



Cite this: *Analyst*, 2018, **143**, 175

An ionic liquid-magnetic graphene composite for magnet dispersive solid-phase extraction of triazine herbicides in surface water followed by high performance liquid chromatography

Hua Zhang,^a Yanan Yuan,^a Yunyun Sun,^a Can Niu,^{*a} Fengxia Qiao^b and Hongyuan Yan ^{*a}

A new ionic liquid-magnetic graphene (IL-MG) composite was used as the adsorbent in magnetic dispersive solid-phase extraction to rapidly extract and isolate triazine herbicides from surface water. IL-MG was synthesized by a simple and time-saving one-pot strategy where the synthesis of magnetic Fe₃O₄, the modification with an IL, and the reduction of graphene oxide to graphene were conducted at the same time. An IL was applied to enrich the interaction mechanism between IL-MG and analytes (π - π , hydrophobic interaction, and electrostatic interaction). Moreover, the IL and Fe₃O₄ nanoparticles acted as spacers, inserting between the layers of graphene to prevent the aggregation of graphene, which improved the adsorption ability because of the large specific surface area of IL-MG. The resultant IL-MG had hierarchical flake structures and showed a high adsorption capacity (8266.0–12 324.1 $\mu\text{g g}^{-1}$) toward triazine herbicides. Under suitable conditions, the linearity for triazine herbicides was achieved in the range of 0.55–500 ng mL^{-1} with a detection limit of 0.09–0.15 ng mL^{-1} and a quantitation limit of 0.31–0.51 ng mL^{-1} , and the enrichment factor was 83-fold, which indicated that the proposed method could be successfully applied for the determination of triazine herbicides in surface water.

Received 3rd August 2017,
Accepted 1st November 2017

DOI: 10.1039/c7an01290j

rs.c.li/analyst

Introduction

Triazine herbicides, one common type of herbicide with high activity to inhibit photosynthesis in plants or the germination of weeds, are widely used in agriculture to protect crops. The extensive use of triazine herbicides has led to their presence in the environment, such as surface water, soil, and so on.^{1,2} In view of their persistence, triazine herbicides and their metabolites can migrate from the environment into food chains, resulting in potential harm to human health.^{3,4} Some countries and regions have legislated to limit the levels of triazine residues. However, the amounts of the residues often exceed these levels because of insufficient control or monitoring. Therefore, it is necessary to develop a simple and effective method to detect triazine herbicides in environmental samples.⁵

Several methods have been developed for determining triazine herbicides in various environmental matrices, including liquid chromatography,⁶ liquid chromatography-mass spec-

trometry,⁷ gas chromatography-mass spectrometry,⁸ and capillary electrophoresis.⁹ However, due to the low levels of analytes, sample pretreatment and enrichment processes were the most complex and crucial steps in the analytical procedures. Until now, the pretreatment methods associated with the isolation of triazine herbicides have mainly included solid-phase extraction (SPE),^{10–12} dispersive liquid-liquid microextraction,¹³ solid-phase microextraction (SPME),¹⁴ dispersant-assisted dynamic microwave extraction (DA-DME),¹⁵ and stir bar sorptive extraction.¹⁶ Among these methods, SPE is one of the most widely applied pretreatment techniques because of its high recovery, reproducibility, and simple operation. However, most of the SPE procedures are tedious, laborious, and time consuming, so the exploration of a simple and time-saving pattern of SPE is imperative.

As a new mode of SPE, magnetic dispersive solid-phase extraction (MDSPE) is performed under an external magnetic field without tedious centrifugation or filtration procedures.^{17–19} Magnetic adsorbents dispersed into the sample solution can increase the contact interface with analytes significantly, and this was beneficial to facilitate the mass transfer of analytes. Moreover, magnetic adsorbents can be easily retrieved, which makes the sample pretreatment procedure more convenient, time-saving, and economical.²⁰

^aKey Laboratory of Medicinal Chemistry and Molecular Diagnosis, Ministry of Education & Hebei University, Baoding, 071002, China

^bDepartment of Biochemistry, Baoding University, Baoding, 071000, China.
E-mail: niucanwo@126.com, yanhy@hbu.edu.cn; Tel: +86-312-5079788

Graphene has a two-dimensional single-atom sheet structure and a high specific surface area, which make it possess excellent adsorption ability and become a promising candidate as an adsorbent.^{21–23} There has been an increasing application of magnetic graphene in MDSPE,^{24–26} because the composite materials combine the merits of magnetic adsorbents and graphene. However, graphene materials exhibit a tendency to agglomerate and have a single adsorption mechanism, indicating the limitation of their practical applications. Ionic liquids (ILs), composed of an organic cation and an organic or inorganic anion, exhibit beneficial characteristics, such as high chemical and thermal stability, negligible vapor pressure, non-flammability, and high ionic conductivity.^{27–29} Recently, an IL was introduced in the synthesis process of magnetic graphene to improve the adsorption capacity of the obtained materials and prevent aggregation, because the adsorption mechanism of IL-based materials usually involves multiple interactions, including electrostatic, π - π , hydrogen bonding, and ion exchange interactions, and the IL can enhance the dispersion stability of prepared materials in aqueous solution.^{30–32} However, the synthesis of magnetic graphene oxide and its subsequent modification are always completed step by step, which are complex and time consuming. Therefore, an easy and time-saving preparation strategy is yet to be developed.

In this work, a new IL-MG was synthesized by a simple and time-saving one-pot strategy where the synthesis of magnetic Fe₃O₄, the modification with an IL, and the reduction of graphene oxide (GO) to graphene were conducted at the same time to provide multiple binding sites. The IL-MG has hierarchical flake structures because the IL and Fe₃O₄ nanoparticles can act as spacers inserting between the layers of graphene and effectively enlarge the layer spacing. The proposed IL-MG-MDSPE method combines the excellent adsorption ability of IL-MG and the rapid extraction efficiency of MDSPE, which was successfully applied to rapidly isolate and analyze triazine herbicides from surface water.

Experimental

Chemicals and reagents

Cyanazine, ametryn and atrazine were purchased from Sigma-Aldrich Co., Ltd (St Louis, USA). 1-(3-Aminopropyl)-3-methylimidazolium chloride hydrochloride (ApMeIm⁺Cl⁻) was purchased from Shanghai Chengjie Chemical Co. Ltd (Shanghai, China). Graphite powder (500 mesh, 99.95%), iron(III) chloride hexahydrate (FeCl₃·6H₂O), iron(II) chloride tetrahydrate (FeCl₂·4H₂O), potassium hydroxide (KOH), potassium permanganate (KMnO₄), hydrochloric acid (HCl), and hydrogen peroxide (H₂O₂) were purchased from Huadong Chemical Reagent Co. Ltd (Tianjin, China). Methanol, acetonitrile, acetone and trifluoroacetic acid (TFA) were obtained from Kermel Chemical Co., Ltd (Tianjin, China). Methanol (HPLC grade) was obtained from Xingke Biochemistry Co., Ltd (Shanghai, China). The water used was double-deionized and filtered with a 0.45 μ m filter membrane.

Instruments and conditions

HPLC analysis was performed by using an UltiMate 3000 equipped with a VWD-3100 UV-VIS detector (Thermo Fisher Scientific, Massachusetts, USA). The morphological evaluation was performed by using a Phenom Pro scanning electron microscope (SEM) (Phenom, Eindhoven, Netherland). The specific surface area was evaluated by using a TriStar II 3020 pore size and surface area analyzer (Micromeritics, Norcross, USA). The freeze dryer was purchased from SIM International Group (California, USA). The detection wavelength of the detector was set at 224 nm. The analytical column was purchased from Welch Materials, Inc. (Ultimate AQ-C₁₈, 4.6 mm \times 250 mm, 5 μ m). The mobile phase was water-methanol (4 : 6, v/v, containing 0.5% TFA) with a flow rate of 1.0 mL min⁻¹.

Synthesis of IL-MG composite

Graphite powder (1.00 g) was added into 45 mL of H₂SO₄ with stirring in an ice-bath for 30 min. After dispersing sufficiently, 3.00 g of KMnO₄ was added gradually, and the mixture was kept stirring for 24 h at room temperature. After that, 60 mL of water was slowly added in the ice-bath with stirring for 1 h. Then 3 mL of H₂O₂ was added to the mixture with stirring for 1 h. Finally, the mixture was washed with 5% HCl to remove metal ions, followed by washing with water to achieve pH 7, and graphite oxide was obtained.

Graphite oxide (0.40 g) was dissolved in 320 mL of water under ultrasonication for 3 h to form a homogeneous dispersion. Then, ApMeIm⁺Cl⁻ (1.60 g) was added and the mixture was sonicated for 20 min followed by adding FeCl₂·4H₂O (0.56 g) and FeCl₃·6H₂O (1.50 g). Then KOH (1.40 g) was dissolved in 2 mL of water and the solution was added dropwise to the above mixture solution, and then the mixture solution was stirred at 700 rpm for 24 h (80 °C). Subsequently, the obtained IL-MG was washed with water to achieve pH 7 and freeze-dried under vacuum.

Rebinding experiment: for static adsorption experiment, IL-MG (10 mg) was dispersed in 10 mL of mixed solution with a concentration of 1–20 μ g mL⁻¹. After being shaken for 24 h, the supernatant solution was collected by using a magnet and then filtered through a 0.45 μ m filter membrane. The concentration of the supernatant solution was determined to calculate the adsorption capacity of IL-MG. For dynamic adsorption experiment, six parts of IL-MG (10 mg) were dispersed in 10 mL of mixed solution with a concentration of 20 μ g mL⁻¹, respectively. After being shaken for 1–12 h, the supernatant solution was collected at different times and filtered through a 0.45 μ m filter membrane. The concentration of the supernatant solution was detected to evaluate the adsorption capacity of IL-MG.

Procedure of MDSPE

The procedure of MDSPE is illustrated in Fig. 1a. IL-MG (40 mg) was dispersed in a 100 mL conical flask and 50 mL of sample aqueous solution was added. After vortexing for 15 min, IL-MG was isolated with a magnet and then the supernatant solution was collected. Elution was performed with



Fig. 1 Procedure of MDSPE (a), and one-pot synthesis of IL-MG (b).

acetone (6 mL) by vortexing for 5 min. Subsequently, the eluate was collected and evaporated to dryness at 40 °C. Finally, the residues were re-dissolved with 0.6 mL of water-methanol (1 : 5, v/v) for further HPLC analysis.

Results and discussion

Synthesis of IL-MG composite

As shown in Fig. 1b, one-pot synthesis of IL-MG can avoid the tedious process where the synthesis of magnetic Fe₃O₄, the modification with an IL, and the reduction of GO to graphene were conducted simultaneously in the presence of KOH. The whole operation was easy and time-saving. The surface of GO possesses abundant active epoxy groups, which can undergo a ring-opening addition reaction with the amino groups (–NH₂) of ApMeIm⁺Cl[–] under catalysis by KOH.^{33,34} As a result, Fe₃O₄ nanoparticles were doped in graphene sheets by the *in situ* ultrasonic-assisted co-precipitation of iron ions in alkaline solution in the presence of GO, and the IL was incorporated to the surface of graphene by covalent binding, not only enhancing the dispersion stability of the prepared materials in aqueous solution, but also improving the affinity with the analytes by providing multiple adsorption mechanism such as π–π interaction, electrostatic interaction, and ion exchange interaction. The IL and Fe₃O₄ nanoparticles were also able to alleviate the aggregation of graphene, because they can attach to graphene sheets serving as the spacer that effectively enlarged the layer spacing and significantly enhanced the extraction efficiency of the graphene-based composites.

The effects of the amount of IL and Fe₃O₄ nanoparticles on the adsorption capacity were evaluated by performing MDSPE. Briefly, IL-MG (10 mg) was dispersed in 10 mL of mixed solution (atrazine, ametryn, and cyanazine) with a concentration of 25.0 μg mL^{–1}. The supernatant solution was collected after vortexing (10 min) for HPLC analysis to detect the adsorption amount of the analytes. For IL-MG, the amount of IL is a key parameter to increase the adsorption capability. Fig. 2a shows that along with the increasing amount of IL, the adsorption capacity of the triazine herbicides increased and then decreased when the amount of IL was more than 1.60 g. The

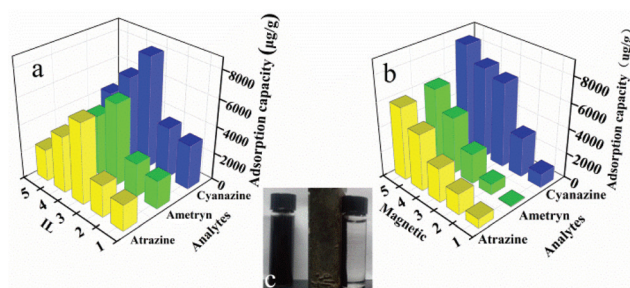


Fig. 2 Effect of the amount of IL (a) and magnetic Fe₃O₄ (b). The magnetic separation behavior of IL-MG (c). (a) 1: 0 g; 2: 0.32 g; 3: 1.60 g; 4: 8.00 g; 5: 32.00 g and b 1: 9.04 g FeCl₂·4H₂O + 24.00 g FeCl₃·6H₂O; 2: 4.52 g FeCl₂·4H₂O + 12.00 g FeCl₃·6H₂O; 3: 2.26 g FeCl₂·4H₂O + 6.00 g FeCl₃·6H₂O; 4: 1.13 g FeCl₂·4H₂O + 3.00 g FeCl₃·6H₂O; 5: 0.56 g FeCl₂·4H₂O + 1.50 g FeCl₃·6H₂O).

reason may be that the low amount of IL increased the dispersion stability of the prepared materials in aqueous solution for avoiding aggregation, but the excessive IL induced an increase in viscosity which was not conducive for the dispersion of graphene, therefore leading to the aggregation of graphene and affecting the interaction between IL-MG and analytes. In Fig. 2b, the function of Fe₃O₄ is presented. With the increase of magnetic quantities, the adsorption ability gradually weakened. The reason may be that excess amounts of Fe₃O₄ would cover the graphene sheets, reducing the surface area of graphene and affecting the adsorption property of IL-MG. However, if the amount of Fe₃O₄ is insufficient, the obtained magnetic intensity is weak, which makes no contribution to the rapid separation of the materials under an external magnetic field. The results indicated that FeCl₂·4H₂O (0.56 g), FeCl₃·6H₂O (1.50 g) and IL (1.60 g) were the optimal amounts to obtain the best extraction efficiency for IL-MG. Besides, Fig. 2c clearly shows that the well-dispersed IL-MG could be collected under a magnetic field within 60 s and redispersed quickly with a slight shake once the external magnetic field was removed. The excellent magnetic redispersibility made the IL-MG a promising candidate for practical applications in the MDSPE field.

Characterization of IL-MG and MG

The morphologies of IL-MG and MG were investigated by SEM, and the results are shown in Fig. 3. The SEM image of MG showed a compact block structure (Fig. 3b), while the introduction of the IL enabled IL-MG to obtain a porous structure (Fig. 3a). Besides, the hierarchical architecture of the thin layers of graphene can be observed in Fig. 3c. The SEM images illustrated that the IL existing between the graphene sheets effectively increased the dispersion stability of graphene, which contributed to the prevention of agglomeration. Meanwhile, the specific surface areas of IL-MG and MG were determined using the nitrogen adsorption test (BET). The specific surface areas of IL-MG and MG were 227 m² g^{–1} and 131 m² g^{–1}, respectively. The IL can avoid the aggregation of graphene and increase the basal space between graphene sheets, so the specific surface

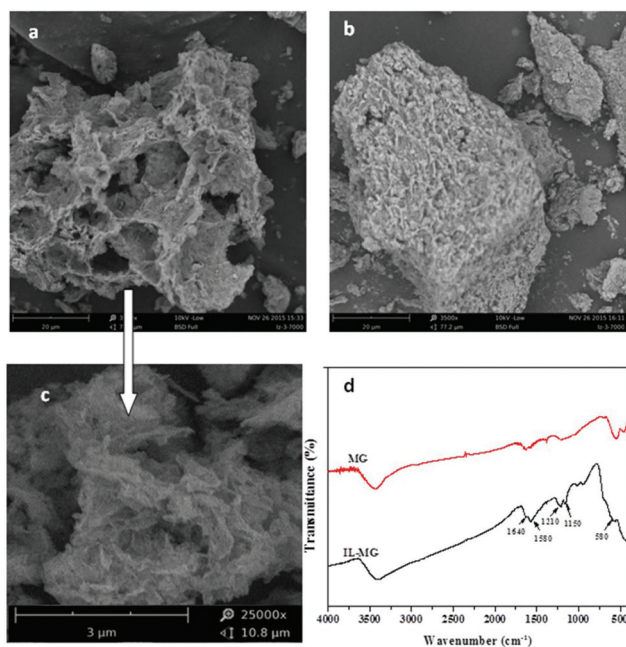


Fig. 3 Scanning electron micrograph of IL-MG (a, c), MG (b) (a: $\times 3500$; b: $\times 3500$; c: $\times 25\,000$) and FT-IR spectra of MG and IL-MG (d).

area of IL-MG was larger than MG, improving the adsorption property of IL-MG. Fig. 3d shows the FT-IR spectra of MG and IL-MG. The adsorption peak at 580 cm^{-1} belonged to the Fe-O vibration, which implied that the Fe_3O_4 nanoparticles were successfully attached to IL-MG. The bands for the imidazole ring (1150 cm^{-1}), N-H (1210 cm^{-1}), and C-N (1580 cm^{-1}) of the IL were observed, confirming the successful introduction of the IL into IL-MG.

Adsorption characteristics of IL-MG

To demonstrate the adsorption ability of IL-MG, static adsorption experiment and dynamic adsorption experiment were conducted. The results of the static adsorption experiment (Fig. 4a) showed that the adsorption capacity of triazine herbi-

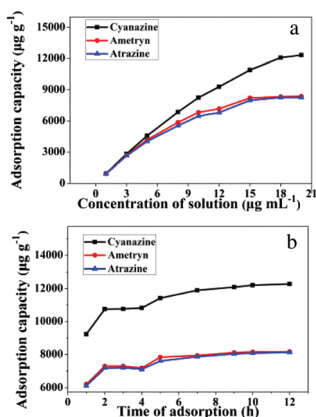


Fig. 4 Binding isotherm of triazines on IL-MG. (a: static adsorption; b: dynamic adsorption).

cides increased along with the increasing concentration of analytes, and the adsorption capacity tended to saturate when the concentration of triazine herbicides was larger than $18.0\text{ }\mu\text{g mL}^{-1}$. The adsorption amounts of cyanazine, ametryn and atrazine were $12\,324.1$, 8374.7 and $8266.0\text{ }\mu\text{g g}^{-1}$, respectively. The results of dynamic adsorption (Fig. 4b) showed that the adsorption capacity slightly increased after 2 h and tended to balance at approximately 7 h. Furthermore, two models Langmuir and Freundlich were used to analyze the equilibrium adsorption data. The Langmuir and Freundlich adsorption equations were calculated using eqn (1) and (2):

$$\frac{C_e}{Q_e} = \frac{C_e}{Q_m} + \frac{1}{Q_m b} \quad (1)$$

$$\ln Q_e = \frac{1}{n} \ln C_e + \ln K_F \quad (2)$$

Herein, C_e is the equilibrium concentration of the analyte in solution ($\mu\text{g mL}^{-1}$), Q_e is the adsorption amount of the adsorbent at equilibrium ($\mu\text{g mg}^{-1}$), Q_m is the maximum adsorption capacity of the adsorbent ($\mu\text{g mg}^{-1}$), b is the Langmuir adsorption equilibrium constant ($\text{mL }\mu\text{g}^{-1}$), and K_F and $1/n$ are the Freundlich characteristic constants. The parameters of Langmuir and Freundlich adsorption are shown in Table 1. The correlation coefficients for both Langmuir and Freundlich were close to 1, suggesting that the adsorption of analytes on IL-MG was both physical adsorption and chemical adsorption.³⁵ For the Langmuir adsorption equilibrium of the three analytes on the IL-MG surface, the three correlation coefficients were all larger than 0.99 indicating that the adsorption was a monolayer process. The fit of the experimental isotherm data to the Langmuir equation (0.9960–0.9982) was closer to 1 than that to the Freundlich equation (0.9710–0.9792). Therefore, the Langmuir model represented the experimental data better on the basis of the values of correlation coefficient.³⁶

Optimization of IL-MG-MDSPE method

The IL-MG possessed an aromatic ring and charged imidazole ring, so there were mainly three interactions (π - π , hydrophobic interaction, and electrostatic interaction) between triazine herbicides and IL-MG, which influenced the MDSPE process. In order to obtain optimum conditions for rapid isolation and extraction of triazine herbicides from the water sample, various parameters of MDSPE including the adsorbent amount, the adsorption and desorption time, and the kind and volume of elution solvent were investigated.

Table 1 Langmuir and Freundlich data for the adsorption of analytes on IL-MG at $25\text{ }^\circ\text{C}$

Analytes	Langmuir			Freundlich		
	Q_m (ng mL^{-1})	b	r	$1/n$	K_F	r
Cyanazine	13.63	1.08	0.9963	0.486	5.61	0.9766
Ametryn	8.98	1.16	0.9982	0.404	3.78	0.9710
Atrazine	9.01	0.89	0.9960	0.418	3.49	0.9792

Firstly, the amounts of IL-MG ranging from 10.0 to 60.0 mg were evaluated. IL-MG was dispersed in 50.0 mL of mixed standard solution (atrazine, ametryn, and cyanazine) with a concentration of 100 ng mL^{-1} . The supernatant solution was collected after vortexing (10 min) for HPLC analysis to detect the loss of the analytes. As shown in Fig. 5a, the loss rate of triazine herbicides was progressively decreased from 10.0 to 40.0 mg of the adsorbent and then remained almost constant with a further increase of the amount of IL-MG. Consequently, 40.0 mg of IL-MG was used as the optimum amount for MDSPE. The adsorption time was evaluated by changing the vortexing time (1 to 30 min). Fig. 5b shows that IL-MG has a large adsorption capacity, and extraction equilibrium was attained rapidly within 15 min, so 15 min was selected as the adsorption time for further work.

A suitable elution solvent should be utilized to elute the analytes from IL-MG and four types of elution solvents were evaluated by performing MDSPE, including methanol, acetonitrile, acetone, and chloroform. As shown in Fig. 5c, acetone can obtain the highest recovery for all three triazine herbicides. And the recoveries of the three triazine herbicides increased until the volume of acetone reached 6.0 mL and then remained unchanged. Hence, the optimal elution solvent was 6.0 mL of acetone. The effect of the desorption time on the recoveries of triazine herbicides was evaluated by changing the elution time (1 to 20 min). Fig. 5d shows that desorption equilibrium was attained rapidly within 5 min, so 5 min was selected as the desorption time for further work.

Validation of IL-MG-MDSPE method

The proposed IL-MG-MDSPE method was validated by linearity, precision, repeatability, limits of detection (LOD) and quantitation (LOQ), and intra-assay and inter-assay deviation as presented in Table 2. A calibration curve was constructed by least-squares linear regression analysis of the peak area and the concentration of triazine herbicides at seven increasing spiked levels in the range of $0.55\text{--}500 \text{ ng mL}^{-1}$ with the correlation coefficient (r) ≥ 0.9996 . Based on the signal-to-noise ratio of 3 and 10, the LOD and LOQ of the method for triazine herbicides were $0.09\text{--}0.15 \text{ ng mL}^{-1}$ and $0.31\text{--}0.51 \text{ ng mL}^{-1}$, respectively. The precision and accuracy were assessed by analyzing the spiked samples in the same day ($n = 3$) and three consecutive days ($n = 3$). The intra-assay and inter-assay precision expressed as relative standard deviations (RSDs) were 3.1–3.5% and 4.8–6.8%, respectively. To investigate the effect of the sample matrix on the accuracy of the IL-MG-MDSPE method for real samples application, recovery experiment was carried out by spiking three levels of triazine herbicides (1, 50, and 500 ng mL^{-1}) into blank water samples. The results (Table 3) demonstrated that the recovery of the IL-MG-MDSPE method was 97.0–100.8% in surface water with RSDs $\leq 3.8\%$.

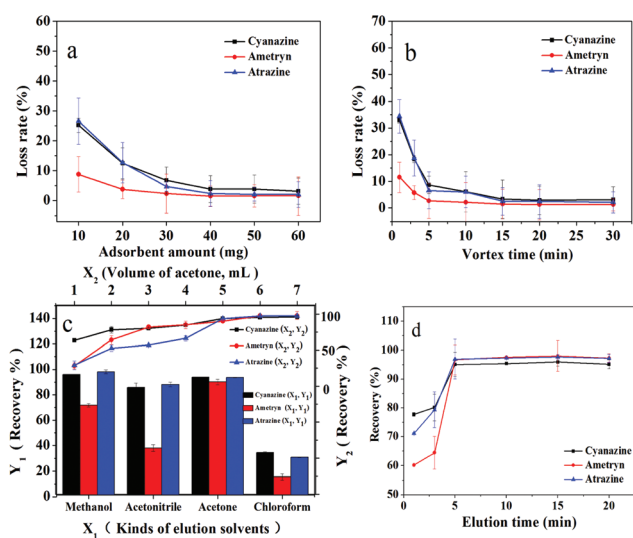


Fig. 5 Optimization of MDSPE conditions: a: adsorbent amount; b: adsorption time; c: kinds and volume of elution solvents; d: desorption time.

Table 2 Features of the IL-MG-MDSPE-HPLC method

Analytes	Regression equation	r	Linearity (ng mL^{-1})	LOD (ng mL^{-1})	LOQ (ng mL^{-1})	RSDs (%)	
						Intra-day	Inter-day
Cyanazine	$Y = 0.19x + 0.68$	0.9998	0.55–500	0.15	0.51	3.1	6.8
Ametryn	$Y = 0.21x + 0.06$	0.9998	0.55–500	0.09	0.31	3.4	5.1
Atrazine	$Y = 0.25x + 0.03$	0.9996	0.55–500	0.12	0.39	3.5	4.8

Table 3 Recoveries of cyanazine, ametryn, and atrazine from spiked water samples

Analytes	1 ng mL^{-1}		50 ng mL^{-1}		500 ng mL^{-1}	
	Recovery (%)	RSDs (%)	Recovery (%)	RSDs (%)	Recovery (%)	RSDs (%)
Cyanazine	98.8	3.2	97.0	3.1	100.8	2.0
Ametryn	97.5	3.7	100.1	3.4	99.2	1.9
Atrazine	100.1	3.8	100.7	3.5	99.8	1.6

Table 4 Comparison of IL-MG-MDSPE-HPLC with other methods

Pretreatment method	Adsorbent	Linearity	LOD	Recovery (%)	RSD (%)	Ref.
SPE	Multiwalled carbon nanotubes	0.50–80.0 ng mL ⁻¹	0.42–0.47 ng mL ⁻¹	88.0–124.0	7.6	8
SPME	Ametryn-imprinted polymer	100–5000 ng mL ⁻¹	14–74 ng mL ⁻¹	93.6–99.8	9.18	10
SPE	Atrazine-imprinted polymer	10.0–500.0 ng g ⁻¹	1.6–3.3 ng g ⁻¹	80.2–119.1	14.6	12
DA-DME	Ethyl acetate	10–100 ng mL ⁻¹	0.62–1.79 ng mL ⁻¹	84.3–101.0	6.2	15
SPME	IL-calixarene-coated fibers	25–5000 ng g ⁻¹	3.3–13.0 ng g ⁻¹	71.5–96.9	9.9	37
SPE	Hollow molecularly imprinted microspheres	0.5–250 ng g ⁻¹	0.08–0.16 ng g ⁻¹	81.0–96.0	9.8	38
MDSPE	IL-MG	0.55–500 ng mL ⁻¹	0.09–0.15 ng mL ⁻¹	97.0–100.8	3.8	This work

Comparison of IL-MG-MDSPE with other methods

By considering the linear range, LOD, recovery, and RSD, the developed method was compared with the published methods for the extraction and determination of triazine herbicides. Compared with the reported methods in Table 4, the presented method had higher recovery, and a lower LOD and RSD which proved its better accuracy, sensitivity, and precision. Compared with other adsorbents, IL-MG has excellent adsorption ability and reaches adsorption equilibrium (15 min) and desorption equilibrium (5 min) in a short time. Besides, the analytes adsorbed on the surface of the IL-MG adsorbent can be easily isolated from the sample solution by using an external magnet, which reduced the pretreatment time and made the sample pretreatment procedure more convenient. These characters enabled IL-MG-MDSPE to become a simple and fast pretreatment method for the extraction and isolation of triazine herbicides from surface water.

Assay of triazine herbicides in surface water

To evaluate the application of the presented IL-MG-MDSPE method, five surface water samples including river water, pond water, and farmland surface water were collected from a local city and processed under the optimized conditions. The chromatograms in Fig. 6a show that the proposed IL-MG-MDSPE has a good enrichment effect (83-fold) and there was no matrix interference at the retention time of triazine herbicides. In addition, the trace level of atrazine was detected in one of the farmland surface water samples (Fig. 6b). The result implied that the present IL-MG-MDSPE method was reliable for the extraction and determination of triazine herbicides from surface water.

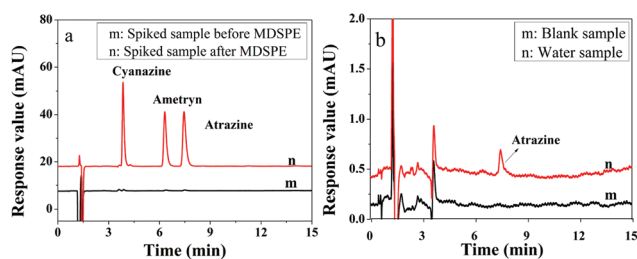


Fig. 6 Chromatograms of the spiked sample before and after MDSPE (a) and real water sample after MDSPE (b).

Conclusions

In the present work, IL-MG was synthesized by a simple and time-saving one-pot strategy wherein the tedious processes such as the synthesis of magnetic Fe₃O₄, modification with an IL, and reduction of GO to graphene were conducted at the same time. IL-MG exhibited excellent adsorption ability owing to these two reasons: (1) IL and Fe₃O₄ nanoparticles inserted between graphene sheets that effectively inhibited the stacking of graphene sheets and enlarged the surface area of graphene; (2) IL provided multiple binding sites between IL-MG and analytes (π - π , hydrophobic interaction, and electrostatic interaction). The IL-MG-MDSPE method combined the excellent adsorption ability of IL-MG and the rapid extraction efficiency of MDSPE, and was successfully applied to the rapid isolation and determination of the trace levels of triazine herbicides from surface water.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

This work was supported by the National Natural Science Foundation of China [21575033]; the Natural Science Foundation of Hebei Province [B2015201132, B2015104013]; the Natural Science Foundation of Education Department of Hebei Province [ZD2015036]; and the third graduate education and teaching reform program of Hebei University [Yjs2016-39].

Notes and references

- 1 A. C. F. Vida, D. J. Cocovi-Solberg, E. A. G. Zagatto and M. Miró, *Talanta*, 2016, **156**–157, 71–78.
- 2 D. Chen, Y. Zhang, H. Miao, Y. Zhao and Y. Wu, *J. Agric. Food Chem.*, 2015, **63**, 9855–9862.
- 3 H. Geng, S. Miao, S. Jin and H. Yang, *Anal. Bioanal. Chem.*, 2015, **407**, 8803–8812.
- 4 T. Liu, P. Cao, J. Geng, J. Li, M. Wang, M. Wang, X. Li and D. Yin, *Food Chem.*, 2014, **142**, 358–364.

- 5 S. Y. Panshin, D. S. Carter and E. R. Bayless, *Environ. Sci. Technol.*, 2000, **34**, 2131–2137.
- 6 N. Rodríguez-González, E. Beceiro-González, M. J. González-Castro and M. F. Alpendurada, *J. Chromatogr., A*, 2016, **1470**, 33–41.
- 7 F. N. Andrade, Á. J. Santos-Neto and F. M. Lanças, *J. Sep. Sci.*, 2014, **37**, 3150–3156.
- 8 M. M. Sanagi, S. S. Muhammad, I. Hussain, W. A. W. Ibrahim and I. Ali, *J. Sep. Sci.*, 2015, **38**, 433–438.
- 9 A. A. Elbashir and H. Y. Aboul-Enein, *Biomed. Chromatogr.*, 2014, **59**, 570–576.
- 10 D. Djozan, M. Mahkam and B. Ebrahimi, *J. Chromatogr., A*, 2009, **1216**, 2211–2219.
- 11 S. Xu, H. Lu and L. Chen, *J. Chromatogr., A*, 2014, **1350**, 23–29.
- 12 F. N. Andrade, C. E. D. Nazario, Á. J. Santos-Neto and F. M. Lanças, *Anal. Methods*, 2016, **8**, 1181–1186.
- 13 M. M. Sanagi, H. H. Abbas, W. A. W. Ibrahim and H. Y. Aboul-Enien, *Food Chem.*, 2012, **133**, 557–562.
- 14 E. Turiel, M. Díaz-Álvarez and A. Martín-Esteban, *J. Chromatogr., A*, 2016, **1432**, 1–6.
- 15 D. Li, Z. Zhang, N. Li, K. Wang, K. Zang, J. Jiang, A. Yu, H. Zhang and X. Li, *Anal. Methods*, 2016, **8**, 3788–3794.
- 16 Q. Zhong, Y. Hu, Y. Hu and G. Li, *J. Sep. Sci.*, 2012, **35**, 3396–3402.
- 17 A. Keramat and R. Zare-Dorabei, *Ultrason. Sonochem.*, 2017, **38**, 421–429.
- 18 M. Wierucka and M. Biziuk, *TrAC, Trends Anal. Chem.*, 2014, **59**, 50–58.
- 19 V. Singh, A. K. Purohit, S. Chinthakindi, D. G. Raghavender, V. Tak, D. Pardasani, A. R. Shrivastava and D. K. Dubey, *J. Chromatogr., A*, 2016, **1448**, 32–41.
- 20 J. Wu, D. Xiao, H. Zhao, H. He, J. Peng, C. Wang, C. Zhang and J. He, *Microchim. Acta*, 2015, **182**, 2299–2306.
- 21 L. Wang, M. Wang, H. Yan, Y. Yuan and J. Tian, *J. Chromatogr., A*, 2014, **1368**, 37–43.
- 22 Y. Li, Z. Li, W. Wang, S. Zhong, J. Chen and A. Wang, *J. Chromatogr., A*, 2016, **1447**, 17–25.
- 23 W. Fan, H. Man, L. You, X. Zhu, B. Chen and B. Hu, *J. Chromatogr., A*, 2016, **1443**, 1–9.
- 24 Z. Liu, Y. Wang, R. Deng, L. Yang, S. Yu, S. Xu and W. Xu, *ACS Appl. Mater. Interfaces*, 2016, **8**, 14160–14168.
- 25 Y. Li, X. Wu, Z. Li, S. Zhong, W. Wang, A. Wang and J. Chen, *Talanta*, 2015, **144**, 1295–1299.
- 26 E. Ziaei, A. Mehdinia and A. Jabbari, *Anal. Chim. Acta*, 2014, **850**, 49–56.
- 27 H. Yan, C. Yang, Y. Sun and K. H. Row, *J. Chromatogr., A*, 2014, **1361**, 53–59.
- 28 J. Wang, S. Huang, P. Wang and Y. Yang, *Food Control*, 2016, **67**, 278–284.
- 29 R. Leyma, S. Platzer, F. Jirsa, W. Kandioller, R. Krachler and B. K. Keppler, *J. Hazard. Mater.*, 2016, **314**, 164–171.
- 30 H. Duan, X. Wang, Y. Wang, Y. Sun, J. Li and C. Luo, *Anal. Chim. Acta*, 2016, **918**, 89–96.
- 31 M. Q. Cai, J. Su, J. Q. Hu, Q. Wang, C. Y. Dong, S. D. Pan and M. C. Jin, *J. Chromatogr., A*, 2016, **1459**, 38–46.
- 32 E. Aliyari, M. Alvand and F. Shemirani, *RSC Adv.*, 2016, **6**, 64193–64202.
- 33 C. Wang, B. Lin, G. Qiao, L. Wang, L. Zhu, F. Chu, T. Feng, N. Yuan and J. Ding, *Mater. Lett.*, 2016, **173**, 219–222.
- 34 J. Wu, H. Zhao, D. Xiao, P. H. Chuong, J. He and H. He, *J. Chromatogr., A*, 2016, **1454**, 1–8.
- 35 H. L. Huang, X. H. Wang, H. Ge and M. Xu, *ACS Sustainable Chem. Eng.*, 2016, **4**, 3334–3343.
- 36 A. Afkhami, R. Moosavi and T. Madrakian, *Talanta*, 2010, **82**, 785–789.
- 37 M. Tian, R. Cheng, J. Ye, X. Liu and Q. Jia, *Food Chem.*, 2014, **145**, 28–33.
- 38 Q. Zhao, H. Li, Y. Xu, F. Zhang, J. Zhao, L. Wang, J. Hou, H. Ding, Y. Li, H. Jin and L. Ding, *J. Chromatogr., A*, 2015, **1376**, 26–34.