


Cite this: *RSC Adv.*, 2020, 10, 37168

Network pharmacology-based study of the mechanisms of action of anti-diabetic triterpenoids from *Cyclocarya paliurus*†

Zixin Lin,^{ab} Yingpeng Tong,^{bc} Na Li,^b Ziping Zhu^b and Junmin Li^{id} *^{bc}

Diabetes is a complex illness requiring long-term therapy. *Cyclocarya paliurus*, a recently confirmed new food resource, shows significant hypoglycemic and hypolipidemic effects in type II diabetes. Triterpenoid saponins are considered as the effective medicinal components of *C. paliurus* and are useful for the treatment of diabetes mellitus. However, little is known regarding their specific mechanism of actions. In this study, we used active ingredient screening and target prediction techniques to determine the components of *C. paliurus* responsible for its anti-diabetic effects as well as their targets. In addition, we used bioinformatics technology and molecular docking analysis to determine the mechanisms underlying their anti-diabetic effects. A total of 39 triterpenes were identified through a literature search and 1 triterpene compound by experiments. In all, 33 potential target proteins associated with 36 pathways were predicted to be related to diabetes. Finally, 7 compounds, 15 target proteins, and 15 signaling pathways were found to play important roles in the therapeutic effects of *C. paliurus* against diabetes. These results provide a theoretical framework for the use of *C. paliurus* against diabetes. Moreover, molecular docking verification showed that more than 90% of the active ingredients had binding activity when tested against key target proteins, and a literature search showed that the active ingredients identified had anti-diabetic effects, indicating that the results were highly reliable.

Received 8th August 2020
Accepted 24th September 2020

DOI: 10.1039/d0ra06846b

rsc.li/rsc-advances

1. Introduction

Diabetes is a disorder of glucose metabolism that leads to high blood sugar levels and to a series of complications which reduce the quality of life of patients, increasing their mortality.¹ Based on its pathogenesis, it can be divided into type 1 diabetes (T1DM) and type 2 diabetes (T2DM), with T2DM accounting for more than 90% of cases.² At present, diabetes is treated with drugs, such as sulfonylureas, DPP-4 inhibitors, oral antidiabetic agents, sodium-glucose co-transport inhibitors, and glucokinase activators,^{3–7} but many drugs have side effects.⁸ Therefore, it is urgent to find new, safe and effective early intervention medicinal compounds for diabetes.⁹

Traditional botanical and herbal medicines have been widely used for the treatment of diabetes,^{10,11} e.g. *Lobelia chinensis* Lour.,⁷ *Hydrangea macrophylla* var. *thunbergii*,¹² and *Cyclocarya paliurus* (Batal.) Iljinskaja.¹³ *C. paliurus* is a medicinal plant native to southern China. Its leaves have been traditionally used in the form of herbal tea for their beneficial effects against

diabetes. Various bioactive compounds have been extracted from the leaves of *C. paliurus*, including flavonoids, triterpenoids, polysaccharides, and organic acids, which may contribute to its antihyperglycemic, antihyperlipidemic, and antihypertensive effects.^{13–24} Although the mechanisms underlying the anti-diabetic effects of the ethanol and aqueous extracts of *C. paliurus* leaves have been explored,¹⁸ the specific bioactive constituents of *C. paliurus* and their targets are still unknown. Due to their safety and efficacy, plant-derived triterpenoids have attracted attention for the treatment of diabetes.^{25–27} For example, new cucurbitane-type triterpenoids have shown potential for the prevention and management of diabetes by improving insulin sensitivity and glucose homeostasis.²⁸ Triterpenoids, such as cyclocaric acid B and cyclocarioside H, extracted from *C. paliurus* leaves, promote glucose uptake in the absence of insulin, and ameliorate inflammation by inhibiting the insulin receptor substrate 1 (IRS-1)/phosphoinositide 3-kinase (PI3K)/protein kinase B (Akt) pathway.²⁹ However, no *C. paliurus* triterpenoids with anti-diabetic effects or their specific targets have been described.

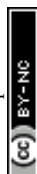
Network pharmacology is a new systematic method to study target-drug interactions and their influence on disease. Combined with pharmacology and pharmacodynamics, network pharmacology has been successfully applied to explore the mechanisms of action of traditional botanical or herbal medicines at the molecular level,³⁰ to dissect the relationship

^aSchool of Life Science, Shanghai Normal University, Shanghai 200234, China

^bZhejiang Provincial Key Laboratory of Evolutionary Ecology and Conservation, Taizhou University, Taizhou 318000, China. E-mail: lijmtzc@126.com

^cSchool of Advanced Study, Taizhou University, Taizhou 318000, China

† Electronic supplementary information (ESI) available. See DOI: 10.1039/d0ra06846b



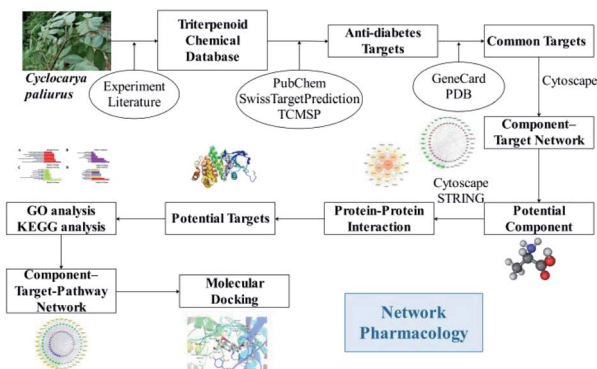


Fig. 1 The flowchart of the network pharmacological analysis approach.

between medicines and diseases, and to identify signaling pathways. This approach makes it feasible to explore the therapeutic mechanisms of multi-component and multi-target traditional botanical or herbal medicines.¹³ Investigated the beneficial effects of *C. paliurus* against diabetic dyslipidemia and its mechanisms of action using lipidomics, serum pharmacology, and network pharmacology approaches.¹³ Based on network pharmacology analysis, they identified the predicted targets of *C. paliurus*'s active ingredients (ALOX12, APP, BCL2, CYP2C9, PTPN1) and their linked lipidome targets [PLD2, PLA2G(s), and PI3K(s) families], which could be responsible for the effects of *C. paliurus* leaf extracts against diabetic dyslipidemia.¹³ In this study, we used network pharmacology to explore the effects of *C. paliurus* triterpenoids on diabetes and elucidate their mechanisms of action. The detailed procedures are shown in Fig. 1.

2. Materials and methods

2.1 Plant species

C. paliurus is commonly called "tian sha Chun" in China, meaning "sweat tree", due to the flavor of its leaves. This tree can reach 30 m and grows in moist forests on mountains at altitudes between 400 and 2500 m. It is distributed in many regions of China with temperatures between 7.8 °C and 19.9 °C and with precipitation between 825.9 mm and 2394.5 mm,^{31,32} including Anhui, Fujian, Guangdong, Guangxi, Guizhou, Hainan, Hubei, Hunan, Jiangsu, Jiangxi, Sichuan, Taiwan, Yunnan, and Zhejiang Provinces. The leaves of this plant are used in traditional Chinese medicine (TCM) for treating disorders like hypertension, high cholesterol, and diabetes, as well as for improving the function of the immune system.^{15,33} The first health tea from China approved by the U. S. Food and Drug Administration (FDA) was made with the leaves of *C. paliurus*.³¹

2.2 Chemical ingredients database

Leaves of *C. paliurus* obtained from Taizhou University in October 2018 were air-dried (total weight, 1.6 kg). Dried leaves of *C. paliurus* (1.6 kg) were pulverized and extracted three times with 90% ethanol (8.0 L) at room temperature. After filtration, the solvents were removed under vacuum to obtain a crude

extract (1.1 kg), which was resuspended in H₂O (1.0 L) and then partitioned successively with petroleum ether (1.0 L), ethyl acetate (EtOAc, 1.0 L) and butyl alcohol (1.0 L) three times. After removal of solvent, the entire EtOAc extract was fractionated by silica gel column chromatography using a gradient of petroleum ether (PE)/EtOAc (20 : 1 to 0 : 1, v/v) and then methanol to obtain a pure component.

With the exception of the previous triterpenoids, other triterpenoids compounds from *C. paliurus* were identified by reviewing previously published studies (ESI Table 1†) and the Traditional Chinese Medicine Systems Pharmacology database (TCMSP, <http://lsp.nwu.edu.cn/tcmsp.php>).³⁴ The specific structures were obtained from Pubchem databases (<https://pubchem.ncbi.nlm.nih.gov/>). If the specific structures could not be found in the databases, the original research articles were reviewed. In addition, two natural products were isolated from the *C. paliurus* extract. Then, all triterpenes were subjected to network pharmacologic prediction. ChemDraw 14.0 software (<https://www.chemdraw.com.cn/xiazai.html>) was used to draw the structures, and the files were saved in mol and mol2 formats.

2.3 Target protein database

Proteins involved in diabetes were selected from the literature and from the following public databases: PubChem (<https://pubchem.ncbi.nlm.nih.gov/>), which provides information about chemical substances and their biological activities as a public repository; Genecards database (<https://www.genecards.org/>), which is a unique bioinformatics and chemical informatics resource; Potential Drug Target Database (<http://dddc.ac.cn/pdtd/index.php>), which is a protein database for drug target recognition; and RCSB Database (<http://www.rcsb.org/pdb/home/home.do>), which provides information about therapeutic proteins, nucleic acid targets, targeted diseases, pathways, and the corresponding drugs for each of these targets. Based on the determination of the protein-ligand complex and crystal structures, protein targets were obtained. The crystal structures of the target protein complexes with the original ligands were downloaded from the Protein Data Bank (PDB) database, which is a free database containing structural data of biological macromolecules. These proteins were used as receptors in molecular docking experiments. The structures of the compounds were imported into the Prediction Target of Swiss Target Prediction network database as a *.mol format³⁵ (<http://www.swisstargetprediction.ch/>), setting the species to "*Homo sapiens*" to predict active targets. This platform automatically conducts a docking simulation between the compound and the background target database, selecting 15 potential active targets with the highest affinity. Predicted targets showing high probability were selected and duplicates were deleted. Diabetes mellitus was searched in the GeneCards online database (<http://www.genecards.org/>) to identify genes related to diabetes. Common targets between the database and *C. paliurus* were identified by using excel. The results were imported into Cytoscape 3.6.1 software and the component-target interaction network was constructed and analyzed.



This journal is © The Royal Society of Chemistry 2020

isolated and identified in our study and 38 other reported compounds. These were used to create a library of compounds for the network analysis (Table 1). The ADME data for the compounds is shown in ESI Table 2.† We used SwissTargetPrediction to predict the targets of the active compounds, and a total of 585 targets proteins were identified. A total of 33 potential target proteins of *C. paliurus* anti-diabetes compounds were identified, as shown in Table 2.

3.2 Analysis of interactions between active components and target proteins

Cytoscape was used to visualize the component-target interactions. The component-target network consists of a total of 73 nodes and 213 edges; 33 of these nodes represent target proteins and 39 nodes active ingredients directly related to the

anti-diabetic effects of *C. paliurus* (Fig. 2). The network shows that the anti-diabetic effects of *C. paliurus* may be the result of multi-component and multi-target protein effects. The top 7 components based on degree values were arjunglucoside II (M26), α -boswellic acid (M28), 3 β ,23-trihydroxyurs-11-oxo-12-ene-28-oic acid (M24), 3 β ,23,27-trihydroxy-1-oxo-olean-12-ene-28-oic acid (M24), dihydroxy-1,12-dioxo-olean-28-oic acid (M22), 23-3 β ,23,27-trihydroxy-1-oxo-olean-12-ene-28-oic acid (M23), 2 α ,3 β ,23-trihydroxyoleana-11,13(18)-dien-28-oic acid (M33) and cyclocarioside K (M36). Of these, four compounds, M23, M24, M25 and M34, complied with the "Lipinski's Rule of Five". These components are the key active ingredients underlying the antipyretic effects of *C. paliurus* and their structural formulas are shown in ESI Fig. 2.† The component-targets network revealed that several target proteins could interact

Table 1 The information of 39 triterpenoid compounds in *Cyclocarya paliurus*^a

MOL ID	Compound	Degree	Betweenness centrality	Molecular formula	CAS
M1	2 α ,3 α ,23-Trihydroxyurs-12-en-28-oic acid	5	0.0017618	C ₃₀ H ₄₈ O ₅	103974-74-9
M2	3 β -Hydroxy-urs-11-en-28,13-lactone	5	0.0017618	C ₃₀ H ₄₆ O ₃	35959-05-8
M3	2 α -Hydroxyursolic acid	5	0.0017618	C ₃₀ H ₄₈ O ₄	72881-13-1
M4	2 α ,3 α ,23-Trihydroxyurs-12,20(30)-dien-28-oic acid	5	0.0017618	C ₃₀ H ₄₆ O ₅	143839-01-4
M5	2 α ,3 β ,23-Trihydroxy-12-ene-28-ursolic acid	5	0.0017618	C ₃₀ H ₄₈ O ₅	464-92-6
M6	3 β -O-trans-Caffeoyl-morolic acid	4	0.00594168	C ₃₉ H ₅₄ O ₆	97534-10-6
M7	Actinidic acid	4	0.00111799	C ₃₀ H ₄₆ O ₅	341971-45-7
M8	Arjunolic acid	4	0.00111799	C ₃₀ H ₄₈ O ₅	465-00-9
M9	Corosolic acid	5	0.00508509	C ₃₀ H ₄₈ O ₄	4547-24-4
M10	Cyclocaric acid B	5	0.0017618	C ₃₀ H ₄₆ O ₅	182315-46-4
M11	Daucosterol	5	0.00979396	C ₃₅ H ₆₀ O ₆	474-58-8
M12	Hederagenin	5	0.0017618	C ₃₀ H ₄₈ O ₄	465-99-6
M13	Maslinic acid	5	0.0017618	C ₃₀ H ₄₈ O ₄	4373-41-5
M14	Olean-12-en-28-oic acid	5	0.0017618	C ₃₀ H ₄₈ O ₂	17990-43-1
M15	Oleanolic acid	5	0.0017618	C ₃₀ H ₄₈ O ₃	508-02-1
M16	Taraxerol	5	0.00598173	C ₃₀ H ₅₀ O ₀	127-22-0
M17	β -Amyrin	5	0.00590412	C ₃₀ H ₅₀ O	559-70-6
M18	β -Amyrone	4	0.00397423	C ₃₀ H ₄₈ O	638-97-1
M19	β -Sitosterol	5	0.00270752	C ₂₉ H ₅₀ O	83-46-5
M20	Cyclocariosides I	6	0.01121781	C ₃₅ H ₅₆ O ₈	1644624-82-7
M21	Cyclocarioside N	2	0.00019562	C ₄₄ H ₇₄ O ₁₃	2093058-24-1
M22*	3 β ,23-Dihydroxy-1,12-dioxo-olean-28-oic acid	6	0.01175987	C ₃₀ H ₄₆ O ₆	2093058-20-7
M23*	3 β ,23,27-Trihydroxy-1-oxo-olean-12-ene-28-oic acid	6	0.01175987	C ₃₀ H ₄₆ O ₆	2093058-21-8
M24*	2 α ,3 β ,23-Trihydroxyurs-11-oxo-12-ene-28-oic acid	6	0.00987916	C ₃₀ H ₄₆ O ₆	107302-99-8
M25	Cyclocarioside II	3	0.00464055	C ₃₅ H ₃₆ O ₈	173294-76-3
M26*	Arjunglucoside II	7	0.01868779	C ₃₆ H ₅₈ O ₁₀	62369-72-6
M27	Quadranoside IV	6	0.00588117	C ₃₆ H ₅₈ O ₁₀	267001-55-8
M28*	α -Boswellic acid	7	0.0127971	C ₃₀ H ₄₈ O ₃	471-66-9
M29	3 α ,4 β ,18 α -3-Hydroxyurs-12-en-23-oic acid	6	0.00860482	C ₃₀ H ₄₈ O ₃	2243454-82-0
M30	β -Boswellic acid	6	0.00860482	C ₃₀ H ₄₈ O ₃	631-69-6
M31	Cyclocarioside H	2	0.0001863	C ₄₃ H ₇₂ O ₁₃	1403937-87-0
M32	Cyclocarioside J	2	0.0001863	C ₃₅ H ₅₈ O ₉	1644624-86-1
M33*	2 α ,3 β ,23-Trihydroxyoleana-11,13(18)-dien-28-oic acid	5	0.01046971	C ₃₀ H ₄₆ O ₅	6790-76-7
M34	Cyclocaric acid A	5	0.00786033	C ₃₀ H ₄₆ O ₃	21754-17-6
M35	(+)-Betulinic acid	4	0.00368899	C ₃₀ H ₄₈ O ₃	472-15-1
M36*	Cyclocarioside K	5	0.01444275	C ₃₆ H ₅₈ O ₈	1644624-87-2
M37	Cyclocarioside III	2	0.00041134	C ₃₆ H ₆₀ O ₉	173294-77-4
M38	Cyclocarioside L	2	0.00041134	C ₃₈ H ₆₂ O ₁₂	2093058-22-9
M39	Cyclocarioside M	1	0	C ₄₀ H ₆₄ O ₁₃	2093058-23-0

^a * means the core active ingredients.

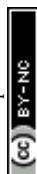


Table 2 The information of the target proteins used in the manuscript

UniProt IDs	Protein name	Gene name	Betweenness centrality	Degree
P24666	Low molecular weight phosphotyrosine protein phosphatase	ACP1	0.04820819	16
P15121	Aldose reductase	AKR1B1	0.00271300	2
P10275	Androgen receptor	AR	0.01510033	6
P42574	Caspase-3	CASP3	0.00164939	2
P21554	Cannabinoid receptor 1 (by homology)	CNR1	0.01369327	2
P11511	Cytochrome P450 19A1	CYP19A1	0.02352865	8
P03372	Estrogen receptor alpha	ESR1	0.01221787	5
P00734	Thrombin	F2	0.00971275	5
P12104	Fatty acid binding protein intestinal	FABP2	0.00305104	3
P15090	Fatty acid binding protein adipocyte	FABP4	0.00487387	4
P09038	Basic fibroblast growth factor	FGF2	0.02471924	3
P04035	HMG-CoA reductase	HMGCR	0.01564161	6
P28845	11-Beta-hydroxysteroid dehydrogenase 1	HSD11B1	0.14538756	24
P80365	11-Beta-hydroxysteroid dehydrogenase 2	HSD11B2	0.00419333	3
O14920	Inhibitor of nuclear factor kappa B kinase beta subunit	IKBKB	0.00271300	2
P60568	Interleukin-2	IL2	0.01315293	6
P35968	Vascular endothelial growth factor receptor 2	KDR	0.00265707	2
Q16539	MAP kinase p38 alpha	MAPK14	0.01369327	2
P35228	Nitric oxide synthase, inducible	NOS2	0.03106597	13
Q96RI1	Bile acid receptor FXR	NR1H4	0.00625265	4
P04150	Glucocorticoid receptor	NR3C1	0.00661281	4
P42336	PI3-kinase p110-alpha subunit	PIK3CA	0.00164939	2
Q07869	Peroxisome proliferator-activated receptor alpha	PPARA	0.00487387	4
Q03181	Peroxisome proliferator-activated receptor delta	PPARD	0.00487387	4
P37231	Peroxisome proliferator-activated receptor gamma	PPARG	0.00487387	4
P35354	Cyclooxygenase-2	PTGS2	0.00176559	2
P18031	Protein-tyrosine phosphatase 1B	PTPN1	0.20514580	28
P10586	Receptor-type tyrosine-protein phosphatase F (LAR)	PTPRF	0.03748757	15
O00767	Acyl-CoA desaturase	SCD	0.00139742	2
P04278	Testis-specific androgen-binding protein	SHBG	0.01774612	7
P40763	Signal transducer and activator of transcription 3	STAT3	0.04628878	9
P11473	Vitamin D receptor	VDR	0.07454926	10
P15692	Vascular endothelial growth factor A	VEGFA	0.03145277	4

with multiple compounds. Network values for active components and target proteins are listed in Tables 1 and 2.

3.3 Analysis of target protein PPI network

The interaction network for the anti-diabetic target proteins of *C. paliurus*, including 33 nodes and 153 edges, was constructed based on an interaction reliability greater than mid-range (Fig. 3). In protein interaction networks, the position of node proteins is not the same. The most important nodes are key nodes. The network node degree parameter is a common method for evaluating key nodes of the network. The PPI network showed that the median value of betweenness centrality and closeness centrality was greater than the median with high degree for a total of 15 target proteins. These 15 target proteins (Table 3) can be considered as core target proteins for the treatment of diabetes and may play an important role in the treatment of diabetes. Among them, PTGS2, VEGFA, CASP3, PPARG, STAT3, and NR3C1 are centrally located in the PPI network (Fig. 3), indicating that these proteins are involved in the pathogenesis of diabetes mellitus.

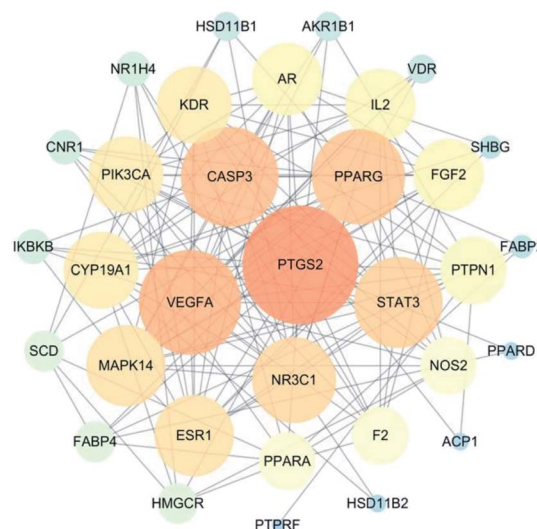


Fig. 3 *Cyclocarya paliurus*'s potential target protein interaction network for treatment of diabetes. The size and color of the node represent the size of the degree value. The darker the color, the larger the node and the more important it is in the network.



Table 3 Features of *Cyclocarya paliurus*'s protein–protein interaction network

Gene symbol	Name	Betweenness centrality	Closeness centrality	Degree
PTGS2	Prostaglandin-endoperoxide synthase 2	0.15607784	0.76190476	22
VEGFA	Vascular endothelial growth factor A	0.05924204	0.71111111	19
CASP3	Caspase 3	0.04400629	0.68085106	18
PPARG	Peroxisome proliferator activated receptor gamma	0.14458119	0.68085106	17
STAT3	Signal transducer and activator of transcription 3	0.02315961	0.66666667	16
NR3C1	Nuclear receptor subfamily 3 group C member 1	0.07062907	0.62745098	15
MAPK14	Mitogen-activated protein kinase 14	0.01666107	0.64000000	14
ESR1	Estrogen receptor 1	0.04629238	0.62745098	14
KDR	Kinase insert domain receptor	0.04561908	0.60377358	13
PIK3CA	Phosphatidylinositol-4,5-bisphosphate 3-kinase catalytic subunit alpha	0.01214958	0.60377358	13
CYP19A1	Cytochrome P450 family 19 subfamily a member 1	0.07381888	0.60377358	13
IL2	Interleukin 2	0.01306559	0.57142857	12
AR	Androgen receptor	0.01695036	0.58181818	12
PTPN1	Protein tyrosine phosphatase non-receptor type 1	0.09420179	0.59259259	11
PPARA	Peroxisome proliferator activated receptor alpha	0.03537093	0.54237288	9

3.4 GO analysis of target proteins

GO enrichment analysis results (Fig. 4) showed that the numbers of target proteins involved in the BP, MF, and CC categories were 116 (70.7%), 37 (22.5%), and 11 (6.7%), respectively. In the BP category, the target proteins were mainly involved in positive regulation of transcription from RNA polymerase II promoter (13, 39.4%), signal transduction (9, 29.3%), and transcription, DNA-templated (9, 29.3%). In the MF category, the target proteins were mainly involved in protein binding (23, 69.7%), zinc ion binding (9, 27.3%) and transcription factor activity, sequence-specific DNA binding (9, 27.3%). In the CC category, target proteins were mainly associated with the nucleus (17, 51.5%), cytoplasm (14, 42.4%), nucleoplasm (12, 36.4%) and cytosol (12, 36.4%). These results demonstrate that *C. paliurus* acts on diabetes probably by engaging the above mentioned pathways.

3.5 KEGG classification of target proteins

KEGG pathway annotation showed that 27 of the 33 (81.8%) potential target proteins were enriched and involved in 41

pathways; of these, 35 pathways were significantly correlated with the target proteins ($P \leq 0.05$). The following pathways included the largest number of proteins: cancer pathways (11, 40.7%), proteoglycans in cancer (8, 29.6%), insulin resistance (6, 22.2%), acute myeloid leukemia (6, 22.2%), PI3K-Akt signaling pathway (6, 22.2%), PPAR signaling pathway (6, 22.2%), and so on. The top 10 pathways with the largest number of proteins involved are shown in Fig. 4, and the complete KEGG classification results are shown in Table 4. The target proteins involved in cancer pathways included

AR, CASP3, PPARG, PTGS2, VEGFA, PPARG, PIK3CA, NOS2, IKBKB, FGF2 and STAT3; the target proteins involved in insulin resistance included PPARA, PTPRF, PIK3CA, PTPN1, IKBKB and STAT3; and the target proteins involved in the PI3K-Akt signaling pathway included VEGFA, PIK3CA, IKBKB, FGF2, KDR and IL2. As can be seen, there are multiple target proteins in a pathway and the same target protein exists in multiple pathways. These results suggest that *C. paliurus* may exert its effects on diabetes by regulating these pathways *via* core target proteins.

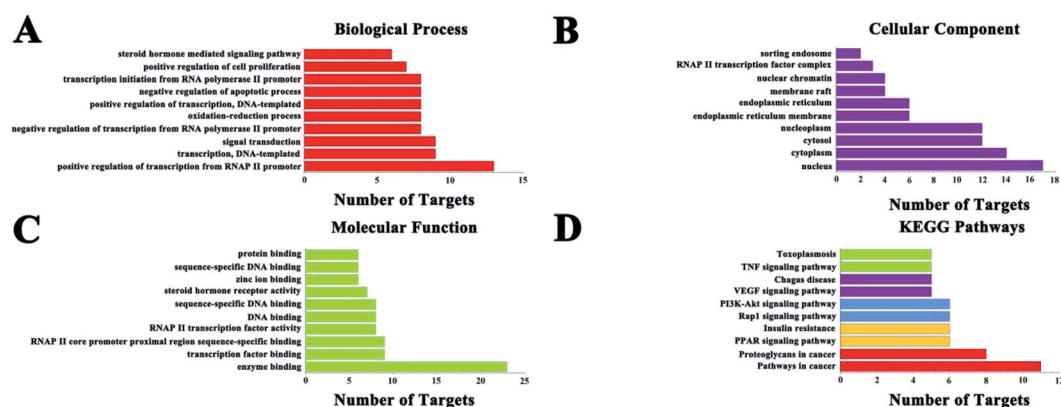


Fig. 4 GO and KEGG pathways analyses of the target proteins by using the DAVID database. Data show the top 10 remarkably enriched items in the biological process (A), cell component (B), molecular function (C), and KEGG pathways (D).

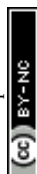


Table 4 The result of KEGG Classification of target proteins^a

Term	Count	Percent%	P value	Genes
hsa05200:Pathways in cancer	11	33.33	0.00	AR, CASP3, PPARD, PTGS2, VEGFA, PPARG, PIK3CA, NOS2, IKBKB, FGF2, STAT3
hsa05205:Proteoglycans in cancer	8	24.24	0.00	CASP3, MAPK14, VEGFA, ESR1, PIK3CA, FGF2, STAT3, KDR
hsa03320:PPAR signaling pathway	6	18.18	0.00	PPARA, PPARD, SCD, PPARG, FABP4, FABP2
hsa04931:Insulin resistance	6	18.18	0.00	PPARA, PTPRF, PIK3CA, PTPN1, IKBKB, STAT3
hsa04015:Rap1 signaling pathway	6	18.18	0.00	MAPK14, CNR1, VEGFA, PIK3CA, FGF2, KDR
hsa04151:PI3K-Akt signaling pathway	6	18.18	0.02	VEGFA, PIK3CA, IKBKB, FGF2, KDR, IL2
hsa04370:VEGF signaling pathway	5	15.15	0.00	PTGS2, MAPK14, VEGFA, PIK3CA, KDR
hsa05142:Chagas disease	5	15.15	0.00	MAPK14, PIK3CA, NOS2, IKBKB, IL2
hsa04668:TNF signaling pathway	5	15.15	0.00	CASP3, PTGS2, MAPK14, PIK3CA, IKBKB
hsa05145:Toxoplasmosis	5	15.15	0.00	CASP3, MAPK14, NOS2, IKBKB, STAT3
hsa05160:Hepatitis C	5	15.15	0.00	PPARA, MAPK14, PIK3CA, IKBKB, STAT3
hsa04014:Ras signaling pathway	5	15.15	0.02	VEGFA, PIK3CA, IKBKB, FGF2, KDR
hsa05206:MicroRNAs in cancer	5	15.15	0.03	CASP3, PTGS2, VEGFA, IKBKB, STAT3
hsa05221:Acute myeloid leukemia	4	12.12	0.00	PPARD, PIK3CA, IKBKB, STAT3
hsa05212:Pancreatic cancer	4	12.12	0.00	VEGFA, PIK3CA, IKBKB, STAT3
hsa04917:Prolactin signaling pathway	4	12.12	0.00	MAPK14, ESR1, PIK3CA, STAT3
hsa05222:Small cell lung cancer	4	12.12	0.01	PTGS2, PIK3CA, NOS2, IKBKB
hsa04066:HIF-1 signaling pathway	4	12.12	0.01	VEGFA, PIK3CA, NOS2, STAT3
hsa04660:T cell receptor signaling pathway	4	12.12	0.01	MAPK14, PIK3CA, IKBKB, IL2
hsa05169:Epstein-Barr virus infection	4	12.12	0.02	MAPK14, PIK3CA, IKBKB, STAT3
hsa04152:AMPK signaling pathway	4	12.12	0.02	HMGCR, SCD, PPARG, PIK3CA
hsa04380:Osteoclast differentiation	4	12.12	0.02	MAPK14, PPARG, PIK3CA, IKBKB
hsa04068:FoxO signaling pathway	4	12.12	0.02	MAPK14, PIK3CA, IKBKB, STAT3
hsa04910:Insulin signaling pathway	4	12.12	0.02	PTPRF, PIK3CA, PTPN1, IKBKB
hsa04550:Signaling pathways regulating pluripotency of stem cells	4	12.12	0.02	MAPK14, PIK3CA, FGF2, STAT3
hsa05161:Hepatitis B	4	12.12	0.02	CASP3, PIK3CA, IKBKB, STAT3
hsa04932:Non-alcoholic fatty liver disease (NAFLD)	4	12.12	0.03	PPARA, CASP3, PIK3CA, IKBKB
hsa05152:Tuberculosis	4	12.12	0.04	VDR, CASP3, MAPK14, NOS2
hsa04923:Regulation of lipolysis in adipocytes	3	9.09	0.02	PTGS2, PIK3CA, FABP4
hsa00140:Steroid hormone biosynthesis	3	9.09	0.03	HSD11B1, HSD11B2, CYP19A1
hsa04210:Apoptosis	3	9.09	0.03	CASP3, PIK3CA, IKBKB
hsa05120:Epithelial cell signaling in <i>Helicobacter pylori</i> infection	3	9.09	0.03	CASP3, MAPK14, IKBKB
hsa04920:Adipocytokine signaling pathway	3	9.09	0.04	PPARA, IKBKB, STAT3
hsa05140:Leishmaniasis	3	9.09	0.04	PTGS2, MAPK14, NOS2
hsa05133:Pertussis	3	9.09	0.04	CASP3, MAPK14, NOS2

^a Bold text means the important pathways reported and involved in diabetes.

3.6 Component-target-pathway network

The component-target-pathway network was constructed to visualize all interactions between target proteins and the anti-diabetic-related pathways. Based on the previous KEGG enrichment analysis, a component-target-pathway network was generated by connecting compounds, targets and pathways (Fig. 5). This network included 106 nodes (39 active compound nodes, 33 composite target protein nodes and 34 pathways nodes) and 745 edges. The component-target-pathway network results are shown in ESI Table 2 and ESI Fig. 1.[†]

The network analysis showed that the median value of betweenness centrality and closeness centrality was greater than the median with high degree for a total of 15 pathways, including cancer (hsa05200), insulin resistance (hsa04931), HIF-1 signaling pathway (hsa04066), PI3K-Akt signaling pathway (hsa04151) *etc.* These 15 pathways with several diabetes-associated target proteins can be considered as core

pathways for the treatment of diabetes. Among these pathways, insulin resistance, HIF-1 signaling, and PI3K-Akt signaling pathways have been shown to have a clear link with the occurrence of diabetes. As shown in Fig. 6, the active anti-inflammatory and anti-diabetic components present in *C. pal-iurus* can synergize with multiple target proteins within these pathways to form a multi-component-multi-target-multi-pathway mechanism.

3.7 Docking analysis

The protein structure was set to a rigid macromolecule, and the algorithm to local search parameters. Then, the object of molecular docking visualization was selected according to the lowest binding energy for docking. The core compounds and diabetes target proteins were ranked according to the docking binding energy. The top 10 core target proteins in the previous ranking were docked with 7 key medicinal components (Table



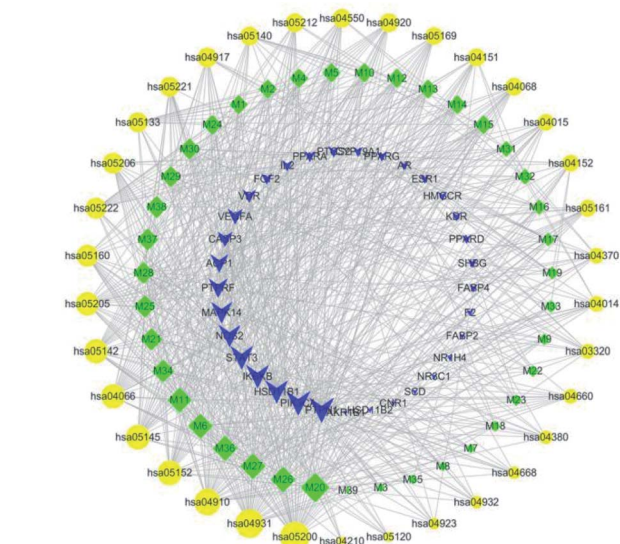


Fig. 5 *Cyclocarya paliurus*'s component-target-pathway network. The green diamond-shaped node represents the active ingredient, the blue triangular node represents the target, the yellow circular node represents the pathway, and the size of the node represent the degree value. Lines represent the relationships between the compounds, targets, and pathways.

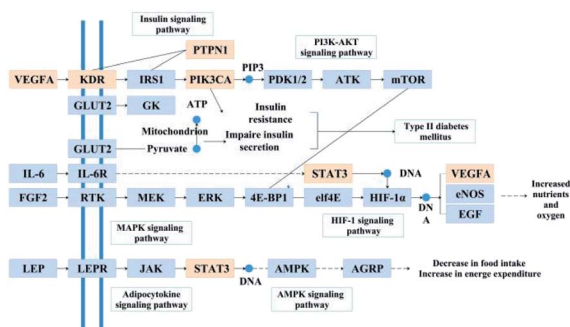


Fig. 6 Distribution of the target proteins of *Cyclocarya paliurus* on the predicted pathway. The orange boxes are potential target proteins of *C. paliurus*, while the blue boxes are relevant targets in the pathway.

5). The docking results showed that the lowest binding energy for the 7 core medicinal components and CASP3 were all below -5.0 kJ mol^{-1} , indicating that the 7 key medicinal components bound better to the CASP3 target protein. With the exception of $2\alpha,3\beta,23$ -trihydroxyoleana-11,13(18)-dien-28-oic acid, the other 6 components showed the lowest docking binding energies with MAPK14 target protein when compared with the other 9 target proteins. Interactions between the 7 core components of *C. paliurus* and the previously mentioned 10 core target proteins with the lowest binding energy and the smallest binding energy were visualized using Pymol (Fig. 7 and 8).

4. Discussion

Used as a health tea, *C. paliurus* has a long history in the treatment of diabetes. In this study, 39 triterpenoids were

selected to create a library of compounds, and network pharmacology technology was employed to explore the multiple pharmacological effects of triterpenoids from *C. paliurus* in the treatment of diabetes. We found that seven core ingredients found in the leaves of *C. paliurus*, including arjunglucoside II, α -boswellic acid, $3\beta,23$ -dihydroxy-1,12-dioxo-olean-28-oic acid, 23 -trihydroxyurs-11-oxo-12-ene-28-oic acid, $3\beta,23,27$ -trihydroxy-1-oxo-olean-12-ene-28-oic acid, $2\alpha,3\beta,23$ -trihydroxyoleana-11,13(18)-dien-28-oic acid and cyclocarioside K, showed potential for the treatment of diabetes.

Among them, α -boswellic acid has been shown to exert some anti-diabetic effects by suppressing the expression of proinflammatory cytokines.⁴⁵ Cyclocarioside K is a new epoxydammarene triterpenoid saponin isolated from the ethanol leaf extracts of *C. paliurus*.⁴⁶ Arjunglucoside II is a common glucoside derivative of arjunic acid, found in *Terminalia arjuna*.⁴⁷ The DPP-IV inhibitory activity of arjunic acid and its derivatives may have demonstrable therapeutic benefits in diabetes patients with cardiovascular comorbidities.⁴⁸ $3\beta,23$ -Dihydroxy-1,12-dioxo-olean-28-oic acid and $3\beta,23,27$ -trihydroxy-1-oxo-olean-12-ene-28-oic acid are new triterpenoid saponins isolated from the CH_3Cl -soluble extract of *C. paliurus* leaves, whereas 23 -trihydroxyurs-11-oxo-12-ene-28-oic acid and $2\alpha,3\beta,23$ -trihydroxyoleana-11,13(18)-dien-28-oic acid are also known triterpenoid saponins isolated from the CH_3Cl -soluble extract of the leaves of *C. paliurus*.⁴⁹ These triterpenoid saponins may have an effect on diabetes by inhibiting apolipoprotein B48 overproduction.^{49,50} Based on the interaction network of *C. paliurus*'s anti-diabetes target proteins, we found that STAT3, PTPN1, PTGS2, VEGFA, CASP3, etc. were the key target proteins. These targets may play an important role in *C. paliurus*'s anti-diabetes effects. According to the results of the component-target and PPI networks, STAT3, CASP3, ACP1, PTPN1, HSD11B1 and other targets were regulated by multiple components, such as arjunglucoside II, cyclocarioside K, quadranoside IV, corosolic acid, cyclocariosides I and so on. Some of these ingredients have been proven to have strong anti-diabetic effects. For example, corosolic acid can reduce glucose levels in human hepatocellular carcinoma cells, as well as in zebrafish and in rats.⁵¹ Molecular docking is one of the most widely used methods to investigate binding of a compound to a target protein.³⁹⁻⁴² We used AutoDock software to analyze molecular docking. Compounds with docking energies below -5 kJ mol^{-1} were considered to bind well to their targets. In the PPI network diagram, betweenness is defined as the number of shortest paths between pairs of nodes that run through nodes, and degree centrality is the most direct measure of the importance of a network node. The larger the node degree value, the more important the node is in the network. We used two indicators (betweenness centrality and closeness centrality) as the criteria for screening targets. Only target proteins with betweenness centrality and closeness centrality values greater than the median for all target proteins were selected. Then, all target proteins were sorted according to the degree value. Based on the docking results between the top ten target proteins and core compounds, we observed that although the docking energy of some core compounds to the target proteins was very small



Table 5 The lowest binding energy of the active ingredients of *Cyclocarya paliurus* to the ten core target proteins for treating diabetes (unit: kcal mol⁻¹)^a

No.	Compound name	PTGS2	VEGFA	CASP3	PPARG	STAT3	NR3C1	MAPK14	ESR1	KDR	PIK3CA
1	3β,23-Dihydroxy-1,12-dioxo-olean-28-oic acid	-5.2	-4.5	-6.1	2.9	-5.8	-2.4	-6.7	-4.7	-5	-5.3
2	3β,23,27-Trihydroxy-1-oxo-olean-12-ene-28-oic acid	-5.2	-4.5	-6.1	3	-5.8	-2.4	-6.7	-4.7	-4.9	-5.3
3	2α,3β,23-Trihydroxyurs-11-oxo-12-ene-28-oic acid	-4.9	-4.6	-5.2	0	-5.2	-3.6	-8.7	-2.9	-5	-6.1
4	Arjunglucoside II	-4.8	-4.6	-5.5	2.3	-5.4	8.2	-6.7	-4.4	-4.6	-5.1
5	α-Boswellic acid	-5.7	-5.3	-6.1	-1.7	-6	-6.3	-9.1	-6.2	-5.2	-5.4
6	2α,3β,23-Trihydroxyoleana-11,13(18)-dien-28-oic acid	-4.5	-4	-6	1.7	-5.3	16.5	-0.3	3.1	-4.7	-2.4
7	Cyclocarioside K	-4.3	-4.9	-4.9	-2.9	-5.7	4.5	-8	-5.4	-4.1	-6.7
8	Ligand	-3.1	-3.2	-3.4	-5.5	-6.1	-11.9	-12.7	-11.4	-3	-7.9

^a Ligand means a substance that has the ability to recognize the receptor and can bind to it. The docking method is rigid docking. Algorithm is all local search parameters.

and the docking results were good, there were also poor docking results. Only STAT3 showed docking energies for all compounds below -5.0 kJ mol⁻¹. STAT3 is an important signal transduction factor that participates in the signal transduction process of various cytokines, such as interferon, interleukins and growth factors, and it forms part of the JAK2/STAT3 signaling pathway.^{52,53} Many studies have shown that the JAK2/STAT3 signaling pathway is an important pathway that mediates the signal transduction process of various cytokines and growth factors, and can regulate cell growth, differentiation, migration, apoptosis, autophagy, immunity, and metabolism.^{54,55} It is considered to play an important role in the development of diabetes.^{56,57} STAT3 may regulate diabetes by acting on SOCS3 and thereby affecting the activity of insulin receptor substrate 1 (IRS-1).⁵⁸ SOCS3 can mediate central leptin resistance in obese patients.⁵⁹ Leptin signal transduction in the hypothalamus can regulate liver glucose and lipid metabolism⁶⁰ as well as liver gluconeogenesis.⁶¹ It seems evident that STAT3 plays an important role regulating disorders of glucose metabolism. In network pharmacology analysis, component-target networks are mainly used to screen core compounds and targets. Betweenness centrality and closeness centrality values greater than the median are also the main criteria for screening core target proteins. All eligible target proteins were sorted according to the degree value, and the one with the highest value was PTPN1. In component-target networks, the higher the degree of a target protein, the more the compounds that regulate it. The more active compounds bind to a protein target, the greater the possibility of producing a drug effect, and the higher the possibility of becoming the key node. PTPN1 is a non-transmembrane protein tyrosine phosphatase which functions mainly by regulating the levels of protein tyrosine phosphorylation in cells, and is involved in the regulation of multiple cell signaling pathways modulating cell proliferation, growth and migration.⁶² Many studies have found correlations between DNA sequence variations in the PTPN1 gene, SNP and T2DM.⁶³ PTPN1 can regulate insulin signaling by modulating the

expression of the PTP1B enzyme. PTP1B is a key factor that regulates various metabolic processes in the body and is closely associated with the occurrence and development of diseases, especially type 2 diabetes.⁶⁴ Studies have shown that inhibiting the expression of the PTP1B enzyme can regulate insulin sensitivity in diabetic mice and improve insulin resistance, producing a therapeutic effect in diabetes.⁶⁵ Therefore, we can study the potential pharmacological effects of *C. paliurus* on diabetes by analyzing the relationship between active compounds and core targets. According to the results of the component-target-protein network analysis, the insulin resistance pathway, the PI3K-Akt signaling pathway and the HIF-1 pathway may play an important role in the anti-diabetes effects of *C. paliurus*. As shown in Fig. 5, the active anti-diabetes ingredients of *C. paliurus* can synergize with various target proteins in these pathways, resulting in a multi-component, multi-target and multi-pathway mechanism of action. Insulin resistance, a condition in which cells are resistant to the action of insulin, is often found in obese and diabetic patients and is closely related with the occurrence of diabetes.⁶⁶ It is associated with increased activity of phosphatases, including PTPs, PTEN, and PP2A, and decreased activation of signaling molecules, such as PI3K/AKT, resulting in reduced GLUT4 translocation, glucose uptake and glycogen synthesis in skeletal muscle, as well as increased hepatic gluconeogenesis and decreased glycogen synthesis in the liver.^{67,68} Many studies have shown that this pathway is activated by changes in the levels of enzymes such as IL-6, Akt and IRS-1, or other abnormalities.^{69,70}

HIF-1, a transcription factor which functions as a major regulator of oxygen homeostasis,⁷¹ is associated with diabetic nephropathy. HIF-1 expression indirectly affects renal oxygen metabolism.^{72,73} In hypoxic conditions, it can reduce damage to the body by regulating the expression of target genes coding for downstream factors related with the hypoxic response, and by modulating cell energy metabolism, glucose metabolism and apoptosis.⁷⁴ Studies have shown that oxidative stress and



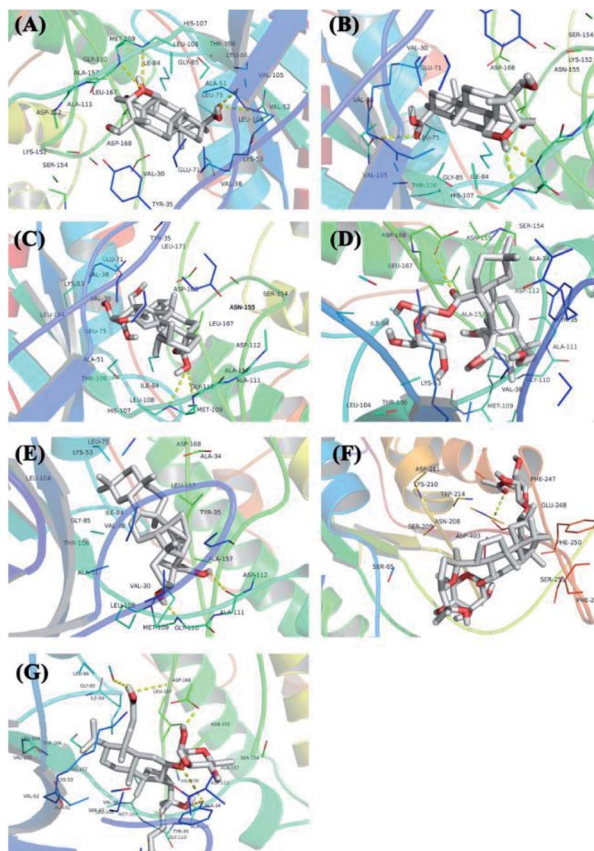


Fig. 7 The 3D docking results of ten core protein molecules and seven core components are visualized. The structure of the compound is represented by a stick, the different branches of the protein are represented by different colors, and the yellow dotted line represents its hydrogen bond, which marks the position of the hydrogen bond and the compound in the compound. (A) Interaction between 3 β ,23-dihydroxy-1,12-dioxo-olean-28-oic acid and MAPK14; (B) interaction between 3 β ,23,27-trihydroxy-1-oxo-olean-12-ene-28-oic acid and MAPK14; (C) interaction between 2 α ,3 β ,23-trihydroxyurs-11-oxo-12-ene-28-oic acid and MAPK14; (D) interaction between arjunglucoside II and MAPK14; (E) interaction between α -boswellic acid and MAPK14; (F) interaction between 2 α ,3 β ,23-trihydroxyoleana-11,13(18)-dien-28-oic acid and CASP3; (G) interaction between cyclocarioside K and MAPK14.

microcirculatory disorders are important mechanisms underlying the development of diabetic nephropathy and other diseases.⁷⁵ Antioxidant therapy can increase the expression of medullary HIF in diabetic kidneys and improve renal oxidative damage.⁷⁶ HIF activation can attenuate changes in renal oxygen metabolism and mitochondrial function caused by diabetes, thereby reducing proteinuria and improving renal tubular interstitial damage.⁷⁷ The PI3K-Akt signaling pathway plays an important role maintaining insulin stability.⁷⁵ This signaling pathway is activated by many types of cellular stimuli or toxic insults and regulates fundamental cellular functions, such as transcription, translation, proliferation, growth, and survival.⁷⁶ At present, there are studies showing that selective activation of the PI3K/AKT signaling pathway can modulate neuroprotection, angiogenesis, and islet β cell survival in diabetic rats.⁷⁵ Many

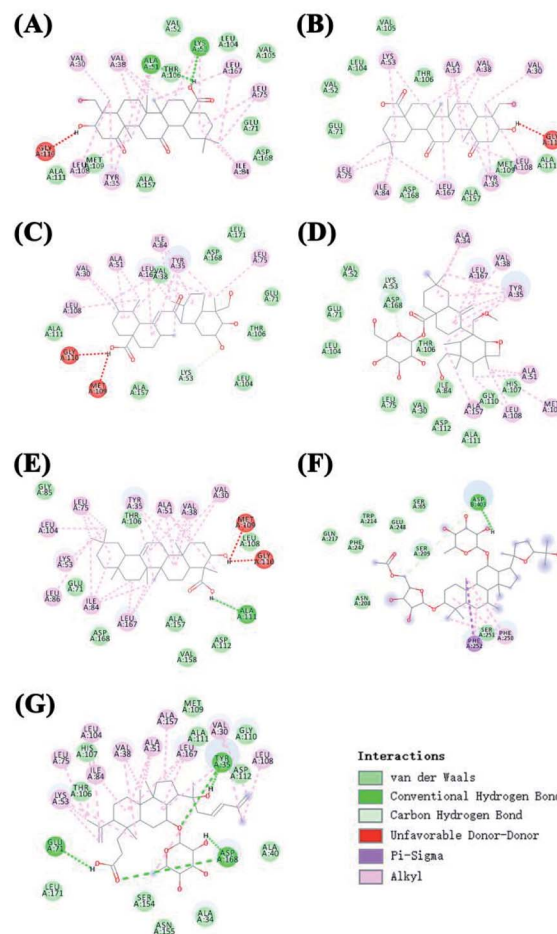


Fig. 8 The 2D docking results of ten core protein molecules and seven core components are visualized. (A) Interaction between 3 β ,23-dihydroxy-1,12-dioxo-olean-28-oic acid and MAPK14; (B) interaction between 3 β ,23,27-trihydroxy-1-oxo-olean-12-ene-28-oic acid and MAPK14; (C) interaction between 2 α ,3 β ,23-trihydroxyurs-11-oxo-12-ene-28-oic acid and MAPK14; (D) interaction between arjunglucoside II and MAPK14; (E) interaction between α -boswellic acid and MAPK14; (F) interaction between 2 α ,3 β ,23-trihydroxyoleana-11,13(18)-dien-28-oic acid and CASP3; (G) interaction between cyclocarioside K and MAPK14.

studies indicate that after insulin binds to its receptor, it will self-phosphorylate and phosphorylate IRS-2 tyrosine sites.⁷⁷ Phosphorylated IRS-2 binds to the p85 subunit of PI3K to further activate PI3K and to activate Akt after phosphorylation.⁷⁸ Akt2 is a subtype of Akt, a serine/threonine kinase and an important signaling molecule located downstream of PI3K^{79,80} which can be regulated by modulating Gsk-3 β , GLUT-4, *etc.* A series of downstream molecules promote glycogen synthesis, glucose transport, and other pathways to regulate glucose metabolism, thereby regulating diabetes.⁷⁰

Receptor theory is the basic theory of pharmacodynamics. It postulates that the combination of drugs and target proteins is the basis of pharmacodynamics. Molecular docking is a method to evaluate binding of active drug ingredients to target proteins. In network pharmacology research, molecular docking is commonly used to study interactions between small molecules and key target proteins of the network, and to validate key target



proteins identified in the network based on the interaction between active ingredients and target proteins.⁸¹ As shown in Fig. 7 and 8, molecular docking analysis showed that hydrogen bonds can form spontaneously between the 7 core compounds and the target proteins. These studies showed that the number of hydrogen bonds between the 4 core compounds and the target proteins surpassed 4, indicating that the core components bound well to the key target proteins of the network. Although the molecular docking results support the key node network findings, more *in vivo* or *in vitro* experiments are needed to verify the effects of ingredients on target proteins.

5. Conclusions

In summary, in this study we applied the network pharmacological approach to investigate the main target proteins of *C. paliurus* compounds with anti-diabetes activity by constructing a target interaction network, and used molecular docking methods to validate the key findings. Our results indicate that seven compounds, including arjunglucoside II, α -boswellic acid, 3 β ,23-dihydroxy-1,12-dioxo-olean-28-oic acid, 23-trihydroxyurs-11-oxo-12-ene-28-oic acid, 3 β ,23,27-trihydroxy-1-oxo-olean-12-ene-28-oic acid, 2 α ,3 β ,23-trihydroxyoleana-11,13(18)-dien-28-oic acid and cyclocarioside K show potential for the treatment of diabetes. These triterpene compounds were predicted to interact with PTGS2, VEGFA, CASP3, and other enzymes. These results provide valuable insights into the synergistic mechanism of action of natural medicines. In addition, biological process and pathway enrichment analysis of the target proteins of the active ingredients of *C. paliurus* enhanced our understanding of its mechanism of action in diabetes. Our study provides scientific evidence supporting the use of *C. paliurus* for the treatment of diabetes and confirms that modern technologies can be used to explore the therapeutic value of natural medicines.

Data availability statement

All datasets generated for this study are included in the article/ESI.†

Author contributions statement

Z. L., Y. T., N. L., Z. Z., and J. L. conceived the experiment. Z. L. collected data, conducted the analytical part, wrote the first version of the manuscript. Y. T., N. L., Z. Z., and J. L. revised the manuscript. J. L. finalized the manuscript. All authors read and approved the final manuscript.

Conflicts of interest

The authors declare that they have no conflict of interest.

Acknowledgements

This research work was financially supported by Zhejiang Provincial Key Research and Development Program

(2018C02021) and the Ten Thousand Talent Program of Zhejiang Province (2019).

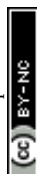
Notes and references

- 1 J. M. Baena-Díez, J. Peñafiel, I. Subirana, R. Ramos, R. Elosua, A. Marín-Ibañez, M. J. Guembe, F. Rigo, M. J. Tormo-Díaz, C. Moreno-Iribas, J. J. Cabré, A. Segura, M. García-Lareo, A. Gómez De La Cámara, J. Lapetra, M. Quesada, J. Marrugat, M. J. Medrano, J. Berjón, G. Frontera, D. Gavrila, A. Barricarte, J. Basora, J. M. García, N. C. Pavone, D. Lora-Pablos, E. Mayoral, J. Franch, M. Mata, C. Castell, A. Frances and M. Grau, Risk of Cause-Specific Death in Individuals with Diabetes: A Competing Risks Analysis, *Diabetes Care*, 2016, **39**, 1987–1995.
- 2 N. M. Maruthur, M. O. Gribble, W. L. Bennett, S. Bolen, L. M. Wilson, P. Balakrishnan, A. Sahu, E. Bass, W. H. L. Kao and J. M. Clark, The Pharmacogenetics of Type 2 Diabetes: A Systematic Review, *Diabetes Care*, 2014, **37**, 876.
- 3 C. I. Ezema, E. Omeh, O. Onyeso, C. C. Anyachukwu, M. J. Nwankwo, A. Amaeze, J. U. Ugwulebor, E. O. Nna, E. O. Ohotu and I. Ugwuanyi, The Effect of an Aerobic Exercise Programme On Blood Glucose Level, Cardiovascular Parameters, Peripheral Oxygen Saturation, and Body Mass Index Among Southern Nigerians with Type 2 Diabetes Mellitus, Undergoing Concurrent Sulfonylurea and Metformin Treatment, *Malays. J. Med. Sci.*, 2019, **26**, 88–97.
- 4 S. H. Havale and M. Pal, Medicinal Chemistry Approaches to the Inhibition of Dipeptidyl Peptidase-4 for the Treatment of Type 2 Diabetes, *Bioorg. Med. Chem.*, 2009, **17**, 1783–1802.
- 5 A. J. Krentz and C. J. Bailey, Oral Antidiabetic Agents, *Drugs*, 2005, **65**, 385–411.
- 6 J. J. Neumiller, J. R. White and R. K. Campbell, Sodium-Glucose Co-Transport Inhibitors, *Drugs*, 2010, **70**, 377–385.
- 7 P. Kaur, A. Mittal, S. Nayak, M. Vyas, V. Mishra and G. Khatik, Current Strategies and Drug Targets in the Management of Type 2 Diabetes Mellitus, *Curr. Drug Targets*, 2018, **19**, 1738–1766.
- 8 Z. Zhang, Y. Cao, Y. J. Tao, E. Meng, J. H. Tang, Y. C. Liu and F. P. Li, Sulfonylurea and Fracture Risk in Patients with Type 2 Diabetes Mellitus: A Meta-Analysis, *Diabetes Res. Clin. Pract.*, 2019, **159**, S168–S8227.
- 9 L. Wang, P. Gao, M. Zhang, Z. Huang, D. Zhang, Q. Deng, Y. Li, Z. Zhao, X. Qin, D. Jin, M. Zhou, X. Tang, Y. Hu and L. Wang, Prevalence and Ethnic Pattern of Diabetes and Prediabetes in China in 2013, *JAMA*, 2017, **317**, 2515–2523.
- 10 S. S. Kolhe and P. R. Rachh, Review On Potent Anti-Diabetic Plants Or Herbs From Traditional Medicine, *J. Drug Deliv. Therapeut.*, 2018, **8**, 92–98.
- 11 Q. Ge, L. Chen, Y. Yuan, L. Liu, F. Feng, P. Lv, S. Ma, K. Chen and Q. Yao, Network Pharmacology-Based Dissection of the Anti-Diabetic Mechanism of *Lobelia Chinensis*, *Front. Pharmacol.*, 2020, **11**, 347.



- 12 H. Zhang, H. Matsuda, A. Kumahara, Y. Ito, S. Nakamura and M. Yoshikawa, New Type of Anti-Diabetic Compounds From the Processed Leaves of *Hydrangea Macrophylla* Var. *Thunbergii* (Hydrangeae Dulcis Folium), *Bioorg. Med. Chem. Lett.*, 2007, **17**, 4972–4976.
- 13 L. Zhai, Z. Ning, T. Huang, B. Wen, C. Liao, C. Lin, L. Zhao, H. Xiao and Z. Bian, *Cyclocarya paliurus* Leaves Tea Improves Dyslipidemia in Diabetic Mice: A Lipidomics-Based Network Pharmacology Study, *Front. Pharmacol.*, 2018, **9**, 973.
- 14 Z. Yang, K. Ouyang, J. Zhao, H. Chen, L. Xiong and W. Wang, Structural Characterization and Hypolipidemic Effect of *Cyclocarya Paliurus* Polysaccharide in Rat, *Int. J. Biol. Macromol.*, 2016, **91**, 1073–1080.
- 15 H. Kurihara, H. Fukami, A. Kusumoto, Y. Toyoda, H. Shibata, Y. Matsui, S. Asami and T. Tanaka, Hypoglycemic Action of *Cyclocarya paliurus* (Batal.) Iljinskaja in Normal and Diabetic Mice, *Biosci., Biotechnol., Biochem.*, 2003, **67**, 877–880.
- 16 S. Li, J. Li, X. Guan, J. Li, S. Deng, L. Li, M. Tang, J. Huang, Z. Chen and R. Yang, Hypoglycemic Effects and Constituents of the Barks of *Cyclocarya Paliurus* and their Inhibiting Activities to Glucosidase and Glycogen Phosphorylase, *Fitoterapia*, 2011, **82**, 1081–1085.
- 17 Y. Ma, C. Jiang, N. Yao, Y. Li and Q. Wang, Antihyperlipidemic Effect of *Cyclocarya Paliurus* (Batal.) Iljinskaja Extract and Inhibition of Apolipoprotein B48 Overproduction in Hyperlipidemic Mice, *J. Ethnopharmacol.*, 2015, **166**, 286–296.
- 18 Q. Wang, C. Jiang, S. Fang, J. Wang, Y. Ji, X. Shang, Y. Ni, Z. Yin and J. Zhang, Antihyperglycemic, Antihyperlipidemic and Antioxidant Effects of Ethanol and Aqueous Extracts of *Cyclocarya Paliurus* Leaves in Type 2 Diabetic Rats, *J. Ethnopharmacol.*, 2013, **150**, 1119–1127.
- 19 J. Xie, D. Cai-jun, N. Shao-Ping, L. Feng, W. Zhi-Jun, S. Ming-Yue and M. Xie, Extraction, Chemical Composition and Antioxidant Activity of Flavonoids From *Cyclocarya Paliurus* (Batal.) Iljinskaja Leaves, *Food Chem.*, 2015, **186**, 186–197.
- 20 J. Xie, M. Xie, S. Nie, M. Shen, Y. Wang and C. Li, Isolation, Chemical Composition and Antioxidant Activities of a Water-Soluble Polysaccharide From *Cyclocarya Paliurus* (Batal.) Iljinskaja, *Food Chem.*, 2010, **119**, 1626–1632.
- 21 H. Yoshitomi, R. Tsuru, L. Li, J. Zhou, M. Kudo, T. Liu and M. Gao, *Cyclocarya Paliurus* Extract Activates Insulin Signaling Via Sirtuin1 in C2C12 Myotubes and Decreases Blood Glucose Level in Mice with Impaired Insulin Secretion, *PLoS One*, 2017, **12**, e183988.
- 22 C. Jiang, Q. Wang, Y. Wei, N. Yao, Z. Wu, Y. Ma, Z. Lin, M. Zhao, C. Che, X. Yao, J. Zhang and Z. Yin, Cholesterol-Lowering Effects and Potential Mechanisms of Different Polar Extracts From *Cyclocarya Paliurus* Leave in Hyperlipidemic Mice, *J. Ethnopharmacol.*, 2015, **176**, 17–26.
- 23 W. Lin, Y. Li, Q. Lu, H. Lu and J. Li, Combined Analysis of the Metabolome and Transcriptome Identified Candidate Genes Involved in Phenolic Acid Biosynthesis in the Leaves of *Cyclocarya Paliurus*, *Int. J. Mol. Sci.*, 2020, **21**, 1337.
- 24 C. Sun, X. Shang, H. Ding, Y. Cao and S. Fang, Natural Variations in Flavonoids and Triterpenoids of *Cyclocarya Paliurus* Leaves, *J. For. Res.*, 2020, **31**, DOI: 10.1007/s11676-020-01139-1.
- 25 J. G. A. W. Du, M. Zhang and Li Chen, Compound K, a Final Intestinal Metabolite of Ginsenosides, Enhances Insulin Secretion in MIN6 Pancreatic β -Cells by Upregulation of GLUT2, *Fitoterapia*, 2013, **87**, 84–88.
- 26 A. E. Ardiles, Á. González-Rodríguez, M. J. N. Ez, N. R. Perestelo, V. Pardo, I. A. Jiménez, Á. M. Valverde and I. L. Bazzocchi, Studies of Naturally Occurring Friedelane Triterpenoids as Insulin Sensitizers in the Treatment Type 2 Diabetes Mellitus, *Phytochemistry*, 2012, **84**, 116–124.
- 27 D. K. Burdi, S. N. Qureshi and A. B. Ghanghro, An Overview of Available Hypoglycemic Triterpenoids and Saponins to Cure Diabetes Mellitus, *Adv. Life Sci.*, 2014, **1**, 119–128.
- 28 J. Han, N. Tuan, M. Park, K. Quan, J. Oh, K. Heo, M. Na and C. Myung, Cucurbitane Triterpenoids From the Fruits of *Momordica Charantia* Improve Insulin Sensitivity and Glucose Homeostasis in Streptozotocin-Induced Diabetic Mice, *Mol. Nutr. Food Res.*, 2018, **62**, 1700769.
- 29 K. N. Zhu, C. H. Jiang, Y. S. Tian, N. Xiao, Z. F. Wu, Y. L. Ma, Z. Lin, S. Z. Fang, X. L. Shang, K. Liu, J. Zhang, B. L. Liu and Z. Q. Yin, Two Triterpenoids From *Cyclocarya Paliurus* (Batal) Iljinsk (Juglandaceae) Promote Glucose Uptake in 3T3-L1 Adipocytes: The Relationship to AMPK Activation, *Phytomedicine*, 2015, **22**, 837–846.
- 30 S. Yue, L. Xin, Y. Fan, S. Li, Y. Tang, J. Duan, H. Guan and C. Wang, Herb Pair Danggui-Honghua: Mechanisms Underlying Blood Stasis Syndrome by System Pharmacology Approach, *Sci. Rep.*, 2017, **7**, 40318.
- 31 M. U. Kakar, M. Naveed, M. Saeed, S. Zhao, M. Rasheed, S. Firdoos, R. Manzoor, Y. Deng and R. Dai, A Review On Structure, Extraction, and Biological Activities of Polysaccharides Isolated From *Cyclocarya Paliurus* (Batalin) Iljinskaja, *Int. J. Biol. Macromol.*, 2020, **156**, 420–429.
- 32 Z. Wu and T. Ding, *Seed Plants of China*, Kunming: Yunnan Science and Technology Press., 1999.
- 33 Z. Li, X. Ma, D. Wang, Y. Li, C. Wang and X. Jin, Evolution of Plastid Genomes of *Holcoglossum* (Orchidaceae) with Recent Radiation, *BMC Evol. Biol.*, 2019, **19**, 63.
- 34 J. Ru, P. Li, J. Wang, W. Zhou, B. Li, C. Huang, P. Li, Z. Guo, W. Tao, Y. Yang, X. Xu, Y. Li, Y. Wang and L. Yang, TCMSP: A Database of Systems Pharmacology for Drug Discovery From Herbal Medicines, *J. Cheminf.*, 2014, **6**, 13.
- 35 D. Gfeller, O. Michielin and V. Zoete, Shaping the Interaction Landscape of Bioactive Molecules, *Bioinformatics*, 2013, **29**, 3073–3079.
- 36 X. Liu, J. Wu, D. Zhang, K. Wang, X. Duan and X. Zhang, A Network Pharmacology Approach to Uncover the Multiple Mechanisms of *Hedyotis Diffusa* Willd. On Colorectal Cancer, *J. Cheminf.*, 2018, **2018**, 6517034.
- 37 C. von Mering, L. J. Jensen, B. Snel, S. D. Hooper, M. Krupp, M. Foglierini, N. Jouffre, M. A. Huynen and P. Bork, STRING: Known and Predicted Protein-Protein Associations, Integrated and Transferred Across Organisms, *Nucleic Acids Res.*, 2004, **33**, D433–D437.
- 38 R. Yu, L. Chen, R. Lan, R. Shen and P. Li, Computational Screening of Antagonists Against the SARS-CoV-2 (COVID-

- 19) Coronavirus by Molecular Docking, *Int. J. Antimicrob. Agents*, 2020, **56**, 106012.
- 39 X. G. Liu, M. C. Lv, M. Y. Huang, Y. Q. Sun, P. Y. Gao and D. Q. Li, A Network Pharmacology Study On the Triterpene Saponins from *Medicago Sativa* L. For the Treatment of Neurodegenerative Diseases, *J. Food Biochem.*, 2019, **43**, e12955.
- 40 J. Li, X. Ma, C. Liu, H. Li, J. Zhuang, C. Gao, C. Zhou, L. Liu, K. Wang and C. Sun, Exploring the Mechanism of Danshen against Myelofibrosis by Network Pharmacology and Molecular Docking, *Evid. base Compl. Alternative Med.*, 2018, **2018**, 1–11.
- 41 R. Liu, C. Su, Y. Xu, K. Shang, K. Sun, C. Li and J. Lu, Identifying Potential Active Components of Walnut Leaf that Action Diabetes Mellitus through Integration of UHPLC-Q-Orbitrap HRMS and Network Pharmacology Analysis, *J. Ethnopharmacol.*, 2020, **253**, 112659.
- 42 Q. Luo, X. Shi, J. Ding, Z. Ma, X. Chen, Y. Leng, X. Zhang and Y. Liu, Network Pharmacology Integrated Molecular Docking Reveals the Antiosteosarcoma Mechanism of *Biochanin A*, *Evid. base Compl. Alternative Med.*, 2019, **2019**, 1–10.
- 43 Y. Xiong, Y. Yang, W. Xiong, Y. Yao, H. Wu and M. Zhang, Network Pharmacology-Based Research On the Active Component and Mechanism of the Antihepatoma Effect of *Rubia Cordifolia* L., *J. Cell. Biochem.*, 2019, **120**, 12461–12472.
- 44 C. Mu, Y. Sheng, Q. Wang, A. Amin and Y. Xie, Potential Compound From Herbal Food of *Rhizoma Polygonati* for Treatment of COVID-19 Analyzed by Network Pharmacology and Molecular Docking Technology, *J. Funct. Foods*, 2020, 104149.
- 45 H. P. T. Ammon, Boswellic Extracts and 11-Keto- β -Boswellic Acids Prevent Type 1 and Type 2 Diabetes Mellitus by Suppressing the Expression of Proinflammatory Cytokines, *Phytomedicine*, 2019, **63**, 153002.
- 46 B. Cui and S. Li, New Triterpenoid Saponins From the Leaves of *Cyclocarya Paliurus*, *Chin. Chem. Lett.*, 2015, **26**, 585–589.
- 47 T. Honda, T. Murae, T. Tsuyuki, T. Takahashi and M. Sawai, Arjungenin, Arjunglucoside I, and Arjunglucoside II. A New Triterpene and New Triterpene Glucosides From *Terminalia Arjuna*, *Bull. Chem. Soc. Jpn.*, 1976, **49**, 3213–3218.
- 48 I. R. Mohanty, M. Borde, S. Kumar C and U. Maheshwari, Dipeptidyl Peptidase IV Inhibitory Activity of *Terminalia Arjuna* Attributes to its Cardioprotective Effects in Experimental Diabetes: In Silico, in Vitro and in Vivo Analyses, *Phytomedicine*, 2019, **57**, 158–165.
- 49 Z. Wu, F. Meng, L. Cao, C. Jiang, M. Zhao, X. Shang, S. Fang, W. Ye, Q. Zhang, J. Zhang and Z. Yin, Triterpenoids From *Cyclocarya Paliurus* and their Inhibitory Effect On the Secretion of Apolipoprotein B48 in Caco-2 Cells, *Phytochemistry*, 2017, **142**, 76–84.
- 50 C. Phillips, G. Murugasu, D. Owens, P. Collins, A. Johnson and G. H. Tomkin, Improved Metabolic Control Reduces the Number of Postprandial Apolipoprotein B-48-containing Particles in Type 2 Diabetes, *Atherosclerosis*, 2000, **148**, 283–291.
- 51 S. Xu, G. Wang, W. Peng, Y. Xu, Y. Zhang, Y. Ge, Y. Jing and Z. Gong, Corosolic Acid Isolated From *Eriobotrya Japonica* Leaves Reduces Glucose Level in Human Hepatocellular Carcinoma Cells, Zebrafish and Rats, *Sci. Rep.*, 2019, **9**, 4388.
- 52 J. Jiang, C. Yu, X. Guo, H. Zhang, S. Tian, K. Cai, Z. He and C. Sun, G Protein-Coupled Receptor GPR87 Promotes the Expansion of PDA Stem Cells through Activating JAK2/STAT3, *Mol. Ther.–Oncolytics*, 2020, **17**, 384–393.
- 53 S. J. Thomas, J. A. Snowden, M. P. Zeidler and S. J. Danson, The Role of JAK/STAT Signalling in the Pathogenesis, Prognosis and Treatment of Solid Tumours, *Br. J. Cancer*, 2015, **113**, 365–371.
- 54 J. Zhou, Y. Jiang, J. Zhao, H. Zhang, J. Fu, P. Luo, Y. Ma, D. Zou, H. Gao, J. Hu, Y. Zhang and Z. Jing, Dp44mT, an Iron Chelator, Suppresses Growth and Induces Apoptosis Via RORA-mediated NDRG2-IL6/JAK2/STAT3 Signaling in Glioma, *Cell. Oncol.*, 2020, **43**, 461–475.
- 55 G. L. Zhao, L. M. Yu, W. L. Gao, W. X. Duan, B. Jiang, X. D. Liu, B. Zhang, Z. H. Liu, M. E. Zhai, Z. X. Jin, S. Q. Yu and Y. Wang, Berberine Protects Rat Heart From Ischemia/Reperfusion Injury Via Activating JAK2/STAT3 Signaling and Attenuating Endoplasmic Reticulum Stress, *Acta Pharmacol. Sin.*, 2016, **37**, 354–367.
- 56 C. M. Taniguchi, B. Emanuelli and C. R. Kahn, Critical Nodes in Signalling Pathways: Insights Into Insulin Action, *Nat. Rev. Mol. Cell Biol.*, 2006, **7**, 85–96.
- 57 Y. Gao, M. Zhang, R. Zhang, L. You, T. Li and R. H. Liu, Whole Grain Brown Rice Extrudate Ameliorates the Symptoms of Diabetes by Activating the IRS1/PI3K/AKT Insulin Pathway in Db/Db Mice, *J. Agric. Food Chem.*, 2019, **67**, 11657–11664.
- 58 L. Rui, M. Yuan, D. Frantz, S. Shoelson and M. F. White, SOCS-1 and SOCS-3 Block Insulin Signaling by Ubiquitin-Mediated Degradation of IRS1 and IRS2, *J. Biol. Chem.*, 2002, **277**, 42394–42398.
- 59 A. L. Rodrigues, E. G. de Moura, M. C. F. Passos, I. H. Trevenzoli, E. P. S. da Coceição, I. T. Bonono, J. F. N. Neto and P. C. Lisboa, Postnatal Early Overfeeding Induces Hypothalamic Higher SOCS3 Expression and Lower STAT3 Activity in Adult Rats, *J. Nutr. Biochem.*, 2011, **22**, 109–117.
- 60 G. J. Morton and M. W. Schwartz, Leptin and the Central Nervous System Control of Glucose Metabolism, *Physiol. Rev.*, 2011, **91**, 389–411.
- 61 L. Cheng, Y. Yu, A. Szabo, Y. Wu, H. Wang, D. Camer and X. Huang, Palmitic Acid Induces Central Leptin Resistance and Impairs Hepatic Glucose and Lipid Metabolism in Male Mice, *J. Nutr. Biochem.*, 2015, **26**, 541–548.
- 62 C. E. Nunes-Xavier, O. Aurtenetxe, L. Zaldumbide, R. López-Almaraz, A. Erramuzpe, J. M. Cortés, J. I. López and R. Pulido, Protein Tyrosine Phosphatase PTPN1 Modulates Cell Growth and Associates with Poor Outcome in Human Neuroblastoma, *Diagn. Pathol.*, 2019, **14**, 134.
- 63 J. Xu, L. Li, Z. Qian, J. Hong, S. Shen and W. Huang, Reduction of PTP1B by RNAi Upregulates the Activity of Insulin Controlled Fatty Acid Synthase Promoter, *Biochem. Biophys. Res. Commun.*, 2005, **329**, 538–543.
- 64 S. Legeay, P. Fautrat, J. B. Norman, G. Antonova, S. Kennard, T. Bruder-Nascimento, V. S. Patel, S. Faure and E. J. B. de



- Chantemèle, Selective Deficiency in Endothelial PTP1B Protects From Diabetes and Endoplasmic Reticulum Stress-Associated Endothelial Dysfunction Via Preventing Endothelial Cell Apoptosis, *Biomed. Pharmacother.*, 2020, **127**, 110200.
- 65 C. M. Rondinone, J. M. Trevillyan, J. Clampit, R. J. Gum, C. Berg, P. Kroeger, L. Frost, B. A. Zinker, R. Reilly, R. Ulrich, M. Butler, B. P. Monia, M. R. Jirousek and J. F. Waring, Protein Tyrosine Phosphatase 1B Reduction Regulates Adiposity and Expression of Genes Involved in Lipogenesis, *Diabetes*, 2002, **51**, 2405–2411.
- 66 A. Janus, E. Szahidewicz-Krupska, G. Mazur and A. Doroszko, Insulin Resistance and Endothelial Dysfunction Constitute a Common Therapeutic Target in Cardiometabolic Disorders, *Mediators Inflammation*, 2016, **2016**, 3634948.
- 67 D. Leto and A. R. Saltiel, Regulation of Glucose Transport by Insulin: Traffic Control of GLUT4, *Nat. Rev. Mol. Cell Biol.*, 2012, **13**, 383–396.
- 68 P. R. Shepherd and B. B. Kahn, Glucose Transporters and Insulin Action-Implications for Insulin Resistance and Diabetes Mellitus, *N. Engl. J. Med.*, 1999, **341**, 248–257.
- 69 S. Polsky and S. L. Ellis, Obesity, Insulin Resistance, and Type 1 Diabetes Mellitus, *Curr. Opin. Endocrinol. Diabetes Obes.*, 2015, **22**, 277.
- 70 Y. Gao, M. Zhang, R. Zhang, L. You, T. Li and R. H. Liu, Whole Grain Brown Rice Extrudate Ameliorates the Symptoms of Diabetes by Activating the IRS1/PI3K/AKT Insulin Pathway in Db/Db Mice, *J. Agric. Food Chem.*, 2019, **67**, 11657–11664.
- 71 E. S. Popravka, N. S. Linkova, S. V. Trofimova and V. K. Khavinson, HIF-1 as a Marker of Age-Related Diseases Associated with Tissue Hypoxia, *Biol. Bull.*, 2018, **8**, 497–508.
- 72 M. T. Coughlan, A. L. Mibus and J. M. Forbes, Oxidative Stress and Advanced Glycation in Diabetic Nephropathy, *Ann. N. Y. Acad. Sci.*, 2008, **1126**, 190–193.
- 73 L. Nordquist, M. Friederich-Persson, A. Fasching, P. Liss, K. Shoji, M. Nangaku, P. Hansell and F. Palm, Activation of Hypoxia-Inducible Factors Prevents Diabetic Nephropathy, *J. Am. Soc. Nephrol.*, 2015, **26**, 328.
- 74 C. Rosenberger, M. Khamaisi, Z. Abassi, V. Shilo, S. Zangen, M. Goldfarb, A. Shina, F. Zibertrest, K. Eckardt, S. Rosen and S. Heyman, Adaptation to Hypoxia in the Diabetic Rat Kidney, *Kidney Int.*, 2008, **73**, 34–42.
- 75 R. U. Ostrovskaya, S. V. Ivanov, T. A. Gudasheva and S. B. Seredenin, A Novel Dipeptide NGF Mimetic GK-2 Selectively Activating the PI3K/AKT Signaling Pathway Promotes the Survival of Pancreatic β -Cells in a Rat Model of Diabetes, *Acta Naturae*, 2019, **11**, 48–57.
- 76 X. Li, H. Liu, J. Wang, J. Qin, Z. Bai, B. Chi, W. Yan and X. Chen, Curcumol Induces Cell Cycle Arrest and Apoptosis by Inhibiting IGF-1R/PI3K/Akt Signaling Pathway in Human Nasopharyngeal Carcinoma CNE-2 Cells, *Phytother. Res.*, 2018, **32**, 2214–2225.
- 77 J. F. Wojtaszewski, B. F. Hansen, B. Kiens and E. A. Richter, Insulin Signaling in Human Skeletal Muscle: Time Course and Effect of Exercise, *Diabetes*, 1997, **46**, 1775–1781.
- 78 L. R. Pearce, D. Komander and D. R. Alessi, The Nuts and Bolts of AGC Protein Kinases, *Nat. Rev. Mol. Cell Biol.*, 2010, **11**, 9–22.
- 79 G. Xi, X. Shen, C. Wai, M. White and D. Clemmons, Hyperglycemia Induces Vascular Smooth Muscle Cell Dedifferentiation by Suppressing Insulin Receptor Substrate-1-Mediated p53/KLF4 Complex Stabilization, *J. Biol. Chem.*, 2018, **294**, A118–A5398.
- 80 C. Kim, J. Lee, M. Kim and J. Hwang, Hypoglycemic Effect of Whole Grain Diet in C57BL/KsJ-db/db Mice by Activating PI3K/Akt and AMPK Pathways, *J. Food Sci. Biotechnol.*, 2019, **28**, 895–905.
- 81 W. Feng, H. Ao, S. Yue and C. Peng, Systems Pharmacology Reveals the Unique Mechanism Features of Shenzhu Capsule for Treatment of Ulcerative Colitis in Comparison with Synthetic Drugs, *Sci. Rep.*, 2018, **8**, 16160.

