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Plasma source mass spectrometry for radioactive waste characterisation in support of nuclear decommissioning: a review

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The efficient characterization of nuclear waste materials represents a significant challenge during nuclear site decommissioning, with a range of radionuclides requiring measurement in varied and often complex sample matrices. Of the available measurement techniques, inductively coupled plasma mass spectrometry (ICP-MS) has traditionally been applied to long-lived radionuclides, particularly in the actinide series. With recent advances in the technique, both the sensitivities achievable and number of radionuclides potentially measurable has expanded, with the reduced procedural time offering significant economic benefits to nuclear site waste characterization compared with traditional radiometric (typically alpha and beta spectrometry) techniques. This review provides a broad assessment of recent developments, improvements in capability and describes the advantages and drawbacks of ICP-MS with regards to sample introduction and instrument design. The review will be of interest to international agencies concerned with nuclear decommissioning as well as nuclear site laboratories, project managers and sites involved in environmental monitoring and nuclear forensics.

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1 Introduction

Over the last 30 years, the application of ICP-MS for radionuclide quantification has grown significantly. Initially, ICP-MS techniques focused on longer lived radionuclides where their low specific activities favored atom-counting over radiometric techniques (typically alpha and beta spectrometry). For example, U, Th, Pu, ⁹⁹Tc, and ²³⁷Np have been measured using ICP-MS since its early days. With the shift to decommissioning, other long lived but less abundant radionuclides such as ⁹³Zr have also been quantified using ICP-MS. In addition, improvements in instrument sensitivity achieved through advances in sample introduction, mass spectrometry configuration and design and vacuum pump technologies, has opened up the possibility of using ICP-MS for quantification of shorter-lived radionuclides such as ⁹⁰Sr, significantly reducing the analytical time for such analyses (Fig. 1).

Historically, radionuclide analysis within the nuclear sector has supported environmental monitoring programs, health physics, process control, effluent and waste characterization and personnel monitoring. Analytical programs tended to focus on relatively short-lived radionuclides that were likely to contribute significantly to personnel and public doses or contamination of the workplace, or which provided information on reactor performance. However, in recent years, many first and second generation nuclear facilities worldwide have either entered or are approaching shutdown and decommissioning phases. This has led to a rapidly increasing demand for radionuclide analysis to characterize the wastes arising from site decommissioning programs (*e.g.* plant contamination assessments, radioactively contaminated land *etc.*)

As well as focusing on radionuclides that contribute to radiological worker dose and waste activity inventories in the short term, analytical strategies are now also required to quantify the low-abundance long lived radionuclides that will impact on waste repository safety cases over 10³ to 10⁶ years. The change in emphasis to decommissioning has resulted in a number of analytical challenges. These include the need for rapid radionuclide characterization of wastes prior to sentencing, the provision of techniques capable of measuring low-abundance long-lived radionuclides in the presence of other significantly higher abundance radionuclides and the requirement to analyze diverse and complex matrices. In all these cases, mass spectrometric techniques, and particularly ICP-MS, offer some unique capabilities, which help to address these challenges. Given the increasing expectations facing radioanalytical science arising from decommissioning and the expanding programme of decommissioning worldwide, it is timely to review the state-of-the-art regarding ICP-MS analysis of radionuclides and to explore how the technique could be more widely applied in the future.

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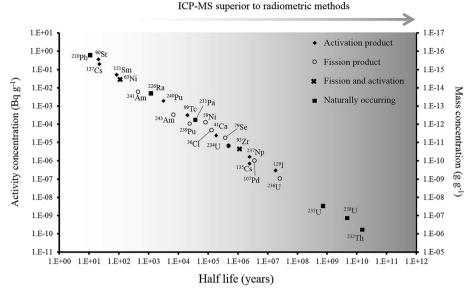


Fig. 1 Half-life versus minimum detectable activity (activity that gives count rate that is >3 times standard deviation of the background count rate) and concentration of selected radionuclides, labelled according to their method of production. Adapted from Russell et al. (2014).

History 2

There have been numerous developments in the field of mass spectrometry.2-20 In 1983 a significant advance was the introduction of the first commercial ICP-QMS, 21,22 instrument that allowed elements and isotope rations to be

measured at high sensitivity. The ICP-QMS uses an inductively coupled argon plasma as an excitation source to ionize the sample and a quadrupole mass spectrometer as an analyser to separate and selectively transmit analyte ions of a single mass-to-charge ratio (m/z) to the detector. During this process, sample ions rapidly undergo large temperature

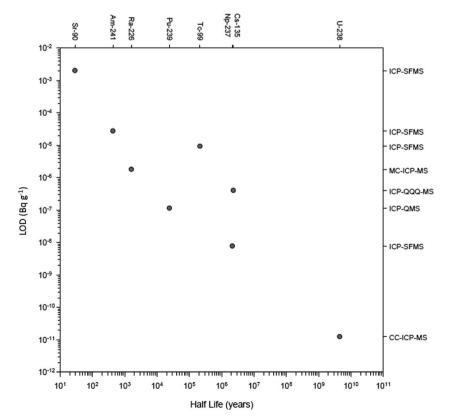


Fig. 2 Recent detection limits achieved for multiple radionuclides as a function of their half-life.

(6000 K-to-room temp.) and pressure (760 to 10⁻⁶ Torr) reductions. In essence, ions are systematically transferred from the plasma to the detector in a highly controlled electrostatic field within a dynamically increasing vacuum which was followed several years later by early measurements of radionuclides.²²⁻²⁷ The rapidity of ICP-QMS and ability to simultaneously measure multiple radionuclides were established as major advantages compared to alpha and beta counting techniques,²⁸⁻³⁰ whilst the robustness of the technique better-suited to routine analysis compared to alternative mass spectrometric techniques, specifically thermal ionization mass spectrometry (TIMS).^{30,31} Additionally, sample introduction into ICP-QMS could be achieved from a solid, liquid or gas.³²

Early studies were critical in establishing the uncertainties associated with radionuclide detection by ICP-QMS and considerations for instrumental setup. This included sample pre-treatment prior to sample introduction to improve detection limits,23 the impact of sample introduction on sensitivity and interference removal, 27,29,33-35 the importance of abundance sensitivity in removing peak tailing,26 and the use of internal standardization to account for matrix effects.36 As well as advances in quadrupole instrument design, the development of other ICP-MS setups including sector field (ICP-SFMS), collision/reaction cell instruments and multiple detector systems (MC-ICP-MS) has increased the sensitivity, interference-removal capability, and the number of nuclides measurable (Fig. 2). This has significantly expanded the toolbox for the radioanalytical chemist with regards to nuclear waste characterization and decommissioning.

3 Mass spectrometry vs. radiometric analysis

The specific activity (the rate of radioactive decay for a given mass of isotope) for a radionuclide is inversely proportional to the half-life and hence, for long lived radionuclides, more sensitive measurements can potentially be achieved by determining the concentration rather than the activity of the radionuclide (Fig. 1). More recently, there has been a growing interest in quantifying long-lived, low abundance radionuclides (e.g. ⁴¹Ca, ⁵⁹Ni, ⁶³Ni, ⁹³Zr, ¹³⁵Cs, ¹⁵¹Sm) formed through fission or neutron activation. These radionuclides were not considered significant during operational phases as their contribution to operator dose was significantly lower than for the short lived radionuclides such as 90Sr and 137Cs (Fig. 3). However, such radionuclides contribute significantly to the long-term nuclear waste repository dose estimates. Measurement of these radionuclides radiometrically is challenging as their emissions are often masked by the more abundant short-lived isotopes. For example, in fresh fission wastes the ¹³⁵Cs: ¹³⁷Cs atomic ratio is approximately 1:1 whereas the activity ratio is 1:80 000. In such instances, mass spectrometric techniques are the best analytical approach and bring significant benefits through analytical cost saving and sample throughput.

In some cases, radiometric techniques are limited to measurement of radionuclides with relatively short half-lives. Consequently, isotope ratio measurements are limited to monitoring of nuclear incidents shortly after the event (*e.g.* ¹³⁴Cs/¹³⁷Cs measurements at Fukushima), and are no longer applicable to samples affected by atmospheric weapons test fallout or

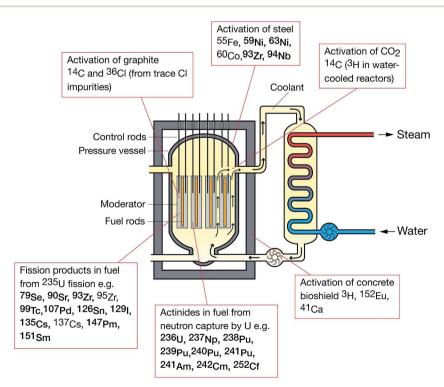


Fig. 3 Formation route of medium to long-lived radionuclides in a nuclear reactor.

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Table 1 Summary of sample introduction techniques for ICP-MS

Technique	Comments	Radionuclide applications ref.
Solution nebulization	 Range of designs, some with high tolerance to solid content Sample uptake rate as low as 50 µL min⁻¹ Relatively high oxide and hydride formation Sample to plasma transfer efficiency can be 1-2% 	45-52
Desolvating sample introduction	 Sample to plasma transfer efficiency can be 1–2% Reduced solvent loading, low oxide and hydride formation Ultrasonic nebulisers have high sample uptake rate (~1 mL min⁻¹) 	53-57
Direct injection	 100% sample to plasma efficiency Increased solvent loading into plasma increases oxide and hydride formation 	34, 41, 48 and 58
Flow injection (FI)	 Direct, real-time measurements Reduced sample preparation compared to offline separation 100% sample to plasma efficiency High maintenance Ultra-trace measurement difficult 	59–62
Sequential injection (SI)	 Evolution of flow injection Delivery of eluents, washing solutions and standards without reconfiguring the manifold Potential cross-contamination using a single manifold 	63
Lab on chip	 Downscaling of sequential injection Reagent-based assay to sub-μL levels 	64
Laser ablation (LA-ICP-MS)	 Direct measurement of solid samples Surface and depth profiling, or measurement of single particles Reduced hydride and oxide formation because of 'dry' plasma Solution nebulization preferable for bulk sample composition Lack of reference materials 	65–76
Electro thermal vaporization (ETV)	 High analyte transport efficiency (20-80%) Low oxide and hydride formation Can handle complex sample matrices Inferior detection limit compared to solution nebulization 	77-81
Glow discharge	 Complete material characterization Interferences can arise from discharge gas and sample matrix Isobaric interferences prevents direct determination of radionuclides 	82 and 83
High performance liquid chromatography (HPLC)	Rapid separation compared to offline chemical separationSeparation and detection of multiple radionuclides from	48, 74, 84 and 85
Cold plasma	 Rapid isobaric interference separation compared to offline chemical separation Sensitivity dependent on sample matrix 	86 and 87
Capillary electrophoresis	 Several potential applications <i>e.g.</i> ⁷⁹Se/⁷⁹Br, ¹²⁶Sn/¹²⁶Te, ¹²⁹I/¹²⁹Xe Low sample volumes (nL to μL) Rapid separation compared to offline chemical separation 	88–90

Chernobyl. Advances in ICP-MS have put this technique in a position where ¹³⁵Cs (2.3 million year half-life) is measurable, enabling determination of the ¹³⁵Cs/¹³⁷Cs ratio, expanding measurement options over longer timescales. In other cases, radiometric techniques are unable to separate isotopes with similar decay energies *e.g.* ²³⁹Pu and ²⁴⁰Pu, whereas ICP-MS is capable of accurate measurement of the ²³⁹Pu/²⁴⁰Pu ratio, which can vary significantly depending on the source of contamination, and therefore represents a significant advance with regards to routine monitoring and nuclear forensics.

The significant reduction in counting time compared to radiometric techniques was identified as an early advantage of ICP-MS for particular radionuclides. As the technique has advanced, there has been a focus on reducing the sample preparation time using techniques such as online chemical separation coupled directly to the instrument, interference separation using an integrated collision or reaction cell, and

improvements in sensitivity and isotope ratio accuracies using sector-field and multi-collector instruments, respectively.

Accurate and low uncertainty measurements of the decay properties of radionuclides is of importance in numerous fields including decay heat calculations in the nuclear industry and calibration of instruments.^{37,38} ICP-MS can achieve low uncertainty measurement of the number of atoms in a sample, which combined with radiometric activity measurements allow determination of the decay constant and half-life. This information is used as part of development of technical standards, which improves measurement with regards to measurement quality, reproducibility and comparison between studies^{39,40}

4 Sample introduction

Development of high sensitivity sample introduction has been a vital part of improving detection limits, and is potentially a key factor in reducing isobaric, polyatomic and tailing interferences depending on the instrumental setup. ^{41,42} A number of studies have compared the performance of different sample introduction techniques ^{41,43,44} and these are summarised in Table 1.

5 Quadrupole ICP-MS (ICP-QMS)

ICP-QMS achieved the first successful measurements of radionuclides by ICP-MS. Early instruments were unable to achieve separation of isobaric and polyatomic interferences because of their limited abundance sensitivity. Their use was originally focused on higher mass radionuclides such as ²³⁸U that do not suffer from the same number of interferences as lower mass radionuclides. However, advances in instrumental sensitivity, versatility with regards to sample introduction, and equipping certain instruments with a collision/reaction cell has improved detection limits and expanded applications over time.

MicroMass Ltd. (Wilmslow, UK) introduced an early version of a collision cell instrument (Platform ICP-MS), with the aim of thermalizing ions and dissociating disturbing molecular ions such as argides. Compared to operating without a collision cell, ion transmission, sensitivity and isotope ratio precision can be improved. However, there are conflicting views on the impact of ICP-CC-MS on abundance sensitivity. On the one hand, collisions in the cell can reduce the ion kinetic energy, potentially improving the abundance sensitivity by increasing the residence time of ions in the mass analyser which results in better mass separation. However, the collision gas increases pressure in the mass analyser, with residual gas ions leading to scattering of ions in the quadrupole, which at higher gas flow rates can have a negative impact on the abundance sensitivity.

The Perkin Elmer Elan 6100 Dynamic Reaction Cell (DRC) (based on Elan 6000 ICP-QMS) is an early example of a reaction cell instrument, capable of operating with multiple gases including NH₃, CH₄, H₂ and He. DRC instruments are equipped with a Bandpass Tuning feature, which offers mass discrimination against interfering by-products formed in the cell, whilst allowing analyte transmission. This compares to a collision cell setup that often operates with an energy filter to prevent newly formed interferences from leaving the cell. This can lead to an energy overlap between the analyte and interferences that may ultimately increase instrument backgrounds, decrease analyte signal and adversely impact detection limits. ⁹¹ Bandura *et al.* (2005) extensively studied cell-based separation of

radionuclides from overlapping isotopes using a Perkin Elmer Elan DRC, with the results outlining the cell gas used and the likelihood of the reaction occurring. 92 The DRC series has been updated with the NexION series, which is equipped with a quadrupole ion deflector that turns the ion beam 90° prior to the entrance to the collision/reaction cell. The NexION series is also equipped with an additional hyper skimmer cone to improve the removal of unionized material.

The Agilent 8800 Triple Quadrupole ICP-MS (ICP-QQQ-MS) consists of two quadrupoles positioned either side of a collision-reaction cell (termed the Octopole Reaction System, ORS). It is in effect an ICP-MS/MS or tandem mass spectrometer. Positioning a quadrupole before the entrance to the cell means the ion beam can be mass filtered prior to the cell entrance, enabling greater control over the ions entering the cell, preventing undesirable secondary polyatomic ions from forming in the cell. Secondly, the additional quadrupole improves the abundance sensitivity, with a theoretical value of 10⁻¹⁴. This is advantageous for radionuclides affected by tailing from a stable isotope of the same element e.g. ⁸⁸Sr on ⁹⁰Sr, and ¹²⁷I on ¹²⁹I. The 8800 has recently been superseded by the 8900, offering improvements including more rapid sample acquisition, measurement at higher masses (beneficial for actinide-based cell products) and introduction of samples with up to 25% total dissolved solid content.

6 Sector field ICP-MS (ICP-SFMS)

The introduction of ICP-SFMS offered lower background and higher sensitivity compared with ICP-QMS, enabling lower limits of detection for radionuclide measurement. The counting efficiency of ICP-SFMS is generally on the order of \sim 0.1%, with low background signals of <1 counts per second, compared to typical ICP-QMS values of 0.01% counting efficiency and several counts per second background. 93,94 ICP-SFMS can therefore theoretically analyze smaller bulk samples with lower analyte concentrations.95 Operating at higher mass resolution can be used to reduce or remove isobaric and polyatomic interferences, however, this is at the expense of ion transmission and therefore sensitivity.94 Reproducibility can also be affected, with typical values at low resolution of <0.02%, compared to <0.1% at medium and high resolution.96 Additionally, overlapping peaks from stable isobars affecting radionuclide detection cannot be resolved, even at high resolution. Therefore, ICP-SFMS is

Table 2 Summary of main commercially available ICP-MS instruments

Instrument type	Thermo Scientific	Agilent	Perkin Elmer	Nu	Spectro
Collision/reaction cell	iCap Q	7800 7900 8900	NexION 300 series		
Sector field	Element 2 Element XR			Attom	Spectro MS
Multi-collector	Neptune Plus			Plasma II Plasma 1700	

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generally operated at low mass resolution to maximize instrumental sensitivity, 97,98 with interference removal dependent on sample introduction and/or chemical separation.

Multi-collector instruments (MC-ICP-MS) are fitted with multiple ion counting detectors, which eliminates the need to cycle a number of small ion beams through a single detector. In addition, the effects of ion beam instability are eliminated, and flat-topped peaks and high precision isotope ratios ($\sim 0.001\%$) are achievable, which is not possible using single detector instruments. 51,54,99 For example, the Thermo Neptune (equipped with eight Faraday cups that are interchangeable with ion counting detectors) are capable of flat-topped peaks at medium mass resolution (4000), unlike single detector ICP-SFMS instruments. Multiple collectors also increase the signal in proportion to the number of available ion counting channels. For example, Taylor et al. (2003) demonstrated that with two ion-counting channels, the dwell time of ²⁴⁰Pu/²³⁹Pu is doubled, or only half the amount of plutonium is needed to give similar counting statistics to the peak jumping method.51

Finally, the Spectro-MS is a fully simultaneous, double focusing ICP-SFMS consisting of an entrance slit, ESA, energy slit, magnetic sector field and a solid state detector split into 4800 channels, which is capable of measuring the entire mass system. 100 For every analysis, the entire mass spectrum is captured, rather than focusing over a single mass unit. A summary of commercially available ICP-MS is given in Table 2.

Future developments in hardware

7.1 Miniaturization

Current applications of miniature instruments include drug screening of people, packages, luggage and vehicles, as well as forensics and environmental analysis. 101-103 The instruments developed can be self-contained with volumes of approximately 50 cm³, although this may exclude ionization sources and compressed gas cylinders. 103 There are a number of ionization techniques (Table 3) and commercially available instruments. 104 The pump and sample introduction system have a significant influence on instrument size and weight, whilst miniaturization can lead to a reduction in performance. Instruments typically have detection limits in the ppm to ppb range, 105,106 and to date there have been no applications for nuclear decommissioning. However, this approach potentially offers no sample preparation, and on-site, real-time analysis without having to return samples to an off-site laboratory.

7.2 Extreme environments

There have been a limited number of studies into instrument contamination following handling of active samples. Oak Ridge National Laboratory investigated contamination of ICP-QMS from 90Sr and 137Cs in the pump oil, and swabs of solid components.107 No activity was measured on the detector, with the majority in the interface region. Whilst any activity on the sample cone was burned off by the torch, some was detected on the skimmer cone. Two separate studies found contamination on the quadrupoles. 108,109

Table 3 Summary of atmospheric pressure ionization sources

Abbreviation	Full name
DBDI	Dielectric barrier discharge ion source
DESI	Desorption electrospray ionization
DART	Direct analysis in real time
LEMS	Laser electrospray mass spectrometry
FAPA	Flowing atmospheric pressure afterglow
LTP	Low temperature plasma
LP-DBDI	Low pressure dielectric barrier discharge ionization

There are a number of variables that must be considered, including the ion transmission, which is likely to vary depending on the age of the instrument. Contamination is also likely to vary with sample introduction system, instrument design, radionuclide and activity analyzed, and the sample throughput. 110 It is therefore good practice to monitor solid components such as cones, detectors and quadrupoles when they are replaced, prior to disposal. Some instruments have been modified in order to handle higher activity samples, with the nebulizer, torch and sliding interface situated inside a glovebox. 97,111-116

Application to radionuclide measurement

The following radionuclides, of direct interest to the nuclear decommissioning, waste characterisation and repository safety cases, have been considered as part of this review: ³⁶Cl, ⁴¹Ca, ⁵⁹Ni and ⁶³Ni, ⁷⁹Se, ⁹⁰Sr, ⁹³Zr, ⁹⁹Tc, ¹⁰⁷Pd, ¹²⁹I, ¹³⁵Cs and ¹³⁷Cs, ¹⁵¹Sm, ²¹⁰Pb, ²²⁶Ra and ²²⁸Ra, ²³¹Pa, Th and U isotopes, ²³⁷Np, Pu isotopes, ²⁴¹Am, and isotopes of Cm and Cf. The generalised origin of these radionuclides is indicated in Fig. 3.

8.1 Sample preparation

A range of possible dissolution methods exist (Table 4), with the choice of technique guided by the sample matrix and radionuclide(s) of interest. Some materials that will dissolve easily or with persistence in mineral acids (or in some cases using microwaveinduced heating in pressurised vessels) followed that approach. Up to the mid-1990s, radioanalytical practitioners encountering samples that contained components resistant to acid digestion procedures would have followed a fusion-based approach where the sample matrix was completely opened-out using fluxes such as alkali carbonates and fluorides. In 1996, Croudace and co-workers, 117 effectively demonstrated the significant benefits of using lithium borate fusion as a rapid and effective digestion method to the radioanalytical community. Prior to this, borate fusion was almost exclusively employed for elemental analysis (X-ray fluorescence analysis and ICP-OES) where it was established for its broad sample dissolution capability in an effective, safe, rapid manner and where the glasses formed could be readily dissolved in mineral acids. Up to the 1996 study referred to, radioanalytical practitioners tended to use classical (and less effective) methods such as mineral acid leaching or fusions with alkali carbonates, fluorides or hydroxides. Croudace et al. (1998) noted that the chemical nature of the radionuclides and their

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 Table 4
 Summary of sample digestion methods available adopted by radioanalytical practitioners

	Digestion or analyte extraction method	Problems/comments	Silicates	Oxides	Sulphates	Carbonates	Borates	Phosphates	Metals, carbides, silicides ^a	References
Acids & alkalis	HCl and/or HNO ₃	Microwave digestion, heating in PTFE or PFA pressure vessels may be effective. Full recovery of analytes potentially low. Oxidation of sample may be required to prevent volatilization. Difficult to achieve full disolution. Possible volatility issues				×	×		×	120
	HF/HClO ₄ acid mix	with: As, Ge, Po, S, Su, Se, Tc Only small sample masses readily treatable. HF needs to be removed prior to analysis. Insoluble fluoride precipitates in large sample volumes. Perchlorates potentially explosive. Frequently requires the use of HCl and/or HNO ₃ . Possible volatility issues with:	×	*		×	*	*	*	121
	HF/H ₂ SO ₄ acid mix	As, B, Ge, Po, Sb, Tc Small sample volumes treatable. HF needs to be removed prior to analysis. Many evaporation stages							×	120
Traditional alkali fusions	Caustic digests NaOH fusion	Widely applied to dissolve halogens, Tc Opens out mineral lattices but requires lengthy post fusion treatment. Dissolution of Pr hardware nossible	×			*			×	122
	NaCO ₃ fusion	Opens out mineral lattices but requires lengthy treatment. Dissolution of Pt hardware possible. Elevated Pb or Fe(II) will alloy with Pt hardware. Possible volatility	*			×	*		×	123
	Na_2O_2 fusion or sinter with acid digestion	Assues with: As, Hg, Po, It, II, Se Attack of Pt hardware possible. Typical fusion temperature of 250–500 °C. Small sample volumes treatable. Time intensive procedure to dissolve the alkaline fusion cake Dossible volatility issues with Au & Ru	*						×	124
	Alkali fluoride followed by pyrosulfate	Hazardous as HP produced, requires treatment with pyrosulfate to remove fluorides. Will attack Pt hardware	×			*				125
Borate fusions	Borate fusion ± acid digestion	Flexible method with no problems. Effectively digests most materials (some may require an oxidant pretreatment) and is ideal for many elemental and isotopic analysis purposes. High purity lithium borate fluxes used to ensure low analytical blanks. Sample size can vary from 0.1–10 g. Sample: flux ratios from 1:1 upward. Pt–Au crucibles used which are easily cleaned. Typical fusion temperature <1000–1200 °C. Possible volatility issues with: Hg, Pb Po & Tl	*	×	×	×			*	7117

Table 4 (Contd.)

	Digestion or analyte extraction method	Problems/comments	Silicates	Oxides	Sulphates	Carbonates	Borates	Metals, ca Silicates Oxides Sulphates Carbonates Borates Phosphates silicides $^{\alpha}$	Metals, carbides, silicides ^a	References
Fluxless fusion	Fluxless fusion \pm acid digestion digestion	Small sample volumes treatable. Conducted in inert Ar atmosphere. Typical fusion temperature >1300 °C. May require addition of SiO ₂ and MgO if silicate poor to help glass formation. Possible volatility issues with Hg, Pb & Tl	*	*	×	×	*	×	*	126
Thermal oxidation Laser ablation	Analyte(s) trapped in bubbler or condenser Direct ablation, Viridiscan, LIBS/ICP- MS	Used to liberate volatile radionuclides e.g. pyrolyser, HBO ₂ Effective in liberating small quantities of elemental material in a controlled manner. Ablation can occur in carrier gas, by trapping in a filter medium or under acid to reduce later digestion problems	× ×	× ×	× ×	× ×	× ×	× ×	× ×	127-129

Some samples (metals, carbides and disilicides may require an oxidative pretreatment prior to borate fusion. Adapted from Croudace et al. 2016. 119

location (lattice-bound or adsorbed) needed to be considered and that some actinides such as Pu can often be digested using mineral acids, whereas high fired Pu was more intransigent and required a total dissolution techniques *e.g.* borate fusion. Sinilarly, whilst ¹³⁵Cs and ¹³⁷Cs have been recovered by acid leaching for a range of sample matrices, it was proven that complete recovery in clay-rich sediments was only achievable using lithium borate fusion. Additionally, certain radionuclides (⁹⁹Tc and ¹²⁹I) suffer from losses due to volatility, which must be considered during sample preparation. A summary of sample digestion methods available for radionuclides has recently been published. 119

8.2 Chlorine-36

Chlorine-36 (half-life 3.02×10^5 years) is formed by neutron activation of stable 35Cl, which is present as an impurity in concrete and other reactor components. 130,131 Concrete and graphite wastes generated from decommissioning are the main sample matrices of interest, with mg kg⁻¹ concentrations of ³⁵Cl in concrete, combined with the high neutron capture crosssection (43 barns) and high volume of waste concrete that must be characterised. 130 Chlorine-36 is sometimes measured alongside 129I, and the volatility of both nuclides must be considered during sample preparation. Past separation techniques include leaching from concrete, followed by addition of oxidants to convert Cl and I to halides, which are then trapped in sodium hydroxide. 130 Alternatively, precipitation of Cl as AgCl followed by ion exchange chromatography has been applied to concrete, aluminium and graphite samples, 132 whilst an extraction chromatographic material (CL resin) was developed by Triskem International that is applicable to separation of ³⁶Cl and ¹²⁹I from decommissioning samples.133

Chlorine-36 is a beta emitting radionuclide ($E_{\text{max}} = 0.71 \text{ MeV}$) that can be measured by liquid scintillation counting (LSC), with Ashton et al. (1999) achieving a minimum detectable activity of 9.7 mBq g $^{-1}$ in concrete samples when measured in combination with $^{129}\mathrm{I.^{130}}$ In a separate study, Hou *et al.* (2007) achieved a LOD of 14 mBq in various decommissioning samples, with the Cl chemical yield assessed by ICP-SFMS (MicroMass Plasma Trace 2).132 The LOD for ICP-MS measurement for stable Cl was 0.01 mg kg⁻¹, equivalent to 12.2 Bq kg⁻¹ 36 Cl. The specific activity of 36 Cl (1.07 \times 10 9 Bq g $^{-1}$) makes it well suited to mass spectrometric measurement, however, there is no known ICP-MS procedure for measurement of ³⁶Cl. Aside from isobaric ³⁶S (36.0% abundance), peak tailing from ³⁵Cl, and isobaric overlap ³⁶Ar in the plasma gas must be overcome, which will be dependent on the abundance sensitivity of the instrument and the use of a collision or reaction cell, respectively. AMS is capable of separating 36Cl from the 36S isobar, and has achieved a detection limit of 0.1 Bq kg⁻¹ in food for sample sizes of 3-4 g,131 with good reproducibility (<2%) for samples with a ³⁶Cl/Cl ratio >10⁻¹². ¹³⁴

8.3 Calcium-41

Calcium-41 ($t_{1/2} = 1.03 \times 10^5$ years) is formed through thermal neutron capture of stable ⁴⁰Ca (natural abundance 96.94%), and

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also naturally through cosmic ray spallation. The latter has led to interest in measurement in the fields of cosmochemistry and geomorphology, whilst the high bioavailability is of interest in biomedical tracing in the progress of bone disease. 135-141 Calcium-41 is present in reactor shield concrete, and is therefore a key radionuclide with regards to high-volume low and intermediate level waste originating from decommissioning, as well as in longterm monitoring of nuclear waste repositories. 140,142-144

Decay of ⁴¹Ca to ⁴¹K ground state by electron capture emits low energy X-rays and Auger electrons (0.3-0.6 keV), and is therefore measurable by X-ray spectrometry or LSC. 139,142 X-ray spectrometry is straightforward but the low counting efficiency and low abundance of X-rays (11.4% for 3.31 keV) results in low sensitivity.142 This also impacts measurement by LSC, which requires separation of other radionuclides prior to measurement. 142,143 A detection limit of 0.02 Bq g⁻¹ was achieved for a 5 g concrete sample following a 60 minute count time. 142 Calcium-41 has been successfully separated from the bulk matrix and radiometric interferences by numerous techniques including ion exchange, liquid-liquid extraction, liquid membrane separation, and precipitations including calcium fluoride, carbonate and oxalate. 139,143,145

The long half-life of 41Ca and low energy X-ray and Auger electrons makes it well-suited to mass spectrometric determination, with an additional interest in measurement of the ⁴¹Ca: ⁴⁰Ca ratio in nuclear samples. There is no known ICP-MS application for detection of 41Ca, due to significant tailing effects from ⁴⁰Ca, as well as ⁴⁰Ar (99.60% natural abundance) in the plasma gas, with additional interferences from isobaric 41K (natural abundance 6.73%) and instrument-generated ⁴⁰CaH. ¹³⁶ Even in highly contaminated samples, a 41Ca isotopic abundance of 10⁻⁹ relative to ⁴⁰Ca must be determined ¹¹⁸ (compared to natural terrestrial ratios of $\sim 10^{-14}$ to 10^{-15}). 137,139 A half-life value of 9.94 \pm 0.15 \times 10⁴ years was recently determined using multiple techniques including TIMS.144 A 42Ca-48Ca double spike as well as two NIST reference materials of known isotopic composition were used to correct for isotopic fractionation. A mathematical correction was applied to account for tailing from ⁴⁰Ca, and isobaric ⁴⁰K using ³⁹K (natural abundance 93.26%) by monitoring at mass 41 when processing natural Ca samples.

RIMS is a highly sensitive technique for isotopic ⁴¹Ca/⁴⁰Ca measurements in the 10^{-10} to 10^{-11} range, ^{136,139} whilst AMS is capable of isotope ratio measurements as low as 10⁻¹⁵.139,145 Given the difficulty in routinely measuring ⁴¹Ca, Nottoli et al. (2013) estimated the activity from measurement of ⁶⁰Co in ion exchange resins used for primary fluid purification in pressurized water reactors (PWR's).145 AMS has been compared to LSC for detection of 41Ca in concrete and other solid samples including sediment and soil. 139,143 Given the fundamental difference in the two techniques, there was good agreement between results in both studies, however Hampe et al. noted significant deviations at lower activities. 139

Nickel-59 and nickel-63

Neutron irradiation of Ni, Ni alloys and stainless steels result in the formation of two long-lived Ni radioisotopes, ⁵⁹Ni

 $(t_{1/2}=7.6 imes10^4\ {
m years})$ and $^{63}{
m Ni}\ (t_{1/2}=99\ {
m years}).$ Irradiation of ⁶³Cu can also result in the formation of ⁶³Ni via ⁶³Cu(n,p)⁶³Ni reaction. Typical 59 Ni/ 63 Ni activity ratios are \sim 1 : 100 and most published methodologies have focused on the quantification of ⁶³Ni as the dominant radionuclide in recently irradiated materials. However, the long-lived ⁵⁹Ni will have a more long-term impact on waste radionuclide inventories and its characterization is becoming of greater interest. Measurement of ⁶³Ni is relatively straightforward through measurement of the 66.95 keV beta emissions by liquid scintillation analysis, with detection limits of typically 14 mBq.146 Thin window beta counting has also been used, achieving limits of detection of 1 mBq for a 3000 min count time.147 Measurement of 59Ni is more complicated. Theoretically, it should be possible to detect the associated X-ray emissions (5.88-6.49 keV) using liquid scintillation analysis, but in practice these emissions are indistinguishable from the more abundant beta emissions arising from the associated ⁶³Ni. X-ray spectrometry using LEGe of Si(Li) detectors has therefore typically been employed for ⁵⁹Ni measurement although such approaches are relatively insensitive with detection limits of 1-2 Bq being reported.148 In all cases, chemical purification of the Ni fraction is required prior to measurement of the Ni radioisotopes. Purification of Ni is most commonly achieved using dimethylglyoxime (DMG) at pH 8-9, either via precipitation or solvent extraction of the Ni-DMG complex, or via adsorption of Ni onto an extraction chromatographic material incorporating DMG loaded onto an inert support. Co-adsorption of other transition metals (notably Co, Fe and Cu) can be prevented by including ammonium acetate in the load solution. Ni/Co decontamination factors of 10³ have been reported using such separations.146 Further purification of Ni from Co can be achieved using anion exchange chromatography, with Co being retained from 9 M HCl solutions, with Ni/Co decontamination factors approaching 104.149 Purification of Ni prior to AMS measurement through the formation of volatile Ni(CO)4 has also been described and this could have significance in the development of robust Ni/Co separation procedures prior to ICP-MS measurement. 150

Given the long half-life of ⁵⁹Ni, the radionuclide should be measurable with reasonable sensitivity using mass spectrometric techniques. In practice, 59Ni is always associated with stable Ni with a maximum ⁵⁹Ni/Ni ratio of 10⁻⁷. In addition, mass spectrometric measurement of 59Ni is impacted by an isobaric interference from the only stable isotope of Co, ⁵⁹Co. Measurement of 59Ni and 63Ni by AMS following chemical separation of the Co interference has been reported. 150,151 AMS was considered in preference to ICP-MS for the measurement of ⁵⁹Ni to detect nuclear waste container leaks in the Kara Sea¹⁵² with proposed detection limits of 2-5 fg (6-15 μBq). Although less sensitive, ICP-MS should still be of use for general waste characterization, yet application of ICP-MS to 59Ni measurement has not been reported to date. This most likely reflects the challenges in measuring the low abundance ⁵⁹Ni in the presence of a significantly higher signal of 58Ni and the associated abundance sensitivity interference along with the isobaric interference from ⁵⁹Co. The extremely low abundance sensitivities (theoretically 10⁻¹⁰) associated with ICP-QQQ-MS

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may ultimately permit measurement of ⁵⁹Ni by ICP-MS. Isobaric interference from ⁵⁹Co cannot be resolved using high resolution mass spectrometry as a resolution of >50 000 would be required to achieve this. Effective separation of Ni and Co must therefore be achieved either through chemical separation prior to ICP-MS measurement or via reaction gas technologies to remove the ⁵⁹Co interference. To date, effective separation of Ni and Co using dynamic reaction cell-based techniques has not been extensively investigated. Bandura et al. (2006) suggested N2O as a reaction gas for Co/Ni separation. Co⁺ reacts at a faster rate with N₂O compared with Ni⁺. 92 However, formation of NiO⁺ was still observed and a Co/Ni separation factor of only 6 was determined. Efficient off-line chemical separation of Ni and Co therefore appears critical to the application of ICP-MS for ⁵⁹Ni determination.

8.5 Selenium-79

Selenium-79 is a long lived ($t_{1/2} = 6.0 \times 10^5$ years) fission product with a low energy beta emission ($E_{\text{max}} = 0.151 \text{ MeV}$). The radionuclide is of interest for post-closure repository performance assessments given its potentially mobile nature. Measurement of 79Se is potentially well suited to ICP-MS, however this is problematic given its low abundance in nuclear wastes, its relatively high ionisation potential (9.75 eV) and isobaric interference arising from ⁷⁹Br (50.6% abundance), as well as polyatomic interferences arising from 39K40Ar and ³⁸Ar⁴⁰ArH. Aguerre and Frechou (2006) developed a scheme to effectively separate Se from Br and associated fission products (although 125Sb was co-extracted with the Se) with final Se measurement by ICP-QMS (Perkin Elmer Elan 6000).153 Plasma power was increased to 1300 W to maximise sensitivity, with a detection limit of 0.15 μ g L⁻¹, equivalent to 50 Bq L⁻¹ ⁷⁹Se. There is also the potential to use cold plasma conditions (reduced RF power) in combination with chemical separation to separate ⁷⁹Se from ⁷⁹Br, although there is no known application of this approach. Compte et al. (1995) developed a method for determination of ⁷⁹Se in high activity fission product solutions $(10^{10} \text{ Bq L}^{-1})$ using ETV-ICP-MS, following single stage chemical separation, with ⁷⁹Se measured at a concentration of 0.43 mg L^{-1} (120.01 Bq L^{-1}). 154 ETV-ICP-MS has also been used in combination with LSC for determining the half-life of ⁷⁹Se, following separation from a reprocessing solution using liquidliquid extraction and ion exchange chromatography. 155

8.6 Strontium-90

Strontium-90 is a beta-emitting radionuclide (decay energy 0.546 MeV) with a half-life of 28.8 years that decays to 90Y (halflife 2.67 days) and then on to stable 90Zr. Strontium-90 is of critical importance in nuclear waste management, environmental monitoring and radiation protection. Additionally, 90Sr is a mobile element that can accumulate in soils and plants via precipitation and ion exchange mechanisms, 62,77,86 as well as in bones and teeth if inhaled or ingested, because of its similar chemical properties to calcium. This increases the risk of leukemia and bone cancer.86,87,156 An extensive review of sample preparation and measurement techniques for 90Sr is presented

elsewhere,62 with a recent review paper focusing on mass spectrometric measurement of 90Sr, along with 135Cs and 137Cs.157

The primary limitation affecting ICP-MS measurement of ⁹⁰Sr is an isobaric interference from stable ⁹⁰Zr (natural abundance 51.45%). The similarity in mass requires a resolution of \sim 30 000 for effective separation, which is beyond the capabilities of ICP-SFMS.86 Even in highly 90Sr-contaminated soils surrounding the Chernobyl Nuclear Power Plant, a 90Zr decontamination factor of $\sim 10^6$ is still required. An additional interference is peak tailing from stable 88Sr (natural abundance 82.6%), which is present at high concentrations in environmental samples (20-300 mg kg⁻¹ in soils, 7-9 mg L⁻¹ in seawater).87 Finally, multiple polyatomic interferences can arise from reactions of elements in the plasma.⁵⁴

Beta-counting techniques are applicable to highly sensitive detection of 90 Sr, either through direct measurement of 90 Sr, or *via* 90Y.62 The overlapping spectra of these two radionuclides can be resolved by calculating an ingrowth of 90Y, or more commonly waiting for 2-3 weeks for the establishment of secular equilibrium, followed by long count times, depending on the detection limits required. The short measurement time for ICP-MS means the concentration of 90Y daughter product is negligible (\sim 0.02% of the total intensity at m/z = 90 in nuclear fuel samples) and will not interfere with 90 Sr determination. 158

Chemical separation is commonly based on extraction chromatography using commercially available Sr-resin (Triskem International). For more complex sample matrices, this is commonly preceded by a pre-concentration stage such as calcium oxalate precipitation.77,86,158-160 Online chemical separation is advantageous when a rapid response is required, for example in the prompt assessment of contamination following the Fukushima accident in 2011.160,161 A lab-on-valve based setup successfully removed 99.8% Zr using Sr-resin prior to measurement with a Perkin Elmer Elan II DRC-ICP-MS, whilst an automated solid phase elution (SPE) system coupled to the same instrument reduced the chemical separation and analysis time by \sim 50% compared to offline separation. Grinberg et al. (2007) investigated ETV-based separation of Sr from Zr and Y using a commercial graphite tube in combination with DRC-ICP-MS.⁷⁷ In Zr-free samples, ETV using a graphite tube achieved a Sr detection limit of 1.8 ng kg⁻¹ (9.2 \times 103 Bq kg⁻¹), compared to 0.1 ng kg⁻¹ (513.2 Bq kg⁻¹) using solution nebulization. However, in the presence of 100 mg kg⁻¹ Zr, the detection limit for solution nebulization increased to 89 ng kg⁻¹ $(4.5 \times 10^5 \text{ Bq kg}^{-1}) \text{ compared to 2.9 ng kg}^{-1} (1.4 \times 10^4 \text{ Bq kg}^{-1})$ using ETV.

A Perkin Elmer Elan II DRC-ICP-MS is the most commonly applied instrument to 90Sr measurement (Table 5). With O2 as the reaction gas, 90Zr is oxidized to 90Zr16O, while the same oxidation reaction with 90Sr is not energetically favorable and would require a stronger oxidant such as N2O, or a higher O2 flow rate. 87,160 DRC-ICP-MS has been applied to detection of 90 Sr in soil samples in the vicinity of the Chernobyl nuclear power plant. 158 A Zr decontamination factor of >107 was achieved by a combination of Sr-resin and DRC-ICP-MS. The abundance sensitivity was calculated as $\sim 3 \times 10^{-9}$, with a detection limit

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of 4 pg L⁻¹ in Zr-free solutions, increasing to 200 pg L⁻¹ (1026.5 Bg L^{-1}) in digested soils. Strontium-90 has also been measured in soil contaminated by the accident at the Fukushima nuclear power plant, using on-line Sr-resin separation combined with DRC-ICP-MS, with a detection limit of 0.77 pg L^{-1} (4.0 Bq L⁻¹). Taylor et al. (2007) measured ⁹⁰Sr by DRC-ICP-MS in samples collected from Perch Lake, Ontario, with detection limits of 0.1 ng kg $^{-1}$ (513.2 Bq kg $^{-1}$) 0.04 ng kg $^{-1}$ $(205.3 \text{ Bq kg}^{-1})$ and 0.003 ng L^{-1} (15.4 Bq L⁻¹) for sediments, plants and water, respectively.159 There was a good agreement in the values measured for a standard reference material by Cerenkov counting and DRC-ICP-MS, with detection limits for the two techniques of 13.7 pg kg $^{-1}$ (70.3 Bq kg $^{-1}$) and 97.7 pg kg $^{-1}$ (501.4 Bq kg⁻¹), respectively. A recent study by Amr et al. (2016) measured 90Sr (along with 137Cs and Pu isotopes) by triple quadrupole ICP-MS (Agilent 8800) in Qatar soils and sediments.162 Following acid leaching and extraction chromatography using Sr-resin (Triskem International), 90Sr was separated from 90Zr using oxygen in the collision-reaction cell, with ⁹⁰Sr measured at an average concentration of 0.61 fg g⁻¹ $(3.1 \text{ Bq g}^{-1}).$

A less common approach to 90Sr measurement is ICP-SFMS operating with cold plasma conditions (forward power = 650-750 W) and medium mass resolution (R = 4000-4500). 86,87 Cold plasma conditions are applicable to separation of Sr from Zr based on the difference in ionization potential (5.7 eV and 6.8 eV, respectively), although sensitivity becomes more dependent on the sample matrix under these conditions. Medium mass resolution reduces instrument sensitivity compared to low resolution, but also reduces tailing from stable ⁸⁸Sr. ¹³² When a combination of medium mass resolution and cold plasma are applied, peak tailing of 88Sr is considered the critical factor affecting accurate detection of 90Sr. Abundance sensitivity values of 6×10^{-7} and 0.8×10^{-7} have been recorded at medium resolution, which is an improvement of 2 orders of magnitude compared to low mass resolution, 76 but is lower than that of DRC-ICP-MS. A detection limit of 0.4 pg L^{-1} (2.1 Bq L^{-1}) was achieved for urine samples by ICP-SFMS, compared to 2 ng L^{-1} (10.5 Bq L^{-1}) for ICP-QMS operating with a collision cell (Platform, Micromass Ltd).86

8.7 Zirconium-93

Zirconium-93 is a fission product (6.35% thermal neutron fission yield) and is also produced via the neutron irradiation of 92 Zr (natural abundance 17.1% $\sigma_{
m therm}$ – 0.2 barns). Although theoretically predicted previously, 93Zr was first positively identified in irradiated uranium by Steinberg and Glendenin in 1950. The long half-life ($t_{1/2} = 1.53 \times 10^6$ years) and corresponding low specific activity (9.3 \times 10⁷ Bq g⁻¹) means that the nuclide is ideally suited to mass spectrometric detection. In addition, the low beta energy ($E_{\text{max}} = 56 \text{ keV}$) and lack of gamma emission and, in recently irradiated materials, potential interference from 95Zr makes radiometric detection less attractive. Given the low ionisation efficiency of Zr, ICP-MS-based techniques are preferable to TIMS-based approaches. Measurement of atomic percentages of a range of fission products, including ⁹³Zr, in spent nuclear fuel dissolver solutions by direct measurement has been reported using a Perkin Elmer Elan 250 ICP-MS,164 although isobaric interferences were identified as an issue. In general, isobaric interferences arising from ⁹³Nb (100% abundance), ^{93m}Nb and ⁹³Mo (both activation products) and polyatomic 92MoH+ must be mitigated by chemically separating the interfering species from 93Zr prior to measurement. Chemical separations of 93Zr/93Nb have been reported based on solvent extraction into benzoylphenylhydroxylaminetrichloromethane-HCl165 and ion-exchange chromatography in HCl/HF media. 166 Isobaric interference suppression on-line using ICP-DRC-MS has not been employed. Bandura et al. (2006) noted that no reactions had been identified that would permit separation of 93Nb/93Mo interferences in a reaction cell.92

Chartier et al. (2008) reported an isotope-dilution MC-ICP-MS (Isoprobe) approach using a 96Zr spike. 165 A mass bias of 3% amu⁻¹ was observed for Zr and corrected for through measurement of 94 Zr/ 90 Zr ratios (reference value 0.3378 \pm 0.0002) in a natural Zr standard.165 The relative uncertainty of measured [93 Zr] was estimated as 0.6% (k=2). The procedure was subsequently applied to the determination of the half-life of 93Zr.167

8.8 Technetium-99

Technetium-99 is a pure-beta emitting radionuclide with a halflife of 2.13×10^5 years, and is a high yield fission product (6.1%) and 5.9% fission yield from thermal neutron fission of 235U and ²³⁹Pu, respectively) of significant interest with regards to nuclear waste characterization and decommissioning. The relatively high mobility is a concern with regards to waste storage and disposal, whilst the high solubility in seawater (as TcO₄-) makes it an important seawater tracer. ¹⁶⁸ A comprehensive review of analytical methods for 99Tc is published elsewhere.169

Low level gas flow GM counter or LSC are applicable to 99Tc measurement. The long half-life and low decay energy ($E_{\text{max}} =$ 292 keV) means long count times for LSC if detection limits in the pg g⁻¹ range are required, 169 whilst interferences from ¹⁰³Ru, ¹⁰⁶Ru and ⁹⁰Sr must be removed prior to measurement. ICP-MS measurement of 99Tc is impacted by an isobaric interference from stable 99Ru (natural abundance 12.76%), which can be corrected for by chemical separation, or monitoring ¹⁰¹Ru (natural abundance 17.06%). Several isotopes have been used for drift correction, including 103Rh and 115In. Issues with ¹⁰³Rh are memory effects, and instability in dilute HNO₃ for long analytical runs. 168,170 By comparison, 115 In does not suffer from memory effects and has minimal disturbance from ⁹⁹Tc¹⁶O. ¹⁶⁹ There are no stable isotopes of Tc, although longlived 97 Tc and 98 Tc (both with 4.2×10^6 year half-lives) can be used as both a yield tracer and internal standard.169

Solvent extraction, ion exchange chromatography and extraction chromatography are all effective approaches to separation of ⁹⁹Tc, with extraction chromatography using TEVA resin the most commonly used.169 The volatility of Tc must be considered during sample preparation, with losses noted when

Table 5 Summary of recent procedures for ⁹⁰Sr measured by ICP-MS

Reference	Instrument	Model	Matrix	Chemical separation	LOD, pg L^{-1} (Bq L^{-1})
86	SFMS	Element	Urine	CaPO ₄ pptate, Sr-resin, medium res, cold plasma	0.4 (2.1)
64	SFMS	Element	Groundwater	Medium res, cold plasma	11 (56.5)
161	DRC-MS	Perkin Elmer Elan DRC II	Soils	On-line solid phase elution, reaction cell	600 (3079.4)
160	DRC-MS	Perkin Elmer Elan DRC II	Soils	Sr resin, reaction cell	0.8 (3.9)
77	DRC-MS		Environmental samples	Sr resin, ETV sample introduction, reaction cell	1800 (9238.3)
158	DRC-MS	Perkin Elmer Elan DRC II	Soils	Sr resin, reaction cell	200 (1026.5)
	CC-QMS			Collision cell	900 (4619.1)
86	CC-QMS	Platform, Micromass Ltd	Urine	CaPO ₄ pptate, Sr-resin, collision cell	$2000 (1.0 \times 10^4)$

the temperature of a HNO₃ solution (up to 8 M concentration) increased from 100 °C to 150 °C.169 Such losses must also be considered when dry ashing samples to decompose organic matter in solid samples.171 No significant losses were observed when ashing at 800 °C for less than 6 hours for a seaweed sample. Technetium-99 has been measured in a range of environmental and biological samples by ETV-ICP-MS (Perkin Elmer HGA-600 MS), using NH₄OH as matrix modifier to form an alkaline solution and minimize loss of Tc during drying and ashing. Depending on the daily performance, the detection limit ranged from 0.5-1.0 pg mL⁻¹ (3.2 \times 10⁻⁴ to 6.3 \times 10⁻⁴ Bq mL⁻¹) compared to 5-50 pg mL⁻¹ (3.2 \times 10⁻⁴ to 3.2 \times 10⁻² Bq mL⁻¹) using radiochemical techniques.

A modular automated separator (MARS Tc-99; Northstar Technologies, Madison WI, USA) was developed for rapid analysis of groundwater samples from boreholes at a low and intermediate waste repository. The system was coupled to ICP-QMS (Varian 810), with an activity of 1658 \pm 7 mBq L⁻¹ (2.6 ng mL⁻¹) measured for a method standard, which was in good agreement with the certified value of 1598 \pm 40 mBq L⁻¹ (2.5 ng mL⁻¹). The detection limit of the procedure developed was 0.12 mBq mL⁻¹ (0.19 pg mL⁻¹), whilst the system developed was potentially applicable to other radionuclides. In a separate study, large volume water samples were measured by ICP-QMS (Thermo X-series II) following chemical separation by ferrous hydroxide co-precipitation, and online SI using 2 TEVA columns. 168 Decontamination factors of 1×10^6 and 5×10^5 were achieved for Ru and Mo, with a detection limit of 11.5 pg m⁻³ (7.5 mBq m⁻³) for a 200 L sample. Slightly lower decontamination factors of 4×10^4 and 1×10^5 for Mo and Ru, respectively, were recorded following 2 TEVA column-separations of environmental samples prior to ICP-QMS (Thermo X-series II) measurement, with a detection limit of 0.15 mBq g⁻¹ (0.24 pg mL⁻¹).¹⁷¹ Samples were left for 1 week following separation to allow for decay of 99mTc (half-life 6.0 hours).

Measurement of 99Tc in waste stream samples was carried out at the Lan-Yu low level waste storage site, Taiwan, where Tc was previously estimated from a scaling factor based on the activity of 137Cs.173 Following acid digestion and TEVA resin separation, samples were measured by ICP-QMS (Agilent 7500a), with a detection limit of 0.021 ng g^{-1} (0.013 Bq g^{-1}). Accurate quantification meant a revision of the scaling factors being used was recommended, to the extent that reclassification of historical waste should also be carried out. The performance of ICP-QMS (Thermo X-series II) and ICP-SFMS (Thermo Element 2) was compared for low-level radioactive waste following digestion and separation using TEVA extraction chromatography resin. 170 Instrumental detection limits of 0.024 ng L^{-1} (0.015 Bg L^{-1}) and 0.015 ng L^{-1} (0.0095 Bg L^{-1}) were measured for ICP-QMS and ICP-SFMS, respectively. The minimum detectable activity for LLW samples was <8.5 mBq g^{-1} (13.4 pg g^{-1}) for ICP-QMS, <5.9 mBq g^{-1} (9.3 pg g^{-1}) for ICP-SFMS and <414-511 mBq g^{-1} (806.8 pg g^{-1}) for a gas proportional counter, whilst the method detection limit (for a 0.1 g waste sample) was 13.6 ng L^{-1} (8.5 Bq L^{-1}) for ICP-QMS, and 1.19 ng g⁻¹ (745 mBq g⁻¹) for LSC (no value was given for ICP-SFMS).

8.9 Palladium-107

Palladium-107 is a long lived ($t_{1/2} = 6.5 \times 10^6$ years) low energy beta emitting radionuclide ($E_{\text{max}} = 33 \text{ keV}$). The radionuclide is produced with a thermal neutron fission yield of 0.15%. The specific activity of 1.9×10^7 Bq g⁻¹ indicates that mass spectrometric measurement would be favorable. Isobaric interferences would arise from natural 107Ag (51.8% abundance), as well as a potential polyatomic interference from ⁹¹ZrO⁺. There are few published procedures based on ICP-MS for measurement of 107Pd. Bibler et al. (1997) used a ICP-QMS (Fisons/VG Plasmaquad) to measure fission products including 107Pd in HLW sludge's and glass.174 It was noted that direct measurement of ¹⁰⁷Pd was not possible due to isobaric interference from silver which is used to scavenge iodine and is therefore present in the waste stream. Bannochie et al. (2009) noted a similar problem and calculated 107Pd activities via measurement of ¹⁰⁵Pd and correcting for the relative fission yield of the two isotopes.175 Interference from Ag could also be overcome through chemical separation, with Bienvenu et al. (1998) noting that Pd could be separated from Ag using liquid-liquid extraction into triphenyl phosphine in nitro-2-phenylpentyl ether from 2 M HNO3 with Pd being back-extracted into 4 M ammonia.176 However, decontamination factors were not sufficient for high Ag containing samples, and an electrothermal vaporization approach was subsequently developed whereby Ag was volatilized at <1000 °C and Pd volatilized at 1500 °C.

8.10 Iodine-129

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Iodine-129 is a fission product (0.6% yield), and is also produced naturally via cosmic-ray-induced spallation of Xe, spontaneous fission of ²³⁸U, neutron bombardment of Te in the geosphere, and via thermal neutron-induced fission of stable I in the lithosphere.^{177–180} Iodine-129 decays with a half-life of 1.57×10^7 years to stable ¹²⁹Xe, and is a weak beta-gamma emitter with maximum beta decay energy of 29.78 keV (37.2%) and gamma energy of 39.58 keV (7.42%).

Iodine is a redox-sensitive element, which controls speciation and exchange between the ocean/atmosphere terrestrial environment. The high concentration and solubility in water makes 129 I a useful oceanographic tracer, makes also been used in distinguishing the discharge pathways from European reprocessing facilities at Sellafield and La Hague, Measurement of 129 I in Fukushima-contaminated samples has also been applied as an analogue for reconstruction of the distribution and activity of shorter-lived 131 (half-life 8.02 days). 184

Traditionally, radiochemical neutron activation analysis (RNAA) was the only method applied to 129I detection in environmental samples for a number of years. 185 Beta, gamma and X-ray-based techniques are limited by interferences and shielding issues, as well as long count times for low activity samples.191 For mass spectrometric determination, the primary concern is accurate detection of 129 I in the presence of high concentrations of stable ¹²⁷I (natural abundance 100%). Isotopic ¹²⁹I/¹²⁷I ratios range from 10^{-13} to 10^{-12} prior to anthropogenic nuclear activities, compared to 10^{-11} to 10^{-9} in the marine and terrestrial environment, with ratios as high as 10^{-8} to 10^{-3} at reprocessing sites. 178,180,182,186-188 The 129 I/127 I ratio measurable by the instrument is therefore a critical factor. Decomposition of solid samples and chemical separation prior to quantification are additional challenges, with losses due to volatization and/or transformation to an undesirable chemical form. 197 Separation techniques include ion chromatography, and phyrohydrolysis and solvent extraction for soil samples. 179,184,189

Until recently, ICP-MS was generally used for detection of stable 127I, while AMS was the favored technique for 129I detection, because it can achieve instrumental background ratios of 10^{-15} to 10^{-13} (Table 6). 181,182,184-186 ICP-MS was considered not to be capable of distinguishing between iodine isotopes, with applications limited to more contaminated samples with $^{129}\text{I}/^{127}\text{I}$ ratios greater than $\sim 10^{10}.^{177,180,187,189}$ Additional interferences include the low ionization efficiency of I (33.9%) owing to its high first ionization energy (10.45 eV), 190 isobaric ¹²⁹Xe (natural abundance 26.40%) present as an impurity in the plasma gas, and polyatomic interferences (particularly from collision or reaction cell instruments) including ¹²⁷IH₂, ⁹⁷MoO₂, ¹¹³CdO, ¹¹⁵In¹⁴N and ⁸⁹Y⁴⁰Ar. ^{179,191}An additional challenge is selection of an appropriate internal standard, with possible options including Cs, Re and Te. 179 One study selected Te owing to the similarity in ionization energy with I (9.01 eV), 190 whilst another did not use an internal standard owing to limitations with the elements listed.179

Iodine-129 was detected in vegetation to a detection limit of 1.4 ng kg⁻¹ (9.2 mBq kg⁻¹) using a Plasma Quad PQ2 ICP-QMS,¹⁹¹ with the ¹²⁹Xe interference corrected for using ¹³¹Xe (natural abundance 21.23%). The volatility of iodine was seen as an advantage that partially overcame the low transport efficiency (2-4% with pneumatic nebulization), as the formation of droplets in the spray chamber increased surface area and rate of vaporization, with vapors then taken up into the ICP. More recently, a Perkin Elmer Elan DRCe was applied to measurement of soils in land surrounding a fuel reprocessing site in Tokai, Japan. 187 Using oxygen as a reactive gas, the 129Xe interference was reduced, however ion transport was generally suppressed, and the instrument $^{129}\text{I}/^{127}\text{I}$ background of $\sim 10^{-8}$ was not improved in DRC mode due to 127 IH2 formation with trace impurities in the reaction gas. The background was improved to 10^{-10} using Axial Field Technology, which applies an accelerating axial field to ions in the DRC, improving instrument stability and reducing matrix effects. Fukushimacontaminated soil samples were measured using an Agilent 8800 ICP-QQQ-MS, 189 with O2 used for 129 Xe removal, and in-cell polyatomic interferences reduced by the additional quadrupole mass filter positioned before the entrance to the cell. A sample standard bracketing technique was used to correct for mass discrimination and improve reproducibility, with good agreement between ICP-MS and AMS.

ICP-SFMS (Thermo Element 2) detection of ¹²⁹I was achieved in Fukushima-contaminated Greenland snow samples. 179 Clean laboratory conditions (class 10-100) were used for sample preparation, whilst separation of Mo, In and Cd using online ion chromatography reduced polyatomic interference formation. A desolvating sample introduction system (Apex-Q (ESI)) tripled the sensitivity compared a cyclonic Peltier-cooled spray chamber, however the nitrogen carrier gas increased Xe background by an order of magnitude. The formation of 127 I2H was controlled by the instrumental setup e.g. sample uptake and gas flow rate, and plasma temperature. This is compared to an 'on-peak' baseline subtraction by Ohno et al. (2013) when using ICP-QQQ-MS, with a $^{127}IH_2/^{127}I$ ratio of 3 \times 10⁻⁸ calculated from aspiration of a 100 mg L⁻¹ stable I solution. The inability of ICP-SFMS to eliminate 129Xe was resolved by monitoring changes in the background Xe signal in Ar plasma gas to determine whether a background correction was necessary.

8.11 Caesium 135 and 137

High yield fission products 135 Cs and 137 Cs (6.58% and 6.22%, respectively) are present in environmental samples as a result of releases from nuclear power plants and reprocessing sites, nuclear accidents, and fallout from atmospheric weapons testing. Caesium-137 ($t_{1/2}=30.07$ years) is established as an important radionuclide in radiation protection, environmental monitoring and waste disposal. By comparison, 135 Cs is a long-lived radioisotope ($t_{1/2}=2.3\times10^6$ years) with a comparatively low radiation risk; however it is a significant contributor to the long term radiological risk associated with deep geological disposal. Furthermore, the 135 Cs/ 137 Cs ratio varies with reactor, weapon and fuel type, and recent studies have achieved precise

determination of 135Cs/137Cs ratios as a forensic tool to identify the source of radioactive contamination. 195-200

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Caesium-137 decays by beta emission to short-lived metastable isomer 137mBa, with maximum energies of 514 keV (94.4% yield) and 1175 keV (5.4% yield). This is immediately accompanied by gamma ray emission of 661.7 keV (85.1% yield) to form ¹³⁷Ba. Gamma spectrometry is generally favored because it exploits the good gamma vield of the 661.6 keV energy that is not susceptible to significant absorption, whilst the ability to directly count most samples without any chemical separation is also beneficial. Caesium-135 decays with a maximum beta particle energy of 269 keV, however, measurement by beta counting is restricted by any ¹³⁷Cs also present in the sample with a radioactivity concentration that is typically 5 orders of magnitude higher. Measurement of ¹³⁵Cs by mass spectrometry offers a considerable advantage because of the larger atom population compared with radioactive decays. A review of measurement of ¹³⁵Cs and ¹³⁷Cs and ¹³⁵Cs/¹³⁷Cs isotope ratios in environmental samples has been published recently,²⁰¹ and Table 7 gives a summary of recent procedures for ¹³⁵Cs/¹³⁷Cs measurement.

A key challenge for mass spectrometric measurement is removal of isobaric interferences from naturally occurring ¹³⁵Ba and ¹³⁷Ba (isotopic abundances 6.6% and 11.2%, respectively), peak tailing from stable 133Cs (isotopic abundance 100%), and polyatomic interferences including 95Mo40Ar, 97Mo40Ar, ¹¹⁹Sn¹⁶O and ¹²¹Sb¹⁶O. For mass spectrometric measurement, Cs separation has commonly been achieved using ammonium molybdophosphate (AMP) followed by anion exchange and/or cation exchange chromatography, most commonly Dowex AG1 Dowex AG50W (both Bio-Rad, USA), respectively. 1,195,197,198,200,202 Other techniques include extraction chromatography using calixarenes,203 and potassium nickel hexacyanoferrate (KNiFC).200 Barium-138 (natural abundance 71.7%) has been used to assess the effectiveness of separation at masses 135 and 137, however any interference correction will reduce the accuracy of 135Cs/137Cs values. Caesium-137 values can be verified by gamma spectrometry, however if a significant correction is required this will not validate the ¹³⁵Cs result. ^{204–206}

Online separation has been applied to Cs and Ba separation prior to ICP-MS analysis. For example, Isnard et al. (2009) achieved a 135 Cs and 137 Cs detection limit of 2 pg L^{-1} (8.2 × 10 $^{-8}$ Bq L^{-1})

Table 6 Summary of recent procedures for measurement of ¹²⁷I/¹²⁹I

Reference	Iodine isotope	Instrument	Model	Matrix	Chemical separation	LOD, pg L ⁻¹ (Bq L ⁻¹)
192	127	QMS	X series II	Lake sediments		
	129	AMS				
181	127	AMS		Aerosol particles		
	129					
193	127	AMS		Environmental samples		
	129					
177	127	QMS	X series II	Scandinavian soil and	Sequential extraction into	
	129	AMS		sediment	soil/sediment fractions	
178	127	QMS	X series II	Baltic sea seawater		
	129	AMS				
187	127	DRC-MS	Elan DRCe	Soil	O ₂ -based extraction with heating,	
	129				extraction from activated carbon	
188	127	CC-QMS	Agilent	Baltic sea, lake and rain	Anion exchange, solvent extraction	
	129	AMS	7500ce/cx	waters from Finland		
182	127	QMS		Seawater and aerosol filters	Liquid-liquid exchange for air filters	
	129	AMS			Seawater samples anion exchange	
					Matrix adjustment for I-127, but no	
					matrix separation	
194	127	AMS		Pacific ocean and Japan	Solvent extraction	
	129			seawater		
190	127	SFMS	Element 2	Japanese coastal seawater		2300 (0.015)
	129			(pre-Fukushima)		
183	127	QMS	X series II	Fukushima seawater samples	Anion exchange	
	129	AMS				
189	127	QQQ-MS	Agilent 8800	Fukushima soil samples	Pyrohydrolysis and solvent	
	129				extraction	
189	127	ORS-QMS	Agilent	Fukushima soil samples	Pyrohydrolysis and solvent	1500 (0.0098)
	129		7700X		extraction	
179	127	SFMS	Element 2	Snow samples from Greenland	Online IC separation	700 (0.0046)
	129			(Fukushima contaminated)	(anion exchange)	
184	127	ORS-QMS	Agilent 7700	Fukushima soil samples	Pyrohydrolysis	
		QQQ-MS	Agilent 8800			
	129	AMS				
180	127	AMS		Woodward iodine corporation		
	129					

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and 1 pg L $^{-1}$ (3.2 imes 10 $^{-3}$ Bq L $^{-1}$), respectively, using on-line chromatographic separation with a CG3 IonPac guard column (Dionex, USA) for a spiked groundwater matrix. 207 Following AMP and anion exchange separation, Taylor *et al.* (2008) achieved online Cs/Ba separation in sediment samples by attaching a Dionex CS12A 3 imes 150 mm cation exchange column onto a high performance liquid chromatography (HPLC) module, and coupling this system with ICP-MS. 195

Sample introduction-based separation has also been applied to Cs/Ba separation. Song $et\ al.$ used ETV with potassium thiocyanate (KSCN) as a chemical modifier. ^9 At a volatilization temperature of 1100 °C, Cs was vaporized in a 10^4 excess of Ba, with a Ba signal increase at mass 135 of only 1%. The detection limit of 135 Cs using QMS (Perkin Elmer Elan 5000) was calculated as 0.2 ng L $^{-1}$ (0.01 Bq L $^{-1}$) An alternative online separation technique is capillary electrophoresis (CE-ICP-MS), which was effectively applied to 135 Cs/ 137 Cs measurement in plutonium uranium redox extraction (PUREX) and mixed oxide fuel (MOX) samples. 208 The procedure achieved 133 Cs detection limits of 6000 ng L $^{-1}$ (245.0 Bq L $^{-1}$) and 4 ng L $^{-1}$ (0.2 Bq L $^{-1}$) in ICP-QMS (Perkin Elmer Elan 5000) and ICP-SFMS (Thermo Element 2), respectively.

A reaction cell has been applied to Cs separation, with N₂O, and CH3Cl both effectively used as reactive gases. 195,209 For example, kinetic data from Lavrov et al. (2004) indicates the reaction for Ba with N₂O has an efficiency of 32%, compared to 0.01% for Cs.210 An Agilent 8800 ICP-QQQ-MS has been used for ¹³⁵Cs/¹³⁷Cs detection in rainwater and environmental samples contaminated following the accident at the Fukushima Daiichi NPP. 198-200 Using N2O as a reactive gas, a 135Cs and 137Cs detection limit of 10 pg L^{-1} (equivalent to 4.1×10^{-7} Bq L^{-1} and 3.2×10^{-2} Bq L⁻¹, respectively) was achieved in a Ba-free solution, compared to 100 pg L^{-1} (4.1 \times 10⁻⁶ Bq L^{-1}) and 270 pg L^{-1} (8.6 × $10^{-1} \text{ Bq L}^{-1}$) for ¹³⁵Cs and ¹³⁷Cs, respectively, in solutions containing 100 μg L⁻¹ Ba. These values are higher than the instrumental and method detection limits achievable using ICP-SFMS.1 However, the ICP-QQQ-MS offers the combination of chemical and instrument-based separation, whilst the two quadrupoles have measured a 133Cs/135Cs abundance sensitivity of $\sim 10^{-10}$.

A procedure using a Thermo Element 2XR ICP-SFMS achieved an instrumental detection limit of 1 pg L⁻¹ for stable ¹³³Cs. The absence of instrument-based separation meant highly efficient chemical separation techniques using high purity reagents and clean laboratory conditions was required to minimize Ba contamination prior to sample introduction. Following chemical separation, a higher method detection limit of 20 pg L^{-1} was calculated (equivalent to 8.2×10^{-7} Bq L^{-1} and 6.4×10^{-2} Bq L⁻¹ for ¹³⁵Cs and ¹³⁷Cs, respectively). Time independent fractionation is an issue affecting accurate isotope ratio measurements by ICP-MS. One cause of this is space charge effects, as lighter ions are deflected to a greater extent than heavier ions during ion extraction.207 The absence of a 135Cs/137Cs certified reference material makes accurate mass bias correction challenging, and limits the ability to compare results between studies. Isnard et al. (2009) applied a sample standard bracketing technique using natural 121Sb, 123Sb and ¹⁵¹Eu, ¹⁵³Eu to correct for mass bias, as the average mass of these isotopes is close to that of ¹³⁷Cs.²⁰⁷ The isotopic fractionation affecting TIMS and ICP-MS, as well as the absence of a certified reference material to verify accuracy of ¹³⁵Cs/¹³⁷Cs measurements, led to a comparison of the two techniques for uranium oxide and mixed oxide fuel samples.

Of the alternative mass spectrometric techniques, Lee *et al.* (1993) were the first to develop a procedure for measuring ¹³⁵Cs/¹³⁷Cs in environmental samples using TIMS.²¹¹ There have been multiple recent TIMS applications including sediment core samples to highlight fractionation of ¹³⁵Cs and ¹³⁷Cs following aboveground nuclear tests at the Nevada Nuclear test site, ¹⁹⁶ and environmental samples contaminated by the Fukushima Daiichi accident.²⁰² TIMS is affected by time-dependent isotopic fractionation, as lighter isotopes are preferentially evaporated from the filament. The lack of an available certified Cs isotopic standard makes it difficult to measure mass fractionation, however has been anticipated to be small relative to estimated uncertainties.¹⁹⁶ Alternatively, mass fractionation can be corrected by measuring ¹³³Cs/¹³⁷Cs and ¹³⁵Cs/¹³⁷Cs ratios during the analysis.²⁰⁷

Pibida *et al.* used both RIMS and TIMS to determine the chronological age of nuclear fuel burn up samples through measurement of ¹³⁵Cs/¹³⁷Cs.²¹² For low-level detection in environmental samples, improvements in overall efficiency and chemical separation were required, whilst the absence of a commercially available instrument and time-consuming optimization of instrument efficiency and chemical preparation are the main limitations to this approach. AMS measurement is limited by the high energies required to separate Cs and Ba isobars, which makes separation a complex and costly process.²⁰⁶ A method to produce a beam of Cs anions, and production of suitable yield tracers to determine the efficiency of the process are also required.²¹³

8.12 Samarium-151

Samarium-151 is a lanthanide element fission product, with a 235 U thermal neutron fission yield of 0.53%. The radionuclide has a half-life of 94.7 years giving a specific activity of 9.25 \times 10¹¹ Bq g⁻¹. Quantification of 151 Sm is typically required in determining nuclear fuel burn up and in nuclear waste characterization. Samarium-151 is a pure beta emitting radionuclide ($E_{\rm max}=76$ keV), which is typically analyzed using liquid scintillation counting following separation of 151 Sm from other rare earth radionuclides and 241 Am. Separation of Sm from other lanthanides is usually achieved using either extraction chromatographic materials such as HDEHP-based resins, 218 ion exchange chromatography, or HPLC^{219,220} based techniques. More recently, isotachophoretic separation coupled with ICP-MS has been developed for lanthanide quantification of spent nuclear fuel. 221

There are only a limited number of papers relating to measurement of ¹⁵¹Sm by ICP-MS, although ICP-MS has also been used in nuclear fuel burn up assessments to measure the variation in stable Sm isotopic composition.⁸⁵ Isobaric interferences arise from stable ¹⁵¹Eu (47.81%) and polyatomic species ¹¹⁹Sn ¹⁶O₂, ¹³³Cs ¹⁸O, ¹³⁴Ba ¹⁷O, ¹³⁵Ba ¹⁶O and ¹⁵⁰Nd ¹H. Due to the potential for interference from Eu and Nd and their

Table 7 Summary of recent procedures for measurement of ¹³⁵Cs/¹³⁷Cs

Reference	Instrument	Model	Matrix	Chemical separation	LOD, pg L^{-1} (Bq L^{-1} 135 Cs/ 137 Cs)
214	QMS		Spent fuel pellets and solutions	Cation exchange	
215	QMS	VG Elemental Plasma Quad 2	Waste tank sludge and supernatants	Ba(OH) ₂ precipitation	$209\ 000\ \big(8.53/6.7\times 10^5\big)$
216	QMS	Perkin Elmer Elan 5000	High activity waste	Cation exchange	$16\ 000\ (0.65/5.12\times 10^5)$
79	QMS	Perkin Elmer Elan 5000	Cs-137 standard	ETV sample introduction	200 (0.01/640.1)
217	QMS	PQ Excell	Groundwater	On-line cation exchange	$2(8.2 \times 10^{-5}/6.4)$
207 and 209	MC-ICP-MS	GV Isoprobe	Spent fuel pellets	Anion exchange and HPLC, reaction cell	, ,
1	SFMS	Element 2XR	Fission product standard	AMP, cation exchange, extraction chromatography	$50 \ \big(2.0 \times 10^{-3}/160.0\big)$
208	SFMS	Perkin Elmer Elan 5000	Simulated spent fuel	Capillary electrophoresis	$4000 (0.16/1.3 \times 10^4)$
195	DRC-MS	Perkin Elmer Elan DRC II	Soil and sediment	AMP, anion exchange, online cation exchange, reaction cell	200 (0.01/640.1)
198	QQQ-MS	Agilent 8800	Rainwater	Reaction cell	270 (0.01/864.2)
199	QQQ-MS	Agilent 8800	Environmental samples	AMP, anion exchange, cation exchange, reaction cell	,
200	QQQ-MS	Agilent 8800	Environmental samples	AMP, anion exchange, cation exchange, reaction cell	$10/6 \; \big(4.1 \times 10^{-4}/19.2\big)$

chemical similarity to Sm, reliable ¹⁵¹Sm measurements by ICP-MS rely on robust chemical separation of the rare earth elements.

Alonso *et al.* (1995) measured actinides and fission products in spent nuclear fuel using HPLC coupled to an Elan 5000 ICP-QMS. ²¹⁴ Lanthanide separation was performed using CG5/CS5 guard/separation columns and isocratic elution with 0.1 M oxalic acid and 0.19 M LiOH. The HPLC eluent was introduced into the ICP-MS via a cross flow nebulizer and Scott type, double pass spray chamber. Plasma RF power was optimized at 1050 W. Spent nuclear fuel ¹⁵¹Sm/¹⁵⁰Sm values of 0.0119–0.0175 were reported. The same experimental set-up was used to measure fission products, including ¹⁵¹Sm, in UO₂ and MOX fuel pellets. ²⁰⁴

Wolf *et al.* (2004) coupled HPLC and ICP-MS to separate rare earth elements prior to measurement.⁸⁴ HPLC separation was achieved using an Amberlite CG50 ($2 \times 50 \text{ mm}^2$) guard column coupled with a Dionex CS5A ($2 \times 250 \text{ mm}^2$) analytical column, a linear gradient elution of 0.040.0.26 M hydroxyisobutyric acid (HIBA) with a 0.30 mL min⁻¹ flow rate. The eluent from the HPLC was introduced directly into the ICP-MS *via* a CETAC MCN-100, Model M-2 pneumatic microconcentric nebulizer. Measurements were performed using a Fison PlasmaQuad 2+quadrupole ICP-MS. The within-sample precision of ¹⁵¹Sm measurement was quoted as 6%.

Isnard *et al.* (2005) separated Sm and Gd off-line using HPLC, followed by MC-ICP-MS measurement of Sm isotopes, including ¹⁵¹Sm, using a GV Isoprobe-N MC-ICP-MS.²²² Sample introduction was *via* a microconcentric nebulizer and cyclonic spray chamber. Plasma RF power was set at 1350 W, with measurements performed *via* static Faraday detectors. Analysis of MOX spent nuclear fuel gave mean ¹⁵¹Sm/¹⁵⁰Sm atom ratios of 0.04319 with a standard deviation of 0.2%. Pitois *et al.* (2008) coupled capillary electrophoretic separation with both ICP-QMS

(PerkinElmer ELAN 5000) and ICP-SFMS (Thermo Element 2) to measure rare earth fission products although not $^{151}\mathrm{Sm.}^{208}$ Detection limits of 8 ng mL $^{-1}$ (ICP-QMS) and 7 pg mL $^{-1}$ (ICP-SFMS) were reported for rare earth elements, which would be equivalent to 6500 Bq mL $^{-1}$ and 6.5 Bq mL $^{-1}$ of $^{151}\mathrm{Sm.}$

8.13 Lead-210

The longest lived radioisotopes of Pb are 202 Pb ($t_{1/2}=5.3\times10^4$ years), 205 Pb ($t_{1/2} = 1.5 \times 10^7$ years), and 210 Pb ($t_{1/2} = 22.23$ years). Lead-210 is naturally occurring as part of the ²³⁸U decay series, and has been measured in the fields of geochronology, air flux measurements of radon^{223,224} radiation protection associated with uranium mining, and environmental contamination from the iron and steel industry.225 Lead-210 is a beta emitting radionuclide (Emax 63.5 keV, 100% intensity), and is also a weak alpha (decay energy 3.72 MeV, 1.9×10^{-6} % intensity) and gamma emitter (12.56 keV, 22.0% intensity). Measurement is therefore potentially achievable by multiple radiometric techniques. Gamma spectrometry measurement is affected by limited sample throughput, and prior knowledge of the sample geometry and composition is required. The low energy gamma ray, selfabsorption and interferences from other gamma emitters and X-rays must also be considered.²²⁴ Lead-210 measurement by beta counting using a GM counter is achieved through measuring the ²¹⁰Bi daughter (1.2 MeV Emax), following an ingrowth period of 8-9 days. Finally, measurement by alpha spectrometry is achievable through measurement of ²¹⁰Po, however an ingrowth time of greater than 3 months is required.224

Mass spectrometric measurement of ²¹⁰Pb is affected by peak tailing from stable ²⁰⁸Pb (52.4% abundance), and polyatomic interferences associated with Pt, Hg, Bi and Pb. ²²⁶Polyatomic interferences combined with the relatively short half-life of ²¹⁰Pb have been highlighted as the causes for measurement precision being inferior to that of alpha

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spectrometry.²²³ Pre-concentration of ²¹⁰Pb prior to measurement is achievable by co-precipitation with cobalt-ammonium pyrrolidine dithiocarbonate,226 or as lead sulphate,225 whilst chemical separation from interferences can be performed by extraction chromatography using Sr-resin.226 Given the Pt-based interferences, Pt sampler and skimmer cones should be avoided, as high backgrounds have been reported at m/z = 210.225

Larivière et al. compared ICP-QMS (Perkin Elmer Elan 5000), ICP-DRC-MS (Perkin Elmer Elan 6100) and ICP-SFMS (Thermo Element 2) for measurement of ²¹⁰Pb in water. ²²⁶ All instruments were equipped with an Apex-Q sample introduction system, and a detection limit of 90 mBq L⁻¹ (0.03 pg L⁻¹) was recorded for final measurements. The tailing removal was greatest using ICP-DRC-MS (>3 \times 10⁹), compared to >2 \times 10⁶ for QMS, and 1×10^5 for SFMS. In a separate study, CC-ICP-MS (Agilent 7500ce) was applied to measurement of standards and solutions spiked with ²¹⁰Pb, and sediment samples from waste associated with an on oil and gas production field in Egypt.224 The recovery was assessed using isotope dilution with 206Pb. The results showed good agreement with those obtained by gamma spectrometry, with a detection limit of 0.21 ng L^{-1} (593.24 Bq L^{-1}). The formation of polyatomic interferences was reduced using krypton as a collision gas at a flow rate of 0.4 mL min⁻¹, which was determined by assessing the 208Pb/206Pb ratio with the change in gas flow rate. It was also noted that potential lead hydride formation could be removed using a desolvating sample introduction system. Technologically enhanced 210Pb as a result of iron and steel manufacture has been measured along with ²¹⁰Po using a combination of ICP-QMS (Perkin Elmer Elan 9000), gamma spectrometry and alpha spectrometry, 225 with a Pb activity range of <7 Bq kg⁻¹ (2.5 pg kg⁻¹) to 4240 Bq kg⁻¹ (1.5 ng kg⁻¹).

8.14 Radium-226 and 228

Radium-226 and 228 are naturally occurring radioactive materials, with large amounts produced from mining and extraction of fossil fuels, as well as from the phosphate industry and uranium mine tailings.^{227,228} Radium-228 is also continually produced from the decay of 232Th in shelf sediments.229 The natural occurrence of these nuclides has led to application as a tracer for ocean circulation, seawater sediment fluxes and particulate residence times.²²⁹ Radium-226 decays with a halflife of 1599 years, with maximum alpha decay energy of 4.79 MeV (94.6%) and gamma emission of 186 keV (32.8%). The half-life of ²²⁸Ra is comparatively short (5.76 years), with a maximum beta decay energy of 39 keV.

Radiometric detection is achievable by alpha or gamma spectrometry, however both require a lengthy ingrowth period for secular equilibrium to be established between 228Ra and ²²⁸Th. ^{229,230} Whilst ICP-MS measurement needs lengthy sample treatment to remove multiple potential polyatomic interferences prior to sample introduction (including 88Sr138Ba, ⁸⁷Sr¹³⁹La, ⁸⁶Sr¹⁴⁰Ce, ²⁰⁸Pb¹⁸O, and ¹⁸⁶W⁴⁰Ar), no ingrowth time is required, and the sample analysis time is also significantly lower than radiometric measurements. 228,229 Chemical separation is generally achieved by diffusive gradients in thin films (DGT) or manganese dioxide (MnO2) combined with ion exchange and/or extraction chromatography. 227,230-233

Measurement of the ²²⁸Ra is challenging given the significantly shorter half-life and lower concentration in environmental samples compared to ²²⁶Ra. Recent applications have taken advantage of the high instrumental sensitivity of ICP-SFMS for ²²⁶Ra/²²⁸Ra detection, using either single^{227,231–233} or multi-collector instruments^{229,230,233} (Table 8). Instruments have been successfully operated at both low resolution, 231-233 and medium resolution,227 with the latter setup improving polyatomic interference removal at the expense of one order of magnitude sensitivity loss when analysing seawater samples (1200 cps per ng L^{-1} at low resolution compared to 100 cps per ng L⁻¹ at medium resolution). A recent study of ²²⁶Ra in high salinity seawater using a Perkin Elmer NexION 300x achieved a detection limit of 100 pg L^{-1} (3.7 Bq L^{-1}), ²²⁸ which is several orders of magnitude higher than that of ICP-SFMS. Multiple elements have been applied for mass bias correction, given the absence of a certified 226Ra/228Ra reference material, including $^{207}\text{Pb}/^{208}\text{Pb}$, $^{238}\text{U}/^{235}\text{U}$, and $^{229}\text{Th}/^{232}\text{Th}$. 229,230

8.15 Protactinium-231

Protactinium-231 is the longest lived intermediate daughter of 235 U, with a half-life of 3.28 \times 10⁴ years, and can also be produced as a by-product in the thorium fuel cycle via a fast neutron reaction with 232Th or 232U. Protactinium-231 is an alpha emitting radionuclide (5.149 MeV), decaying to ²²⁷Ac. A major application of 231Pa is in precise isotope ratio calculations for uranium-series age dating, and as an indicator of climate change, and is commonly measured in combination with Th and U isotopes. This required high precision measurements of isotopic ratios, which can be achieved using TIMS (typically 2-3% precision). 234-236 However, extensive matrix separation, and challenges associated with sample loading limit sample throughput.234 Alpha spectrometry has been used for measurement of 231Pa in seawater samples, however cubicmetre levels of sample are required, followed by extensive chemical separation, with low final measurement precision (5-10%).²³⁴ ICP-MS measurement can rival the precision of TIMS, whilst achieving a significant improvement in sample throughput. The main interference is peak tailing from ²³²Th, which can be removed by chemical separation prior to measurement if necessary. Protactinium-234 ($t_{1/2} = 26.97$ days) has been used a spike in some studies,234,237 however, consideration must be given to decay to ²³³U, leading to changes in the ²³¹Pa/²³³Pa ratio over time, with losses of Pa to beaker walls also highlighted as an issue.

Applications include a fossil coral fragment measured by MC-ICP-MS (FISONS PLASMA 54), with samples introduced by a CETAC MCN6000 desolvating nebulizer.²³⁸ The reduced sample handling, sample size and increased throughput compared to TIMS were highlighted as advantages. Choi et al. (2001) measured 231Pa along with 230Th in seawater by ICP-SFMS (Element) equipped with a desolvating nebulizer (Cetac MCN-6000), with a detection limit of 0.4 fg, corresponding to 0.02 fg g⁻¹ $(3.5 \times 10^{-8} \text{ Bq g}^{-1})$ in 20 L seawater samples.²³⁴ In a separate

Table 8 Summary of recent procedures for measurement of ²²⁶Ra

Reference	Instrument	Model	Matrix	Chemical separation	$LOD pg L^{-1} (Bq L^{-1})$
228	CC-QMS	Perkin Elmer NexION 300x	High salinity wastewater	AG50 (IE)	100 (3.7)
227	SFMS	Element II	Seawater		
231	SFMS	Element II	Water and sediment pore water	DGT	0.5 (0.02)
232	SFMS	Element II	Water and sediments	DGT, MnO ₂ pptate, Sr resin (EC)	0.5 (0.02)
233	MC-ICP-MS	Nu plasma	Natural waters	MnO ₂ , AG50 (IE), Sr resin (EC)	0.05 (0.002) 0.05 (0.002)
230 229	MC-ICP-MS MC-ICP-MS	Neptune Nu instruments	Seawater and suspended particles Seawater	MnO ₂ , AG50, AG1, Sr resin	0.091 (0.003)

study, silicate rock samples were measured following anion exchange and extraction chromatography (TRU (Triskem International)) separation by MC-ICP-MS (Thermo Neptune) equipped with a desolvating nebulizer (Cetac Aridus), with a detection limit of 200 fg $\rm L^{-1}$ (3.5 \times 10 $^{-6}$ Bq $\rm L^{-1}).^{237}$

8.16 Thorium isotopes

Thorium is a naturally occurring element, with an abundance in the earth's crust 3-4 times higher than uranium, and has also been used as an alternative fuel for nuclear fission. The most abundant isotope is 232 Th with a half-life of 14.02×10^9 years, and undergoes alpha decay to ²²⁸Ra. Additional radioisotopes with long half-lives measurable by ICP-MS are 229 Th ($t_{1/2} = 7340$ years) and 230 Th $(t_{1/2} = 7.5 \times 10^4 \text{ years})$. Thorium is commonly measured as ²³²Th along with isotopes of U and Pu in bioassay samples, as well as with uranium in chronology, paleoclimatology, archaeology, hydrology, geochemistry and oceanography.239,240 Alternative mass spectrometric measurement techniques include TIMS and SIMS.241 When Th is measured by ICP-MS in combination with U, the 232Th tailing can impact measurement of ²³³U, as can the formation of ²³²Th¹H.²⁴² A summary of recent measurement of Th isotopes by ICP-MS is given in Table 9, with a review of mass spectrometric determination of Th published elsewhere.243

As instrument sensitivity has improved, the low-level measurement of Th is potentially impacted by environmental contamination during sample preparation. The concentrations of Th, U and their progenies in the reagents and labware used has become an increasingly important issue, with clean laboratory conditions and cleaning of materials prior to use required for some applications. Hoppe *et al.* (2013) investigated the use of low background materials and maintenance of low background levels of Th and U.²⁴⁴ It was concluded that sample preparation is the limiting factor affecting sensitivity for very low level measurements, and thorough cleaning and acid leaching of these materials has made very sensitive measurements possible.

Several studies have measured Th in combination with U and/or Pu in urine. ^{242,245,246} Becker *et al.* (2004) measured Th and U using LA-ICP-MS, which minimized sample preparation, whilst quantification issues associated with laser ablation were resolved with matrix-matched standards. ²⁴⁵ Measurements were performed by both ICP-QMS (Perkin Elmer Elan 6000) and

ICP-SFMS (Thermo Element) coupled to laser ablation (CETAC LSX 200). The Th recovery ranged from 97-104% at doping concentrations of 0.52-2.49 ng L^{-1} (2.1 \times 10⁻⁶ Bq L^{-1} to $1.0 \times 10^{-5} \,\mathrm{Bg} \,\mathrm{L}^{-1}$) with detection limits of 0.4 $\mathrm{ng} \,\mathrm{L}^{-1}$ (3.5 \times 10⁻⁸ Bq g^{-1}) and 0.2 ng L^{-1} (8.1 \times 10⁻⁷ Bq g^{-1}) for ICP-QMS and ICP-SFMS, respectively. The study highlighted the advantages of ICP-MS for actinide measurement over alpha spectrometry, which was a less sensitive approach that required more extensive sample preparation. In a separate study, Shi et al. (2013) measured Th, U and Pu in urine samples with regards to dose assessment, as well as measuring decommissioning samples including tape and paint.242 Samples were measured by ICP-SFMS (Thermo Element XR) with an Elemental Scientific Apex desolvating sample introduction system, which reduced the $^{232}\mathrm{Th}^{1}\mathrm{H}$ formation rate to 1.3 \times 10⁻⁵, compared to 6.6 \times 10⁻⁵ for solution nebulization using a Micromist nebulizer. Cozzella and Pelttirossi (2013) measured Th, U and Pu in urine by ICP-MS rather than alpha spectrometry, highlighting the ability to process a large number of samples whilst maintaining an acceptable measurement uncertainty.246 Samples were measured by ICP-QMS (Thermo X-series) with a Burgener nebulizer. Prior to sample introduction, Th was stabilized in urine with Triton X-100 and then mixed with MICROTENE-TOPO, shaken vigorously and then loaded onto a column prior to elution. Thorium recovery ranged from 93 \pm 0.2% to $120 \pm 1.2\%$ over a spiked concentration range of 0.5–2 $\mu g L^{-1}$ $(2.0 \times 10^{-3} \text{ to } 8.1 \times 10^{-3} \text{ Bq g}^{-1}).$

Thorium and U were measured along with multiple rare earth elements in natural spring water samples in Brazil, in relation to supplying safe potable water to nearby towns.247 Water samples were measured by ICP-DRC-MS (Perkin Elmer ELAN DRC-e) equipped with a sea spray nebulizer and cyclonic spray chamber. The method detection limit was 0.5 ng L^{-1} (2.0 \times 10⁻⁶ Bq g⁻¹) Avivar et al. (2011) measured Th and U in multiple environmental samples (different water samples, a phosphogypsum sample, and a channel sediment reference material) using multi-syringe FIA (MSFIA), using UTEVA resin (Triskem International) for online separation and pre-concentration.248 The system was coupled to ICP-DRC-MS (Perkin Elmer Elan DRC-e) equipped with a Scott spray chamber and cross-flow nebulizer. Combination of LOV with MSFIA achieved a 232 Th detection limit of 2.8 ng L^{-1} $(1.1 \times 10^{-5} \text{ Bg L}^{-1})$, compared to 120 ng L⁻¹ $(4.9 \times 10^{-4} \text{ Bg L}^{-1})$ using FIA only. Tuovinen (2015) compared ICP-MS to XRF, ERD,

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EPMA, gamma spectrometry and alpha spectrometry for determination of Th and U in ore and mill tailing samples collected in Finland. Thorium was measured over a concentration range of 7–157 mg $\rm L^{-1}$ (2.8–637.2 Bq $\rm L^{-1})$ by ICP-QMS (Agilent 7500 ce/cx), with the results showing generally good agreement with other techniques. The lowest values were measured by ICP-MS compared to other techniques, which was the result of challenges with sample preparation.

8.17 Uranium isotopes

Uranium is naturally occurring, with an average concentration of $\sim\!\!4~\mu g~g^{-1}$ in the terrestrial crust, 3 $\mu g~L^{-1}$ in seawater (uniformly distributed in the world's oceans) and ranging from 0.5–500 $\mu g~L^{-1}$ in surface freshwater depending on the extent of contamination. The environmental occurrence of uranium is mostly in hexavalent form, associated with oxygen in nature as the uranyl ion $U{O_2}^{2^+}$. Under strongly reducing conditions, uranium is present in tetravalent form in strongly reducing medium such as high organic material (UO₂). Uranium-238 is the major isotope (99.27%), with additional minor isotopes of 234 U (0.006%) and 235 U (0.72%).

Uranium was one of the early nuclides to be measured by ICP-MS, with studies initially focusing on the elemental concentration of ²³⁸U. As instrumental performance improved, there was an increasing focus on isotopic ratio measurements, initially ²³⁵U/²³⁸U, and more recently ²³⁴U/²³⁸U, and ²³⁶U/²³⁸U, with isotopic analysis enabling distinction between exposure to natural and anthropogenic U sources. Reviews of uranium determination using atomic spectrometric techniques, and in environmental samples, are published elsewhere, ^{78,251} and Table 10 gives a summary of recent ICP-MS procedures for U isotopic measurements.

Several spectral interferences must be considered for uranium analysis, with Ir, Bi, Th or Ru effectively used for interference correction.⁷⁸ Potential platinum-argide interferences from the use of platinum cones can be avoided by using nickel cones,²⁵² whilst chloride-based interferences with

elements including Au and Hg must also be considered.78 Uranium-236 detection is potentially affected by both ²³⁵U¹H formation, and tailing from 238U. Sector field instruments are a popular choice for uranium determination due to low background, high sensitivity at low resolution, and ability to remove interferences at high resolution.78 Additionally, the simultaneous data acquisition of MC-ICP-MS combined with sector field sensitivity leads to more precise isotope ratio determination.253 Determination of uranium concentration and/or isotopic ratios in bioassay samples is commonly carried out through measurement of urine samples, as it is non-invasive, whilst the complexity of the sample matrix represents a challenge for isotope ratio determination. 252-254 For example, uranium was measured as part of the Baltimore VA Depleted Uranium Clinical Follow-up Program by ICP-SFMS (Thermo Element 2).252 Measurement of 235U/238U and 236U/238U was compared for a quartz concentric nebulizer with a cyclonic spray chamber, and APEX Q (Elemental Scientific) sample introduction system. Superior sensitivity was recorded using the Apex O, which was preferable for low U concentrations, whilst the concentric nebulizer and cyclonic spray chamber was more robust and suitable for higher U concentrations. The limited abundance sensitivity and ²³⁵UH formation meant the background equivalent concentration of 236U was 25 times higher than for ²³⁵U and ²³⁸U, with more scattered ²³⁶U/²³⁸U ratios at total uranium concentrations <10 ng L-1. Additionally, the accuracy of 235U measurements was poor when total U concentrations were less than 5 ng L^{-1} .

In a separate study, Arnason *et al.* (2015) carried out an interlaboratory comparison for uranium concentration and multiple isotope ratio values in urine, with results from sites using ICP-QMS or ICP-SFMS.²⁵⁴ As the total uranium concentration decreased, the concentration measured by isotope dilution only were higher than those that used chemical separation or digestion prior to measurement. At a concentration of 50 ng $\rm L^{-1}$, the predicted $\rm ^{234}U/^{238}U$ value was 0.000053. Sites that carried out digestion and chemical separation prior to measurement yielded values from $\rm 5.2 \times 10^{-5}$ to $\rm 7.2 \times 10^{-5}$

Table 9 Summary of recent procedures for measurement of Th isotopes

Reference	Instrument	Model	Matrix	Chemical separation	LOD^a , pg L^{-1} (Bq L^{-1})
246	QMS	X-series	Urine	MICROTENE-TOPO chromatography	
250	QMS	X-series II	River water	Stacked TEVA and DGA	²²⁹ Th: 4490 (34.9) ²³⁰ Th: 4490 (3.4) ²³² Th: 694 000 (2.8 × 10 ⁻³)
249	QMS	Agilent 7500 ce/cx	Ores and mill tailings		,
245	QMS	Perkin Elmer Elan 6000	Urine	TiCl ₄ precipitation and Dowex 1-x8	QMS-400 (1.6 \times 10 ⁻⁶)
	SFMS	Element			SFMS-200 (8.1 \times 10 ⁻⁷)
242	SFMS	Element 2XR	Urine, decommissioning samples including paint and tape		,
247	DRC-MS	Perkin Elmer DRC-e	Natural waters		$500~(2.0\times10^{-6})$
248	DRC-MS	Perkin Elmer DRC-e	Environmental samples	FIA, UTEVA	$2800 (1.1 \times 10^{-5})$
/ C	232				

^a LOD for ²³²Th unless indicated otherwise.

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compared to 1.5 \times 10⁻³ to 1.8 \times 10⁻² for isotope-dilution-only methods. A significant positive bias was also seen for ²³⁵U/²³⁸U for dilution only-methods, accounted for by polyatomic interference at m/z = 235. Finally, for $^{236}\text{U}/^{238}\text{U}$, only studies that applied chemical separation were considered, and, of these five studies, four either did not report a result or reported a nondetect result. The conclusion from this study was that accurate and precise isotope ratio measurements was more dependent on the analytical methodology and instrument capability than measurement of total U concentration. Arnason et al. also measured uranium isotopes in urine samples at volumes from 1-8 mL by ICP-SFMS (Element 2) equipped with a Cetac Aridus desolvating sample introduction system, following chemical separation using UTEVA extraction chromatography separation. 253 At spike concentrations of 2.5-25 ng L^{-1} , 235 $U/^{238}U$ was successfully detected, with an improvement in precision as U concentration increased. By comparison, ²³⁴U/²³⁸U was only detected in samples spiked with 25 ng L⁻¹. It was also realized that whilst a higher urine volume achieved better precision, the uranium recovery was lower.

Liu et al. (2011) applied extractive electrospray ionization mass spectrometry (EESI-MS) to determination of ²³⁵U/²³⁸U in uranyl nitrate solutions, prepared from samples including natural water, uranium ore and soil.255 EESI-MS is typically used for organic compounds, and in this study samples were directly measured without pre-concentration or separation. The resulting mass spectrum looked for detection of uranyl nitrate at m/z $456 \, [^{238}UO_2(NO_3)_3^{-1}]$, as well as peaks for 234 and 235. There was a relative error in isotope ratios of 0.21-0.25%, and RSD of 1.54-1.81%. This approach overcomes the extensive sample preparation that can lead to U losses, and also offers fast analysis speed (~5 minutes per sample). Results were comparable between EESI-MS and ICP-QMS (Agilent 7500ce) over a U concentration range of \sim 2.6 µg L⁻¹ to \sim 3.1 mg L⁻¹. Results from EESI-MS returned a RSD of 1.25-3.26%, compared to 0.71-1.46% for ICP-MS. The EESI used in this study was homemade, and it was suggested that improvements in RSD would be realized using a commercial EESI source.

Uranium-236 was measured in soil samples in the vicinity of Chernobyl by ICP-SFMS (Thermo Element), ICP-QMS (Perkin Elmer Elan 6000) and ICP-CC-MS (Micromass Platform ICP), with high precision isotope ratio measurements were performed using a MC-ICP-MS (Nu Instruments).256 Multiple sample introduction systems (micro-concentric Micromist nebulizer, Q-DIHEN or MCN equipped with an Aridus desolvating sample introduction (CETAC)) were investigated. An abundance sensitivity for $^{236}\text{U}/^{238}\text{U}$ of 5 \times 10⁻⁶, 3 \times 10⁻⁷, 6×10^{-7} and 3×10^{-7} was measured for ICP-SFMS, MC-ICP-MS, ICP-QMS and ICP-CC-MS, respectively. The precision ranged from 0.28-0.34% for ²³⁶U/²³⁸U for single collector instruments, improving to up to 0.07% for MC-ICP-MS equipped with USN or MCN with an Aridus. Ultra-low ²³⁶U/²³⁸U ratios were recently measured at isotopic ratios of <10⁻⁷ by ICP-QQQ-MS (Agilent 8800).257 The two mass filters effectively removed tailing from the peak at ²³⁸U, whilst the ²³⁵U¹H interference was removed by reacting UH with O2 in the collision-reaction cell. The interference-removal capability meant that accurate isotopic ratios

could be measured without the need for spectral interference correction.

8.18 Neptunium-237

Neptunium-237 is an alpha emitting radionuclide (maximum decay energy 4.78 MeV (47.6%) with a half-life of 2.1 million years. Neptunium is present in the environment as a result of atmospheric weapons test fallout and nuclear fuel reprocessing. A review of ²³⁷Np measurement in nuclear and environmental samples was recently published.266 Samples investigated include soils, sediment, groundwater, seawater and various other environmental samples.82,267-277

Neptunium occurs in both tetra- and pentavalent states, and the variation in oxidation state depending on the reagents and conditions used makes sample preparation challenging. 278 Bulk separation from the sample matrix has been achieved by techincluding iron hydroxide precipitation²⁷² and lanthanum hydroxide precipitation, 275 followed by ion exchange or extraction chromatography. 82,268-272,275,278 Neptunium is often measured along with Pu following chemical separation, given the similar chemical behavior of the two elements in HNO₃ and HCl on anion exchange and extraction chromatography resins. 63,268-271,274-276,279 A micro-flow injection sample introduction system combined with membrane desolvation improved the instrument sensitivity, and the detection limit by concentrating the analyte into a smaller volume.276 In a separate study, Qiao et al. incorporated TEVA resin into a sequential injection (SI) system for simultaneous Np and Pu measurement in environmental samples, or Bio-RAD AG MP-1M for soil, sediment and seaweed.279

The major interference impacting ²³⁷Np detection is tailing from ²³⁸U, which must be removed by chemical separation, or by the instrument if abundance sensitivity is sufficient.276 The ²³⁷Np detection limit has been noted to increase with increasing U concentration (0.32 fg (8.3 \times 10⁻⁹ Bq) with no uranium, compared to 10 fg $(2.6 \times 10^{-7} \text{ Bq})$ with 2500 pg of U, with a negative impact on detection limit once the U concentration exceeded 30 pg).

The lack of a suitable tracer for ²³⁷Np has been identified as a limitation, 266,276 particularly for isotope dilution applications. 274,280 Neptunium-236 (154 000 year half-life) is potentially suitable,269,274 but suffers from a lack of commercial availability and production difficulties. 280,281 Alternative isotopes of neptunium (239Np and 235Np) are less well suited given their short half-lives (2.35 days and 396 days, respectively) and suffer from potential isobaric interferences from 239Pu and 235U, respectively. 63 Plutonium-242 (half-life 3.75 × 105 years) has been used as a tracer for determination of both 237Np and Pu isotopes,276 given its similar chemical behavior and commercial availability.269,270,274,275,280 An issue with this is variation in chemical fractionation between Np and Pu during analysis, which will increase uncertainty,280 and an isotope of the same element is desirable.

Neptunium-237 has frequently been determined using ICP-SFMS^{268,270-274} (Table 11). Kim et al. (2004) simultaneously measured ²³⁷Np with ²³⁹Pu and ²⁴⁰Pu in environmental samples

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Table 10 Summary of recent procedures for measurement of U isotopes

Reference	Instrument	Model	Sample matrix	Chemical separation	${ m LOD}^a, { m pg} \ { m L}^{-1} \left({ m Bq} \ { m L}^{-1} \right)$
78	QMS	Plasma Quad 2+	Environmental samples (oyster tissue and pine needles)		Pneumatic nebulizer: $5400~(6.7 \times 10^{-5})$ to $48~000~(6.0 \times 10^{-4})$ ETV: $900~(1.1 \times 10^{-5})$ to $21~000$ (2.6×10^{-4})
255 97 258	Ion trap MS QMS QMS	LTQ-XL Elan 5000	Water, U ore, soil Uranium oxide leachate Environmental samples	EESI Anion exchange TEVA resin	$30\ 000\ (3.7 \times 10^{-4})$ $100\ (1.2 \times 10^{-6})$
259 250	QMS QMS	rokogawa FMS-2000 Perkin Elmer Elan 6000 X-series II	Urine River water	Stacked TEVA and DGA resins	$4000 (5.0 \times 10^{-5})$ $^{233}U: 6.5 (2.3 \times 10^{-3})$ $^{234}U: 6.5 (1.5 \times 10^{-3})$
260	ÓMS	Agilent 7500	Urine	Ca phosphate, stacked TEVA, TRU, DGA	$^{2.3}$ U: $23.8 \left(1.9 \times 10^{-9} \right)$ 236 U: $6.5 \left(1.6 \times 10^{-5} \right)$ 238 U: $2820 \left(3.5 \times 10^{-5} \right)$
261	DRC-MS	Perkin Elmer Elan 6100 DRC	Urine	Samule dilution only	Instrument LOD: 4 (5.0×10^{-8}) Method LOD: 22 (2.2×10^{-1}) 200 (2.5×10^{-6})
263 263 49	SFMS SFMS	Element Element	Standard solutions Standard solutions Soil samples 4–20 km north and west of Chernohyl	Anion exchange	Soil: $40 (5.0 \times 10^{-7})$ Water: $0.2 (5.5 \times 10^{-9})$
259	SFMS	Element	Urine		Instrument LOD: $60 (7.5 \times 10^{-7})$ Method LOD: $<3000 (3.7 \times 10^{-5})$
264	SFMS CC-MC-ICP-MS MC-ICP-MS	Plasma Trace 2 Micromass Isoprobe VG Elemental P54	Urine	Phosphate precipitation $2 \times TRU$ UTEVA resin	$5 (6.2 \times 10^{-8})$ 1 (1.2×10^{-8}) 200 (2.5×10^{-6})
265	SFMS	Element XR	Swab samples	Stacked ion exchange and extraction chromatography	
265 266	SFMS SFMS	Element 2 Element 2	Urine Urine	Ca and Mg co-precipitation UTEVA resin	235 U: 0.8 (6.4×10^{-8}) 236 U: 0.05 (1.2×10^{-7}) 238 U: 100 (1.2×10^{-9})

Table 11 Summary of recent procedures for measurement of ²³⁷Np

Reference	Instrument	Model	Matrix	Chemical separation	LOD, pg L^{-1} (Bq L^{-1})
283	QMS	Perkin Elmer Elan 5000	Natural groundwater	Capillary electrophoresis	$50\ 000\ 000\ (1.3\times 10^3)$
260	QMS	Agilent 7500	Urine	Ca phosphate, stacked TEVA, TRU, DGA resin	
275	QMS	Agilent 7500	Large soil samples		8700 (0.2)
63	QMS	X-series II	Environmental samples	Hydroxide precipitate, sequential injection (TEVA resin)	
279	QMS	X-series II	Soil, sediment, seaweed	Iron hydroxide precipitation, sequential injection (anion exchange)	$1.5\ (3.9\times 10^{-5})$
267	QMS	VG Elemental PlasmaQuad PQ2+	Environmental samples	TOA extraction chromatography	$460~(1.2\times 10^{-2})$
268	SFMS	Finnigan MAT Element	Environmental samples	TRU resin	
82	SFMS	Not given	Irish sea sediments	Glow discharge	80 000 (2.1)
269	SFMS	Finnigan-MAT Element	Environmental samples	La(OH) ₃ co-precipitation, TEVA resin	$0.5(1.3 imes10^{-5})$
270	SFMS	Micromass PlasmaTrace 2	Environmental samples	Automated sequential injection (TEVA resin)	$2.5~(6.5\times 10^{-5})$
272	SFMS	Finnigan MAT Element 2	Soil	TEVA resin	$0.2-0.4~(5.2\times10^{-6}~{ m to}~1.0\times10^{-5})$
284	SFMS	Element 2	Soils and sediments from river Yenisei	TEVA resin	

by ICP-SF-MS (Micromass Plasma Trace 2) equipped with an Aridus desolvating sample introduction system and T1-H microconcentric nebulizer. Chemical separation of Np and Pu was achieved using an automated SI system using TEVA extraction chromatography resin, with a $^{237}\rm{Np}$ detection limit of 2.5 pg L $^{-1}$ (6.5 \times 10 $^{-5}$ Bq L $^{-1}$). In a separate study, an Agilent 7500 QMS equipped with an Octopole Reaction System was operated in 'no gas' mode for Np measurement. However, there was evidence that Np would behave similarly to Pu with CO $_2$ as the reactive gas, which could be used as a basis for separation of Np and Pu from U. Rescaled that applying chemical separation prior to ICP-SFMS measurement could lower the detection limit.

Sediments have been measured by glow discharge mass spectrometry. Set The instrument was operated at a mass resolution of around 6000, resulting in high transmission (>75%) and removal of tailing interferences, as well as reduced sample preparation time compared to chemical separation. Irish Sea sediment samples were measured after being compacted into pellets. Neptunium was determined in groundwater at the Nevada National Security Site (NNSS) by ICP-SFMS (NuPlasma) at concentrations ranging from $^{\rm 4}\times 10^{-4}$ to 2.6 mBq L $^{\rm -1}$ (0.015–99.7 pg L $^{\rm -1}$) with all values below the US Environment Protection Agency drinking limits of 560 mBq L $^{\rm -1}$. The outcome was used to evaluate retardation of Np relative to other radionuclides in NNSS groundwater.

8.19 Plutonium isotopes

Plutonium is present in the environment as a result of nuclear weapons tests, reactor accidents, and discharges from reprocessing facilities, and is arguably the most frequently studied transuranic element. Isotopes of particular interest with regards to ICP-MS measurement are 238 Pu ($t_{1/2}=87.7$ years), 239 Pu ($t_{1/2}=24$ 110 years) and 240 Pu ($t_{1/2}=6561$ years) (Table 13). Several reviews on the determination of plutonium isotopes are given elsewhere. 95,120,285

Isotope ratio measurements by ICP-MS (most commonly ²⁴⁰Pu/²³⁹Pu) have been effectively used to determine the source of nuclear contamination²⁸⁶ (Table 12). This is a significant advance over traditional alpha spectrometry detection, which is unable to resolve the similar alpha energies of ²³⁹Pu and ²⁴⁰Pu (5.24 and 5.25 MeV, respectively). Measurement of Pu isotopes by alpha spectrometry also requires extensive chemical separation and counting times on the order of days to weeks.

Measurement of 239 Pu is affected by tailing from 238 U, with decontamination factors of 10^8 or higher required depending on the sample matrix. 285,286 A detection limit of 1.0 fg $(2.3 \times 10^{-6}$ Bq) was achieved for 239 Pu at U concentrations from 0–30 pg using ICP-SFMS (Thermo Element XR), compared to 8.6 fg $(2.0 \times 10^{-5}$ Bq) when U concentration was increased to 2500 pg. 276 There is an additional polyatomic interference from 238 U 1 H, which can be removed by U/Pu separation, or by reducing hydride formation using a desolvating sample introduction system. Plutonium-238 measurement is challenging owing to low concentrations, and an isobaric interference from 238 U, whilst 241 Pu can be affected by isobaric 241 Am. Chemical separation of Pu prior to sample introduction is generally a 2

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Table 12 240 Pu : 239 Pu atom ratios for different sources 286,299

Source	²⁴⁰ Pu/ ²³⁹ Pu
Integrated weapon test fallout	0.18
Weapon production	0.01-0.07
Chernobyl accident	0.40
MAGNOX reactor	0.23^{a}
Pressurized heavy water reactor	0.41^{a}
Advanced gas-cooled reactor (AGR)	0.57^{a}
Pressure tube boiling water reactor	0.67^{a}
Boiling water reactor (BWR)	0.40^{a}
Pressurized water reactor (PWR)	0.43^{a}
Fukushima prefecture coast	0.19-0.26
sediments	

^a After fuel burn up (the amount of energy extracted from the primary fuel source).48,287

stage process-bulk matrix co-precipitation, followed by actinide pre-concentration and separation using extraction chromatography. 63,275,282,288-290 More rapid FI or SI techniques have been effectively applied to urine,291 soils,275,289 and environmental samples. 63 A number of Pu isotopes including 236 Pu, 242 Pu and ²⁴⁴Pu have been effectively used as tracers, ⁶³ with isotope dilution a common technique that can also address issues with plasma instability and ion intensity drift.95

Carbon dioxide has been used as a reactive gas for U separation from Pu for a ²³⁵U target using CC-ICP-MS (GV Isoprobe MC-ICP-MS).²⁸² At CO₂ gas flow rates >0.5 mL min⁻¹, U is present as ${\rm UO}^+(\sim\!95\%)$ and ${\rm UO_2}^+(\sim\!5\%)$, compared to Pu, which was present as Pu^{+} (40%), PuO^{+} (~60%), and PuO_{2}^{+} (~2%). A recent study used lab-on-valve separation of Pu using bead injection extraction chromatography micro-flow system was coupled to ICP-QMS (Thermo X-series II) equipped with Xs skimmer cone and Burgener nebulizer.292 The design enabled processing of large urine volumes (1 L), Pu chemical yields >90%, and completion of the analytical procedure in less than 3 hours, compared to 1-2 days for manual processing. A detection limit of 1.0–1.5 pg L^{-1} (equivalent to 2.3×10^{-3} to 3.4×10^{-3} Bq L^{-1} for ^{239}Pu and 1.5 \times 10^{-4} to 2.2 \times 10^{-4} Bq L^{-1} for ²⁴²Pu) was achieved for ²³⁹Pu and ²⁴²Pu, however, potential improvements to the procedure were suggested, including overcoming the limited flow rate (<1 mL min⁻¹) due to backpressure build up and settling of beads within the system, which could lead to leakage and/or malfunctioning for longterm operations.

Plutonium isotope ratios (239Pu/240Pu) were determined from alpha planchettes originally prepared from samples from the Mayak nuclear facility, Russia.293 The aim was to show that direct analysis of samples was possible without the timeconsuming dissolution and chemical separation associated with solution ICP-MS. A potential advantage is that this approach can be applied to planchette samples round the world, to generate a large amount of new data from existing samples. Samples had previously been separated from the sample matrix, and were measured by ICP-QMS (Agilent 7700X). If the 239+240Pu activity on the planchette was <1 Bq, the

statistical variation in isotope ratios was too high and reliable determination of ²⁴⁰Pu was not possible. It was noted that the likelihood of distinguishing between sources of Pu could be improved by chemical separation to remove U from the sample prior to measurement.

Varga et al. investigated rapid and direct measurement of U and Pu, 235U/239Pu and 236U/240Pu chronometers without chemical separation using a Thermo Element 2 ICP-SFMS equipped in a glovebox.294 The instrument was run in low resolution mode with a low-flow micro concentric nebulizer, and a quartz glass spray chamber. A second method incorporating extraction chromatography prior to measurement was also tested, with multiple U/Pu ratios measured. Results from two Pu CRM's were in good agreement with archive purification

Bu et al. measured Pu isotopes in lichen, kelp, moss and horse mussel collected from Alaska by ICP-SF-MS (Thermo Element XR).295 The instrument was operated in low resolution, equipped with an Apex (Elemental Scientific) sample introduction system and PFA nebulizer. A 242Pu tracer was used, and a correction factor was applied to correct for uranium hydride interference, however this was expected to be low as chemical separation was applied prior to sample introduction. Mass discrimination was determined from measurement of 235 U/238 U in a natural U solution, with the overall procedure validated by measuring $^{239+240}$ Pu in a CRM. A measured value of 5.18 \pm $0.10 \; \mathrm{Bq \; kg^{-1}}$ was in good agreement with the certified value of 5.30 ± 0.16 Bg kg⁻¹. The total ²³⁹⁺²⁴⁰Pu content in sample measured ranged from 3.8 to 573 Bq kg⁻¹ dry weight. The difference in isotope ratio values measured showed isotopically heavier 240Pu/239Pu values in marine samples compared to terrestrial samples, accounted for as being a large input of Pu into the Pacific Ocean, most likely Marshall Islands high yield

A comparison of liquid scintillation and ICP-SFMS for ²⁴¹Pu measurement in nuclear waste slurries was reported by Jäggi et al. (2012).²⁹⁶ ICP-MS measurements were performed using a double focusing ICP-SFMS (Thermo Element 2). Samples were introduced via an Apex nebulizing system connected to an ACM desolvation system and a self-aspirating PFA-ST nebulizer. Correction for tailing and hydride formation were performed following measurement of a 238U standard. Correction factors for mass 239, 240 and 241 arising from 238U were typically 9×10^{-6} , 1×10^{-6} and 3×10^{-7} . A detection limit of 0.27 Bq g⁻¹ (117.6 pg g^{-1}) for a 0.1 g slurry sample was achieved.

Isotope ratios were measured from multiple environmental samples collected in Finland by ICP-SFMS (Thermo Element 2) operating in low resolution, fitted with a coaxial nebulizer and cyclonic spray chamber.297 Instrument performance was carried out using a certified reference material containing natural U, and quality of 240Pu/239Pu measurements was validated from a CRM. For surface air samples collected in 1963, values mostly fell in the range 0.15-0.25, indicative of global weapons fallout. Results from environmental samples showed a wider range in values from 0.13 \pm 0.1 to 0.53 \pm 0.03, indicative of both global fallout and the Chernobyl accident. As well as digestion and

Summary of recent procedures for measurement of Pu isotopes Table 13

2.2 SPMS Plasma Trace 2 Environmental samples Stream and TEXA resin Stream and TEXA resin 3 (6.5 × 10 ⁻³) Eg L ⁻¹ 2.9 QMS Perfection filter Ella 5000 Nation gooundental samples Capillary electrophoresis 500 000 000 (1.2 × 10 ⁻³) 2.9 QMS Perfection filter Ella 6000 Environmental samples Town injection (TEVA resin) 4 (6.5 × 10 ⁻³) 4 (6.5 × 10 ⁻³) 2.0 QMS Applient 7500 Invironmental samples Town injection (TEVA resin) 4 (6.5 × 10 ⁻³) 4 (6.5 × 10 ⁻³) 2.5 QMS Applient 7500 Invironmental samples Town injection (TEVA resin) 4 (6.5 × 10 ⁻³) 4 (6.5 × 10 ⁻³) 2.5 QMS Applient 7500 Lyrine mental samples Town injection (TEVA resin) 4 (6.5 × 10 ⁻³) 4 (6.5 × 10 ⁻³) 2.5 QMS Applient 7500 Univer water Soil samples Town injection (TEVA resin) 4 (6.5 × 10 ⁻³) 4 (6.5 × 10 ⁻³) 2.5 QMS Averies II River water River water Axion exchange 4 (6.5 × 10 ⁻³) 4 (6.5 × 10 ⁻³) 2.5 QMS		HISH HIHEHL	Model	Matrix	Chemical separation	LOD, pg L^{-1} (Bq L^{-1}) ^a
QMS Petrichia Elmare Elan 5000 Natural groundwater Capillary electrophoresis QMS Potario Elmer Elan 5000 Environmental samples Flow injection (UTEVA resin) QMS Agrient 7500 Urine TRVA resin TRVA resin QMS Agrient 7500 Urine TRVA resin Ca phosphate precipitation, stacked QMS Agrient 7500 Urine TRVA resin Ca phosphate precipitation, stacked QMS Agrient 7500 Urine TRVA resin Ca phosphate precipitation, stacked QMS X-series II Environmental samples, sequential Hydroxide precipitation, stacked QMS X-series II Rovinomental samples, sequential Hydroxide precipitation, stacked QMS X-series II Rover water Stacked TRVA and DGA resin QMS Elan 5000 Uraniuno oxide leachate Capliary electrophoresis QMS Frinkiam MAT Element During and stacked TRVA resin SFMS Flement 2 Uraniunomental samples Anion exchange SFMS Asion SC Environmental samples Caphosphate precipitat	42	SFMS	Plasma Trace 2	Environmental samples	Sr resin and TEVA resin	$3~(6.5 imes 10^{-5}~{ m Bq~L^{-1}})$
QMS PlasmaQuad2+ Pol Savell-s- Soll Environmental samples Poly Savell-s- Soll Flow injection (UTFVA resin) Flow injection (UTFVA resin) QMS Varian 810 MS Environmental samples Flow injection (UTFVA resin) QMS Agllent 7500 Environmental samples, sequential TFVA, TRU, DGA resins QMS X-series II Soll, sediment, seaweed, sequential Hydroxide precipitation, stacked QMS X-series II Soll, sediment, seaweed, sequential Hydroxide precipitation, stacked QMS X-series II Soll, sediment, seaweed, sequential Injection (TFVA resin) QMS X-series II River water Stacked TFVA and DGA resin QMS X-series II River water Stacked TFVA and DGA resin QMS Prelimigan MAT Element Stacked TFVA and DGA resin QMS Planigan MAT Element Soll samples Anion exchange SFMS Element Chernolyl Anion exchange SFMS Micromass Plasmafrace 2 Environmental samples Anion exchange SFMS Thermo Element Urine None SFMS Micromass Pl	283	QMS	Perkin Elmer Elan 5000	Natural groundwater	Capillary electrophoresis	$50000000(1.2 imes10^5)$
QMS PQ Excell-Is Soil Profession Profession QMS Varian 810 MS Environmental samples Flow injection (UTEVA resin) QMS Agilent 7500 Urine Environmental samples TEVA resin QMS X-series II Environmental samples TEVA, TRU, DGA resins QMS X-series II Soil, sediment, seaweed, sequential Hydroxide precipitation, sequential QMS X-series II Soil, sediment, seaweed, sequential Iron injection QMS X-series II Rivier water Adjuiction, St-based QMS Perkins Elmer Elan 5000 Uranium oxide leachate Anion exchange QMS Perkins Elmer Elan 5000 Uranium oxide leachate Anion exchange CC-QMS GV Isoprobe U-235 target TRU resin SFMS Element Soil samples TRU resin SFMS Benent Urine Anion exchange SFMS Micromass Plasmafrace 2 Environmental samples Anion exchange SFMS Thermo Element Urine Ca phosphate precipitation and TEVA <td>300</td> <td>QMS</td> <td>PlasmaQuad2+</td> <td>Environmental samples</td> <td></td> <td>$^{239/240} \mathrm{Pu}$: $10 \left(2.3 imes 10^{-2} / 8.4 imes 10^{-2} ight)$</td>	300	QMS	PlasmaQuad2+	Environmental samples		$^{239/240} \mathrm{Pu}$: $10 \left(2.3 imes 10^{-2} / 8.4 imes 10^{-2} ight)$
QMS Varian 810 MS Previoumental samples Flow injection (TEVA resin) QMS Agilent 7500 Environmental samples TEVA resin QMS Agilent 7500 Large soil samples TEVA, TRU, DGA resins QMS Agilent 7500 Large soil samples, sequential Hydroxide precipitation, stacked QMS X-series II Environmental samples, sequential Hydroxide precipitation, sequential QMS X-series II River water Stacked TEVA, TRU, DGA resins QMS X-series II River water Stacked TEVA resin) QMS Perkin Elmer Elan 5000 Uranium oxide leachate Capillary electrophoresis CC-QMS Ferkin Elmer Elan 5000 Uranium oxide leachate Capillary electrophoresis CC-QMS Ferkin Elmer Elan 5000 Uranium oxide leachate Capillary electrophoresis SFMS Filement Soil samples 4-20 km north and west of Anion exchange SFMS Micromass Plasmafrace 2 Environmental samples Anion exchange SFMS Thermo Element Urine Caphosphate precipitation and TEVA SFMS	289	QMS	PQ Excell-s	Soil	Flow injection (UTEVA resin)	$4.3~(9.9 imes 10^{-3})$
QMS Perkin Elmer Elan 6000 Environmental samples TRVA resin QMS Agilent 7500 Urine TRVA, TRU, Doda resins QMS X-series II Environmental samples, sequential Hydroxide precipitation, sequential injection QMS X-series II Soil, sediment, seaweed, sequential injection Hydroxide precipitation, sequential injection QMS X-series II River water Stacked TRVA and DGA resin QMS X-series II River water Stacked TRVA and DGA resin QMS Perkin Elmer Elan 5000 Urranium oxide leachate Anion exchange QMS Perkin Elmer Elan 5000 Urranium oxide leachate Capiliary electrophoresis QMS Perkin Elmer Elan 5000 Urranium oxide leachate Capiliary electrophoresis SPMS Finnipan MAT Element Soil samples TRV resin SFMS Micromass PlasmaTrace 2 Environmental samples Automated sequential injection (TEVA SFMS Thermo Element Urrine Environmental samples Automated sequential injection (TEVA SFMS Element 2 Environmental samples Cap phospha	301	QMS	Varian 810 MS	Environmental samples	Flow injection (TEVA resin)	$3~(6.9 imes 10^{-3})$
QMS Agilent 7500 Urine Ca phosphate precipitation, stacked QMS X-series II Ingestoil samples Hydroxide precipitation, sequential injection QMS X-series II Soil, sediment, seaweed, sequential injection Hydroxide precipitation, sequential injection QMS X-series II River water Stacked TEVA resin QMS Perkin Ellen 5000 Uranium oxide leachate Anio exchange QMS Perkin Ellen Ellan 5000 Uranium oxide leachate Anio exchange QMS Perkin Ellen Ellan 5000 Uranium oxide leachate Capillary electrophorics is CC-QMS GV Isoprobe U-2.35 target Capillary electrophores is CC-QMS Finnigan MAT Element Environmental samples TRU resin SFMS Element 2 Urine Chine SFMS Micromass PlasmaTrace 2 Environmental samples Automated sequential injection (TEVA and TRU resin) SFMS Element 2 Urine Capination, UTEVA and TRU resin SFMS Element 2 Environmental samples Car ₂ precipitation, UTEVA and TRU resin SFMS	302	QMS	Perkin Elmer Elan 6000	Environmental samples	TEVA resin	
QMS Agilent 7500 Large soil samples Hydroxide precipitation, sequential injection Hydroxide precipitation, sequential injection QMS X-series II Soil, sediment, seaweed, sequential injection Injection Soil, sediment, seaweed, sequential injection (TEVA resin) QMS X-series II River water Action Action by droxide precipitation, SI-based injection QMS Perkin Elmer Elan 5000 Uranium oxide leachate Anion exchange Action oxchange QMS Perkin Elmer Elan 5000 Uranium oxide leachate Anion exchange Anion exchange CC-QMS GV Isoprobe U-235 target TRU resin TRU resin SFMS Element Chain samples TRU resin Anion exchange SFMS Micromass PlasmaTrace 2 Environmental samples Anion exchange SFMS Axiom SC Environmental samples Action exchange SFMS Axiom SC Environmental samples Caphosphate precipitation, UTEVA and TRU resin SFMS Element 2 Drine Car ₂ precipitation, UTEVA and TRU resin SFMS Element 2 Marine sediments	260	ÓМS	Agilent 7500	Urine	Ca phosphate precipitation, stacked TEVA, TRU, DGA resins	
QMS X-series II Environmental samples, sequential Hydroxide precipitation, sequential injection QMS X-series II Soil, sediment, seaweed, sequential injection Injection (TEVA resin) QMS X-series II River water AG1 QMS Perkin Elmer Elan 5000 Uranium oxide leachate Anion exchange QMS Perkin Elmer Elan 5000 Uranium oxide leachate Anion exchange CC-QMS GV Isoprobe U-235 target Anion exchange CC-QMS Finnigan MAT Element Brivampete 4-20 km north and west of Chemobyl Anion exchange SFMS Element 2 Urine None SFMS Axiom SC Environmental samples Ponne SFMS Axiom SC Environmental samples Ca phosphate precipitation and TEVA SFMS Hiement 2 Brivinonmental samples Ca phosphate precipitation and TEVA SFMS Hiement 2 Marine sediments Calcination, UTEVA and TRU resin SFMS Micromass PlasmaTrace 2 Marine sediments Calcination, anion exchange SFMS Hiement 2 Marine sedi	275	QMS	Agilent 7500	Large soil samples		700 (1.6)
QMS X-series II injection injection injection (TEVA resin) QMS X-series II River water AG1 QMS X-series II River water AG1 QMS Elan 5000 Uranium oxide leachate Anion exchange QMS Perkin Elnner Ellan 5000 Uranium oxide leachate Capillary electrophoresis CC-QMS GV Isoprobe U-2.35 target TRU resin SFMS Finnigan MAT Element Soil samples 4-20 km north and west of Chemobyl Anion exchange SFMS Element 2 Urine None SFMS Axiom SC Environmental samples Automated sequential injection (TEVA resin) SFMS Axiom SC Environmental samples Automated sequential injection (TEVA resin) SFMS Thermo Element Urine Ca phosphate precipitation, UTEVA and TRU resin SFMS Element 2 Marine sediments Capicination, UTEVA and TRU resin SFMS Element 2 Marine sediments Capicination, UTEVA and TRU resin	63	QMS	X-series II	Environmental samples, sequential	Hydroxide precipitation, sequential	
QMS X-series II Soil, sediment, seaweed, sequential Iron hydroxide precipitation, SI-based injection QMS X-series II Rever water Stacked TEVA and DGA resin QMS Perkin Elmer Elan 5000 Uranium oxide leachate Anion exchange QMS Perkin Elmer Elan 5000 Uranium oxide leachate Anion exchange CC-QMS GV Isoprobe U-235 arguentargroundwater Capillary electrophoresis SFMS Finnigan MAT Element Element Capillary electrophoresis SFMS Element 2 Soil samples 4-20 km north and west of Chemobyl Anion exchange SFMS Micromass PlasmaTrace 2 Environmental samples Automated sequential injection (TEVA and TRU resin) SFMS Axiom SC Environmental samples resin) SFMS Micromass PlasmaTrace 2 Environmental samples Ca phosphate precipitation and TEVA resin SFMS Micromass PlasmaTrace 2 Marine sediments Caphosphate precipitation, UTEVA and TRU resin SFMS Micromass PlasmaTrace 2 Marine sediments Caphosphate precipitation, UTEVA and TRU resin				injection	injection (TEVA resin)	
QMS X-series II River water QMS Elan 5000 Uranium oxide leachate Anion exchange QMS Perkin Elmer Elan 5000 Uranium oxide leachate Anion exchange CC-QMS GV Isoprobe U-235 target TRU resin SFMS Element Element Element SFMS Element 2 Urine None SFMS Micromass PlasmaTrace 2 Environmental samples Automated sequential injection (TEVA resin) SFMS Axiom SC Environmental samples Resin) SFMS Thermo Element Urine Ca phosphate precipitation and TEVA resin SFMS Element 2 Environmental samples Ca phosphate precipitation and TEVA resin SFMS Micromass PlasmaTrace 2 Marine sediments Carlination, urine exchange chromatography SFMS Element 2 Environmental samples Calcination, anion exchange chromatography SFMS Element 2 Barine sediments Calcination, anion exchange chromatography	279	QMS	X-series II	Soil, sediment, seaweed, sequential	Iron hydroxide precipitation, SI-based	$1.5~(3.5\times 10^{-3})$
QMS Elan 5000 Uranium oxide leachate Anion exchange QMS Perkin Elmer Elan 5000 Uranium oxide leachate Anion exchange CC-QMS GV Isoprobe U-235 target Capillary electrophoresis SFMS Finnigan MAT Element Soil samples 4-20 km north and west of Chemobyl Anion exchange SFMS Element 2 Urine None SFMS Axiom SC Environmental samples Automated sequential injection (TEVA resin) SFMS Axiom SC Environmental samples Automated sequential injection (TEVA resin) SFMS Thermo Element Urine Ca phosphate precipitation and TEVA resin SFMS Hicromass PlasmaTrace 2 Marine sediments Calcination, anion exchange chromatography SFMS Element 2 Marine sediments from river Yenisei TEVA resin	050	OMC	X-comioc II	mjecuon Diver water	AGI Stocked TRWA and DGA resin	239/240/242 D ₁₁ . 0 3 (6.3 × 10 ⁻⁴ 2.3 × 10 ⁻³
QMSElan 5000Uranium oxide leachateAnion exchangeQMSPerkin Elmer Elan 5000Natural groundwaterCapillary electrophoresisCC-QMSGV IsoprobeU-235 targetSFMSFinnigan MAT ElementElement annoth and west of ChernobylTRU resinSFMSElement 2ChernobylNoneSFMSMicromass PlasmaTrace 2Environmental samplesNoneSFMSAxiom SCEnvironmental samplesResinSFMSThermo ElementUrineCa phosphate precipitation and TEVA resinSFMSMicromass PlasmaTrace 2Marine sedimentsCarb, precipitation, UTEVA and TRU resinSFMSMicromass PlasmaTrace 2Marine sedimentsCalcination, anion exchange chromatographySFMSElement 2Marine sedimentsCalcination, anion exchange chromatography						$4.0 imes 10^{-5}$
QMSPerkin Elmer Elan 5000Natural groundwaterCapillary electrophoresisCC-QMSGV IsoprobeU-235 targetTRU resinSFMSFinnigan MAT ElementElement 2TRU resinSFMSElement 2UrineNoneSFMSAxiom SCEnvironmental samplesNoneSFMSAxiom SCEnvironmental samplesAutomated sequential injection (TEVA resin)SFMSThermo ElementUrineAutomated sequential injection (TEVA resin)SFMSThermo ElementUrineCa phosphate precipitation and TEVA resinSFMSElement 2Marine sedimentsCalcination, UTEVA and TRU resinSFMSElement 2Marine sedimentsCalcination, anion exchange chromatographySFMSElement 2Marine sedimentsCalcination, anion exchange chromatography	97	ÓMS	Elan 5000	Uranium oxide leachate	Anion exchange	30 000 (68)
CC-QMSGV Isoprobe SFMSU-235 target Element SFMSU-235 target Element Chernobyl Micromass PlasmaTrace 2U-235 target Solls and sediments from river YeniseiTRU resin Anion exchange Anion exchange Anion exchange Chernobyl NoneTRU resin Anion exchange Anion exchange Anion exchange Automated sequential injection (TEVA resin)SFMSAxiom SCEnvironmental samplesCa phosphate precipitation and TEVA resinSFMSElement 2Environmental samplesCa phosphate precipitation and TEVA resinSFMSElement 2Marine sedimentsCalcination, anion exchange chromatographySFMSElement 2Soils and sediments from river YeniseiTEVA resin	283	ÓMS	Perkin Elmer Elan 5000		Capillary electrophoresis	$50\ 000\ 000\ (1.2 imes 10^5)$
SFMSFinnigan MAT ElementEnvironmental samplesTRU resinSFMSElementSoil samples 4–20 km north and west of ChernobylAnion exchangeSFMSElement 2UrineNoneSFMSMicromass PlasmaTrace 2Environmental samplesAutomated sequential injection (TEVA resin)SFMSAxiom SCEnvironmental samplesCa phosphate precipitation and TEVA resinSFMSElement 2Environmental samplesCa phosphate precipitation and TEVA resinSFMSMicromass PlasmaTrace 2Marine sedimentsCalcination, anion exchange chromatographySFMSElement 2Marine sediments from river YeniseiTEVA resin	282	CC-QMS	GV Isoprobe	U-235 target	•	,
SFMS Element 2 Orine Automated sequential injection (TEVA resin) SFMS Axiom SC Environmental samples Automated sequential injection (TEVA resin) SFMS Axiom SC Environmental samples Ca phosphate precipitation and TEVA resin SFMS Element 2 Environmental samples Cars, precipitation, UTEVA and TRU resin SFMS Element 2 Marine sediments Cars, precipitation, UTEVA and TRU resin SFMS Marine sediments Cars, precipitation, UTEVA and TRU resin SFMS Marine sediments Cars, precipitation, anion exchange chromatography SFMS Element 2 Soils and sediments from river Yenisei TEVA resin	268	SFMS	Finnigan MAT Element	Environmental samples	TRU resin	
SFMS Element 2 Urine None SFMS Axiom SC Environmental samples Automated sequential injection (TEVA resin) SFMS Axiom SC Environmental samples Ca phosphate precipitation and TEVA resin SFMS Element 2 Environmental samples Calcination, UTEVA and TRU resin SFMS Micromass PlasmaTrace 2 Marine sediments Calcination, anion exchange chromatography SFMS Element 2 Marine sediments Calcination, anion exchange chromatography SFMS Element 2 Soils and sediments from river Yenisei TEVA resin	49	SFMS	Element	Soil samples 4–20 km north and west of	Anion exchange	
SFMS Element 2 Urine None SFMS Micromass PlasmaTrace 2 Environmental samples Automated sequential injection (TEVA resin) SFMS Axiom SC Environmental samples Ca phosphate precipitation and TEVA resin SFMS Element 2 Environmental samples Calcination, UTEVA and TRU resin SFMS Marine sediments Calcination, anion exchange chromatography SFMS Element 2 Marine sediments SFMS Soils and sediments from river Yenisei TEVA resin				Chernobyl		ć
SFMS Micromass PlasmaTrace 2 Environmental samples Automated sequential injection (TEVA resin) SFMS Axiom SC Environmental samples Ca phosphate precipitation and TEVA resin SFMS Element 2 Environmental samples CaF2 precipitation, UTEVA and TRU resin SFMS Micromass PlasmaTrace 2 Marine sediments Calcination, anion exchange chromatography SFMS Element 2 Soils and sediments from river Yenisei TEVA resin	291	SFMS	Element 2	Urine	None	$4.7~(1.1 imes10^{-2})$
SFMS Axiom SC Environmental samples Ca phosphate precipitation and TEVA resin SFMS Thermo Element 2 Environmental samples CaP2, precipitation, UTEVA and TRU resin SFMS Micromass PlasmaTrace 2 Marine sediments Calcination, anion exchange chromatography SFMS Element 2 Soils and sediments from river Yenisei TEVA resin	270	SFMS	Micromass PlasmaTrace 2	Environmental samples	Automated sequential injection (TEVA resin)	$^{2.1}\left(4.8\times10^{-3} ight)^{240} \mathrm{Pu:}~0.42~(9.7 imes10^{-4})$
SFMS Thermo Element 2 Environmental samples Ca phosphate precipitation and TEVA resin SFMS Element 2 Environmental samples CaF2 precipitation, UTEVA and TRU resin SFMS Marine sediments Calcination, anion exchange chromatography SFMS Soils and sediments from river Yenisei TEVA resin	300	SFMS	Axiom SC	Environmental samples	`	SFMS: $^{239/240}$ Pu: 0.1 (2.3 \times 10 ⁻⁴ /8.4 \times 10 ⁻⁴)
SFMS Thermo Element Urine Ca phosphate precipitation and TEVA resin SFMS Element 2 Environmental samples CaF2 precipitation, UTEVA and TRU resin SFMS Micromass PlasmaTrace 2 Marine sediments Calcination, anion exchange chromatography Chromatography SFMS Element 2 Soils and sediments from river Yenisei TEVA resin						241 Pu: 0.05 (0.2)
SFMS Element 2 Environmental samples CaF2 precipitation, UTEVA and TRU resin SFMS Micromass PlasmaTrace 2 Marine sediments Calcination, anion exchange chromatography SFMS Soils and sediments from river Yenisei TEVA resin	303	SFMS	Thermo Element	Urine	Ca phosphate precipitation and TEVA resin	$1.0 imes 10^{-3} (2.3 imes 10^{-9})$
cnromatography SFMS Element 2 Soils and sediments from river Yenisei TEVA resin	57 304	SFMS SFMS	Element 2 Micromass PlasmaTrace 2	Environmental samples Marine sediments	CaF ₂ precipitation, UTEVA and TRU resin Calcination, anion exchange	9.2 (2.1×10^{-2}) to $15 (3.5 \times 10^{-2})$ $^{240/241/242}$ Pu: $3-4 (2.5 \times 10^{-2})$ to 3.4×10^{-2}
SFMS Element 2 Soils and sediments from river Yenisei TEVA resin					cnromatograpny	10^{-7} 11.4-15.2/4.4 × 10 $^{-10}$ 5.9 × 10 $^{239/240}$ Pu: 8-9 $(1.8 \times 10^{-2}$ to 2.1 × $10^{-2}/6.7 \times 10^{-2}$ to 7.6 × 10 $^{-2}$
	284	SFMS	Element 2	Soils and sediments from river Yenisei	TEVA resin	

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chemical separation, old alpha planchettes were wet-ashed and purified by extraction chromatography prior to measurement.

Both airborne Pu and U originating from the Fukushima Daiichi NPP were identified in the atmosphere 120 km from the site through measurement of aerosol samples, with a sampling time of approximately one week.298 Following ashing and chemical separation, samples were analyzed by a combination of AMS and ICP-SFMS. Whilst the amount of environmental U and Pu increased, the dose from airborne Pu was negligible. ICP-SFMS was used for analysis of ²³⁴U/²³⁸U and ²³⁵U/²³⁸U, however AMS was advantageous for Pu measurement due to excellent suppression of hydride interferences. No Pu spike was added because of the low levels being investigated, and the potential negative impact on detection limit that adding a spike could have. Zheng et al. presented the first data on distribution of Pu isotopes in surface sediments 30 km off Fukushima.²⁸⁷ The ²⁴⁰Pu/²³⁹Pu ratios were measured using ICP-SFMS with Apex (Elemental Scientific) sample introduction. Chemical separation was performed using anion exchange chromatography, and several reference materials were used for method validation. Isotope ratio results were compared to background data in Japanese estuaries and the western North Pacific, and it was found that no Pu contamination was detected outside the 30 km zone around the plant. A more comprehensive assessment of Pu isotope measurements in seawater and sediments within the 30 km zone was recommended, to improve understanding of marine environmental behavior.

Detection limits in the attogram range in sediment and seaweed reference materials was achieved using ICP-SFMS (Element XR) equipped with the Jet interface, 56 comparable to sensitivities achievable by AMS. Despite the larger orifice of the Jet sample cone, the Jet interface did not increase the hydride formation rate when operating with a desolvating sample introduction system. Without the Jet interface, the precision of 240 Pu/ 239 Pu was 20.5% and accuracy -3.3%, compared to 5.0% and 0.83% when equipped with the Jet interface and Cetac Aridus II desolvating sample introduction system. Plutonium has also been determined in urine samples by multiple techniques-alpha spectrometry, ICP-SFMS (Thermo Element XR) and AMS.299 The minimum detectable activity for SF-ICP-MS (23 fg) was two times better than alpha spectrometry (50 fg), but inferior to AMS (0.44 fg).

Americium-241

Americium-241 is produced as an activation product, and from beta decay of ²⁴¹Pu, with a half-life of 432.2 years. Americium-241 is an alpha emitter (5.44 MeV, 12.8% yield and 5.49 MeV, 85.2% yield), and gamma emitter (59.5 keV, 36% abundance), with gamma spectrometry considered inadequate for low-level ²⁴¹Am determination. ⁶² Alpha spectrometry is the most sensitive and frequently used approach, and measurement is generally combined with chemical yield tracer 243 Am ($t_{1/2} = 7.4 \times$ 10³ years), which decays by alpha emission (5.28 MeV, 87.4% yield and 5.23 MeV, 11% yield), and gamma emission (74.7 keV, 62.8% abundance). A comprehensive review of analytical

americium determination published methods for elsewhere.62,305

Measurement of ²⁴¹Am is affected by an isobaric interference from ²⁴¹Pu, and from ²⁴⁰PuH⁺, with Pu typically present at comparable or higher concentrations than that of 241Am in materials of reactor origin. There are also multiple polyatomic interferences, and ²⁴³Am (half-life 3730 years) has been applied as an internal standard, 57,306 with 242Pu used to assess 241Pu contamination.98 Alternatively, a mixed elemental standard can be used for external calibration of the instrument. 116 Chemical separation techniques prior to sample introduction include precipitation/co-precipitation and/or liquid-liquid extraction, ion exchange chromatography and extraction chromatography. 57,306 Providing these interferences can be effectively removed, ICP-SFMS can match the detection limits of alpha spectrometry.62

Krachler et al. (2014) noted the absence of matrix-matched reference materials, leading to determination of Am in spent nuclear fuels using both high resolution ICP-OES and ICP-SFMS (Thermo Element 2) installed in a glovebox. 116 The absence of a certified reference material meant instrumental mass bias was estimated based on the difference in sensitivity between 232Th and 238U in the internal standard, and assuming the same instrument sensitivity for all actinides. The mean concentration differed by a maximum of 4% between ICP-OES and ICP-MS.

Isotope-dilution ICP-SFMS has been used for the determination of pg kg⁻¹ concentrations of ²⁴¹Am in sediments (Agarande et al., 2001).307 Measurements of 241 Am relative to a 243 Am spike were performed using a VG Elemental Axiom ICP-MS with single collector and MCN6000 desolvating microconcentric nebulizer. Plutonium-242 was also added to confirm effective separation of Pu and Am prior to measurement. Measurements were performed in low resolution mode and mass bias was corrected using the U isotopic standard IRMM-72/1. Limits of detection of 0.2 Bg kg $^{-1}$ (1.6 pg kg $^{-1}$) were reported. Americium was analyzed along with Pu in Chernobyl-contaminated samples by ICP-SFMS (Thermo Element 2) in comparison to alpha spectrometry.62 Good agreement was observed between the two techniques for Certified Reference Materials (IAEA-384 and IAEA-385). The measurement time for ICP-SFMS was several minutes, compared to count times of days by alpha spectrometry. ICP-SFMS achieved a detection limit of 13.2 μBq g⁻¹ (0.1 fg g^{-1}) for ICP-SFMS, with a precision of 0.8–3%, compared to 4 μ Bq g⁻¹ (0.03 fg g⁻¹) and a precision of 1–5% by alpha spectrometry. Mass discrimination was again determined by applying a linear correction using a natural uranium solution. The combined measurement of Pu and Am allowed calculation of date of contamination based on the 241Pu/241Am ratio, assuming release of radionuclides was over a short time rather than continuous, as ²⁴¹Am is also produced by decay of ²⁴¹Pu. In a separate study, Varga measured 241Am in two environmental reference materials (sediment and seaweed) by ICP-SFMS (Thermo Element),98 with alpha spectrometry used to validate the procedure. The removal of polyatomic interferences was tested using stable element standards of Bi, Pb, Hg and Tl, whilst isobaric ²⁴¹Pu removal was monitored using a ²⁴²Pu isotopic tracer. A detection limit of 0.86 fg g⁻¹ (0.11 mBq g⁻¹)

was achieved, which was comparable to that of alpha spectrometry 0.79 fg (0.10 mBq).

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A method for ²⁴¹Am determination in urine was developed and validated at the Centre for Disease Control and Prevention (CDC).306 Measurements were performed using ICP-SFMS (Thermo Element XR) equipped with a CETAC Aridus desolvating sample introduction system, with a detection limit of 0.22 pg L^{-1} (0.028 Bq L⁻¹). These values were in good agreement with liquid scintillation counting (LSC), and NIST CRM target values. The analytical bias from -0.3 to 1.7% for observed values compared to target values, and 2.1-3% for samples run in an internal comparison with LSC. The previous procedure using LSC returned a detection limit of 32.3 pg L^{-1} (4.2 Bq L^{-1}), higher than the CDC action level of 0.73 pg L^{-1} (0.09 Bq L^{-1}). It was noted that if U concentrations exceeded 10 μ g L⁻¹, more aggressive rinsing was required to eliminate U from solution, with analytical bias increasing to between -0.62 and -5.61%compared to NIST target values.

On-line extraction chromatographic separation of actinides, including 241 Am, from urine matrices coupled with ICP-QMS has also been evaluated (Hang *et al.*, 2004). Detection limits of 0.15 pg mL $^{-1}$ (0.02 Bq mL $^{-1}$) were reported for a 25 mL urine sample volume. Coupling of flow injection analysis with ICP-SFMS has also been reported for the analysis of environmental samples, with detection limits down to 0.6 fg being reported. Separation of actinides, including the samples of actinides and separation of actinides, including the sample of the analysis of environmental samples, with detection limits down to 0.6 fg being reported.

Wang *et al.* recently developed a method for 241 Am measurement in large (2–20 g) soil samples using ICP-SFMS (Thermo Element XR) equipped with a high efficiency nebulizer (HEN). 309 Multiple chemical separation techniques were investigated for separation of Am from soil matrix components, rare earth elements, and ICP-MS interferences (Bi, Tl, Hg, Pb, Hf and Pt). A good chemical recovery of Am (76–82%) and detection limit of 0.012 mBq g $^{-1}$ was achieved, whilst decontamination factors of 7 × 10 5 for Pu was the highest reported for 241 Am studies, enabling measurement in Fukushima sourced soils contaminated with 241 Pu.

8.21 Curium/californium

Curium isotopes are produced via successive neutron capture by Am and subsequent beta decay. The main Cm isotopes produced in nuclear reactors are 242 Cm ($t_{1/2} = 162.9$ days), 243 Cm ($t_{1/2} = 28.9$ years), 244 Cm ($t_{1/2} = 18.2$ years), 245 Cm $(t_{1/2} = 8480 \text{ years}), ^{246}\text{Cm} (t_{1/2} = 4760 \text{ years}) \text{ and } ^{248}\text{Cm}$ $(t_{1/2} = 3.48 \times 10^5 \text{ years})$. For Cf, the main isotopes are ²⁴⁹Cf $(t_{1/2} = 351 \text{ years}), {}^{250}\text{Cf}(t_{1/2} = 13.1 \text{ years}), {}^{251}\text{Cf}(t_{1/2} = 898 \text{ years})$ and 252 Cf ($t_{1/2} = 2.6$ years). To date, there are no reported applications of ICP-MS for Cm or Cf measurement in nuclear wastes. Kurosaki et al. (2014) reported the measurement of ²⁴³Cm, ²⁴⁴Cm, ²⁴⁵Cm, ²⁴⁶Cm and ²⁴⁰Pu using a Thermo Finigan Element 2 for nuclear forensics applications. 310 Gourgiotis et al. (2010) used ICP-QMS (Thermo Electron X-series) for Cm and Cf isotope measurement in transmutation studies.311 Sample introduction was via a quartz Peltier-cooled impact bead spray chamber (natural aspiration) and micro concentric nebulizer. Two peak jump routines were employed to separately measure high abundance (248Cm/246Cm) and low abundance

 $(^{245}\text{Cm}/^{246}\text{Cm},\ ^{247}\text{Cm}/^{246}\text{Cm},\ ^{249}\text{Ch},\ ^{249}\text{(Bk}+\text{Cf)}/^{251}\text{Cf},\ ^{250}\text{Cf}/^{251}\text{Cf},\$ and $^{252}\text{Cf}/^{251}\text{Cf})$ isotope ratios. High precision isotope ratio measurement was achieved using corrected peak centering (3 points per peak with the central point associated with the maximum count rate). Dead time and mass bias corrections (using sample standard bracketing) were also applied. Peak tailing and hydride interferences were corrected by measuring the $^{237/238}\text{U}$ ratio and $^{238}\text{UH}/^{238}\text{U}$: ratio for a U010 standard. Abundance sensitivities of 1.97 \pm 0.02 ppm and hydride formation of (35.7 \pm 0.1) \times 10 $^{-6}$ were reported. Chromatographic separation of ^{249}Bk and ^{249}Cf using Dionex HPLC was developed to permit separate determination of $^{249}\text{Bk}/^{248}\text{Cm}$ and $^{249}\text{Cf}/^{248}\text{Cm}$ ratios. 312

9 Conclusions

Advances in the capability and sensitivity of recent ICP-MS instruments have stimulated new interest from the nuclear sector. This stems from the potential of these instruments to routinely and swiftly measure a range of key radionuclides considered important in nuclear decommissioning programmes. ICP-MS has proven itself as a versatile technique with regards to sample introduction and instrument setup, both of which can be used to improve sensitivity and/or interference removal. For the majority of radionuclides suitable for ICP-MS measurement, interference removal is the critical aspect affecting optimised detection limits. Effective measurement must be combined with robust, effective sample digestion and chemical separation (either through wet chemistry techniques, sample introduction and/or using an instrument with collision/reaction cell capabilities).

For long-lived radionuclides, ICP-MS offers significant benefits in sample throughput compared to the traditional radiometric methods (alpha and beta counting techniques). These benefits arise from reduced measurement time per sample, and reduced preparation time with the use of on-line separation and collision/reaction cell instruments and provide significant potential economic benefits to nuclear sites. The improvements in ICP-MS means that a number of long-lived radionuclides are now measureable that are very challenging or even not measurable by radiometric techniques e.g. 93Zr, 107Pd, ¹³⁵Cs, ⁴¹Ca and ⁵⁹Ni. These radionuclides are of increasing interest to regulatory agencies concerned with long-term nuclear waste storage and disposal. Additionally, ICP-MS (particularly multi-collector instruments) offers the ability to accurately measure isotopic ratios e.g. 239Pu/240Pu and ²³⁶U/²³⁸U, enabling the determination of the source of contamination, rather than the activity concentration alone.

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References

- 1 B. C. Russell, I. W. Croudace, P. E. Warwick and J. A. Milton, Anal. Chem., 2014, 86, 8719-8726.
- 2 W. Wien, Ann. Phys., 1898, 65, 440.
- 3 J. J. Thomson, Philos. Mag., 1897, 44, 29.
- 4 A. J. Dempster, Phys. Rev., 1918, 11, 316.
- 5 F. W. Aston, Philos. Mag., 1919, 38, 709.
- 6 F. W. Aston, Philos. Mag., 1920, 39, 449.
- 7 F. W. Aston, Proc. R. Soc. A, 1927, 115, 487.
- 8 W. Bartky and A. J. Dempster, *Phys. Rev.*, 1929, 33, 1019.
- 9 J. Mattauch and R. Hertzog, Z. Phys., 1934, 89, 786.
- 10 K. T. Bainbridge and E. B. Jordan, Phys. Rev., 1936, 50, 282.
- 11 A. O. Nier, Rev. Sci. Instrum., 1940, 11, 212.
- 12 A. O. Nier, Rev. Sci. Instrum., 1947, 18, 39.
- 13 J. A. Hipple, J. Appl. Phys., 1942, 13, 551.
- 14 H. Ewald and H. Hintenberger, Methoden und Anwendungen der Massenspektroskopie, Verlag Chemie GmbH, Weinheim, Germany, 1953.
- 15 S. Penner, Rev. Sci. Instrum., 1961, 32, 150.
- 16 H. Ewald, E. Konecny, H. Opower and H. Rösler, Z. Naturforsch., C: Biochem., Biophys., Biol., Virol., 1964, 19, 194.
- 17 S. Fukumoto, T. Matsuo and H. Matsuda, J. Phys. Soc. Jpn., 1968, 25, 946.
- 18 J. Frantzen, Jahrbuch der Wittheit zu Bremen 2006/2007, Hauschild Verlag, Bremen, Germany, 2008, vol. 181, ISBN 978-3-89757-529-5.
- 19 A History of European Mass Spectrometry, ed. K. R. Jennings, IM Publications LLP, Chichester, UK, 2012, ISBN 978-1-906715-04-5, pp. 285.
- 20 K. S. Sharma, Int. J. Mass Spectrom., 2013, 349-350, 3-8.
- 21 G. P. Russ and J. M. Bazan, Spectrochim. Acta, Part B, 1987, 42, 49-62.
- 22 D. W. Boomer and M. J. Powell, Anal. Chem., 1987, 59, 2810-2813.
- 23 R. M. Brown, S. E. Long and C. J. Pickford, Sci. Total Environ., 1988, 70, 265-274.
- 24 G. L. Beck and O. T. Farmer, J. Anal. At. Spectrom., 1988, 3, 771-773.
- 25 C. K. Kim, A. Takaku, M. Yamamoto, H. Kawamura, K. Shiraishi, Y. Igarashi, S. Igarashi, H. Takayama and N. Ikeda, J. Radioanal. Nucl. Chem., 1989, 132(1), 131–137.
- 26 A. S. Hursthouse, M. S. Baxter, K. McKay and F. R. Livens, J. Radioanal. Nucl. Chem., 1992, 157(2), 281-294.
- 27 C. K. Kim, R. Seki and S. Morita, J. Anal. At. Spectrom., 1991, 6, 205-209.
- 28 J. S. Crain and B. L. Mikesell, Appl. Spectrosc., 1992, 46(10), 1498-1502.
- 29 C. Riglet, O. Provitina, J. L. Dautheribes and D. Revy, J. Anal. At. Spectrom., 1992, 7, 923-927.
- 30 M. R. Smith, E. J. Wyse and D. W. Koppenaal, J. Radioanal. Nucl. Chem., 1992, 160(2), 341-354.
- 31 R. J. Rosenberg, J. Radioanal. Nucl. Chem., 1993, 171(2), 465-482.
- 32 J. S. Crain, J. S. Yaeger, F. P. Smith, J. A. Alvarado, L. L. Smith, J. T. Kiely and D. G. Graczyk, Argonne

- NIST, National Laboratory **CIRMS** Meeting, Gaithersberg, 28-30 Nov 1995.
- 33 J. S. Crain and J. Alvarado, J. Anal. At. Spectrom., 1994, 9, 1223-1227.
- 34 E. J. Wyse and D. R. Fisher, *Radiat. Prot. Dosim.*, 1994, 55(3), 199-206.
- 35 J. S. Crain, L. L. Smith, J. S. Yaeger and J. A. Alvarado, J. Radioanal. Nucl. Chem., 1995, 194(1), 133-139.
- 36 Y. Igarashi, C. K. Kim, Y. Takaku, K. Shiraishi, M. Yamamooto and N. Ikeda, Anal. Sci., 1990, 6, 157-164.
- 37 S. Collins, S. Pommé, S. M. Jerome and A. K. Pearce, Appl. Radiat. Isot., 2015, 104, 203-211.
- 38 S. M. Collins, A. K. Pearce, K. Ferreira, A. Fenwick, P. H. Regan and J. Keightley, Appl. Radiat. Isot., 2015, 99,
- 39 NPL Radioactivity Group, http://www.npl.co.uk/sciencetechnology/radioactivity/products-and-services/primary-andsecondary-standards, accessed 17/08/16.
- 40 NIST Radionuclide Half-Life measurements, http:// www.nist.gov/pml/data/halflife.cfm, accessed 17/08/16.
- 41 J. S. Becker and H. J. Dietz, Fresenius' J. Anal. Chem., 1999, 364, 482-488.
- 42 C. S. Kim, C. U. Kim, J. I. Lee and K. J. Lee, J. Anal. At. Spectrom., 2000, 15, 247-255.
- 43 S. F. Boulyga and J. S. Becker, Fresenius' J. Anal. Chem., 2001, 370, 612-617.
- 44 K. K. Falkner, G. P. Klinkhammer, C. Ungerer and D. M. Christie, Annu. Rev. Earth Planet. Sci., 1995, 23, 409-
- 45 C. M. Barshick, D. C. Duckworth and D. H. Smith, Inorganic Mass Spectrometry Fundamentals and Applications, Oak Ridge National Laboratory, Marcel Dekker Inc., Oak Ridge, Tennessee, 2000.
- 46 R. Thomas, A Beginner's Guide to ICP-MS, Part II: The Sample Introduction System, Spectroscopy, 2001, 16(5), 56 - 60.
- 47 W. Kerl, J. S. Becker, H.-J. Dietze and W. Dannecker, Fresenius' J. Anal. Chem., 1997, 359, 407-409.
- 48 J. S. Becker and H. J. Dietz, J. Anal. At. Spectrom., 1999, 14, 1493-1500.
- 49 S. F. Boulyga and J. S. Becker, J. Anal. At. Spectrom., 2002, 17, 1143-1147.
- 50 N. Jakubowski, L. Moens and F. Vanhaecke, Spectrochim. Acta, Part B, 1998, 53(12), 1739-1763.
- 51 R. N. Taylor, T. Warneke, J. A. Milton, I. W. Croudace, P. E. Warwick and R. W. Nesbitt, J. Anal. At. Spectrom., 2003, 18, 480-484.
- 52 J. A. McLean, J. S. Becker, S. F. Boulyga, H. J. Dietze and A. Montaser, Int. J. Mass Spectrom., 2001, 208, 193.
- 53 S. F. Boulyga, C. Testa, D. Desideri and J. S. Becker, J. Anal. At. Spectrom., 2001, 16, 1283-1289.
- 54 D. Larivière, V. F. Taylor, R. D. Evans and R. J. Cornett, Spectrochim. Acta, Part B, 2006, 61(8), 877-904.
- 55 M. G. Minnich and R. S. Houk, J. Anal. At. Spectrom., 1998,
- 56 J. Zheng, J. Nucl. Radiochem. Sci., 2015, 15(1), 7-13.

- 57 Z. Varga, G. Surányi, N. Vajda and Z. Stefánka, *Microchem. J.*, 2006, 85, 39–45.
- 58 J. S. Becker, Spectrochim. Acta, Part B, 2003, 58, 1757–1784.
- 59 M. Hollenbach, J. Grohs, S. Mamich and M. Kroft, *J. Anal. At. Spectrom.*, 1994, **9**, 927–933.
- 60 J. H. Aldstadt, J. M. Kuo, L. L. Smith and M. D. Erickson, *Anal. Chim. Acta*, 1996, **319**, 135–143.
- 61 E. H. Hansen and M. Miró, Trends Anal. Chem., 2007, 26(1), 18–26.
- 62 N. Vajda and C.-K. Kim, J. Radioanal. Chem., 2010, 284, 341-366.
- 63 J. Qiao, X. Hou, P. Roos and M. Miró, *J. Anal. At. Spectrom.*, 2010, 25, 1769–1779.
- 64 J. Ruzicka, Analyst, 2000, 125, 1053-1060.
- 65 S. F. Boulyga, D. Desideri, M. A. Meli, C. Testa and J. S. Becker, *Int. J. Mass Spectrom.*, 2003, 226, 329–339.
- 66 M. D. Seltzer, Appl. Spectrosc., 2003, 57(9), 1173-1177.
- 67 J. S. Becker, Int. J. Mass Spectrom., 2005, 242, 183-195.
- 68 L.-S. Chen, T. Wang, Y.-K. Hsieh, C.-H. Hsu, J. C.-T. Lin and C.-F. Wang, *J. Membr. Sci.*, 2014, **456**, 202–208.
- 69 S. Bürger and L. R. Riciputi, *J. Environ. Radioact.*, 2009, **100**(11), 970–976.
- 70 Z. Varga, Anal. Chim. Acta, 2008, 625, 1-7.
- 71 F. Pointurier, A. Hubert and A. C. Pottin, *J. Radioanal. Nucl. Chem.*, 2013, **296**, 609–616.
- 72 J. V. Cizdziel, M. E. Ketterer, D. Farmer, S. H. Faller and V. F. Hodge, *Anal. Bioanal. Chem.*, 2008, **390**, 521–530.
- 73 J. S. Becker and H. J. Dietz, Fresenius' J. Anal. Chem., 2000, 368, 23–30.
- 74 J. S. Becker, C. Pickhardt and H.-J. Dietze, *Int. J. Mass Spectrom.*, 2000, **202**, 283–297.
- 75 Viridiscan, http://www.viridian-tc.co.uk/viridiscan.html, accessed 17/08/16.
- 76 R. C. Marin, J. E. S. Sarkis and M. R. L. Nascimento, J. Radioanal. Nucl. Chem., 2013, 295, 99-104.
- 77 P. Grinberg, S. Willie and R. E. Sturgeon, *J. Anal. At. Spectrom.*, 2007, 22(11), 1409–1414.
- 78 J. S. Santos, L. S. G. Teixeira, W. N. L. dos Santos, V. A. Lemos, J. M. Godoy and S. L. C. Ferreira, *Anal. Chim. Acta*, 2010, 674, 143–156.
- 79 M. Song, T. U. Probst and N. G. Berryman, *Fresenius' J. Anal. Chem.*, 2001, **370**(6), 744–751.
- 80 J. B. Truscott, L. Bromley, P. Jones, E. Hywel Evans, J. Turner and B. Fairman, *J. Anal. At. Spectrom.*, 1999, **14**, 627–631.
- 81 P. Grinberg, S. Willie and R. E. Sturgeon, *J. Anal. At. Spectrom.*, 2005, **20**, 717–723.
- 82 L. A. De las Heras, E. Hrnecek, O. Bildstein and M. Betti, *J. Anal. At. Spectrom.*, 2002, 17, 1011–1014.
- 83 M. Betti and L. A. de las Heras, *Spectrochim. Acta, Part B*, 2004, 59(9), 1359–1376.
- 84 S. F. Wolf, D. L. Bowers and J. C. Cunnane, *J. Radioanal. Nucl. Chem.*, 2005, **263**(3), 581–586.
- 85 I. Günther-Leopold, N. Kivel, J. K. Waldes and B. Wernli, *Anal. Bioanal. Chem.*, 2008, **309**, 503–510.
- 86 A. P. Vonderheide, M. V. Zoriy, A. V. Izmer, C. Pickhardt, J. A. Caruso, P. Ostapczuk, R. Hille and J. S. Becker, J. Anal. At. Spectrom., 2004, 19(5), 675–680.

- 87 M. V. Zoriy, P. Ostapczuk, L. Halicz, R. Hille and J. S. Becker, *Int. J. Mass Spectrom.*, 2005, 242(2–3), 203–209.
- 88 A. R. Timerbaev and R. M. Timerbeav, *Trends Anal. Chem.*, 2013, **51**, 44–50.
- 89 C. Möser, R. Kautenburger and H. P. Beck, *Electrophoresis*, 2012, 33(9–10), 1482–1487.
- 90 C. Bresson, E. Ansoborlo and C. Vidaud, *J. Anal. At. Spectrom.*, 2011, 26, 593-601.
- 91 Perkin Elmer, Elan II DRC brochure, http://www.esc.cam.ac.uk/esc/files/Department/facilities/icp-ms/drcii-b.pdf, accessed 18/05/2016.
- 92 D. R. Bandura, V. I. Baranov, A. E. Litherland and S. D. Tanner, *Int. J. Mass Spectrom.*, 2006, 235–236, 312–327.
- 93 U. Nygren, I. Rodushkin, C. Nilsson and D. C. Baxter, J. Anal. At. Spectrom., 2003, 18(12), 1426–1434.
- 94 M. Moldovan, E. M. Krupp, A. E. Holliday and O. F. X. Donard, *J. Anal. At. Spectrom.*, 2004, **1**(9), 815–822.
- 95 M. E. Ketterer and S. C. Szechenyi, *Spectrochim. Acta, Part B*, 2008, **63**, 719–737.
- 96 M. Hamester, D. Wiederin, J. Wills, W. Kerl and C. B. Douthitt, *Fresenius' J. Anal. Chem.*, 1999, 364(5), 495–498.
- 97 D. Solatie, P. Carbol, M. Betti, F. Bocci, T. Hiernaut, V. V. Rondinella and J. Cobos, *Fresenius' J. Anal. Chem.*, 2000, **368**, 88–94.
- 98 Z. Varga, Anal. Chim. Acta, 2007, 587, 165-169.
- 99 M. E. Wieser and J. B. Schwieters, *Int. J. Mass Spectrom.*, 2005, 242(2-3), 97-115.
- 100 Spectro, http://www.spectro.com/products/icp-ms-spectro meters, Overview of ICP-MS spectrometers, accessed 18/ 05/2016.
- 101 N. L. Sanders, S. Kothari, G. Huang, G. Salazar and R. G. Cooks, *Anal. Chem.*, 2010, **82**, 5313–5316.
- 102 J. K. Dalgleish, K. Hou, Z. Ouyang and R. G. Cooks, *Anal. Lett.*, 2012, 45, 1440–1446.
- 103 J. M. Wells, M. J. Roth, A. D. Keil, J. W. Grossenbacher, D. R. Justes, G. E. Patterson and D. J. Barket Jr, *J. Am. Soc. Mass Spectrom.*, 2008, 19(10), 1419–1424.
- 104 A. Malcolm, S. Wright, R. R. A. Syms, N. Dash, M.-A. Schwab and A. Finlay, *Anal. Chem.*, 2010, **82**, 1751–1758.
- 105 P. I. Hendricks, J. K. Dalgleish, J. T. Shelley, M. A. Kirleis, M. T. McNicholas, L. Li, T.-C. Chen, C.-H. Chen, J. S. Duncan, F. Boudreau, R. J. Noll, J. P. Denton, T. A. Roach, Z. Ouyang and R. G. Cooks, *Anal. Chem.*, 2014, 86, 2900–2908.
- 106 S. Kumano, M. Sugiyama, M. Yamada, K. Nishimura, H. Hasegawa, H. Morokuma, H. Inoue§ and Y. Hashimoto, *Anal. Chem.*, 2013, **85**(10), 5033–5039.
- 107 J. M. Giaquinto, J. M. Keller and A. M. Meeks, J. Radioanal. Nucl. Chem., 1998, 234(1-2), 137-141.
- 108 F. Pilon, S. Lorthioir, J. Birolleau and S. Lafontan, *J. Anal. At. Spectrom.*, 1996, **11**(9), 759–764.
- 109 J. I. G. Alonso, D. Thoby-Schultzendorff, B. Giovanonne, L. Koch and H. Wiesmann, *J. Anal. At. Spectrom.*, 1993, **8**, 673–679.
- 110 J. D. Burraston, PhD thesis, *GAU-Radioanalytical, Ocean and Earth Science*, University of Southampton, National Oceanography Centre, 2014.

111 R. Forsyth and U.-B. Eklund, *Spent nuclear fuel corrosion: SKB Technical Report 95-04*, Swedish Nuclear Fuel and Waste Management Company, 1995.

- 112 M. Betti, G. Rasmussen, T. Hiernaut and L. Koch, *J. Anal. At. Spectrom.*, 1994, 9, 385–391.
- 113 M. R. Smith, O. T. Farmer III, J. H. Reeves and D. W. Koppenaal, *J. Radioanal. Nucl. Chem.*, 1995, **194**(1), 7–13.
- 114 J. M. Barrero Moreno, M. Betti and J. I. Garcia Alonso, J. Anal. At. Spectrom., 1997, 12, 355–361.
- 115 O. B. Egorov, M. J. O'Hara, O. T. Farmer III and J. W. Grate, *Analyst*, 2000, **126**, 1594–1601.
- 116 M. Krachler, R. Alvarez-Sarandes and S. V. Winckel, *J. Anal. At. Spectrom.*, 2014, 29, 817–824.
- 117 I. W. Croudace, P. E. Warwick, R. Taylor and S. Dee, *Anal. Chim. Acta*, 1998, **371**, 217–225.
- 118 C. S. Kim, C.-K. Kim, P. Martin and U. Sansone, *J. Anal. At. Spectrom.*, 2007, 22, 827–841.
- 119 I. W. Croudace, P. E. Warwick, D. G. Reading and B. C. Russell, *Trends Anal. Chem.*, 2016, **85**, 120–129.
- 120 R. Bock, *A Handbook of Decomposition Methods in Analytical Chemistry*, International Textbook Company Limited, London, 1979.
- 121 B. Parsa, J. Radioanal. Nucl. Chem., 1992, 157, 65-73.
- 122 U.S. Environmental Protection Agency, Rapid Method for Sodium Hydroxide/Sodium Peroxide Fusion of Radioisotope Thermoelectric Generator Materials in Water and Air Filter Matrices Prior to Plutonium Analyses for Environmental Remediation Following Radiological Incidents, 2014, EPA/600/F-14/284.
- 123 S. Fisher and R. Kunin, Anal. Chem., 1957, 29, 400-402.
- 124 C. Galindo, L. Mougin and A. Nourreddine, *Appl. Radiat. Isot.*, 2007, **65**, 9–16.
- 125 C. W. Sill, K. W. Puphal and F. D. Hindman, *Anal. Chem.*, 1974, **46**, 1725–1737.
- 126 J. S. Fedorowich, J. P. Richards, J. C. Cain, R. Kerrich and J. Fan, *Chem. Geol.*, 1993, **106**, 229–249.
- 127 M. Dong, D. Oropeza, J. Chirinos, J. J. González, J. Lu, X. Mao and R. E. Russo, *Spectrochim. Acta,Part B*, 2015, **109**, 44–50.
- 128 K. Subedi, T. Trjos and J. Almirall, *Spectrochim. Acta, Part B*, 2015, **103–104**, 76–83.
- 129 Y. Lee, S.-H. Nam, K.-S. Ham, J. Gonzalez, D. Oropeza, D. Quarles Jr, J. Yoo and R. E. Rosso, *Spectrochim. Acta, Part B*, 2016, **118**, 102–111.
- 130 L. Ashton, P. Warwick and D. Giddings, *Analyst*, 1999, **124**, 627–632.
- 131 M. Baxter, L. Castle, H. M. Crews, M. Rose, C. Garner, G. Lappin and D. Leong, *Food Addit. Contam., Part A*, 2009, **26**, 139–144.
- 132 X. Hou, L. F. Ostergaard and S. P. Nielsen, *Anal. Chem.*, 2007, **79**, 3126–3134.
- 133 A. Zulaf, S. Happel, P. Warwick, B. M. Mokili, A. Bombard and H. Junglas, http://www.triskem-international.com/fr/iso_album/poster_rrmc_2011_cl_resin.pdf, accessed 17/08/16.

- 134 M. Martschini, P. Andersson, O. Fortsner, R. Golser, D. Hanstorp, A. O. Lindahl, W. Kutschera, S. Pavetich, A. Priller, J. Rohlén, P. Steier, M. Suter and A. Wallner, Nucl. Instrum. Methods Phys. Res., Sect. B, 2013, 294, 115– 120.
- 135 P. W. Kubik, D. Elmore, N. J. Conard, K. Nishiizumi and J. R. Arnold, *Nature*, 1986, **319**, 568–570.
- 136 P. Müller, B. A. Bushaw, K. Blaum, S. Diel, C. Geppert, A. Nähler, N. Trautmann, W. Nörtershäuser and K. Wendt, Anal. Chem., 2001, 370, 508–512.
- 137 K. D. A. Wendt, C. Geppert, M. Miyabe, P. Müller, W. Nörtershäuser and N. Trautmann, *J. Nucl. Sci. Technol.*, 2002, **39**(4), 303–307.
- 138 C. Hennessy, M. Berglund, M. Ostermann, T. Walczyk, H.-A. Synal, C. Geppert, K. Wendt and P. D. P. Taylor, Nucl. Instrum. Methods Phys. Res., Sect. B, 2005, 299, 281– 292.
- 139 D. Hampe, B. Gleisberg, S. Akhmadaliev, G. Rugel and S. Merchel, *J. Radioanal. Nucl. Chem.*, 2013, **296**, 617–624.
- 140 G. S. Jackson, D. J. Hillegonds, P. Muzikar and B. Goehring, Nucl. Instrum. Methods Phys. Res., Sect. B, 2013, 313, 14–20.
- 141 S. Merchel, L. Benedetti, D. L. Bourlès, R. Braucher, A. Deqald, T. Faestermann, R. C. Finkel, G. Korschinek, J. Masarik, M. Poutivtsev, P. Rochette, G. Rugel and K.-O. Zell, *Nucl. Instrum. Methods Phys. Res., Sect. B*, 2010, 268, 1179–1184.
- 142 X. Hou, Radiochim. Acta, 2005, 93, 611-617.
- 143 P. E. Warwick, I. W. Croudace and D. J. Hillegonds, *Anal. Chem.*, 2009, 81, 1901–1906.
- 144 G. Jörg, Y. Amelin, K. Kossert, C. Lierse and L. V. Gostomski, Geochim. Cosmochim. Acta, 2012, 88, 51– 65.
- 145 E. Nottoli, D. Bourlès, P. Bienvenu, A. Labet, M. Arnold and M. Bertaux, *Appl. Radiat. Isot.*, 2013, **82**, 340–346.
- 146 X. Hou, L. F. Østergaard and S. P. Nielsen, *Anal. Chim. Acta*, 2005, 535(1–2), 297–307.
- 147 E. Holm, P. Roos and B. Skwarzec, *Appl. Radiat. Isot.*, 1992, 43(1–2), 371–376.
- 148 I. Gresits and S. Tölgyesi, *J. Radioanal. Nucl. Chem.*, 2003, 258(1), 107–122.
- 149 P. E. Warwick and I. W. Croudace, *Anal. Chim. Acta*, 2006, **567**, 277–285.
- 150 A. A. Marchetti, L. J. Hainsworth, J. E. McAninch, M. R. Leivers, P. R. Jones, I. D. Proctor and T. Straume, Nucl. Instrum. Methods Phys. Res., Sect. B, 1997, 123(1-4), 230-234.
- 151 P. Persson, M. Kiisk, B. Erlandsson, M. Faarinen, R. Hellborg, G. Skog and K. Stenström, *Nucl. Instrum. Methods Phys. Res., Sect. B*, 2000, 172(1-4), 188-192.
- 152 M. E. Mount, D. W. Layton, N. M. Lynn and T. F. Hamilton, Report UCRL-JC-130412, Lawrence Livermore National Laboratory, USA, 1998.
- 153 S. Aguerre and C. Frechou, *Talanta*, 2006, **69**(3), 565–571.
- 154 J. Compte, P. Bienvenu, E. Brochard, J.-M. Fernandez and G. Andreoletti, *J. Anal. At. Spectrom.*, 2003, **18**, 702–707.
- 155 P. Bienvenu, P. Cassette, G. Andreoletti, M.-M. Be, J. Comte and M.-C. Lépy, *Appl. Radiat. Isot.*, 2007, **65**(3), 355–364.

- 156 S. Brun, S. Bessac, D. Uridat and B. Boursier, J. Radioanal. Nucl. Chem., 2002, 253(2), 191-197.
- 157 W. Bu, J. Zheng, X. Liu, K. Long, S. Hu and S. Uchida, Spectrochim. Acta, Part B, 2016, 119, 65-75.
- 158 J. Feuerstein, S. F. Boulyga, P. Galler, G. Stingeder and T. Prohaska, J. Environ. Radioact., 2008, 99(11), 1764-1769.
- 159 V. Taylor, R. D. Evans and R. J. Cornett, Anal. Bioanal. Chem., 2007, 387(1), 343-350.
- 160 Y. Takagai, M. Furukawa, Y. Kameo and K. Suzuki, Anal. Methods, 2014, 6(2), 355-362.
- 161 M. Sakama, Y. Nagano, T. Saze, S. Higaki, T. Kitade, N. Izawa, O. Shikino and S. Nakayama, Appl. Radiat. Isot., 2013, 81, 201-207.
- 162 M. A. Amr, A.-F. I. Helal, A. T. Al-Kinani and P. Balakrishnan, J. Environ. Radioact., 2016, 153, 73-87.
- 163 E. P. Steinberg and L. E. Glendenin, Phys. Rev., 1950, 78,
- 164 J. I. G. Alonso, D. Thoby-Schultzendorff, B. Giovanonne, J.-P. Glatz, G. Pagliosa and L. Koch, J. Anal. At. Spectrom., 1994, 9, 1209-1215.
- 165 F. Chartier, H. Isnard, J. P. Degros, A. L. Faure and C. Fréchou, Int. J. Mass Spectrom., 2008, 270(3), 127-133.
- 166 S. Dulanská, B. Remenec, V. Gardoňová and L. Mátel, J. Radioanal. Nucl. Chem., 2012, 293, 635-640.
- 167 P. Cassette, F. Chartier, H. Isnard, C. Fréchou, I. Laszak, J. P. Degros, M. M. Bé, M. C. Lépy and I. Tartes, Appl. Radiat. Isot., 2010, 68, 122-130.
- 168 K. Shi, J. Qiao, W. Wu, P. Roos and X. Hou, Anal. Chem., 2012, 84, 6783-6789.
- 169 K. Shi, X. Hou, P. Roos and W. Wu, Anal. Chim. Acta, 2012, **709**, 1-20.
- 170 T. Y. Su, T. L. Tsay, H. C. Wu and L. C. Men, J. Radioanal. Nucl. Chem., 2015, 303, 1245-1248.
- 171 K. Shi, X. Hou, P. Roos and W. Wu, Anal. Chem., 2012, 84, 2009-2016.
- 172 K. H. Chung, S. D. Choi, G. S. Choi and M. J. Kang, Appl. Radiat. Isot., 2013, 81, 57-61.
- 173 L. S. Chen, T. H. Wang, Y. K. Hsieh, L. W. Jian, W. H. Chen, T. L. Tsai and C. F. Wang, J. Radioanal. Nucl. Chem., 2014, 299, 1883-1889.
- 174 N. E. Bibler, W. T. Boyce, C. J. Coleman and W. F. Kinard, Report WSRC-MS-97-0011, Office of Scientific Technical Information, Oak Ridge, USA, 1997.
- 175 C. J. Bannochie, N. E. Bibler and D. P. DiPrete, WM2009 conference proceedings, March 1-5 2009, Phoenix, USA, 2009.
- 176 P. G. Bienvenu, E. A. Brochard and E. A. Excoffier, in Application of inductively coupled plasma mass spectrometry to radionuclide determinations: second volume, ed. R.W. Morrow and J.S. Crain, ASTM, West Conshohocken, USA,
- 177 V. Hansen, P. Roos, A. Aldahan, X. Hou and G. Possnert, J. Environ. Radioact., 2011, 102, 1096-1104.
- 178 V. Hansen, P. Yi, X. Hou, A. Aldahan, P. Roos and G. Possnert, Sci. Total Environ., 2011, 412-413, 296-303.
- 179 Ž. Ežerinskis, A. Spolaor, T. Kirchgeorg, G. Cozzi, P. Vallelonga, H. A. Kjaer, J. Šapolaité, C. Barbante and R. Druteikiené, J. Anal. At. Spectrom., 2014, 29, 1827-1834.

- 180 C. Vockenhuber, N. Casacuberta, M. Christl and H.-A. Synal, Nucl. Instrum. Methods Phys. Res., Sect. B, 2015, 361, 510-516.
- 181 E. Englund, A. Aldahan, X. L. Hou, G. Possnert and C. Söderström, Nucl. Instrum. Methods Phys. Res., Sect. B, 2010, 268, 1139-1141.
- 182 R. Michel, A. Daraoui, M. Gorny, D. Jakob, R. Sachse, L. Tosch, H. Nies, I. Goronocy, J. Herrman, H. A. Synal, M. Stocker and V. Alfimov, Sci. Total Environ., 2012, 419, 151-169.
- 183 X. Hou, P. Povinec, L. Zhang, K. Shi, D. Biddulph, C.-C. Chang, Y. Fan, R. Golser, Y. Hou, M. Ješkovský, A. J. Tim Jull, Q. Liu, M. Luo, P. Steier and W. Zu, Environ. Sci. Technol., 2013, 47(7), 3091-3098.
- 184 Y. Muramatsu, H. Matsuzaki, C. Toyama and T. Ohno, J. Environ. Radioact., 2015, 139, 344-350.
- 185 J. M. Gómez-Guzmán, S. M. Enamorado-Báez, A. R. Pinto-Goméz and J. M. Abril-Hernández, Int. J. Mass Spectrom., 2011, 303, 103-108.
- 186 V. Alfimov and H.-A. Synal, Nucl. Instrum. Methods Phys. Res., Sect. B, 2010, 268, 769-772.
- 187 H. Fujiwara, K. Kawabata, J. Suzuki and O. Shikino, J. Anal. At. Spectrom., 2011, 26, 2528-2533.
- 188 J. Lehto, T. Räty, X. Hou, J. Paatero, A. Aldahan, G. Possnert, J. Flinkman and H. Kankaanpää, Sci. Total Environ., 2012, **419**, 60-67.
- 189 T. Ohno, Y. Muramatsu, Y. Shikamori, C. Toyama, N. Okabe and H. Matsuzaki, J. Anal. At. Spectrom., 2013, 28, 1283-
- 190 J. Zheng, H. Takata, K. Tagami, T. Aono, K. Fujita and S. Uchida, *Microchem. J.*, 2012, **100**, 42–47.
- 191 R. J. Cox, C. J. Pickford and M. Thompson, J. Anal. At. Spectrom., 1992, 7, 635-640.
- 192 E. Englund, A. Aldahan, X. L. Hou, R. Petersen and G. Possnert, Nucl. Instrum. Methods Phys. Res., Sect. B, 2010, 268, 1102-1105.
- 193 X. Hou, W. Zhou, N. Chen, L. Zhang, Q. Liu, M. Luo, Y. Fan, W. Liang and Y. Fu, Anal. Chem., 2010, 82, 7713-7721.
- 194 T. Suzuki, S. Otosaka, J. Kubawara, H. Kawamura and T. Kobayashi, Biogeosciences, 2013, 10, 3839-3847.
- 195 V. Taylor, R. Evans and R. Cornett, J. Environ. Radioact., 2008, 99(1), 109-118.
- 196 J. E. Delmore, D. C. Snyder, T. Tranter and N. R. Mann, J. Environ. Radioact., 2011, 102(11), 1008-1011.
- 197 D. C. Snyder, J. E. Delmore, T. Tranter, N. R. Mann, M. L. Abbott and J. E. Olson, J. Environ. Radioact., 2012, **110**, 46-52.
- 198 T. Ohno and Y. Muramatsu, J. Anal. At. Spectrom., 2014, **29**(2), 347–351.
- 199 J. Zheng, K. Tagami, W. Bu, S. Uchida, Y. Watanabe, Y. Kubota, S. Fuma and S. Ihara, Environ. Sci. Technol., 2014, 48, 5433-5438.
- 200 J. Zheng, W. Bu, K. Tagami, Y. Shikamori, K. Nakano, S. Uchida and N. Ishii, Anal. Chem., 2014, 86(14), 7103-
- 201 B. C. Russell, I. W. Croudace and P. E. Warwick, Anal. Chim. Acta, 2015, 890, 7-20.

202 Y. Shibahara, T. Kubota, T. Fuji, S. Fukutani, T. Ohta, K. Takamiya, R. Okumura, S. Mizuno and H. Yamana, *J. Nucl. Sci. Technol.*, 2014, **51**(5), 575–579.

- 203 B. C. Russell, P. E. Warwick and I. W. Croudace, *Anal. Chem.*, 2014, **86**(23), 11890–11896.
- 204 J. M. B. Moreno, J. I. G. Alonso, P. Arbore, G. Nicolaou and L. Koch, *J. Anal. At. Spectrom.*, 1996, 11(10), 929–935.
- 205 S. Asai, Y. Hanzawa, K. Okamura, N. Shinohara, J. Inagawa, S. Hotoku, K. Suzuki and S. Kaneko, *J. Nucl. Sci. Technol.*, 2011, **48**(5), 851–854.
- 206 J. Eliades, X.-L. Zhao, A. E. Litherland and W. E. Kieser, *Nucl. Instrum. Methods Phys. Res., Sect. B*, 2013, **294**, 361–363.
- 207 H. Isnard, M. Granet, C. Caussignac, E. Ducarme, A. Nonell, B. Tran and F. Chartier, *Spectrochim. Acta, Part B*, 2009, **64**(11–12), 1280–1286.
- 208 A. Pitois, L. A. De Las Heras and M. Betti, *Int. J. Mass Spectrom.*, 2008, **270**, 118–126.
- 209 M. Granet, A. Nonell, G. Favre, F. Chartier, H. Isnard, J. Moureau, C. Caussignac and B. Tran, *Spectrochim. Acta, Part B*, 2008, **63**(11), 1309–1314.
- 210 V. V. Lavrov, V. Blagojevic, G. K. Koyanagi, G. Orlova and D. K. Bohme, *J. Phys. Chem. A*, 2004, **108**(26), 5610–5624.
- 211 T. Lee, K. The-Lung, L. Hsiao-Ling and C. Ju-Chin, *Geochim. Cosmochim. Acta*, 1993, 57(14), 3493–3497.
- 212 L. Pibida, C. McMahon and B. A. Bushaw, *Appl. Radiat. Isot.*, 2004, **60**(2–4), 567–570.
- 213 C. Macdonald, C. R. J. Charles, R. J. Cornett, X. L. Zhao, W. E. Kieser and A. E. Litherland, *Congress de l'ACP, Contribution ID: 155 Type: oral, Session Classification : (T2–9) Instrumentation-DIMP/Instrumentation-DPIM*, 2014.
- 214 J. I. G. Alonso, F. Sena, P. Arbore, M. Betti and L. Koch, *J. Anal. At. Spectrom.*, 1995, **10**, 381–393.
- 215 A. Meeks, J. Giaquinto and J. Keller, *J. Radioanal. Nucl. Chem.*, 1998, 234(1), 131–136.
- 216 J. M. B. Moreno, M. Betti and G. Nicolaou, *J. Anal. At. Spectrom.*, 1999, **14**(5), 875–879.
- 217 M. Liezers, O. T. Farmer and M. Thomas, *J. Radioanal. Nucl. Chem.*, 2009, **282**(1), 309–313.
- 218 J. E. Martin, in *Environmental Radiochemical Analysis*, ed. G. W. A. Newton, Royal Society of Chemistry, London, UK, 1999.
- 219 S. Sumiya, N. Hayashi, H. Katagiri and O. Narita, *Sci. Total Environ.*, 1993, **130–131**, 303–315.
- 220 M. Yoshida, S. Sumiya, H. Watanabe and K. Tobit, *J. Radioanal. Nucl. Chem.*, 1995, **197**, 219–227.
- 221 L. Vio, G. Crétier, F. Chartier, V. Geertsen, A. Gourgiotis, H. Isnard and J.-L. Rocca, *Talanta*, 2012, **99**, 586–593.
- 222 H. Isnard, R. Brennetot, C. Caussignac, N. Caussignac and F. Chartier, *Int. J. Mass Spectrom.*, 2005, **246**, 66–73.
- 223 M. Baskaran, J. Environ. Radioact., 2011, 102, 500-513.
- 224 M. A. Amr, K. A. Al-Saad and A. I. Helal, *Nucl. Instrum. Methods Phys. Res., Sect. A*, 2010, **615**, 237–241.
- 225 A. E. M. Khater and W. F. Bakr, *J. Environ. Radioact.*, 2011, **102**, 527–530.
- 226 D. Larivière, K. M. Reiber, R. D. Evans and R. J. Cornett, *Anal. Chim. Acta*, 2005, **549**, 188–196.

- 227 Z. Varga, Anal. Bioanal. Chem., 2008, 390, 511-519.
- 228 T. Zhang, D. Bain, R. Hammack and R. D. Vidic, *Environ. Sci. Technol.*, 2015, **49**(5), 2969–2976.
- 229 Y.-T. Hsieh and G. M. Henderson, *J. Anal. At. Spectrom.*, 2011, **26**, 1338–1346.
- 230 M. Bourquin, P. Van Beek, J. L. Reyss, J. Riotte and R. Freydier, *Mar. Chem.*, 2011, **126**, 132–138.
- 231 M. Leermakers, Y. Gao, J. Navez, A. Poffijn, K. Croes and W. Baeyens, *J. Anal. At. Spectrom.*, 2009, **24**, 1115–1117.
- 232 Y. Gao, W. Baeyens, S. De Galan, A. Poffijn and M. Leermakers, *Environ. Pollut.*, 2010, **158**, 2439–2445.
- 233 G. Sharabi, B. Lazar, Y. Kolodny, N. Teplyakov and L. Halicz, *Int. J. Mass Spectrom.*, 2010, **294**, 112–115.
- 234 M. S. Choi, R. Francois, K. Sims, M. P. Bacon, S. Brown-Leger, A. P. Fleer, L. Ball, D. Schneider and S. Pichat, *Mar. Chem.*, 2001, **76**, 99–112.
- 235 D. A. Pickett, M. T. Murrell and R. W. Williams, *Anal. Chem.*, 1994, **66**(7), 1044–1049.
- 236 C.-C. Shen, H. Cheng, R. L. Edwards, S. B. Moran, H. N. Edmonds, J. A. Hoff and R. B. Thomas, *Chem. Geol.*, 2002, 3–4, 165–178.
- 237 M. Regelous, S. P. Turner, T. R. Elliott, K. Rostami and C. J. Hawkesworth, *Anal. Chem.*, 2004, **76**, 3584–3589.
- 238 R. A. Mortlock, R. G. Fairbanks, T.-C. Chiu and J. Rubenstone, *Geochim. Cosmochim. Acta*, 2005, **69**(3), 649–657.
- 239 C.-C. Shen, R. L. Edwards, H. Cheng, J. A. Dorale, R. B. Thomas, S. B. Moran, S. E. Weinstein and H. N. Edmonds, *Chem. Geol.*, 2002, **185**, 165–178.
- 240 H. Cheng, R. L. Edwards, C.-C. Shen, V. J. Polyak, Y. Asmerom, J. Woodhead, J. Hellstrom, Y. Wang, X. Kong, C. Spötl, X. Wang and E. C. Alexander Jr, *Earth Planet. Sci. Lett.*, 2013, 371–372, 82–91.
- 241 H. Cheng, R. L. Edwards, J. Hoff, C. D. Gallup, D. A. Richards and Y. Asmerom, *Chem. Geol.*, 2000, **169**, 17–33.
- 242 Y. Shi, R. Collins and C. Broome, *J. Radioanal. Nucl. Chem.*, 2013, **269**, 509–515.
- 243 S. K. Aggarwal, Radiochim. Acta, 2016, 104(7), 445-455.
- 244 E. W. Hoppe, N. R. Overman and B. D. LaFerriere, Low Radioactivity Techniques, *AIP Conf. Proc.*, 2013, **1549**, 58–65.
- 245 J. S. Becker, M. Burow, M. V. Zoriy, C. Pickhardt, P. Ostapczuk and R. Hille, *J. Anal. At. Spectrom.*, 2004, 25(5), 197–202.
- 246 M. L. Cozzella and R. Pettirossi, *Radiation Protection Institute ENEA*, Casaccia, Rome, Italy, 2012.
- 247 H. E. L. Palmieri, E. A. N. Knupp, L. M. L. A. Auler, L. M. F. Gomes and C. C. Windmöller, *International Nuclear Atlantic Conference-INAC*, 2011.
- 248 J. Avivar, L. Ferrer, M. Cass and V. Cerà, *J. Anal. At. Spectrom.*, 2012, 27, 327–334.
- 249 H. Tuovinen, D. Vesterbacka, E. Pohjolainen, D. Read, D. Solatie and J. Lehto, J. Geochem. Explor., 2015, 148, 174–180
- 250 A. Habibi, B. Boulet, M. Gleizes, D. Larrriiivière and G. Cote, *Anal. Chim. Acta*, 2015, **883**, 109–116.

- 251 A. Borylo, J. Radioanal. Nucl. Chem., 2013, 295, 621-631.
- 252 P. J. Gray, L. Zhang, H. Zu, M. McDiarmid, K. Squibb and J. A. Centeno, *Microchem. J.*, 2012, **105**, 94–100.
- 253 J. G. Arnason, C. N. Pellegri and P. J. Parsons, *J. Anal. At. Spectrom.*, 2013, **28**, 1410–1419.
- 254 J. G. Arnason, C. N. Pellegri and P. J. Parsons, *J. Anal. At. Spectrom.*, 2015, **30**, 126–138.
- 255 C. Liu, B. Hu, J. Shi, J. Li, X. Zhang and H. Chen, *J. Anal. At. Spectrom.*, 2011, **26**, 2045–2051.
- 256 S. F. Boulyga, J. L. Matusevich, V. P. Mironov, V. P. Kudrjashov, L. Halicz, I. Segal, J. A. McLean, A. Montaser and J. S. Becker, *J. Anal. At. Spectrom.*, 2002, 17, 958–964.
- 257 M. Tanimizu, N. Sugiyama, E. Ponzevera and G. Bayon, *J. Anal. At. Spectrom.*, 2013, **28**, 1372–1376.
- 258 S. Uchida, R. García-Tenorio, K. Tagami and M. García-León, *J. Anal. At. Spectrom.*, 2000, **15**, 889–892.
- 259 R. S. Pappas, B. G. Ting, J. M. Jarrett, D. C. Paschal, S. P. Caudill and D. T. Miller, *J. Anal. At. Spectrom.*, 2002, 17, 131–134.
- 260 S. L. Maxwell III and V. D. Jones, *Talanta*, 2009, **80**, 143-150.
- 261 J. W. Ejnik, T. I. Todorov, F. G. Mullick, K. Squibb, M. A. McDiarmid and J. A. Centeno, *Anal. Bioanal. Chem.*, 2005, **382**, 73–79.
- 262 P. Krystek and R. Ritsema, Anal. Bioanal. Chem., 2002, 374, 226–229.
- 263 M. Magara, T. Sakakibara, S. Kurosawa, M. Takahashi, S. Sakurai, Y. Hanzawa, F. Esaka, K. Watanabe and S. Usada, J. Anal. At. Spectrom., 2002, 17, 1157–1160.
- 264 R. R. Parrish, M. F. Thirlwall, C. Pickford, M. Horstwood, A. Gerdes, J. Anderson and D. Coggon, *Health Phys.*, 2006, **90**(2), 127–138.
- 265 X. Dai and S. K. Tremblay, *J. Radioanal. Nucl. Chem.*, 2011, **289**, 461–466.
- 266 P. Thakur and G. P. Mulholland, Appl. Radiat. Isot., 2012, 70, 1747–1778.
- 267 Y.-Q. Ji, J.-Y. Li, S.-G. Luo, T. Wu and J.-L. Liu, Fresenius' J. Anal. Chem., 2001, 371, 49-53.
- 268 J. B. Truscott, P. Jones, B. E. Fairman and E. H. Evans, *Anal. Chim. Acta*, 2001, **433**, 245–253.
- 269 T. C. Kenna, J. Anal. At. Spectrom., 2002, 17, 1471-1479.
- 270 C. S. Kim, C. K. Kim and K. J. Lee, *J. Anal. At. Spectrom.*, 2004, **19**, 743–750.
- 271 Q. C. Chen, H. Dahlgaard, S. P. Nielsen and A. Aarkrog, *J. Radioanal. Nucl. Chem.*, 2002, **253**(3), 451–458.
- 272 M. Ayranov, U. Krähenbühl, H. Sahli, S. Röllin and M. Burger, *Radiochim. Acta*, 2005, **93**, 631–635.
- 273 P. Lindahl, P. Roos, E. Holm and H. Dahlgaard, *J. Environ. Radioact.*, 2005, **82**, 285–301.
- 274 T. C. Kenna, J. Environ. Radioact., 2009, 100, 547-557.
- 275 S. L. Maxwell, B. K. Culligan, V. D. Jones, S. T. Nichols, M. A. Bernard and G. W. Noyes, *Anal. Chim. Acta*, 2010, 682, 130–136.
- 276 Y. Shi, X. Dai, C. Li, R. Collins, S. Kramer-Tremblay, R. Riopel and C. Broome, *J. Anal. At. Spectrom.*, 2014, 29, 1708–1713.

- 277 P. Zhao, R. M. Tinnacher, M. Zavarin and A. B. Kersting, J. Environ. Radioact., 2014, 137, 163–172.
- 278 N. Guérin, M.-A. Langevin, K. Nadeau, C. Labrecque, A. Gagnè and D. Larivière, Appl. Radiat. Isot., 2010, 68, 2132–2139.
- 279 J. Qiao, X. Hou, P. Roos and M. Miró, *Anal. Chem.*, 2011, **83**, 374–381.
- 280 B. S. Matteson, S. K. Hanson, J. L. Miller and W. J. Oldham Jr, *J. Environ. Radioact.*, 2015, **142**, 62–67.
- 281 S. Jerome, P. Ivanov, C. Larijani, D. H. Parker and P. H. Regan, *J. Environ. Radioact.*, 2014, **138**, 315–322.
- 282 A. Gourgiotis, M. Granet, H. Isnard, A. Nonell, C. Gautier, G. Stadelmann, M. Aubert, D. Durand, S. Legand and F. Chartier, *J. Anal. At. Spectrom.*, 2010, 25, 1939–1945.
- 283 B. Kuczewski, C. M. Marquardt, A. Seibert, H. Geckeis, J. V. Kratz and N. Trautmann, *Anal. Chem.*, 2003, 75(24), 6769–6774.
- 284 S. Röllin, H. Sahli, R. Holzer, M. Astner and M. Burger, *Appl. Radiat. Isot.*, 2009, **67**, 821–827.
- 285 J. Qiao, X. Hou, M. Miró and P. Roos, Anal. Chim. Acta, 2009, 652, 66–84.
- 286 T. Warneke, PhD thesis, *GAU-Radioanalytical, Ocean and Earth Science*, Universityof Southampton, 2002.
- 287 J. Zheng, T. Aono, S. Uchida, J. Zhang and M. C. Honda, *Geochem. J.*, 2012, **46**, 361–369.
- 288 N. Erdmann, G. Hermann, G. Huber, S. Köhler, J. V. Kratz, A. Mansel, M. Nunneman, G. Passler, N. Trautmann, A. Turchin and A. Waldek, *Fresenius' J. Anal. Chem.*, 1997, 359, 378–381.
- 289 Y. Ohtsuka, Y. Takaku, K. Nishimura, J. Kimura, S. Hisamatsu and J. Inaba, *Anal. Sci.*, 2006, 22, 309–311.
- 290 D. Larivière, T. A. Cumming, S. Kiser, C. Li and R. J. Cornett, *J. Anal. At. Spectrom.*, 2007, 22, 1–9.
- 291 B. G. Ting, R. S. Pappas and D. C. Paschal, *J. Anal. At. Spectrom.*, 2003, **18**, 795–797.
- 292 J. Qiao, X. Hou, P. Roos and M. Miró, *Anal. Chem.*, 2013, **85**, 2853–2859.
- 293 S. Cagno, K. Hellemans, O. C. Lind, L. Skipperud, K. Janssens and B. Salbu, *Environ. Sci.: Processes Impacts*, 2014, **16**, 306–312.
- 294 Z. Varga, A. Nicholl, M. Wallenius and K. Mayer, J. Radioanal. Nucl. Chem., 2015, 307(3), 1919–1926.
- 295 K. Bu, J. V. Cizdziel and D. Dasher, *J. Environ. Radioact.*, 2013, **124**, 29–36.
- 296 M. Jäggi, S. Röllin, J. A. C. Alvaredo and J. Eikenberg, *Appl. Radiat. Isot.*, 2012, **70**, 360–364.
- 297 S. Salminen-Paatero, U. Nygren and J. Paatero, *J. Environ. Radioact.*, 2012, **113**, 163–170.
- 298 T. Shinonaga, P. Steier, M. Lagos and T. Ohkura, *Environ. Sci. Technol.*, 2014, 48, 3608–3814.
- 299 H. Hernández-Mendoza, E. Chamizo, A. Delgado, M. García-León and A. Yllera, J. Anal. At. Spectrom., 2011, 26, 1509–1513.
- 300 F. N. Pointurier, N. Baglan and P. Hémet, *Appl. Radiat. Isot.*, 2004, **60**, 561–566.
- 301 V. N. Epov, R. D. Evans, J. Zheng, O. F. X. Donard and M. Yamada, *J. Anal. At. Spectrom.*, 2007, 22, 1131–1137.

302 M. L. D. P. Godoy, J. M. Godoy and L. A. Roldão, *J. Environ. Radioact.*, 2007, **97**, 124–136.

- 303 M. V. Zoriy, L. Halicz, M. E. Ketterer, C. Pickhardt, P. Ostapczuk and J. S. Becker, *J. Anal. At. Spectrom.*, 2004, 19, 362.
- 304 O. F. X. Donard, F. Bruneau, M. Moldovan, H. Garraud, V. N. Epov and D. Boust, *Anal. Chim. Acta*, 2007, **587**, 170–179.
- 305 S. K. Aggarwal, *Mass Spectrom. Rev.*, 2016, DOI: 10.1002/mas.21506.
- 306 G. Xiao, D. Saunders, R. L. Jones and K. L. Caldwell, *J. Radioanal. Nucl. Chem.*, 2014, **301**, 285–291.
- 307 M. Agarande, S. Benzoubir, P. Bouisset and D. Calmet, *Appl. Radiat. Isot.*, 2001, 55(2), 161–165.

- 308 W. Hang, L. Zhu, W. Zhong and C. Mahan, *J. Anal. At. Spectrom.*, 2004, **19**, 966–972.
- 309 Z. Wang, J. Zheng, L. Cao, K. Tagami and S. Uchida, *Anal. Chem.*, 2016, **88**(14), 7387–7394.
- 310 H. Kurosaki, J. R. Cadieux and S. B. Clark, *J. Anal. At. Spectrom.*, 2014, **29**, 2419–2423.
- 311 A. Gourgiotis, H. Isnard, M. Aubert, E. Dupont, I. AlMahamid, G. Tiang, L. Rao, W. Lukens, P. Cassette, S. Panebianco, A. Letourneau and F. Chartier, *Int. J. Mass Spectrom.*, 2010, 291(3), 101–107.
- 312 A. Gourgiotis, H. Isnard, A. Nonell, M. Aubert,
 G. Stadelmann, E. Dupont, I. AlMahamid, G. Tiang,
 L. Rao, W. Lukens, P. Cassette, S. Panebianco,
 A. Letourneau and F. Chartier, *Talanta*, 2013, 106, 39–44.