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Photocatalytic oxidation of glycerol with red light employing quinacridone sensitized TiO₂ nanoparticles†

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Photocatalytic nanomaterials combining organic dyes and inorganic semiconductor nanoparticles (NPs) are extensively investigated for light-driven production of solar fuels and for conversion of organic feedstocks. However, their applications for the valorization of abundant raw materials by exploiting low-energy visible light remain limited. In this study, we report a facile preparation of TiO₂ nanoparticles sensitized with a quinacridone (QA) industrial pigment for the aqueous oxidation of glycerol to glyceraldehyde with red light ($\lambda = 620$ nm), reaching $47.5 \pm 5.0 \mu\text{mol g}_{\text{NP}}^{-1} \text{h}^{-1}$ of productivity and 80% selectivity in the presence of TEMPO co-catalyst. The hybrid material outperforms the single components and shows recyclability up to at least 5 additional times under red light while maintaining intact productivity; furthermore, it demonstrates versatility by operating also under green, yellow or white light irradiation. We believe that this work will provide a new avenue for using industrial pigment-sensitized materials in photocatalysis exploiting low energy light, providing novel strategies for the future development of this field.

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Introduction

Photocatalysis offers promising solutions for conducting chemical transformations under safe and sustainable conditions, ideally exploiting the inexhaustible power from the sun. Applications of light-catalyzed processes include solar fuel production, removal of pollutants, and development of organic processes.^{1–9} Photocatalysis can be conducted under homogeneous conditions, at the surface of electrodes or with heterogeneous, photoactive nanomaterials.¹⁰ The latter offer significant advantages: (i) the opportunity to integrate in a single component the three main functional modules in photocatalysis, *i.e.*, light absorption, charge separation and redox catalytic routines; (ii) wireless operation, without the application of external bias; (iii) possible photocatalyst recovery and recyclability typical of heterogeneous catalysis.¹¹ Indeed, back in 2014, mixed colloid photocatalysts were considered by H. B. Gray and co-workers as the most viable technology for large scale applications and scalability purposes of photochemical water splitting,¹² and since then significant advancement in the field has been reached.^{13–18}

In addition to fully inorganic photocatalysts, hybrid nanomaterials combining organic dyes and semiconductor nanoparticles (NPs) can provide further benefits, such as a separate

tuning of the two components, with easy modulation of light absorption and of energy levels.¹⁹ These hybrid nanocomposites typically exploit visible light and display enhanced charge separation efficiency, due to the location of the charge carriers on two different chemical entities (*i.e.*, the dye and the semiconductor). Moreover, their design takes advantage of the endless arsenal of organic chemistry modulation concerning the dye component. The typical operating principles involve light absorption by the organic dye and electron injection in the conduction band of the semiconductor, inducing the formation of a charge separated state. This may lead to the conduction of the desired redox reactions occurring simultaneously at the surface of the material, possibly taking advantage of co-catalysts.²⁰

To date, organic dye-sensitized NPs have found applications in photocatalytic hydrogen production^{21–23} and, more recently, in the reduction of carbon dioxide^{24–26} in the presence of sacrificial electron donors; exploitation of the oxidative route for selective conversion of organic chemicals is less explored.^{27–29}

In 2008, Ma, Zhao and co-workers reported TiO₂ NPs sensitized with Alizarin Red S (ARS) dye coupled to a homogeneous 2,2,6,6-tetramethyl-1-piperidinyloxy (TEMPO) free radical co-catalyst for the aerobic oxidation of alcohols to carbonyl compounds in benzotrifluoride solvent and exploiting 400–500 nm light, with concomitant reduction of dioxygen to superoxide anion O₂^{•−}.³⁰ A similar photochemical system for the oxidation of alcohols to carbonyls was reported combining TiO₂ with Eosin Y dye.³¹ A recent breakthrough was reported by Vauthey, Coutsolelos, Odobel and co-workers, on diketopyrrolopyrrole-sensitized TiO₂ NPs for the oxidation of benzyl alcohols coupled to hydrogen evolution,

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exploiting aminoxyl radical and Pt⁰ co-catalysts, operating in water with green light at 525 nm.²⁸ Other oxidative processes that were targeted with photocatalytic nanomaterials include the oxidation of amines,³² the conversion of sulfides to sulfoxides,³³ the activation of sp² C–H bonds through thiocyanation and cyclization reactions,³⁴ benzylic C–H activation with aerobic oxidation of benzyl ethers,³⁵ and decarboxylation and alkyl addition to tetrahydroisoquinolines.³⁶

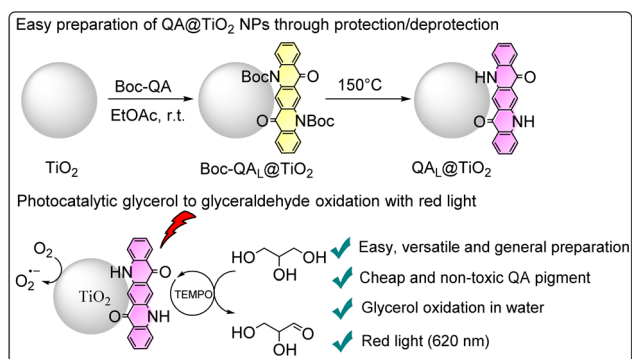
From the current state-of-the-art, three main points need to be addressed for further development and application of dye-sensitized semiconductor NPs: (i) the use of cheap and available organic dyes, and their easy integration onto the surface of the semiconductor, possibly avoiding the usage of anchoring functional groups³⁷ that requires additional synthetic steps; (ii) photocatalytic applications in water towards the conversion of abundant raw materials into valuable chemicals, with one relevant example being the oxidation of glycerol as a major by-product of soap manufacturing and of biodiesel processing;^{38–41} (iii) the use of low-energy visible light, and in particular in the red region of the electromagnetic spectrum,⁴² a long-sought but still limited goal in photocatalysis.^{43–46} Besides integrating visible light-absorbing dyes on semiconductors, a strategy to further extend the absorption of low energy radiation is to exploit dyes in aggregated states,⁴⁷ for which applications in oxidation reactions are even more rare.^{48–54}

With these premises, in this work we selected the quinacridone (QA) industrial pigment as a cheap, non-toxic and versatile tool in photocatalysis applications,⁵⁵ and sought to incorporate QA into a hybrid TiO₂ NP system (QA@TiO₂) through an easy protection/deprotection method.⁵⁶ We applied the photocatalytic QA@TiO₂ nanomaterial to the challenging and ambitious selective photocatalytic oxidation of glycerol to high added-value glyceraldehyde in water using low energy photons (up to red light at 620 nm, Scheme 1), and showing recyclability up to five additional cycles.

Results and discussion

Preparation of QA_L@TiO₂ NPs

We sought to first derivatize the parent QA pigment with a transient protecting group, such as *tert*-butoxycarbonyl

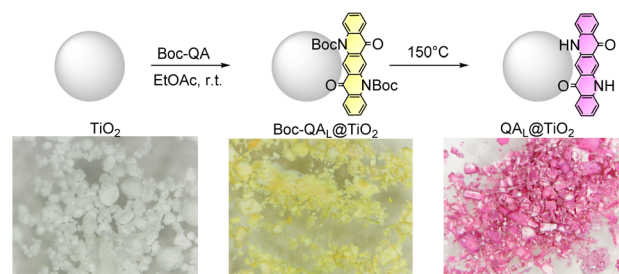


Scheme 1 Synthetic strategy of QA@TiO₂ NPs and their application for photocatalysis in water and glycerol oxidation by exploiting red light, highlighting the figures of merit of this work.

(Boc), to give Boc-QA.⁵⁶ Then, the increased processability of Boc-QA⁵⁷ can be used to adsorb the QA dye on NPs, followed by easy deprotection to obtain the dye-doped hybrid. This approach should complement the literature that took advantage of sol-gel deposition or impregnation methods to prepare QA/TiO₂ composites onto a silica support.^{58,59} Advantages of this method are the flexibility and generality of the protocol, being extendable to other dyes and to other materials, while easily controlling the dye loading (*vide infra*).

In the design of the hybrid nanomaterial, we selected commercially available titanium dioxide nanopowder (80%/20% anatase/rutile, 21 nm primary particle size). The sensitization of TiO₂ NPs exploits a simple stirring overnight of a suspension of Boc-QA, followed by the elimination of the solvent by evaporation under reduced pressure. This leads to the quantitative physisorption of the Boc-QA dye on the surface of TiO₂, to produce bright yellow Boc-QA_L@TiO₂ NPs, with the suffix *L* indicating the loading of the dye expressed in μmol per gram of TiO₂ (Scheme 2, *L* between 16 and 624). Heating the Boc-QA_L@TiO₂ NPs in an oven (150 °C for *ca.* 30 minutes) allows the removal of the Boc protecting group obtaining the QA_L@TiO₂ NPs. During this step, the color of the hybrid material clearly changes from yellow to purple (Scheme 2) as a consequence of the Boc-QA → QA transformation upon deprotection. Indeed, diffuse reflectance spectroscopy on the particles shows a shift of the absorption bands from Boc-QA₃₁₂@TiO₂ NPs (broad band between 400 and 550 nm) to QA₃₁₂@TiO₂ NPs (broad band between 450 and 650 nm), see Fig. S3 in the ESI.† We anticipate that the QA₃₁₂@TiO₂ NPs with *L* = 312 are the most active photocatalyst (*vide infra*), thus the characterization will refer to this material as a representative case.

The loss of Boc protecting group was confirmed by thermogravimetric analysis: this reveals a two-step weight loss for Boc-QA₃₁₂@TiO₂ NPs, the first taking place between 100 and 200 °C (first derivative curve presenting a maximum peak at 153 °C) due to Boc loss,⁶⁰ and the second mainly occurring between



Scheme 2 Preparation of quinacridone-sensitized NPs (QA_L@TiO₂; *L* = 16–624 μmol g_{TiO₂}^{−1}), with digital photographs of the materials at each stage of the protocol. General procedure: 20 mg of TiO₂ NPs were stirred overnight in 2 mL of ethyl acetate in the presence of Boc-QA. The solvent was then removed by evaporation under vacuum, and the resulting material heated in an oven at 150 °C for *ca.* 30 minutes (Boc deprotection). The procedure was also extended to other dyes (indigo, cibacron) that were used as photocatalytic materials in control experiments (*vide infra*).



350 and 500 °C (first derivative curve presenting a maximum peak at 430 °C) due to QA loss (Fig. S4†). In contrast, for QA₃₁₂@TiO₂ NPs only the second weight loss process is observed, being consistent with the nominal loading of QA (Fig. S4†). It is worth mentioning that, different from the protected Boc-QA dye, the QA pigment can form hydrogen bonds²³ with the semiconductor surface through the N-H groups,⁶¹ thus providing additional supramolecular interactions between the pigment and the semiconductor component. The advantages of this methodology for the preparation of dye-sensitized NPs are (i) the easy control of the relative amount of QA and TiO₂, and (ii) the unnecessary introduction on the dye of permanent functional groups for its covalent anchorage to the surface.^{37,62,63}

Characterization of QA_L@TiO₂ NPs

The hybrid dye-sensitized NPs were characterized by combining several techniques providing complementary information. Transmission electron microscopy (TEM) images of pristine TiO₂ NPs and of QA_L@TiO₂ are reported in Fig. S5,† showing that the functionalization with the QA dye is only slightly impacting the size and shape of the NPs, in line with a spherical model where the QA layer is expected to be lower than 2 nm even at its highest loading (ESI†). We then conducted a scanning transmission electron microscopy (STEM) coupled to energy dispersive X-ray (EDX) tomography to trace the elemental map composition. As can be seen from the secondary electron images (Fig. 1a), the QA₃₁₂@TiO₂ NPs appear smooth and uniform (see Fig. S6† reporting the same analysis on the QA₁₆@TiO₂ hybrid and on TiO₂ materials; high-angle annular dark-field (HAADF)-STEM images are also reported in Fig. S6†). The homogeneous distribution of the coating of QA is confirmed by the EDX-mapping of C, N, O and Ti (Fig. 1b).

X-ray photoelectron spectroscopy (XPS) spectra of QA₃₁₂@TiO₂ NPs were interpreted in comparison of bare TiO₂ and of QA. In the survey scan, only the signals of the expected elements were observed, while the Cu signal from the support was minimal (Fig. S7†). For both QA₃₁₂@TiO₂ and TiO₂ NPs, the Ti 2p photoemission line consists of a doublet with a sharper Ti 2p_{3/2} peak and a broader Ti 2p_{1/2} peak in a 2 : 1 area ratio (Fig. S8†). No shape changes can be appreciated in the presence of QA, suggesting that there is no surface chemical reaction between the TiO₂ and the QA film. In QA₃₁₂@TiO₂, the C 1s spectra was found to be consistent with QA in terms of energy position and full width at half maximum, Fig. 1c. After a multi-peak analysis, the C 1s photoemission line revealed the expected three components related to QA. Besides the sp² C=C at 284.6 eV, another peak due to the carbon in alpha to the nitrogen (C-NH-C) was found at a binding energy of 285.6–285.7 eV, while the chemically shifted component associated with the C of C=O groups was found between 286.5 and 286.6 eV (Fig. 1c). The reliability of the deconvolution is also given by the consistency of the N 1s/C 1s ratio across the QA molecule (10.4%) and QA₃₁₂@TiO₂ sample (9.2%), excluding the O-C=O contamination.^{64,65} The N 1s core level is relatively narrow (full width at half maximum = 1.6 eV) and rather symmetrical indicating a single nitrogen type. The peak is

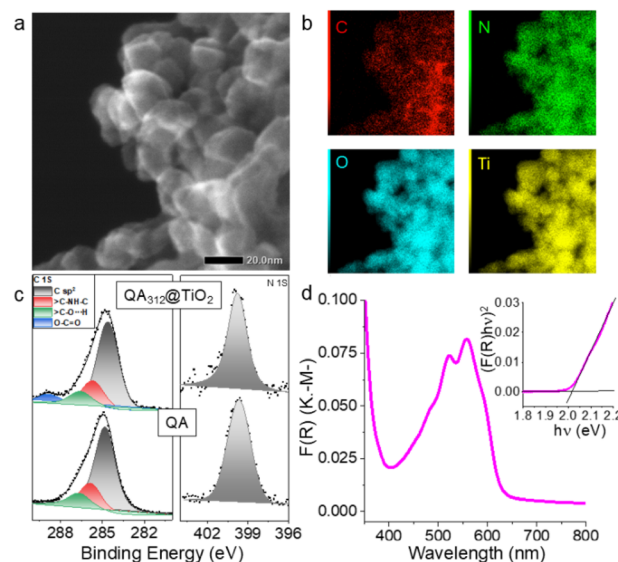


Fig. 1 Characterization of QA₃₁₂@TiO₂. (a) STEM (scale bar 20 nm). (b) EDX analysis (C, N, O, Ti). (c) XPS (C 1s and N 1s regions) of QA₃₁₂@TiO₂ (top) and of QA (bottom); due to charging effects, the energy scale is reported by setting the chemically shifted C 1s component of C sp² at 284.6 eV. (d) Diffuse reflectance spectra of QA₃₁₂@TiO₂ reported in Kubelka–Munk units; inset: Tauc plot analysis.

centered at 400.0 eV for the QA molecule and at 399.8–399.9 eV for the QA₃₁₂@TiO₂, aligning with the expected binding energies of aromatic amines (Fig. 1c). Similar features are observed in the analysis of QA₁₆@TiO₂, except for a lower intensity of the C and N signals due to the lower amount of QA in this material (Fig. S9†).

The diffuse reflectance spectrum of the QA₃₁₂@TiO₂ representative material is reported in Kubelka–Munk units in Fig. 1d (see Fig. S10† for all spectra). The spectra show an absorption feature below 400 nm ascribable to the TiO₂ semiconductor and a broad band in the 450–650 nm region, typical of the aggregated states of QA.⁶¹ Increasing the amount of QA in the particles leads to an increase in the overall absorption (Fig. S10†),⁶⁶ together with a redshift of the band. The Kubelka–Munk's analysis reveals an optical band gap comprised between 2.0 and 2.1 eV (inset in Fig. 1d), in agreement with the one observed for QA films on mesoporous surfaces (Fig. S10†).^{55,61} The combination of the optical band gap with the half-wave potential under anodic scan of 1.23 ± 0.02 V vs. the Reversible Hydrogen Electrode (RHE) for QA films⁶¹ indicates a proper energy level alignment to achieve photoinduced electron injection from QA to TiO₂ (conduction band at -0.16 V vs. RHE⁶⁷), Fig. S11.† In particular, while further photochemical characterization on QA@TiO₂ NPs is hampered by scattering, aggregated QA films are characterized by lifetimes up to the nanosecond timescale, while photoinduced electron transfer onto TiO₂ occurs in a ps timescale in photoelectrode slides.⁶¹

Given that we observed aggregated QA in the QA_L@TiO₂ samples, we performed an XRD analysis and resonant Raman experiments to identify the crystal structure and the possible presence of polymorphs.^{68,69} The diffractogram of QA₃₁₂@TiO₂



shows the same characteristics of the TiO_2 starting material, maintaining the mixture of anatase and rutile phases after sensitization. No shape changes can be appreciated in the presence of the QA film, supporting that it does not affect the bulk crystal structure of the NPs (Fig. S12†). In $\text{QA}_{312}@\text{TiO}_2$, no signals attributable to QA are detected, probably due to the small amount.

The $\text{QA}_{312}@\text{TiO}_2$ resonant Raman spectra show the features of both TiO_2 (Raman shifts at 138, 395, 514 and 637 cm^{-1})⁷⁰ and of the QA pigment (Fig. S13†). Particularly informative is the single peak at 226 cm^{-1} suggesting the presence of the γ -polymorph of QA, while the β -polymorph shows a pair of bands in this region.⁶⁸ In $\text{QA}_L@\text{TiO}_2$ hybrids, the appearance of a fluorescence band of the QA pigment is also observed at wavenumbers above 1000 cm^{-1} . Interestingly, this band is much more intense when QA is physisorbed onto SiO_2 instead of TiO_2 particles, prepared by the same protection-deprotection protocol (Fig. S13†). This indicates a partial quenching of the QA pigment when physisorbed onto TiO_2 with respect to SiO_2 , consistent with the postulated photoinduced $\text{QA} \rightarrow \text{TiO}_2$ electron injection. This encouraged us to exploit $\text{QA}_L@\text{TiO}_2$ in photochemical oxidations.

Photocatalysis with $\text{QA}_L@\text{TiO}_2$ nanoparticles

Light driven oxidation of 3,3',5,5'-tetramethylbenzidine (TMB). 3,3',5,5'-Tetramethylbenzidine (TMB) was selected as a probe substrate since its oxidation yields colored products and can be easily followed by optical spectroscopy. This electron-rich aromatic diamine can be oxidized by a single electron transfer to give the corresponding radical cation $\text{TMB}^{\cdot+}$, which undergoes a disproportionation equilibrium to form a blue charge transfer complex (absorption features at 370 and 652 nm) between TMB and its doubly oxidized form[†] (Fig. 2a).^{71–73}

Light-induced oxidation of TMB was then conducted by employing a custom-made photocatalytic reactor with 16 positions (Fig. 2b), emitting visible light (33 mW cm^{-2}) from the bottom of 8 mL glass vials containing the reaction mixtures (2 mL of 0.2 mM TMB in 0.1 M aqueous acetate buffer, pH 5, and 1 mg mL^{-1} of NPs). Images of reaction vials are reported in Fig. 2c at increasing irradiation time (in a timeframe of 10 minutes), showing the different activity of $\text{QA}_{312}@\text{TiO}_2$ NPs (right vial) with respect to the TiO_2 control (left vial). A typical outcome for $\text{QA}_{312}@\text{TiO}_2$ NPs is reported in Fig. 2d, which shows the rising of the broad band centered at 650 nm typical of the charge transfer complex as the oxidized product of TMB (formation of the radical cation $\text{TMB}^{\cdot+}$ was also confirmed by electron paramagnetic resonance spectroscopy,⁷¹ see Fig. S14†). The absorbance at 650 nm can be plotted over time to provide the TMB oxidation outcome for the different materials tested, as shown for some traces in Fig. 2e. The results can be discussed as follows: (i) the $\text{QA}_L@\text{TiO}_2$ nanoparticles drive the photochemical oxidation of TMB with a reactivity that depends on the QA loading (compare traces for $\text{QA}_{312}@\text{TiO}_2$ and $\text{QA}_{16}@\text{TiO}_2$ in Fig.

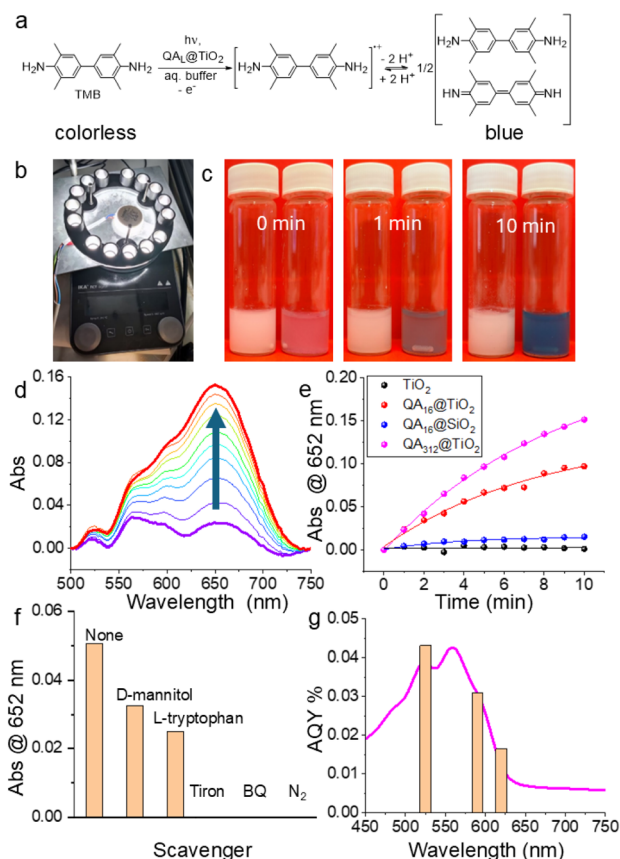


Fig. 2 (a) Light driven oxidation of 0.2 mM TMB with 1 mg per mL $\text{QA}_L@\text{TiO}_2$ NPs in 2 mL of 0.1 M aqueous acetate buffer, pH 5. (b and c) Images of the photoreactor and of the reaction vials over ten minutes containing $\text{QA}_{312}@\text{TiO}_2$ (right) and TiO_2 control (left). (d) Spectral evolution obtained for photochemical oxidation of TMB with $\text{QA}_{312}@\text{TiO}_2$ NPs. (e) Plot of the absorbance at 652 nm over time for the NPs investigated. The spectroscopic response was recorded with a plate reader, sampling aliquots of $100\text{ }\mu\text{L}$ of the irradiated solution every minute. (f) Control tests in the presence of scavengers (7.5 mM); the absorbance at 652 nm reported in the histograms refers to the $\text{QA}_{16}@\text{TiO}_2$ particles after 4 minutes irradiation. (g) Apparent Quantum Yield (AQY%) superimposed to the diffuse reflectance spectrum of $\text{QA}_{312}@\text{TiO}_2$ NPs.

2e); (ii) control with bare TiO_2 provides a null reactivity; (iii) $\text{QA}_{16}@\text{SiO}_2$ – employed as a material to elucidate the activity of the sole QA pigment§ – gives a ten times lower reactivity with respect to $\text{QA}_{16}@\text{TiO}_2$. These experiments support the role of the QA pigment in light absorption and electron injection into the conduction band of TiO_2 as the photoinduced events leading to the charge-separated state responsible for TMB oxidation.⁶¹ Moreover, the absence of reactivity under a nitrogen atmosphere suggested the involvement of reactive oxygen species (ROS). This prompted us to further investigate the photochemical reactivity in the presence of ROS scavengers: D-

† If oxidation of the radical cation of TMB proceeds further towards a second step, formation of the yellow diimine occurs, showing absorption maximum at 450 nm .

§ We used a $\text{QA}@\text{SiO}_2$ hybrid as a control test to elucidate the sole role of QA, since this was prepared in the same way as the $\text{QA}@\text{TiO}_2$ material, while SiO_2 is an inert support. Direct use of QA is more challenging given the necessity to properly disperse QA.



mannitol (scavenger for $\text{OH}^\bullet$), L-tryptophan (scavenger for $^1\text{O}_2$), sodium 4,5-dihydroxybenzene-1,3-disulfonate hydrate (Tiron monohydrate) and *p*-benzoquinone (scavengers for $\text{O}_2^{\bullet-}$), Fig. 2f.⁷⁴ The results show a major abatement of reactivity in the presence of Tiron monohydrate and *p*-benzoquinone (BQ), suggesting that superoxide $\text{O}_2^{\bullet-}$ may be involved in the oxidation process.

Finally, we evaluated the apparent quantum yield (AQY) of the process by running light-driven TMB oxidation with $\text{QA}_{312}@\text{TiO}_2$ NPs employing light emitting diodes at 525 (green), 590 (yellow) and 620 nm (red), Table S1.† As shown in Fig. 2g, the AQY reaches 0.043% at 525 nm, while decreasing to 0.031% and 0.016% at 590 and 620 nm, respectively, resembling the diffuse reflectance spectrum of $\text{QA}_{312}@\text{TiO}_2$, and confirming that the particles are photoactive up to 620 nm. A control test employing irradiation at 740 nm, where the particles do not absorb, provides indeed a null reactivity.

Light driven oxidation of glycerol. Given the promising properties of $\text{QA}_x@\text{TiO}_2$ in the photochemical oxidation of TMB, we aimed at developing an appealing oxidative process by considering the conversion of glycerol as an abundant feedstock into valuable chemicals.³⁸ Oxidation of glycerol into high added-value products has been recently investigated through plasma technologies,⁷⁵ electrocatalysis,^{76–78} photo^{79–90} and photoelectrocatalysis^{40,41,91–94} (see Table S2 in the ESI† for an overview of the photochemical and photoelectrochemical systems reported in the literature). Light-driven oxidation of glycerol (40 mM) was conducted in water under an aerobic atmosphere in the presence of the NPs and of 2,2,6,6-tetramethyl-1-piperidinyloxy free radical (TEMPO, 7.5 mM) as a co-catalyst,⁹⁵ as shown in Fig. 3. This design exploited the previous indication of activation of TEMPO upon photo-irradiation of QA-sensitized semiconductive electrodes.⁶¹

An initial screening with white light (200 mW cm^{-2}) showed that the photocatalytic reactivity was dependent on the QA pigment loading and identified $\text{QA}_{312}@\text{TiO}_2$ as the most active material at an optimal NP loading of 2 mg mL^{-1} (Table S3 in the ESI†). Under these conditions, glyceraldehyde (GLAD, estimated market value of 177\$ per g)⁷⁹ was formed as a valuable primary oxidation product after 20 h, with a productivity of $58 \mu\text{mol g}_{\text{NP}}^{-1} \text{ h}^{-1}$ and a selectivity of ca. 80% among solution products as revealed by HPLC analysis where also dihydroxyacetone (DHA), glycolic acid (GA) and formic acid (FA) were detected (control tests confirmed that TEMPO was necessary for the reactivity; TiO_2 NPs⁸³ showed a GLA productivity of $12 \mu\text{mol g}_{\text{NP}}^{-1} \text{ h}^{-1}$ and a selectivity of 67%). The preferential formation of GLAD results from the oxidation of the secondary position of glycerol typically observed in the case of photocatalysis with aminoxyl radical catalysts.⁴¹

The productivity of GLAD with $\text{QA}_{312}@\text{TiO}_2$ NPs was verified by also exploiting green (525 nm , 120 mW cm^{-2} ; GLAD productivity of $38 \mu\text{mol g}_{\text{NP}}^{-1} \text{ h}^{-1}$), yellow (590 nm , 200 mW cm^{-2} ; GLAD productivity of $46 \mu\text{mol g}_{\text{NP}}^{-1} \text{ h}^{-1}$) and red light (620 nm , 33 mW cm^{-2} , GLAD productivity of $47.5 \pm 5.0 \mu\text{mol g}_{\text{NP}}^{-1} \text{ h}^{-1}$), still maintaining a selectivity among solution products in the range 77–85%, see Fig. 3, S15 and Table S3.† Under these conditions, the reactivity of bare TiO_2 NPs is almost

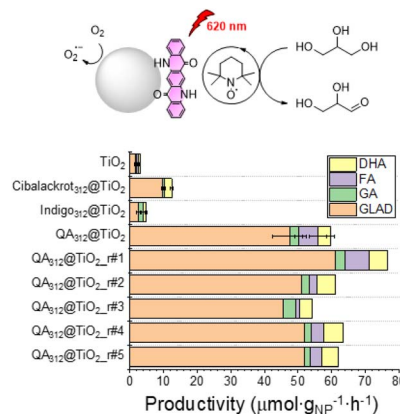


Fig. 3 Light driven oxidation of glycerol with $\text{QA}_{312}@\text{TiO}_2$ NPs co-catalyzed by TEMPO. General conditions: 40 mM glycerol in water, 7.5 mM TEMPO, 3.0 mg photocatalyst in 1.5 mL, irradiation with red light (620 nm, 33 mW cm^{-2}).

entirely suppressed (GLAD productivity of $1.75 \pm 0.25 \mu\text{mol g}_{\text{NP}}^{-1} \text{ h}^{-1}$ with red light), Fig. 3. NPs sensitized with other industrial pigments such as indigo₃₁₂@TiO₂ and cibalakrot₃₁₂@TiO₂ provided a lower reactivity than $\text{QA}_{312}@\text{TiO}_2$ (2.5 and $9.8 \mu\text{mol g}_{\text{NP}}^{-1} \text{ h}^{-1}$, respectively), Fig. 3.

The photocatalytic activity of $\text{QA}_{312}@\text{TiO}_2$ was maintained up to five additional cycles for a total of 120 h irradiation (Fig. 3), confirming that NPs sensitized with QA pigment combined photocatalytic activity with pigment stability (indeed, XPS spectra of recovered $\text{QA}_{312}@\text{TiO}_2$ were superimposable with the ones of the pristine material, Fig. S16†). Over these consecutive runs, 20 turnovers were obtained for GLAD production based on the amount of QA in the $\text{QA}_{312}@\text{TiO}_2$ material. This performance is in line with the value of 50 obtained for diketopyrrolopyrrole-sensitized TiO₂ NPs, but was reported for oxidation of benzyl alcohols with green light.²⁸

Conclusions and perspectives

In this work, we introduced an easy and general procedure for enabling the use of the industrial pigment QA to sensitize TiO₂ nanoparticles, and showed that the hybrid nanomaterials provide a boosted photochemical reactivity with respect to the separate components, enabling access to red light exploitation (up to 620 nm). We ultimately used these hybrid composites in combination with a TEMPO co-catalyst in the aqueous oxidation of glycerol to glyceraldehyde, as a significant example of conversion of an abundant and cheap raw material into a high added-value chemical. Glyceraldehyde productivity reaches $47.5 \pm 5.0 \mu\text{mol g}_{\text{NP}}^{-1} \text{ h}^{-1}$ with 80% selectivity under red light, with recyclability of the hybrid nanomaterial up to further five times without loss of activity. Turnover number for the QA pigment reaches 20, while the materials show versatility, maintaining activity also under green, yellow and white light irradiation.

Besides the facility of preparation of the hybrid nanomaterials, their versatile reactivity can be expanded to other target reactions, since TEMPO can catalyze several organic



transformations, while the reductive side can be expanded to proton or carbon dioxide reduction by considering the integration of suitable co-catalysts. These strategies are currently under investigation in our groups, together with a nano-engineering of the semiconductive support.

We thus believe that this work can pave the way for new strategies in the development of industrial pigment-based photocatalytic materials taming redox reactions in the absence of sacrificial electron donors or acceptors.

Experimental section

Synthesis of Boc-QA

312 mg (1 mmol) of QA was suspended in 50 mL of CH_2Cl_2 , and 1.06 g (4.8 mmol) of di-*tert*-butyl carbonate and 254 mg (2.1 mmol) of 4-dimethyl-aminopyridine were added thereto. The reaction mixture was stirred at room temperature for 48 hours under a nitrogen atmosphere. After the specified time, the reaction mixture showed a weak green fluorescence. The mixture was dried by rotary evaporation. Boc-QA was purified by silica gel column chromatography (the eluent gradient was 80 : 20 petroleum ether/ethyl acetate until pure ethyl acetate) and the product was obtained as a yellow powder (230 mg, yield 45%). $^1\text{H-NMR}$ (400 MHz, chloroform- d): δ 8.75 (s, 1H), 8.41 (dd, $J = 8.0, 1.6$ Hz, 1H), 7.84 (d, $J = 8.6$ Hz, 1H), 7.72 (ddd, $J = 8.7, 7.0, 1.7$ Hz, 1H), 7.38 (ddd, $J = 8.0, 7.0, 1.0$ Hz, 1H), 1.75 (s, 9H).

Preparation of $\text{QA}_L\text{@TiO}_2$ NPs

The required amount of Boc-QA and 20 mg of titanium dioxide (Merck, product code 718467) were added to an 8 mL vial, and then 2 mL of ethyl acetate was added. After ultrasonication for 1 hour, the suspension was stirred at 400 rpm overnight. The solvent was then evaporated using a rotary evaporator to obtain Boc-QA@ TiO_2 NPs. These were then placed in an oven at 150 °C for 30 minutes to convert Boc-QA into QA. The same procedure was used to prepare $\text{QA}_{16}\text{@SiO}_2$ NPs, using SiO_2 from Supelco (60737, 40–63 μm particle size, 60 Å pore size).

Characterization of NPs

TEM images were recorded with the help of Federico Caicci at the Biology Department of the University of Padova, using an FEI Tecnai Instrument G2 with a side-mounted Olympus Veleta camera and a bottom-mounted TVIPS F114 camera. Particles were suspended in ethanol before being deposited onto the grid.

STEM imaging was performed using a Jeol cold-FEG S/TEM F200 facility, at the Department of Chemical Sciences of the University of Padova.

X-ray photoelectron spectroscopy (XPS) spectra of QA, bare TiO_2 and $\text{QA}_{312}\text{@TiO}_2$ NPs were acquired in a custom-designed UHV-system with a 10^{-10} mbar base pressure, equipped with a dual Al-Mg anode X-ray source, and an Omicron EA125 electron analyzer. Core level photoemission spectra were collected with a non-monochromatic Al K α X-ray source (1486.6 eV), using an energy step of 0.1 eV, 0.5 s of dwell time, 20 eV pass energy. The data were collected by smearing the powders on the Cu substrate.

The multipeak analysis of the C 1s, N 1s, and Ti 2p photoemission lines was performed *via* subtraction of Shirley backgrounds and linear combination of Voigt functions using the KolXP software.

Diffuse reflectance spectra were recorded on a Shimadzu UV-2600i equipped with an ISR-2600Plus integrating sphere. Ultra-fine BaSO_4 powder supplied by Shimadzu Company was used as a reference. Powders of the samples were pressed in holders of ca. 3 cm diameter and 5 mm depth. The powders of the nanoparticles were mixed with the reflectance reference (BaSO_4) in a weight ratio of 3 : 100, before recording the diffuse reflectance spectra. The spectra were recorded between 1000 nm and 200 nm and transformed through the Kubelka–Munk function.

Powder X-ray diffraction was carried out on a Bruker D8 Advance Plus diffractometer, the diffraction data were acquired in Bragg–Brentano geometry by employing the Cu K α radiation (Cu anode supplied with 40 kV and a current of 40 mA) and a LYNXEYE XE-T detector with 192 measuring channels in 1D mode.

Raman measurements were conducted using a Micro-Raman setup. An argon ion laser emitting a single-line served as the excitation light source, featuring two primary lines at 488 and 514.5 nm (Spectra Physics Stabilite 2017 with an output power of 1 W). The 488 nm radiation was filtered out, and a half-wave plate was employed to control the polarization of the incident light. Optical density filters were strategically placed on a remotely controlled reel to regulate the intensity of light reaching the sample. The laser beam was coupled to a microscope (Olympus BX 40) and directed onto the sample through a 20 $\times$ or 50 $\times$ objective (Olympus SLMPL, NA D 0 : 75), resulting in a typical spot diameter of 3 or ~ 1 μm at the focus, respectively. The back-scattered Raman signal, distinct from Rayleigh scattering, was separated using an edge filter and subsequently analyzed through a 320 mm focal length imaging spectrograph (TRIAx-320 ISA) and a liquid nitrogen-cooled CCD camera (Spectrum One, Jobin Yvon). Each Raman spectrum was recorded utilizing the 50 $\times$ microscope objective, and the spectrograph slit was set at 100 μm . Spectra were generated by averaging ten repeated measurements, each with an acquisition time of 10 seconds (10 seconds $\times$ 10 times).

Photocatalysis

Photocatalysis experiments were conducted in 8 mL commercial glass vials under stirring (600 rpm), using: (i) a homemade photoreactor with 16 positions emitting white light (33 mW cm^{-2} corresponding to 0.33 sun irradiation); or (ii) a homemade photoreactor with LED at different wavelengths: red light (620 nm, 33 mW cm^{-2}); yellow light (590 nm, 200 mW cm^{-2}); green light (525 nm, 120 mW cm^{-2}); or (iii) a Godox-SL600IID lamp as the light source, and the irradiance (radiant power per unit area) was calculated at each wavelength using a calibrated silicon photodiode. The distance to the sample was 3.5 cm (2 sun irradiation, irradiance = 200 mW cm^{-2}). Further details are available in the ESI.†

Oxidation of 3,3',5,5'-tetramethylbenzidine (TMB)

1 mg TMB was dissolved in 1 mL DMSO to prepare a 4 mM stock solution in DMSO, this was then diluted into 19 mL of 0.1 M



acetate buffer (pH 5) to get 0.2 mM TMB in acetate buffer. For every reaction, 2 mL of this solution was transferred into 8 mL vials containing 2 mg of nanoparticles, then ultrasonicated for 1 hour. The vial was then subjected to irradiation under white light (0.33 sun intensity), and aliquots of 100 μ L were sampled every minute to be analyzed with a UV-vis Infinite 200 Pro plate reader (wavelength range from 300 to 800 nm, step 2 nm, read 10 flashes for every step, settle time 1 ms). The EPR experiment was recorded on a Bruker ESR5000 operating at a microwave frequency of 9.4 GHz. The sample was 1 mM TMB in 0.1 M acetate buffer (pH 5), prepared by adding 100 μ L of 4 mM TMB DMSO solution into 300 μ L 0.1 M acetate buffer with 1 mg per mL QA₁₆@TiO₂ nanoparticles.

Oxidation of glycerol

92.1 mg of glycerol (final concentration 40 mM) and 29.3 mg of TEMPO (final concentration 7.5 mM) were added into a 25 mL volumetric flask, filled with water. 1.5 mL of this solution was transferred into an 8 mL vial containing 3 mg of the nanoparticle catalysts (2 mg mL⁻¹), then inserted in an ultrasonic bath for 1 hour. The vial was subjected to irradiation for 20 hours. After the photochemical reaction, the particles were separated by centrifugation, and the solution was analyzed by HPLC.

Analysis of the oxidation products of glycerol

For quantification of the products of glycerol oxidation, high performance liquid chromatography (HPLC) was used. The instrument used was an Agilent 1260 Infinity II, equipped with a diode array detector; the instrument was equipped with a Hi-Plex H column 300 $\times$ 7.7 mm. Conditions of analysis: [H₂SO₄] = 5 mM in Milli-Q water as the eluent, isocratic gradient, flow rate 0.7 mL min⁻¹, injected volume 20 μ L, temperature 50 °C. Different concentrations of glyceraldehyde, glycolic acid, formic acid, dihydroxyacetone, and glyceric acid aqueous solutions were configured to obtain a standard curve. The solution of the photoreaction product was then centrifuged to separate the nanoparticles obtained. The solution was directly detected using HPLC, and the product concentration was converted between the detection area and the standard curve.

Data availability

The data supporting this article have been included as part of the ESI.†

Author contributions

Y. Y.: investigation, data analysis; M. N.: investigation, data analysis; A. B.: investigation, data analysis; R. B.: investigation; R. S.: investigation, data analysis; S. A.: data analysis, supervision; L. D.: conceptualization, supervision, writing, funding acquisition; A. S.: conceptualization, supervision, writing, funding acquisition.

Conflicts of interest

There are no conflicts to declare.

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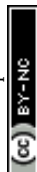
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Notes and references

- 1 S. Wu, J. Kaur, T. A. Karl, X. Tian and J. P. Barham, *Angew. Chem., Int. Ed.*, 2022, **61**, e202107811.
- 2 J. P. Barham and B. König, *Angew. Chem., Int. Ed.*, 2020, **59**, 11732–11747.
- 3 A. Vega-Peñaloza, J. Mateos, X. Companyó, M. Escudero-Casao and L. Dell'Amico, *Angew. Chem., Int. Ed.*, 2021, **60**, 1082–1097.
- 4 G. E. M. Crisenza, D. Mazzarella and P. Melchiorre, *J. Am. Chem. Soc.*, 2020, **142**, 5461–5476.
- 5 G. E. M. Crisenza and P. Melchiorre, *Nat. Commun.*, 2020, **11**, 8–11.
- 6 N. A. Romero and D. A. Nicewicz, *Chem. Rev.*, 2016, **116**, 10075–10166.
- 7 P. R. D. Murray, J. H. Cox, N. D. Chiappini, C. B. Roos, E. A. McLoughlin, B. G. Hejna, S. T. Nguyen, H. H. Ripberger, J. M. Ganley, E. Tsui, N. Y. Shin, B. Koronkiewicz, G. Qiu and R. R. Knowles, *Chem. Rev.*, 2022, **122**, 2017–2291.
- 8 R. Cao, D. Xiao, M. Wang, Y. Gao and D. Ma, *Appl. Catal., B*, 2024, **341**, 123357.
- 9 D. Spasiano, R. Marotta, S. Malato, P. Fernandez-Ibañez and I. Di Somma, *Appl. Catal., B*, 2015, **170–171**, 90–123.
- 10 M. A. Hassaan, M. A. El-Nemr, M. R. Elkatory, S. Ragab, V. C. Niculescu and A. El Nemr, *Principles of Photocatalysts and Their Different Applications: A Review*, Springer International Publishing, 2023, vol. 381.
- 11 Y. Qu and X. Duan, *Chem. Soc. Rev.*, 2013, **42**, 2568–2580.
- 12 J. R. McKone, N. S. Lewis and H. B. Gray, *Chem. Mater.*, 2014, **26**, 407–414.
- 13 Y. Goto, T. Hisatomi, Q. Wang, T. Higashi, K. Ishikiriya, T. Maeda, Y. Sakata, S. Okunaka, H. Tokudome, M. Katayama, S. Akiyama, H. Nishiyama, Y. Inoue, T. Takewaki, T. Setoyama, T. Minegishi, T. Takata, T. Yamada and K. Domen, *Joule*, 2018, **2**, 509–520.



- 14 K. Maeda, A. Xiong, T. Yoshinaga, T. Ikeda, N. Sakamoto, T. Hisatomi, M. Takashima, D. Lu, M. Kanehara, T. Setoyama, T. Teranishi and K. Domen, *Angew. Chem., Int. Ed.*, 2010, **49**, 4096–4099.
- 15 Q. Wang and K. Domen, *Chem. Rev.*, 2020, **120**, 919–985.
- 16 T. Takata, J. Jiang, Y. Sakata, M. Nakabayashi, N. Shibata, V. Nandal, K. Seki, T. Hisatomi and K. Domen, *Nature*, 2020, **581**, 411–414.
- 17 H. Nishiyama, T. Yamada, M. Nakabayashi, Y. Maehara, M. Yamaguchi, Y. Kuromiya, Y. Nagatsuma, H. Tokudome, S. Akiyama, T. Watanabe, R. Narushima, S. Okunaka, N. Shibata, T. Takata, T. Hisatomi and K. Domen, *Nature*, 2021, **598**, 304–307.
- 18 P. Zhou, I. A. Navid, Y. Ma, Y. Xiao, P. Wang, Z. Ye, B. Zhou, K. Sun and Z. Mi, *Nature*, 2023, **613**, 66–70.
- 19 D. A. Kader and S. J. Mohammed, *RSC Adv.*, 2023, **13**, 26484–26508.
- 20 C. Bie, L. Wang and J. Yu, *Chem*, 2022, **8**, 1567–1574.
- 21 N. Manfredi, B. Cecconi, V. Calabrese, A. Minotti, F. Peri, R. Ruffo, M. Monai, I. Romero-Ocaña, T. Montini, P. Fornasiero and A. Abbotto, *Chem. Commun.*, 2016, **52**, 6977–6980.
- 22 L. Zani, M. Melchionna, T. Montini and P. Fornasiero, *J. Phys.: Energy*, 2021, **3**, 031001.
- 23 X. Li, X. Lv, Q. Zhang, B. Huang, P. Wang, X. Qin, X. Zhang and Y. Dai, *J. Colloid Interface Sci.*, 2018, **525**, 136–142.
- 24 V. Nikolaou, C. Govind, E. Balanikas, J. Bharti, S. Diring, E. Vauthey, M. Robert and F. Odobel, *Angew. Chem., Int. Ed.*, 2024, **63**, e202318299.
- 25 S. Yoshino, T. Takayama, Y. Yamaguchi, A. Iwase and A. Kudo, *Acc. Chem. Res.*, 2022, **55**, 966–977.
- 26 V. Nikolaou, P. B. Pati, H. Terrisse, M. Robert and F. Odobel, *EES Catal.*, 2024, 1314–1319.
- 27 A. E. B. S. Stone, A. Fortunato, X. Wang, E. Saggioro, R. Q. Snurr, J. T. Hupp, F. Arcudi and L. Dordevic, *Adv. Mater.*, 2024, 2408658.
- 28 D. Romito, C. Govind, V. Nikolau, R. J. Fernandez-Teran, A. Stoumpidi, E. Agapaki, G. Charalambidis, S. Diring, E. Vauthey, A. G. Coutsolelos and F. Odobel, *Angew. Chem., Int. Ed.*, 2024, e202318868.
- 29 D. Franchi and Z. Amara, *ACS Sustain. Chem. Eng.*, 2020, **8**, 15405–15429.
- 30 M. Zhang, C. Chen, W. Ma and J. Zhao, *Angew. Chem., Int. Ed.*, 2008, **47**, 9730–9733.
- 31 Y. Zhang, Z. Wang and X. Lang, *Catal. Sci. Technol.*, 2017, **7**, 4955–4963.
- 32 X. Li, H. Hao and X. Lang, *J. Colloid Interface Sci.*, 2021, **581**, 826–835.
- 33 F. Huang, H. Hao, W. Sheng and X. Lang, *Chem. Eng. J.*, 2021, **423**, 129419.
- 34 M. Koohgard, Z. Hosseinpour, A. M. Sarvestani and M. Hosseini-Sarvari, *Catal. Sci. Technol.*, 2020, **10**, 1401–1407.
- 35 L. Ren, M. M. Yang, C. H. Tung, L. Z. Wu and H. Cong, *ACS Catal.*, 2017, **7**, 8134–8138.
- 36 L. Ren and H. Cong, *Org. Lett.*, 2018, **20**, 3225–3228.
- 37 K. L. Materna, R. H. Crabtree and G. W. Brudvig, *Chem. Soc. Rev.*, 2017, **46**, 6099–6110.
- 38 G. Dodekatos, S. Schünemann and H. Tüysüz, *ACS Catal.*, 2018, **8**, 6301–6333.
- 39 M. R. K. Estahbanati, M. Feilizadeh, F. Attar and M. C. Iliuta, *React. Chem. Eng.*, 2021, **6**, 197–219.
- 40 Y. Xiao, M. Wang, D. Liu, J. Gao, J. Ding, H. Wang, H. B. Yang, F. Li, M. Chen, Y. Xu, D. Xu, Y. X. Zhang, S. Fang, X. Ao, J. Wang, C. Su and B. Liu, *Angew. Chem., Int. Ed.*, 2024, **63**, e202319685.
- 41 D. Bruggeman, A. Laoprte, R. J. Detz, S. Mathew and J. N. H. Reek, *Angew. Chem., Int. Ed.*, 2022, **61**, e202200175.
- 42 T. Gobatto, G. A. Volpato, A. Sartorel and M. Bonchio, *Chem. Sci.*, 2023, **14**, 12402–12429.
- 43 H. Zhao, Z. Zhou, X. Feng, C. Liu, H. Wu, W. Zhou and H. Wang, *Nano Res.*, 2023, **16**, 8809–8816.
- 44 K. Zhang, X. Dong, B. Zeng, K. Xiong and X. Lang, *J. Colloid Interface Sci.*, 2023, **651**, 622–632.
- 45 B. F. Buksh, S. D. Knutson, J. V. Oakley, N. B. Bissonnette, D. G. Oblinsky, M. P. Schwoerer, C. P. Seath, J. B. Geri, F. P. Rodriguez-Rivera, D. L. Parker, G. D. Scholes, A. Ploss and D. W. C. MacMillan, *J. Am. Chem. Soc.*, 2022, **144**, 6154–6162.
- 46 A. H. Schade and L. Mei, *Org. Biomol. Chem.*, 2023, **21**, 2472–2485.
- 47 D. Cappelletti, M. Barbieri, A. Aliprandi, M. Maggini and L. Đorđević, *Nanoscale*, 2024, **16**, 9153–9168.
- 48 Y. Sheng, W. Li, Y. Zhu and L. Zhang, *Appl. Catal., B*, 2021, **298**, 120585.
- 49 L. Liu, J. Liu, S. Zong and Z. Huang, *Int. J. Hydrogen Energy*, 2022, **47**, 39486–39498.
- 50 Q. Zhang, L. Jiang, J. Wang, Y. Zhu, Y. Pu and W. Dai, *Appl. Catal., B*, 2020, **277**, 119122.
- 51 Q. Gao, J. Xu, Z. Wang and Y. Zhu, *Appl. Catal., B*, 2020, **271**, 118933.
- 52 J. Wang, W. Shi, D. Liu, Z. Zhang, Y. Zhu and D. Wang, *Appl. Catal., B*, 2017, **202**, 289–297.
- 53 J. Wang, D. Liu, Y. Zhu, S. Zhou and S. Guan, *Appl. Catal., B*, 2018, **231**, 251–261.
- 54 W. Wei, Z. Wei, D. Liu and Y. Zhu, *Appl. Catal., B*, 2018, **230**, 49–57.
- 55 E. Rossin, Y. Yang, M. Chirico, G. Rossi, P. Galloni and A. Sartorel, *Energy Adv.*, 2024, **3**, 1894–1904.
- 56 M. Sytnyk, E. D. Głowacki, S. Yakunin, G. Voss, W. Schöffberger, D. Kriegner, J. Stangl, R. Trotta, C. Gollner, S. Tollabimazraehno, G. Romanazzi, Z. Bozkurt, M. Havlicek, N. S. Sariciftci and W. Heiss, *J. Am. Chem. Soc.*, 2014, **136**, 16522–16532.
- 57 M. Forchetta, V. Baia, F. Sabuzi, V. Conte and P. Galloni, *Eur. J. Org. Chem.*, 2023, **26**, e202300523.
- 58 H. Ding, H. Sun and Y. Shan, *J. Photochem. Photobiol., A*, 2005, **169**, 101–107.
- 59 C.-J. Lin, Y.-H. Liou, S.-Y. Chen and M.-C. Tsai, *Sustain. Environ. Res.*, 2012, **22**, 167–172.
- 60 T. Marangoni, S. A. Mezzasalma, A. Llanes-Pallas, K. Yoosaf, N. Armaroli and D. Bonifazi, *Langmuir*, 2011, **27**, 1513–1523.



- 61 Y. Yang, G. A. Volpato, E. Rossin, N. Peruffo, F. Tumbarello, C. Nicoletti, R. Bonetto, L. Paoloni, P. Umari, E. Colusso, L. Dell'Amico, S. Berardi, E. Collini, S. Caramori, S. Agnoli and A. Sartorel, *ChemSusChem*, 2023, e202201980.
- 62 G. A. Volpato, M. Marasi, T. Gobatto, F. Valentini, F. Sabuzi, V. Gagliardi, A. Bonetto, A. Marcomini, S. Berardi, V. Conte, M. Bonchio and S. Caramori, *Chem. Commun.*, 2020, **56**, 2248–2251.
- 63 G. A. Volpato, E. Colusso, L. Paoloni, M. Forchetta, F. Sgarbossa, V. Cristino, M. Lunardon, S. Berardi, S. Caramori, S. Agnoli, F. Sabuzi, P. Umari, A. Martucci, P. Galloni and A. Sartorel, *Photochem. Photobiol. Sci.*, 2021, **20**, 1243–1255.
- 64 Y. Yu, B. Mao, A. Geller, R. Chang, K. Gaskell, Z. Liu and B. W. Eichorn, *Phys. Chem. Chem. Phys.*, 2014, **16**, 11633–11639.
- 65 S. Chenakin and N. Kruse, *J. Catal.*, 2018, **358**, 224–236.
- 66 H. Yoon, D. Kim, M. Park, J. Kim, J. Kim, W. Srituravanich, B. Shin, Y. Jung and S. Jeon, *J. Phys. Chem. C*, 2018, **122**, 12114–12121.
- 67 J. L. Peper, N. E. Gentry, B. Boudy and J. M. Mayer, *Inorg. Chem.*, 2022, **61**, 767–777.
- 68 T. Salzillo, A. Rivalta, N. Castagnetti, S. D'Agostino, M. Masino, F. Grepioni, E. Venuti, A. Brillante and A. Girlando, *CrystEngComm*, 2019, **21**, 3702–3708.
- 69 A. Giunchi, L. Pandolfi, R. G. D. Valle, T. Salzillo, E. Venuti and A. Girlando, *Cryst. Growth Des.*, 2023, **23**, 6765–6773.
- 70 H. C. Choi, Y. M. Jung and S. B. Kim, *Vib. Spectrosc.*, 2005, **37**, 33–38.
- 71 P. D. Josephy, T. Eling and R. P. Mason, *J. Biol. Chem.*, 1982, **257**, 3669–3675.
- 72 (a) F. Arcudi, L. Đorđević, N. Schweitzer, S. I. Stupp and E. A. Weiss, *Nat. Chem.*, 2022, **14**, 1007–1013; (b) L. Đorđević, T. J. Jaynes, H. Sai, M. Barbieri, J. E. Kupferberg, N. A. Sather, S. Weigand and S. I. Stupp, *Adv. Mater.*, 2025, 2418137.
- 73 C. Wang, *ACS Sens.*, 2024, **9**, 3808–3809.
- 74 Y. Jiao, L. Đorđević, H. Mao, R. M. Young, T. Jaynes, H. Chen, Y. Qiu, K. Cai, L. Zhang, X. Chen, Y. Feng, M. R. Wasielewski, S. I. Stupp and J. F. Stoddart, *J. Am. Chem. Soc.*, 2021, **143**, 8000–8010.
- 75 S. Bang, R. Snoeckx and M. S. Cha, *ChemSusChem*, 2024, **17**, e202300925.
- 76 W. Chen, L. Zhang, L. Xu, Y. He, H. Pang, S. Wang and Y. Zou, *Nat. Commun.*, 2024, **15**, 2420.
- 77 S. Zhong, B. He, S. Wei, R. Wang, R. Zhang and R. Liu, *Appl. Catal., B*, 2025, **362**, 124743.
- 78 Z. Huang, H. Ren, J. Guo, Y. Tang, D. Ye, J. Zhang and H. Zhao, *Appl. Catal., B*, 2024, **351**, 123986.
- 79 H. H. Kuo, T. G. Vo and Y. J. Hsu, *J. Photochem. Photobiol., C*, 2024, **58**, 100649.
- 80 M. R. K. Estahbanati, M. Feilizadeh, F. Attar and M. C. Iliuta, *Ind. Eng. Chem. Res.*, 2020, **59**, 22330–22352.
- 81 Y. Liu, B. Zhang, D. Yan and X. Xiang, *Green Chem.*, 2024, **26**, 2505–2524.
- 82 P. Limpachanangkul, P. Nimmmanterdwong, L. Liu, M. Hunsom, K. Pruksathorn, P. Piumsomboon and B. Chalermisinsuwan, *Sci. Rep.*, 2023, **13**, 14936.
- 83 M. Liu, H. Liu, N. Li, C. Zhang, J. Zhang and F. Wang, *ChemSusChem*, 2022, **15**, e202201068.
- 84 X. Liu, Y. Zou and J. Jiang, *Appl. Catal., B*, 2024, **350**, 123927.
- 85 E. Tacchi, G. Rossi, M. Natali, L. Đorđević and A. Sartorel, *Adv. Sustain. Syst.*, 2024, 2400538.
- 86 K. Meng, J. Zhang, B. Cheng, X. Ren, Z. Xia and F. Xu, *Adv. Mater.*, 2024, **36**, 2406460.
- 87 H. Bajpai, T. R. Nivedhitha, E. Dais, S. S. Kanungo and C. S. Gopinath, *J. Catal.*, 2024, **437**, 115644.
- 88 H. Bajpai, I. Chauhan, K. N. Salgaonkar, N. B. Mhamane and C. Gopinath, *RSC Sustain.*, 2023, **1**, 481–493.
- 89 T. R. Nivedhitha, H. Bajpai, J. V. Oommen, A. Abraham, I. Chauhan and C. S. Gopinath, *ACS Sustain. Chem. Eng.*, 2024, **12**, 14841–14853.
- 90 Y. He, Y. Wang, J. Qian, K. Xu, B. Lu, S. Tang, Y. Liu and J. Shen, *Appl. Catal., B*, 2025, **361**, 124565.
- 91 D. Liu, J. C. Liu, W. Cai, J. Ma, H. B. Yang, H. Xiao, J. Li, Y. Xiong, Y. Huang and B. Liu, *Nat. Commun.*, 2019, **10**, 1779.
- 92 H. Tateno, S. Y. Chen, Y. Miseki, T. Nakajima, T. Mochizuki and K. Sayama, *ACS Sustain. Chem. Eng.*, 2022, **10**, 7586–7594.
- 93 Y. Miao, Z. Li and M. Shao, *ChemCatChem*, 2024, **16**, 1–9.
- 94 Á. Balog, E. Kecsenovity, G. F. Samu, J. He, D. Fekete and C. Janáky, *Nat. Catal.*, 2024, **7**, 522–535.
- 95 X. Zhao, J. Yang and J. Cheng, *J. Org. Chem.*, 2023, **88**, 540–547.

