

Cite this: *Chem. Sci.*, 2023, 14, 13419

All publication charges for this article have been paid for by the Royal Society of Chemistry

Controlling primary chain dispersity in network polymers: elucidating the effect of dispersity on degradation†

Takanori Shimizu,^{ac} Richard Whitfield,^{id}*^a Glen R. Jones,^a Ibrahim O. Raji,^b Dominik Konkolewicz,^{id}^b Nghia P. Truong,^{id}^a and Athina Anastasaki^{id}*^a

Although dispersity has been demonstrated to be instrumental in determining many polymer properties, current synthetic strategies predominantly focus on tailoring the dispersity of linear polymers. In contrast, controlling the primary chain dispersity in network polymers is much more challenging, in part due to the complex nature of the reactions, which has limited the exploration of properties and applications. Here, a one-step method to prepare networks with precisely tuned primary chain dispersity is presented. By using an acid-switchable chain transfer agent and a degradable crosslinker in PET-RAFT polymerization, the *in situ* crosslinking of the propagating polymer chains was achieved in a quantitative manner. The incorporation of a degradable crosslinker, not only enables the accurate quantification of the various primary chain dispersities, post-synthesis, but also allows the investigation and comparison of their respective degradation profiles. Notably, the highest dispersity networks resulted in a 40% increase in degradation time when compared to their lower dispersity analogues, demonstrating that primary chain dispersity has a substantial impact on the network degradation rate. Our experimental findings were further supported by simulations, which emphasized the importance of higher molecular weight polymer chains, found within the high dispersity materials, in extending the lifetime of the network. This methodology presents a new and promising avenue to precisely tune primary chain dispersity within networks and demonstrates that polymer dispersity is an important parameter to consider when designing degradable materials.

Received 2nd October 2023
Accepted 9th November 2023

DOI: 10.1039/d3sc05203f

rsc.li/chemical-science

Introduction

Control over polymer molar mass distributions (MMDs) has recently received considerable attention, with the development of numerous successful methods that allow for polymers with specific dispersities to be synthesized while targeting precise molecular weights and maintaining high end-group functionality.^{1–9} Historically, blending methods were commonplace,¹⁰ where a desired molar mass distribution could be obtained by mixing a number of polymers of different molecular weights, but more recently a number of one-pot strategies have been developed. Fors and coworkers pioneered a method where the molar mass distribution could be precisely controlled by gradually adding initiator into various controlled

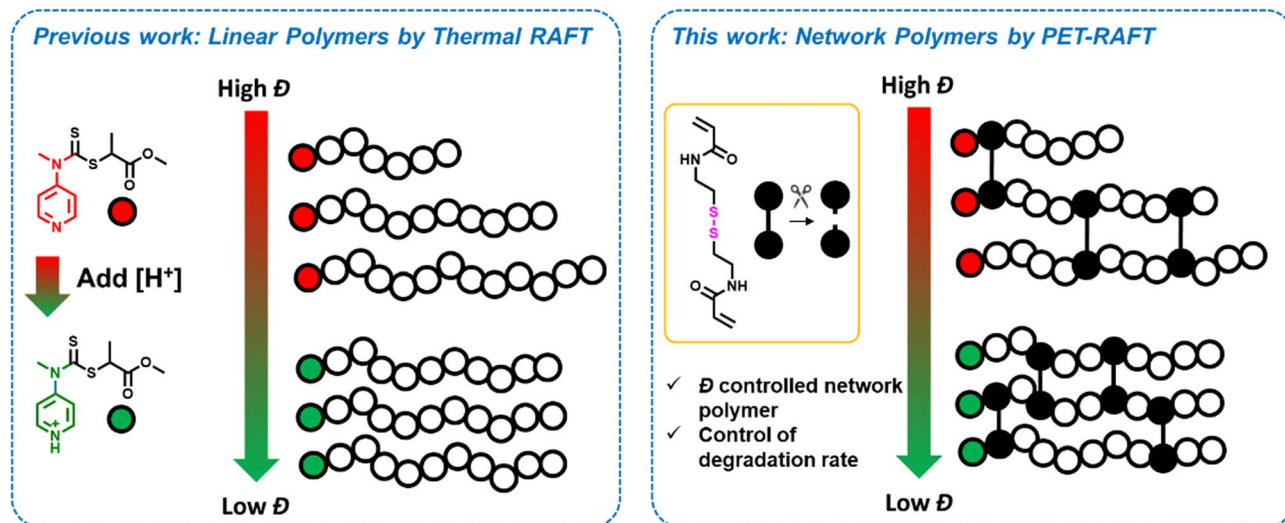
polymerizations.^{11–16} In addition, flow chemistry has been combined with mathematical models to allow specific molar mass distributions to be designed and prepared.^{17–26} Finally, a range of chemical methods have been developed which allow polymerization equilibria to be directly manipulated and the dispersity to be systematically tuned. For example, Chiu and coworkers utilized the photo-isomerism of initiators in cationic polymerization to vary dispersity.²⁷ In the area of controlled radical polymerization, both our group and the Matyjaszewski group illustrated that by lowering the catalyst concentration in atom transfer radical polymerization (ATRP), the dispersity of the resulting polymers could be incrementally increased.^{28–32} In addition, it has been demonstrated that dispersity control was possible by adding a comonomer in organocatalyzed living radical polymerization (O-LRP)³³ and that by mixing chain transfer agents (CTAs) in reversible addition-fragmentation chain transfer (RAFT) polymerization, dispersity could be tuned while maintaining high end-group fidelity.^{34–36} Most recently, our group expanded dispersity control to multiblock copolymers using a switchable CTA in thermal RAFT and by simply adding acid or base alongside each monomer, a low, intermediate or high dispersity block could be obtained.^{37,38}

^aLaboratory of Polymeric Materials, Department of Materials, ETH Zurich, Vladimir Prelog Weg 5, 8093 Zurich, Switzerland. E-mail: richard.whitfield@mat.ethz.ch; Athina.Anastasaki@mat.ethz.ch

^bDepartment of Chemistry and Biochemistry, Miami University, 651 E High St, Oxford, OH 45056, USA

^cScience & Innovation Center, Mitsubishi Chemical Corporation, 1000 Kamoshida-cho, Aoba-ku, Yokohama-shi, Kanagawa 227-8502, Japan

† Electronic supplementary information (ESI) available. See DOI: <https://doi.org/10.1039/d3sc05203f>



Scheme 1 Previous approaches to tune dispersity using a switchable RAFT agent and our current approach to time the primary chain dispersity within gels to investigate their degradation.

The motivation behind these recent achievements was to elucidate the effect of dispersity on a number of different polymer properties. As such, various mechanical, thermal, rheological and self-assembly effects have been reported.^{11,20,39,40} For instance, the Fors group elegantly demonstrated that variations in the molar mass distribution influenced chain stiffness, yield and tensile strengths, toughness and hysteresis energy.^{41,42} In addition, the same group illustrated the role of dispersity in tuning the domain spacing of self-assembled block copolymer thin films,⁴³ and most recently, the Junkers group revealed that the dispersity of the core-forming polymer block has a significant effect on polymer self-assembly.⁴⁴ Very interestingly, by using higher dispersity core-forming blocks, lower dispersity nanoparticles could be prepared. In addition, dispersity has been shown to play a significant role in determining polymer brush behaviour, with effects on the interfacial and mechanical properties of brush-grafted nanoparticles demonstrated.^{45–52}

Despite these impressive advances in dispersity control, the vast majority of current reports focus almost exclusively on linear polymers, while the development of methods to control primary chain dispersity within polymer networks and any potential effect on their properties remains far less explored.^{53–55} This is because controlling dispersity in polymer networks presents several challenges. For example, blending and feeding/flow methods may be incompatible in tuning dispersity in networks, unless multistep procedures are utilized.^{53,56} Chemical methods also face the challenge of maintaining control as the system becomes highly viscous and then heterogeneous after gelation.⁵⁷ In particular, deactivation in ATRP and degenerative chain transfer in RAFT are essential in maintaining control, but these are both potentially compromised at high viscosities, thus creating the potential for additional termination.⁵⁸ As such, the synthesis of networks with tuneable primary chain dispersity in a single step would be a significant development. However, to the best of our knowledge, there are only limited examples where

dispersity-controlled networks have been prepared on variation of the catalyst concentration in ATRP.^{59,60} Matyjaszewski and coworkers demonstrated that decreasing copper concentration in activator regenerated by electron transfer (ARGET) ATRP resulted in a higher primary chain dispersity and an earlier gel point.⁶⁰ However, the primary chain dispersity of the resulting networks was not measured. Most recently, Kopeck and coworkers reported the primary chain dispersity of networks by using dispersity values obtained from analogous linear polymerization reaction conditions.⁵⁹ There is therefore an ongoing challenge in terms of both the synthesis and characterization of these materials, so any influence of primary chain dispersity on many properties has yet to be ascertained. This includes the effect on degradability, which is arguably one of the most useful and important properties that can be incorporated into networks, with precise tuning of network degradation essential for numerous applications including drug delivery, tissue engineering, wound healing, pollution control, agriculture (water and nutrient retention in soil) and personal care (cosmetics and skin care).^{61–67}

In this work, we sought to develop a high-yielding synthetic methodology to prepare primary dispersity-controlled network polymers, with full incorporation of starting monomer, crosslinker and polymer chains. By utilising a degradable crosslinker within these materials, we were able to directly characterize the various primary chain dispersity materials and then accurately study the effect on their degradation rate (Scheme 1). We demonstrate that dispersity has a significant effect on the rate of degradation, with higher dispersity polymers degrading more slowly than their lower dispersity analogues. Finally, we perform simulations to further understand and verify this difference.

Results and discussion

In search for an ideal system to tune dispersity in network polymers, there were three essential criteria that it was



necessary to fulfil. First, excellent control over these complex syntheses must be achieved so as to allow uniform networks to be prepared. Second, quantitative conversions should be obtained in order to enable the complete incorporation of all the starting materials (*i.e.* monomer and crosslinker). Last, but not least, a simple handle to systematically vary the dispersity must be established while high livingness is ensured. We selected photoinduced electron/energy transfer reversible addition-fragmentation chain transfer polymerization (PET-RAFT) polymerization as this has previously been shown to afford lower amounts of termination and higher uniformity networks upon *in situ* crosslinking of the propagating polymer chains.^{58,68} Inspired by seminal work on thermal RAFT polymerization, we envisaged that the addition of various amounts of acid in the presence of the switchable CTA would enable the desired control over dispersity.³⁷ However, given that the switchable CTA has not previously been employed to tune dispersity in PET-RAFT and the effect of the acid on the photocatalyst activity was unclear, the first objective was to develop suitable reaction conditions for a model linear polymerization system. Dimethylacrylamide (DMA) was selected as the model monomer, methyl 2-[methyl(4-pyridinyl)carbamothioylthio]propionate as the pH switchable CTA and eosin Y (EY) as the photocatalyst (Fig. 1a).^{37,69–71} All polymerizations were performed in a mixture of water and DMF under blue light irradiation, and first the amount of EY was varied in the absence of acid (0.005, 0.02 and 0.04 equivalents with respect to the CTA). It is expected that

without acid, the CTA is in its “low activity” (low chain transfer constant state), and as such high dispersity polymers should be prepared. Indeed, after 4 hours of polymerization, broad, yet monomodal molar mass distributions were obtained. In all cases, with high final dispersity values ($\bar{D} = 1.58$ – 1.69 , Fig. 1b, S1 and Table S1†). Even though 0.04 equivalents had previously been reported as optimal for a trithiocarbonate CTA,⁷² for the switchable CTA, lower equivalents of EY (0.005 and 0.02 equivalents) were necessary to ensure near-quantitative conversions ($\sim 95\%$) and an excellent agreement between theoretical and obtained molecular weights (Table S1,† entries 2 and 3). Full incorporation of the CTA on the polymer chains was evidenced by UV detector analysis from size exclusion chromatography (SEC) and the high livingness achieved under these polymerization conditions was further confirmed by performing a successful chain extension experiment (Fig. 1c, s2 and Table S2†). It is also worth highlighting that precisely obtaining the targeted molecular weight, even at high monomer conversion is not usually the case for many previously developed dispersity-controlled methodologies where a huge deviation from the theoretical values is typically observed when targeting high dispersity polymers.^{28,30,31} As this work aims to subsequently apply the optimized conditions to the synthesis of network polymers, it is necessary to be able to attain comparable molecular weights regardless of the targeted primary chain dispersity as networks with comparable compositions are the ultimate goal.

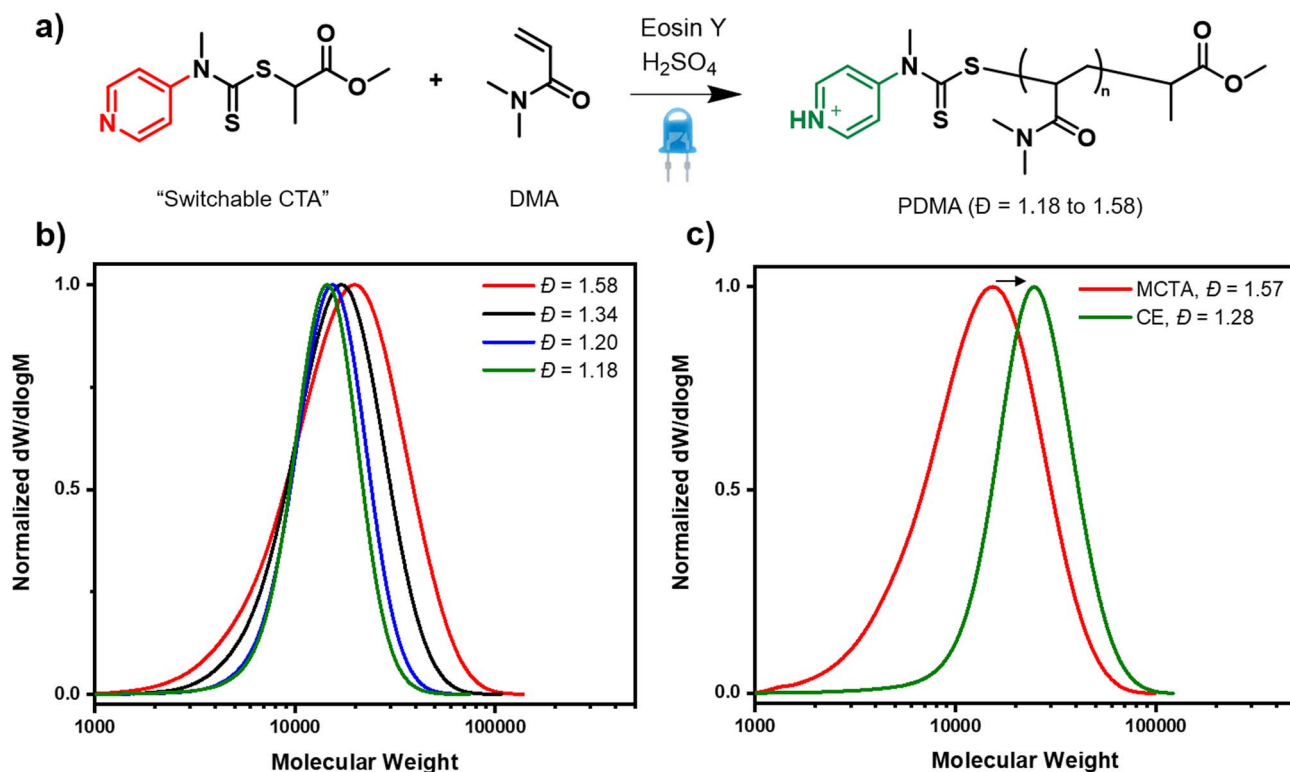


Fig. 1 Dispersity control for linear PDMA prepared with a switchable CTA and PET-RAFT. In (a) a scheme of the polymerization reaction is shown. SEC traces show the molar mass distributions of (b) various dispersity homopolymers obtained with different amounts of acid and (c) a successful chain extension of high dispersity PDMA in the presence of acid.

Our next aim was to obtain networks with various lower primary chain dispersities, so we conducted three further syntheses with various acid concentrations (0.25, 0.5 and 3 equivalents w.r.t. CTA). In all cases, all monomer, crosslinker and polymer were quantitatively incorporated and after extraction, all acid and dye were successfully removed (Fig. S10[†]). On cleavage of the crosslinker with DTT, monomodal molar mass distributions were observed and the primary chain dispersity was measured to be 1.55, 1.40 and 1.28, respectively (Table 1, entries 2–4). This demonstrated the overall effectiveness of the

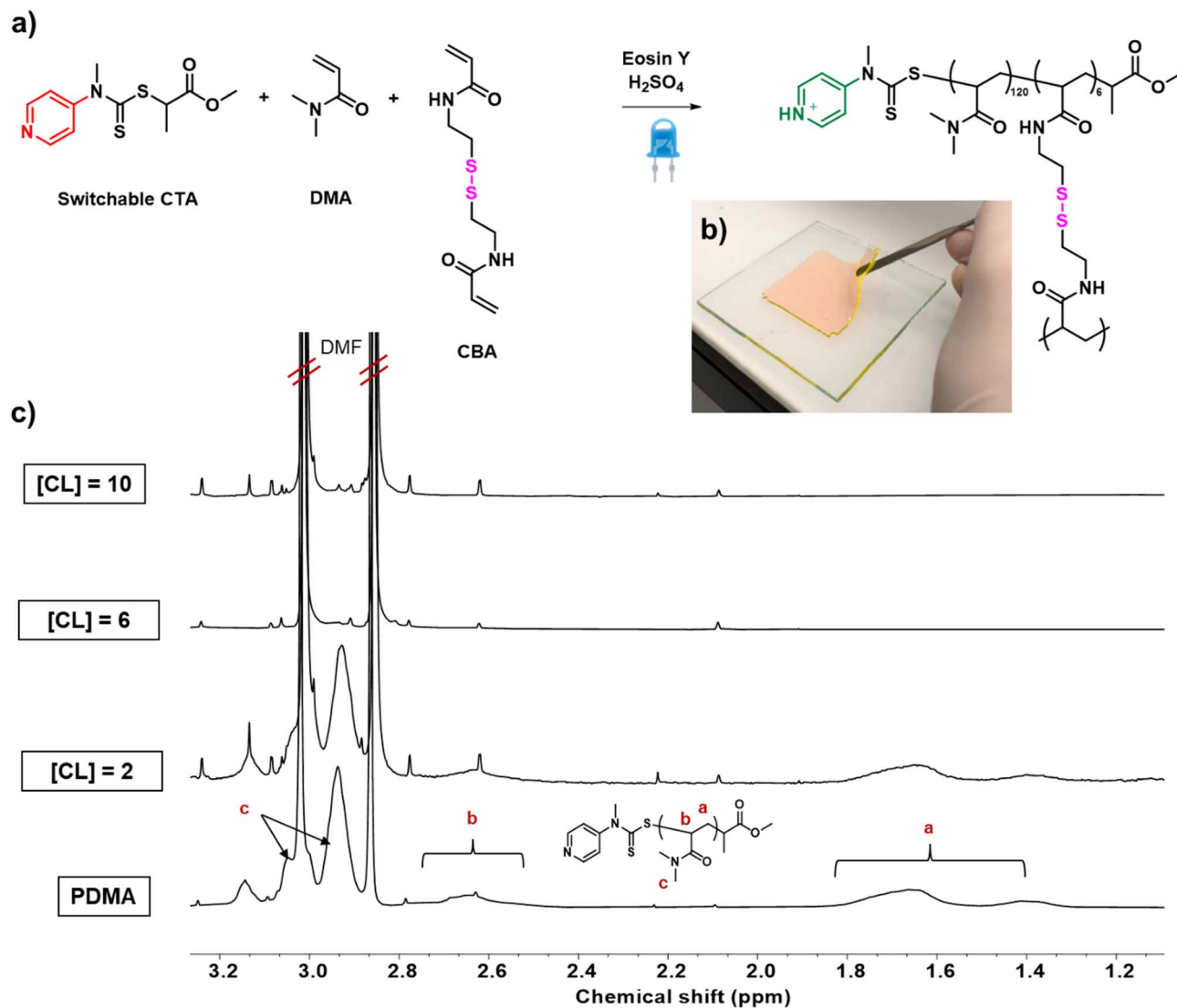


Fig. 2 The preparation of networks with various primary chain dispersities. (a) is a scheme of the polymer gelation reaction, (b) is a photo of the obtained gel and (c) shows ¹H NMR analysis of the extractions obtained from syntheses with various equivalents of crosslinker (CL).

acid-switchable CTA in tuning the dispersity within network polymers (Fig. 3b). These reactions were simple to perform, carried out in one-pot and required just a change in the acid concentration to obtain the various materials. There was a deviation between the theoretical and experimentally obtained molecular weights after cleavage of the networks and a slightly higher dispersity value was observed for the lowest dispersity material, when compared to the analogous linear polymer synthesis (e.g. 18 400, $\bar{D} = 1.28$ vs. 12 500 Da, $\bar{D} = 1.20$, Table 1 and Fig. S11†). We attribute this difference to the closer proximity of the propagating polymer chains within the network, which could result in a higher probability of termination through intramolecular combination reactions and also the reduced efficiency of deactivation by chain transfer due to the highly viscous nature of the synthesis.^{59,74} This is evidenced by a small amount of high molecular weight shouldering in the molar mass distributions of the various cleaved polymer

networks (Fig. 3b) and demonstrates that analogous linear and network polymer syntheses do not yield identical materials. To ensure our gels were comparable, we analysed their equilibrium water contents, by immersing each of the networks in excess water for 24 hours. They were then weighed before and after freeze-drying. For all of our various primary chain dispersity networks, the solvent adsorption capacity was comparable (87–89%), indicating that each material had comparable crosslink densities and compositions (Table 1, column 3). The swelling ratios of the gels were also obtained to determine the uniformity of the crosslinked network materials. As shown in Table 1 column 4, the networks have excellent swelling ratios (6.8–8.4) which suggests good network homogeneity, even though a slight increase in swelling ratio with decreasing primary chain dispersity was observed.

We, therefore, proceeded to measure the degradability of our various primary chain dispersity networks, by performing

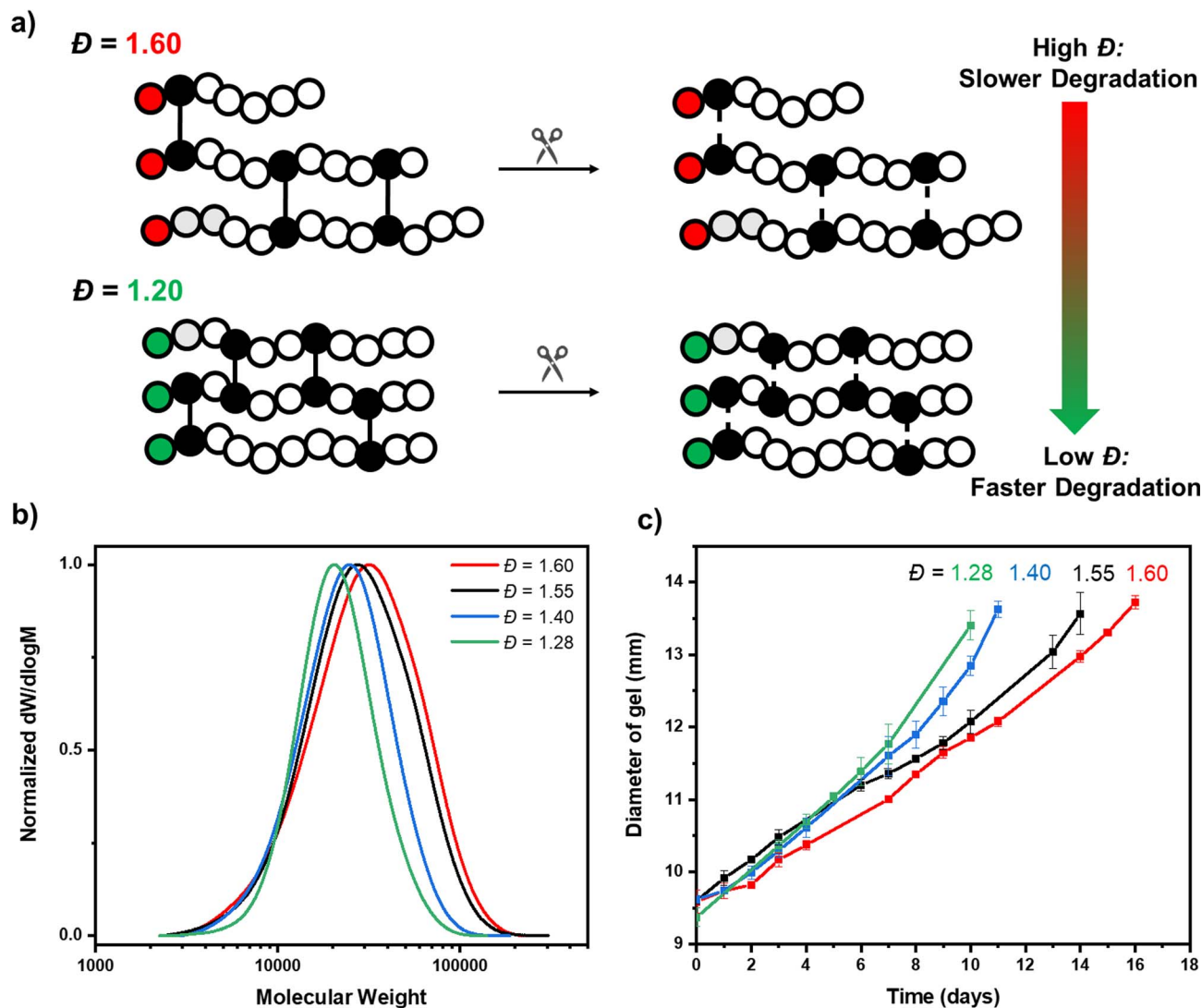


Fig. 3 The degradation of various primary chain dispersity networks. (a) is a scheme illustrating the degradation of the crosslinker, (b) shows SEC analysis of linear polymers obtained after cleavage of the various materials with DTT, and (c) shows the kinetics of degradation of the various dispersity networks with glutathione.

Table 1 Experimental data obtained from the degradation of networks prepared with various amounts of acid. The acid concentration from the synthesis corresponds to the primary chain dispersity of the resulting polymers

Entry	[H ₂ SO ₄]	EW ^a (%)	SR ^b	<i>M_n</i> (SEC)	<i>M_w</i> (SEC)	<i>M_p</i> (SEC)	$\bar{D}^c$
1	0	87.2	6.8	22 900	36 700	31 700	1.60
2	0.25	86.9	6.6	21 800	33 800	27 100	1.55
3	0.5	89.4	8.4	19 200	26 700	24 600	1.40
4	0.3	88.5	7.7	18 400	23 600	20 600	1.28

^a Equilibrium water content (EW). ^b Swelling Ratio (SR) were calculated based on the ratio of swollen and dried gel weight.

^c Cleavage was performed with DTT in dimethylacetamide.

experiments in glutathione solution at room temperature.^{75,76} We first took 9.5 mm discs of our highest primary chain dispersity ($\bar{D} = 1.60$) network and performed degradation

experiments in triplicate. Over time, degradation of the crosslinker occurred, as evidenced by a gradual increase in the diameter of the discs. The average size was 11 mm after 7 days, 12 mm after 12 days and 13 mm after 15 days (Table S5† and Fig. 3c). This swelling was due to the random degradation of crosslinkers increasing the distance between the remaining intact crosslinkers. This resulted in the network structure becoming less rigid and consequently it swells better. By day 17, a sufficient amount of crosslinker had been degraded for the network to reach its gel-sol transition point where it fully dissolved. The degradation experiment was then repeated with the various primary chain dispersity networks that we had previously prepared ($\bar{D} = 1.55, 1.40, 1.28$). Notably, a distinct trend in degradation rate was observed depending on the initial primary chain dispersity. It was observed that the lower the dispersity of the network, the faster the rate of diameter increase and the earlier the time point of full dissolution. The network with

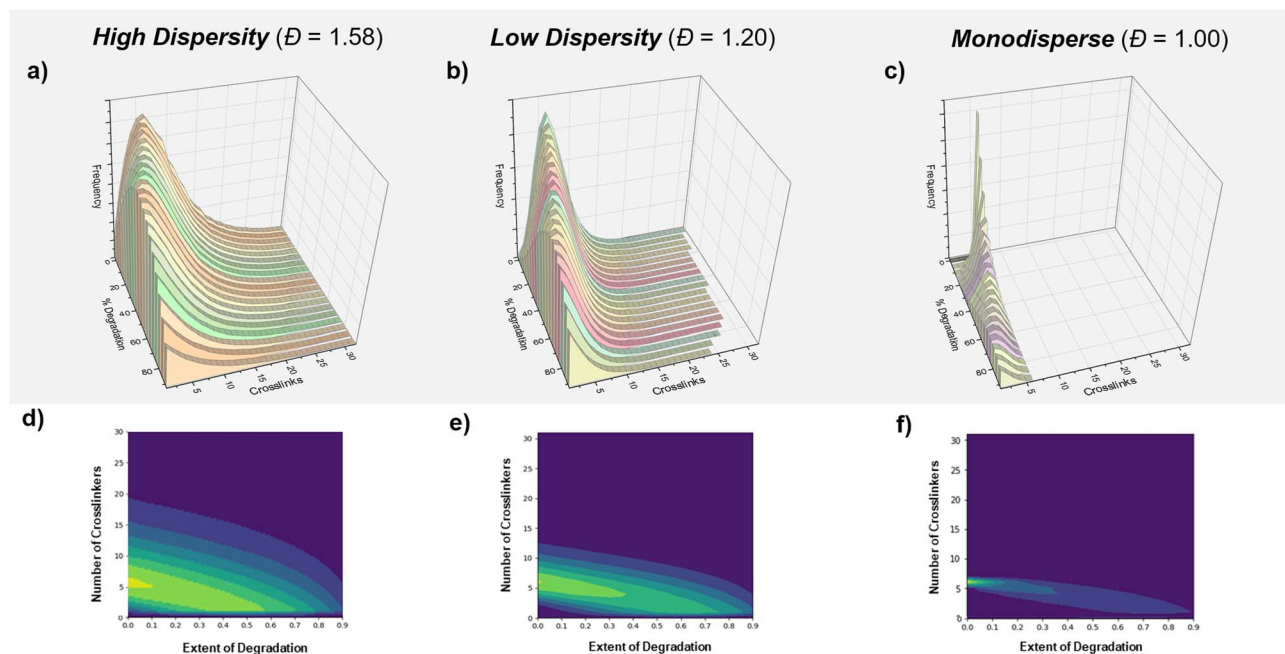
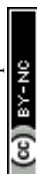


Fig. 4 Modelling of the effect of primary chain dispersity on degradation. The number of crosslinks per chain is modelled for a network of (a) high, (b) low and (c) mono primary chain dispersity. In (d–f) corresponding heat maps are shown to correlate the number of crosslinkers at various extents of degradation.

primary chain dispersity of 1.55 fully dissolved on day 15, whereas the 1.40 and 1.28 dispersity networks required only 12 and 11 days, respectively (Tables S6–S8†). This illustrates that by carefully tuning the primary chain dispersity, the rate of degradation was directly affected. In terms of degradation time, there was a significant difference between our highest and lowest dispersity values of almost 40% (17 vs. 11 days, Fig. 3c). All experiments were performed in triplicate and the degradation was shown to be accurately reproducible with error bars illustrated in Fig. 3c. It should be highlighted that there were some small deviations in the molecular weight of the chains within our various primary chain dispersity networks ($M_n = 18\,500$ – $22\,900$ Da), so additional control experiments were performed. A range of different primary molecular weight networks were synthesized, while keeping the dispersity constant (either $D = 1.28$ or $D = 1.40$, Tables S9, S10 and Fig. S8†) and on degradation with glutathione, no molecular weight trend was observed (Tables S11–S15 and Fig. S12–S13†). Overall, it was concluded that primary chain dispersity significantly impacts degradation and this is a parameter that should be considered when designing degradable polymeric networks. In addition, our method provides a facile route to obtain such dispersity-controlled materials, creating many opportunities in applications where controlled degradation is required.

Finally, we were interested in further investigating and understanding the origin of this dispersity effect. We hypothesized that there could be two possible causes; (i) a positional effect based on the distribution of the crosslinkers within our various primary chain dispersity materials and/or (ii) an effect from different rates of glutathione diffusion. Simulations were therefore performed to understand the effect of the crosslinker

distribution by converting the molar mass distributions of the highest and lowest primary chain dispersity materials to distributions of crosslinker (Fig. 4a–c and S14†). It was observed from this data that even though both materials had the same average number of crosslinkers per chain, the high dispersity sample had a significantly larger maximum number of crosslinkers per chain (60 vs. 30) and also many more chains with just 1 or 2 crosslinkers. The degradation of these materials ($\sim 1\,000\,000$ polymer chains in total) was next simulated using a simple Python program, where crosslinkers were randomly selected and broken one by one, before a snapshot of the distribution was taken at increments of 5% degradation (or 50 000 bonds, Fig. 4d and e). Throughout the simulation, there was a consistent theme with the high dispersity network containing more chains with larger numbers of crosslinkers, but also larger numbers of chains that had fully dissociated from the material. For example, at 80% degradation, the high dispersity material contained $\sim 80\,000$ chains that had six or more crosslinks (31%) and $\sim 30\,000$ chains (12%) with ten or more crosslinkers compared to $\sim 55\,000$ (14%) with more than 6 crosslinkers and ~ 8000 chains (2%) with more than 10 crosslinkers in the low dispersity sample. In addition, when evaluating how many chains no longer had a single crosslink and had therefore dissociated from the network, $\sim 75\%$ percent of the original chains had fully dissociated from the high primary chain dispersity network compared to $\sim 63\%$ for the low dispersity material (Tables S16 and S17†). Coupled with the experimental data, we propose that the higher molecular weight chains are essential in extending the degradation time of the network. For example, when comparing chains with 6 and 60 crosslinkers, those with 60 crosslinkers required a much higher percentage



supervision, methodology, visualization, writing – original draft. G. R. J. investigation: simulations, writing – reviewing and editing. I. R. investigation: ANOVA and swelling ratio calculations, conceptualization, writing – reviewing and editing. D. K. conceptualization, writing – reviewing and editing. N. P. T. conceptualization, writing – reviewing and editing. A. A. conceptualization, project administration, supervision, funding acquisition, writing – reviewing and editing.

The authors declare no conflicts of interest.

A. A. gratefully acknowledges ETH Zurich (Switzerland) and the European Research Council (ERC) under the European Union's Horizon 2020 research and innovation programme (DEPO: Grant no. 949219) for financial support. T. S. acknowledges Mitsubishi Chemical Corporation for generous support. N. P. T. acknowledges the award of a DECRA Fellowship from the ARC (DE180100076).

We have established a one-pot method to tune the primary chain dispersity in polymer networks and demonstrated a significant effect of primary chain dispersity on gel degradation. PET-RAFT polymerization was performed with a degradable crosslinker, a switchable CTA and various amounts of acid, which allowed the dispersity to be precisely tuned, while incorporating all monomer, crosslinker and polymer chains within the materials. On crosslinker degradation, the precise primary chain dispersity values were obtained and correlated to the degradation profiles. The key observation was that the higher the primary chain dispersity of the network, the slower the degradation. This was supported by simulations, which emphasized the importance of the higher molecular weight chains in the higher dispersity materials, in preserving the network until a higher extent of degradation had been reached. This work creates many new research opportunities in applications where precise tuning of network degradation is necessary.

- 1 D. T. Gentekos, R. J. Sifri and B. P. Fors, *Nat. Rev. Mater.*, 2019, 1–14.
- 2 R. Whitfield, N. P. Truong, D. Messmer, K. Parkatzidis, M. Rolland and A. Anastasaki, *Chem. Sci.*, 2019, **10**, 8724–8734.
- 3 T. Junkers, *Macromol. Chem. Phys.*, 2020, **221**, 2000234.
- 4 M. R. Martinez and K. Matyjaszewski, *CCS Chem.*, 2022, **4**, 2176–2211.
- 5 R. Whitfield, N. Truong and A. Anastasaki, *Angew. Chem., Int. Ed.*, 2021, **60**, 19383–19388.
- 6 M. Nardi, E. Blasco and C. Barner-Kowollik, *J. Am. Chem. Soc.*, 2022, **144**, 1094–1098.
- 7 M. Chen, J. Li, K. Ma, G. Jin, X. Pan, Z. Zhang and J. Zhu, *Angew. Chem.*, 2021, **133**, 19857–19861.
- 8 H. S. Wang, K. Parkatzidis, S. Harrisson, N. P. Truong and A. Anastasaki, *Chem. Sci.*, 2021, **12**, 14376–14382.
- 9 K. Parkatzidis, M. Rolland, N. T. Phuoc and A. Anastasaki, *Polym. Chem.*, 2021, **12**, 5583–5588.
- 10 T. Takamatsu, S. Shioya and Y. Okada, *Ind. Eng. Chem. Res.*, 1988, **27**, 93–99.
- 11 V. Kottisch, D. T. Gentekos and B. P. Fors, *ACS Macro Lett.*, 2016, **5**, 796–800.
- 12 D. T. Gentekos, L. N. Dupuis and B. P. Fors, *J. Am. Chem. Soc.*, 2016, **138**, 1848–1851.
- 13 M. Nadgorny, D. T. Gentekos, Z. Xiao, S. P. Singleton, B. P. Fors and L. A. Connal, *Macromol. Rapid Commun.*, 2017, **38**, 1700352.
- 14 R. J. Sifri, O. Padilla-Vélez, G. W. Coates and B. P. Fors, *J. Am. Chem. Soc.*, 2019, **142**, 1443–1448.
- 15 S. Domanskyi, D. T. Gentekos, V. Privman and B. P. Fors, *Polym. Chem.*, 2020, **11**, 326–336.

Additional experimental details and data are provided in the ESI.† This includes SEC, UV-vis and ^1H NMR data, alongside photographs of the experimental set-up and raw data from the simulations.

T. S. methodology, investigation: polymer synthesis and degradation, validation, visualization, writing – reviewing and editing. R. W. conceptualization, project administration,

- 16 S. Rosenbloom, R. Sifri and B. Fors, *Polym. Chem.*, 2021, **12**, 4910–4915.
- 17 N. Corrigan, A. Almasri, W. Taillades, J. Xu and C. Boyer, *Macromolecules*, 2017, **50**, 8438–8448.
- 18 N. Corrigan, R. Manahan, Z. T. Lew, J. Yeow, J. Xu and C. Boyer, *Macromolecules*, 2018, **51**, 4553–4563.
- 19 K. Liu, N. Corrigan, A. Postma, G. Moad and C. Boyer, *Macromolecules*, 2020, **53**, 8867–8882.
- 20 S. Xu, F. J. Trujillo, J. Xu, C. Boyer and N. Corrigan, *Macromol. Rapid Commun.*, 2021, **42**, 2100212.
- 21 M. Rubens and T. Junkers, *Polym. Chem.*, 2019, **10**, 6315–6323.
- 22 M. Rubens and T. Junkers, *Polym. Chem.*, 2019, **10**, 5721–5725.
- 23 T. Junkers and J. H. Vrijsen, *Eur. Polym. J.*, 2020, **134**, 109834.
- 24 N. Corrigan and C. Boyer, *Macromolecules*, 2022, **55**, 8960–8969.
- 25 M. H. Reis, T. P. Varner and F. A. Leibfarth, *Macromolecules*, 2019, **52**, 3551–3557.
- 26 D. J. Walsh, D. A. Schinski, R. A. Schneider and D. Guironnet, *Nat. Commun.*, 2020, **11**, 1–9.
- 27 D. Liu, A. D. Sponza, D. Yang and M. Chiu, *Angew. Chem.*, 2019, **58**, 16210–16216.
- 28 R. Whitfield, K. Parkatzidis, M. Rolland, N. P. Truong and A. Anastasaki, *Angew. Chem., Int. Ed.*, 2019, **58**, 13323–13328.
- 29 Z. Wang, J. Yan, T. Liu, Q. Wei, S. Li, M. Olszewski, J. Wu, J. Sobieski, M. Fantin and M. R. Bockstaller, *ACS Macro Lett.*, 2019, **8**, 859–864.
- 30 T. Shimizu, N. P. Truong, R. Whitfield and A. Anastasaki, *ACS Polym. Au*, 2021, **1**, 187–195.
- 31 M. Rolland, N. P. Truong, R. Whitfield and A. Anastasaki, *ACS Macro Lett.*, 2020, **9**, 459–463.
- 32 M. Rolland, V. Lohmann, R. Whitfield, N. P. Truong and A. Anastasaki, *J. Polym. Sci.*, 2021, **59**, 2502–2509.
- 33 X. Liu, C. G. Wang and A. Goto, *Angew. Chem.*, 2019, **131**, 5654–5659.
- 34 R. Whitfield, K. Parkatzidis, N. P. Truong, T. Junkers and A. Anastasaki, *Chem*, 2020, **6**, 1340–1352.
- 35 K. Parkatzidis, N. P. Truong, M. N. Antonopoulou, R. Whitfield, D. Konkolewicz and A. Anastasaki, *Polym. Chem.*, 2020, **11**, 4968–4972.
- 36 T. Nwoko, N. D. A. Watuthanthrige, B. Parnitzke, K. Yehl and D. Konkolewicz, *Polym. Chem.*, 2021, **12**, 6761–6770.
- 37 M.-N. Antonopoulou, R. Whitfield, N. P. Truong, D. Wyers, S. Harisson, T. Junkers and A. Anastasaki, *Nat. Chem.*, 2022, **14**, 304–312.
- 38 M.-N. Antonopoulou, R. Whitfield, N. P. Truong and A. Anastasaki, *Eur. Polym. J.*, 2022, **174**, 111326.
- 39 S. I. Rosenbloom, J. H. Hsu and B. P. Fors, *J. Polym. Sci.*, 2022, **60**, 1291–1299.
- 40 O. Terreau, L. Luo and A. Eisenberg, *Langmuir*, 2003, **19**, 5601–5607.
- 41 S. I. Rosenbloom and B. P. Fors, *Macromolecules*, 2020, **53**, 7479–7486.
- 42 S. I. Rosenbloom, D. T. Gentekos, M. N. Silberstein and B. P. Fors, *Chem. Sci.*, 2020, **11**, 1361–1367.
- 43 D. T. Gentekos, J. Jia, E. S. Tirado, K. P. Barteau, D.-M. Smilgies, R. A. DiStasio Jr and B. P. Fors, *J. Am. Chem. Soc.*, 2018, **140**, 4639–4648.
- 44 A.-L. Buckinx, M. Rubens, N. R. Cameron, C. Bakkali-Hassani, A. Sokolova and T. Junkers, *Polym. Chem.*, 2022, **13**, 3444–3450.
- 45 Z. Wang, J. Yan, T. Liu, Q. Wei, S. Li, M. Olszewski, J. Wu, J. Sobieski, M. Fantin, M. R. Bockstaller and K. Matyjaszewski, *ACS Macro Lett.*, 2019, **8**, 859–864.
- 46 R. Yin, Z. Wang, M. R. Bockstaller and K. Matyjaszewski, *Polym. Chem.*, 2021, **12**, 6071–6082.
- 47 M. Romio, B. Grob, L. Trachsel, A. Mattarei, G. Morgese, S. N. Ramakrishna, F. Niccolai, E. Guazzelli, C. Paradisi and E. Martinelli, *J. Am. Chem. Soc.*, 2021, **143**, 19067–19077.
- 48 V. Yadav, N. Hashmi, W. Ding, T.-H. Li, M. K. Mahanthappa, J. C. Conrad and M. L. Robertson, *Polym. Chem.*, 2018, **9**, 4332–4342.
- 49 T.-H. Li, V. Yadav, J. C. Conrad and M. L. Robertson, *ACS Macro Lett.*, 2021, **10**, 518–524.
- 50 C.-G. Wang, A. M. L. Chong and A. Goto, *ACS Macro Lett.*, 2021, **10**, 584–590.
- 51 J. C. Conrad and M. L. Robertson, *JACS Au*, 2023, **3**, 333–343.
- 52 F. M. Benedetti, Y.-C. M. Wu, S. Lin, Y. He, E. Flear, K. R. Storme, C. Liu, Y. Zhao, T. M. Swager and Z. P. Smith, *JACS Au*, 2022, **2**, 1610–1615.
- 53 D. Merckle and A. C. Weems, *Polym. Chem.*, 2023, **14**, 3587–3599.
- 54 N. D. A. Watuthanthrige, P. Chakma and D. Konkolewicz, *Trends Chem.*, 2021, **3**, 231–247.
- 55 I. O. Raji, O. J. Dodo, N. K. Saha, M. Eisenhart, K. M. Miller, R. Whitfield, A. Anastasaki and D. Konkolewicz, *ChemRxiv*, 2023, preprint, DOI: [10.26434/chemrxiv-2023-6ngm1](https://doi.org/10.26434/chemrxiv-2023-6ngm1).
- 56 J. J. Lessard, K. A. Stewart and B. S. Sumerlin, *Macromolecules*, 2022, **55**, 10052–10061.
- 57 C. W. A. Bainbridge, A. Wangsadijaya, N. Broderick and J. Jin, *Polym. Chem.*, 2022, **13**, 1484–1494.
- 58 J. Cuthbert, S. V. Wanasinghe, K. Matyjaszewski and D. Konkolewicz, *Macromolecules*, 2021, **54**, 8331–8340.
- 59 F. Dawson, H. Jafari, V. Rimkevicius and M. Kopeć, *Macromolecules*, 2023, **56**, 2009–2016.
- 60 W. Li, H. Gao and K. Matyjaszewski, *Macromolecules*, 2009, **42**, 927–932.
- 61 A. D. Metcalfe and M. W. Ferguson, *J. R. Soc., Interface*, 2007, **4**, 413–437.
- 62 K. A. Davis and K. S. Anseth, *Crit. Rev. Ther. Drug Carrier Syst.*, 2002, **19**, 385–423.
- 63 K. S. Anseth, A. T. Metters, S. J. Bryant, P. J. Martens, J. H. Elisseeff and C. N. Bowman, *J. Controlled Release*, 2002, **78**, 199–209.
- 64 M. Grosjean, L. Gangolphe and B. Nottelet, *Adv. Funct. Mater.*, 2023, **33**, 2205315.
- 65 G. X. Wang, D. Huang, J. H. Ji, C. Völker and F. R. Wurm, *Advanced Science*, 2021, **8**, 2001121.
- 66 J. W. DuBose, C. Cutshall and A. T. Metters, *J. Biomed. Mater. Res., Part A*, 2005, **74**, 104–116.



- 67 N. De Alwis Watuthanthrige, D. Dunn, M. Dolan, J. L. Sparks, Z. Ye, M. B. Zanjani and D. Konkolewicz, *ACS Appl. Polym. Mater.*, 2022, **4**, 1475–1486.
- 68 S. V. Wanasinghe, M. Sun, K. Yehl, J. Cuthbert, K. Matyjaszewski and D. Konkolewicz, *ACS Macro Lett.*, 2022, **11**, 1156–1161.
- 69 G. Opiyo and J. Jin, *Eur. Polym. J.*, 2021, **159**, 110713.
- 70 J. Xu, S. Shanmugam, H. T. Duong and C. Boyer, *Polym. Chem.*, 2015, **6**, 5615–5624.
- 71 Y. Xiu, V. A. Bobrin, N. Corrigan, Y. Yao, J. Zhang and C. Boyer, *J. Polym. Sci.*, 2023, DOI: [10.1002/pol.20230634](https://doi.org/10.1002/pol.20230634).
- 72 A. Bagheri, C. W. A. Bainbridge, K. E. Engel, G. G. Qiao, J. Xu, C. Boyer and J. Jin, *ACS Appl. Polym. Mater.*, 2020, **2**, 782–790.
- 73 W. Konigsberg, in *Methods in enzymology*, Elsevier, 1972, vol. 25, pp. 185–188.
- 74 K. Matyjaszewski and T. P. Davis, *Handbook of radical polymerization*, John Wiley & Sons, Hoboken, 2002.
- 75 D. J. Phillips and M. I. Gibson, *Biomacromolecules*, 2012, **13**, 3200–3208.
- 76 J. F. Quinn, M. R. Whittaker and T. P. Davis, *Polym. Chem.*, 2017, **8**, 97–126.
- 77 P. J. Flory, *J. Am. Chem. Soc.*, 1947, **69**, 30–35.
- 78 W. H. Stockmayer, *J. Chem. Phys.*, 1943, **11**, 45–55.
- 79 W. H. Stockmayer, *J. Chem. Phys.*, 1944, **12**, 125–131.

