



Showcasing research from Dr Constantina Papatriantafyllopoulou's laboratory, School of Chemistry, National University of Ireland Galway, Galway, Ireland.

A biocompatible ZnNa_2 -based metal-organic framework with high ibuprofen, nitric oxide and metal uptake capacity

The Papatriantafyllopoulou research group is focused on the development of new metal-organic frameworks (MOFs) with biomedical and/or environmental applications. Herein, the synthesis and characterization of a ZnNa_2 MOF (**NUIG1**) with a new framework topology is reported; **NUIG1** exhibits an exceptionally high drug (ibuprofen, NO) adsorption capacity and metal encapsulation potential. The magnetic properties of **NUIG1** are affected by the type and quantity of the encapsulated species.

As featured in:



See Liam Morrison, George Froudakis, Constantina Papatriantafyllopoulou *et al.*, *Mater. Adv.*, 2020, 1, 2248.

Cite this: *Mater. Adv.*, 2020,
1, 2248

A biocompatible ZnNa₂-based metal–organic framework with high ibuprofen, nitric oxide and metal uptake capacity†

Meghan Winterlich,^{ab} Constantinos G. Efthymiou,^b Wassillios Papawassiliou,^c Jose P. Carvalho,^c Andrew J. Pell,^{id} Julia Mayans,^d Albert Escuer,^{id} Michael P. Carty,^f Patrick McArdle,^b Emmanuel Tylianakis,^g Liam Morrison,^{*h} George Froudakis^{*i} and Constantina Papatriantafyllopoulou^{id} ^{*abj}

Metal organic frameworks (MOFs) have received significant attention in recent years in the areas of biomedical and environmental applications. Among them, mixed metal MOFs, although promising, are relatively few in number in comparison with their homometallic analogues. The employment of benzophenone-4,4'-dicarboxylic acid (bphdcH₂) in mixed metal MOF chemistry provided access to a 3D MOF, [Na₂Zn(bphdc)₂(DMF)₂]_n (**NUIG1**). **NUIG1** displays a new topology and is a rare example of a mixed metal MOF based on 1D rod secondary building units. UV-vis, HPLC, TGA, XRPD, solid state NMR and computational studies indicated that **NUIG1** exhibits an exceptionally high ibuprofen (Ibu) and nitric oxide adsorption capacity. The MCF-7 cell line was used to assess the toxicity of **NUIG1** and Ibu@**NUIG1**, revealing that both species are non-toxic (cell viability > 70%). **NUIG1** exhibits good performance in the adsorption of metal ions (Co^{II}, Ni^{II}, Cu^{II}) from aqueous environments, as was demonstrated by UV-vis, EDX, ICP, SEM and direct and alternate current magnetic susceptibility studies. The colour and the magnetic properties of the M@**NUIG1** species depend strongly on the kind and the amount of the encapsulated metal ion in the MOF pores.

Received 24th June 2020,
Accepted 19th August 2020

DOI: 10.1039/d0ma00450b

rsc.li/materials-advances

Introduction

Metal–organic frameworks (MOFs) are a family of hybrid porous materials that have attracted immense research interest in recent decades due to their appealing structural features.^{1,2} They often possess large surface area, high porosity, flexible structure, an amphiphilic internal microenvironment, and the possibility of introducing functional groups in the pores and frameworks in a spatially controlled way, which make these materials especially suitable for encapsulating a large variety of guest molecules. As such, MOFs can be used in a wide range of applications, e.g. gas storage and/or separation,³ magnetism, catalysis, sensing, drug delivery and imaging related applications.^{4,5}

In relation to the use of MOFs in drug delivery applications, they were introduced into this field nearly ten years ago, with one of the first reports being by Horcajada *et al.*, who showed that the mesoporous MOFs MIL-100 and MIL-101 display remarkable ibuprofen adsorption and release performance.⁶ Since then, many MOFs with high drug uptake have been reported, representative examples being other members of the MIL family, Bio-MOFs, CD-MOFs, UiO-MOFs, ZIF-MOFs, Zr MOFs, *etc.*^{7–9} MOFs offer significant advantages over the currently used drug delivery systems that are based on liposomes, micelles, dendrimers,

^a CÚRAM Centre for Research in Medical Devices, Ireland^b School of Chemistry, College of Science and Engineering, National University of Ireland Galway, H91 TK 33 Galway, Ireland.

E-mail: constantina.papatriantafyllopo@nuigalway.ie; Tel: +353 91 493462

^c Department of Materials and Environmental Chemistry, Stockholm University, Sweden^d Instituto de Ciencia Molecular (ICMol), Universidad de Valencia, Catedrático José Beltrán 2, 46980 Paterna, Valencia, Spain^e Departament de Química Inorgànica i Orgànica, Secció Inorgànica and Institut de Nanociència i Nanotecnologia (IN2UB), Universitat de Barcelona, Martí i Franquès 1-11, 08028 Barcelona, Spain^f Biochemistry, School of Natural Sciences, National University of Ireland Galway, H91 TK 33 Galway, Ireland^g Department of Materials Science and Technology, University of Crete, Voutes Campus,

GR-70013 Heraklion, Crete, Greece

^h Earth and Ocean Sciences and Ryan Institute, School of Natural Sciences, National University of Ireland Galway, H91 TK 33 Galway, Irelandⁱ Department of Chemistry, University of Crete, Voutes Campus, GR-70013 Heraklion, Crete, Greece^j SSPC, Synthesis and Solid State Pharmaceutical Centre, Ireland

† Electronic supplementary information (ESI) available: Experimental data, graphs and tables with fitting parameters for the adsorption studies; thermal analysis graphs. CCDC 2008513. For ESI and crystallographic data in CIF or other electronic format see DOI: 10.1039/d0ma00450b



gold/iron/silica nanoparticles, carbon nanotubes, and quantum dots.^{10–12} MOFs often exhibit high drug upload capacity, biocompatibility, thermal stability, inexpensive and rapid scale-up synthesis, and thus can address major challenges related to the poor active pharmaceutical ingredient (API) stability and/or solubility, burst effect and toxicity.^{13,14} The use of MOFs in drug delivery has recently been expanded to anticancer drugs and the targeted, and controlled delivery has been achieved by a variety of different techniques, such as pH-controlled, photo-dynamic and magnetically triggered drug release.^{15–17}

In a similar vein, MOFs can capture and sense toxic compounds in the environment.^{18–20} The adsorption process may occur *via* a variety of mechanisms, such as interactions between acid and base, electrostatic interactions between adsorbates and adsorbent, H-bonding and π - π stacking, and hydrophobic interactions.^{20c} The removal of toxic metals from water is important and contemporary techniques involve chemical precipitation, ion exchange, membrane filtration and adsorption, with the latter which can employ MOFs being the most preferred as it does not require a high operating temperature. The adsorbents that are currently used either possess high porosity but are prone to aggregation, which decreases their efficiency (*e.g.* magnetic nanoparticles), or have high stability and efficiency, but low surface area (*e.g.* zeolites).²¹ MOFs display all the desirable features to provide an efficient solution to the removal of toxic metals from water, and as such many MOF families (MIL, UiO-66, HKUST, *etc.*) have now been reported for their ability to adsorb toxic metals.²² MOFs often have sensing properties, which is based on the effect of the toxic/guest metal ion to the physical properties (colour, electrochemical, luminescence, *etc.*) of the MOF.^{23,24} One recent such example is the FJI-C8 MOF,²⁴ which exhibits a high sensitivity for Fe³⁺ with the detection limit being 0.0233 mM; it contains a π -conjugated aromatic ligand with free/uncoordinated N- and O- atoms, which interact strongly with the encapsulated Fe³⁺ ions affecting the emission spectrum of the anionic FJI-C8 MOF.

The advantages that MOFs offer in important biomedical and environmental applications, constitute an increasing need for the isolation of new such species with high stability, porosity and fine-tuning properties. One recent approach towards this direction involves the presence of a second metal ion in the MOF structure;²⁵ this often increases the robustness of the framework, which results to high thermal and water stability. Furthermore, the synergism between the different metal ions has been proven to enhance MOF features and functionalities, including, among others, the breathing effect, catalytic activity, *etc.*²⁶ It is noteworthy that although *ca.* 6000 new MOFs are reported per year, relatively few of them are mixed-metal, with the mixed metal MOF field being still in its infancy.²⁵

With the above mentioned in mind, we decided to expand the family of mixed metal MOFs by using benzophenone-4,4'-dicarboxylic acid (bphdcH₂, Scheme 1) as the organic linker; this ligand has been extensively used for the synthesis of homometallic MOFs, yet its employment towards new mixed metal MOFs is limited.²⁷ In particular, the combination of bphdcH₂ with other bridging ligands, including 4,4'-bipyridine (bpy),



Scheme 1 A schematic representation of benzophenone-4,4'-dicarboxylic acid (bphdcH₂) and the two different coordination modes that it adopts in **NUIG1**: $\eta^2:\eta^1:\eta^1:\eta^1:\mu_5$ (**Type A**, left) and $\eta^1:\eta^1:\eta^1:\eta^1:\mu_4$ (**Type B**, right).

4-diazabicyclo[2.2.2]octane (dadco), and others, in Zn^{II} and Cu^{II} coordination chemistry has yielded new 3D MOFs with interesting properties; among them, [Cu₂(bphdc)₂(bpy)]_n and [Zn₂(bphdc)₂(dadco)]_n display very high H₂ and CO₂ adsorption.^{27a} Herein, we report the synthesis and characterization of [Na₂Zn(bphdc)₂(DMF)₂]_n (**NUIG1**), which is a rare example of a mixed-metal MOF based on an 1D rod SBU. Herein, the drug uptake performance of **NUIG1** have been studied and discussed in detail for Ibuprofen and nitric oxide. The metal adsorption capacity of **NUIG1** was also examined using a variety of techniques, including UV-vis, EDX, ICP, SEM and magnetism.

Experimental

All manipulations were performed under aerobic conditions using materials as received.

Synthesis of [Na₂Zn(bphdc)₂(DMF)₂]_n (**NUIG1**)

1 M NaOH (0.5 mL) and Zn(CH₃CO₂)₂·2H₂O (0.03 g, 0.1 mmol) were added to a solution of bphdcH₂ (0.07 g, 0.3 mmol) in DMF/EtOH (10/8 mL) and left under magnetic stirring for 5 min at room temperature. Then, the vial was placed in the oven at 100 °C for 24 hours, after which time white polyhedral crystals of **NUIG1** were observed. The crystals were kept in mother liquor for X-ray analysis or collected by filtration for other solid-state studies. Yield: ~70%. Anal. calcd (found) for **NUIG1**: C, 54.46 (54.71); H, 3.81 (3.66); N, 3.53 (3.64)%. Selected IR data (KBr, cm⁻¹): 3437(b), 2928(b), 1655(s), 1614(m), 1558(w), 1498(w), 1438(w), 1361(m), 1298(w), 1256(m), 1127(w), 1094(s), 1063(w), 1016(w), 933(m), 880(m), 833(s), 811(w), 786(w), 734(s), 705(w), 661(m).

Physical studies

Elemental analyses (C, H, N) were performed by the in-house facilities of the National University of Ireland Galway, School of Chemistry. IR spectra (4000–400 cm⁻¹) were recorded using a PerkinElmer 16PC FT-IR spectrometer with samples prepared as KBr pellets. Powder X-ray diffraction data (XRPD) were collected using an Inex Equinox 6000 diffractometer. Solid-state, variable-temperature and variable-field magnetic data were collected on powdered samples using an MPMS5 Quantum Design magnetometer operating at 0.03 T in the 300 to 2.0 K range for the magnetic susceptibility and at 2.0 K in the 0 to 5 T range for the magnetization measurements. Diamagnetic corrections were applied to the observed susceptibilities using Pascal's constants. TGA experiments were performed on a STA625 thermal analyser from Rheometric Scientific (Piscataway, New Jersey). The heating



rate was kept constant at $10\text{ }^{\circ}\text{C min}^{-1}$, and all runs were carried out between 20 and $600\text{ }^{\circ}\text{C}$. The measurements were made in open aluminium crucibles, nitrogen was purged in ambient mode, and calibration was performed using an indium standard.

X-ray crystallography

Crystallographic data for **NUIG1** were collected in an Oxford Diffraction Xcalibur CCD diffractometer using graphite-monochromatic Mo K α radiation ($\lambda = 0.71073\text{ \AA}$) at room temperature. The structures were solved using SHELXT,²⁸ embedded in the OSCALE software.²⁹ The non-H atoms were treated anisotropically, whereas the hydrogen atoms were placed in calculated, ideal positions and refined as riding on their respective carbon atoms. The program SQUEEZE,³⁰ a part of the PLATON package of crystallographic software, was used to remove contribution of highly disordered solvent molecules.

Unit cell data and structure refinement details are listed in Table S1 (ESI†). CCDC 2008513.†

Drug adsorption and release experiments

The Ibu adsorption capacity of **NUIG1** was investigated as described below: drug (0.23 g, 0.70 mmol) was added to a glass vial containing EtOH (10 mL) and stirred until all solid is dissolved. Solid **NUIG1** (0.05 g, 0.07 mmol) was then added and the mixture was left stirring at room temperature. For the kinetic study, small volumes of aliquots were taken at designated time intervals, centrifuged, and the drug content in the supernatant solution was determined by spectroscopic (UV-vis) and chromatographic (HPLC) techniques. For the thermodynamic study, the same procedure was repeated with varying **NUIG1**: drug ratios; the mixture was stirred for 120 hours, filtered, and the filtrate was analysed for its drug content. The drug release properties of **NUIG1** were studied in H₂O and phosphate buffer solution (PBS): Ibu@**NUIG1** (0.05 g) was added to a glass vial containing solvent (10 mL) and left stirring at $37\text{ }^{\circ}\text{C}$. Batch studies were performed at designated time intervals, and the amount of the released drug was determined by UV-vis and HPLC techniques.

Metal adsorption kinetic and thermodynamic studies

The metal adsorption capacity of **NUIG1** was investigated using the same method to the drug adsorption: the hydrated acetate salt of a metal ion (0.10 g, 0.40 mmol for Co²⁺ and Ni²⁺; 0.05 g, 0.22 mmol for Cu²⁺) was added to a glass vial containing distilled H₂O (10 mL, for Co²⁺ and Ni²⁺; 20 mL, for Cu²⁺) and stirred until all solid is dissolved. Solid **NUIG1** (0.10 g, 0.14 mmol, Co²⁺ and Ni²⁺; 0.05 g, 0.07 mmol, Cu²⁺) was then added and the mixture was left stirring at room temperature. For the kinetic study, the sample was centrifuged at designated time intervals, and the metal content in the supernatant was determined by spectroscopic (UV-vis) techniques and by using an Elan DRC-e Inductively Coupled Plasma–Mass Spectrometry, ICP-MS, [PerkinElmer, USA] in standard mode in a class 1000 clean room (ISO 6). For the thermodynamic study, the same procedure was repeated with varying **NUIG1**: metal ratios; the mixture was stirred for 30 minutes, filtered, and the filtrate was analysed for its metal content.

Solid state NMR

All solid-state NMR spectra were collected on a Bruker Avance III spectrometer with magnetic field strength of 9.4 T operating at frequencies of 400 MHz for ¹H, 100.6 MHz for ¹³C, and 105.842 MHz for ²³Na, respectively. For the acquisition of the data a 2.5 mm HX probe was used, employing magic-angle spinning (MAS) frequencies of up to 30 kHz.

The ²³Na spectra were collected at 30 kHz MAS. The one-pulse spectra were acquired using a RF pulse with a 136 kHz nutation frequency and small flip angles (0.44 μs) and a recycle delay of 16 s. Two-dimensional one-pulse double-quantum-filtered satellite transition MAS (TOP-DQF-STMAS) spectra^{31a,b} were acquired with a sampling interval of 33.33 μs and 128 increments in the indirect dimension, with 192 scans per increment for the MOF and 96 scans for the Ibuprofen loaded MOF with a recycle delay of 4 s. Pulses with a nutation frequency of 136 kHz and lengths of 1.1 μs and 1.2 μs were used for satellite-transition coherence excitation, and central-transition reconversion respectively. The central-transition-selective pulse had a nutation frequency of 12 kHz and a length of 10.4 μs . Solid NaCl was used for RF pulse calibration and for referencing the chemical shifts at 7.21 ppm. For the one-dimensional ¹³C spectra, cross polarization (CP) ¹H \rightarrow ¹³C experiments were performed at 10 kHz MAS. Acquisitions employed a ¹H excitation pulse of 71 kHz nutation frequency and 3.5 μs length followed by matched spin-lock fields of 81 kHz for ¹H and 71 kHz for ¹³C, and a contact time of 1250 ms. SPINAL64^{31c} decoupling of 71 kHz was used during acquisition. Between 8192 and 16384 transients were collected, depending on the sample, with a 4 s recycle delay. The two-dimensional heteronuclear correlation (HETCOR) experiment^{31d} was acquired using the same experimental conditions as CP, but with 80 increments in the indirect (¹H) dimension and 256 scans per increment. Adamantane was used for rf pulse calibrations and chemical shifts referencing for both ¹H and ¹³C.

Computational studies

In order to explore the ability of **NUIG1** to adsorb NO and Ibu, Grand Canonical ensemble Monte Carlo simulations were conducted. According to these simulations, the chemical potential (μ), the volume of the system (V), and the temperature (T) are kept constant, whereas the number of particles of the system, N could fluctuate. During the simulations trial moves are attempted: particle insertion or deletion, particle translation and particle rotation with each trial move having the same probability of selection.³²

The chemical potential was determined for NO by using the Peng–Robinson equation of state^{33a} with the appropriate physical constants *i.e.* critical pressure, P_c , critical temperature, T_c , and acentric factor. Temperature was set to 121 K. The Lennard Jones potential was used to describe the van der Waals interactions between framework atoms and guest molecules. Parameters set for the various atoms were taken from the Dreiding force field,^{33b} whereas the Lorentz–Berthelot mixing rules were used for cross parameters. The minimum image convention was used, placing the atom under examination at a



centre of a sphere having radius equal to the potential cut off distance, *i.e.* 12.8 Å. Interactions of the atom under examination with atoms lying outside this sphere were not taken into consideration. The potential beyond this distance was truncated and no tail corrections were used. A super cell of $2 \times 2 \times 2$ unit cells was used for the simulations to ensure that each direction, *x*, *y* and *z* is at least twice the cut off distance.

Together with Lennard Jones for the van der Waals interactions, the Coulomb potential was considered for describing the electrostatic interactions between the framework and fluid atoms. Point charges for the atoms of the framework were calculated using the Gaussian 03 software package and applying the cluster method applied: An appropriate part of the framework was cut and this part was first geometrically optimized applying Density Functional Theory at the B3LYP level.³⁴ The charges of the atoms for the optimized part were then calculated using the CHELPG method.³⁵

N_2 : The TraPPE force field^{36a} was used for describing the interactions of nitrogen molecules according to which the molecule is described as a three site model, one for each atom and one for the centre of mass. A charge of $q = -0.482$ was assigned to each atom with Lennard Jones parameters $\epsilon = 36$ K and $\sigma = 3.31$ Å. A $q = +0.964$ was placed to the centre of mass without any LJ interactions.

NO : For nitric oxide a two centre model was used, both centres having LJ parameters and Coulomb interactions.^{36b} The N–O bond length was kept fixed at 1.15 Å; for oxygen, $\epsilon = 96.986$ K and $\sigma = 2.875$ Å and for nitrogen, $\epsilon = 79.538$ K and $\sigma = 3.014$ Å. These parameters were able to reproduce the heat of vaporization during *NVT* simulations. In addition, prior to proceeding with simulations, these parameters successfully reproduced the NO bulk density at all thermodynamic states used for the GCMC simulations.

Ibuprofen: Ibuprofen molecule was modelled following the methodology reported by Bernini *et al.*^{36c} The parameters for the atoms or beads of the drug were taken from the corresponding references of the TraPPE Force. The molecule was treated as flexible.

Cytotoxicity measurements using the MTT assay

The MTT assay involves the reduction of a yellow tetrazolium salt, [3-(4,5-dimethylthazol-2-yl)-2,5-diphenyltetrazolium bromide] tetrazolium, to an insoluble formazan crystal by the metabolic activity of living cells. For this analysis, MTT assay was used to investigate the cytotoxicity of **NUIG1**, and Ibu@**NUIG1**. MCF-7 cells were seeded at a density of 1000 cells per mL in a 96-well micro assay culture plate and allowed to grow over a period of 24 h at 37 °C in a 5% CO_2 incubator. **NUIG1** in fresh culture medium was added into each well with different concentrations from 0.01–30 μM and incubated for 72 hours. Culture medium only was employed as the control group, and wells containing culture media without cells were used as blanks. 20 μL of a solution containing 43.2 mg MTT dissolved in 8.6 mL RPMI was added to each well, and the cells were incubated for another 3 hours. The excess MTT solution was then carefully removed from each well, and the formed

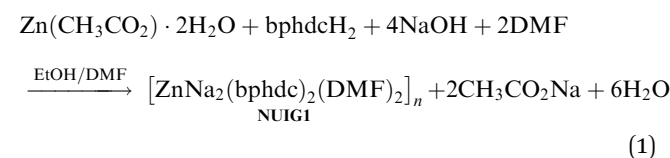
formazan was dissolved in 100 μL of DMSO. The optical density of each well was then measured at a wavelength of 550 nm using a microplate reader (Bio-Rad, xMark). The results from the three individual experiments were averaged. The following formula was used to calculate the viability:

$$\text{Viability (\%)} = (\text{mean of absorbance value of treatment group} - \text{blank}) / (\text{mean absorbance value of control} - \text{blank}) \times 100.$$

Results and discussion

Synthesis

Various reactions with differing reagent ratios, metal sources, solvents and other reaction conditions have been systematically explored towards the isolation of new non-toxic, mixed metal MOFs. The reaction of $\text{Zn}(\text{CH}_3\text{CO}_2)_2 \cdot 2\text{H}_2\text{O}$, bphdcH₂ and NaOH in a 1:2:1.5 molar ratio in EtOH/DMF (1:1.25) at 100 °C for 24 hours provided access to colourless cubic crystals of $[\text{ZnNa}_2(\text{bphdc})_2(\text{DMF})_2]_n$ (**NUIG1**). The stoichiometric equation of the reaction that led to the formation of **NUIG1** is presented in eqn (1).



It is noteworthy that the nature of the solvent affects the crystallinity of the product; for example, reactions in DMF yielded white microcrystalline powder of **NUIG1** (IR evidence). Similarly, the source of Na^+ ions does not play a crucial role in the identity of the product, but it affects its crystallinity and the yield of the reaction. In particular, when $\text{Na}(\text{CH}_3\text{CO}_2) \cdot 3\text{H}_2\text{O}$ was used instead of NaOH, **NUIG1** was produced in polycrystalline form in lower yield. The employment of significantly different molar ratios resulted in amorphous precipitates that could not be further characterized. Several bands appear in the 1655–1360 cm^{-1} region in the IR spectrum of **NUIG1**. Contributions from carboxylate $\nu_{\text{as}}(\text{CO}_2)$ and $\nu_{\text{s}}(\text{CO}_2)$ modes would be expected in this region, but overlap with the stretching vibrations of the aromatic ring and the carbonyl group renders assignment difficult.

Description of structure

Representations of the ZnNa_2 repeating unit, the rod SBU and the 3D network of **NUIG1** are shown in Fig. 1 and 2. **NUIG1** crystallizes in the monoclinic space group *C2/c* and its structure contains three crystallographically inequivalent metal ions (Zn1 , Na1 and Na2) and bphdc²⁻ ligands that adopt two different coordination modes, $\eta^2:\eta^1:\eta^1:\eta^1:\mu_5$ (**Type A**, Scheme 1), and $\eta^1:\eta^1:\eta^1:\eta^1:\mu_4$ (**Type B**, Scheme 1). The metal ions within the ZnNa_2 repeating unit are held together through two **Type B** and two **Type A** ligands with one of the latter being coordinated through its μ_2 end, whereas the other through its μ_3 end. Furthermore, three **Type A** and two **Type B** bphdc²⁻ ligands





Fig. 1 Representation of the ZnNa_2 repeating unit (top) and a part of the 1D zig-zag Zn/Na SBU (bottom) in **NUIG1**. Color code: N (dark blue), Zn (light blue), Na (green), O (red).

are linked to Na1 and Na2 and bridge the ZnNa_2 with five neighboring units. This results in the formation of a Zn/Na 1D chain, which is the secondary building unit (SBU) of **NUIG1** (Fig. 1, bottom); all the Na atoms are aligned in a collinear fashion, with the Zn atoms lying in a zig-zag configuration. Each 1D chain is cross-linked to four adjacent rod SBUs through the organic bphdc^{2-} ligands, leading to the formation of the 3D microporous network (Fig. 2).

Zn1 adopts a tetrahedral coordination geometry being linked to four carboxylic O atoms, that come from four different bphdc^{2-} ligands. Both Na atoms are hexacoordinate with a distorted octahedral coordination geometry. Na1 is coordinated to four oxygen atoms (equatorial plane), which come from four different organic ligands, as well as to two terminal DMF molecules in the axial positions. The coordination sphere of Na2 consists of six carboxylic O atoms from six different bphdc^{2-} ligands.

Employing a standard representation of the crystal structure of **NUIG1** and taking into account the Na atoms, both types of ligands and all metal ions can be considered as connection points;³⁷ this gives rise to the formation of a novel three dimensional 4,4,4,5,6-c network, with point symbol $\{4^2 \cdot 8^3 \cdot 10\}\{4^2 \cdot 8^4\}2\{4^4 \cdot 6^2\}2\{4^4 \cdot 6^3 \cdot 8^3\}2\{4^8 \cdot 6^6 \cdot 8\}$ (Fig. S1, ESI[†]); the latter has never been reported in the past, thus **NUIG1** exhibits a network with a unique architecture. Alternatively, the determination of the **NUIG1** topology can be performed without taking into account the Na atoms as they are weakly bonded to the organic ligands. In this case, the structure consists of polycatenated 2-periodic four-coordinated networks of undulated shape with an **sql** topology. The latter is common with *ca.* 100 000 such structures in different representations (complete, standard or cluster) been currently known. The solvent-accessible volume of **NUIG1**, calculated by PLATON (excluding all solvents from the pores), is 4001.9 \AA^3 and corresponds to the 46.5% of the unit-cell volume (8610.8 \AA^3). A representation of the voids using the structure visualization program MERCURY reveals a complex 3D pore network, where the large compartmentalized cavities communicate through relatively wide channels with a diameter $\geq 4 \text{ \AA}$.

NUIG1 is a new MOF with an aesthetically pleasing crystal structure and interesting structural features. It is a new addition to the growing family of mixed-metal MOFs, being the first



Fig. 2 Representation of the 3d structure of **NUIG1** along the *a* axis (top) and its pore network shown in yellow (bottom). Color code: N (dark blue), Zn (light blue), Na (green), O (red), C (grey).

Zn/Na MOF that contains the ligand bphdc^{2-} . It is a rare example of a mixed metal MOF based on a rod SBU, being the first such Zn/Na species.³⁸ It is worth to mention that the vast majority of the rod-based MOFs are homometallic and exhibit special properties, such as breathing and forbidden catenation; their SBU can be either a simple rode, *i.e.* equally spaced points on a straight line, or it can be more complicated, *e.g.* displaying a zig-zag topology or being based on edge sharing tetrahedra, *etc.*³⁸ Furthermore, **NUIG1** displays a novel three dimensional 4,4,4,5,6-c network, whose topology has not appeared in the past in a MOF.

Drug adsorption and release studies

The inherent porosity in **NUIG1**, and the existence of open channels in its structure, prompted us to assess its potential for drug delivery applications. Ibuprofen (Ibu) was chosen as a proof of concept model drug due to the availability of literature reports that would allow the evaluation of **NUIG1** as drug carrier. The MOF crystals were activated prior to the drug encapsulation in order to reduce the amount of solvent present in the pores, and thus facilitate the drug adsorption; this was



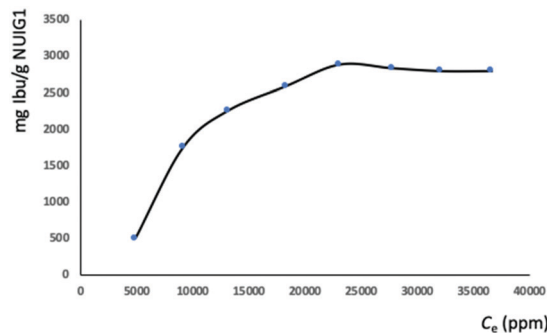


Fig. 3 Equilibrium data for the Ibu adsorption by **NUIG1** (contact time: 144 hours). The solid line is a guide to the eye.

performed by stirring the MOF in DMF for several hours and then exchanging this solvent with more volatile acetone, which is easily removed at 80 °C. The adsorption of Ibu by **NUIG1** was initially confirmed by IR spectroscopy with the appearance of peaks at 2800 and 2900 cm^{-1} , which correspond to the $\nu(\text{C-H})$ vibrations of the methyl groups from the Ibu. There is an increased intensity at the broad peak at 3100–3500 cm^{-1} due to the additional aromatic rings coming from the Ibu ($\nu(\text{Ar-H})$ at $\sim 3100 \text{ cm}^{-1}$) and the increased amount of hydrogen bonding interactions. Furthermore, there is a typical increase in the intensity of the peaks at 1500, 1430 and 1040 cm^{-1} , characteristic of the COO^- groups.

The impact of the Ibu: MOF molar ratio on the **NUIG1** drug loading performance was studied; it was found that the Ibu uptake (mg Ibu per g **NUIG1**) increases as this ratio increases from 1 : 1 to 1 : 6, where it reaches its maximum capacity (Fig. 3). The latter was determined by UV-vis and HPLC (Fig. S2, ESI[†]), which revealed that the maximum Ibu upload is 70% by weight, corresponding to 2.8 g Ibu per g **NUIG1**.

NUIG1 possesses an exceptionally high Ibu uptake capacity, which is higher than previously reported for Ibu or any other drug.⁶ This unusual behaviour is attributed to an enhanced breathing effect in **NUIG1**, *i.e.* enlargement of its pore diameter upon the encapsulation of a guest molecule.^{39,40} In order to further investigate the breathing property of **NUIG1**, XRPD studies were carried out; Fig. 4 shows the comparison of the XRPD pattern of a physical mixture of **NUIG1** and Ibu with that of Ibu@**NUIG1**, as well as that of the activated **NUIG1**. Ibu@**NUIG1** shows a similar pattern to **NUIG1** with additional characteristic peaks attributed to Ibu. The peaks that correspond to **NUIG1** have been shifted in the PXRD pattern of Ibu@**NUIG1**, indicating a change in the cell volume and/or symmetry of the MOF after the drug encapsulation.^{7a} The latter is a strong indication of the breathing property of **NUIG1**, which is further supported by the slow kinetics of Ibu encapsulation (Fig. S3, ESI[†]); in fact only the 15% wt of the drug is adsorbed in the first 24 hours. There is a gradual increase in the adsorption in the first 40 hours, after which a sharp increase is observed in the next 24 hours, reaching the amount of 2600 mg Ibu per g **NUIG1**.

The encapsulation rate then slows down slightly before it reaches a plateau after 144 hours. The initial low drug uptake rate, followed by a sharp increase, is related to the breathing effect in **NUIG1**, indicating that the pore expansion is initiated

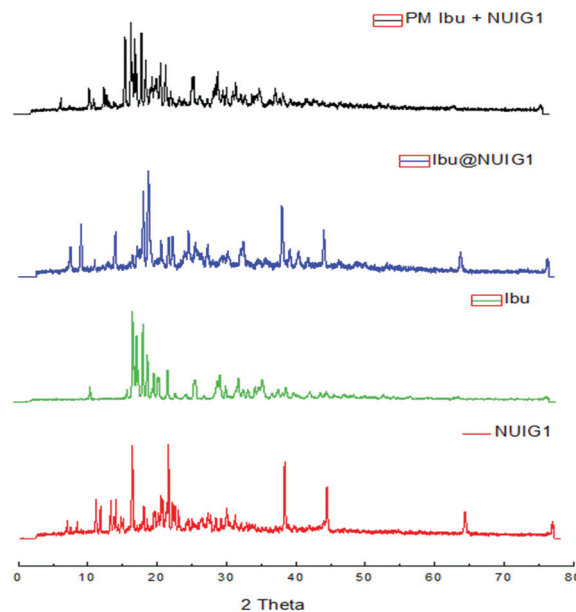


Fig. 4 XRPD patterns of **NUIG1**, Ibu, Ibu@**NUIG1**, and a physical mixture of **NUIG1** and Ibu.

after the encapsulation of a small amount of Ibu over the first 40 hours.

To confirm the unprecedentedly high drug adsorption capacity of **NUIG1** and gain insight into the interactions between **NUIG1** framework and Ibu, thermal stability (Fig. S4, ESI[†]) and solid state NMR studies were pursued. Solid-state NMR has proven to be able to provide significant information on the local environments of host systems such as MOFs, zeolites and covalent organic frameworks, and subsequently probe the host-guest interaction of those systems, ranging from adsorbed CO_2 to encapsulated drug molecules.^{41,42}

The $^1\text{H} \rightarrow ^{13}\text{C}$ CP spectra are shown in Fig. 5. There is a distinct difference in the chemical shifts due to the carboxyl functional groups of the samples, with deprotonated COO^- (173 ppm) being observed in the unloaded MOF, whilst the protonated $-\text{COOH}$ is seen in crystalline ibuprofen (183 ppm).⁴³

Both resonances are seen in the spectrum of the loaded MOF, with a very low intensity COOH resonance being slightly shifted to 181.5 ppm. Additionally, peaks at 21.8 ppm and 16.3 ppm, corresponding to the aliphatic region of Ibuprofen, are also visible in the spectrum of the loaded **NUIG** (but not for the unloaded MOF), whose correlation with the respective proton shift at 0.9 ppm is further manifested in the HETCOR experiment (Fig. S5, ESI[†]). The presence of both sets of resonances in the spectrum of Ibu@**NUIG1** indicate that ibuprofen is indeed taken up into the MOF. The comparatively small change in chemical shift of the carboxyl resonance of ibuprofen on adsorption suggests that there is no deprotonation of the ibuprofen, and no covalent bonding to the MOF network, suggesting that the adsorption mechanism is predominantly physisorption.^{42b}

To obtain a better understanding of the mechanism, ^{23}Na MAS NMR experiments were carried out. ^{23}Na is a quadrupolar nucleus (spin 3/2), whose quadrupolar charge moment interacts





Fig. 5 (top): The ibuprofen molecule, reproduced with VESTA.^{42a} (bottom) The 1-D $^1\text{H} \rightarrow ^{13}\text{C}$ cross polarization spectra for Ibu (blue), pristine **NUIG1** (red), and Ibu@**NUIG1** (green). Inset: The expanded spectral regions, highlighting the aliphatic and carboxylic species that are present in both the Ibuprofen and the Ibu loaded MOF. Spinning sidebands are labelled with asterisks.

with the electric field gradient of the local electronic environment. The one-dimensional ^{23}Na MAS NMR spectrum of the unloaded MOF (Fig. 6) exhibits a resonance broadened by the quadrupolar interaction centred on -20 ppm, with two sharp peaks likely due to sodium salt impurities. The TOP-DQF-STMAS spectra (Fig. 6) clearly show that this broad feature is actually two distinct resonances, due to the two different sodium sites present in the structures of **NUIG1**. These sodium sites lack the distinctive features of the second order quadrupolar lineshapes, which indicates a structural disorder that induces a distribution of quadrupolar coupling constants. The TOP-DQF-STMAS spectrum of Ibu@**NUIG1** (Fig. S6, ESI[†]) is largely unchanged, and the two sets of resonances yield chemical shifts and quadrupolar products that are the same within experimental error (Table S2, ESI[†]), which suggests that the sodium sites are not extensively affected by the presence of Ibuprofen. This result corroborates with the full hexacoordination of both Na sites and confirms that physisorption is the dominant mechanism of adsorption of Ibu by **NUIG1**.



Fig. 6 Top: ^{23}Na solid-state NMR spectrum of unloaded (red) and loaded (green) MOF. Bottom: DQF-TOP-STMAS ^{23}Na spectrum of the loaded MOF. Two separate resonances (1) and (2) can be identified. Horizontal extracted slices of these resonances are also represented. The arrow indicates the direction of the second order quadrupolar induced shift (QIS), which accounts for the small differences in resonance position between the one-dimensional spectra (top) and the two-dimensional spectrum (bottom).

The drug release performance **NUIG1** was also studied to H_2O and PBS at 37°C in order to simulate physiological conditions. 50 mg of the loaded Ibu@**NUIG1** was immersed in the corresponding solution and left under stirring; at certain time intervals a sample from the suspension was removed and analysed using UV-vis. Fig. S7 (ESI[†]) shows the Ibu release kinetics in both H_2O and PBS. In the first case, the maximum amount of the released Ibu is 73% with the release being completed within 50 hours; 40% of the drug is released over the first 35 hours, after which an increase in the release rate being observed. The remaining 33% of the drug is released during the next 15 hours. On the other hand, a faster release is observed in PBS conditions with the 30% drug loss being evident in the first 5 hours. A plateau is reached after 24 hours, where the 55% of the drug has been released.

Cytotoxicity studies

The MCF-7 (breast cancer) cell line was used to assess the toxicity of **NUIG1** and Ibu@**NUIG1**. The cytotoxicity was determined using the MTT assay revealing that both **NUIG1** and Ibu@**NUIG1** are non-toxic (cell viability $> 70\%$, Fig. S8, ESI[†]). This is not surprising as previously reported Zn^{II} MOFs have been proven to be non-toxic, whereas ibuprofen does not show anticancer activity against breast cancer cells,^{44a} but it has been



found to show a chemopreventive effect against the development of breast cancer, only under regular use.^{44b}

Metal uptake studies

The outstanding drug delivery performance of **NUIG1** prompted us to study its metal adsorption potential. The metal encapsulation studies were carried out by soaking activated crystals of **NUIG1** into aqueous solutions of Co^{2+} , Cu^{2+} and Ni^{2+} , respectively. Immediately, the MOF crystals underwent a color change from white (**NUIG1**) to purple (Co@NUIG1), blue (Cu@NUIG1), or green (Ni@NUIG1), Fig. S9 (ESI[†]); the colour change is directly related to the adsorbed metal; this property can be used for the naked-eye detection of a range of metal ions. The metal encapsulation was initially investigated by batch studies using UV-vis spectroscopy, (Fig. S10, ESI[†]). The maximum loading capacity obtained for Co^{2+} , Cu^{2+} and Ni^{2+} were 6% wt (equivalent to 67 mg Co^{2+} per g **NUIG1**), 8.3% wt (equivalent to 81 mg Cu^{2+} per g **NUIG1**), and 3.8% wt (equivalent to 37 mg Ni^{2+} per g **NUIG1**), respectively.

The metal adsorption by **NUIG1** exhibits fast kinetics; there is a smooth increase in the adsorption capacity over time for the three metals, which after 20 min reaches a plateau (Fig. 7, top). There is no metal adsorption after 20 min. In order to get a better insight into the metal adsorption mechanism, the experimental kinetic data were fitted to a theoretical model;⁴⁵ pseudo-first order and pseudo-second order kinetic models were used according to eqn (2) and (3), respectively.



Fig. 7 Top: Metal adsorption capacity (mg g^{-1}) versus time (h) plot. The solid lines is a guide to the eye; bottom: simulation of the experimental data to the pseudo-second order kinetic model. The solid lines represent the fitting of the data.



Fig. 8 Equilibrium data for the metal adsorption by **NUIG1** (contact time: 30 min). The solid lines are a guide to the eye.

$$\ln(q_e - q_t) = \ln q_e - k_1 t \quad (2)$$

$$\frac{t}{q_t} = \frac{1}{k_2 q_e^2} + \frac{1}{q_e} t, \quad (3)$$

where k_1 and k_2 are the rate constants for the pseudo-first, and pseudo-second kinetic models, respectively. A good fit was obtained for the pseudo-second kinetic model (Fig. 7, bottom), which is indicative of a chemisorption mechanism, *i.e.* the formation of a strong interaction or coordination bond between the encapsulated metal ions and **NUIG1**. The corresponding fitting parameters are listed in Table S3 (ESI[†]).

The metal adsorption equilibrium data are plotted in Fig. 8. The best description of the data is provided by the Langmuir model (Fig. S11, ESI[†]),⁴⁶ considering a monolayer adsorption with a finite number of homogeneous and equivalent active sites (eqn (4)):

$$\frac{c_e}{q_e} = \frac{c_e}{q_s} + \frac{1}{q_s K_L}, \quad (4)$$

where q (mg g^{-1}) is the amount of metal ion per gram of **NUIG1** at the equilibrium concentration C_e (ppm of metal ion remaining in solution), q_m is the maximum adsorption capacity of the **NUIG1**, and K_L is the Langmuir constant related to the free energy of the adsorption; the fitting parameters are listed in Table 1.

The metal adsorption capacity of **NUIG1** was further investigated by EDX, ICP, TGA and SEM studies. Fig. 9 shows the EDX spectra of **NUIG1**, Co@NUIG1 , Cu@NUIG1 , and Ni@NUIG1 samples. Zn, Na, C and O are detected in the four samples, with the spectra of the M@NUIG1 ($M = \text{Co}, \text{Cu}, \text{Ni}$) displaying one additional peak corresponding to the metal ion that has been adsorbed, hence confirming the metal uptake. This was further confirmed by ICP studies, from which it was found that the maximum % wt M content in the M@NUIG1 samples is 5.0%Co (in Co@NUIG1), 6.8%Cu (in Cu@NUIG1),

Table 1 Fitting parameters of the metal adsorption data to the Langmuir model

Metal ion	q_s (mg g^{-1})	K_L (10^4 L mol^{-1})	R^2
Co^{2+}	76.92	1.9696	0.9979
Cu^{2+}	70.42	20.2860	0.9956
Ni^{2+}	42.55	5.0000	0.9985



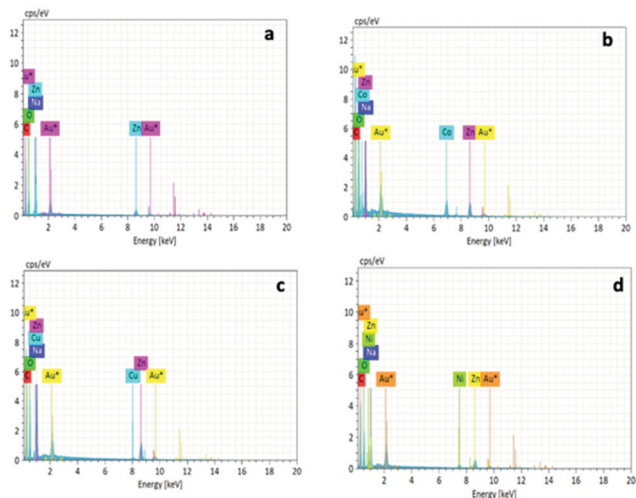


Fig. 9 EDX spectra of **NUIG1** (a), **Co@NUIG1** (b), **Cu@NUIG1** (c), and **Ni@NUIG1** (d).

and 3.8%Ni (in **Ni@NUIG1**); this is in good agreement with the results from the UV-vis studies.

The **M@NUIG1** samples are decomposed in two steps (Fig. S12, ESI[†]), with the step at 450 °C corresponding to the framework breakdown. The additional drop at 250 °C is evidence of the absorbed metal, with the mass loss at this temperature being less than 10%, in accordance with the % wt M content in the **M@NUIG1** species that has been determined by the UV-vis and ICP studies.

The SEM images of the four MOF species are depicted in Fig. 10. **NUIG1** comprises a significant number of parallel nanosheets. It is noteworthy that, upon metal encapsulation, sponge-like nanospheres are formed, with the ratio of the coexistent nanosheets: nanospheres depending on the kind of the metal ion that has been adsorbed. In particular, the number of nanospheres is high in **Co@NUIG1**, and considerably lower in the case of **Cu@NUIG1** and **Ni@NUIG1**, whose morphology is very similar to that of **NUIG1**.



Fig. 10 The SEM images of **NUIG1**, **Co@NUIG1**, **Cu@NUIG1**, and **Ni@NUIG1**.

The regeneration of **NUIG1** by **M@NUIG1** is not feasible possibly due to the strong interactions between the metal ions and the framework, as suggested by the kinetic studies. Several attempts were performed, that included washing of **M@NUIG1** with a concentrated aqueous solution of EDTA or 1 M HCl. While the guest metal was removed in all cases, XRPD showed degradation of the framework.

Magnetism studies

The magnetism properties of polycrystalline **M@NUIG1** samples were measured as a means to confirm the metal uptake capacity of **NUIG1**, and also preform an initial investigation of the potential of this technique to be used for the development of novel MOF-based sensors. Note that **NUIG1** is diamagnetic, hence any paramagnetic component would be due to the presence of paramagnetic metal ions into its pores. Direct current magnetic susceptibility measurements were carried out for **Co@NUIG1**, **Ni@NUIG1** and **Cu@NUIG1** in the 2–300 K temperature range (Fig. 11).

A closer inspection in Fig. 11 reveals that the presence of the metal ions into the **NUIG1** pores affects the magnetic properties of the MOF with the observed $\chi_M T$ value depending on the kind and the amount of the encapsulated metal ion; this is a very desirable property for the magnetic sensing of environmentally hazardous species. The values of the $\chi_M T$ products at room temperature indicate that the %M w/w uptake is in good agreement with the values derived from the previous characterization techniques: 0.6 cm³ mol⁻¹ K for Ni with a $g = 2.30$; 0.25 cm³ mol⁻¹ K for Cu, with a $g = 2$; 2.3 for Co^{II} which is slightly higher than the expected value taking into account the anisotropy of the Co^{II} ion ($g = 2.45$). The $\chi_M T$ value remains essentially stable until low temperature indicating weak interactions between the metal ions. There is an increase at the $\chi_M T$ at lower temperatures which can be attributed to a ferromagnetic coupling between the cations. The decay of the curve at very low temperatures for the Cu and Ni cations can be attributed to intramolecular interactions, whereas in the case of the Co analogue this is due to the presence of axial zero field splitting. The ferromagnetic response of the three complexes



Fig. 11 $\chi_M T$ vs. T plot for the three **M@NUIG1** species.



evidences that the cations are not isolated inside the pores, forming polynuclear aggregates that according the low temperature data should be of medium/low nuclearity.

Looking at the static magnetic properties of Co@NUIG1 and Ni@NUIG1, we decided to investigate the dynamic magnetic properties and the slow relaxation of the magnetization potential. Both aggregates present clear out-of-phase peaks (Fig. S13, ESI[†]), which are observed at relatively high temperature for the Ni case. In both cases, there are frequency dependent χ_M'' signals in absence of an external dc external field, which indicates an SMM behaviour for the complexes.

Computational studies

The BET surface area for NUIG1 was calculated by simulating the N₂ adsorption; the N₂ uptake isotherm was calculated at 77 K (Fig. 12, top) and the data were analysed as previously reported in the literature.⁴⁷ The pressure limit, where the Nitrogen monolayer is formed, was determined by the $V \cdot (P_0 - P)$ (V = Volumetric uptake) vs. P graph. After defining the pressure limit, the BET surface area for NUIG1 was calculated from the slope of the diagram in Fig. 12, bottom, and was found to be 1040 m² g⁻¹. The void volume for NUIG1 was also determined by using the helium void volume method⁴⁸ and was found to be 44%, resulting to a gravimetric pore volume of 0.45 cm³ g⁻¹. This is in perfect agreement with the solvent accessible area as calculated by PLATON.

NUIG1 was also tested for its ability to adsorb nitric oxide. Nitric oxide is known for its therapeutic properties, including among others its relaxing effect on smooth muscle dilating blood vessels, especially in the lungs, while recent reports show that NO might play an important role in the treatment



Fig. 12 N₂ volumetric uptake isotherm at 77 K for the calculation of the BET surface area of NUIG1 (top), and plot of the selected linear region that satisfies the consistency criterion (bottom).⁴⁸



Fig. 13 Gravimetric (top) and volumetric (bottom) uptake isotherms for NO at 121 K.

of COVID-19;⁴⁹ as such, the development of materials that can adsorb and deliver NO to the cells in a controlled way is very important. The isotherm (Fig. 13, top) was from GCMC simulations at 121 K and for pressures up to P_0 . To the best of our best knowledge, there are no similar studies at 121 K in literature. In contrast there are some studies at room temperature. For this reason and for the sake of comparison of our results with results from similar studies we also performed similar simulations at 298 K. The gravimetric uptake is presented in Fig. 13, bottom; the gravimetric uptake is 1.3 mmol g⁻¹ and 6 mmol g⁻¹ at 1 and 10 bar, respectively. These values are significantly higher than the corresponding values of ca. 0.5 mmol g⁻¹ and 3.6 mmol g⁻¹ for the top performing materials,^{50a} which were studied at the same temperature. This value for the top performing material^{50b} at 1 bar is ca. 1 mol kg⁻¹ but this study was at 313 K. Fig. S14 (ESI[†]) shows the isosteric heat of adsorption for NO as taken from the simulations at 298 K. At low loading this value is ca. 20 kJ mol⁻¹. This value is also very high showing the strong interaction between fluid molecules and MOF framework and is consistent with the high adsorption capacity of the material. This is also consistent with the fact that this value is considerably higher than the largest values taken from the top performing materials of the two studies referred earlier which are 10 and 20 kJ mol⁻¹, respectively.

Similar Monte Carlo simulations were conducted for ibuprofen to find the maximum uptake by the MOF. The simulations were done at various fugacity values, instead of pressure, since it is not possible to apply the equation of state for ibuprofen. The calculations were performed considering that there is an almost proportional dependence of the adsorbed amount of drug on the gravimetric pore volume of the material.⁵¹ Literature data were used in order to produce this linear dependence



of the uptake on the pore volume (Fig. S15, ESI†).⁵² According to this, the maximum calculated gravimetric loading is 230 mg drug per g **NUIG1**, which is significantly lower than the maximum drug uptake that has been experimentally determined. There are several possible explanations for this including the fact that the breathing property is not considered in the theoretical model, as well as to possible structural distortions that the MOF undergoes during activation, *e.g.* it is likely that not only the solvents that occupy the pores of the MOF are removed, but also the terminally ligated solvents, which will result in a considerable expansion of the MOF's pores, and hence its drug uptake capacity increases. Another possible reason is that a percentage of the amount of the drug is likely adsorbed by the external surface of the MOF and does not enter in to its pores.

Conclusions

The discovery of new materials with improved performance in environmental and biomedical applications, including drug delivery, and adsorption and removal of hazardous species from aqueous systems is very important. Mixed metal MOFs, although they display desirable features for these applications, have not been systematically investigated so far. **NUIG1** is a new addition to the relatively small family of mixed-metal MOFs, being a rare example of such a species whose structure is based on rod SBUs. It is non-toxic and displays a new network topology. **NUIG1** exhibits an outstanding performance in Ibu and NO adsorption capacity. The mechanism of adsorption was studied in detail by solid-state NMR, which revealed that the dominant mechanism is physisorption. Furthermore, **NUIG1** possesses a considerable metal uptake capacity for Co, Ni and Cu. Dc magnetic susceptibility studies revealed that the magnetic properties of the M@**NUIG1** species change significantly according to the kind and the amount of the encapsulated metal ion.

This work establishes the mixed-metal MOFs as excellent candidates in important biomedical and environmental applications. It also reveals that magnetism can be a powerful technique for the detection of environmentally hazardous chemicals, and this can be useful especially for species that do not affect the photoluminescence properties or the colour of a compound, hence they cannot be detected by the commonly used sensors. Further work is currently in progress towards this direction and will be reported in due course. The isolation of analogues of **NUIG1** with paramagnetic metal ions instead of Zn is also of special interest in order to investigate the impact on the adsorption capacity and the magnetism-based sensing properties.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

This work has been funded by the Science Foundation Ireland. W. P., J. P. C., and A. J. P. were supported by the Swedish Research Council (project no. 2016-03441).

References

- (a) H.-C. J. Zhou and S. Kitagawa, *Chem. Soc. Rev.*, 2014, **43**, 5415; (b) M. Eddaoudi, D. B. Moler, H. Li, B. Chen, T. M. Reineke, M. O'Keeffe and O. Yaghi, *Acc. Chem. Res.*, 2001, **34**, 319; (c) S. Yuan, L. Feng, K. Wang, J. Pang, M. Bosch, C. Lollar, Y. Sun, J. Qin, X. Yang, P. Zhang, Q. Wang, L. Zou, Y. Zhang, L. Zhang, Y. Fang, J. Li and H.-C. Zhou, *Adv. Mater.*, 2018, **37**, 1704303.
- (a) H. Furukawa, K. E. Cordova, M. O'Keeffe and O. M. Yaghi, *Science*, 2013, **341**, 1230444; (b) A. Schneemann, V. Bon, I. Schwedler, I. Senkowska, S. Kaskel and A. R. Fischer, *Chem. Soc. Rev.*, 2014, **43**, 6062.
- N. L. Rosi, J. Eckert, M. Eddaoudi, D. T. Vodak, J. Kim, M. O'Keeffe and O. Yaghi, *Science*, 2003, **300**, 1127.
- (a) M. Kurmoo, *Chem. Soc. Rev.*, 2009, **38**, 1353; (b) J. Lee, O. K. Farha, J. Roberts, K. A. Scheidt, S. T. Nguyen and J. T. Hupp, *Chem. Soc. Rev.*, 2009, **38**, 1450.
- (a) P. Kumar, A. Deep and K.-H. Kim, *TrAC, Trends Anal. Chem.*, 2015, **73**, 39; (b) P. Horcajada, T. Chalati, C. Serre, B. Gillet, C. Sebrie, T. Baati, J. F. Eubank, D. Heurtaux, P. Clayette, C. Kreuz, J.-S. Chang, Y. K. Hwang, V. Marsaud, P.-N. Bories, L. Cynober, S. Gil, G. Férey, P. Couvreur and R. Gref, *Nat. Mater.*, 2010, **9**, 172.
- P. Horcajada, C. Serre, M. Vallet-Regí, M. Sebban, F. Taulelle and G. Férey, *Angew. Chem., Int. Ed.*, 2006, **45**, 5974.
- (a) P. Horcajada, C. Serre, G. Maurin, N. A. Ramsahye, F. Balas, M. Vallet-Regí, M. Sebban, F. Taulelle and G. Férey, *J. Am. Chem. Soc.*, 2008, **130**, 6774; (b) A. C. McKinlay, R. E. Morris, P. Horcajada, G. Férey, R. Gref, P. Couvreur and C. Serre, *Angew. Chem., Int. Ed.*, 2010, **49**, 6260.
- (a) Y. Han, W. Liu, J. Huang, S. Qiu, H. Zhong, D. Liu and J. Liu, *Pharmaceutics*, 2018, **10**, 271; (b) C. He, K. Lu, D. Liu and W. Lin, *J. Am. Chem. Soc.*, 2014, **136**, 5181.
- (a) C.-Y. Sun, C. Qin, X.-L. Wang, G.-S. Yang, K.-Z. Shao, Y.-Q. Lan, Z.-M. Su, P. Huang, C.-G. Wang and E.-B. Wang, *Dalton Trans.*, 2012, **41**, 6906; (b) I. A. Lazarov and R. S. Forgan, *Coord. Chem. Rev.*, 2019, **380**, 230.
- (a) L. Sercombe, T. Veerati, F. Moheimani, S. Y. Wu, A. K. Sood and S. Hua, *Front. Pharmacol.*, 2015, **6**, 286; (b) W. Xu, P. Ling and A. Zhang, *J. Drug Delivery*, 2013, 340315.
- (a) E. Abbasi, S. F. Aval, A. Akbarzadeh, M. Milani, H. T. Nasrabadi, S. W. Joo, Y. Hanifehpour, K. Nejati-Koshki and R. Pashaei-Asl, *Nanoscale Res. Lett.*, 2014, **9**, 247; (b) W. H. De Jong and P. J. A. Borm, *Int. J. Nanomed.*, 2008, **3**, 133; (c) A. Bianco, K. Kostarelos and M. Prato, *Curr. Opin. Chem. Biol.*, 2005, **9**, 674.
- (a) P. Zrazhevskiy, M. Sena and X. Gao, *Chem. Soc. Rev.*, 2010, **39**, 4326; (b) A. Z. Wilczewska, K. Niemirowicz, K. H. Markiewicz and H. Car, *Pharmacol. Rep.*, 2012, **64**, 1020.
- (a) M. Hoop, C. F. Walde, R. Riccò, F. Mushtaq, A. Terzopoulou, X.-Z. Chen, A. J. deMello, C. J. Doonan, P. Falcaro, B. J. Nelson, J. Puigmartí-Luis and S. Pané, *Appl. Mater. Today*, 2018, **11**, 13; (b) K. S. Park, Z. Ni, A. P. Côté, J. Y. Choi, R. Huang, F. J. Uribe-Romo, H. K. Chae, M. O'Keeffe and O. M. Yaghi, *Proc. Natl. Acad. Sci. U. S. A.*, 2006, **103**, 10186.



- 14 (a) M. Witman, S. Ling, A. Gladysiak, K. C. Stylianou, B. Smit, B. Slater and M. J. Haranczyk, *J. Phys. Chem. C*, 2017, **121**, 1171; (b) X. Huang and C. S. O. Brazel, *J. Controlled Release*, 2001, **73**, 121.
- 15 (a) K. Roth Stefaniak, C. C. Epley, J. J. Novak, M. L. McAndrew, H. D. Cornell, J. Zhu, D. K. McDaniel, J. L. Davis, I. C. Allen, A. J. Morris and T. Z. Grove, *Chem. Commun.*, 2018, **54**, 7617; (b) H. Zheng, Y. Zhang, L. Liu, W. Wan, P. Guo, A. M. Nyström and X. Zou, *J. Am. Chem. Soc.*, 2016, **138**, 962.
- 16 Z. Zou, S. Li, D. He, X. He, K. Wang, L. Li, X. Yang and H. Li, *J. Mater. Chem. B*, 2017, **5**, 2126.
- 17 F. Ke, Y.-P. Yuan, L.-G. Qiu, Y.-H. Shen, A.-J. Xie, J.-F. Zhu, X.-Y. Tian and L.-D. Zhang, *J. Mater. Chem.*, 2011, **21**, 3843.
- 18 (a) R. Ricco, M. J. Styles and P. Falcaro, *Metal-Organic Frameworks (MOFs) for Environmental Applications*, ed. S. K Ghosh, Elsevier, 2019, pp. 383–426; (b) H. Molavi, A. Hakimian, A. Shojaei and M. Raeiszadeh, *Appl. Surf. Sci.*, 2018, **445**, 424.
- 19 (a) P. A. Kobielska, A. J. Howarth, O. K. Farha and S. Nayak, *Coord. Chem. Rev.*, 2018, **358**, 92; (b) K. Vellingiri, J. E. Szulejko, P. Kumar, E. E. Kwon, K.-H. Kim, A. Deep, D. W. Boukhvalov and R. J. C. Brown, *Sci. Rep.*, 2016, **6**, 27813.
- 20 (a) I. Ahmed and S. H. Jhung, *J. Hazard. Mater.*, 2016, **301**, 259; (b) P. W. Seo, B. N. Bhadra, I. Ahmed, N. A. Khan and S. H. Jhung, *Sci. Rep.*, 2016, **6**, 34462; (c) M. S. Khan, M. Khalid and M. Shahid, *Mater. Adv.*, 2020, DOI: 10.1039/d0ma00291g.
- 21 (a) J. Yang, B. Hou, J. Wang, B. Tian, J. Bi, N. Wang, X. Li and X. Huang, *Nanomaterials*, 2019, **9**, 424; (b) E. Zanin, J. Scapinello, M. de Oliveira, C. L. Rambo, F. Franscescon, L. Freitas, J. M. M. de Mello, M. A. Fiori, J. V. Oliveira and J. Magro, *Process Saf. Environ. Prot.*, 2017, **105**, 194.
- 22 (a) E. M. Dias and C. Petit, *J. Mater. Chem. A*, 2015, **3**, 22484; (b) L. Zhao, M. R. Azhar, X. Li, X. Duan, H. Sun, S. Wang and X. Fang, *J. Colloid Interface Sci.*, 2019, **542**, 421; (c) J. Barreto, M. D. G. Xavier, R. P. P. L. Ribeiro, D. Martins, I. A. A. C. Esteves, M. Branco, T. Tirolien, J. P. B. Mota and G. Bonfait, *J. Chem. Eng. Data*, 2019, **64**, 5407; (d) S. Ramanayaka, M. Vithanage, A. Sarmah, T. An, K.-H. Kim and Y. Sik Ok, *RSC Adv.*, 2019, **9**, 34359.
- 23 W. P. Lustig, S. Mukherjee, N. D. Rudd, A. V. Desai, J. Li and S. K. Ghosh, *Chem. Soc. Rev.*, 2017, **46**, 3242.
- 24 C.-H. Chen, X.-S. Wang, L. Li, Y.-B. Huang and R. Cao, *Dalton Trans.*, 2018, **47**, 3452.
- 25 S. Abednatanzi, P. Gohari Derakhshandeh, H. Depauw, F.-X. Coudert, H. Vrielinck, P. Van Der Voort and K. Leus, *Chem. Soc. Rev.*, 2019, **48**, 2535.
- 26 (a) M. Y. Masoomi, A. Morsali, A. Dhakshinamoorthy and H. Garcia, *Angew. Chem., Int. Ed.*, 2019, **58**, 15188; (b) Q.-G. Zhai, X. Bu, C. Mao, X. Zhao and P. Feng, *J. Am. Chem. Soc.*, 2016, **138**(8), 2524.
- 27 (a) H. J. Park and M. P. Suh, *Chem. – Eur. J.*, 2008, **14**, 8812; (b) Y. Ge, N.-Y. Li, W. Ma, D. Liu and X.-Y. Tang, *CrystEngComm*, 2014, **16**, 10949; (c) H. J. Park and M. P. Suh, *Chem. Commun.*, 2010, **46**, 610; (d) V. Bon, I. Senkonsva, D. Wallacher, D. M. Toebeens, I. Zizak, R. Feyerharm, U. Mueller and S. Kaskel, *Inorg. Chem.*, 2014, **53**, 1513.
- 28 G. M. Sheldrick, *Acta Crystallogr., Sect. A: Found. Adv.*, 2015, **71**, 3.
- 29 P. McArdle, K. Gilligan, D. Cunningham, R. Dark and M. Mahon, *CrystEngComm*, 2004, **6**, 303.
- 30 P. Van der Sluis and A. L. Spek, *Acta Crystallogr., Sect. A: Found. Crystallogr.*, 1990, **46**, 194.
- 31 (a) Z. Gan, *J. Am. Chem. Soc.*, 2000, **122**, 3242; (b) J. P. Carvalho, A. Jaworski, M. Brady and A. J. Pell, *Magn. Reson. Chem.*, 2020, in press; (c) B. M. Fung, A. K. Khitrin and K. Ermolaev, *J. Magn. Reson.*, 2000, **142**, 97; (d) P. Caravatti, L. Braunschweiler and R. R. Ernst, *Chem. Phys. Lett.*, 1983, **100**, 305.
- 32 E. Tylianakis and G. E. Froudakis, *J. Comput. Theor. Nanosci.*, 2009, **6**, 335.
- 33 (a) D.-Y. Peng and D. B. Robinson, *Ind. Eng. Chem. Fundam.*, 1976, **15**, 59; (b) S. L. Mayo, B. D. Olafson and W. A. Goddard, *J. Phys. Chem.*, 1990, **94**, 8897.
- 34 (a) A. D. Becke, *Phys. Rev. A*, 1988, **38**, 3098; (b) A. D. Becke, *J. Chem. Phys.*, 1993, **98**, 5648.
- 35 C. M. Breneman and K. B. Wiberg, *J. Comput. Chem.*, 1990, **11**, 361.
- 36 (a) J. J. Potoff and J. I. Siepmann, *AIChE J.*, 2001, **47**, 1676; (b) J. Yang, Y. Ren, A. Tian and H. Sun, *J. Phys. Chem. B*, 2000, **104**, 4951; (c) M. C. Bernini, D. Fairen-Jimenez, M. Pasinetti, A. J. Ramirez-Pastor and R. Q. Snurr, *J. Mater. Chem. B*, 2014, **2**, 766.
- 37 (a) V. A. Blatov, A. P. Shevchenko and D. M. Proserpio, *Cryst. Growth Des.*, 2014, **14**, 3576; (b) M. O’Keeffe, M. A. Peskov, S. J. Ramsden and O. M. Yaghi, *Acc. Chem. Res.*, 2008, **41**, 1782; (c) E. V. Alexandrov, V. A. Blatov, A. V. Kochetkov and D. M. Proserpio, *CrystEngComm*, 2011, **13**, 3947.
- 38 A. Schoedel, M. Li, D. Li, M. O’Keeffe and O. Yaghi, *Chem. Rev.*, 2016, **116**, 12466.
- 39 M. Alhamami, H. Doan and C.-H. Cheng, *Materials*, 2014, **7**, 3198.
- 40 F. Nouar, T. Devic, H. Chevreau, N. Guillou, E. Gibson, G. Clet, M. Daturi, A. Vimont, J. M. Grenèche, M. I. Breeze, R. I. Walton, P. L. Llewellyne and C. Serre, *Chem. Commun.*, 2012, **48**, 10237.
- 41 (a) S. Devautour-Vinot, G. Maurin, C. Serre, P. Horcajada, D. Paula da Cunha, V. Guillerme, E. de Souza Costa, F. Taulelle and C. Martineau, *Chem. Mater.*, 2012, **24**, 2168; (b) M. Castro, P. Losch, W. Park, M. Haouas, F. Taulelle, C. Loerbroks, G. Brabants, E. Breynaert, C. E. A. Kirschhock, R. Ryoo and W. Schmidt, *Chem. Mater.*, 2018, **30**, 2676; (c) S.-Y. Ding, M. Dong, Y.-W. Wang, Y.-T. Chen, H.-Z. Wang, C.-Y. Su and W. Wang, *J. Am. Chem. Soc.*, 2016, **138**, 3031.
- 42 (a) P. Rzepka, Z. Bacsik, A. J. Pell, N. Hedin and A. A. Jaworski, *J. Phys. Chem. C*, 2019, **123**, 21497; (b) N. Liédana, A. Galve, C. Rubio, C. Téllez and J. Coronas, *ACS Appl. Mater. Interfaces*, 2012, **4**, 5016; (c) N. Rasenack and B. W. Müller, *Drug Dev. Ind. Pharm.*, 2002, **28**, 1077; (d) K. Momma and F. Izumi, *J. Appl. Crystallogr.*, 2011, **44**, 1272.
- 43 T. Azaïs, C. Tourné-Péteilh, F. Aussenac, N. Baccile, C. Coelho, J.-M. Devoisselle and F. Babonneau, *Chem. Mater.*, 2006, **18**, 6382.
- 44 (a) S. Rojas, T. Devic and P. Horcajada, *J. Mater. Chem. B*, 2017, **14**, 2560; (b) R. E. Harris, R. T. Chlebowski,



- R. D. Jackson, D. J. Frid, J. L. Ascenseo, G. Anderson, A. Loar, R. J. Rodabough, E. White and A. McTiernan, *Cancer Res.*, 2003, **63**, 6096.
- 45 (a) S. Lagergren, *Zur Theorie der Sogenannten Absorption Gelöster Stoffe*, PA Norstedt & Söner, Stockholm, Sweden, 1898, pp. 1–39; (b) Y. S. Ho and G. McKay, *Process Biochem.*, 1999, **34**, 451; (c) K.-D. Zhang, F.-C. Tsai, N. Ma, Y. Xia, H.-L. Liu, X.-Q. Zhan, X.-Y. Yu, X.-Z. Zeng, T. Jiang, D. Shi and C.-J. Chang, *Materials*, 2017, **10**, 205.
- 46 (a) I. Langmuir, *J. Am. Chem. Soc.*, 1915, **38**, 102; (b) N. Ayawei, A. N. Ebelegi and D. Wankasi, *J. Chem.*, 2017, 3039817.
- 47 K. S. Walton and R. Q. Snurr, *J. Am. Chem. Soc.*, 2007, **129**, 8552.
- 48 M. A. Lyers and P. A. Monson, *Langmuir*, 2002, **18**, 10261.
- 49 (a) J. Kobayashi and I. Murata, *Ann. Intensive Care*, 2020, **10**, 61; (b) J.-L. Burgaud, E. Ongini and P. Del Soldato, *Ann. N. Y. Acad. Sci.*, 2002, **962**, 360.
- 50 (a) L. Wang, L. Wang, J. Zhao and T. Yan, *J. Appl. Phys.*, 2012, **111**, 112628; (b) W. Sun, L.-C. Lin, X. Peng and B. Smit, *AIChE J.*, 2014, **60**, 2314.
- 51 I. Erucar and S. Keskin, *Ind. Eng. Chem. Res.*, 2016, **55**, 1929.
- 52 (a) M. C. Bernini, D. Fairen-Jimenez, M. Pasinetti, A. J. Ramirez-Pastor and R. Q. Snurr, *J. Mater. Chem. B*, 2014, **2**, 766; (b) R. Babarao and J. J. Jiang, *J. Phys. Chem. C*, 2009, **113**, 18287; (c) B. Liu, Y. Lian, Z. Li and G. Chen, *Acta Chim. Sin.*, 2014, **72**, 942.

