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CdTe/CdS quantum dot-labeled fluorescent immunochromatography test strips for rapid detection of *Escherichia coli* O157:H7

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Immunochromatography test strips have been widely used to detect foodborne pathogens due to their high sensitivity, stability and specificity, and the selection of the label is an important factor. In this study, CdTe/CdS quantum dots with a high fluorescence were synthesized as a fluorescence label to conjugate with antibodies to prepare an immunochromatography test strip for the detection of *Escherichia coli* O157:H7 (*E. coli* O157:H7). Antibody-quantum dot conjugation was coated onto a glass fiber membrane, which played the role of the conjugate pad. *E. coli* O157:H7 polyclonal antibodies and goat anti-rabbit immunoglobulin G (Ig G) were used to create the test line and the control line, respectively. The detection limit was determined as 10^4 CFU mL⁻¹ and no cross-reactions were discovered with other *Escherichia coli* and other strains. In addition, the strips detected effectively after storing at room temperature in the dark for 35 d. The detection was easy to perform, rapidly and sensitively, and the total detection time was less than 12 min to obtain accurate results, and this method has the potential to be a powerful tool to detect bacteria quantitatively.

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Introduction

Escherichia coli O157:H7 (*E. coli* O157:H7) is a serious foodborne pathogen that can result in paroxysmal and life-threatening diseases, such as acute bloody diarrhea, abdominal spasm, and hemorrhagic enteritis.¹ This pathogen, since first discovered in the stools of infected people in Oregon and Michigan in 1982, is responsible for many outbreaks, cases and deaths around the world.² Considering the thoroughly imperfect therapeutic measures and the limit of valid vaccines, as well as no internationally accepted method, the most probable number (MPN), which is labor-intensive and often requires 3–4 days to obtain confirmed results, rapid, simple and sensitive methods are urgently needed to protect public from infection. In recent years, many methods have been established for detecting *E. coli* O157:H7, such as enzyme-linked immunosorbent assay (ELISA), gene chip, polymerase chain reaction (PCR) and biosensor techniques.^{3,4} Although these methods make it possible to gain a rapid and sensitive analysis, the cost of expensive reagents and instruments in some ways limits out-of laboratory applications for fast screening.

Quantum dots (QDs), known as semiconductor nanocrystals, possess unique luminescence and photo-physical properties, including broadband excitation, narrow and tunable emission spectra, high fluorescence and stability.⁵ Furthermore, it has been testified that the photoluminescence properties of core/shell QDs are better than those of the bare core ones because of the presence of a shell, which reduces surface defects and renders protection to the core from photo-initiated degradation.⁶ Recently, a huge research interest in QDs has been garnered in biological and biomedical fields as cell imaging and biological labeling to detect the influenza virus,⁷ cancer cells,⁸ heart attacks,⁹ organophosphate pesticides,^{10,11} mycotoxins¹² and polyphenols.¹³ Although the applications of QDs have made great progress, less attention and effort are concentrated on the determination of pathogenic microorganisms.

Immunochromatography test strips have been widely used to detect pathogenic microorganisms and toxic contaminants in food and drinks, because the technique is simple, rapid, sensitive, and has no requirements for special reagents and instruments, ideally suitable for one-site testing by untrained personnel. The results can be judged by the naked eye and readily preserved. A colloidal gold-labeled test strip is regarded publicly as the most frequently-used for the quick diagnosis of infectious diseases and fast examination of biological antiteriorism with a detection limit (LOD) of 10^5 – 10^6 CFU mL⁻¹, but sensitivity sometimes is seriously limited.^{14,15}

In our study, CdTe/CdS QDs were synthesized as the fluorescence label to conjugate with polyclonal antibodies to prepare immunochromatography test strips for the quantitative detection of *E. coli* O157:H7, taking advantage of the specificity

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and sensitivity of antigen-antibody reactions and the accuracy of optics. The detection limit was 10^4 CFU mL⁻¹. Besides, to assess the effectiveness of the test strip, we used the strips to detect milk samples with different concentrations of *E. coli* O157:H7. The detection was performed easily and rapidly within 12 min. The strips showed good specificity and stability at room temperature in the dark.

Experimental

Materials and reagents

The microorganisms used in this study were *E. coli* O157:H7 (ATCC 51659), *Candida albicans* (ATCC 90028), *Staphylococcus aureus* (ATCC 25923), *Vibrio parahaemolyticus* (ATCC 17802), *Listeria monocytogenes* (ATCC 15313), *Enterobacter sakazakii* (ATCC 51329), *Enterobacter aerogenes* (CMCC 45103), *Proteus mirabilis* (CMCC 49005), *Salmonella paratyphi* A (ATCC 50001), *Shigella boydii* (ATCC 51315), *E. coli* (ATCC 25922), *E. coli* (ATCC 35218), and *E. coli* (ATCC 8099). All the strains and colloidal gold-labeled test strips (Romer Labs Inc. Newark. DE. USA) were provided by the Center for Disease Control and Prevention of HuBei. In the preparation of the antigens, these strains were heat-inactivated for 2 h in boiling water, then centrifuged under 10 000 rpm, 5 min for three times, washed with sterile saline and stored at -20 °C with an adjusted concentration of 10^9 CFU mL⁻¹. New Zealand female rabbits were also supplied by the Center for Disease Control and Prevention of HuBei with an average weight 2.5 kg. The total amount of rabbit diets was 150 g twice a day and enough pure water was provided. Common reagents and solvents were of analytical grade, and mostly obtained from Sinopharm Chemical Reagent Co., Ltd (Shanghai, China).

Preparation of water-soluble CdTe/CdS QDs

CdTe/CdS QDs was synthesized *via* an aqueous phase synthesis method with some modification.¹⁶ Briefly, 100 mg of sodium borohydride and 63.8 mg of tellurium powder were rapidly added in reaction bulbs with 3 mL of high purity water at 40 °C with magnetic stirring until the solutions turned to purple with a white sediment, which indicated the sodium hydrogen telluride precursor was successfully prepared. Cadmium chloride (91.34 mg) was dissolved in high purity water (100.0 mL) in a round bottom flask with four necks, with mercaptopropionic acid (MPA, 104 μ L) as stabilizer and then adjusting the pH value to 10. After aerating with nitrogen for 30 min, sodium hydrogen telluride (0.6 mL) was added and heated at 95 °C for 1 h. Then thioacetamide (1 mL, 0.05 M) was added slowly and reaction was continued at 95 °C for 1 h, and so CdTe/CdS QDs were prepared.

Preparation of *E. coli* O157:H7 polyclonal antibodies

The *E. coli* O157:H7 polyclonal antibodies were prepared by our own laboratory. We injected heat-inactivated *E. coli* O157:H7, namely antigens, into the New Zealand female rabbits through multiple-site subcutaneous injection. The initial immune dose was 10^7 CFU mL⁻¹ (1 mL), changed to 10^8 CFU mL⁻¹ (1 mL) in

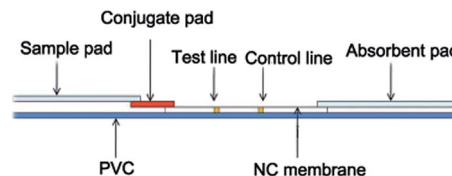


Fig. 1 A schematic diagram of the fluorescence test strips.

the following immunization. The interval of immunization was 2 weeks for 6 times. The titer of antibodies was 1 : 25 600 through ELISA. The molecular weight of the final purified antibodies was 152 kDa by sodium dodecyl sulfate polyacrylamide gel electrophoresis. All the animal work was carried out in strict compliance with the guidelines of the China Experimental Animal Welfare Ethics Committee, and the Scientific Ethics Committee for experimental animals at Huazhong Agricultural University approved all the studies.

Conjugation of QDs with antibodies

A modified amino acid-thiol covalent method was adopted to conjugate QDs with the antibodies.¹⁷ 100 μ L of purified CdTe/CdS QDs were mixed with 700 μ L of reconstituted fluid (40 μ L 0.1 M 1-ethyl-(3-dimethyl aminopropyl) carbodiimide hydrochloride, 20 μ L 0.1 M *N*-hydroxysuccinimide and 640 μ L, pH 7.4, 0.01 M phosphate buffer). Subsequently, antibodies (100 μ L, 1 mg mL⁻¹) were added at 37 °C in the dark. After shaking for 2 h, glycine (100 μ L) was added to terminate the reactions. Then uncoupled QDs were removed through 100 kDa ultra-filtrated membranes and the precipitates were stored in 0.01 M phosphate buffer (pH 7.4) at 4 °C in the dark.

Fabrication of fluorescent test strips

The test strip included a sample pad, conjugate pad, nitrocellulose membrane (NC membrane), absorbent pad, and polyvinyl chloride (PVC), schematically shown in Fig. 1. Among these components, a glass fiber membrane (0.6 cm \times 0.7 cm) was used as conjugate pad with 1 mL QD-antibody conjugations. *E. coli* O157:H7 polyclonal antibodies (1 μ L) and goat anti-rabbit Ig G (1 μ L) were dotted onto the NC membrane (0.6 cm \times 3.5 cm) as the test line (T line) and control line (C line), respectively, with 1 cm distance kept between two lines. No treatments were required for both the sample pad (0.6 cm \times 2.9 cm) and the absorbent pad (0.6 cm \times 3.2 cm). During the assembly, a 2 mm overlapped was used to ensure the performance of immunochromatography.

Results and discussion

Fluorescence of water-soluble CdTe/CdS QDs

As shown in Fig. 2A and B, CdTe/CdS QDs gave a clearly orange and yellow fluorescence under the visible and ultraviolet light (UV light), respectively. Fig. 2C presents the transmission electron microscope image of CdTe/CdS QDs, the average diameter was about 3 nm and the QDs were spherical particles with an even dispersion. Ultraviolet-visible (UV-vis) absorption



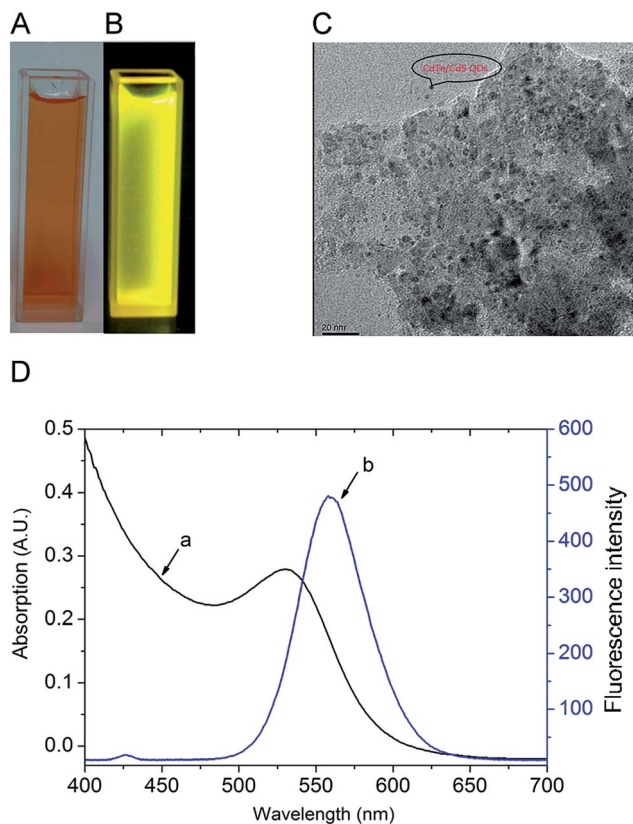


Fig. 2 Fluorescence of CdTe/CdS QDs when exposed to visible light (A) and UV light (365 nm) (B). (C) The transmission electron microscope image of CdTe/CdS QDs. (D) The absorption spectrum (a) was recorded at room temperature with a scanned range from 300 to 800 nm and the fluorescence emission spectrum (b) was obtained in the range of 400 to 700 nm.

and fluorescence emission spectra of the QDs were recorded at room temperature with a Shimadzu UV-1077 ultraviolet spectrometer (scanning wavelength was from 300 to 800 nm) and a Shimadzu RF-5301pc fluorescence spectrometer (scanning range: 400–800 nm, excitation wavelength: 370 nm), respectively. Fig. 2D shows the UV-vis absorption (curve a) and fluorescence emission spectra (curve b) of the QDs. It was concluded that the optical absorption had a feature at 530 nm and the narrow and symmetric emission peak was at 556 nm. Furthermore, from the fluorescence emission spectra, the full width at half-maximum was about 50 nm, indicating that during the growth of the QDs, the electronic defect sites had been mostly reduced.

Bio-activity of antibodies after conjugation

A dot blotting assay was carried to evaluate the bio-activity of antibodies after conjugating with the QDs. The bio-activity was defined as the fluorescence on the NC membrane under UV light and the intensity was related to the concentration of heat-inactivated *E. coli* O157:H7; phosphate buffer (pH 7.4) was used as negative control. Fig. 3 demonstrated that the bio-activity of the antibodies remained after coupling with the QDs and the

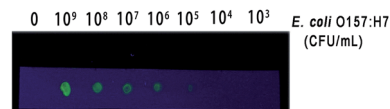


Fig. 3 Photo of dot blots for detecting the bio-activity of antibodies under UV light. Phosphate buffer was selected as the blank control.

fluorescence intensity weakened as the concentration of heat-inactivated *E. coli* O157:H7 decreased.

Properties of fluorescent test strips

The detection limit, specificity and stability of the fluorescent test strips were tested for three times to characterize the prepared fluorescent test strips. The positive result was that the control lines and test lines showed yellow-green fluorescence and a negative result was that only the control line showed fluorescence. If no fluorescence was emitted at the control line and test line or only at the test line, this indicated that the prepared test strips were useless.

Detection limit (LOD) of strips

In order to determine the detection limit (LOD) of strips, different concentrations of the heat-inactivated *E. coli* O157:H7

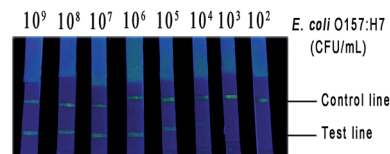


Fig. 4 Detection limit of fluorescence test strips for *E. coli* O157:H7 (1×10^2 to 1×10^9 CFU mL⁻¹).

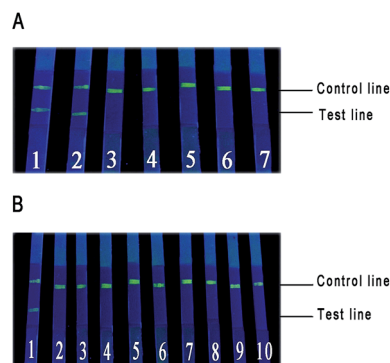


Fig. 5 Specificity of *E. coli* O157:H7 detection by fluorescence test strips. (A) Typical fluorescence responses of the strips with different kinds of *E. coli*. (1) Isolate of *E. coli* O157:H7; (2) *E. coli* O157:H7 antigen; (3) *E. coli* ATCC 25922; (4) *E. coli* ATCC 35218; (5) *E. coli* ATCC 8099; (6 and 7) Different isolates of *E. coli*. (B) Image of test strips prepared using 10 strains. (1) *E. coli* O157:H7 ATCC 51659; (2) *C. albicans* ATCC 90028; (3) *S. aureus* ATCC 25923; (4) *V. parahaemolyticus* ATCC 17802; (5) *L. monocytogenes* ATCC 15313; (6) *E. aerogenes* CMCC 45103; (7) *P. mirabilis* CMCC 49005; (8) *S. paratyphi* A ATCC 50001; (9) *E. sakazakii* ATCC 51329; (10) *S. boydii* ATCC 51315.



(10^9 CFU mL⁻¹, 10^8 CFU mL⁻¹, 10^7 CFU mL⁻¹, 10^6 CFU mL⁻¹, 10^5 CFU mL⁻¹, 10^4 CFU mL⁻¹, 10^3 CFU mL⁻¹ and 10^2 CFU mL⁻¹) were dropped onto the sample pad to perform the chromatography reactions. Sample solutions would transfer to the conjugate pad due to capillary action and the immunoreactions would complete on the test line (Fig. 1B). As shown in Fig. 4, the visual LOD of the strips was 10^4 CFU mL⁻¹; compared with conventional colloidal gold-labeled test strips, the visual LOD was 10–100 times lower (10^5 – 10^6 CFU mL⁻¹).^{14,15} In recent years, some development of gold-labeled test strips has been reported, such as Zhang *et al.*¹⁸ demonstrated a kind of hierarchical tipped flowerlike gold nanoparticles to label immunochromatography test strips to detect *E. coli* O157:H7 with

a visual LOD of 10^4 – 10^6 CFU mL⁻¹ at different sizes. Cui *et al.*¹⁹ also reported that the LOD could be highly improved *via* optimizing the size and uniformity of gold nanoparticles with a bioassay reader. Wang *et al.*²⁰ studied LOD improvement by adding enhancers and the LOD was 5×10^3 CFU mL⁻¹ by a strip reader, but whether the addition of enhancers had some influence on the stability of the strips is unknown and the detection time was 20 min, which is quite long. Besides, for other test strips, such as immunomagnetic bead-labeled and fluorescent microsphere-labeled ones, to our knowledge, the LOD of test strips based on immunomagnetic beads to detect *E. coli* O157:H7 was about 10^5 CFU mL⁻¹;²¹ moreover, immunomagnetic beads required conjugated antibodies with a high affinity while QDs could couple with a variety of molecules, such as proteins and streptavidin, as well as antibodies, easily after some chemical modifications due to its good biocompatibility. Fluorescent microspheres are a kind of solid particle that fluorescent materials, such as QDs, are covered by polymeric substances which emit fluorescence when stimulated by external energies, and the LOD of *E. coli* O157:H7 was 10^4 CFU mL⁻¹.¹⁴ In our study, the LOD of test strips based on only QDs was also up to 10^4 CFU mL⁻¹. So, we thought the sensitivity of our test strips was ideal to apply to the detection *E. coli* O157:H7 in real lives.

Specificity of strips

We studied the specificity of strips by using isolated strains of *E. coli* O157:H7, as well as other *E. coli* (*E. coli* ATCC 25922, *E. coli* ATCC 35218 and *E. coli* ATCC 8099) with 10^8 CFU mL⁻¹. The results were shown in Fig. 5A; except for *E. coli* O157:H7 and its isolated strains, no response was found for other *E. coli*, which meant that the strips were unable to determine other *E. coli*. Then, we continued to choose another nine standard strains to evaluate the cross-reactivity. Only the strip that detected *E. coli* O157:H7 exhibited fluorescence in the test line (Fig. 5B), which further proved that the strips had a high specificity in our study.

Stability of strips

The stability of test strips was determined to detect different concentrations of heat-inactivated *E. coli* O157:H7 (10^6 CFU mL⁻¹, 10^5 CFU mL⁻¹ and 10^4 CFU mL⁻¹) after storing for 0 d, 7 d, 14 d, 21 d, 28 d and 35 d. The results were shown in Fig. 6A. The strips, after storing for 35 d, were still able to realize detection. However, the fluorescence of the strips that detected

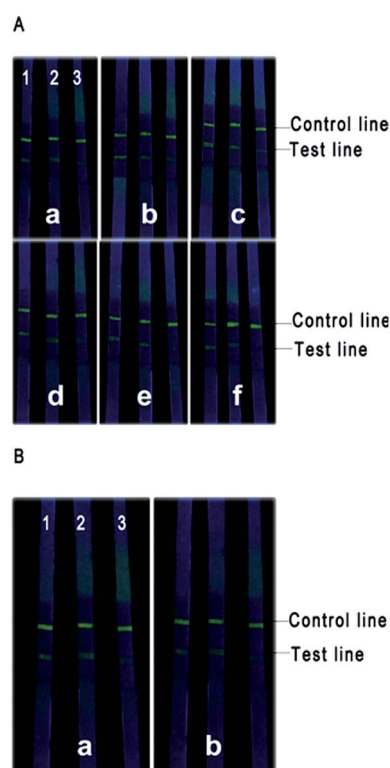


Fig. 6 The stability of strips. 1, 2 and 3 represented the different concentrations of *E. coli* O157:H7, namely 10^6 CFU mL⁻¹, 10^5 CFU mL⁻¹ and 10^4 CFU mL⁻¹, respectively (A) The results of strips for the detection of *E. coli* O157:H7 after storing for 0 d (a), 7 d (b), 14 d (c), 21 d (d), 28 d (e) and 35 d (f) (B) the diagrams of the used strips after storing for 0 d (a) and 20 d (b).

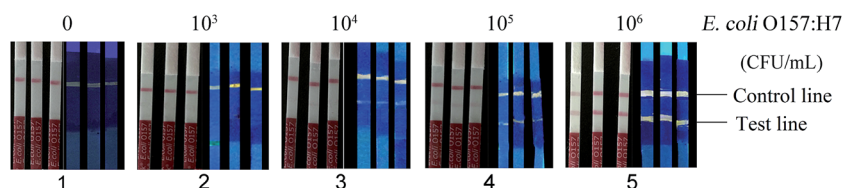


Fig. 7 Detection of *E. coli* O157:H7 in milk which was inoculated with *E. coli* O157:H7 at different concentrations (0 CFU mL⁻¹, 10^3 CFU mL⁻¹, 10^4 CFU mL⁻¹, 10^5 CFU mL⁻¹ and 10^6 CFU mL⁻¹) by CdTe/CdS QD-labeled test strips and colloidal gold-labeled test strips. From numbers 1–5, among every image, the left was the detection results of colloidal gold-labeled test strips and the right was the detection results of CdTe/CdS QD-labeled test strips.



10^4 CFU mL⁻¹ *E. coli* O157:H7 became faint after storing for 21 d and completely disappeared in the subsequent week. Furthermore, we happened to determine that the used strips had no changes after storing for 20 d at room temperature in the dark for reviewing at a certain time (Fig. 6B).

Detection of *E. coli* O157:H7 in real samples

To evaluate the potential application of our test strips, cultured *E. coli* O157:H7 were inoculated in milk with concentrations of 0 CFU mL⁻¹, 10^3 CFU mL⁻¹, 10^4 CFU mL⁻¹, 10^5 CFU mL⁻¹ and 10^6 CFU mL⁻¹. Every concentration corresponded to three samples. The samples were detected by our CdTe/CdS QD-labeled test strips and colloidal gold-labeled test strips (Romer Labs Inc. Newark. DE. USA), respectively. The results were shown in Fig. 7; when the concentration of *E. coli* O157:H7 was 10^4 CFU mL⁻¹, there were no red line in the test line of the colloidal gold-labeled test strips while weak fluorescence could be observed in the test line of QD-labeled test strips, which further determined the results of the LOD test.

Conclusions

Immunochromatographic test strips labeled by nanoparticles have been applied widely in many fields to meet the requirements of simplicity, sensitivity and rapidity, especially for those localities lacking instruments and technical detectors. In this work, we developed core/shell QDs with a high fluorescence as a novel label to conjugate with *E. coli* O157:H7 polyclonal antibodies to rapidly detect *E. coli* O157:H7. The visual LOD of the strips was 10^4 CFU mL⁻¹. Furthermore, the strips could be placed at room temperature in the dark for 35 d. The whole detection time was less than 12 min and this method would be a useful and novel protocol for performing immunoassays.

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References

- 1 S. M. Chekabab, V. J. Paquin, C. M. Dozois and J. Harel, *FEMS Microbiol. Lett.*, 2013, **341**, 1–12.
- 2 H. Pennington, *Lancet*, 2010, **376**, 1428–1435.
- 3 Y. Li, R. Afrasiabi, F. Fathi, N. Wang, C. Xiang, R. Love, Z. She and H. B. Kraatz, *Biosens. Bioelectron.*, 2014, **58**, 193–199.
- 4 Y. H. Lin, S. H. Chen, Y. C. Chuang, Y. C. Lu, T. Y. Shen, C. A. Chang and C. S. Lin, *Biosens. Bioelectron.*, 2008, **23**, 1832–1837.
- 5 Q. Ma, Y. Nakane, Y. Mori, M. Hasegawa, Y. Yoshioka, T. M. Watanabe, K. Gonda, N. Ohuchi and T. Jin, *Biomaterials*, 2012, **33**, 8486–8494.
- 6 X. Fu, K. Huang and S. Liu, *J. Microbiol. Methods*, 2009, **79**, 367–370.
- 7 O. Adegoke, T. Kato and E. Y. Park, *Biosens. Bioelectron.*, 2016, **80**, 483–490.
- 8 Q. Wu, L. Chen, L. Huang, J. Wang, J. Liu, C. Hu and H. Han, *Biosens. Bioelectron.*, 2015, **74**, 16–23.
- 9 D. Bhatnagar, V. Kumar, A. Kumar and I. Kaur, *Biosens. Bioelectron.*, 2016, **79**, 495–499.
- 10 H. Liang, D. Song and J. Gong, *Biosens. Bioelectron.*, 2014, **53**, 363–369.
- 11 X. Yan, H. Li, X. Han and X. Su, *Biosens. Bioelectron.*, 2015, **74**, 277–283.
- 12 V. V. Gofman, T. Aubert, D. V. Ginsté, R. D. Van, N. V. Beloglazova, Z. Hens, S. S. De and I. Y. Goryacheva, *Biosens. Bioelectron.*, 2016, **79**, 476–481.
- 13 U. S. Akshath, L. R. Shubha, P. Bhatt and M. S. Thakur, *Biosens. Bioelectron.*, 2014, **57**, 317–323.
- 14 Q. Y. Xie, Y. H. Wu, Q. R. Xiong, H. Y. Xu, Y. H. Xiong, K. Liu, Y. Jin and W. H. Lai, *Biosens. Bioelectron.*, 2014, **54**, 262–265.
- 15 Y. Ju, H. J. Hao, G. H. Xiong, H. R. Geng, Y. L. Zheng, J. Wang, Y. Cao, Y. H. Yang, X. H. Cai and Y. Jiang, *Vet. Immunol. Immunopathol.*, 2010, **133**, 207–211.
- 16 Y. L. Yu, L. R. Xu, J. Chen, H. Y. Gao, S. Wang, J. Fang and S. K. Xu, *Colloids Surf., B*, 2012, **95**, 247–253.
- 17 C. Shi, X. Y. Huang, C. Q. Dong, H. J. Chen and J. C. Ren, *Chin. Chem. Lett.*, 2009, **20**, 1119–1122.
- 18 L. Zhang, Y. J. Huang, J. Y. Wang, Y. Rong, W. H. Lai, J. W. Zhang and T. Chen, *Langmuir*, 2015, **31**, 5537–5544.
- 19 X. Cui, Y. J. Huang, J. Y. Wang, L. Zhang, Y. Rong, W. H. Lai and T. Chen, *RSC Adv.*, 2015, **5**, 45092–45097.
- 20 J. Y. Wang, M. H. Chen, Z. C. Sheng, D. F. Liu, S. S. Wua and W. H. Lai, *RSC Adv.*, 2015, **5**, 62300–62305.
- 21 H. Qi, Z. Zhong, H. X. Zhou, C. Y. Deng, H. Zhu, J. F. Li, X. L. Wang and F. R. Li, *Int. J. Nanomed.*, 2011, **6**, 3033–3039.

