



Cite this: *Soft Matter*, 2016, 12, 2887

Supramolecular polymerisation in water; elucidating the role of hydrophobic and hydrogen-bond interactions†

Christianus M. A. Leenders, Matthew B. Baker, Imke A. B. Pijpers, René P. M. Lafleur, Lorenzo Albertazzi, Anja R. A. Palmans* and E. W. Meijer*

Understanding the self-assembly of small molecules in water is crucial for the development of responsive, biocompatible soft materials. Here, a family of benzene-1,3,5-tricarboxamide (BTA) derivatives that comprise a BTA moiety connected to an amphiphilic chain is synthesised with the aim to elucidate the role of hydrophobic and hydrogen-bonding interactions in the self-assembly of these BTAs. The amphiphilic chain consists of an alkyl chain with a length of 10, 11, or 12 methylene units, connected to a tetraethylene glycol (at the periphery). The results show that an undecyl spacer is the minimum length required for these BTAs to self-assemble into supramolecular polymers. Interestingly, exchange studies reveal only minor differences in exchange rates between BTAs containing undecyl or dodecyl spacers. Additionally, IR spectroscopy provides the first experimental evidence that hydrogen-bonding is operative and contributes to the stabilisation of the supramolecular polymers in water.

Received 19th November 2015,
 Accepted 10th February 2016

DOI: 10.1039/c5sm02843d

www.rsc.org/softmatter

Introduction

Self-assembling systems in aqueous solution^{1–5} and low molecular weight (LMW) hydrogelators^{6–10} that display dynamic and responsive behaviour have great potential as scaffolds in biomedical applications. For the development of such systems it is important to thoroughly understand the self-assembly process. In particular, it is important to understand the role of the different non-covalent interactions, as water molecules compete with hydrogen-bonding motifs and solvophobic effects are particularly strong in water. In linear (non-ionic) amphiphiles, the role of hydrophobic domain size in micelle formation has been extensively studied, and a direct correlation has been found between the length of the hydrophobic domain and the critical micelle concentration (CMC).^{11–16} Predicting the self-assembly properties of molecules relying on more than amphiphilicity is difficult, let alone predicting the exchange dynamics of monomers between aggregates. Nevertheless, examples of aqueous self-assembly that combine hydrophobic effects with π - π stacking^{17–22} or make use of directional hydrogen bonds to drive self-assembly have been reported.^{23–31} To further develop general design rules for aqueous self-assembly, studies are

needed which relate systematic modifications of the molecular structure to the self-assembly behaviour and exchange dynamics.

In the last two decades, our group and others have studied the aggregation and phase behaviour of *N,N,N'*-trialkyl-benzene-1,3,5-tricarboxamides (BTAs) in great detail.³² The simple and synthetically accessible structure of the BTA moiety allowed the establishment of structure/property relationships for bulk materials,^{33–38} and unravelling of the mechanism and dynamics of BTA self-assembly in dilute solution.^{39,40} In apolar organic media, the self-assembly of the alkyl-BTAs is dominated by cooperative hydrogen-bond formation, which results in supramolecular polymers that are stabilised by a triple helical array of hydrogen bonds.^{40,41} Recently, we redesigned the BTAs to self-assemble in water, by conjugating either water-soluble metal complexes²⁸ or tetraethylene glycol⁴² (compound **1a**, Scheme 1) to their periphery. Detailed studies, both experimentally as well as computationally, on amphiphilic BTA **1a** have been conducted. The dodecyl chains of **1a** were sufficiently hydrophobic to induce aggregation while the hydrophilic ethylene glycols at the periphery ensured compatibility of the aggregates with water. With microscopic techniques, long and dynamic supramolecular polymers of high aspect ratio were observed, and the mechanism and time-scale of exchange of monomers between different polymers was elucidated.^{43,44} Furthermore, computational studies supported that the supramolecular polymers formed by **1a** were stabilised by hydrogen-bond formation.⁴⁵

Here, we vary the length of the aliphatic spacer from a dodecyl (**1a**) to an undecyl (**1b**), to a decyl spacer (**1c**), respectively,

Institute for Complex Molecular Systems, Eindhoven University of Technology, P.O. Box 513, 5600 MB Eindhoven, The Netherlands. E-mail: A.Palmans@tue.nl, E.W.Meijer@tue.nl

† Electronic supplementary information (ESI) available: Experimental details, characterization by IR and UV spectroscopy and dynamic light scattering, video files of optical microscopy imaging. See DOI: 10.1039/c5sm02843d



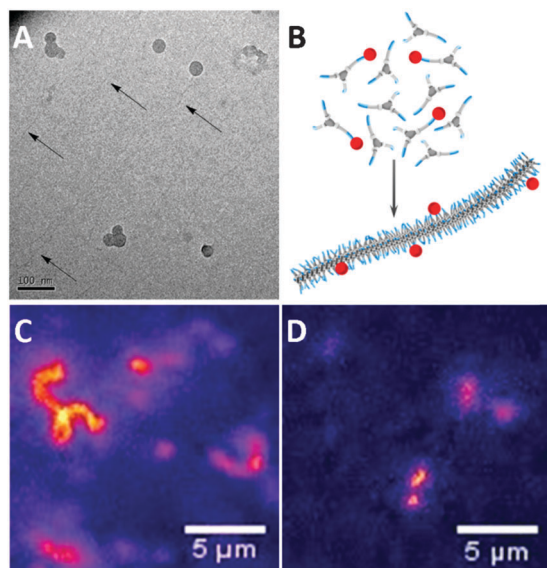


Fig. 2 (A) CryoTEM image of **1b** (0.5 mg mL^{-1}) in H_2O revealing the presence of supramolecular polymers. (B) Schematic representation of mixing a small amount of dye labelled BTAs with unlabelled BTAs prior to injection into water, to allow the assembly of the BTAs into fluorescently labelled fibres. Snapshots from movies taken during fluorescence microscopy (samples were equilibrated at $1 \times 10^{-5} \text{ M}$, and diluted to $1 \times 10^{-6} \text{ M}$ prior to imaging. See ESI† for sample preparation) show (C) fibres formed by **1a** with 5% **3a** and (D) fibres formed by **1b** with 5% **3b** with lengths close to the resolution limit.

we established previously.^{43–45} With this procedure, the dye labelled BTAs incorporate randomly into the formed aggregates, creating a fluorescently labelled structure. When imaging the solution, differences between the samples are observed. The snapshots from full movies (see ESI†) clearly show the presence of supramolecular polymers in samples of **1a** (Fig. 2C). Because of the resolution limit (250 nm) the fibres appear much wider than those observed with cryoTEM. Samples of **1b** also show the presence of elongated aggregates although these were more difficult to capture in a snapshot due to fast diffusion. (Fig. 2D). Furthermore, in line with cryoTEM, no resolved self-assembled structures of **1c** were observed.

The self-assembly of **1a–c** was evaluated in more detail by spectroscopy. UV spectroscopy and fluorescence spectroscopy using the dye Nile Red were applied to identify aggregate formation and the presence of a hydrophobic pocket, respectively. Solutions of **1a–c** were prepared by injecting a small aliquot of a concentrated methanol solution into water to obtain the desired final concentration ($5 \times 10^{-5} \text{ M}$). In all cases, the solutions were optically transparent. Similar to previous results⁴² the UV spectrum of **1a** at 20°C shows two absorption bands at 211 and 226 nm (Fig. 3A), indicating aggregation. In acetonitrile, a solvent in which BTAs are molecularly dissolved,³⁹ the absorption spectrum of **1a** displays a maximum at 207 nm (Fig. 3A). The UV spectrum of **1b** closely resembles that of **1a** at 20°C with two absorption maxima at 211 nm and 226 nm, respectively (Fig. 3A). In contrast, the UV spectrum of **1c** shows only a single absorption maximum at 209 nm at 20°C (Fig. 3A), similar to **1a** in acetonitrile.

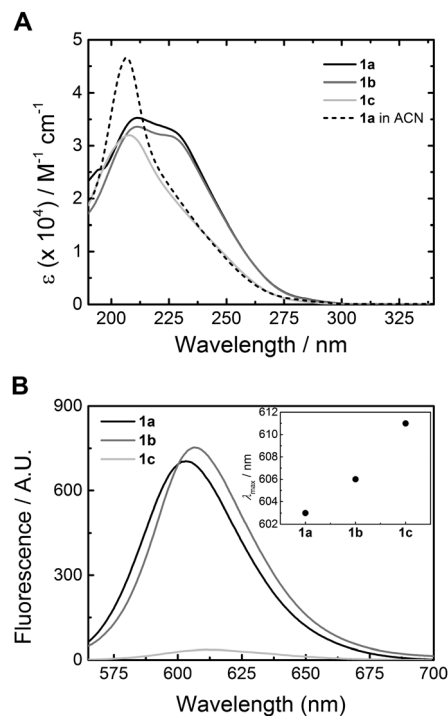


Fig. 3 (A) UV-vis spectra at 20°C of **1a**, **1b**, **1c** in H_2O ($c = 5 \times 10^{-5} \text{ M}$, $\lambda_{\text{ex}} = 550 \text{ nm}$), and **1a** in acetonitrile ($c = 1 \times 10^{-5} \text{ M}$) (B) Fluorescence emission spectra of Nile Red in solutions of **1a–c** in H_2O ($c_{\text{BTA}} = 1 \times 10^{-5} \text{ M}$, $c_{\text{NR}} = 5 \times 10^{-6} \text{ M}$, $\lambda_{\text{ex}} = 550 \text{ nm}$) at 20°C . The inset shows the $\lambda_{\text{max,em}}$ of NR in BTAs **1a–c**.

This indicates that **1b** does, but **1c** does not self-assemble at this concentration and temperature in water.

The formation of a hydrophobic pocket upon self-assembly by **1a–c** was probed using the solvatochromic dye Nile Red.⁴⁹ In pure water, Nile Red shows low fluorescence intensity with λ_{max} at 650 nm. In a more apolar environment, the fluorescence intensity increases and the λ_{max} emission shifts to shorter wavelengths.¹⁰ Solutions of **1a–c** were prepared by injection from methanol into water following the same procedure as described before ($c_{\text{BTA}} = 1 \times 10^{-5} \text{ M}$), a concentrated solution of Nile Red in methanol ($5 \mu\text{L}$ $c_{\text{NR}} = 2.5 \times 10^{-3} \text{ M}$) was added ($c_{\text{NRfinal}} = 5 \times 10^{-6} \text{ M}$), and the solutions were allowed to equilibrate. Compared to water ($\lambda_{\text{max}} = 647 \text{ nm}$, intensity 1.8 a.u.), the fluorescence emission intensity of Nile Red in solutions of **1a** and **1b** increases by two orders of magnitude. This is accompanied by a large blue-shift to $\lambda_{\text{max}} = 603$ and 607 nm for **1a** and **1b**, respectively (Fig. 3B). On the other hand, the emission intensity of **1c** is very low and the emission maximum is less blue-shifted ($\lambda_{\text{max}} = 612 \text{ nm}$). The intensity increase and similar shift in λ_{max} in the solutions of **1a** and **1b** indicate aggregation and the formation of a hydrophobic pocket in both cases, while in the case of **1c** no significant hydrophobic pocket is formed.

Hydrogen-bond formation in water probed by FT-IR spectroscopy

The above results show that BTAs **1a** and **1b** self-assemble into supramolecular polymers of high aspect ratio while **1c** does not.



This indicates that a critical hydrophobic spacer length of 11 methylenes is required for the formation of these supra-molecular polymers. To investigate whether hydrogen bonds form between the BTAs in water, IR measurements were carried out. We selected D₂O as the solvent instead of H₂O since the O–H vibrations of water obscure the amide vibrations. Because of the increased mass of deuterium compared to hydrogen, the frequencies of the vibrations change. To assign the amide vibrations, we first investigated the deuterium-amide vibrations in bulk by exchanging all labile OH and NH protons of **1a** with deuterium to afford **1a-d₆**. The bulk FT-IR spectrum of **1a-d₆** recorded at 25 °C (Fig. 4, black curve, Fig. S3, ESI†) was compared to that of **1a** (Fig. S1, ESI†). In addition, we measured the IR spectrum of **1a** in MeOD (*c* = 50 mg mL⁻¹), a solvent in which BTAs are molecularly dissolved and intermolecular hydrogen bonding is absent (Fig. 4, grey curve).

In the bulk, the behaviour of **1a-d₆** is similar to that of **1a** (Fig. S1, ESI†). At 25 °C the N–D stretch vibration at 2363 cm⁻¹ (Fig. S3, ESI†) is indicative for intermolecular hydrogen-bond formation. The amide I vibration at 25 °C is split up into two very sharp peaks at 1634 and 1620 cm⁻¹, respectively, whereas the amide II vibration has shifted to 1468 cm⁻¹. The origin of the split in the vibration at ~1600 cm⁻¹ for **1a-d₆** is unclear at the moment and warrants more detailed investigations. The IR spectrum of **1a** in MeOD shows an amide I vibration at a frequency of 1648 cm⁻¹, a value indicative of the absence of intermolecular hydrogen bonds.

With these spectra as reference, the FT-IR spectrum of a solution of **1a** in D₂O (*c* = 50 mg mL⁻¹) was measured (Fig. 4, blue curve). In D₂O, the amide I gives a particularly sharp vibration at 1635 cm⁻¹, which overlaps perfectly with one of the amide I vibrations observed in the bulk. This vibration – when compared to the amide I vibration at 1648 cm⁻¹ in MeOD solution – clearly indicates the presence of intermolecular hydrogen bonds. The amide II vibration in D₂O solution has

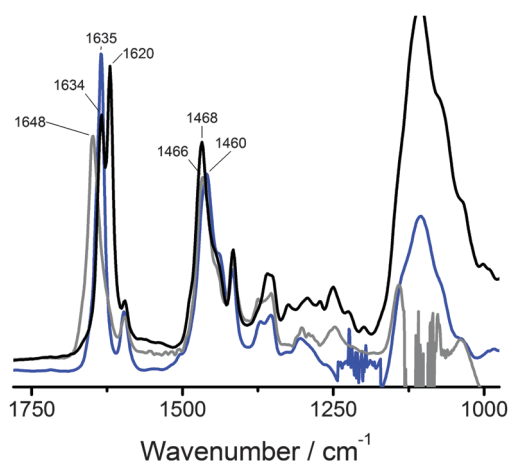


Fig. 4 FT-IR spectrum of **1a-d₆** in the bulk at 25 °C (black), FT-IR spectrum of a solution of **1a** (*c* = 50 mg mL⁻¹) at room temperature in D₂O (blue) displaying an amide I vibration indicative for hydrogen bonding and in MeOD (grey) displaying an amide I vibration indicative for the absence of hydrogen bonding.

a frequency of 1460 cm⁻¹ and is more difficult to interpret. In the field of protein secondary structure determination by FT-IR spectroscopy, the amide I vibration is commonly considered to be highly informative while a correlation between the amide II vibration and the secondary structure is not well established due to its low sensitivity to the secondary structure.^{50–52} Furthermore, upon N-deuteration the amide II vibration overlaps with aliphatic CH₂ vibrations⁵³ and with the bending mode of HOD.^{54,55} This may cause the position of the amide II vibration to be different for **1a** in the solid state compared to **1a-d₆** in the solid state and the same position for **1a-d₆** in the solid state and for **1a** in D₂O. Therefore, as is common in secondary structure determination of peptides, solely the amide I vibration is regarded informative for the presence of hydrogen bonds. As such, these results provide the first convincing experimental evidence for the presence of intermolecular hydrogen bonds stabilising BTA assemblies in water.

Thermal stability of supramolecular polymers in water

The thermal stability of the assemblies formed by **1a** and **1b** was assessed by variable temperature UV measurements (Fig. S5, ESI†). At 20 °C, the UV spectrum of **1a** shows two absorption bands at 211 nm and 226 nm (Fig. 2A). Only after heating to above 50 °C the absorption spectrum changes, which finally results in an absorption band at 207 nm at 70 °C. Around this temperature the lower critical solution temperature (LCST) of **1a** is reached. We hypothesise that at 70 °C the assemblies are disrupted but the molecularly dissolved state is not reached. Rather, small aggregates held together by the amphiphilic nature of the molecules and hydrophobicity resulting from the LCST are present. Similar to **1a**, the UV spectrum of **1b** is stable up to 50 °C. At 60 °C, the intensity around 226 nm significantly drops resulting in a single absorption maximum at 209 nm. This implies that the self-assembly of **1b** is somewhat more sensitive to temperature than the self-assembly of **1a** and indicates a somewhat lower stability of the assemblies. These observations are in line with the results of variable temperature IR measurements of **1a** and **1b** in the bulk, which show a higher thermal stability of the hydrogen bonds in **1a**; in the bulk of **1a** the hydrogen bonds are lost above 45 °C compared to 35 °C for **1b** (Fig. S1 and S2 (ESI†), respectively).

Fluorescence exchange studies

After studying the effect of varying the spacer length on the self-assembly properties, we were interested in the monomer exchange rates within the series **1a-c**. Förster resonance energy transfer (FRET) has been widely relied on to probe dynamics and exchange processes,^{56–60} and has recently been introduced to follow the reversible assembly and disassembly of BTA-based systems.^{43,45} By co-assembling either **2a-c** (FRET donor) or **3a-c** (FRET acceptor) (5 mol%) with the parent BTA **1a-c** fibres labelled with either Cy3 or Cy5 were prepared (Fig. 5A, *c*_{BTA} = 1 × 10⁻⁵ M). Mixing these solutions and monitoring the FRET ratio allowed the determination of the rate of equilibration. Both systems of **1a** and **1b** show a rise in the FRET ratio upon mixing, indicating monomer exchange between the stacks (Fig. 5B). This increase in



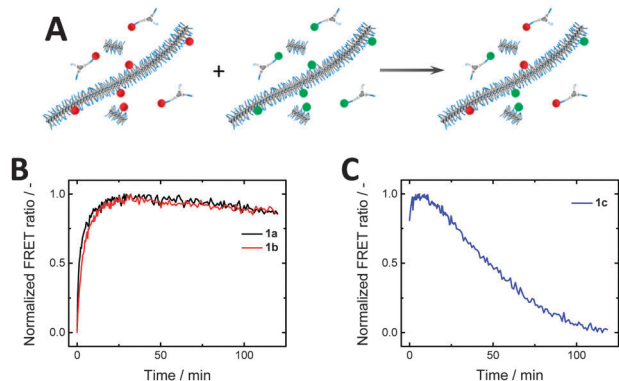


Fig. 5 (A) Schematic of FRET exchange experiment. (B) FRET experiments of **1a** and **1b** show similar exchange kinetics, while (C) **1c** shows erratic behaviour due to poor self-assembly ($c_{\text{BTA}} = 1 \times 10^{-5}$ M).

FRET ratio follows similar kinetics for **1a** and **1b**, plateauing around 40 minutes. Both of these curves can be fit by a single exponential, resulting in similar time constants (Fig. S6, ESI†). In contrast, for **1c** the FRET ratio as function of time proved very erratic and highly irreproducible (Fig. 5C), likely due to poor self-assembly.

Discussion

The influence of the number of carbons in the aliphatic spacer on the self-assembly of BTAs in water is noteworthy and a C10 spacer corresponds to the limiting case in which self-assembly does not occur. BTAs **1a** and **1b** self-assemble in water into supramolecular polymers of high aspect ratio. Both form fibres of micrometres in length and of a comparable diameter of less than 10 nm. In sharp contrast, **1c** does not form fibrillar aggregates at the same conditions. This indicates that a critical hydrophobic spacer length of 11 carbon atoms is required in order for these BTAs to self-assemble in the concentration regime studied. If we consider **1a–c** as simple ethylene-oxide based amphiphiles, the removal of one $-\text{CH}_2-$ unit is expected to increase the CMC by less than an order of magnitude and this effect becomes smaller with increasing chain length.^{12–16} Going from **1b** to **1c** three $-\text{CH}_2-$ units are removed (one per side chain), which according to this trend would increase the CMC by roughly two orders of magnitude. Although we have not been able to determine the CMC of **1a** and **1b**, it is below 10^{-6} M for both compounds, while at two orders of magnitude higher concentration no aggregation was observed for **1c**. This suggests that our system does not behave like simple (non-ionic) amphiphiles. Instead, we propose that the hydrophobic–hydrophilic balance in the BTA molecular design plays a double role. First the hydrophobic collapse represents a primary driving force for aggregation, this was also concluded from MD simulations.⁴⁵ When the hydrophobic domain is too small, the molecules become too soluble in water, whereas the molecules become insoluble when the domain is too large. Secondly, although hydrophobic collapse may suffice to induce aggregation, a sufficiently large hydrophobic pocket around the benzene-tricarboxamide core stabilises amide–amide

intermolecular hydrogen-bonding interactions, and shields these interactions from the surrounding water. In that case, intermolecular hydrogen bonding further stabilises the formed aggregates. This is supported by FT-IR measurements on solutions of **1a** in aqueous solution, which confirm the formation of hydrogen bonds. The Nile Red experiments show that the polarity of the hydrophobic local domain in aggregates of **1a** and **1b** is very similar, indicating that the BTA core is effectively shielded from surrounding water in both systems. In the case of **1c**, the hydrophobic collapse and hydrophobic shielding appear insufficient to enable interactions between the BTAs needed for the formation of supramolecular polymers.

The difference of the thermodynamic stability in water as well as the thermal stability in bulk between **1a** and **1b** is more subtle. The loss of hydrogen bonds in IR in the bulk occurs at 45 °C and 35 °C for **1a** and **1b**, respectively, and the stability of the self-assembled structures formed by **1b** is lower. Interestingly, the exchange dynamics between supramolecular polymers of BTAs with C11 (**1b**) or C12 (**1a**) spacers seem to depend less on the spacer length. In a previous study the rate of monomer exchange between BTA-based supramolecular polymers in water was shown to depend on the character of the hydrophobic spacer.⁴⁵ The stereogenic nature of that modification was concluded to be the major cause for the change in exchange dynamics. It is of note that within these experiments we observe the exchange of the dye labelled monomers, and not the unlabelled monomers. Despite this limitation, the similar profiles in the FRET experiment observed for **1a** and **1b** suggest that the extension of the hydrophobic substituent by one CH_2 unit does not necessarily affect the exchange dynamics.

Conclusions

We have studied the role of hydrophobic and hydrogen-bonding interactions on the aqueous self-assembly behaviour and exchange dynamics of BTA derivatives **1a–c**. The results stress the crucial importance of a delicate balance between hydrophobic and hydrophilic units within the BTA: a decyl spacer is too short to provide the hydrophobicity required for forming ordered assemblies, while the undecyl and dodecyl spacer do result in stable supramolecular polymer formation in water. Furthermore, FT-IR spectroscopy provides the first experimental evidence for stabilisation through hydrogen-bond formation in these supramolecular polymers in aqueous solution and confirms previous computational findings. Finally, while variation of the hydrophobic spacer results in differences in aggregate size and thermal stability, the exchange dynamics of the supramolecular polymers appear to not show such a direct relation with the spacer length. This is exemplified by the equally fast exchange rates observed for **1a** and **1b**.

Acknowledgements

We gratefully acknowledge financial support from the Dutch Ministry of Education, Culture and Science (Gravity program 024.001.035) and the European Research Council (FP7/2007-2013, ERC Grant Agreement 246829). M. B. B. and L. A. are grateful for



financial support from the Netherlands Institute for Regenerative Medicine. L. A. is grateful for financial support from the Netherlands Organization for Scientific Research (NWO – VENI Grant: 722.014.010). The ICMS animation studio is acknowledged for providing the artwork.

References

- 1 A. R. Hirst, B. Escuder, J. F. Miravet and D. K. Smith, *Angew. Chem., Int. Ed.*, 2008, **47**, 8002–8018.
- 2 A. Noro, M. Hayashi and Y. Matsushita, *Soft Matter*, 2012, **8**, 6416–6429.
- 3 E. A. Appel, J. del Barrio, X. J. Loh and O. A. Scherman, *Chem. Soc. Rev.*, 2012, **41**, 6195–6214.
- 4 J. H. Collier, *Soft Matter*, 2008, **4**, 2310–2315.
- 5 B. Rybtchinski, *ACS Nano*, 2011, **5**, 6791–6818.
- 6 T. Rehm and C. Schmuck, *Chem. Commun.*, 2008, 801–813.
- 7 L. A. Estroff and A. D. Hamilton, *Chem. Rev.*, 2004, **104**, 1201–1217.
- 8 M. de Loos, J. H. van Esch, R. M. Kellogg and B. L. Feringa, *Tetrahedron*, 2007, **63**, 7285–7301.
- 9 Y. Yan, Y. Lin, Y. Qiao and J. Huang, *Soft Matter*, 2011, **7**, 6385–6398.
- 10 J. Boekhoven, A. M. Brizard, P. van Rijn, M. C. A. Stuart, R. Eelkema and J. H. van Esch, *Angew. Chem., Int. Ed.*, 2011, **50**, 12285–12289.
- 11 K. Esumi and M. Ueno, *Structure-Performance Relationships in Surfactants*, CRC Press, 2003.
- 12 P. D. T. Huibers, V. S. Lobanov, A. R. Katritzky, D. O. Shah and M. Karelson, *Langmuir*, 1996, **12**, 1462–1470.
- 13 A. R. Katritzky, L. M. Pacureanu, S. H. Slavov, D. A. Dobchev and M. Karelson, *Ind. Eng. Chem. Res.*, 2008, **47**, 9687–9695.
- 14 M. Mattei, G. M. Kontogeorgis and R. Gani, *Ind. Eng. Chem. Res.*, 2013, **52**, 12236–12246.
- 15 L.-J. Chen, S.-Y. Lin and C.-C. Huang, *J. Phys. Chem. B*, 1998, **102**, 4350–4356.
- 16 D. Chandler, *Nature*, 2005, **437**, 640–647.
- 17 D. Görl, X. Zhang and F. Würthner, *Angew. Chem., Int. Ed.*, 2012, **51**, 6328–6348.
- 18 Y. Tidhar, H. Weissman, S. G. Wolf, A. Gulino and B. Rybtchinski, *Chem. – Eur. J.*, 2011, **17**, 6068–6075.
- 19 V. Villari, P. Mineo, E. Scamporrino and N. Micali, *Chem. Phys.*, 2012, **409**, 23–31.
- 20 J. M. Kroon, R. B. M. Koehorst, M. van Dijk, G. M. Sanders and E. J. R. Sudholter, *J. Mater. Chem.*, 1997, **7**, 615–624.
- 21 L. Brunsveld, B. G. G. Lohmeijer, J. Vekemans and E. W. Meijer, *Chem. Commun.*, 2000, 2305–2306.
- 22 D.-W. Lee, T. Kim, I.-S. Park, Z. Huang and M. Lee, *J. Am. Chem. Soc.*, 2012, **134**, 14722–14725.
- 23 A. Pal, S. Karthikeyan and R. P. Sijbesma, *J. Am. Chem. Soc.*, 2010, **132**, 7842–7843.
- 24 E. Obert, M. Bellot, L. Bouteiller, F. Andrioletti, C. Lehen-Ferrenbach and F. Boue, *J. Am. Chem. Soc.*, 2007, **129**, 15601–15605.
- 25 J. D. Hartgerink, E. Beniash and S. I. Stupp, *Science*, 2001, **294**, 1684–1688.
- 26 B. J. Cafferty, I. Gállego, M. C. Chen, K. I. Farley, R. Eritja and N. V. Hud, *J. Am. Chem. Soc.*, 2013, **135**, 2447–2450.
- 27 S. Han, S. Cao, Y. Wang, J. Wang, D. Xia, H. Xu, X. Zhao and J. R. Lu, *Chem. – Eur. J.*, 2011, **17**, 13095–13102.
- 28 P. Besenius, G. Portale, P. H. H. Bomans, H. M. Janssen, A. R. A. Palmans and E. W. Meijer, *Proc. Natl. Acad. Sci. U. S. A.*, 2010, **107**, 17888–17893.
- 29 K. J. C. van Bommel, C. van der Pol, I. Muizebelt, A. Friggeri, A. Heeres, A. Meetsma, B. L. Feringa and J. van Esch, *Angew. Chem., Int. Ed.*, 2004, **43**, 1663–1667.
- 30 N. Chebotareva, P. H. H. Bomans, P. M. Frederik, N. Sommerdijk and R. P. Sijbesma, *Chem. Commun.*, 2005, 4967–4969.
- 31 P. Y. W. Dankers, T. M. Hermans, T. W. Baughman, Y. Kamikawa, R. E. Kieltyka, M. M. C. Bastings, H. M. Janssen, N. A. J. M. Sommerdijk, A. Larsen, M. J. A. van Luyn, A. W. Bosman, E. R. Popa, G. Fytas and E. W. Meijer, *Adv. Mater.*, 2012, **24**, 2703–2709.
- 32 S. Cantekin, T. F. A. de Greef and A. R. A. Palmans, *Chem. Soc. Rev.*, 2012, **41**, 6125–6137.
- 33 A. Timme, R. Kress, R. Q. Albuquerque and H. W. Schmidt, *Chem. – Eur. J.*, 2012, **18**, 8329–8339.
- 34 M. Blomenhofer, S. Ganzleben, D. Hanft, H.-W. Schmidt, M. Kristiansen, P. Smith, K. Stoll, D. Mäder and K. Hoffmann, *Macromolecules*, 2005, **38**, 3688–3695.
- 35 F. Abraham and H.-W. Schmidt, *Polymer*, 2010, **51**, 913–921.
- 36 M. Masuda, P. Jonkheijm, R. P. Sijbesma and E. W. Meijer, *J. Am. Chem. Soc.*, 2003, **125**, 15935–15940.
- 37 C. F. C. Fitie, W. S. C. Roelofs, P. Magusin, M. Wubbenhorst, M. Kemerink and R. P. Sijbesma, *J. Phys. Chem. B*, 2012, **116**, 3928–3937.
- 38 T. Mes, M. M. J. Smulders, A. R. A. Palmans and E. W. Meijer, *Macromolecules*, 2010, **43**, 1981–1991.
- 39 M. M. J. Smulders, A. P. H. J. Schenning and E. W. Meijer, *J. Am. Chem. Soc.*, 2008, **130**, 606–611.
- 40 P. J. M. Stals, M. M. J. Smulders, R. Martin-Rapun, A. R. A. Palmans and E. W. Meijer, *Chem. – Eur. J.*, 2009, **15**, 2071–2080.
- 41 I. A. W. Filot, A. R. A. Palmans, P. A. J. Hilbers, R. A. van Santen, E. A. Pidko and T. F. A. de Greef, *J. Phys. Chem. B*, 2010, **114**, 13667–13674.
- 42 C. M. A. Leenders, L. Albertazzi, T. Mes, M. M. E. Koenigs, A. R. A. Palmans and E. W. Meijer, *Chem. Commun.*, 2013, **49**, 1963–1965.
- 43 L. Albertazzi, F. J. Martinez-Veracochea, C. M. A. Leenders, I. K. Voets, D. Frenkel and E. W. Meijer, *Proc. Natl. Acad. Sci. U. S. A.*, 2013, **110**, 12203–12208.
- 44 L. Albertazzi, D. van der Zwaag, C. M. A. Leenders, R. Fitzner, R. W. van der Hofstad and E. W. Meijer, *Science*, 2014, **344**, 491–495.
- 45 M. B. Baker, L. Albertazzi, I. K. Voets, C. M. A. Leenders, A. R. A. Palmans, G. M. Pavan and E. W. Meijer, *Nat. Commun.*, 2015, **6**, DOI: 10.1038/ncomms7234.
- 46 E. E. Dormidontova, *Macromolecules*, 2004, **37**, 7747–7761.
- 47 L. Brunsveld, A. Schenning, M. A. C. Broeren, H. M. Janssen, J. Vekemans and E. W. Meijer, *Chem. Lett.*, 2000, 292–293.



- 48 P. J. M. Stals, J. F. Haveman, R. Martin-Rapun, C. F. C. Fitie, A. R. A. Palmans and E. W. Meijer, *J. Mater. Chem.*, 2009, **19**, 124–130.
- 49 M. C. A. Stuart, J. C. van de Pas and J. B. F. N. Engberts, *J. Phys. Org. Chem.*, 2005, **18**, 929–934.
- 50 W. K. Surewicz and H. H. Mantsch, *Biochim. Biophys. Acta, Protein Struct. Mol. Enzymol.*, 1988, **952**, 115–130.
- 51 M. Jackson and H. H. Mantsch, *Crit. Rev. Biochem. Mol. Biol.*, 1995, **30**, 95–120.
- 52 J. Kong and S. Yu, *Acta Biochim. Biophys. Sin.*, 2007, **39**, 549–559.
- 53 M. Beer, H. B. Kessler and G. B. B. M. Sutherland, *J. Chem. Phys.*, 1958, **29**, 1097–1104.
- 54 H. Susi, S. N. Timasheff and L. Stevens, *J. Biol. Chem.*, 1967, **242**, 5460–5466.
- 55 C. A. Swenson, *Spectrochim. Acta*, 1965, **21**, 987–993.
- 56 J.-Y. Kim, C. Kim and N. K. Lee, *Nat. Commun.*, 2015, **6**, 6992.
- 57 Y. Zhao, P. Schapotschnikow, T. Skajaa, T. J. Vlugt, W. J. Mulder, C. de Mello Donega and A. Meijerink, *Small*, 2014, **10**, 1163–1170.
- 58 J. Bai and R. E. Pagano, *Biochemistry*, 1997, **36**, 8840–8848.
- 59 E. S. Barrett, T. J. Dale and J. Rebek, Jr., *J. Am. Chem. Soc.*, 2007, **129**, 8818–8824.
- 60 K. Petkau-Milroy, D. A. Uhlenheuer, A. J. H. Spiering, J. A. J. M. Vekemans and L. Brunsveld, *Chem. Sci.*, 2013, **4**, 2886–2891.

