

Cite this: *RSC Adv.*, 2015, 5, 40324

Use of herbs, spices and their bioactive compounds in active food packaging

A. Valdés, A. C. Mellinas, M. Ramos, N. Burgos, A. Jiménez and M. C. Garrigós*

The interest and societal demand on the use of natural, biodegradable and renewable resources has increased in the last few years. In addition, food producers and consumers have improved their requirements for the quality of processed food, particularly in the field of increasing shelf-life while preserving organoleptic and nutritional properties. Active packaging technologies have greatly developed in the last decade by trying to satisfy the need for long-life processed food in addition to antioxidant/antimicrobial components in the packaging material. These components are intended to be released in a controlled way to food. These rising trends have been reflected in the field of food packaging by the use of chemicals extracted and obtained from plants in active packaging formulations. Herbs and spices have shown great potential to be used as renewable, biodegradable and valuable sources of chemicals, such as polyphenols, with high antioxidant/antimicrobial performance. This review aims to present the latest published work in this area.

Received 31st December 2014
Accepted 21st April 2015

DOI: 10.1039/c4ra17286h

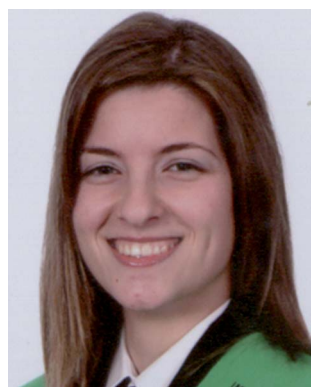
www.rsc.org/advances

1. Introduction

It is well known that antioxidants are present in many plants as a defence towards oxidative degradation coming from the environment. These bio-based and natural antioxidants have improved their use in food-related applications through their positive contributions to human health. Many plants have been considered as potential antioxidant sources and some of them have been studied in recent years. In this context, numerous herbs and spices are the most important and studied targets in

the search for natural antioxidants.¹ The terms 'herbs' and 'spices' have more than one definition in common languages, but the most commonly used are those which consider herbs to be obtained from the green parts of plants, such as the stem and leaves, while spices are usually obtained from the bark, roots, seeds or flowers. Most of the common spices are usually extracted from plants grown in warm or tropical climates while herbal plants can be grown in nearly all conditions and geographical areas around the world. In addition, spices have a more intense taste and are used in small amounts.² Based on data reported by the Food and Agriculture Organization of the United Nations (FAO), the worldwide spice production has

University of Alicante, Dpt. Analytical Chemistry, Nutrition & Food Sciences, 03690, San Vicente del Raspeig, Spain. E-mail: mc.garrigos@ua.es



Arantazu Valdés received her PhD in Chemistry in 2014 and she is currently working as a post-doc researcher at the University of Alicante (Analytical Chemistry, Nutrition and Food Science Department). Her research interests include studies on the oxidative stability of fatty food; active packaging, natural antioxidants and antimicrobials; sustainable additives for polymers; characterization of polymers for food packaging; properties and degradation studies; revalorization of agricultural residues and development of analytical methods for the determination of additives in polymers.



Ana Cristina Mellinas, graduated in chemistry from the University of Alicante in 2012. She currently serves as a senior technician in the Analysis of Polymers and Nanomaterials Research Group, Department of Analytical Chemistry, Nutrition and Food Science at the University of Alicante.

increased significantly in the last few years³ whereas the global production of herbs has not been reported in the literature.

Since ancient times, herbs and spices have been extensively used to enhance or improve the flavour of food and by their preservative properties.⁴ Several popular herbs and spices are known to have beneficial effects for human health, including digestive stimulant, anti-inflammatory, antimicrobial, antioxidant and anti-carcinogenic activities, which are mostly attributed to polyphenols present in their chemical composition. Moreover, other volatile constituents in herbs and spices (particularly essential oils) can also contribute to the plant's biological activity, resulting in benefits for human health. In general terms, essential oils are chemically constituted by more than 70 different compounds, mostly polyphenols, terpenes, monoterpenes and sesquiterpenes, some of them representing more than 85% of the total oil content.⁵ Essential oils and their components are gaining attention by their wide acceptance by consumers and the possibility of exploitation for potential multi-purpose functional uses.

Phytochemicals extracted from various herbs and spices, including polyphenols, flavonoids, flavonols, tannins and terpenoids, could help to minimise tumour development by inhibiting the initiation and promotion of carcinogenesis, inducing tumour cell differentiation and apoptosis, while suppressing tumour angiogenesis. They are also able to prevent and reverse many of the processes that underlie chronic diseases. These phytochemicals can be broadly classified as carotenoids, phenolics, alkaloids, nitrogen-containing compounds and organosulfur compounds.⁶ Some studies have been recently reported to state the presence and activity of antioxidants in herbs, to determine their total phenolic contents and to relate the total antioxidant and antimicrobial activities of methanolic extracts, infusions and decoctions of several plants; such as peppermint, thyme, lemon balm, basil, rosemary and sage.^{6,7}

On the other hand, antioxidants present in spices have gained importance by their role in the prevention of atherogenesis. Although spices have been recognized by their beneficial health effects and have been used in traditional medicine



Marina Ramos graduated in Chemistry from the University of Alicante (Spain). She is a PhD student in the "Polymer and Nanomaterials Analysis" group at the same University. Her current research is focused on the development of antioxidant and antimicrobials active packaging systems based on conventional polymers and biopolymers. She has demonstrated excellent results when

natural additives such as thymol and silver nanoparticles are incorporated into a polymer matrix achieving antioxidant and antimicrobial properties.



Alfonso Jiménez, Chemistry PhD (1996). Full Professor in Analytical Chemistry and Food Science at the University of Alicante from 2001. Head of the Polymer and Nanomaterials Analysis Group. Editor of 15 books on Sustainable Polymers. Author of 103 research papers published in journals in Analytical Chemistry, Food Technology and Polymer Science. The main research areas are: Chemical

modification of biopolymers; Natural additives for active packaging; TPUs obtained from vegetable oils; Valorization of agro-food residues.



Nuria Burgos, graduated in Chemical Engineering in 2002 and received her PhD in Chemistry in 2013 at the University of Alicante (Spain). She is working as a Senior Researcher in the Polymer and Nanomaterials Analysis group (Department of Analytical Chemistry, Nutrition and Food Sciences, University of Alicante). Her main research areas are focused on the development of new sustainable

composites based on biopolymers and natural compounds (plasticizers, nanofillers and active additives) for their use as food packaging materials. She has published several research papers in journals relating to Food and Polymer Technology and she has presented their results at different international conferences.



María del Carmen Garrigós, PhD in Chemistry in the University of Alicante (2003). Assistant Professor in Analytical Chemistry at the University of Alicante from 2008. Author of 32 research papers published in journals on Analytical Chemistry, Food Technology and Polymer Science. The main research areas are: Chemical modification of biopolymers; Natural additives for active

packaging; Edible films; TPUs obtained from vegetable oils; Valorization of agro-food residues; Carbohydrate-based advanced biomaterials; Extraction of bioactive compounds; Quality control methods and multivariate analysis for food authentication.

since ancient times, their attributes in health protection have been experimentally verified only in the last three decades. In this sense, the antioxidant effects of turmeric/curcumin, clove/eugenol, onion, garlic, ginger/gingerol, red pepper/capsaicin, black pepper/piperine and fenugreek, have been recently reviewed.⁸

Beneficial effects of chemicals obtained from herbs and spices can be used not only directly into food but also in packaging materials to increase food shelf-life and quality. In recent years, the increase in consumer's demand for minimally processed foods, the change in retail and distribution practices associated with global markets, new product logistics and distribution trends (such as on-line shopping), automatic handling systems and strict requirements regarding consumer's health and safety have been the major driving force for innovation in food packaging technologies.⁹ In this context, packaging of fat food is a particular issue, since lipid oxidation after microbial growth is the main cause of food spoilage.¹⁰ In particular, food with high content in poly-unsaturated fatty acids is more prone to oxidative deterioration resulting in the development of off-flavours, such as toxic aldehydes, and rendering products unacceptable for human consumption by their loss in nutritional quality.¹¹ The use of active packaging systems with antioxidant activity is a promising approach to improve food quality by the incorporation of antioxidant agents to the packaging material increasing the stability of oxidation-sensitive food. However, the use of synthetic antioxidants in these active systems is questioned due to potential toxicological risks, resulting in requirements for strict controls. The alternative based on the use of natural antioxidants, particularly polyphenols, extracted from plants and essential oils from herbs and spices¹² and from agricultural by-products¹³ is one of the current trending topics in food packaging research. In this sense, natural extracts have been incorporated into plastic packaging materials in order to achieve antioxidant properties. For instance, Li *et al.*¹⁴ developed active gelatin films containing natural antioxidants extracted from different sources (green tea, grape seed polyphenols, grape seed proanthocyanidins, ginger and ginkgo leaf) for food packaging applications, showing different effects on the food packaged depending on the incorporated extract. Among them, the ginkgo leaf extract induced the highest radical-scavenging activity whereas the green tea extract resulted in increased protection against moisture. Quince seed mucilage films containing oregano essential oils were prepared and their antioxidant properties were evaluated by Jouki *et al.*¹⁵ These films exhibited high level of radical-scavenging activity (45, 57 and 61% for 1, 1.5 and 2 wt% of essential oils, respectively). In an interesting work recently reported by Castro *et al.*,¹⁶ good feasibility of catechins and quercetin released from green tea natural extracts incorporated into polypropylene films was observed.

Several different methods have been proposed to determine the antioxidant performance of active packaging systems. Some of them are based on measuring the degree of lipid oxidation of food in direct contact with packaging materials. Several parameters, such as peroxide index, conjugated dienes, conjugated triene hydroperoxides, free fatty acids, thiobarbituric acid

index and *p*-anisidine value (AV) can be determined to assess lipid hydrolysis and primary and secondary lipid oxidation in fat food. In addition, the antioxidant activity provided by films for food packaging can be evaluated by measuring the radical scavenging ability in food simulants. Several procedures have been proposed, most of them based on producing and determining oxidative free-radicals, to evaluate the antioxidant properties of additives used in packaging materials. This is the case of oxygen radical absorbance capacities (ORAC), ferric reducing antioxidant power (FRAP), ABTS and DPPH tests.¹⁷

Some studies have been published on the antimicrobial performance of essential oils extracted from plants against food-borne pathogens. Since essential oils are rich in volatile terpenoids and phenolics, they show high potential to inhibit a wide spectrum of microorganisms. The active components of plant essential oils are able to inhibit microorganisms' proliferation through disturbance of the cytoplasmic membrane, disrupting the proton motive force, electron flow, active transport and inhibition of protein synthesis.¹⁸

Antimicrobial and/or antioxidant active packaging systems are getting attention from food and packaging industries due to the increasing consumer demands for minimally processed and preservative-free food products. Some plastic films allow the controlled release of additives onto food to cover the whole storage and distribution operations, while limiting undesirable effects in flavour and taste. A raising trend in the use of antimicrobial additives, such as those derived from essential oils, to minimise contamination in different types of food, such as meat, fruits and vegetables, has been reported and will be discussed in this review article. For instance, Ramos *et al.* reported the development of antimicrobial active films based on polypropylene by incorporating thymol and carvacrol at three different concentrations (4, 6 and 8 wt%) demonstrating their potential to be used as active additives.¹⁹ Pereira de Abreu *et al.*²⁰ reported the potential use of films containing antioxidant extracts from barley husks for the commercialization of frozen cod fillets and the effect on lipid oxidation during prolonged frozen storage. This increasing interest in the use of natural extracts obtained from plants in active packaging technologies is the main reason of this review, where the current situation in the potential use of herbs, spices and their bioactive compounds in antioxidant and antimicrobial active food packaging systems is discussed.

2. Herbs

The antioxidant/antimicrobial properties of herbs have been known since ancient times and they have been used in many applications, such as traditional medicine, food preservation and others. But the raising trend for the use of active packaging in processed food has resulted in new applications of herbs and/or extracted chemicals in food technology. It is known that antimicrobial/antioxidant agents can be incorporated directly into the active packaging systems (either as aqueous or ethanolic extracts). Some examples are summarized in Table 1.

Among them, green tea has been considered for years as a good source of polyphenolic compounds. Catechins (also

Table 1 Examples of the direct incorporation of herb extracts into polymer matrices

Herb extract	Polymer	Effect on food packaging	References
Ginseng	LDPE, PVC	Antimicrobial	22
	Alginate-based	Antioxidant	41
Rosemary	LDPE	Antioxidant	23
	LDPE	Lipid oxidation	24 and 25
	PCL, methylcellulose	Antimicrobial	42
Murta	Carboxymethylcellulose	Antioxidant/antimicrobial	12, 43 and 44
Green tea	Fish skin gelatin	Antioxidant	30
	Silver carp skin gelatin	Antioxidant	31
	Multilayer PET/PE/EVOH/PE	Antioxidant/antimicrobial	45
	EVOH	Antioxidant	46
	Fish skin gelatin	Antioxidant	30
	Agar-fish gelatin	Antioxidant	32
	EVOH	Lipid oxidation	47
	Agar with glycerol	Antimicrobial	33
	Chitosan	Lipid oxidation	35 and 36
	Cassava starch with glycerol	Antioxidant	48
Ginkgo leaf	Fish skin gelatin	Antioxidant	30
Mint	Chitosan and polyvinyl alcohol	Antioxidant/antimicrobial	21

known as flavanols), including epicatechin, epigallocatechin, epicatechin gallate, epigallocatechin gallate, catechin, gallic acid, catechin gallate and gallic acid, are the dominant phenolic compounds in tea leaves and most of them have been extracted from green tea leaves and other parts of the plant.²¹

Some commodity polymers such as low density polyethylene (LDPE),^{22–26} poly(vinyl chloride) (PVC)²² and polypropylene (PP)^{27–29} have been tested as adequate matrices to host different herb extracts. But in recent years biopolymers based on starch, cellulose derivatives, chitosan/chitin, gums, proteins obtained from animal or plant-sources and lipids have emerged as environmentally-friendly alternatives to synthetic polymers as carriers of herb extracts (Table 1). Biopolymers could be processed by wet methods using water and other solvents, avoiding the degradation and loss of volatile active compounds in the final material due to the severe thermo-mechanical conditions applied in conventional techniques for polymer processing (extrusion, injection, blow molding and heat pressing). These biomaterials show additional advantages, such as their biocompatibility, edibility, barrier to moisture and/or gases, non-toxicity, mechanical integrity and relatively low cost.¹² Fish gelatin^{30–34} and chitosan^{35–40} have been widely used as biopolymer matrices by their own intrinsic antimicrobial activity getting synergies with natural additives.

Different methods have been proposed to evaluate the antioxidant effectiveness of active materials¹⁷ directly in food or food simulants. The antioxidant activity provided by polymer films can be evaluated by measuring the radical scavenging ability in food simulants, by using the DPPH (2,2-diphenyl-1-picrylhydrazyl) test.^{21,23–41,46,47} Other similar tests were based on the ferric reducing antioxidant power (FRAP)³² and the antioxidant activity evaluated by complexation of 2,2-azino-bis-3-ethylbenzothiazoline-6-sulfonic acid (ABTS).^{32–46}

Other test methods are based on the measurement of the degree of lipid oxidation, particularly by the determination of the initial autoxidation products, *i.e.* hydroperoxides, which decompose to low molecular-weight compounds resulting in the development of rancid flavours and odours.⁴⁹ Frequently, both the primary and secondary oxidation products are monitored by the peroxide and TBAR indices^{22–24,47,48} in shelf-life assays directly applied to food.

In a recent report, authors have not only incorporated aqueous extracts obtained from herbs to polymer matrices but also in the form of volatile essential oils (EOs), normally found in special cells or groups of cells present in leaves and stems, and commonly concentrated in leaves, bark or fruits.⁵⁰ Some results in the use of EOs extracted from herbs are presented in Table 2.

Oregano is one of the most studied plants by its high antioxidant and antimicrobial properties.^{40,51} Oregano is a popular culinary herb with high content in EOs containing terpenoids, mainly the monoterpenoid phenols of thymol(5-methyl-2-[1-methylethyl]phenol) and carvacrol(5-isopropyl-2-methylphenol).⁵⁰ One important feature of the EOs obtained from oregano is their high antimicrobial activity, which is usually determined by the agar plate diffusion method, where the inhibition on solid media was used to determine the antimicrobial effects of active films against different bacteria.²⁶ Other studies have highlighted the antimicrobial activity of EOs directly on food, such as meat or fish.^{52,53} Oregano and other herbs have shown good antimicrobial performance depending on the type of food bacteria. For example, oregano EOs have been reported as effective against Gram-positive bacteria, such as *Staphylococcus aureus*, *Bacillus cereus*, *Streptococcus mutans*, *L. monocytogenes* and *Enterococcus faecalis*; as well as against Gram-negative bacteria, such as *Escherichia coli* and *Salmonella enterica* subsp.^{54,55} Results showed positive inhibition for EOs concentrations higher than 5% (w/w) for all bacteria.^{27,52–55}

Table 2 Examples of the incorporation of essential oils from herbs into polymer matrices

Essential oil	Polymer	Effect on food packaging	References
<i>Zataria multiflora</i>	Kappa-carrageenan (KC)	Antimicrobial	56 and 57
	Soluble soybean polysaccharide	Antioxidant/antimicrobial	58
<i>Satureja hortensis</i>	KC	Antioxidant/antimicrobial	59
<i>Mentha pulegium</i>	KC	Antioxidant/antimicrobial	56
	Soluble soybean polysaccharide	Antioxidant/antimicrobial	58
Oregano	Ethylene vinyl alcohol copolymer	Antioxidant/antimicrobial	51
	PET	Antimicrobial	27
	Mucilage	Antioxidant/antimicrobial	60
	Thermoplastic starch with poly (butylene adipate-co-terephthalate)	Antioxidant/antimicrobial	61
	Chitosan	Antioxidant/antimicrobial	54
	PP	Antimicrobial	27
	Methylcellulose	Antimicrobial	62
	PP	Insect-repellent	28
	Polyvinyl alcohol (PVA)	Antimicrobial	55
	PP/EVOH	Antimicrobial	52–63
	Multilayer PET/PE/EVOH/PE	Antioxidant/antimicrobial	45
	Isolated soy protein	Antioxidant	40
	PP	Antifungal	29
	LDPE	Antimicrobial	26
	PLA-CNC	Antimicrobial	53
	Cellulose and protein isolate	Antimicrobial	64
Green tea	Ethylene vinyl alcohol copolymer (EVOH)	Antioxidant/antimicrobial	51
Rosemary	PP	Insect-repellent	28
	Cellulose acetate	Antimicrobial	65
	Chitosan	Antioxidant/antimicrobial	37 and 38
Sage	Cellulose and protein isolate	Antimicrobial	64
Basil	Isolated fish skin gelatin	Antimicrobial	34
Citronella	PP	Insect-repellent	28
Thyme	Isolated soy protein	Antioxidant	40
	Pectin	Antimicrobial	66
	Chitosan	Antimicrobial	39

3. Spices

Spices have been traditionally used as food additives to improve their organoleptic characteristics as well as to extend shelf-life by the inhibition of pathogenic bacteria, yeast and moulds and the reduction of lipid oxidation, which is associated to the presence of terpenoids, flavonoids and phenolic compounds.^{67,68} Therefore, the use of spice extracts as active agents in food packaging has been suggested and studied since they provide antioxidant and antimicrobial properties to packaging materials⁶⁹ acting as potential alternatives to food synthetic preservatives.⁷⁰ The bioactive effects of the main spice extracts incorporated to food packaging films or coatings in the last few years are summarized in Table 3.

Spice extracts have been usually incorporated as EOs into natural or synthetic polymers in food packaging applications. Some authors evaluated the direct incorporation of spice powders or their aqueous extracts into natural polymer matrices for food protective packaging, but no satisfactory results were reported up to now. For instance, Li *et al.*¹⁴ studied the effect of ginger extract (water soluble) into fish gelatins with no improvement in their antioxidant properties (measured by the DPPH radical scavenging method). In other study, Kechichian *et al.*⁷¹ evaluated the antimicrobial effect of cassava starch films with clove and cinnamon powders in contact with bread slices.

They concluded that the antimicrobial effect of this combination of spices could not be clearly assessed since water activity of the bread slices increased considerably during storage, resulting in good conditions for microorganisms' proliferation.

As in the case of herbs, biopolymers such as starch, cellulose derivatives, chitosan, proteins or gums have been proposed as carrier matrices for spice extracts (Table 3). EOs from spices have been usually incorporated into polymer matrices by casting^{72–77} or surface coating.^{29,55,78,79} New active packaging materials consisting of self-adhesive PP labels with cinnamon EOs inside PET trays were used to extend the shelf-life in late-maturing peach fruits.⁸⁰ Authors reported the high decrease in the number of infected fruits after 12 days at room temperature when the active label was used (13%), in comparison with the non-active packaged peaches (86%).

The extraction method selected to obtain EOs from spices plays a key role in their composition and final quality,⁹² modifying their antimicrobial activity.⁹³ Nevertheless, the current use of EOs extracted from spices in food packaging is still limited by their volatility and low stability against oxygen and light during processing and storage. Microencapsulation technologies have emerged as a promising alternative to control the release of EOs onto food surfaces and to increase stability against environmental factors. Kim *et al.*^{78,79} developed laminated films based on PP and LDPE coated with a printing ink containing

Table 3 Spice extracts or essential oils and their application in active food packaging

Spice extract	Polymer	Effect on food packaging	References
Cinnamon EO	Cellulose acetate	Alteration of microstructures and mechanical properties	72
	PP	Sensory evaluation: increase in shelf-life from 3–10 days	81
	Cassava starch	Antimicrobial	76 and 77
	Self-adhesive PP active label inside a PET tray	Antioxidant, antifungal and inhibition of oxidative enzymes	80
	Chitosan	Antimicrobial	82
	PP coated with an organic-base formulation with EO	Antimicrobial, antifungal	29
Cinnamon EO fortified	PP	Antifungal, antimycotoxigenic	83
Cinnamon EO microencapsulated	LDPE-PP	Insect-repelling agent to protect food from Indian meal moth (<i>Plodia interpunctella</i>)	78 and 79
Cinnamon EO nanoliposomes	Fish gelatin	Antimicrobial stability and decrease of release rate	84
Ginger extract	Fish skin gelatin/glycerol 30% (w/w)	Antioxidant, physical and mechanical changes	34
Turmeric oleoresin encapsulated	Gelatin–gum Arabic	Improve stability to light	85
Clove EO	Chicken feather protein/gelatin	Antimicrobial and antioxidant activity on smoked salmon	86
	Fish protein	Antioxidant/antimicrobial	74
	Cassava bagasse/PVA/glycerol (trays)	Antimicrobial	55
	Sunflower protein	Antioxidant, antimicrobial and lipid oxidation on sardine patties	87
	Gelatin/chitosan	Antimicrobial on fish during chilled storage	88
Clove EO (coarse and nanoemulsion)	Methylcellulose/PEG	Antimicrobial activity on sliced bread	62
Cinnamon and lavender (EOs)	NaAlg/glycerol	Antimicrobial, antifungal	73
Cinnamon and cumin (EOs)	Whey protein	Antimicrobial activity on fresh beef	75
Cinnamon, clove and ginger (EOs)	PP	Antioxidant	89
Cinnamon, clove and red pepper powders	Cassava starch	Antimicrobial effect on bread slices	71
Allspice, cinnamon and clove bud EOs	Edible apple films	Antimicrobial activity	90
Asian (nutmeg, lemongrass and citral)	Methylcellulose (MC), PCL/ALG	Antibacterial effect on pre-cut broccoli	42
and Italian spice EOs (oregano and lemongrass)	PCL/MC (EO)/PCL	Antimicrobial activity on pre-cut broccoli	91

microencapsulated cinnamon EO to protect food from the Indian meal moth (*Plodia interpunctella*). Authors reported that these films effectively inhibited the invasion of moth larvae in cookies, milk, chocolate and caramel, acting as insect-resistant films. They observed that microencapsulation decreased the release rate of cinnamaldehyde EO and increased their thermal stability, with no effect on the tensile and moisture barrier properties of the active film. Zuanon *et al.*⁸⁵ observed that the complex coacervation of turmeric oleoresin (a natural pigment) using gelatin and Arabic gum as wall materials was a feasible process to produce microcapsules with suitable colour, physical attributes and high curcumin retention. However, this process did not preserve the pigment under natural radiation.

Other strategies to incorporate EOs into water-soluble polymers to form antimicrobial films have been reported. Otoni *et al.*⁶² prepared coarse emulsions (1.3–1.9 μm diameter) and nanoemulsions (180–250 nm diameter) of clove bud EOs through low-speed mixing and ultrasonication, respectively, to be incorporated into methylcellulose matrices. They observed that droplet size reduction provided further improvement in

antimicrobial properties against yeasts and moulds in sliced bread. In addition, low EOs contents might be used if encapsulated in smaller particles to keep antimicrobial efficiency. However, the EO emulsions reduced the rigidity and increased the extensibility of films and these effects were more pronounced for nanodroplets. Some increase in the antimicrobial stability with the decrease in cinnamon EOs release rate was observed for gelatin films incorporated with cinnamon EO nanoliposomes.⁸⁴ In order to minimize organoleptic effects, Bentayeb *et al.*⁸⁹ prepared films with higher thickness and low concentration of EOs.

EOs extracted from cinnamon and cloves were studied with successful incorporation into natural and synthetic polymers, providing bioactive properties to films (Table 3). Cinnamon oil is mainly composed of cinnamaldehyde (60.4%), with eugenol, linalool and 1,8-cineole as minor compounds.⁸² These EOs have been reported to show antimicrobial, antibacterial, antifungal, insect-repellent and antioxidant activities.^{69,94}

In the case of clove EOs, the major component is eugenol, with eugenyl-acetate, β -caryophyllene and 2-heptanone as

minor compounds, with variable proportions depending on the type and origin of the plant and the extraction method.⁹⁵ Clove oil exhibited antimicrobial activity against moulds, yeasts and bacteria, and also antifungal and antioxidant properties when incorporated into natural polymers (Table 3).

The addition of spice extracts with antioxidant and/or antimicrobial properties into a polymer matrix could affect the physical, morphological, mechanical, thermal and gas barrier properties of the film. In this sense, significant decrease in tensile strength and elongation at break of films with spice extracts were reported.^{34,73,74,76} However, Wu *et al.*⁸⁴ studied the incorporation of cinnamon EOs to fish gelatin resulting in more flexible films. On the other hand, some positive effects related with the decrease in water solubility and water vapour permeability of films were observed by some authors.^{74,76,84,87}

4. Active compounds

As previously reported, essential oils have been thoroughly evaluated in their antimicrobial, antifungal, antiviral, insecticidal and antioxidant properties, but the most innovative approaches in this area come from the use of their bioactive compounds.^{12,69,96} For this reason, current trends in food packaging research include the development of active systems based on materials able to host a variety of bioactive compounds, extracted from herbs and spices.^{69,98} The addition of bioactive compounds with antioxidant/antimicrobial properties directly to food or packaging materials depends primarily on their activity against the targeted microorganisms and oxidative degradation, and their ability to be released at controlled rate during storage and distribution, resulting in extension of food shelf-life.⁹⁹ Nevertheless, other factors, including specific activity of bioactive chemicals, resistance of microorganisms to their action, controlled release mechanisms, chemical nature of the packaged food, physical and mechanical properties of packaging materials, organoleptic properties and potential toxicity of additives should be considered to design packaging systems with bioactive compounds.^{17,69} The mechanical, gas barrier, thermal and morphological properties of films can be altered by the incorporation of bioactive additives. In consequence, the evaluation of these properties will help to understand the interaction of polymer matrices with additives as well as to set processing parameters for fabrication of bioactive films.

Bastarrachea *et al.*¹⁰⁰ reported a significant change in tensile properties of polymer films after the incorporation of some antimicrobials. Barrier properties were evaluated with different gases with significant effect of additives, mostly but not always negative. Authors discussed the addition of antimicrobials and their influence in permeability to gases by changing their solubility or due to the creation of pinholes in the packaging structure depending on the chemical structure of the antimicrobial agent.¹⁰⁰ In addition, some modifications in crystallinity and glass transition temperatures were observed.¹⁹ Scanning electron microscopy studies helped to explain structural modifications induced by the incorporation of additives to

polymer matrices, by detecting formation of pores, thereby influencing the tensile and gas barrier properties of the film.¹⁰¹

Diffusion of additive molecules through films has been also studied and mathematical models were proposed to describe the mass transfer of additive molecules and other substances through the polymer bulk to the surface. Most of these models were based on Fick's laws helping in the assessment of compliance with specific migration limits and describing the effective transport of migrating species with time.¹⁰² Modifications in diffusion coefficients of additive molecules through the polymer structure can be related with formation/rupture of hydrogen bonds, hydrophobic and/or electrostatic interactions. Some authors proposed kinetic models where high initial rates were observed with further observation of a plateau at different times and temperatures.^{103–105} This behaviour was attributed to hydrogen bonding effects between additives and polymer matrices and/or to tortuosity effects within polymer macromolecular structures.¹⁰⁶

Many active packaging materials have been developed in the last few years with bioactive compounds extracted from spices and herbs. Typically, in antimicrobial and antioxidant packaging systems, additives are incorporated in different ways:¹⁰⁷ (a) incorporation by melt-blending before extrusion where a masterbatch is obtained with further injection to obtain films; (b) solvent-compounding and solution coating to prevent the loss of volatile additives¹⁰⁷ where solutions can be incorporated to the polymer matrix as thin layers or coatings by spraying, casting or lamination after extrusion;^{17,108} and (c) encapsulation¹⁰⁹ to permit the formation of a physical barrier between the external media and sensitive active compounds to be protected against oxidation and changes in moisture and pH. Encapsulated microparticles can be produced by spray-drying, spray-cooling/chilling, coacervation, gelation, solvent evaporation, extrusion methods and supercritical fluid expansion;⁷⁹ (d), grafting,¹¹⁰ to permit the covalent binding of active compounds to natural and synthetic polymers.

Fig. 1 summarizes the chemical structures of the main bioactive compounds incorporated into or coated onto packaging films in the last years. The most common bioactive additives are those with antioxidant performance, particularly phenols, such as carvacrol, tocopherols and thymol. These natural compounds can be found in fruits, vegetables, cereals, tea, oils, and many herbs and spices.¹¹¹ Their action against the lipidic auto-oxidation is based on the phenols' ability to release H-atoms from their hydroxyl groups and further reaction with peroxy radicals to produce stable phenoxyl radicals to terminate lipid peroxidation chain reactions.^{112,113} The efficiency of the extracted chemicals in their antioxidant activity depends on the electronic and steric effects of the phenol ring, their substituents and the strength of the hydrogen-bonding interactions between phenol and solvents.⁹⁸

Manzanarez-López *et al.* developed new films for food packaging based on PLA with 2.58 wt% of α -tocopherol.⁹⁹ They evaluated the optical and thermal properties of these films as well as the kinetics of the antioxidant diffusion from the PLA matrix to ethanol and vegetable oil. Results showed that the diffusion of α -tocopherol into oil was slower than to ethanol

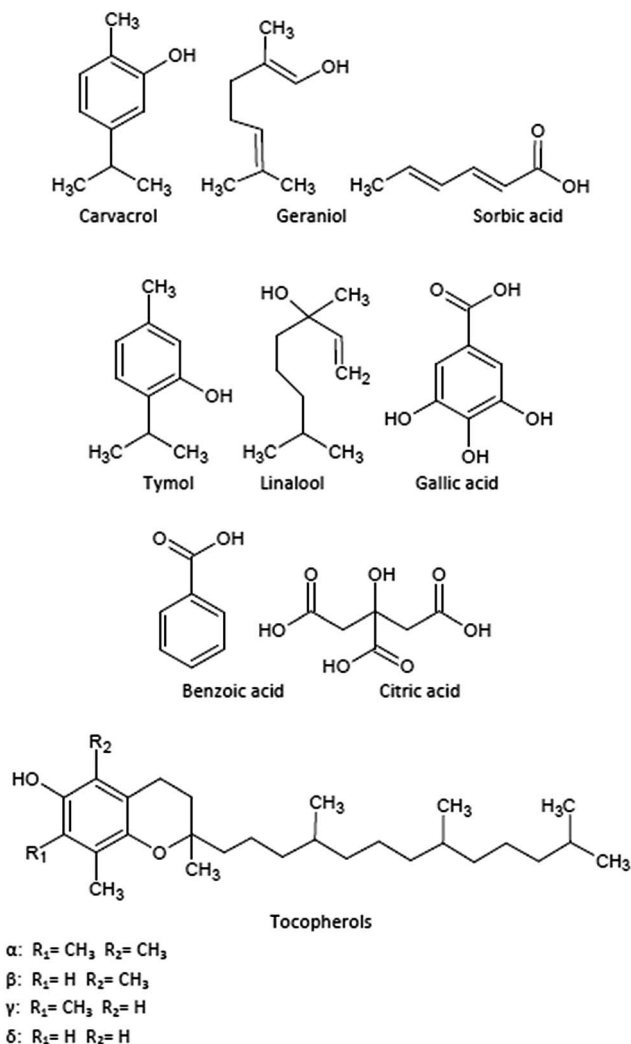


Fig. 1 Main bioactive compounds used in polymer-based films.

with 5.1 and 12.9%, respectively, after 10 days of contact time. They also studied the release of α -tocopherol from films to soybean oil with the aim to delay the induction of oxidation in food stored at 20 and 30 °C.⁹⁹

Another interesting alternative in the use of PLA in active packaging formulations by using casting processing was reported by Jamshidian *et al.*¹¹⁴ who obtained films with natural antioxidants including α -tocopherol and synthetic commercial phenols, such as butylated hydroxyanisole (BHA), butylated hydroxytoluene (BHT), propyl gallate or *tert*-butylhydroquinone (TBHQ). Their release in different food simulants was studied to evaluate the kinetic coefficients to support the continuous control of the oxidation reactions in diverse foodstuff.

Other biopolymer matrices have been tested as host for active components. For example, edible polymer films comprise polysaccharides, lipids, proteins or blends of these macromolecules. Cellulose-based edible films, such as methylcellulose or chitosan,¹¹⁵ were recently proposed to develop antioxidant active packaging materials based on the incorporation of α -tocopherol to this blend by encapsulation of this additive to the chitosan matrix.¹¹⁶ Other authors have proposed poly

(ϵ -caprolactone) (PCL) nanocapsule suspensions to permit a longer permanence of the additive into the polymer matrix.¹¹⁷

As previously stated, carvacrol and thymol are two phenolic monoterpenes obtained from oregano and with high potential as antioxidant active compounds.¹¹⁸ They have received attention due to their natural origin and broad antimicrobial and antifungal spectra against different microorganisms, high antioxidant activity and heat stability when incorporated into packaging materials.^{50,119,120} A wide variety of polymers were used to host carvacrol and/or thymol, and different properties in these active materials were evaluated. Thermal, mechanical and optical performance in these blends showed to be competitive in comparison with common polymers used in food packaging, while the release of these additives from polymer matrices showed adequate rate to understand the transport mechanisms of these molecules to go through polymer matrices and to reach food. LDPE in combination with an organo-modified montmorillonite (MMT) and carvacrol has been proposed as a potential antimicrobial active packaging material.¹²¹ These blends proved to be more effective against *E. coli* than LDPE with just carvacrol without the addition of MMT. This effect was ascribed to the significantly higher carvacrol content permitted by the presence of the nanoclay. Similar results were obtained by Efrati *et al.* for nanocomposites based on LDPE with thymol and different organo-modified montmorillonites.¹²² They found a dependence between the concentration of clay in the film and the desorption mechanism of thymol. Ramos *et al.* prepared PP-based nanocomposites with thymol and carvacrol, reporting that both additives were more effective against gram negative (*E. coli*) than against gram positive (*S. aureus*)¹⁹ bacteria.

A wide variety of antimicrobial materials based on biopolymer matrices has been developed. Some authors based their studies in chitosan with carvacrol as antimicrobial or antioxidant agent.^{123–126} Different procedures were proposed to determine the antimicrobial and antioxidant activities and release in different simulants to determine their performance as materials for active food packaging. Kavosi *et al.* used gelatin solutions with carvacrol at different concentrations (1–5 wt%) to develop antioxidant and antimicrobial films.¹²⁷ They stated that these films exhibited excellent antioxidant and antibacterial properties against both gram positive and gram negative bacteria.

Thymol has been also incorporated into different matrices to enhance their antimicrobial and antioxidant performance to increase biosafety of the packed food.¹²⁸ Wu *et al.* used PLA/PCL blends prepared by solvent casting,¹³⁰ while Ramos *et al.* prepared PLA-based films by melt-blending and hot-pressing with improved thermal, mechanical and gas barrier properties.^{98,101,129} In addition, these films showed enhanced inhibition to different food borne bacteria and excellent antioxidant capacity.

Geraniol and linalool are monoterpenes traditionally used as fragrance/flavour additives in food industry with properties to tackle infectious diseases and/or to preserve food due to their remarkable properties as insect repellent, antimicrobial, antioxidant, anti-inflammatory and anticancer.^{130,131} These are

natural components of EOs of citronella and coriander, respectively, and they have been used to develop antimicrobial active packaging materials with evaluation of their stability and physico-mechanical and optical properties. Kayaci *et al.* based their studies in the development of encapsulation strategies with cyclodextrins to extend the product shelf-life by improving the additives stability.¹²³ Thermogravimetric analysis results showed higher thermal stability in geraniol complexed with cyclodextrins than in pure geraniol permitting an easier incorporation into the polymer matrices. Linalool at relatively low (0.75 ± 0.08 wt%), medium (1.08 ± 0.04 wt%) and high concentrations (3.20 ± 0.29 wt%) were incorporated into thermoplastic starch (TPS).¹³² Authors reported that the direct incorporation of this additive into TPS films did not adversely affect the film properties, particularly at low linalool concentration. These studies need to go deeper into the antimicrobial effect of these films, but some preliminary studies showed their potential application in functional food packaging and other food or medical related applications.

Organic acids, such as benzoic, gallic, citric and sorbic, also showed strong antimicrobial and/or antioxidant activities and they have been used as food preservatives.¹³³ They have been also proposed as active additives in food packaging materials, since they have been included in a positive list by the Food and Drug Administration (FDA) and they are compounds generally recognized as safe (GRAS).^{134,135} A recent study showed that the incorporation of citric acid to linear LDPE, besides showing enhanced antimicrobial and antioxidant properties in meat samples stored under refrigeration conditions, could also act as plasticizer in films.¹³⁵ Sorbic and benzoic acids and their salts are also active against yeast, moulds and many bacteria.¹¹⁵ Da Rocha *et al.* found that protein-based edible films with 1.50 wt% of sorbic or benzoic acids inhibited the proliferation of *E. coli* O157:H7, *Listeria monocytogenes* and *Salmonella enteritidis*; but they did not inhibit the growing of *S. aureus*.¹³⁴ Manab *et al.* showed that benzoic acid incorporated to whey protein-based films significantly inhibited *E. coli* and *Salmonellas*.¹³⁶

5. Final remarks

The rising number of studies presented in the last years on the incorporation of natural active additives to polymer and more specifically to biopolymer matrices has shown the great potential of these formulations to improve selected properties in food packaging materials and to protect the packaged food in a more efficient way. This review focused on studies with natural additives obtained from herbs and spices, either getting directly some of the valuable components from plants or, more commonly, by extraction of chemicals with antioxidant/antimicrobial properties. As a conclusion of this exhaustive study, it should be highlighted that herbs and spices have shown great potential as sources of highly valuable compounds to be further used in different applications, particularly in active food packaging. Nevertheless deeper insight in the interactions of these additives with biopolymer matrices is necessary and further studies are currently on-going.

Acknowledgements

Authors would like to thank the Spanish Ministry of Economy and Competitiveness (MINECO) by their financial support (Project MAT2011-28648-C02-01). Moreover, Marina Ramos would like to thank University of Alicante (Spain) for UAFPU2011-48539721S predoctoral research grant.

References

- 1 M. Viuda-Martos, E. S. Zapata, J. Fernández-López and J. A. Pérez-Álvarez, *Flavour Fragrance J.*, 2010, **25**, 13–19.
- 2 B. Ranton, *Handbook of herbs and spices*, CRC Press, 2004.
- 3 FAO, Last access December 2014.
- 4 A. Vallverdú-Queralt, J. Regueiro, M. Martínez-Huélamo, J. F. Rinaldi Alvarenga, L. N. Leal and R. M. Lamuela-Raventos, *Food Chem.*, 2014, **154**, 299–307.
- 5 T. Kulisic, A. Radonic, V. Katalinic and M. Milos, *Food Chem.*, 2004, **85**, 633–640.
- 6 R. I. Shobha, C. U. Rajeshwari and B. Andallu, in *Cancer*, ed. V. Preedy, Academic Press, San Diego, 2014, pp. 91–100.
- 7 S. Kamiloglu, E. Capanoglu, O. Yilmaz, A. F. Duran and D. Boyacioglu, *Qual. Assur. Saf. Crops Foods*, 2014, **6**, 151–158.
- 8 K. Srinivasan, *Crit. Rev. Food Sci. Nutr.*, 2014, **54**, 352–372.
- 9 P. Singh, A. A. Wani and S. Saengerlaub, *Nutr. Food Sci.*, 2011, **41**, 249–260.
- 10 P. Sanchez-Bel, I. Egea, M. T. Pretel, F. B. Flores, F. Romojaro and M. C. Martínez-Madrid, *Food Sci. Technol. Int.*, 2011, **17**, 529–540.
- 11 A. Valdés, A. Beltrán and M. C. Garrigós, *J. Food Sci.*, 2013, **78**, C138–C144.
- 12 A. Silva-Weiss, M. Ihl, P. J. A. Sobral, M. C. Gómez-Guillén and V. Bifani, *Food Eng. Rev.*, 2013, **5**, 200–216.
- 13 A. Valdés, A. C. Mellinas, M. Ramos, M. C. Garrigós and A. Jiménez, *Front. Chem.*, 2014, **2**, 1–10.
- 14 J. H. Li, J. Miao, J. L. Wu, S. F. Chen and Q. Q. Zhang, *Food Hydrocolloids*, 2014, **37**, 166–173.
- 15 M. Jouki, F. T. Yazdi, S. A. Mortazavi and A. Koocheki, *Food Hydrocolloids*, 2014, **36**, 9–19.
- 16 M. M. Castro, J. M. López and M. V. González, *Food Chem.*, 2014, **150**, 119–127.
- 17 J. Gómez-Estaca, C. López-de-Dicastillo, P. Hernández-Muñoz, R. Catalá and R. Gavara, *Trends Food Sci. Technol.*, 2014, **35**, 42–51.
- 18 S.-Y. Sung, L. T. Sin, T.-T. Tee, S.-T. Bee, A. R. Rahmat, W. A. W. A. Rahman, A.-C. Tan and M. Vikhraman, *Trends Food Sci. Technol.*, 2013, **33**, 110–123.
- 19 M. Ramos, A. Jiménez, M. Peltzer and M. C. Garrigós, *J. Food Eng.*, 2012, **109**, 513–519.
- 20 D. A. Pereira de Abreu, J. Maroto, K. Villalba Rodríguez and J. M. Cruz, *J. Sci. Food Agric.*, 2012, **92**, 427–432.
- 21 S. R. Kanatt, M. S. Rao, S. P. Chawla and A. Sharma, *Food Hydrocolloids*, 2012, **29**, 290–297.
- 22 W. Hu, P. Xu and T. Uchino, *J. Sci. Food Agric.*, 2005, **85**, 2475–2481.

- 23 L. Barbosa-Pereira, G. P. Aurrekoetxea, I. Angulo, P. Paseiro-Losada and J. M. Cruz, *Meat Sci.*, 2014, **97**, 249–254.
- 24 T. Bolumar, M. L. Andersen and V. Orlien, *Food Chem.*, 2011, **129**, 1406–1412.
- 25 O. E. Çoban and O. P. Can, *J. Aquat. Food Prod. Technol.*, 2013, **22**, 361–370.
- 26 A. C. V. Solano and C. R. de Gante, *Food Bioprocess Technol.*, 2012, **5**, 2522–2528.
- 27 V. Otero, R. Becerril, J. A. Santos, J. M. Rodríguez-Calleja, C. Nerín and M.-L. García-López, *Food Control*, 2014, **42**, 296–302.
- 28 F. Licciardello, G. Muratore, P. Suma, A. Russo and C. Nerín, *Innovative Food Sci. Emerging Technol.*, 2013, **19**, 173–180.
- 29 S. Manso, R. Becerril, C. Nerín and R. Gómez-Lus, *Food Control*, 2015, **47**, 20–26.
- 30 J.-H. Li, J. Miao, J.-L. Wu, S.-F. Chen and Q.-Q. Zhang, *Food Hydrocolloids*, 2014, **37**, 166–173.
- 31 J. Wu, S. Chen, S. Ge, J. Miao, J. Li and Q. Zhang, *Food Hydrocolloids*, 2013, **32**, 42–51.
- 32 B. Giménez, A. López de Lacey, E. Pérez-Santín, M. E. López-Caballero and P. Montero, *Food Hydrocolloids*, 2013, **30**, 264–271.
- 33 A. M. López de Lacey, M. E. López-Caballero and P. Montero, *LWT-Food Sci. Technol.*, 2014, **55**, 559–564.
- 34 Y. A. Arfat, S. Benjakul, T. Prodpran, P. Sumpavapol and P. Songtipya, *Food Hydrocolloids*, 2014, **41**, 265–273.
- 35 U. Siripatrawan and S. Noipha, *Food Hydrocolloids*, 2012, **27**, 102–108.
- 36 U. Siripatrawan and B. R. Harte, *Food Hydrocolloids*, 2010, **24**, 770–775.
- 37 M. Abdollahi, M. Rezaei and G. Farzi, *Int. J. Food Sci. Technol.*, 2012, **47**, 847–853.
- 38 M. Abdollahi, M. Rezaei and G. Farzi, *J. Food Eng.*, 2012, **111**, 343–350.
- 39 M. H. Hosseini, S. H. Razavi, S. M. A. Mousavi, S. A. S. Yasaghi and A. G. Hasansaraei, *J. Appl. Sci.*, 2008, **8**, 2895–2900.
- 40 B. Kodal Coşkun, E. Çalikoğlu, Z. Karagöz Emiroğlu and K. Candoğan, *J. Food Qual.*, 2014, **37**, 203–212.
- 41 K. Norajit, K. M. Kim and G. H. Ryu, *J. Food Eng.*, 2010, **98**, 377–384.
- 42 P. N. Takala, K. D. Vu, S. Salmieri, R. A. Khan and M. Lacroix, *LWT-Food Sci. Technol.*, 2013, **53**, 499–506.
- 43 C. Ramírez, I. Gallegos, M. Ihl and V. Bifani, *J. Food Eng.*, 2012, **109**, 424–429.
- 44 M. Quilaqueo Gutiérrez, I. Echeverría, M. Ihl, V. Bifani and A. N. Mauri, *Carbohydr. Polym.*, 2012, **87**, 1495–1502.
- 45 J. M. Lorenzo, R. Batlle and M. Gómez, *LWT-Food Sci. Technol.*, 2014, **59**, 181–188.
- 46 C. López De Dicastillo, C. Nerín, P. Alfaro, R. Catalá, R. Gavara and P. Hernández-Muñoz, *J. Agric. Food Chem.*, 2011, **59**, 7832–7840.
- 47 C. López-de-Dicastillo, J. Gómez-Estaca, R. Catalá, R. Gavara and P. Hernández-Muñoz, *Food Chem.*, 2012, **131**, 1376–1384.
- 48 K. K. N. L. C. L. Perazzo, A. C. V. Conceição, J. C. P. Dos Santos, D. D. J. Assis, C. O. Souza and J. I. Druzian, *PLoS One*, 2014, **9**, DOI: 10.1371/journal.pone.0105199.
- 49 F. Tian, E. A. Decker and J. M. Goddard, *Food Funct.*, 2013, **4**, 669–680.
- 50 K. K. Kuorwel, M. J. Cran, K. Sonneveld, J. Miltz and S. W. Bigger, *J. Food Sci.*, 2011, **76**, R164–R177.
- 51 V. Muriel-Galet, M. J. Cran, S. W. Bigger, P. Hernández-Muñoz and R. Gavara, *J. Food Eng.*, 2015, **149**, 9–16.
- 52 V. Muriel-Galet, J. P. Cerisuelo, G. López-Carballo, S. Aucejo, R. Gavara and P. Hernández-Muñoz, *Food Control*, 2013, **30**, 137–143.
- 53 S. Salmieri, F. Islam, R. A. Khan, F. M. Hossain, H. M. M. Ibrahim, C. Miao, W. Y. Hamad and M. Lacroix, *Cellulose*, 2014, **21**, 4271–4285.
- 54 N. M. Hromiš, V. L. Lazić, S. L. Markov, Ž. G. Vaštag, S. Z. Popović, D. Z. Šuput and N. R. Džinić, *Acta Period. Technol.*, 2014, **45**, 33–43.
- 55 F. Debiagi, R. K. T. Kobayashi, G. Nakazato, L. A. Panagio and S. Mali, *Ind. Crops Prod.*, 2014, **52**, 664–670.
- 56 S. Shojaei-Aliabadi, H. Hosseini, M. A. Mohammadifar, A. Mohammadi, M. Ghasemlou, S. M. Hosseini and R. Khaksar, *Carbohydr. Polym.*, 2014, **101**, 582–591.
- 57 S. Shojaei-Aliabadi, M. A. Mohammadifar, H. Hosseini, A. Mohammadi, M. Ghasemlou, S. M. Hosseini, M. Haghshenas and R. Khaksar, *Int. J. Biol. Macromol.*, 2014, **69**, 282–289.
- 58 D. Salarbashi, S. Tajik, S. Shojaei-Aliabadi, M. Ghasemlou, H. Moayyed, R. Khaksar and M. S. Noghabi, *Food Chem.*, 2014, **146**, 614–622.
- 59 S. Shojaei-Aliabadi, H. Hosseini, M. A. Mohammadifar, A. Mohammadi, M. Ghasemlou, S. M. Ojagh, S. M. Hosseini and R. Khaksar, *Int. J. Biol. Macromol.*, 2013, **52**, 116–124.
- 60 M. Jouki, F. T. Yazdi, S. A. Mortazavi and A. Koocheki, *Food Hydrocolloids*, 2014, **36**, 9–19.
- 61 L. A. Cestari, R. C. Gaiotto, J. L. Antigo, M. R. S. Scapim, G. S. Madrona, F. Yamashita, M. S. S. Pozza and I. N. Prado, *J. Food Sci. Technol.*, 2014, DOI: 10.1007/s13197-014-1357-z.
- 62 C. G. Otoni, S. F. O. Pontes, E. A. A. Medeiros and N. d. F. F. Soares, *J. Agric. Food Chem.*, 2014, **62**, 5214–5219.
- 63 V. Muriel-Galet, J. P. Cerisuelo, G. López-Carballo, M. Lara, R. Gavara and P. Hernández-Muñoz, *Int. J. Food Microbiol.*, 2012, **157**, 195–201.
- 64 M. Royo, I. Fernández-Pan and J. I. Maté, *J. Sci. Food Agric.*, 2010, **90**, 1513–1519.
- 65 A. A. M. de Melo, R. M. Geraldine, M. F. A. Silveira, M. C. L. Torres, C. S. M. e. Rezende, T. H. Fernandes and A. N. de Oliveira, *Braz. J. Microbiol.*, 2012, **43**, 1419–1427.
- 66 P. J. P. Espitia, R. J. Avena-Bustillos, W.-X. Du, R. F. Teófilo, N. F. F. Soares and T. H. McHugh, *Food Packaging and Shelf Life*, 2014, **2**, pp. 38–49.
- 67 P. S. Negi, *Int. J. Food Microbiol.*, 2012, **156**, 7–17.
- 68 M. M. Tajkarimi, S. A. Ibrahim and D. O. Cliver, *Food Control*, 2010, **21**, 1199–1218.

- 69 F. Sadaka, C. Nguimjeu, C. H. Brachais, I. Vroman, L. Tighzert and J. P. Couvercelle, *Innovative Food Sci. Emerging Technol.*, 2014, DOI: 10.1016/j.ifset.2014.03.002.
- 70 J. Gómez-Estaca, C. López-de-Dicastillo, P. Hernández-Muñoz, R. Catalá and R. Gavara, *Trends Food Sci. Technol.*, 2014, **35**, 42–51.
- 71 V. Kechichian, C. Ditchfield, P. Veiga-Santos and C. C. Tadini, *LWT–Food Sci. Technol.*, 2010, **43**, 1088–1094.
- 72 P. J. P. Espitia, N. D. F. F. Soares, L. C. M. Botti and W. A. Silva, *Macromol. Symp.*, 2011, **299–300**, 199–205.
- 73 I. Liakos, L. Rizzello, D. J. Scurr, P. P. Pompa, I. S. Bayer and A. Athanassiou, *Int. J. Pharm.*, 2014, **463**, 137–145.
- 74 B. Teixeira, A. Marques, C. Pires, C. Ramos, I. Batista, J. A. Saraiva and M. L. Nunes, *LWT–Food Sci. Technol.*, 2014, **59**, 533–539.
- 75 K. R. Badr, Z. S. Ahmed and M. S. El Gamal, *Int. J. Agric. Res.*, 2014, **9**, 242–250.
- 76 S. Bahram, M. Rezaei, M. Soltani, A. Kamali, S. M. Ojagh and M. Abdollahi, *J. Food Process. Preserv.*, 2014, **38**, 1251–1258.
- 77 A. C. Souza, G. E. O. Goto, J. A. Mainardi, A. C. V. Coelho and C. C. Tadini, *LWT–Food Sci. Technol.*, 2013, **54**, 346–352.
- 78 I.-H. Kim, J. Han, J. H. Na, P.-S. Chang, M. S. Chung, K. H. Park and S. C. Min, *J. Food Sci.*, 2013, **78**, E229–E237.
- 79 I.-H. Kim, A. Y. Song, J. Han, K. H. Park and S. C. Min, *J. Food Sci.*, 2014, **79**, E2023–E2030.
- 80 P. Montero-Prado, A. Rodríguez-Lafuente and C. Nerin, *Postharvest Biol. Technol.*, 2011, **60**, 211–219.
- 81 L. Gutiérrez, C. Sánchez, R. Batlle and C. Nerin, *Trends Food Sci. Technol.*, 2009, **20**, 92–99.
- 82 S. M. Ojagh, M. Rezaei, S. H. Razavi and S. M. H. Hosseini, *Food Chem.*, 2010, **122**, 161–166.
- 83 S. Manso, D. Pezo, R. Gómez-Lus and C. Nerin, *Food Control*, 2014, **45**, 101–108.
- 84 J. Wu, H. Liu, S. Ge, S. Wang, Z. Qin, L. Chen, Q. Zheng, Q. Liu and Q. Zhang, *Food Hydrocolloids*, 2015, **43**, 427–435.
- 85 L. A. C. Zuanon, C. R. Malacrida and V. R. N. Telis, *J. Food Process Eng.*, 2013, **36**, 364–373.
- 86 N.-B. Song, J.-H. Lee, M. Al Mijan and K. B. Song, *LWT–Food Sci. Technol.*, 2014, **57**, 453–460.
- 87 P. R. Salgado, M. E. López-Caballero, M. C. Gómez-Guillén, A. N. Mauri and M. P. Montero, *Food Hydrocolloids*, 2013, **33**, 74–84.
- 88 J. Gómez-Estaca, A. López de Lacey, M. E. López-Caballero, M. C. Gómez-Guillén and P. Montero, *Food Microbiol.*, 2010, **27**, 889–896.
- 89 K. Bentayeb, P. Vera, C. Rubio and C. Nerin, *Anal. Bioanal. Chem.*, 2009, **394**, 903–910.
- 90 W. X. Du, C. W. Olsen, R. J. Avena-Bustillos, T. H. McHugh, C. E. Levin and M. Friedman, *J. Food Sci.*, 2009, **74**, M372–M378.
- 91 P. N. Takala, S. Salmieri, A. Boumail, R. A. Khan, K. D. Vu, G. Chauve, J. Bouchard and M. Lacroix, *J. Food Eng.*, 2013, **116**, 648–655.
- 92 M. Bendahou, A. Muselli, M. Grignon-Dubois, M. Benyoucef, J.-M. Desjobert, A.-F. Bernardini and J. Costa, *Food Chem.*, 2008, **106**, 132–139.
- 93 M. Lahlou, *Phytother. Res.*, 2004, **18**, 435–448.
- 94 H. J. Jo, K. M. Park, S. C. Min, J. H. Na, K. H. Park and J. Han, *J. Food Sci.*, 2013, **78**, E1713–E1720.
- 95 K. Chaieb, H. Hajlaoui, T. Zmantar, A. B. Kahla-Nakbi, M. Rouabhia, K. Mahdouani and A. Bakhrouf, *Phytother. Res.*, 2007, **21**, 501–506.
- 96 S. Burt, *Int. J. Food Microbiol.*, 2004, **94**, 223–253.
- 97 S. Kordali, A. Cakir, H. Ozer, R. Cakmakci, M. Kesdek and E. Mete, *Bioresour. Technol.*, 2008, **99**, 8788–8795.
- 98 M. Ramos, A. Jiménez, M. Peltzer and M. C. Garrigós, *Food Chem.*, 2014, **162**, 149–155.
- 99 F. Manzanarez-López, H. Soto-Valdez, R. Auras and E. Peralta, *J. Food Eng.*, 2011, **104**, 508–517.
- 100 L. Bastarrachea, S. Dhawan and S. Sablani, *Food Eng. Rev.*, 2011, **3**, 79–93.
- 101 M. Ramos, E. Fortunati, M. Peltzer, F. Dominici, A. Jiménez, M. d. C. Garrigós and J. M. Kenny, *Polym. Degrad. Stab.*, 2014, **108**, 158–165.
- 102 M. F. Poças, J. C. Oliveira, F. A. R. Oliveira and T. Hogg, *Crit. Rev. Food Sci. Nutr.*, 2008, **48**, 913–928.
- 103 A. Z. Graciano-Verdugo, H. Soto-Valdez, E. Peralta, P. Cruz-Zárte, A. R. Islas-Rubio, S. Sánchez-Valdes, A. Sánchez-Escalante, N. González-Méndez and H. González-Ríos, *Food Res. Int.*, 2010, **43**, 1073–1078.
- 104 F. Iñiguez-Franco, H. Soto-Valdez, E. Peralta, J. F. Ayala-Zavala, R. Auras and N. Gámez-Meza, *J. Agric. Food Chem.*, 2012, **60**, 6515–6523.
- 105 M. Ramos, A. Beltrán, M. Peltzer, A. J. M. Valente and M. d. C. Garrigós, *LWT–Food Sci. Technol.*, 2014, **58**, 470–477.
- 106 K. K. Kuorwel, M. J. Cran, K. Sonneveld, J. Miltz and S. W. Bigger, *LWT–Food Sci. Technol.*, 2013, **50**, 432–438.
- 107 *Innovations in Food Packaging*, ed. J. H. Han, Elsevier Academic Press, Canada, 2005.
- 108 M. Ramos, A. Beltran, A. Valdes, M. A. Peltzer, A. Jimenez and M. C. Garrigos, *J. Bioequivalence Bioavailability*, 2013, **5**, 154–160.
- 109 H. M. C. Marques, *Flavour Fragrance J.*, 2010, **25**, 313–326.
- 110 S. B. Schreiber, J. J. Bozell, D. G. Hayes and S. Zivanovic, *Food Hydrocolloids*, 2013, **33**, 207–214.
- 111 J. Mastelic, I. Jerkovic, I. Blazevic, M. Poljak-Blazi, S. Borovic, I. Ivancic-Bace, V. Smrecki, N. Zarkovic, K. Brcic-Kostic, D. Vikić-Topic and N. Müller, *J. Agric. Food Chem.*, 2008, **56**, 3989–3996.
- 112 L. Barbosa-Pereira, J. M. Cruz, R. Sendón, A. Rodríguez Bernaldo de Quirós, A. Ares, M. Castro-López, M. J. Abad, J. Maroto and P. Paseiro-Losada, *Food Control*, 2013, **31**, 236–243.
- 113 J. Hughes, R. Thomas, Y. Byun and S. Whiteside, *Carbohydr. Polym.*, 2012, **88**, 165–172.
- 114 M. Jamshidian, E. Tehrany and S. Desobry, *Food Bioprocess Technol.*, 2013, **6**, 1450–1463.
- 115 M. C. Cruz-Romero, T. Murphy, M. Morris, E. Cummins and J. P. Kerry, *Food Control*, 2013, **34**, 393–397.
- 116 M. V. Dias, V. Machado Azevedo, S. V. Borges, N. d. F. F. Soares, R. V. de Barros Fernandes,

- J. J. Marques and É. A. A. Medeiros, *Food Chem.*, 2014, **165**, 323–329.
- 117 C. M. Noronha, S. M. de Carvalho, R. C. Lino and P. L. M. Barreto, *Food Chem.*, 2014, **159**, 529–535.
- 118 G. Al-Bandak and V. Oreopoulou, *Eur. J. Lipid Sci. Technol.*, 2007, **109**, 247–255.
- 119 N. Didry, L. Dubreuil and M. Pinkas, *Pharm. Acta Helv.*, 1994, **69**, 25–28.
- 120 H. J. D. Dorman and S. G. Deans, *J. Appl. Microbiol.*, 2000, **88**, 308–316.
- 121 R. Shemesh, D. Goldman, M. Krepker, Y. Danin-Poleg, Y. Kashi, A. Vaxman and E. Segal, *J. Appl. Polym. Sci.*, 2015, **132**, n/a-n/a.
- 122 R. Efrati, M. Natan, A. Pelah, A. Haberer, E. Banin, A. Dotan and A. Ophir, *J. Appl. Polym. Sci.*, 2014, **131**, 40564.
- 123 L. Higuera, G. López-Carballo, P. Hernández-Muñoz, R. Catalá and R. Gavara, *Int. J. Food Microbiol.*, 2014, **188**, 53–59.
- 124 M. Kurek, A. Guinault, A. Voilley, K. Galić and F. Debeaufort, *Food Chem.*, 2014, **144**, 9–17.
- 125 M. López-Mata, S. Ruiz-Cruz, N. Silva-Beltrán, J. Ornelas-Paz, P. Zamudio-Flores and S. Burrueal-Ibarra, *Molecules*, 2013, **18**, 13735–13753.
- 126 M. Kurek, S. Moundanga, C. Favier, K. Galić and F. Debeaufort, *Food Control*, 2013, **32**, 168–175.
- 127 G. Kavosi, S. M. M. Dadfar, A. Mohammadi Purfard and R. Mehrabi, *J. Food Saf.*, 2013, **33**, 423–432.
- 128 N. Khairuddin and I. I. Muhamad, *Sciences and Engineering*, 2013, **62**, 31–34.
- 129 Y. Wu, Y. Qin, M. Yuan, L. Li, H. Chen, J. Cao and J. Yang, *Polym. Adv. Technol.*, 2014, **25**, 948–954.
- 130 N. G. Sahib, F. Anwar, A.-H. Gilani, A. A. Hamid, N. Saari and K. M. Alkharfy, *Phytother. Res.*, 2013, **27**, 1439–1456.
- 131 F. Kayaci, H. S. Sen, E. Durgun and T. Uyar, *Food Res. Int.*, 2014, **62**, 424–431.
- 132 K. K. Kuorwel, M. J. Cran, K. Sonneveld, J. Miltz and S. W. Bigger, *Packag. Technol. Sci.*, 2014, **27**, 149–159.
- 133 K. A. Hammer, C. F. Carson and T. V. Riley, *J. Appl. Microbiol.*, 1999, **86**, 985–990.
- 134 M. d. Rocha, M. R. Loiko, E. C. Tondo and C. Prentice, *Food Hydrocolloids*, 2014, **37**, 213–220.
- 135 A. Júnior, N. Fronza, F. Foralosso, D. Dezen, E. Huber, J. dos Santos, R. Machado and M. Quadri, *Food Bioprocess Technol.*, 2014, 1–13.
- 136 A. Manab, M. E. Sawitri, K. U. Al Awwaly and H. Purnomo, *Afr. J. Food Sci.*, 2011, **1**, 6–11.