ChemComm



COMMUNICATION

View Article Online



Cite this: Chem. Commun., 2024, 60, 7164

Received 30th March 2024 Accepted 7th May 2024

DOI: 10.1039/d4cc01462f

rsc.li/chemcomm

9-Step synthesis of (-)-larikaempferic acid methyl ester enabled by skeletal rearrangement†

Mario E. Rivera, Lei Li, Aditya Kolisetti, Nina Chi and Mingji Dai **D**

We report here a concise synthesis of the anti-tumor-promoting (-)-larikaempferic acid methyl ester, a novel and rearranged abietane-type diterpene natural product containing a unique tetracyclic skeleton with a trans-hydrindane, an oxabicyclo[3.2.1]octane, and six stereogenic centers. Our synthesis starts with the cheap and abundant abietic acid and features an oxidative C-C bond cleavage followed by a transannular aldol reaction to skeletally rearrange the 6-6-6 tricyclic carbon skeleton of abietic acid to the desired 6-5-7 tricyclic carbon skeleton and an intramolecular oxa-Michael addition to form the oxa bridge. This skeletal rearrangement strategy enabled us to synthesize (-)-larikaempferic acid methyl ester in 9 steps.

Larikaempferic acid (1, Fig. 1) was first isolated as its methyl ester form (2) by Tanaka and co-workers from the leaves of Larix kaempferi in 1999 with 0.0006% isolation yield. Its structure was elucidated by HRMS, comprehensive NMR analysis, and circular dichroism spectroscopy. It was identified as a structurally rearranged abietane-type diterpene natural product with a unique tetracyclic skeleton featuring a trans-hydrindane, an oxabicyclo[3.2.1]octane, and six stereogenic centers. Interestingly, Zhao et al. recently discovered larikaempferic acid from the root bark of *Pinus massoniana* as well.² Biologically, larikaempferic acid methyl ester was found to have potent inhibitory effect on Epstein-Barr virus early antigen (EBV-EA) activation in Raji cells induced by 12-O-tetradecanoylphorbol 13-acetate (TPA). It's more effective than β-carotene,³ which has been heavily investigated in cancer prevention in animal models. Thus, larikaempferic acid methyl ester is a promising anti-tumor-promoting lead for cancer prevention development.

8α,9α,14α-Diepoxyabietan-18-oic acid (3) is another natural product isolated from the same leaves of Larix kaempferi, which

led Tanaka and Matsunaga to propose 3 as a biosynthesis precursor for larikaempferic acid (1) via intermediates including 4, 5, and 6.4 As shown in Fig. 1(A), 3 could be converted to 4 via a sequence of reductive epoxide ring opening with a hydride, dehydration, and a nucleophilic epoxide ring opening with water to form a 1,2-diol (see 4). Oxidative cleavage of the 1,2-diol would convert 4 to 5 with a ten-membered diketone. An enzymatic and stereoselective transannular aldol cyclization would deliver 6 for a subsequent intramolecular etherification to form the oxa bridge and produce larikaempferic acid. While this plausible biosynthesis proposal has not been validated yet, it inspired us to propose a synthetic approach using a transannular aldol reaction⁵⁻¹¹ as the key step to build the tricyclic carbon skeleton with a trans-hydrindane and a trans 5,7-fused ring system. As shown in Fig. 1(B), larikaempferic acid methyl ester (2) could be potentially synthesized from enone 7 with the desired 6-5-7 tricyclic ring system via an intramolecular oxa-Michael addition. The 6-5-7 tricyclic carbon framework could then be generated via a transannular aldol reaction of the trans 6,10-fused diketone 8. Diketone 8 could be synthesized from compound 9 with a 6-6-6 tricyclic carbon skeleton, which can be derived from abietic acid (10), a cheap and abundant starting material. At the planning stage, we were aware of a few challenges. First, in addition to the proposed C7-C9 bond formation to form the desired 6-5-7 tricyclic ring system, the transannular aldol reaction could happen between C14 and C9 or C11 and C8 to form two different 6-7-5 tricyclic systems. Second, it might be difficult to control the stereochemistry of the newly formed 5,7-fused ring junction from the transannular aldol reaction, which turned out to be very substrate dependent. Third, it would be ideal to keep the C13-C15 double bond which could migrate into conjugation with the C8 ketone for the oxa-Michael addition, but selective oxidative cleavage of the C8-C9 double bond in presence of the C13-C15 double bond would be challenging. Fourth, if we reduce the C13-C15 double bond, how the stereochemistry at C13 would control the transannular aldol reaction and the following steps was unclear. Overall, we were intrigued by the strategy of structurally rearranging a readily available 6-6-6 tricyclic ring system to the

^a Department of Chemistry, Emory University, Atlanta, GA 30022, USA. E-mail: mingji.dai@emory.edu; Tel: 001-404-727-4299

^b Department of Pharmacology and Chemical Biology, Emory University, Atlanta, GA 30022, USA

[†] Electronic supplementary information (ESI) available. CCDC 2326256 (14a), 2326255 (14b) and 2326254 (19). For ESI and crystallographic data in CIF or other electronic format see DOI: https://doi.org/10.1039/d4cc01462f

Communication ChemComm

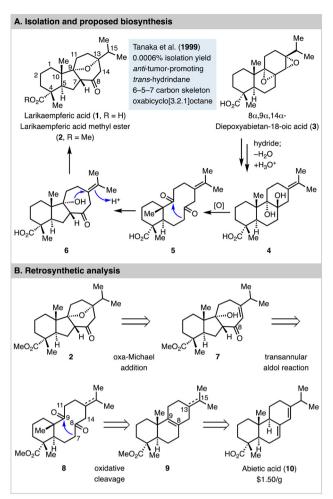


Fig. 1 Larikaempferic acid, proposed biosynthesis, and retrosynthetic analysis

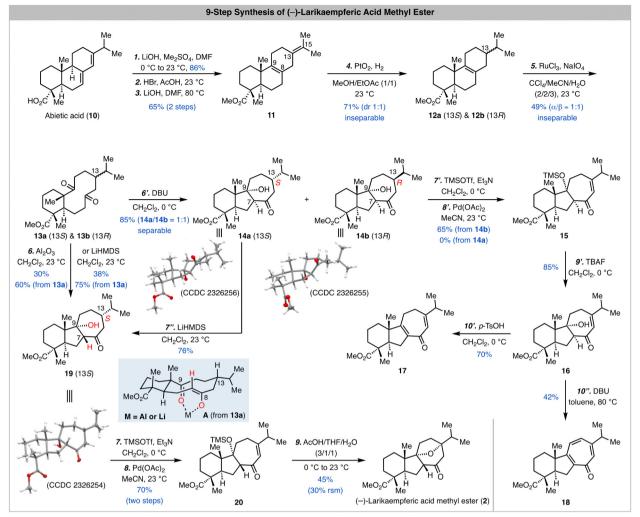
challenging 6-5-7 tricyclic ring system of larikaempferic acid. Herein, we report the details of our explorations which led to a concise 9-step synthesis of (-)-larikaempferic acid methyl ester from abietic acid.

As shown in Scheme 1, our synthesis started from commercially available abietic acid, which can be advanced to 11 via a reported three-step procedure. 12,13 As expected, selective oxidative cleavage of the C8-C9 double bond in presence of the C13-C15 double bond turned out to be problematic. Thus, we selectively reduced the C13-C15 double bond with a PtO₂catalyzed hydrogenation in presence of the C8-C9 double bond and a 1/1 mixture of inseparable 12a and 12b was obtained in 71% yield. At this stage, oxidative cleavage of the C8-C9 double bond with a combination of RuCl₃ and NaIO₄ developed by Sharpless and co-workers¹⁴ gave a 1/1 mixture of inseparable 13a and 13b in 49% yield. We next used this mixture to explore the transannular aldol reaction. When the mixture was treated with DBU in CH₂Cl₂, a 1/1 mixture of 14a and 14b was obtained in 85% total yield and separated. The structures of both 14a and 14b were unambiguously established by X-ray crystallographic analysis (CCDC 2326256 and CCDC 2326255, respectively).† It turned out that a cis 5,7-fused ring system with desired

stereochemistry at C9 but undesired stereochemistry at C7 was formed during the transannular aldol reaction. Since C7 is potentially epimerizable, we decided to move forward with 14a and 14b for the next Saegusa-Ito oxidation. 15 Interestingly, while 14b could be converted to enone 15 in 65% yield, 14a failed presumably because the oxoallyl-palladium is formed at the opposite face of the C13 hydrogen atom thus prohibiting the βhydride elimination to form the enone. Several one-step oxidations including the IBX oxidation16 and Mukaiyama dehydrogenation¹⁷ were explored as well but unsuccessful. After removal of the TMS group on the tertiary alcohol to produce 16, we explored different basic and acidic conditions to epimerize the C7 stereocenter and promote the oxa-Michael addition. Unfortunately, these endeavours were not fruitful. Notably, when 16 was treated with p-TsOH, dehydration product 17 was obtained in 70% yield; when it was treated with DBU in toluene at elevated temperature (80 °C), tropone 18 was produced in 42% yield presumably via a sequence of dehydration and oxidation with air.

The failure of converting 16 to larikaempferic acid methyl ester (2) indicates that the trans ring junction stereochemistry of the 5,7-fused ring system is important for the oxa-Michael addition. Thus, we needed to tune the transannular aldol reaction to provide the desired trans 5,7-fused system. We suspected that adding a Lewis acid to chelate with the C8 enolate (trans form) and C9 ketone can help to control the transannular aldol reaction (see the chelation model A derived from 13a). To our delight, when the 1/1 mixture of 13a and 13b was treated with Al_2O_3 , 18,19 product 19 (CCDC 2326254)† with desired trans stereochemistry at the ring junction of the 5,7-fused ring system was obtained in 60% based on the amount of 13a used. Interestingly, 13b didn't lead to the formation of the corresponding transannular aldol product but decomposed, which is presumably due to the high energy of the transition state with the isopropyl resides in the pseudo axial position (structure not shown; see model A but switch the position of the hydrogen atom and isopropyl group on C13). We also discovered that treating the mixture of 13a and 13b with LiHMDS gave similar result and 19 was obtained in higher yield (75% based on 13a). During this reaction, we observed that both 19 and 14a were produced at an early stage and 19 was the dominant product at the end of the reaction, which indicates that 14a could be converted to 19 via a sequence of retro-aldol reaction and chelation controlled transannular aldol cyclization. Indeed, when 14a was subjected to the LiHMDS conditions, 19 was produced in 76% yield. Notably, 14b didn't undergo a similar process to epimerize the C7 stereocenter, but elimination of the C9 alcohol.

With a better understanding of the transannular aldol reaction and 19 in hand, we moved forward to complete the synthesis of larikaempferic acid methyl ester (2). Saegusa-Ito oxidation of 19 occurred smoothly to deliver enone 20 in 70% yield over two steps. We then developed a one-step procedure to remove the TMS group and trigger the oxa-Michael addition. When 20 was treated with a mixture of AcOH/THF/H₂O (3/1/1), (-)-larikaempferic acid methyl ester was obtained in 45% yield. The NMR, Mass Spec, and optical rotation data of our synthetic sample matched well with the ones reported for the natural sample.



Scheme 1 9-Step synthesis of (–)-larikaempferic acid methyl ester and related investigations

In summary, using a skeletal rearrangement²⁰ and transannular cyclization strategy, ^{21,22} we developed an efficient synthesis of (—)-larikaempferic acid methyl ester from abietic acid. The key steps include an oxidative C8–C9 double bond cleavage to deliver a 10-membered diketone, a chelation-controlled transannular aldol cyclization to build the *trans* 5,7-fused ring system, and an oxa-Michael addition to form the oxabicyclo[3.2.1]octane. These enabling transformations led us to (—)-larikaempferic acid methyl ester in 9 steps. In addition, this synthesis provides support to the proposed biosynthetic pathway and suggests the oxa-Michael reaction could be an alternative process for the oxa bridge formation.

This work was financially supported by NIH GM128570. We thank Dr John Bacsa for collecting the XRD data for **14a** (CCDC 2326256), **14b** (CCDC 2326255), and **19** (CCDC 2326254).† We also thank Dr Bing Wang and Dr Shaoxiong Wu for NMR measurements and Dr Frederick Strobel for high resolution Mass Spectrometry analysis.

Conflicts of interest

There are no conflicts to declare.

Notes and references

- 1 H. Ohtsu, R. Tanaka, S. Matsunaga, H. Tokuda and H. Nishino, *Planta Med.*, 1999, **65**, 664–666.
- 2 Y. Fu, X. Ding, X. Zhang, X. Shao, J. Zhao, Y. Xu, X. Luo and W. Zhao, J. Nat. Prod., 2020, 83, 1229–1237.
- 3 A. Murakami, H. Ohigashi and K. Koshimizu, *Biosci., Biotechnol., Biochem.*, 1996, **60**, 1–8.
- 4 R. Tanaka and S. Matsunaga, Yakugaku Zasshi, 1999, 119(5), 319-339.
- 5 M. Inoue, T. Sato and M. Hirama, J. Am. Chem. Soc., 2003, 125, 10772-10773.
- 6 M. Inoue, T. Sato and M. Hirama, Angew. Chem., Int. Ed., 2006, 45, 4843–4848.
- 7 A. J. Catino, A. Sherlock, P. Shieh, J. S. Wzorek and D. A. Evans, *Org. Lett.*, 2013, 15, 3330–3333.
- 8 C. L. Chandler and B. List, J. Am. Chem. Soc., 2008, 130, 6737-6739.
- 9 J. Liu, Y. Zhou, J. Zhu and Z.-X. Yu, Org. Lett., 2021, 23, 7566-7570.
- 10 L. Jiao, C. Yuan and Z.-X. Yu, J. Am. Chem. Soc., 2008, 130, 4421–4430.
- 11 L.-P. Zhong, R. Feng, J.-J. Wang and C.-C. Li, J. Am. Chem. Soc., 2023, 145, 2098–2103.
- 12 A. Abad, M. Arno, L. R. Domingo and R. J. Zaragoza, *Tetrahedron*, 1985, 41, 4937–4940.
- 13 M. A. González, J. Correa-Royero, L. Agudelo, A. Mesa and L. Betancur-Galvis, Eur. J. Med. Chem., 2009, 44, 2468–2472.
- 14 P. H. J. Carlsen, T. Katsuki, V. S. Martin and K. B. Sharpless, J. Org. Chem., 1981, 46, 3936–3938.

Communication ChemComm

- 15 Y. Ito, T. Hirao and T. Saegusa, J. Org. Chem., 1978, 43, 1011-1013.
- 16 K. C. Nicolaou, T. Montagnon and P. S. Baran, Angew. Chem., Int. Ed., 2002, 41, 993-996.
- 17 T. Mukaiyama, J.-i. Matsuo and H. Kitagawa, Chem. Lett., 2000, 1250-1251.
- 18 A. D. Payne, B. W. Skelton, D. Wege and A. H. White, Eur. J. Org. Chem., 2007, 1184-1195.
- 19 V. A. Ignatenko, Y. Han and G. P. Tochtrop, J. Org. Chem., 2013, 78, 12229-12235.
- 20 L. Li, W. Liang, M. E. Rivera, Y.-C. Wang and M. Dai, J. Am. Chem. Soc., 2023, **145**, 53-57.
- 21 P. A. Clarke, A. T. Reeder and J. Winn, Synthesis, 2009, 691-709.
- 22 E. Reyes, L. Prieto, L. Carrillo, U. Uria and J. L. Vicario, Synthesis, 2022, 54, 4167-4183.