

## REVIEW

View Article Online

View Journal | View Issue



Cite this: *Mater. Chem. Front.*, 2021,  
5, 2173

# A brief review on novel pyrene based fluorometric and colorimetric chemosensors for the detection of Cu<sup>2+</sup>

Zannatul Kowser,<sup>ab</sup> Ummey Rayhan,<sup>ac</sup> Thamina Akther,<sup>a</sup> Carl Redshaw<sup>d</sup> and Takehiko Yamato<sup>id</sup> <sup>★a</sup>

The development of colorimetric and fluorometric chemosensors that capable of detecting Cu<sup>2+</sup> ions by a change in colour and fluorescence intensity has been described. Herein, chemosensors having pyrene functional groups as a signaling moiety are discussed in detail as pyrene derivatives show significant photophysical properties being superior to those of other commonly used scaffolds. This review article provides a detailed overview of pyrene containing chemosensors based on fluorescence mechanisms, such as excimer/excimer formation, photoinduced electron transfer (PET), photoinduced charge transfer (PCT), aggregation induced emission (AIE), ligand to metal charge transfer process (LMCT), chelation enhanced quenching mechanism (CHEQ), Cu<sup>2+</sup>-selective reactions for the selective and sensitive detection of Cu<sup>2+</sup>. Potential future applications are also discussed because of the fact Cu<sup>2+</sup> ion recognition has a great significance in the biological, environmental and medical sectors.

Received 2nd December 2020,  
Accepted 11th January 2021

DOI: 10.1039/d0qm01008a

rsc.li/frontiers-materials

## 1. Introduction

The development of fluorescent sensors has received increasing attention in recent years due to their simplicity and low detection limits.<sup>1</sup> For the determination of copper ions, various techniques such as atomic absorption spectrometry,<sup>2</sup> inductively coupled plasma mass spectrometry (ICPMS),<sup>3</sup> inductively coupled plasma atomic emission spectrometry (ICP-AES),<sup>4</sup> and voltammetry<sup>5</sup> and anodic stripping voltammetry (ASV)<sup>6</sup> have been applied. However, these methods cause operational difficulties, requiring well-trained personnel and high-cost analytical instrumentation. These techniques also lead to the destruction of the cells of biological samples and cannot be used to visualize biological phenomena *in situ*.<sup>7</sup> For this reason, fluorescent sensors have emerged as a useful technique for high sensitivity, simplicity and for the non-destructive imaging of intracellular distribution for the detection of cations, anions and neutral species that are of great importance in the biological and environmental sectors.<sup>8–10</sup>

Chemosensors are molecules of abiotic origin, which show changes of one or more properties such as colour

(colorimetric chemosensors), fluorescence (fluorescent chemosensors) or redox potentials (electrochemical sensors) upon interaction with guest species.<sup>1,11–13</sup> These sensors are mainly organic molecules, and may be categorized into four classes: (i) as fluorescent ligands which have identical receptor (the recognition site) and fluorophore (the signal source) (ii) as fluoroionophores where the fluorophore and receptor are involved in direct electronic conjugation, (iii) fluoroionophores combined *via* fluorophore-spacer-receptor systems, (iv) excimer or excimer forming probes (EPs) where, the fluorophore and receptor units can construct an intramolecular excimer or excimer (Fig. 1). Herein, strong intramolecular geometry changes are observed after binding with the analyte by increasing or decreasing the ratio of excimer-to-monomer emission.<sup>14,15</sup> To be an ideal fluorescent chemosensor, the receptor must have the strongest affinity for the relevant target (binding-selectivity) and the fluorescence signal should avoid any environmental interference (signal-selectivity).<sup>16</sup>

Among the heavy and transition metal ions, copper is one of the important trace elements for both plants and animals, including humans.<sup>17</sup> When levels of Cu<sup>2+</sup> exceeds cellular needs, it can be considered as toxic to biological systems.<sup>18</sup> The US Environmental Protection Agency (EPA) has set the limit of 1.3 ppm (~20 μM) for copper in drinking water.<sup>14</sup> For this reason, improved fluorescent chemosensors for the selective and sensitive detection of Cu<sup>2+</sup> are of great importance.<sup>19–22</sup> It is noteworthy that Cu<sup>2+</sup> ion detection poses some challenges when designing fluorescence turn-on (fluorescence intensity increases)

<sup>a</sup> Department of Applied Chemistry, Faculty of Science and Engineering, Saga University, Honjo-machi 1, Saga 840-8502, Japan  
E-mail: yamatot@cc.saga-u.ac.jp

<sup>b</sup> Department of Chemistry, Faculty of Science, Jashore University of Science and Technology, Jashore-7408, Bangladesh

<sup>c</sup> Department of Chemistry, Dhaka University of Engineering & Technology, Gazipur-1700, Bangladesh

<sup>d</sup> Department of Chemistry, The University of Hull, HU6 7RX, UK

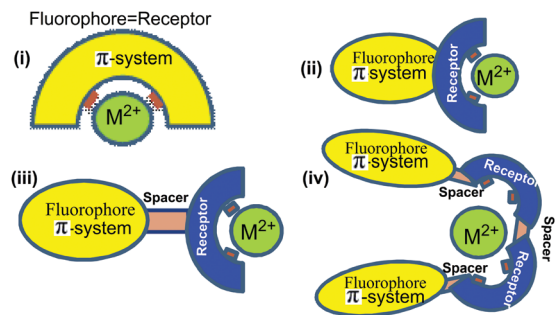


Fig. 1 Schematic presentation of fluorosensors with metal ions. (i) Fluorescent ligand (ii) intrinsic fluorescent probe (iii) fluorophore-spacer-receptor system (iv) exciplex or excimer forming probe.

sensors due to its paramagnetic nature with an unfilled d orbital. Moreover, according to the Irving-Williams rule,  $\text{Cu}^{2+}$  has the strongest binding ability *versus* any other divalent metal ion of the first transition series. The paramagnetic cations  $\text{Cu}^{2+}$ ,  $\text{Ni}^{2+}$  and  $\text{Co}^{2+}$  are usually more strongly bound than the diamagnetic ions  $\text{Zn}^{2+}$  or  $\text{Cd}^{2+}$ , which is determined on the basis of the ionic radius and the second ionization potential.<sup>14,23</sup> Therefore, strong fluorescence “turn-off” (fluorescence intensity decreases) sensors were predominantly observed due to fluorescence quenching, upon addition of  $\text{Cu}^{2+}$ .<sup>24,25</sup> Recently, many turn-on sensors have been studied for  $\text{Cu}^{2+}$  detection by using the concept of ion-induced changes in the geometry or the flexibility of the ligand as well as from the availability of certain functional groups involved in fluorescence quenching for the ligand in the unbound state.<sup>14</sup> In this review, we discuss the sensing mechanisms of  $\text{Cu}^{2+}$  with pyrene based fluorescent sensors. This will help shape the design of new pyrene chemosensors for copper ion determination based on different mechanisms such as excimer/exciplex formation, photo-induced electron transfer (PET), photoinduced charge transfer (PCT), aggregation induced emission (AIE), ligand to metal charge transfer process (LMCT), chelation enhanced quenching mechanism (CHEQ) and  $\text{Cu}^{2+}$ -selective reactions.

## 2. The sources, applications, effects of $\text{Cu}^{2+}$

### 2.1. Sources of copper

Copper is the third most abundant essential transition metal ion after iron and zinc in the human body.<sup>26</sup> It is found in both natural sources and all body tissues. In the body, the liver, brain, heart, kidneys, and skeletal muscle contain the maximum amount of copper. High doses of copper are found in a commercially available multivitamins. Moreover, naturally rich sources of copper are exhibited in oysters, sesame seeds, tahini, cocoa powder, chocolate, nuts, calamari, lobster, sunflower seeds, sun dried tomatoes, roasted pumpkin, squash seeds and dried herbs, *etc.* Additionally, because of copper plumbing, it is present in water. In most of the vegetarian diets, copper is present in the food items.<sup>27</sup>

### 2.2. Applications of copper

Copper is an important trace element for human metabolism and plays a vital role in the physiological processes of organisms. It contributes to the formation of red blood cells and in maintaining nerve cells and the immune system. It also helps to generate energy in the body and plays a significant role for collagen production and the absorption of iron. Copper intake also reduces the chance of cardiovascular disease and osteoporosis. Therefore, for normal development and proper working of the brain and as a cofactor of many enzymes, the presence of copper is essential.<sup>27</sup>

### 2.3. Deficiency and toxicity of copper

Copper present at low or high concentrations can increase the risk to health. Free  $\text{Cu}^{2+}$  ion is both acutely and chronically toxic for aquatic life and microorganisms, even at micromolar concentrations. Copper deficiency is associated with growth failure and can affect deterioration of the nervous system. However, an excessive accumulation of copper may lead to neurodegenerative disorders, abnormalities in red blood cells and heart problems. Gene mutations are responsible for the two major genetic disorders of copper metabolism in humans such as Menkes' disease and Wilson's disease, and it is found that these diseases are the result of excessive intracellular copper transport. Increased serum copper levels have been linked to a higher risk of cardiovascular disease. Different diseases, including Alzheimer's disease, Indian childhood cirrhosis (ICC) and prion disease are also associated with the toxicity of  $\text{Cu}^{2+}$ . Besides, copper is one of the most common metal pollutants because of its huge applications in our daily lives. For adults (19 years and above), the upper limit is 10 mg a day, above this it is considered toxic. Moreover,  $\text{Cu}^{2+}$  ions are often essential components in biochemical reactions such as catalysis, transport or biosynthesis at trace levels ( $<1 \mu\text{M}$ ). However, in the presence of large amounts of  $\text{Cu}^{2+}$ , unhealthy interactions occur in biochemical redox processes and can, for instance result in the inhibition of enzyme activity or nephrotoxicity.<sup>27,28</sup> Therefore, it is necessary to design a technique that is effective for a rapid response toward  $\text{Cu}^{2+}$  even at micromolar concentrations. The design and synthesis of fluorescent chemosensors for copper ion determination could help resolve these issues.

## 3. Pyrene and pyrene derivatives as fluorophore

Pyrene and pyrene derivatives have interesting photophysical properties notably a long lifetime, high quantum yield ( $\tau_{\text{M}} = 450 \text{ ns}$  and  $\Phi_{\text{M}} = 0.60$  in cyclohexane) and expanded  $\pi$  electron delocalization of the pyrene monomers. Pyrene derivatives generally display five well-shaped and fine absorption bands between 210 and 330 nm and the emission spectra usually consist of a broad band centered at 400 nm. Fluorescence quenching of a pyrene fluorophore can easily occur in the presence of a variety of quenchers (heavy metals, anionic species and NPs), which may be due to through-bond and through-space energy transfer between pyrene and the quenchers.<sup>29</sup> A pyrene based

fluorescent “off-on” chemosensor has been developed for detection of cyanide *via* ligand to  $\text{Cu}^{2+}$  complex as  $\text{Cu}^{2+}$  generally performs as a fluorescence quencher *via* a PET mechanism.<sup>30</sup> Pyrene also can perform as either an electron donor or acceptor during the energy transfer depending on the substituents attached to it, and as a result blue or red shifted emission bands are easily observed from the pyrene monomer emission band. One of the most interesting features of pyrene derivatives is that a pyrene monomer ( $\lambda_{\text{abs}} = 350 \text{ nm}$ ,  $\lambda_{\text{em}} = 398 \text{ nm}$ ) can combine with another pyrene monomer to form an excimer (or excited state dimer,  $\lambda_{\text{em}} = 485 \text{ nm}$ ) because of its affinity for strong  $\pi$ - $\pi$  interactions.<sup>31</sup> The changes in the emission properties of the pyrene excimer from a monomer has been used for designing sensors.<sup>32–34</sup> A pyrene-based triazole ligand was reported which exhibited self-assembly in the presence of  $\text{ZnCl}_2$ . The free ligand showed monomer emission bands at 382 and 402 nm with a broad shoulder at 420 nm. After addition of  $\text{ZnCl}_2$ , it can bind with  $\text{ZnCl}_2$  in a 2 : 1 ratio due to strong excimer emission at 410 nm.<sup>35</sup>

## 4. Mechanism of $\text{Cu}^{2+}$ sensing

Fluorescent chemosensors for  $\text{Cu}^{2+}$  detection have been designed based on different binding mechanisms which fulfill the criteria of affinity, selectivity and sensitivity (Table 1). In general, the chemosensor is designed such that it contains oxygen or nitrogen donor atoms for co-ordination of the  $\text{Cu}^{2+}$ .<sup>36</sup> In most of the cases, fluorescent sensors bind  $\text{Cu}^{2+}$  by the fluorescence quenching process due to its paramagnetic nature which opens up the excited state de-excitation pathways by enhancing the rate of non-radioactive processes such as electronic energy transfer (EET) and/or PET process involving the metal centers. Also, the free electron in the orbital is likely to quench the fluorescence *via* spin-orbit coupling.<sup>37</sup> There is also an enhancement of the fluorescence intensity in turn-on sensors, which are more desirable than turn-off sensors, with a red or blue shift of the emission after  $\text{Cu}^{2+}$  ion binding with ligand. Recently, some new strategies have been developed for fluorescence off-on sensors for  $\text{Cu}^{2+}$  detection. The classical sensing mechanisms such as PET,<sup>38,39</sup> PCT,<sup>40,41</sup> excimer/exciplex formation<sup>42</sup> are mainly mentioned when investigating binding phenomenon. There are also some mechanisms which control the response of a fluorophore to substrate binding including AIE, LMCT, CHEQ and  $\text{Cu}^{2+}$ -assisted reactions.

### 4.1. Excimer emission mechanism

When an excited fluorophore during its lifetime in an excited state interacts with a ground state fluorophore, an excimer is formed.<sup>43</sup> Excimer emission typically shows a red shifted broad fluorescence band in most aromatic molecules,<sup>44</sup> by about  $6000 \text{ cm}^{-1}$  to lower energies than the uncomplexed (“monomer”) fluorophore emission. There are two kinds of excimers based on the origin of the pyrene dimer: a dynamic excimer and a static excimer. The dynamic or static excimer formation depends on the distance between the two pyrene units (Fig. 2).<sup>45</sup> The conventional dynamic excimers are produced due

to the interaction of the diffused ground state fluorophore with the electronic excited state fluorophore during its lifetime within van der Waals contact distances. On the other hand, in case of the static excimer, initially a pyrene dimer is formed in the ground state. After photo-excitation, the monomer units interact with each other at very close distances (3.5–3.9 Å in the case of polycyclic hydrocarbons) to form the excimer. Importantly, metal ion co-ordination with the ligand controls the separation and relative orientation of multiple fluorophore units attached to the ligands. For this reason, cation recognition is investigated by the monomer to excimer fluorescence intensity ratio.<sup>1,46–50</sup>

### 4.2. Photoinduced electron transfer (PET)

In the PET process, an excited electron is transferred from the donor to the acceptor. These ideas can also be explained with the help of molecular orbital energy diagrams when the molecular device is non-luminescent and luminescent (Fig. 3). The emission of photons from LUMO to HOMO after excitation of electrons is called fluorescence and the molecule is termed as a fluorophore. However, PET occurs when one electron from the HOMO of the electron donor is transferred efficiently to the hole in the HOMO of the fluorophore and the initially excited electron of the LUMO moves to the HOMO of the electron donor. This event happens if the HOMO of the electron donor is higher in energy than the singly occupied orbital of the fluorophore HOMO. The PET mechanism leads to nonradiative deactivation of the excited state, and as a result the emission intensity is decreased or “quenching” of fluorescence is observed.<sup>38,51–55</sup> However, after interaction with metal ions, the donor electron containing orbital shifts its position from higher energy to lower energy and inhibits the electron-transfer process, resulting a radiative emission. In the case of metal ion binding, this effect is referred to as chelation-enhanced fluorescence (CHEF). It is observed that in detection of  $\text{Cu}^{2+}$  by the PET method, the pyrene containing chemosensor mostly exhibits a weak fluorescence due to fluorescence quenching from the nitrogen lone pairs onto pyrene. The binding of  $\text{Cu}^{2+}$  with the ligand prevents the PET mechanism, resulting in a significant enhancement in pyrene fluorescence.<sup>56,57</sup>

### 4.3. Photoinduced charge transfer (PCT)

The chemosensors based on PCT or ICT are designed in which an electron withdrawing unit is conjugated with electron-donating substituents in the same molecule (Fig. 4). Therefore, the “push-pull”  $\pi$  electron system of the chemosensor occurs in the excited state<sup>58,59</sup> and the charge transfer can occur over long distances through conjugation and is related with major dipole moment changes. This phenomenon is used for cation sensing, as the close interaction of cations with the donor or the acceptor moiety affects the photophysical properties of the fluorophore.<sup>60</sup> Cation complexation of an electron donor group within a fluorophore decreases the electron-donating character of the donor group and produces a blue shift in the spectrum due to reduction of conjugation. On the other hand, a red-shifted absorption spectrum is observed for metal ion binding to the

Table 1 Comparison of important features of reported Cu<sup>2+</sup> sensors discussed above

| Probe no. | Solvent medium                                                 | Mechanism                                     | Association constant  | Limit of detection     | $\lambda_{\text{ex}}/\lambda_{\text{em}}$ (nm) | Stoichiometry (sensor/Cu <sup>2+</sup> ) | Application                                        | Ref. |
|-----------|----------------------------------------------------------------|-----------------------------------------------|-----------------------|------------------------|------------------------------------------------|------------------------------------------|----------------------------------------------------|------|
| 1         | EtOH                                                           | Monomer and excimer emission                  | $3.5 \times 10^5$     | $1.44 \times 10^{-7}$  | 344/379, 397, 484                              | 1:1                                      | NA                                                 | 57   |
| 2         | CH <sub>3</sub> CN/H <sub>2</sub> O                            | Monomer and excimer emission                  | NA                    | $1.87 \times 10^{-8}$  | 343/396, 485                                   | 1:1                                      | NA                                                 | 58   |
| 3         | CH <sub>3</sub> CN/CH <sub>2</sub> Cl <sub>2</sub>             | Monomer and excimer emission                  | $3.57 \times 10^5$    | NA                     | 367/415, 518                                   | 1:1                                      | NA                                                 | 59   |
| 4         | Tris-HNO <sub>3</sub> buffer solution                          | Monomer and excimer emission                  | $2.18 \times 10^4$    | $2 \times 10^{-8}$     | 342/375, 460                                   | 2:1                                      | Environmental samples, live cells (HeLa cells)     | 60   |
| 5         | Dioxane                                                        | Monomer and excimer emission                  | NA                    | NA                     | 360/420                                        | 1:1                                      | NA                                                 | 61   |
| 6         | Dioxane                                                        | Monomer and excimer emission                  | $3.10 \times 10^{-2}$ | NA                     | 360/420, 500                                   | 1:1                                      | NA                                                 | 61   |
| 7         | CH <sub>2</sub> Cl <sub>2</sub>                                | Dynamic excimer to static excimer             | 4.4                   | NA                     | 335/440                                        | 1:1                                      | NA                                                 | 62   |
| 8         | CH <sub>3</sub> CN/H <sub>2</sub> O                            | Static excimer emission                       | $2.8 \times 10^4$     | NA                     | 342/375, 455                                   | 2:1                                      | NA                                                 | 63   |
| 9         | CH <sub>3</sub> CN                                             | Static excimer emission                       | $5.42 \times 10^5$    | NA                     | 360/388, 460                                   | 2:1                                      | NA                                                 | 64   |
| 10        | CH <sub>3</sub> CN                                             | NA                                            | NA                    | NA                     | NA                                             | NA                                       | NA                                                 | 64   |
| 11        | CH <sub>3</sub> CN                                             | NA                                            | NA                    | NA                     | NA                                             | NA                                       | NA                                                 | 64   |
| 12        | Tris-HCl buffer containing CH <sub>3</sub> CN/H <sub>2</sub> O | Static excimer emission                       | $4.583 \times 10^3$   | $4 \times 10^{-8}$     | 350/452                                        | 2:1                                      | Drinking water                                     | 65   |
| 13        | CH <sub>3</sub> CN                                             | Excimer emission                              | $1.96 \times 10^6$    | $9.72 \times 10^{-7}$  | 395/455                                        | 2:1                                      | NA                                                 | 66   |
| 14        | CH <sub>3</sub> CN                                             | Dynamic excimer to static excimer             | 65 600                | NA                     | 342/447                                        | 2:1                                      | NA                                                 | 67   |
| 15        | EtOH/H <sub>2</sub> O                                          | Excimer to monomer emission                   | 11.53                 | $40 \times 10^{-9}$    | 340/378, 466                                   | 1:2                                      | NA                                                 | 68   |
| 16        | CH <sub>3</sub> OH/H <sub>2</sub> O                            | Excimer switch-off                            | $4.8 \times 10^6$     | NA                     | 410/466, 520–560                               | 1:1                                      | Live cell (HEK 293 cells)                          | 69   |
| 17        | Mixed liposomes                                                | Monomer and excimer emission                  | NA                    | NA                     | 342/395, 470                                   | NA                                       | NA                                                 | 70   |
| 18        | Mixed liposomes                                                | Monomer and excimer emission                  | NA                    | NA                     | 342/395, 471                                   | NA                                       | NA                                                 | 70   |
| 19        | Mixed liposomes                                                | Monomer and excimer emission                  | NA                    | NA                     | 342/395, 472                                   | NA                                       | NA                                                 | 70   |
| 20        | Mixed liposomes                                                | Monomer and excimer emission                  | NA                    | NA                     | 342/395, 473                                   | NA                                       | NA                                                 | 70   |
| 21        | CH <sub>3</sub> CN                                             | Monomer and excimer emission                  | $1.89 \times 10^5$    | NA                     | 343/482                                        | 1:1                                      | NA                                                 | 71   |
| 22        | CH <sub>3</sub> CN/CH <sub>2</sub> Cl <sub>2</sub>             | PET                                           | $1.29 \times 10^5$    | $8.80 \times 10^{-8}$  | 367/405                                        | 1:1                                      | NA                                                 | 72   |
| 23        | CH <sub>3</sub> CN/CH <sub>2</sub> Cl <sub>2</sub>             | PET                                           | $1.55 \times 10^4$    | $4.94 \times 10^{-7}$  | 367/405                                        | 1:1                                      | NA                                                 | 72   |
| 24        | CH <sub>3</sub> CN/CH <sub>2</sub> Cl <sub>2</sub>             | PET                                           | NA                    | NA                     | 367/405                                        | 1:1                                      | NA                                                 | 72   |
| 25        | CH <sub>3</sub> CN/H <sub>2</sub> O                            | PET                                           | $5 \times 10^8$       | $2.73 \times 10^{-6}$  | 385/468                                        | 2:1                                      | Live cell (RAW 264.7 cells)                        | 73   |
| 26        | CH <sub>3</sub> CN                                             | PET                                           | $5.71 \times 10^5$    | $3.91 \times 10^{-6}$  | 350/429                                        | 1:1                                      | NA                                                 | 74   |
| 27        | CH <sub>3</sub> OH/H <sub>2</sub> O                            | PET                                           | $2.75 \times 10^3$    | NA                     | 360/455                                        | 1:1                                      | Live cell                                          | 75   |
| 28        | CH <sub>3</sub> OH/H <sub>2</sub> O                            | PET                                           | $5.55 \times 10^3$    | NA                     | 350/417                                        | 1:1                                      | NA                                                 | 76   |
| 29        | CH <sub>3</sub> CN/H <sub>2</sub> O                            | PET                                           | $1.0 \times 10^4$     | $0.04 \times 10^{-6}$  | 346/389                                        | 1:1                                      | Live cell (RAW 264.7 cells)                        | 77   |
| 30        | DMSO-H <sub>2</sub> O                                          | PET                                           | $1.16 \times 10^4$    | $0.26 \times 10^{-6}$  | 393/463                                        | 1:1                                      | Live cell (RAW 264.7 cells)                        | 78   |
| 31        | CH <sub>3</sub> CN/H <sub>2</sub> O                            | PET                                           | NA                    | $7.8 \times 10^{-9}$   | 305/444                                        | 1:1                                      | Live cell (HeLa cells), real samples               | 79   |
| 32        | CH <sub>3</sub> CN                                             | PET                                           | $7.74 \times 10^6$    | $4.5 \times 10^{-6}$   | 350/388, 409, 473                              | 1:1                                      | NA                                                 | 80   |
| 33        | CH <sub>3</sub> CN                                             | PET                                           | NA                    | $3.60 \times 10^{-7}$  | 340/444                                        | 1:1                                      | NA                                                 | 81   |
| 34        | CH <sub>3</sub> CN                                             | PCT                                           | NA                    | NA                     | 343/470                                        | 1:1                                      | NA                                                 | 82   |
| 35        | DMF/HEPES buffer                                               | LMCT                                          | NA                    | $8.5 \times 10^{-6}$   | 305/370                                        | 1:1                                      | NA                                                 | 84   |
| 36        | CH <sub>3</sub> CN/H <sub>2</sub> O                            | LMCT                                          | $5.65 \times 10^5$    | $0.503 \times 10^{-6}$ | 384/429                                        | 2:2                                      | Real samples                                       | 85   |
| 37        | DMSO/H <sub>2</sub> O                                          | ILCT                                          | $6.5789 \times 10^4$  | $6.865 \times 10^{-8}$ | 370/390, 412                                   | 2:1                                      | Real samples                                       | 86   |
| 38        | H <sub>2</sub> O/DMF                                           | AEE                                           | $1.89 \times 10^9$    | $35 \times 10^{-9}$    | 370/455                                        | 2:1                                      | Live cell (HeLa cells)                             | 89   |
| 39        | CH <sub>3</sub> CN, CH <sub>3</sub> CN/H <sub>2</sub> O        | PET, AIE                                      | NA                    | $2.5 \times 10^{-6}$   | 393/440, 454.5, 509                            | 1:1                                      | Antioxidant property                               | 90   |
| 40        | 10% aqueous Tris-buffered DMSO solution                        | Cu <sup>2+</sup> induced catalytic hydrolysis | NA                    | $5.93 \times 10^{-8}$  | 340/392                                        | NA                                       | Simulated semiconductor waste water                | 92   |
| 41        | CH <sub>3</sub> CN/HEPES buffer solution                       | Cu <sup>2+</sup> induced ring opening process | $2.5 \times 10^4$     | NA                     | 520/575                                        | 1:1                                      | NA                                                 | 94   |
| 42        | DMSO/H <sub>2</sub> O                                          | Paramagnetic nature, CHEQ process             | $2.3 \times 10^5$     | $0.73 \times 10^{-9}$  | 440/510                                        | 1:1                                      | Live cell (A549 cells) and real water samples      | 96   |
| 43        | CH <sub>3</sub> OH solution                                    | Heavy atom effect                             | $1.48 \times 10^5$    | $10^{-6}$ M            | 340/392                                        | 1:1                                      | NA                                                 | 97   |
| 44        | Aqueous HEPES buffer (H <sub>2</sub> O/CH <sub>3</sub> CN)     | Paramagnetic nature                           | $2.4 \times 10^4$     | NA                     | 353/435                                        | NA                                       | Live cell (COS-7 kidney cells), biological samples | 98   |

Table 1 (continued)

| Probe no. | Solvent medium                         | Mechanism                        | Association constant | Limit of detection    | $\lambda_{ex}/\lambda_{em}$ , (nm) | Stoichiometry (sensor/ $Cu^{2+}$ ) | Application           | Ref. |
|-----------|----------------------------------------|----------------------------------|----------------------|-----------------------|------------------------------------|------------------------------------|-----------------------|------|
| 45        | Aqueous HEPES buffer ( $H_2O/CH_3CN$ ) | NA                               | NA                   | NA                    | NA                                 | NA                                 | NA                    | 98   |
| 46        | $CH_2Cl_2/DMSO$                        | Paramagnetic nature, reverse PET | $3.3 \times 10^5$    | NA                    | 343/393, 486                       | 1:1                                | NA                    | 99   |
| 47        | $CH_2Cl_2/CH_3CN$                      | Reverse PET                      | 4.939                | NA                    | 342/377                            | 1:1                                | NA                    | 101  |
| 48        | $CH_2Cl_2/CH_3CN$                      | Monomer and excimer emission     | 10.3086              | NA                    | 342/375, 467                       | 1:2                                | NA                    | 101  |
| 49        | $DMSO/H_2O$                            | $Cu^{2+}$ -Assisted-complexation | NA                   | $2.17 \times 10^{-6}$ | NA                                 | 1:1                                | Environmental systems | 102  |
| 50        | $DMSO/H_2O$                            | $Cu^{2+}$ induced complexation   | $1.09 \times 10^4$   | $1.0 \times 10^{-4}$  | 348/396, 439                       | 1:1                                | NA                    | 103  |
| 51        | $CH_3CN/H_2O$                          | Inhibition of ICT process        | NA                   | NA                    | NA                                 | NA                                 | NA                    | 104  |
| 52        | $CH_3CN/H_2O$                          | Inhibition of ICT process        | $5.2 \times 10^3$    | NA                    | 340/420, 590                       | NA                                 | NA                    | 104  |
| 53        | $CH_3CN/HEPES$ buffer                  | CHEF and ICT process             | NA                   | $1.21 \times 10^{-6}$ | 330/414                            | 1:1                                | Live cell             | 105  |

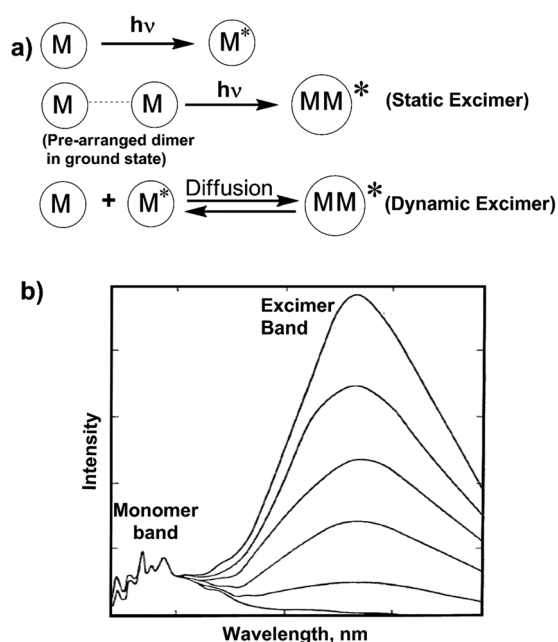


Fig. 2 (a) Mechanism of static and dynamic excimer formation. (b) The monomer and excimer emission band of pyrene at various concentration in cyclohexane.

acceptor group that enhances its electron-withdrawing character.<sup>61</sup> The fluorescence spectra should be shifted in the same direction as the absorption spectra. Receptors for the determination of  $Cu^{2+}$  sensing are designed from this PCT mechanism by using either the electron-donating, electron withdrawing ability or  $\pi$ -conjugation degree of the fluorophores.

## 5. Chemosensors for sensing $Cu^{2+}$

### 5.1. Chemosensors based on excimer emission

Calixarenes are an important class of phenol-based macrocycles having some unique properties, specifically a nonpolar

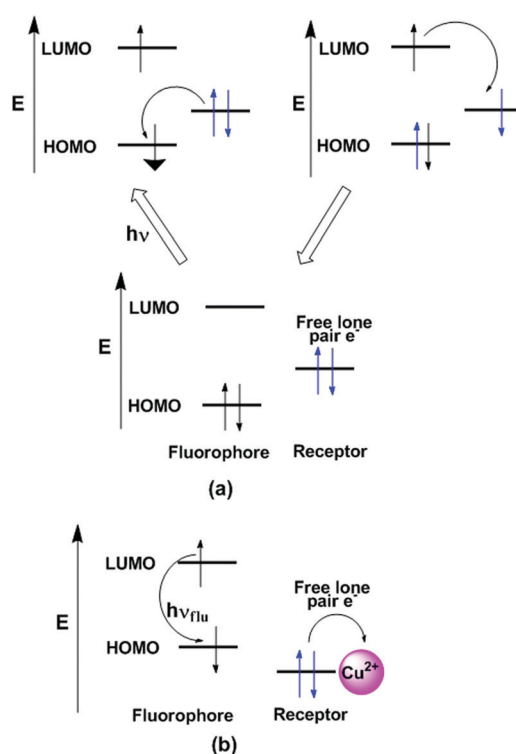


Fig. 3 Mechanisms for (a) PET (metal free receptor) and (b) CHEF (metal bound receptor) systems with the help of HOMO and LUMO of the molecular orbital energy diagrams of the fluorophore and an external molecular orbital.

cavity, that are able to encapsulate various guest species, and possess well-defined conformations with tunable functionalization of their lower and upper rims. Like calixarenes, thiacalix[4]arenes containing bridging sulfur atoms with their 3-dimensional structure are an ideal platform for the development of chemosensors.<sup>62</sup> Thiacalix[4]arenes appended with a pyrene moiety have been successfully designed utilizing the

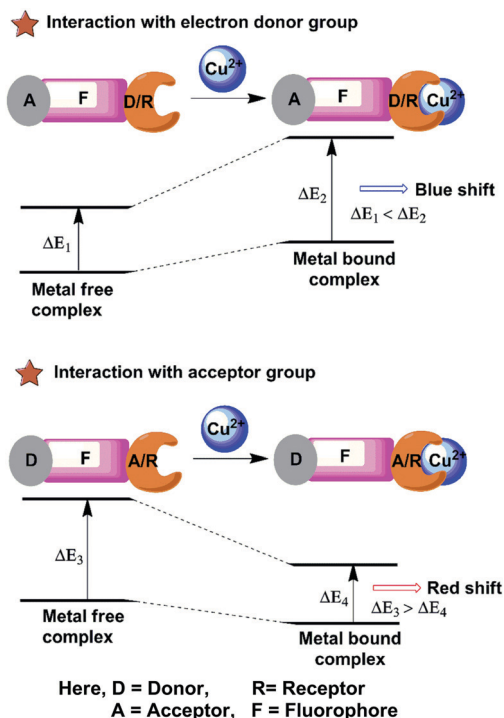


Fig. 4 Schematic representation of PCT or ICT for fluorescent probes with  $\text{Cu}^{2+}$  complex.

intensity ratio of the monomer to excimer emission ( $I_{\text{M}}/I_{\text{E}}$ ) of the pyrene moiety for the recognition of cations, anions or neutral molecules.<sup>63</sup> In 2013, Kumar *et al.* developed a heteroditopic receptor based on a thiacalix[4]arene possessing 1,3-alternate conformation. The receptor has two urea linked pyrene moieties and a crown-ether moiety at the opposite sides of the thiacalix[4]arene cavity, and this exhibited ratiometric fluorescence for the selective recognition of  $\text{F}^-$  and  $\text{CN}^-$  ions in THF.<sup>64</sup> In 2014, the Yamato group established a heterodimeric system based on a thiacalix[4]arene possessing a 1,3-alternate conformation which was capable of binding  $\text{K}^+$  ions and various anions *via* the crown-5-ring moiety and the two urea linked pyrene moieties in a  $\text{CH}_2\text{Cl}_2$ -DMSO solvent system. Moreover, a positive allosteric effect was observed for the formation of a heterogeneous dinuclear complex with  $\text{Br}^-$  and  $\text{K}^+$  ions and a negative allosteric effect induces the decomplexation of the  $\text{K}^+$  ion from the crown-5 ring for the recognition of  $\text{Cl}^-$ .<sup>65</sup> However, very few ratiometric fluorescence sensors based on a thiacalix[4]arene have been designed for the recognition of  $\text{Cu}^{2+}$ . For this purpose, in 2015 the Yamato group developed a ratiometric fluorescent chemosensor **1**, in which an acetate group was introduced between the thiacalix[4]arene and triazolyl-pyrenyl groups in a 1,3-alternate conformation, which displays high selectivity toward  $\text{Cu}^{2+}$  ions (Fig. 5).<sup>66</sup> The free ligand **1** exhibits strong face-to-face  $\pi$ - $\pi$  stacking because of excimer emission at 484 nm and weaker monomer emissions at 379 and 397 nm. After gradual addition of  $\text{Cu}^{2+}$  ions (up to 20 equiv.), the monomer to excimer relative emission intensity ratio of **1** +  $\text{Cu}^{2+}$  increased 228-fold *versus* the monomer to excimer intensity ratio of the

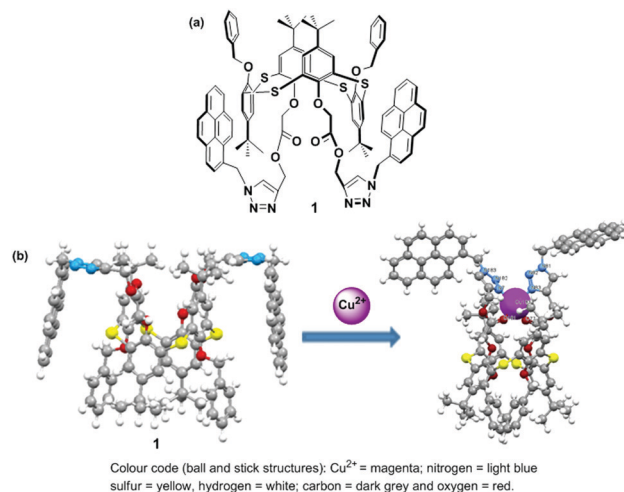


Fig. 5 (a) The molecular structure of ligand **1**; (b) geometry-optimized structures of **1** for  $\text{Cu}^{2+}$  in gas phase. Reprinted from ref. 66, with permission of Copyright 2015, Elsevier.

free ligand **1**, which was 0.54. The ligand **1** at low concentrations causes excimer quenching and monomer enhancing due to the coordination bond of a  $\text{Cu}^{2+}$  ions between the nitrogen atoms of the triazole ring and the adjacent oxygen atoms of **1**. This event forces the pyrenyl groups of **1** to move away from each other resulting in a conformational change. Notably, the monomer emission intensity of **1** was vividly reduced on addition of 5 to 45 equiv. of  $\text{Cu}^{2+}$  ions in EtOH solution, and this can be explained by a reverse PET from the pyrene unit to the nitrogen atoms of the triazole ring or to a heavy atom effect at high ionic solution strength. A Job's plot for the ligand to  $\text{Cu}^{2+}$  complexation revealed a 1:1 stoichiometry with the detection limit  $1.44 \times 10^{-7}$  M. The IR spectra of **1** revealed that the strong absorption band for the triazole group had disappeared and the band for the  $-\text{COO}-$  group changed to a weak absorption after binding with  $\text{Cu}^{2+}$ . Moreover,  $^1\text{H}$  NMR titration experiments for **1** were also consistent with reversibility characteristics as tested with  $\text{Cu}^{2+}$  and ethylenediamine. By using Density Functional Theory (DFT) computational studies, the geometry-optimized energies of **1** with  $\text{Cu}^{2+}$  complexes were calculated.

Homooxacalix[3]-arenes with a basic  $C_3$ -symmetric cavity are related to calixarenes and crown ethers. Recently, homooxacalix[3]-arenes appended with pyrene functionality have also been used for the development of novel fluorescence chemosensors.<sup>67</sup> Our group reported a new type of fluorescent chemosensor based on a homooxacalix[3]arene which is connected with a pyrene moiety through a triazole group. The sensor exhibits a great sensitivity and selectivity for the recognition of  $\text{Pb}^{2+}$  compared with most other competitive metal ions apart from  $\text{Cu}^{2+}$  where quenching was observed in an aqueous organic solvent system.<sup>68</sup> Bearing this in mind, we developed a novel ratiometric chemosensor **2** using a pyrene linked triazole modified homooxacalix[3]arene for the recognition of  $\text{Hg}^{2+}$  and  $\text{Cu}^{2+}$ .<sup>69</sup> The free ligand **2** exhibited fluorescence monomer and excimer emissions at wavelengths

of 396 and 485 nm, respectively. The fluorescence spectral changes of the chemosensor **2** showed that the excimer and monomer emission of pyrene was dramatically quenched in the presence of  $\text{Cu}^{2+}$  and  $\text{Hg}^{2+}$  ions in pure acetonitrile solution. Interestingly, the monomer emission of **2** appeared to enhance with the addition of  $\text{Cu}^{2+}$  and  $\text{Hg}^{2+}$  to the organic/aqueous solutions. The time-dependent fluorescence spectral changes explained the detailed information about monomer enhancement of the  $2 + \text{Hg}^{2+}$  and  $2 + \text{Cu}^{2+}$  complexes at 396 nm in the presence of 5% water in acetonitrile solution. The binding phenomenon was further confirmed by matrix-assisted laser desorption ionization time-of-flight mass spectrometry (MALDI-TOF) and by  $^1\text{H}$  NMR spectroscopic titration experiments. The participation of water molecules in the complexation procedure of **2** with  $\text{Hg}^{2+}$  and  $\text{Cu}^{2+}$  inhibits the heavy atom effect and thereby the enhancement of monomer emission is observed. Similar phenomenon was observed using  $\text{CH}_3\text{OH}$ ,  $\text{CH}_3\text{CH}_2\text{OH}$  or 4,4'-bipyridine as solvent/melt for the  $2 + \text{Cu}^{2+}$  and  $2 + \text{Hg}^{2+}$  complexes (Fig. 6).

Hexahomotrioxacalixarenes are defined as a class of synthetic macrocycles with phenolic units connected by  $\text{CH}_2\text{OCH}_2$  bridges. By introducing different ionophores, homotrioxacalix[3]arenes have been used as potential receptors.<sup>70</sup> The Yamato group incorporated a 2,2'-bipyridyl group attached *via* a carbonyl group at the upper rim and a diethylacetamide group at the lower rim of the hexahomotrioxacalix[3]arene that served as a tritopic receptor for  $\text{Ag}^+$ ,  $\text{Li}^+$  and  $\text{Na}^+$  ions in a cooperative fashion.<sup>71</sup> Another representative example was reported by taking advantage of the excellent fluorescent properties of pyrene. In particular, we

synthesized a pyrene-armed hexahomotrioxacalix[3]arene **3** as a ratiometric fluorescent sensor for the selective and sensitive detection of  $\text{Cu}^{2+}$  *via* a  $\text{Zn}^{2+}$  or  $\text{Cd}^{2+}$  triggered synergistic effect in a  $\text{CH}_3\text{CN}/\text{CH}_2\text{Cl}_2$  solvent system.<sup>72</sup> Furthermore, the ligand **3** with  $\text{Cu}^{2+}$  plays a vital role as an indirect sensor for  $\text{F}^-$  recognition through demetallation. In this case, the fluorescence spectra of **3** in  $\text{CH}_3\text{CN}/\text{CH}_2\text{Cl}_2$  exhibit a comparatively strong excimer emission at 518 nm and a weak monomer emission at 415 nm, with an intensity ratio of monomer to excimer emission of 0.09. The addition of  $\text{Cu}^{2+}$  ions to a solution of **3** leads to a significant increase in the monomer emission and a comparative decrease in the excimer emission to reveal a ratiometric change from 0.09 to 4.36. These spectral changes of sensor **3** with  $\text{Cu}^{2+}$  can be ascribed to the cooperating effect of the geometrical structural changes and the reduced PET effect. The complex follows a 1:1 stoichiometry with a binding constant  $(3.57 \pm 0.1) \times 10^5 \text{ M}^{-1}$ . Furthermore, the  $3 + \text{Cu}^{2+}$  complex displays a highly sensitive response at 415 nm to both  $\text{Zn}^{2+}$  and  $\text{Cd}^{2+}$  through a synergistic effect. In this case, the quantum yield of the  $3 + \text{Cu}^{2+}$  complex is considerably greater than before (from 0.05 to 0.19) upon addition of  $\text{Zn}^{2+}$  or  $\text{Cd}^{2+}$ . From the fluorescence and NMR spectroscopic titration experiments, the following can be proposed: (i) the nitrogen atom of ligand **3** interacts with both  $\text{Cu}^{2+}$  and  $\text{Zn}^{2+}$ . The  $\text{Cu}^{2+}$  coordination with nitrogen of **3** inhibits the electron transfer from the nitrogen to the photo-excited pyrene moieties, resulting in the enhancement of monomer emission. (ii) The presence of both  $\text{Zn}^{2+}$  and  $\text{Cu}^{2+}$  further decreases the electron density at the nitrogen atom which leads to the significant increase in the monomer emission intensity of the pyrene moieties and forms a trimer complex. However, spectral changes suggest that ligand **3** can sequentially recognize  $\text{Cu}^{2+}$  and  $\text{F}^-$  through a two-step process, namely a complexation approach between **3** and  $\text{Cu}^{2+}$ , and then a  $\text{Cu}^{2+}$  displacement approach by  $\text{F}^-$  to form  $\text{CuF}_2$ . Therefore, sensor **3** can act as a multifunctional molecular device by constructing a fluorescence-based combinational logic gate using the  $\text{Cu}^{2+}$ ,  $\text{Zn}^{2+}/\text{Cd}^{2+}$  and  $\text{F}^-$  fluorescence responses (Fig. 7).

Li and co-workers have reported a pyrene based novel ratiometric fluorescent chemosensor for  $\text{Cu}^{2+}$  containing

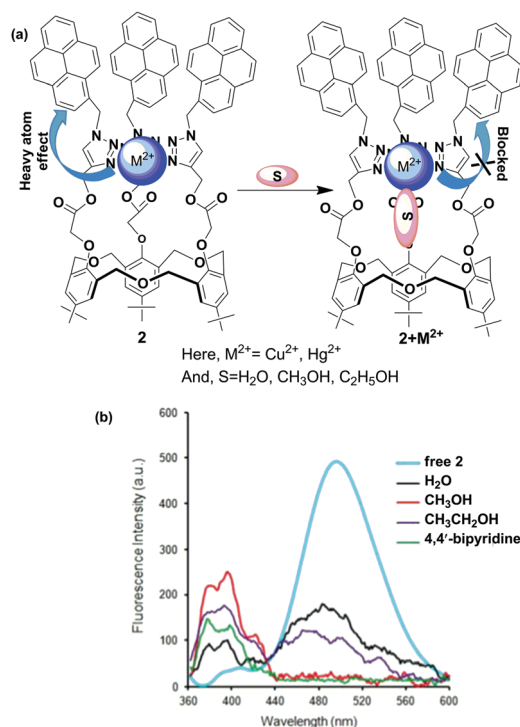


Fig. 6 (a) The heavy atom effect of  $2 + \text{M}^{2+}$  complex blocked by solvent S; (b) fluorescence response of  $2 + \text{M}^{2+}$  upon interaction with various solvents.

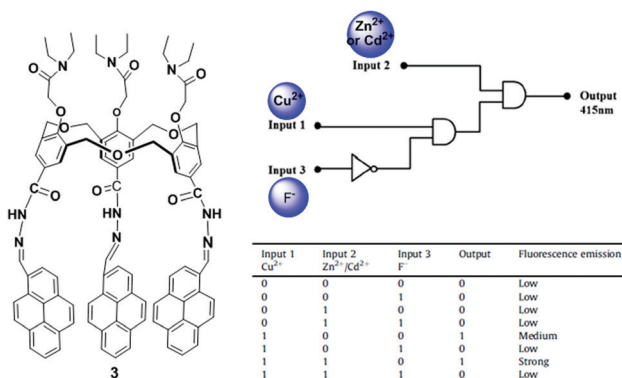


Fig. 7 The molecular structure of receptor **3** and the combinational logic gate for **3** with truth table using  $\text{Cu}^{2+}$ ,  $\text{Cd}^{2+}/\text{Zn}^{2+}$  and  $\text{F}^-$  as chemical inputs. Reproduced from ref. 72 with permission from the Elsevier, copyright 2015.

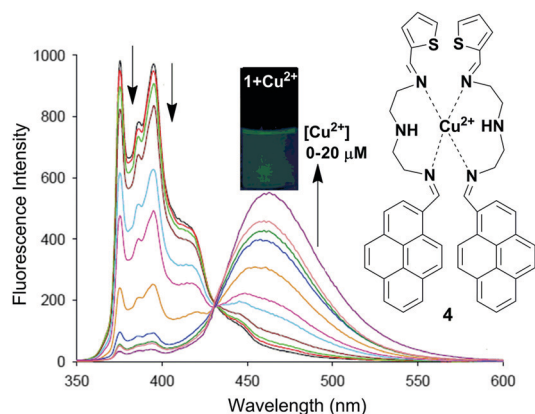


Fig. 8 Proposed binding mechanism of chemosensor **4** with fluorescence colour changes. Fluorescence intensity changes of compound **4** (10  $\mu\text{M}$ ) with different concentrations of  $\text{Cu}^{2+}$  in Tris- $\text{HNO}_3$  buffer solution at  $\lambda_{\text{exc}} = 342 \text{ nm}$ . Reproduced from ref. 73 with permission from the Elsevier, copyright 2016.

thiophene, namely compound **4** (Fig. 8).<sup>73</sup> The monomer to excimer conversion of chemosensor **4** occurred on gradual addition of  $\text{Cu}^{2+}$ , owing to intermolecular  $\pi$ - $\pi$  stacked dimerization of the two pyrenes. The fluorescent emission of **4** (10  $\mu\text{M}$ ) in Tris- $\text{HNO}_3$  buffer solution with high sensitivity and selectivity increases 127-fold with 2 equiv. of  $\text{Cu}^{2+}$  (20  $\mu\text{M}$ ). In addition, the detection of  $\text{Cu}^{2+}$  present in environmental samples such as river and pond water has been determined by using **4**. Moreover, cytotoxicity tests on **4** (from 0  $\mu\text{M}$  to 20  $\mu\text{M}$ ) using cells showed that more than 96% of the cells were viable which established the real applicability of the chemosensor **4** in biological samples. The association constant ( $K_a$ ) for  $\text{Cu}^{2+}$  binding by chemosensor **4** was determined to be  $2.18 \times 10^4 \text{ M}^{-1}$ . There was a good linear relationship covering  $1.0 \times 10^{-7}$  to  $2.0 \times 10^{-5} \text{ M}$  between the fluorescence ratio ( $I_{460}/I_{375}$ ) of **4** and the concentration of  $\text{Cu}^{2+}$  with a detection limit of  $2 \times 10^{-8} \text{ M}$ .

Two new chemosensors bearing either a single **5** or two **6** pyrene units linked by a flexible polyoxaethylene bridge have been developed that are sensitive to water and metal cations (Fig. 9).<sup>74</sup> Compound **5** in dioxane showed a single monomer absorption and emission band, whilst compound **6** gave an absorption band in the 400–500 nm region with an emission band with maxima at  $\sim 500 \text{ nm}$ . In dioxane/water mixtures with  $x\text{H}_2\text{O} = 0.29$ , the two probes exhibited a new band (with maxima varying from 405 to 490 nm) due to exciplex formation. Time-resolved experiments have been used to explain a two-state system for ligand **5**, which involves the monomer and a charged species, which can potentially be assigned to an exciplex-like species, whereas with **6**, a three-state system is involved. After addition of the metal ions ( $\text{Cu}^{2+}$ ,  $\text{Zn}^{2+}$  and  $\text{Ag}^+$ ) to probe **5** in dioxane followed by absorption and emission studies, a gradual quenching effect of the monomer emission was observed and this was significantly selective for  $\text{Cu}^{2+}$ . In case of ligand **6**, after the interaction with metal cations, the emission band decreases at approximately 550 nm and the

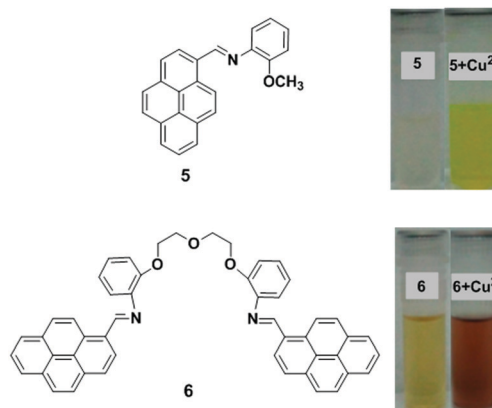


Fig. 9 Molecular structures of receptors **5** and **6**. Observed colour change **5** and **6** with  $\text{Cu}^{2+}$ . Reproduced from ref. 74 with permission from the American Chemical Society, copyright 2013.

monomer emission band increases at  $\sim 450 \text{ nm}$ . The binding ratio of ligand (**5** and **6**) to metal was proposed to be a 1 : 1 stoichiometry.

A pentiptycene-bispyrenyl system **7** was developed from the reaction of pentiptycene hydroquinone with 1-bromomethylpyrene (Fig. 10).<sup>75</sup> Yang *et al.* reported compound **7** as a selective and sensitive fluorescent chemosensor for  $\text{Ca}^{2+}$ ,  $\text{Cd}^{2+}$  and  $\text{Cu}^{2+}$ . The fluorescence spectra of compound **7** exhibited typical monomer emission at 375 and 395 nm and an excimer emission at 475 nm in  $\text{CH}_2\text{Cl}_2$ . In particular, upon addition of  $\text{Cu}^{2+}$ , a strong blue shifted (from 475 to 440 nm) pyrene excimer emission was observed with increased intensity. This behaviour of sensor **7** was clarified by the excitation spectra which clearly showed a significant red shift at  $\text{Cu}^{2+}$ -induced 440 nm than 375 nm, due to the conversion of a partially overlapping static excimer from a sandwich-like dynamic excimer. Moreover, the **7** +  $\text{Cu}^{2+}$  complex at 440 nm has a short fluorescence lifetime (approximately 16-times) in comparison with 475 nm excimer emissions, which was explained by observation of the fluorescence decay times. Therefore, the radiative decay rate constant was larger for the static emission at 440 nm than the dynamic emission at 475 nm. In this case,  $\text{Cu}^{2+}$  binding

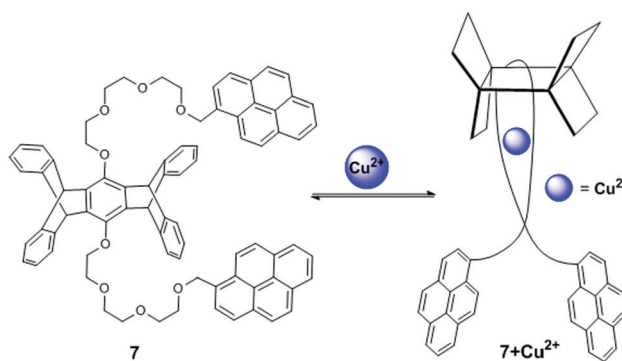


Fig. 10 Structure of chemosensor **7** and possible binding modes for  $\text{Cu}^{2+}$ . Reproduced from ref. 75 with permission from the American Chemical Society, copyright 2001.

with **7** brings the pyrene groups together in the ground state for the formation of static excimers and minimizes the relaxation event in the excited state leading to a blue-shifted excimer emission. The UV-vis absorption spectra of ligand **7** also exhibited a clear broadening of the spectrum with four defined isobestic points at 325, 331, 339 and 348 nm and suggested a 1:1 complex with a binding constant of 4.4.

Kim *et al.* reported the mono-pyrenylalkylamine derivative **8**.<sup>76</sup> Upon addition of  $\text{Cu}^{2+}$ , chemosensor **8** exhibited band broadening and a red-shift (from 342 to 455 nm) in a mixed solvent system of  $\text{CH}_3\text{CN}/\text{H}_2\text{O}$  (1:1) (Fig. 11). These observations were explained by favourable intermolecular  $\pi$ - $\pi$  stacking dimerization of the two pyrenes in the ground state. In the Job's plot measurements, the maximum point appeared at the mole fraction of 0.6, which suggested a 2:1 ligand-to-metal complex. Fluorescence spectral changes of **8** revealed high selectivity toward  $\text{Cu}^{2+}$  over other competitive species ( $\text{Li}^+$ ,  $\text{Na}^+$ ,  $\text{K}^+$ ,  $\text{Rb}^+$ ,  $\text{Cs}^+$ ,  $\text{Mg}^{2+}$ ,  $\text{Ba}^{2+}$ ,  $\text{Ca}^{2+}$ ,  $\text{Sr}^{2+}$ ,  $\text{Ag}^+$ ,  $\text{Zn}^{2+}$ ,  $\text{Cd}^{2+}$ ,  $\text{Hg}^{2+}$  and  $\text{Pb}^{2+}$ ). Moreover, the theoretical DFT calculations revealed that the pre-organized cavity is essential with two proximate nitrogen atoms of the sulfonamide groups to stabilize the complex of **8** with  $\text{Cu}^{2+}$  for static excimer emission.

The design and synthesis of new pyrene-derived fluorescent sensors containing quinolinylamide groups (**9–11**) was also reported by Kim and coworkers. The coordination behaviour was examined with variations in length of the methylene groups ( $n = 0, 1, 3$ ) between the pyrene and quinolinylamide groups of **9–11** toward  $\text{Cu}^{2+}$  by fluorescence spectroscopy and theoretical DFT calculations (Fig. 12).<sup>77</sup> Addition of  $\text{Cu}^{2+}$  to ligand **9** ( $n = 0$ ) produced a new broad emission band which supports strong static excimer emission at 460 nm, whilst the binding constant was found to be  $5.42 \times 10^5 \text{ M}^{-1}$ . Moreover, compound **9** interacts with  $\text{Cu}^{2+}$  following a 2:1 stoichiometry as deduced from the Job's plot analysis, mass spectra and fluorescence behaviour. A DFT study confirmed that the minimum intramolecular distance between the pyrene and quinoline amide is the main factor for  $\text{Cu}^{2+}$  ion detection.

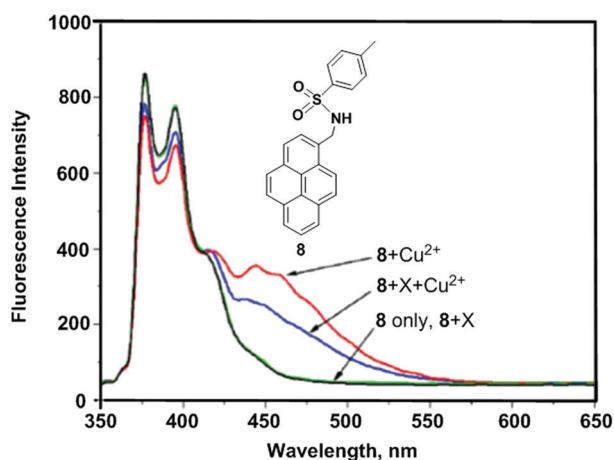


Fig. 11 The fluorescence spectra of **8** in the presence of  $\text{Cu}^{2+}$  and miscellaneous cations, **X** (10 equiv.). Reproduced from ref. 76 with permission from the American Chemical Society, copyright 2008.

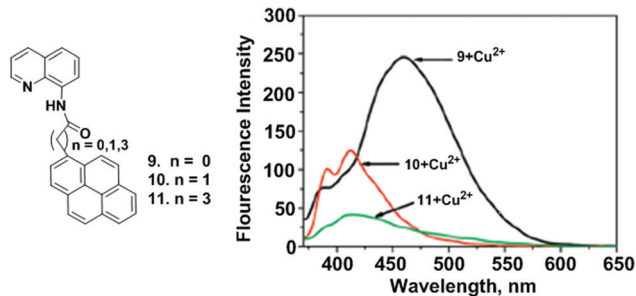


Fig. 12 The structures of the molecular receptors **9**, **10** and **11**. Fluorescence spectra of **9–11** upon addition of  $\text{Cu}(\text{ClO}_4)_2$  (10 equiv.). Reproduced from ref. 77 with permission from the American Chemical Society, copyright 2009.

In the presence of 10 equiv. of  $\text{Cu}^{2+}$ , the excimer emission intensity of **9** became approximately 8 times greater than that of probe **10** and **11** ( $n = 1, 3$ ). Thus, it was concluded that the methylene spacers between the pyrene and carbonyl unit have a great impact on strong intermolecular Py-Py\* formation in order to show an intense static excimer band.

A pyrene containing Schiff base fluorosensor **12** has been developed which showed a high selectivity and sensitivity towards  $\text{Cu}^{2+}$  ions.<sup>78</sup> This was due to the hindrance of the PET process upon complexation of the pyrene moiety with  $\text{Cu}^{2+}$  through interaction of the nitrogen lone pair electron of **12** with  $\text{Cu}^{2+}$  that induced the intermolecular excimer formation (Fig. 13). The free ligand **12** exhibited two weak broad emissions centred at 385 nm and 452 nm in Tris-HCl (10 mM, pH = 7) buffer containing  $\text{CH}_3\text{CN}-\text{H}_2\text{O}$  (v/v = 1:1) medium which were assigned to the monomer and excimer of the pyrene moieties respectively. Moreover, the presence of  $\text{Cu}^{2+}$  triggered a noticeable enhancement (10 fold) of the excimer peak at 452 nm along with two prominent monomer peaks at 378 and 396 nm. The detection limit of the sensor **12** was found to be  $4 \times 10^{-8} \text{ M}$ . The emission property of **12** (5  $\mu\text{M}$ ) was dependent on the nature of the solvent, and was enhanced on increasing the concentration of water up to 80% and there after reduced at 90% water fraction. Moreover, since some active groups (e.g.  $-\text{NH}_2$ ) are present in the probe which are sensitive to  $\text{H}^+$  ions, there is an effect of pH on the fluorescent response of the probe.

Lin and co-workers have designed a simple pyrene containing derivative as a  $\text{Cu}^{2+}$  turn-on chemosensor **13**.<sup>79</sup> The probe

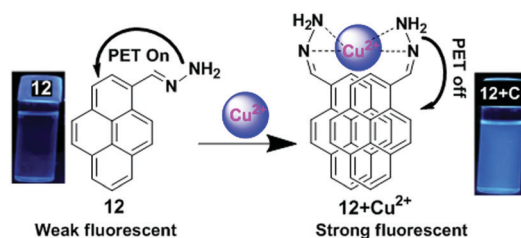


Fig. 13 Schematic representation of  $\text{Cu}^{2+}$  sensing by fluorosensor **12** and emission photograph of **12** and **12** +  $\text{Cu}^{2+}$  under UV radiation. Reproduced from ref. 78, with permission from the Royal Society of Chemistry.

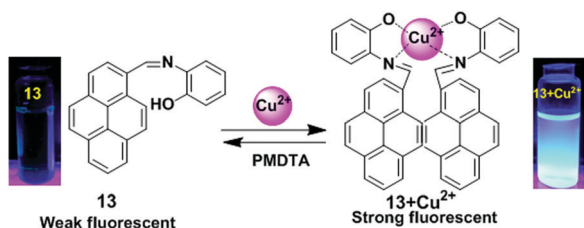


Fig. 14 Schematic presentation for detection of  $\text{Cu}^{2+}$  using receptors **13**. Photographs of **13** and **13** +  $\text{Cu}^{2+}$  visualized under UV-light irradiations. Reproduced from ref. 79, with permission from the Royal Society of Chemistry.

**13** in  $\text{CH}_3\text{CN}$  revealed a greater photoluminescence intensity enhancement at 459 nm for  $\text{Cu}^{2+}$  ( $\Phi = 0.284$ ) versus other different metal ions ( $\text{Li}^+$ ,  $\text{Ag}^+$ ,  $\text{K}^+$ ,  $\text{Na}^+$ ,  $\text{Cs}^+$ ,  $\text{Ni}^{2+}$ ,  $\text{Fe}^{3+}$ ,  $\text{Co}^{2+}$ ,  $\text{Zn}^{2+}$ ,  $\text{Cd}^{2+}$ ,  $\text{Pb}^{2+}$ ,  $\text{Ca}^{2+}$ ,  $\text{Cr}^{3+}$ ,  $\text{Mg}^{2+}$ ,  $\text{Cu}^{2+}$ ,  $\text{Mn}^{2+}$ ,  $\text{Hg}^{2+}$ ,  $\text{Fe}^{2+}$  and  $\text{Ag}^+$ ). The sensing mechanism involves heteroatoms (O and N) to form the excimer in which one  $\text{Cu}^{2+}$  binds with two N and two O (one N and one O from one molecule) (Fig. 14). The 2 : 1 stoichiometry of the **13** +  $\text{Cu}^{2+}$  complex was calculated from Job's plots based on UV-vis absorption titrations. Moreover, the **13** +  $\text{Cu}^{2+}$  sensor was found to be active over a wide range of pH (1–14). After 10 minutes, the relative fluorescence intensity changes for **13** +  $\text{Cu}^{2+}$  reached a maximum value and so the system is effective with respect to time (0–10 minutes). The detection limit (LOD) of **13** with  $\text{Cu}^{2+}$  was calculated at  $9.72 \times 10^{-7}$  M. Similarly, based on fluorescent binding isotherms, the association constant ( $K_a$ ) of the complex was estimated at  $1.96 \times 10^6 \text{ M}^{-1}$ .

A novel fluorescent chemosensor **14**, made up of pyrene units connected by a binaphthyl-crown derivative was designed and prepared for the selective detection of  $\text{Cu}^{2+}$  in the presence of other metal cations ( $\text{Ca}^{2+}$ ,  $\text{Cd}^{2+}$ ,  $\text{Co}^{2+}$ ,  $\text{Cs}^+$ ,  $\text{Cu}^{2+}$ ,  $\text{Hg}^{2+}$ ,  $\text{K}^+$ ,  $\text{Mg}^{2+}$ ,  $\text{Mn}^{2+}$ ,  $\text{Na}^+$ ,  $\text{Ni}^{2+}$ ,  $\text{Pb}^{2+}$ , and  $\text{Zn}^{2+}$ ) in acetonitrile solution (Fig. 15).<sup>80</sup> Compound **14** (1  $\mu\text{M}$ ) shows monomer and excimer emissions at 376 and 477 nm respectively for excitation at 342 nm.

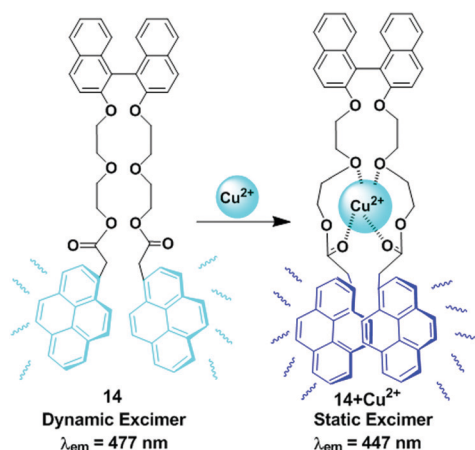


Fig. 15 Schematic representation of  $\text{Cu}^{2+}$  sensing by pyrene appended-binaphthyl-crown derivative compound, **14**. Reproduced from ref. 80 with permission from the Elsevier, copyright 2006.

Addition of  $\text{Cu}^{2+}$  produced a new blue shifted excimer emission band centered at 447 nm with increasing intensity maximum up to 3-fold. The sensor **14** exhibits static excimer emission characteristics upon addition of  $\text{Cu}^{2+}$  that can be easily understood from excitation spectra. Here, the emission at 477 nm resulted from a dynamic excimer due to the identical excitation spectrum at both the monomer wavelength (376 nm) and the excimer wavelength (477 nm). The excitation spectrum of ligand **14** with  $\text{Cu}^{2+}$  monitored at 447 nm was significantly red-shifted compared with the spectrum taken at 376 nm which was attributed to the 447 nm from the static excimer. Moreover, from the fluorescence titration experiments, the association constant of **14** with  $\text{Cu}^{2+}$  was determined to be  $65\,600 \text{ M}^{-1}$ .

Kumar *et al.* have developed a new heteroditopic receptor **15**, based on the 1,3-alternate conformation of thiacalix[4]arene bearing amine groups appended with pyrene moieties and a crown-4 ring (Fig. 16).<sup>81</sup> The receptor **15** can detect  $\text{Cu}^{2+}$  ions selectively among other metal ions with a detection limit of  $40 \times 10^{-9} \text{ mol L}^{-1}$  and follows a 1 : 2 stoichiometry of ligand to  $\text{Cu}^{2+}$  complexation. It shows negative allosteric behaviour between  $\text{Cu}^{2+}/\text{Li}^+$  ions in mixed aqueous media. The binding behaviour of compound **15** was studied by UV-vis and fluorescence spectroscopy which suggests the  $\text{Cu}^{2+}$  ions interact with the amino nitrogen centres and  $\text{Li}^+$  ions co-ordinate to the crown ether ring of receptor **15**. The excimer emission band of receptor **15** (1.0  $\mu\text{M}$ ;  $\text{EtOH-H}_2\text{O}$ ; 8 : 2, v/v) for the pyrene moieties was clearly observed at 466 nm due to the fully overlapped position of the two pyrene units resulting in the folded conformation of **15**. Notably, after gradual addition of only  $\text{Cu}^{2+}$  ions, the monomer fluorescence emission at 378 nm shows a major enhancement with a remarkable decrease in the excimer emission intensity and the variation of these emissions led to the formation of an isoemissive point at 430 nm. This ratiometric behaviour of receptor **15** in the presence of  $\text{Cu}^{2+}$  ions was due to the decreased electron density at nitrogen leading to the conformational changes that moved the overlapped pyrene units away from each other.

A novel pyrene containing chemosensor **16** which possesses a self-assembled 3D crystal structure *via* C-H... $\pi$ ,  $\pi$ ... $\pi$  and different types of H-bonding interactions, has been designed

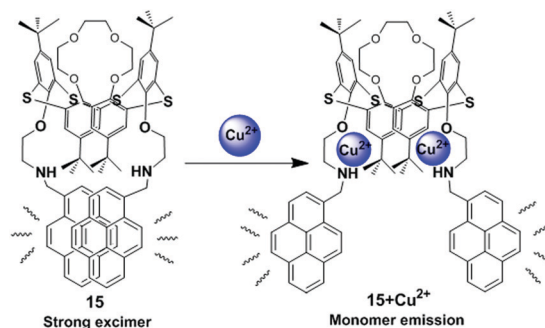


Fig. 16 Schematic representation of possible sensing mechanism for **15** on interaction with  $\text{Cu}^{2+}$ . Reproduced from ref. 81 with permission from the Royal Society of Chemistry.

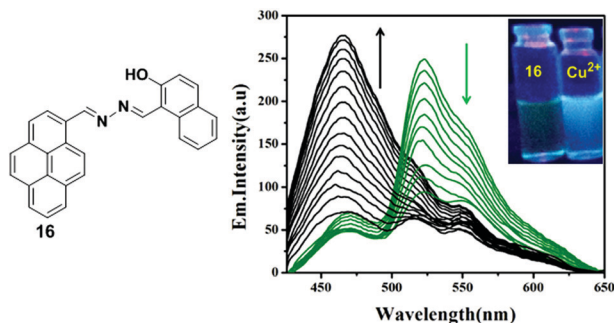


Fig. 17 Structure of compound **16** showing fluorescence emission spectra of **16** with the addition of  $\text{Cu}^{2+}$ . Inset: Visual fluorescence changes of **16** and upon addition of  $\text{Cu}^{2+}$  using a handheld UV lamp. Reproduced from ref. 82, with permission from the Royal Society of Chemistry.

for the efficient and selective detection of  $\text{Cu}^{2+}$  and  $\text{F}^{-}$  ions in a dual sensing mode which do not interfere with each other.<sup>82</sup> The sensor **16** can recognize  $\text{Cu}^{2+}$  by fluorimetric experiments and by visual colour changes under UV-light. The emission spectra of sensor **16** ( $1 \times 10^{-7}$  M) in a methanol–water mixture (methanol:water 7:3 v/v) shows, on changing the concentration, a local emission band at 466 nm which remains almost unchanged, whilst the band at 530 nm was continuously altered indicating the formation of excimers. After addition of  $\text{Cu}^{2+}$  to the sensor **16**, a decrease of the excimer bands at 520 nm and 560 nm was initially observed as a result of interaction of the copper with the sensor and then the **16** +  $\text{Cu}^{2+}$  complex causes an increment of the band at 466 nm (Fig. 17). The above phenomenon can be explained by an excimer switch-off mechanism for promoting complexation with  $\text{Cu}^{2+}$ . A Benesi–Hildebrand plot and Job's plot from UV-vis titrations demonstrated that the stoichiometry of the complex formed by **16** and  $\text{Cu}^{2+}$  is 1:1 with a binding constant of  $4.8 \times 10^6 \text{ M}^{-1}$ . In order to confirm the binding phenomena, the structure and electronic properties of **16** and its copper complex **16** +  $\text{Cu}^{2+}$  have been investigated by Density Functional Theory (DFT). Moreover, the sensor **16** has potential for biological applications when detecting  $\text{Cu}^{2+}$  as it is transported across mammalian cell membranes (HEK 293 cells).

Mallik *et al.* have reported the development of four new saturated, pyrene-containing, metal-chelating lipids as fluorescent sensors (**17–20**).<sup>83</sup> These lipids can interact with transition metal ions *via* a variety of metal-chelating head groups such as iminodiacetate (IDA), EDTA, DTPA (Fig. 18). However, all the lipids (**17–20**) in DMSO solution failed to show the presence of any excimers in the emission spectra. They can form liposomes ( $2.0 \text{ mg mL}^{-1}$ ) under standard liposome formation conditions. The emission spectrum of **17** showed that both pyrene monomers (395 nm) and excimers (470 nm) were present in the liposomes in which the excimer to monomer intensity ratio (0.69) clearly indicated that lipid **17** was aggregated in the liposomes. Upon treatment with  $\text{Cu}^{2+}$ , the overall fluorescence intensity of the liposomes decreased due to quenching of the excited states of the pyrene monomer by  $\text{Cu}^{2+}$  compared to the pyrene excimer, leading to an increase in excimer-to-monomer ratio.

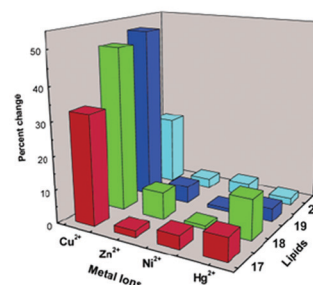
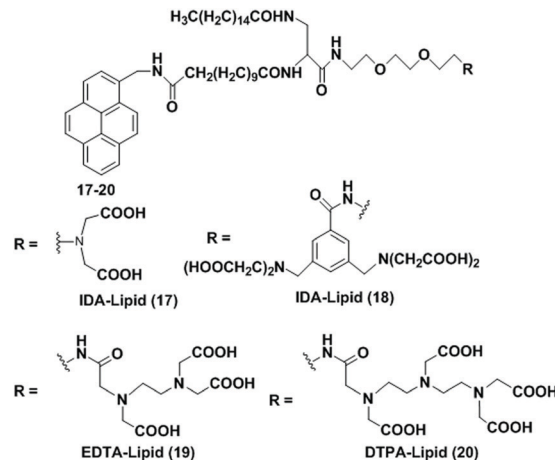


Fig. 18 Structures of molecular receptor **17–20**. Relative increase of the  $I_{470}/I_{395}$  ratio with transition metal ions for the saturated lipids. Reproduced from ref. 83 with permission from the American Chemical Society, copyright 2003.

Similar behaviour was also observed for liposomes from **17–19** except **20** which showed very little change. Other transition metal ions ( $\text{Ni}^{2+}$ ,  $\text{Hg}^{2+}$  and  $\text{Zn}^{2+}$ ) did not show any response in the excimer to monomer intensity ratio.

Another pyrene-appended ratiometric fluorescent chemosensor **21** has been reported by Yamato and coworkers in 2015. In this case, the homooxacalix[3]arene acts as a molecular spacer incorporating three pyrene fluorophore at the upper rim and three substituted triazole arms as ionophore at the lower rim (Fig. 19).<sup>84</sup> The sensor **21** can serve as a ratiometric chemosensor for heavy and transition metal (HTM) ions ( $\text{Cu}^{2+}$ ,  $\text{Hg}^{2+}$ ,  $\text{Pb}^{2+}$  and  $\text{Zn}^{2+}$ ) resulting in a conformational change for **21**. The fluorescence emission spectra of free **21** show a characteristic pyrene excimer band at 482 nm in  $\text{CH}_3\text{CN}$  solution. The interaction of **21** with  $\text{Cu}^{2+}$  and other HTM ions led to the enhancement of monomer emission accompanied by excimer quenching. Herein, the ditopic calixarene as a molecular spacer prevents the heavy atom effect of the HTM ion by insulating the fluorophore from the ionophore through a long distance. The association constant for 1:1 complexation of **21** +  $\text{Cu}^{2+}$  was calculated to be  $1.89 \times 10^5 \text{ M}^{-1}$ . FT-IR spectra of the chemosensor **21** have been recorded, and the absorption band of C–O present on the lower rim is shifted at  $1184 \text{ cm}^{-1}$  whereas no absorption band change is observed for C=O at the upper rim of **21** upon complexation with  $\text{Cu}^{2+}$ . The authors stated that three substituted triazole moieties at the lower rim

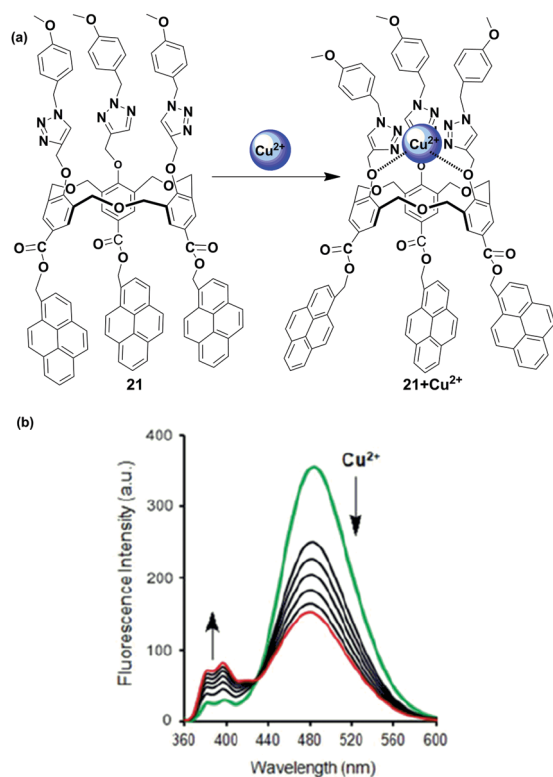


Fig. 19 (a) The structure of ratiometric sensor **21** and its binding mode with  $\text{Cu}^{2+}$ ; (b) showing fluorescence for the  $\text{Cu}^{2+}$  ion in  $\text{CH}_3\text{CN}$ . Reproduced from ref. 84 with permission from the Taylor & Francis, copyright 2015.

bind with  $\text{Cu}^{2+}$  and cause conformational changes that make the pyrene units move far away from one another.

## 5.2. Chemosensors based on PET mechanism

Receptor molecules have been designed for the enhancement of fluorescence intensity upon binding with a metal ion through the PET mechanism.<sup>85</sup> In 2015, based on the PET mechanism, our group developed 7-*tert*-butylpyrene-appended fluorescent sensors containing dipicolylamine (Dpa) linkages.<sup>86</sup> Both the mono-chelate and bis-chelate ligand showed selectivity for  $\text{Zn}^{2+}$  and  $\text{Cd}^{2+}$  ions. Herein, we explain that the binding phenomenon of both ligands towards  $\text{Zn}^{2+}$  and  $\text{Cd}^{2+}$  varied on changing the solvent system. However, the addition of  $\text{Cu}^{2+}$  showed almost no fluorescence enhancement yet  $\text{Cu}^{2+}$  quenched the fluorescence of the bis-chelate ligand and  $\text{Zn}^{2+}$  complex, which can be explained by the paramagnetic nature of  $\text{Cu}^{2+}$ . For the selective and sensitive detection of  $\text{Cu}^{2+}$ , our group designed and synthesized three new fluorescence 'off-on' chemosensors **22–24** based on pyrene containing Schiff base derivatives with different chain lengths (Fig. 20).<sup>87</sup> Compound **22** was synthesized from the condensation reaction of 1-pyrene carbaldehyde with 4-(hydrazidocarbonyl)(*N,N*-diethylaminocarbonylmethoxy)benzene. The other compounds **22** and **23** were easily synthesized *via* condensation reactions, and were compared against **22** towards  $\text{Cu}^{2+}$  in terms of binding ability. The probe **22** (1.0  $\mu\text{M}$ ) was highly sensitive for

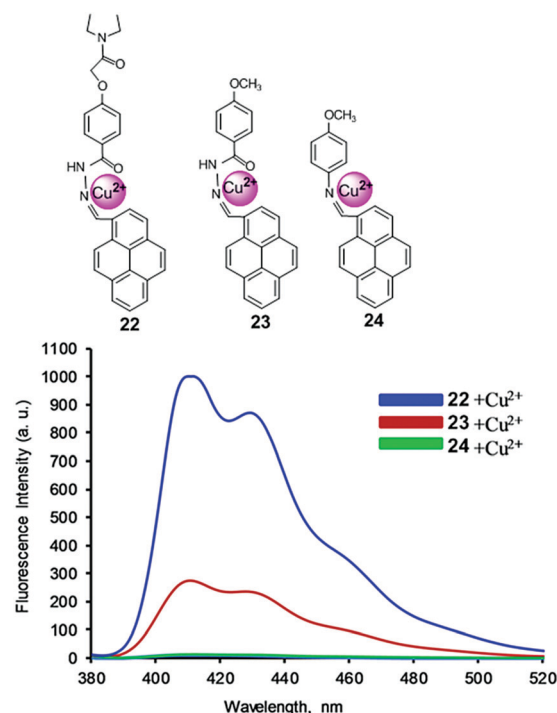


Fig. 20 Structure of chemosensors **22**, **23** and **24**. Fluorescence response of ligands upon addition of  $\text{Cu}^{2+}$  ions in  $\text{CH}_3\text{CN}/\text{CH}_2\text{Cl}_2$ . Reproduced from ref. 87 with permission from the Elsevier, copyright 2016.

$\text{Cu}^{2+}$  detection due to strong inhibition of PET. After addition of  $\text{Cu}^{2+}$ , the emission intensity was enhanced approximately 65 times more ( $\Phi = 0.31$ ) than that of the free ligand **22** ( $\Phi = 0.01$ ), and was 57 times and 40 times greater than free **23** and **24**, respectively. A Job's plot analysis was carried out at 405 nm in which the fluorescence intensity showed a maximum at the mole fraction 0.5 which corresponds to a 1:1 ratio of ligand and  $\text{Cu}^{2+}$  complex. The resulting association constant for **22** was found to be  $1.29 \times 10^5 \pm 0.32 \text{ M}^{-1}$ . A DFT computational study together with  $^1\text{H}$  NMR and  $^{13}\text{C}$  NMR spectroscopic titration experiments revealed that  $\text{Cu}^{2+}$  was bound at the imine nitrogen atom and the amide carbonyl oxygen of ligands **22** and **23**. The diethylaminocarbonylmethoxy group of **22** provides a contribution for **22** and  $\text{Cu}^{2+}$  complexation through an inductive effect.

A pyrene derivative containing a benzothiazolenhydrazone receptor as a fluorescent sensor **25** was reported by Wang and Wu (Fig. 21).<sup>88</sup> Upon addition of  $\text{Cu}^{2+}$  (25  $\mu\text{M}$ ), probe **25** (50  $\mu\text{M}$ ) exhibited a significant fluorescence enhancement at 468 nm in acetonitrile–water ( $v/v = 3/1$ , 5 mM HEPES, pH 7.0) from which the limit of detection of chemosensor **25** was estimated to be 2.73  $\mu\text{M}$ . The coordination of  $\text{Cu}^{2+}$  ions inhibited the PET process resulting in a significant enhancement of fluorescence intensity. The Job's plot experiments revealed a 2:1 stoichiometry for ligand **25** to metal complex. Density functional theory (DFT) calculations determined that two chemosensors **25** bind  $\text{Cu}^{2+}$  using four nitrogen atoms. Moreover, the fluorescence and bright-field images indicated that the probe **25** is useful for the detection of  $\text{Cu}^{2+}$  ions in living cells.

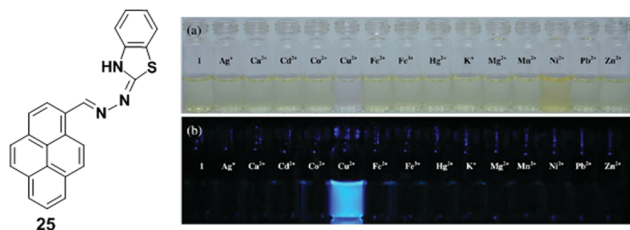


Fig. 21 Structures of probe **25** and colour (a) and fluorescence (b) changes of chemosensor **25** (500  $\mu\text{M}$ ) after addition of various metal ions (500  $\mu\text{M}$ ). Reproduced from ref. 88 with permission from the Elsevier, copyright 2013.

Molina *et al.* designed the chemosensor **26** in which a 2-aza-1,3-diene moiety was used as the ionophore for the recognition of  $\text{Cu}^{2+}$  (Fig. 22).<sup>89</sup> The absorption spectrum of chemosensor **26** exhibited two well-defined isosbestic points at 275 and 440 nm with a gradual increase of a new red shifted band at 496 nm in the presence of  $\text{Cu}^{2+}$  ion. This was responsible for the change of colour from yellowish to deep orange. The fluorescence intensity of **26** ( $2.5 \times 10^{-5} \text{ M}$ ) in  $\text{CH}_3\text{CN}$  increased upon addition of  $\text{Cu}^{2+}$  ( $I_{\text{complex}}/I_{\text{free ligand}} = 22\text{-fold}$ ), where the excimer emission shifted from 450 to 429 nm. From the emission intensity data, the association constant of the chemosensor **26** +  $\text{Cu}^{2+}$  system was found to be  $5.71 \times 10^5 \text{ M}^{-1}$ . The stoichiometry of the ligand **26** to metal complex system was estimated to be 1 : 1. The response of the fluorescence of **26** was also studied in  $\text{CH}_3\text{CN}/\text{H}_2\text{O}$  (70/30). Under these conditions, titration experiments demonstrated that the  $\text{Cu}^{2+}$  ions yielded a 10-fold enhancement of quantum yield along with a slight red shift (10 nm) of the excimer emission band. The titration data indicated the calculated detection limit of **26** was  $3.91 \times 10^{-6} \text{ M}$  for  $\text{Cu}^{2+}$ .

A pyrene based fluorescent chemosensor **27** containing a picolinohydrazide receptor was developed by Wu and co-workers (Fig. 23).<sup>90</sup> Binding with  $\text{Cu}^{2+}$  ions in mixed aqueous media ( $\text{CH}_3\text{OH} : \text{H}_2\text{O} = 7 : 3$ ) to the chemosensor **27** blocks PET and greatly enhanced the fluorescence of pyrene. After gradual addition of  $\text{Cu}^{2+}$  to the chemosensor **27** (25  $\mu\text{M}$ ), a new emission band appeared at 455 nm and the quantum yield of that emission band was 0.267, which is 20-fold that of chemosensor **27**, 0.013. The Job's plot experiment indicated that the binding ratio for the chemosensor **27** +  $\text{Cu}^{2+}$  complex was 1 : 1. It was also confirmed from  $^1\text{H}$  NMR and IR spectroscopy that the  $\text{Cu}^{2+}$  was bound to one nitrogen atom from a pyridine and

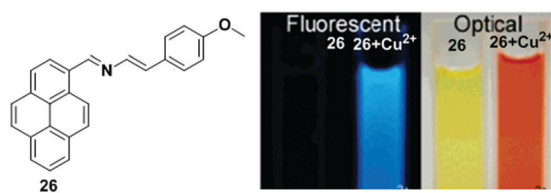


Fig. 22 Structure of chemosensor **26** and colour and fluorescence changes of **26** with  $\text{Cu}^{2+}$  in  $\text{CH}_3\text{CN}$ . Reproduced from ref. 89 with permission from the American Chemical Society, copyright 2006.

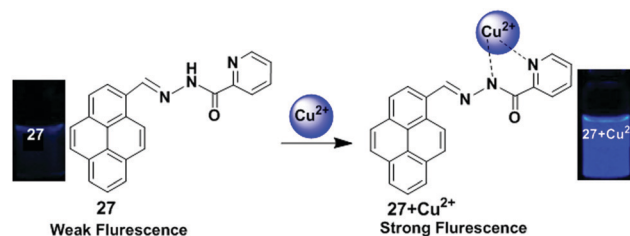


Fig. 23 Schematic representation of the sensing mechanism of probe **27** with  $\text{Cu}^{2+}$ . Reproduced from ref. 90 with permission from the Springer Nature, copyright 2012.

one nitrogen atom from an amide. Moreover, fluorescence signals determined by a fluorescence microscope indicated that chemosensor **27** possessed good cell-membrane permeability and could detect  $\text{Cu}^{2+}$  present in living cells.

A pyrene-based chemosensor **28** in which diaminomaleonitrile acted as a chelator, was synthesized and designed for  $\text{Cu}^{2+}$  ion detection (Fig. 24).<sup>91</sup> It was observed from UV-visible spectroscopy that the absorbance at 420 nm was reduced and a new band appeared at 355 nm after gradual addition of  $\text{Cu}^{2+}$  to chemosensor **28**. The colour of **28** also changed from yellow to colourless. Moreover, free ligand **28** exhibited very weak fluorescence ( $\Phi = 0.0045$ ) due to a PET process from the lone pair electrons of the two-nitrile groups to pyrene. However, the  $\text{Cu}^{2+}$ -bound probe **28** inhibited the electron withdrawing ability of the two nitrile groups and showed a sharp emission peak at 417 nm ( $\Phi = 0.59$ ) under excitation of 350 nm in acetonitrile-water.

A newly designed fluorescence chemosensor **29** that contains a pyrene motif with a hydrazinylpyridine moiety has been reported.<sup>92</sup> The sensor **29** revealed high sensitivity towards  $\text{Cu}^{2+}$  ions in a  $\text{CH}_3\text{CN}/\text{H}_2\text{O}$  solvent system over the pH range of 5.0–10. The interaction of  $\text{Cu}^{2+}$  inhibited the PET mechanism from the nitrogen lone pairs to the pyrene, resulting in significant enhancement in pyrene fluorescence (Fig. 25). The quantum yield of ligand **29** to  $\text{Cu}^{2+}$  complex at the emission band 389 nm was 0.56, which was 560 times greater than that of the free ligand **29** at 0.001. The association constant ( $K_a$ ) of **29** with  $\text{Cu}^{2+}$  was found to be  $1.0 \times 10^4 \text{ M}^{-1}$ . On binding with  $\text{Cu}^{2+}$ , the absorption spectra exhibited a 42 nm blue shift because of hindered conjugation between the double bonds of **29**.

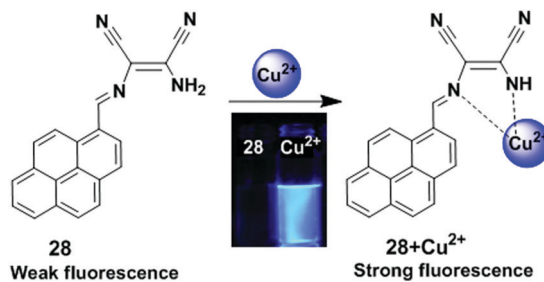


Fig. 24 Structure of chemosensor **28** and possible binding modes for  $\text{Cu}^{2+}$ . Reproduced from ref. 91 with permission from the Elsevier, copyright 2010.

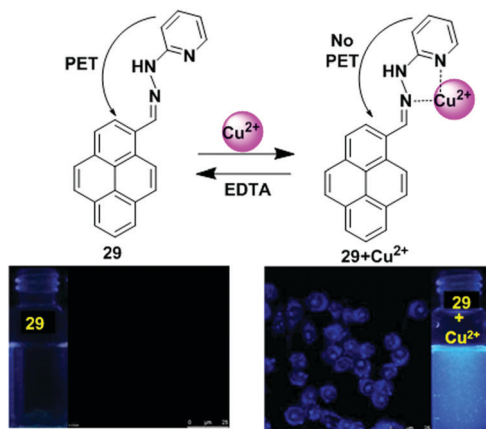


Fig. 25 Possible binding modes and fluorescence images of macrophage (RAW 264.7) cells treated with **29** (left) and then **29** +  $\text{Cu}^{2+}$  (right). Inset: Fluorescence colour changes of solution **29** before and after addition of  $\text{Cu}^{2+}$ . Reproduced from ref. 92 with permission from the Royal Society of Chemistry.

The colour also changed from light yellow to colourless. Moreover, confocal fluorescence microscopy imaging for detecting  $\text{Cu}^{2+}$  in living cells showed that chemosensor **29** can be used as an effective fluorescent probe due to its ( $<30\ \mu\text{M}$ ) low cytotoxicity and ability to easily penetrate cell membranes.

A Schiff-base fluorescent compound **30** was developed by exploiting the PET process for the detection  $\text{Cu}^{2+}$  ions (Fig. 26).<sup>93</sup> Ligand **30** ( $4 \times 10^{-6}\ \text{M}$ ) shows weak fluorescence

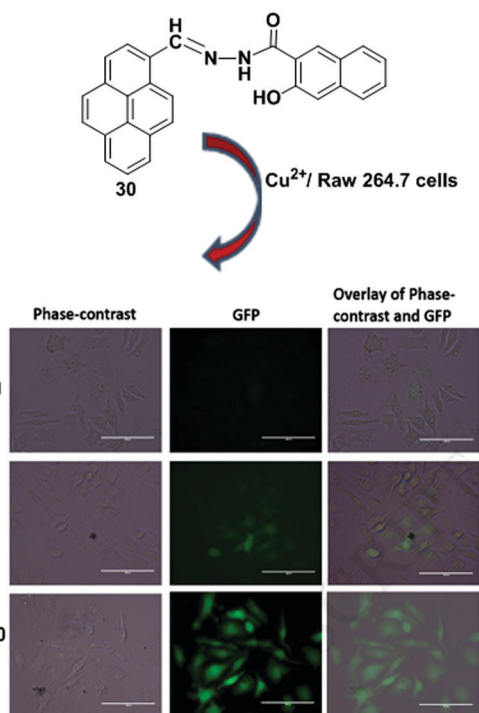


Fig. 26 Structure of ligand **30**. Fluorescence images of RAW 264.7 cells treated with chemosensor **30** and  $\text{Cu}^{2+}$  ions. Phase-contrast (left); GFP image ( $\lambda_{\text{ex}} = 393\ \text{nm}$ ) (middle); and overlay of phase-contrast and GFP (right). Reproduced from ref. 93 with permission from the Elsevier, copyright 2018.

because of PET. The fluorescence intensity was remarkably enhanced after gradual addition of  $\text{Cu}^{2+}$  ions (0–65 equiv.) to the chemosensor **30** in DMSO– $\text{H}_2\text{O}$  (1:1 v/v, HEPES = 50 mM, pH = 7.4). From the emission intensity data, the detection limit of chemosensor **30** +  $\text{Cu}^{2+}$  system was found to be  $0.26 \times 10^{-6}\ \text{M}$  with an association constant  $1.16 \times 10^4\ \text{M}^{-1}$ . A Job's plot experiment showed that the **30** to  $\text{Cu}^{2+}$  complex followed a 1:1 binding model. Treatment of chemosensor **30** with various metal ions such as  $\text{Pb}^{2+}$ ,  $\text{Ce}^{3+}$ ,  $\text{Cd}^{2+}$ ,  $\text{La}^{3+}$ ,  $\text{Mg}^{2+}$ ,  $\text{Zn}^{2+}$ ,  $\text{Ba}^{2+}$ ,  $\text{Bi}^{2+}$ ,  $\text{Hg}^{2+}$ ,  $\text{Fe}^{3+}$ ,  $\text{Ag}^+$ ,  $\text{Ni}^{2+}$ ,  $\text{Cr}^{3+}$ ,  $\text{Ca}^{2+}$ ,  $\text{Mn}^{2+}$ ,  $\text{Na}^+$ ,  $\text{K}^+$ ,  $\text{Co}^{2+}$ ,  $\text{Zr}^{2+}$ ,  $\text{Li}^+$ ,  $\text{Sr}^{2+}$ ,  $\text{Fe}^{2+}$ ,  $\text{Al}^{3+}$  and  $\text{Th}^{4+}$  (100 equiv.) resulted in insignificant changes in the fluorescence intensity, which indicate high selectivity of **30** towards the  $\text{Cu}^{2+}$  ion. Time-dependence fluorescence intensity indicated that sensor **30** completely binds with  $\text{Cu}^{2+}$  ion within 5 min. Moreover, the bio-imaging and spectroscopic methods confirmed its detection ability for  $\text{Cu}^{2+}$  ions in living cells.

Patra and co-workers have developed a chemosensor **31** containing a benzilmonohydrazone moiety for the detection of  $\text{Cu}^{2+}$ .<sup>94</sup> The chemosensor **31** exhibited a significant colour change from yellow to colourless in the presence of 10 equiv. of  $\text{Cu}^{2+}$  ions. The binding properties of **31** were further investigated by fluorescence titration experiments in acetonitrile–water (2/1, v/v). The fluorescence quantum yield increased from 0.035 to 0.67 after gradual addition of  $\text{Cu}^{2+}$ . In this case,  $\text{Cu}^{2+}$  is coordinated with the outer azino nitrogen atom of the ligand **31**, which inhibits the PET mechanism (Fig. 27). The detection limit of **31** based on fluorescent-titration experiments was found to be 7.8 nM for  $\text{Cu}^{2+}$ . The 1:1 stoichiometric complexation between **31** and  $\text{Cu}^{2+}$  was observed by  $^1\text{H}$  NMR spectroscopic data, the Job's plot and the ESI-MS spectrum. The ligand **31** was efficiently applied to real samples for the recognition of  $\text{Cu}^{2+}$  over the wide pH range of 4–11.

A new 2-aza-1,3-butadiene ionophore which connects two pyrene groups has been synthesized and studied as a colorimetric and fluorescent chemosensor **32** for  $\text{Cu}^{2+}$  ions (Fig. 28).<sup>95</sup> The UV-vis and fluorescence measurements illustrated the binding mechanism of receptor **32** ( $c = 2.5 \times 10^{-5}\ \text{M}$ ) with several metal

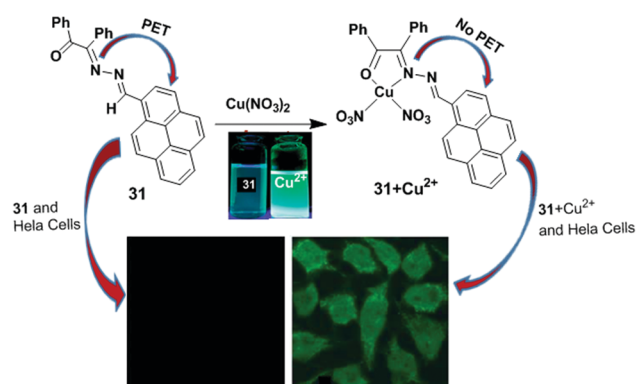


Fig. 27 Representation of possible sensing mechanism of probe **31**. Fluorescence images of HeLa cells; cells incubated with probe **31** only and cells incubated with **31** +  $\text{Cu}^{2+}$ . Reproduced from ref. 94 with permission from the Royal Society of Chemistry.

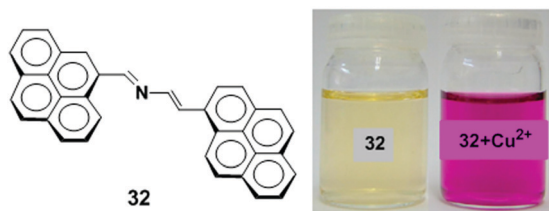


Fig. 28 Molecular structure of receptor **32**. Visual changes in the colour of **32** (left) and after addition of Cu<sup>2+</sup> ion (right). Reproduced from ref. 95 with permission from the Elsevier, copyright 2010.

cations (Li<sup>+</sup>, Na<sup>+</sup>, K<sup>+</sup>, Mg<sup>2+</sup>, Ca<sup>2+</sup>, Ni<sup>2+</sup>, Cu<sup>2+</sup>, Zn<sup>2+</sup>, Cd<sup>2+</sup>, Hg<sup>2+</sup>, Pb<sup>2+</sup>, Sm<sup>3+</sup>, Eu<sup>3+</sup>, Yb<sup>3+</sup> and Lu<sup>3+</sup>) in CH<sub>3</sub>CN. The receptor **32** showed selective changes for Cu<sup>2+</sup> and Hg<sup>2+</sup> ions with a visible colour change by naked eye detection. Compound **32** exhibits a very weak fluorescence due to PET quenching from the lone pair electrons on the nitrogen atom in the 2-azadiene bridge to the excited state of the pyrene moiety when excited at 350 nm ( $\Phi = 0.014$ ). Upon addition of Cu<sup>2+</sup> to **32**, a chelation enhanced fluorescence (CHEF) as well as excimer emission band was observed which was 27 times stronger in acetonitrile and 2.7 times stronger in an acetonitrile/water (7:3) solvent system. Moreover, the detection limit in both solvent systems was 10<sup>-6</sup> M with a 1:1 stoichiometric for the **32** to Cu<sup>2+</sup> complex.

A new fluorescent chemosensor **33** incorporating 1-nitronyl nitroxide pyrene has been developed as an off-on sensor for the selective recognition of Cu<sup>2+</sup> (Fig. 29).<sup>96</sup> As a Cu<sup>2+</sup> selective sensor, the fluorogenic behaviour of **33** (10 mM) was investigated in CH<sub>3</sub>CN in which a weak fluorescence signal at 378 nm for the free solution of **33** remarkably increased at 444 nm after addition of 10.0 equiv. of Cu<sup>2+</sup>. This phenomenon can be explained by the inhibition of PET from nitronyl nitroxide to the pyrene moiety. An ESR experiment confirmed the inhibition caused by the coordination between Cu<sup>2+</sup> and nitronyl nitroxide of **33**.

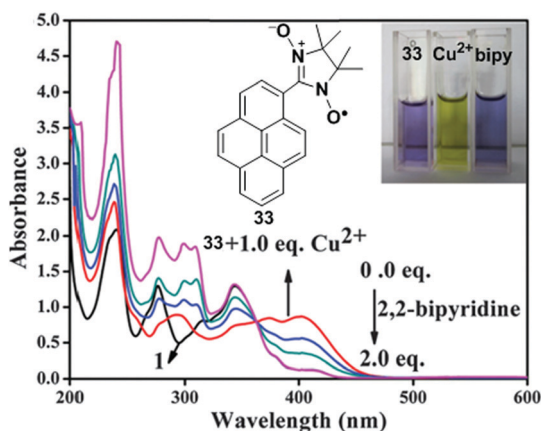


Fig. 29 The UV-vis absorption spectra of **33** + Cu<sup>2+</sup> and upon addition of 2,2'-bipyridine in CH<sub>3</sub>CN. Inset: Visual changes of the colour of compound **33**; compound **33** + Cu<sup>2+</sup>; compound **33** + Cu<sup>2+</sup> + 2,2'-bipyridine (from left to right). Reproduced from ref. 96 with permission from the Centre National de la Recherche Scientifique (CNRS) and the Royal Society of Chemistry.

On the other hand, the fluorescence intensity of **33** showed no significant changes with various other metal ions (Li<sup>+</sup>, Na<sup>+</sup>, Ba<sup>2+</sup>, Fe<sup>3+</sup>, Fe<sup>2+</sup>, Mn<sup>2+</sup>, Mg<sup>2+</sup>, Ag<sup>+</sup>, Pb<sup>2+</sup>, Cd<sup>2+</sup>, Co<sup>2+</sup>, Ni<sup>2+</sup>, Zn<sup>2+</sup>, Hg<sup>2+</sup>). Moreover, the absorbances at 345 nm and 276 nm reduced sharply and increased at 402 and 295 nm and were accompanied with four well-defined isosbestic points, which indicated **33** + Cu<sup>2+</sup> complexation; the colour changed from purple to bright yellow. In contrast, a reversible absorption response was observed at 345 nm by titration of 2,2'-bipyridine with the **33** + Cu<sup>2+</sup> complex, which resulted in the recovery of the colour which is probably due to a stronger coordination between Cu<sup>2+</sup> and 2,2'-bipyridine. Furthermore, the Job's plot calculated a 1:1 binding ratio between Cu<sup>2+</sup> and **33** with a detection limit of 3.60 × 10<sup>-7</sup> M in CH<sub>3</sub>CN.

### 5.3. Chemosensor based on PCT mechanism

The calix[4] crown fluorescent chemosensor **34** bearing two facing pyreneamide groups has been designed based on the PCT mechanism (Fig. 30).<sup>97</sup> After addition of Pb<sup>2+</sup> or Cu<sup>2+</sup>, probe **34** showed fluorescence quenching in both the excimer and monomer emissions owing to reverse PET and conformational changes. The **34**/Cu<sup>2+</sup> complex gave a significant red-shifted excitation spectrum in an acetonitrile solvent system at the excimer wavelength 470 nm in comparison with monomer wavelength 380 nm, which suggests that the two-pyrene groups form the static excimer. Moreover, in both the fluorescence and absorption spectra, wavelength shifts of **34** are noted on binding of Cu<sup>2+</sup> with the nitrogen atoms of the amide groups, resulting in a PCT mechanism.

### 5.4. Chemosensor based on LMCT process

The LMCT process occurs in a complex when electrons are transferred from a molecular orbital ( $\sigma$ ,  $\sigma^*$ ,  $\pi$ ,  $\pi^*$  and non-bonding) of the ligand to the empty or partially filled metal d-orbitals. Therefore, a reduction of the metal occurs in LMCT transitions. In this case, this type of transfer predominantly occurs in the case of ligands having relatively high energy lone pairs atoms (example O, S or Se) or when the metal has low lying empty orbitals. The LMCT based fluorescent sensors

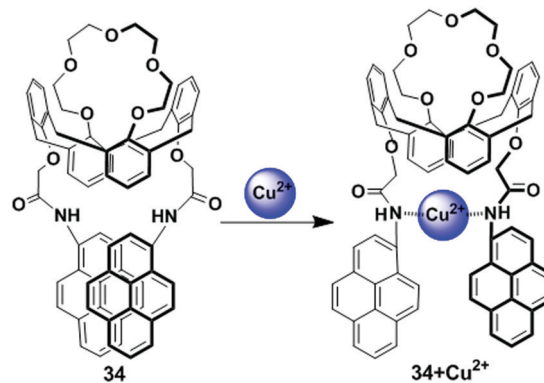


Fig. 30 Schematic representation of proposed sensing mechanism for probe **34** in presence of Cu<sup>2+</sup>. Reproduced from ref. 97 with permission from the American Chemical Society, copyright 2006.

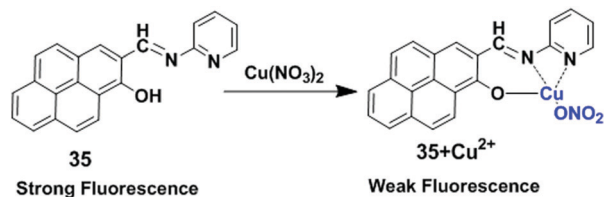


Fig. 31 The possible binding modes of **35** with  $\text{Cu}^{2+}$ . Reproduced from ref. 99 with permission from the Elsevier, copyright 2017.

depend on the selection of chromophore such as a pyrene unit, along with the nature and location of the receptor. The metal ion binding with a receptor will generally lead to a change in the absorption spectra or quenching of the fluorescence of the pyrene fluorophore.<sup>98</sup>

In 2017, Lu and co-workers developed chemosensor **35** by combining pyrene with a pyridine unit based on LMCT which exhibited high selectivity for  $\text{Cu}^{2+}$  and  $\text{Fe}^{3+}$  over other ions (Fig. 31).<sup>99</sup> Sensor **35** exhibited stable absorption or fluorescence intensity over a wide range of pH from 3 to 12. Addition of  $\text{Cu}^{2+}$  to ligand **35** partially quenched the emission of the fluorophore through electron and/or energy transfer processes due to its paramagnetic nature with an unfilled d orbital. DFT calculations of **35** +  $\text{Cu}^{2+}$  also fully confirmed the LUMO was distributed more over the C=N bond and receptor rather than the pyrene unit after binding with  $\text{Cu}^{2+}$ , than that of the free compound **35**. The UV-vis absorption spectra of **35** in DMF-HEPES buffer (2 : 8, v/v, pH = 7.4) showed a red shift (21 nm) from 352 nm to 373 nm after addition of 40 equiv. of  $\text{Cu}^{2+}$  with an optical colour change from colourless to pink. The limit of detection toward  $\text{Cu}^{2+}$  was 8.5  $\mu\text{M}$ . The Job's plot experiment revealed a 1 : 1 binding stoichiometry for **35** and  $\text{Cu}^{2+}$ .

Another example of the LMCT mechanism was shown in the pyrene-based turn-off chemosensor **36** that was synthesized from the condensation reaction of 1-aminopyrene and 2,4-dihydroxy benzaldehyde (Fig. 32).<sup>100</sup> Sensor **36** exhibited a high

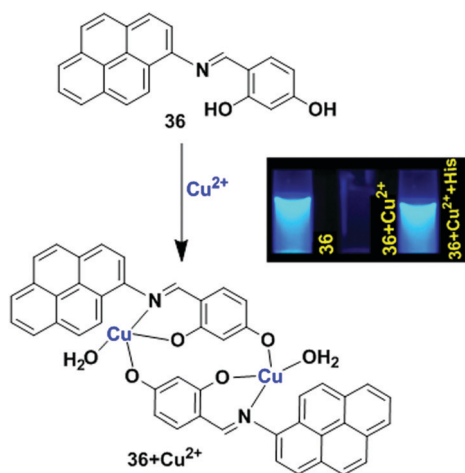


Fig. 32 Schematic representation of possible sensing mechanism of probe **36** with  $\text{Cu}^{2+}$ . Reproduced from ref. 100 with permission from the John Wiley and Sons, copyright 2018.

selectivity for recognizing  $\text{Cu}^{2+}$  ion in the presence of other metal ions. The UV-vis absorption spectroscopy of **36** with  $\text{Cu}^{2+}$  exhibited a hypsochromic shift for the absorption peaks at 368 nm and 340 nm with the formation of two isosbestic points in  $\text{CH}_3\text{CN}/\text{H}_2\text{O}$  (9 : 1, v/v). Meanwhile, in aqueous medium, the solution colour of **36** changed from yellow to brown even at lower concentrations of  $\text{Cu}^{2+}$ , and this was also directly observed by the naked eye. Moreover, the emission peak at 429 nm of **36** (20  $\mu\text{M}$ ) was gradually quenched after treatment with  $\text{Cu}^{2+}$  in which the quenching efficiency was 93%. This phenomenon is due to the LMCT characteristics resulting in the quenching of the fluorescence chemosensor **36** with  $\text{Cu}^{2+}$ . The time-resolved fluorescence spectroscopy also confirmed the static quenching mechanism for **36** +  $\text{Cu}^{2+}$ . A Job's plot experiment and ESI-mass spectroscopic analysis revealed a 2 : 2 stoichiometry for binding of **36** with  $\text{Cu}^{2+}$ . The lower limit of detection of  $\text{Cu}^{2+}$  by **36** is 0.503  $\mu\text{M}$ . The sensor **36** +  $\text{Cu}^{2+}$  underwent a reversibility test in the presence of histidine and regained its original colour. Therefore, the sensor **36** can be utilized in real samples to investigate trace amounts of copper.

### 5.5. Chemosensor based on ILCT mechanism

A ligand sometimes has a donor and an acceptor site simultaneously. The ILCT takes place within the ligands mediated by a metal. In most instances, these complexes exhibit long-lived excited states with low-energy absorptions. Generally, this kind of chemosensor is fluorescent in the presence of a metal ion.<sup>101</sup>

Using the above technique, Mukherjee *et al.* reported a new simple, reversible, turn-on luminescent chemosensor, namely the hydrazone based pyrene derivative **37** for the detection of  $\text{Cu}^{2+}$  (Fig. 33).<sup>102</sup> The binding of the receptor **37** with  $\text{Cu}^{2+}$  was studied in DMSO/ $\text{H}_2\text{O}$  (2 : 1) medium, and the generation of new peaks at 512 nm (broad) and 345 nm through isosbestic points was evident with a colour change from yellow to reddish brown. The emission intensity of **37** was enhanced at 495 nm up to 44-fold upon addition of  $\text{Cu}^{2+}$  in DMSO solvent due to ILCT process. The other metal ions ( $\text{Hg}^{2+}$ ,  $\text{Pb}^{2+}$ ,  $\text{Cd}^{2+}$ ,  $\text{Ni}^{2+}$ ,  $\text{Co}^{2+}$ ,  $\text{Fe}^{2+}$ ,  $\text{Fe}^{3+}$ ,  $\text{Mn}^{2+}$ ,  $\text{Zn}^{2+}$ ,  $\text{Al}^{3+}$  and  $\text{Cr}^{3+}$ ) caused no significant change in the emission intensity of **37**. The limit of detection (LOD) was found to be of the order of  $10^{-8}$  M. DFT experiments revealed that the electron spin density was focused only on the pyrene moiety for all the frontier molecular orbitals which also

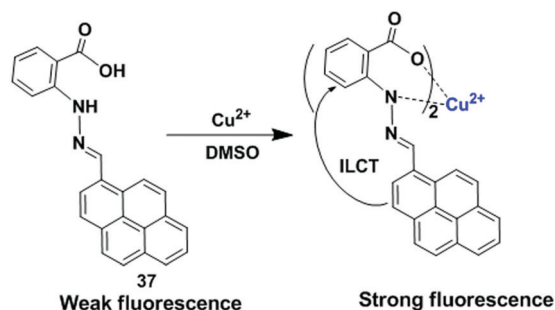


Fig. 33 The sensing mechanism of **37** with  $\text{Cu}^{2+}$ . Reproduced from ref. 102 with permission from the Elsevier, copyright 2018.

supports the ILCT process. The compound **37** can extract  $\text{Cu}^{2+}$  with 94% extraction efficiency over the pH range of 6.5–11 from an aqueous mixture of metal ions or from real samples by a selective two-phase liquid–liquid extraction using a water–dichloromethane mixture (1:1). During extraction, the **37** +  $\text{Cu}^{2+}$  complex was found to have a 2:1 ratio of ligand and  $\text{Cu}^{2+}$ . Good recyclability and reusability were observed for chemosensor **37** as the fluorescence emission reversed to its original condition in the presence of  $\text{C}_2\text{O}_4^{2-}$  and the **37** +  $\text{Cu}^{2+}$  complex, and this characteristic was used to “INHIBIT” the logic gate application. As an analysis tool, a smartphone can also be used for detecting colour change of **37** during  $\text{Cu}^{2+}$  extraction.

### 5.6. Chemosensor based on AIE mechanism

In an AIE luminogen, the intramolecular rotations of aromatic compounds are active in dilute solutions, which serve as a relaxation channel for its excitons resulting in non-radiative decay. However, in the aggregate state, the intramolecular rotations are suppressed because of the physical constraints, which opens the radiative pathway to emit efficiently. In the crystalline state, multiple C–H– $\pi$  hydrogen bonds are formed between the hydrogen atoms of the aromatic rings of one ligand and the  $\pi$  electrons of the aromatic rings of another ligand. These hydrogen bonds stiffen the conformations of the ligand and enhance their light emission.<sup>103</sup>

For these special characteristics of light emission, Wang and co-workers have used AIE or an AEE luminogen to develop chemosensors of type **38** for  $\text{Cu}^{2+}$  ion detection.<sup>104</sup> In this case, pyrene acted as the strong fluorophore and was combined with Schiff bases which were responsible for the AIE properties in semi-aqueous solution toward  $\text{Cu}^{2+}$  (Fig. 34). The AEE behaviour of **38** (10  $\mu\text{M}$ ) was explained by measurement of the fluorescence spectra in different volumes of water in mixed  $\text{H}_2\text{O}/\text{DMF}$  (from 0% to 100%) solution. The experiment suggests that with increasing percentage of water the peak continually showed a red shift because of amide–amidic acid tautomerization with water in the aggregation state of **38**. Time-resolved fluorescence measurements supported the above observations. Sensor **38** became strongly emissive at 455 nm in the presence of  $\text{Cu}^{2+}$  (5 eq.) with a quantum yield  $\Phi$  from 0.09 to 0.58 in  $\text{H}_2\text{O}/\text{DMF}$  (1:1, v/v). The UV-vis absorption spectroscopy also established that self-assembly of **38** (10  $\mu\text{M}$ ) prompted by  $\text{Cu}^{2+}$  involves a coordination interaction and changes in the aggregation form. A Job's plot experiment evaluated from the fluorescence spectra confirmed that the binding of **38** to  $\text{Cu}^{2+}$  followed a 2:1 stoichiometry.

Another example of aggregate formation was observed in a pyrene-based hydrophobic hydrocarbon framework in a binary water–solvent system.<sup>105</sup> Das *et al.* have reported a chemosensor **39** in which the pyrene part acts as a fluorescence reporter and the antipyrene part containing the pyrazolone unit plays the role of a chelating moiety (Fig. 35). Sensor **39** exhibited a very weak fluorescence due to a PET process between N donors in the imine bond and the pyrene ring. Upon interaction with  $\text{Cu}^{2+}$ , a fluorescence enhancement of **39**

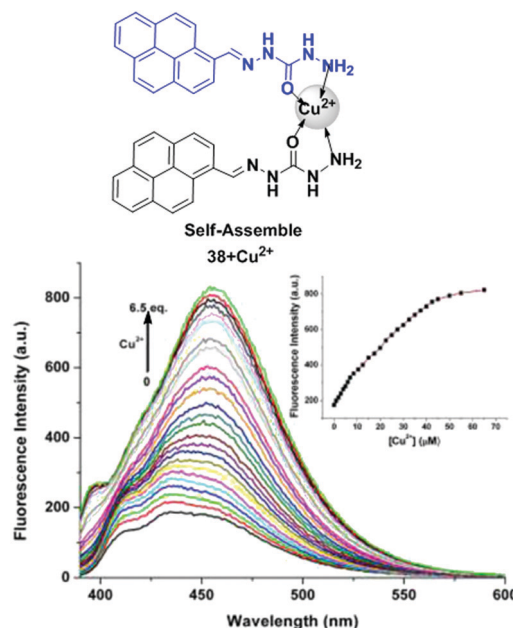


Fig. 34 The AIE luminescence mechanism of **38** and  $\text{Cu}^{2+}$ . The fluorescence spectra with  $\text{Cu}^{2+}$  ions. Inset: Emission intensity at 455 nm with  $\text{Cu}^{2+}$  concentration. Reproduced from ref. 104 with permission from the Elsevier, copyright 2018.

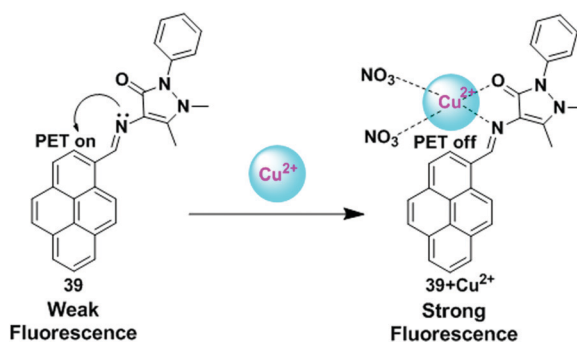


Fig. 35 Schematic representation of  $\text{Cu}^{2+}$  sensing by ligand **39**. Reproduced from ref. 105 with permission from the Elsevier, copyright 2018.

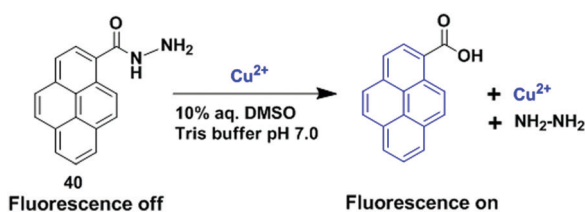
was observed at 454.5 nm along with 440 nm and 509 nm, which indicated complex formation due to prevention of the PET process by an N donor site in acetonitrile solvent. Time fluorescence titration revealed the maximum fluorescence enhancement of **39** was observed at 432 nm (blue shift) up to the addition of 150  $\mu\text{M}$  of  $\text{Cu}^{2+}$  ion. However, with an increasing percentage of water in the acetonitrile solvent system, the fluorescence emission intensity of **39** gradually increased at 454.5 nm with a red shift with poor intensity in pure acetonitrile. At 80% of water fraction, there was 230 times increase in the emission intensity with a red shift of 16 nm. This was because of the AIE characteristics of probe **39**. The critical aggregate concentration of ligand **39** in this solvent system was calculated to be 23.4  $\mu\text{M}$ . Moreover, the antioxidant nature of the compound **39** was confirmed from UV studies.

### 5.7. $\text{Cu}^{2+}$ -Promoted reaction based chemosensors

The development of chemodosimeters that operates on the basis of analyte selective chemical reactions has attracted a great attention in the last few years. Among them,  $\text{Cu}^{2+}$ -selective reaction-based probes show a significant fluorescence emission enhancement with high selectivity. In this case,  $\text{Cu}^{2+}$ -assisted hydrolysis, oxidation, reduction and spirolactam ring-opening methods have been used for building  $\text{Cu}^{2+}$ -selective reaction-based probes. Generally, fluorescent sensors of this type lead to non-emissive precursors to fluorescent products through irreversible chemical reactions. It is noticeable that  $\text{Cu}^{2+}$  induced-catalytic reactions to develop fluorescent sensors are the easiest way to avoid the consideration of the paramagnetic nature of  $\text{Cu}^{2+}$  as fluorescent products have a slight attraction to  $\text{Cu}^{2+}$ .<sup>106</sup>

Using this concept, Chang *et al.* have reported a chemosensor **40** in which the pyrene moiety acts as a fluorophore and a fluorescence-quenching hydrazide moiety plays the role of a signaling handle (Fig. 36).<sup>107</sup> The pyrenecarbohydrazide probe **40** displayed a high selectivity towards  $\text{Cu}^{2+}$  ions *via*  $\text{Cu}^{2+}$ -induced catalytic hydrolysis to pyrenecarboxylic acid and hydrazine. <sup>1</sup>H NMR spectroscopy and mass spectrometry also confirmed the hydrolysis of probe **40**. It exhibited a very weak fluorescence due to PET which was remarkably enhanced (130-fold) at 392 nm upon addition of  $\text{Cu}^{2+}$  in 10% aqueous DMSO solution (Tris-buffered at pH 7.0). Moreover, the  $\text{Cu}^{2+}$  selective fluorescence signaling behaviour of **40** was less prominent as the space between the hydrazide functionality and the pyrene fluorophore increased. Probe **40** has a wide application in environmentally related samples, especially for semiconductor waste water sample over a wide pH range. The detection limit of **40** for  $\text{Cu}^{2+}$  was  $5.93 \times 10^{-8}$  M and  $6.93 \times 10^{-8}$  M (0.005 ppm) in the waste water sample.

Rhodamine derivatives as chemosensors for  $\text{Cu}^{2+}$  detection were first utilized by Czarnik in 1997. Even now, they are used for the detection of cations and anions given their excellent photophysical properties and the ring opening process. In general, the carbonyl group in the rhodamine spirolactam form gets activated by complexation with specific metal ions using certain solvent systems and pH.<sup>108</sup> Kim *et al.* introduced a novel fluorescent sensor, a rhodamine based derivative bearing a 1,8-naphthalimide group which performed as a dual-mode sensor for  $\text{Cu}^{2+}$  using two mechanisms, one is the rhodamine ring-opening mechanism and the other, a ratiometric displacement



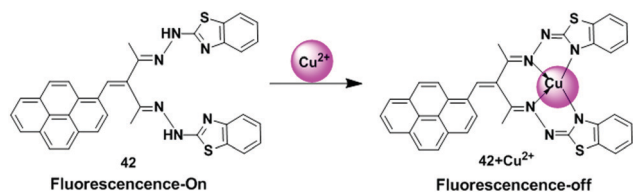


Fig. 38 Proposed sensing mechanism of probe **42** for  $\text{Cu}^{2+}$ . Reproduced from ref. 112 with permission from the John Wiley and Sons, copyright 2020.

Upon complexation with the paramagnetic  $\text{Cu}^{2+}$  ions, quenching interactions probably dominate the emission characteristics of these complexes. The probe **42** displays a high sensitivity and selectivity in DMSO- $\text{H}_2\text{O}$  (8:2 v/v) for  $\text{Cu}^{2+}$  due to chelation with **42**. The ligand **42** +  $\text{Cu}^{2+}$  ensemble shows a high sensitivity towards  $\text{S}^{2-}$  ions. The UV-vis titration of **42** showed that the absorption bands at 395 and 410 nm gradually reduced and the band at 455 nm increased upon addition of  $\text{Cu}^{2+}$  (0–100  $\mu\text{M}$ ). The association constant ( $K_a$ ) of **42** with  $\text{Cu}^{2+}$  ions was found to be  $2.3 \times 10^5 \text{ M}^{-1}$ . The change in the appearance of solution **42** from the yellow to brown colour was observed after the addition of  $\text{Cu}^{2+}$  ions (100  $\mu\text{M}$ ) which can be seen by the naked eye. Moreover, the addition of  $\text{Cu}^{2+}$  to the solution of **42** gradually quenched the fluorescence emission intensity at 510 nm which was explained by the paramagnetic nature and the CHEQ effect of the  $\text{Cu}^{2+}$  ions. The ligand **42** was cell-permeable, and was efficiently utilized for the recognition of copper ions in living cells and in real water samples.

Similarly, quenching is promoted by paramagnetic  $\text{Cu}^{2+}$  ions *via* the heavy atom effect in a pyrene-based dipicolylamine derivative sensor **43** (Fig. 39).<sup>113</sup> The compound **43** selectively recognized  $\text{Cu}^{2+}$  and  $\text{Fe}^{3+}$  over the other metal cations  $\text{Ni}^{2+}$ ,  $\text{Mg}^{2+}$ ,  $\text{Cd}^{2+}$ ,  $\text{Hg}^{2+}$ ,  $\text{Na}^+$ ,  $\text{K}^+$ ,  $\text{Ca}^{2+}$ ,  $\text{Co}^{2+}$ ,  $\text{Cr}^{3+}$ ,  $\text{Pb}^{2+}$  and  $\text{Zn}^{2+}$ . Upon treatment of  $\text{Cu}^{2+}$  ions, sensor **43** exhibited well-defined absorption bands in MeOH solution in which a newly appeared band at 660 nm with red shifted peaks at 290, 333 and 358 nm were observed owing to the d-d transition of  $\text{Cu}(\text{II})$ . This phenomenon was distinguished by naked eye detection. From the results of UV-vis experiments, the stoichiometry of the Job's plot was found to be 1:1 with the binding constant

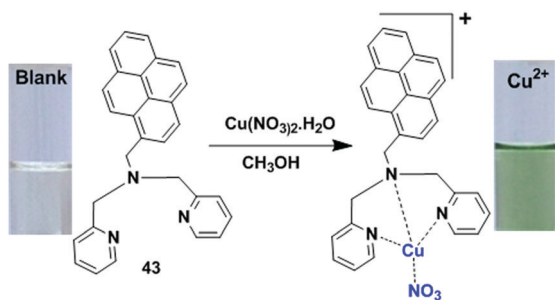


Fig. 39 Possible binding interaction and colorimetric change of ligand **43** with  $\text{Cu}^{2+}$ . Reproduced from ref. 113 with permission from the Elsevier, copyright 2017.

$1.48 \times 10^5 \text{ M}^{-1}$  for the **43** +  $\text{Cu}^{2+}$  system. Moreover,  $^1\text{H}$  NMR spectroscopic titration experiments in  $\text{CD}_3\text{OD}$  revealed that the seventeen aromatic protons of free receptor **43** in the 7.00–8.85 ppm region disappeared in the presence of  $\text{Cu}^{2+}$  ions due to the paramagnetic effect of the  $\text{Cu}^{2+}$  ( $d^9$  system). The fluorescence spectrum of chemosensor **43** in methanol solution showed an emission band at 392 nm ( $\Phi \sim 0.31$ ) which was quenched significantly ( $\Phi \sim 0.05$ ). This behaviour was observed because of the efficient non-radiative deactivation by  $\text{Cu}^{2+}$ , which resulted in the enhanced spin-orbit coupling associated with the heavy atom effect of the complexed  $\text{Cu}^{2+}$ .

Pandey *et al.* investigated the difference in the photophysical properties of probes **44**, where coumarin-pyrene conjugates form with the non-conjugated form of probe **45** for the selective detection of  $\text{Cu}^{2+}$  (Fig. 40).<sup>114</sup> Herein, it was observed that the paramagnetic nature of the  $\text{Cu}^{2+}$  ions have a great influence on the conjugated system of **44** rather than **45**. The probe **44** has a conjugated system exhibiting C-H- $\pi$ ,  $\pi$ - $\pi$  interactions, and H-bonding interactions. Successive addition of  $\text{Cu}^{2+}$  ions (1–10  $\mu\text{M}$ ) results in the gradual quenching of fluorescence at 430 nm indicating the high sensitivity of probe **44** towards  $\text{Cu}^{2+}$  ions. The phenomenon behind the quenching of fluorescence can be attributed to the combined effect of donation of electrons from the fluorophore fragment to the adjacent metal ion and is due to the paramagnetic nature of the  $\text{Cu}^{2+}$  ions. The binding constant of **44** with  $\text{Cu}^{2+}$  was calculated to be  $2.4 \times 10^4 \text{ M}^{-1}$ . Besides, sensor **45** showed no prominent change due to its non-conjugated form. Moreover, sensor **44** can be effectively applied in medical sciences for the detection of  $\text{Cu}^{2+}$  levels in kidney cell lining using its ability for quenching fluorescence.

Yamato and co-workers have described the reverse PET mechanism for the detection of paramagnetic  $\text{Cu}^{2+}$  ions by the heteroditopic receptor **46**.<sup>115</sup> As  $\text{Cu}^{2+}$  has an unfilled d orbital, it probably quenches the emission of the fluorophore *via* electron transfer.<sup>116</sup> The receptor **46** has a thiacalix[4]arene moiety with two different side arms in which two pyrene-appended triazole rings are incorporated at one side of the

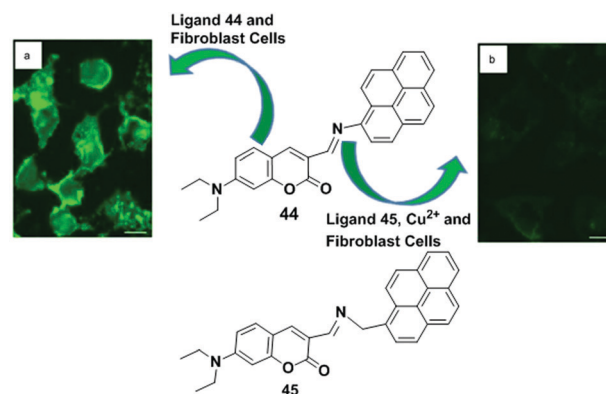


Fig. 40 Structure of chemosensors **44** and **45**. (a) Fluorescence images of **44** in a kidney cell line: cos-7 cells treated with **44** (5 mM, 20 min); (b) loss of fluorescence after addition of  $\text{CuCl}_2$  (10 mM, 30 min). Reproduced from ref. 114 with permission from the Elsevier, copyright 2016.

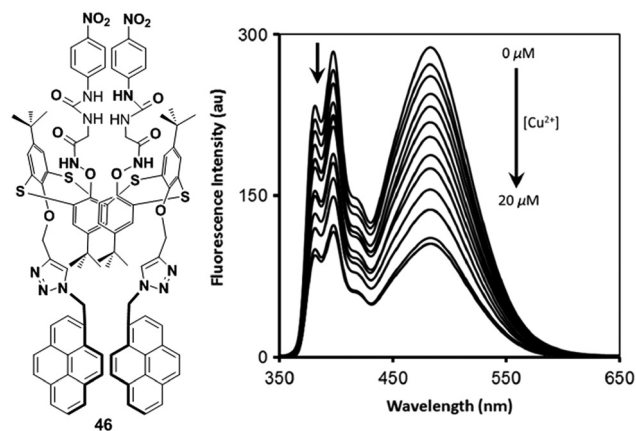


Fig. 41 The molecular structure of fluorescence chemosensor **46**, showing fluorescence quenching in the presence of  $\text{Cu}^{2+}$  ions. Reproduced from ref. 115 with permission from the Elsevier, copyright 2014.

thiacalix[4]arene cavity and the other side contains two urea moieties with various phenyl groups (Fig. 41). Herein, an effective positive allosteric effect of **46** was also observed for  $\text{Ag}^+$  with  $\text{Cl}^-$ . Besides, the UV-vis absorption, fluorescence spectra and  $^1\text{H}$  NMR spectroscopic titration experiments of **46** showed changes in the presence of transition metal cations ( $\text{Ag}^+$ ,  $\text{Cu}^{2+}$  and  $\text{Hg}^{2+}$ ) and anions in the  $\text{CH}_2\text{Cl}_2$ -DMSO solvent system. The interaction of ligand **46** with  $\text{Ag}^+$  ion increased the monomer emission at 393 nm and decreased the excimer emission at 486 nm with a 1 : 1 binding stoichiometry. Furthermore, upon addition of  $\text{Cu}^{2+}$ , monomer and excimer emissions of free **46** were significantly quenched. This was because the binding with the pyrene-appended triazole groups caused a reverse PET from the pyrene moieties to the triazole groups. The association constant for the complexation of **46** +  $\text{Cu}^{2+}$  was found to be  $330\,000 \pm 23\,100\text{ M}^{-1}$ .

### 5.9. Miscellaneous

As discussed in the preceding part of this review, chemosensors operating using different well-known mechanisms have been reported. However, there are still other types of chemosensors which do not belong to the above classified mechanisms or they fall into the category of following several mechanisms for  $\text{Cu}^{2+}$  detection. Herein, we will try to briefly discuss these sensors.

Two new fluorescent sensors **47**, **48** based on thiacalix[4]arenes bearing two pyrene groups were developed by Kumar and co-workers for the recognition of  $\text{Cu}^{2+}$  and  $\text{CN}^-$  ions (Fig. 42).<sup>117</sup> The authors reported that the former receptor **47** undergoes a reverse PET process, whereas the latter **48** follows the monomer and excimer emission mechanism with a ratiometric response for  $\text{Cu}^{2+}$  detection in  $\text{CH}_2\text{Cl}_2/\text{CH}_3\text{CN}$  (1 : 1, v/v). Moreover, the UV-vis absorption spectra of both receptors upon interaction with  $\text{Cu}^{2+}$  showed a decrease of the blue shift band and the formation of a new red shift band. This is because of the collaboration between the  $\text{Cu}^{2+}$  ion with the nitrogen atoms of the amide groups which reduces the electron donating ability of the N atom and induces the red shift absorption spectra due to PCT mechanism. The strong blue fluorescence of

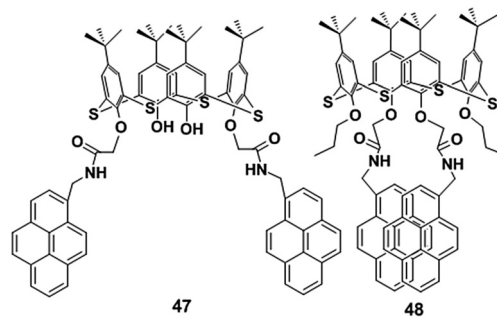


Fig. 42 Molecular structure of receptors **47** and **48**.

compound **47** indicates the typical monomer emission of pyrene at 377 nm which confirms there is no  $\pi$ - $\pi$  interaction between two pyrene units. Upon addition of  $\text{Cu}^{2+}$  ions (6.0–600  $\mu\text{M}$ ) to the solution of **47** (6.0  $\mu\text{M}$ ), the fluorescence emission intensity significantly decreased due to reverse PET from the pyrene units to the nitrogen atom. The association constant  $K_a$  of **47** with  $\text{Cu}^{2+}$  was calculated to be  $8.55 \times 10^5\text{ M}^{-1}$ . On the other hand, the fluorescence properties of compound **48** were completely different compared with compound **47**. The compound **48** showed a weak monomer emission at 375 nm and a strong intramolecular excimer emission at 467 nm. Upon treatment with 60 mM (10 equiv.) of  $\text{Cu}^{2+}$  ions, the titration profile of **48** exhibited a ratiometric response with monomer enhancement and a excimer emission quenching. The relative intensity ratio of monomer to excimer emission ( $I_M/I_E$ ) of free ligand **48** was 0.47 and it increased by 11.7-fold to 5.52 on addition of  $\text{Cu}^{2+}$  ion. A selectivity test of **47** and **48** was carried out in the presence of various metal ions with  $\text{Cu}^{2+}$  which revealed **47** +  $\text{Cu}^{2+}$  and **48** +  $\text{Cu}^{2+}$  complexes to be selective chemosensors. This observation was in agreement with the 1 : 1 and 1 : 2 (H/G) binding model for **47** +  $\text{Cu}^{2+}$  and **48** +  $\text{Cu}^{2+}$  complexes respectively, which was also confirmed by the method of continuous variation (Job's plot).

A pyrene containing Schiff base colorimetric sensor **49** was synthesized *via* a simple one-pot reaction (Fig. 43).<sup>118</sup> The probe **49** displayed a high sensitivity and selectivity towards  $\text{Cu}^{2+}$  and the sensitivity was not affected with by the presence of other relevant metal ions. In the solvent mixture DMSO/ $\text{H}_2\text{O}$  (v/v = 8/2, buffered with HEPES, pH = 7.4), the absorption peaks of probe **49** at 355 and 452 nm were slowly decreased on gradual addition of  $\text{Cu}^{2+}$  and the peaks were blue-shifted to 326 and

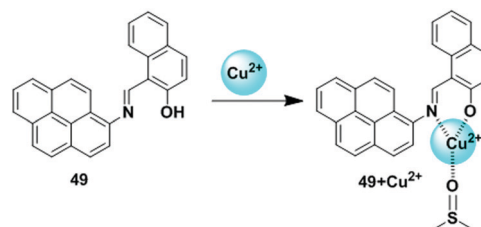


Fig. 43 Schematic representation of possible sensing mechanism of probe **49** for  $\text{Cu}^{2+}$ . Reproduced from ref. 118 with permission from the Elsevier, copyright 2014.

410 nm with two isosbestic points at 349 and 414 nm, respectively with changes of colour from yellow to pale lemon. This phenomenon is due to the formation of the  $\text{Cu}^{2+}$ -assisted  $49\text{-Cu}^{2+}$ -DMSO complex. The absorption spectrum on changing the  $\text{Cu}^{2+}$  concentration indicated a 1:1 stoichiometry between the host **49** and the  $\text{Cu}^{2+}$  ions.  $^1\text{H}$  NMR spectroscopic titration experiments of **49** also found that the proton signal of the OH became broad and slightly shifted downfield with increasing  $\text{Cu}^{2+}$  concentration. The detection limit of **49** for  $\text{Cu}^{2+}$  ions was calculated to be  $(2.17 \pm 0.02) \times 10^{-6}$  M.

In 2010, Yen and coworkers synthesized a novel colorimetric and fluorometric receptor **50** containing a pyrene unit and a 4-methylphenylthiourea moiety for selectively sensing  $\text{Cu}^{2+}$  and  $\text{Hg}^{2+}$  (Fig. 44).<sup>119</sup> The coordination with  $\text{Cu}^{2+}$  and  $\text{Hg}^{2+}$  ions show colour and fluorescence changes of **50** in aqueous solution (DMSO/ $\text{H}_2\text{O}$  = 4/1, buffered with HEPES, pH 7.8) which allowed them to be distinguished from other metal ions. Moreover, the designed sensor **50** can recognize  $\text{Cu}^{2+}$  through binding with the thiourea group. For this reason, the typical pyrene absorption bands of **50** in the region of 235–350 nm gradually increased at 278, 334 nm with a shoulder peak at 388 nm and decreased at 348 nm after addition of  $\text{Cu}^{2+}$ . The colour of the **50** +  $\text{Cu}^{2+}$  solution changed from pale yellow to green-yellow. Moreover, the enhancement of emission intensity (45 times more) at 396 and 439 nm was observed for **50** upon treatment with  $\text{Cu}^{2+}$  ions, with a colour change from pale to strong blue. The  $\text{Cu}^{2+}$  ion-induced complexation was also confirmed by NMR and ESI-MS spectra. The Job's plot determined the 1:1 stoichiometry with  $1.09 \times 10^4$   $\text{M}^{-1}$  binding constant for the **50** +  $\text{Cu}^{2+}$  complex.

Liu *et al.* have developed the pyrene based diaminomaleonitrile chemosensors **51** and **52** ( $1 \times 10^{-5}$  M) which can effectively sense  $\text{Cu}^{2+}$  in acetonitrile–water solution (v/v = 1:1, 10 mM HEPES, pH = 7) (Fig. 45).<sup>120</sup> The absorption spectra of **51** exhibits a strong red-shifted band at approximately 421 nm from that of pure pyrene at around 355 nm. On the other hand, compound **52** showed the characteristic absorption band of pyrene at 342 nm and another strong band at 380 nm, owing to the ICT mechanism. Upon gradual addition of  $\text{Cu}^{2+}$  to **52** (10  $\mu\text{M}$ ), the absorbance at 380 nm gradually decreased and the band centered at 280 nm increased, because of the  $\text{Cu}^{2+}$  interaction with diaminomaleonitrile moiety which prevents

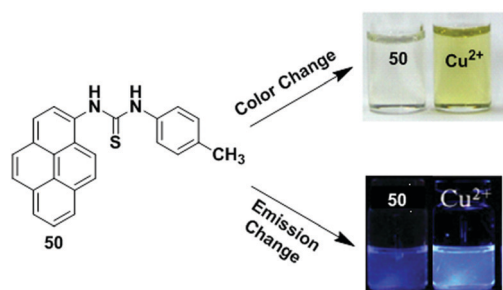


Fig. 44 The colour and fluorescence changes of chemosensor **50** upon addition of  $\text{Cu}^{2+}$  ions. Reproduced from ref. 119 with permission from the Elsevier, copyright 2010.

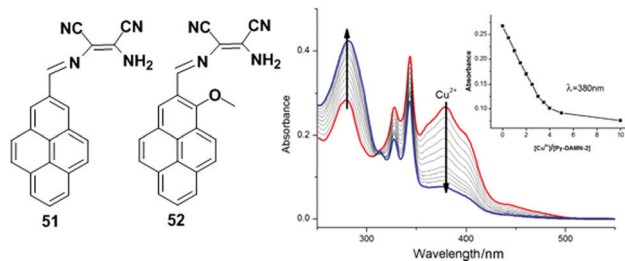


Fig. 45 The structure of probes **51** and **52**. Absorption spectra of **52** in presence of various amounts of  $\text{Cu}^{2+}$  in acetonitrile–water solution. Reproduced from ref. 120 with permission from the Elsevier, copyright 2015.

the charge-transfer of pyrene to the electron withdrawing diaminomaleonitrile moiety. This result is consistent with the spectral changes of receptor **51**.<sup>91</sup> The receptors **51** and **52** exhibited weak fluorescence (quantum yields in acetonitrile are 0.005 and 0.02, respectively) due to the presence of the Schiff base moiety and the overlap of the emission and absorption spectra. However, only ligand **52** gave a red-shifted band at around 590 nm in addition to the other band close to the characteristic emission band of pyrene. A significant fluorescence enhancement of **52** was observed at the emission band of pyrene on titration of  $\text{Cu}^{2+}$ . The quantum yield of the fluorescence emission band of **52** +  $\text{Cu}^{2+}$  complex was 0.42, approximately 20-times more than that of free **52**. The apparent association constant ( $K_a$ ) of  $\text{Cu}^{2+}$  binding to **52** was estimated to be  $5.2 \times 10^3$   $\text{M}^{-1}$ . However, the authors observed that on changing the solvent system from  $\text{CH}_3\text{CN}/\text{H}_2\text{O}$  to PBS/DMF, the receptor cannot bind with  $\text{Cu}^{2+}$  as DMF trapped the  $\text{Cu}^{2+}$  and hindered the interaction between the ligand **52** and  $\text{Cu}^{2+}$ .

In 2014, Goswami and co-workers designed and synthesized a new pyrene-based fluorescence probe **53** which was capable of working in both prokaryotic and eukaryotic living cells for the fluorogenic detection of  $\text{Cu}^{2+}$ . The complexation of receptor **53** with  $\text{Cu}^{2+}$  occurs through a chelation-enhanced fluorescence (CHEF) mechanism and several other reasons (Fig. 46).<sup>121</sup> In a  $\text{CH}_3\text{CN}$ -HEPES buffer (7:3, v/v, at pH 7.5) protic solvent system, the free receptor **53** shows a stronger fluorescence compared with aprotic  $\text{CH}_3\text{CN}$  solvent. This happens because of the hydrogen bonding interaction of the solvent with the N and O lone electron pairs of **53** which weakens the

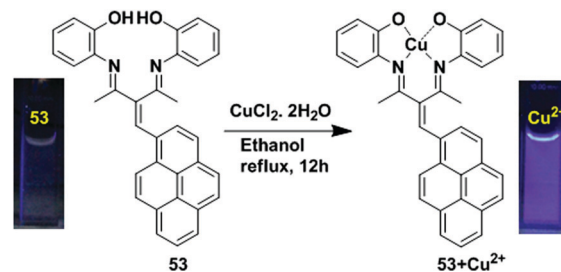


Fig. 46 The binding modes of **53** +  $\text{Cu}^{2+}$  and visual colour change of **53** with the addition of 2 equiv. of  $\text{CuCl}_2 \cdot 2\text{H}_2\text{O}$  under UV light. Reproduced from ref. 121 with permission from the Royal Society of Chemistry.

intramolecular radiationless transition leading to the red shifted emission maxima ( $\lambda_{em}$ ) on increasing protonation by the solvent. Moreover, the fluorescence behaviour of **53** upon addition of  $Cu^{2+}$  was enriched owing to the reduced energy gap between the ground state and the excited state of the metal bound species by possible metal–ligand charge transfer (ICT) and chelation. In addition, ESI LC-MS spectral analysis and the Job's plot confirmed the formation of a mononuclear complex of **53** with  $Cu^{2+}$ . The fluorescence titration experiments confirmed the minimum detection limit of copper was 1.21  $\mu M$  using 10  $\mu M$  of the ligand **53**. The selectivity of the fluorescence enhancement of **53** with  $Cu^{2+}$  (2.0 equivalents) was investigated in the presence of other metal ions (8.0 equivalents). With the exception of  $Cd^{2+}$  and  $Co^{2+}$ , no other competing metal ions inhibited the detection of  $Cu^{2+}$  by **53**. The NMR spectroscopic data in  $DMSO-d_6$  also demonstrated that the adduct formation between  $Cu^{2+}$  and **53** results in the disappearance of the phenolic protons and downfield shift of the protons of aromatic ring. The receptor **53** is cell membrane permeable and would enable detection of intracellular copper present in a biological system being incubated with copper perchlorate salt (1 mg  $mL^{-1}$ ) for 45 minutes.

## 6. Conclusion and future perspectives

This review article focuses on a particular type of fluorescent probe that contains a pyrene functional group and that have become interesting tools in modern biology and in environmental work. We have focused on the design of fluorescent sensors based on different mechanisms including monomer and excimer emission, PET, PCT, AIE, LMCT, CHEQ,  $Cu^{2+}$ -assisted reaction and others for the detection of  $Cu^{2+}$ . Interestingly, there have been a large number of fluorescent sensors which show fluorescence enhancement following monomer and excimer emission and inhibition of PET for sensing  $Cu^{2+}$ . In the monomer and excimer emission mechanism, the fluorescence response can experience three main spectroscopic signals as monomer, static or dynamic excimer emission processes depending on the binding nature of the ligand to  $Cu^{2+}$ . Based on the monomer and excimer emission mechanism, Yamato and co-workers as well as Kumar *et al.* have developed several ratiometric chemosensors which can easily diagnose  $Cu^{2+}$  ions. Moreover, a  $Cu^{2+}$ -promoted reaction based chemosensor in protic solvent systems was reported by Chang *et al.* and Yoon *et al.* and such systems may exhibit unique advantages and find special applications in environmental systems. Some probes based on the pyrene moiety show fluorescence quenching due to the paramagnetic nature of  $Cu^{2+}$  ions and LMCT processes or reverse PET mechanisms with prominent selective and sensitive responses for  $Cu^{2+}$  ions. It is noteworthy to mention that N- and/or O- and/or S-containing ligands hold more promise for the detection of  $Cu^{2+}$  ions. However, there are very few chemosensors described with reversible properties, which is a weakness in this type of host–guest chemistry. Therefore, it is necessary to consider the need for recycle of these resources to avoid waste generation. Besides, the design and

synthesis of fully water soluble efficient pyrene chemosensor remains a challenging task for the detection of  $Cu^{2+}$ . Most of the pyrene sensors in this article are found to be only partially soluble in water, on mixing with organic solvents like  $CH_3CN$ ,  $CH_3OH$ ,  $CH_2Cl_2$ . The introduction of polar functional groups or polar moieties to the chemosensor may increase the water solubility and make it more efficient for sensing  $Cu^{2+}$  in biological and environmental arenas. This review article could help to explore new concepts for designing and synthesizing chemosensors based on pyrene-derivatives and more compelling advances are anticipated in the near future.

## Conflicts of interest

There are no conflicts of interest to declare.

## Acknowledgements

CR thanks the EPSRC (EP/R023816/1) for an overseas travel grant.

## Notes and references

- (a) B. Valeur, *Molecular Fluorescence: Principles and Applications*, Wiley-VCH, Weinheim, Germany, 2002; (b) A. P. de Silva, H. Q. N. Gunaratne, T. Gunnlaugsson, A. J. M. Huxley, C. P. McCoy, J. T. Rademacher and T. E. Rice, Signaling recognition events with fluorescent sensors and switches, *Chem. Rev.*, 1997, **97**, 1515.
- A. P. S. Gonzales, M. A. Firmino, C. S. Nomura, F. R. P. Rocha, P. V. Oliveira and I. Gaubeur, Peat as a natural solid-phase for copper preconcentration and determination in a multicommuted flow system coupled to flame atomic absorption spectrometry, *Anal. Chim. Acta*, 2009, **636**, 198–204.
- J. S. Becker, A. Matusch, C. Depboylu, J. Dobrowolska and M. V. Zoriy, Quantitative imaging of selenium, copper, and zinc in thin sections of biological tissues (slugs–genus arion) measured by laser ablation inductively coupled plasma mass spectrometry, *Anal. Chem.*, 2007, **79**, 6074–6080.
- Y. Liu, P. Liang and L. Guo, Nanometer titanium dioxide immobilized on silica gel as sorbent for preconcentration of metal ions prior to their determination by inductively coupled plasma atomic emission spectrometry, *Talanta*, 2005, **68**, 25–30.
- A. A. Ensafi, T. Khayamian, A. Benvidi and E. Mirmomtaz, Simultaneous determination of copper, lead and cadmium by cathodic adsorptive stripping voltammetry using artificial neural network, *Anal. Chim. Acta*, 2006, **561**, 225–232.
- J. S. Becker, M. V. Zoriy, C. Pickhardt, N. Palomero-Gallagher and K. Zilles, Imaging of copper, zinc, and other elements in thin section of human brain samples (hippocampus) by laser ablation inductively coupled plasma mass spectrometry, *Anal. Chem.*, 2005, **77**, 3208–3216.
- N. Mekjinda, S. Phunnarungsi, V. Ruangpornvisuti, R. J. Ritchie, I. Hamachi, A. Ojida and J. Wongkongkatep,

- Masking phosphate with rare earth elements enables selective detection of arsenate by dipicolylamine-Zn<sup>II</sup> chemosensor, *Sci. Rep.*, 2020, **10**, 2656.
- 8 (a) I. Takashima, M. Kinoshita, R. Kawagoe, S. Nakagawa, M. Sugimoto, I. Hamachi and A. Ojida, Design of ratio-metric fluorescent probes based on arene-metal-ion interactions and their application to Cd<sup>II</sup> and hydrogen sulfide imaging in living cells, *Chem. – Eur. J.*, 2014, **20**, 2184–2192; (b) J. Guan, P. Zhang, T.-B. Wei, Q. Lin, H. Yao and Y.-M. Zhang, A highly selective PET-based chemosensor for instant detecting of Zn<sup>2+</sup>, *RSC Adv.*, 2014, **4**, 35797–35802; (c) S. Uchinomiya, N. Matsunaga, K. Kamoda, R. Kawagoe, A. Tsuruta, S. Ohdo and A. Ojida, Fluorescence detection of metabolic activity of the fatty acid beta oxidation pathway in living cells, *Chem. Commun.*, 2020, **56**, 3023–3026; (d) S. I. Reja, N. Sharma, M. Gupta, P. Bajaj, V. Bhalla, R. D. Parihar, P. Ohri, G. Kaur and M. Kumar, A highly selective fluorescent probe for detection of hydrogen sulfide in living systems: in vitro and in vivo applications, *Chem. – Eur. J.*, 2017, **23**, 9872–9878; (e) A. Kathiravan, A. Gowri, T. Khamrang, M. D. Kumar, N. Dhenadhayalan, K.-C. Lin, M. Velusamy and M. Jaccob, Pyrene-based chemosensor for picric acid—fundamentals to smartphone device design, *Anal. Chem.*, 2019, **91**, 13244–13250.
  - 9 (a) S. Lee, K. K. Y. Yuen, K. A. Jolliffe and J. Yoon, Fluorescent and colorimetric chemosensors for pyrophosphate, *Chem. Soc. Rev.*, 2015, **44**, 1749–1762; (b) H. N. Lee, Z. Xu, S. K. Kim, K. M. K. Swamy, Y. Kim, S.-J. Kim and J. Yoon, Pyrophosphate-selective fluorescent chemosensor at physiological pH: formation of a unique excimer upon addition of pyrophosphate, *J. Am. Chem. Soc.*, 2007, **129**, 3828–3829.
  - 10 (a) W. Shi and H. Ma, Spectroscopic probes with changeable  $\pi$ -conjugated systems, *Chem. Commun.*, 2012, **48**, 8732–8744; (b) *Bioinorganic Chemistry*, ed. A. X. Trautwein, Wiley-VCH, Weinheim, 1997; (c) *Metals and Their Compounds in the Environment*, ed. E. Merian, VCH, Weinheim, 1991; (d) Z. Yang, J. Cao, Y. He, J. H. Yang, T. Kim, X. Peng and J. S. Kim, Macro-/micro-environment-sensitive chemosensing and biological imaging, *Chem. Soc. Rev.*, 2014, **43**, 4563.
  - 11 (a) N. Kaur and S. Kumar, Colorimetric metal ion sensors, *Tetrahedron*, 2011, **67**, 9233–9264; (b) E. V. Anslyn, Supramolecular Analytical Chemistry, *J. Org. Chem.*, 2007, **72**, 687–699; (c) P. D. Beer, Transition-metal receptor systems for the selective recognition and sensing of anionic guest species, *Acc. Chem. Res.*, 1998, **31**, 71–80; (d) C. Bargossi, M. C. Fiorini, M. Montalti, L. Prodi and N. Zaccheroni, Recent developments in transition metal ion detection by luminescent chemosensors, *Coord. Chem. Rev.*, 2000, **208**, 17–32.
  - 12 A. B. Ellis and D. R. Walt, Guest editorial, *Chem. Rev.*, 2000, **100**, 2477–2478.
  - 13 L. Prodi, F. Bolletta, M. Montalti and N. Zaccheroni, Luminescent chemosensors for transition metal ions, *Coord. Chem. Rev.*, 2000, **205**, 59–83.
  - 14 K. Rurack, Flipping the light switch 'on'-the design of sensor molecules that show cation-induced fluorescence enhancement with heavy and transition metal ions, *Spectrochim. Acta, Part A*, 2001, **57**, 2161–2195.
  - 15 (a) A. T. Afaneh and G. Schreckenbach, Fluorescence enhancement/quenching based on metal orbital control: computational studies of a 6-thienyllumazine-Based mercury sensor, *J. Phys. Chem. A*, 2015, **119**, 8106–8116; (b) J.-P. Desvergne, F. Fages, H. Bouas-Laurent and P. Marsau, tunable photoresponsive supramolecular systems, *Pure Appl. Chem.*, 1992, **64**, 1231.
  - 16 Z. Xu, J. Yoon and D. R. Spring, Fluorescent chemosensors for Zn<sup>2+</sup>, *Chem. Soc. Rev.*, 2010, **39**, 1996–2006.
  - 17 D. G. Barceloux and D. D. Barceloux, Copper, *Clin. Toxicol.*, 1999, **37**, 217–230.
  - 18 (a) Z. L. Harris and J. D. Gitlin, Genetic and molecular basis for copper toxicity, *Am. J. Clin. Nutr.*, 1996, **63**, 836S–841S; (b) I. H. Scheinberg and I. Sternlieb, Wilson disease and idiopathic copper toxicosis, *Am. J. Clin. Nutr.*, 1996, **63**, 842S–845S.
  - 19 (a) Y. Xiang, A. Tong, P. Jin and Y. Ju, New fluorescent rhodamine hydrazone chemosensor for Cu(II) with high selectivity and sensitivity, *Org. Lett.*, 2006, **8**, 2863–2866; (b) X. Zhang, Y. Shiraishi and T. Hirai, Cu(II)-selective green fluorescence of a rhodamine–diacetic acid conjugate, *Org. Lett.*, 2007, **9**, 5039–5042.
  - 20 G. Li, Z. Xu, C. Chen and Z. Huang, A highly efficient and selective turn-on fluorescent sensor for Cu<sup>2+</sup> ion based on calix[4]arene bearing four iminoquinoline subunits on the upper rim, *Chem. Commun.*, 2008, 1774–1776.
  - 21 H. S. Jung, P. S. Kwon, J. W. Lee, J. I. Kim, C. S. Hong, J. W. Kim, S. Yan, J. Y. Lee, J. H. Lee, T. Joo and J. S. Kim, Coumarin-derived Cu<sup>2+</sup>-selective fluorescence sensor: synthesis, mechanisms, and applications in Living Cells, *J. Am. Chem. Soc.*, 2009, **131**, 2008–2012.
  - 22 S.-P. Wu and S. R. Liu, A new water-soluble fluorescent Cu(II) chemosensor based on tetrapeptide histidyl-glycyl-glycyl-glycine (HGGG), *Sens. Actuators, B*, 2009, **141**, 187–191.
  - 23 H. Irving and R. J. P. Williams, Order of Stability of Metal Complexes, *Nature*, 1948, **162**, 746–747.
  - 24 T. L. Banfield and D. Husain, Electronic energy transfer from triplet state acridine to paramagnetic ions, *Trans. Faraday Soc.*, 1969, **65**, 1985–1991.
  - 25 A. W. Varnes, R. B. Dodson and E. L. Wehry, Interactions of transition-metal ions with photoexcited states of flavines. Fluorescence quenching studies, *J. Am. Chem. Soc.*, 1972, **94**, 946–950.
  - 26 P. C. Bull, G. R. Thomas, J. M. Rommens, J. R. Forbes and D. W. Cox, The Wilson disease gene is a putative copper transporting P-type ATPase similar to the Menkes gene, *Nat. Genet.*, 1993, **5**, 327–337.
  - 27 (a) D. Udhayakumari, S. Naha and S. Velmathi, Colorimetric and fluorescent chemosensors for Cu<sup>2+</sup>. A comprehensive review from the years 2013–15, *Anal. Methods*, 2017, **9**, 552–578; (b) M. Ware, Health benefits and risks

- of copper, *Medical News Today*, October 23, 2017; (c) B. Sarkar, Treatment of Wilson and Menkes diseases, *Chem. Rev.*, 1999, **99**, 2535–2544; (d) J. A. Cowan, *Inorganic Biochemistry: An Introduction*, Wiley-VCH, New York, NY, USA, 2nd edn, 1997, pp. 133–134; (e) M. DiDonato and B. Sarkar, Copper transport and its alterations in Menkes and Wilson diseases, *Biochim. Biophys. Acta*, 1997, **1360**, 3–16.
- 28 (a) K. J. Barnham, C. L. Masters and A. I. Bush, Neurodegenerative diseases and oxidative stress, *Nat. Rev. Drug Discovery*, 2004, **3**, 205–214; (b) S. H. Hahn, M. S. Tanner, D. M. Danke and W. A. Gahl, Normal metallothionein synthesis in fibroblasts obtained from children with Indian childhood cirrhosis or copper-associated childhood cirrhosis, *Biochem. Mol. Med.*, 1995, **54**, 142–145; (c) D. R. Brown, Copper and prion disease, *Brain Res. Bull.*, 2001, **55**, 165–173; (d) D. Beyersmann, The significance of interactions in metal essentiality and toxicity, in *Metals and Their Compounds in the Environment*, ed. E. Merian, VCH, Weinheim, 1991, p. 491.
- 29 (a) M. Kaur, P. Kaur, V. Dhuna, S. Singh and K. Singh, A ferrocene-pyrene based ‘turn-on’ chemodosimeter for  $\text{Cr}^{3+}$ -application in bioimaging, *Dalton Trans.*, 2014, **43**, 5707–5712; (b) I. Berlmán, *Handbook of Fluorescence Spectra of Aromatic Molecules*, Academic Press, New York, 2nd edn, 1971.
- 30 M. Wang, J. Xu, X. Liu and H. Wang, A highly selective pyrene based “off-on” fluorescent chemosensor for cyanide, *New J. Chem.*, 2013, **37**, 3869–3872.
- 31 (a) D. Udhayakumari, Chromogenic and fluorogenic chemosensors for lethal cyanide ion. A comprehensive review of the year 2016, *Sens. Actuators, B*, 2018, **259**, 1022–1057; (b) M.-H. Yang, P. Thirupathi and K.-H. Lee, Selective and sensitive ratiometric detection of  $\text{Hg}(\text{II})$  Ions using a simple amino acid based sensor, *Org. Lett.*, 2011, **13**, 5028–5031.
- 32 (a) X.-L. Ni, S. Wang, X. Zeng, Z. Tao and T. Yamato, Pyrene-linked triazole-modified homooxalix[3]arene: A unique  $C_3$  symmetry ratiometric fluorescent chemosensor for  $\text{Pb}^{2+}$ , *Org. Lett.*, 2011, **13**, 552–555; (b) Q. Dai, W. Liu, X. Zhuang, J. Wu, H. Zhang and P. Wang, Ratiometric fluorescence sensor based on a pyrene derivative and quantification detection of heparin in aqueous solution and serum, *Anal. Chem.*, 2011, **83**, 6559–6564; (c) G. Sivaraman, T. Anand and D. Chellappa, Development of a pyrene based “turn on” fluorescent chemosensor for  $\text{Hg}^{2+}$ , *RSC Adv.*, 2012, **2**, 10605–10609.
- 33 F. Wang, R. Nandhakumar, J. H. Moon, K. M. Kim, J. Y. Lee and J. Yoon, Ratiometric fluorescent chemosensor for silver ion at physiological pH, *Inorg. Chem.*, 2011, **50**, 2240–2245.
- 34 L. Wang, M. Yu, Z. Liu, W. Zhao, Z. Li, Z. Ni, C. Li and L. Wei, A visible light excitable “on-off” and “green-red” fluorescent chemodosimeter for  $\text{Ni}^{2+}/\text{Pb}^{2+}$ , *New J. Chem.*, 2012, **36**, 2176–2179.
- 35 E. Manandhar, J. H. Broome, J. Myrick, W. Lagrone, P. J. Cragg and K. J. Wallace, A pyrene-based fluorescent sensor for  $\text{Zn}^{2+}$  ions: a molecular ‘butterfly’, *Chem. Commun.*, 2011, **47**, 8796–8798.
- 36 H. Irving and R. J. P. Williams, The stability of transition-metal complexes, *J. Chem. Soc.*, 1953, 3192–3210.
- 37 (a) A. W. Varnes, R. B. Dodson and E. L. Whery, Interactions of transition-metal ions with photoexcited states of flavines. Fluorescence quenching studies, *J. Am. Chem. Soc.*, 1972, **94**, 946–950; (b) G. Sivaraman, M. Iniya, T. Anand, N. G. Kotla, O. Sunnapu, S. Singaravadeivel, A. Gulyani and D. Chellappa, Chemically diverse small molecule fluorescent chemosensors for copper ion, *Coord. Chem. Rev.*, 2018, **357**, 50–104; (c) K. Rurack, U. Resch, M. Sensoer and S. Daehne, A new fluorescence probe for trace metal ions: Cation-dependent spectroscopic properties, *J. Fluoresc.*, 1993, **3**, 141–143; (d) K. Li, N. Li, X. Chen and A. Tong, A ratiometric fluorescent chemodosimeter for  $\text{Cu}(\text{II})$  in water with high selectivity and sensitivity, *Anal. Chim. Acta*, 2012, **712**, 115–119; (e) X. Lou, D. Ou, Q. Li and Z. Li, An indirect approach for anion detection: the displacement strategy and its application, *Chem. Commun.*, 2012, **48**, 8462–8477.
- 38 (a) I. Aoki, H. Kawabata, K. Nakashima and S. Shinkai, Fluorescent calix[4]arene which responds to solvent polarity and metal ions, *J. Chem. Soc., Chem. Commun.*, 1991, 1771–1773; (b) I. Aoki, T. Sakaki and S. Shinkai, A new metal sensory system based on intramolecular fluorescence quenching on the ionophoric calix[4]arene ring, *J. Chem. Soc., Chem. Commun.*, 1992, 730–732.
- 39 (a) H.-F. Ji, R. Dabestani, G. M. Brown and R. A. Sachleben, A new highly selective calix[4]crown-6 fluorescent caesium probe, *Chem. Commun.*, 2000, 833–834; (b) H.-F. Ji, R. Dabestani, G. M. Brown and R. L. Hettich, Synthesis and sensing behavior of cyanoanthracene modified 1,3-alternate calix[4]benzocrown-6: a new class of  $\text{Cs}^+$  selective optical sensors, *J. Chem. Soc., Perkin Trans. 2*, 2001, 585–591.
- 40 (a) C. D. Gutsche, in *Calixarenes in Monographs in Supramolecular Chemistry*, ed. J. F. Stoddart, Royal Society of Chemistry, Cambridge, UK, 1989, vol. 1; (b) J. D. V. Loon, W. Verboom and D. N. Reinhoudt, Selective functionalization and conformational properties of calix[4]arenes: A review, *Org. Prep. Proced. Int.*, 1992, **24**, 437–462.
- 41 (a) R. Ungaro and A. Pochini, in *Frontiers in Supramolecular Organic Chemistry and Photochemistry*, ed. H.-J. Schneider, VCH, Weinheim, Germany, 1991, pp. 57–81; (b) T. Jin, K. Ichikawa and T. Koyama, A fluorescent calix[4]arene as an intramolecular excimer-forming  $\text{Na}^+$  sensor in non-aqueous solution, *J. Chem. Soc., Chem. Commun.*, 1992, 499–501.
- 42 S. Nishizawa, H. Kaneda, T. Uchida and N. Teramae, Anion sensing by a donor-spacer-acceptor system: an intramolecular exciplex emission enhanced by hydrogen bond-mediated complexation, *J. Chem. Soc., Perkin Trans. 2*, 1998, 2325–2328.
- 43 (a) S. K. Kim, S. H. Lee, J. Y. Lee, J. Y. Lee, R. A. Bartsch and J. S. Kim, An excimer-based, binuclear, on-off switchable

- calix[4]crown chemosensor, *J. Am. Chem. Soc.*, 2004, **126**, 16499–16506; (b) F. M. Winnik, Photophysics of preassociated pyrenes in aqueous polymer solutions and in other organized media, *Chem. Rev.*, 1993, **93**, 587–614; (c) S. H. Lee, S. H. Kim, S. K. Kim, J. H. Jung and J. S. Kim, Fluorescence ratiometry of monomer/excimer emissions in a space-through PET system, *J. Org. Chem.*, 2005, **70**, 9288–9295; (d) J. Y. Lee, S. K. Kim, J. H. Jung and J. S. Kim, Bifunctional fluorescent calix[4]arene chemosensor for both a cation and an anion, *J. Org. Chem.*, 2005, **70**, 1463–1466.
- 44 J. B. Briks, Excimers, *Rep. Prog. Phys.*, 1975, **38**, 903–974.
- 45 (a) J. K. Choi, S. H. Kim, J. Yoon, K.-H. Lee, R. A. Bartsch and J. S. Kim, A PCT-based, pyrene-armed calix[4]crown fluoroionophore, *J. Org. Chem.*, 2006, **71**, 8011–8015; (b) J.-S. Yang, C.-S. Lin and C.-Y. Hwang, Cu<sup>2+</sup>-induced blue shift of the pyrene excimer emission: A new signal transduction mode of pyrene probes, *Org. Lett.*, 2001, **3**, 889–892; (c) H. J. Kim, S. K. Kim, J. Y. Lee and J. S. Kim, Fluoride-sensing calix-luminophores based on regioselective binding, *J. Org. Chem.*, 2006, **71**, 6611–6614.
- 46 (a) J. B. Birks, *Organic Molecular Photophysics*, J. Wiley, London, New York, 1973–1975; (b) J. B. Birks, *Photophysics of Aromatic Molecules*, Wiley-Interscience, London, 1970.
- 47 S. Sarkar, S. Roy, A. Sikdar, R. N. Saha and S. S. Panja, A pyrene-based simple but highly selective fluorescence sensor for Cu<sup>2+</sup> ions via a static excimer mechanism, *Analyst*, 2013, **138**, 7119–7126.
- 48 (a) B. Valeur and I. Leray, Ion-responsive supramolecular fluorescent systems based on multichromophoric calixarenes: A review, *Inorg. Chim. Acta*, 2007, **360**, 765–774; (b) S. H. Kim, H. J. Kim, J. Yoon and J. S. Kim, Fluorescent chemosensors, in *Calixarenes in the nanoworld*, ed. J. Vicens and J. Harrowfield, Springer, Dordrecht, The Netherlands, 2007, pp. 311–334.
- 49 (a) F. M. Winnik, Photophysics of preassociated pyrenes in aqueous polymer solutions and in other organized media, *Chem. Rev.*, 1993, **93**, 587–614; (b) A. Okamoto, T. Ichiba and I. Saito, Pyrene-labeled oligodeoxynucleotide probe for detecting base insertion by excimer fluorescence emission, *J. Am. Chem. Soc.*, 2004, **126**, 8364–8365.
- 50 H. Nohta, H. Satozono, K. Koiso, H. Yoshida, J. Ishida and M. Yamaguchi, Highly selective fluorometric determination of polyamines based on intramolecular excimer-forming derivatization with a pyrene-labeling reagent, *Anal. Chem.*, 2000, **72**, 4199–4204.
- 51 A. P. de Silva, T. S. Moody and G. D. Wright, Fluorescent PET (Photoinduced Electron Transfer) sensors as potent analytical tools, *Analyst*, 2009, **134**, 2385–2395.
- 52 M. Sauer, Single-Molecule-Sensitive Fluorescent sensors Based on photoinduced intramolecular charge transfer, *Angew. Chem., Int. Ed.*, 2003, **42**, 1790–1793.
- 53 Y. Bao, B. Liu, F. Du, J. Tian, H. Wang and R. Bai, A new strategy for highly selective fluorescent sensing of F<sup>−</sup> and Zn<sup>2+</sup> with dual output modes, *J. Mater. Chem.*, 2012, **22**, 5291–5294.
- 54 H.-F. Ji, G. M. Brown and R. Dabestani, Optical sensing of cesium using 1,3-Alternate calix[4]-mono- and di(anthrylmethyl)aza-crown-6, *Chem. Commun.*, 1999, 609–610.
- 55 I. Leray, F. O'Reilly, J.-L. Habib Jiwan, J.-P. Soumilion and B. Valeur, A new calix[4]arene-based fluorescent sensor for sodium ion, *Chem. Commun.*, 1999, 795–796.
- 56 J. S. Kim, K. H. Noh, S. H. Lee, S. K. Kim, S. K. Kim and J. Yoon, Molecular taekwondo. 2. A new calix[4]azacrown bearing two different binding sites as a new fluorescent ionophore, *J. Org. Chem.*, 2003, **68**, 597–600.
- 57 J. S. Kim, O. J. Shon, J. A. Rim, S. K. Kim and J. Yoon, Pyrene-armed calix[4]azacrowns as new fluorescent ionophores: “molecular taekowndo” process via fluorescence change, *J. Org. Chem.*, 2002, **67**, 2348–2351.
- 58 Z. R. Grabowski and J. Dobkowski, Twisted intramolecular charge transfer (TICT) excited states: energy and molecular structure, *Pure Appl. Chem.*, 1983, **55**, 245–252.
- 59 M. Liu, X. Yu, M. Li, N. Liao, A. Bi, Y. Jiang, S. Liu, Z. Gong and W. Zeng, Fluorescent probes for the detection of magnesium ions (Mg<sup>2+</sup>): from design to application, *RSC Adv.*, 2018, **8**, 12573–12587.
- 60 W. Rettig and R. Lapouyade, Probe design and chemical sensing, in *Topics in Fluorescence Spectroscopy*, ed. J. R. Lakowicz, Plenum Press, New York, 1994, vol. 4, pp. 109–149.
- 61 H.-G. Löhr and F. Vögtle, Chromo- and fluoroionophores. A new class of dye reagents, *Acc. Chem. Res.*, 1985, **18**, 65–72.
- 62 (a) P. Neri, J. L. Sessler and M.-X. Wang, *Calixarenes and Beyond*, Springer, 2016; (b) R. Kumar, A. Sharma, H. Singh, P. Suating, H. S. Kim, K. Sunwoo, I. Shim, B. C. Gibb and J. S. Kim, Revisiting fluorescent calixarenes: from molecular sensors to smart materials, *Chem. Rev.*, 2019, **16**, 9657–9721; (c) J. S. Kim and D. T. Quang, Calixarene-derived fluorescent probes, *Chem. Rev.*, 2007, **107**, 3780–3799; (d) K. Sharma and P. Cragg, Calixarene based chemical sensors, *Chem. Senses*, 2011, **1**, 1–18; (e) R. Kumar, Y. O. Lee, V. Bhalla, M. Kumar and J. S. Kim, Recent developments of thiacalixarene based molecular motifs, *Chem. Soc. Rev.*, 2014, **43**, 4824–4870; (f) J.-Y. Fu, L. Mu, X. Zeng, J.-L. Zhao, C. Redshaw, X.-L. Ni and T. Yamato, An NBD-armed thiacalix[4]arene-derived colorimetric and fluorometric chemosensor for Ag<sup>+</sup>: a metal–ligand receptor of anions, *Dalton Trans.*, 2013, **42**, 3552–3560.
- 63 (a) J.-L. Zhao, H. Tomiyasu, X.-L. Ni, X. Zeng, M. R. J. Elsegood, C. Redshaw, S. Rahman, P. E. Georghiou and T. Yamato, Synthesis and evaluation of a novel ionophore based on a thiacalix[4]arene derivative bearing imidazole units, *New J. Chem.*, 2014, **38**, 6041–6049; (b) J.-L. Zhao, C. Wu, X. Zeng, S. Rahman, P. E. Georghiou, M. R. J. Elsegood, T. G. Warwick, C. Redshaw, S. J. Teat and T. Yamato, Thiacalix[4]arene derivatives bearing imidazole units: A ditopic hard/soft receptor for Na<sup>+</sup> and K<sup>+</sup>/Ag<sup>+</sup> with an allosteric effect and

- a reusable extractant for dichromate anions, *Chemistry-Select*, 2016, **1**, 1541–1547; (c) J.-L. Zhao, C. Wu, H. Tomiyasu, X. Zeng, M. R. J. Elsegood, C. Redshaw and T. Yamato, A rare and exclusive endoperoxide photoproduct derived from a thiacalix[4]arene crown-shaped derivative bearing a 9, 10-substituted anthracene moiety, *Chem. – Asian J.*, 2016, **11**, 1606–1612; (d) S. Rahman, H. Tomiyasu, H. Kawazoe, J.-L. Zhao, H. Cong, X.-L. Ni, X. Zeng, M. R. J. Elsegood, T. G. Warwick, S. J. Teat, C. Redshaw, P. E. Georghiou and T. Yamato, A study of anion binding behaviour of 1,3-alternate thiacalix[4]arene-based receptors bearing urea moieties, *New J. Chem.*, 2016, **40**, 9245–9251; (e) M. Kumar, A. Dhir and V. Bhalla, On–Off switchable binuclear chemosensor based on thiacalix[4]-crown armed with pyrene moieties, *Eur. J. Org. Chem.*, 2009, 4534–4540.
- 64 M. Kumar, R. Kumar and V. Bhalla, Differential fluorogenic sensing of  $F^-$  versus  $CN^-$  based on thiacalix[4]arene derivatives, *Tetrahedron Lett.*, 2013, **54**, 1524–1527.
- 65 H. Tomiyasu, X.-L. Ni, X. Zeng, C. Redshaw and T. Yamato, A study of allosteric binding behaviour of a 1,3-alternate thiacalix[4]arene-based receptor using fluorescence signal, *Org. Biomol. Chem.*, 2014, **12**, 4917–4923.
- 66 J.-L. Zhao, H. Tomiyasu, C. Wu, H. Cong, X. Zeng, S. Rahman, P. E. Georghiou, D. L. Hughes, C. Redshaw and T. Yamato, Synthesis, crystal structure and complexation behaviour study of an efficient  $Cu^{2+}$  ratiometric fluorescent chemosensor based on thiacalix[4]arene, *Tetrahedron*, 2015, **71**, 8521–8527.
- 67 (a) X.-L. Ni, X. Zeng and T. Yamato, Ratiometric fluorescent receptors for both  $Zn^{2+}$  and  $H_2PO_4^-$  ions based on a pyrenyl-linked triazole-modified homooxacalix[3]arene: A potential molecular traffic signal with an R-S latch logic circuit, *J. Org. Chem.*, 2011, **76**, 5696–5702; (b) X.-L. Ni, S. Rahman, S. Wang, C.-C. Jin, X. Zeng, D. Hughes, C. Redshaw and T. Yamato, hexahomotrioxacalix[3]arene derivatives as ionophores for molecular recognition of dopamine, serotonin and phenylethylamine, *Org. Biomol. Chem.*, 2012, **10**, 4618–4626.
- 68 X.-L. Ni, S. Wang, X. Zeng and T. Yamato, Pyrene-linked triazole-modified homooxacalix[3]arene: A unique  $C_3$  symmetry ratiometric fluorescent chemosensor for  $Pb^{2+}$ , *Org. Lett.*, 2011, **13**, 552–555.
- 69 X.-L. Ni, Y. Wu, C. Redshaw and T. Yamato, Direct evidence of a blocking heavy atom effect on the water-assisted fluorescence enhancement detection of  $Hg^{2+}$  based on ratiometric chemosensor, *Dalton Trans.*, 2014, **43**, 12633–12638.
- 70 (a) X.-L. Ni, C.-C. Jin, X.-K. Jiang, M. Takimoto, S. Rahman, X. Zeng, D. L. Hughes, C. Redshaw and T. Yamato, Tri-substituted hexahomotrioxacalix[3]arene derivatives bearing imidazole units: synthesis and extraction properties for cations and chromate anions, *Org. Biomol. Chem.*, 2013, **11**, 5435–5442; (b) C.-C. Jin, T. Kinoshita, H. Cong, X.-L. Ni, X. Zeng, D. L. Hughes, C. Redshaw and T. Yamato, Synthesis and inclusion properties of  $C_3$ -symmetric triazole derivatives based on hexahomotrioxacalix[3]arene, *New J. Chem.*, 2012, **36**, 2580–2586; (c) X.-L. Ni, X. Zeng, D. L. Hughes, C. Redshaw and T. Yamato, Novel ion-pair receptors based on hexahomotrioxacalix[3]arene derivatives, *Org. Biomol. Chem.*, 2011, **9**, 6535–6541.
- 71 C.-C. Jin, H. Cong, X.-L. Ni, X. Zeng, C. Redshaw and T. Yamato, Synthesis and inclusion behavior of a hetero-tritopic receptor based on hexahomotrioxacalix[3]arene, *RSC Adv.*, 2014, **4**, 31469–31475.
- 72 C.-C. Jin, M. Fukuda, C. Wu, X. Jiang, X.-L. Ni, X. Zeng, C. Redshaw and T. Yamato, A pyrene-armed hexahomotrioxacalix[3]arene as a multi-sensor via synergistic and demetallation effects, *Tetrahedron*, 2015, **71**, 9593–9597.
- 73 Y.-S. Wu, C.-Y. Li, Y.-F. Li, D. Li and Z. Li, Development of a simple pyrene-based ratiometric fluorescent chemosensor for copper ion in living cells, *Sens. Actuators, B*, 2016, **222**, 1226–1232.
- 74 J. Fernández-Lodeiro, C. Núñez, J. S. S. de Melo, J. S. Capelo and L. Carlos, Steady-state and time-resolved investigations on pyrene-based chemosensors, *Inorg. Chem.*, 2013, **52**, 121–129.
- 75 J.-S. Yang, C.-S. Lin and C.-Y. Hwang,  $Cu^{2+}$ -induced blue shift of the pyrene excimer emission: A new signal transduction mode of pyrene probes, *Org. Lett.*, 2001, **3**, 889–892.
- 76 H. J. Kim, J. Hong, A. Hong, S. Ham, J. H. Lee and J. S. Kim,  $Cu^{2+}$ -induced intermolecular static excimer formation of pyrenealkylamine, *Org. Lett.*, 2008, **10**, 1963–1966.
- 77 H. S. Jung, M. Park, D. Y. Han, E. Kim, C. Lee, S. Ham and J. S. Kim,  $Cu^{2+}$  ion-induced self-assembly of pyrenylquinoline with a pyrenyle Excimer formation, *Org. Lett.*, 2009, **11**, 3378–3381.
- 78 S. Sarkar, S. Roy, A. Sikdar, R. N. Saha and S. S. Panja, A pyrene-based simple but highly selective fluorescence sensor for  $Cu^{2+}$  ions via a static excimer mechanism, *Analyst*, 2013, **138**, 7119–7126.
- 79 M. Shellaiah, Y.-H. Wu, A. Singh, M. V. R. Raju and H.-C. Lin, Novel pyrene-and anthracene-based Schiff base derivatives as  $Cu^{2+}$  and  $Fe^{3+}$  fluorescence turn-on sensors and for aggregation induced emissions, *J. Mater. Chem. A*, 2013, **1**, 1310–1318.
- 80 E. J. Jun, H. N. Won, J. S. Kim, K.-H. Lee and J. Yoon, Unique blue shift due to the formation of static pyrene excimer: highly selective fluorescent chemosensor for  $Cu^{2+}$ , *Tetrahedron Lett.*, 2006, **47**, 4577–4580.
- 81 M. Kumar, N. Kumar and V. Bhalla, Ratiometric nanomolar detection of  $Cu^{2+}$  ions in mixed aqueous media: a  $Cu^{2+}/Li^+$  ions switchable allosteric system based on thiacalix[4]crown, *Dalton Trans.*, 2012, **41**, 10189–10193.
- 82 S. Ghosh, A. Ganguly, M. R. Uddin, S. Mandal, M. A. Alam and N. Guchhait, Dual mode selective chemosensor for copper and fluoride ions: a fluorometric, colorimetric and theoretical investigation, *Dalton Trans.*, 2016, **45**, 11042–11051.
- 83 B. C. Roy, B. Chandra, D. Hromas and S. Mallik, Synthesis of new, pyrene-containing, metal-chelating lipids and sensing of cupric ions, *Org. Lett.*, 2003, **5**, 11–14.

- 84 Y. Wu, X.-L. Ni, L. Mou, C.-C. Jin, C. Redshaw and T. Yamato, Synthesis of a ditopic homooxacalix[3]arene for fluorescence enhanced detection of heavy and transition metal ions, *Supramol. Chem.*, 2015, **27**, 501–507.
- 85 (a) S. Sumiya, Y. Shiraishi and T. Hirai, Mechanism for different fluorescence response of a coumarin–amide–dipicolylamine linkage to Zn(II) and Cd(II) in water, *J. Phys. Chem. A*, 2013, **117**, 1474–1482; (b) D. T. Shi, B. Zhang, Y. X. Yang, C. C. Guan, X. P. He, Y. C. Li, G. R. Chen and K. X. Chen, Bis-triazolyl indoleamines as unique “off–approach–on” chemosensors for copper and fluorine, *Analyst*, 2013, **138**, 2808–2811; (c) C. Gao, H. Zhu, M. Zhang, T. Tan, J. Chen and H. Qiu, A new highly Zn<sup>2+</sup>-selective and “off–on” fluorescent chemosensor based on the pyrene group, *Anal. Methods*, 2015, **7**, 8172–8176; (d) X. Sun, Q. Xu, G. Kim, S. E. Flower, J. P. Lowe, J. Yoon, J. S. Fossey, X. Qian, S. D. Bull and T. D. James, A water-soluble boronate-based fluorescent probe for the selective detection of peroxynitrite and imaging in living cells, *Chem. Sci.*, 2014, **5**, 3368–3373.
- 86 Z. Kowser, H. Tomiyasu, X. Jiang, U. Rayhan, C. Redshaw and T. Yamato, Solvent effect and fluorescence response of the 7-tert-butylpyrene–dipicolylamine linkage for the selective and sensitive response toward Zn(II) and Cd(II) ions, *New J. Chem.*, 2015, **39**, 4055–4062.
- 87 Z. Kowser, C.-C. Jin, X. Jiang, S. Rahman, P. E. Georghiou, X.-L. Ni, X. Zeng, C. Redshaw and T. Yamato, Fluorescent turn-on sensors based on pyrene-containing Schiff base derivatives for Cu<sup>2+</sup> recognition: spectroscopic and DFT computational studies, *Tetrahedron*, 2016, **72**, 4575–4581.
- 88 H.-F. Wang and S.-P. Wu, A pyrene-based highly selective turn-on fluorescent sensor for copper(II) ions and its application in living cell imaging, *Sens. Actuators, B*, 2013, **181**, 743–748.
- 89 R. Martínez, F. Zapata, A. Caballero, A. Espinosa, A. Tárraga and P. Molina, 2-Aza-1,3-butadiene Derivatives Featuring an Anthracene or Pyrene Unit: Highly Selective Colorimetric and Fluorescent Signaling of Cu<sup>2+</sup> Cation, *Org. Lett.*, 2006, **8**, 3235–3238.
- 90 S.-P. Wu, Z.-M. Huang, S.-R. Liu and P. K. Chung, A pyrene-based highly selective turn-on fluorescent sensor for copper(II) ion and its application in live cell imaging, *J. Fluoresc.*, 2012, **22**, 253–259.
- 91 S.-P. Wu, T.-H. Wang and S.-R. Liu, A highly selective turn-on fluorescent chemosensor for copper(II) ion, *Tetrahedron*, 2010, **66**, 9655–9658.
- 92 P. Venkatesan and S.-P. Wu, A turn-on fluorescent pyrene-based chemosensor for Cu(II) with live cell application, *RSC Adv.*, 2015, **5**, 42591–42596.
- 93 A. Saravanan, G. Subashini, S. Shyamsivappan, T. Suresh, K. Kadirvelu, N. Bhuvanesh, R. Nandhakumar and P. S. Mohan, A selective fluorescence chemosensor: pyrene motif Schiff base derivative for detection of Cu<sup>2+</sup> ions in living cells, *J. Photochem. Photobiol., A*, 2018, **364**, 424–432.
- 94 A. Ghorai, J. Mondal, A. K. Manna, S. Chowdhury and G. K. Patra, A novel pyrene based highly selective reversible fluorescent-colorimetric sensor for the rapid detection of Cu<sup>2+</sup> ions: application in bio-imaging, *Anal. Methods*, 2018, **10**, 1063–1073.
- 95 R. Martínez, A. Espinosa, A. Tárraga and P. Molina, A new bis (pyrenyl) azadiene-based probe for the colorimetric and fluorescent sensing of Cu(II) and Hg(II), *Tetrahedron*, 2010, **66**, 3662–3667.
- 96 H. Han, M. Wang and H. Wang, 1-Nitronyl nitroxide pyrene as a new off–on fluorescent chemosensor for Cu<sup>2+</sup>, *New J. Chem.*, 2014, **38**, 914–917.
- 97 J. K. Choi, S. H. Kim, J. Yoon, K.-H. Lee, R. A. Bartsch and J. S. Kim, A PCT-based, pyrene-armed calix[4]crown fluorophore, *J. Org. Chem.*, 2006, **71**, 8011–8015.
- 98 L. Mohapatra and K. Parida, A review of solar and visible light active oxo-bridged materials for energy and environment, *Catal. Sci. Technol.*, 2017, **7**, 2153–2164.
- 99 Y. Guo, L. Wang, J. Zhuo, B. Xu, X. Li, J. Zhang, Z. Zhang, H. Chi, Y. Dong and G. Lu, A pyrene-based dual chemosensor for colorimetric detection of Cu<sup>2+</sup> and fluorescent detection of Fe<sup>3+</sup>, *Tetrahedron Lett.*, 2017, **58**, 3951–3956.
- 100 S. Dalbera, S. Kulovi and S. Dalai, Pyrene-based Schiff base as selective chemosensor for copper(II) and sulfide Ions, *ChemistrySelect*, 2018, **3**, 6561–6569.
- 101 A. Vogler and H. Kunkely, Ligand-to-ligand and intraligand charge transfer and their relation to charge transfer interactions in organic zwitterions, *Coord. Chem. Rev.*, 2007, **251**, 577–583.
- 102 S. Mukherjee and S. Betal, Sensing phenomena, extraction and recovery of Cu<sup>2+</sup> followed by smart phone application using a luminescent pyrene based chemosensor, *J. Lumin.*, 2018, **204**, 145–153.
- 103 (a) Y. Hong, J. W. Y. Lam and B. Z. Tang, Aggregation-induced emission, *Chem. Soc. Rev.*, 2011, **40**, 5361–5388; (b) T. Han, Y. Hong, N. Xie, S. Chen, N. Zhao, E. Zhao, J. W. Y. Lam, H. H. Y. Sung, Y. Dong, B. Tong and B. Z. Tang, Defect-sensitive crystals based on diaminomaleonitrile-functionalized Schiff base with aggregation-enhanced emission, *J. Mater. Chem. C*, 2013, **1**, 7314–7320; (c) T. Han, X. Gu, J. W. Y. Lam, A. C. S. Leung, R. T. K. Kwok, T. Han, B. Tong, J. Shi, Y. Dong and B. Z. Tang, Diaminomaleonitrile-based Schiff bases: aggregation-enhanced emission, red fluorescence, mechanochromism and bioimaging applications, *J. Mater. Chem. C*, 2016, **4**, 10430–10434.
- 104 W.-N. Wu, P.-D. Mao, Y. Wang, X.-J. Mao, Z.-Q. Xu, Z.-H. Xu, X.-L. Zhao, Y.-C. Fan and X.-F. Hou, AEE active Schiff base-bearing pyrene unit and further Cu<sup>2+</sup>-induced self-assembly process, *Sens. Actuators, B*, 2018, **258**, 393–401.
- 105 N. Chakraborty, A. Chakraborty and S. Das, A pyrene based fluorescent turn on chemosensor for detection of Cu<sup>2+</sup> ions with antioxidant nature, *J. Lumin.*, 2018, **199**, 302–309.
- 106 (a) N. Li, Y. Xiang and A. Tong, Highly sensitive and selective “turn-on” fluorescent chemodosimeter for Cu<sup>2+</sup> in water via Cu<sup>2+</sup>-promoted hydrolysis of lactone moiety in coumarin, *Chem. Commun.*, 2010, **46**, 3363–3365; (b) S. Liu,

- Y.-M. Wang and J. Han, Fluorescent chemosensors for copper(II) ion: Structure, mechanism and application, *J. Photochem. Photobiol., C*, 2017, **32**, 78–103.
- 107 H. Ryu, J. H. Baek, M. G. Choi, J. C. Lee and S.-K. Chang, Cu<sup>2+</sup>-selective turn-on fluorescence signaling based on metal-induced hydrolysis of pyrenecarbohydrazide, *Tetrahedron Lett.*, 2017, **58**, 2927–2930.
- 108 (a) V. Dujols, F. Ford and A. W. Czarnik, A long-wavelength, fluorescent chemodosimeter selective for Cu(II) ion in water, *J. Am. Chem. Soc.*, 1997, **119**, 7386–7387; (b) A. Mokhir and R. Kramer, Double discrimination by binding and reactivity in fluorescent metal ion detection, *Chem. Commun.*, 2005, 2244–2246; (c) J. Kovacs, T. Rödler and A. Mokhir, Chemodosimeter for Cu<sup>II</sup> detection based on cyclic peptide nucleic acids, *Angew. Chem., Int. Ed.*, 2006, **45**, 7815–7817; (d) J. Kovacs and A. Mokhir, Catalytic hydrolysis of esters of 2-hydroxypyridine derivatives for Cu<sup>2+</sup> detection, *Inorg. Chem.*, 2008, **47**, 1880–1882; (e) R. M. Kierat and R. Kraemer, A fluorogenic and chromogenic probe that detects the esterase activity of trace copper(II), *Bioorg. Med. Chem. Lett.*, 2005, **15**, 4824–4827; (f) L. Zeng, E. W. Miller, A. Pralle, E. Y. Isacoff and C. J. Chang, A selective turn-on fluorescent sensor for imaging copper in living cells, *J. Am. Chem. Soc.*, 2006, **128**, 10–11; (g) X. Qi, E. J. Jun, L. Xu, S.-J. Kim, J. S. J. Hong, Y. J. Yoon and J. Yoon, New BODIPY derivatives as OFF-ON fluorescent chemosensor and fluorescent chemodosimeter for Cu<sup>2+</sup>: cooperative selectivity enhancement toward Cu<sup>2+</sup>, *J. Org. Chem.*, 2006, **71**, 2881–2884; (h) M. H. Kim, H. H. Jang, S. Yi, S.-K. Chang and M. S. Han, Coumarin-derivative-based off-on catalytic chemodosimeter for Cu<sup>2+</sup> ions, *Chem. Commun.*, 2009, 4838–4840; (i) E. J. Corey and S. Knapp, Facile conversion of *N,N*-dimethylhydrazones to carbonyl compounds by cupric ioncatalyzed hydrolysis, *Tetrahedron Lett.*, 1976, **17**, 3667–3668; (j) Q. Wu and E. V. Anslyn, Catalytic signal amplification using a Heck reaction. An example in the fluorescence sensing of Cu(II), *J. Am. Chem. Soc.*, 2004, **126**, 14682–14683; (k) Y. Xiang and A. Tong, Ratiometric and selective fluorescent chemodosimeter for Cu(II) by Cu(II)-induced oxidation, *Luminescence*, 2008, **23**, 28–31; (l) L. Mei, Y. Xiang, N. Li and A. Tong, A new fluorescent probe of rhodamine B derivative for the detection of copper ion, *Talanta*, 2007, **72**, 1717–1722.
- 109 J. F. Zhang, Y. Zhou, J. Yoon, Y. Kim, S. J. Kim and J. S. Kim, Naphthalimide modified rhodamine derivative: ratiometric and selective fluorescent sensor for Cu<sup>2+</sup> based on two different approaches, *Org. Lett.*, 2010, **12**, 3852–3855.
- 110 Y. Zhou, F. Wang, Y. Kim, S.-J. Kim and J. Yoon, Cu<sup>2+</sup>-selective ratiometric and “Off-On” sensor based on the rhodamine derivative bearing pyrene group, *Org. Lett.*, 2009, **11**, 4442–4445.
- 111 (a) T. L. Banfield and D. Husain, Electronic energy transfer from triplet state acridine to paramagnetic ions, *Trans. Faraday Soc.*, 1969, **65**, 1985; (b) A. W. Varnes, R. B. Dodson and E. L. Wehry, Interactions of transition-metal ions with photoexcited states of flavines. Fluorescence quenching studies, *J. Am. Chem. Soc.*, 1972, **94**, 946.
- 112 D. Rajasekaran, K. Venkatachalam and V. Periasamy, “On-off-on” pyrene-based fluorescent chemosensor for the selective recognition of Cu<sup>2+</sup> and S<sub>2</sub><sup>−</sup> ions and its utilization in live cell imaging, *Appl. Organomet. Chem.*, 2020, **34**, 1–9.
- 113 D. Phapale, A. Gaikwad and D. Das, Selective recognition of Cu(II) and Fe(III) using a pyrene based chemosensor, *Spectrochim. Acta, Part A*, 2017, **178**, 160–165.
- 114 M. A. Wani, P. K. Singh, R. Pandey and M. D. Pandey, Coumarin–pyrene conjugate: synthesis, structure and Cu-selective fluorescent sensing in mammalian kidney cells, *J. Lumin.*, 2016, **171**, 159–165.
- 115 H. Tomiyasu, N. Shigyo, X.-L. Ni, X. Zeng, C. Redshaw and T. Yamato, Positive allosteric binding behavior of pyrene-appended triazole-modified thiacalix[4]arene-based fluorescent receptors, *Tetrahedron*, 2014, **70**, 7893–7899.
- 116 G. Huang, C. Li, X. Han, S. O. Aderinto, K. Shen, S. Mao and H. Wu, Sensitive and selective detection of Cu(II) ion: A new effective 1,8-naphthalimide-based fluorescence ‘turn off’ sensor, *Luminescence*, 2018, 1–10.
- 117 R. Kumar, V. Bhalla and M. Kumar, Cu<sup>2+</sup> and CN-selective fluorogenic sensors based on pyrene-appended thiacalix[4]arenes, *Tetrahedron*, 2008, **64**, 8095–8101.
- 118 Y. R. Bhorge, H.-T. Tsai, K.-F. Huang, A. J. Pape, S. N. Janaki and Y.-P. Yen, A new pyrene-based Schiff-base: a selective colorimetric and fluorescent chemosensor for detection of Cu(II) and Fe(III), *Spectrochim. Acta, Part A*, 2014, **130**, 7–12.
- 119 W.-C. Lin, C.-Y. Wu, Z.-H. Liu, C.-Y. Lin and Y.-P. Yen, A new selective colorimetric and fluorescent sensor for Hg<sup>2+</sup> and Cu<sup>2+</sup> based on a thiourea featuring a pyrene unit, *Talanta*, 2010, **81**, 1209–1215.
- 120 Y. Chen, Q. Lv, Z. Liu and Q. Fang, Diaminomaleonitrile substituted pyrene as a solvent-dependent chemosensor for copper(II) ion and hypochlorite, *Inorg. Chem. Commun.*, 2015, **52**, 38–40.
- 121 S. Goswami, S. Chakraborty, S. Paul, S. Halder, S. Panja and S. K. Mukhopadhyay, A new pyrene based highly sensitive fluorescence probe for copper(II) and fluoride with living cell application, *Org. Biomol. Chem.*, 2014, **12**, 3037–3044.