Nanoscale



PAPER

View Article Online



Cite this: Nanoscale, 2023, 15, 4282

Self-assembly of β-cyclodextrin-pillar[5] arene molecules into supramolecular nanoassemblies: morphology control by stimulus responsiveness and host-quest interactions†

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Macrocyclic molecules have attracted considerable attention as new functional materials owing to their unique pore size structure and excellent host-quest properties. With the development of macrocyclic compounds, the properties of mono-modified macrocyclic materials can be improved by incorporating pillar[n]arene or cyclodextrin derivatives through bridge bonds. Herein, we report the self-assembly of amphiphilic di-macrocyclic host molecules (H_{1-2}) based on β -cyclodextrin and pillar[5]arene units linked by azophenyl or biphenyl groups. In a $H_2O/DMSO$ (19:1, v/v) mixed polar solvent, an amphiphile H_1 with an azophenyl group self-assembled into unique nanorings and exhibited an obvious photoresponsive colour change. This photochromic behaviour makes H_1 suitable for application in carbon paper materials on which arbitrary patterns can be erased and rewritten. The amphiphile H_2 , with a biphenyl unit, selfassembled into spherical micelles. These differences indicate that various linker units lead to changes in the intermolecular and hydrophilic-hydrophobic interactions. In a CHCl₃/DMSO (19:1, v/v) mixed lowpolarity solvent, the amphiphile H_1 self-assembled into fibrous aggregates, whereas the molecule H_2 assembled into unique nanoring aggregates. In this CHCl₃/DMSO mixed solvent system, small nanosheet aggregates were formed by the addition of a guest molecule (G) composed of tetraphenylethene and hexanenitrile groups. With prolonged aggregation time, the small sheet aggregates further aggregated into cross-linked nanoribbons and eventually formed large nanosheet aggregates. The data reveal that the morphology of H_{1-2} can be controlled by tuning the intermolecular interactions of the molecules via the formation of host-guest complexes. Moreover, the polyhydroxy cyclodextrin unit on H_{1-2} can be strongly adsorbed on the stationary phase in column chromatography via multiple hydrogen bonds, and the singly modified pillar[5]arenes can be successfully separated by host-guest interactions.

Received 18th December 2022, Accepted 1st February 2023 DOI: 10.1039/d2nr07097a

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Introduction

Macrocyclic molecules, a class of cyclic molecules with specific pore sizes, have attracted the attention of many chemists and materials scientists in recent years as excellent host molecules in host-guest chemistry. 1-8 Dimacrocyclic host molecules (DHM), as the name suggests, are a type of double host molecule formed by the covalent bridging of two macrocyclic molecules. 9-12 As the host molecules, DHMs not only retain the original host-guest properties of macrocyclic compounds, but also display new properties based on the selection of specific bridging agents. For example, photoresponsive azo-

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† Electronic supplementary information (ESI) available. See DOI: https://doi.org/ 10.1039/d2nr07097a

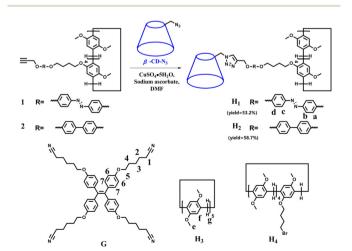
benzene can be selected as a bridging agent to confer specific photoresponsive properties to the designed materials. 13-19 Different types of macrocyclic molecules can also be selected for bridging to form functional materials.²⁰ This provides macrocyclic molecules with a more specific recognition ability to form host-guest molecules, leading to more selective separation or recognition properties for the fabrication of functional materials.

Cyclodextrins (CD), which have a hydrophilic outer cavity and hydrophobic inner cavity, are ideal host molecules, similar to enzymes, and have been widely applied in the fields of catalysis, separation, food, and medicine. 21-29 For example, Liu et al. constructed hyaluronidase-responsive polysaccharide supramolecular assemblies using chlorambucil, β-CD with hexylimidazolium units, and adamantyl-grafted hyaluronic acid. These aggregates can specifically target cancer cells and respond to hyaluronidase to induce the release of anticancer drugs. Interestingly, the functional β-CD obtained by hydroNanoscale

lysis can form a stable 1:1 host-guest complex with ATP, which considerably inhibits the hydrolysis of ATP and cuts off linker structure on the morphology of the self-assemblies, we the energy supply of cancer cells thus effectively alleviating also synthesized a molecule H containing hiphenyl units

which considerably inhibits the hydrolysis of ATP and cuts off the energy supply of cancer cells, thus effectively alleviating the multidrug resistance of cancer cells.³⁰ Pillar[n]arenes formed by methylene bridges linking p-dialkoxy benzene were first reported by Ogoshi in 2008, as a new host family of macrocyclic molecules.³¹ Although pillar[n]arene molecules have unique columnar structures and specific pore sizes, their poor solubility in water considerably limits their applications in materials science. To overcome these shortcomings, studies have attempted to modify the pillar[n]arene matrix via different methods to change the properties of pillar[n] arenes and expand their application range. 32-38 For example, we previously reported the synthesis and self-assembly of an amphiphilic macromolecule composed of pillar[5]arene, biphenyl, and hydrophilic polyethylene glycol monomethyl ether groups, which can detect aliphatic diamines in aqueous solutions.39 Huang et al. linked crown ethers with pillar[5]arenes through a click reaction, and formed a dynamic rotane with guest molecules through a pH response.40 However, there are few examples of pillar[n]arenes and CD linked together via covalent bonds. To the best of our knowledge, only Xin et al. used a click reaction to bridge azide-modified α -CD and pillar [5] arene groups to form hybrid molecules. These molecules can be effectively inserted into a lipid bilayer to form artificial ion channels. As the length of the linker increases, the selectivity of the cation transport changes, where the hybrid molecules with the shortest channel have a specific transmembrane transport preference for K⁺. These results provide a biomimetic alternative to natural K⁺ channels. ⁴¹ To date, there is no report on the synthesis of molecules based on β-CD and pillar[5] arene units connected by a covalent bond, as a linker.

With this in mind, in this study, we synthesised an amphiphilic β -CD-pillar[5]arene molecule H_1 (Scheme 1), which is formed by the linkage of β -CD and pillar[5]arenes through azobenzene. Notably, the azobenzene unit, as a linker, leads to the photoisomerization of H_1 for the construction of various



Scheme 1 Synthesis of H_{1-2} and the chemical structures of $G,\ H_3,$ and $H_4.$

nanostructural aggregates. To investigate the effect of the linker structure on the morphology of the self-assemblies, we also synthesised a molecule \mathbf{H}_2 containing biphenyl units (Scheme 1). Moreover, sheet-like supramolecular aggregates are constructed from the host-guest complex formed by \mathbf{H}_{1-2} and the guest molecule \mathbf{G} with a tetraphenylethene group, having strong aggregation-induced emission (AIE) properties.

Results and discussion

Synthesis of H₁ and H₂ and the formation of nanoassemblies in H₂O/DMSO or CHCl₃/DMSO

In recent years, bridged pillar[n]arene derivatives 42-44 and bridged cyclodextrin derivatives 45-48 have been widely reported for applications in materials science. These molecules are either covalently bridged by two or more pillar [n] arene units or covalently bridged by CD units. However, there are few reports of pillar[n]arenes and cyclodextrins linked together by covalent bonds. Therefore, amphiphilic molecules H1 and H2 were synthesised by the linkage of β-CD and pillar[5]arene groups using azobenzene or biphenyl, respectively. Molecules H₁ and H2 were obtained from a click reaction between an azide-modified β-CD and an azobenzene-functionalized pillar[5]arene or a biphenyl-functionalized pillar[5]arene. The structures of H₁ and H₂ were characterised using ¹H-NMR and ¹³C-NMR spectroscopy (Fig. S27, S28, S29, and S30†), elemental analysis, and MALDI-TOF-MS (Fig. S31 and S32†). Analysis of the experimental data showed that the molecular structure was consistent with that shown in Scheme 1.

Molecules H₁ and H₂ are composed of hydrophilic β-CD and hydrophobic rod segments that contain an azobenzenefunctionalized pillar[5]arene or a biphenyl-functionalized pillar[5]arene. The strong amphiphilic characteristics of the β-CD unit enable it to exhibit a strong self-assembly behaviour through hydrophilic/hydrophobic interactions in polar solvents, as well as strong hydrogen bonding interactions induced by the large number of hydroxyl groups on β-CD in low-polarity solvents. Therefore, $H_2O/DMSO$ (v/v = 19:1) with high polarity and $CHCl_3/DMSO$ (v/v = 19:1) with low polarity were selected to study the aggregation behaviour of the molecules. Fig. 1 and Fig. S33† show the UV-vis and fluorescence (FL) spectra of H_1 and H_2 in DMSO, $H_2O/DMSO$ (v/v = 19:1), and CHCl₃/DMSO (v/v = 19:1) solutions, respectively (3.03 \times 10⁻⁶ mol L⁻¹). Compared with that of H₁ in DMSO solution, the UV-vis spectra (Fig. 1a and Fig. S33a†) of H1 exhibited a blue shift in H₂O/DMSO and CHCl₃/DMSO solutions. This indicates that the hydrophobic rod segment of H1 tends to adopt H-type aggregation. 49-51 However, compared with that of H₂ in DMSO, the UV-vis spectra (Fig. 1c and Fig. S33c†) of H₂ exhibited a red shift in H2O/DMSO solution and a blue shift in CHCl₃/DMSO solution. It is proposed that this shift is due to the different dipole moments of the azobenzene and biphenyl units of the H₁ and H₂ molecules, leading to differences in the strength of the hydrophilic-hydrophobic interactions and steric hindrance of the two molecules. Compared with the

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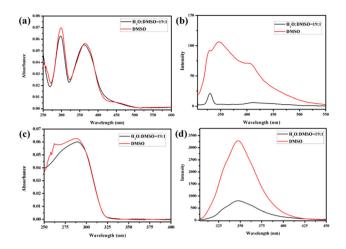


Fig. 1 The absorption spectra of (a) H_1 and (c) H_2 , and emission (excited at 298 nm, Ex bandwidth: 2.5 nm; Em bandwidth: 5 nm) spectra of (b) H_1 and (d) H_2 in DMSO and H_2 O/DMSO (v/v = 19:1) solutions (3.03 \times 10⁻⁶ mol L⁻¹).

molecules in DMSO solution, obvious fluorescence quenching of $\mathbf{H_1}$ or $\mathbf{H_2}$ was observed in both $\mathbf{H_2O}/\mathrm{DMSO}$ and $\mathrm{CHCl_3}/\mathrm{DMSO}$ solutions. This further indicates the formation of molecular aggregates in the selected solvents. To determine the size of such aggregates, dynamic light scattering (DLS) data were acquired for the $\mathbf{H_2O}/\mathrm{DMSO}$ and $\mathrm{CHCl_3}/\mathrm{DMSO}$ solutions of $\mathbf{H_1}$ and $\mathbf{H_2}$. Molecules $\mathbf{H_1}$ and $\mathbf{H_2}$ formed aggregates with diameters of approximately 21 nm and 20 nm, respectively, in $\mathbf{H_2O}/\mathrm{DMSO}$ solution (Fig. S34†), and formed aggregates with diameters of approximately 20 nm and 18 nm, respectively, in $\mathrm{CHCl_3}/\mathrm{DMSO}$ solution.

Atomic force microscopy (AFM) and transmission electron microscopy (TEM) were used to obtain detailed information about the morphology of the aggregates formed by $\mathbf{H_1}$ and $\mathbf{H_2}$. $\mathbf{H_1}$ self-aggregated into nanorings with a uniform diameter and width of \sim 7 nm and \sim 20 nm (Fig. 2a and b), respectively,

(d) (e) (f)

Fig. 2 The TEM image of (a) H_1 and (d) H_2 and AFM image of (b) H_1 and (e) H_2 obtained from 3.03×10^{-6} mol L^{-1} H_2 O/DMSO (v/v = 19:1) solutions. The TEM image of (c) H_1 and (f) H_2 obtained from 3.03×10^{-6} mol L^{-1} CHCl₃/DMSO (v/v = 19:1) solution.

in $H_2O/DMSO$ (v/v = 19:1) solutions. Additionally, the full spread length of H₁ was found to be 3.3 nm (Fig. S35a†) using CPK modelling, which is consistent with the observed half-diameter. This implies that the rigid hydrophobic segments of H₁ are arranged in a circular shape surrounded by hydrophilic β-CD units to minimise the surface tension of the molecule in aqueous solution (Fig. 3a). Unlike the structure in H₂O/DMSO solution, in CHCl₃/DMSO (v/v = 19:1) solution, H₁ selfassembled into a nanofibre structure with a diameter of approximately 4.5 nm, which is between one and two times the molecular length when the molecules are fully unfolded. It is proposed that the hydrophilic β-CD units were staggered into rings through hydrogen bonding and surrounded by hydrophobic pillar[5]arene units to minimise the surface tension of the molecules in the CHCl3/DMSO solution (Fig. 3b). This may also be due to the weak hydrophilic-hydrophobic interactions of the molecules and stronger hydrogen bond interactions between the β -CD units in the less polar CHCl₃/DMSO solution. For H₂ with a biphenyl unit as the linker group, the dipole moment is weaker than that of molecule H1 with azobenzene as the linker unit. Thus, H2 undergoes weaker hydrophilic-hydrophobic interactions in the H₂O/ DMSO solution with relatively high polarity, leading to assembly into spherical micelles with a diameter of approximately 20 nm, as shown in Fig. 2(d and e). This is considerably longer than the length of 3.1 nm (Fig. S35b†) when the molecule is fully unfolded. Thus, in an aqueous solution, the rigid hydrophobic segments of H₂ first aggregate into a nucleus, which is wrapped by hydrophilic β-CD units, and then gradually aggregate to form stable spherical micelles by synergistic driving forces such as hydrogen bonds, π - π stacking, and hydrophobic interactions (Fig. 3c). Compared with the azobenzene units, the biphenyl units with weaker dipole moments undergo weaker π - π stacking interactions in CHCl₃/DMSO solution; thus, H2 was assembled into circular aggregates with a

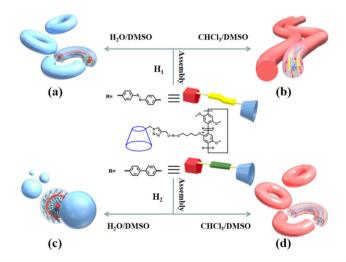


Fig. 3 Diagrammatic representation of the self-assembly of molecules H_1 (a and b) and H_2 (c and d) in $H_2O/DMSO$ and $CHCl_3/DMSO$ solutions, respectively.

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uniform diameter and width of ~5 nm and ~18 nm (Fig. 2f), respectively. Unlike the assembly arrangement of H_1 , the hydrophilic β-CD units of H₂ are staggered into rings through hydrogen bonding and surrounded by hydrophobic pillar[5] arene units in CHCl₃/DMSO solution (Fig. 3d).

Construction of reversible photoresponsive nanostructures of H₁ in aqueous solution

The trans azobenzene derivatives can form cis isomers under ultraviolet (UV) irradiation, which can be restored to the trans form upon irradiation with visible light or by increasing the temperature. 52-54 Thus, the photoresponsive properties of H₁ in DMSO solution were initially studied by ¹H NMR spectral analysis. As shown in Fig. 4, the signals of the protons on the azobenzene of the trans isomer were shifted upfield, whereas those of the protons on the pillar[5] arene units or β -CD units were not shifted when H₁ was irradiated with 365 nm UV light for 20 min. This is attributed to the photo-induced transformation of the trans azobenzene units into the cis isomers (cis-H₁). Interestingly, the DMSO solution of H₁ changed from pale-vellow to an orange-vellow solution of cis-H₁ upon irradiation with UV light (Fig. S36†), confirming the occurrence of photoisomerization. UV-vis spectroscopy was used to further examine the photoisomerization of H₁ in H₂O/DMSO (v/v = 19:1) mixed solution. Before UV irradiation, the $H_2O/$ DMSO solution of H₁ at 25 °C exhibited two characteristic absorption bands (Fig. 5a) at 298 nm and 362 nm, which are related to the $n-\pi^*$ transitions of the pillar[5] arene units and the π - π * transitions of the *trans* isomers of the azobenzene moieties, respectively. After irradiation with 365 nm UV light, the n-π* absorption band around 298 nm corresponding to the pillar[5]arene units was almost unchanged; however, a noteworthy decrease in the π - π * absorption band at 362 nm and a new $n-\pi^*$ absorption band at approximately 450 nm were observed, with a clear isosbestic point at 436 nm (Fig. 5a). This indicates trans-to-cis isomerisation of the azo-

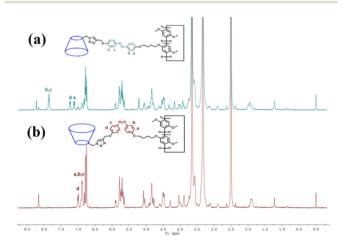


Fig. 4 1 H NMR spectra (500 MHz, DMSO- d_{6} , 25 $^{\circ}$ C) of (a) H₁ (2.72 \times 10^{-3} mol L⁻¹) and (b) H₁ (2.72 × 10^{-3} mol L⁻¹) after irradiation with 365 nm UV light for 20 min.

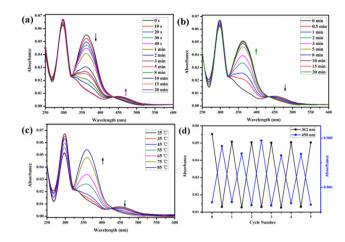


Fig. 5 UV-vis spectra of H_1 in $H_2O/DMSO$ (v/v = 19:1) solution upon exposure to different stimuli: (a) duration of UV irradiation was set to 0-20 min at 365 nm; (b) duration of vis irradiation was set to 0-20 min; (c) different temperatures of H_1 in $H_2O/DMSO$ (v/v = 19:1) solution after 20 min of UV exposure. (d) Absorbance at wavelengths of 362 nm and 450 nm under alternate UV (365 nm) irradiation for 20 min and vis irradiation for 20 min.

benzene units. When the solution of cis-H1 was irradiated with visible light, the $n-\pi^*$ absorption band at 362 nm gradually increased and the π - π * absorption band at 450 nm gradually decreased (Fig. 5b). This phenomenon became more obvious as the visible-light exposure time increased. A similar phenomenon occurred when the solution of cis-H1 was heated (Fig. 5c), suggesting cis-to-trans isomerisation of the azobenzene units. The trans-to-cis or cis-trans isomerisation was cycled by alternate illumination with 365 nm UV light or visible light (Fig. 5d). Taking advantage of this property of H₁, as well as the obvious colour change of H₁ shown in Fig. S36,† a H₂O/DMSO solution of H₁ was applied to ordinary copy paper to fabricate photosensitive erasable writing materials. As shown in Fig. S37,† the copy paper coated with H₁ was erasable, and specific pictures and text were rewritten with arbitrary patterns.

TEM and AFM were used to intuitively show the effect of UV light on the morphology of the molecular aggregates in solution. Fig. 6 shows the TEM and AFM images of cis-H₁. The images show annular aggregates with an outer diameter of

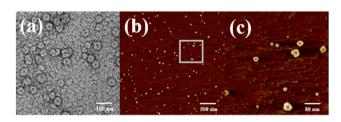


Fig. 6 (a) Negative-staining TEM images and (b) AFM images of cis-H₁ (H₁ after irradiation with 365 nm UV light for 20 min) obtained from 3.03 $\times 10^{-6}$ mol L⁻¹ H₂O/DMSO (v/v = 19:1) solutions. (c) Enlarged image of part b.

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approximately 38 nm. The size of the aggregates increased compared with that before illumination (Fig. 2a and b) when the H₂O/DMSO solution of the dimacrocyclic molecule H₁ was irradiated with 365 nm UV light for 20 min. This may result from the synergistic driving force of the molecular arrangement. In this system, the polarity of cis-H₁ is larger than that of trans-H₁, which enhances the hydrophilic-hydrophobic interactions in H₂O/DMSO solution. In contrast, the increased polarity leads to increased intermolecular force to form a larger-scale arrangement.

Control of the nanostructures of H₁₋₂ by host-guest complexes

It has been reported that a stable host-guest complex can be formed between alkylnitrile molecules and pillar[5]arene units. Tetraphenyl ethylene, a small organic molecule with AIE properties, has been widely used in advanced materials in recent years owing to its unique luminescence properties. 55-57 Based on this, we prepared a guest molecule G containing tetraphenylethylene and four alkylnitrile units that are strongly associated with pillar[5]arene units. The formation of hostguest complexes between H₁-2 and G was investigated using ¹H NMR and FL spectroscopy. The above results show that H₁ and H2 undergo very strong aggregation in chloroform solution. This induces poor solubility of molecules H₁ and H₂ in the chloroform solution and limits detailed investigation of the host-guest interaction in chloroform. Furthermore, because the aggregation behaviour leads to weak absorbance of H₁ and H₂ and fluorescence quenching, it is impossible to determine the association constant of the host-guest complex by UV and fluorescence titration. Therefore, H₃ was selected as a model for H_{1-2} to study the host-guest interaction between the pillar[5] arene unit and alkyl nitrile. In the ¹H NMR spectrum of the chloroform solution of G and 4 molar equivalents of H₃, the signals of the H_{e,f} protons of H₃ were shifted from 6.75 ppm and 3.64 ppm to 6.79 ppm and 3.66 ppm, respectively, whereas the signals of the alkyl H₁₋₅ protons on molecule G were both shifted upfield (Fig. S38†). The shift suggests the formation of a host-guest complex in chloroform. To further confirm this conjecture, ¹H-¹H NOESY NMR experiments were carried out. As shown in Fig. 7, the H_{e,f} protons of H₃ and the H₁₋₅ alkyl protons of G were significantly correlated, which indicates that there is a strong host-guest interaction between molecular H_3 and G. To calculate the value of the association, the equilibrium constant was calculated from the FL titration in CDCl₃ (Fig. S39a†).⁵⁸ When the concentration of G was 0.05 mM, the fluorescence intensity gradually increased with an increase in H₃. According to the Benesi-Hildebrand equation, the association constant K_a for the interaction of the **G** and H_3 molecules was calculated to be 1.22 \times 10¹⁷ M⁻⁴ (Fig. S39b†). This higher association constant is reasonable because compound G has four recognition sites for complexation. As shown in Fig. S40,† the plot of $1/\Delta X \nu s$. $1/[host]^4$ had a good linear least-squares fit, with a better correlation coefficient compared to that of $1/\Delta X \nu s$. $1/[host]^n$ (n = 1, 2, 3), indicating that the stoichiometry of the inclusion complex between H_3 and compound G was 4:1. The stoichiometry was also con-

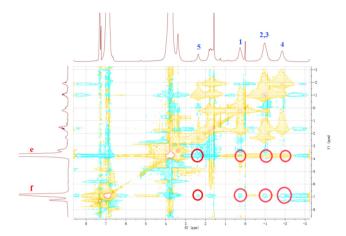


Fig. 7 ¹H-¹H NOESY NMR spectrum (500 MHz) of the complex of G with H₃ at a 1:4 molar ratio in CDCl₃ at 25 °C.

firmed using the continuous variation method (Job's plot), as shown in Fig. S40.†

Subsequently, G molecules were added to H₁ and H₂, and the ¹H NMR spectra were recorded to monitor the host-guest interactions in CDCl₃: DMSO- d_6 (v/v = 1:1) solution. As shown in Fig. 8 and S41,† the signals of the proton on the benzene ring of the pillar[5]arene unit in H1 or H2 were shifted downfield, and the signals of the alkyl H₁₋₅ protons on molecule G were both shifted upfield. The chemical shift values were consistent with those of molecule G in deuterated chloroform solution of H4, indicating that H1 or H2 and the guest molecule G formed complexes through strong host-guest interactions. The signals of the protons on the cyclodextrin unit did not change during this process (Fig. S42†), which further confirmed that a stable 4:1 host-guest complex $(H_1 \subset G \text{ or } H_2 \subset G)$ was formed between the pillar[5] arene unit and the G molecule in CHCl₃/DMSO.

Since the guest molecule G itself is an excellent aggregation-induced luminescent material, fluorescence spectroscopy

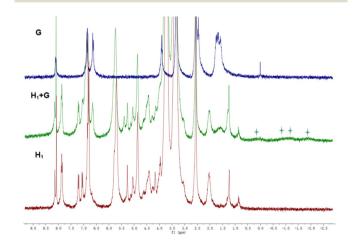


Fig. 8 1 H NMR spectra of G (blue line), $H_1 + G$ (green line), and H_1 (red line) in CDCl₃: DMSO- d_6 (v/v = 1:1) at 25 °C.

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was used to further verify the formation of the host-guest complex. However, as illustrated in Fig. S43(a),† no obvious fluorescence emission was observed when 1 equiv. of the guest molecule G was added to a solution of CHCl₃/DMSO (v/v = 19:1) containing 4 equiv. of H₁. Interestingly, when the guest molecule G was added to the solution of CHCl₃/DMSO (v/v = 19:1) of H₂, a strong fluorescence emission peak appeared at 488 nm (Fig. S43b†). This means that $H_2 \subset G$ exhibits strong AIE, compared to $H_1 \subset G$, which is useful for application in materials science. The fluorescence quenching of $H_1 \subset G$ is attributed to the absorption of the emission of molecule G by the photo-responsive azobenzyl group in the H₁ molecule, which suppresses the strong fluorescence of the tetraphenylethene unit, consistent with the reports by other groups.⁵⁹ To clearly observe such fluorescence changes with the naked eye, 1 equiv. of the guest molecule G was added to 4 equiv. of H₁ or H_2 in CHCl₃/DMSO (v/v = 5:1). Under irradiation with 365 nm ultraviolet light, the solution of G presented a weak light-blue fluorescence (Fig. S44†), whereas H₁CG in solution appeared orange-yellow without obvious fluorescence. Interestingly, the H₂⊂G solution showed bright-white fluorescence when irradiated with UV light, which is consistent with the results of the fluorescence spectroscopy analysis. Notably, the presence of the H₁ molecule significantly quenched the fluorescence; thus, this amphiphilic molecule can potentially be used as a sunscreen material.

In order to intuitively demonstrate the influence of the host-guest complex on the aggregate morphology in solution, TEM images of $H_1 \subset G$ and $H_2 \subset G$ were acquired. The experimental data are presented in Fig. 9. When the host-guest complex $H_1 \subset G$ (7.58 × 10⁻⁷ mol L⁻¹) was placed in CHCl₃/ DMSO (v/v = 19:1) solution for 1 h, an irregular circular small sheet structure with a diameter of approximately 100 nm was observed. When the solution was allowed to stand for 4 h, the aggregate changed from an irregular circular sheet structure to a cross-linked nanoribbon aggregate. After standing for 24 h,

Fig. 9 TEM images of molecule $H_1 \subset G$ (c/c = 4:1) obtained from $CHCl_3/DMSO$ (v/v = 19:1) solutions after (a) 1 h, (b) 4 h and (c) 24 h. TEM images of molecule $H_2 \subset G$ (c/c = 4:1) obtained from CHCl₃/DMSO (v/v = 19:1) solution after (d) 1 h, (e) 4 h and (f) 24 h.

larger nanosheet aggregates were observed. The height of this lamellar aggregate was approximately 7 nm, as measured by AFM (Fig. S45a and c†), which is similar to the sum of the H₁ and G lengths when the molecules are fully extended (Fig. S35c†). Compared to H1, which formed nanofibres in CHCl₃/DMSO, the supramolecular H₁⊂G molecule composed of rigid pillar[5]arenes and tetraphenylethene through a hostguest interaction undergoes a strong π - π stacking interaction to induce the formation of nanosheet-like aggregates with less curvature than the aggregates of H₁ in CHCl₃/DMSO solution. In CHCl₃/DMSO (v/v = 19:1) solution, tetraphenylethene, azobenzene, and the rigid segment of the pillar[5] arenes units are connected by an alkyl nitrile group arranged in the inner sheet through strong π - π stacking interactions, and are surrounded by four cyclodextrin units at the outer side of the supramolecule (Fig. 10a).

Similarly, after immersion of the host-guest complex $H_2 \subset G$ $(7.58 \times 10^{-7} \text{ mol L}^{-1})$ in CHCl₃/DMSO (v/v = 19:1) solution for 1 h, small nanosheet-like aggregates with a diameter of approximately 50 nm were obtained, where the size was smaller than that of the aggregates of $H_1 \subset G$ (Fig. 9d). We speculate that this is because the linker of H₁ contains azobenzene units with a larger dipole moment than that of the biphenyl units. The different driving forces of π - π stacking between the two molecules result in slightly different aggregate sizes of the molecules in solution. With an increase in the immersion time to 4 h, cross-linked nanoribbon aggregates were also found (Fig. 9e), and when the immersion time was extended to 24 h, large nanosheet aggregates with a height of ~7 nm (Fig. 9f and S45b and d†) were observed. The experimental results revealed that various nanostructural aggregates can be constructed by the formation of host-guest supramolecular molecules by precisely tuning the intermolecular interactions (Fig. 10b).

In the synthesis of singly modified pillar[5]arene H₄, owing to the excessive addition of p-dimethoxybenzene and the low solubility of the by-product H₃, a large amount of the byproduct H₃ was generated. Since the side-chain bromobutoxy

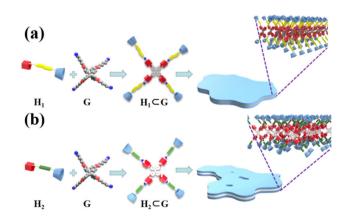


Fig. 10 Formation of host-quest complexes (a) $H_1 \subset G$ and (b) $H_2 \subset G$, and self-assembly in CHCl₃/DMSO solution, respectively.

group in H₄ can form a host-guest complex⁶⁰ with H₃, separating these molecules by column chromatography presents challenges. Accordingly, we considered the application of host-guest complexes in the separation of H₃ and H₄. Due to the strong intermolecular hydrogen bonds between the multiple hydroxyl units on cyclodextrin and ordinary column chromatography silica gel or neutral aluminium oxide (N-Al₂O₃), compounds containing cyclodextrin cannot be separated by ordinary column chromatography using silica gel and N-Al₂O₃. Taking advantage of this property, H₁ or H₂ was thoroughly stirred with N-Al₂O₃ in DMSO as the stationary phase for column chromatography, and then fully flushed with chloroform solution. After standing for 24 h, a mixture of chloroform and petroleum ether (v/v = 1:8) was used to separate the mixture of H₃ and H₄. After separation using this chromatographic column, relatively pure H₄ was obtained. It is proposed that the effective separation of H₃ and H₄ was achieved due to the side-chain bromobutoxy group in H4, which can form host-guest complexes with the pillar[5] arene unit of H₁ or H_2 . However, for molecule H_3 , there is only a weak π - π stacking interaction between H₃ and H₁ or H₂. This interaction is much weaker than the host-guest interaction of H₁ or H₂. Infrared (IR) spectroscopy was used to confirm the strong absorption between H₁ or H₂ and N-Al₂O₃ (Fig. S46(a)†). Compared with the separated column material, the strength of the stretching vibration peak of the hydroxyl group of H₁ decreased, and the absorption peak moved to a lower wavenumber, indicating that H₁ interacts with N-Al₂O₃ by hydrogen bonding. The stationary phase composed of H₂ and N-Al₂O₃ also showed the same phenomenon in the IR tests (Fig. S46b†). Thin-layer chromatography (TLC) was used to clarify the separation process. As shown in Fig. S47,† when the TLC plate did not contain H₁ or H₂, the R_f values of H₃ and H₄ were almost the same; however, when the TLC plate contained H₁ or H₂, molecules H₃ and H₄ were separated owing to the host-guest interactions of H_1/H_2 and H_4 . We believe that this material may be effective for separating molecules such as alkyl amines and halogenated hydrocarbons, which can form

Experimental section

host-guest complexes with H_1 or H_2 .

Molecules 1, 2, H_3 , H_4 , β -CD- N_3 , and G (Scheme 1 and Scheme S1†) were prepared according to previously described methods. All the structural characterisation data are summarised in the ESI (Fig. S1–S32†).

Synthesis of H₁ and H₂

These compounds were synthesised according to the same procedure. The synthesis of $\mathbf{H_1}$ is described as a representative example.

Synthesis of H_1 . β -CD- N_3 (0.101 g, 0.087 mmol) and 1 (0.1 g, 0.096 mmol) were dispersed in 10 ml of DMF and the mixture was degassed. CuSO₄·5H₂O (0.024 g, 0.096 mmol) and sodium ascorbate (0.017 g, 0.096 mmol) were added, and the mixture

was heated to 90 °C for 48 h under nitrogen. After DMF was removed, water was added, and the mixture was extracted with dichloromethane. Finally, 0.102 g of the pure target molecule **H**₁ was obtained after purification using Sephadex LH-20.

Compound H₁. (Yellow solid, yield: 53.2%, M.p.: 273 °C), ¹H NMR (300 MHz, DMSO- d_6 , δ , ppm): 8.23 (s, 1H), 7.90–7.84 (m, 4H), 7.24 (d, J = 8.4 Hz, 2H), 7.12 (d, J = 8.1 Hz, 2H), 6.89–6.75 (m, 10H), 5.93-5.91 (m, 1H), 5.81-5.66 (m, 14H), 5.23-5.17 (m, 2H), 5.07-5.05 (m, 1H), 4.97-4.78 (m, 8H), 4.60-4.46 (m, 6H), 4.36-4.31 (m, 1H), 4.19-4.14 (m, 2H), 4.06-3.51 (m, 73H), 3.17–3.10 (m, 2H), 2.93–2.85 (m, 2H), 1.99–1.87 (m, 4H). ¹³C NMR (125 MHz, DMSO- d_6 , δ , ppm): 161.40, 160.83, 150.44, 150.39, 149.71, 146.84, 146.63, 142.61, 128.00, 127.95, 127.90, 126.05, 124.64, 124.59, 115.72, 115.42, 114.58, 113.81, 102.70, 102.48, 102.36, 101.74, 83.99, 82.58, 82.13, 82.00, 81.93, 81.86, 81.45, 73.71, 73.62, 73.45, 73.34, 73.11, 72.91, 72.83, 72.70, 72.54, 72.34, 70.51, 68.18, 67.83, 61.83, 60.61, 60.55, 60.45, 60.40, 60.32, 59.47, 55.92, 55.88, 55.36, 29.48, 26.27. MALDI-TOF-MS: m/z [M + Na]⁺ 2224.8. Elemental analysis for $C_{105}H_{135}N_5O_{46}$ (2203.20 g mol⁻¹): C: 57.24%, H: 6.18%, N: 3.18%, found: C: 57.18%, H: 6.26%, N: 3.20%.

Compound H₂. (White solid, yield: 58.7%, M.p.: 288 °C), ¹H NMR (500 MHz, DMSO- d_6 , δ , ppm): 8.19 (s, 1H), 7.56–7.55 (m, 4H), 7.11 (d, J = 7.5 Hz, 2H), 7.00 (d, J = 8.0 Hz, 2H), 6.90–6.70 (m, 10H), 5.90-5.88 (m, 1H), 5.78-5.62 (m, 14H), 5.16-5.11 (m, 2H), 5.06-5.05 (m, 1H), 4.94-4.78 (m, 8H), 4.63-4.46 (m, 6H), 4.33-4.29 (m, 1H), 4.10-4.08 (m, 2H), 4.02-3.99 (m, 2H), 3.92-3.90 (m, 2H), 3.81-3.34 (m, 69H), 3.16-3.12 (m, 2H), 2.93-2.89 (m, 2H), 1.99-1.87 (m, 4H). ¹³C NMR (125 MHz, DMSO- d_6 , δ , ppm): 158.25, 157.75, 150.43, 150.36, 149.70, 143.01, 133.07, 132.69, 128.53, 127.99, 127.94, 127.73, 125.97, 115.55, 115.26, 114.55, 113.78, 102.68, 102.47, 102.36, 101.74, 83.95, 82.54, 82.08, 82.01, 81.91, 81.87, 81.46, 73.70, 73.60, 73.44, 73.33, 73.13, 72.92, 72.84, 72.70, 72.63, 72.24, 70.48, 67.88, 67.74, 61.50, 60.65, 60.49, 60.43, 60.36, 60.32, 59.47, 55.91, 55.84, 55.61, 29.47, 26.21. MALDI-TOF-MS: m/z [M + $[Na]^{+}$ 2196.97. Elemental analysis for $C_{105}H_{135}N_{3}O_{46}$ (2175.19 g mol⁻¹): C: 57.98%, H: 6.26%, N: 1.93%, found: C: 58.06%, H: 6.17%, N: 1.91%.

Conclusions

Amphiphilic bridged β -cyclodextrin-pillar[5]arene molecules $\mathbf{H_1}$ and $\mathbf{H_2}$ were successfully synthesised, and their self-assembly behaviour was investigated. Molecule $\mathbf{H_1}$ contains a photoresponsive azophenyl group, whereas $\mathbf{H_2}$ contains a biphenyl group as a linker. In the $\mathbf{H_2O/DMSO}$ mixed solvent, $\mathbf{H_1}$ self-assembled into nanorings, whereas $\mathbf{H_2}$ self-assembled into spherical micelles. Similarly, in CHCl₃/DMSO, $\mathbf{H_1}$ self-assembled into fibrous aggregates, whereas $\mathbf{H_2}$ assembled into nanoring aggregates, revealing that a slight modification of the chemical structure of the bridging bond can precisely control the morphology of the nanoaggregates. The effect of UV irradiation on the self-assembly of $\mathbf{H_1}$ in $\mathbf{H_2O/DMSO}$ solution was investigated. The synergistic interaction between π - π

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stacking and hydrophilic-hydrophobic interactions of H₁ in solution led to larger nanoaggregates of cis-H1 than those of trans-H₁ in the H₂O/DMSO mixed system. Notably, the photoresponsive colour change of H₁ enabled its application in carbon paper materials for the fabrication of photosensitive erasable writing materials. Analysis of the self-assembly behaviour of the host-guest complex formed between H₁-2 and G in CHCl₃/DMSO solution showed that H₁ and G form stable hostguest complexes, which assemble into small sheet aggregates under the action of strong hydrogen bonding and π - π stacking. With prolonged assembly time, the small sheet aggregates further aggregate into cross-linked nanoribbon aggregates and finally assemble into large sheet aggregates. Similarly, the host-guest complex between H2 and G eventually aggregates into large sheet-like aggregates. Notably, because the molecule contains polyhydroxyl β-cyclodextrin units, the molecules can be bound to the neutral alumina filler through hydrogen bonding. Interestingly, the molecular filler can be effectively used in the separation of singly modified pillar[5]arenes from the derivatives of pillar[5] arenes. We expect that these functional molecules will be widely applied in the fields of molecular recognition, sensing, and separation.

Author contributions

J. Lu: methodology, validation, formal analysis, conducting the experiments, analysis of the experimental results, and writing – original draft. Y. Deng and P. Liu: assisted in the preparation of the molecules. Q. Han: performed the TEM and AFM experiments and analyzed the experimental results. Long Yi Jin: conceptualization, methodology, investigation, resources, supervision, project administration, funding acquisition, and writing – review and editing.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

This work was supported by the National Natural Science Foundation of China (grant numbers 21961041 and 21562043) and the Higher Education Discipline Innovation Project (D18012).

References

- 1 Y. Mei, Q.-W. Zhang, Q. Gu, Z. Liu, X. He and Y. Tian, J. Am. Chem. Soc., 2022, 144, 2351–2359.
- 2 H. Zhu, L. Shangguan, D. Xia, J. H. Mondal and B. Shi, *Nanoscale*, 2017, **9**, 8913–8917.
- 3 H. B. Jeon, S. Park, K. R. Ryu, S. K. Ghosh, J. Jung, K. M. Park and J. W. Ha, *Chem. Sci.*, 2021, 12, 7115–7124.

- 4 Y.-F. Li, Z. Li, Q. Lin and Y.-W. Yang, *Nanoscale*, 2020, **12**, 2180–2200.
- 5 H. Aramoto, M. Osaki, S. Konishi, C. Ueda, Y. Kobayashi, Y. Takashima, A. Harada and H. Yamaguchi, *Chem. Sci.*, 2020, **11**, 4322–4331.
- 6 T. Xiao, L. Xu, L. Zhou, X.-Q. Sun, C. Lin and L. Wang, J. Mater. Chem. B, 2019, 7, 1526–1540.
- 7 C. Tu, W. Wu, W. Liang, D. Zhang, W. Xu, S. Wan, W. Lu and C. Yang, *Angew. Chem., Int. Ed.*, 2022, **61**, e202203541.
- 8 Y.-G. Jia, M. Zhang and X. X. Zhu, *Macromolecules*, 2017, **50**, 9696–9701.
- 9 K.-X. Teng, L.-Y. Niu and Q.-Z. Yang, Chem. Sci., 2022, 13, 5951-5956.
- 10 H. Li, Y. Yang, F. Xu, Z. Duan, R. Li, H. Wen and W. Tian, *Org. Chem. Front.*, 2021, **8**, 1117–1124.
- 11 H. Kitagishi, S. Kurosawa and K. Kano, *Chem. Asian J.*, 2016, 11, 3213–3219.
- 12 Y. Liu, Z. Su, S. Jiang, H. Sun, H. Lyu and Z. Xie, *Polym. Chem.*, 2021, **12**, 6123–6133.
- 13 L. Zhang, J. Gu, X. Luo, Z. Tang, Y. Qu, C. Zhang, H. Liu, J. Liu, C. Xie and Z. Wu, *Nanoscale*, 2022, **14**, 976–983.
- 14 M. Fujiwara, M. Akiyama, M. Hata, K. Shiokawa and R. Nomura, ACS Nano, 2008, 2, 1671–1681.
- 15 L. Dong, Y. Feng, L. Wang and W. Feng, Chem. Soc. Rev., 2018, 47, 7339–7368.
- 16 M. Zheng and J. Yuan, Org. Biomol. Chem., 2022, 20, 749–767.
- 17 A. Abdollahi, H. Roghani-Mamaqani, B. Razavi and M. Salami-Kalajahi, *Polym. Chem.*, 2019, **10**, 5686–5720.
- 18 G. K. Joshi, K. N. Blodgett, B. B. Muhoberac, M. A. Johnson, K. A. Smith and R. Sardar, *Nano Lett.*, 2014, 14, 532–540.
- 19 T. Ogoshi, K. Yoshikoshi, T. Aoki and T.-a. Yamagishi, *Chem. Commun.*, 2013, **49**, 8785–8787.
- 20 X. Yan, D. Xu, J. Chen, M. Zhang, B. Hu, Y. Yu and F. Huang, *Polym. Chem.*, 2013, **4**, 3312–3322.
- 21 L. Yue, H. Ai, Y. Yang, W. Lu and L. Wu, *Chem. Commun.*, 2013, **49**, 9770–9772.
- 22 W. C. E. Schofield, C. D. Bain and J. P. S. Badyal, *Chem. Mater.*, 2012, 24, 1645–1653.
- 23 F.-F. Shen, Y. Chen, X. Dai, H.-Y. Zhang, B. Zhang, Y. Liu and Y. Liu, *Chem. Sci.*, 2021, 12, 1851–1857.
- 24 F. Seidi, A. A. Shamsabadi, M. Amini, M. Shabanian and D. Crespy, *Polym. Chem.*, 2019, **10**, 3674–3711.
- 25 S. Datz, B. Illes, D. Gößl, C. v. Schirnding, H. Engelke and T. Bein, *Nanoscale*, 2018, **10**, 16284–16292.
- 26 B. Zhang, W. Guan, S. Zhang, B. Li and L. Wu, *Chem. Commun.*, 2016, 52, 5308–5311.
- 27 D. J. DeDora, C. Suhrland, S. Goenka, S. M. Chowdhury, G. Lalwani, L. R. Mujica-Parodi and B. Sitharaman, J. Biomed. Mater. Res., Part B, 2016, 104, 1457–1464.
- 28 Z. Liu, W. Zhou, J. Li, H. Zhang, X. Dai, Y. Liu and Y. Liu, *Chem. Sci.*, 2020, **11**, 4791–4800.
- 29 L. Zhou and J. Yu, J. Sep. Sci., 2022, 45, 2310-2320.
- 30 C. Chen, Y. Chen, X. Dai, J. Li, S. Jia, S. Wang and Y. Liu, *Chem. Commun.*, 2021, 57, 2812–2815.

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- 31 T. Ogoshi, S. Kanai, S. Fujinami, T.-a. Yamagishi and Y. Nakamoto, J. Am. Chem. Soc., 2008, 130, 5022-
- 32 T. Ogoshi, T.-a. Yamagishi and Y. Nakamoto, Chem. Rev., 2016, 116, 7937-8002.
- 33 H. Lei, X. Chen, L. Xue, L. Sun, J. Chen, Z. Tan, Z.-G. Zhang, Y. Li and G. Fang, Nanoscale, 2018, 10, 8088-
- 34 Z.-Y. Li, Y. Zhang, C.-W. Zhang, L.-J. Chen, C. Wang, H. Tan, Y. Yu, X. Lo and H.-B. Yang, J. Am. Chem. Soc., 2014, 136, 8577-8589.
- 35 T. Ogoshi, K. Maruyama, Y. Sakatsume, T. Kakuta, T.-a. Yamagishi, T. Ichikawa and M. Mizuno, J. Am. Chem. Soc., 2019, 141, 785-789.
- 36 T. Xiao, W. Zhong, L. Xu, X.-Q. Sun, X.-Y. Hu and L. Wang, Org. Biomol. Chem., 2019, 17, 1336-1350.
- 37 T. Ogoshi, T. Kakuta and T.-a. Yamagishi, Angew. Chem., Int. Ed., 2019, 58, 2197-2206.
- 38 T. Ogoshi, R. Shiga and T.-a. Yamagishi, J. Am. Chem. Soc., 2012, 134, 4577-4580.
- 39 J. Lu, X. Gou, Y. Deng, Y.-R. Pei, Z. Huang and L. Y. Jin, Dyes Pigm., 2022, 199, 110052.
- 40 X. Yan, P. Wei, Z. Li, B. Zheng, S. Dong, F. Huang and Q. Zhou, Chem. Commun., 2013, 49, 2512-2514.
- 41 P. Xin, H. Kong, Y. Sun, L. Zhao, H. Fang, H. Zhu, T. Jiang, J. Guo, Q. Zhang, W. Dong and C.-P. Chen, Angew. Chem., Int. Ed., 2019, 58, 2779-2784.
- 42 Y. Cai, X. Yan, S. Wang, Z. Zhu, M. Cen, C. Ou, Q. Zhao, Q. Yan, J. Wang and Y. Yao, Inorg. Chem., 2021, 60, 2883-2887.
- 43 Y. Wang, M.-Z. Lv, N. Song, Z.-J. Liu, C. Wang and Y.-W. Yang, Macromolecules, 2017, 50, 5759-5766.
- 44 S. Sun, X.-Y. Hu, D. Chen, J. Shi, Y. Dong, C. Lin, Y. Pan and L. Wang, Polym. Chem., 2013, 4, 2224-2229.

- 45 X. Dai, X. Dong, Z. Liu, G. Liu and Y. Liu, Biomacromolecules, 2020, 21, 5369-5379.
- 46 Y. Liu, Y. Chen, L. Li, H. Y. Zhang, S. X. Liu and X. D. Guan, J. Org. Chem., 2001, 66, 8518-8527.
- 47 Y. Liu, G.-S. Chen, L. Li, H.-Y. Zhang, D.-X. Cao and Y.-J. Yuan, I. Med. Chem., 2003, 46, 4634-4637.
- 48 X. Guan, D. Zhang, T. Jia, Y. Zhang, L. Meng, Q. Jin, H. Ma, D. Lu, S. Lai and Z. Lei, Ind. Eng. Chem. Res., 2017, 56, 3913-3919.
- 49 N. Ye, Y.-r. Pei, Q. Han, M. Lee and L. Y. Jin, Soft Matter, 2021, 17, 6661-6668.
- 50 S. Yu, R. Shan, G.-Y. Sun, T. Chen, L. Wu and L. Y. Jin, ACS Appl. Mater. Interfaces, 2018, 10, 22529-22536.
- 51 J. Lu, Y. Deng, K. Zhong, Z. Huang and L. Y. Jin, Polym. Int., 2022, 71, 478-486.
- 52 S. Yu, T. Yang, M. Sui, G.-Y. Sun, T. Chen and L. Y. Jin, Dyes Pigm., 2019, 171, 107694.
- 53 S. Qian, S. Li, W. Xiong, H. Khan, J. Huang and W. Zhang, Polym. Chem., 2019, 10, 5001-5009.
- 54 A. Adam and G. Haberhauer, J. Am. Chem. Soc., 2017, 139, 9708-9713.
- 55 G. Zhang, B. Li, Y. Zhou, X. Chen, B. Li, Z.-Y. Lu and L. Wu, Nat. Commun., 2020, 11, 425.
- 56 S. Wang, Y. Wang, Z. Chen, Y. Lin, L. Weng, K. Han, J. Li, X. Jia and C. Li, Chem. Commun., 2015, 51, 3434-3437.
- 57 N. Song, D.-X. Chen, Y.-C. Qiu, X.-Y. Yang, B. Xu, W. Tian and Y.-W. Yang, Chem. Commun., 2014, 50, 8231-8234.
- 58 H. Zhang, K. T. Nguyen, X. Ma, H. Yan, J. Guo, L. Zhu and Y. Zhao, Org. Biomol. Chem., 2013, 11, 2070-2074.
- 59 R. Arumugaperumal, W.-L. Hua, P. Raghunath, M.-C. Lin and W.-S. Chung, ACS Appl. Mater. Interfaces, 2020, 12, 29650-29660.
- 60 X. Wang, K. Han, J. Li, X. Jia and C. Li, Polym. Chem., 2013, 4, 3998-4003.