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Rhodamine B derivatives-modified upconversion nanoparticles as a fluorescent turn-off–on sensor for the highly sensitive detection of Cu²⁺ and pyrophosphate†

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Rhodamine B derivatives (RBP)-modified UCNPs (UCNPs@mSiO₂-RBP) were developed as a fluorescent turn-off–on sensor based on FRET and IFE to detect Cu²⁺ and pyrophosphate (PPI) with a wide linear response range (0–10 μM for Cu²⁺ and 5–35 μM for PPI, much wider than that reported previously) and high sensibility (117 nM for Cu²⁺ and 70 nM for PPI). The MTT experiments and the bioimaging experiments show its promising prospect in tissue imaging.

Introduction

Copper ions and pyrophosphate (P₂O₇⁴⁻, PPI) are critical to human bodies. Excess and lack of Cu²⁺ ions both can lead to severe diseases such as Alzheimer's disease, while pyrophosphate (PPI) plays an important role in various life activities. Hence, developing an effective method to detect Cu²⁺ and PPI in environmental and biological samples is very important as they are important bio-functional anions.

Fluorescent sensing strategy is widely used for bio-detection due to its simple, real-time and *in vivo* detection. The recognition of PPI is generally performed by utilizing metal ion complexes as binding sites because PPI lacks UV-Vis absorbance and fluorescence. The complexing of metal ions with PPI can generate the strong UV-Vis absorbance and thus result in changes in fluorescence signal. Therefore, fluorescence techniques have been employed in the detection of PPI in the past decades.^{1–3} Cu²⁺ ions can be used to detect PPI due to the strongly complexing ability of PPI to Cu²⁺.¹ Several fluorescence probes have been developed to sequentially detect Cu²⁺ and PPI.^{4–7} However, most fluorescence probes respond mainly to relatively short wavelengths of light irradiation (*e.g.* UV). This may cause photodamage to tissues, and the lower penetration

depth hampers their wide applications to some extent. Recently, upconversion fluorescence probes have earned people's attention due to their unique abilities, which can minimize autofluorescence from the biosamples and enhance the penetration depth in tissues.

To date, hybrids of upconversion nanoparticles (UCNPs) with chromophores have been fabricated for the detection of important biological species and toxins, such as DNA,⁸ CN⁻,⁹ Hg²⁺,¹⁰ Zn²⁺,¹¹ and H₂S.¹² These hybrid sensors are based on the process of energy-transfer from UCNPs to chromophores to “turn on” or “turn off” the upconversion luminescence. Recently, several rhodamine B derivatives RBH-modified UCNP sensors have been reported to detect Cu²⁺ ions.^{13–15} However, there are very few reports on the use of UCNPs to detect PPI. Wang *et al.* reported that branched polyethyleneimine (PEI)-capped NaGdF₄:Yb/Tm UCNPs can be used as a fluorescence probe for the sequential detection of Cu²⁺ and pyrophosphate (P₂O₇⁴⁻, PPI).⁴ However, the sensor exhibited a narrow linear response range (0.1–2 μM for Cu²⁺ and 0.5–8 μM for PPI). Therefore, it is of importance to develop upconversion probes with a broad linear range.

Herein, we developed a rhodamine B derivative RBP- (RBP was synthesized according to literature,^{16,17} as shown in Fig. S1†) modified UCNPs as a sensor to detect Cu²⁺ and PPI. RBP was usually used to serve as a “turn-on” fluorescent sensor for Al³⁺ with strong fluorescent response, but it was rarely reported as a fluorescent sensor for Cu²⁺ due to its weak fluorescent response. It was found that RBP had only weak UV-Vis absorbance but showed strong fluorescence response to Al³⁺. In contrast, the response of RBP to the copper is very different from the Al³⁺, the strong UV-Vis absorbance will appear when Cu²⁺ was added to RBP solution due to the paramagnetism of copper ions, but the fluorescence response to Cu²⁺ is weak,

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† Electronic supplementary information (ESI) available: ¹H NMR spectra of RBH and RBP, TEM images of UCNPs and UCNPs@mSiO₂, powder-XRD, EDX spectrum and elemental mapping of the UCNPs, IR spectra and emission intensity of the UCNPs@mSiO₂, absorption spectra of the PBP and RBH, and selectivity test for the RBP to the Cu²⁺, and the linear plot of detection (absorbance) of Cu²⁺. See DOI: 10.1039/c8ra08090a



which shows that RBP has higher responding speed and intensity of UV-Vis absorbance than the traditional probe RBH to detect Cu^{2+} .^{13–15}

In this study, we aimed to achieve efficient FRET (fluorescence resonance energy transfer) and IFE (internal filter effect) from UCNP to RBP, which was expected to have strong UV-Vis absorbance response. Hence, RBP was selected as a chromophore to modify UCNP owing to its strong UV-Vis absorbance response towards Cu^{2+} . In the hybrid RBP-UCNP sensing system, the added Cu^{2+} will coordinate with RBP and lead to a strong quenching of the upconversion luminescence. When PPI was present, it could bind strongly with Cu^{2+} and thus turn on the fluorescence of the UCNP (Scheme 1).

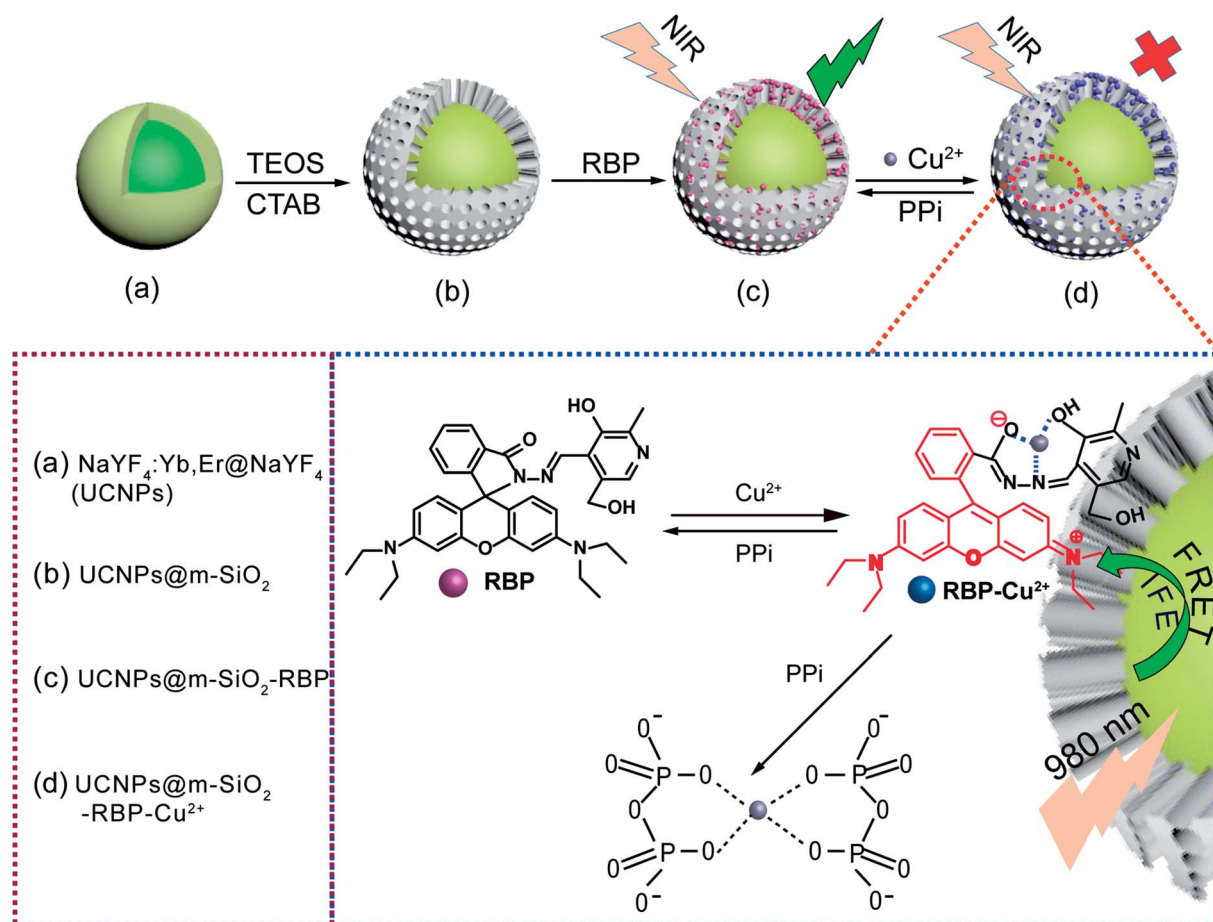
Results and discussions

Preparation of UCNP and the UCNP@mSiO₂-RBP composite probe

$\text{NaYF}_4:\text{Yb}^{3+},\text{Er}^{3+}$ UCNP were synthesized by the reported coprecipitation method.¹⁸ To enhance the emission intensity of the UCNP, an inert layer of NaYF_4 was coated onto their surface by a typical core-shell approach.^{19,20} The TEM images of the as-prepared $\text{NaYF}_4:\text{Yb}^{3+},\text{Er}^{3+}$ and $\text{NaYF}_4:\text{Yb}^{3+},\text{Er}^{3+}@\text{NaYF}_4$ UCNP displayed uniform shape and size, and the corresponding XRD

patterns shown in Fig. S3c† were well indexed to the standard card of $\beta\text{-NaYF}_4$ (JCPDS 16-0334). In addition, the luminescence spectra in Fig. S3d† indicate that core-shell strategy significantly improved the luminous intensity. The core-shell structure exhibits strong green emissions at 525 nm and 546 nm, originating from the transition of $^2\text{H}_{11/2} \rightarrow ^4\text{I}_{15/2}$ and $^4\text{S}_{3/2} \rightarrow ^4\text{I}_{15/2}$ of Er^{3+} . The EDX spectrum of $\text{NaYF}_4:\text{Yb}^{3+},\text{Er}^{3+}$ illustrated in Fig. S4(a)† implies that the main constituents in the studied samples are F, Na, Y, Yb and Er. Furthermore, the elemental mapping of F, Na, Y, Yb and Er (Fig. S4(c)–(g)†) and the total image formed by overlapping the elemental maps also indicate that various elements are well distributed in the entire range of nanoparticles (Fig. S4(b)†).

Aimed towards loading RBP and detecting PPI in aqueous solution, the UCNP were coated by mesoporous silica (mSiO_2). UCNP@mSiO₂ was characterized by TEM, FT-IR and luminescence spectroscopy. The TEM images in Fig. S5a and b† clearly demonstrate the formation of the SiO₂ layer on the surface of UCNP. Furthermore, through ion exchange with ammonium nitrate, CTAB molecules were removed and mSiO_2 were formed. In the FT-IR spectra in Fig. S5c,† the stretching vibration peaks disappeared, which further confirmed the removal of CTAB. The emission spectra in Fig. S5d† disclosed that the removal of CTAB has a minor effect on the emission



Scheme 1 Schematic of the detection of Cu^{2+} and PPI by RBP attached with UCNP. (a) Core-shell UCNP (b) UCNP@m-SiO₂ (c) UCNP@m-SiO₂ absorbed the RBP to form the composite probe (d) composite probe with Cu^{2+} .



properties of UCNPs. Then, a certain amount of the as-prepared UCNPs@mSiO₂ was added to the 20 μM rhodamine B derivative RBP solution and stirred overnight to obtain the fluorescent probe UCNPs@mSiO₂-RBP.

As shown in Fig. 1a, UCNPs@mSiO₂-RBP showed strong green emissions at 525 nm and 546 nm and a weak red emission, similar to that of unmodified UCNPs, because RBP has no evident absorbance in the visible region. However, green emission intensity was significantly quenched after the addition of Cu²⁺ due to the strong absorbance of RBP-Cu²⁺ from 520 nm to 600 nm. The strong affinity between Cu²⁺ and PPI leads to the formation of the Cu²⁺-PPI complex and results in the detachment of Cu²⁺ from the UCNPs@mSiO₂-RBP probe; thus, the fluorescence is triggered by the addition of PPI. A significant decrease in the Er³⁺ ⁴S_{3/2} → ⁴I_{15/2} lifetime was observed when Cu²⁺ was added to UCNPs@mSiO₂-RBP from 0 μM to 10 μM (Fig. 1b, measured with the modified Edinburgh FS5 fluorescence spectrometer under a frequency of 20 Hz and a pulse width of 300 μs to modulate the 980 nm laser), which is solid evidence for FRET. However, the lifetime of Er³⁺ emission changed slightly when the concentration of Cu²⁺ was increased from 10 μM to 15 μM, while the PL intensity of the composite probe decreased significantly (Fig. 2b), indicating that IFE also worked in this quenching process. These results suggest that FRET and IFE are the two main processes in the detection process.

Detecting Cu²⁺ and PPI

First, the sensing ability of UCNPs@mSiO₂-RBP towards Cu²⁺ was studied by UV-Vis absorbance spectroscopy with different concentrations of Cu²⁺. From the absorbance spectra (Fig. 2a), it can be found that the absorbance intensity at the peak (500–600 nm) increased with the increase in concentration of the Cu²⁺ ion. Fig. S6† shows the linear plot of the concentration of Cu²⁺ versus the absorbance intensity.

To certify the energy transfer of UCNPs emission to RBP, the effect of Cu²⁺ concentration on the upconversion emission intensity of the UCNPs@mSiO₂-RBP sensing system was explored. Fig. 2b shows a gradual decrease of green upconversion emission intensity with the increase in amount of Cu²⁺ in

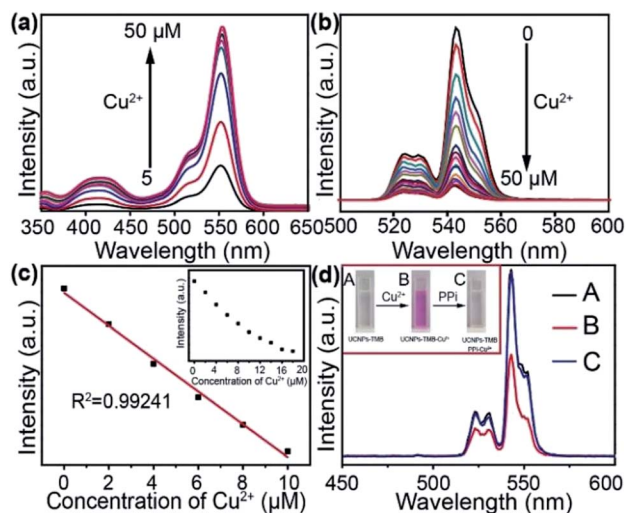


Fig. 2 (a) UV-Vis spectra of UCNPs@mSiO₂-RBP dispersion with different concentrations of Cu²⁺ (0–50 μM) in the solution of ethanol–water. (b) Fluorescence spectra of UCNPs@mSiO₂-RBP dispersion upon the addition of different concentrations of Cu²⁺ (0–50 μM) in the solution of ethanol–water. (c) The linear plot of the detection (emission) of Cu²⁺, the inset is scatter plot for different concentrations of Cu²⁺ (0–20 μM). (d) The digital images of colour changes after the addition of Cu²⁺ and PPI in sequence and its emission spectra in the ethanol–water solution.

the range of 0–50 μM. When the Cu²⁺ concentration reached 50 μM, the upconversion emission at 540 nm quenched by about 92% of the initial emission intensity of UCNPs@mSiO₂-RBP. As shown in Fig. 2c, the concentration of Cu²⁺ exhibited a linear correlation with the emission intensity of UCNPs@mSiO₂-RBP with an R² of 0.99241 in the concentration range of 0–10 μM, which was much wider than that for the recently reported fluorescence methods.⁴ The limit of detection for the concentration of Cu²⁺ was calculated to be 117 nM based on 3σ/slop, where σ was the standard deviation of the blank samples and slop was the slope of the calibration curve. Herein, RBP has relatively broader linear range than previously reported RBH, which can be reasoned by the different intensities of UV-Vis of RBP and RBH toward Cu²⁺, shown in Fig. S2.† The high sensibility for Cu²⁺ of RBP can be attributed to its molecular structure. RBP was synthesized by RBH and pyridoxal hydrochloride (as shown in Fig. S1.†), which has larger electron cloud density than RBH and thus, it can combine with Cu²⁺ more easily.

To prove the high selectivity of the developed sensor for Cu²⁺, the effect of other cations, including K⁺ Ca²⁺ Na⁺ Zn²⁺ Ba²⁺, Al³⁺ and Mn²⁺, on the UV-Vis absorbance spectrum was investigated. When the concentration of Cu²⁺ was maintained at 20 μM and that of other cations was 100 μM, no apparent signal change was observed in contrast to Cu²⁺ except for the signal change due to Al³⁺, as shown in Fig. S7.† which displayed a slight response to UV-Vis irradiation at 520–560 nm.

The fluorescence intensity of the system could be recovered on adding PPI to the system, which can be attributed to the stronger coordination effect between PPI with Cu²⁺ than RBP since PPI can effectively withdraw the Cu²⁺ from the complexes of UCNPs@mSiO₂-RBP-Cu²⁺. In fact, the same mechanism for

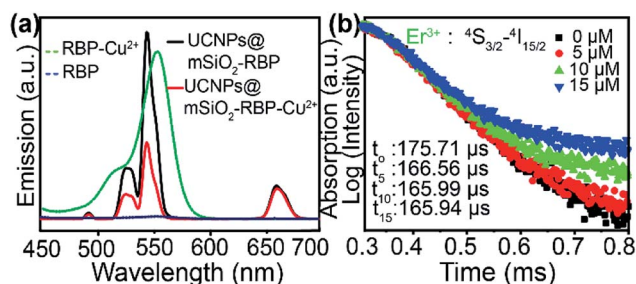


Fig. 1 (a) The absorbance spectrum of RBP-Cu²⁺ and RBP and the emission spectrum of UCNPs@mSiO₂-RBP and UCNPs@mSiO₂-RBP-Cu²⁺ in the ethanol–water solution. (b) Er³⁺ decay curves recorded for UCNPs@mSiO₂-RBP with different concentrations of Cu²⁺ from 0 to 15 μM in the ethanol–water solution.



detecting PPI has been described in previously reported studies.^{6,21,22} The photographs of UCNPs@mSiO₂-RBP and the emission spectra of UCNPs@mSiO₂-RBP after adding Cu²⁺ and PPI in sequence (Fig. 2d) clearly showed this fluorescence “turn-off-on” process.

At the same time, we also studied the sensing ability of UCNPs@mSiO₂-RBP-Cu²⁺ toward PPI *via* UV-Vis absorbance spectroscopy. The absorbance intensity of UCNPs@mSiO₂-RBP-Cu²⁺ decreased linearly in a certain range (0–15 μM, Fig. 3b) with the increase in concentration of PPI (Fig. 3a), which approved that the strong complexing of PPI with Cu²⁺ plays a crucial role in this process.

To examine the feasibility of using UCNPs@mSiO₂-RBP-Cu²⁺ as the turn-on sensing probes for PPI, different concentrations of PPI were added into the solutions containing UCNPs@mSiO₂-RBP (RBP: 20 μM, UCNPs@mSiO₂: 10 mg mL⁻¹) and Cu²⁺ (20 μM) under optimal conditions. Fig. 3c shows that the fluorescence intensity of the system could be gradually recovered on increasing the concentration of PPI, and almost 100% of the initial fluorescence intensity recovered when the concentration of PPI reached 45 μM. In addition, a linear fitting curve of fluorescence intensity *versus* concentration of PPI was obtained in the range of 5–35 μM, with an *R*² value of 0.99672 (Fig. 3d). The limit of detection (LOD) was estimated to be 70 nM based on 3σ/slop. Compared with the reported results of the detection of Cu²⁺ and PPI (Table S1 and S2[†]), our results also show the good ability for ion detection.

The specificity of UCNPs@mSiO₂-RBP-Cu²⁺ toward PPI was also evaluated; the fluorescence response of UCNPs@mSiO₂-RBP-Cu²⁺ toward PPI was examined with the existence of PO₄³⁻, HPO₄²⁻, H₂PO₄⁻, Ac⁻, SO₄²⁻, HSO₄⁻ or F⁻ inions under the

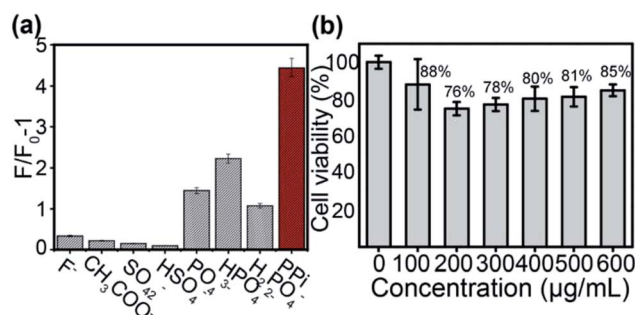


Fig. 4 (a) Selectivity for PPI detection in the ethanol–water solution. (The concentration of PPI and the other anions is 100 μM, where F_0 is the fluorescence intensity of UCNPs@mSiO₂-RBP-Cu²⁺, and F is the fluorescence intensity of the UCNPs@mSiO₂-RBP-Cu²⁺ with the added anions.) (b) Cell viabilities of HeLa cells incubated with RBP-capped UCNPs at different concentrations (0, 100, 200, 300, 400, 500, 600 μg mL⁻¹) for 24 h.

same concentration as PPI. Fig. 4a shows the intensity of ($F/F_0 - 1$) against the concentrations of PPI and other anions. It is evident that only PPI resulted in drastic fluorescence recovery. The anions PO₄³⁻, HPO₄²⁻, H₂PO₄⁻ also caused fluorescence recovery to some degree, which may be due to the similar chemical structures to form stable metal chelates. The influence of phosphate anions could be considered to be acceptable due to the much weaker recovered fluorescence intensity than that recovered by PPI, while other anions had slight effects on changes in fluorescence intensity. These results indicated that our probe had good specificity for PPI detection.

Cell cytotoxicity and *in vivo* imaging of the composite probe

The effect of UCNPs@mSiO₂-RBP on the proliferation of HeLa cells was investigated by a standard MTT assay. As shown in Fig. 4b, the cell viability is still satisfactory when the concentration of the nanoprobe is raised to 600 μg mL⁻¹. These results show that the cytotoxicity of the composite probe is weak, and can be applied in bioimaging.

Furthermore, bioimaging experiments were performed. All procedures and operations were performed according to the Guide for the Care and Use of Laboratory Animal Resources and the National Research Council, and were approved by the Institutional Animal Care and Use Committee of the NIH.

Nude mice (~10 g) were purchased from the Dalian Medical University (Dalian, China) and were used in imaging studies. 100 μL of 0.9% NaCl saline solution containing the RBP-coated UCNPs (10 mg mL⁻¹) were subcutaneously injected into the back area of nude mice (~10 g) anesthetized with chloral hydrate. The mice were then imaged using a modified Night-OWL II LB983 small animal *in vivo* imaging system equipped with a sensitive charge-coupled device (CCD) camera with the excitation of a 980 nm laser (c.w.). Other mice underwent the same process except with an injection of 100 μL of Cu²⁺ solution (50 μM) in the same place after injecting UCNPs@mSiO₂-RBP. As shown in Fig. 5, *in vivo* imaging shows the promising prospect in tissue imaging, and the decrease in the imaging intensity from 1141 to 718 after injecting Cu²⁺ indicated that UCNPs@mSiO₂-RBP has good ability to detect Cu²⁺ *in vivo*.

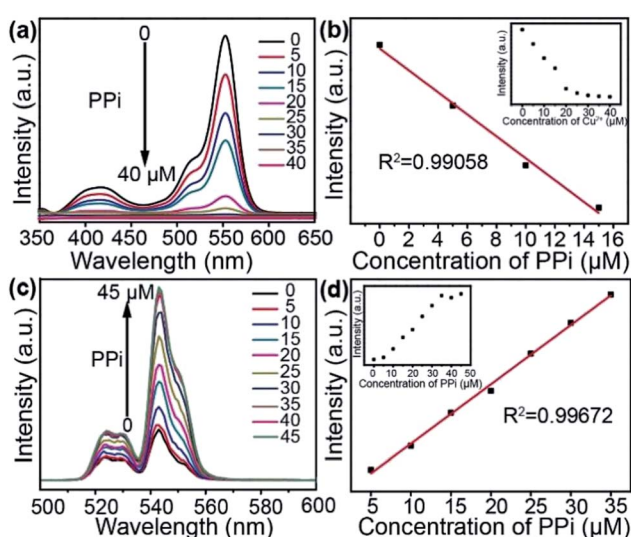


Fig. 3 (a) UV-Vis spectra of UCNPs@mSiO₂-RBP dispersion with different concentrations of PPI. (0–40 μM) in the ethanol–water solution. (b) The linear plot of detection (absorbance) of PPI, the inset is the scatter plot of different concentrations of PPI (0–40 μM). (c) The fluorescence spectra of UCNPs@mSiO₂-RBP-Cu²⁺ with different concentrations of PPI (0–45 μM) in the ethanol–water solution. (d) The linear plot of detection (emission) of PPI, the inset is a scatter plot of different concentrations of PPI.



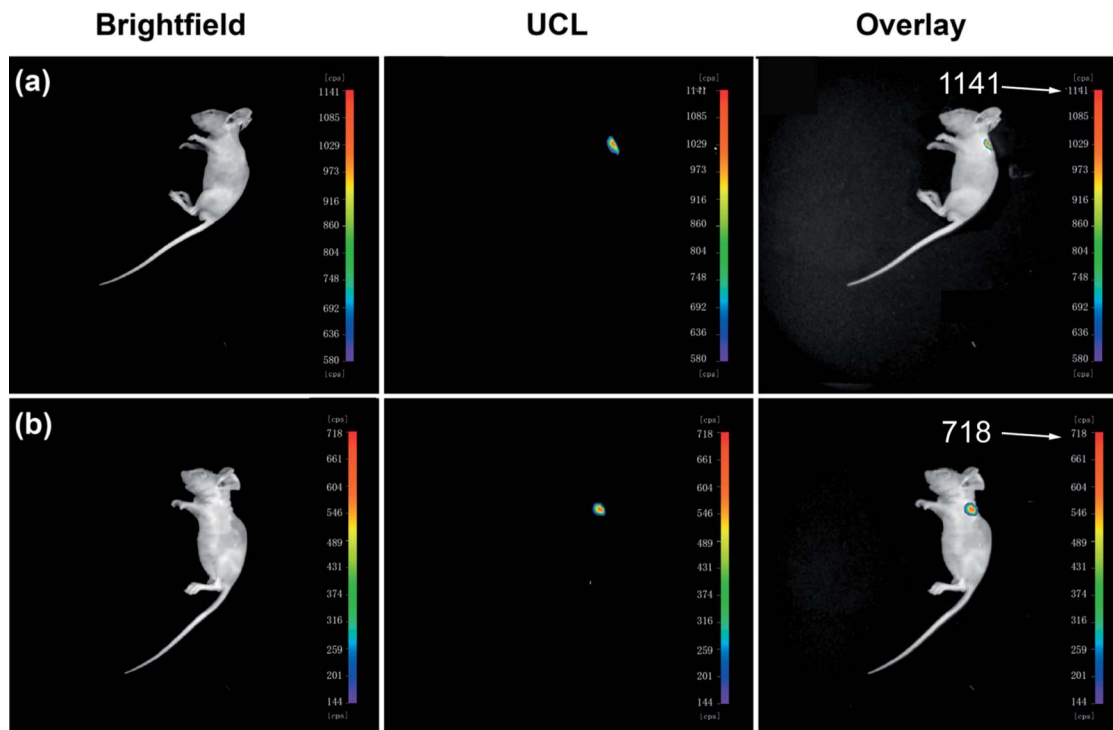


Fig. 5 *In vivo* upconversion luminescence (UCL) imaging of nude mice with (a) subcutaneous injection of UCNPs@mSiO₂-RBP (10 mg mL⁻¹) (b) subcutaneous injection of UCNPs@mSiO₂-RBP (10 mg mL⁻¹) and Cu²⁺ (50 μM), sequentially.

Conclusions

In summary, in this study, we used RBP-modified UCNPs@mSiO₂ as a probe to detect Cu²⁺ and PPI. The results show that the fluorescence intensity of UCNPs@mSiO₂-RBP is almost linear with the concentrations of Cu²⁺ ions in the range of 0–10 μM. The detection limit is 117 nM. We further used the UCNPs@mSiO₂-Cu²⁺ complex as a probe to detect PPI because of the strong interactions between PPI and Cu²⁺. These results show that the fluorescence intensity of UCNPs@mSiO₂-RBP-Cu²⁺ has a strong linear correlation with the concentration of PPI ion in the range of 5–35 μM, and the detection limit is about 70 nM. The cytotoxicity of this sensing system is weak, and *in vivo* imaging shows its promising prospect in tissue imaging and good ability to detect Cu²⁺ *in vivo*.

Experimental

Experimental details and characterization data can be found in the ESI.†

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

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Notes and references

- 1 Z. S. Qian, L. J. Chai, Y. Y. Huang, C. Tang, J. Jia Shen, J. R. Chen and H. Feng, *Biosens. Bioelectron.*, 2015, **68**, 675–680.
- 2 S. Kim, M. S. Eom, S. K. Kim, S. H. Seo and M. S. Han, *Chem. Commun.*, 2013, **49**, 152–154.
- 3 X. Feng, Y. An, Z. Yao, C. Li and G. Shi, *ACS Appl. Mater. Interfaces*, 2012, **4**, 614–618.
- 4 F. Wang, C. Zhang, Q. Xue, H. Li and Y. Xian, *Biosens. Bioelectron.*, 2017, **95**, 21–26.
- 5 L. Wang, Q. Song, Q. Liu, D. He and J. Ouyang, *Adv. Funct. Mater.*, 2015, **25**, 7017–7027.
- 6 S. Lee, K. K. Y. Yuen, K. A. Jolliffe and J. Yoon, *Chem. Soc. Rev.*, 2015, **44**, 1749–1762.
- 7 X. Huang, Z. Guo, W. Zhu, Y. Xie and H. Tian, *Chem. Commun.*, 2008, 5143–5145.
- 8 M. Kumar, Y. Guo and P. Zhang, *Biosens. Bioelectron.*, 2009, **24**, 1522–1526.
- 9 L. Yao, J. Zhou, J. Liu, W. Feng and F. Li, *Adv. Funct. Mater.*, 2012, **22**, 2667–2672.
- 10 Q. Liu, J. Peng, L. Sun and F. Li, *ACS Nano*, 2011, **5**, 8040–8048.
- 11 J. Peng, W. Xu, C. L. Teoh, S. Han, B. Kim, A. Samanta, J. C. Er, L. Wang, L. Yuan, X. Liu and Y. T. Chang, *J. Am. Chem. Soc.*, 2015, **137**, 2336–2342.
- 12 J. Peng, C. L. Teoh, X. Zeng, A. Samanta, L. Wang, W. Xu, D. Su, L. Yuan, X. Liu and Y.-T. Chang, *Adv. Funct. Mater.*, 2016, **26**, 191–199.



- 13 B. Gu, M. Ye, L. Nie, Y. Fang, Z. Wang, X. Zhang, H. Zhang, Y. Zhou and Q. Zhang, *ACS Appl. Mater. Interfaces*, 2018, **10**, 1028–1032.
- 14 Y. Xu, H. Li, X. Meng, J. Liu, L. Sun, X. Fan and L. Shi, *New J. Chem.*, 2016, **40**, 3543–3551.
- 15 C. Li, J. Liu, S. Alonso, F. Li and Y. Zhang, *Nanoscale*, 2012, **4**, 6065–6071.
- 16 Q. Huang, Q. Zhang, E. Wang, Y. Zhou, H. Qiao, L. Pang and F. Yu, *Spectrochim. Acta, Part A*, 2016, **152**, 70–76.
- 17 Y. Xiang, L. Mei, N. Li and A. Tong, *Anal. Chim. Acta*, 2007, **581**, 132–136.
- 18 F. Wang, R. Deng and X. Liu, *Nat. Protoc.*, 2014, **9**, 1634–1644.
- 19 P. Ghosh, J. Oliva, E. D. I. Rosa, K. K. Haldar, D. Solis and A. Patra, *J. Phys. Chem. C*, 2008, **112**, 9650–9658.
- 20 P. Hu, X. Wu, S. Hu, Z. Chen, H. Yan, Z. Xi, Y. Yu, G. Dai and Y. Liu, *Photochem. Photobiol. Sci.*, 2016, **15**, 260–265.
- 21 K. Xu, Z. Chen, L. Zhou, O. Zheng, X. Wu, L. Guo, B. Qiu, Z. Lin and G. Chen, *Anal. Chem.*, 2015, **87**, 816–820.
- 22 K. Selvakrishnan and Y.-C. Chen, *Biosens. Bioelectron.*, 2014, **61**, 88–94.

