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Emerging per- and polyfluoroalkyl substances (PFAS) in human milk from Sweden and China †

Raed Awad,^a Yihui Zhou,^c Elisabeth Nyberg,^d Shahla Namazkar,^a Wu Yongning,^e Qianfen Xiao,^c Yaijie Sun,^f Zhiliang Zhu,^f Åke Bergman^{a,c,g} and Jonathan P. Benskin^a

Twenty per- and polyfluoroalkyl substances (PFAS) were determined in human milk from residents of three Chinese cities (Shanghai, Jiaxing, and Shaoxing; [$n = 10$ individuals per city]), sampled between 2010 and 2016. These data were compared to a combination of new and previously reported PFAS concentrations in human milk from Stockholm, Sweden, collected in 2016 ($n = 10$ individuals). Across the three Chinese cities, perfluorooctanoate (PFOA; sum isomers), 9-chlorohexadecafluoro-3-oxanone-1-sulfonic acid (9Cl-PF3ONS; also known as 6:2 Cl-PFESA or by its trade name "F53-B"), and perfluorooctane sulfonate (PFOS; sum isomers) occurred at the highest concentrations among all PFAS (up to 411, 976, and 321 pg mL^{-1} , respectively), while in Stockholm, PFOA and PFOS were dominant (up to 89 and 72 pg mL^{-1} , respectively). 3H-Perfluoro-3-[(3-methoxy-propoxy)propanoic acid] (ADONA) was intermittently detected but at concentrations below the method quantification limit (i.e. $<10 \text{ pg mL}^{-1}$) in Chinese samples, and was non-detectable in Swedish milk. The extremely high concentrations of F53-B in Chinese milk suggest that human exposure assessments focused only on legacy substances may severely underestimate overall PFAS exposure in breastfeeding infants.

Introduction

Per- and polyfluoroalkyl substances (PFAS) are a large and diverse group of anthropogenic chemicals that have been

Environmental significance

Human milk is a significant source of exposure to per- and polyfluoroalkyl substances (PFAS) in breastfed infants. While many legacy PFAS are no longer produced, there are a growing number of alternative PFAS which have been introduced as replacements. To investigate the potential for infant exposure to these chemicals, we measured legacy and emerging PFAS in human milk from three cities in China (Shanghai, Jiaxing, and Shaoxing) and one city in Sweden (Stockholm). The legacy PFAS perfluorooctanoate (PFOA), perfluorooctane sulfonate (PFOS), along with the PFOS-replacement, 9-chlorohexadecafluoro-3-oxanone-1-sulfonic acid (trade name 'F53-B') occurred at the highest concentrations among all PFAS, with the latter substance only detected in Chinese cities. This work suggests that human exposure assessments focused only on legacy substances may severely underestimate overall exposure to PFAS.

manufactured since the 1940s.¹ Approximately 5000 PFAS-related CAS numbers are known to exist globally.² Many PFAS contain perfluoroalkyl chains which impart unique properties, including extreme stability and combined oil/water repellency. These characteristics have made PFAS attractive for use in many industrial processes and consumer products. Unfortunately, due to their widespread use and recalcitrant nature, PFAS now occur in the blood of humans globally.³

For the most widely studied PFAS (i.e. the legacy PFAS perfluorooctane sulfonate (PFOS) and perfluorooctanoate (PFOA)), the main exposure routes for non-occupationally-exposed adults include food,^{4–7} drinking water,⁸ air⁹ and house dust.^{9,10} In infants, exposure to legacy PFAS occurs primarily from placental transfer while in the womb^{11–14} and breast milk thereafter.^{15–17} Even in children up to 3–5 years of age, PFAAs obtained from breast milk as an infant represent a significant fraction of their overall PFAS burden.^{18,19} This is clearly a concern, given the links between adverse health effects and PFAS exposure in children.^{20–22}

Recent investigations of human milk from Stockholm, Sweden, revealed highly contrasting PFAS profiles for the time periods 1972–1996, 2000–2012, and 2013–2016, reflecting shifting exposure over time.^{17,23} Of particular note was the

^aDepartment of Environmental Science (ACES), Stockholm University, 106 91 Stockholm, Sweden. E-mail: Raed.Awad@ivl.se; Jon.Benskin@aces.su.se

^bSwedish Environmental Research Institute (IVL), 114 28 Stockholm, Sweden

^cState Key Laboratory of Pollution Control and Resource Reuse, College of Environmental Science and Engineering, Tongji University, Shanghai 200092, China

^dDepartment of Contaminants, Swedish Environmental Protection Agency, Virkesvägen 2, SE-106 48 Stockholm, Sweden

^eNHC Key Laboratory of Food Safety Risk Assessment, China National Center for Food Safety Risk Assessment, Beijing, 100021, China

^fKey Laboratory of Yangtze River Water Environment (Ministry of Education), College of Environmental Science and Engineering, Tongji University, Shanghai 200092, China

^gDepartment of Science and Technology, Örebro University, 701 82 Örebro, Sweden

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steadily increase in concentrations of certain long-chain perfluoroalkyl carboxylic acids (PFCAs) and decline in concentrations of PFOS, PFOA, and perfluorooctane sulfonamide (FOSA). The latter result likely reflects regulation and phase-out initiatives while the former may indicate ongoing use of long-chain PFCAs and/or their precursors. Similar trends have been reported in human serum from countries around the world,^{24–27} including Sweden.²⁸ However, in China, where PFOS production has continued under Stockholm Convention manufacture and use exemptions, a decline in the concentrations of PFOS in human blood has not been observed.²⁹

Relatively little is known about human exposure to emerging PFAS ‘alternatives’ which have been introduced as replacements to substances such as PFOS and PFOA. The few data which exist suggest that these substances may already occur throughout the environment. For example, 2,3,3,3-tetrafluoro-2-(1,1,2,2,3,3,3-heptafluoropropoxy)propanoic acid (HFPO-DA; trade name ‘Gen-X’) was introduced in 2010 as a PFOA alternative and has been reported in surface water globally.^{30–33} Similarly, the replacement emulsifier 3H-perfluoro-3-[(3-methoxy-propoxy)propanoic acid] (ADONA), which has been used as a PFOA replacement since 2008 has been reported in both surface water and human serum from Germany.^{33,34} In China, the chlorinated polyfluorinated ether sulfonates (Cl-PFESAs) 9-chlorohexadecafluoro-3-oxanone-1-sulfonic acid (9Cl-PF3ONS; also known as 6:2 Cl-PFESA or the trade name ‘F-53B’) and 11-chloroeicosafluoro-3-oxaundecane-1-sulfonic acid (11Cl-PF3OUDS; also known as 8:2 Cl-PFESA), are also an emerging concern. F-53B is a PFOS alternative, and has been used for over 30 years as a commercial mist suppressant by the Chinese electroplating industry.³⁵ In addition to its widespread occurrence in surface water from China,³³ Cl-PFESAs were observed in >98% of metal plating workers and high fish consumers sampled from the provinces of Shandong and Hubei at concentrations up to 5040 ng mL^{−1}.³⁶ Most recently, Cl-PFESAs have been detected in marine mammal livers from Greenland^{37,38} and Sweden,³⁸ polar bear serum from Hudson Bay and the Beaufort Sea,³⁹ and human milk from Hangzhou, China.⁴⁰

To shed further light on PFAS exposure in breastfed newborns, in particular to emerging PFAS, the current study investigated 20 PFAS (including the alternatives F53-B and ADONA) in human milk from residents of three Chinese cities (Shanghai, Jiaying, and Shaoxing), sampled between 2010 and 2016. These data were compared to a combination of new and previously reported PFAS data from human milk from Stockholm, Sweden, collected in 2016. These data provide further insight on the variability in PFAS concentrations and profiles between cities and countries.

Experimental

Sampling

Samples from Stockholm ($n = 10$) were purchased in 2016 from the Mothers’ Milk Centre at Stockholm South General Hospital. Milk was donated anonymously from healthy native Swedish mothers, which were non-smokers and predominantly primiparous. Sampling was carried out between 2 weeks and 3 months

after delivery. Data from these samples were previously reported in Nyberg *et al.*,¹⁷ but were re-analyzed in the present work for F53-B and ADONA. Further information is provided in Table S1 of the ESI†. Samples from Jiaying ($n = 10$) and Shanghai ($n = 10$) were collected from 2015–2016, while samples from Shaoxing ($n = 10$) were collected in 2010. All participants were informed of the objectives of the study and signed a participant information and consent form. The study was approved by, and performed in accordance with guidelines from the Ethics Committee of China National Center for Food Safety Risk Assessment. Samples were initially stored at $-18\text{ }^{\circ}\text{C}$ in polyethylene sampling bags and bottles. Upon being shipped to the Environmental Specimen Bank (ESB) at the Swedish Museum of Natural History in Stockholm Sweden and the Yangtze Environmental Specimen Bank (YESB) at the Tongji Jiaying Research Institution in Jiaying China, samples were thawed, transferred to pre-washed glass bottles with lids covered with aluminum foil, and stored at $-20\text{ }^{\circ}\text{C}$ prior to analysis.

Standards and reagents

Linear isomer standards of perfluoroheptanoate (PFHpA), PFOA, perfluorononanoate (PFNA), perfluorodecanoate (PFDA), perfluorundecanoate (PFUnDA), perfluorododecanoate (PFDoDA), perfluorotridecanoate (PFTTrDA), perfluorotetradecanoate (PFTTeDA), 3-perfluoroheptyl propanoic acid (7:3 FTCA), perfluorobutane sulfonate (PFBS), perfluorohexane sulfonate (PFHxS), PFOS, perfluorodecane sulfonate (PFDS), perfluorooctanesulfonamide (FOSA), *N*-methyl perfluorooctane sulfonamidoacetic acid (MeFOSAA), *N*-ethyl perfluorooctane sulfonamidoacetic acid (EtFOSAA), potassium 9-chlorohexadecafluoro-3-oxanonane-1-sulfonate (9Cl-PF3ONS), potassium 11-chloroeicosafluoro-3-oxaundecane-1-sulfonate (11Cl-PF3OUDS), and sodium dodecafluoro-3H-4,8-dioxanonanoate (ADONA) were obtained from Wellington Labs (Guelph, ON, Canada). Perfluoropentanoate (PFPeA) and perfluorohexanoate (PFHxA) were investigated as part of Nyberg *et al.*¹⁷ (*i.e.* Stockholm samples from 2016), but since they were not measured in the Chinese samples, they were left out of the present analysis to ensure comparability of sum PFAS concentrations between Chinese and Swedish samples. All isotopically labeled standards were obtained from Wellington Labs (a full list is provided in Table S2, ESI†). Formic acid, ammonium sulfate, and ammonium acetate were obtained from Merck. Potassium hydroxide was purchased from VWR, methyl *tert*-butyl ether (HPLC grade) was purchased from RATHBURN (Walkerburn, Scotland) and acetonitrile was obtained from Honeywell. Standard Reference Material (SRM) 1954: Organic Contaminants in Fortified Human Milk, was obtained from the National Institute of Standards and Technology (NIST).

Chemical analysis

Sample extraction was carried out using a previously published method.^{17,23} Briefly, 2 mL of each sample was acidified with formic acid and then spiked with isotopically labeled internal standards. After vortexing, the sample was fortified with 600 μL of saturated ammonium sulfate, vortexed again, then fortified



with 7 mL acetonitrile, and finally placed on a mechanical shaker. After 30 min of shaking, the samples were centrifuged and the top organic layer was transferred into a polypropylene tube and evaporated at 40 °C. The extract was then diluted with 300 µL of water and vortexed before adding 500 µL of 1 M potassium hydroxide. The tube was vortexed and then supplemented with 7 mL of methyl *tert*-butyl ether, then placed in a mechanical shaker for 20 minutes followed by centrifugation. The organic layer was subsequently transferred into a clean polypropylene tube. The extracts were evaporated at 40 °C and then fortified with 200 µL of buffer (1 : 2 : 1 ammonium acetate : acetonitrile : water). After vortexing and centrifugation, the lower layer was transferred to an Eppendorf tube and recovery standard was added. After vortexing, the mixture was transferred to microvial for analysis by UPLC-MS/MS.

Instrumental analysis was carried out on a Waters ultra-performance liquid chromatograph (UPLC) coupled to a Waters Xevo TQ-S triple quadrupole mass spectrometer operated in negative electrospray ionization, selected reaction monitoring mode. Instrumental parameters are provided in Table S2 (ESI†). Extracts were chromatographed on a BEH C18 analytical column (2.1 × 50 mm, 1.7 µm particle size, Waters) operated at a flow rate of 0.4 mL min⁻¹, using a mobile phase composition of 90% water/10% acetonitrile containing 2 mM ammonium acetate (solvent A) and 100% acetonitrile containing 2 mM ammonium acetate (solvent B). The gradient profile is provided in Table S3 (ESI†). A total of two precursor/product ion transitions were monitored per analyte; one for quantification and the other for qualification (Table S2, ESI†).

Quantitative determination of target compounds was carried out by isotope dilution or an internal standard approach using a linear calibration curve with 1/*x* weighting. Branched isomers were determined semi-quantitatively using the calibration curve for the linear isomer. In almost all cases, the most sensitive ion was used for quantification. However, for PFOA, we observed an interference in the *m/z* 413/369 ion; therefore, the *m/z* 413/169 ion was used for quantification. For PFOS, concentrations are reported as the average of *m/z* 499/80 and 499/99 ions for both branched and linear isomers, as recommended by Riddell *et al.*⁴¹ Sum (Σ) PFOS concentrations were obtained by summing the concentrations of branched and linear PFOS.

Quality control

Laboratory background contamination was monitored by including procedural blanks (2 mL Milli-Q water, *n* = 6) during the processing of human milk samples. Blank concentrations were generally below method detection limits for all targets but PFOA and FOSA (see Tables S4–S6, ESI†). For these substances, a higher reporting limit was adopted (see *Data handling* section). Method accuracy and precision were assessed using replicate samples of bovine milk analyzed with a low- (1–20 ng; *n* = 4), or high- (4–60 ng; *n* = 4) spike of PFAS (Table S7, ESI†). In most cases, mean percent recoveries ranged from 68 to 103% and 70 to 101% at low- and high-spiking levels, respectively. The exceptions were for PFBS and 7:3 FTCA, which displayed lower recoveries of 34/67%, 3.3/19% (low/high fortification levels),

respectively, which was attributed to a lack of exactly-matched, isotopically-labelled internal standards. Finally, external validation of the method was achieved through comparison of PFAS concentrations measured in SRM 1954 (*n* = 3) to those reported by Nyberg *et al.*¹⁷ and by Keller *et al.*⁴² (Table S5, ESI†). In general, concentrations reported in SRM 1954 were consistent with Nyberg *et al.*¹⁷ and at the lower end of the range reported by Keller *et al.* (2010).⁴²

Data handling

The limits of detection and quantification (LOD and LOQ, respectively) were defined as the concentration produced at a signal-to-noise ratio of 3 and 10, respectively, and are similar to those reported previously for PFAS in breast milk.^{23,40,43,44} In most cases, concentrations are reported down to the LOD. The exceptions were for PFOA and FOSA in Chinese samples, which were detectable in blanks, and therefore higher reporting limits were used (average blank + 3 × the standard deviation of the blanks). Notably, reporting limits in Nyberg *et al.* were slightly lower than the present work, therefore PFAS in Swedish samples are in some cases detectable despite being below reporting limits used for Chinese samples. In all cases, values below the reporting limit were replaced with 0 for calculation of sum PFAS concentrations, estimated daily intakes (EDIs) and estimated weekly intakes (EWIs). For EDI and EWI calculations we assumed an infant body weight of 7 kg and a milk consumption rate of 600 mL day⁻¹, as described previously.¹⁵

Results and discussion

PFAS profiles

Average Σ_{20} PFAS concentrations, together with relative profiles and the concentrations of the top 9 most frequently detected PFAS are summarized in Fig. 1. Means, ranges, and detection frequencies for all PFAS are summarized in Table 1 (raw data in ESI Tables S4–S6†). The average Σ_{20} PFAS concentrations in human milk (HM) were highest in Jiaxing (765 pg mL⁻¹) followed by Shanghai (462 pg mL⁻¹), Shaoxing (430 pg mL⁻¹), and Stockholm (130 pg mL⁻¹). The year of sampling (2015–2016 for Shanghai and Jiaxing; 2010 for Shaoxing; and 2016 for Stockholm) as well as maternal parity (*n* = 1 or 2 children for individuals from Shanghai and Jiaxing and *n* = 1 child for individuals from Shaoxing and Stockholm) may play an important role in the observed PFAS concentrations and profiles.^{17,45} However, due to the small number of individuals included in this study, the importance of these factors could not be investigated in detail. Despite this potential source of variability, some similarities were observed in PFAS profiles across all cities. For example, profiles in Jiaxing, Shanghai, Shaoxing, and Stockholm were all dominated by PFOS, PFOA, and long chain PFCAs (*i.e.* C9–C14), but in Chinese cities, 9Cl-PF3ONS (the main component of F53-B) was also prevalent, occurring in 100% of samples at concentrations of up to 976 pg mL⁻¹. 11Cl-PF3OUDS (a minor component of F53-B) was also detected in most samples from Jiaxing and Shaoxing (up to 25 pg mL⁻¹) and 3 samples from Shanghai (up to 87 pg mL⁻¹). Neither 9Cl-



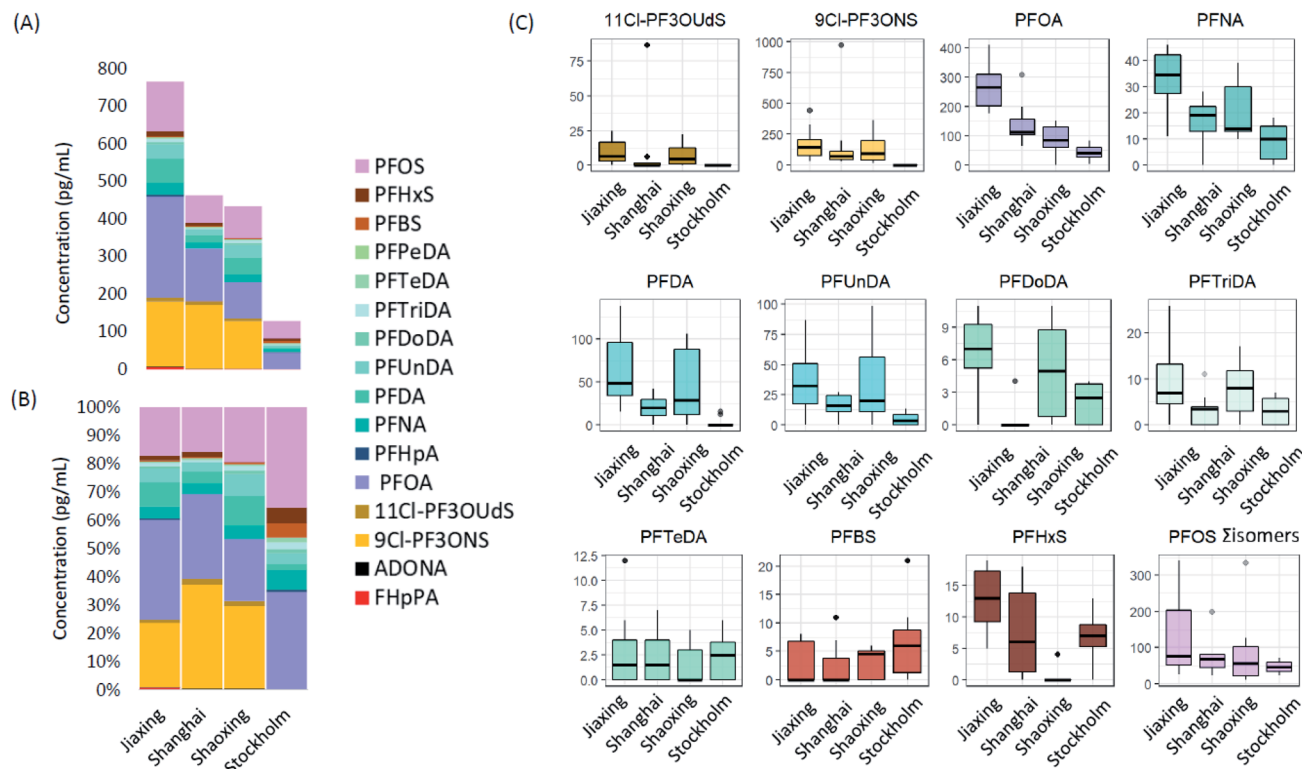


Fig. 1 Sum mean concentrations (pg mL⁻¹; Panel A) and profiles normalized to 100% (Panel B) for PFAS determined in human milk samples from Shanghai, Jiaxing, and Shaoxing (China) and Stockholm (Sweden). Box-whisker plots of individual concentrations are shown in Panel C. Boxes display 1st and 3rd quartiles bisected by the median while whiskers represent the interquartile range (excluding the median). For all panels, values below limits of detection were replaced with '0'.

PF3ONS or 11Cl-PF3OUdS were observed in samples from Stockholm. The occurrence of F53-B components exclusively in Chinese samples is not surprising since this product is produced and used exclusively in China. However, the highly elevated concentrations of 9Cl-PF3ONS, which exceeded PFOS in some samples, was unexpected and highlights the extreme bioaccumulative properties of this substance.³⁶

Among the remaining PFAS, concentrations of PFOS, PFOA, PFNA, PFDA and PFUnDA were generally higher in Chinese cities compared to Stockholm, while concentrations of PFDoDA and PFTriDA were comparable (Fig. 1). Surprisingly, mean PFBS concentrations in Stockholm were slightly higher than Chinese cities, while PFHxS concentrations in Stockholm were comparable to Jiaxing and Shanghai and considerably higher than Shaoxing (Fig. 1). This may point to more prevalent use of PFBS and PFHxS-based formulations in Sweden compared to some locations in China. Contamination of drinking water with PFHxS and PFBS in some Swedish cities is also known to have occurred,⁸ which may (at least in part) explain the occurrence of these substances in Sweden. All other PFAS (*i.e.* PFHpA, Br-PFOA, L-PFPeDA, Br-PFHxS, PFDS, FOSA, MeFOSAA, EtFOSAA, 7:3 FTCA, and ADONA) were detected in <25% of all samples (*i.e.* from China and Sweden), and not exclusively in one city (Table 1). The infrequent detection of these substances precluded any firm conclusions regarding relative differences in exposure among Chinese cities, or between China and Sweden. However,

it is notable that ADONA was detected (but below limits of quantification) in 8 of the 30 Chinese samples but none from Stockholm, while L-FOSA was detected in 3 of the 10 samples from Stockholm but none from China. To the best of our knowledge, this is the first observation of ADONA in human milk. However, given the very low concentrations (*i.e.* <8 pg mL⁻¹), which fell below the LOQ (10 pg mL⁻¹) this result remains tentative at this time.

To the best of our knowledge, Cl-PFESAs have only been reported in human milk once previously.⁴⁰ In that work, samples from 174 women from Hangzhou, China (2018 and 2019) contained $\sum_{19}$ PFAS concentrations ranging from 9–1755 pg mL⁻¹ (mean 203 pg mL⁻¹).⁴⁰ 9Cl-PF3ONS was detected in 100% of samples (consistent with the present study) while 11Cl-PF3OUdS was detected in only 20% of samples. Based on mean concentrations, PFOA was the dominant substance (87 pg mL⁻¹) followed by PFHxA (41 pg mL⁻¹; not measured in the present work), 9Cl-PF3ONS (28 pg mL⁻¹), PFOS (25 pg mL⁻¹), and perfluorobutanoate (24 pg mL⁻¹; not measured in the present work). Mean concentrations of C9–C11 PFCAs ranged from 12–13 pg mL⁻¹. Overall, the PFAS concentrations and profiles reported in Jin *et al.* (2020)⁴⁰ are very similar to those reported in the present work for Shaoxing, which is perhaps not surprising considering the close proximity of these cities. Our work builds on this initial finding by showing that the occurrence of F-53B in human milk is widespread in China.



Table 1 Average concentrations (pg mL^{-1}) $\pm$ stdev and detection frequency (DF) of PFAS in mother's milk from Shanghai, Jiaying and Shaoxing, China, and from Stockholm, Sweden. Range shown in parentheses

Analyte	DF (%; all samples)	Shanghai ($n = 10$)		Jiaying ($n = 10$)		Shaoxing ($n = 10$)		Stockholm ($n = 10$)	
		Conc.	DF (%)	Conc.	DF (%)	Conc.	DF (%)	Conc.	DF (%)
L-PFHpA	10	(<10)	0	(<10–17)	30	(<10)	5	(<10–11)	2.5
L-PFOA	97.5	139 $\pm$ 36 (64–308)	100	266 $\pm$ 37 (177–411)	100	94 $\pm$ 21 (33–151)	47.5	42 $\pm$ 12 (<2–81)	25
Br-PFOA	22.5	(<2–3)	0	4 $\pm$ 1 (<2–8)	50	2 $\pm$ 1 (<2–5)	10	2 $\pm$ 1 (<2–6)	7.5
L-PFNA	90	17 $\pm$ 4 (<5–28)	90	32 $\pm$ 6 (11–46)	100	21 $\pm$ 6 (10–39)	42.5	10 $\pm$ 3 (<5–18)	17.5
L-PFDA	70	21 $\pm$ 6 (<10–42)	80	64 $\pm$ 21 (15–139)	100	47 $\pm$ 21 (<10–106)	27.5	<10 (<10–15)	5
L-PFUnDA	75	16 $\pm$ 4 (<5–27)	80	38 $\pm$ 14 (<5–87)	90	34 $\pm$ 16 (<5–99)	35	7 $\pm$ 2 (<5–13)	12.5
L-PFDoDA	55	(<2–4)	10	7 $\pm$ 2 (2–11)	80	6 $\pm$ 2 (<2–11)	32.5	2 $\pm$ 1 (<2–4)	15
L-PFTriDA	70	4 $\pm$ 1 (<2–11)	60	10 $\pm$ 4 (<2–26)	80	8 $\pm$ 3 (<2–17)	35	4 $\pm$ 1 (<2–7)	15
L-PFTeDA	50	3 $\pm$ 1 (<2–7)	50	4 $\pm$ 2 (<2–12)	50	3 $\pm$ 1 (<2–5)	25	3 $\pm$ 1 (<2–6)	15
L-PFPeDA	10	(<2–3)	10	(<2–4)	10	2 $\pm$ 1 (<2–4)	7.5	(<2)	0
L-PFBS	50	4 $\pm$ 1 (<4–11)	30	5 $\pm$ 1 (<4–8)	40	4 $\pm$ 1 (<4–6)	32.5	7 $\pm$ 3 (<4–21)	17.5
L-PFHxS	65	8 $\pm$ 3 (<4–18)	70	13 $\pm$ 3 (5–19)	100	3 $\pm$ 0.3 (<4–4)	22.5	7 $\pm$ 2 (<4–13)	20
Br-PFHxS	22.5	<2 (<2–3)	30	<2 (<2–5)	30	(<2)	10	(<2–4)	7.5
L-PFOS	100	65 $\pm$ 22 (16–177)	100	119 $\pm$ 52 (21–321)	100	77 $\pm$ 45 (7–308)	50	39 $\pm$ 7 (23–58)	25
Br-PFOS	87.5	8 $\pm$ 3 (<2–22)	90	12 $\pm$ 4 (3–26)	100	7 $\pm$ 4 (1–27)	40	7 $\pm$ 2 (<1–14)	20
L-PFDS	2.5	(<2)	0	(<2)	0	(<2)	5	(<2–3)	2.5
Br-PFDS	0	(<2)	0	(<2)	0	(<2)	2.5	(<2)	0
L-FOSA	7.5	(<2–4)	0	2 $\pm$ 0.5 (<2–4)	0	(<2–4)	7.5	(<2–4)	7.5
Br-FOSA	0	(<2)	0	(<2)	0	(<2)	2.5	(<2)	0
L-MeFOSAA	0	(<5)	0	(<5)	0	(<5)	2.5	(<5)	0
Br-MeFOSAA	0	(<5)	0	(<5)	0	(<5)	2.5	(<5)	0
L-EtFOSAA	0	(<5)	0	(<5)	0	(<5–8)	5	(<5)	0
Br-EtFOSAA	0	(<5)	0	(<5)	0	(<5)	2.5	(<5)	0
7:3 FTCA	20	(<2–4)	10	7 $\pm$ 4 (<2–27)	40	2 $\pm$ 1 (<2–8)	10	(<2–3)	2.5
ADONA	20	3 $\pm$ 1 (<3–8)	30	3 $\pm$ 1 (<3–8)	30	(<3–5)	7.5	(<3)	0
9Cl-PF3ONS	75	170 $\pm$ 144 (31–976)	100	173 $\pm$ 65 (30–443)	100	126 $\pm$ 56 (17–364)	27.5	(<2)	0
11Cl-PF3OUDS	45	11 $\pm$ 13 (<2–87)	30	10 $\pm$ 5 (<2–25)	80	8 $\pm$ 4 (<2–22)	20	(<2)	0

Estimated daily and weekly intakes and potential health risks

Mean Σ_{20} PFAS EDIs were highest in Jiaying (66 $\text{ng kg}^{-1} \text{bw d}^{-1}$; range 30–116 $\text{ng kg}^{-1} \text{bw d}^{-1}$) followed by Shanghai (40 $\text{ng kg}^{-1} \text{bw d}^{-1}$; range 14–144 $\text{ng kg}^{-1} \text{bw d}^{-1}$), Shaoxing (37 $\text{ng kg}^{-1} \text{bw d}^{-1}$; range 5–71 $\text{ng kg}^{-1} \text{bw d}^{-1}$), and finally Stockholm (11 $\text{ng kg}^{-1} \text{bw d}^{-1}$; range 5–17 $\text{ng kg}^{-1} \text{bw d}^{-1}$). In comparison, Winkens *et al.*⁴⁶ reported Σ PFAS EDIs of 43 $\text{ng kg}^{-1} \text{bw d}^{-1}$ for HM sampled from Spain (2008)⁴⁷ and 29 $\text{ng kg}^{-1} \text{bw d}^{-1}$ for HM from Sweden (2004)⁴⁸ while Tao *et al.*⁴⁹ estimated a Σ PFAS EDI of up to 87 $\text{ng kg}^{-1} \text{bw d}^{-1}$ for HM collected in USA in 2004. The range of EDIs reported here for Shanghai are consistent with the results of Liu *et al.*,⁴³ who reported a Σ_6 PFAS EDI of up to 129.1 $\text{ng kg}^{-1} \text{bw d}^{-1}$ (median = 17.2 $\text{ng kg}^{-1} \text{d}^{-1}$) for human milk sampled in 2007 from this city.

In 2008, the European Food Safety Authority (EFSA) established tolerable daily intakes (TDIs) for PFOS and PFOA of 150 and 1500 $\text{ng kg}^{-1} \text{bw d}^{-1}$, respectively.⁴⁹ These values were modified to tolerable weekly intakes (TWIs) of 13 and 6 $\text{ng kg}^{-1} \text{bw w}^{-1}$, respectively, in 2018 (1.8 and 0.8 $\text{ng kg}^{-1} \text{bw d}^{-1}$, respectively, assuming chronic intake),⁵⁰ and most recently (2020) to a single TWI of 8 $\text{ng kg}^{-1} \text{bw w}^{-1}$ based on the sum of four PFAS (PFOA, PFNA, PFHxS and PFOS). A comparison of the most recent TWIs to estimated weekly intakes (EWIs; based on Σ_4 PFAS; Table S9†) in the present work revealed that all samples exceeded the TWI of 8 $\text{ng kg}^{-1} \text{bw w}^{-1}$. The highest EWI was for Jiaying (average 260 $\text{ng kg}^{-1} \text{bw w}^{-1}$; range 163–384 $\text{ng kg}^{-1} \text{bw}$

w^{-1}), followed by Shanghai (average 138 $\text{ng kg}^{-1} \text{bw w}^{-1}$; range 63–314 $\text{ng kg}^{-1} \text{bw w}^{-1}$), Shaoxing (average 118 $\text{ng kg}^{-1} \text{bw w}^{-1}$; range 39–268 $\text{ng kg}^{-1} \text{bw w}^{-1}$) and Stockholm (average 60 $\text{ng kg}^{-1} \text{bw w}^{-1}$; range 26–89 $\text{ng kg}^{-1} \text{bw w}^{-1}$). Unfortunately, TWIs have not been determined for most of the substances analyzed in this study. Such data are particularly important for the main components of F-53B which displayed very high concentrations in some Chinese HM samples. We note that some agencies have already started to define Σ PFAS threshold values based on a larger number of PFAS than what was used in the most recent EFSA estimates (*e.g.* the Swedish Drinking Water Guideline).⁵¹ Re-calculation of EWIs from the present work using Σ_{20} PFAS concentrations revealed averages of 442, 271 and 250 $\text{ng kg}^{-1} \text{bw w}^{-1}$ for Jiaying, Shanghai and Shaoxing, respectively, and 73 $\text{ng kg}^{-1} \text{bw w}^{-1}$ for Stockholm (Table 2), considerably higher than when only using Σ_4 PFAS concentrations.

Conclusions

Our analysis of human milk from China and Sweden revealed the presence of 9Cl-PF3ONS, 11Cl-PF3OUDS, and ADONA in human milk from China. To the best of our knowledge, there is only one prior study which has reported Cl-PFESAs in human milk.⁴⁰ 9-Cl-PF3ONS occurred at extremely high concentrations across all Chinese samples and suggest that human exposure



Table 2 Range, mean, and median estimated weekly intakes (EWIs; ng kg⁻¹ bw w⁻¹) based on $\sum_{20}$ PFAS concentrations and assuming consumption of 600 mL day⁻¹ and a body weight of 7 kg. Values below LOQ were set to 0 for EDI calculations

	Shanghai (n = 10)	Jiaxing (n = 10)	Shaoxing (n = 10)	Stockholm (n = 10)
Range	98–978	206–789	58–487	33–112
Mean	271	447	255	75
Median	204	391	227	72

assessments focused only on legacy substances may severely underestimate overall PFAS exposure in breastfeeding infants.

Conflicts of interest

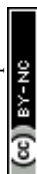
There are no conflicts to declare.

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