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Enhanced annihilation electrochemiluminescence by nanofluidic confinement†

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Microfabricated nanofluidic electrochemical devices offer a highly controlled nanochannel geometry; they confine the volume of chemical reactions to the nanoscale and enable greatly amplified electrochemical detection. Here, the generation of stable light emission by electrochemiluminescence (ECL) in transparent nanofluidic devices is demonstrated for the first time by exploiting nanogap amplification. Through continuous oxidation and reduction of $[\text{Ru}(\text{bpy})_3]^{2+}$ luminophores at electrodes positioned at opposite walls of a 100 nm nanochannel, we compare classic redox cycling and ECL annihilation. Enhanced ECL light emission of attomole luminophore quantities is evidenced under ambient conditions due to the spatial confinement in a 10 femtoliter volume, resulting in a short diffusion timescale and highly efficient ECL reaction pathways at the nanoscale.

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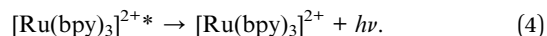
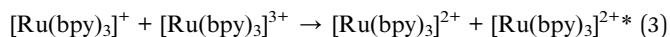
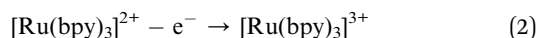
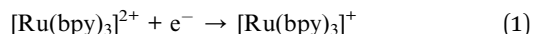
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Introduction

Electrochemiluminescence (ECL) is the emission of light by electrochemical means: reactive species are generated at the electrode surface and undergo a highly exergonic electron-transfer reaction producing the excited state of the luminophore.¹ It relaxes to the ground state, emitting a photon. The possibility to regenerate the reactive species *in situ* as well as not requiring an external light source for excitation, gives this method many advantages for (bio)sensing applications, single object imaging,^{2–4} and light-emitting devices.⁵ These advantages include a straightforward experimental setup, a very high sensitivity and a wide detection range. Thus, applications and fundamental aspects of ECL have been researched extensively.⁵

Whereas the precise mechanism of light generation depends on the species involved, there are two dominant pathways: the annihilation and the co-reactant pathways.¹ Here, we focus on the annihilation pathway in a nanochannel, in which light is generated through a reaction between the oxidized and reduced forms of the original compound. We selected tris(bipyridine)

ruthenium(II), $[\text{Ru}(\text{bpy})_3]^{2+}$, one of the most well-studied ECL compounds^{6,7} due to its relatively high quantum yield, water solubility and long-lived triplet excited state. Its annihilation pathway is:⁸



These reactions were initially investigated by rapidly pulsing a single electrode between the oxidizing and reducing potential.^{9,10} However, this technique suffers from drawbacks¹¹ such as large charging currents. Using two electrodes, both $[\text{Ru}(\text{bpy})_3]^+$ and $[\text{Ru}(\text{bpy})_3]^{3+}$ can be generated in close vicinity under steady-state conditions. Various two-electrode setups have been employed, including ring and disk electrodes,¹² thin layer cells,¹³ band electrodes,^{14–16} and scanning electrochemical microscopy.^{17,18} These studies have shown that annihilation occurs more efficiently at smaller inter-electrode distances^{16,18} as the diffusional distance between electrogenerated $[\text{Ru}(\text{bpy})_3]^+$ and $[\text{Ru}(\text{bpy})_3]^{3+}$ is decreased. Indeed, the stability of the reduced form, $[\text{Ru}(\text{bpy})_3]^+$, is a key issue limiting the efficiency of the annihilation ECL. Yet, observation of light emission so far has been limited to a minimum inter-electrode distance of 2 μm .^{18,19} In addition, due to the sensitivity of annihilation ECL towards impurities such as dioxygen and water, experiments are performed in drastic conditions: dried and distilled solvents just before use, recrystallized and dried luminescent compounds and supporting electrolytes, under an inert

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atmosphere or in a dry box.^{15,18,20} This is especially true for platinum electrodes as the reduction potential for water/dioxygen competes with that of $[\text{Ru}(\text{bpy})_3]^{2+}$.

Here, ECL emission is observed under ambient conditions using platinum electrodes in electrochemical nanogap devices.²¹ We exploit the spatial confinement of such devices to enhance the annihilation ECL-reaction of $[\text{Ru}(\text{bpy})_3]^{2+}$ and to reduce the effects of impurities. Microfabrication allows precise control of a nanofluidic channel geometry; two individually addressable electrodes positioned at the top and bottom of a nanochannel are separated by a distance of 100 nm or less (see Fig. 1).

The nanoscale distance between electrodes results in short diffusion times between them and gives rise to highly amplified currents through electrochemical redox cycling. Reversibly electrochemically active analytes are repeatedly oxidized and reduced as they diffuse between both electrodes, shuttling electrons across the nanochannel. At diffusion times of $\sim 10 \mu\text{s}$, each analyte molecule contributes thousands of electrons per seconds to the detected limiting current, which, thus, is highly amplified. Due to efficient redox cycling and a confined geometry, nanogap transducers and similar geometries have been employed for, *e.g.*, studying adsorption,²² migration,^{23,24} biosensing,²⁵ single-molecule electrochemistry,²⁶ and spectroelectrochemistry.²⁷

Herein, for the first time, annihilation ECL is examined in transparent nanofluidic electrochemical devices. We report enhanced light emission due to highly efficient reaction pathways at the nanoscale. Light emission is stable at ambient conditions due to reduced degradation by contaminants.

Results and discussion

Nanogap devices were fabricated as described previously,^{28,29} except that a transparent glass substrate was used. Due to the transparency and lower heat conductivity of glass, most fabrication steps had to be altered completely. In brief, structures comprising a thin 20 nm Pt bottom electrode, 100 nm Cr sacrificial layer (forming the nanochannel volume) and 120 nm

Pt top electrode were patterned on a 10 cm borosilicate glass wafer using UV lithography followed by evaporation deposition and lift-off. A 500 nm SiO_2/SiN passivation layer, deposited using chemical vapor deposition, encased the devices. Access holes were etched through the passivation layer by reactive ion etching. Finally, the wafer was diced into individual chips. Before experimentation, a polydimethylsiloxane fluid reservoir was placed on top of the device, and the sacrificial Cr layer was selectively wet-etched to form the channel. In Fig. 1a, a device is shown with a $5 \mu\text{m} \times 32 \mu\text{m} \times 100 \text{ nm}$ channel volume, and an effective area of overlap between the electrodes of $3 \mu\text{m} \times 20 \mu\text{m}$.

Electrochemical measurements were performed with a bipotentiostat (Autolab – PGSTAT30). Functioning simultaneously as both reference electrode and counter electrode, we used an $\text{Ag}/\text{AgCl}/\text{KCl}$ (3 M) electrode in water, or an Ag wire in acetonitrile, respectively. For all experiments, freshly etched devices were cleaned by cycling the potential of both electrodes between -0.15 V and 1.2 V vs. Ag/AgCl in 50 mM H_2SO_4 . Channels were thoroughly flushed with Milli-Q water, and subsequently with acetonitrile, which was then replaced by 10 mM $[\text{Ru}(\text{bpy})_3](\text{PF}_6)_2$ and 0.1 M TBAPF₆ in acetonitrile. This solution was used for all experiments.

We investigated two electrochemical ‘modes’ in the nanofluidic channel (see Fig. 2). In redox cycling, the top electrode is biased at an oxidizing potential generating $[\text{Ru}(\text{bpy})_3]^{3+}$. Biasing the bottom electrode at 0 V leads to the one-electron reduction of $[\text{Ru}(\text{bpy})_3]^{3+}$ and, thus, regeneration of $[\text{Ru}(\text{bpy})_3]^{2+}$. In annihilation ECL, the same potential is imposed at the top electrode whereas the bottom electrode is placed at a sufficiently cathodic potential to reduce $[\text{Ru}(\text{bpy})_3]^{2+}$ to $[\text{Ru}(\text{bpy})_3]^+$. The annihilation reaction (3) takes place, and ECL light is emitted (4). Conceptually, this process is similar to redox cycling as it involves the constant consumption and regeneration of $[\text{Ru}(\text{bpy})_3]^{2+}$. However, instead of taking place between the two electrode surfaces, regeneration (annihilation) occurs homogeneously in the channel bulk, where $[\text{Ru}(\text{bpy})_3]^+$ and $[\text{Ru}(\text{bpy})_3]^{3+}$ meet. In the top region, redox cycling between $[\text{Ru}(\text{bpy})_3]^{2+}$ and its oxidized form occurs, whereas the bottom region shows cycling of $[\text{Ru}(\text{bpy})_3]^{2+}$ with the reduced form. The channel is therefore effectively split, and anodic and cathodic redox cycling occur simultaneously (Fig. 2b). Therefore, as diffusion times scale inversely with the length of the diffusive path, switching from redox cycling to annihilation ECL mode should result in a higher faradaic current.



Fig. 1 (a) Top view optical micrograph of an electrochemical nanogap device. (b) Schematic cross section of a device with access holes to reservoir and two individually addressable Pt electrodes. The sacrificial Cr layer is etched before the experiments to form the nanochannel.

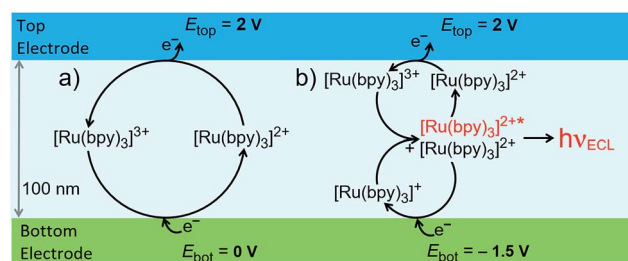


Fig. 2 Schematic of two reaction pathways in the nanochannel: (a) redox cycling, and (b) annihilation leading to ECL emission.



ECL-emission is produced in annihilation mode inside a nanochannel and was imaged and spatially resolved as shown in Fig. 3 using an epifluorescence microscope (DMI6000, Leica). Photon emission was collected by an inverted 40 \times microscope objective and detected by an Electron Multiplying Charge Coupled Device (EM-CCD) Camera (Hamamatsu, 9100-13). Light emission from a luminophore quantity of 60 attomoles present in the active area of the device was intense enough to be recorded through the 20 nm-thick semi-transparent Pt bottom electrode, due to nanogap amplification. The overlay image (Fig. 3a, right) shows that ECL intensity is uniform along the channel and decreases rapidly at the edges. No emission is seen from the bottom electrode area under the access holes. Furthermore, ECL emission is limited to a region narrower than the 5 μm wide channel. This indicates that emission is caused exclusively by reaction in the 3 $\mu\text{m} \times 20 \mu\text{m}$ effective area between the electrodes (as observed for purely electrochemical cycling³⁰). Full-width-at-half-maxima of longitudinal and lateral intensity profiles amount to 20 μm and 4.3 μm , respectively. Presumably, the optical resolution of the 40 \times objective as well as inhomogeneous transparency of Pt and glass lead to a slightly wider profile than expected. Clearly, ECL emission is confined to the active area of the nanochannel, where reactions (1–4) (Fig. 2b) take place.

The constant ECL intensity along the nanochannel (Fig. 3b, blue curve) indicates that only negligible degradation occurs. If $[\text{Ru}(\text{bpy})_3]^+$ or $[\text{Ru}(\text{bpy})_3]^{3+}$ would degrade, the intensity would be strongly attenuated in the center of the channel because the chance of encountering contaminants increases quadratically as luminophores diffuse towards the center.

Fig. 4a displays the chronoamperometric currents at both electrodes when stepping between redox cycling and

annihilation ECL modes. In redox cycling mode, the oxidation and reduction currents are symmetric and constant. The observed currents of $\sim 0.65 \mu\text{A}$ are in agreement with an expected analytical estimate³¹ of $I_{\text{lim}} = \frac{FADc}{h} = 0.58 \mu\text{A}$ (F : faraday constant; A : active area of 3 μm by 20 μm ; $D = 1 \times 10^{-9} \text{ m}^2 \text{ s}^{-1}$ diffusion coefficient, c : 10 mM $[\text{Ru}(\text{bpy})_3]^{2+}$ concentration, h : 100 nm channel height).

ECL intensity was measured simultaneously by using a photomultiplier tube (Hamamatsu R4632). The signal was amplified by a Keithley Picoammeter before acquisition with the second input signal of a $\mu\text{Autolab}$ type II potentiostat. Emission was not observed in redox cycling mode (Fig. 4b). Upon switching to annihilation mode, an initial transient current increase can be ascribed to adsorption of analytes onto the electrode surfaces due to the high surface-to-volume ratio of the nanogap devices.^{22,29,32} Oxidation and reduction currents are symmetric as expected for reactions (1–4). In annihilation mode, the light intensity (Fig. 4b) exhibits a strong transient behavior before approaching a steady state. Understanding this transient behavior will require further studies, we partially attribute the complex transient to desorption when stepping the potential as well as accumulating $[\text{Ru}(\text{bpy})_3]^{3+}$ during redox cycling.

Upon switching from redox cycling to annihilation mode, an expected increase in the current is observed as the diffusive path for cycling molecules halves to 50 nm. Concentration profiles of all $[\text{Ru}(\text{bpy})_3]$ species are illustrated in Fig. 5 (COM-SOL Multiphysics, see ESI[†]). Considering the symmetry of the nanofluidic channel and the annihilation reaction (*i.e.*, double cycling redox process), ECL is emitted in the middle of the



Fig. 3 (a) From left to right: bright-field micrograph of the nanochannel, ECL emission recorded in the dark, and overlay of both images (dashed lines: direction of profiles in (b)). (b) ECL intensity profiles along (blue) and across (orange) the center of the channel, using a solution of 10 mM $\text{Ru}(\text{bpy})_3(\text{PF}_6)_2$ and 0.1 M TBAPF_6 in acetonitrile. The top electrode was biased at 2 V, the bottom electrode at -1.5 V vs. Ag.



Fig. 4 (a) Measured chronoamperometric currents at the top (blue) and bottom (green) electrodes, and (b) corresponding ECL intensity. The top electrode was maintained at 2.3 V while the bottom electrode was pulsed between 0 V and -1.5 V vs. Ag, highlighted in white (redox cycling mode) and grey (annihilation ECL mode), respectively. Solution consisted of 10 mM $\text{Ru}(\text{bpy})_3(\text{PF}_6)_2$ and 0.1 M TBAPF_6 in acetonitrile.



nanogap where both $[\text{Ru}(\text{bpy})_3]^{3+}$ and $[\text{Ru}(\text{bpy})_3]^+$ react to form $[\text{Ru}(\text{bpy})_3]^{2+*}$, assuming similar diffusion coefficients for $[\text{Ru}(\text{bpy})_3]^{3+}$ and $[\text{Ru}(\text{bpy})_3]^+$. The ECL emission should occur in the vertical symmetry plane of the device and be confined to the electroactive area of the nanochannel (see Fig. 5d). However, it is not possible to resolve this with optical microscopy due to the nanometric dimension of the inter-electrode gap.

The simulated diffusive fluxes (*i.e.*, slopes of concentration profiles) at electrode surfaces double when switching from redox cycling to annihilation. However, a ratio of only 1.2 for the steady-state redox cycling and annihilation currents is observed (Fig. 4a). Understanding this low ratio requires further studies. As origin we rule out degradation (see above) and a limited annihilation rate (as a two-electron electrode reaction of $[\text{Ru}(\text{bpy})_3]^+$ and $[\text{Ru}(\text{bpy})_3]^{3+}$ would exactly compensate for every annihilation not taking place^{18,33,34}). Furthermore, any influences on the currents by unequal diffusivities as well as by adsorption are balanced by coupling to the bulk reservoir.^{29,31} We speculate that more complex concentration profiles are caused by contribution of electrical migration (at an overall potential drop of 3.5 V over 100 nm).

Classically, annihilation ECL experiments are performed using highly inert conditions (*vide supra*).^{15,18,20} In the present work, we imaged an intense ECL signal under ambient conditions using platinum electrodes. In the nanochannel, neither strong degradation of $[\text{Ru}(\text{bpy})_3]^+$ nor greatly reduced annihilation occurs (previously reduction of the annihilation rate from $>10^7 \text{ M}^{-1} \text{ s}^{-1}$ to $\sim 10^3 \text{ M}^{-1} \text{ s}^{-1}$ was observed under ambient conditions^{18,35}). Negligible degradation is caused by the short diffusive path, which greatly reduces the chance to encounter contaminants such as dioxygen, and/or by depletion of contaminants in the nanochannel by electrochemical deactivation at the electrodes (in the ESI† we further explore the effect of channel height, lifetime of the excited state and possible

degradation on concentration profile by finite element calculations).

Conclusions

ECL-emission was investigated in transparent nanofluidic devices for the first time. Light emission was shown to occur through an annihilation reaction pathway when both electrodes were biased at appropriate potentials. Light emission is enhanced in two ways: first, the close vicinity of the two electrodes leads to efficient annihilation reactions due to very fast diffusion. Second, stable light emission is achieved as the very short diffusive distances protect from degradation by contaminants, and our results indicate that measurements in nano-devices are possible under ambient conditions, which cannot be achieved using classical setups. Microfabricated nanodevices combine the properties of exact control of an inter-electrode distance, a highly reproducible device geometry and strong confinement of analytes in the channel volume. Therefore, we believe that nanogap transducers are powerful tools for investigating pathways in ECL and also to exploit their unique nanometric properties to develop multicolor ECL emission systems.

Conflicts of interest

There are no conflicts to declare.

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References

- 1 A. Bard, *Electrogenerated chemiluminescence*, Marcel Dekker, New York, 2004.
- 2 A. J. Wilson, K. Marchuk and K. A. Willets, *Nano Lett.*, 2015, **15**, 6110–6115.
- 3 J. E. Dick, C. Renault, B. K. Kim and A. J. Bard, *Angew. Chem., Int. Ed.*, 2014, **53**, 11859–11862.
- 4 G. Valenti, S. Scarabino, B. Goudeau, A. Lesch, M. Jović, E. Villani, M. Sentic, S. Rapino, S. Arbault, F. Paolucci and N. Sojic, *J. Am. Chem. Soc.*, 2017, **139**, 16830–16837.
- 5 L. Hu and G. Xu, *Chem. Soc. Rev.*, 2010, **39**, 3275–3304.
- 6 B. A. Gorman, P. S. Francis and N. W. Barnett, *Analyst*, 2006, **131**, 616–639.
- 7 X. B. Yin, S. Dong and E. Wang, *TrAC, Trends Anal. Chem.*, 2004, **23**, 432–441.
- 8 R. J. Forster, P. Bertoncello and T. E. Keyes, *Annu. Rev. Anal. Chem.*, 2009, **2**, 359–385.
- 9 R. Breslow, R. Hill and E. Wasserman, *J. Am. Chem. Soc.*, 1964, **83**, 5350–5351.
- 10 K. S. V. Santhanam and A. J. Bard, *J. Am. Chem. Soc.*, 1965, **87**, 139–140.

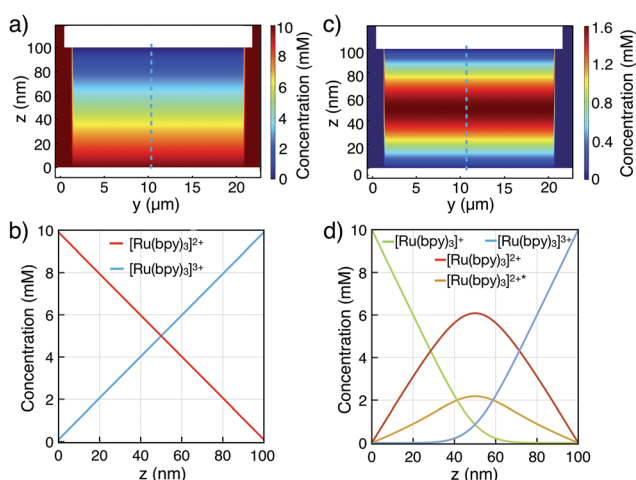


Fig. 5 Finite element simulations of concentration profiles for a 10 mM $[\text{Ru}(\text{bpy})_3]^{2+}$ bulk concentration in the nanochannel. Two-dimensional $[\text{Ru}(\text{bpy})_3]^{2+}$ profiles in (a) redox cycling mode, and (c) annihilation mode. (b and d) Corresponding cross-sectional profiles in the channel center (indicated by the dashed line in both modes, respectively).



- 11 F. F. Fan, in *Electrogenerated Chemiluminescence*, ed. A. J. Bard, CRC Press, New York, 1st edn, 2004, pp. 23–99.
- 12 J. T. Maloy, K. B. Prater and A. J. Bard, *J. Am. Chem. Soc.*, 1971, **93**, 5959–5968.
- 13 G. H. Brilmyer and A. J. Bard, *J. Electrochem. Soc.*, 1980, **127**, 104–110.
- 14 J. E. Bartelt, S. M. Drew and R. M. Wightman, *J. Electrochem. Soc.*, 1992, **139**, 70–74.
- 15 C. Amatore, C. Pebay, L. Servant, N. Sojic, S. Szunerits and L. Thouin, *ChemPhysChem*, 2006, **7**, 1322–1327.
- 16 G. C. Fiaccabrino, M. Koudelka-Hep, Y. T. Hsueh, S. D. Collins and R. L. Smith, *Anal. Chem.*, 1998, **70**, 4157–4161.
- 17 F.-R. F. Fan, D. Cliffler and A. J. Bard, *Anal. Chem.*, 1998, **70**, 2941–2948.
- 18 J. Rodríguez-López, M. Shen, A. B. Nepomnyashchii and A. J. Bard, *J. Am. Chem. Soc.*, 2012, **134**, 9240–9250.
- 19 M. Shen, N. Arroyo-Currás and A. J. Bard, *Anal. Chem.*, 2011, **83**, 9082–9085.
- 20 A. Kapturkiewicz and G. Angulo, *Dalton Trans.*, 2003, 3907–3913.
- 21 M. A. G. Zevenbergen, D. Krapf, M. R. Zuiddam and S. G. Lemay, *Nano Lett.*, 2007, **7**, 384–388.
- 22 D. Mampallil, K. Mathwig, S. Kang and S. G. Lemay, *J. Phys. Chem. Lett.*, 2014, **5**, 636–640.
- 23 Q. Chen, K. McKelvey, M. A. Edwards and H. S. White, *J. Phys. Chem. C*, 2016, **120**, 17251–17260.
- 24 C. Ma, W. Xu, W. R. A. Wichert and P. W. Bohn, *ACS Nano*, 2016, **10**, 3658–3664.
- 25 L. Rassaei, K. Mathwig, S. Kang, H. A. Heering and S. G. Lemay, *ACS Nano*, 2014, **8**, 8278–8284.
- 26 S. Kang, A. F. Nieuwenhuis, K. Mathwig, D. Mampallil, Z. Kostiuhenko and S. G. Lemay, *Faraday Discuss.*, 2016, **193**, 41–50.
- 27 D. Han, G. M. Crouch, K. Fu, L. P. Zaino III and P. W. Bohn, *Chem. Sci.*, 2017, **8**, 5345–5355.
- 28 K. Mathwig and S. G. Lemay, *Micromachines*, 2013, **4**, 138–148.
- 29 S. Kang, K. Mathwig and S. G. Lemay, *Lab Chip*, 2012, **12**, 1262–1267.
- 30 S. Kang, A. F. Nieuwenhuis, K. Mathwig, D. Mampallil and S. G. Lemay, *ACS Nano*, 2013, **7**, 10931–10937.
- 31 D. Mampallil, K. Mathwig, S. Kang and S. G. Lemay, *Anal. Chem.*, 2013, **85**, 6053–6058.
- 32 E. Kätelhön, K. J. Krause, K. Mathwig, S. G. Lemay and B. Wolfrum, *ACS Nano*, 2014, **8**, 4924–4930.
- 33 Q. Wang, J. Rodríguez-López and A. J. Bard, *ChemPhysChem*, 2010, **11**, 2969–2978.
- 34 C. Amatore, F. BonhSomme, J. L. Bruneel, L. Servant and L. Thouin, *J. Electroanal. Chem.*, 2000, **484**, 1–17.
- 35 M. M. Collinson, R. M. Wightman and P. Pastore, *J. Phys. Chem.*, 1994, **98**, 11942–11947.

