



Cite this: *Chem. Commun.*, 2018, 54, 8466

Received 30th May 2018,
Accepted 26th June 2018

DOI: 10.1039/c8cc04316g

rsc.li/chemcomm

'AND'-based fluorescence scaffold for the detection of ROS/RNS and a second analyte†

Maria L. Odyniec,[‡] Adam C. Sedgwick,^{‡*} Alexander H. Swan,[‡] Maria Weber,[‡] T. M. Simon Tang,[‡] Jordan E. Gardiner,[‡] Miao Zhang,^b Yun-Bao Jiang,^b Gabriele Kociok-Kohn,^a Robert B. P. Elmes,^c Steven D. Bull,^{‡*} Xiao-Peng He[‡] and Tony D. James^{‡*ae}

Traditionally, fluorescence probes have focused on the detection of a single biomarker for a specific process. In this work, we set out to develop a number of fluorescence probes that enable the detection of a chosen analyte in the presence of reactive oxygen/nitrogen species (ROS/RNS). These fluorescence probes when activated result in the formation of the highly fluorescent pink dye, resorufin. Therefore, we have labelled these fluorescent probes as 'Pinkments'. Our first 'Pinkment' was shown to detect biologically relevant concentrations of ONOO⁻ and have an excellent selectivity against other ROS/RNS. Pinkment-OH was developed to provide a core unit which could be easily functionalised to produce a range of 'AND' based fluorescence probes for the detection of ROS/RNS and a second analyte. For proof of concept, we synthesised Pinkment-OTBS and Pinkment-OAc. These 'AND'-based probes were successfully shown to detect ROS/RNS and F⁻ or esterase, respectively.

Since the discovery of the first fluorescence-based probe in 1867, fluorescence probes have revolutionised the understanding of biological systems.¹⁻⁴ Historically, these probes have focused on the detection of a single analyte or biomarker. However, biological systems are complex with more than one chemical species being released/present during any biological processes. For example, glutathione (GSH) accumulates at the

nucleus during the cell cycle to aid transcription factors binding to DNA⁵ and the pathological role of Zn²⁺ is believed to be associated with the glutamate system.⁶ Furthermore, the sensitivity of a cell towards peroxynitrite (ONOO⁻) largely depends on the concentration of intracellular GSH.⁷⁻¹⁰ Therefore, in order to further understand cellular functions and the root causes of disease it is important to be able to study biologically important species simultaneously.

Alongside the development of the field of fluorescence probes, the field of molecular logic gates has developed.¹¹ Molecular logic gates are molecules that have the ability to bind or react with multiple analytes (input) and turn it into a measurable optical output. Consequently, these attractive molecules are now emerging in the literature demonstrating the ability to simultaneously detect multiple analytes in biological systems.¹²⁻²¹ Within our research group, we are interested in developing reaction-based fluorescence probes including 'AND'-based fluorescence probes for the detection of biologically important analytes.²²⁻²⁴ Dual responsive ('AND') fluorescence probes require both analytes being present to produce a fluorescence response. In this work, we identified a previously reported boronate-based fluorescence probe developed by Chang *et al.* **PR1**, with a free amino group attached (Fig. 1). Boronates have well-known reactivity towards ONOO⁻ and hydrogen peroxide (H₂O₂),²⁵ therefore we believed **PR1** could provide a suitable

^a Department of Chemistry, University of Bath, Bath, BA2 7AY, UK.
E-mail: t.d.james@bath.ac.uk, s.d.bull@bath.ac.uk

^b Department of Chemistry, College of Chemistry and Chemical Engineering, the MOE Key Laboratory of Spectrochemical Analysis and Instrumentation, and iChEM, Xiamen University, Xiamen 361005, China

^c Department of Chemistry, Maynooth University, Maynooth, Co. Kildare, Ireland

^d Key Laboratory for Advanced Materials & Feringa Nobel Prize Scientist Joint, Research Center, East China University of Science and Technology, 130 Meilong Rd., Shanghai 200237, P. R. China

^e Department of Materials and Life Sciences, Faculty of Science and Technology Sophia University, 7-1 Kioi-cho, Chiyoda-ku, Tokyo 102-8554, Japan

† Electronic supplementary information (ESI) available. CCDC 1844385. For ESI and crystallographic data in CIF or other electronic format see DOI: 10.1039/c8cc04316g

‡ A. C. S. and M. L. O. contributed equally, A. C. S. – organic synthesis, M. L. O. – fluorescence analysis.



Fig. 1 **PR1** and **Pinkment** fluorescence probes for the detection of reactive oxygen and reactive nitrogen species (ROS/RNS). **Pinkment-Bn** – demonstrates the requirement elimination to produce a free N–H.



Foundation of China (21722801), the Science and Technology Commission of Shanghai Municipality (15540723800) and the Shanghai Rising-Star Program (16QA1401400) for financial support. TDJ, MZ and YBJ thank the Royal Society for funding an International Joint Project (IE121564). NMR characterisation facilities were provided through the Chemical Characterisation and Analysis Facility (CCAF) at the University of Bath (www.bath.ac.uk/ccaf). The EPSRC UK National Mass Spectrometry Facility at Swansea University is thanked for analyses. All data supporting this study are provided as ESI† accompanying this paper.

Conflicts of interest

No conflicts of interest.

Notes and references

- 1 D. Wu, A. C. Sedgwick, T. Gunnlaugsson, E. U. Akkaya, J. Yoon and T. D. James, *Chem. Soc. Rev.*, 2017, **46**, 7105–7123.
- 2 J. Chan, S. C. Dodani and C. J. Chang, *Nat. Chem.*, 2012, **4**, 973–984.
- 3 Y. M. Yang, Q. Zhao, W. Feng and F. Y. Li, *Chem. Rev.*, 2013, **113**, 192–270.
- 4 X. P. He and H. Tian, *Chem*, 2018, **4**, 246–268.
- 5 J. Markovic, C. Borrás, A. Ortega, J. Sastre, J. Vina and F. V. Pallardo, *J. Biol. Chem.*, 2007, **282**, 20416–20424.
- 6 B. Pochwat, G. Nowak and B. Szewczyk, *Neural Plast.*, 2015, **2015**, 591563.
- 7 K. A. Marshall, R. Reist, P. Jenner and B. Halliwell, *Free Radical Biol. Med.*, 1999, **27**, 515–520.
- 8 P. Pacher, J. S. Beckman and L. Liaudet, *Physiol. Rev.*, 2007, **87**, 315–424.
- 9 J. P. Bolanos, S. J. R. Heales, J. M. Land and J. B. Clark, *J. Neurochem.*, 1995, **64**, 1965–1972.
- 10 M. Nakamura, V. H. Thourani, R. S. Ronson, D. A. Velez, X. L. Ma, S. Katzmark, J. Robinson, L. S. Schmarkey, Z. Q. Zhao, N. P. Wang, R. A. Guyton and J. Vinten-Johansen, *Circulation*, 2000, **102**, 332–338.
- 11 S. Erbas-Cakmak, S. Kolemen, A. C. Sedgwick, T. Gunnlaugsson, T. D. James, J. Yoon and E. U. Akkaya, *Chem. Soc. Rev.*, 2018, **47**, 2228–2248.
- 12 C. Y. Ang, S. Y. Tan, S. J. Wu, Q. Y. Qu, M. F. E. Wong, Z. Luo, P. Z. Li, S. T. Selvan and Y. L. Zhao, *J. Mater. Chem. C*, 2016, **4**, 2761–2774.
- 13 S. Debieu and A. Romieu, *Org. Biomol. Chem.*, 2015, **13**, 10348–10361.
- 14 G. C. Van de Bittner, C. R. Bertozzi and C. J. Chang, *J. Am. Chem. Soc.*, 2013, **135**, 1783–1795.
- 15 F. B. Yu, P. Li, B. S. Wang and K. L. Han, *J. Am. Chem. Soc.*, 2013, **135**, 7674–7680.
- 16 X. F. Yang, Q. Huang, Y. G. Zhong, Z. Li, H. Li, M. Lowry, J. O. Escobedo and R. M. Strongin, *Chem. Sci.*, 2014, **5**, 2177–2183.
- 17 S. Resa, A. Orte, D. Miguel, J. M. Paredes, V. Puente-Munoz, R. Salto, M. D. Giron, M. J. Ruedas-Rama, J. M. Cuerva, J. M. Alvarez-Pez and L. Crovetto, *Chem. – Eur. J.*, 2015, **21**, 14772–14779.
- 18 X. P. He, X. L. Hu, T. D. James, J. Yoon and H. Tian, *Chem. Soc. Rev.*, 2017, **46**, 6687–6696.
- 19 L. Yu, S. L. Wang, K. Z. Huang, Z. G. Liu, F. Gao and W. B. Zeng, *Tetrahedron*, 2015, **71**, 4679–4706.
- 20 J. L. Kolanowski, F. Liu and E. J. New, *Chem. Soc. Rev.*, 2018, **47**, 195–208.
- 21 A. Romieu, *Org. Biomol. Chem.*, 2015, **13**, 1294–1306.
- 22 A. C. Sedgwick, H. H. Han, J. E. Gardiner, S. D. Bull, X. P. He and T. D. James, *Chem. Commun.*, 2017, **53**, 12822–12825.
- 23 A. C. Sedgwick, R. S. L. Chapman, J. E. Gardiner, L. R. Peacock, G. Kim, J. Yoon, S. D. Bull and T. D. James, *Chem. Commun.*, 2017, **53**, 10441–10443.
- 24 A. C. Sedgwick, H. H. Han, J. E. Gardiner, S. D. Bull, X. P. He and T. D. James, *Chem. Sci.*, 2018, **9**, 3672–3676.
- 25 A. Sikora, J. Zielonka, M. Lopez, J. Joseph and B. Kalyanaraman, *Free Radical Biol. Med.*, 2009, **47**, 1401–1407.
- 26 E. W. Miller, A. E. Albers, A. Pralle, E. Y. Isacoff and C. J. Chang, *J. Am. Chem. Soc.*, 2005, **127**, 16652–16659.
- 27 X. M. Chen, B. H. Zhou, T. T. Yan, H. Wu, J. H. Feng, H. S. Chen, C. Gao, T. Peng, D. Yang and J. G. Shen, *Free Radical Biol. Med.*, 2018, **117**, 158–167.
- 28 H. Fujigaki, K. Saito, F. Lin, S. Fujigaki, K. Takahashi, B. M. Martin, C. Y. Chen, J. Masuda, J. Kowalak, O. Takikawa, M. Seishima and S. P. Markey, *J. Immunol.*, 2006, **176**, 372–379.
- 29 X. F. Yang, S. J. Ye, Q. Bai and X. Q. Wang, *J. Fluoresc.*, 2007, **17**, 81–87.
- 30 J. Cao, C. C. Zhao, P. Feng, Y. L. Zhang and W. H. Zhu, *RSC Adv.*, 2012, **2**, 418–420.
- 31 D. Yang, Y. C. Tang, J. Chen, X. C. Wang, M. D. Bartberger, K. N. Houk and L. Olson, *J. Am. Chem. Soc.*, 1999, **121**, 11976–11983.
- 32 M. Abo, Y. Urano, K. Hanaoka, T. Terai, T. Komatsu and T. Nagano, *J. Am. Chem. Soc.*, 2011, **133**, 10629–10637.
- 33 J. Yang, X. L. Zhang, P. Yuan, Y. G. Xu, J. Grutzendler, Y. H. Shao, A. Moore and C. Z. Ran, *Proc. Natl. Acad. Sci. U. S. A.*, 2017, **114**, 12384–12389.
- 34 M. Santra, S. K. Ko, I. Shin and K. H. Ahn, *Chem. Commun.*, 2010, **46**, 3964–3966.
- 35 F. L. Song, S. Watanabe, P. E. Floreancig and K. Koide, *J. Am. Chem. Soc.*, 2008, **130**, 16460–16461.
- 36 B. J. Shenker, T. L. Guo and I. M. Shapiro, *Environ. Res.*, 2000, **84**, 89–99.
- 37 A. E. A. Moneim, *Neural Regener. Res.*, 2015, **10**, 881–882.
- 38 A. E. A. Moneim, *Metab. Brain Dis.*, 2015, **30**, 935–942.
- 39 S. Alarifi, D. Ali, S. Alkahtani and R. S. Almeer, *Oxid. Med. Cell. Longevity*, 2017, DOI: 10.1155/2017/8439098.

