

ChemComm

Chemical Communications

rsc.li/chemcomm



ISSN 1359-7345



ROYAL SOCIETY
OF CHEMISTRY

Celebrating
IYPT 2019

COMMUNICATION

Normand Voyer *et al.*

Total synthesis and antimalarial activity of mortiamides A–D





Total synthesis and antimalarial activity of mortiamides A–D†

Cite this: *Chem. Commun.*, 2019, 55, 7434

Received 15th April 2019,
Accepted 18th May 2019

DOI: 10.1039/c9cc02864a

rsc.li/chemcomm

Christopher Bérubé,^a Dominic Gagnon,^b Alexandre Borgia,^a Dave Richard ^b and
Normand Voyer ^{*,a}

Mortiamides A–D (1–4) are head-to-tail cyclic heptapeptides that were identified from a novel *Mortierella* sp. isolate obtained from marine sediments from Northern Canada. Herein we report the first total synthesis of mortiamides A–D (1–4) on a solid support by concomitant cyclization/cleavage without any oligomerization side reactions, and overall yields up to 48%. We also report on the antiplasmodial activity of mortiamides A–D (1–4). We show that three out of the four tested mortiamides (A, B and D) have moderate antiplasmodial activity, while mortiamide D (4) exhibits low micro-molar activity.

Malaria is one of the most serious infectious diseases worldwide. In 2016, the World Health Organization estimated about 216 million cases of malaria in 91 countries. There were five million new cases observed in 2015 alone. Among the five *Plasmodium* parasite species that cause malaria in humans, two of them, *P. falciparum* and *P. vivax*, are responsible for about half a million deaths per year. The increasing resistance of *Plasmodium* species to drug therapies has created an urgent need to develop new antimalarial agents. Natural products have been a source of amazing drugs for efficient treatment of a variety of diseases and illnesses, including malaria. The planet's biodiversity has long been recognized as an abundant drug cabinet, offering medicinal chemists an immense range of molecular structures serving as inspiring sources for drug discovery.¹ From very small structures, such as urea, to complex molecules like taxol, natural products originate from a wide range of organisms. Though tropical plants have been long known as a source for medicinal compounds, organisms from extreme ecosystems such as the polar regions, are largely unexplored for the discovery of new metabolites with useful

bioactivities.² Northern ecosystems' severe environmental conditions force living organisms to adapt to face high UV radiation, low temperatures, low nutrient inputs, and other stresses. In addition, deep sea organisms are subjected to high pressure, low oxygen levels, and the absence of light.³ These factors contribute to gene modifications and to other adaptive processes producing unique secondary metabolites. Notably, arctic fungi have been underexplored so far. Along this line, Kerr's group isolated recently such species from deep sea sediments from Frobisher Bay in Northern Canada.⁴ BlastN investigation permitted researchers to place this isolate within the genus *Mortierella* based on a nucleotide sequence that has 96% similarity with *Mortierella antarctica*. To date, though nearly 100 *Mortierella* species are known,⁵ few natural products have been isolated from them (Fig. 1).^{4b,6} Interestingly, Kerr's group identified from this *Mortierella* species four natural cyclic heptapeptides mortiamides A to D (1–4) (Fig. 1).

Analysis of the structures of mortiamides A to D (1–4), composed of D/L hydrophobic amino acids, revealed that they have similarities with natural products having antiplasmodial activity. Indeed, several natural head-to-tail macrocyclic peptides exhibit antiplasmodial activity.⁷ Among those, several have seven to eight L- α -amino acids, of which only one is polar.⁸ For example, mahafacyclin B, a hydrophobic cyclic heptapeptide containing four L-phenylalanines, two glycines, and one threonine shows good antimalarial activity (2.2 μ M).^{7c,9}

On that basis, we sought to investigate the antiplasmodial potential of mortiamides A to D (1–4). In order to do so, we have developed an efficient synthetic methodology on a solid support which, for the first time, quickly produces the desired cyclic heptapeptides in sufficient quantities for biological investigations.

Since the amino acid sequence influences the oligomerization/cyclization process, we examined the primary sequence of mortiamides A to D (1–4) to identify potential linear precursors.

Those cyclic heptapeptides are characterized by a 21-membered peptide ring containing L- and D-branched and aromatic amino acids. The absence of β -turn-inducing proline and flexible glycine

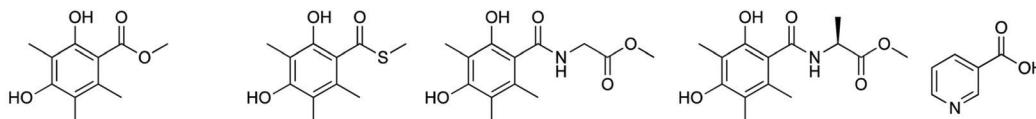
^a Département de Chimie and PROTEO, Université Laval, Faculté des sciences et de génie, 1045 avenue de la Médecine, Québec, QC, G1V 0A6, Canada.

E-mail: normand.voyer@chm.ulaval.ca

^b Département de microbiologie-infectiologie et d'immunologie, Université Laval, Québec, G1V 0A6, Canada

† Electronic supplementary information (ESI) available: Synthetic procedures, mortiamides characterization. See DOI: 10.1039/c9cc02864a

Non-peptidic natural products



methyl 2,4-dihydroxy-3,5,6-trimethylbenzoate

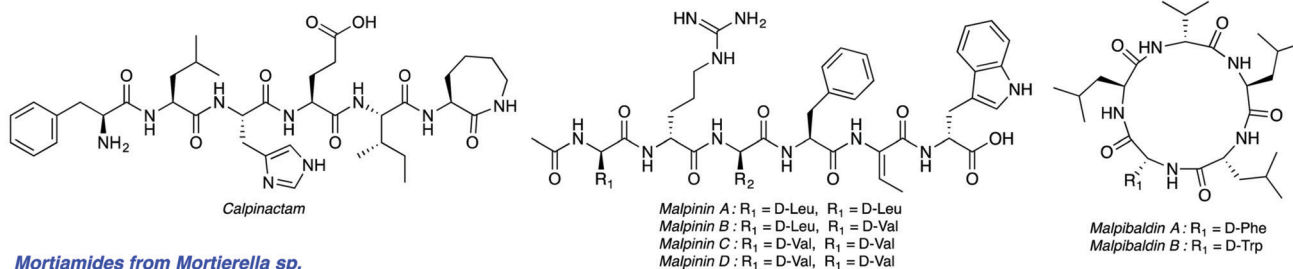
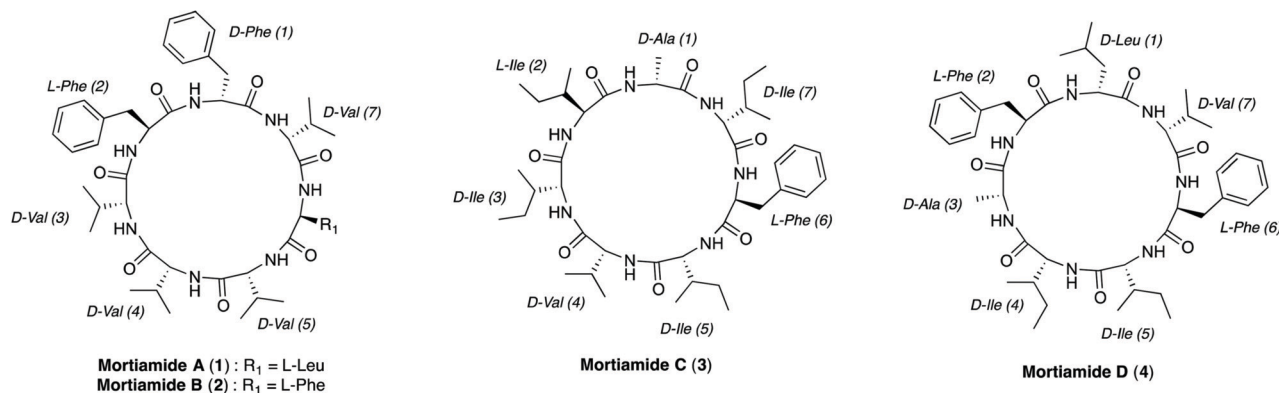
Mortivinacin A

Mortivinacin B

Mortivinacin C

Nicotinic acid

Peptide natural products

Mortiamides from *Mortierella* sp.Fig. 1 Structure of natural products isolated from *Mortierella* species.

in the sequence makes it more challenging to avoid oligomerization side reactions during cyclization reaction. In addition, mortiamides A to D (1–4) contain five D-amino acids at position 1, 3, 4, 5, and 7, of which three are contiguous. Positions 1 to 4 contain amino acids with alternate configurations, whereas sites 2 and 6 contain L-amino acids. Moreover, the primary sequence of compounds 1–4 contains seven hydrophobic amino acids. Consequently, the on-resin linear precursor could form extended strongly aggregating chains, decreasing the probability of cyclization occurring.¹⁰ Inspired by precedents in the literature¹¹ that use oxime resin¹² in peptide macrocyclization, we selected a linear precursor to facilitate the peptide folding and cyclization to obtain mortiamides A to D (1–4) by an on-resin acid-catalyzed head-to-tail concomitant cyclization/cleavage. Examples of sequence-dependent cyclization/cleavages that favour the intramolecular process on oxime resin have been described.¹³ In any macrocyclization process, it is always challenging to avoid undesired oligomerization and epimerization side reactions. However, anchoring the linear peptide on a solid support can reduce the rate of diffusion, thus decreasing the probability of dimerization and oligomerization (pseudodilution effect).¹⁴ Hence, we chose to incorporate a D-amino acid (site 1) at the anchor point with the objective to facilitate peptide folding by inserting L-amino acids at positions 2 and 6. Interestingly, incorporating alternating D- and L-amino

acids in linear primary sequences causes turn-inducing effects and has been employed to improve the yield of peptide cyclizations.¹⁵ With the above considerations in mind, mortiamides A to D (1–4) were synthesized on oxime resin using standard protocols with a 0.43 mmol g^{−1} loading rate (Fig. 2).¹⁶ Briefly, the first amino acid was coupled for 3 h using DIC as a coupling reagent. The N-Boc protecting group was removed using a mixture of 1:1 trifluoroacetic acid (TFA)/dichloromethane, while the second amino acid was activated with hydroxybenzotriazole (6-Cl-HOBT) and 1-[bis(dimethylamino)methylene]-1H-1,2,3-triazolo[4,5-b]pyridinium 3-oxide hexafluorophosphate (HCTU). After completion of the linear precursor on the oxime resin, we proceeded with the key step of the synthesis. The linear peptide was simultaneously cyclized and cleaved from the resin in the presence of diisopropylethylamine (DIEA; 2.5 equiv.) and acetic acid (AcOH; 5 equiv.) in dichloromethane in a respectable 10^{−2} M cyclization concentration (Fig. 2), leading to very good macrocyclization yields of 1–4 of 35% to 48% without any oligomerization side reactions (>99% of cyclic monomers).

Taking advantage of this powerful synthetic methodology that yields the cyclic monomer almost exclusively, simple triturations with cold ether gave pure mortiamides 1–4. Spectroscopic data of synthetic mortiamides A–D (1–4) matched entirely those reported for the isolated natural product, taking



Fig. 2 Mortiamides A to D synthetic procedures.

into account the possible conformers equilibrium (see ESI† for details).

In order to investigate whether mortiamides A to D (1–4) exhibit an inhibitory effect on *P. falciparum* proliferation, *in vitro* susceptibility assays were conducted and their IC₅₀ were determined (Fig. 3 and 4). Assays were performed on the commonly used lab strain 3D7 and on the chloroquine-mefloquine-pyrimethamine multiresistant strain Dd2.¹⁷ The mortiamide A (1) IC₅₀ with 3D7 and Dd2 were 7.85 ± 0.97 μM (*n* = 4) and 5.31 ± 0.24 μM (*n* = 5) respectively. The mortiamide B (2) IC₅₀ with 3D7 and Dd2 values were 3.16 ± 0.65 μM (*n* = 4) and 2.10 ± 0.18 μM (*n* = 4) respectively. The IC₅₀ value for mortiamide C (3) could not be determined as the molecule was not effective enough at the tested concentrations. The mortiamide D IC₅₀ with 3D7 and Dd2 were 1.31 ± 0.12 μM (*n* = 11) and 0.94 ± 0.07 μM (*n* = 13)

respectively (Fig. 4). To ensure that the observed growth inhibition effect was not due to red blood cell (RBC) lysis but to a biological effect on the parasite itself, hemolysis assays were performed with mortiamides A, B, and D. The absorbance reading at 405 nm after 72 h of incubation revealed no increase of hemoglobin release in the culture medium supernatant for mortiamides A and B at all tested concentrations (0.78 nM to 25.6 μM) when compared to untreated RBCs. Mortiamide D (4) only showed a mild RBC lysis effect of 19.7% at 25.6 μM and was reduced to 4.4% at 12.8 μM when compared to the untreated sample. Those concentrations are much higher than the IC₅₀ values obtained with mortiamide D (4) proving its activity against the parasite.

We achieved the first total synthesis of mortiamides A to D (1–4) on a solid support by concomitant cyclization/cleavage process without any oligomerization side reactions, and with impressive overall yield of 35 to 48%. Access to a sufficient amount of mortiamides allowed us to perform the first antiparasitological investigation on those natural macrocyclic peptides. Results revealed that mortiamides A and B (1 and 2) have a moderate antiparasitological activity, while mortiamide D (4) exhibits low micromolar activity. Interestingly, the results demonstrate that small variations on the side chains significantly impact the activity. Although the mechanism of action is not known yet, the results reported can be used to prepare analogs of mortiamide D, to improve its potency against *P. falciparum*.

This work was supported by the NSERC of Canada, the FRQNT of Quebec, PROTEO, Sentinelle Nord and Université Laval. CB wishes to thank PROTEO, FRQNT and NSERC for

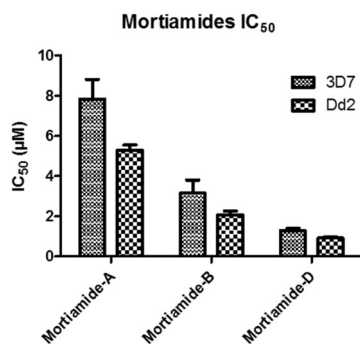


Fig. 3 IC₅₀ of mortiamides A, B, and D (1, 2, and 4).

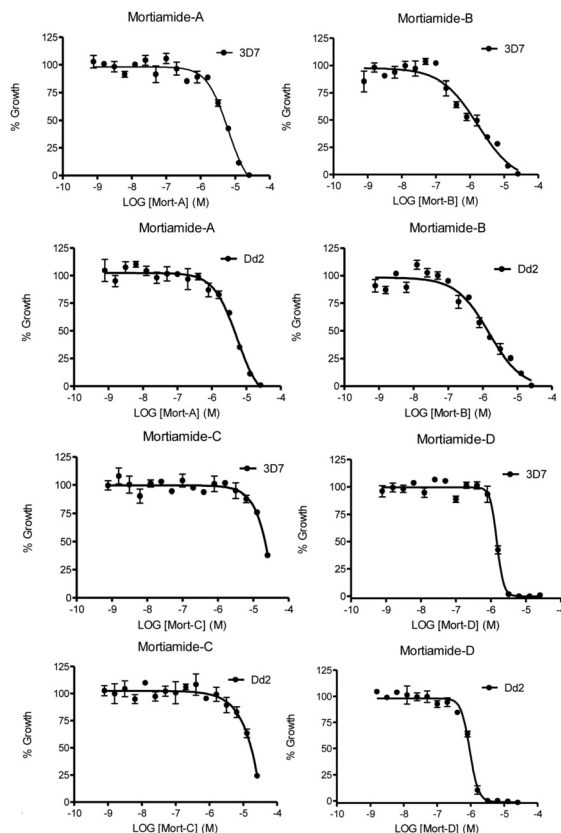


Fig. 4 *In vitro* 72 h viability assays. Shown here are representative assays for both strains treated with each mortiamide.

postgraduate scholarships. AB thanks the NSERC of Canada for a USRA scholarship. Work in Richard's lab was supported by a Canadian Institutes for Health Research Operating grant (MOP 130359). DR is a Fonds de la recherche du Québec-Santé Junior 2 Fellow. The authors are also thankful to Pierre Audet and François Otis for useful advices in mass spectrometry and HPLC analyses, respectively.

Conflicts of interest

There are no conflicts to declare.

Notes and references

- (a) T. Rodrigues, D. Reker, P. Schneider and G. Schneider, *Nat. Chem.*, 2016, **8**, 531; (b) D. J. Newman and G. M. Cragg, *J. Nat. Prod.*, 2016, **79**, 629.
- (a) Y. Tian, Y. Li and F. Zhao, *Mar. Drugs*, 2017, **15**, 28; (b) S. Abbas, M. Kelly, J. Bowling, J. Sims, A. Waters and M. Hamann, *Mar. Drugs*, 2011, **9**, 2423; (c) Z. E. Wilson and M. A. Brimble, *Nat. Prod. Rep.*, 2009, **26**, 44.
- D. Skropeta and L. Wei, *Nat. Prod. Rep.*, 2014, **31**, 999.
- (a) A. W. Robertson, N. G. McCarville, L. W. MacIntyre, H. Correa, B. Haltli, D. H. Marchbank and R. G. Kerr, *J. Nat. Prod.*, 2018, **81**, 858; (b) A. L. Grunwald, F. Berrue, A. W. Robertson, D. P. Overy and R. G. Kerr, *J. Nat. Prod.*, 2017, **80**, 2677.
- L. Wagner, B. Stielow, K. Hoffmann, T. Petkovits, T. Papp, C. Vagvolgyi, G. de Hoog, G. Verkley and K. Voigt, *Persoonia*, 2013, **30**, 77.
- (a) K. Nobuhiro, K. Shigenobu, N. Kenichi, M. Rokuro, M. Makoto, O. Satoshi and T. Hiroshi, *J. Antibiot.*, 2010, **63**, 183; (b) A. G. Soman, J. Gloer and D. Wicklow, *J. Nat. Prod.*, 1999, **62**, 386; (c) F. Baldeweg, P. Warncke, D. Fischer and M. Gressler, *Org. Lett.*, 2019, **21**, 1444.
- (a) A. Lawer, J. Tai, K. A. Jolliffe, S. Fletcher, V. M. Avery and L. Hunter, *Bioorg. Med. Chem. Lett.*, 2014, **24**, 2645; (b) P. Barbie and U. Kazmaier, *Org. Biomol. Chem.*, 2016, **14**, 6036; (c) C. Baraguey, A. Blond, F. Cavelier, J.-L. Pousset, B. Bodo and C. Auvin-Guette, *J. Chem. Soc., Perkin Trans. 1*, 2001, 2098, DOI: 10.1039/B008538N; (d) V. Rukachaisirikul, S. Chantaruk, C. Tansakul, S. Saithong, L. Chaicharemwimonkoon, C. Pakawatchai, M. Isaka and K. Intereya, *J. Nat. Prod.*, 2006, **69**, 305; (e) C. Auvin-Guette, C. Baraguey, A. Blond, H. S. Xavier, J.-L. Pousset and B. Bodo, *Tetrahedron*, 1999, **55**, 11495; (f) C. Baraguey, C. Auvin-Guette, A. Blond, F. Cavelier, F. Lezenven, J.-L. Pousset and B. Bodo, *J. Chem. Soc., Perkin Trans. 1*, 1998, 3033, DOI: 10.1039/A804003F; (g) C. H. M. Kocken, A. van der Wel, B. Rosenwirth and A. W. Thomas, *Exp. Parasitol.*, 1996, **84**, 439; (h) M. E. F. Pinto, J. M. Batista, J. Koehbach, P. Gaur, A. Sharma, M. Nakabashi, E. M. Cilli, G. M. Giesel, H. Verli, C. W. Gruber, E. W. Blanch, J. F. Tavares, M. S. D. Silva, C. R. S. Garcia and V. S. Bolzani, *J. Nat. Prod.*, 2015, **78**, 374; (i) C. Auvin, C. Baraguey, A. Blond, F. Lezenven, J.-L. Pousset and B. Bodo, *Tetrahedron Lett.*, 1997, **38**, 2845; (j) C. Auvin-Guette, C. Baraguey, A. Blond, J.-L. Pousset and B. Bodo, *J. Nat. Prod.*, 1997, **60**, 1155; (k) C. Baraguey, A. Blond, I. Correia, J.-L. Pousset, B. Bodo and C. Auvin-Guette, *Tetrahedron Lett.*, 2000, **41**, 325.
- S. Ramalho, M. Pinto, D. Ferreira and V. Bolzani, *Planta Med.*, 2018, **84**, 558.
- A. J. J. van den Berg, S. F. A. J. Horsten, J. J. K.-V. Bosch, B. H. Kroes, C. J. Beukelman, B. R. Leeftang and R. P. Labadie, *FEBS Lett.*, 1995, **358**, 215.
- E. T. Kaiser, *Acc. Chem. Res.*, 1989, **22**, 47.
- (a) A. K. Yudin, *Chem. Sci.*, 2015, **6**, 30; (b) C. J. White and A. K. Yudin, *Nat. Chem.*, 2011, **3**, 509; (c) J. S. Davies, *J. Pept. Sci.*, 2003, **9**, 471; (d) J. M. Humphrey and A. R. Chamberlin, *Chem. Rev.*, 1997, **97**, 2243.
- (a) W. F. DeGrado and E. T. Kaiser, *J. Org. Chem.*, 1980, **45**, 1295; (b) W. F. DeGrado and E. T. Kaiser, *J. Org. Chem.*, 1982, **47**, 3258.
- (a) N. Nishino, M. Xu, H. Mihara and T. Fujimoto, *Bull. Chem. Soc. Jpn.*, 1992, **65**, 991; (b) N. Nishino, M. Xu, H. Mihara, T. Fujimoto, Y. Ueno and H. Kumagai, *Tetrahedron Lett.*, 1992, **33**, 1479.
- (a) L. T. Scott, J. Rebek, L. Ovsyanko and C. L. Sims, *J. Am. Chem. Soc.*, 1977, **99**, 625; (b) S. Mazur and P. Jayalekshmy, *J. Am. Chem. Soc.*, 1979, **101**, 677.
- (a) H. Kessler and B. Haase, *Int. J. Pept. Protein Res.*, 1992, **39**, 36; (b) M. Tamaki, S. Akabori and I. Muramatsu, *J. Am. Chem. Soc.*, 1993, **115**, 10492.
- (a) C. Bérubé, A. Borgia and N. Voyer, *Org. Biomol. Chem.*, 2018, **16**, 9117; (b) C. Bérubé, A. Borgia and N. Voyer, *Tetrahedron Lett.*, 2018, **59**, 4176; (c) C. Bérubé, A. Borgia, G. Simon, D. Grenier and N. Voyer, *Phytochem. Lett.*, 2018, **26**, 101.
- (a) G. Françoise, A. D. James, F. Hisashi, B. K. David, M. Olga, C. K. David, A. Masamichi, B. V. Akhil and E. W. Thomas, *J. Cell Biol.*, 1996, **135**, 269; (b) X.-Z. Su, M. T. Ferdig, Y. Huang, C. Q. Huynh, A. Liu, J. You, J. C. Wootton and T. E. Wellems, *Science*, 1999, **286**, 1351.