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Emma L. Schymanski *et al.*

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Connecting environmental exposure and neurodegeneration using cheminformatics and high resolution mass spectrometry: potential and challenges†

Emma L. Schymanski,^{ID}*^a Nancy C. Baker,^{ID}^b Antony J. Williams,^{ID}^c Randolph R. Singh,^{ID}^{ad} Jean-Pierre Trezzi,^{ID}^{ef} Paul Wilmes,^{ID}^f Pierre L. Kolber,^{gh} Rejko Kruger,^{ID}^{gh} Nicole Paczia,ⁱ Carole L. Linster,^{ID}ⁱ and Rudi Balling,^{ID}^j

Connecting chemical exposures over a lifetime to complex chronic diseases with multifactorial causes such as neurodegenerative diseases is an immense challenge requiring a long-term, interdisciplinary approach. Rapid developments in analytical and data technologies, such as non-target high resolution mass spectrometry (NT-HR-MS), have opened up new possibilities to accomplish this, inconceivable 20 years ago. While NT-HR-MS is being applied to increasingly complex research questions, there are still many unidentified chemicals and uncertainties in linking exposures to human health outcomes and environmental impacts. In this perspective, we explore the possibilities and challenges involved in using cheminformatics and NT-HR-MS to answer complex questions that cross many scientific disciplines, taking the identification of potential (small molecule) neurotoxins in environmental or biological matrices as a case study. We explore capturing literature knowledge and patient exposure information in a form amenable to high-throughput data mining, and the related cheminformatic challenges. We then briefly cover which sample matrices are available, which method(s) could potentially be used to detect these chemicals in various matrices and what remains beyond the reach of NT-HR-MS. We touch on the potential for biological validation systems to contribute to mechanistic understanding of observations and explore which sampling and data archiving strategies may be required to form an accurate, sustained picture of small molecule signatures on extensive cohorts of patients with chronic neurodegenerative disorders. Finally, we reflect on how NT-HR-MS can support unravelling the contribution of the environment to complex diseases.

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Environmental significance

Non-target high resolution mass spectrometry has attracted immense interest regarding the potential for increased characterisation of chemicals and exposures in environmental studies. However, high quality studies remain difficult to perform and often yield fewer successful identifications than desired. We present our multi-disciplinary perspective on tackling potential causes for this discrepancy and the complexity involved in capturing the knowledge needed to investigate the impact of chemicals on human health in the context of long-term diseases. We look at how to mine patient and expert knowledge to find, measure and validate potential neurotoxins and explore the potential for NT-HR-MS to provide small molecule data signatures in cohorts and build the knowledge required for greater future understanding of environmental impacts on complex diseases.

^aEnvironmental Cheminformatics Group, Luxembourg Centre for Systems Biomedicine (LCSB), University of Luxembourg, 6 Avenue du Swing, L-4367 Belvaux, Luxembourg. E-mail: emma.schymanski@uni.lu

^bLeidos, Research Triangle Park, North Carolina, USA

^cNational Centre for Computational Toxicity (NCCT), United States Environmental Protection Agency, Research Triangle Park, North Carolina, USA

^dOak Ridge Institute for Science and Education Research Fellow, United States Environmental Protection Agency, Research Triangle Park, North Carolina, USA

^eIntegrated Biobank of Luxembourg, Luxembourg Institute of Health, 1 rue Louis Rech, L-3555 Dudelange, Luxembourg

^fEco-Systems Biology Group, Luxembourg Centre for Systems Biomedicine (LCSB), University of Luxembourg, 6 Avenue du Swing, L-4367 Belvaux, Luxembourg

^gClinical and Experimental Neuroscience Group, Luxembourg Centre for Systems Biomedicine (LCSB), University of Luxembourg, 6 Avenue du Swing, L-4367 Belvaux, Luxembourg

^hNeurology, Centre Hospitalier de Luxembourg, Luxembourg City, Luxembourg

ⁱEnzymology and Metabolism Group, Luxembourg Centre for Systems Biomedicine (LCSB), University of Luxembourg, 6 Avenue du Swing, L-4367 Belvaux, Luxembourg

^jSystems Biomedicine, Luxembourg Centre for Systems Biomedicine (LCSB), University of Luxembourg, 6 Avenue du Swing, L-4367 Belvaux, Luxembourg

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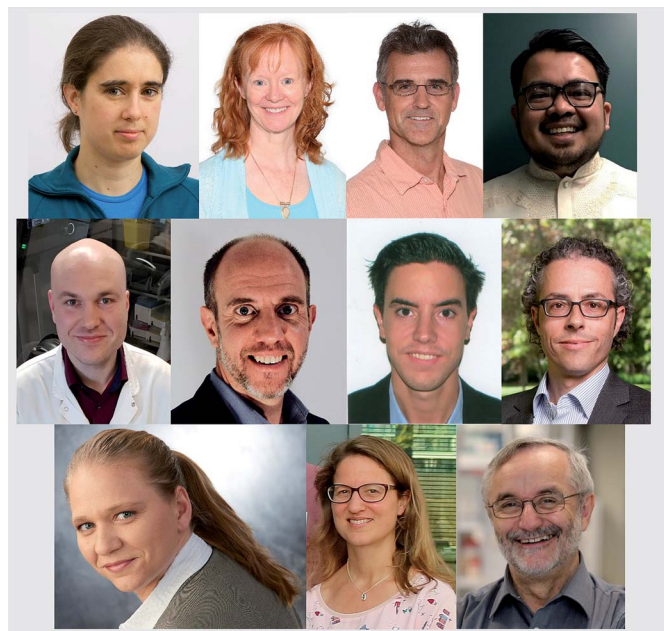
Introduction

Rapid developments in analytical techniques over the last decades have opened up new opportunities such as non-target high resolution mass spectrometry (NT-HR-MS) to explore the impact of humans on the environment,¹ as well as the effect of environmental contamination on humans^{2,3} and other organisms.^{4,5} With growing accessibility and maturity in NT-HR-MS analytical methods, these are being applied to increasingly

complex research questions and hypotheses.^{6–8} In this perspective article, we take a close look at the challenges in identifying “neurotoxicants” in the context of chronic neuro-degenerative diseases and use this case study to explore the potential and limitations of NT-HR-MS to support answering these questions. Ultimately, solving complex challenges such as these will need a systems approach,^{9,10} requiring knowledge that crosses many disciplines. While the main focus of this article, determining which (relevant) chemicals are present in environmental or biological samples (let alone their mode of action), is already a daunting task, it is a necessary foundation to enable the elucidation of functional effects and generation of hypotheses for a greater understanding of the causes of complex diseases and environmental impacts.

NT-HR-MS, typically coupled with separation techniques such as liquid or gas chromatography, is becoming an increasingly popular approach for the broad screening of complex environmental samples.¹ NT-HR-MS can be performed on low sample amounts and reach ppb (or lower) detection limits in many cases, with data acquisition now often able to capture both known and unknown chemicals in a single measurement.¹¹ Ideally in NT-HR-MS, post-acquisition data processing is used to identify (1) the known chemicals, *i.e.* the “targets” or also “biomarkers” in the context of metabolomics and disease; (2) potential chemicals of interest based on prior knowledge, *i.e.* lists of chemicals (“suspects”) and (3) the relevant “unknowns” or “non-targets” using some form of prioritisation.¹ Data generated in NT-HR-MS can be archived for later use and exploration (also termed retrospective screening).¹² While NT-HR-MS has been declared “ready to go”,¹ in this article we explore how to gather the knowledge required to take NT-HR-MS to the next level of helping unravel the cause and mechanisms of environmental exposure in the context of late-developing complex diseases, where a lifetime of exposure to thousands of chemicals may potentially impact disease progression. Forming epidemiological connections, however, does not just involve the detection of chemicals of interest, but also gathering information on qualitative aspects and characteristics of potential neurotoxicants, as well as time and duration of exposure.

To date, NT-HR-MS of environmental samples has been commonly directed by substance classes. Several examples include using suspect screening approaches to investigate classes such as pharmaceuticals,^{13,14} pesticides^{15,16} or antibiotics.^{17,18} Further studies are now using publicly-available suspect lists in their screening efforts.^{19,20} Other early efforts include comprehensive, exploratory characterisation efforts of wastewater that went beyond compound classes.^{21,22} As NT-HR-MS matures, effect-directed analysis (EDA) studies are beginning to show some level of success with established bioassay endpoints such as mutagenicity.²³ The growing popularity of “big data” techniques has seen the expansion into more data exploration-based methods such as virtual EDA, clustering, trend analysis and fingerprinting, summarized elsewhere,¹ inspired partially by metabolomics efforts. From a metabolomics perspective, great investments have been made into the detection of biomarkers for certain diseases (discussed further



Top: E. L. Schymanski, N. C. Baker, A. J. Williams,
R. R. Singh Middle: J.-P. Trezzi, P. Wilmes, P. L. Kolber,
R. Krüger Bottom: N. Paczia, C. L. Linster, R. Balling

Associate Professor Emma Schymanski is a Luxembourg National Research Fund (FNR) ATTRACT Fellow and head of the Environmental Cheminformatics group at the Luxembourg Centre for Systems Biomedicine (LCSB), University of Luxembourg. Dr Randolph Singh is a postdoctoral fellow in the same group. Dr Nancy Baker is a consultant at Leidos, Research Triangle Park, USA. Dr Antony Williams is a Computational Chemist, National Center of Computational Toxicology, US EPA. Dr Jean-Pierre Trezzi is postdoctoral fellow affiliated with the Integrated Biobank of Luxembourg, Luxembourg Institute of Health as well as the Eco-Systems Biology group at LCSB, which is headed by Associate Professor Paul Wilmes. Dr med. Pierre Kolber, MD is a member of Neurology department at the Centre Hospitalier de Luxembourg and the Clinical and Experimental Neuroscience Group at LCSB, which is headed by Professor Dr med. Rejko Krüger, also an FNR PEARL Fellow and Director of Transversal Translational Medicine at the Luxembourg Institute of Health. Dr Nicole Paczia, former joint-manager of the Metabolomics Platform at LCSB, now heads the Metabolomics Core facility at the Max Planck Institute in Marburg. Dr Carole Linster is head of the Enzymology and Metabolism Group, LCSB. Professor Rudi Balling is Director of the LCSB.



As mentioned above, the key to successful NT-HR-MS is finding chemicals of high relevance to the study question in an efficient manner (prioritisation). A well-designed suspect screening, *i.e.* searching for a discrete list of chemicals relevant to the study question, is an ideal way to quickly find masses (features) and thus candidates of particular interest. A number of lists of neurotoxicants were compiled for the purposes of this perspective, summarized in Table 1 and described further in the ESI.[†] These include small, carefully validated lists explained extensively in the source publications (DNTEFFECTS⁴⁹ (https://comptox.epa.gov/dashboard/chemical_lists/dnteffects), DNTINVIVO⁵⁰ (https://comptox.epa.gov/dashboard/chemical_lists/dntinvivo), and the European Union Reference List (EURL) for the identification of neurotoxic substances (EURL-INTOX) (https://comptox.epa.gov/dashboard/chemical_lists/eurl-intox)).

comptox.epa.gov/dashboard/chemical_lists/dntinvivo) and HUMANNEUROTOX⁵¹ (https://comptox.epa.gov/dashboard/chemical_lists/humanneurotox), where few suspect hits will ever be found in environmental samples. The DNTPOTNEG⁵⁰ (https://comptox.epa.gov/dashboard/chemical_lists/dntpoteq) list contains potential negative controls for neurotoxicity which, if found, should not be associated with neurotoxic effects, enabling a second layer of data quality verification of neurotoxicant findings. Two larger lists (NEUROTOXINS⁵² (https://comptox.epa.gov/dashboard/chemical_lists/neurotoxins) and LITMINEDNEURO^{53,54} (https://comptox.epa.gov/dashboard/chemical_lists/litminedneuro)) have also been compiled for this perspective. The NEUROTOXINS list was compiled from public resources including ChEBI,^{55,56} Wikipedia,^{57,58} T3DB,⁴⁰ with further details given in the ESI.†

LITMINEDNEURO contains chemicals associated with neurotoxicity compiled through systematic literature mining of PubMed⁵⁹ using Medical Subject Heading (MeSH) terms⁶⁰ and associating these with single chemical substances (where possible) using previously published methods.⁶¹ Articles were identified in which a nervous system disease was annotated with the MeSH node C10 through the MeSH tree with disease subheading “chemically induced” and subheading “toxicity”, “poisoning”, or “adverse effects”. Nerve diseases caused by trauma and manually identified “common English terms” that could not be associated with any specific chemicals (*e.g.* “particulate matter”, “contrast media”) were omitted. In total 4528 chemicals were identified; all chemicals with 5 or more literature references were registered in the Dashboard and included in the final list. The output of this processing was exported to Microsoft® Excel and is included as ESI† and available on FigShare.⁵³ The CASRN and the CompTox Chemicals Dashboard substance identifier (DTXSID) were included in the spreadsheet for chemicals for which this relationship was captured and associated with MeSH identifiers. The

overview tab of this workbook contains 1250 chemicals (1243 unique DTXSIDs) and the co-annotations with 554 nervous system diseases in over 53 000 chemical-disease pairs (“Detail” tab). These relationships were described in 38 192 articles. A batch search of the Dashboard by DTXSID will return all related chemical information needed for generating suspect lists with subject-specific reference scores for disease or effect subsets of this list.

Automated text-mining techniques such as those described here have the advantage of being easy to run. Since they encompass the large and fast-growing PubMed corpus (28 million citations as of October 2018), the approach can identify chemicals that may cause disease and which may not have yet achieved visibility through other means. On the other hand, data extracted through automated text-mining has also not passed through rigorous manual vetting and is likely to contain various types of errors. For instance, some articles discuss more than one chemical and more than one disease, while the algorithms that associate each disease with each chemical may not construct valid pairs. Typing errors and ambiguous synonyms in the original literature may confuse proper mapping of the chemical identities. Additionally, the MeSH annotations do not capture negative results (a chemical not causing a particular neurotoxic effect) and therefore it can be difficult to disambiguate positive from negative reports. For instance, caffeine (<https://comptox.epa.gov/dashboard/dsstoxdb/results?search=DTXSID0020232&abbreviation=DNT EFFECTS>) is documented with data demonstrating effects on neurodevelopment, yet potentially neuroprotective in the context of Parkinson's disease (PD);⁷⁰⁻⁷² this can be traced *via* the Excel macro provided in the ESI† in the entry for caffeine. Likewise nicotine is documented in all the neurotoxicant lists in Table 1 (except the negative control list DNTPOTNEG (https://comptox.epa.gov/dashboard/chemical_lists/dntpoteq)), yet smoking may be protective in some cases in the context of

Table 1 A range of lists for performing suspect screening of neurotoxicants, compiled for this perspective. These lists are also available as a single collection on Zenodo⁶²

List code	Entries and references	Description
DNTEFFECTS (https://comptox.epa.gov/dashboard/chemical_lists/dnteffects)	96 (ref. 49 and 63)	Chemicals with data demonstrating effects on neurodevelopment
DNTINVIVO (https://comptox.epa.gov/dashboard/chemical_lists/dntinvivo)	33 (ref. 50 and 64)	A (non-exhaustive) list of compounds documented to trigger developmental neurotoxicity (DNT) in at least two different laboratories
DNTPOTNEG (https://comptox.epa.gov/dashboard/chemical_lists/dntpoteq)	41 (ref. 50 and 65)	Suggested potential negative controls for developmental neurotoxicity (DNT) assays. Statins can also be used, see https://comptox.epa.gov/dashboard/chemical_lists/statins
HUMANNEUROTOX (https://comptox.epa.gov/dashboard/chemical_lists/humanneurotox)	190 (ref. 51, 66 and 67)	A set of chemicals identified as <i>potential</i> neurotoxicants by the authors using literature searching, not necessarily active neurotoxicants
NEUROTOXINS (https://comptox.epa.gov/dashboard/chemical_lists/neurotoxins)	511 (ref. 52, 62 and 68)	A list of chemicals reported as neurotoxicants, compiled from public resources (source file with details on Zenodo ⁶²)
LITMINEDNEURO (https://comptox.epa.gov/dashboard/chemical_lists/litminedneuro)	1243 (ref. 53, 54 and 69)	Chemicals associated with neurotoxicity compiled through automated literature mining of PubMed using MeSH terms (node C10, subheadings “chemically induced”, “toxicity”, “poisoning”, or “adverse effects”) and associating these with single chemical substances

Parkinson's disease. For example, non-smoking carriers of the LRRK2 Gly2385Arg gene have increased risk of developing PD;⁷³ while another study (published in German) discusses the balance between neurotoxic and neuroprotective effects of nicotine and smoking.⁷⁴ Understanding of the context is critical and the interface through the Excel file provided as ESI† is one method to follow-up the literature references for specific chemicals and understand the relevance of the suspect for their particular study question (see examples for nicotine and caffeine above).

Related structures: mapping “chemicals” and “substances”

Reviewing some of the lists in Table 1 *via* the Dashboard reveals several interesting cases where new cheminformatics approaches are needed and already under development to

capture less well-defined substance information in the form of discrete chemical structures suitable for NT-HR-MS screening studies. Several examples are given in Fig. 1 and more extensive details given in the ESI.†

One of the most interesting challenges associated with capturing chemical information for NT-HR-MS related to neurotoxicity (or any potential health impact) is relating what we as humans consume relative to discrete chemical components. For instance, while toxicity testing is most commonly performed on a chemical such as caffeine, patients will instead report coffee (or tea or energy drink) consumption. Coffee, for instance, is documented to contain over 1000 chemical constituents,⁷⁶ which require a variety of analytical techniques for detection.⁷⁷ Nicotine is another common example where the chemical tested is often nicotine (or an associated salt or mixture), but patients will instead report *e.g.* smoking habits. It



Fig. 1 Top: Nicotine (red box) (https://comptox.epa.gov/dashboard/dsstoxdb/mixture_search?cid=930) and selected mixtures, salts and components. Middle: Diazinon (red box) (<https://comptox.epa.gov/dashboard/dsstoxdb/results?search=DTXSID9020407&abbreviation=HUMANNEUROTOX#related-substances>) and selected mapped transformation products. Bottom row: The PCB class (red box) (<https://comptox.epa.gov/dashboard/dsstoxdb/results?search=PCBs#related-substances>) and selected mapped individual PCB compounds. While the generic PCB representation is suboptimal visually, it allows a full, automated enumeration of all members of the PCB class *via* the ChemAxon Markush Enumeration module to create “Markush child” entries (see label) – an important consideration for upscaling these cheminformatics efforts. All chemicals display the structure, DTXSID, number of assays (active/total) in TOXCAST⁷⁵ (https://comptox.epa.gov/dashboard/chemical_lists/toxcast) and the data sources.



is well known that nicotine is not the only chemical in cigarettes that may cause detrimental health effects. Several thousand chemicals have been identified in cigarettes^{78,79} with 599 chemicals listed as additives.⁸⁰ Capturing such knowledge (*e.g.* *via* cross-mapping and adding as lists or related substances in databases) will be increasingly important to help reconcile NT-HR-MS results in the future, yet expand suspect lists even further. The connection of individual chemical structures to mixtures⁴³ and the associated metadata can enable high throughput screening of NT-HR-MS data, including *e.g.* toxicity and product information,⁸¹ which are often associated with mixtures. It also enables better data interpretation downstream. Furthermore, the collection of chemicals into lists combined with well-selected metadata can ensure rapid prioritization by score and can help rapidly pinpoint highly promising candidates amongst hundreds to thousands of possible masses and chemical structures, as demonstrated in Fig. 2. More specific details about how such information can be included in NT-HR-MS studies are given in the ESI.†

Capturing patient and medical knowledge

While the preceding sections discuss capturing and exploring the documented chemical and medical knowledge in the context of neurotoxicity and using this during identification in NT-HR-MS, capturing the patient knowledge is an incredibly important part of connecting chemicals to disease. For instance, neurodegeneration in chronic diseases such as Parkinson's disease takes place for many years before the first motor manifestations appear that define the clinical diagnosis of this movement disorder.⁸³ The time span between the diagnosis of PD and the appearance of the initial non-motor symptoms such as depression, REM sleep behavior disorder, hyposmia or chronic constipation (defining the prodromal phase of PD), can range from approximately 5 up to 30 years. Even before this prodromal phase of the disease, neuronal dysfunction and neuronal cell death are already ongoing and might precede the prodromal phase by many years or even

decades.⁸⁴ Thus, in order to analyze the potential neurotoxicants and environmental factors that might lead to neurodegenerative diseases, one has to retrace the exposure of patients long before the diagnosis of the disease, better still before the first neuronal dysfunctions appear. Furthermore, as discussed below, there may no longer be any traces of the chemical to which the patient was exposed in cohort samples and thus the patient's memory of their chemical exposures may be the only documentation that could provide clues to potential causative agents. In the few longitudinal PD patient cohorts that exist, such as the Luxembourg Parkinson's study,⁸⁵ the first step in analyzing the environmental risk factors leading to neurodegeneration is by using detailed validated patient questionnaires retrospectively trying to trace the patient's professional or leisure activities throughout their entire life (see Fig. 3 for a screenshot from the REDCap⁸⁶ system (<https://www.project-redcap.org/>)).

These classical epidemiological strategies apply self-reporting questionnaires that are not without flaws, as there is a non-negligible recall bias, especially considering the fact that patients with neurodegenerative diseases are mainly elderly people and to some degree affected by more or less pronounced cognitive defects. Additional confusion can occur due to misunderstanding of the terminology, for instance weed killers approved for household use are not always understood as a herbicide by patients and thus pesticide/herbicide exposure might be neglected by patients in some questionnaires. Nevertheless, these questionnaires can provide first hints towards potential environmental risk factors using the knowledge of the patients themselves. Anecdotal cases such as the one mentioned above are highly subjective and difficult to avoid with a generic survey form as in Fig. 3. However, the association of chemicals with products (*e.g.* *via* synonym linking such as aspirin/acetylsalicylic acid (<https://comptox.epa.gov/dashboard/dsstoxdb/results?search=aspirin>), plus ingredients listing such as the cigarette example above) and products with exposure scores, such as CPDat,⁸¹ may help connect what patients report and the information required for chemical analysis. Obtaining chemical and approved product information from authorities and linking this to *e.g.* agricultural land use can support



Fig. 2 An example of how metadata can help candidate selection in high throughput NT-HR-MS studies, using nicotine in MetFrag.⁸² Further details in the ESI.†



Residential history, age 18 through age 25	
B1) From age 18 through age 25, where did you live for the longest time? List: City/Town, State, Zip Code, Country	<input type="text"/> <small>This field is retired, use the next 3 questions below instead</small>
B1b) From age 18 through age 25, in what country did you live for the longest time?	<input type="text"/>
B1c) What city/town?	<input type="text"/>
B1d) What zip/postal code?	<input type="text"/>
B2) At the time you lived there, was this residence located in a ...	<input type="text"/>
B3) Was your main source of drinking water at this residence a private well?	<input type="text"/>
B4) Was this residence located near farm fields (within 0.25 mile)?	<input type="text"/>
B5) Was there pesticide spraying at or around this residence when you lived there?	<input type="text"/>
B5a) How often did the spraying happen?	<input type="text"/>
Residential history, age 26 through age 35	
C1) From age 26 through age 35, where did you live for the longest time? List: City/Town, State, Zip Code, Country	<input type="text"/> <small>This field is retired, use the next 3 questions below instead</small>
C1b) From age 26 through age 35, in what country did you live for the longest time?	<input type="text"/>
C1c) What city/town?	<input type="text"/>
C1d) What zip/postal code?	<input type="text"/>

Fig. 3 Screenshot of the "Tanner survey" used to capture patient information, using REDCap (<https://www.project-redcap.org/>).^{86,87}

epidemiological studies^{87,88} as well as ecological assessments.⁸⁹ Product information from authorities, while often kept confidential, is essential for improved connections between NT-HR-MS and epidemiology. As such, greater efforts to share this information in the public domain are desperately needed and several authorities are now supportive of information sharing – as evidenced for example by the many lists now available on the CompTox Chemicals Dashboard⁹⁰ and the NORMAN Suspect Exchange.^{41,42}

Using current knowledge, one can hypothesise that neurotoxicant exposure might precede neurodegenerative diseases such as PD by many decades, potentially during young adulthood or childhood (or even *in utero*), launching a cascade of malfunctions progressively leading to apoptosis of dopaminergic neurons. This makes finding a representative sample from patients very challenging, as ideally the sample used should contain the neurotoxicants from that initial exposure period, which will also potentially transform over time. See discussion in the next section. Considering PD specifically, the risk for PD due to the usage of hydrocarbon-based compounds such as pesticides, herbicides or fungicides is quite established by now, especially professional pesticide usage.^{91,92}

However, metal-based compounds are also suspected to contribute to an increased risk for PD or atypical parkinsonian syndromes (aPS), such as progressive supranuclear palsy (PSP) – a rapidly progressive aPS with prominent falls and gaze palsy as major clinical hallmarks. Some studies hypothesise a role of toxic metals and metalloids (such as arsenic) in the pathogenesis of PSP.⁹³ Exposure to heavy metals (such as lead) is suggested to be associated with an increased risk for PD. However, difficulties in making differential diagnoses between these disorders can confound these conclusions. A further factor to consider is epigenetics, where epigenetic imprints may reflect exposure in the preceding generation and could add some prior knowledge with respect to the effects observed, if not the actual chemical exposure itself.⁹⁴ Thus, while surveys can help capture patient knowledge, in the days of "big data" and with the trend towards personalised medicine, it is important to consider how "on the spot" analysis and digital archiving of small molecule data during cohort sample collection may enable a long term generation of a baseline dataset for greater understanding of disease progression and diagnosis in the future.



When studying disease-related neurotoxicant exposure, human-derived samples are of primary interest as these depict the pathological changes and can be monitored over time. However, not all of these matrices can be obtained easily and depending on the matrix used, the results can vary significantly. For instance, traces of many compounds are very short-lived in complex matrices like blood, as they are quickly metabolised and excreted by the kidneys. If one analyzes the urine, the window of detection can be delayed (the time for the compound to be filtered out from the blood by the kidneys), but is equally short-lived in the context of long-term diseases. The individual's kidney function, hydration status and metabolism complicate this aspect, as the amount of urine and the frequency of the mictions vary hugely between each individual patient. Not only does the time window of detection vary, but also the concentration of the compound detected in the matrix can vary hugely from one time period to another and if the peak of concentration is missed, the results can become insignificant. Additionally, if one analyzes, for example, the total 24 h diuresis in an attempt not to miss the detection window, the compound can become so diluted that it can falsely render the exposure as insignificant. Furthermore, for those substances where the metabolites are not known or documented, the analyst will have to use additional experiments and data processing approaches to identify potentially relevant metabolites. An impressive recent effort shows the possibility for NT-HR-MS and advanced data processing to provide much broader coverage than the typical targeted urine analysis.⁹⁵ Yet another very recent effort reports initial developments towards non-invasive drug monitoring *via* NT-HR-MS analysis of skin.⁹⁶ However, the transient nature of some compounds and matrices has to be kept in mind while analysing potential neurotoxicants in human samples. Samples containing the compounds for a longer period of time seem to be, in this regard, more valuable. Bone is an interesting matrix for metals exposure and, for example, lead concentration can be measured with ¹⁰⁹Cd-excited K-shell X-ray fluorescence, representing a biomarker of cumulative lead exposure.⁹⁷ Teeth have also been used to estimate long-term exposure to metals and selected organic compounds, with especially recent studies drawing conclusions for various aspects of neurotoxicity.^{98–101} Hair sampling is a relatively non-invasive longer-term sample, due to the continual release of fat-soluble contaminants that have been proven to provide biomarkers for monitoring human exposures to environmental chemicals such as pesticides and metals.¹⁰² In addition, information obtained from hair analysis is more representative of an individual's level of exposure than other matrices such as urine and blood.¹⁰³ As many (easily available) biological matrices are so transient, linking chemical exposure to affected pathways with *e.g.* metabolomics approaches will be increasingly relevant.^{2,3} As recent research has indicated a potential connection between the gut microbiome and disease progression (see text below), there is also increasing

Apart from human-derived samples, environmental samples can also be studied for the presence of neurotoxicants as surrogates for human samples, knowing that biological matrices can be challenging to access. Wastewater samples, for example, may reveal group exposure to specific contaminants as this matrix is a collection from a population and presents an idea of what the bulk population of a specific area is currently (or was) exposed to.¹¹ As wastewater is released back into the environment after treatment, studying local fish, crustacea or even resting eggs¹⁰⁴ may show which specific neurotoxicants have been present over a longer time frame and have bio-accumulative potential. Sediments and soil cores, on the other hand, may give an idea of historical exposure that may have happened over the last few years or decades,¹⁰⁵ although degradation and transformation must be taken into account. Even groundwater transects can provide interesting historical trends.⁷ Such samples will generally reveal only population trends. Dust samples are typically more representative of individual indoor exposure, with many targeted studies now showing health-related conclusions.¹⁰⁶ Increasing numbers of non-target studies are being published, including several collaborative and ring trials,^{29,107} showing this to be a matrix of great interest for future efforts, although high quality data processing is still challenging for routine use.

The quality of the samples of interest, and thus downstream analysis and data analysis, is highly impacted by pre-analytical variations. Ideally, according to biobank protocols, tissue samples should be snap-frozen in liquid nitrogen immediately after sampling. However, for logistical reasons, it is often not feasible to respect these recommendations. Thereby, it is crucial to monitor the pre-analytical variations that can occur after sampling, such as the time between the sampling and the processing of the sample, temperature, pH among others. Especially for biomarker discovery/validation studies, it is crucial to monitor these changes. In this regard, biobanks have established a standard pre-analytical coding for biospecimens to enable standardised documentation of the sample collection and processing to limit pre-analytical variations.¹⁰⁸ This becomes crucial in multi-center collection studies. In general, biobanks have stringent protocols for sample preservation and storage, together with strict quality controls. Biobanks typically store biospecimens derived from human individuals, such as blood derivatives (plasma, serum), cerebrospinal fluid, urine, saliva, stool and cells,¹⁰⁸ and are all potential matrices for neurotoxicant analyses depending on the study interest. Together with the samples, metadata on medical variables, dietary information, as well as environmental data are collected and made available for research. In terms of creating a digital “snapshot” of small molecule conditions, analysis as close to time of receipt will be a distinct advantage to prevent sample degradation, with the downside of creating large variabilities and batch effects in downstream data analysis. However, with sufficient quality controls and strategies to best capture batch effects to allow correction (standard measurements, internal

standards, pooled samples), sophisticated data processing techniques are now available to perform such corrections.

Analysing the chemical space of neurotoxicants

Chemical properties and analytical methods

Not all analytical methods are equally efficient and, as evidenced by the variety in the physico-chemical properties of potential neurotoxicants (see Fig. 4), it is not realistic to expect a “one size fits all” analytical method to perform a full screen of all potential neurotoxicants in a given matrix. A detailed description of the list and the corresponding diversity in physico-chemical properties can be found in the ESI.† Already the choice of solvents during extraction will have an impact on the chemical nature of the analytes to be found in the data acquisition. Multiple extractions as well as measurement protocols have to be applied to enable a comprehensive picture of the possible neurotoxicant composition of the matrix of interest, especially in NT-HR-MS. To create long term records, a minimum of analytical measurements capturing the maximum amount of information possible within the compromise of time, cost and effort will be needed to provide enough information to direct future, more sophisticated analysis on the frozen and archived (physically and digitally) samples. In the next section we explore some of the analytical options available to cover this extraordinarily broad range of

contaminants within the category “potential neurotoxicants”. As there are many excellent reviews on analysis,¹¹ including one specifically on PD,¹⁰⁹ the text below covers major analytical aspects and decisions that we consider most likely to affect the quality of NT-HR-MS data processing that can be performed, while the ESI† contains some additional discussion.

When performing potentially long term studies of complex samples, careful sample pre-processing steps are required for the analysis, such as the removal of highly abundant matrix components such as lipids and/or pigments that could interfere with the analysis. However, the use of any sample clean-up technique, as well as the choice of the extraction method, carries the risk of eliminating the actual compounds of interest. Furthermore, careful but simple sample preparation and pre-treatment is required that will enable minimum interferences and appropriate normalisation of samples for data processing and subsequent identification efforts over years of data collection. This is especially challenging for unknowns, as the usual techniques applied to targeted compounds (internal standard, recovery and matrix correction) cannot be applied; furthermore as the structure is not known it is challenging to select closely-matching substances. Comparing potential neurotoxicants in samples (especially *e.g.* faecal matter¹¹⁰ or house dust) from different groups in cohort studies requires the normalisation of the qualitative or quantitative signal. For instance, the concentration measured in an extract of stool is affected by the properties of the sample (water content, percentage of undigested matter, *etc.*) beyond even the complexity of wastewater samples, which have been “averaged” in a way throughout the journey to the treatment plant. In the metabolomics context, the retrospective normalisation of the measurement signal to the overall metabolite content (total ion chromatogram or TIC normalisation) or the dry weight of the sample are most common. While the TIC normalisation works only under the (theoretical) assumption that the overall metabolic profile is constant and comparable in all samples of the sequence, the retrospective normalisation to the sample dry weight makes it impossible to adjust sample preparation steps, like the amount of extraction fluid, to achieve best comparability between samples. Such issues make high quality non-targeted data processing and the generation of robust statistics extremely challenging, and will require some dedicated efforts in the coming years, as it is easy for such issues to be inadvertently disregarded, especially in automated workflows.

Measuring the chemical diversity of potential neurotoxicants

In terms of measurement, a large percentage of the neurotoxicants have polarities and ionizable groups that make these molecules amenable to analysis by liquid chromatography high resolution mass spectrometry (LC-HRMS). API techniques like electrospray ionisation (ESI) and atmospheric chemical ionisation (APCI) has dramatically improved compound discovery because of enhanced ionisation efficiency, as well as preventing the need for derivatisation (often the case with GC-MS) that may complicate data interpretation. Analytes that easily lose protons, mainly due to their gas phase



Fig. 4 Box plots showing the wide range of physicochemical properties of the NEUROTOXINS list (https://comptox.epa.gov/dashboard/chemical_lists/neurotoxins),⁵² according to the OPERA predictions⁵⁷ and intrinsic properties from the Dashboard. (a) log values of the biodegradation constant (log BD), octanol–water coefficient (log K_{ow}), water solubility in mol L⁻¹ (log Sol), vapour pressure in mm Hg (log VP) and bioconcentration factor (log BCF) plus (b) boiling point (°C) and monoisotopic mass (Da). The monoisotopic mass was cut-off at 1000 Da for display purposes; 11 entries had a mass between 1000 and 5040 Da.



acidity, can be analyzed by negative ESI, while strongly basic compounds in the gas phase are ionised with ease in positive mode.^{111,112} Weaker bases, on the other hand, are better detected with APCI ionisation.¹¹³ Some potential neurotoxicants that can be analyzed by APCI are hexamethylmelamine (<https://comptox.epa.gov/dashboard/dsstoxdb/results?search=DTXSID4022579>), almitrine (<https://comptox.epa.gov/dashboard/dsstoxdb/results?search=DTXSID4057899>), and diphenylhydantoin (<https://comptox.epa.gov/dashboard/dsstoxdb/results?search=DTXSID8020541>), which have functional groups that are known to ionise better using APCI (triazine for the first two and phenylurea for the last). Endosulfan (<https://comptox.epa.gov/dashboard/dsstoxdb/results?search=DTXSID1020560>) has also been shown to be APCI amenable, which provides better precursor ion information compared to EI analysis.¹¹⁴ Another technique that may find more utility in the future is atmospheric pressure photo-ionisation (APPI) for the analysis of compounds that ionise poorly using the ionisation techniques mentioned thus far.¹¹⁵

Another lesson learned from the neurotoxicant mapping is that many of these compounds are very polar and thus tend to elute at void volume when using reversed phase LC. Because unretained matrix components like macromolecules and salts also come out at or near the void time, hydrophilic molecules tend to suffer from ionisation suppression. Some specific examples include amygdalin (<https://comptox.epa.gov/dashboard/dsstoxdb/results?search=DTXSID00897159>), stavudine (<https://comptox.epa.gov/dashboard/dsstoxdb/results?search=DTXSID1023819>), fludarabine (<https://comptox.epa.gov/dashboard/dsstoxdb/results?search=DTXSID4039657>) and allopurinol (<https://comptox.epa.gov/dashboard/dsstoxdb/results?search=DTXSID4022573>). Hydrophilic interaction liquid chromatography (HILIC) is often able to retain these compounds better, thus providing better analyte signal integrity. HILIC would potentially open up unique chemical space that has received very little attention in environmental analysis up until recent years.¹ Enabling a better separation of compounds of interest will be critical to obtaining sufficient and clean fragmentation information to assist in identification and confirmation, but requires the addition of yet another method to the NT-HR-MS toolbox.

While derivatisation in LC-MS may sometimes be necessary to improve the analytical signal¹¹⁶ and can be used to identify compounds with certain toxicophores,²³ complicated data interpretation often ensues. Measurements made with and without derivatisation, followed by customised data processing, may ease this process in the coming years. Another group of neurotoxicants that require special attention are the toxicants with masses above 1000 Da. For these molecules, multiple charging can be expected in NT-HR-MS and care must be taken when doing data analysis. While multiple charges can be detected in many non-target workflows, these are not commonly investigated in great detail and more work needs to be done in the mass spectrometric analysis of these analytes (see also discussion below).

Further inspection of the neurotoxicant list reveals that many entries are either heavy metals or contain heavy metals. Metals like chromium, lead, mercury, nickel, silver, and

thallium, organometallics like dimethyl and ethyl mercury, and the metalloid arsenic have all been reported to have neurotoxic effects. However, many of these compounds are neither LC nor GC amenable. For these compounds one needs to perform a spectrophotometric analysis or use an inductively coupled plasma to convert these analytes to the gas phase. Preliminary results in the context of the ENTACT project revealed that some arsenic-containing organometallic compounds can indeed be detected by LC-NT-HR-MS with very informative fragmentation patterns, showing that these should certainly be considered in the interpretation of NT-HR-MS data, although not currently covered by many workflows – this integration will be critical in future years.

Looking beyond small molecules (toxic proteins)

Apart from small molecules, exposure to toxic bio-macromolecules (>1000 Da) may also be highly relevant in the context of neurotoxicity. In this context, neurotoxic as well as amyloidogenic (<https://en.wikipedia.org/wiki/Amyloid>) proteins are of particular relevance as they have been implicated as causal factors in Alzheimer's and Parkinson's diseases.¹¹⁷ In particular, prions (misfolded proteins with infectious properties) have been implicated in triggering amyloid-beta, alpha-synuclein and tau misfolding. Once the misfolding of endogenous proteins occurs, propagation of misfolding may occur through mechanisms similar to those that underlie prion pathogenesis.^{118,119} Importantly, the causal agent for the initial misfolding may be distant from the brain regions that are ultimately affected by the diseases.^{120,121} In this context, the enteric nervous system has been implicated.¹²² Based on these observations, it has been postulated that molecules produced by the gut microbiota may trigger a slowly ascending pathological process.^{123,124} For instance, the gut microbiota of newly diagnosed Parkinson's disease patients is enriched in known mucus-foraging bacteria.¹²⁵ Furthermore, apart from other bacterial toxins,¹²⁶ proteins have been posited to trigger the misfolding of proteins in enteric neurons.¹²⁷ Thus, a combination of mucus erosion and a resulting exposure to higher levels of microbial amyloidogenic proteins might at least play a role in Parkinson's disease. To obtain pointers to causal mechanisms, it is therefore essential to not only consider small molecules as potential causal agents but also proteins. This means that integrated multi-omic analyses will be highly relevant for resolving potential combinatorial mechanisms involved in neurotoxicity.

As mentioned above, it is possible that protein and peptide signals may be observed in small molecule measurements. This is a challenge that computational workflows will have to tackle increasingly in the future, with respect to the multiple charge states, large masses and also dealing with this information in compatible ways. While multi-omic integration is becoming a common buzzword, the reality is that workflows and formats are still largely incompatible. For instance, the protein structures for download in the Database of Bacterial ExoToxins for Human¹²⁸ (DBETH, <http://www.hpppi.iicb.res.in/btox/>) are not in a format compatible with compound databases for small



molecules such as the CompTox Chemicals Dashboard or any common small molecule cheminformatics formats. Thus, even obtaining chemical information such as masses and formulas needed for typical suspect or non-target screening workflows is already a challenge with this style of database and better integration will be needed in the future.

Relating chemical analytical results from NT-HR-MS to biology

Biological confirmation of neurotoxic effects

While a critical part of NT-HR-MS is identification and analytical confirmation of suspected chemicals with reference standards (where possible), the results must be related back to the original study question. In environmental studies this is quite often performed using bioassays, as in effect-directed analysis. A recent article contains extensive material discussing approaches for screening neurotoxicity from an ecological standpoint (“econeurotoxicity”) and we refer readers to Legradi *et al.*¹²⁹ rather than reproducing a summarised form of this discussion here. The material below provides additional perspectives.

Specific features that distinguish the central nervous system (CNS) from other organs need to be considered when choosing the biological systems, read-outs and methods to test the potential neurotoxic effects of environmental chemicals. These features include the presence of the blood-brain barrier, the high lipid content of the nervous tissue, the high energy requirement of neurons, the particular intercellular signaling system (synaptic transmission), the neural cell structure (long axonal projections), the presence of specific reactive endogenous molecules (*e.g.* dopamine) and the post-mitotic nature of neuronal cells (making them more sensitive to age-related accumulation of cellular damage¹³⁰). In addition, the heterogeneity of the brain tissue and the sometimes highly selective susceptibility of certain neuronal cell types to only one or another neurotoxic substance calls for the use of more than one test system to maximise the chances of identifying hazardous effects of a potentially new neurotoxic compound.

A current tendency in the field of (neuro)toxicity testing is to transition from more observational projects using rodents to more mechanistic studies involving the use of *in vitro* (cellular) test systems.¹³¹ The latter include primary neural cells, immortalised neural cell lines and stem cells of non-human or human origin. Given species-specific responses to neurotoxic substances, even between rodents and humans, working with human derived cells (as opposed to cells from non-human origin) maximises the chances for making predictions on neurotoxicity relevant to the human nervous system. However, access to human primary cells is limited, given that they are derived from aborted foetuses or from brain surgery resections. Although less limited, isolation of primary cells from rodents is laborious and requires animal sacrifices for each new experiment. This explains the widespread use of rodent or human cell lines for neurotoxicity testing, even though the physiological relevance of the results generated in these systems are more

questionable due to their tumorigenic origin or the immortalisation process applied (e.g. overexpression of oncogenes or telomerase). A very dynamic and expanding field is the use of mouse or human derived embryonic stem cells (ESCs) and of human induced pluripotent stem cells (iPSCs), which have been derived more recently by reprogramming of human somatic cells.^{132,133} A panoply of protocols have been developed to differentiate iPSCs into different neural cell types.¹³⁴ Exposing these cells to potential neurotoxicants during the differentiation process allows testing more specifically for *developmental* neurotoxicity. Finally, rapid progress is ongoing in the development of 3D organoid models of the brain, which are also derived from iPSCs.^{135–137} Here the aim is to better mimic the complexity of the human brain, or certain parts of it (e.g. midbrain, relevant for Parkinson's disease research), by creating *in vitro* tissue structures including several cell types in a spatial organisation that reflects closely the physiological situation, but that offer easy access for experimental manipulations.

Common endpoints for *in vitro* neurotoxicity testing and the analytical approaches used for measuring those endpoints have been reviewed in Schmidt *et al.*¹³¹ with a focus on those methods that allow for high-throughput screening. Briefly, the endpoints can be grouped into viability, morphological and functional read-outs. Viability assays allow testing for neurotoxicity by determining whether one or several types of neural cells are more sensitive to compounds of interest than other cell types. At sub-cytotoxic concentrations, compounds can be further tested for their impact on neural cell morphology or function with endpoints including the monitoring of neurite outgrowth, spontaneous electrical activity of neuronal networks, receptor signaling and cell communication, reactive oxygen species formation, cell migration, mitochondrial transport, calcium storage and release, cell membrane potential, gliosis (*i.e.* proliferation of glial cells), myelination, network formation, and synaptogenesis. A number of these endpoints, as well as cell viability, can be measured by high-throughput screens based on absorbance, fluorescence or luminescence measurements in multi-well plates. An interesting, more recent development are microelectrode arrays (MEA) that allow for the measurement of extracellular electrical fields of neurons that grow on them. The mean firing rate has been proposed to be the most sensitive parameter for neurotoxicity screening *via* this method, which allows measurement of the spontaneous electrical activity arising in neural networks at relatively high throughput (24-well MEA plates).¹³⁸ High-content imaging, *i.e.* automated fluorescent imaging of fixed or live cells in a high-throughput manner, is a technique that continues to undergo important developments, but has already become a central part of neuroscience and related compound screens.¹³⁹ Transcriptomics, miRNA profiling and metabolomics are used as analytical tools to characterise molecular changes that occur in response to neurotoxicant exposure. These techniques hold promise for defining molecular signatures that are specific for certain chemicals, providing a read-out that can be more sensitive than morphological changes.^{140,141}

With this perspective we have started a conversation amongst several research groups at one institute and close collaborators to scope the breadth of experience needed to begin to perform a prospective, high quality, comprehensive assessment of potential small molecule neurotoxicants in environmental and biological samples using cheminformatics and non-target high

resolution mass spectrometry, and what would be required to link this back to the disease state. Screening the thousands of chemicals potentially involved is a huge task that can be eased significantly by using some carefully-considered cheminformatics, literature-based and data processing methods and we hope users find the resources prepared in this article a useful way to start to explore different approaches for data processing. While we have prepared and described the methods using resources we use and develop, the data files and concepts are transferable to the many excellent workflows and approaches already available. Realistically, non-target MS-based analysis will have to evolve towards a few harmonised methods to ensure sufficient coverage of substances of interest in different contexts. However, compounds not covered by small molecule HR-MS analysis also play a defining role in diseases, such as larger molecules (requiring multi-omics approaches) or metals in salt forms (requiring alternative analysis) and these also need consideration in the bigger picture. Despite the compilation of many chemical lists, it is also clear that these chemicals transform and, although not covered in detail here, the candidate space for NT-HR-MS can and should be extended to screen for predicted transformation products and metabolites of potential neurotoxicants. Another area to explore in future work is a better connection to the concept of disease maps, such as the Parkinson's disease map,¹⁵⁴ which can be queried computationally – and thus potentially integrated into mass spectral workflows – *via* interfaces such as MINERVA.¹⁵⁵

While hypothesis generation can begin already from data snapshots on patients and controls at certain time points, in reality the prioritisation and discovery of “driving” neurotoxicants will depend on forming high quality, long term datasets involving cohorts where participants are recruited even before any symptoms are displayed, so that long term trends become clear. Ideally, this would be extended across multiple cohorts to allow hypothesis development in one cohort and replication in another based on chemical-environmental stratification. Alternatively, the increasing development of sensors and portable devices that can be given to patients and the general population open up new opportunities for citizen science, crowd sourcing of data and greater collaborative efforts to form detailed sampling campaigns that would provide complementary information to cohort data. The increasing willingness to share data in open resources such as the Global Natural Products Social Molecular Networking (GNPS)¹⁵⁶ repository will open up many new opportunities in the coming years. This includes greater availability of larger public datasets on which to develop machine learning and artificial intelligence approaches to improve identification and classification of relevant chemicals and exposures.¹⁵⁷ High resolution mass spectrometry of small molecules is certainly a well-placed technique to start building the knowledge to help answer these extremely complicated questions, assuming the long term support is available to create and maintain such extremely valuable datasets. In the end, however, this will provide only a starting point for further investigations. In addition to hands on biological experiments to elucidate possible exposure routes or gene–environment interactions, additional efforts in

computational modeling and simulation using mathematical and mechanistic models will be needed to turn identified chemicals into a greater understanding of an extremely complex system and potential environmental causes of chronic diseases.

Author contributions

ELS conceptualised the perspective upon invitation and coordinated the writing. General contributions: cheminformatics: NCB (literature mining), AJW (registration, lists) and ELS (NT-HR-MS); analytical/samples: RS, JPT, PLK, ELS. Domain-specific knowledge: JPT (biobanking, metabolomics), PLK (medical), RK (clinical/molecular neuroscience), PW (multi-omics), CLL (biology), NP (metabolomics), RB (systems biomedicine). All authors made essential contributions to discussions and material presented in this perspective.

Conflicts of interest

There are no conflicts to declare.

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