

The versatility of “click” reactions: molecular recognition at interfaces†

Cite this: *RSC Adv.*, 2014, 4, 17694

Thomas Heinrich,^{ab} Christoph H.-H. Traulsen,^a Erik Darlatt,^b Sebastian Richter,^a Johannes Poppenberg,^a Nora L. Traulsen,^a Igor Linder,^a Andreas Lippitz,^b Paul M. Dietrich,^b Baha Dib,^a Wolfgang E. S. Unger^{*b} and Christoph A. Schalley^{*a}

In order to investigate molecular recognition on surfaces, an azide-functionalized monolayer was deposited on gold. The monolayer was characterized by X-ray photoelectron spectroscopy (XPS) and angle-resolved near-edge X-ray absorption fine structure (NEXAFS) experiments and the decomposition of the azide upon irradiation with X-ray beams was investigated. Subsequently, various alkyne-functionalized host and guest molecules were attached to the azide by 1,3-dipolar cycloaddition. These modified surfaces and their host–guest chemistry were analysed by XPS and angle-resolved NEXAFS. The reversibility of guest binding was shown for one example as a proof of principle.

Received 27th February 2014
 Accepted 28th March 2014

DOI: 10.1039/c4ra01730g

www.rsc.org/advances

Introduction

The controlled modification of interfaces with various functional groups is a main topic in modern physical, materials and bioorganic chemistry.^{1–9} Therefore, a variety of approaches has been developed. The covalent attachment of organic compounds on surfaces yields highly stable interfaces.¹⁰ Among others, the silanization of silica substrates^{11,12} and the chemisorption of organic thiols¹³ or liponic acids¹⁴ on gold substrates is a far more convenient approach. Numerous examples exist, using the formation of stable self-assembled monolayers (SAMs) on solid supports with different terminal groups such as hydroxides,¹⁵ carboxylates,¹⁶ pyridines,¹⁷ terpyridines,^{18–22} and amines.^{23–28} These terminal groups enable the attachment of additional layers on top using for example the formation of thiourea²⁴ or non-covalent complex formation with transition metal ions.^{17,19,29,30} Further terminal groups which have lately gained attention in this context are 1,3-dipoles such as nitrile oxides and azides which react with alkynes in what has been coined the “click” reaction.^{31–34} Another approach is the non-covalent physisorption of Langmuir–Blodgett films *e.g.* the non-covalent attachment of saturated and non-saturated carboxylates.^{35,36} While covalently attached SAMs are somewhat limited by reduced reversibility of and long reaction times for self-

assembly, Langmuir–Blodgett films require amphiphilic molecules and are often less stable against polar solvents and/or reactive species.³⁷

The transfer of supramolecular structures and processes from solution to solid supports is still a challenging task in modern physical and biochemistry.^{38–40} Our approach presented here combines covalent generation of SAMs that can further be functionalized by host or guest molecules which allows the specific non-covalent attachment of another layer through molecular recognition. An azide-functionalized SAM is used here to graft these molecules onto the surface by a copper-catalysed 1,3-dipolar cycloaddition of azides and alkynes forming triazole rings.⁴¹ The simple experimental setup, mild reaction conditions and fast conversion of the “click” reaction in solution remain more or less unchanged features of surface-bound triazole formation as shown earlier for the immobilization of terpyridines,³¹ polymers,⁴² ferrocene,³⁴ nanoparticle multilayers,⁴³ and metal organic frameworks.⁴⁴ An advantage of the modification of surfaces *via* “click” reactions is the orthogonality of the azide and alkyne with most other functional groups. This broadens the variety of molecules significantly, which can be attached. Triazole formation also often gives rise to densely packed consecutive layers.³²

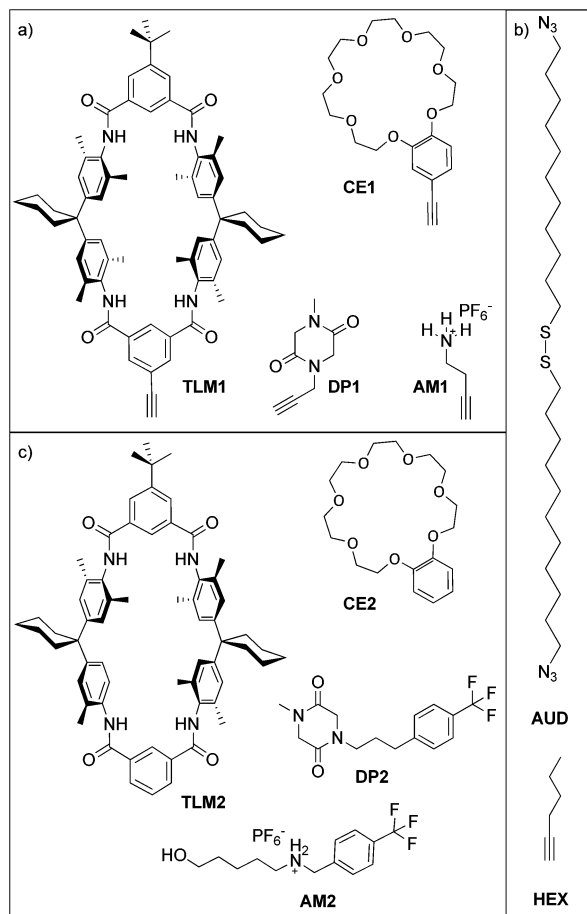
Here, we report the formation of an azide-terminated monolayer which is subsequently functionalized with benzo[21] crown-7 ether **CE1** and tetralactam macrocycle **TLM1** (ref. 45) as specific host molecules and with primary ammonium **AM1** and diketopiperazine **DP1** as the corresponding guest molecules (Scheme 1). For each surface-attached molecule, a counterpart (**CE2**, **TLM2**, **AM2** and **DP2**) exists which does not carry the alkyne and can be used for on-surface host–guest experiments. These molecules were selected as both binding motifs were thoroughly studied in solution. The tetralactam macrocycle

^aInstitut für Chemie und Biochemie der Freien Universität Berlin, Takustraße 3, 14195 Berlin, Germany. E-mail: c.schalley@fu-berlin.de

^bBAM Federal Institute for Materials Research and Testing, Unter den Eichen 44-46, 12203 Berlin, Germany. E-mail: wolfgang.unger@bam.de

† Electronic supplementary information (ESI) available: Full experimental procedures and characterization of new compounds, copies of ¹H, ¹³C and ¹⁹F NMR spectra, multilayer preparation, instrumentation and data processing used for the described experiments, additional transmission, XPS and NEXAFS data for the characterization of samples. See DOI: 10.1039/c4ra01730g





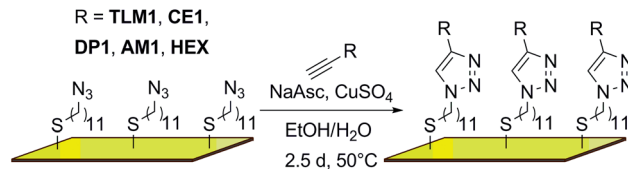
Scheme 1 (a) Alkyne-functionalized host and guests, (b) control compound HEX and SAM-forming disulphide AUD, and (c) the corresponding guests and host. The CF₃ groups in DP2 and AM2 serve as labels for easy detection by XPS.

nically binds diketopiperazines in its cavity through four hydrogen bonds between the guest's carbonyl groups and the converging amide NH groups of the macrocycle.^{46,47} Benzo[21]crown-7 is just large enough to allow pseudorotaxane formation with secondary alkyl ammonium axes.⁴⁸ Hexyne **HEX** was used as a reference to investigate triazole formation during the "click" reaction. In former studies, we have demonstrated the utility of copper(i)-catalysed 1,3-dipolar cycloadditions on azide-terminated SAMs assembled from 1,2-bis(11-azidoundecyl)-disulfane (**AUD**) for the covalent attachment of functionalized alkynes. Since **AUD** is analogous to alkanethiol SAMs, it should form densely packed and well-ordered SAMs on gold substrates.^{31,32}

Results and discussion

Analysis of azide-decomposition upon irradiation

In the first step, an **AUD** SAM was prepared which forms the basis for further functionalization with alkyne-substituted organic molecules as shown in Scheme 2. The **AUD** SAM was characterized by quite surface-sensitive synchrotron radiation X-ray photoelectron spectroscopy (SR-XPS; 500 eV excitation



Scheme 2 Surface attachment of organic molecules via "click" chemistry (NaAsc = sodium ascorbate).

energy, XPS 95% information depth $z_{95\%} \sim 2.1$ nm) and NEXAFS spectroscopy as described earlier. Angle-resolved NEXAFS experiments provide evidence for a predominant upright orientation of the **AUD** molecules.^{31,32}

A major characterization problem of surface-bound azides by XPS is the decomposition of the azides upon X-ray radiation. This irradiation damage yields a new nitrogen component at lower binding energy (BE) in the N 1s XP spectrum of a pristine azide-terminated SAM. Non-damaged SAM molecules should provide three components of equal peak areas diagnostic for azide moieties as displayed in Fig. 1a. Two major components at lower BE (400.9 and 400.3 eV) and a single component at higher BE (404.2 eV) can be assigned to electron-rich and electron-poor azide nitrogen atoms.^{31,37} During the decomposition process, a new signal appears at a lower BE of 399.0 eV (Fig. 1a). In order to gain more detailed insight into this process and in order to be able to clearly assign all nitrogen species which may arise after the "click" reaction, 35 N 1s spectra were recorded in 4 min intervals at the same spot to follow the time course of the decomposition reaction resulting in a total irradiation time of 142 minutes. In Fig. 1b, the normalized N 1s peak areas of the overall (black line), the azide (green line) and the decomposition products (blue line) are plotted. As expected, the decomposition process is accompanied by an exponential decay of the azide nitrogens. The overall amount of nitrogen does not remain constant, but decreases to about a third of the starting nitrogen peak area at the end of the irradiation time with a half-life of 34 min. Molecular nitrogen is thus released during the decomposition reaction. Nitrogen release follows a first-order rate law ($k = 3 \times 10^{-4} \text{ s}^{-1}$) as determined by plotting the natural logarithm of the azide nitrogen mole fraction over the irradiation time resulting in a linear fit (Fig. 1c).

Mechanistically, the decomposition can follow different pathways (Scheme 3). The initially formed highly reactive nitrene intermediate can either react by insertion into a C–H bond of a neighbouring **AUD** molecule. This reaction would give rise to secondary amines, which is however inconsistent with the binding energy of the new nitrogen species.²⁹ Consequently, this reaction does not play a prominent role. The second pathway would be dimerization of two nitrenes to yield an azo-compound, which might react through nitrogen loss forming two alkyl radicals. However, no XPS peak for azo-groups is observed and therefore, we also rule out this pathway as the major decomposition route.²⁸ A third typical reaction of electron sextet compounds such as nitrenes is a 1,2-hydrogen shift from the adjacent carbon to the atom with the electron sextet, here the nitrene nitrogen atom. This hydrogen shift leads to an



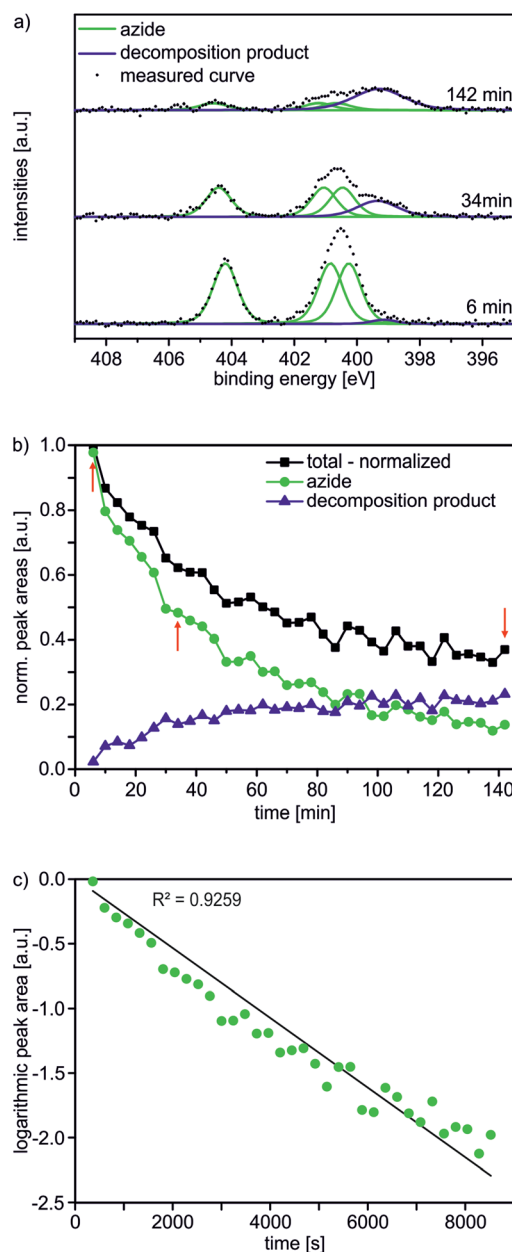
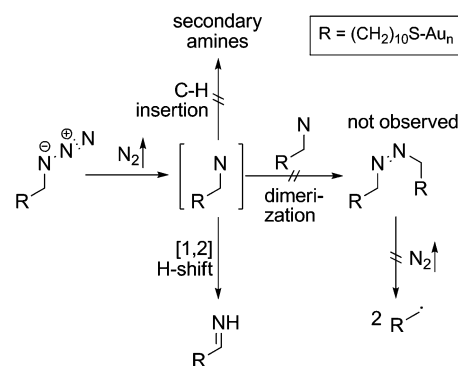


Fig. 1 *In situ* damage of the AUD SAM caused by monochromatic Al K α radiation (X-ray excitation energy 1486 eV) with (a) peak fits of the N 1s XP spectra, (b) quantitative analysis of the individual peak fits. Red arrows indicate the data points corresponding to the spectra in (a). (c) Logarithmic plot of normalised azide peak areas over time. All peak areas are normalized to the total peak area of the first spectrum after 6 min irradiation time.

imine. The binding energy of 399.0 eV for the new component is consistent with the formation of an imine. Also, the π^* resonance at 398.3 eV observed in the N K-edge NEXAFS spectroscopy suggests imine formation.²⁸ The 1,2-hydrogen shift pathway is consequently the major reaction following nitrogen loss and nitrene formation. However, it is not possible to determine whether N₂ loss and the 1,2-hydrogen shift are subsequent steps or occur in a concerted manner due to the rather long intervals between the spectra.



Scheme 3 Possible azide decay-mechanisms with a nitrene as the central intermediate.

As a second conclusion, these data reveal azide decomposition to occur rather slowly so that it does not significantly hamper the analysis of azide-functionalized surfaces. This is also important for the following experiments, in which alkynes are clicked to the azides, as for example a sterically demanding molecule such as tetralactam macrocycle **TLM1** will like leave azides “unclicked” underneath the macrocycle layer.

Cycloaddition experiments

N 1s XP and N K-edge NEXAFS spectra. On-surface “click” reactions were performed using alkyne-substituted **TLM1**, **CE1**, **DP1** and **AM1**. In order to clearly assign all observed XPS peaks, the unfunctionalized alkyne **HEX** was included in this series. **HEX** does not only contain no additional nitrogen components, but has also a small steric demand. Therefore, **HEX** is well suited as a reference.

For all five surfaces, the Au-S interface is stable under the experimental conditions of the “click” reaction as indicated by the main S 2p_{3/2} species that is detected at ~162.0 eV (ESI†).³²

Fig. 2 (left) summarises the N 1s XP spectra of these five surfaces in comparison to that of the pristine **AUD** SAM. In order to reduce irradiation damage, irradiation times were kept below three minutes. Clearly, the azide signals of the latter vanish completely, when **HEX** is added. Instead, three nitrogen signals for the triazole (BE = 399.6, 400.3, and 401.6 eV) with equal peak areas and equal FWHM values can be fitted into the broad N 1s signal. A more or less quantitative “click” reaction was achieved as indicated by the absence of the azide signal at 404.8 eV. A very similar result is obtained for the surface covered with crown ether **CE1**, which does not bear any additional nitrogen atoms.

The picture changes for **AM1**, which was “clicked” to the **AUD** SAM as the free amine followed by treatment with HPF₆. This molecule contains an additional ammonium nitrogen atom. Consequently, the triazole signals are superimposed by an additional peak at a BE of 400.3 eV. This binding energy is surprisingly the same as that of free amines as observed for the same surface before protonation, while no new signal for ammonium is observed at the expected BE of 401.8 eV. Thus, only a small protonation yield is obtained. We refrained from using H₂SO₄ for protonation as reported earlier for surface-



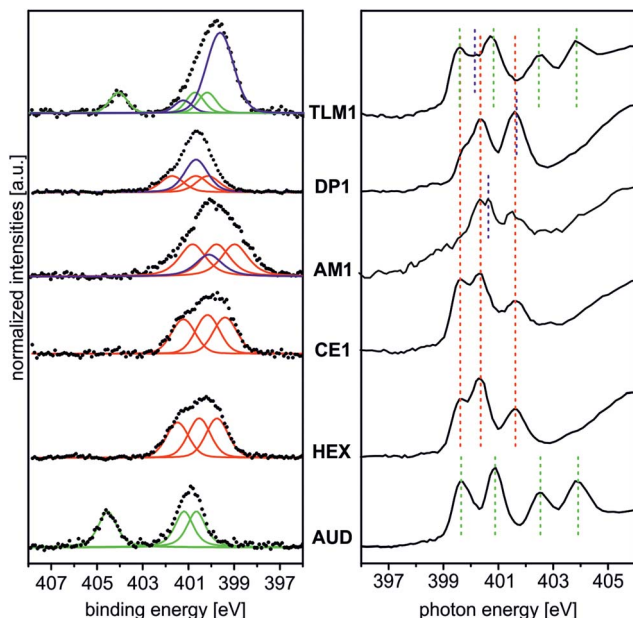


Fig. 2 Left: qualitative N 1s XP spectra of the surfaces of HEX, CE1, AM1, DP1 and TLM1 in comparison to that of the pristine AUD SAM ($h\nu = 500$ eV and 0° emission angle). Black dotted lines: experimental spectra; green and red curves: contributions of azide and triazole. Blue curves correspond to amine (AM1) and amide nitrogen atoms (DP1, TLM1). To facilitate direct comparison, the backgrounds of the spectra were subtracted followed by normalization to the maximum count rate. Right: π^* region of the N K-edge NEXAFS spectra at 55° incident synchrotron light. For qualitative comparison, the spectra were normalized to their maximum count rates. The different photon energies for azide (green), triazole (red), amine and amide nitrogens (blue) are indicated by dotted lines.

bound pyridines,¹⁷ as strong ion pairing between the doubly charged sulphate and ammonium ions will likely compete too strongly with crown ether binding in the host-guest studies.

The surface bearing DP1 also exhibits a new signal in addition to the triazole nitrogen atoms which corresponds to the two amide nitrogen atoms of the diketopiperazine and is detected at a binding energy of 400.8 eV.⁴⁰ Finally, tetralactam macrocycle TLM1 exhibits the triazole signals superimposed by a large peak at BE = 400.3 eV for the four amide nitrogen atoms.²⁸ The azide signal at BE = 404.8 eV is also clearly visible. This indicates the steric bulk of TLM1 to leave unreacted azides underneath the macrocycle layer. Not unexpectedly, the triazole nitrogen atoms exhibit small peak shift changes depending on the exact chemical environment provided by the different molecules attached.

These XPS data are supported by N K-edge NEXAFS experiments (Fig. 2, right). Clearly, azide signals (green dotted lines) appear for the pristine AUD SAM and for the TLM1-functionalized surface. The N K-edge spectrum of the AUD SAM exhibits four pronounced signals in the π^* region at 399.6, 400.9, 402.5 and 403.9 eV (Fig. 2, bottom right). Electron transitions from the N 1s energy level of the central azide nitrogen into two different π^* orbitals are the reason for the resonances at higher photon energy (PEs). The other two signals at lower PEs can be

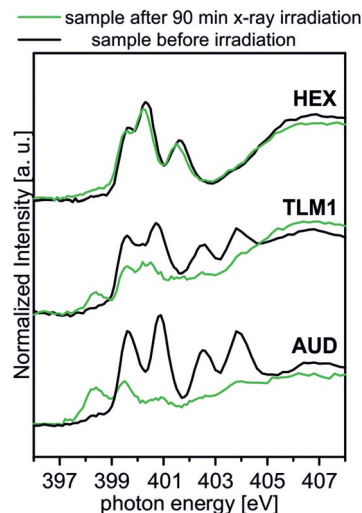


Fig. 3 Behaviour of TLM1 attached to the surface upon radiation in comparison to the AUD SAM and the surface attached HEX.

assigned to the two adjacent nitrogen atoms of the azide moiety.³¹

Spectral contributions attributed to the triazole group (red lines) are visible for all surfaces except the AUD SAM. The assignment of the signals at 399.6 eV, 400.3 eV and 401.5 eV is in good agreement with DFT calculations reported earlier.³¹ The additional amine and amide nitrogen atoms of AM1, DP1 and TLM1 lead to the observed changes in the structure of the triazole peaks (e.g. PE = 401.6 eV for the tertiary amide π^* resonances in DP1 (ref. 49) or PE = 400.4 eV for the secondary amides in TLM1 (ref. 29)). The presence of remaining azide groups underneath the TLM1 layer are also supported by NEXAFS spectra (Fig. 3): when the surface is irradiated for longer intervals, it clearly reveals the same radiation-induced azide decomposition as the pristine AUD SAM, while similar changes do not occur for the HEX-functionalised surface.

These experiments provide clear evidence for the successful attachment of the alkyne-substituted molecules under study here in copper-catalysed click reactions. The surface coverage is likely high as indicated by the fact that only the largest molecule, TLM1, leaves visible signals for unreacted azide group.

C 1s XP spectra. More detailed insight into triazole formation with different alkynes can be obtained by analysing the C 1s BE region of the XP spectra (Fig. 4). The major carbon component for the aliphatic carbon atoms in HEX and DP1 is observed at BE = 284.8 eV. DP1 additionally exhibits a major component at 286.6 eV which can be assigned to the C–N carbon atoms.⁴⁹ Similarly, the crown ether CH₂ groups that are flanked by oxygen atoms appear at BE = 286.7 eV (C–O). All carbon atoms can be classified into those adjacent to electron-withdrawing heteroatoms (C–N, C–O; BE \sim 286 eV), amide carbons (BE 288–289 eV) and all remaining aliphatic and aromatic carbon atoms (BE \sim 285 eV). The relative peak areas and theoretical ratios calculated for the different components are summarized in Table 1 and are in good agreement with each other – clearly supporting a successful “click” reaction with all compounds under study.



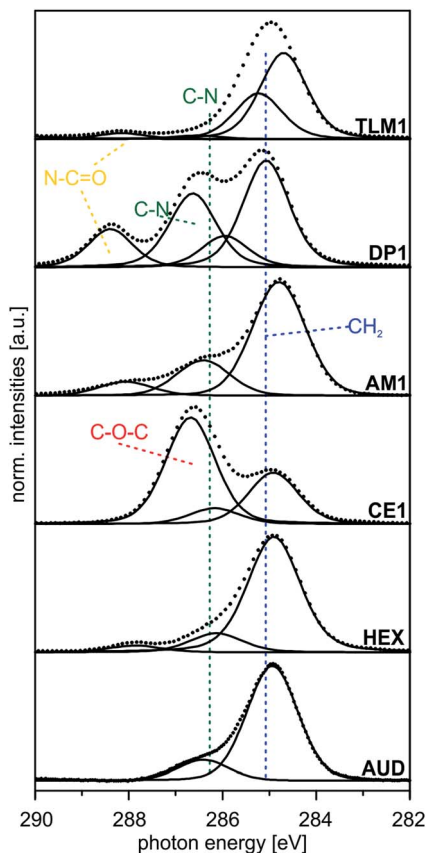


Fig. 4 Peak fits of C 1s SR XP spectra ($h\nu = 350$ eV, emission angle = 0°) obtained at high surface sensitivity of XPS, 95% information depth ($z_{95\%} \sim 2$ nm).⁵⁶

Angle-resolved C K-edge NEXAFS spectra. Angle-resolved NEXAFS spectra (Fig. 5) can be used to determine, whether the molecules on the surface exhibit a preferential orientation. In

Table 1 Summary of the C 1s peak fits. Theoretical peak areas are calculated according to the corresponding sum formula

Sample	BE ^a [eV] (rel. peak area)	Theor. ratios	Assigned carbon atoms	FWHM [eV]
AUD	284.9 (85)	82	-CH ₂ -	1.3
	286.4 (15)	8	-CH ₂ -N ₃ , -CH ₂ -S	
HEX	284.9 (82)	82	-CH ₂ -	1.3
	286.1 (14)	18	C _{sp²} -N, N-CH ₂ -	
CE1	284.9 (30)	34	-CH ₂ -	1.3
	286.2 (9)	10	C _{sp²} -N, N-CH ₂ -O-	
AM1	286.7 (61)	56	CH ₂ -, -O-C(sp ²)-	1.3
	284.8 (70)	71	-CH ₂ -	
DP1	286.4 (25)	29	C _{sp²} -N, N-CH ₂ -	1.2
	285.1 (52)	52	-CH ₂ -	
TLM1	285.9 (10)	11	N-CH ₂ -, N-CH ₃	1.2
	286.6 (27)	26	OCN-CH ₂ -, C _{sp²} -N	
	288.4 (11)	11	O=C-N-	
	284.5 (52)	48	C _{sp²}	
	285.2 (37)	44	-CH ₂ -, C-C	
	286.6 (2)	3	C-N, C _{sp²} -N	
	288.1 (4)	5	O=C-N	

^a Binding energy reference: Au 4f_{7/2} signal at BE = 84.0 eV.

this case, linear dichroism effects are observed, while the NEXAFS spectra obtained for unordered surfaces do not depend on the angle of the incident light.

In the angle-resolved NEXAFS spectra of the pristine AUD SAM, clear linear dichroism effects are observed. The C-H* resonance decreases significantly when the angle of the incident linear polarized synchrotron light is changed from 90° to 30° . In contrast, the σ^* resonance increases. This is typical for well-ordered SAMs of long alkyl chains that are oriented in an upright fashion on the surface.^{13,50} The same effect is observed for HEX-modified AUD SAMs. Therefore, we conclude the HEX SAM to be highly ordered as well as the AUD precursor SAM.

Linear dichroism effects are also observed for the CE1-functionalized surface. In addition to the effect of the underlying AUD SAM, one observes a linear dichroism effect for the peak corresponding to the σ^* resonance of the CH₂-O groups, which appears at 289.3 eV.^{51,52} This effect points to crown ethers with a preferential orientation and thus ordered crown ether layers even though it remains unclear, what their exact packing pattern on the surface is. Furthermore, the DP1-covered surface exhibits preferentially oriented diketopiperazines as indicated by the linear dichroism in the π^* (C=O) resonance at 288.4 eV.⁵¹ In marked contrast, the linear dichroism effects are small for the surfaces loaded with AM1 and TLM1. It is therefore likely that AM1 is not well ordered. The quite complex TLM1 structure with its many aromatic rings that are oriented in different angles relative to the surface, one would expect only a small linear dichroism effect. It is not possible to clearly state whether

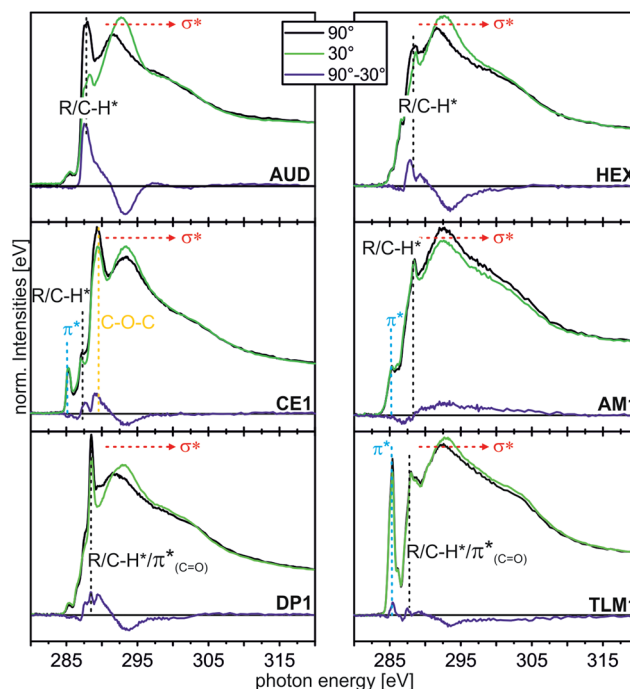


Fig. 5 C K-edge NEXAFS spectra (for experimental details, see ESI†) of the AUD SAM and the click-functionalized surfaces with different moieties at incidence angles of 90° (black) and 30° (green). The difference spectra are illustrated by the blue line for every sample. The characteristic resonances are marked by dotted lines.



the small effects observed have their origin in isotropically distributed macrocycles on the surface or in an ordered layer of macrocycles with aromatic rings mutually compensating each other's linear dichroism effects.

As a conclusion, these angle-resolved NEXAFS experiments provide evidence for preferentially ordered molecules in layers of **AUD**, **HEX**, **CE1** and **DP1**, while “clicking” **AM1** to the surface leads to a more or less isotropic arrangement. The situation after reaction with **TLM1** is too complex to be described.

On-surface host–guest experiments

N 1s XPS and N K-edge NEXAFS analysis of the host–guest complexes on the surface. Four different host–guest experiments can be performed with the molecules and surfaces under study: surface-bound **CE1** can be tested for complexation of the secondary ammonium axle **AM2**. The opposite combination with **AM1** on the surface and **CE2** as the binding partner in solution is also possible. Similarly, the two pairs of surface-deposited **TLM1** and **DP2** and **DP1** and **TLM2** are available. In order to gain evidence for host–guest interactions, the surfaces were immersed for 60 minutes in a 1 mM solution of the corresponding component in dichloromethane followed by rinsing with dichloromethane and drying. The surfaces were characterised by XPS and NEXAFS experiments as described above (Fig. 6).

Beside the triazole nitrogen peaks, the **CE1–AM2** pair clearly shows additional features at 401.7 eV BE in the N 1s XP spectrum and at 405.8 eV PE in the N K-edge NEXAFS spectrum. These signals can be assigned to the secondary ammonium group of the axle²⁸ and thus provide evidence for its presence even after thoroughly rinsing the surface. We therefore conclude the axle to be bound to surface by threading into the cavity of the crown ether. The opposite pair, **AM1** and **CE2**, exhibits an N 1s XP spectrum, which does not differ much from that of the **AM1**-covered surface before crown deposition. This is not surprising because no further nitrogen is introduced and the protonation degree is low according to the results discussed above. This becomes also clear, when one compares the binding energies of the nitrogen incorporated in **AM1** (BE = 400.3 eV for an amine nitrogen) with that of **AM2** (BE = 401.7 eV for the ammonium nitrogen) in the experiment above.

Deposition of the tetralactam macrocycle in the **DP1–TLM2** host–guest combination can already be clearly observed in the N 1s XP spectra. The intensity of the peak at BE = 400.8 eV representing the amides in **DP1** as well as **TLM2** is significantly higher compared to the triazole peaks of the **DP1**-covered surface before **TLM1** deposition (Fig. 2). For the opposite combination of **TLM1** and **DP2**, the situation is not as clear, and C 1s XP spectra will provide more detailed insight.

C 1s XPS analysis. The C 1s XPS data are in good agreement with the analysis of the N 1s and N K-edge spectra in Fig. 6. The

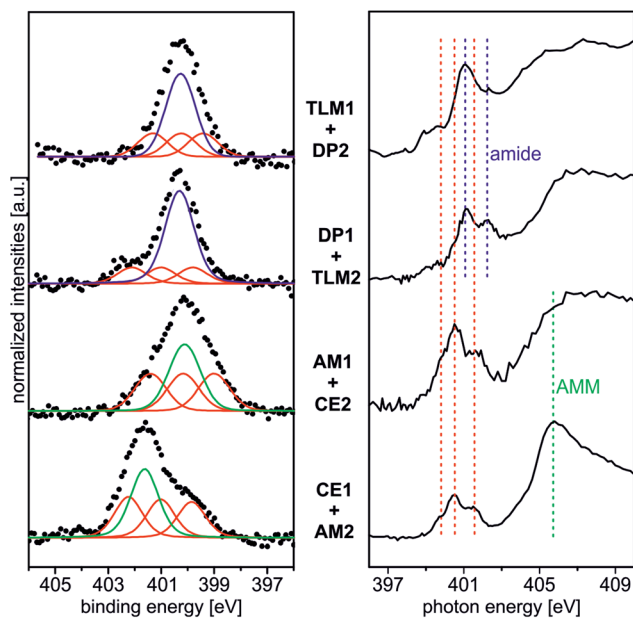


Fig. 6 Left: qualitative N 1s XP spectra of the four host–guest pairs ($h\nu = 500$ eV and 0° emission angle). Black dotted lines: experimental spectra; red curves: triazoles, blue curves: amide groups (**DP1**, **TLM1**), green curves display ammonium (**CE1**) or amine nitrogen atoms (**AM1**). To facilitate comparison, the backgrounds of the spectra were subtracted followed by normalization to the maximum count rate. Right: π^* region of the N K-edge NEXAFS spectra taken at 55° incident synchrotron light. For qualitative comparison the spectra were normalized to their maximum count rates. The different photon energies for triazole resonances (red), amides (blue), and ammonium nitrogen atoms (green) are marked by dotted lines.

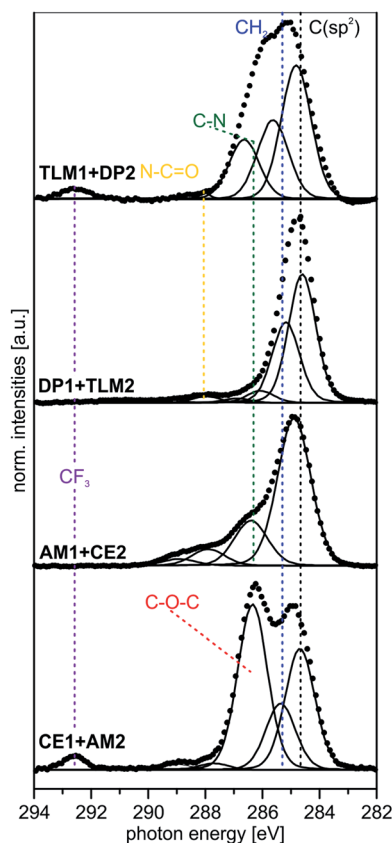


Fig. 7 Peak fits of C 1s SR-XPS spectra ($h\nu = 350$ eV, emission angle = 0°) for the four different host–guest pairs.



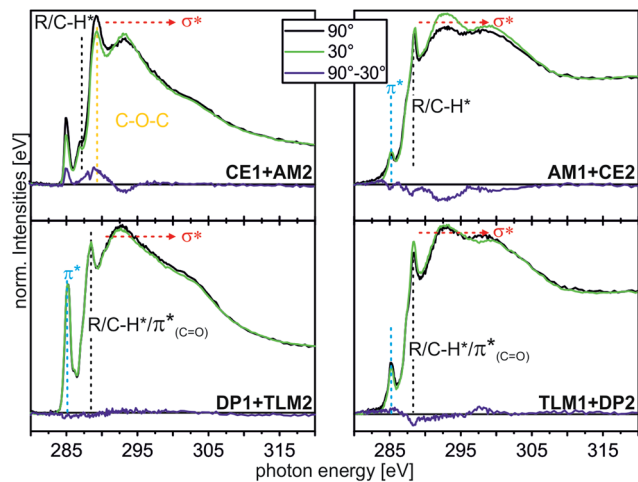


Fig. 8 C K-edge NEXAFS spectra (for experimental details, see ESI†) of the host-guest experiments with different moieties at incident angles of 90° (black) and 30° (green). The differences between the spectra are illustrated by the blue lines. Characteristic resonances are marked by dotted lines.

presence of **AM2** and **DP2** (bottom and top spectra in Fig. 7) is clearly evidenced by the signal for the CF_3 carbon atom, which appears significantly shifted in comparison to all other carbon signals at $\text{BE} = 292.5 \text{ eV}$.⁵³ Also, the signal ratios agree with complex formation. For example, the signals for aromatic carbon ($\text{C}(\text{sp}^2)$) and $\text{CH}_2\text{-N}$ grow relative to the crown ether's $\text{CH}_2\text{-O}$ signal for the **CE1-AM2** host-guest pair. Again, the **AM1-CE2** pair is the exception. The corresponding C 1s XP spectrum remains almost unchanged relative to that of **AM1** before crown ether deposition (Fig. 4). Consequently, we can conclude all host-guest complexes are formed with the exception of the **AM1-CE2** pair, which suffers from the protonation difficulties discussed before.

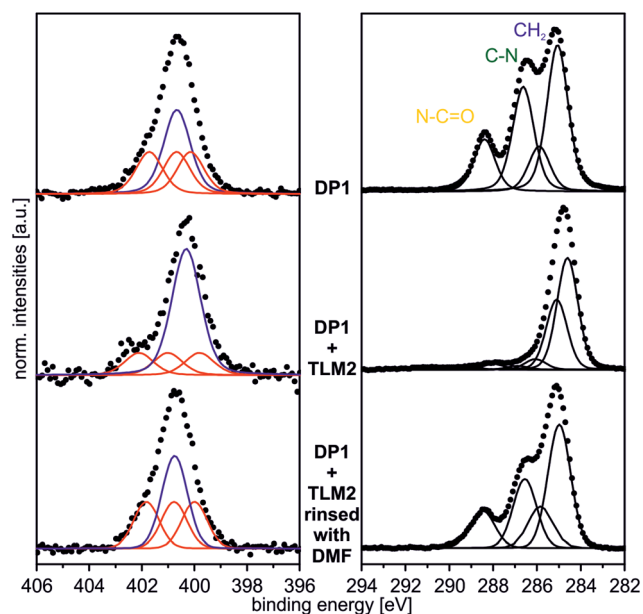


Fig. 9 N 1s ($h\nu = 500 \text{ eV}$; 0°) and C 1s ($h\nu = 350 \text{ eV}$; 0°) spectrum of **DP1 + TLM2** rinsed vigorously with DMF.

Angle-resolved C K-edge NEXAFS Experiments. Again, angle-resolved C K-edge NEXAFS experiments have been performed with the three successful host-guest pairs in order to establish whether the layers are ordered after complex formation (Fig. 8).

The **CE1-AM2** pair clearly exhibits linear dichroism effects on the $\sigma^*(\text{CH}_2\text{O})$ resonance of the crown ether methylene groups. Consequently, the crown ethers still have a preferred orientation. In addition, a new linear dichroism effect is observed for the π^* resonance of the aromatic rings. As this was not present in the corresponding spectra of surface-attached **CE1** prior to **AM2** deposition (Fig. 5), we attribute this effect to the aromatic ring incorporated in the ammonium axle. Consequently, **CE1** on the surface does not only retain its orientational preference, but also induces a preferred orientation into the deposited guest molecule **AM2**.

The other two pairs, **DP1-TLM2** and **TLM1-DP2**, do not exhibit any significant angle-dependence. These two host-guest pairs thus either remain unordered (**TLM1-DP2**) or even lose their orientational preference upon complex formation (**DP1-TLM2**). In particular the latter host-guest pair shows that **DP1** is capable of adapting its position to the steric requirements of its complex with **TLM2** because the dichroism effect of the $\text{C}=\text{O}$ band at 288.4 eV vanishes upon addition of **TLM2**.

Reversibility of the on-surface molecular recognition events

In order to provide a proof of principle that the host-guest complexation is reversible, the **DP1-TLM2** pair was chosen. First of all, the hydrogen bonding between these two molecules is strongly affected in solution by competitive solvents. Second, it is not associated with a similarly high dissociation barrier as the benzo[21]crown-7/secondary ammonium ion motif.

Consequently, a sample surface covered with **DP1** and subsequently treated with **TLM2** for complex formation was rinsed vigorously with dimethylformamide which is a quite strong hydrogen-bond competitor. The XP N 1s and C 1s spectra of the surface after this treatment are displayed in Fig. 9. Both, the N 1s and the C 1s spectra are almost identical to those of the untreated **DP1**-functionalized surface (Fig. 2 and 4). The characteristic features for **DP1** “clicked” to the surface can be observed and furthermore, the relative intensities are similar to those of the pristine **DP1**. Consequently, the host-guest complex can be dissociated with competitive solvents providing evidence for its reversible formation.

Conclusions

In this contribution, click chemistry was utilized to attach host and guest molecules to Au surfaces covered with a self-assembled monolayer with azide head groups. Prior to the cycloaddition reactions, the X-ray irradiation-induced decomposition of the azide groups was investigated in detail by N 1s XPS in order to ensure that XP and NEXAFS analysis delivers valid data for the characterization of the “clicked” surfaces. X-ray damage experiments clearly suggest that molecular nitrogen is lost from the azide to the nitrene intermediate followed by a 1,2-hydrogen shift to the resulting imine.



Covalent anchoring of four host or guest molecules to the above mentioned azide monolayer was obtained by copper-catalysed 1,3-dipolar cycloaddition. The resulting samples were investigated *via* XPS and NEXAFS which confirmed the cycloadditions to be successful for all functional molecules. Surface-attached crown ether **CE1** and diketopiperazine **DP1** exhibit a preferred orientation within the monolayer.

Finally, the attached molecules – except for the primary amine – are found to be able to build host–guest complexes with their corresponding counterpart on the surface as again shown by XPS and NEXAFS experiments. The reversibility of complex formation was shown for the **DP1–TLM2** complex.

Acknowledgements

The authors gratefully acknowledge the Deutsche Forschungsgemeinschaft (SCHA 893/9-1, UN 80/8-1), the Freie Universität Berlin and the Fonds der Chemischen Industrie (FCI) for financial support. S. R. thanks the FCI for a Chemiefonds Ph.D. fellowship. We are grateful to D. Treu and Dr T. Gross for operating the XPS instrument at BAM 6.8. Support by the staff at BESSY II and the HE-SGM CRG (Dr A. Nefedov, KIT) during our activities at the HE-SGM beamline is gratefully acknowledged.

Notes and references

- C. Nicosia and J. Huskens, *Mater. Horiz.*, 2014, **1**, 32–45.
- J. J. Gooding and S. Ciampi, *Chem. Soc. Rev.*, 2011, **40**, 2704–2718.
- D. Samanta and A. Sarkar, *Chem. Soc. Rev.*, 2011, **40**, 2567–2592.
- C. Haensch, S. Hoepfener and U. S. Schubert, *Chem. Soc. Rev.*, 2010, **39**, 2323–2334.
- G. Fioravanti, N. Haraszkiwicz, E. R. Kay, S. M. Mendoza, C. Bruno, M. Marcaccio, P. G. Wiering, F. Paolucci, P. Rudolf, A. M. Brouwer and D. A. Leigh, *J. Am. Chem. Soc.*, 2008, **130**, 2593–2601.
- A. F. M. Altaalar, I. Klinkert, K. Jalink, R. P. J. de Lange, R. A. H. Adan, R. M. A. Heeren and S. R. Piersma, *Anal. Chem.*, 2005, **78**, 734–742.
- D. Wasserberg, C. Nicosia, E. E. Tromp, V. Subramaniam, J. Huskens and P. Jonkheijm, *J. Am. Chem. Soc.*, 2013, **135**, 3104–3111.
- T. Chang, D. I. Rozkiewicz, B. J. Ravoo, E. W. Meijer and D. N. Reinhoudt, *Nano Lett.*, 2007, **7**, 978–980.
- L. Tauk, A. P. Schröder, G. Decher and N. Giuseppone, *Nat. Chem.*, 2009, **1**, 649–656.
- A. Ulman, *Chem. Rev.*, 1996, **96**, 1533–1554.
- M. Li, Y. Wang, V. Pinon III and M. Weck, *Chem. Commun.*, 2011, **47**, 2802–2804.
- Y. Liu, S. Yamazaki and S. Izuhara, *J. Organomet. Chem.*, 2006, **691**, 5809–5824.
- C. Vericat, M. E. Vela, G. Benitez, P. Carro and R. C. Salvarezza, *Chem. Soc. Rev.*, 2010, **39**, 1805–1834.
- K. Kim, W. S. Jeon, J.-K. Kang, J. W. Lee, S. Y. Jon, T. Kim and K. Kim, *Angew. Chem., Int. Ed.*, 2003, **42**, 2293–2296.
- A. Pulsipher, N. P. Westcott, W. Luo and M. N. Yousaf, *Adv. Mater.*, 2009, **21**, 3082–3086.
- M. J. Shuster, A. Vaish, M. L. Gilbert, M. Martinez-Rivera, R. M. Nezarati, P. S. Weiss and A. M. Andrews, *J. Phys. Chem. C*, 2011, **115**, 24778–24787.
- J. Poppenberg, S. Richter, E. Darlatt, C. H.-H. Traulsen, H. Min, W. E. S. Unger and C. A. Schalley, *Surf. Sci.*, 2012, **606**, 367–377.
- A. Auditore, N. Tuccitto, G. Marzanni, S. Quici, F. Puntoriero, S. Campagna and A. Licciardello, *Chem. Commun.*, 2003, 2494–2495.
- C. H.-H. Traulsen, E. Darlatt, S. Richter, J. Poppenberg, S. Hoof, W. E. S. Unger and C. A. Schalley, *Langmuir*, 2012, **28**, 10755–10763.
- N. Tuccitto, V. Ferri, M. Cavazzini, S. Quici, G. Zhavnerko, A. Licciardello and M. A. Rampi, *Nat. Mater.*, 2009, **8**, 41–46.
- N. Tuccitto, I. Delfanti, V. Torrisi, F. Scandola, C. Chiorboli, V. Stepanenko, F. Wurthner and A. Licciardello, *Phys. Chem. Chem. Phys.*, 2009, **11**, 4033–4038.
- C. Battocchio, G. Polzonetti, L. Gambino, N. Tuccitto, A. Licciardello and G. Marletta, *Nucl. Instrum. Methods Phys. Res., Sect. B*, 2006, **246**, 145–150.
- P. M. Dietrich, N. Graf, T. Gross, A. Lippitz, S. Krakert, B. Schüpbach, A. Terfort and W. E. S. Unger, *Surf. Interface Anal.*, 2010, **42**, 1184–1187.
- P. M. Dietrich, N. Graf, T. Gross, A. Lippitz, B. Schüpbach, A. Bashir, C. Wöll, A. Terfort and W. E. S. Unger, *Langmuir*, 2010, **26**, 3949–3954.
- N. Graf, E. Yegen, T. Gross, A. Lippitz, W. Weigel, S. Krakert, A. Terfort and W. E. S. Unger, *Surf. Sci.*, 2009, **603**, 2849–2860.
- M. R. Lockett, M. R. Shortreed and L. M. Smith, *Langmuir*, 2008, **24**, 9198–9203.
- Y. Luo, M. Piantek, J. Miguel, M. Bernien, W. Kuch and R. Haag, *Appl. Phys. A*, 2008, **93**, 293–301.
- X. Song, Y. Ma, C. Wang, P. M. Dietrich, W. E. S. Unger and Y. Luo, *J. Phys. Chem. C*, 2012, **116**, 12649–12654.
- S. Richter, J. Poppenberg, C. H.-H. Traulsen, E. Darlatt, A. Sokolowski, D. Sattler, W. E. S. Unger and C. A. Schalley, *J. Am. Chem. Soc.*, 2012, **134**, 16289–16297.
- C. H.-H. Traulsen, V. Kunz, T. Heinrich, S. Richter, M. Holzweber, A. Schulz, L. K. S. von Krbek, U. T. J. Scheuschner, J. Poppenberg, W. E. S. Unger and C. A. Schalley, *Langmuir*, 2013, **29**, 14284–14292.
- E. Darlatt, A. Nefedov, C. H.-H. Traulsen, J. Poppenberg, S. Richter, P. M. Dietrich, A. Lippitz, R. Illgen, J. Kühn, C. A. Schalley, C. Wöll and W. E. S. Unger, *J. Electron Spectrosc. Relat. Phenom.*, 2012, **185**, 621–624.
- E. Darlatt, C. H.-H. Traulsen, J. Poppenberg, S. Richter, J. Kühn, C. A. Schalley and W. E. S. Unger, *J. Electron Spectrosc. Relat. Phenom.*, 2012, **185**, 85–89.
- I. F. Gallardo and L. J. Webb, *Langmuir*, 2010, **26**, 18959–18966.
- J. P. Collman, N. K. Devaraj and C. E. D. Chidsey, *Langmuir*, 2004, **20**, 1051–1053.
- D. Zacher, R. Schmid, C. Wöll and R. A. Fischer, *Angew. Chem., Int. Ed.*, 2011, **50**, 176–199.



- 36 J. Berná, D. A. Leigh, M. Lubomska, S. M. Mendoza, E. M. Pérez, P. Rudolf, G. Teobaldi and F. Zerbetto, *Nat. Mater.*, 2005, **4**, 704–710.
- 37 A. Ulman, *An Introduction to Ultrathin Organic Films-From Langmuir-Blodgettto Self-Assembly*, Academic Press, London, UK, 1991.
- 38 A. C. Fahrenbach, S. C. Warren, J. T. Incorvati, A.-J. Avestro, J. C. Barnes, J. F. Stoddart and B. A. Grzybowski, *Adv. Mater.*, 2013, **25**, 331–348.
- 39 J. Poppenberg, S. Richter, C. H.-H. Traulsen, E. Darlatt, B. Baytekin, T. Heinrich, P. M. Deutinger, K. Huth, W. E. S. Unger and C. A. Schalley, *Chem. Sci.*, 2013, **4**, 3131–3139.
- 40 S. Richter, C. H.-H. Traulsen, T. Heinrich, J. Poppenberg, C. Leppich, M. Holzweber, W. E. S. Unger and C. A. Schalley, *J. Phys. Chem. C*, 2013, **117**, 18980–18985.
- 41 H. C. Kolb, M. G. Finn and K. B. Sharpless, *Angew. Chem., Int. Ed.*, 2001, **40**, 2004–2021.
- 42 G. K. Such, J. F. Quinn, A. Quinn, E. Tjipto and F. Caruso, *J. Am. Chem. Soc.*, 2006, **128**, 9318–9319.
- 43 A. P. Upadhyay, D. K. Behara, G. P. Sharma, A. Bajpai, N. Sharac, R. Ragan, R. G. S. Pala and S. Sivakumar, *ACS Appl. Mater. Interfaces*, 2013, **5**, 9554–9562.
- 44 Z. Wang, J. Liu, H. K. Arslan, S. Grosjean, T. Hagendorn, H. Gliemann, S. Bräse and C. Wöll, *Langmuir*, 2013, **29**, 15958–15964.
- 45 B. Baytekin, S. S. Zhu, B. Brusilowskij, J. Illigen, J. Ranta, J. Huuskonen, L. Russo, K. Rissanen, L. Kaufmann and C. A. Schalley, *Chem.-Eur. J.*, 2008, **14**, 10012–10028.
- 46 C. Allott, H. Adams, C. A. Hunter, J. A. Thomas, P. L. Bernad Jr and C. Rotger, *Chem. Commun.*, 1998, 2449–2450.
- 47 E. V. Dzyuba, L. Kaufmann, N. L. Löw, A. K. Meyer, H. D. F. Winkler, K. Rissanen and C. A. Schalley, *Org. Lett.*, 2011, **13**, 4838–4841.
- 48 W. Jiang, H. D. F. Winkler and C. A. Schalley, *J. Am. Chem. Soc.*, 2008, **130**, 13852–13853.
- 49 Y. Zubavichus, M. Zharnikov, A. Schaporenko and M. Grunze, *J. Electron Spectrosc. Relat. Phenom.*, 2004, **134**, 25–33.
- 50 L. Strong and G. M. Whitesides, *Langmuir*, 1988, **4**, 546–558.
- 51 K. C. Prince, R. Richter, M. de Simone, M. Alagia and M. Coreno, *J. Phys. Chem. A*, 2003, **107**, 1955–1963.
- 52 D. J. Menzies, B. Cowie, C. Fong, J. S. Forsythe, T. R. Gengenbach, K. M. McLean, L. Puskar, M. Textor, L. Thomsen, M. Tobin and B. W. Muir, *Langmuir*, 2010, **26**, 13987–13994.
- 53 D. Brete, D. Przyrembel, C. Eickhoff, R. Carley, W. Freyer, K. Reuter, C. Gahl and M. Weinelt, *J. Phys.: Condens. Matter*, 2012, **24**, 394015.

