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Furosemide's one little hydrogen atom: NMR crystallography structure verification of powdered molecular organics†

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The potential of NMR crystallography to verify molecular crystal structures deposited in structural databases is evaluated, with two structures of the pharmaceutical furosemide serving as examples. While the structures differ in the placement of one H atom, using this approach, we verify one of the structures in the Cambridge Structural Database using quantitative tools, while establishing that the other structure does not meet the verification criteria.

Significant progress has been made in solid-state nuclear magnetic resonance (SSNMR). With the development of high applied magnetic fields¹ and fast magic-angle spinning (MAS) probes,^{2,3} studies of chemical systems that would have been intractable using SSNMR as recently as 10 years ago are becoming routine. An area of consistent recent development is 'NMR crystallography'.^{4–6} As the name suggests, it offers insights into the structures of crystalline materials, but is distinct from diffraction methods as it uses SSNMR experiments to collect system data. Alongside the SSNMR advances, developments in density functional theory (DFT) software operating under periodic boundary conditions that use pseudopotentials to describe core electron states and plane waves to describe the valence electrons have enabled the efficient computation of NMR parameters for periodic systems.^{4,7,8} When relativistic effects may be ignored, the accuracy of these quantum chemical tools is such that, when paired with SSNMR data, this combined approach has been able to refine and determine molecular crystal structures *in lieu* of diffraction,^{9–11} although this approach is not yet a general method for crystal structure determinations.

Diffraction-based crystal structure determination methods have been crucial in the advancement of chemistry.¹² Crystal structure databases, such as the Cambridge Structural Database (CSD), are important repositories of crystal structures determined using diffraction methods. While metrics are available to the crystallographic community to quantify the quality of a crystal structure refinement (*e.g.*, *R*-factors), systematic studies of the CSD have shown that a significant portion of deposited structures possess unrealistic structural features.^{13–15} Independent verification of crystal structures determined using diffraction methods could be of significant general value. Indeed, recent work by van de Streek and Neumann has shown that dispersion-corrected DFT (dc-DFT) offers the potential to verify crystal structures by observing the changes in the non-H atomic positions under a full structural relaxation.¹⁶ However, due to the difficulties in locating hydrogen atom positions by X-ray diffraction (XRD), prior studies have not included them in their verification procedures.

In principle, new single-crystal XRD (scXRD) experiments may be performed to confirm a crystal structure, although this can be a significant challenge for systems that do not readily form sufficiently large single crystals. Secondly, the use of XRD methods to verify XRD structures, *e.g.* by comparison against established distributions of interatomic distances, is obviously cyclical in nature. NMR crystallography methods can address both these aspects. As NMR experiments are sensitive local probes of structure and dynamics, they probe matter in a different fashion than XRD, and do not require single crystal samples. Based upon the successes of powder NMR crystallography for structure determinations,^{2,9–11} refinements¹⁷ and distinctions,^{18,19} NMR may have an important role in the verification of crystal structures deposited in chemical structure databases.

As a test case of NMR crystallography methods to verify crystal structures, we consider furosemide, which is an important pharmaceutical listed in the World Health Organization's List of Essential Medicines.²⁰ Furosemide has therapeutic applications related to relieving fluid accumulation in the heart, liver and kidney due to the partial or total failure of these organs,²¹

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† Electronic supplementary information (ESI) available: Molecular label specifications, full experimental disclosure, pXRD data for **S-A**, crystal structure data for **FURSEM-NEW**, tables of chemical shifts, ¹⁵N CP/MAS NMR spectrum, additional ¹H–¹³C HETCOR spectra, details of computational results (shielding values and energies), shift/shielding correlation plots, computed ¹³C NMR spectrum, sample RMSD calculation. CCDC 1458460. Original research data can be accessed through data DOI: 10.15128/s1784k724. For ESI and crystallographic data in CIF or other electronic format see DOI: 10.1039/c6cc02171a





Fig. 1 An overlay of the asymmetric units of **FURSEM01** (C atoms in grey) and **FURSEM17** (C atoms in green). H atoms are generally omitted, but a critical difference in the carboxyl H atom positioning is shown. Arrow head points toward the H position of **FURSEM17**.

and in treating hypertension.²² Due to its poor bioavailability, recent studies have focused on the synthesis of furosemide co-crystals,^{23–26} but prior SSNMR data are known for pure furosemide (form 1).^{24,27} There are several crystal structure determinations in the CSD for furosemide (*ca.* 10), which provides the opportunity to compare and contrast selected structures using NMR crystallography methods.

We highlight the structures of two determinations of furosemide (Fig. 1), referred to here using their CSD reference codes, **FURSEM01**²⁸ and **FURSEM17**.²⁹ The diffraction measurement temperatures were similar between both structures (ambient), and both have relatively low *R*-factors (6.8% and 5.7%, respectively). This eliminates complicating factors, such as temperature-dependent polymorphism and obviously poor structures (*i.e.*, *R*-factor > 10%).³⁰

A review of furosemide structures in the CSD by Karami *et al.*³¹ used simulated powder XRD (pXRD) patterns to show that several deposited structures were essentially the same polymorphic form (form 1), and were most correctly described by **FURSEM01**. In contrast to the two other known polymorphic forms of furosemide (forms 2 and 3),³² the structure of **FURSEM17** is very similar to **FURSEM01**, but contains a chemically non-intuitive hydrogen bonding motif where one of the COOH hydrogen atoms is directed away from forming an O–H···O dimer. The refined structures possess very similar unit cell dimensions and non-H atomic positions (non-H atomic root-mean-squared difference (RMSD) value of 0.015 Å for 16 overlaid molecules), with *Z'* = 2. As the major difference between these crystal structures lies in the placement of a single hydrogen atom, (Fig. 1) the calculated pXRD patterns are nearly identical (ESI[†]), and thus pXRD cannot distinguish between these two structures. In contrast, NMR experiments are expected to be sensitive to this structural difference, given their local nature. However, it is unknown if the sensitivity to this single local difference is sufficient to distinguish between **FURSEM01** and **FURSEM17** over the total crystal structure.

We obtained powders of furosemide from two sources. The first was used as-is from Sigma-Aldrich (**S-A**), with phase purity

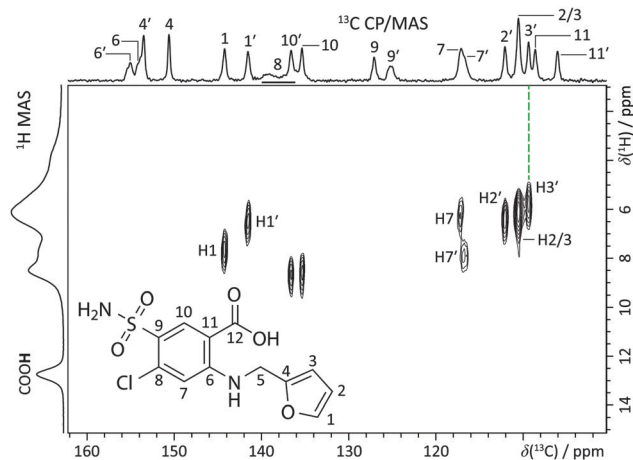


Fig. 2 Partial ¹H–¹³C refocused dipolar FSLG-HETCOR NMR spectrum ($B_0 = 11.7$ T, $T = 293(1)$ K; $\nu_{\text{MAS}} = 13.24$ kHz, contact time = 100 μ s) with site assignments for the furosemide molecule (inset, **S-A** sample). The top horizontal axis shows the ¹³C CP/MAS NMR spectrum, while the left vertical axis corresponds to the 60 kHz ¹H MAS NMR spectrum of the same material. See ESI[†] for a detailed account of the assignment.

verified by pXRD (ESI[†]). A second sample was obtained by following the re-crystallisation procedure advocated by the authors who obtained the structure of **FURSEM17** (**ReCryst**). This yielded a sufficiently large single crystal to solve its crystal structure, which is referred to as **FURSEM-NEW**.[‡]

Fast ¹H MAS, ¹³C and ¹⁵N cross-polarization (CP)/MAS, and ¹H–¹³C refocused frequency-switched Lee-Goldburg dipolar heteronuclear correlation (FSLG-HETCOR) SSNMR experiments were used to achieve a reasonably complete assignment of the NMR peaks to sites in the furosemide molecules (Fig. 2 and ESI[†]). The present assignment is largely consistent with that of Longhi *et al.*,²⁷ however the peak at *ca.* 138 ppm, assigned here to C–Cl groups, was apparently not observed in the prior account, leading to a misassignment. By using variable contact time FSLG-HETCOR experiments (ESI[†]), and analysing systematic trends in computed magnetic shielding values from gauge-including projector augmented-wave density functional theory (GIPAW DFT) calculations (ESI[†]),³³ we could distinguish between nearly all of the H and C chemical sites in the two furosemide molecules of the asymmetric unit. Further ¹³C–¹³C correlation experiments could provide data for a complete assignment, but as the ¹H spin-lattice relaxation value (T_1) is over 40 s at room temperature (ESI[†]), this experiment is not feasible without ¹³C isotopic enrichment. Signal enhancement methods such as dynamic nuclear polarization (DNP),^{34,35} or the addition of paramagnetic dopants may also prove fruitful.

The assignment facilitates the comparison between experimentally measured isotropic chemical shift values (δ_{iso}) with those calculated using GIPAW DFT, quantified through an RMSD. This contrasts with earlier work where differences in H atom positioning could be distinguished by simple qualitative comparisons of either ¹H fast MAS spectra¹⁸ or ¹³C NMR spectra.¹⁹ The quantitative ensemble approach presented here has been used in NMR crystallography structure determinations of organics,^{9,10}





Fig. 3 RMSD values for $\delta_{\text{iso}}(^1\text{H})$ (a) and $\delta_{\text{iso}}(^{13}\text{C})$ (b) between experimentally measured δ_{iso} of **S-A** and **ReCryst** with those computed using GIPAW DFT on the H-optimised structures indicated. Grey bands correspond to RMSD ranges established using a series of benchmark organics. \S The ^1H RMSD is highly sensitive to the COOH hydrogen atom position, as shown by RMSD values which include (blue) and do not include (red) this site. (c) compares calculated (**FURSEM01**/**FURSEM17**) and experimental (**S-A**) ^1H NMR spectra. Red traces in (c) correspond to the carboxyl H atom from each crystallographically unique furosemide molecule.

but has not previously been used to verify hydrogen positioning. ^1H RMSD values of 0.33 ± 0.16 ppm, and ^{13}C RMSD values of 1.9 ± 0.4 ppm are ranges typical for valid molecular organic structures. \S Accordingly, the crystal structures of **FURSEM01**, **FURSEM17**, and **FURSEM-NEW** were H-optimised, followed by GIPAW DFT calculations using CASTEP. $^{36-38}$ The calculated δ_{iso} values for ^1H and ^{13}C were compared against those measured experimentally to yield δ_{iso} RMSD values, Fig. 3(a and b). In Fig. 3c we compare the calculated ^1H NMR spectra (see ESI, \dagger for calculated vs. experimental correlation plots) for **FURSEM01** and **FURSEM17** with the experimental spectrum of **S-A**.

Although there is some disagreement when considering calculated and experimental $\delta_{\text{iso}}(^1\text{H})$ values for the OH sites

in **FURSEM01**, this is not sufficient to invalidate its structure; as discussed in the ESI, \dagger structure changes of a few pm can significantly reduce the ^1H RMSD value. Importantly, the overall agreement between the calculated and experimental data is clearly much better for the structures of **FURSEM01** and **FURSEM-NEW**, while the calculated values using the **FURSEM17** structure are in poor agreement with experimental values. Further, the differences in the computed ^1H and ^{13}C RMSD values for the **FURSEM01** and **FURSEM17** structures allow them to be distinguished using NMR crystallography, with a significance of *ca.* 2σ in both ^1H and ^{13}C . For **FURSEM01** and **FURSEM-NEW** however, the differences between both the atomic positions (non-H atomic RMSD over the unit cell = 0.08 \AA), and the computed δ_{iso} values are minor, especially when noting that **FURSEM-NEW** was determined at 120 K, while **FURSEM01** was determined at room temperature. As such these two structures cannot be distinguished by NMR crystallography, although the slightly lower RMSD values for **FURSEM-NEW** may reflect the lower temperature at which the data were acquired. 39 Despite following the crystallisation procedure of the authors of **FURSEM17**, we do not arrive at the same crystal structure.

Due to the good agreement between calculation and experiment for **FURSEM01** (RMSD(^1H) = 0.46 ppm and RMSD(^{13}C) = 2.01 ppm), we state that its structure (and that of **FURSEM-NEW**) is verified using NMR crystallography; however, we are unable to verify the structure of **FURSEM17**, as both associated RMSD values are quite large (RMSD(^1H) = 0.77 ppm and RMSD(^{13}C) = 2.78 ppm, Fig. 3(a and b)). The largest contributions to the ^1H RMSD value of **FURSEM17** (ESI, \dagger) are due to the ^1H shift value for the COOH hydrogen from one of the two furosemide molecules (see the variation in the computed values in the spectra provided in Fig. 3c). This is consistent with the previously highlighted structural difference between **FURSEM01** and **FURSEM17**. Omitting this datum from the ^1H RMSD calculations significantly reduces the calculated ^1H RMSD value for **FURSEM17** (Fig. 3a, red columns).

Being unable to verify a crystal structure using NMR crystallography does not infer a structure is incorrect. Similarly, the approach we outline should complement existing verification methods, and does not serve as a replacement. We consider additional computational information to assess the likelihood that **FURSEM17** is an isolable polymorph of furosemide. From the dc-DFT structure verification method of van de Streek and Neumann, 16 **FURSEM01** is validated, as its non-H atomic positions, when subjected to a dc-DFT calculation that optimises the unit cell and all atoms, do not change significantly (non-H atomic RMSD for one unit cell = 0.093 \AA , where deviations greater than 0.25 \AA are considered significant). From a dc-DFT geometry optimisation, **FURSEM17** is found to be close to a local energy minimum and hence a theoretically viable furosemide polymorph. However, the non-H atomic RMSD after optimisation is 0.327 \AA over the unit cell, and so **FURSEM17** should be flagged for closer inspection. NMR crystallography provides an obvious way to perform such an inspection according to the above dc-DFT protocol.

Finally, the DFT-calculated enthalpies for the fully-optimised structures of **FURSEM01** and **FURSEM17** are distinct, with the



optimised structure of **FURSEM17** being *ca.* 25 kJ mol⁻¹ molecule⁻¹ above **FURSEM01** (ESI[†]). Based upon recent findings of the crystal structure prediction (CSP) community,^{40,41} such a high energy structure would be very unlikely to occur under typical conditions. Indeed, this would establish a new record for an energy difference between two non-conformational polymorphic forms by a margin of 7 kJ mol⁻¹ molecule⁻¹.

In conclusion, we have demonstrated the potential value of NMR crystallography for the verification of crystal structures. We verify the structure of **FURSEM01** *via* NMR crystallography, and show that this conclusion is consistent with other methods, such as energy calculations using dispersion-corrected DFT. However, for the crystal structure of **FURSEM17**, the DFT-computed chemical shift data is inconsistent with NMR experiments, and hence its structure is not verified. This conclusion is consistent with current protocols from the CSP community, and also when using the method of van de Streek and Neumann.¹⁶ We are also able to distinguish between the **FURSEM01** and **FURSEM17** structures with *ca.* 2σ confidence, which is noteworthy as the structures are essentially identical save for the position of one hydrogen atom. While the diffraction community has many available structure verification tools, the ability to ensure the quality of crystal structures in databases in a way which is independent of the chosen experimental method is potentially important, as these structures are often starting points when screening for new pharmaceuticals, testing computational methods, and designing new materials. We are refining and extending this protocol as part of a systematic study on how to verify crystal structures using NMR crystallography tools.

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Notes and references

† CCDC 1458460.

§ These values were established by benchmarking *ca.* 30 molecular systems of known correct structure using the GIPAW DFT method in CASTEP. The grey banded areas correspond to regions equal to ±1σ about the mean values (dashed lines). Details for these systems are in preparation for submission.

- 1 K. Hashi, S. Ohki, S. Matsumoto, G. Nishijima, A. Goto, K. Deguchi, K. Yamada, T. Noguchi, S. Sakai, M. Takahashi, Y. Yanagisawa, S. Iguchi, T. Yamazaki, H. Maeda, R. Tanaka, T. Nemoto, H. Suematsu, T. Miki, K. Saito and T. Shimizu, *J. Magn. Reson.*, 2015, **256**, 30–33.
- 2 V. Agarwal, S. Penzel, K. Szekely, R. Cadalbert, E. Testori, A. Oss, J. Past, A. Samoson, M. Ernst, A. Böckmann and B. H. Meier, *Angew. Chem., Int. Ed.*, 2014, **53**, 12253–12256.

- 3 R. Zhang, M. K. Pandey, Y. Nishiyama and A. Ramamoorthy, *Sci. Rep.*, 2015, **5**, 11810.
- 4 *NMR Crystallography*, ed. R. K. Harris, R. E. Wasylshen and M. J. Duer, John Wiley & Sons Ltd, Chichester, West Sussex, United Kingdom, 2009.
- 5 C. Martineau, J. Senker and F. Taulelle, *Annu. Rep. NMR Spectrosc.*, 2014, **82**, 1–57.
- 6 C. Martineau, *Solid State Nucl. Magn. Reson.*, 2014, **63–64**, 1–12.
- 7 C. Bonhomme, C. Gervais, F. Babonneau, C. Coelho, F. Pourpoint, T. Azais, S. E. Ashbrook, J. M. Griffin, J. R. Yates, F. Mauri and C. J. Pickard, *Chem. Rev.*, 2012, **112**, 5733–5779.
- 8 T. Charpentier, *Solid State Nucl. Magn. Reson.*, 2011, **40**, 1–20.
- 9 M. Baías, C. M. Widdifield, J.-N. Dumez, H. P. G. Thompson, T. G. Cooper, E. Salager, S. Bassil, R. S. Stein, A. Lesage, G. M. Day and L. Emsley, *Phys. Chem. Chem. Phys.*, 2013, **15**, 8069–8080.
- 10 M. Baías, J.-N. Dumez, P. H. Svensson, S. Schantz, G. M. Day and L. Emsley, *J. Am. Chem. Soc.*, 2013, **135**, 17501–17507.
- 11 E. Salager, G. M. Day, R. S. Stein, C. J. Pickard, B. Elena and L. Emsley, *J. Am. Chem. Soc.*, 2010, **132**, 2564–2566.
- 12 T. Sumner, *Science*, 2014, **343**, 1092–1093.
- 13 B. H. Toby, *Powder Diffr.*, 2006, **21**, 67–70.
- 14 Y. L. Slovokhotov, *Cryst. Growth Des.*, 2014, **14**, 6205–6216.
- 15 J. van de Streek, *Acta Crystallogr., Sect. B: Struct. Sci.*, 2006, **B62**, 567–579.
- 16 J. van de Streek and M. A. Neumann, *Acta Crystallogr., Sect. B: Struct. Sci.*, 2010, **B66**, 544–558.
- 17 T. Pawlak and M. J. Potrzebowski, *J. Phys. Chem. B*, 2014, **118**, 3298–3309.
- 18 M. Kibalchenko, D. Lee, L. Shao, M. C. Payne, J. J. Titman and J. R. Yates, *Chem. Phys. Lett.*, 2010, **498**, 270–276.
- 19 R. K. Harris, P. Hodgkinson, V. Zorin, J.-N. Dumez, B. Elena-Herrmann, L. Emsley, E. Salager and R. S. Stein, *Magn. Reson. Chem.*, 2010, **48**, S103–S112.
- 20 www.who.int/medicines/publications/essentialmedicines.
- 21 K. M. Ho and B. M. Power, *Anaesthesia*, 2010, **65**, 283–293.
- 22 F. Andreasen, O. L. Pedersen and E. Mikkelsen, *Eur. J. Clin. Pharmacol.*, 1978, **14**, 237–244.
- 23 H. E. Kerr, L. K. Softley, K. Suresh, A. Nangia, P. Hodgkinson and I. R. Evans, *CrystEngComm*, 2015, **17**, 6707–6715.
- 24 N. R. Goud, S. Gangavaram, K. Suresh, S. Pal, S. G. Manjunatha, S. Nambiar and A. Nangia, *J. Pharm. Sci.*, 2012, **101**, 664–680.
- 25 B. I. Harriss, L. Vella-Zarb, C. Wilson and I. R. Evans, *Cryst. Growth Des.*, 2014, **14**, 783–791.
- 26 T. Ueto, N. Takata, N. Muroyama, A. Nedu, A. Sasaki, S. Tanida and K. Terada, *Cryst. Growth Des.*, 2012, **12**, 485–494.
- 27 C. Garnerio, A. K. Chattah and M. Longhi, *Carbohydr. Polym.*, 2013, **94**, 292–300.
- 28 J. Lamotte, H. Campsteyn, L. Dupont and M. Vermeire, *Acta Crystallogr.*, 1978, **B34**, 1657–1661.
- 29 O. Bolukbasi and A. Yilmaz, *Vib. Spectrosc.*, 2012, **62**, 42–49.
- 30 J. van de Streek and S. Motherwell, *Acta Crystallogr., Sect. B: Struct. Sci.*, 2005, **B61**, 504–510.
- 31 S. Karami, Y. Li, D. S. Hughes, M. B. Hursthouse, A. E. Russell, T. L. Threlfall, M. Claybourn and R. Roberts, *Acta Crystallogr., Sect. B: Struct. Sci.*, 2006, **B62**, 689–691.
- 32 N. J. Babu, S. Cherukuvada, R. Thakuria and A. Nangia, *Cryst. Growth Des.*, 2010, **10**, 1979–1989.
- 33 X. Filip and C. Filip, *Solid State Nucl. Magn. Reson.*, 2015, **65**, 21–28.
- 34 Q. Z. Ni, E. Daviso, T. V. Can, E. Markhasin, S. K. Jawla, T. M. Swager, R. J. Temkin, J. Herzfeld and R. G. Griffin, *Acc. Chem. Res.*, 2013, **46**, 1933–1941.
- 35 A. J. Rossini, A. Zagdoun, M. Lelli, A. Lesage, C. Copéret and L. Emsley, *Acc. Chem. Res.*, 2013, **46**, 1942–1951.
- 36 S. J. Clark, M. D. Segall, C. J. Pickard, P. J. Hasnip, M. I. J. Probert, K. Refson and M. C. Payne, *Z. Kristallogr.*, 2005, **220**, 567–570.
- 37 C. J. Pickard and F. Mauri, *Phys. Rev. B: Condens. Matter Mater. Phys.*, 2001, **63**, 245101.
- 38 J. R. Yates, C. J. Pickard and F. Mauri, *Phys. Rev. B: Condens. Matter Mater. Phys.*, 2007, **76**, 024401.
- 39 M. Dračinský and P. Hodgkinson, *CrystEngComm*, 2013, **15**, 8705–8712.
- 40 H. P. G. Thompson and G. M. Day, *Chem. Sci.*, 2014, **5**, 3173–3182.
- 41 J. Nyman and G. M. Day, *CrystEngComm*, 2015, **17**, 5154–5165.

