

MINIREVIEW

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Designing nanodiscs as versatile platforms for on-demand therapy

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Nowadays, there has been an increasing utilization of nanomedicines for disease treatment. Nanodiscs (NDs) have emerged as a novel platform technology that garners significant attention in biomedical research and drug discovery. NDs are nanoscale phospholipid bilayer discs capable of incorporating membrane proteins and lipids within a native-like environment. They are assembled using amphiphilic biomacromolecular materials, such as apolipoprotein A1 or membrane scaffold proteins (MSPs), peptides, and styrene–maleic acid polymers (SMAs). NDs possess well-defined sizes and shapes, offering a stable, homogeneous, and biologically relevant environment for studying membrane proteins and lipids. Their unique properties have made them highly desirable for diverse applications, including cancer immunotherapy, vaccine development, antibacterial and antiviral therapy, and treating Alzheimer's disease (AD) and diabetes-related conditions. This review discusses the classifications, advantages, and applications of NDs in disease therapy.

1. Introduction

Nanomaterials have made a significant impact as drug delivery carriers, with nanodiscs (NDs) emerging as a forefront class of nanoparticles (NPs) for both treatment and diagnosis of various ailments. NDs are synthetic phospholipid particles with a unique morphology and size, which heightens their effectiveness in drug delivery. Pioneered by Sligar *et al.* in the early 2000s, these model membrane systems have dimensions of roughly 10 nm in diameter and a thickness ranging from 4.6 to 5.6 nm.¹ NDs resemble high-density lipoprotein (HDL) in structure. HDL, with a diameter of approximately 10 nm,

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functions as a carrier for different types of lipids, such as cholesterol and triglycerides.² HDL is made up of a variety of apolipoproteins, the most important of which is apolipoprotein A-1 (ApoA-1); HDL is a good model for loading poorly soluble small molecules. For addressing the solubility challenge, scientists have modified ApoA-1 proteins, leading to an array of membrane scaffold proteins (MSPs) and peptides, which can then be assembled into NDs of similar size.³

A standout application of NDs is seen in EVOQ therapeutics' nanodisc technology. This advanced system is designed to target disease-specific antigens directly to lymph nodes, presenting a promising avenue for autoimmune disease treatments. Notably, Amgen and subsequently Gilead Sciences have recognized the value of nanodiscs and invested \$660 million on January 3, 2023, for the development of novel therapies targeting autoimmune diseases like systemic lupus erythematosus and rheumatoid arthritis; this demonstrates the significant potential of NDs in disease treatment. This review summarizes the classifications and benefits of NDs and introduces their applications in disease treatment.

2. Classification of NDs

Since the FDA approved the first nanoparticles, known as INFeD (Sanofi Aventis, Allergan), for clinical use in 1974, there has been a steadily increasing interest in exploring nanomedicines as a potential treatment for various diseases.⁴ Research findings indicate that NPs can be broadly categorized into three main types: polymer-based nanoparticles, lipid-based nanoparticles, and inorganic particles (Fig. 1).⁵ Among these, lipid-based nanoparticles composed of lipids have gained significant attention for their ability to deliver hydrophobic drugs and nucleic acids, which results in prolonged circulation and enhanced drug delivery capabilities.^{6–8} Polymer-based nanoparticles, assembled using amphiphilic copolymers such as polymersomes, micelles, and dendrimers, which are typically formed with materials like

polyethylene glycol (PEG) and polydimethylsiloxane (PDMS), offer improved stability and cargo-retention efficiency, making them well-suited for drug delivery.^{9,10} Dendrimers, formed from charged polymers such as polyethylenimine (PEI) and polyamidoamine (PAMAM), have applications as theranostic and contrast agents.^{11,12} Inorganic particles, created from materials such as gold, iron, and silica, exhibit unique physical, magnetic, and optical properties and have been extensively studied for diagnostics, imaging, and photothermal therapy.^{13–16} Each type of NP possesses distinct advantages for disease therapy, but they also come with notable drawbacks.

Nanoparticles typically range in size from 1 to 100 nm, making them susceptible to uptake by the liver and spleen. However, non-spherical NPs offer advantages over their spherical counterparts, including an extended blood circulation half-life and a higher cellular endocytosis rate. In recent years, significant attention has been directed toward the development of NDs, which are non-spherical NPs coated with amphiphilic molecules like proteins, synthetic polymers, or short-chain lipids. NDs draw inspiration from the transportation of lipid nanocarriers in the human bloodstream. Among these nanocarriers, high-density lipoprotein (HDL) plays a pivotal role in the transport and metabolism of lipids, particularly cholesterol and triglycerides.¹⁷ NDs can efficiently carry various substances, including drugs, nucleic acids, signal-emitting molecules, or dyes, making them highly effective nanocarriers for therapeutic applications or medical diagnostics.¹⁸ This versatility and capability make NDs promising tools in the field of nanomedicine.

NDs are considered ideal nanoformulations due to their enhanced anisotropic characteristics and multifunctional functionalization potential. Based on their composition (as illustrated in Fig. 2), NDs can be classified into four categories:¹⁹

(1) Lipid nanodiscs (LNDs). LNDs consist of amphiphilic biomacromolecules such as membrane scaffold proteins (MSPs),²⁰ apolipoprotein A1 (ApoA-1),²¹ and peptides modified according to ApoA-1 (Fig. 2A).²²



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Fig. 1 Classification of NPs. (A) Polymer-based nanoparticles assembled using amphiphilic copolymers such as polymersomes, micelles, and dendrimers. (B) Lipid-based nanoparticles. (C) Inorganic particles: gold NPs, mesoporous NPs, and silicon NPs.

(2) Styrene–maleic acid (SMA) lipid nanoparticles (SMALPs) and novel SMALPs. SMALPs are formed through the copolymerization of styrene and maleic anhydride monomers. They are the hydrolyzed form of styrene–maleic anhydride (SMAnh) copolymers (Fig. 2B).²³

(3) Polymer nanodiscs (PNDs). PNDs are assembled from MSPs or other biomacromolecules in the presence of amphiphilic block copolymers such as the bacterial ATP-binding cassette (ABC) transporter MsbA (Fig. 2C).^{24–26}

(4) Hybrid nanodiscs (HNDs). HNDs encompass hybrid bicelles (HBs), edge-modified hybrid bicellars (EHBs), plane-modified hybrid bicellars (PHBs), and similar structures. HBs are an extension of bicelles and are composed of a long-chain cerasome-forming lipid and a short-chain phospholipid. The

edge- and plane-modified hybrid bicellar nanodiscs are referred to as EHB and PHB, respectively (Fig. 2D).²⁷

3. Advantages of NDs

NDs present numerous advantages over other nanoparticle platforms, such as uncomplicated formulation, consistent reproducibility, enhanced stability, and superior biocompatibility. When employed *in vivo*, NDs demonstrate remarkable pharmacokinetics and biological distribution, allowing them to exhibit heightened specificity to target tissues and cellular receptors. This specificity enables more accurate drug release or lipid exchange. As a multifaceted platform, NDs have a broad spectrum of potential applications. To sum up, the benefits of NDs significantly overshadow those of traditional NPs.

3.1 Size control and therapeutic targeting

One of the primary attributes of NDs is their uniform and adjustable size, which plays a crucial role in therapeutic targeting. NDs can be fabricated from diverse materials, resulting in varying sizes. For instance, NDs offer precise size control by leveraging specific membrane scaffold protein (MSP) structures, usually from 10 nm to 20 nm. In recent years, the development of cyclized MSP proteins (cMSP) has made it possible to obtain covalently circularized NDs (cNDs) with a size larger than 80 nm, which have better thermal stability and long-term stability; they can load drugs and membrane proteins more efficiently and achieve better targeted drug delivery, and achieve better applications in structural analysis.^{28–30} Assembled with styrene–maleic acid (SMA), NDs typically possess a size of around 20 nm—ideal for drug delivery and transporting small molecular probes across the blood–brain barrier (BBB).^{31,32} Additionally, strategic ligand modifications,



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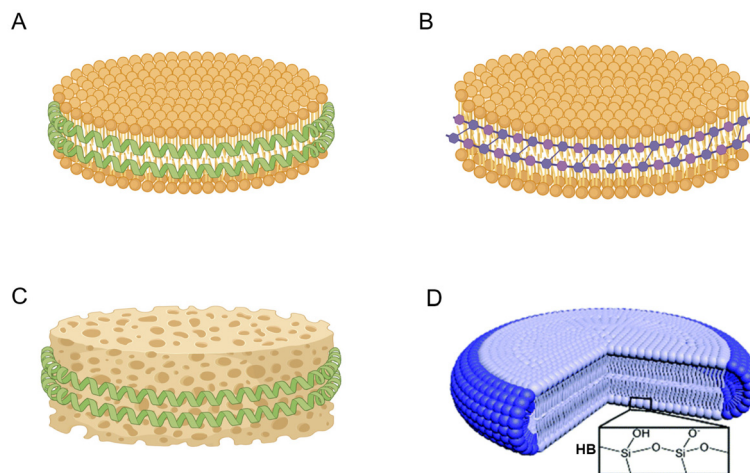


Fig. 2 Different forms of NDs. (A) Lipid nanodiscs (LNDs) consist of amphiphilic biomacromolecules. (B) Styrene-maleic acid (SMA) lipid nanoparticles (SMALPs) formed through the copolymerization of styrene and maleic anhydride monomers. (C) Polymer nanodiscs (PNDs) assembled from MSPs or other biomacromolecules in the presence of amphiphilic block copolymers. (D) Hybrid nanodiscs (HNDs) composed of a long-chain cerasome-forming lipid and a short-chain phospholipid.²⁷

such as planar and edge adjustments on NDs, can amplify the anisotropic effect, thereby bolstering targeting efficiency.³³

In recent studies, Zhang's team, in 2023, innovatively used bacterial outer membranes to design cell-based nanodiscs (OM-NDs) as an antibacterial vaccination platform. Using *Pseudomonas aeruginosa* as a model pathogen, they demonstrated that OM-NDs efficiently reached the lymph nodes *in vivo* due to their petite size. These NDs also interacted effectively with antigen-presenting cells (APCs), which sped up uptake and intensified immune responses. Such findings underscore NDs' ideal sizing and adaptability for various material loadings.³⁴ In 2017, Zhang Qiang's group delved into how nanomaterials' biological attributes could be modulated by altering their shape and ligand modifications. They designed and engineered four types of nanoparticles, including hybrid nanospheres, NDs, and NDs modified with octahedral arginine or edge or planar modification (R8) sequences. The researchers discovered that the anisotropy induced by R8 modification significantly impacted endocytosis, intracellular transport, and even tissue penetration of nanoparticles. The maximum increase in cell uptake, comparing the planar modification to marginal modification of R8, reached up to 17-fold. The team also compared the cell uptake differences across six different cell lines. Their findings revealed that shape anisotropy and ligand alterations profoundly influence the endocytosis, internal transport, and tissue penetration capacities of nanoparticles.²⁷ NDs, with their capacity for fine-tuned size adjustment and enhanced targeting, have emerged as potent assets in oncological research and treatments. These unique characteristics of NDs hold great promise for evolving imaging probes and delivery mechanisms for chemotherapeutics, vaccines, and anti-cancer agents.

3.2 Binding affinity and adhesion

Compared to spherical NPs, disk-shaped NDs offer distinctive perks. Their expansive surface area leads to prolonged circula-

tion, augmented cellular intake, and refined biological distribution. Additionally, NDs' flattened geometry instigates a torque within blood flow, propelling them to rotate and roll. This dynamism amplifies their proximity to vessel walls, reinforcing vascular adherence.^{35,36}

Recent advancements have highlighted challenges in systemically delivering STING agonists to tumors. However, a marked improvement in tumor penetration was observed when STING-activated cyclic dinucleotides (CDNs) were integrated into NDs for intravenous administration. This approach ensures that most tumor cells are exposed to STING agonists.³⁷ Likewise, another investigation showed that disc-shaped nanocarriers decorated with *N*-methylated glutamate (CR(NMe)EKA) and loaded with Cys-Arg-Glu-Lys-Ala exhibited superior tumor targeting and penetration compared to spherical nanoparticles. This enhanced delivery system effectively transported tumor necrosis factor-associated apoptosis-inducing ligand (TRAIL) and paclitaxel (PTX) to lung melanoma tumor sites.³⁸

3.3 Biocompatibility and biosafety

The foundation of NDs lies in predominantly biocompatible constituents. For example, the primary elements that comprise LNDs or PNDs are modifications of the human ApoA-1 protein, like membrane scaffold proteins (MSPs) or peptides. Studies have consistently vouched for the commendable pharmacokinetic traits and biocompatibility of LNDs and PNDs formulated with MSPs or ApoA-1 analogs.^{39,40} SMA, an amphiphilic biomacromolecule utilized in assembling SMALPs, has been extensively investigated for its biosafety. Furthermore, innovations are underway to create sturdier SMA copolymers that remain stable under acidic conditions or amid polyvalent ions.⁴¹ Silica, a key component in the construction of HNDs, is classified as "generally considered safe" by the US Food and Drug Administration (ID Code: 14808-60-7).³³ These biosafe materials and their metabolic pathways contribute to the

overall biocompatibility of NDs, making them a promising platform for various biomedical applications.

The comprehensive benefits of NDs encompass size adaptability, targeting proficiency, biocompatibility, and seamless drug modification potential. Although considerable efforts have been made in preclinical research, a significant portion of ND-related studies remains in the validation phase, awaiting concrete clinical data. To harness NDs' full potential in clinical contexts, continued research, validation, and clinical trials are imperative. As ND research continues to evolve, it is pivotal to rigorously assess NDs through clinical examinations to confirm their efficacy and safety, ensuring that they become a trustworthy instrument in clinical applications.

4. Application of NDs in disease therapy

NDs present a remarkably versatile molecular delivery platform, amenable to extensive modifications that enhance disease targeting and enable personalized tumor treatments. Numerous studies have underscored the utility of NDs in disease therapy, illuminating their remarkable versatility and untapped potential.^{42–47} The unique disc-shaped structure of NDs allows for efficient loading of poorly soluble drugs, thereby improving their therapeutic efficacy. Furthermore, NDs can be modified with vaccine adjuvants or anti-tumor drugs to facilitate tumor immunotherapy.¹⁹ Functional small molecules can also be loaded onto NDs to investigate transmembrane signaling pathways and employ visualization tracing techniques. In subsequent discussions, we will delve into the specific applications of NDs in disease therapy, providing comprehensive insights into their therapeutic potential and highlighting their contributions to advancing medical research and personalized medicine.

4.1 Cancer immunotherapy

Cancer immunotherapy faces two formidable challenges: the insufficient infiltration of CD8⁺ T cells and the overexpression of programmed cell death protein 1 (PD-1) on the surfaces of activated T cells.⁴⁸ Extensive research has focused on small-molecule drugs targeting the stimulator of interferon genes (STING), immune checkpoints, and toll-like receptors (TLR) as potential tumor therapy agents. Although these drugs exhibit promise against various cancers, their efficacy is hampered by the daunting barriers posed by tumor heterogeneity, the immunosuppressive tumor microenvironment (TME), and the blood–brain barrier.^{44,49,50} Thanks to their unique advantages, NDs have emerged as a promising platform for cancer immunotherapy. One approach involves utilizing NDs to deliver tumor antigens to dendritic cells, specialized immune cells that can activate T cells to recognize and eliminate cancer cells. NDs can be engineered with tumor antigens and adjuvants, such as toll-like receptor agonists, to enhance the immune response. By delivering these immunogenic components to dendritic cells, NDs facilitate the generation of a

robust anti-tumor immune response.^{51–53} The advantages of employing NDs in cancer immunotherapy are manifold. Firstly, the disk-shaped structure of NDs provides a larger surface area for loading tumor antigens and adjuvants, enabling efficient delivery to dendritic cells. Secondly, the size and surface modification of NDs can be fine-tuned with precision to augment cellular uptake and activate immune cells. Thirdly, NDs can protect the encapsulated antigens and adjuvants from degradation and premature release, ensuring their effective delivery to the target cells. By harnessing NDs' capabilities, researchers aim to overcome the limitations of current cancer immunotherapy approaches and achieve improved therapeutic outcomes. Ongoing research endeavors are imperative to optimize ND-based immunotherapeutic strategies and validate their efficacy in preclinical and clinical settings.

Targeting the STING signaling pathway with innate immune stimuli has demonstrated potential in bolstering anti-tumor immunity. However, the effective delivery of STING agonists directly to tumors remains challenging. To tackle this, researchers have conjugated STING agonists, specifically cyclic dinucleotides (CDNs), with PEGylated lipids, creating CDN-PEG-lipids. When these are incorporated into LNDs, the resultant LNDs (as depicted in Fig. 3A) have shown improved tumor penetration upon intravenous administration. This led to improved co-localization of CDNs and tumor antigens in dendritic cells, increased uptake of LND-CDNs by cancer cells, and enhanced T cell activation.^{44,54} As carriers, LNDs ensure efficient compound delivery within solid tumors, reinforcing their potential to augment immunotherapy. Integrating chemotherapy drugs with immunotherapies has shown a promising synergy in immune activation. For instance, the commonly employed chemotherapy agent, adriamycin (DOX), has been channeled through NDs as delivery vehicles. This modality triggers immunogenic cell death in malignant cells, exerting an anti-tumor effect (Fig. 3B).⁴⁶ Another notable study focused on NDs for chemoimmunotherapy targeting glioblastoma (GBM), wherein the vector, packed with docetaxel (DTX) and CpG (a toll-like receptor 9 agonist), efficiently delivered the chemotherapy drug to the GBM microenvironment, causing notable tumor regression (Fig. 3C).⁵⁰

Highly heterogeneous tumors represent a formidable obstacle to targeted therapies. Within this context, photothermal therapy (PTT) has emerged as a promising treatment modality. Ye's group utilized phage display technology to identify an osteosarcoma (OS) targeting peptide (OTP). Subsequently, they ingeniously integrated OTP and Cy7-TCF (2-dicyanomethylene-3-cyano-4,5,5-trimethyl-2,5-dihydrofuran, TCF) into NDs, resulting in the creation of T-NDs (illustrated in Fig. 3D) tailored for heterogeneous tumors. Experimental results demonstrated that these specialized NDs effectively enhanced photothermal tumor ablation under near-infrared light and prolonged tumor retention.⁵⁵

These studies highlight the potential of ND-based cancer therapy as a novel and adaptable approach for priming anti-tumor immunity and enhancing tumor sensitivity to immune

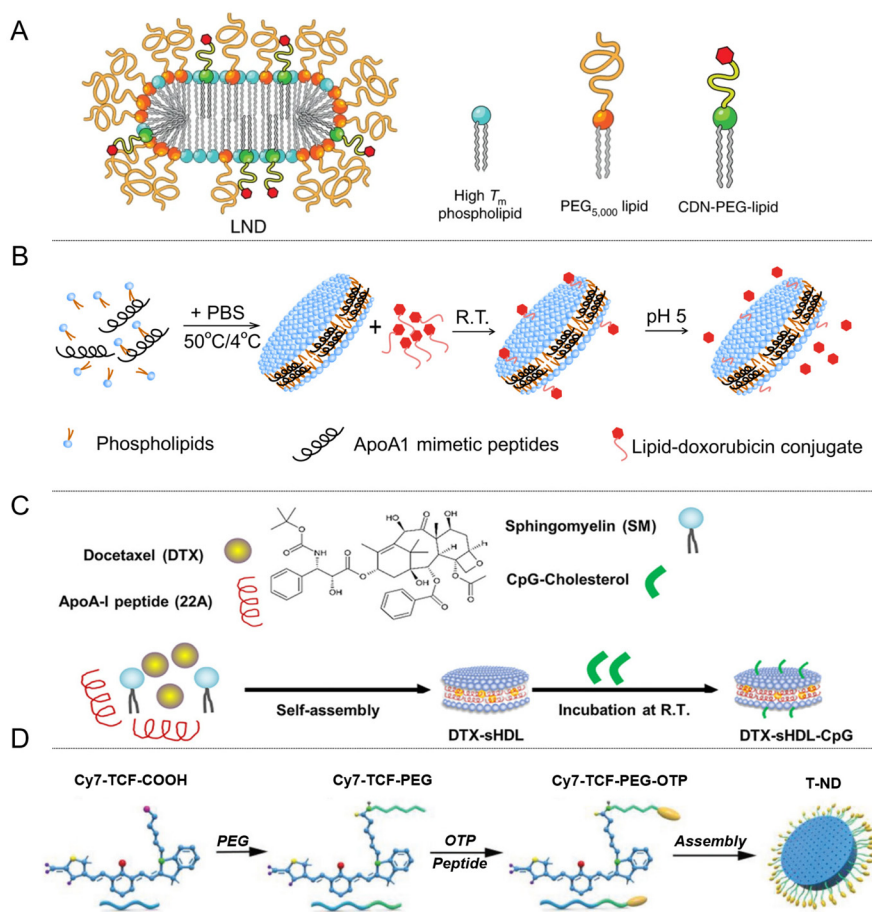


Fig. 3 (A) LNDs conjugated with STING agonists, CDNs and PEGylated lipids to improve their delivery. (B) The chemotherapy drug DOX, which is widely used, delivered using NDs as carriers. (C) DTX-sHDL-CpG loaded with DTX and CpG, a toll-like receptor 9 (TLR9) agonist. (D) T-NDs modified with OTP and Cy7-TCF (2-dicyanomethylene-3-cyano-4,5,5-trimethyl-2,5-dihydrofuran, TCF).^{44,46,50,55}

checkpoint blockade. This provides a theoretical foundation for the precise treatment of highly heterogeneous tumors. By leveraging NDs' unique targeting capabilities and photothermal properties, researchers aim to develop innovative strategies for combating complex and diverse tumor types. Further exploration and validation of these approaches are essential for advancing the field of precision medicine in the treatment of heterogeneous tumors.

4.2 Vaccine development

Based on soluble antigens paired with classic adjuvants, conventional cancer vaccines have demonstrated limited performance in clinical settings. This underscores the imperative need for innovative vaccine technologies to stimulate robust T cell responses for effective antitumor activity. To date, many reports have been published on using nanovesicles to integrate nanovaccines, which can achieve personalized tumor immunotherapy.^{56,57} Whether sourced from diverse membranes or loaded with antigenic peptides and tumor markers, NDs present a rich tapestry of antigenic features and inherent immunogenic attributes. They stand out as promising candi-

dates for devising vaccines for tumor immunotherapy and countering infectious diseases.

Extracting membrane proteins directly from the natural membrane environment to assemble NDs has great potential and can well retain the structure and activity of membrane proteins. In 2018, Tsyr-Yan Yu's group extracted bacteriorhodopsin from the natural membrane environment and assembled it into a covalently circularized nanodisc; the natural structure and activity of the protein are well retained.⁵⁸ Zhang's group has successfully constructed a cellular ND, harnessing the bacterial outer membrane of *Pseudomonas aeruginosa* (OM-NDs) (Fig. 4A) as a platform for antimicrobial vaccine. Owing to their diminutive size, these NDs can be efficiently delivered to lymph nodes, forging effective interactions with antigen-presenting cells, resulting in accelerated uptake and improved immune stimulation. This approach has demonstrated the ability to elicit strong humoral and cellular immune responses against *Pseudomonas aeruginosa*, as confirmed in a mouse model of pneumonia.³⁴ Furthermore, NDs offer the advantage of being loaded with antigenic peptides or specific tumor markers, enhancing their immunogenicity. Studies conducted in mice and nonhuman primates have validated the efficacy

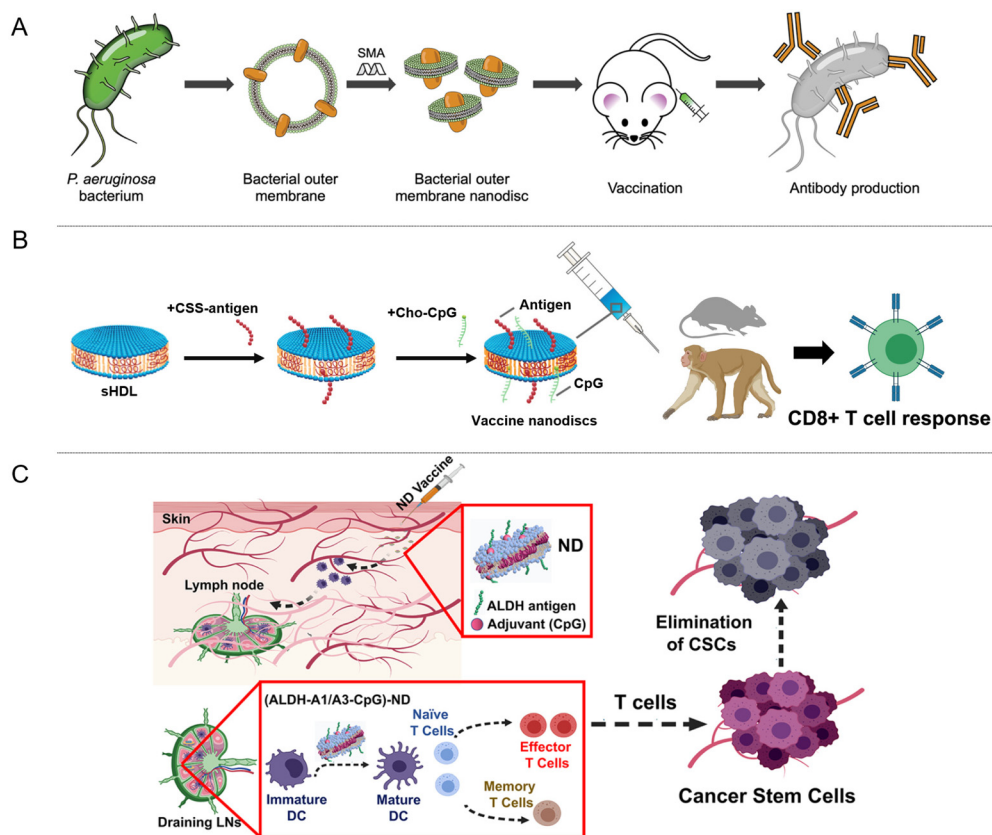


Fig. 4 (A) OM-ND formulation used as a nanovaccine to protect against bacterial infection. (B) Vaccination of NDs loaded with antigenic peptides or specific tumor markers conducted in mice and nonhuman primates has validated the efficacy and safety. (C) Illustration of ND vaccination against ALDH^{high} CSCs, ALDH-A1/A3-CpG-NDs facilitate antigen transport to lymph nodes and induce robust ALDH-specific T cell responses.^{34,45,60,61}

and safety of NDs loaded with antigenic peptides (Fig. 4B).^{49,59–61} Aldehyde dehydrogenase (ALDH) has been widely utilized as a marker for isolating cancer stem cells (CSCs), which proliferate extensively and drive tumor metastasis and recurrence and have been demonstrated to be present in more than 20 tumor types to date.^{62,63} James J. Moon's group developed synthetic NDs for vaccination against ALDH^{high} CSCs. NDs facilitate antigen transport to lymph nodes and induce robust ALDH-specific T cell responses, presenting a renewed avenue for cancer immunotherapy aimed at CSCs (Fig. 4C).⁴⁵

Furthermore, there have been studies on loading the HIV envelope glycoprotein (Env) membrane protein onto NDs. These studies have successfully resolved the complex structure formed by Env and the membrane-proximal external region (MPER)-targeting antibody 10E8. Through the exploration of the molecular mechanism of the Env-MPER antibody complex, it was discovered that the tilt of Env plays a crucial role in the neutralization mechanism of MPER-targeted antibodies.⁶⁴ A work studied the structure of MPER NDs combined with 4E10, and also found that membrane fusion was abolished by changing MPER dynamics and modifying average ectodomain tilt, providing information for future vaccine development.⁶⁵ In another research study, MPER-TMD (MPER and the adjacent

transmembrane domain) was assembled into NDs, and the structure of MPER-TMD and its mechanism of action combined with 10E8 were explored to provide a structural basis for vaccine design.⁶⁶ These findings provide a solid structural foundation for developing vaccines based on the ND platform, with the potential to advance HIV immunotherapy.

4.3 Antibacterial and antiviral therapy

NDs have emerged as promising platforms for antimicrobial and antiviral therapies, aiming to enhance the targeting and eradication of bacterial and viral pathogens. The utilization of NDs for delivering antimicrobial peptides or drugs offers several benefits. Firstly, it protects antibacterial drugs from degradation, ensuring their stability and prolonging their effectiveness. Additionally, NDs facilitate the efficient uptake of these agents by pathogens, thereby improving their potency and selectivity. In the context of antiviral therapy, NDs can display viral or bacterial antigens, triggering robust and specific immune responses against targeted pathogens. This immune stimulation is critical in combating viral infections. To further optimize therapy, NDs can be functionalized with antigens, adjuvants, and targeting ligands such as antibodies or peptides. These modifications enhance the immunogenicity

and specificity of the therapeutic approach, ultimately improving its overall efficacy.

The misuse and overuse of antibiotics have contributed to the emergence of drug resistance during treating bacterial infections. Additionally, inflammation poses another significant challenge in antimicrobial therapy. Infections often trigger the recruitment of inflammatory cells, producing high levels of reactive oxygen species (ROS), which can cause oxidative damage to healthy tissues.⁶⁷ Most antimicrobial agents currently available lack anti-inflammatory properties, highlighting the need to develop new therapeutic platforms capable of selectively targeting microbes while also possessing anti-inflammatory activity. Although non-antibiotic drugs, such as cationic peptides or metal-based nanomaterials, have been explored as antimicrobial agents, they often suffer from poor selectivity and may induce cytotoxicity in normal human somatic cells, leading to severe adverse effects.⁶⁸ However, recent research has demonstrated the tremendous potential of Bi₂Se₃-NDs in treating bacterial infections. Bi₂Se₃-NDs exhibit several desirable properties. They effectively bind to lipoteichoic acid found in Gram-positive bacteria, physically disrupting the bacterial wall/membrane. Additionally, these nanosheets possess intrinsic antioxidant activity, allowing them to scavenge ROS generated by macrophages during bacterial infection. Importantly, Bi₂Se₃-NDs (Fig. 5A) demonstrate low off-target side effects and are effective against drug-resistant bacteria. They also exhibit high stability and biocompatibility, opening up new possibilities for developing non-antibiotic drugs.⁶⁹ In addition, another study also reported that Bi₂Se₃-NDs can clear intestinal reactive oxygen and nitrogen species (RONS), effectively reduce intestinal inflammation, and have a good therapeutic effect in the treatment of inflammatory bowel disease.⁷⁰

Antiviral drugs that target the viral membrane offer the advantage of creating a barrier against drug resistance and exhibiting broad-spectrum activity across different viruses, irrespective of their specific viral membranes. However, there is a current lack of antiviral drugs that selectively target and destroy viral membranes, inhibiting a wide range of enveloped viruses.^{71–73} In a significant development, Dae-Hyuk Kwon has pioneered using NDs containing a decoy virus receptor to

inhibit viral infections. The antiviral activity of these NDs is enhanced when a viral receptor, such as sialic acid, is incorporated into the nanoparticle. This incorporation allows the NDs to disrupt the viral envelope within endosomes, particularly at low pH levels. Consequently, the viral RNA becomes trapped within endolysosomes, undergoing enzymatic hydrolysis. In experimental mouse models, NDs effectively inhibit influenza virus infection, reducing morbidity and mortality without causing noticeable toxicity (Fig. 5B).⁷⁴

Antimicrobial peptides (AMPs) play a crucial role in the innate immune system of animals, plants, fungi, and prokaryotes. These cationic and amphiphilic peptides directly target bacterial membranes, causing damage and compromising their integrity. AMPs have shown promising prospects in combating antibiotic-resistant infections. In a study conducted by Michael T. Marty *et al.*, the interaction between different lipid bilayer NDs and AMP-lipids was investigated to gain insights into the potential mechanism of action of AMPs.⁷⁵ Furthermore, Tomohiro Imura *et al.* isolated an antimicrobial peptide called PXT-5 and its modified version, modified PXT-5, from the skin of diploid *Xenopus tropicalis*. Through experiments, it was demonstrated that these antimicrobial peptides could be efficiently incorporated into NDs, highlighting the potential for their application in antimicrobial therapies.⁷⁶

In conclusion, NDs offer substantial potential as versatile platforms for antibacterial and antiviral therapies. Their capability to deliver antimicrobial agents, induce specific immune responses, and be modified with various components significantly enhances the immunogenicity, specificity, and effectiveness of these therapeutic approaches. The studies mentioned above contribute to our understanding of the interaction between NDs and AMPs, furthering the development of innovative antimicrobial strategies.

4.4 Treatment of other diseases

4.4.1 Alzheimer's disease (type III diabetes). Alzheimer's disease (AD), a prevalent neurodegenerative disease, currently lacks effective treatment options, recent clinical and basic research has reported that AD and diabetes share common pathophysiological changes and signaling pathways, and the

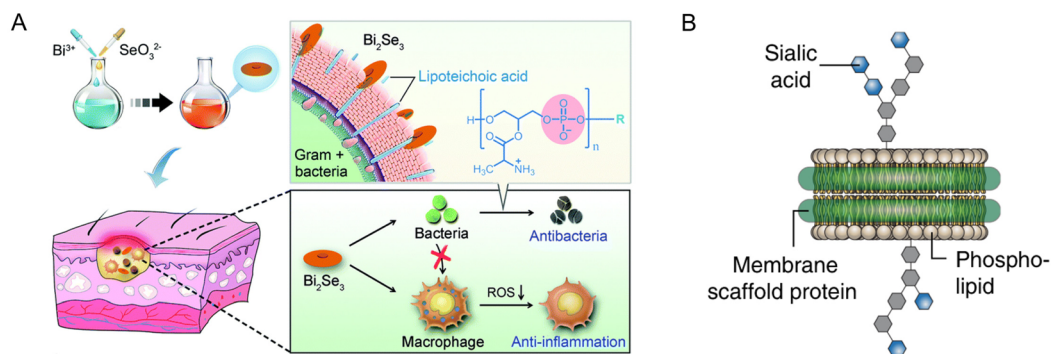


Fig. 5 (A) Illustration of the preparation of Bi₂Se₃-NDs and the combined antibacterial and anti-inflammatory therapy of bacterial infections. (B) The total ganglioside-embedded nanodisc (NDTG) contains a decoy virus receptor to inhibit viral infections.^{69,74}

term “type-3-diabetes” for AD has been proposed.^{77,78} Two common hypotheses for AD pathogenesis are the amyloid hypothesis and the acetylcholine hypothesis. Research indicates that in late-onset AD, there is a global impairment of amyloid-beta ($A\beta$) clearance while $A\beta$ production remains unimpaired.^{79,80} The imbalance between $A\beta$ production and clearance leads to the abnormal aggregation of $A\beta$, resulting in the formation of amyloid fibers and plaques.⁸¹ These plaques exhibit neurotoxicity, trigger neurite damage, and elicit proinflammatory responses.^{80,82,83} Additionally, studies have revealed a link between AD pathogenesis and the dysfunction of acetylcholinesterase (AChE). AChE-induced imbalanced hydrolysis of acetylcholine can accelerate $A\beta$ deposition and contribute to cognitive decline associated with inflammation.^{84,85} Hence, developing immunotherapies and acetylcholinesterase inhibitors to enhance $A\beta$ clearance is considered a promising treatment strategy for AD.⁸⁶ Nanotechnology provides a valuable approach for targeted therapy against $A\beta$ by serving as an efficient drug delivery system. Curcumin-modified nanoliposomes and lipid nanocarriers based on phosphatidic acid and cardiolipin have demonstrated a high affinity for $A\beta$ and its polymers, offering potential for AD treatment.^{87–89} However, most nanocarriers face challenges penetrating the blood–brain barrier (BBB). Thus, developing nanocarriers with BBB permeability and

strong $A\beta$ targeting capabilities is essential. In this context, NDs have shown remarkable advantages.

In a study conducted by Gao's group in 2015, a nanostructure called monosialotetrahexosylganglioside (GM1)-modified recombinant high-density lipoprotein (GM1-rHDL) was reported (Fig. 6A). This nanostructure exhibited a high antibody-like affinity to $A\beta$ and facilitated the degradation of $A\beta$ by microglia and its efflux across the BBB. When administered intranasally, NDs demonstrated efficient distribution in the brain. The researchers loaded a neuroprotective peptide called NAP onto NDs, resulting in RNAP-GM1-rHDL. This formulation exhibited enhanced neuroprotection, reduced $A\beta$ deposition, and improved neurological changes in AD model mice.³¹ In 2016, Gao's group developed a biomimetic nanocarrier called APOE-reconstituted high-density lipoprotein nanocarrier (ANC). Confocal microscopy studies demonstrated that ANC could cross cerebral blood vessels and accumulate around $A\beta$ aggregates. Subsequently, the researchers loaded α -mangogenin (α -M) onto ANC, forming ANC- α -M (Fig. 6B). α -M is a polyphenolic drug that inhibits the formation of $A\beta$ oligomers and fibers, and enhances the cellular degradation of $A\beta$. ANC- α -M exhibited a strong affinity for $A\beta$ and effectively reduced amyloid deposition, decreased microglia proliferation, and improved memory deficits in SAMP8 mice.³² The BBB poses a challenge in delivering therapeutic and diagnostic



Fig. 6 (A) The assembly process of GM1-rHDL, GM1-rHDL exhibited a high antibody-like affinity to $A\beta$ and facilitated the degradation of $A\beta$. (B) The assembly process of ANC- α -M, which can enhance $A\beta$ uptake and degradation in mouse models.^{31,32}

drugs to the central nervous system (CNS). However, NDs with high BBB permeability and a solid binding affinity for A β have been developed, offering a promising platform for drug delivery in AD. These advancements hold the potential for effective treatment strategies for AD.

4.4.2 Diabetes-related diseases. Diabetes-related diseases, such as diabetic nephropathy (DN), have been extensively studied due to their high prevalence and impact on renal function. Epidemiological studies have identified DN as the leading cause of end-stage renal disease.⁹⁰ The development of DN is associated with lipid metabolism disorders, characterized by elevated levels of very low-density lipoprotein (VLDL), low-density lipoprotein (LDL), and triglycerides (TG), as well as decreased levels of high-density lipoprotein.^{91,92} Impaired renal cholesterol efflux has also been implicated in the risk of DN.^{93,94} LXRs (liver X receptors) are intracellular cholesterol sensors and their agonists play a crucial role in activating cholesterol transporters.⁹⁵ LXR agonists have shown renal protective effects in DN by inhibiting mesangial dilation and reducing excessive lipid accumulation in macrophages within kidney tissues. This, in turn, leads to the suppression of various signaling pathways and a decrease in pro-inflammatory cytokine levels such as IL-6 and TNF- α .^{96,97} However, the clinical application of LXR agonists is hindered by their liver side effects, which result in excessive triglyceride secretion into the systemic circulation.⁹⁸ NP-based drug delivery systems offer a promising approach to overcome these challenges. By utilizing NPs, drug half-life can be extended and drug efficacy and targeting can be improved.^{99,100} Specifically, in the context of DN, nanoparticles with sizes ranging from 10 to 50 nanometers have been developed to facilitate targeted treatment. These nanoparticles can readily enter the kidney through the initial barrier of kidney filtration and deliver drugs directly to the disease site, thereby avoiding excretion through urine.^{17,101,102}

Anna Schwendeman's research group has significantly advanced in developing a renal-targeted therapy strategy for DN. They have modified synthetic NDs by incorporating a renal targeting ligand called KT peptide and encapsulating LXR agonists within the NDs. This approach aims to specifically target the kidneys and deliver LXR agonists to address the underlying mechanisms of DN. The LXR agonists delivered by NDs play a crucial role in promoting the removal of excess

lipids from mesangial cells, improving inflammation, and restoring normal kidney function. In experimental studies, the data demonstrated that the modified NDs, known as KT-sHDL/TO, effectively facilitated cholesterol efflux and inhibited mesangial cell proliferation. Moreover, these NDs exhibited significant therapeutic effects in a DN mouse model, showcasing their potential for future clinical translation in the treatment of DN. The research conducted by Anna Schwendeman's group provides promising insights into the development of targeted therapies for DN, offering new avenues for managing this prevalent complication of diabetes (Fig. 7A).¹⁰²

Therapeutic approaches utilizing endogenous stem cell mobilizers have shown promise in providing effective cell-free therapy for various ischemic diseases. Substance P (SP), a peptide consisting of 11 sequences, has been identified as an effective endogenous mobilizer for mesenchymal stem cells (MSCs) and endothelial progenitor cells (EPCs), leading to wound healing and angiogenesis induction. It has been considered a potential treatment for severe ischemic diseases, including diabetic hind limb ischemia, ulcers, and wounds, without needing *in vitro* cell culture and expansion.^{103,104} However, the SP peptide has a short half-life and is rapidly degraded in the body. Diabetic patients, in particular, experience increased levels of neutral endopeptidase (NEP), an enzyme responsible for degrading SP. This results in a half-life of SP of less than 1 minute in diabetic mice, limiting its therapeutic potential.¹⁰⁵ To overcome this challenge, Seung-Woo Cho's group has developed HDL-SP-simulated NDs. By combining SP with HDL NDs, the *in vitro* and *in vivo* half-life of SP is significantly extended, showing an over thousand-fold increase. This innovative approach improves the retention of SP in the bone marrow, leading to enhanced angiogenesis and restoration of blood perfusion in a diabetic limb ischemia model. As a result, limb retention is significantly improved, offering a new and promising strategy for treating diabetic ischemia (Fig. 7B).¹⁰⁶

Indeed, studies have demonstrated that NDs are highly efficient drug carriers with several advantageous properties. One notable feature is their ability to enhance the stability and prolong the half-life of unstable polypeptide or protein drugs. Encapsulating these fragile drugs within NDs can reduce their susceptibility to degradation, thereby improving their thera-



Fig. 7 (A) KT-sHDL/TO loaded with a renal targeting ligand called KT peptide and LXR agonists, which can facilitate cholesterol efflux and inhibit mesangial cell proliferation. (B) Schematic illustration of HDL-SP NDs formulation, NDs extend the half-life of SP and improve the retention of SP in the bone marrow.^{102,106}

peutic potential. One of the key advantages of NDs is their ultra-small size, typically around 10 nanometers. This nanoscale dimension allows them to traverse biological barriers and reach target tissues more effectively. Additionally, NDs exhibit high *in vivo* tolerance, meaning that they are well tolerated by the body and do not elicit significant adverse effects. This is crucial for their safe and efficient application as drug carriers. Furthermore, NDs possess a prolonged cycle half-life, allowing them to circulate in the body for an extended period. This extended circulation time increases the chances of drug delivery to target sites, enhancing the efficacy of the treatment.^{17,107} NDs have demonstrated remarkable potential as drug carriers due to their small size, biocompatibility, prolonged circulation, and high targeting capabilities. These properties make them efficient and versatile platforms for delivering various types of drugs to target tissues, thereby enhancing drug stability, improving pharmacokinetics, and optimizing therapeutic outcomes.

4.4.3 Treatment of retina. Age-related macular degeneration (AMD) is the leading cause of vision loss in developed countries. Research has demonstrated that nutritional supplementation, including vitamins C and E, β -carotene, lutein, and zeaxanthin, can slow the progression of AMD and reduce the rate of vision loss.¹⁰⁸ Lutein, an essential nutrient for eye health, cannot be synthesized by the human body and must be obtained from the diet. However, lutein's poor water solubility challenges its oral bioavailability.^{109,110} NDs have successfully loaded poorly soluble drugs such as polyphenols, curcumin, and antibiotics.^{111–113} Robert O. Ryan *et al.* assembled lutein into NDs to protect photoreceptor cells from light-induced damage.¹¹⁴ Additionally, some studies have explored the development of photosensitive NDs and aimed to create a retinal simulation platform for researching vision restoration treatments.¹¹⁵

4.5 Novel biotechnology

Nanotechnology has been extensively explored for drug delivery and various emerging biological technologies.^{116–119} Yet, it is essential to note that traditional liposome-based drug delivery systems have intrinsic drawbacks, such as brief circulation duration, suboptimal drug encapsulation, and instability in biological media.¹²⁰ NDs, have emerged as a solution, offering promise not only in disease treatment but also in a range of other innovative biological applications.^{121,122}

4.5.1 Imaging technique. Albumin, as a crucial plasma carrier and a major soluble protein involved in nutrient transport and osmotic pressure regulation, plays a significant role in various biological processes. Bovine serum albumin (BSA) is often utilized as a model protein in research studies due to its structural similarity to human serum albumin (HSA). It has been observed that BSA-loaded NDs can effectively enhance tumor penetration, leading to improved therapeutic

outcomes.^{123,124} In a study conducted in 2019, BSA was incorporated into NDs along with a fluorescent probe called fluorescein isothiocyanate (FITC). This innovative design allows for the fluorescent labeling of NDs with any protein, facilitating their visualization and detection using imaging and fluorescence spectroscopy techniques.¹²⁵ Furthermore, researchers have successfully developed density-based fluorescent sensors known as isenNDs using NDs. A study reported in 2023 obtained covalently circularized NDs (cNDs) through the engineering of a self-cycling “autocyclase” protein, compared with both linear and cyclised NDs equipped with imaging agents, the *in vivo* imaging results showed that cNDs have better biodistribution and metabolism.¹²⁶ These NDs function as powerful tools for reporting membrane biochemical reactions, making them valuable in various applications.⁴³

4.5.2 Magnetic field nanoscalpel. The surgical resection of tumors is a widely employed method for treating malignant tumors. However, achieving complete tumor removal while minimizing damage to healthy tissues poses a significant surgical challenge.^{127,128} To address this challenge, the concept of a “nanoscalpel” has emerged. This approach involves remotely controllable magnetic nanosheets exhibiting high affinity and specificity for tumor cells. The nanosheets have anisotropic properties and a discoidal shape, facilitating their movement through blood vessels and enabling them to penetrate tumor tissue *via* endothelial cells easily. This innovative strategy holds the potential to offer targeted therapy for cancer patients under visual guidance, thereby addressing the issue of precision in cancer treatment.¹²⁹

5. Conclusions

NDs are a versatile and innovative platform that can be used to deliver drugs or peptides for various disease treatments. NDs can protect the drugs or peptides from degradation, improve their pharmacokinetics and selectivity, and enhance their therapeutic efficacy. NDs can also be engineered to display or release enzymes, growth factors, or drugs that can promote tissue repair or regeneration, or to act as adhesives or scaffolds that can facilitate cell migration or attachment. NDs have been explored in various disease treatments, such as cancer immunotherapy, vaccine development, antibacterial and antiviral therapy, and treatment of diabetes mellitus and Alzheimer's disease. NDs have also shown promise as imaging agents and platforms for microsurgery.

Despite the promising potential of NDs in various disease treatments, several challenges and limitations need to be addressed to translate them into clinical applications. First, the scalability and reproducibility of ND production and purification need to be improved to ensure the consistency and quality of NDs. Second, the safety and toxicity of NDs need to be thoroughly evaluated, especially in terms of their long-term effects on the immune system, the liver, and the kidney. Third, the pharmacokinetics and biodistribution of NDs need to be optimized to ensure the efficient delivery and clearance of

drugs or peptides. Fourth, the targeting and specificity of NDs need to be enhanced to ensure the selective and effective accumulation of drugs or peptides in the diseased tissues or cells. Fifth, the cost-effectiveness and regulatory compliance of NDs need to be considered to ensure the affordability and accessibility of the therapy.

In the future, several strategies can be explored to overcome these challenges and improve the clinical potential of NDs. First, new MSP variants or alternative scaffold proteins can be developed to enhance the stability, homogeneity, and scalability of NDs. Second, new surface modifications or coatings can be introduced to enhance the biocompatibility, immunogenicity, and targeting of NDs. Third, new encapsulation or loading strategies can be developed to enhance the loading efficiency, stability, and specificity of drugs or peptides. Fourth, new imaging or diagnostic modalities can be developed to enhance the real-time monitoring and evaluation of the therapy. Fifth, new collaborations or partnerships can be established to facilitate the translation and commercialization of the technology. Overall, NDs hold great promise as a novel disease treatment platform and have the potential to revolutionize the field of medicine in the years to come.

Author contributions

The manuscript was written through the contributions of all authors. All authors have given approval to the final version of the manuscript.

Conflicts of interest

There are no conflicts to declare.

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References

- 1 A. Nath, W. M. Atkins and S. G. Sligar, *Biochemistry*, 2007, **46**, 2059–2069.
- 2 S. Ben-Aicha, L. Badimon and G. Vilahur, *Int. J. Mol. Sci.*, 2020, **21**, 732.
- 3 M. C. Phillips, *J. Lipid Res.*, 2013, **54**, 2034–2048.
- 4 H. Park, A. Otte and K. Park, *J. Controlled Release*, 2022, **342**, 53–65.
- 5 M. J. Mitchell, M. M. Billingsley, R. M. Haley, M. E. Wechsler, N. A. Peppas and R. Langer, *Nat. Rev. Drug Discovery*, 2021, **20**, 101–124.
- 6 B. Fonseca-Santos, M. P. Gremião and M. Chorilli, *Int. J. Nanomed.*, 2015, **10**, 4981–5003.
- 7 R. Alyautdin, I. Khalin, M. I. Nafeeza, M. H. Haron and D. Kuznetsov, *Int. J. Nanomed.*, 2014, **9**, 795–811.
- 8 I. Vhora, R. Lalani, P. Bhatt, S. Patil and A. Misra, *Int. J. Pharm.*, 2019, **563**, 324–336.
- 9 E. Rideau, R. Dimova, P. Schwiller, F. R. Wurm and K. Landfester, *Chem. Soc. Rev.*, 2018, **47**, 8572–8610.
- 10 S. W. Lee, Y. M. Kim, C. H. Cho, Y. T. Kim, S. M. Kim, S. Y. Hur, J. H. Kim, B. G. Kim, S. C. Kim, H. S. Ryu and S. B. Kang, *Cancer Res. Treat.*, 2018, **50**, 195–203.
- 11 L. Palmerston Mendes, J. Pan and V. P. Torchilin, *Molecules*, 2017, **22**, 1401.
- 12 A. R. Menjoge, R. M. Kannan and D. A. Tomalia, *Drug Discovery Today*, 2010, **15**, 171–185.
- 13 J. Wang, A. M. Potocny, J. Rosenthal and E. S. Day, *ACS Omega*, 2020, **5**, 926–940.
- 14 L. S. Arias, J. P. Pessan, A. P. M. Vieira, T. M. T. Lima, A. C. B. Delbem and D. R. Monteiro, *Antibiotics*, 2018, **7**, 46.
- 15 K. W. Huang, F. F. Hsu, J. T. Qiu, G. J. Chern, Y. A. Lee, C. C. Chang, Y. T. Huang, Y. C. Sung, C. C. Chiang, R. L. Huang, C. C. Lin, T. K. Dinh, H. C. Huang, Y. C. Shih, D. Alson, C. Y. Lin, Y. C. Lin, P. C. Chang, S. Y. Lin and Y. Chen, *Sci. Adv.*, 2020, **6**, eaax5032.
- 16 C. Xu, J. Nam, H. Hong, Y. Xu and J. J. Moon, *ACS Nano*, 2019, **13**, 12148–12161.
- 17 R. Kuai, D. Li, Y. E. Chen, J. J. Moon and A. Schwendeman, *ACS Nano*, 2016, **10**, 3015–3041.
- 18 S. Ben-Aicha, L. Badimon and G. Vilahur, *Int. J. Mol. Sci.*, 2020, **21**, 732.
- 19 J. Bariwal, H. Ma, G. A. Altenberg and H. Liang, *Chem. Soc. Rev.*, 2022, **51**, 1702–1728.
- 20 T. H. Bayburt and S. G. Sligar, *FEBS Lett.*, 2010, **584**, 1721–1727.
- 21 T. Nakayama, J. S. Butler, A. Sehgal, M. Severgnini, T. Racie, J. Sharman, F. Ding, S. S. Morskaya, J. Brodsky, L. Tchangov, V. Kosovrasti, M. Meys, L. Nechev, G. Wang, C. G. Peng, Y. Fang, M. Maier, K. G. Rajeev, R. Li, J. Hettinger, S. Barros, V. Clausen, X. Zhang, Q. Wang, R. Hutabarat, N. V. Dokholyan, C. Wolfrum, M. Manoharan, V. Kotelianski, M. Stoffel and D. W. Sah, *Mol. Ther.*, 2012, **20**, 1582–1589.
- 22 R. Kuai, L. J. Ochyl, K. S. Bahjat, A. Schwendeman and J. J. Moon, *Nat. Mater.*, 2017, **16**, 489–496.
- 23 J. M. Dörr, S. Scheidelaar, M. C. Koorengevel, J. J. Dominguez, M. Schäfer, C. A. van Walree and J. A. Killian, *Eur. Biophys. J.*, 2016, **45**, 3–21.
- 24 M. C. Fiori, Y. Jiang, W. Zheng, M. Anzaldúa, M. J. Borgnia, G. A. Altenberg and H. Liang, *Sci. Rep.*, 2017, **7**, 15227.
- 25 M. E. Zoghbi, R. S. Cooper and G. A. Altenberg, *J. Biol. Chem.*, 2016, **291**, 4453–4461.
- 26 R. S. Cooper and G. A. Altenberg, *J. Biol. Chem.*, 2013, **288**, 20785–20796.
- 27 X. Y. Wang, L. Lin, R. F. Liu, M. Chen, B. L. Chen, B. He, B. He, X. L. Liang, W. B. Dai, H. Zhang, X. Q. Wang,

- Y. G. Wang, Z. F. Dai and Q. Zhang, *Adv. Funct. Mater.*, 2017, **27**, 1701027.
- 28 M. L. Nasr, D. Baptista, M. Strauss, Z.-Y. J. Sun, S. Grigoriu, S. Huser, A. Plückthun, F. Hagn, T. Walz, J. M. Hogle and G. Wagner, *Nat. Methods*, 2017, **14**, 49–52.
- 29 J. Miehl, D. Goricanec and F. Hagn, *ChemBioChem*, 2018, **19**, 1927–1933.
- 30 Y. Yusuf, J. Massiot, Y.-T. Chang, P.-H. Wu, V. Yeh, P.-C. Kuo, J. Shiue and T.-Y. Yu, *Langmuir*, 2018, **34**, 3525–3532.
- 31 M. Huang, M. Hu, Q. Song, H. Song, J. Huang, X. Gu, X. Wang, J. Chen, T. Kang, X. Feng, D. Jiang, G. Zheng, H. Chen and X. Gao, *ACS Nano*, 2015, **9**, 10801–10816.
- 32 Q. Song, H. Song, J. Xu, J. Huang, M. Hu, X. Gu, J. Chen, G. Zheng, H. Chen and X. Gao, *Mol. Pharm.*, 2016, **13**, 3976–3987.
- 33 Q. Chen, G. Guan, F. Deng, D. Yang, P. Wu, S. Kang, R. Sun, X. Wang, D. Zhou, W. Dai, X. Wang, H. Zhang, B. He, D. Chen and Q. Zhang, *Biomaterials*, 2020, **251**, 120008.
- 34 I. Noh, Z. Guo, J. Zhou, W. Gao, R. H. Fang and L. Zhang, *ACS Nano*, 2022, 2c08360, DOI: [10.1021/acsnano](https://doi.org/10.1021/acsnano).
- 35 L. Yuan, Q. Chen, J. E. Riviere and Z. Lin, *J. Drug Delivery Sci. Technol.*, 2023, **83**, 104404.
- 36 G. Sharma, D. T. Valenta, Y. Altman, S. Harvey, H. Xie, S. Mitragotri and J. W. Smith, *J. Controlled Release*, 2010, **147**, 408–412.
- 37 E. L. Dane, A. Belessiotis-Richards, C. Backlund, J. Wang, K. Hidaka, L. E. Milling, S. Bhagchandani, M. B. Melo, S. Wu, N. Li, N. Donahue, K. Ni, L. Ma, M. Okaniwa, M. M. Stevens, A. Alexander-Katz and D. J. Irvine, *Nat. Mater.*, 2022, **21**, 710–720.
- 38 S. Huang, L. Deng, H. Zhang, L. Wang, Y. Zhang, Q. Lin, T. Gong, X. Sun, Z. Zhang and L. Zhang, *Nano Res.*, 2022, **15**, 728–737.
- 39 A. Tahmasbi Rad, C. W. Chen, W. Aresh, Y. Xia, P. S. Lai and M. P. Nieh, *ACS Appl. Mater. Interfaces*, 2019, **11**, 10505–10519.
- 40 H. Patel, B. Ding, K. Ernst, L. Shen, W. Yuan, J. Tang, L. R. Drake, J. Kang, Y. Li, Z. Chen and A. Schwendeman, *Int. J. Nanomed.*, 2019, **14**, 3069–3086.
- 41 M. C. Fiori, Y. Jiang, G. A. Altenberg and H. Liang, *Sci. Rep.*, 2017, **7**, 7432.
- 42 L. Chen, C. Yu, W. Xu, Y. Xiong, P. Cheng, Z. Lin, Z. Zhang, L. Knoedler, A. C. Panayi, S. Knoedler, J. Wang, B. Mi and G. Liu, *ACS Nano*, 2023, **17**, 3153–3167.
- 43 Q. Ren, S. Zhang and H. Bao, *Commun. Biol.*, 2022, **5**, 507.
- 44 E. L. Dane, A. Belessiotis-Richards, C. Backlund, J. Wang, K. Hidaka, L. E. Milling, S. Bhagchandani, M. B. Melo, S. Wu, N. Li, N. Donahue, K. Ni, L. Ma, M. Okaniwa, M. M. Stevens, A. Alexander-Katz and D. J. Irvine, *Nat. Mater.*, 2022, **21**, 710–720.
- 45 A. Hassani Najafabadi, J. Zhang, M. E. Aikins, Z. I. Najaf Abadiare, F. Liao, Y. Qin, E. B. Okeke, L. M. Scheetz, J. Nam, Y. Xu, D. Adams, P. Lester, T. Hetrick, A. Schwendeman, M. S. Wicha, A. E. Chang, Q. Li and J. J. Moon, *Nano Lett.*, 2020, **20**, 7783–7792.
- 46 R. Kuai, W. M. Yuan, S. Son, J. Nam, Y. Xu, Y. C. Fan, A. Schwendeman and J. J. Moon, *Sci. Adv.*, 2018, **4**, eaao1736.
- 47 R. Patwa, N. Soundararajan, N. Mulchandani, S. M. Bhasney, M. Shah, S. Kumar, A. Kumar and V. Katiyar, *Biopolymers*, 2018, **109**, e23231.
- 48 P. S. Hegde and D. S. Chen, *Immunity*, 2020, **52**, 17–35.
- 49 R. Kuai, X. Sun, W. Yuan, L. J. Ochyl, Y. Xu, A. Hassani Najafabadi, L. Scheetz, M. Z. Yu, I. Balwani, A. Schwendeman and J. J. Moon, *J. Controlled Release*, 2018, **282**, 131–139.
- 50 P. Kadiyala, D. Li, F. M. Nuñez, D. Altshuler, R. Doherty, R. Kuai, M. Yu, N. Kamran, M. Edwards, J. J. Moon, P. R. Lowenstein, M. G. Castro and A. Schwendeman, *ACS Nano*, 2019, **13**, 1365–1384.
- 51 A. Harari, M. Graciotti, M. Bassani-Sternberg and L. E. Kandalaf, *Nat. Rev. Drug Discovery*, 2020, **19**, 635–652.
- 52 W. Li, X. Zhang, C. Zhang, J. Yan, X. Hou, S. Du, C. Zeng, W. Zhao, B. Deng, D. W. McComb, Y. Zhang, D. D. Kang, J. Li, W. E. Carson 3rd and Y. Dong, *Nat. Commun.*, 2021, **12**, 7264.
- 53 J. Zhang, B. Fan, G. Cao, W. Huang, F. Jia, G. Nie and H. Wang, *Adv. Mater.*, 2022, **34**, e2205950.
- 54 N. Gong and M. J. Mitchell, *Nat. Mater.*, 2022, **21**, 616–617.
- 55 P. Lin, Y. Xue, X. Mu, Y. Shao, Q. Lu, X. Jin, E. Yinwang, Z. Zhang, H. Zhou, W. Teng, H. Sun, W. Chen, W. Shi, C. Shi, X. Zhou, X. Jiang, X. Yu and Z. Ye, *Small*, 2022, **18**, e2200179.
- 56 X. Liu, C. Liu, Z. Zheng, S. Chen, X. Pang, X. Xiang, J. Tang, E. Ren, Y. Chen, M. You, X. Wang, X. Chen, W. Luo, G. Liu and N. Xia, *Adv. Mater.*, 2019, **31**, e1808294.
- 57 C. Liu, X. Liu, X. Xiang, X. Pang, S. Chen, Y. Zhang, E. Ren, L. Zhang, X. Liu, P. Lv, X. Wang, W. Luo, N. Xia, X. Chen and G. Liu, *Nat. Nanotechnol.*, 2022, **17**, 531–540.
- 58 V. Yeh, T.-Y. Lee, C.-W. Chen, P.-C. Kuo, J. Shiue, L.-K. Chu and T.-Y. Yu, *Sci. Rep.*, 2018, **8**, 13501.
- 59 R. Kuai, L. J. Ochyl, K. S. Bahjat, A. Schwendeman and J. J. Moon, *Nat. Mater.*, 2017, **16**, 489–496.
- 60 R. Kuai, X. Sun, W. Yuan, Y. Xu, A. Schwendeman and J. J. Moon, *Bioconjug Chem.*, 2018, **29**, 771–775.
- 61 A. H. Najafabadi, Z. I. N. Abadi, M. E. Aikins, K. E. Foulds, M. M. Donaldson, W. Yuan, E. B. Okeke, J. Nam, Y. Xu, P. Weerappuli, T. Hetrick, D. Adams, P. A. Lester, A. M. Salazar, D. H. Barouch, A. Schwendeman, R. A. Seder and J. J. Moon, *J. Controlled Release*, 2021, **337**, 168–178.
- 62 P. Marcato, C. A. Dean, C. A. Giacomantonio and P. W. Lee, *Cell Cycle*, 2011, **10**, 1378–1384.
- 63 C. Ginestier, M. H. Hur, E. Charafe-Jauffret, F. Monville, J. Dutcher, M. Brown, J. Jacquemier, P. Viens, C. G. Kleer,

- S. Liu, A. Schott, D. Hayes, D. Birnbaum, M. S. Wicha and G. Dontu, *Cell Stem Cell*, 2007, **1**, 555–567.
- 64 K. Rantalainen, Z. T. Berndsen, A. Antanasijevic, T. Schiffner, X. Zhang, W. H. Lee, J. L. Torres, L. Zhang, A. Irimia, J. Copps, K. H. Zhou, Y. D. Kwon, W. H. Law, C. A. Schramm, R. Verardi, S. J. Krebs, P. D. Kwong, N. A. Doria-Rose, I. A. Wilson, M. B. Zwick, J. R. Yates 3rd, W. R. Schief and A. B. Ward, *Cell Rep.*, 2020, **31**, 107583.
- 65 S. Yang, G. Hiotis, Y. Wang, J. Chen, J. H. Wang, M. Kim, E. L. Reinherz and T. Walz, *Nat. Commun.*, 2022, **13**, 6393.
- 66 Y. Wang, P. Kaur, Z. J. Sun, M. A. Elbahnasawy, Z. Hayati, Z. S. Qiao, N. N. Bui, C. Chile, M. L. Nasr, G. Wagner, J. H. Wang, L. Song, E. L. Reinherz and M. Kim, *Proc. Natl. Acad. Sci. U. S. A.*, 2019, **116**, 22556–22566.
- 67 J. Wang, Y. Zhang, E. Archibong, F. S. Ligler and Z. Gu, *Adv. Biosyst.*, 2017, **1**, e1700084.
- 68 Z. Xu, Z. Qiu, Q. Liu, Y. Huang, D. Li, X. Shen, K. Fan, J. Xi, Y. Gu, Y. Tang, J. Jiang, J. Xu, J. He, X. Gao, Y. Liu, H. Koo, X. Yan and L. Gao, *Nat. Commun.*, 2018, **9**, 3713.
- 69 J. Ouyang, M. Wen, W. Chen, Y. Tan, Z. Liu, Q. Xu, K. Zeng, L. Deng and Y. N. Liu, *Chem. Commun.*, 2019, **55**, 4877–4880.
- 70 C. Zhang, Q. Li, J. Shan, J. Xing, X. Liu, Y. Ma, H. Qian, X. Chen, X. Wang, L. M. Wu and Y. Yu, *Acta Biomater.*, 2023, **160**, 252–264.
- 71 S. Speerstra, A. A. Chistov, G. V. Proskurin, A. V. Aralov, E. A. Ulashchik, P. P. Streshnev, V. V. Shmanai, V. A. Korshun and L. M. Schang, *Antiviral Res.*, 2018, **149**, 164–173.
- 72 M. C. Wolf, A. N. Freiberg, T. Zhang, Z. Akyol-Ataman, A. Grock, P. W. Hong, J. Li, N. F. Watson, A. Q. Fang, H. C. Aguilar, M. Porotto, A. N. Honko, R. Damoiseaux, J. P. Miller, S. E. Woodson, S. Chantasirivisal, V. Fontanes, O. A. Negrete, P. Krogstad, A. Dasgupta, A. Moscona, L. E. Hensley, S. P. Whelan, K. F. Faull, M. R. Holbrook, M. E. Jung and B. Lee, *Proc. Natl. Acad. Sci. U. S. A.*, 2010, **107**, 3157–3162.
- 73 S. Skalickova, Z. Heger, L. Krejcova, V. Pekarik, K. Bastl, J. Janda, F. Kostolansky, E. Vareckova, O. Zitka, V. Adam and R. Kizek, *Viruses*, 2015, **7**, 5428–5442.
- 74 B. Kong, S. Moon, Y. Kim, P. Heo, Y. Jung, S. H. Yu, J. Chung, C. Ban, Y. H. Kim, P. Kim, B. J. Hwang, W. J. Chung, Y. K. Shin, B. L. Seong and D. H. Kweon, *Nat. Commun.*, 2019, **10**, 185.
- 75 L. R. Walker and M. T. Marty, *Biochemistry*, 2020, **59**, 2135–2142.
- 76 Y. Ikeda, T. Taira, K. Sakai, H. Sakai, Y. Shigeri and T. Imura, *J. Oleo Sci.*, 2018, **67**, 1035–1041.
- 77 V. W. Henderson and C. E. Finch, *J. Neurosurg.*, 1989, **70**, 335–353.
- 78 R. Kandimalla, V. Thirumala and P. H. Reddy, *Biochim. Biophys. Acta, Mol. Basis Dis.*, 2017, **1863**, 1078–1089.
- 79 K. G. Mawuenyega, W. Sigurdson, V. Ovod, L. Munsell, T. Kasten, J. C. Morris, K. E. Yarasheski and R. J. Bateman, *Science*, 2010, **330**, 1774.
- 80 J. M. Castellano, J. Kim, F. R. Stewart, H. Jiang, R. B. DeMattos, B. W. Patterson, A. M. Fagan, J. C. Morris, K. G. Mawuenyega, C. Cruchaga, A. M. Goate, K. R. Bales, S. M. Paul, R. J. Bateman and D. M. Holtzman, *Sci. Transl. Med.*, 2011, **3**, 89ra57.
- 81 K. R. Wildsmith, M. Holley, J. C. Savage, R. Skerrett and G. E. Landreth, *Alzheimers Res. Ther.*, 2013, **5**, 33.
- 82 P. Zhang, Y. Kishimoto, I. Grammatikakis, K. Gottimukkala, R. G. Cutler, S. Zhang, K. Abdelmohsen, V. A. Bohr, J. Misra Sen, M. Gorospe and M. P. Mattson, *Nat. Neurosci.*, 2019, **22**, 719–728.
- 83 Y. Zhao, J. Cai, Z. Liu, Y. Li, C. Zheng, Y. Zheng, Q. Chen, H. Chen, F. Ma, Y. An, L. Xiao, C. Jiang, L. Shi, C. Kang and Y. Liu, *Nano Lett.*, 2019, **19**, 674–683.
- 84 T. H. Ferreira-Vieira, I. M. Guimaraes, F. R. Silva and F. M. Ribeiro, *Curr. Neuropharmacol.*, 2016, **14**, 101–115.
- 85 N. N. Nalivaeva and A. J. Turner, *Chem.-Biol. Interact.*, 2016, **259**, 301–306.
- 86 T. Wisniewski and F. Goñi, *Neuron*, 2015, **85**, 1162–1176.
- 87 S. Mourtas, M. Canovi, C. Zona, D. Aurilia, A. Niarakis, B. La Ferla, M. Salmona, F. Nicotra, M. Gobbi and S. G. Antimisiaris, *Biomaterials*, 2011, **32**, 1635–1645.
- 88 D. Brambilla, R. Verpillot, B. Le Droumaguet, J. Nicolas, M. Taverna, J. Kóna, B. Lettiero, S. H. Hashemi, L. De Kimpe, M. Canovi, M. Gobbi, V. Nicolas, W. Scheper, S. M. Moghimi, I. Tvaroška, P. Couvreur and K. Andrieux, *ACS Nano*, 2012, **6**, 5897–5908.
- 89 M. Gobbi, F. Re, M. Canovi, M. Beeg, M. Gregori, S. Sesana, S. Sonnino, D. Brogioli, C. Musicanti, P. Gasco, M. Salmona and M. E. Masserini, *Biomaterials*, 2010, **31**, 6519–6529.
- 90 J. G. Tsun, S. Yung, M. K. Chau, S. W. Shiu, T. M. Chan and K. C. Tan, *PLoS One*, 2014, **9**, e105787.
- 91 G. Russo, P. Piscitelli, A. Giandalia, F. Viazzi, R. Pontremoli, P. Fioretto and S. De Cosmo, *J. Nephrol.*, 2020, **33**, 1001–1008.
- 92 T. Toyama, M. Shimizu, K. Furuichi, S. Kaneko and T. Wada, *Clin. Exp. Nephrol.*, 2014, **18**, 201–205.
- 93 G. M. Ducasa, A. Mitrofanova and A. Fornoni, *Curr. Diab. Rep.*, 2019, **19**, 144.
- 94 P. Liu, L. Peng, H. Zhang, P. M. Tang, T. Zhao, M. Yan, H. Zhao, X. Huang, H. Lan and P. Li, *Front. Physiol.*, 2018, **9**, 343.
- 95 E. E. Morin, Y. Guo, H. He, W. Yuan, W. N. Souery, M. V. Fawaz, Y. E. Chen and A. Schwendeman, *Front. Pharmacol.*, 2020, **11**, 513031.
- 96 E. Kiss, B. Kränzlin, K. Wagenblaß, M. Bonrouhi, J. Thiery, E. Gröne, V. Nordström, D. Teupser, N. Gretz, E. Malle and H. J. Gröne, *Am. J. Pathol.*, 2013, **182**, 727–741.
- 97 Y. Wang, S. Xiao, S. Zhou, R. Zhang, H. Liu, Y. Lin and P. Yu, *Front. Physiol.*, 2020, **11**, 552483.
- 98 K. Feng, C. Ma, Y. Liu, X. Yang, Z. Yang, Y. Chen, T. Xu, C. Yang, S. Zhang, Q. Li, Z. Wei, D. Zhao, P. Zeng, J. Han, J. Gao, Y. Chen and Y. Duan, *Theranostics*, 2021, **11**, 2634–2654.

- 99 F. Oroojalian, F. Charbgoos, M. Hashemi, A. Amani, R. Yazdian-Robati, A. Mokhtarzadeh, M. Ramezani and M. R. Hamblin, *J. Controlled Release*, 2020, **321**, 442–462.
- 100 G. Wang, Q. Li, D. Chen, B. Wu, Y. Wu, W. Tong and P. Huang, *Theranostics*, 2019, **9**, 6191–6208.
- 101 H. He, R. Adili, L. Liu, K. Hong, M. Holinstat and A. Schwendeman, *Sci. Adv.*, 2020, **6**, eabd0130.
- 102 H. He, T. A. Halseth, L. Mei, C. Shen, L. Liu and A. Schwendeman, *J. Controlled Release*, 2022, **348**, 1016–1027.
- 103 S. Amadesi, C. Reni, R. Katore, M. Meloni, A. Oikawa, A. P. Beltrami, E. Avolio, D. Cesselli, O. Fortunato, G. Spinetti, R. Ascione, E. Cangiano, M. Valgimigli, S. P. Hunt, C. Emanuelli and P. Madeddu, *Circulation*, 2012, **125**, 1774–1786.
- 104 H. S. Hong, S. Kim, S. Nam, J. Um, Y. H. Kim and Y. Son, *Wound Repair Regen.*, 2015, **23**, 268–277.
- 105 J. J. Bowden, A. M. Garland, P. Baluk, P. Lefevre, E. F. Grady, S. R. Vigna, N. W. Bunnett and D. M. McDonald, *Proc. Natl. Acad. Sci. U. S. A.*, 1994, **91**, 8964–8968.
- 106 H. J. Park, R. Kuai, E. J. Jeon, Y. Seo, Y. Jung, J. J. Moon, A. Schwendeman and S. W. Cho, *Biomaterials*, 2018, **161**, 69–80.
- 107 M. Hoekstra, *Atherosclerosis*, 2017, **258**, 153–161.
- 108 W. L. Wong, X. Su, X. Li, C. M. Cheung, R. Klein, C. Y. Cheng and T. Y. Wong, *Lancet Global Health*, 2014, **2**, e106–e116.
- 109 E. Loane, J. M. Nolan and S. Beatty, *Invest. Ophthalmol. Vis. Sci.*, 2010, **51**, 5897–5905.
- 110 Y. Wang, S. L. Connor, W. Wang, E. J. Johnson and W. E. Connor, *Exp. Eye Res.*, 2007, **84**, 591–598.
- 111 M. N. Oda, P. L. Hargreaves, J. A. Beckstead, K. A. Redmond, R. van Antwerpen and R. O. Ryan, *J. Lipid Res.*, 2006, **47**, 260–267.
- 112 M. Ghosh, A. T. Singh, W. Xu, T. Sulchek, L. I. Gordon and R. O. Ryan, *Nanomedicine*, 2011, **7**, 162–167.
- 113 A. Moschetti, L. N. Vine, K. Lethcoe, R. K. Dagda, P. Ellison and R. O. Ryan, *Lipids*, 2020, **55**, 141–149.
- 114 A. Moschetti, C. A. Fox, S. McGowen and R. O. Ryan, *Front. Nanotechnol.*, 2022, **4**, 955022.
- 115 B. Park, Y. K. Cha, J. Kwak, K. S. Hwang, H. S. Kim, S. Park, Y. Pak, T. H. Park, H. S. Song and J. H. Kim, *Nano Lett.*, 2022, **22**, 6825–6832.
- 116 T. Murakami, *Biotechnol. J.*, 2012, **7**, 762–767.
- 117 M. Niu, Y. Lu, L. Hovgaard and W. Wu, *Int. J. Nanomed.*, 2011, **6**, 1155–1166.
- 118 N. T. Huynh, C. Passirani, P. Saulnier and J. P. Benoit, *Int. J. Pharm.*, 2009, **379**, 201–209.
- 119 S. Krol, R. Ellis-Behnke and P. Marchetti, *Maturitas*, 2012, **73**, 61–67.
- 120 A. K. Agrawal, H. Harde, K. Thanki and S. Jain, *Biomacromolecules*, 2014, **15**, 350–360.
- 121 A. Nath, W. M. Atkins and S. G. Sligar, *Biochemistry*, 2007, **46**, 2059–2069.
- 122 A. Nath, A. J. Trexler, P. Koo, A. D. Miranker, W. M. Atkins and E. Rhoades, *Methods Enzymol.*, 2010, **472**, 89–117.
- 123 D. C. Carter and J. X. Ho, *Adv. Protein. Chem.*, 1994, **45**, 153–203.
- 124 J. S. Mandeville and H. A. Tajmir-Riahi, *Biomacromolecules*, 2010, **11**, 465–472.
- 125 S. Damiati, A. Scheberl, S. Zayni, S. A. Damiati, B. Schuster and U. B. Kompella, *Biophys. Chem.*, 2019, **251**, 106178.
- 126 X. Jia, Y. K. Y. Chin, A. H. Zhang, T. Crawford, Y. Zhu, N. L. Fletcher, Z. Zhou, B. R. Hamilton, M. Stroet, K. J. Thurecht and M. Mobli, *Commun. Chem.*, 2023, **6**, 48.
- 127 J. Ferlay, E. Steliarova-Foucher, J. Lortet-Tieulent, S. Rosso, J. W. Coebergh, H. Comber, D. Forman and F. Bray, *Eur. J. Cancer*, 2013, **49**, 1374–1403.
- 128 G. C. Barnett, C. M. West, A. M. Dunning, R. M. Elliott, C. E. Coles, P. D. Pharoah and N. G. Burnet, *Nat. Rev. Cancer*, 2009, **9**, 134–142.
- 129 T. N. Zamay, V. S. Prokopenko, S. S. Zamay, K. A. Lukyanenko, O. S. Kolovskaya, V. A. Orlov, G. S. Zamay, R. G. Galeev, A. A. Narodov and A. S. Kichkailo, *Nanomaterials*, 2021, **11**, 1459.