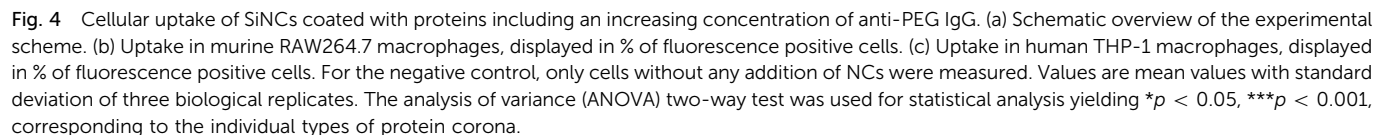


Fig. 3 Investigation of anti-PEG IgG presence in the protein corona. Values are mean values with a standard deviation of three replicates. (a) Protein concentration in the protein corona depending on the presence of PEG on the NC surface, for a normalized NC surface area of 0.05 m^2 per sample. Purple: overall protein concentration of all plasma proteins as analyzed by a Pierce assay, blue: total immunoglobulin concentration as analyzed with LC-MS (percentage of total proteins converted to concentration based on Pierce assay), cyan: anti-PEG IgG concentration analyzed by ELISA. (b) Fraction of anti-PEG IgG concentration compared to the total concentration of immunoglobulin. The analysis of variance (ANOVA) one-way test was used for statistical analysis yielding *** $p < 0.001$, corresponding to the difference between CTMA-Cl and PEG-stabilized NCs.

In general, IgG acts as an opsonin, meaning when present in the corona it promotes the internalization of NCs into phagocytosing cells.⁴⁰ Accordingly, our observed increased uptake in macrophages might not be anti-PEG antibody specific, but a general result of an increased presence of immunoglobulins in the protein corona. Due to sensitivity reasons, we were not able to analyze the presence of IgM in the protein corona and its consequences for cellular uptake. However, we expect a similar trend but with generally lower concentrations. Many studies reported that anti-PEG antibodies can be elicited by PEGylated



The composition of the protein corona depends on various factors – primarily the physico-chemical properties of the NCs. Many researchers attempt to control the composition of the proteins adsorbed to NCs to prevent clearance by immune cells.⁵¹ To obtain further insight into the role of anti-PEG antibodies and generalize their effect, additional PEGylated and unPEGylated NC systems need to be investigated. Due to the high concentration of anti-PEG antibodies in the blood-stream, they might also accumulate non-specifically in the protein corona. Thus, it is also of great interest to explore more widely, which antibody types are found among the immunoglobulins in the protein corona.

Due to the high prevalence of anti-PEG antibodies in the plasma samples, we investigated their concentration in the protein corona of nanocarriers with varying degrees of PEGylation. Anti-PEG antibodies were indeed enriched in the protein corona of PEGylated nanocarriers with increasing PEG chain

Our findings have significant implications regarding the use and design of nanomedicines and contribute to further our understanding of nano-bio interactions.

Mareike F. S. Deuker: data curation, formal analysis, investigation, methodology, validation, visualization, writing – original draft. Volker Mailänder: supervision, methodology. Svenja Morsbach: conceptualization, methodology, project administration, writing – review & editing, supervision. Katharina Landfester: conceptualization, supervision, funding acquisition, writing – review & editing.

There are no conflicts to declare.

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