

Volume 11
Number 30
14 August 2023
Pages 6985–7240

Journal of Materials Chemistry B

Materials for biology and medicine

rsc.li/materials-b



ISSN 2050-750X



ROYAL SOCIETY
OF CHEMISTRY

REVIEW ARTICLE

Sara Rojas, Antonio Rodríguez-Diéguez *et al.*
Therapy and diagnosis of Alzheimer's disease: from discrete
metal complexes to metal-organic frameworks





Cite this: *J. Mater. Chem. B*, 2023, 11, 7024

Therapy and diagnosis of Alzheimer's disease: from discrete metal complexes to metal–organic frameworks

Amalia García-García,^{ab} Sara Rojas^{id}*^a and Antonio Rodríguez-Diéguez^{id}*^a

Alzheimer's disease (AD) is a neurodegenerative disorder affecting 44 million people worldwide. Although many issues (pathogenesis, genetics, clinical features, and pathological aspects) are still unknown, this disease is characterized by noticeable hallmarks such as the formation of β -amyloid plaques, hyperphosphorylation of tau proteins, the overproduction of reactive oxygen species, and the reduction of acetylcholine levels. There is still no cure for AD and the current treatments are aimed at regulating the cholinesterase levels, attenuating symptoms temporarily rather than preventing the AD progression. In this context, coordination compounds are regarded as a promising tool in AD treatment and/or diagnosis. Coordination compounds (discrete or polymeric) possess several features that make them an interesting option for developing new drugs for AD (good biocompatibility, porosity, synergetic effects of ligand–metal, fluorescence, particle size, homogeneity, monodispersity, etc.). This review discusses the recent progress in the development of novel discrete metal complexes and metal–organic frameworks (MOFs) for the treatment, diagnosis and theragnosis of AD. These advanced therapies for AD treatment are organized according to the target: A β peptides, hyperphosphorylated tau proteins, synaptic dysfunction, and mitochondrial failure with subsequent oxidative stress.

Received 27th February 2023,
Accepted 31st May 2023

DOI: 10.1039/d3tb00427a

rsc.li/materials-b

Introduction

Alzheimer's disease (AD), the most common form of dementia, is an irreversible and progressive neurodegenerative disorder, and a global public health issue, according to the World Health

^a Department of Inorganic Chemistry, Faculty of Science, University of Granada, Av. Fuentenueva S/N, 18071 Granada, Spain. E-mail: srojas@ugr.es, antonio5@ugr.es

^b Centro de Química del Instituto de Ciencias, Benemérita Universidad Autónoma de Puebla, 18 sur & Av. San Claudio, Col. San Manuel, 72570 Puebla, Mexico



Amalia García-García

Dr Amalia García-García currently has a postdoctoral contract Margarita Salas at both the Benemérita Universidad Autónoma de Puebla in the Applied Bioinorganic Laboratory and the University of Granada in the Biochemistry and Electronics as Sensing Technologies group. She obtained her PhD from the University of Granada in 2021 by working on the synthesis of new coordination compounds with biomedical applications.

Her research mainly focuses on the design and synthesis of new coordination metal-complexes with potential applications against worldwide illnesses such as diabetes mellitus, cancer and Alzheimer's disease.



Sara Rojas

Dr Sara Rojas obtained her PhD from the University of Granada (in 2014) with a work focus on the synthesis of porous materials and their application as drug delivery systems. After finishing her PhD, she continued her studies at the Institute Lavoisier in Versailles (France, 2016) and the IMDEA Energy institute (Spain, 2018) under a framework of a Marie Curie Individual Fellowship and Atracción del Talento grant, respectively. In 2021, she joined

the Biochemistry and Electronics as Sensing Technologies group first as Juan de la Cierva Incorporación fellow and now as Ramón y Cajal. Her research focuses on the development of porous materials for biomedical and environmental applications.



Organization (WHO).¹ Given the current aging population, by 2050, the prevalence of dementia is estimated to increase up to 166%,² and therefore, the number of patients with AD is predicted to increase too.³ As there is no effective treatment and the disease's etiology and pathology remain largely unknown, nowadays, AD is the 5th cause of mortality worldwide, affecting about 44 million people in 2020.⁴ This means a global societal cost of 1.3 trillion \$ in 2019, which is expected to surpass 2.8 trillion \$ by 2030.¹ Considering these numbers, one could suggest that AD is an epidemic.

Although the progress in understanding the disease, there are still some controversies, mainly in AD pathologies and the development of efficient therapies. In 1907, Alois Alzheimer reported the first case of AD in a 51 year-old woman in the insane asylum of Frankfurt am Main (Germany). Clinical observation revealed that the symptoms were so odd that it could not be classified as any known disorder. The patient developed a rapid loss of memory, disorientation, mood change, and hallucinations. However, her walk, hands, and reflexes functioned correctly. Interestingly, the post-mortem examination of the brain showed important atrophy and abnormal vascular tissues. Moreover, neurons exhibited fibrils at the surface and inside them that disintegrated cells.⁵

Now, we know that AD is first characterized by a progressive deterioration of the capacity to recall new information, as neurons involved in forming new memories are the first to be damaged. As time passes, other symptoms appear such as memory loss, confusion, trouble understanding, changes in mood, increased anxiety, and difficulty in speaking and writing.⁶ These symptoms might begin 20 years or more after the onset of brain changes.^{7,8}

Pathology of AD

Although Alois Alzheimer reported the presence of abnormal fibrils in the brain of her demented patient, it was not until the end of the 20th century that the pathological hallmarks of AD

were clearly identified. As in many neurodegenerative disorders, AD is also caused by the misfolding and irreversible aggregation of specific proteins, β -amyloid ($A\beta$) and tau, which forms amyloid plaques and neurofibrillary tangles, respectively.^{9,10}

Amyloid plaques

In an attempt to explain the formation of these abnormalities in the brain, Hardy and Higgins proposed the "amyloid cascade hypothesis" in 1992.¹¹ This idea suggests that the deposition of $A\beta$ proteins is the cause of Alzheimer's pathology, whereas the abnormal hyperphosphorylation of tau proteins and thus, neurofibrillary tangle formation, cell loss, vascular damage, and dementia are consequences of the deposition.¹²

$A\beta$ peptides are natural proteolytic products of the transmembrane amyloid precursor protein (APP), and consist of 36–43 amino acids, with $A\beta_{40}$ being the most prevalent one.¹⁴ However, depending on the involved secretase enzyme, two pathways to cleave the APP are possible: the amyloidogenic pathway and the non-amyloidogenic pathway (Fig. 1). In the non-amyloidogenic process, the APP is first cleaved by α -secretase, giving rise to a soluble ectodomain $sAPP\alpha$ and an 83-residue carboxy-terminal fragment (C83). Subsequently, γ -secretase digests C83 into a soluble extracellular peptide (p3) and an APP intracellular domain (AICD). On the other hand, the amyloidogenic pathway begins with the catalysis of the APP by β -secretase 1 (BACE-1), generating the soluble ectodomains $sAPP\beta$ and C99. The following cleavage of C99 by γ -secretase yields the AICD and $A\beta$ peptides of different lengths, increasing the ratio of $A\beta_{42}/A\beta_{40}$, with $A\beta_{42}$ being more toxic than $A\beta_{40}$. Although the reasons of these abnormalities are still unknown, it is believed that in familial AD (*ca.* 5–10% of AD cases)¹⁵ mutations in the gene encoding APP or the γ -secretase subunit proteins presenilins 1 and 2 lead to an increased ratio between longer and shorter forms of $A\beta$.¹⁶ However, in sporadic AD (*ca.* 70% of AD cases), γ -secretase displays normal activity.

At this point, some controversies appear around the amyloid cascade hypothesis, since many individuals present amyloid



Antonio Rodríguez-Diéguez

Prof. Antonio Rodríguez-Diéguez completed his PhD studies in Chemistry at the University of Granada in 2005. Since 2008, he has worked as a Leading Researcher of different projects funded by the Junta de Andalucía and Ministry of Science and Technology and as a Professor at the Inorganic Chemistry Department of the University of Granada. His research interests have included crystalline analysis by XR diffraction, the improvement of solvothermal methods, studies of magnetic and luminescence properties and, recently, the synthesis of metal-organic frameworks for therapeutic applications.

Prof. Antonio Rodríguez-Diéguez completed his PhD studies in Chemistry at the University of Granada in 2005. Since 2008, he has worked as a Leading Researcher of different projects funded by the Junta de Andalucía and Ministry of Science and Technology and as a Professor at the Inorganic Chemistry Department of the University of Granada. His research interests have included crystalline analysis by XR diffraction, the improvement of solvothermal methods, studies of magnetic and luminescence properties and, recently, the synthesis of metal-organic frameworks for therapeutic applications.



Fig. 1 Schematic representation of the two different pathways to cleave the APP. In the non-amyloidogenic pathway (left), the APP is cleaved in the middle by α -secretase, generating the soluble $sAPP\alpha$ and C83. The amyloidogenic pathway (right) is promoted by β -secretase 1 (BACE1) and γ -secretase, giving rise to shorter $A\beta$ species that tends to aggregate. Adapted from ref. 13. Reproduced with permission.



plaques without AD symptoms; or the fact that soluble A β aggregates, rather than insoluble fibrillar aggregates, are thought to be the main factor responsible for the neurotoxic properties.¹⁷

Neurofibrillary tangles

The other neuropathogenic feature in AD is the presence of intracellular neurofibrillary tangles, which are formed by abnormal hyperphosphorylated and aggregated tau proteins in the hippocampus. In healthy brains, tau proteins confer stability to the microtubules of the neuronal axons. However, in AD, tau is hyperphosphorylated, reducing its ability to bind microtubules and self-associate into paired helical filaments, leading to neuronal death.¹⁸

Metals and neurotransmitters

Apart from amyloid plaques and neurofibrillary tangles, other anomalies linked to AD are altered levels of metals and neurotransmitters. Copper, zinc, and iron are essential trace elements involved in many metabolic pathways,¹⁹ and they are also implicated in neurodegenerative processes. These metal ions are found in the amyloid plaques at high concentrations, and *in vitro* studies revealed that they promote A β aggregation and the formation of reactive oxygen species (ROS), which play a key role in AD.^{20–22} On the other hand, neurochemical studies suggest that synapses containing acetylcholine, glutamate, and serotonin in the neocortex and hippocampus are predominantly affected in AD.²³ In presynaptic neurons, choline-acetyltransferase (ChAT) catalyzes the synthesis of acetylcholine (ACh) from choline and acetyl-coenzyme A. Then, ACh is released into the synaptic cleft, where it is bonded to the postsynaptic neuron. Finally, ACh is hydrolyzed in the synaptic cleft by acetylcholinesterase (AChE), which regulates its concentration at the synapse (Fig. 2). Furthermore, in a less-specific pathway, ACh is also hydrolyzed by butyrylcholinesterase (BuChE). However, in AD, the neurotransmitter acetylcholine is gradually lost in the end-stage of the disease.²⁴

Treatments

Since AD discovery, there is no effective treatment that delays or stops its progression. However, a total of 121 new agents are undergoing clinical trials.²⁵ Nowadays, there are only a few common medications approved by the US Food and Drug Administration (FDA): three cholinesterase inhibitors (donepezil, rivastigmine, and galantamine), one glutamate regulator (memantine), and their combination (memantine with donepezil). However, these drugs temporarily attenuate symptoms but do not alter the course of the disease,²⁶ and they produce side effects associated with high dosages and long periods of treatment.²⁷ From 2021, twenty years after the last approved drug, two new treatments were approved: first aducanumab and later lecanemab.^{28,29} These two monoclonal antibodies are the first disease-modifying therapies since they are able to reduce A β plaques and block their formation, respectively.^{30,31}

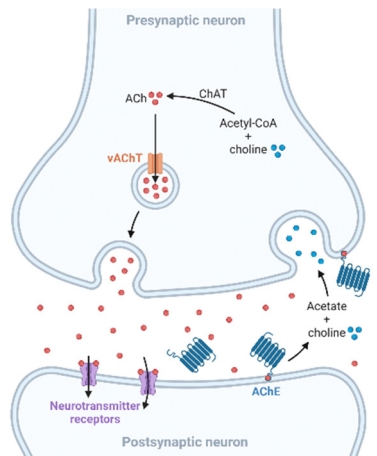


Fig. 2 Scheme of the cholinergic synapse. In the presynaptic neuron, choline-acetyltransferase (ChAT) catalyzes the formation of acetylcholine (ACh) from acetyl-coenzyme A and choline. Then, ACh is released into the synaptic cleft by a vesicular ACh transporter (vAChT), where ACh binds to receptors in the postsynaptic neuron. Finally, ACh is hydrolyzed in the synaptic cleft by acetylcholinesterase (AChE) into choline and acetate.

Both have been authorized by the FDA *via* the accelerated approval pathway, and they are only prescribed for early AD.^{32,33}

Regarding the AD's drug approval, the G8 leaders set a goal for developing effective drugs to treat or prevent AD by 2025,³⁴ and more efforts are being made to investigate other types of treatments, since the use/efficacy of aducanumab and other approved drugs is limited. Currently, the main target is to inhibit brain A β production and aggregation. Several strategies are focused on researching β -secretase inhibitors, the use of immunotherapy through antibodies that bind to A β plaques, tau inhibitors, anti-inflammatory drugs, and agents that chelate copper and iron. Mainly, organic molecules and small peptides are being widely studied for this purpose.^{35,36} Following this trend, coordination compounds could offer advantages as therapeutic agents compared to organic molecules, as the organic moiety can be widely modified to tune its selectivity and reactivity for a particular target, whereas metal ions offer different geometries and variable oxidation states.³⁷ For example, whereas an organic molecule has limited geometry, an octahedral metal center is capable of forming many stereoisomers and, thus, achieving higher protein-binding affinity and selectivity. In addition, these materials could undergo ligand exchange for better reactivity with biomolecular targets (*i.e.*, cisplatin), exhibit photophysical activity, create a 3D porous structure for drug delivery, and possess luminescent and magnetic properties in addition to biological activity.^{38,39}

In this regard, coordination compounds have been approved by the FDA in the treatment and diagnosis of different diseases. Generally, the activity of metal-containing compounds is associated with high toxicity. However, there is a small frontier between the pharmacological efficacy and the toxicity of drugs. Aside from the drug delivery and disposition of the active ingredient, the activity of metal-containing compounds depends on many factors, such as the oxidation state of the metal,



coordination geometry, and functional groups of the organic ligands. For example, mercury and its coordination compounds are believed very toxic, but some complexes such as thiomersal and merbromin are utilized as preservatives for vaccines and antiseptic agents, respectively. To date, many metal complexes based on the elements magnesium, iron, zirconium, palladium, platinum, mercury, and bismuth, among others, have been clinically approved in the US and/or the EU for a wide range of medical uses.³⁸ Some examples are cisplatin and carboplatin (cancer),⁴⁰ sodium stibogluconate and meglumine antimoniate (leishmaniasis),⁴¹ and auranofin (rheumatoid arthritis).⁴² Anyway, the use of biocompatible metals such as zinc, magnesium, and copper, and structures with the controlled delivery of their components is recommendable. With respect to AD, the application of coordination compounds as therapeutic agents is a rather new research field (since 2008), particularly focused on the aggregation and deposition of A β proteins, abnormal processing of the APP, formation of neurofibrillary tangles with hyperphosphorylated tau proteins, synaptic degeneration, and neuronal cell death.²³

Although chemotherapy is demonstrated to be an effective method in the treatment of AD, it has some disadvantages, such as the lack of specificity, significant side effects and the development of acquired resistance to the drug. Importantly, compared to other areas of the organism, the treatment of brain diseases is presently unsuccessful mostly due to the difficulty in crossing the blood–brain barrier (BBB).⁴³ The BBB is formed by the brain capillary endothelium and excludes from the brain *ca.* 100% of large-molecule neurotherapeutics and > 98% of small-molecule drugs.⁴⁴ Furthermore, to date, all the approved FDA drugs have been marked as oral formulations (donepezil, galantamine, and memantine) and one of them has been marked as a transdermal formulation (rivastigmine). So, it is necessary to use high doses to reach the brain, after overcoming oral barriers of adsorption, hepatic metabolism, distribution and finally traversing the BBB. In general, only low molecular weight lipophilic drugs can cross the BBB. However, some solutions are suggested, such as the application of vascular activating compounds that increase the permeability of the BBB, designing drugs with fat solubility, and direct drug delivery to the cerebrospinal fluid (CSF).⁴⁵ In this regard, nanotechnology has provided superior opportunities in the treatment and/or diagnosis of central nervous system diseases. Nanocarriers might be able to cross the BBB, as they can be easily modified and functionalized, increasing the drug concentration in the brain. Furthermore, they have been proposed for theragnosis as they can act as both imaging and therapeutic agents simultaneously.¹⁵

Among all the proposed nanocarriers (polymeric nanoparticles, lipids, liposomes, nanoemulsions, microemulsions, and dendrimers), metal–organic frameworks (MOFs) are an emerging class of materials which have attracted interest in AD treatment and/or diagnosis due to their outstanding properties: (i) large specific surface area and pore volume, associated with high sorption capacities, (ii) biocompatibility, (iii) good stability profiles under physiological conditions, (iv) tunable

particle size, and (v) can be used as imaging agents.^{46,47} Considering their plasticity, different strategies have been proposed in the application of MOFs for AD treatment, like the suppression of A β aggregation, as antioxidative agents or in promoting nerve regeneration.

While there are few reviews about metal-based drugs in AD treatment^{48–50} and MOFs in AD diagnosis have already been documented,^{51–53} the number of reports in this area has sharply increased (34 and 81% published within the last 4 years, according to the Web of Science, February 2023, (i) “coordination” “compounds” “Alzheimer”, and (ii) “metal” “organic” “framework” “Alzheimer”, respectively).

This review will discuss all the developments of coordination compounds, including outstanding discrete compounds and MOFs, published as treatment and/or diagnosis in AD. In order to give a broader picture of opportunities and disadvantages of the use of each compound/material, details of each structure and its properties are also included.

Discrete coordination compounds in AD treatment

Along this review, the reported coordination compounds in AD treatment are organized according to the target: A β peptides, hyperphosphorylated tau proteins, synaptic dysfunction, and mitochondrial failure with subsequent oxidative stress.¹⁴ A summary is shown in Table 1.

Inhibition of β -amyloid aggregation

A β peptides show high affinity for binding metal ions that can modulate peptide aggregation and toxicity. As previously mentioned, metals ions such as Cu^{II}, Zn^{II}, and Fe^{III} interact with A β *via* three histidine residues (6, 13, and 14) modulating the aggregation and toxicity of A β producing reactive oxygen species (ROS). For this reason, most of the reported complexes in AD treatment are centered on inhibiting A β aggregation by coordinating with histidine residues.^{70,71} In 2008, Barnham *et al.* reported the first discrete coordination compounds active against AD, with the general formula [Phen-PtCl₂] (Phen: phenanthroline derivatives; Fig. 3).⁵⁴ These compounds, considered as analogues of cisplatin as they are based on labile chloride ligands and 1,10-phenanthroline derivatives, bind to histidine residues of A β ₄₂ acting as potent inhibitors of A β aggregation and neurotoxicity. In addition, the ability of these compounds to inhibit the metal-dependent biochemical and cellular actions of A β was studied. The authors demonstrated that all prepared compounds can inhibit A β :Cu^{II}-mediated redox chemistry with a half maximal inhibitory concentration (IC₅₀) of 0.3–0.6 μ M.

Following the same trend as for Pt-drugs in cancer treatment, in 2013, novel platinum(II) and (IV) complexes based on 8-(1*H*-benzimidazol-2-yl)-quinoline (8-BQ) were obtained showing high solubility and stability in aqueous media.⁵⁵ In neuronal cell cultures treated with A β ₄₂ (10 μ M, cell viability 77%), compound 4 ([Pt^{II}(*N,N*-dimethyl-2-[2-(quinolin-8-yl)-1*H*-benzimidazol-1-yl]ethanamine)Cl₂], 10 μ M) significantly



Table 1 Overview of the key information of discrete coordination compounds in the literature related to AD treatment

| Coordination compound | Target | Methods | Effectivity | Year ref. |
|--|---------------------------------------|--|---|--------------------|
| [Phen-PtCl ₂] | Aβ ₄₂ and Aβ ₄₀ | Spectroscopic techniques, ThT studies, <i>in vitro</i> , and <i>in vivo</i> | IC ₅₀ : 0.3–0.6 μM for ROS inhibition Cell viability: 80–100% (5 and 10 μM, Aβ ₄₂ 10 μM; BL6J × 129sv mouse cortical neurons) LTP: 150–160% (4 μM, Aβ ₄₂ 2 μM) | 2008 ⁵⁴ |
| [Pt ^{II} (8-BQ)Cl ₂] | Aβ ₄₂ | Spectroscopic techniques, ThT studies, and <i>in vitro</i> | Cell viability: 94% (10 μM, Aβ ₄₂ 10 μM; BL6J × 129sv mouse cortical neurons) LTP: 133% (2 μM, Aβ ₄₂ 2 μM) | 2013 ⁵⁵ |
| [Pt ^{IV} (8-BQ)Cl ₄] | | <i>In vivo</i> | Reduction of 40% in Aβ ₄₂ levels and 26% in Aβ plaques (Tg2576 mice) | |
| [Pt(BPS)Cl ₂] | Aβ ₁₆ and Aβ ₄₂ | Spectroscopic techniques | — | 2013 ⁵⁶ |
| [Pt(φ-MePy)(DMSO)Cl] | Aβ ₂₈ | Mass spectrometry and NMR spectroscopy | — | 2012 ⁵⁷ |
| [Pt(φ-MePy)(DMSO)Cl] | Aβ ₂₈ | ThT studies, mass spectrometry, and NMR spectroscopy | — | 2013 ⁵⁸ |
| [Pt(Phen)Cl ₂] | | | | |
| [Pt(φ-Phen)Cl ₂] | | | | |
| [Pt(Phen)Cl ₂] | Aβ ₁₆ | HPLC, ESI-MS and EPR spectroscopy | — | 2013 ⁵⁹ |
| <i>fac</i> -[Ru(CO) ₃ Cl ₂ (N ¹ -thz)] | Aβ ₂₈ | NMR and CD spectroscopies | — | 2010 ⁶⁰ |
| NAMI A | Aβ ₄₂ | <i>In vitro</i> | LDH released: 20% (PMRU20 40 μM), > 50% (NAMI A and KP1019 40 μM), 90% Aβ ₄₂ (E17 embryos rat neocortex) | 2013 ⁶¹ |
| KP1019 | | | | |
| PMRU20 | | | | |
| [(bpy) ₂ Ru ^{II} (dpp)Pt ^{II} Cl ₂]Cl ₂ | Aβ ₄₂ | Quantum chemical calculations and <i>in vitro</i> (ThT studies and western blot) | — | 2010 ⁶² |
| M(Flav) ₂ and M(X)(Flav) (M = Cu ^{II} and Zn ^{II} ; X: bpy or Phen) | AChE and BuChE | <i>In vitro</i> | 60–80% Inhibition for BuChE and > 90% for AChE | 2016 ⁶³ |
| Phthalocyanine Zn ^{II} and Ga ^{III} complexes | AChE and BuChE | <i>In vitro</i> | IC ₅₀ : 19.24–52.21 μM for BuChE, and 51.32–92.31 μM for AChE | 2021 ⁶⁴ |
| ZnPc ^p /ZnPc ^{np} | AChE and BuChE | <i>In silico</i> calculation and <i>in vitro</i> | Best IC ₅₀ values: AChE: MgPc ^{np} 25.16 μM BuChE: MgPc ^{np} 30.02 μM | 2022 ⁶⁵ |
| MgPc ^p /MgPc ^{np} | | | | |
| Amino acid Schiff base Zn ^{II} complexes | AChE and BuChE | Molecular docking calculations and <i>in vitro</i> | IC ₅₀ : 4.73–11.25 μM for BuChE, and 7.24–18.06 μM for AChE | 2022 ⁶⁶ |
| [Pd(glz) ₂] | AChE, BuChE, and Aβ ₄₂ | <i>In vitro</i> and <i>in vivo</i> | IC ₅₀ : 27 ± 1 μM for BuChE | 2022 ⁶⁷ |
| [Cu ^{II} (gtsm)] | Aβ and tau | <i>In vitro</i> and <i>in vivo</i> | LTP: 121.4% (1 μM, Aβ ₄₂ 2 μM) 48% Improved cognitive performance (10 mg kg ⁻¹ body weight; APP/PS1 transgenic AD mice) | 2009 ⁶⁸ |
| Macrocyclic platiniferous chelators | Aβ ₄₀ | <i>In vitro</i> and <i>in vivo</i> | Cell viability: 65–81% in the M ^{II} -Aβ system (Aβ ₄₀ 10 μM; C57BL6/J mice cortical neurons) | 2012 ⁶⁹ |

AChE, acetylcholinesterase; BPS, 4,7-diphenyl-[1,10]phenanthroline disulfonate; bpy, 2,2'-bipyridine; BuChE, butyrylcholinesterase; CD, circular dichroism; DFT, density functional theory; DMSO, dimethyl sulfoxide; dpp, 2,3-bis(2-pyridyl)pyrazine; EPR, electron paramagnetic resonance; ESI-MS, electrospray ionization mass spectra; Flav, flavanones; glz, gliclazide; gtsm, glyoxalbis(N(4)-methylthiosemicarbazonato); HPLC, high-performance liquid chromatography; LDH, lactic dehydrogenase; LTP, long-term potentiation; φ-MePy, 2-phenyl-5-methyl-pyridine; NMR, nuclear magnetic resonance; np, non-peripheral; p, peripheral; Pc, phthalocyanines; Phen, 1,10-phenanthroline; φ-Phen, 4,7-diphenyl-1,10-phenanthroline; ROS, reactive oxygen species; TEM, transmission electronic microscopy; ThT, thioflavin T; thz, 1,3-thiazole; 8-BQ, 8-(1H-benzoimidazol-2-yl)-quinoline.

inhibited Aβ aggregation and toxicity in neurons, achieving a 94% cell viability. Finally, in a way to prepare orally administered anti-amyloid compounds, the Pt^{IV} complex [Pt^{IV}(N,N-dimethyl-2-[2-(quinolin-8-yl)-1H-benzimidazol-1-yl]thanamine)Cl₄] was synthesized as a prodrug. It is known that the substitution reactions for Pt^{IV} complexes are kinetically very slow, and therefore, stable under acidic environments of the stomach. Then, Pt^{IV} complexes can be reduced by natural reductants (*i.e.*, glutathione), and become active. After the oral administration of the Pt^{IV} compound to the APP/PS1 mouse (used as the model of AD), a 40% reduction in Aβ₄₂ levels was found, demonstrating the potential of Pt^{IV} compounds in AD treatment.

The same year, Streltsov *et al.* reported a novel platinum complex, [Pt(BPS)Cl₂] (BPS: 4,7-diphenyl-[1,10]phenanthroline disulfonate), also active against the inhibition of Aβ₄₂ aggregation by binding to histidine residues.⁵⁶ Through different experimental techniques (X-ray absorption fine structure spectroscopy-XAS, mass spectrometry-MS and dynamic light scattering-DLS) and density functional theory-DFT modelling, the authors demonstrated the interaction of [Pt(BPS)Cl₂] and Aβ₄₂ through histidines 6 and 14, while the non-active cisplatin coordinates predominantly at the S atom of methionine 35. Inspired by Barnham's compounds, Sasaki *et al.* explored the reactivity of a new cyclometallated Pt^{II} complex, [Pt(φ-MePy)(DMSO)Cl] (φ-MePy: 2-phenyl-5-methyl-pyridine, DMSO: dimethyl



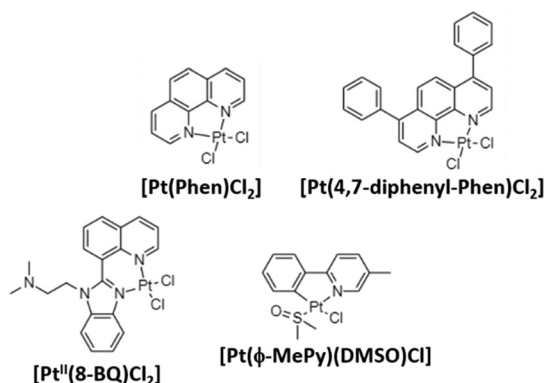


Fig. 3 Some reported Pt^{II} complexes against A β aggregation from Barnham,⁵⁴ Kenche,⁵⁵ and Sasaki *et al.*⁵⁷

sulfoxide),⁵⁷ with A β_{28} , since this peptide includes both the metal binding histidine residues and an hydrophobic part (residues 17–21) responsible for the nucleation process of A β aggregation. Compared to the Barnham [Pt(Phen)Cl₂] compound, this new complex is able to bind to A β_{28} and to modify the Cu^{II} coordination site in the A β :Cu^{II} adduct. Afterwards, the same group investigated the ability of Barnham's and Sasaki's Pt^{II} compounds (Fig. 3) to interfere in Zn^{II} coordination to the A β_{28} peptide, and Cu^{II}-induced ROS production,⁵⁸ but no significant inhibition in A β :Cu^{II} was induced in all the tested Pt^{II} compounds. On the other hand, regarding the Zn^{II}-induced A β_{28} aggregation, the fibril formation depends on the tested compound, decreasing in the following order: [Pt(Phen)Cl₂] and [Pt(φ-Phen)Cl₂] (no active) > [Pt(bpy)Cl₂] (bpy: 2,2-bipyridine) > [Pt(φ-MePy)(DMSO)Cl] and [Pt(DMSO)₂Cl₂] (more potent inhibitors). These differences were attributed to the lability of the linkers, since the more active compounds possess labile ligands (DMSO is more labile than chloride ligands), and thanks to the C–Pt bond in [Pt(φ-MePy)(DMSO)Cl], as well due to a strong *trans* effect.

In a similar work, Ma and coworkers also investigated the interaction between [Pt(Phen)Cl₂] and the adducts A β :Cu^{II} and A β :Zn^{II}, but using A β_{16} .⁵⁹ Their results showed that [Pt(Phen)Cl₂] preferentially targeted Cu^{II}/Zn^{II} binding sites and changed their binding modes, releasing metal ions under some conditions (*i.e.*, acidic media). As Pt^{II} is less reactive than Cu^{II} and Zn^{II}, the chelation of [Pt(Phen)Cl₂] might hamper the A β aggregation. Again, following a similar line of research by using Pt^{II} and Pt^{IV}, Ru^{II} compounds have also been studied as A β aggregation inhibitors. Authors selected this metal as it is well-established that ruthenium compounds are on the average far less toxic than platinum-based ones. Thus, in this work, the complex *fac*-[Ru(CO)₃Cl₂(N¹-thz)] (thz: 1,3-thiazole) was tested against A β_{28} .⁶⁰ The studies revealed that {Ru(CO)₃}²⁺ formed a stable adduct with A β_{28} coordinating with His13 and His14. Following this trend, the same group carried out *in vitro* assays against the AD cellular model with two well-known anticancer Ru^{III} compounds (NAMI A or [ImH][RuCl₄(DMSO)(Im)], KP1019 or [IndH]*trans*-[RuCl₄(Ind)₂], and PMRU20 or [2-AmThH]*trans*-[RuCl₄(2-AmTh)₂] (Im: imidazole; Ind: indazole; 2-AmTh: 2-aminothiazolium).⁶¹ The activity against AD was measured

through the released lactic dehydrogenase (LDH) which is normally delivered during cell damage or death.⁷² A β_{42} produced almost 100% of released LDH compared with the control group, whereas PMRU20 (40 μ M) reduced this release to 20%. Furthermore, PMRU20 showed a significant neuroprotective activity; as long as at the same concentration, NAMI A and KP1019 were poorly (*ca.* 80%) or nearly inactive (*ca.* 95%).

During their research on inhibiting the A β_{42} peptide aggregation, Kumar *et al.* obtained a Ru^{II}–Pt^{II} binuclear metal complex, [(bpy)₂Ru^{II}(dpp)Pt^{II}Cl₂]₂Cl₂ (dpp: 2,3-bis(2-pyridyl)pyrazine).⁶² First, authors demonstrated through *in silico* calculations that the binuclear complex exhibited higher binding affinities in terms of energetic properties than the mononuclear complex [(dpp)Pt^{II}Cl₂], suggesting that a second metal ion could improve the A β inhibition behavior. When tested *in vitro* using the primary mouse neuronal cell culture, this compound is able to inhibit A β_{42} aggregation in a similar manner to Barnham's compound with an IC₅₀ of \sim 4 μ M. Finally, western blot tests demonstrated the absence of fibrils and oligomeric species of A β , whereas Barnham's compound did not prevent the formation of intermediates of aggregation, demonstrating a different way of interaction.

Cholinesterases inhibition

A different mechanism to target AD is to inhibit cholinesterases, mainly acetylcholinesterase (AChE) and butyrylcholinesterase (BuChE) in order to avoid the reduction of acetylcholine levels. The research here is mainly focused on the use of different previously reported active organic moieties (flavonones, phthalocyanines, Schiff bases, and glioclazide; Fig. 4), and their coordination with different metals (Cu, Zn, Ga, and Pd), to obtain novel metallodrugs against AD.

In 2016, Sarria *et al.* reported seventeen Cu^{II} and Zn^{II} complexes based on flavanone derivatives (hesperidin, hesperetin, naringin, and naringenin), known for their neuroprotective effects, and studied their activity against AChE and BuChE.⁶³ Compared with the parent flavanones, nine of these complexes constituted potent, reversible, and selective anti-cholinesterase activity. Comparatively, Cu^{II} complexes possessed

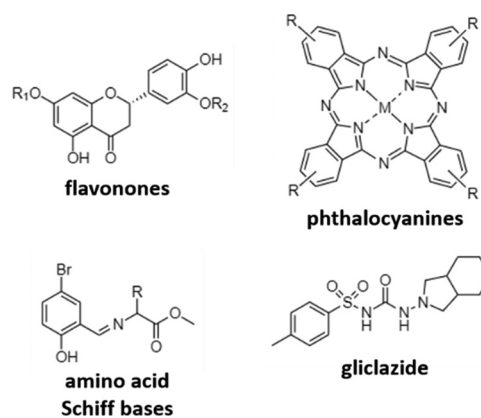


Fig. 4 Organic moieties used to obtain new coordination compounds as cholinesterase inhibitors.



higher inhibitory potency against AChE than the corresponding Zn^{II} complexes. Authors attributed these differences to the d⁹ electron configuration of Cu^{II}, which left two free empty orbitals for interaction with electron donor ligands, such as N atoms of histidine residues in AChE. In contrast, the d¹⁰ electron configuration of Zn^{II} imposes the tetrahedral geometry with no free empty orbitals to coordination. Furthermore, Cu^{II} complexes containing ancillary ligands (bpy or Phen) showed enhanced inhibitory activity, probably due to the coordination sphere of Cu^{II} that favors the π - π stacking interactions. Phthalocyanines are interesting synthetic molecules similar to the heme group with pharmacological and biological applications.⁷³⁻⁷⁶ Based on these molecules, Günsel and coworkers obtained Zn^{II} and Ga^{III} complexes and their water-soluble hydrochloride derivatives, and tested their inhibitory activity against AChE and BuChE.⁶⁴ All complexes showed high BuChE inhibitory activity, with IC₅₀ values in the range of 19.24–52.21 μ M, and 51.32–92.31 μ M for BuChE and AChE, respectively. These results were further improved compared to those obtained for tacrine (178.03 and 141.34 μ M, respectively), which is the first drug introduced into the United States market to manage AD and used here as the standard inhibitor. In a different study using again phthalocyanines, Yalazan *et al.* obtained Zn^{II} and Mg^{II} quinoline-fused phthalocyanine (Pc) compounds (ZnPc^p/ZnPc^{np} and MgPc^p/MgPc^{np}; p: peripheral, np: non-peripheral) in order to combine interesting therapeutic properties with spectroscopic characteristics.⁶⁵ Among them, MgPc^{np} exhibited the highest inhibitory potential against AChE and BuChE with IC₅₀ values of 25.16 and 30.02 μ M, respectively. *In silico* calculations showed that MgPc^{np} had the highest HOMO value and the lowest ΔE energy value, demonstrating that it is the most reactive species. This fact together with its high solubility might explain its great inhibitory effect compared to the other synthesized compounds. However, although MgPc^{np} demonstrated to be efficient in the inhibition of AChE, it presented a lower inhibitory effect than galantamine (an approved drug in the treatment of AD and used as a positive control in this study).

Due to their interesting biological properties, other organic moieties used in the preparation of metallodrugs to treat AD are Schiff bases. In 2021, Şenocak *et al.* synthesized some compounds based on Zn^{II} and amino acid Schiff bases and tested them against AChE and BuChE.⁶⁶ First, amino acid Schiff bases were prepared using 5-bromo-2-hydroxybenzaldehyde and amino acid methyl esters (isoleucine, phenylalanine, and methionine). Once synthesized, the potential inhibition of Zn-based metallodrugs to cholinesterases was explored. The synthesized compounds were found to be highly efficient against BuChE and AChE, obtaining IC₅₀ values in the range of 4.73–11.25 μ M and 7.24–18.06 μ M, respectively, surpassing the inhibitory activity of tacrine.

Recently, our research group published a complex based on palladium and the antidiabetic drug gliclazide (glz), [Pd(glz)₂].⁶⁷ Originally, we selected glz as the organic moiety in the preparation of AD's metallodrugs since several factors associate *Diabetes mellitus* with AD: (i) it is a risk factor for Alzheimer's disease, (ii) it has a similar molecular mechanism of action, and (iii) it is

associated with protein misfolding.^{77,78} Thus, the potential of [Pd(glz)₂] against AD was explored, obtaining the preferential inhibition of BuChE against AChE (91 vs. 58% inhibition), with an IC₅₀ value of 27 μ M for BuChE, as it is capable of interacting with both the free enzyme and the enzyme–substrate complex. Remarkably, its potential activity *in vivo* was also evaluated using *Caenorhabditis elegans* as the animal AD model. Aside from the absence of lethal toxicity, [Pd(glz)₂] was able to increase the percentage of non-paralyzed worms by interacting with A β ₄₂ peptides and inhibiting its aggregation, activity probably associated with the planar structure of the complex (as previously reported for other metallic complexes).

Effects in metal chelation

A β plaques have abnormally high concentrations of metals such as Cu, Zn, and Fe, which promote the A β peptide aggregation. In agreement with these observations, some researchers considered the development of novel metalorganic drugs using known compounds that target abnormal metal accumulation. For example, thiosemicarbazones and bis-thiosemicarbazones have a wide range of pharmacological activities linked to their ability to chelate transition metals.⁷⁹ These compounds elevate intracellular levels of metals, resulting in the activation of phosphoinositol-3-kinase (PI3K) and the inhibition of glycogen synthase kinase 3 (GSK3), which lead to the reduction of neurotoxic A β and tau phosphorylation. In addition, although further research studies are needed, some studies show that these compounds could cross the BBB.^{80,81} One example is the complex [Cu^{II}(gtsm)] (gtsm: glyoxalbis(*N*(4)-methylthiosemicarbazonato), which promotes the expression of A β -degrading pathways (Fig. 5).^{82,83} Both, *in vitro* and *in vivo* studies, demonstrated that [Cu^{II}(gtsm)] increases intracellular Cu^{II}, which is reduced to Cu^I, leading to copper dissociation from the ligand and increasing Cu bioavailability. This induces the cellular pathways that inhibit GSK3 β , which regulates the accumulation of A β peptides and tau phosphorylation. Therefore, this compound is a promising therapy for AD as it attenuates both the A β - and tau-mediated pathologies.⁶⁸

In 2015, Wang *et al.* developed macrocyclic platiniferous chelators containing cyclen (1,4,7,10-tetraazacyclododecane) that bind to metals such as Zn^{II} and Cu^{II}, and the unit [Pt(bpy)Cl₂] that coordinates to histidine residues of A β ₄₀.^{69,84} When tested in the cortical neuronal cell of C57BL6/J mice, these compounds demonstrated the inhibition of A β ₄₀ aggregation induced by Zn^{II} or Cu^{II} and the production of ROS by the Cu^{II}-A β complex. Furthermore, when tested *in vivo*, these compounds reduced the extent of A β aggregation in brain homogenates of transgenic mice. The authors argue that these powerful inhibitions might be promoted by their bifunctional active characters rather than their individual units. In addition, the results were compared with cisplatin, showing that, in contrast, it aggravated the metal-induced A β aggregation as cisplatin coordinates with methionine rather than the metal binding site of A β formed by histidine residues.

Finally, a brief mention to combat AD through metal (Cu, Zn, Fe) chelation with purely organic moieties (organic



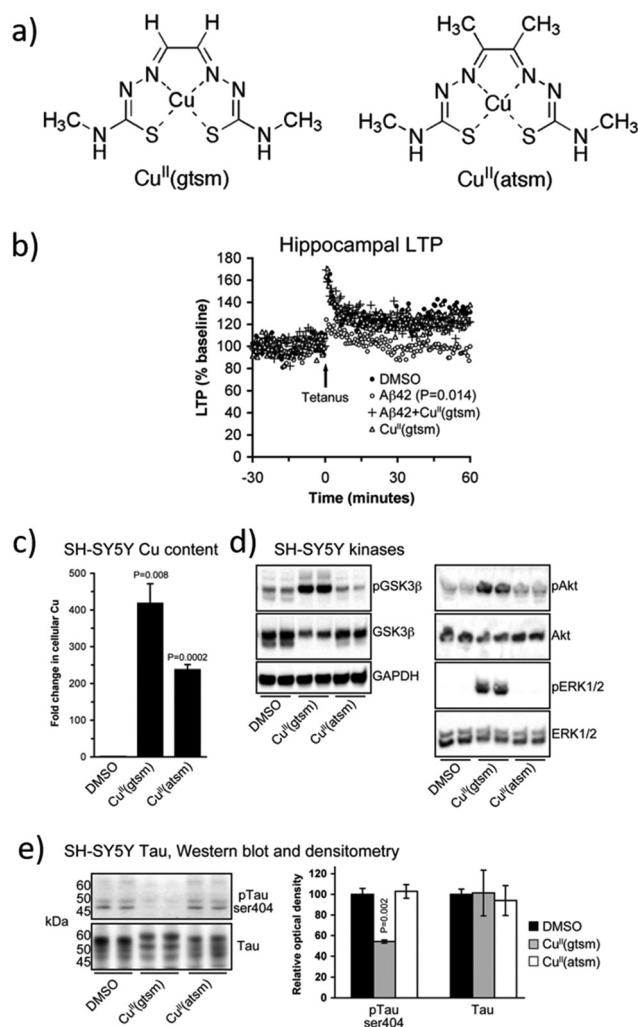


Fig. 5 (a) Structures of [Cu^{II}(gtsm)] and the control [Cu^{II}(atsm)]. (b) Whereas Aβ₄₂ inhibits the hippocampal long-term potentiation (LTP) by 28%, this is prevented when treated with [Cu^{II}(gtsm)]. (c) *In vitro*, [Cu^{II}(gtsm)] increases the cellular levels of Cu. (d) Western blot analyses show that [Cu^{II}(gtsm)] induces the phosphorylation of kinases GSK3β, Akt, and ERK1/2. (e) [Cu^{II}(gtsm)] inhibits tau phosphorylation at ser404 without affecting the abundance of total tau. Adapted from Ref. 68. Reproduced with permission.

drugs^{85–87} and small peptides^{88,89} is made. One outstanding example is the effective use of quinoline derivatives like clioquinol and 8-hydroxyquinoline which have entered clinical trials for AD.^{90,91}

MOFs as alternative platform in AD treatment

MOFs can adsorb a wide range of molecules. Particularly in biomedicine, MOFs have been used to control the dose and delivery kinetics of previously adsorbed active ingredients (drugs, gases, *etc.*),⁴⁶ or as adsorbent detoxifying agents in the treatment of oral overdoses.^{92,93} Very recently, some research groups have focused their efforts in the application of MOFs in AD treatment (Table 2). One of the first examples is the work of Mensinger *et al.* who originally studied the Aβ₄₀ peptide adsorption capability of seven archetypal MOFs (HKUST-1, MIL-53, MIL-69, MIL-88B, MIL-100(Fe), MIL-101(Cr)NDC, and UiO-66).⁹⁴ All MOFs, except MIL-100(Fe), exhibited complete (100%) and quick (10 min) adsorption of Aβ₄₀. Furthermore, authors checked that the intact peptide can be desorbed from HKUST-1 and UiO-66, suggesting that it is adsorbed rather than being hydrolyzed. Although MOFs were demonstrated to be excellent adsorbents of Aβ₄₀, in this work some important issues related to the MOF particle size and their potential passage through the BBB to obtain the peptide have not been considered.

Another approach in the use of MOFs in AD treatment is through the oxygenation of Aβ as a way to suppress its aggregation and neurotoxicity. In 2018, Wang *et al.* proposed the porphyrinic Zr^{IV} metal-organic framework (PCN-224) as a platform in the inhibition of Aβ aggregation by near-infrared (NIR)-activated photooxygenation.⁹⁵ The use of porphyrinic ligands give rise MOFs with great porosity that enhances the accessibility of oxygen and subsequently, the photo-induced ¹O₂ generation. First, the synthesis of PCN-224 was improved to obtain small nanoparticles, as it is known that to cross the BBB the nanoparticle size should be within 10–100 nm. Field-emission scanning electron microscopy (FESEM) images

Table 2 Overview of the key information reported for the application of MOFs in AD treatment

| MOF or MOF composite | Target | Methods | Effectivity | Year ref. |
|--|--|---|---|--------------------|
| HKUST-1, UiO-66, MIL-53, MIL-69, MIL-88B, MIL-100(Fe) and MIL-101(Cr)NDC | Aβ ₄₀ adsorption | <i>In vitro</i> and gel electrophoresis | — | 2020 ⁹⁴ |
| PCN-224 | Aβ ₄₂ photooxygenation | ThT studies, TEM, gel electrophoresis and <i>in vitro</i> | Cell viability: 90% under 650 nm light (PC12 cells) | 2018 ⁹⁵ |
| PMOFs | Aβ ₄₀ photooxygenation Cu ^{II} chelator | Spectroscopic techniques, DFT calculations and <i>in vivo</i> | Hf-MOF results: ROS production: 173 PL intensity Cu-chelating: 0.5 g g ⁻¹ Aβ enrichment: 4.1 μmol mg ⁻¹ | 2019 ⁹⁶ |
| CeRMS | Enhance neurogenesis and reduce ROS | <i>In vitro</i> and <i>in vivo</i> | — | 2020 ⁹⁷ |

DFT, density functional theory; PMOF, porphyrinic metal-organic framework; ROS, reactive oxygen species; TEM, transmission electronic microscopy; ThT, thioflavin T.



demonstrated that the PCN-224 nanoparticle average size was around 70 nm. However, no data about the particle size of PCN-224 in solutions (water, phosphate buffer saline-PBS, etc.) were reported, which is mandatory to discard the potential aggregation of nanoparticles before their possible administration. Results on the photoinduced inhibitory effect of PCN-224 in the aggregation of A β ₄₂ peptides showed that after irradiation under 650 nm light, thioflavin T (used to monitor the aggregation degree) fluorescence intensity remained low; hence, the aggregation of A β ₄₂ was inhibited. Finally, *in vitro* studies in PC12 cells (from transplantable rat pheochromocytoma) showed an increase in cell viability from 40 to 90% when A β ₄₂ was incubated with PCN-224 under 650 nm light irradiation. This fact could be explained since the physical adhesion of A β ₄₂ to the PCN-224 surface may reduce the active sites in A β , and therefore, its aggregation, increasing the efficiency of photooxygenation.

In a further study, Yu and coworkers reported the ability of four already known porphyrinic based MOFs (PMOFs) as Cu^{II} chelating (as porphyrin is a good chelator) and photooxidation agents for inhibiting A β aggregation.⁹⁶ Among the PMOFs studied (Zr-MOF, Al-MOF, Ni-MOF, and Hf-MOF), the Hf-MOF was the best A β oxidant because of its great ability for inducing ROS generation, and the efficient removal of Cu^{II} (0.5 g of Cu per g of the PMOF). In addition, thanks to its porosity, the Hf-MOF enriched A β at a concentration of 4.1 $\mu\text{mol mg}^{-1}$, which can minimize the side effects of ¹O₂ to normal cells, and its oxidation occurred even after binding with Cu^{II}. In the presence of the Hf-MOF, Cu^{II}-induced A β aggregation was reduced and completely suppressed when light was irradiated. Afterwards, the Hf-MOF was modified with a A β -targeting peptide LPFFD (Lys-Pro-Phe-Phe-Asp), thus enhancing specificity to A β and photooxidation efficiency. This modification increased the percentage of oxidized A β (87% vs 66% without the LPFFD), and the reduced cytotoxicity of A β and Cu^{II}-induced A β aggregation. Finally, *in vivo* studies verified the inhibition effects of the LPFFD-modified Hf-MOF by a commonly used AD transgenic *C. elegans* model (Fig. 6).

In another study, the same research group modulated the neuron differentiation and amelioration of oxidative stress by integrating antioxidative nanozymes (ceria) into a MOF for synergically enhanced neurogenesis, which contributes to self-repair after neuron loss.⁹⁷ The main issue is that the high level of oxidative stress in AD promotes the neural stem cell (NSC) death and neuronal apoptosis. The nanoplatfrom, abbreviated as CeRMS (CeNPs/RA@MIL-100/siSOX9), contains Ce (for the elimination of ROS based on the electron transfer Ce^{III}-Ce^{IV}), and MIL-100, which also possesses an antioxidative effect, loaded with siSOX9 (the regulator of the SOX9 protein and an essential transcription factor beneficial to gliogenesis) and retinoic acid (RA, the regulator in the differentiation of the NSC to neurons). First, the RA release was studied in H₂O₂ (1 mM). Under these conditions, CeRMS was destroyed and 40% of RA was released after 15 min of treatment. Furthermore, the ability of CeRMS to modulate the NSC differentiation was tested. Whereas other systems without RA, such as CeM, CeRM, and

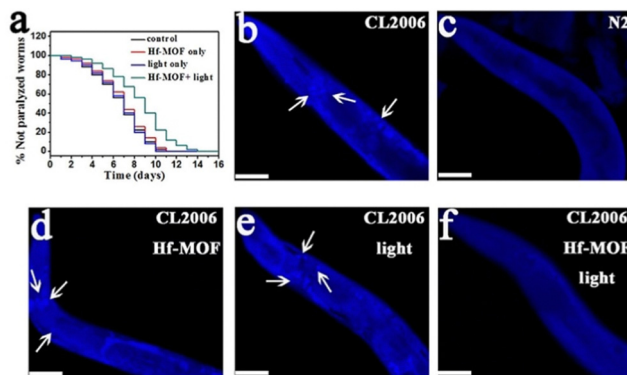


Fig. 6 Effect of the Hf-MOF in *C. elegans*. (a) Kaplan–Meier survival curves of the transgenic strain AD CL2006 show a delay in the A β -triggered paralysis of worms when treated with the Hf-MOF under light. (b–f) Images of the worm's head region of CL2006 (arrows indicate A β deposits): (b) transgenic CL2006 worm, (c) Bristol N2 worm (wild type), (d) CL2006 treated with the Hf-MOF, (e) CL2006 with light, and (f) CL2006 treated with the Hf-MOF under light. Reproduced from ref. 96 with permission.

CeMS, used here as controls, differentiated NSC into glial cells, CeRMS, containing RA, completely differentiated the NSC into neurons. Additionally, under both *in vitro* and *in vivo* conditions, CeRMS reduced the intracellular levels of ROS better than ascorbic acid and, consequently, it decreased the number of apoptotic cells caused by A β ₄₂ aggregation. Overall, this study provides an original synergic strategy, promoting nerve regeneration while protecting from oxidative stress.

MOFs in the detection of the AD

AD progresses asymptotically for years (up to 20 years depending on age, sex, genetic predisposition, etc.) before the appearance of symptoms.⁹⁸ Thus, the early diagnosis of AD might have a major impact on the progress and efficient timely treatment of the disease. As previously mentioned in the Introduction section, AD begins with the deposition of A β , followed by the appearance of tau tangles, and then by cognitive impairment.⁹⁹ Initially, the AD diagnosis was a result of neurological, psychiatric and clinical examinations, neuropsychological tests, and laboratory studies.¹⁰⁰ It was in 2011, and based on neuroimaging techniques and CSF analysis, when the National Institute on Ageing and Alzheimer's Association (NIA-AA) workgroup incorporated into the clinical criteria AD pathology biomarkers, such as A β deposition in the brain, A β ₄₂, and the pathologic tau level in the CSF. Currently, three biomarkers have been well-established and validated to diagnose AD: A β _{1–42}, total tau, and phosphor-tau-181.¹⁰¹

Aside from high concentrations of metal ions (*i.e.*, Cu, Zn, and Fe) in the brain of AD patients, biomolecules such as glutathione (GSH), a key regulator for the redox balance of biological processes;¹⁰² or protein kinases, responsible for the catalysis in the phosphorylation of protein substrates or specific peptides;¹⁰³ have been proposed as biomarkers. However, only amyloid plaques and tau tangles are specific features that



Table 3 Overview of the key information reported in the application of MOFs in AD detection

| MOF | Biomarker | Detection method | Type of sample | Linear range LOD | Year ref. |
|---|------------------------------------|-----------------------------------|-----------------------|--|---------------------|
| Fc@IR-MOF-3 | A β proteins | Electrochemical | Human serum | 10 ⁻⁴ –100 ng mL ⁻¹ 0.03 pg mL ⁻¹ | 2017 ¹⁰⁵ |
| TMBDA-MIL-100(Fe) | Choline Acetylcholine | Photoluminescence | Buffer solution | 0.5–10 μ M 27 nM 0.1–10 μ M 36 nM | 2018 ¹⁰⁷ |
| AuNPs/Cu-MOF | A β O | DPV | aCSF | 1 nM–2 μ M 0.45 nM | 2018 ¹⁰⁸ |
| Cu-BTC/Tb | A β peptide | Fluorescence | Blood | 1–550 nM 0.3 nM | 2018 ¹⁰⁹ |
| Fc@ZIF-8 | A β O | UV-vis and CV | Real samples | 10 ⁻⁵ –10 ² μ M 10 ⁻⁵ μ M | 2019 ¹¹⁰ |
| Ru@MOF | A β proteins | ECL-RET | Buffer solution | 10 ⁻⁵ –500 ng mL ⁻¹ 3.9 fg mL ⁻¹ | 2019 ¹¹¹ |
| Co-MOFs/ABEI | A β proteins | ECL | Human serum | 10 fg mL ⁻¹ –100 ng mL ⁻¹ 3 fg mL ⁻¹ | 2019 ¹⁰⁶ |
| Fe ₃ O ₄ @PPy-Au | A β proteins | SWV | Serum | 10 fg mL ⁻¹ –100 ng mL ⁻¹ 3.3 fg mL ⁻¹ | 2019 ¹¹² |
| UiO-66@PANI-MB | A β proteins | SWV | Serum | 10 fg mL ⁻¹ –100 ng mL ⁻¹ 3.3 fg mL ⁻¹ | 2019 ¹¹² |
| Cu-Al ₂ O ₃ -g-C ₃ N ₄ -Pd | A β proteins | ECL | aCSF | 10 ⁻⁴ –100 ng mL ⁻¹ 13.8 fg mL ⁻¹ | 2019 ¹¹³ |
| [Ru(bpy) ₃][Zn ₂ (C ₂ O ₄) ₃] + Au@NiFe MOF | A β proteins | ECL | aCSF | 10 ⁻⁴ –100 ng mL ⁻¹ 13.8 fg mL ⁻¹ | 2019 ¹¹³ |
| Pd NPs@MIL-53(Al)-NH ₂ | A β proteins | ECL | A β solutions | 10 fg mL ⁻¹ –50 ng mL ⁻¹ 3.4 fg mL ⁻¹ | 2019 ¹¹⁴ |
| Ru@MIL-101(Al) | A β O | Fluorescence | Blood | 1–10 000 pM 0.3 pM | 2020 ¹¹⁵ |
| Ru(bpy) ₃ ²⁺ /NH ₂ -UiO-66 and MIL-101@Au-MoS ₂ | A β proteins | ECL | Buffer solution | 0.00001–50 ng mL ⁻¹ 3.27 fg mL ⁻¹ | 2020 ¹¹⁶ |
| MOF-armed-anti-DNA antibody | A β O | Fluorescence | Blood | 0.001–100 ng mL ⁻¹ 0.4 pg mL ⁻¹ | 2020 ¹¹⁷ |
| ThT@Er-MOF | Presenilin 1 | Fluorescence | CSF | 0–40 nM 0.142 nM | 2020 ¹¹⁸ |
| | A β protein Acetylcholine | Fluorescence Photoluminescence | | 0–33 nM 0.517 nM 0–10 nM 0.03226 nM | |
| AuNPs@CuMOF | A β O | DPV | aCSF | 0.5–500 fM 0.25 fM | 2021 ¹¹⁹ |
| H-USM-SiO ₂ @ZIF-8/BHQ-1 | A β O | Fluorescence | A β O solutions | 0.01–10 μ M 28.4 pM | 2021 ¹²⁰ |
| AuNPs@MIL-88(Fe)-NH ₂ | A β O | ECL | CSF | 0.1–10 pM 71 fM | 2021 ¹²¹ |

ABEI, *N*-(aminobutyl)-*N*-(ethylisoluminol); A β O, β -amyloid oligomer; aCSF, artificial cerebrospinal fluid; BHQ-1, black hole quencher; bpy, 2,2'-bipyridine; BTC, 1,3,5-benzenetricarboxylate; CV, cyclic voltammetry; DPV, differential pulse voltammetry; ECL, electrochemiluminescence; Fc, ferrocene; g-C₃N₄, graphite carbon nitride; H-USM, highly doped upconversion nanoparticles-SiO₂@MOF; LOD, limit of detection; MB, methylene blue; NPs, nanoparticles; PANI, polyaniline; PPy, polypyrrole; RET, resonance energy transfer; SWV, square wave voltammetric; ThT, thioflavine T; TMBDA, *N,N,N',N'*-tetramethyl-1,4-butanediamine.

define AD as a unique neurodegenerative disease among other disorders.¹⁰⁴ In this review, only the use of MOFs in the detection of A β and tau is discussed in detail (Table 3).

In the first example of the potential of MOFs in A β detection reported by Han *et al.*,¹⁰⁵ the postsynthetic modification of the Zn based material IR-MOF-3 was used as the key to access to this specialized application. Particularly, the active ferrocene (Fc, frequently used as the signal tag) is covalently combined in the porous MOF through an amination reaction (Fig. 7a), leading to a composite with high-content Fc signal units well distributed with enhanced electrochemical activity. Furthermore, the encapsulation of Fc into the MOF confers some stability and avoids the leakage of the signal molecule. In a further step, the outer surface of the Fc@IR-MOF-3 composite was modified with gold nanoparticles (AuNPs) as platforms to label with detection antibodies (Ab₂). The stability of Fc@IR-MOF-3 was assessed under extreme conditions, such as strong

acid/base and polar organic solvents, through the relative electrochemical activity of Fc (Fig. 7b). More than 95–97% of the Fc activity was retained when using a pH range from 3 to 10, and different solvents. These results were further better than the ones obtained in the free Fc, where only 15–34% of the Fc activity was retained, respectively, suggesting that the interaction between Fc and the MOF improves the robustness of the signal molecule. The analytical performance of the immunosensor was tested in PBS, achieving a detection limit of 0.03 pg mL⁻¹. Furthermore, the selectivity of the immunoassay for the detection of target A β was analyzed against potential interfering substances (human serum albumin, thrombin, immunoglobulin, *etc.*). Despite the high concentration of the potential interfering substances (100 ng mL⁻¹) compared with A β (10 ng mL⁻¹), no differences were observed in the square wave voltammetry (SWV) responses. Thus, the Fc@IR-MOF-3 composite could selectively discriminate the



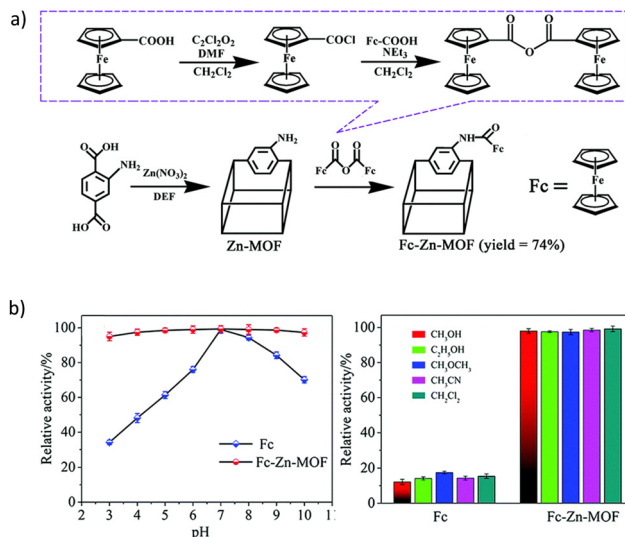


Fig. 7 (a) Schematic diagram of the post-synthetic modification of IR-MOF-3 with Fc. (b) Electrochemical activities of Fc and Fc@IR-MOF-3 at different pHs (3 to 10) and in PBS containing 50% of selected organic solvents. Reproduced from Ref. 105 with permission from the Royal Society of Chemistry.

target A β from other non-target proteins. Finally, the authors confirmed the possible application of the designed immunosensor in human serum samples, obtaining recoveries of 90.0 to 110.3%, which prove the potential of this material for the detection of A β in real samples.

In a subsequent report and using a one-pot synthetic method, an analog of luminol (*N*-(aminobutyl)-*N*-(ethylisoluminol), named ABEI) was incorporated into a Co-based MOF.¹⁰⁶ The potential of luminol and its derivatives in electrochemiluminescence (ECL) lies in their low oxidation potential, high luminous efficiency, and nontoxicity. The obtained Co-MOF/ABEI was decorated with AuNPs as platforms to label with detection antibodies. Incremental ECL intensities were obtained with the increasing A β_{42} concentration, achieving a detection limit (0.003 pg mL⁻¹) ten times better than the first MOF reported for this application. The feasibility of this proposed immunoassay in AD diagnosis was tested in human serum samples, obtaining excellent recoveries (96.0–104.0%).

The sensitivity towards A β oligomers (A β O) was again improved in a further study where an aptasensor was prepared based on a luminescence nanoscaled lanthanum-based MOF.¹¹⁷ The system was designed based on the MOF-armored single-stranded DNA antibody (anti-DNA antibody@MOF) and aptamer magnetic beads (Apt-Fe₃O₄) as capture probes. Again, the strategy followed in the preparation of the signal probe (anti-DNA antibody@MOF) was a bottle around the ship mechanism. The mineralization of the L-MOF around the anti-DNA antibody forms an exoskeletal cage, potentially protecting the encapsulated antibody. This A β O assay demonstrated high sensibility and selectivity, with a detection limit of 0.4 pg mL⁻¹, even in samples with chief components in blood (K⁺, Na⁺, glucose, immunoglobulin, etc.). Compared with other studies on A β O detection based on MOFs, here the reusability of the aptasensor was investigated.

The aptamer was easily regenerated from the complex A β O/Apt-Fe₃O₄ by heating at 50 °C. The aptasensor could be reused 10 times while maintaining its activity. Furthermore, after 7 weeks at room temperature, the aptasensor retained ca. 80% of its fluorescence intensity. This advantage in the long-term preservation activity can potentially solve the problems of manufacturing cost aptasensors.

In a recent study, ZIF-8 loaded with black hole quencher molecules (BHQ-1) was used as a coating for the efficient inactivation of the fluorescence of highly doped upconversion nanoparticles (H-UCNPs) in NH₂-SiO₂ microspheres.¹²⁰ BHQ-1 loaded ZIF-8 exhibited an excellent quenching efficiency for upconversion emission. In contrast, when different concentrations of A β O are present, the upconversion luminescence was recovered due to the decomposition of ZIF-8 and the release of BHQ-1. Authors confirmed that after adding A β O, the competitive binding of Zn^{II} and amino acids of A β O will induce the disassembly of ZIF-8. However, considering that ZIF-8 particles are rather unstable in the serum and most common buffers,^{122–124} a crucial and deep study of the ZIF-8 composite stability in different biological media or in the presence of other competitive biomolecules is necessary for the potential application of this composite material in real samples.

Sensing and treatment for AD (theragnosis)

Theragnosis is the combination of diagnosis and therapy in an effective formulation. Since it is a term fairly new, there is growing interest in this field. Currently, theragnosis is extensively studied in cancer by using mainly nanoparticles and nanocomposites.^{125–127} However, in AD, only few works have been carried out in theragnosis by using both discrete coordination compounds and MOFs (Table 4).

In 2011, Man *et al.* reported the first example of coordination compounds as inhibitors of A β aggregation and luminescent probes for A β_{40} peptides ([Ir(ppy)₂(H₂O)₂]³⁺, [Rh(ppy)₂(H₂O)₂]³⁺, [Ir(bzq)₂(H₂O)₂]³⁺, and [Ir(phq)₂(H₂O)₂]³⁺; ppy: 2-phenylpyridine, bzq: 7,8-benzoquinoline, and phq: 2-phenylquinoline).¹²⁸ They inhibited A β_{40} aggregation *in vitro* since the labile H₂O ligands are replaced by histidine residues and the aromatic ligand form stacking interactions with the peptide. Interestingly, complexes with extended aromatic ligands (*i.e.*, bzq and phq) were less active, probably due to the impediment of the metal center to coordinate with histidine residues. Among them, the Rh^{III} complex showed the highest inhibitory activity with a complex to A β_{40} monomer ratio of 1:10, thus subsequent *in vitro* experiments were carried out. The cytotoxicity study against neuroblastoma cells showed an IC₅₀ value between 25 and 50 μ M, higher than the concentration needed for complete A β_{40} aggregation inhibition. Finally, the luminescence properties of the Ir^{III} complexes showed an increased emission in the presence of histidine solution compared to aqueous buffer.

Methylene blue (MB) is a well-studied tau aggregation inhibitor, but it is quickly dispersed after crossing the BBB, resulting in low bioavailability. In this regard, Zhao and



Table 4 Overview of the key information reported in the application of coordination compounds in AD sensing and treatment

| Complex | Target | Methods | Sensing | Treatment | Year ref. |
|--|---|--|---|---|--|
| [Ir(ppy) ₂ (H ₂ O) ₂] ³⁺ [Rh(ppy) ₂ (H ₂ O) ₂] ³⁺ [Ir(bzq) ₂ (H ₂ O) ₂] ³⁺ [Ir(phq) ₂ (H ₂ O) ₂] ³⁺ | Aβ ₄₀ | <i>In vitro</i> (ThT fluorescence and mass spectrometry) | [Ir(ppy) ₂ (H ₂ O) ₂] ³⁺ 56-fold with Aβ ₄₀ monomer and 134-fold with Aβ ₄₀ fibrils | Rh ^{III} complex decrease Aβ ₄₀ length from 6 to 1.3 μm IC ₅₀ 25–50 μM | 2011 ¹²⁸ |
| Fe-MIL-88B-NH ₂ -NOTA-DMK6240/MB [Tb ₄ (cdda) ₆ (H ₂ O) ₄](DMF) | Tau protein Cu ²⁺ ions, <i>nf-κb</i> , <i>akt</i> , and cytokines | <i>In vitro</i> (MTT and ThT methods) and <i>in vivo</i> <i>In vitro</i> and <i>in vivo</i> | — $K_{sv} = 2,021.8 \text{ M}^{-1}$ | 63% inhibition of tau aggregation <i>nf-κb</i> and <i>akt</i> : ~1 of relative expression. IL-1: 100–125 ng mL ⁻¹ IL-6: 140–160 ng mL ⁻¹ | 2020 ¹²⁹ 2020 ¹³⁰ |
| CeONP-Res-PCM@ZIF-8/PDA | Aβ oligomers | <i>In vitro</i> | 95% of quenching efficiency Detection limit 3.2 nM | 67.23% Res release 93.87% antioxidant effect | 2021 ¹³² |

bzq, 7,8-benzoquinoline; CeONP, ceria nanoparticles; DMF, *N,N*-dimethylformamide; H₂cdda, 4,4'-(9*H*-carbazole-2,7-diyl)dibenzoic acid; K_{sv} , metal ion quenching coefficient; MB, methylene blue; MTT, 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide; NOTA, 1,4,7-triazacyclononane-1,4,7-triacetic acid; PCM, 1-tetradecanol; PDA, polydopamine; phq, 2-phenylquinoline; ppy, 2-phenylpyridine; Res, resveratrol; ThT, thioflavin T.

coworkers synthesized the first MOF containing MB focused on theragnosis against hyperphosphorylated tau proteins.¹²⁹ This nanoscale MOF based on Fe-MIL-88B-NH₂ was obtained as a multifunctional material that integrates magnetic imaging (from Fe atoms), targeted therapy, and drug delivery. The nanoplatform Fe-MIL-88B-NH₂-NOTA-DMK6240/MB was prepared by post-synthetic modification of the surface of Fe-MIL-88B-NH₂ (acts as a nanocarrier and a good magnetic resonance imaging-MRI contrast agent) with NOTA (1,4,7-triazacyclononane-1,4,7-triacetic acid, a macrocyclic linker that binds DMK6240 to the surface), and DMK6240 (hyperphosphorylated tau-directing unit). Finally, MB was loaded into the NMOF, achieving a total particle size of 190–240 nm. After verifying the hyperphosphorylated tau-target ability, MRI capability, and the lack of toxicity, the inhibition of tau aggregation was tested. The nanoplatform could produce 63% of inhibition in tau aggregation and had a neuroprotective effect reaching 80% of cell survival. In addition, learning and memory ability tests *in vivo* indicated that the ability of targeting and the inhibition of tau hyperphosphorylation contributed to improve memory loss in AD model rats. Finally, the degradation assays of the NMOF showed that both Fe and ligand concentrations reached a maximum on the 7th day and then were gradually reduced, demonstrating its biodegradability.

In a subsequent work, a lanthanide 2D polymer based on Tb^{III} was studied due to its interesting luminescence properties as a Cu^{II} sensor.¹³⁰ Here, the compound [Tb₄(cdda)₆(H₂O)₄](DMF) (H₂cdda: 4,4'-(9*H*-carbazole-2,7-diyl)dibenzoic acid, DMF: *N,N*-dimethylformamide) was incubated with several transition metals and the photoluminescence was measured. Whereas all metal ions just improved or reduced the luminescence strength of the complex, Cu^{II} exhibited a strong quenching action with a quenching coefficient of 2021.8 M⁻¹, higher than other Ln-MOF sensors. This effect is a result of the smaller Cu^{II} radius, which allows the easy interaction with the N atom of the ligand cdda and, therefore, decreases the ligands' antenna efficiency and reduces the energy transfer of the compound. On the other hand, since the inflammatory response on neurons

could lead to AD, the anti-inflammatory effect of the compound was tested. After injecting Aβ in rats, the *nf-κb* and *akt* levels were much higher than the control, but when the polymer was administered, these levels were lowered to normal concentrations. Furthermore, this compound also reduced the levels of the cytokines IL-1 and IL-6 in the CSF, which are responsible for the decrease in the production of the inflammatory response.

In the most recent work about theragnosis in AD, Yan *et al.* synthesized a nanocomposite as a platform for the detection and inhibition of Aβ₀ fibrillation.¹³¹ The base of the nanostructure is ZIF-8, since it is a good nanocarrier in biomedicine and has fluorescence quenching properties. The system, CeONP-Res-PCM@ZIF-8/PDA, contains polydopamine (PDA) that enhances the binding ability of ZIF-8 towards biomolecules and acts as a photothermal agent with strong NIR absorption. The nanostructure was loaded with resveratrol (Res) which has a therapeutic effect for AD, 1-tetradecanol (PCM) that acts as a thermal switch to NIR-control Res release, and CeONP that functions as the ROS scavenger based on the electron transfer Ce^{III}-Ce^{IV}. The nanostructure had a particle size of 106 nm, a surface area of 163.82 m² g⁻¹ and a pore size of <1 nm. *In vitro* measurements demonstrated that the compound doped with a specific-Aβ₀ aptamer showed high fluorescence in the presence of Aβ₀ over Aβ monomers and fibrils. In addition, the fluorescence was maintained in SH-SY5Y cells, demonstrating the cell membrane permeability of the nanocomposite. On the other hand, under NIR irradiation, the loading amount of Res depended on the irradiation temperature from 13.9% at 37 °C to 67.23% at 42 °C. The antioxidant activity of the nanocomposite reached 93.87% due to the synergetic effects of ZIF-8, PDA, and CeONP. Finally, the nanostructure was found to both inhibit Aβ monomer aggregation and disaggregate Aβ fibrils. These effects were magnified thanks to the anti-aggregation ability of Res and ZIF-8, the photothermal activity of PDA, and the chemotherapeutic effect of Res. In living cells, studies confirmed the ability of the composite to bring about Aβ aggregation inhibition, the reduction of ROS levels, and the inhibition of neuronal apoptosis under NIR irradiation.



Final considerations

Disease modifying target strategies for AD are still under extensive research. Recent results from clinical trials on anti-A β immunotherapy approaches have indeed suggested that merely reducing the amyloid burden in the brain of patients suffering from AD is insufficient to improve cognitive function. Currently, it appears as though that amyloid deposition may begin up to 20 years before a patient presents any clinical symptoms of AD. Thus, many researchers believe that treatment with anti-amyloid drugs in the mild to moderate stage of the disease is too late, and that for anti-amyloid approaches to be successful, treatment must begin early in the disease process.⁵⁵

Although great progress has been made in the last few years, a deeper understanding of the biological mechanisms of AD is necessary and will contribute to developing more effective therapeutic strategies, since just targeting protein deposits only ameliorates some of the neurological symptoms.¹⁹ Recent studies have integrated multiple new features such as novel biomarkers, new neuropsychological outcomes, treatment of earlier populations, and innovative trial designs. We believe that the future treatment could be determined by the use of different specific agents for every patient, accompanied by particular biomarkers, following a theragnostic approach. Lastly, therapeutic nanomedicine has started to build momentum also in the field, for example improving at the same time diagnosis, prevention, and treatment of various neurological disorders, including AD. Considering the versatility of coordination compounds, its smart functionalization offers the opportunity of obtaining a plethora of configurations, overcoming the main limitations associated with conventional drugs made up by organic molecules. In fact, here, many of the reviewed materials demonstrate that the derived coordination compounds are more active than metals and ligands separately.

Regarding the reported coordination compounds for AD treatment, most of them are discrete complexes, mainly showing two drawbacks: (i) any report evaluates how to eliminate the formed adduct of a complex with the A β peptide, and (ii) there is not a standard protocol in the activity of compounds against AD (*i.e.*, the use of different A β target peptides), making difficult the comparison between the proposed drugs, as is demonstrated by the reports of Collin and Ma *et al.*^{58,59}

Despite MOFs being considered promising candidates in AD theragnosis, they have been mainly described in the detection of the illness, rather than its treatment. The important parameters related to their safety use and efficiency, like the particle size and biocompatibility, should be improved. The particle size of MOFs is a limiting factor for some administration routes. Particularly in AD where the permeation of BBB is the target, MOFs should be synthesized at a size smaller than 100 nm, and stability studies under physiological conditions should be performed in order to obtain stable, homogeneous, and monodispersed MOFs. To date, only the MOF MIL-88B-(CH₃)₄ has been detected in the rats' brain after intravenous

administration of high doses (up to 220 mg Kg⁻¹), suggesting a bypass of the BBB with no evidence of severe cerebral toxicity.¹³³ In summary, the BBB is still a great obstacle for therapeutic agents against AD, and in spite of all reported compounds, in most cases, the permeability of these materials into the brain is not fully studied.

On the other hand, and prior to the potential use of MOFs in AD, the potential toxicity of these materials should be considered. Biocompatibility, and particularly, their interaction between MOFs and living structures, and its resulting effect in biological environments must be studied. Biocompatibility is mainly affected by the chemical composition, structure, morphology, size, and surface properties of MOFs. Therefore, and considering all the above, huge efforts should be made in the preparation of novel biocompatible and nanosized MOFs, or at least to modify the preparation methods to obtain nanosized and stable materials from already known biocompatible MOFs.

Some ambitious and promising examples of discrete metal complexes and MOFs in the treatment, diagnosis and theragnosis of AD have been recently reported. Considering the novelty of this research field and the versatility of coordination compounds, it is expected that they will play an important role in the future development of multifunctional therapies and/or diagnosis. Significant progress is now being achieved in the diversification of MOF bioapplications, which will aid in the continuous exploration of efficient AD treatments.

Abbreviations

| | |
|-------------|--|
| A β | β -Amyloid |
| A β O | A β oligomers |
| ABEI | <i>N</i> -(Aminobutyl)- <i>N</i> -(ethylsoluminol) |
| ACH | Acetylcholine |
| AChE | Acetylcholinesterase |
| aCSF | Artificial cerebrospinal fluid |
| AD | Alzheimer's disease |
| AICD | APP intracellular domain |
| APP | Amyloid precursor protein |
| Apt | Aptamer |
| BACE-1 | β -Secretase 1 |
| BBB | Blood-brain barrier |
| BHQ | Black hole quencher |
| BPS | 4,7-Diphenyl-[1,10]phenanthroline disulfonate |
| bpy | 2,2-Bipyridine |
| BTB | 1,3,5-Tris(4-carboxyphenyl)benzene |
| BuChE | Butyrylcholinesterase |
| bzq | 7,8-Benzoquinoline |
| CD | Circular dichroism |
| CeONP | Ceria nanoparticle |
| ChAT | Choline-acetyltransferase |
| CSF | Cerebrospinal fluid |
| CV | Cyclic voltammetry |
| cyclen | 1,4,7,10-Tetraazacyclododecane |
| DFT | Density functional theory |



| | | | |
|----------------------|--|--------|--|
| DLS | Dynamic light scattering | PMOF | Porphyrinic based MOF |
| DMF | <i>N,N</i> -Dimethylformamide | ppy | 2-Phenylpyridine |
| DMSO | Dimethyl sulfoxide | PPy | Polypyrrole |
| dpp | 2,3-Bis(2-pyridyl)pyrazine | RA | Retinoic acid |
| DPV | Differential pulse voltammetry | RET | Resonance energy transfer |
| ECL | Electrochemiluminescence | Res | Resveratrol |
| EPR | Electron paramagnetic resonance | ROS | Reactive oxygen species |
| ESI-MS | Electrospray ionization mass spectra | SWV | Square wave voltammetry |
| Fc | Ferrocene | TEM | Transmission electronic microscopy |
| FDA | US food and drug administration | ThT | Thioflavin T |
| FESEM | Field-emission scanning electron microscopy | thz | 1,3-Thiazole |
| Flav | Flavanones | vAChT | Vesicular ACh transporter |
| glz | Gliclazide | WHO | World health organization |
| GSH | Glutathione | XAS | X-ray absorption fine structure spectroscopy |
| GSK3 | Glycogen synthase kinase 3 | 2-AmTh | 2-aminothiazolium |
| gtsm | Glyoxalbis(<i>N</i> (4)-methylthiosemicarbazonato) | 8-BQ | 8-(1 <i>H</i> -Benzoimidazol-2-yl)quinoline |
| H ₂ bpydc | 2,2'-Bipyridine-5,5'-dicarboxylic acid | | |
| H ₂ cdda | 4,4'-(9 <i>H</i> -carbazole-2,7-diyl)dibenzoic acid | | |
| HPLC | High-performance liquid chromatography | | |
| H-UCNP | Highly doped upconversion nanoparticle | | |
| H-USM | Highly doped upconversion nanoparticles-SiO ₂ @MOF | | |
| IC ₅₀ | Half maximal inhibitory concentration | | |
| Im | Imidazole | | |
| Ind | Indazole | | |
| K _{sv} | Metal ion quenching coefficient | | |
| LDH | Lactic dehydrogenase | | |
| LTP | Long-term potentiation | | |
| MALDI-TOF MS | Matrix-assisted laser desorption ionization time-of-flight mass spectrometry | | |
| MAPT/ τ | Microtubule-associated protein τ | | |
| MB | Methylene blue | | |
| ϕ -Mepy | 2-Phenyl-5-methyl-pyridine | | |
| MOF | Metal-organic framework | | |
| MRI | Magnetic resonance imaging | | |
| MS | Mass spectrometry | | |
| MTT | 3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide | | |
| NIA-AA | National institute on ageing and Alzheimer's association | | |
| NIR | Near-infrared | | |
| NMOF | Nanoscale MOF | | |
| NMR | Nuclear magnetic resonance | | |
| NOTA | 1,4,7-Triazacyclononane-1,4,7-triacetic acid | | |
| np | Non-peripheral | | |
| NP | Nanoparticle | | |
| NSC | Neural stem cells | | |
| p | Peripheral | | |
| PBS | Phosphate buffer saline | | |
| Pc | Phthalocyanine | | |
| PCM | 1-Tetradecamol | | |
| PDA | Polydopamine | | |
| Phen | Phenanthroline | | |
| ϕ -Phen | 4,7-Diphenyl-1,10-phenanthroline | | |
| phq | 2-Phenylquinoline | | |
| PI3K | Phosphoinositol-3-kinase | | |

Author contributions

All authors contributed equally to this review.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

This work is supported by the MOFSEIDON project (PID2019-104228RB-I00, MCI/AEI/FEDER) and the AgroMOFs project (TED2021-132440B-I00) funded by MCIN/AEI/10.13039/501100011033 and by NextGenerationEU/PRTR and Margarita Salas contract 401 funded by Ministerio de Universidades and Next Generation. B-FQM-394, ProyExcel_00105 and ProyExcel_00386 were funded from la Consejería de Universidad, Investigación e Innovación de la Junta de Andalucía. S.R. is grateful for the grant (RYC2021-032522-I) funded by MCIN/AEI/10.13039/501100011033 and for El FSE invierte en tu futuro. In memory of our grandfathers and grandmothers, with love.

Notes and references

- 1 World Health Organization, Dementia, <https://www.who.int/news-room/fact-sheets/detail/dementia>, (accessed 18 January 2023).
- 2 GBD 2019 Dementia Forecasting Collaborators, *The Lancet Public Health*, 2022, 7, e105–e125.
- 3 Alzheimer's Association, Alzheimer's Disease Facts and Figures, <https://www.alz.org/alzheimers-dementia/facts-figures>, (accessed 1 January 2023).
- 4 P. Mirzayi, P. Shobeiri, A. Kalantari, G. Perry and N. Rezaei, *Mol. Brain*, 2022, 15, 1–14.
- 5 R. A. Stelzmann, H. N. Schnitzlein and F. R. Murtagh, *Clin. Anat.*, 1995, 8, 429–431.
- 6 A. Association, *Alzheimer's Dementia*, 2016, 12, 459–509.
- 7 J. L. Price and J. C. Morris, *Ann. Neurol.*, 1999, 45, 358–368.



- 8 R. J. Bateman, C. Xiong, T. L. S. Benzinger, A. M. Fagan, A. Goate, N. C. Fox, D. S. Marcus, N. J. Cairns, X. Xie, T. M. Blazey, D. M. Holtzman, A. Santacruz, V. Buckles, A. Oliver, K. Moulder, P. S. Aisen, B. Ghetti, W. E. Klunk, E. McDade, R. N. Martins, C. L. Masters, R. Mayeux, J. M. Ringman, M. N. Rossor, P. R. Schofield, R. A. Sperling, S. Salloway and J. C. Morris, *N. Engl. J. Med.*, 2012, **367**, 795–804.
- 9 C. Ferrari and S. Sorbi, *Physiol. Rev.*, 2021, **101**, 1047–1081.
- 10 M. Jucker and L. C. Walker, *Ann. Neurol.*, 2011, **70**, 532–540.
- 11 J. A. Hardy and G. A. Higgins, *Science*, 1992, **256**, 184–185.
- 12 J. Hardy and D. J. Selkoe, *Science*, 2002, **297**, 353–356.
- 13 J. M. Zozzoli, S. Bastias-Candia, M. J. Santos and N. C. Inestrosa, *Front. Aging Neurosci.*, 2014, **6**, 1–12.
- 14 H. W. Querfurth and F. M. LaFerla, *N. Engl. J. Med.*, 2010, **362**, 329–344.
- 15 M. M. Wen, N. S. El-Salamouni, W. M. El-Refaie, H. A. Hazzah, M. M. Ali, G. Tosi, R. M. Farid, M. J. Blanco-Prieto, N. Billa and A. S. Hanafy, *J. Controlled Release*, 2017, **245**, 95–107.
- 16 Y. Zhou, Y. Sun, Q.-H. Ma and Y. Liu, *Cell Stress*, 2018, **2**, 161.
- 17 R. Ricciarelli and E. Fedele, *Curr. Neuropharmacol.*, 2017, **15**, 935.
- 18 C. L. Masters, R. Cappai, K. J. Barnham and V. L. Villemagne, *J. Neurochem.*, 2006, **97**, 1700–1725.
- 19 H. Kozlowski, A. Janicka-Klos, J. Brasun, E. Gaggelli, D. Valensin and G. Valensin, *Coord. Chem. Rev.*, 2009, **253**, 2665–2685.
- 20 P. Faller and C. Hureau, *Dalton Trans.*, 2009, 1080–1094.
- 21 S. Rivera-Mancía, I. Pérez-Neri, C. Ríos, L. Tristán-López, L. Rivera-Espinosa and S. Montes, *Chem.-Biol. Interact.*, 2010, **186**, 184–199.
- 22 K. J. Barnham and A. I. Bush, *Curr. Opin. Chem. Biol.*, 2008, **12**, 222–228.
- 23 J. Grutzendler and J. C. Morris, *Drugs*, 2001, **61**, 41–52.
- 24 H. Soreq and S. Seidman, *Nat. Rev. Neurosci.*, 2001, **2**, 294–302.
- 25 J. Cummings, G. Lee, A. Ritter, M. Sabbagh and K. Zhong, *Alzheimer's Dementia*, 2020, **6**, 1–29.
- 26 X. Fang, J. Zhang, R. J. Roman and F. Fan, *GeroScience*, 2022, **44**, 1879–1883.
- 27 L. S. Schneider, *Dialogues Clin. Neurosci.*, 2000, **2**, 111–128.
- 28 FDA Grants Accelerated Approval for Alzheimer's Drug, <https://www.fda.gov/news-events/press-announcements/fda-grants-accelerated-approval-alzheimers-drug>, (accessed 1 January 2023).
- 29 FDA Grants Accelerated Approval for Alzheimer's Disease Treatment, <https://www.fda.gov/news-events/press-announcements/fda-grants-accelerated-approval-alzheimers-disease-treatment>, (accessed 15 May 2023).
- 30 J. Seigny, P. Chiao, T. Bussière, P. H. Weinreb, L. Williams, M. Maier, R. Dunstan, S. Salloway, T. Chen, Y. Ling, J. O'Gorman, F. Qian, M. Arastu, M. Li, S. Chollate, M. S. Brennan, O. Quintero-Monzon, R. H. Scannevin, H. M. Arnold, T. Engber, K. Rhodes, J. Ferrero, Y. Hang, A. Mikulskis, J. Grimm, C. Hock, R. M. Nitsch and A. Sandrock, *Nature*, 2016, **537**, 50–56.
- 31 C. H. van Dyck, C. J. Swanson, P. Aisen, R. J. Bateman, C. Chen, M. Gee, M. Kanekiyo, D. Li, L. Reyderman, S. Cohen, L. Froelich, S. Katayama, M. Sabbagh, B. Vellas, D. Watson, S. Dhadda, M. Irizarry, L. D. Kramer and T. Iwatsubo, *N. Engl. J. Med.*, 2023, **388**, 9–21.
- 32 J. Cummings, P. Aisen, L. G. Apostolova, A. Atri, S. Salloway and M. Weiner, *J. Prev. Alzheimer's Dis.*, 2021, **8**, 398–410.
- 33 D. A. Wolk, G. D. Rabinovici and B. C. Dickerson, *JAMA Neurology*, 2023, **80**, 429–430.
- 34 J. Cummings, P. S. Aisen, B. DuBois, L. Frölich, C. R. Jack, R. W. Jones, J. C. Morris, J. Raskin, S. A. Dowsett and P. Scheltens, *Alzheimer's Res. Ther.*, 2016, **8**, 1–12.
- 35 K. Blennow, M. J. de Leon and H. Zetterberg, *The Lancet*, 2006, **368**, 387–403.
- 36 M. P. Mattson, *Nature*, 2004, **430**, 631–639.
- 37 D.-L. Ma, H.-Z. He, K.-H. Leung, D. S.-H. Chan and C.-H. Leung, *Angew. Chem., Int. Ed.*, 2013, **52**, 7666–7682.
- 38 J. Karges, R. W. Stokes and S. M. Cohen, *Trends Chem.*, 2021, **3**, 534.
- 39 K. L. Haas and K. J. Franz, *Chem. Rev.*, 2009, **109**, 4921–4960.
- 40 S. Dasari and P. B. Tchounwou, *Eur. J. Pharmacol.*, 2014, **740**, 364–378.
- 41 F. Frézard, C. Demicheli and R. R. Ribeiro, *Molecules*, 2009, **14**, 2317–2336.
- 42 E. V. Capparelli, R. Bricker-Ford, M. J. Rogers, J. H. McKerrow and S. L. Reed, *Antimicrob. Agents Chemother.*, 2017, **61**, 1–8.
- 43 D. M. Teleanu, C. Chircov, A. M. Grumezescu, A. Volceanov and R. I. Teleanu, *Pharmaceutics*, 2018, **10**, 269.
- 44 W. M. Pardridge, *Neurotherapeutics*, 2005, **2**, 3–14.
- 45 M. B. Alahri, R. Arshadizadeh, M. Raeisi, M. Khatami, M. S. Sajadi, W. K. Abdelbasset, R. Akhmadeev and S. Irvani, *Inorg. Chem. Commun.*, 2021, **134**, 108997.
- 46 S. Rojas, A. Arenas-Vivo and P. Horcajada, *Coord. Chem. Rev.*, 2019, **388**, 22–226.
- 47 P. Horcajada, R. Gref, T. Baati, P. K. Allan, G. Maurin, P. Couvreur, G. Férey, R. E. Morris and C. Serre, *Chem. Rev.*, 2012, **112**, 1232–1268.
- 48 D. Valensin, C. Gabbiani and L. Messori, *Coord. Chem. Rev.*, 2012, **256**, 2357–2366.
- 49 D. J. Hayne, S. Lim and P. S. Donnelly, *Chem. Soc. Rev.*, 2014, **43**, 6701–6715.
- 50 J. L. Hickey and P. S. Donnelly, *Coord. Chem. Rev.*, 2012, **256**, 2367–2380.
- 51 Z. Liao, J. Zhang, E. Yu and Y. Cui, *Polyhedron*, 2018, **151**, 554–567.
- 52 F. Zeng, K. Peng, L. Han and J. Yang, *ACS Biomater. Sci. Eng.*, 2021, **7**, 3573–3585.
- 53 J. P. Leite, F. Figueira, R. F. Mendes, F. A. A. Paz and L. Gales, *ACS Sens.*, 2023, **8**, 1033–1053.
- 54 K. J. Barnham, V. B. Kenche, G. D. Ciccotosto, D. P. Smith, D. J. Tew, X. Liu, K. Perez, G. A. Cranston, T. J. Johansen,



- I. Volitakis, A. I. Bush, C. L. Masters, A. R. White, J. P. Smith, R. A. Cherny and R. Cappai, *Proc. Natl. Acad. Sci. U. S. A.*, 2008, **105**, 6813–6818.
- 55 V. B. Kenche, L. W. Hung, K. Perez, I. Volitakes, G. Ciccotosto, J. Kwok, N. Critch, N. Sherratt, M. Cortes, V. Lal, C. L. Masters, K. Murakami, R. Cappai, P. A. Adlard and K. J. Barnham, *Angew. Chem., Int. Ed.*, 2013, **52**, 3374–3378.
- 56 V. A. Streltsov, V. C. Epa, S. A. James, Q. I. Churches, J. M. Caine, V. B. Kenche and K. J. Barnham, *Chem. Commun.*, 2013, **49**, 11364–11366.
- 57 I. Sasaki, C. Bijani, S. Ladeira, V. Bourdon, P. Faller and C. Hureau, *Dalton Trans.*, 2012, **41**, 6404–6407.
- 58 F. Collin, I. Sasaki, H. Eury, P. Faller and C. Hureau, *Chem. Commun.*, 2013, **49**, 2130–2132.
- 59 G. Ma, E. Wang, H. Wei, K. Wei, P. Zhu and Y. Liu, *Metallomics*, 2013, **5**, 879–887.
- 60 D. Valensin, P. Anzini, E. Gaggelli, N. Gaggelli, G. Tamasi, R. Cini, C. Gabbiani, E. Michelucci, L. Messori, H. Kozlowski and G. Valensin, *Inorg. Chem.*, 2010, **49**, 4720–4722.
- 61 L. Messori, M. Camarri, T. Ferraro, C. Gabbiani and D. Franceschini, *ACS Med. Chem. Lett.*, 2013, **4**, 329–332.
- 62 A. Kumar, L. Moody, J. F. Olaivar, N. A. Lewis, R. L. Khade, A. A. Holder, Y. Zhang and V. Rangachari, *ACS Chem. Neurosci.*, 2010, **1**, 691–701.
- 63 A. L. F. Sarria, A. F. L. Vilela, B. M. Frugeri, J. B. Fernandes, R. M. Carlos, M. F. das, G. F. da Silva, Q. B. Cass and C. L. Cardoso, *J. Inorg. Biochem.*, 2016, **164**, 141–149.
- 64 A. Günsel, P. Taslimi, G. Y. Atmaca, A. T. Bilgiçli, H. Pişkin, Y. Ceylan, A. Erdoğan, M. N. Yarasir and İ. Gülçin, *J. Mol. Struct.*, 2021, **1237**, 1–13.
- 65 H. Yalazan, B. Tüzün, D. Akkaya, B. Barut, H. Kantekin and S. Yıldırım, *Appl. Organomet. Chem.*, 2022, **36**, 1–19.
- 66 A. Şenocak, N. A. Taş, P. Taslimi, B. Tüzün, A. Aydin and A. Karadağ, *J. Biochem. Mol. Toxicol.*, 2022, **36**, 1–9.
- 67 A. García-García, S. Rojas, L. Rivas-García, M. D. Navarro-Hortal, J. M. Romero-Márquez, J. G. Fernández-Bolaños, D. Choquesillo-Lazarte, A. Salinas-Castillo, Ó. López, J. L. Quiles and A. Rodríguez-Diéguez, *Chem. Commun.*, 2022, **58**, 1514–1517.
- 68 P. J. Crouch, W. H. Lin, P. A. Adlard, M. Cortes, V. Lal, G. Filiz, K. A. Perez, M. Nurjono, A. Caragounis, T. Du, K. Loughton, I. Volitakis, A. I. Bush, Q.-X. Li, C. L. Masters, R. Cappai, R. A. Cherny, P. S. Donnelly, A. R. White and K. J. Barnham, *Proc. Natl. Acad. Sci. U. S. A.*, 2009, **106**, 381–386.
- 69 X. Wang, X. Wang, C. Zhang, Y. Jiao and Z. Guo, *Chem. Sci.*, 2012, **3**, 1304–1312.
- 70 D. P. Smith, D. G. Smith, C. C. Curtain, J. F. Boas, J. R. Pilbrow, G. D. Ciccotosto, T. L. Lau, D. J. Tew, K. Perez, J. D. Wade, A. I. Bush, S. C. Drew, F. Separovic, C. L. Masters, R. Cappai and K. J. Barnham, *J. Biol. Chem.*, 2006, **281**, 15145–15154.
- 71 A. I. Bush, *Trends Neurosci.*, 2003, **26**, 207–214.
- 72 M. Drent, N. A. M. Cobben, R. F. Henderson, E. F. M. Wouters and M. Van Diejen-Visser, *Eur. Respir. J.*, 1996, **9**, 1736–1742.
- 73 J. B. Pereira, E. F. A. Carvalho, M. A. F. Faustino, R. Fernandes, M. G. P. M. S. Neves, J. A. S. Cavaleiro, N. C. M. Gomes, Â. Cunha, A. Almeida and J. P. C. Tomé, *Photochem. Photobiol.*, 2012, **88**, 537–547.
- 74 Z. Jiang, J. Shao, T. Yang, J. Wang and L. Jia, *J. Pharm. Biomed. Anal.*, 2014, **87**, 98–104.
- 75 A. Günsel, A. Kobyaoglu, A. T. Bilgiçli, B. Tüzün, B. Tosun, G. Arabaci and M. N. Yarasir, *J. Mol. Struct.*, 2020, **1200**, 1–10.
- 76 A. Günsel, G. Y. Atmaca, P. Taslimi, A. T. Bilgiçli, İ. Gülçin, A. Erdoğan and M. N. Yarasir, *J. Photochem. Photobiol., A*, 2020, **396**, 1–9.
- 77 S. Okabayashi, N. Shimosawa, Y. Yasutomi, K. Yanagisawa and N. Kimura, *PLoS ONE*, 2015, **10**, 1–19.
- 78 K. Mittal, R. J. Mani and D. P. Katare, *Sci. Rep.*, 2016, **6**, 1–8.
- 79 B. M. Paterson and P. S. Donnelly, *Chem. Soc. Rev.*, 2011, **40**, 3005–3018.
- 80 M. A. Green, D. L. Klippenstein and J. R. Tennison, *J. Nucl. Med.*, 1988, **29**, 1549–1557.
- 81 Y. Fujibayashi, H. Taniuchi, Y. Yonekura, H. Ohtani, J. Konishi and A. Yokoyama, *J. Nucl. Med.*, 1997, **38**, 1155–1160.
- 82 K. A. Price, A. Caragounis, B. M. Paterson, G. Filiz, I. Volitakis, C. L. Masters, K. J. Barnham, P. S. Donnelly, P. J. Crouch and A. R. White, *J. Med. Chem.*, 2009, **52**, 6606–6620.
- 83 P. S. Donnelly, A. Caragounis, T. Du, K. M. Loughton, I. Volitakis, R. A. Cherny, R. A. Sharples, A. F. Hill, Q.-X. Li, C. L. Masters, K. J. Barnham and A. R. White, *J. Biol. Chem.*, 2008, **283**, 4568–4577.
- 84 X. Wang, X. Wang and Z. Guo, *Acc. Chem. Res.*, 2015, **48**, 2622–2631.
- 85 S. Samanta, K. Rajasekhar, V. Babagond and T. Govindaraju, *ACS Chem. Neurosci.*, 2019, **10**, 3611–3621.
- 86 K. Rajasekhar, K. Mehta and T. Govindaraju, *ACS Chem. Neurosci.*, 2018, **9**, 1432–1440.
- 87 K. Rajasekhar, S. Samanta, V. Bagoband, N. A. Murugan and T. Govindaraju, *iScience*, 2020, **23**, 101005.
- 88 M. Okafor, P. Gonzalez, P. Ronot, I. El Masoudi, A. Boos, S. Ory, S. Chasserot-Golaz, S. Gasman, L. Raibaut, C. Hureau, N. Vitale and P. Faller, *Chem. Sci.*, 2022, **13**, 11829–11840.
- 89 K. Rajasekhar, C. Madhu and T. Govindaraju, *ACS Chem. Neurosci.*, 2016, **7**, 1300–1310.
- 90 M. Di Vaira, C. Bazzicalupi, P. Orioli, L. Messori, B. Bruni and P. Zatta, *Inorg. Chem.*, 2004, **43**, 3795–3797.
- 91 H. Chen, J. Mi, S. Li, Z. Liu, J. Yang, R. Chen, Y. Wang, Y. Ban, Y. Zhou, W. Dong and Z. Sang, *J. Enzyme Inhib. Med. Chem.*, 2023, **38**, 2169682.
- 92 S. Rojas, T. Baati, L. Njim, L. Manchego, F. Neffati, N. Abdejelil, S. Saguem, C. Serre, M. F. M. F. Najjar, A. Zakhama, P. Horcajada, N. Abdeljelil, S. Saguem, C. Serre, M. F. M. F. Najjar, A. Zakhama and P. Horcajada, *J. Am. Chem. Soc.*, 2018, **140**, 9581–9586.
- 93 S. Rojas, N. Guillou and P. Horcajada, *ACS Appl. Mater. Interfaces*, 2019, **11**, 22188–22193.



- 94 Z. L. Mensinger, B. L. Cook and E. L. Wilson, *ACS Omega*, 2020, **5**, 32969–32974.
- 95 J. Wang, Y. Fan, Y. Tan, X. Zhao, Y. Zhang, C. Cheng and M. Yang, *ACS Appl. Mater. Interfaces*, 2018, **10**, 36615–36621.
- 96 D. Yu, Y. Guan, F. Bai, Z. Du, N. Gao, J. Ren and X. Qu, *Chem. – Eur. J.*, 2019, **25**, 3489–3495.
- 97 D. Yu, M. Ma, Z. Liu, Z. Pi, X. Du, J. Ren and X. Qu, *Biomaterials*, 2020, **255**, 1–10.
- 98 L. Vermunt, S. A. M. Sikkens, A. van den Hout, R. Handels, I. Bos, W. M. van der Flier, S. Kern, P. J. Ousset, P. Maruff, I. Skoog, F. R. J. Verhey, Y. Freund-Levi, M. Tsolaki, Å. K. Wallin, M. Olde Rikkert, H. Soininen, L. Spuru, H. Zetterberg, K. Blennow, P. Scheltens, G. Muniz-Terrera and P. J. Visser, *Alzheimer's Dementia*, 2019, **15**, 888–898.
- 99 J. Luo, F. Agboola, E. Grant, C. L. Masters, M. S. Albert, S. C. Johnson, E. M. McDade, J. Vöglein, A. M. Fagan, T. Benzinger, P. Massoumzadeh, J. Hassenstab, R. J. Bateman, J. C. Morris, R. J. Perrin, J. Chhatwal, M. Jucker, B. Ghetti, C. Cruchaga, N. R. Graff-Radford, P. R. Schofield, H. Mori and C. Xiong, *Neurology*, 2020, **95**, e3104–e3116.
- 100 G. McKhann, D. Drachman, M. Folstein, R. Katzman, D. Price and E. M. Stadlan, *Neurology*, 1984, **34**, 939–944.
- 101 C. Humpel, *Trends Biotechnol.*, 2011, **29**, 26–32.
- 102 J. Zhu, T. Xia, Y. Cui, Y. Yang and G. Qian, *J. Solid State Chem.*, 2019, **270**, 317–323.
- 103 J. Bai, L. Liu, C. Jia, Z. Liu, S. Gao, Y. Han and H. Yan, *ACS Appl. Bio Mater.*, 2019, **2**, 6021–6028.
- 104 L. C. Walker, D. G. Lynn and Y. O. Chernoff, *Prion*, 2018, **12**, 261–265.
- 105 J. Han, M. Zhang, G. Chen, Y. Zhang, Q. Wei, Y. Zhuo, G. Xie, R. Yuan and S. Chen, *J. Mater. Chem. B*, 2017, **5**, 8330–8336.
- 106 C. Wang, N. Zhang, Y. Li, L. Yang, D. Wei, T. Yan, H. Ju, B. Du and Q. Wei, *Sens. Actuators, B*, 2019, **291**, 319–328.
- 107 A. H. Valekar, B. S. Batule, M. Il Kim, K.-H. Cho, D.-Y. Hong, U.-H. Lee, J.-S. Chang, H. G. Park and Y. K. Hwang, *Biosens. Bioelectron.*, 2018, **100**, 161–168.
- 108 Y. Zhou, C. Li, X. Li, X. Zhu, B. Ye and M. Xu, *Anal. Methods*, 2018, **10**, 4430–4437.
- 109 B. Liu, H. Shen, Y. Hao, X. Zhu, S. Li, Y. Huang, P. Qu and M. Xu, *Anal. Chem.*, 2018, **90**, 12449–12455.
- 110 J. Qin, M. Cho and Y. Lee, *ACS Appl. Mater. Interfaces*, 2019, **11**, 11743–11748.
- 111 Y. Wang, Y. Zhang, H. Sha, X. Xiong and N. Jia, *ACS Appl. Mater. Interfaces*, 2019, **11**, 36299–36306.
- 112 J. Miao, X. Li, Y. Li, X. Dong, G. Zhao, J. Fang, Q. Wei and W. Cao, *Anal. Chim. Acta*, 2019, **1089**, 48–55.
- 113 G. Zhao, Y. Wang, X. Li, Q. Yue, X. Dong, B. Du, W. Cao and Q. Wei, *Anal. Chem.*, 2019, **91**, 1989–1996.
- 114 J. Fang, G. Zhao, X. Dong, X. Li, J. Miao, Q. Wei and W. Cao, *Biosens. Bioelectron.*, 2019, **142**, 1–6.
- 115 H.-X. Ren, Y.-B. Miao and Y. Zhang, *Microchim. Acta*, 2020, **187**, 1–8.
- 116 X. Dong, G. Zhao, X. Li, J. Fang, J. Miao, Q. Wei and W. Cao, *Talanta*, 2020, **208**, 1–7.
- 117 H.-X. Ren, Q. Zhong, Y.-B. Miao, X.-W. Wen, G.-Y. Wu, H.-L. Wang and Y. Zhang, *Microchim. Acta*, 2020, **187**, 1–8.
- 118 X. Z. Wang, J. Du, N. N. Xiao, Y. Zhang, L. Fei, J. D. LaCoste, Z. Huang, Q. Wang, X. R. Wang and B. Ding, *Analyst*, 2020, **145**, 4646–4663.
- 119 X. Wang, L. Li, X. Gu, B. Yu and M. Jiang, *Microchim. Acta*, 2021, **188**, 1–11.
- 120 W.-K. Fang, L. Liu, L. Zhang, D. Liu, Y. Liu and H.-W. Tang, *Anal. Chem.*, 2021, **93**, 12447–12455.
- 121 L. Yin, Y. Wang, R. Tan, H. Li and Y. Tu, *Microchim. Acta*, 2021, **188**, 1–8.
- 122 A. S. Spitsyna, A. S. Poryvaev, N. E. Sannikova, A. A. Yazikova, I. A. Kirilyuk, S. A. Dobrynin, O. A. Chinak, M. V. Fedin and O. A. Krumkacheva, *Molecules*, 2022, **27**, 1–13.
- 123 M. de, J. Velásquez-Hernández, R. Ricco, F. Carraro, F. T. Limpoco, M. Linares-Moreau, E. Leitner, H. Wiltsche, J. Rattenberger, H. Schröttner, P. Frühwirt, E. M. Stadler, G. Gescheidt, H. Amenitsch, C. J. Doonan and P. Falcaro, *CrystEngComm*, 2019, **21**, 4538–4544.
- 124 A. S. Poryvaev, A. A. Yazikova, D. M. Polyukhov, O. A. Chinak, V. A. Richter, O. A. Krumkacheva and M. V. Fedin, *J. Phys. Chem. C*, 2021, **125**, 15606–15613.
- 125 J. H. Ryu, H. Koo, I. C. Sun, S. H. Yuk, K. Choi, K. Kim and I. C. Kwon, *Adv. Drug Delivery Rev.*, 2012, **64**, 1447–1458.
- 126 K. Kim, J. H. Kim, H. Park, Y. S. Kim, K. Park, H. Nam, S. Lee, J. H. Park, R. W. Park, I. S. Kim, K. Choi, S. Y. Kim, K. Park and I. C. Kwon, *J. Controlled Release*, 2010, **146**, 219–227.
- 127 D. E. Lee, H. Koo, I. C. Sun, J. H. Ryu, K. Kim and I. C. Kwon, *Chem. Soc. Rev.*, 2012, **41**, 2656–2672.
- 128 B. Y.-W. Man, H.-M. Chan, C.-H. Leung, D. S.-H. Chan, L.-P. Bai, Z.-H. Jiang, H.-W. Li and D.-L. Ma, *Chem. Sci.*, 2011, **2**, 917–921.
- 129 J. Zhao, F. Yin, L. Ji, C. Wang, C. Shi, X. Liu, H. Yang, X. Wang and L. Kong, *ACS Appl. Mater. Interfaces*, 2020, **12**, 44447–44458.
- 130 R.-Q. Song, X. Ma and G.-Y. Song, *J. Chin. Chem. Soc.*, 2020, **67**, 1462–1469.
- 131 X. Yan, Y. Pan, L. Ji, J. Gu, Y. Hu, Y. Xia, C. Li, X. Zhou, D. Yang and Y. Yu, *Anal. Chem.*, 2021, **93**, 13823–13834.
- 132 X. Yan, Y. Pan, L. Ji, J. Gu, Y. Hu, Y. Xia, C. Li, X. Zhou, D. Yang and Y. Yu, *Anal. Chem.*, 2021, **93**, 13823–13834.
- 133 T. Baati, L. Njim, F. Neffati, A. Kerkeni, M. Bouttemi, R. Gref, M. F. Najjar, A. Zakhama, P. Couvreur, C. Serre and P. Horcajada, *Chem. Sci.*, 2013, **4**, 1597–1607.

