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Natural Trojan horse inhibitors of aminoacyl-tRNA synthetases†

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For most antimicrobial compounds with intracellular targets, getting inside the cell is the major obstacle limiting their activity. To pass this barrier some antibiotics mimic the compounds of specific interest for the microbe (siderophores, peptides, carbohydrates, etc.) and hijack the transport systems involved in their active uptake followed by the release of a toxic warhead inside the cell. In this review, we summarize the information about the structures, biosynthesis, and transport of natural inhibitors of aminoacyl-tRNA synthetases (albomycin, microcin C-related compounds, and agrocin 84) that rely on such “Trojan horse” strategy to enter the cell. In addition, we provide new data on the composition and distribution of biosynthetic gene clusters reminiscent of those coding for known Trojan horse aminoacyl-tRNA synthetases inhibitors. The products of these clusters are likely new antimicrobials that warrant further investigation.

Introduction

Aminoacyl-tRNA synthetases (aaRSs) are essential cellular enzymes that catalyze the attachment of tRNAs to cognate amino acids and thus ensure correct translation of genetic information into proteins. The tRNA charging proceeds in two steps: first, an amino acid is activated by the formation

of an aminoacyl-adenylate; second, the aminoacyl group is transferred to the 3'-terminal adenosine of cognate tRNA (Fig. 1A). Both steps are performed in a single catalytic center, and some aaRSs bind all three substrates – amino acid, ATP, and tRNA – simultaneously.¹ While the mechanism of tRNA charging is conserved, two distinct aaRS classes with multiple subclasses are distinguished (Fig. 1B). In class I aaRSs, the catalytic domain adopts the Rossmann fold; most enzymes from this class function as monomers. The catalytic domains of class II aaRSs are organized as seven β -strands flanked by α -helices and most of these enzymes form dimers or oligomers.²

As aaRSs perform an essential function, inactivation of any one of them leads to growth inhibition and, eventually, cell death. A number of natural antibiotics targeting tRNAs are

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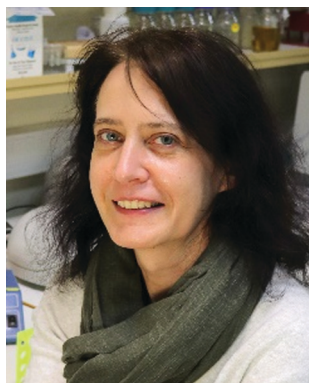


Fig. 1 (A) Aminoacylation of tRNA catalyzed by aminoacyl-tRNA synthetases. (B) The structures of *E. coli* LeuRS (class I, PDB ID: 4AQ7¹³⁶) and *E. coli* ThrRS (class II, PDB ID: 1QF6,¹³⁷ only one of the two monomers is shown) tRNA (blue) complexes. Catalytic domains are shown in orange, anticodon-binding domains – in pink, editing domains – in green, leucine-specific domain of LeuRS – in yellow, C-terminal domain – in red, zinc domain – in violet. Representatives of the two classes of aaRSs are listed together with their known specific inhibitors of bacterial origin (shown in italic). Compounds employing the Trojan-horse strategy and covered in the review are highlighted with red font.

known, two of which are approved for medical³ and veterinary⁴ use. With the exception of purpuromycin,⁵ which acts on

multiple aaRSs, all other natural inhibitors are specific to certain aaRSs.^{6–18} (Fig. 1B). The majority of aaRS-targeting antibiotics bind in the catalytic center and mimic either the cognate amino acid (e.g., indolomycin),¹⁹ aminoacyl-adenylate (e.g., mupirocin)²⁰ or charged tRNA (e.g., halofuginone).²¹ For reviews covering the diversity of natural aaRS inhibitors and their mechanisms of action see Cochrane *et al.*,²² Chopra & Reader²³ and Ho *et al.*²⁴

For many antibiotics with intracellular targets efficient internalization by susceptible bacteria is a key step limiting their bioactivity.²⁵ Little is known about the transport of aaRS inhibitors inside the bacterial cell.^{26,27} Passive transport is especially challenging for molecules with multiple polar and hydrophilic groups, which is the case for aaRS-targeting drugs mimicking aminoacyl-adenylates. In this review, we cover three classes of naturally occurring antimicrobial compounds that target aaRSs and exploit the Trojan horse strategy to get inside the bacterial cell. These molecules comprise two functional



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parts: one that hijacks a specific transporter by mimicking a compound valuable for the cell (siderophore, peptide, opine) and another that acts as a detachable warhead killing the cell once released inside its cytoplasm. We cover existing information about the structures, modes of transport and action, genetics, enzymology of the biosynthesis, and mechanisms of immunity for three known groups of naturally occurring Trojan-horse inhibitors: albomycin, microcin C-related compounds, and agrocin 84. In addition, we reveal several previously overlooked gene clusters that likely encode new aaRS inhibitors.

1. Albomycins

History of discovery and biological activity. In 1947, during the screening of a *Streptomyces griseus* strain collection for the production of antibacterial substances, a new antibiotic, which showed no cross-resistance with previously identified streptomycin was discovered in the Selman Waksman laboratory.²⁸ The compound was designated grisein. In parallel, a compound with potent antibacterial activity against a number of clinically relevant microorganisms was purified from *Streptomyces subtropicus* (former *Actinomyces subtropicus*) by the Gause group in the USSR.²⁹ The compound was named albomycin and found clinical use. In the mid-1950s, when a couple of ampules with albomycin penetrated through the Iron Curtain to Western labs it became evident that albomycin and grisein were identical.³⁰ Albomycin (or its congeners) were rediscovered several times and named alveomycin, LA 5352, Ro 5-2667, etc.³¹

Albomycin is active against a wide range of Gram-positive and Gram-negative bacteria including clinically relevant staphylococci, streptococci and various enterobacteria.^{32–34} Albomycin has a remarkably low (4 to 62 nM) minimal inhibitory concentrations (MIC) for *Streptococcus pneumoniae* strains.^{35,36} The efficacy of albomycin treatment of *S. pneumoniae* infection in a mouse model is similar to that of amoxicillin.³² Extensive preclinical and clinical studies carried out in the early 1950s showed exceptional safety of albomycin and its efficacy in treatment of meningitis and pulmonary infections, including those caused by penicillin-resistant pneumococci.³⁴ Recent elaboration of total chemical synthesis³⁵ allows large-scale production of albomycin and its analogs, opening way for their development into effective antibacterial drugs.

Structure and mechanism of action. The albomycin molecule consists of 4'-thioxylofuranosyl pyrimidine covalently bound to an iron-chelating tri-N5-hydroxy-N5-acetyl-L-ornithine siderophore through a peptide linker (Fig. 2A).³⁷ The three forms of albomycin initially detected by Waksman and later designated as δ_1 , δ_2 , and ϵ differ in substituents at the nucleobase. The major and most potent³⁵ congener δ_2 contains a carbamoyl group at N4 and a methyl group at the N3 position of cytosine. Albomycin ϵ is a biosynthesis intermediate lacking the carbamoyl modification.³⁸ The δ_1 congener is a 3-methyluridin analog of albomycin ϵ and is believed to be a synthesis byproduct.

The iron-chelator portion serves as a vehicle for active delivery of albomycin warhead inside both Gram-positive and Gram-negative bacterial cells through the ferrichrome-specific transporter system.^{36,39} In *E. coli*, albomycin first binds to the

outer membrane receptor FhuA. FhuA is a β -strand barrel buried in the outer membrane. An N-terminal domain located inside the barrel serves as a cork.⁴⁰ Albomycin competes for the binding site with ferrichrome, the natural FhuA substrate (Fig. 2C). Binding of a ligand promotes the FhuA interaction with TonB, which in turn stimulates a rearrangement of the cork domain and the release of the ligand into the periplasmic space. The energy required for translocation is provided by the proton motive force transduced by the TonB-ExbB-ExbD complex.⁴¹ Once in the periplasm, albomycin binds FhuD, a periplasmic subunit of the inner membrane ABC transporter FhuB/FhuC (Fig. 2D).^{42,43} Inside the cell, Fe^{3+} is reduced to Fe^{2+} , which dissociates from the chelator moiety,³⁹ and the siderophore part is cleaved off by the PepN peptidase.⁴⁴ As a result, the toxic nucleoside part is accumulated in the cytoplasm of *E. coli* in ~ 500 -fold excess over the concentration of antibiotic in the medium.³⁹ When added directly to bacterial culture, the nucleoside portion of albomycin does not inhibit cell growth. Thus, albomycin is a Trojan horse agent. The initial rate of Fe^{3+} transport by albomycin inside the cell is only two times lower than that by ferrichrome.³⁹ Thus, the antibiotic not only kills competing bacteria but is also an efficient iron-scavenger for the producer (Fig. 2B).

The target of albomycin was discovered serendipitously during a screening for inhibitors of bacterial aaRSs. SB-217452, a metabolite of *Streptomyces* sp. ATCC 700974, was shown to inhibit *Staphylococcus aureus* seryl-tRNA synthetase (SerRS) at low nanomolar concentration.¹⁵ The compound was identified to be the seryl-linked nucleoside moiety released upon the cleavage of the iron carrier part of albomycin δ_2 . Molecular modeling of SB-217452 in complex with SerRS shows that the N4 carbamoyl group modified cytosine structurally mimics adenine of the natural substrate seryl-AMP. A more extensive network of hydrogen bonding formed by SB-217452 compared to seryl-AMP within the active center of the enzyme ensures its higher affinity.^{45,46}

Biosynthesis and immunity. The biosynthetic gene cluster (BGC) of albomycin in the genome of *Streptomyces* sp. ATCC 700974 comprises 18 genes³⁸ (Fig. 3B). Biosynthesis of albomycin begins with the assembly of ferrichrome, a siderophore based on a nonribosomally synthesized tripeptide. Ferrichrome is produced on nonribosomal peptide synthetase (NRPS) AbmQ, which contains one adenylating (A) and two condensation (C) domains linked *via* thiolation domains (T) (Fig. 3A). The presence of a single adenylating domain is a unique feature of AbmQ and implies that this domain supplies substrates for both condensation domains. The substrate for AbmQ, N5-acetyl-N5-hydroxy-L-ornithine, is generated from L-ornithine by AbmB, a flavin-dependent ornithine monooxygenase, and AbmA, an acetyltransferase. In contrast to most NRPSs, AbmQ does not have a thioesterase domain and the newly synthesized tripeptide is directly transferred from the C-terminal peptidyl-carrier domain to seryl-nucleoside SB-217452.^{38,47} The biosynthetic pathway for SB-217452 is not fully established. It is thought to include several condensation and epimerization steps resulting in 6'-amino-4'-thioheptosyl cytosine linked to L-serine *via* an amide bond



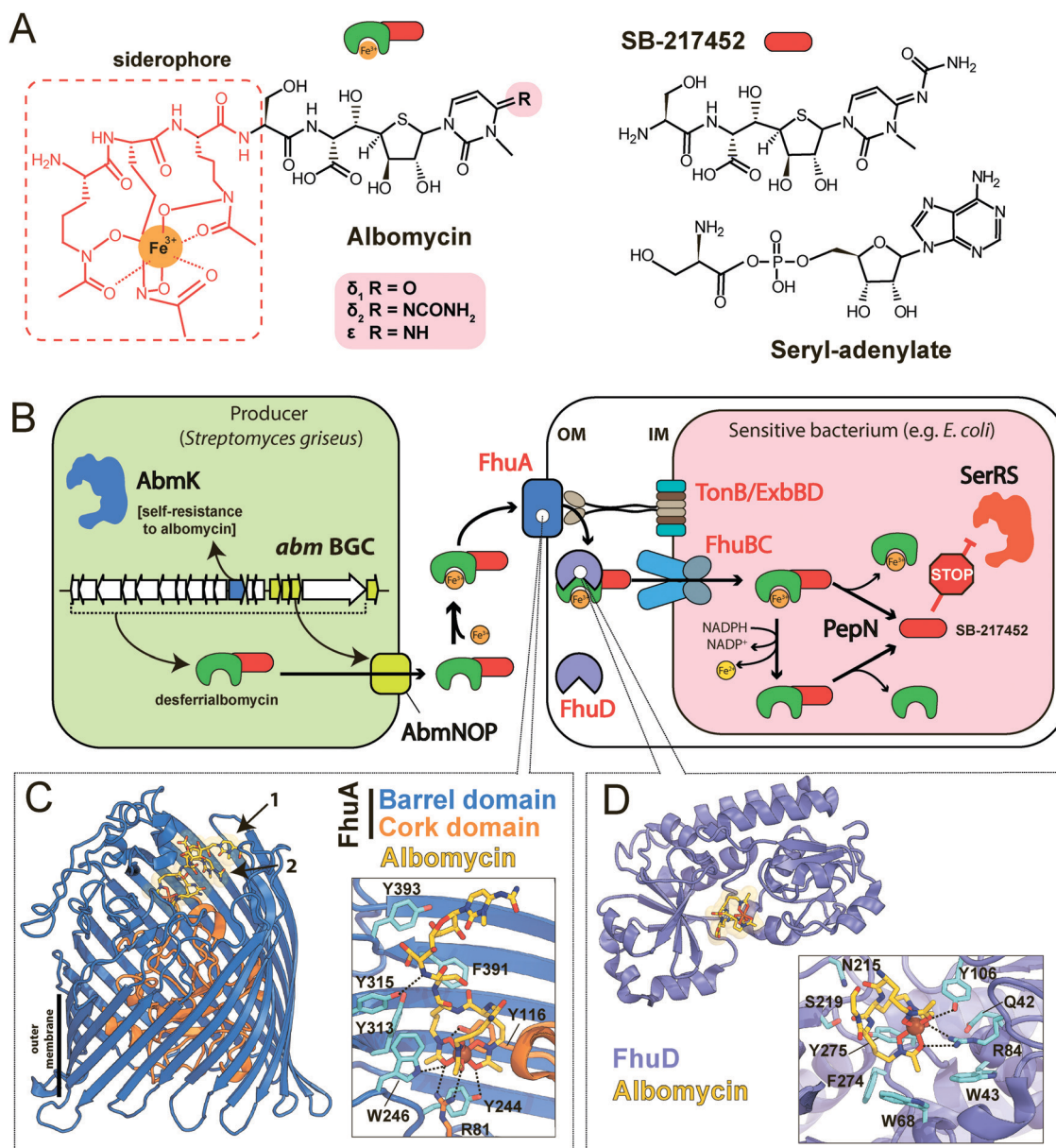


Fig. 2 (A) Structures of albomycins, albomycin toxic moiety (SB-217452) and seryl-adenylate. (B) A scheme showing the biosynthesis, transport and intracellular processing of albomycin. IM – inner membrane, OM – outer membrane, BGC – biosynthetic gene cluster, SerRS – seryl-tRNA-synthetase. (C) The crystal structure of the FhuA outer membrane transporter in complex with albomycin (PDB ID: 1QKC⁴⁰). Two conformational isomers of albomycin are shown (black arrows: 1 – extended form, 2 – compact form). A close-up view of the extended albomycin form binding site is shown, residues involved in the binding are indicated, hydrogen bonds are shown as black dashed lines. (D) The crystal structure of the FhuD periplasmic binding protein in complex with albomycin (PDB ID: 1K7S⁴³). Residues involved in the formation of the albomycin binding site are indicated; hydrogen bonds are shown as black dashed lines. Electron density for the toxic moiety of albomycin is not observed in this structure.

(Fig. 3A).^{47,48} Neither precursors for the synthesis of 5'-oxo-4'-thioribonucleoside nor enzymes involved are known.

The most remarkable part of the albomycin synthesis pathway is the formation of an amide bond between the thionucleoside and the L-serine, which results in SB-217452, a highly toxic compound. *In vitro* reconstruction of this step shows that AbmF uses seryl-adenylate produced by seryl-tRNA synthetases as a seryl donor for the amide bond formation, although a possibility that seryl-tRNA can also serve as an aminoacyl donor was not ruled out.⁴⁷ Interestingly, the albomycin BGC encodes AbmK, which is a

functional SerRS.⁴⁹ Unlike the housekeeping SerRS, AbmK is insensitive to SB-217452 *in vitro* and confers resistance to albomycin in *E. coli* when overexpressed.⁴⁹ *In vitro* *Streptomyces* sp. ATCC 700974 housekeeping SerRS is substantially less efficient in the coupled reaction of thionucleoside biosynthesis since it is inhibited by the end product, SB-217452.⁴⁷ The K_M for serine of AbmK is ~ 20 times higher than that of housekeeping SerRS.⁴⁹ Thus, AbmK may serve a triple function in the producing strain – providing the substrate for albomycin biosynthesis,⁴⁷ conferring immunity to the toxic compound produced,⁴⁹ and,





Fig. 3 (A) Known steps in the biosynthesis of albomycin and the enzymes involved. Domains of NRPS AbmQ are indicated: A – adenylating domain, C – condensation domain, T – thiolation domain. Fragments of the molecule attached at different steps of the biosynthetic pathway are shown in different colors. (B) Biosynthetic gene cluster of albomycin (*abm*, red background) and found BGCs containing subsets of *abm*-like genes. Nomenclature of genes in the *ctj* gene cluster is from Zeng *et al.*³⁸ The color scheme of enzymes in panel A matches that of genes in B. Predicted functions of the gene products are listed below. Protein IDs are shown for AbmF homologs. ABC – ATP binding cassette, CoA – coenzyme A, HAD – haloacid dehalogenase-like, MFS – major facilitator superfamily, PLP – pyridoxal phosphate, SAM – S-adenosyl methionine.

regulating albomycin biosynthesis by restricting it to conditions of high concentrations of intracellular serine.

BGCs of putative seryl-nucleosides. To date, albomycin is the only known representative of its class. We performed a



BLASTP⁵⁰ search of the NCBI protein RefSeq database using the sequence of AbmF as a bait. Top 12 hits with *E*-values less than 1×10^{-35} and query coverage above 85% were hypothetical proteins whose genes were located close to SerRS encoding genes. Within these 12 hits, in addition to trivial cases of BGCs encoding a complete pathway for albomycin biosynthesis from various *Streptomyces*, two additional groups of BGCs were identified (Fig. 3B). The first group contains just two very similar clusters, one from *S. vitaminophilus* ATCC 31673, and another, *ctj* from *Streptomyces* sp. C, that was earlier discovered by Zeng *et al.*³⁸ Though almost a full set of genes required for seryl-thiofuranoside cytosine biosynthesis is present in both clusters, the *abmC* and *abmL* orthologs are absent (Fig. 3B). While the function of AbmL in albomycin biosynthesis is not established, AbmC transfers the ferrichrome moiety to seryl-nucleoside SB-217452 and is thus essential for the production of the Trojan horse compound.⁵¹ The NRPS enzymes encoded in both clusters are highly similar and contain an adenylating domain, a thiolation domain but no condensation domains.³⁸ The product of NRPS may be transferred to the seryl-nucleoside part either by carboxylate-amine ligase family protein (CtjK in *Streptomyces* sp. C), or by CtjH, a protein of unknown function. Presumably, *Streptomyces* sp. C and *S. vitaminophilus* ATCC 31673 produce different SB-217452-like nucleosides, since the *ctj* cluster lacks one of the rSAM enzymes, an ortholog of AbmJ, which is responsible for D-ribo to D-xylo epimerization of the furanose ring in albomycin. It is worth mentioning that the D-ribo form of albomycin has significantly reduced antimicrobial activity.⁴⁸ Both clusters are apparently functional, as the specific enzymatic activity of carbamoyl transferase CtjE and methyltransferase CtjF toward cytidine was confirmed.³⁸ Moreover, *ctjE* complemented the deletion of the *abmE* gene from the albomycin cluster. Interestingly, both clusters encode an HAD-family hydrolase (nucleosidase), which is lacking from other clusters and may serve as an additional immunity gene.

The second group of putative seryl-thiofuranoside cytosine synthesizing BGCs was not previously described (Fig. 3B). Its representatives are found in the genomes of *Proteobacteria* from different classes. Despite major rearrangements in their architecture, these BGCs contain roughly the same set of genes. These include genes encoding cytosine-modifying carbamoyl-transferase (AbmE-like), cytosine methyltransferase (AbmI-like), and thiofuranose-seryl assembly and epimerization enzymes homologous to AbmD, AbmH, and AbmF. The BGC from *Trinickia caryophylli* DSM 50341 also contains an *abmJ* ortholog. In contrast to albomycin BGCs, clusters from *Proteobacteria* entirely lack genes required for assembly of the siderophore part. This implies either a different mechanism of end product uptake by the sensitive cells or the coupling to a siderophore molecule whose biosynthesis is encoded elsewhere in the genome of the producer.

2. Microcin C and related compounds

History of discovery and biological activity. Microcin C (McC, initially named “microcin C7”) was purified from *Escherichia coli* BM7006,⁵² a strain first isolated by Chabbert in 1950.⁵³ The same

compound was independently discovered and described under the name “microcin C51”.⁵⁴ Eventually, it was shown that 1–2% of *E. coli* strains isolated from humans are McC producers.^{55–57}

Enterobacteriaceae such as *Escherichia*, *Salmonella*, and *Shigella*, and some strains of *Klebsiella*, are susceptible to McC at low micromolar concentrations; *Pseudomonas* and Gram-positive bacteria are resistant.⁵⁸ McC may provide competitive advantage to producing strains in natural environments. Moreover, the probiotic activity of *E. coli* strain H22 in a mouse model was attributed to McC production.⁵⁹

Structure and mechanism of action. The *E. coli* McC is the best-studied representative of a family of McC-like compounds that were either experimentally identified or bioinformatically predicted in both Gram-positive and Gram-negative bacteria.^{60,61} The *E. coli* McC consists of a formylated heptapeptide MRTGNAisoN coupled with an adenyl moiety through a phosphoramidate linkage (Fig. 4A).⁶² The phosphate group is decorated with an aminopropyl. Many McC-like compounds are peptidyl-adenylates with no additional decorations.⁶¹ Though peptide parts of some McC-like compounds can be as long as 56 amino acids (Table S1, ESI[†]) most McC-like compounds are based on heptapeptides.⁶¹ The nucleotide part of McC-like compounds can be either an adenyl or cytidyl (Fig. 4A) with various additional modifications.^{63,64}

All studied McC-like compounds share the Trojan horse mechanism of action. The peptide part serves as a vehicle to deliver the isoasparaginyl-nucleotide warhead inside the cell.¹¹ The route of *E. coli* McC entrance into the cell is not fully established. Despite multiple mentions in the literature,⁶⁵ no direct evidence that OmpF is the primary porin responsible for McC passage through the outer membrane of Gram-negative bacteria exists. Most probably, several porins are involved. Once *E. coli* McC reaches the periplasmic space it interacts with YejA, the substrate-binding subunit of the YejABEF ABC transporter (Fig. 4B).⁶⁶ Though exact cellular function of this transporter in *E. coli* is not established, homologs from *Salmonella*, *Brucella* and *Pseudomonas* may be involved in resistance to antimicrobial peptides.^{67–70}

Presumably, the fact that peptide parts of most McC-like compounds are seven amino acids long is explained, at least in part, by the binding preferences of YejA. Using chemically synthesized *E. coli* McC analogs it was shown that shortening the peptide length to six amino acids dramatically reduces bioactivity due to inefficient uptake through YejABEF.⁷¹ Increase of the peptide part length up to 10–12 amino acids stimulated the uptake.⁶¹ Peptidyl-nucleotides with the peptide part lengths around 20–25 amino acids can enter *E. coli* not only through YejABEF but also through SbmA,⁷² a transporter of peptide antibiotics such as microcin B17 (43 amino acids) or microcin J25 (21 amino acids).^{73,74} McC-like compounds with even longer peptide parts, *i.e.*, the 42 amino acid long McC from *Yersinia pseudotuberculosis* (Fig. 4A), undergo endoproteolytic processing prior to export, which leads to bioactive compounds with shorter (11 amino acids in the case of *Y. pseudotuberculosis* McC) peptide parts.⁷⁵

Once *E. coli* McC gets inside the bacterial cytoplasm, its peptide part is deformylated and six N-terminal residues are



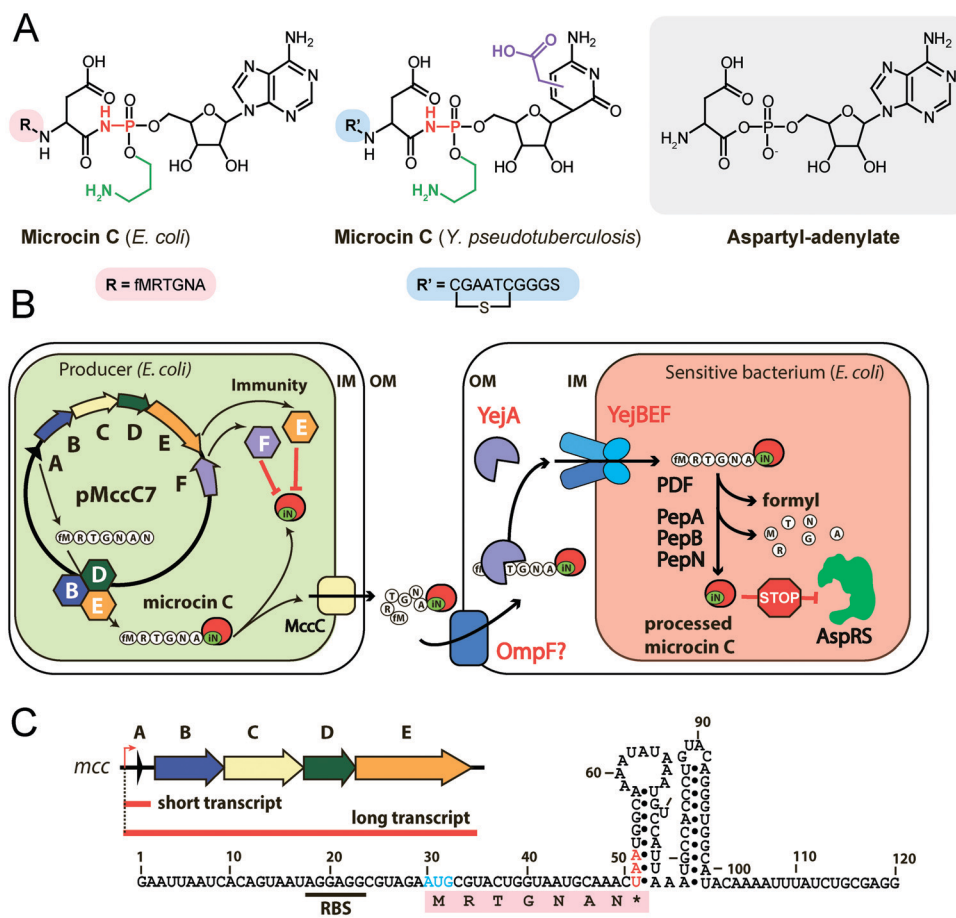


Fig. 4 (A) Chemical structures of peptidyl-nucleotide antibiotics produced by *Escherichia coli* and *Yersinia pseudotuberculosis*. The nonhydrolyzable P–N bond is shown in red, the aminopropyl decoration is green, the carboxymethyl attached to the cytosine ring is violet. The peptide part is shown below in single-letter code. Aspartyl-adenylate is shown on the grey background for comparison. (B) Schematic representation of microcin C biosynthesis, transport and processing. PDF – peptide deformylase, OM – outer membrane, IM – inner membrane, AspRS – aspartyl-tRNA synthetase. (C) Regulation of microcin C production by an internal ribosome-dependent transcription terminator located between the *mccA* and *mccB* genes. The secondary structure of short *mccA* transcript is shown.⁶¹ Start codon is blue, stop codon is red. RBS – ribosome-binding site.

removed by aminopeptidases. In *E. coli*, any one of the three aminopeptidases, PepA, PepB, or PepN, is capable of intracellular McC processing (Fig. 4B).⁷⁶ The final product of processing is an aminopropylated isoasparaginyl-adenylate. Processed *E. coli* McC mimics aspartyl-adenylate, the intermediate of the tRNA^{Asp} aminoacylation reaction (Fig. 4A), and inhibits aspartyl-tRNA synthetase (AspRS).¹¹ Inhibition of AspRS results in the accumulation of uncharged tRNA^{Asp}, translation inhibition, stringent response and cessation of cell growth.^{11,77}

Although the structures of AspRS in complex with several chemically synthesized analogs of processed McC have been determined,⁴⁵ the exact binding mode of processed McC to AspRS is yet to be revealed. Given the variety of known and proposed (see below) auxiliary modifications in the nucleoside part of McC-like compounds, the catalytic center of AspRS should be flexible enough to accommodate structurally diverse aminoacyl-nucleotides.⁴⁵

Biosynthesis and regulation. McC-like compounds belong to the ribosomally synthesized post-translationally modified peptides (RiPPs) family since they are produced by modification

of a gene-encoded peptide precursor by dedicated enzymatic machinery.⁷⁸ Peptide precursors of most RiPPs comprise the leader part, which is required for the specific recognition by the enzymes introducing the post-translational modification, and the core part where modifications are introduced to.⁷⁹ Uniquely, the core peptide of McC-like compounds is represented by a single C-terminal amino acid residue. In contrast to other RiPPs, the fully modified McC-like compounds are exported from the producer cell with complete or partial leader peptide attached to the miniaturized core.

The minimal microcin C BGC (*mcc*) consists of just three genes: *mccA*, encoding the precursor peptide; *mccB*, coding for a ThiF-like nucleoside transferase; and *mccC*, encoding an export pump (Fig. 6 (1)). Auxiliary genes present in many *mcc* BGCs encode enzymes responsible for secondary post-translational modifications or self-immunity, however, the function of most of them is not identified yet (Fig. 6).

When the sequence of the *E. coli* *mcc* gene cluster was determined (Fig. 6 (2)), it became apparent that the *mccA* gene codes for a MRTGNAN heptapeptide, while mature McC has the



fMRTGNA_{isoN} peptide part.⁸⁰ This discrepancy was resolved by work from the Walsh lab, when they showed that nucleoside coupling to the precursor peptide by adenylyltransferase MccB proceeds in two steps through a succinimide intermediate.^{81,82} The opening of the succinimide ring results in the isomerization of the asparagine residue, converting it to isoasparagine. The reaction consumes two molecules of ATP per molecule of peptide-nucleotide synthesized (Fig. 5B). While one would expect that a similar mechanism could be employed with peptide precursors containing C-terminal Gln, however, at least the *E. coli* MccB is unable to modify such a peptide.⁷² In fact, the presence of a C-terminal asparagine is

the only strict requirement shared by known McC-like precursor peptides.

The crystal structure of MccB reveals a homodimer with two active centers (Fig. 5A). Both steps of the adenylation reaction are catalyzed within the same active center. The N-terminal part of one subunit (so-called R_iPP Recognition Element, “RRE”) and the central “crossover loop” of the opposite protomer form the peptide clamp domain responsible for MccA binding.^{82,83} The formyl group on the N-terminal methionine of the precursor heptapeptide strongly improves the efficiency of adenylation by promoting productive binding of the precursor peptide to MccB.⁸⁴



Fig. 5 (A) The structure of the dimeric nucleoside transferase MccB from *E. coli* in complex with *N*-formylated microcin C (PDB ID: 6OM4).⁸⁴ Adenylation domains and N-terminal RREs are shown in different colors, black arrows indicate “crossover loops”. Close-up views: (left) the active site of the adenylation domain with key residues required for efficient catalysis (D214 and Y239) is shown in stick representation, black arrow indicates the P-N bond; (right) the binding site of MccA peptide N-terminus formed by RRE and “crossover loop” (violet), residues 313–347 (catalytic domain) are removed for clarity. (B) The mechanism of P–N bond formation by nucleoside transferase MccB. R stands for the fMRTGNA peptide. Two molecules of ATP consumed per molecule of McC produced are shown in violet, released AMP is blue. (C) Installation of aminopropyl decoration by the MccD and MccE1 enzymes. SAM – *S*-adenosyl methionine, MTA – 5'-methylthioadenosine.



Strikingly, MccB is rather promiscuous in its choice of NTP substrates. *In vitro*, the *E. coli* precursor peptide can be modified with either AMP, CMP, GMP or UMP moieties. *In vivo*, the reaction is highly specific and, depending on the enzyme, exclusively yields either peptidyl-adenylates (e.g., when catalyzed by *E. coli* MccB) or peptidyl-cytidylates (e.g., when catalyzed by *Bacillus amyloliquefaciens* MccB).⁶⁴ All known MccB enzymes catalyzing *in vivo* cytidylation contain a C-terminal methyltransferase domain responsible for the additional decoration of the nucleobase with carboxymethyl. This reaction requires carboxy-*S*-adenosyl methionine (cxSAM) as a substrate. The cxSAM is synthesized from *S*-adenosyl methionine (SAM) and prephenate by MccS, an enzyme encoded exclusively in BGCs producing cytidylated McC-like compounds.^{64,75} The exact position of the carboxymethyl group on the cytidine base remains unknown.

Auxiliary post-translational modifications of RiPPs increase their bioactivity and/or are required to overcome the immunity of target bacteria.⁸⁵ The aforementioned aminopropyl decoration of the phosphate found in *E. coli* McC and in some other McC-like compounds serves both purposes: it increases the toxicity of peptidyl-nucleotides and allows to escape the action of some immunity proteins. Noteworthy, most *Salmonella* species are resistant to McC-like peptidyl-adenylates but sensitive to aminopropylated compounds.⁸⁶ The aminopropyl is introduced *via* a two-step reaction.⁸⁷ During the first step, a class I methyltransferase MccD transfers the 3-carboxy-3-aminopropyl moiety from SAM to the peptidyl-nucleotide substrate, releasing methylthioadenosine (MTA) as a byproduct (Fig. 5C). Since MTA is a potent inhibitor of SAM-binding enzymes, the reaction efficiently proceeds only in the presence of the Mtn nucleosidase, which is encoded in the core genome of many bacteria.⁸⁸ The second step is decarboxylation of the 3-carboxy-3-aminopropylated peptidyl-nucleotide intermediate by MccE1 (in the case of *E. coli* – the N-terminal domain of a fused protein MccE) (Fig. 5C).^{75,87}

E. coli McC is produced in the early stationary phase of growth, and its synthesis is tightly regulated.⁸⁹ Transcription of the *mccABCDE* operon is initiated by the σ^S (σ^{38})-containing RNA polymerase⁹⁰ in a CRP-dependent manner.^{91,92} A ribosome-dependent transcription terminator is commonly found in the *mccA-mccB* intergenic regions of *mcc* operons (Fig. 4C).⁶¹ In *E. coli*, the function of this terminator leads to the production of two types of mRNA molecules: a short one, which contains the *mccA* ORF only, and a long one, comprising the entire operon.⁶¹ Co-transcriptional recognition of the Shine–Dalgarno sequence of *mccA* by the ribosome stimulates transcription termination. The ribosome binding to short mRNA is virtually irreversible, which allows multiple rounds of translation without ribosome dissociation and ensures that enough peptide precursor substrate is produced for the action of the maturation enzymes.⁶¹

Immunity. The McC-producing bacteria are vulnerable to the action of isoasparaginylnucleotide that accumulates inside their cytoplasm.⁷⁷ To prevent the damage to the producer, diverse genes responsible for McC immunity are often incorporated in *mcc*-like BGCs. The Gcn5-related *N*-acetyltransferase (GNAT)

MccE2 provides immunity by acetylating processed McC and many other aminoacyl nucleosides at the alpha-amino group, which likely prevents the binding to cognate aminoacyl-tRNA synthetases (Fig. 7B).^{93,94} MccE2 enzymes appear to be specific to the nucleoside part of their substrates. For instance, *E. coli* MccE2 protects cells from peptide-adenylate compounds, while the *Y. pseudotuberculosis* homolog provides resistance to McCs containing carboxymethylated cytosine.⁶⁴

The MccF protein encoded in the *E. coli mcc* cluster provides self-immunity by cleaving off the aspartate residue or the oligopeptide from, respectively, processed or mature McC (Fig. 7A).⁹⁵ MccF is the S66 family serine peptidase⁹⁶ and is similar to LdcA, a carboxypeptidase, which cleaves amide bonds between α - and β -amino acids during murein recycling.⁹⁷ However, the latter enzyme is unable to protect *E. coli* from McC-like compounds. MccF homologs are encoded in several *mcc*-like BGCs (Fig. 6) and by numerous stand-alone bacterial genes.⁹⁶ The stand-alone MccF ortholog from *B. anthracis* can hydrolyze peptidyl-adenylates and isoasparaginylnucleotides with or without aminopropyl decoration and provides McC resistance to *E. coli* when overexpressed.⁹⁶

Another immunity protein, MccH targets the phosphoramidate linkage of isoasparaginylnucleotides. It is a member of a vast family of Histidine-Triad Hydrolases (HITs). MccH is similar to highly conserved HinT phosphoramidases present in almost all organisms.⁹⁸ The function of HinT is poorly understood and its natural substrates are unknown. HinT from *E. coli* or *Hyalangium minutum* are unable to hydrolyze isoasparaginylnucleotide. In contrast, MccH, which was first identified in the *mcc* BGC from *H. minutum*, evolved to specifically cut the N-P bond in isoasparaginylnucleotide, releasing AMP and isoasparagine (Fig. 7C).⁸⁶ A number of bacterial MccH orthologs not linked to *mcc*-like BGCs were identified. When overexpressed, these enzymes provided resistance to simple McC-like peptidyl-adenylates but not to aminopropylated forms.⁸⁶

BGCs of McC-like compounds. Recently, a number of *mcc*-like BGCs were predicted in bacterial plasmids and chromosomes (Table S1, ESI†).⁶¹ Some of these were validated earlier;⁷² validation of several others is shown in Fig. S1 (ESI†). The majority of the BGCs consist of just three core genes, *mccABC*, and presumably encode the pathway of biosynthesis of simplest peptidyl-nucleotides (Fig. 6 (1)). However, many *mcc*-like BGCs display more complex architectures (Fig. 6).

As already mentioned, clusters encoding MccB proteins with C-terminal carboxymethyltransferase domains also encode cxSAM synthases MccS, which are absent from other *mcc* clusters. Experimentally validated products of such clusters are cytidine-containing antibiotics with carboxymethyl decoration installed by C-terminal carboxymethyltransferase domains of their MccBs (Fig. 6 (14–20), see also above).⁶⁴ Two genes *mccX* (Fig. 6 (15, 18)) and *mccY* (Fig. 6 (16, 17)) are also found exclusively in this group of clusters, suggesting that the proteins they encode may specifically target the carboxymethylated cytosine nucleobase. While MccX has no sequence similarity with any proteins with established function,⁹⁹ MccY is homologous to carbamoyl transferases. Similarly to AbmE from the albomycin



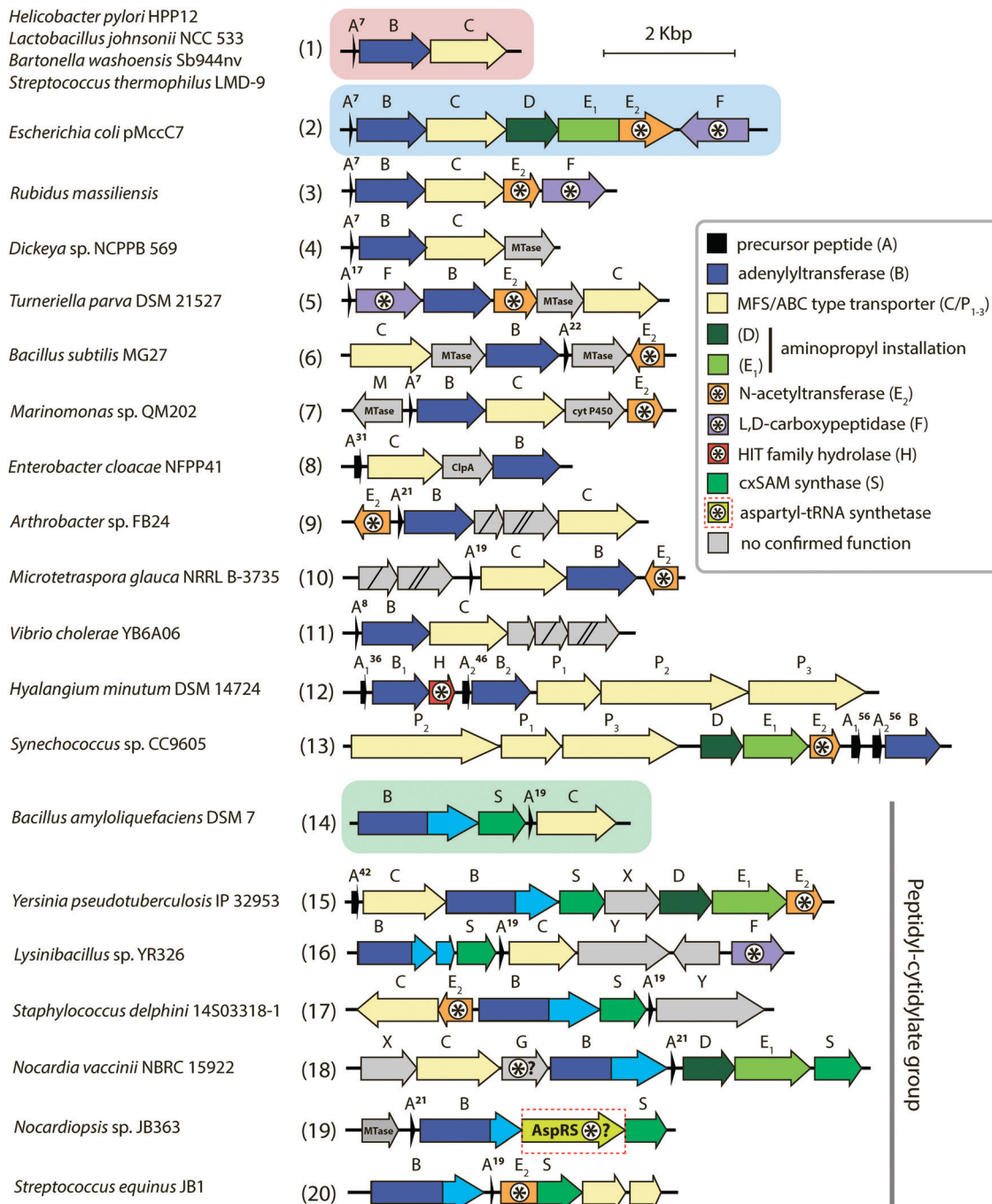


Fig. 6 Biosynthetic gene clusters of microcin C-like compounds found across bacterial genomes. Clusters containing minimal sets of genes required for the production of peptidyl-adenylates and peptidyl-cytidylates are shown on red and green background, respectively. The archetypal and best-studied *mcc* BGC from the *E. coli* pMcc7 plasmid is shown on blue background. The number of amino acid residues in precursor peptides is indicated near the A genes. Genes involved (or proposed to be involved) in self-immunity are indicated with asterisks. Known functions for some gene products are listed in the right. Genes with no identified function are shown in grey. cyt P450 – cytochrome P450, HIT – histidine triad, MTase – methyltransferase.

biosynthesis pathway, MccY may add a carbamoyl decoration to the amino group of the carboxymethylated cytosine nucleobase.

SAM-dependent enzymes comprise the largest and most diverse group of tailoring enzymes encoded by *mcc*-like operons. Besides the already described 3-carboxy-3-aminopropyl transferase (MccD) and carboxymethyltransferase forming the C-terminal domain of some MccB proteins, genes encoding putative

SAM-dependent transferases are identified in many *mcc*-like BGCs (Fig. 6 (4–6)). However, as there are no close homologs with known substrate specificity for these enzymes, secondary post-translational modifications they perform are impossible to predict.

Analysis of the *mcc*-like BGCs composition suggests that the self-immunity strategies employed by Mcc producers extend beyond those already discussed. Some BGCs encode AspN-like



peptidases (Fig. 6 (18)) or ClpA homologs (Fig. 6 (8)), which may hydrolyze isoasparaginyl-nucleotides accumulating inside the producing cell. Several BGCs from *Actinobacteria* include *asnS* (aspartate tRNA synthetase) genes (Fig. 6 (19)), which, similarly to *abmK* from albomycin biosynthetic pathway (see above) or *agnB2* from agrocin 84 BGC (see below) may provide resistance to processed McC-like compounds.

In summary, the abundance and diversity of known or putative strategies of avoidance of self-intoxication by intracellularly accumulating processed McC-like compounds parallels the diversity of secondary post-translational modifications of the basic peptide-nucleotide backbone and suggest that no single strategy is universal: additional decorations attached to the isoasparaginyl-nucleotide core allow some non-cognate



Fig. 7 Proteins providing immunity to microcin C and their mechanisms of action. For each protein, structure, mode of substrate binding, and catalyzed reaction are shown. (A) The structure of *E. coli* carboxypeptidase MccF catalytic mutant (S118A) in complex with processed microcin C (PDB ID: 3TLC).¹³⁸ Hydrogen bonds are shown as black dashed lines, π - π stacking between W186 and adenine nucleobase is indicated, big red arrow points to the peptide bond cleaved by MccF. (B) The structure of *E. coli* acetyltransferase MccE2 (C-terminal domain of MccE, residues 405–589) in complex with coenzyme A and acetylated processed microcin C (PDB ID: 3R9G).⁹⁵ (C) The theoretical model of dimeric HIT-family hydrolase MccH from *Hyalangium minutum* in complex with AMP.⁸⁶



McC-like compounds to overcome the immunity of specific producers.

3. Agrocin 84

History of discovery, biological activity, structure, and mode of action. Crown gall disease caused by soil-borne tumorigenic strains of *Rhizobium radiobacter* (formerly classified as *Agrobacterium tumefaciens*) affects a wide spectrum of dicotyledonous plants and leads to significant losses in nursery stock of stone fruits and nut trees.¹⁰⁰ A nonpathogenic strain *Rhizobium rhizogenes* K84 (formerly *Agrobacterium radiobacter* K84¹⁰¹) isolated from a soil sample derived from an Australian plant nursery in 1970,¹⁰² was shown to prevent the development of agrobacterial infections when added to planting material.^{103,104} For almost half a century, this strain has been used commercially as a biocontrol agent against crown gall disease.¹⁰⁵

The activity of *R. rhizogenes* K84 against tumor-inducing strains is due to the production of an antimicrobial compound agrocin 84, a disubstituted adenine nucleotide containing 3'-deoxyarabinose instead of the ribose.^{106,107} The two moieties attached to the nucleotide core *via* phosphoramidate bonds are D-glucopyranose-2-phosphate at the N6 position of adenine and 2,3-dihydroxy-4-methylpentanamide at the phosphate group (Fig. 8A, left panel).^{108,109} The D-glucose-2-phosphate (Fig. 8A, red dashed frame) is required for the uptake by susceptible strains. Once the uptake moiety is removed, a toxic moiety (TM84, Fig. 8A, right panel) is released, making agrocin 84 a Trojan horse agent.¹¹⁰

Plant-pathogenic agrobacteria insert a specific region (T-DNA) derived from their Ti (Tumor inducing) plasmids into the nuclear genome of the plant cell.¹¹¹ Some T-DNA genes encode enzymes of the biosynthesis of opines – metabolites produced by the transformed plant cells and subsequently imported into the bacterial cells. These compounds serve as a source of carbon, phosphorus and nitrogen for the agrobacteria.^{112,113} The import of opines agrocinopines A and B is determined by the AccABCDE transporter encoded on the Ti plasmid (Fig. 8B). Agrocin 84 hijacks this transport system to get inside the cell of tumor inducing agrobacteria.¹¹⁴ Agrocinopine A is made of sucrose bound to L-arabinose-2-phosphate (Fig. 8A), while agrocinopine B has fructose instead of sucrose.^{115,116} Crystal structures (Fig. 8C) show that the periplasmic binding protein AccA recognizes both agrocin 84 and agrocinopine A through their pyranose-2-phosphate groups (D-glucose-2-phosphate in the case of agrocin and L-arabinose-2-phosphate in the case of agrocinopine, Fig. 8A, red dashed frames).¹⁰⁹ The subsequent processing step required for the release of agrocin 84 toxic moiety is catalyzed by phosphodiesterase AccF (Fig. 8B), which also performs the first step of agrocinopines catabolism.¹¹⁴ Since the *accABCDEF*G (agrocinopine catabolism) locus is part of the Ti-plasmid, agrocin-producing *R. rhizogenes* strains specifically inhibit the growth of tumor inducing strains but have no effect on strains that do not induce tumors.^{106,117} In other words, by mimicking the tumor-derived substance, agrocin 84 is able to find its way specifically into phytopathogenic bacteria harboring the Ti-plasmid.

TM84, the toxic moiety of agrocin 84, inhibits tRNA^{Leu} aminoacylation by leucyl-tRNA synthetase (LeuRS, Fig. 8B).⁶ TM84 is a structural mimic of leucyl-adenylate (Fig. 8A, grey background) with a nonhydrolyzable phosphoramidate bond (Fig. 8A, black arrow) connecting the nucleotide and the modified pentanamide. Unlike other stable leucyl-adenylate analogs, TM84 binds weakly to the LeuRS active site in the absence of tRNA^{Leu}. The 3'-terminal adenosine of tRNA^{Leu} forms a hydrogen bond with TM84, stabilizing the ternary complex (Fig. 8D).¹¹⁸

Biosynthesis and immunity. The production of agrocin 84 by *R. rhizogenes* K84 is associated with the presence of a ~45 kbp plasmid pAgK84, which can be transferred by conjugation to other rhizobia, turning them into agrocin producers and making them resistant to the action of the compound.^{119,120} As the transfer of pAgK84 to phytopathogenic strains can break down biocontrol, a strain lacking a 5.9 kbp fragment of pAgK84 required for the transfer was constructed.¹²¹ This strain, named *R. rhizogenes* K1026, had similar biocontrol properties as the parent strain K84 but was unable to transfer the resistance determinants.¹²² K1026 serves as an active ingredient in commercial biological pesticide NOGALL[®].

The exact pathway of agrocin 84 biosynthesis is unknown. The sequence of the pAgK84 plasmid reported by Kim *et al.*¹²³ allowed the identification of a 17-gene *agn* biosynthetic cluster required for agrocin 84 production and immunity (Fig. 9, pink background). The function of several Agn proteins was proposed based on sequence similarity with known enzymes. AgnA harbors a conserved adenylation domain characteristic of aaRSs and was thus proposed to catalyze the key step in which the precursor of pentanamide moiety is attached to the adenosine monophosphate *via* the phosphoramidate bond.¹²³ Our analysis suggests that AgnC1 has remote structural similarity to prolyl-tRNA synthetases (according to Phyre2¹²⁴ and HHpred¹²⁵) and may thus be another candidate for the enzyme attaching the modified pentanamide to the nucleotide moiety. AgnB1, AgnC3 and AgnC7 are SAM-dependent enzymes proposed to be involved in modified pentanamide biosynthesis. The involvement of SAM in agrocin 84 biosynthesis is supported by an observation that *R. rhizogenes* K84 with mutations in the chromosomal *achY* gene encoding S-adenosyl-L-homocysteine (SAH) hydrolase (an enzyme involved in SAH recycling back to SAM) do not produce agrocin.¹²⁶

The *agn* BGC (Fig. 9, pink background) includes two self-immunity genes.¹²³ *agnG* encodes a transmembrane pump involved in the export of the antibiotic; *agnB2* encodes a leucyl-tRNA-synthetase. Compared to its housekeeping counterpart, the AgnB2 enzyme is ~1000-fold less sensitive to TM84, providing the producer with alternative machinery for tRNA^{Leu} charging.^{6,127}

BGCs of putative agrocin 84-like compounds. Agrocin 434^{128,129} and agrocin 108^{129,130} are narrow-spectrum antimicrobials produced by the strains of the former genus *Agrobacterium* and active against other agrobacteria (Fig. S2, ESI[†]). However, these compounds are structurally distinct from agrocin 84 and no information about their modes of action is available, making





Fig. 8 (A) Chemical structures of agrocin 84, its toxic moiety (TM84), agrocinopine A and leucyl-adenylate. Pyranose-2-phosphate groups shared by agrocin and agrocinopine A are highlighted with red dashed polygons. Groups of atoms that are different between TM84 and leucyl-adenylate are shown in red. The black arrow indicates a nonhydrolyzable P–N bond in TM84. (B) The scheme showing the biosynthesis, transport, and mode of action of agrocin 84. IM – inner membrane, OM – outer membrane, BGC – biosynthetic gene cluster, LeuRS – leucyl-tRNA synthetase. (C) The details of the interaction between agrocin 84/agrocinopine A with the periplasmic solute binding protein (PBP) AccA (PDB IDs: 4ZEC/4ZEB).¹⁰⁹ Agrocinopine is coloured gold, agrocin 84 is green. Amino acid residues interacting with D-glucopyranose-2-phosphate moiety of agrocin are shown in the left corner. Hydrogen bonds are shown as black dashed lines. (D) The crystal structure of the ternary complex of LeuRS, TM84 and tRNA^{Leu} (PDB ID: 3ZGZ).¹¹⁸ Selected amino acid residues involved in the binding of TM84 in the active site of LeuRS are shown in the left corner together with the two 3'-terminal nucleotides of the tRNA^{Leu} acceptor stem. TM84 is green, tRNA^{Leu} is orange.

agrocin 84 the only compound of its class. We performed BLASTP searches for orthologs of proteins encoded in the *agn* biosynthetic gene cluster in the bacterial genomes from the RefSeq database.

A search using AgnA, AgnB1–B2 and AgnC1–C7 sequences as baits found homologs encoded by a BGC in the genome of *Ensifer* sp. ENS09, a representative of *Rhizobiales*. This BGC is analogous to



the *agn* cluster but lacks homologs of *agnE1-agnF* (Fig. 9A). The cluster encodes an export pump that differs from AgnG in the number and composition of predicted transmembrane regions. Strikingly, despite the huge number of various *Rhizobiales* isolated and sequenced to date, the *Ensifer* sp. ENS09 BGC is the first cluster coding for an agrocin 84-like compound identified in the genomes of *Rhizobiales* outside *R. rhizogenes*.

Another interesting finding came from the results of a BLASTP search, which used the sequence of AgnC1 protein as a bait. In addition to trivial hits from *R. rhizogenes* K84 and *Ensifer* sp. ENS09, the top hits with E-values lower than $1e^{-8}$ contained sequences of 19 hypothetical proteins encoded in actinobacterial genomes. Manual analysis of the genomic context of *agnC1* orthologs retrieved by the search revealed genes encoding homologs of other Agn proteins (Fig. 9B). Interestingly a 420-bp ORF preceding *agnC1*, which previously was not annotated as a gene,¹²³ is present in BGCs from *Ensifer* and *Actinobacteria*, suggesting that it may encode a functional partner of AgnC1 (we annotated it as *agnC0*). We propose that identified actinobacterial BGCs encode the biosynthesis of nucleotide-containing natural products structurally close to agrocin 84. The newly identified BGCs share an additional set of genes not found in the *agn* BGC (Fig. 9A, genes with black numbers). The products of these genes may be responsible for additional novel modifications of the nucleotide scaffold. Strikingly, a gene encoding a methionyl-tRNA-synthetase (MetRS) is present in most actinobacterial BGCs (Fig. 9A, red dashed frames). In most of the studied cases, the gene of MetRS associated with the BGC is not the only MetRS encoding gene present in the genome (Fig. 9B). The product of this gene may play the same role as AgnB2, providing immunity to the produced compound. If true, this would imply that unlike agrocin 84, the products of some of the identified BGCs target the housekeeping MetRS.

Conclusions

Albomycin, microcin C-like compounds, and agrocin 84 are aaRSs inhibitors exploiting the “Trojan horse” strategy to enter cells. The transport moieties of these antibiotics mimic three distinct types of compounds actively imported into bacterial cells: a siderophore, peptides, and an opine. The spontaneous resistance to the Trojan horse inhibitors is almost exclusively associated with the loss of function of a dedicated transporter.^{32,66,131} Although the frequency of such resistance mutations is relatively high (within 10^{-4} – 10^{-5} range), it may not undermine the potential of Trojan horse inhibitors to be developed into drugs. For example, the loss of YejABEF or ferrichrome transporters reduces the virulence of bacteria and makes them prone to clearance by the immune system of the host.^{32,68,69} Likewise, the loss of the Ti-plasmid encoding AccABCDE transporter required for the internalization of agrocin 84, leads to a non-phytopathogenic phenotype.¹³¹

Trojan horse inhibitors containing covalently bound siderophore moiety – sideromycins – have inspired researchers to

develop numerous conjugates, each being a combination of a siderophore, a linker, and a known synthetic or natural antibiotic (see Negash *et al.*¹³² or Wencewicz & Miller¹³³ for recent reviews of synthetic sideromycins). In McC-like compounds, the C-terminal residue (isoAsn) enzymatically attached to the nucleoside core through the N–P bond limits the specificity of processed compounds towards AspRS. However, McC analogs containing various terminal amino acids attached to adenosine-5'-sulfamoyl with nonhydrolyzable sulfonamide bond were obtained *via* total chemical synthesis. These compounds retain antibacterial activity and are potent inhibitors of corresponding aaRSs, thus expanding the targeting potential of natural McC variants and validating peptide-nucleotide scaffolds as a platform for the creation of new synthetic antibacterials.^{134,135}

The results of our limited bioinformatics analyses show that the diversity of the three classes of molecules reviewed here is not yet completely tapped. When validated experimentally, these compounds may become viable antibiotics. Most interestingly, the products of putative *agn*-like BGCs revealed by our searches in the genomes of *Actinobacteria* may have different intracellular targets that the known compounds and thus could provide alternative means of biocontrol in agriculture.

Conflicts of interest

Authors declare no conflict of interests.

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References

- 1 L. D. Sherlin and J. J. Perona, *Structure*, 2003, **11**, 591–603.
- 2 M. A. Rubio Gomez and M. Ibba, *RNA*, 2020, **26**, 910–936.
- 3 C. M. Thomas, J. Hothersall, C. L. Willis and T. J. Simpson, *Nat. Rev. Microbiol.*, 2010, **8**, 281–289.
- 4 V. Jain, M. Yogavel, Y. Oshima, H. Kikuchi, B. Touquet, M.-A. Hakimi and A. Sharma, *Structure*, 2015, **23**, 819–829.
- 5 S. Kirillov, L. A. Vitali, B. P. Goldstein, F. Monti, Y. Semenov, V. Makhno, S. Ripa, C. L. Pon and C. O. Gualerzi, *RNA*, 1997, **3**, 905–913.
- 6 J. S. Reader, P. T. Ordoukhanian, J.-G. Kim, V. de Crécy-Lagard, I. Hwang, S. Farrand and P. Schimmel, *Science*, 2005, **309**, 1533.
- 7 A. Ogilvie, K. Wiebauer and W. Kersten, *Biochem. J.*, 1975, **152**, 511–515.
- 8 M. J. Brown, P. S. Carter, A. S. Fenwick, A. P. Fosberry, D. W. Hamprecht, M. J. Hibbs, R. L. Jarvest, L. Mensah,



- P. H. Milner, P. J. O'Hanlon, A. J. Pope, C. M. Richardson, A. West and D. R. Witty, *Bioorg. Med. Chem. Lett.*, 2002, **12**, 3171–3174.
- 9 C. J. Schulze, W. M. Bray, F. Loganzo, M.-H. Lam, T. Szal, A. Villalobos, F. E. Koehn and R. G. Linington, *J. Nat. Prod.*, 2014, **77**, 2570–2574.
- 10 G. Dirheimer and E. E. Creppy, *IARC Sci. Publ.*, 1991, 171–186.
- 11 A. Metlitskaya, T. Kazakov, A. Kommer, O. Pavlova, M. Praetorius-Ibba, M. Ibba, I. Krashennikov, V. Kolb, I. Khmel and K. Severinov, *J. Biol. Chem.*, 2006, **281**, 18033–18042.
- 12 T. A. Scott, S. F. D. Batey, P. Wiencek, G. Chandra, S. Alt, C. S. Francklyn and B. Wilkinson, *ACS Chem. Biol.*, 2019, **14**, 2663–2671.
- 13 M. Uramoto, C. J. Kim, K. Shin-Ya, H. Kusakabe, K. Isono, D. R. Phillips and J. A. McCloskey, *J. Antibiot.*, 1991, **44**, 375–381.
- 14 J. O. Capobianco, D. Zakula, M. L. Coen and R. C. Goldman, *Biochem. Biophys. Res. Commun.*, 1993, **190**, 1037–1044.
- 15 A. L. Stefanska, M. Fulston, C. S. Houge-Frydrych, J. J. Jones and S. R. Warr, *J. Antibiot.*, 2000, **53**, 1346–1353.
- 16 S. Robles, Y. Hu, T. Resto, F. Dean and J. M. Bullard, *Curr. Drug Discovery Technol.*, 2017, **14**, 156–168.
- 17 A. L. Stefanska, N. J. Coates, L. M. Mensah, A. J. Pope, S. J. Ready and S. R. Warr, *J. Antibiot.*, 2000, **53**, 345–350.
- 18 P. Fang, H. Han, J. Wang, K. Chen, X. Chen and M. Guo, *Chem. Biol.*, 2015, **22**, 734–744.
- 19 T. L. Williams, Y. W. Yin and C. W. Carter, *J. Biol. Chem.*, 2016, **291**, 255–265.
- 20 T. Nakama, O. Nureki and S. Yokoyama, *J. Biol. Chem.*, 2001, **276**, 47387–47393.
- 21 H. Zhou, L. Sun, X.-L. Yang and P. Schimmel, *Nature*, 2013, **494**, 121–124.
- 22 R. V. K. Cochrane, A. K. Norquay and J. C. Vederas, *Med-ChemComm*, 2016, **7**, 1535–1545.
- 23 S. Chopra and J. Reader, *Int. J. Mol. Sci.*, 2014, **16**, 321–349.
- 24 J. M. Ho, E. Bakkalbasi, D. Söll and C. A. Miller, *RNA Biol.*, 2018, **15**, 667–677.
- 25 H. I. Zgurskaya, C. A. López and S. Gnanakaran, *ACS Infect. Dis.*, 2015, **1**, 512–522.
- 26 J. O. Capobianco, C. C. Doran and R. C. Goldman, *Antimicrob. Agents Chemother.*, 1989, **33**, 156–163.
- 27 R. G. Werner, *Antimicrob. Agents Chemother.*, 1980, **18**, 858–862.
- 28 D. M. Reynolds, A. Schatz and S. A. Waksman, *Proc. Soc. Exp. Biol. Med.*, 1947, **64**, 50–54.
- 29 G. F. Gause and M. G. Brazhnikova, *Nov. Med.*, 1951, 3–7.
- 30 E. O. Stapley and R. E. Ormond, *Science*, 1957, **125**, 587–589.
- 31 H. Maehr, *Pure Appl. Chem.*, 1971, **28**, 603–636.
- 32 A. Pramanik, U. H. Stroehrer, J. Krejci, A. J. Standish, E. Bohn, J. C. Paton, I. B. Autenrieth and V. Braun, *Int. J. Med. Microbiol.*, 2007, **297**, 459–469.
- 33 D. M. Reynolds and S. A. Waksman, *J. Bacteriol.*, 1948, **55**, 739–752.
- 34 G. F. Gause, *BMJ*, 1955, **2**, 1177–1179.
- 35 Z. Lin, X. Xu, S. Zhao, X. Yang, J. Guo, Q. Zhang, C. Jing, S. Chen and Y. He, *Nat. Commun.*, 2018, **9**, 3445.
- 36 A. Pramanik and V. Braun, *J. Bacteriol.*, 2006, **188**, 3878–3886.
- 37 G. Benz, T. Schröder, J. Kurz, C. Wünsche, W. Karl, G. Steffens, J. Pfitzner and D. Schmidt, *Angew. Chem., Int. Ed. Engl.*, 1982, **21**, 527–528.
- 38 Y. Zeng, A. Kulkarni, Z. Yang, P. B. Patil, W. Zhou, X. Chi, S. Van Lanen and S. Chen, *ACS Chem. Biol.*, 2012, **7**, 1565–1575.
- 39 A. Hartmann, H. P. Fiedler and V. Braun, *Eur. J. Biochem.*, 1979, **99**, 517–524.
- 40 A. D. Ferguson, J. W. Coulton, K. Diederichs, W. Welte, V. Braun and H.-P. Fiedler, *Protein Sci.*, 2000, **9**, 956–963.
- 41 R. Chakraborty, E. Storey and D. van der Helm, *Biomaterials*, 2007, **20**, 263–274.
- 42 M. R. Rohrbach, V. Braun and W. Köster, *J. Bacteriol.*, 1995, **177**, 7186–7193.
- 43 T. E. Clarke, V. Braun, G. Winkelmann, L. W. Tari and H. J. Vogel, *J. Biol. Chem.*, 2002, **277**, 13966–13972.
- 44 V. Braun, K. Günthner, K. Hantke and L. Zimmermann, *J. Bacteriol.*, 1983, **156**, 308–315.
- 45 L. Pang, M. Nautiyal, S. De Graef, B. Gadakh, V. Zorzini, A. Economou, S. V. Strelkov, A. Van Aerschot and S. D. Weeks, *ACS Chem. Biol.*, 2020, **15**, 407–415.
- 46 A. Saha, S. Dutta and N. Nandi, *J. Biomol. Struct. Dyn.*, 2020, **38**, 2440–2454.
- 47 R. Ushimaru, Z. Chen, H. Zhao, P. Fan and H. Liu, *Angew. Chem., Int. Ed.*, 2020, **59**, 3558–3562.
- 48 R. Ushimaru and H. Liu, *J. Am. Chem. Soc.*, 2019, **141**, 2211–2214.
- 49 Y. Zeng, H. H. Roy, P. B. Patil, M. Ibba and S. Chen, *Antimicrob. Agents Chemother.*, 2009, **53**, 4619–4627.
- 50 S. F. Altschul, W. Gish, W. Miller, E. W. Myers and D. J. Lipman, *J. Mol. Biol.*, 1990, **215**, 403–410.
- 51 R. Ushimaru, Z. Chen, H. Zhao, P. Fan and H. Liu, *Angew. Chem., Int. Ed.*, 2020, **59**, 3558–3562.
- 52 F. Baquero, D. Bouanchaud, M. C. Martinez-Perez and C. Fernandez, *J. Bacteriol.*, 1978, **135**, 342–347.
- 53 Y. Chabbert, *Ann. Inst. Pasteur.*, 1950, **79**, 51–59.
- 54 N. E. Kurepina, E. I. Basyuk, A. Z. Metlitskaya, D. A. Zaitsev and I. A. Khmel, *Mol. Gen. Genet.*, 1993, **241**, 700–706.
- 55 D. Smajs, M. Strouhal, P. Matejková, D. Cejtková, L. Cursino, E. Chartone-Souza, J. Smarda and A. M. A. Nascimento, *Plasmid*, 2008, **59**, 1–10.
- 56 D. M. Gordon and C. L. O'Brien, *Microbiology*, 2006, **152**, 3239–3244.
- 57 L. Mícenková, J. Bosák, M. Vrba, A. Ševčíková and D. Šmajs, *BMC Microbiol.*, 2016, **16**, 218.
- 58 J. F. Garcia-Bustos, N. Pezzi and E. Mendez, *Antimicrob. Agents Chemother.*, 1985, **27**, 791–797.
- 59 L. Cursino, D. Smajs, J. Smarda, R. M. D. Nardi, J. R. Nicoli, E. Chartone-Souza and A. M. A. Nascimento, *J. Appl. Microbiol.*, 2006, **100**, 821–829.
- 60 D. Hofreuter and R. Haas, *J. Bacteriol.*, 2002, **184**, 2755–2766.



- 61 I. Zukher, M. Pavlov, D. Tsibulskaya, A. Kulikovskiy, T. Zyubko, D. Bikmetov, M. Serebryakova, S. K. Nair, M. Ehrenberg, S. Dubiley and K. Severinov, *mBio*, 2019, **10**, 11891–11902.
- 62 J. I. Guijarro, J. E. González-Pastor, F. Baleux, J. L. San Millán, M. A. Castilla, M. Rico, F. Moreno and M. Delepierre, *J. Biol. Chem.*, 1995, **270**, 23520–23532.
- 63 O. Bantysh, M. Serebryakova, K. S. Makarova, S. Dubiley, K. A. Datsenko and K. Severinov, *mBio*, 2014, **5**, e01059.
- 64 M. Serebryakova, D. Tsibulskaya, O. Mokina, A. Kulikovskiy, M. Nautiyal, A. Van Aerschot, K. Severinov and S. Dubiley, *J. Am. Chem. Soc.*, 2016, **138**, 15690–15698.
- 65 J. E. González-Pastor, J. L. San Millán, M. A. Castilla, F. Moreno, J. E. Gonzalez-Pastor, J. L. San Millán, M. A. Castilla, F. Moreno, J. E. González-Pastor, J. L. San Millán, M. A. Castilla and F. Moreno, *J. Bacteriol.*, 1995, **177**, 7131–7140.
- 66 M. Novikova, A. Metlitskaya, K. Datsenko, T. Kazakov, A. Kazakov, B. Wanner and K. Severinov, *J. Bacteriol.*, 2007, **189**, 8361–8365.
- 67 U. Qimron, N. Madar, H.-W. Mittrücker, A. Zilka, I. Yosef, N. Bloushtain, S. H. E. Kaufmann, I. Rosenshine, R. N. Apte and A. Porgador, *Cell. Microbiol.*, 2004, **6**, 1057–1070.
- 68 S. M. Eswarappa, K. K. Panguluri, M. Hensel and D. Chakravorty, *Microbiology*, 2008, **154**, 666–678.
- 69 Z. Wang, P. Bie, J. Cheng, L. Lu, B. Cui and Q. Wu, *Sci. Rep.*, 2016, **6**, 31876.
- 70 D. Pletzer, Y. Braun, S. Dubiley, C. Lafon, T. Köhler, M. G. P. Page, M. Mourez, K. Severinov and H. Weingart, *J. Bacteriol.*, 2015, **197**, 2217–2228.
- 71 G. H. M. Vondenhoff, B. Blanchaert, S. Geboers, T. Kazakov, K. A. Datsenko, B. L. Wanner, J. Rozenski, K. Severinov and A. Van Aerschot, *J. Bacteriol.*, 2011, **193**, 3618–3623.
- 72 O. Bantysh, M. Serebryakova, I. Zukher, A. Kulikovskiy, D. Tsibulskaya, S. Dubiley and K. Severinov, *J. Bacteriol.*, 2015, **197**, 3133–3141.
- 73 R. A. Salomón and R. N. Fariás, *J. Bacteriol.*, 1995, **177**, 3323–3325.
- 74 M. Laviña, A. P. Pugsley and F. Moreno, *J. Gen. Microbiol.*, 1986, **132**, 1685–1693.
- 75 D. Tsibulskaya, O. Mokina, A. Kulikovskiy, J. Piskunova, K. Severinov, M. Serebryakova and S. Dubiley, *J. Am. Chem. Soc.*, 2017, **139**, 16178–16187.
- 76 T. Kazakov, G. H. Vondenhoff, K. A. Datsenko, M. Novikova, A. Metlitskaya, B. L. Wanner and K. Severinov, *J. Bacteriol.*, 2008, **190**, 2607–2610.
- 77 J. Piskunova, E. Maisonneuve, E. Germain, K. Gerdes and K. Severinov, *Mol. Microbiol.*, 2017, **104**, 463–471.
- 78 P. G. Arnison, M. J. Bibb, G. Bierbaum, A. A. Bowers, T. S. Bugni, G. Bulaj, J. A. Camarero, D. J. Campopiano, G. L. Challis, J. Clardy, P. D. Cotter, D. J. Craik, M. Dawson, E. Dittmann, S. Donadio, P. C. Dorrestein, K.-D. Entian, M. A. Fischbach, J. S. Garavelli, U. Göransson, C. W. Gruber, D. H. Haft, T. K. Hemscheidt, C. Hertweck, C. Hill, A. R. Horswill, M. Jaspars, W. L. Kelly, J. P. Klinman, O. P. Kuipers, A. J. Link, W. Liu, M. A. Marahiel, D. A. Mitchell, G. N. Moll, B. S. Moore, R. Müller, S. K. Nair, I. F. Nes, G. E. Norris, B. M. Olivera, H. Onaka, M. L. Patchett, J. Piel, M. J. T. Reaney, S. Rebuffat, R. P. Ross, H.-G. Sahl, E. W. Schmidt, M. E. Selsted, K. Severinov, B. Shen, K. Sivonen, L. Smith, T. Stein, R. D. Süßmuth, J. R. Tagg, G.-L. Tang, A. W. Truman, J. C. Vederas, C. T. Walsh, J. D. Walton, S. C. Wenzel, J. M. Willey and W. A. van der Donk, *Nat. Prod. Rep.*, 2013, **30**, 108–160.
- 79 T. J. Oman and W. A. van der Donk, *Nat. Chem. Biol.*, 2010, **6**, 9–18.
- 80 J. E. González-Pastor, J. L. San Millán and F. Moreno, *Nature*, 1994, **369**, 281.
- 81 R. F. Roush, E. M. Nolan, F. Löhr and C. T. Walsh, *J. Am. Chem. Soc.*, 2008, **130**, 3603–3609.
- 82 C. A. Regni, R. F. Roush, D. J. Miller, A. Nourse, C. T. Walsh and B. A. Schulman, *EMBO J.*, 2009, **28**, 1953–1964.
- 83 B. J. Burkhardt, G. A. Hudson, K. L. Dunbar and D. A. Mitchell, *Nat. Chem. Biol.*, 2015, **11**, 564–570.
- 84 S.-H. Dong, A. Kulikovskiy, I. Zukher, P. Estrada, S. Dubiley, K. Severinov and S. K. Nair, *Chem. Sci.*, 2019, **10**, 2391–2395.
- 85 M. A. Funk and W. A. van der Donk, *Acc. Chem. Res.*, 2017, **50**, 1577–1586.
- 86 E. Yagmurov, D. Tsibulskaya, A. Livenskiy, M. Serebryakova, Y. I. Wolf, S. Borukhov, K. Severinov and S. Dubiley, *mBio*, 2020, **11**, e00497-20.
- 87 A. Kulikovskiy, M. Serebryakova, O. Bantysh, A. Metlitskaya, S. Borukhov, K. Severinov and S. Dubiley, *J. Am. Chem. Soc.*, 2014, **136**, 11168–11175.
- 88 N. Parveen and K. A. Cornell, *Mol. Microbiol.*, 2011, **79**, 7–20.
- 89 J. F. García-Bustos, N. Pezzi and C. Asensio, *Biochem. Biophys. Res. Commun.*, 1984, **119**, 779–785.
- 90 L. Díaz-Guerra, F. Moreno and J. L. San Millán, *J. Bacteriol.*, 1989, **171**, 2906–2908.
- 91 F. Moreno, J. E. González-Pastor, M. R. Baquero and D. Bravo, *Biochimie*, 2002, **84**, 521–529.
- 92 D. Fomenko, A. Veselovskii and I. Khmel, *Res. Microbiol.*, 2001, **152**, 469–479.
- 93 V. Agarwal, A. Metlitskaya, K. Severinov and S. K. Nair, *J. Biol. Chem.*, 2011, **286**, 21295–21303.
- 94 M. Novikova, T. Kazakov, G. H. Vondenhoff, E. Semenova, J. Rozenski, A. Metlitskaya, I. Zukher, A. Tikhonov, A. Van Aerschot and K. Severinov, *J. Biol. Chem.*, 2010, **285**, 12662–12669.
- 95 A. Tikhonov, T. Kazakov, E. Semenova, M. Serebryakova, G. Vondenhoff, A. Van Aerschot, J. S. Reader, V. M. Govorun and K. Severinov, *J. Biol. Chem.*, 2010, **285**, 37944–37952.
- 96 B. Nocek, A. Tikhonov, G. Babnigg, M. Gu, M. Zhou, K. S. Makarova, G. Vondenhoff, A. Van Aerschot, K. Kwon, W. F. Anderson, K. Severinov and A. Joachimiak, *J. Mol. Biol.*, 2012, **420**, 366–383.



- 97 M. F. Templin, A. Ursinus and J. V. Höltje, *EMBO J.*, 1999, **18**, 4108–4117.
- 98 C. Brenner, *eLS*, John Wiley & Sons, Ltd, Chichester, UK, 2014, pp. 1–8.
- 99 D. Tsibulskaya, O. Mokina, A. Kulikovskiy, J. Piskunova, K. Severinov, M. Serebryakova and S. Dubiley, *J. Am. Chem. Soc.*, 2017, **139**, 16178–16187.
- 100 J. Pulawska, *J. Plant Pathol.*, 2010, **92**, 87–98.
- 101 E. Velázquez, J. L. Palomo, R. Rivas, H. Guerra, A. Peix, M. E. Trujillo, P. García-Benavides, P. F. Mateos, H. Wabiko and E. Martínez-Molina, *Syst. Appl. Microbiol.*, 2010, **33**, 247–251.
- 102 P. B. New and A. Kerr, *J. Appl. Bacteriol.*, 1972, **35**, 279–287.
- 103 A. Kerr, *J. Appl. Bacteriol.*, 1972, **35**, 493–497.
- 104 K. Htay and A. Kerr, *J. Appl. Bacteriol.*, 1974, **37**, 525–530.
- 105 A. Kerr and G. Bullard, *Agronomy*, 2020, **10**, 1126.
- 106 A. Kerr and K. Htay, *Physiol. Plant Pathol.*, 1974, **4**, 37–44.
- 107 W. P. Roberts, M. E. Tate and A. Kerr, *Nature*, 1977, **265**, 379–381.
- 108 M. E. Tate, P. J. Murphy, W. P. Roberts and A. Kerr, *Nature*, 1979, **280**, 697–699.
- 109 A. El Sahili, S.-Z. Li, J. Lang, C. Virus, S. Planamente, M. Ahmar, B. G. Guimaraes, M. Aumont-Nicaise, A. Vigouroux, L. Soulère, J. Reader, Y. Queneau, D. Faure and S. Moréra, *PLoS Pathog.*, 2015, **11**, e1005071.
- 110 P. J. Murphy, M. E. Tate and A. Kerr, *Eur. J. Biochem.*, 1981, **115**, 539–543.
- 111 H.-H. Hwang, M. Yu and E.-M. Lai, *Arab. B.*, 2017, **15**, e0186.
- 112 E. W. Nester, *Front. Plant Sci.*, 2015, **5**, 1–16.
- 113 I. A. Vladimirov, T. V. Matveeva and L. A. Lutova, *Russ. J. Genet.*, 2015, **51**, 121–129.
- 114 H. Kim and S. K. Farrand, *J. Bacteriol.*, 1997, **179**, 7559–7572.
- 115 J. G. Ellis and P. J. Murphy, *MGG Mol. Gen. Genet.*, 1981, **181**, 36–43.
- 116 M. H. Ryder, M. E. Tate and G. P. Jones, *J. Biol. Chem.*, 1984, **259**, 9704–9710.
- 117 W. P. Roberts and A. Kerr, *Physiol. Plant Pathol.*, 1974, **4**, 81–91.
- 118 S. Chopra, A. Palencia, C. Virus, A. Tripathy, B. R. Temple, A. Velazquez-Campoy, S. Cusack and J. S. Reader, *Nat. Commun.*, 2013, **4**, 1417.
- 119 J. G. Ellis, A. Kerr, M. Van Montagu and J. Schell, *Physiol. Plant Pathol.*, 1979, **15**, 311–319.
- 120 J. E. Slota and S. K. Farrand, *Plasmid*, 1982, **8**, 175–186.
- 121 D. A. Jones, M. H. Ryder, B. G. Clare, S. K. Farrand and A. Kerr, *Mol. Gen. Genet. MGG*, 1988, **212**, 207–214.
- 122 B. Vicedo, R. Peñalver, M. J. Asins and M. M. López, *Appl. Environ. Microbiol.*, 1993, **59**, 309–315.
- 123 J.-G. Kim, B. K. Park, S.-U. Kim, D. Choi, B. H. Nahm, J. S. Moon, J. S. Reader, S. K. Farrand and I. Hwang, *Proc. Natl. Acad. Sci. U. S. A.*, 2006, **103**, 8846–8851.
- 124 L. A. Kelley, S. Mezulis, C. M. Yates, M. N. Wass and M. J. E. Sternberg, *Nat. Protoc.*, 2015, **10**, 845–858.
- 125 L. Zimmermann, A. Stephens, S.-Z. Nam, D. Rau, J. Kübler, M. Lozajic, F. Gabler, J. Söding, A. N. Lupas and V. Alva, *J. Mol. Biol.*, 2018, **430**, 2237–2243.
- 126 R. Penyalver, P. M. Oger, S. Su, B. Alvarez, C. I. Salcedo, M. M. López and S. K. Farrand, *Mol. Plant. Microbe Interact.*, 2009, **22**, 713–724.
- 127 S. Chopra, A. Palencia, C. Virus, S. Schulwitz, B. R. Temple, S. Cusack and J. Reader, *Nat. Commun.*, 2016, **7**, 12928.
- 128 S. C. Donner, D. A. Jones, N. C. McClure, G. M. Rosewarne, M. E. Tate, A. Kerr, N. N. Fajardo and B. G. Clare, *Physiol. Mol. Plant Pathol.*, 1993, **42**, 185–194.
- 129 S. Donner, PhD thesis, The University of Adelaide, *Australia*, 1997.
- 130 C. M. Elvin, R. E. Asenstorfer, M. H. Ryder, S. C. Donner, G. P. Jones and M. E. Tate, *J. Antibiot.*, 2018, **71**, 438–446.
- 131 D. A. Cooksey and L. W. Moore, *Physiol. Plant Pathol.*, 1982, **20**, 129–135.
- 132 K. H. Negash, J. K. S. Norris and J. T. Hodgkinson, *Molecules*, 2019, **24**, 3314.
- 133 T. A. Wenciewicz and M. J. Miller, Sideromycins as Pathogen-Targeted Antibiotics, in *Antibacterials*, ed. J. Fisher, S. Mobashery and M. Miller, Topics in Medicinal Chemistry, Springer, Cham, 2017, vol. 26, DOI: 10.1007/7355_2017_19.
- 134 P. Van de Vijver, G. H. M. Vondenhoff, T. S. Kazakov, E. Semenova, K. Kuznedelov, A. Metlitskaya, A. Van, Aerschot and K. Severinov, *J. Bacteriol.*, 2009, **191**, 6273–6280.
- 135 G. H. M. Vondenhoff, S. Dubiley, K. Severinov, E. Lescrinier, J. Rozenski and A. Van Aerschot, *Bioorg. Med. Chem.*, 2011, **19**, 5462–5467.
- 136 A. Palencia, T. Crépin, M. T. Vu, T. L. Lincecum, S. A. Martinis and S. Cusack, *Nat. Struct. Mol. Biol.*, 2012, **19**, 677–684.
- 137 R. Sankaranarayanan, A. C. Dock-Bregeon, P. Romby, J. Caillet, M. Springer, B. Rees, C. Ehresmann, B. Ehresmann and D. Moras, *Cell*, 1999, **97**, 371–381.
- 138 V. Agarwal, A. Tikhonov, A. Metlitskaya, K. Severinov and S. K. Nair, *Proc. Natl. Acad. Sci. U. S. A.*, 2012, **109**, 4425–4430.
- 139 S. Guindon, J.-F. Dufayard, V. Lefort, M. Anisimova, W. Hordijk and O. Gascuel, *Syst. Biol.*, 2010, **59**, 307–321.

