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Recent advances in carbohydrate-based gelators

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Carbohydrate-based gelators are the emerging new materials having diversified application fields such as drug delivery, environmental remediation, antibacterial agents, tissue engineering, thixotropy and wound healing. They have been established as biocompatible and renewable materials for sustainable development. Natural monosaccharides have versatile structures that are useful in the synthesis of gelators. In this review, recently synthesized low-molecular weight carbohydrate-based gelators from 2014 to 2023 are meticulously discussed with their diverse application in various fields including biomedical applications. The diverse syntheses and applications of carbohydrate-based gelators establish them as new materials that are cleaner, greener and biocompatible. Therefore, this review may be helpful to the researchers working in the field of biocompatible materials for exploring customized materials and their novel future applications.

1. Introduction

Designing and tailoring smart materials with in-built functionalities in molecules are under keen development in all fields of chemistry. Low-molecular weight gelators (LMGs) are molecular assemblies obtained using versatile small molecules, which have been interesting new materials due to their wide plethora

of applications. Such applications include pharmaceuticals, tissue engineering, dye absorption, and drug delivery.^{1–3} LMGs embedded with stimulus-responsive functional groups have been valuable in research.^{3–8} Monosaccharides among carbohydrates have been extensively used in the design and synthesis of molecular gelators.^{9–11} The LMGs yield reversible gels with different solvents and are known as physical gels or supramolecular gels on the basis of their non-covalent bonds such as van der Waals interactions, π - π interactions, hydrophobic interactions, and hydrogen bonding. The gelators give rise to self-assembled, fibrous structures that hold the solvent within their structure, and this property make them unique materials

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having various applications including biomedical. The hydrogel produced by these gelators shows potential to replace the traditional polymer hydrogel in applications requiring biocompatibility. The formation of such hydrogel is reversible and enhances its applicability in various fields.

The formation of supramolecular gels has been extensively studied in diverse fields such as controlled drug delivery,^{1–3} immobilization of enzyme produced in the body,⁴ electronics and optics,^{5–7} and pollution mitigation.^{9,10} Various metallogels have been employed in the formation of smart materials.^{11–14} Extensive research for supramolecular gels has been reported for their biomedical applications^{4–7,9–17} and catalysis.^{18–21} The diverse applications of LMGs encouraged several research studies to study the nature, design, and synthesis of these LMGs.^{22–26} There are varieties of organic compounds such as sugars, amino acids, cholesterol, and urea, which have been employed for the design and synthesis of LMGs. Out of these organic compounds, sugars are biocompatible and have internal chirality, which outclass them for biomedical applications such as controlled release systems and segregation of biomolecules. D-Glucose, D-maltose, sorbitol, D-glucosamine, N-acetyl-D-glucosamine, D-lactose and 2-deoxyribose sugars were used to synthesize carbohydrate based LMGs and low-molecular weight hydrogelators (LMHG), as shown in Fig. 1. Additionally, there are stimulus-responsive gelators that react to light, pH, enzymes, and other factors to stimulate gels. This

review will encompass the recently synthesised carbohydrate-based gelators and their biomedical applications.

2. Monosaccharides and their derivatives

Out of the various organic LMGs, carbohydrates are an important class on the basis of their physical and chemical properties and abundant bioavailability. These properties include complex diversified stereochemical structure and unique biocompatibility, making them a suitable novel class of materials.^{27,28}

2.1. D-Glucose derivatives

Various monosaccharides and their derivatives have been found to be LMGs and LMHG^{29–42} and employed for various applications. A number of LMGs have been synthesised using D-glucose, which is easily accessible, by functionalizing derivatives of α -methyl D-glucose. Diol compounds **1** and their monoesters **2** and **3** and diester derivative **4** were systematically examined for their gelation properties as shown in Fig. 2.³⁶ It was observed that the esters having alkyl chains lacked the properties of good gelators, but the diester **4h** was found to yield gels in H₂O:C₂H₅OH in a volume of 1:1 at 5 mg mL⁻¹, while **3d** which is a 3-ester derivative formed hydrogel at



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Fig. 1 Structurally diverse carbohydrates employed for the design and development of gelators.



Fig. 2 Structures of glucose ester derivatives employed as LMGs.

7 mg mL⁻¹.³⁶ The esters with terminal alkynes **2a–2c**, **3a–3c** and **4a–4c** were found to be gelators for many solvents such as water and hexane and their study helped in design and synthesis of more appropriate gelators.

1-Deoxy glucose diol **5** was synthesized to study the effect of the substituent at the anomeric position on the properties of gels.³⁵ Acyl derivatives, monoesters and diesters **6–8** were also studied for their effect on the properties of gels. It was found that diesters **6a**, **6b** and **6h** showed gelation in various solvents such as ethanol, its aqueous solution and an aqueous solution of DMSO. Such 1-deoxy sugar monoester was reported to be more soluble in organic solvents than 1-methoxy analogues. The researchers found that while long-chain esters were effective gelators, the α -OMe group was crucial in causing this molecule to act as a gelator.³⁵

Carbamates on the basis of their hydrogen bond accepting and donating properties have also been found to be the effective functional groups to make carbohydrates behave as gelators. The carbamate's functionalized carbohydrates have been found to develop readily at low concentrations and to be compatible with a wide range of substituents. Many *O*-linked carbamates **9** were synthesized and studied,³⁸ as shown in Fig. 3.

It was reported here that alkyl derivative **9a** and cyclohexyl derivative **9d** were most effective as gelators in aqueous DMSO and ethyl alcohol. In aqueous ethanol, **9d** showed gelation at 0.91 mg mL⁻¹.³⁸ The carbamates **9b** and **9c** and their aromatic counterparts **9e–9f** exhibited strong gelation in ethyl alcohol and water mixture as well as DMSO and water mixtures. It was also reported that the naphthyl derivative **9g** gave gelation only in aqueous DMSO.



Fig. 3 Carbamate derivatives of α -D-glucopyranosides.

Fig. 4 Carbohydrate-based glycolipid derivatives.

Monosaccharide-based glycolipid derivatives were also reported to exhibit gelation properties in various aliphatic and aromatic solvents⁴³ (Fig. 4). Such glycolipids **10b** and **10g** were gelators in aliphatic solvents with a critical gelation concentration (CGC) of 0.8% (w/v). The compounds **10c**, **10e** and **10h** showed gelation at a CGC of 1.0% (w/v). It was observed in this study that the R' group on aniline is primarily responsible for the gelation properties of the compounds. Nitro and iodo groups on the linear chain **10a–d** showed good gelation properties. The substituent on the aniline ring was responsible for the self-assembly and stability of gels.

4,6-Benzylidene-protected alkyl glucosides⁴⁴ **11a–11h** have also been reported to be organogelators, as shown in Fig. 5. Similarly, phenyl boronic-protected alkyl glucosides **12a–12h** have also been reported as water-sensitive gelators, which are similar to 4,6-benzylidene-protected alkyl glucosides. Such gelators were effective in isopropyl palmitate, ethyl myristate, cyclohexane and toluene. The boronate gelators are water-sensitive and undergo hydrolysis, which destabilize the gel.⁴⁴ The sensitivity of the gelators approved their application as smart materials in topical drug delivery.

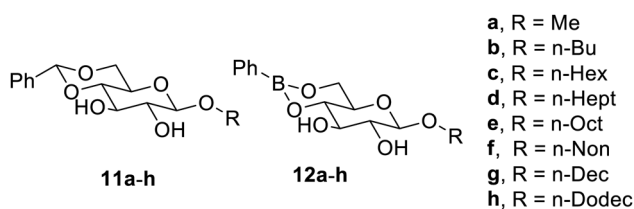


Fig. 5 Phenyl boronic ester-protected alkyl glucoside derivatives.



Fig. 6 Renewable sophorolipids as new gelator materials.

Similar tartaric acid derivatives of phenylboronic acid were reported to be forming a white metallogel in DMSO with LiOH. When D-(+)-glucose was added, this white gel turned red and exhibited enhanced conduction.⁴⁵ Therefore, these metallogels may be employed in the development of smart materials in the field of conduction.

Renewable sophorolipids have been reported to be supramolecular gelators.⁴⁶ The compounds **13a,b** (Fig. 6) have been found to form gels neither with water nor with the organic solvent but with the mixture of methanol, ethanol, DMF, DMSO and water in a ratio of 1 : 5. Compound **13b** undergoes sol–gel transformation at 48 °C. The fibrous structure of these gels was due to π – π interaction and hydrogen bonding.

Glucose-based LMGs have been reported as new materials that form gels in both water and organic solvents.⁴⁷ Such new materials have been found to be methyl α -D-glucopyranosides with an acetal 4,6-protecting group. It was found that the compound **14b** (Fig. 7) formed a silicon oil gel, which is an extruded gel and found to retain the viscoelastic behaviour of the gel. The gel formed by **14b** was able to restore itself once the shearing stress relieved at room temperature.

2.2. Glucosamine and N-acetyl-D-glucosamine derivatives

D-Glucosamine and N-acetyl-D-glucosamine derivatives have been reported to be used for the synthesis of organogelators and hydrogelators. The amino group here may be employed for the functionalization and synthesis of carbamates, urea, amides, *etc.*

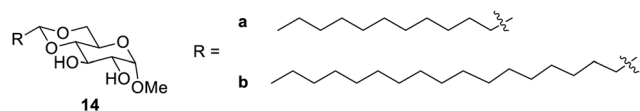
Fig. 7 Methyl α -D-glucopyranosides with acetal 4,6-protecting group as gelators.



Fig. 8 Glucosamine derivatives and dipeptoids.

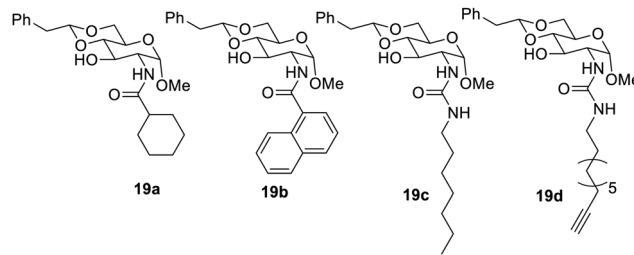


Fig. 11 Amide and urea derivatives as gelators.

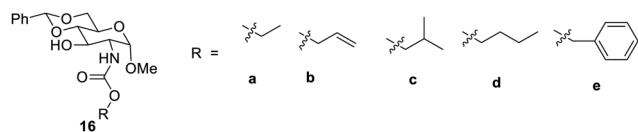


Fig. 9 N-linked carbamate derivatives from glucosamines.

2.2.1. Functionalization at C-2 positions. Dipeptoids⁴¹ **15a–c** were prepared by multicomponent reactions using glucosamine derivatives, as shown in Fig. 8. The synthesized compound showed gelation. The peptoids yielded gels *via* hydrophobic forces and π - π interactions.

N-linked carbamate derivatives³⁸ **16** from glucosamines, as shown in Fig. 9, gave a stable gel. Here the synthesized derivatives **16a–e** were explored for their gelation properties. The derivative **16a** gave a hydrogel at 6.6 mg mL^{-1} . The compound **16c** was found to form gels in aqueous ethanol, aqueous DMSO and hexane. It had minimum gelation concentration in $\text{H}_2\text{O}:\text{DMSO}$ ($v:v:2:1$) but yielded a hydrogel at 0.4%.

Gelation properties were also observed from urea^{48,49} and amide derivatives for a larger span of solvents. Urea derivatives act as a good gelator due to hydrogen bonding, which allows the molecule to congregate in the form of a three-dimensional network. Hydrogen bonding was also found to play a vital role in the gelation properties of amide derivatives.^{50,51} The glucosamine **15** was used for the synthesis³⁷ of amides **17a–n** and urea derivatives **18a–l**, as shown in Fig. 10. It was found that

most of the synthesized compounds formed gel at 33% aqueous solution of DMSO and ethanol and only a few compounds were able to form gels in either organic or aqueous solvents. The compound **17g** yielded a very stable gel at a minimum concentration of 0.7 mg mL^{-1} in aqueous ethyl alcohol. It was found that the compound **17m** behaved as a hydrogelator at a concentration 2 mg mL^{-1} . In this research³⁸ it was reported that a very stable gel was formed by heptyl amide **17d** in aqueous DMSO, which stayed intact for 6 months.

The alkyl derivatives of synthesized urea (**18a–l**) responded for the gel formation at lower concentrations than those of the aromatic derivatives. The compound **18i** formed a stable glassy gel in aqueous solvents. It was also observed in this research that gels formed by ester and carbamates are less firm than that formed by urea and amides.

A 4,6-protective phenyl group was explored with the synthesis of a new series,³⁴ in which the methylene group was flanked by a 4,6-protective site and sugar. The compounds of this synthesized series are shown in Fig. 11. It was observed that amide derivatives yielded gels in aqueous solvents, while the urea derivatives formed good gels in organic solvents. During the study of the gelation property in engine and pump oil, the amide derivatives were more prominent in gelation than urea derivatives. The compound **19a** was most efficient in forming the gel with six different solvents at lower concentrations. Compound **19b** formed a gel in isopropyl alcohol and an aqueous solution of ethyl alcohol and DMSO with a



Fig. 10 Urea and amide derivatives of glucosamines as gelators.



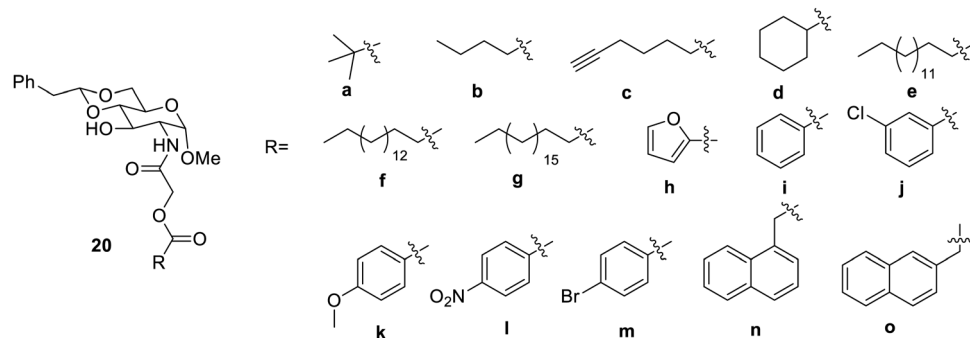


Fig. 12 Ester derivatives of the *N*-acetyl-D-glucosamine.

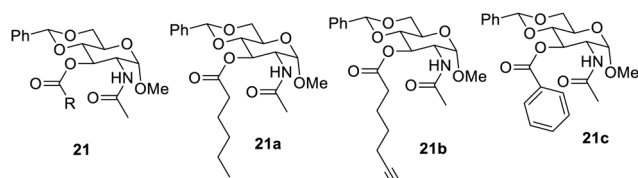


Fig. 13 C-3 ester derivatives of *N*-acetyl-D-glucosamines.

concentration of 3.3 mg mL^{-1} in aqueous ethyl alcohol. The compounds **19c** and **19d** made effective gels in ethylene glycol.

The modification of glucosamines was carried out further by introduction of ester group⁵² in the amide. Some of the synthesized compounds are shown in Fig. 12. The compound **20b** formed an effective gel in aqueous solvents at a concentration of 1.4 mg mL^{-1} . These gelators were found to interact with the enzyme lipase and other basic solutions. Therefore, these gelators were found to be potent new materials for the drug delivery and helpful in dye removal from the environment.

2.2.2. Functionalization at C-3 positions. Another series of ester derivatives⁵³ **21** were prepared by functionalization at position 3 of sugar using *N*-acetyl-glucosamines protected with 4,6-benzylidene (Fig. 13). These esters exhibited good gelation properties and were able to solidify various solvents. Compound **21a** was the most effective gelator able to form a gel with a large number of solvents such as glycerol, ethylene glycol, aqueous ethanol, DMSO and pump oil. Compound **21b** also showed good gelation properties.

Carbamate derivatives⁵⁴ of the sugar were also prepared using *N*-acetyl-D-glucosamines, as shown in Fig. 14. The synthesized compound acted as a gelator in most of the solvents and their binary mixtures. The derivatives **22b–g** formed gels with pump oil at a concentration ranging from 5 to 20 mg mL^{-1} . The aromatic substituted compounds showed gelation properties in aqueous DMSO. Compound **22j** was found to form gels in the presence of metal ions such as Fe^{2+} , Zn^{2+} , Cu^{2+} , and Pb^{2+} . It was observed that compounds **22j** and **22l** showed very good gelation properties, which further adds to the new gelation material.

2.2.3. Using unprotected D-glucosamines. The steroid triamcinalone acetamide, which is employed for treating eye inflammation, has certain drawbacks. Research has been reported⁵⁵ for the development of the prodrug to reduce side effects. In this research, compound **23** has been reported to be a gelator in an aqueous medium, which is formed by attachment of succinate and glucosamines to triamcinalone acetamide. This prodrug may be injected in the body and on the basis of its thixotropy property. The prodrug may be released at the active site on injecting into the body with least toxicity and maximum efficacy. This hydrogel is reported to be an effective new material for the treatment of uveitis (Fig. 15).

The multicomponent gelator^{56,57} is yet another type of the recent material to modify or alter the properties of the gelator. The binary system of the gelators is of three types: (i) one component forms gel and another does not, (b) both components forming gel independently and (iii) both components do

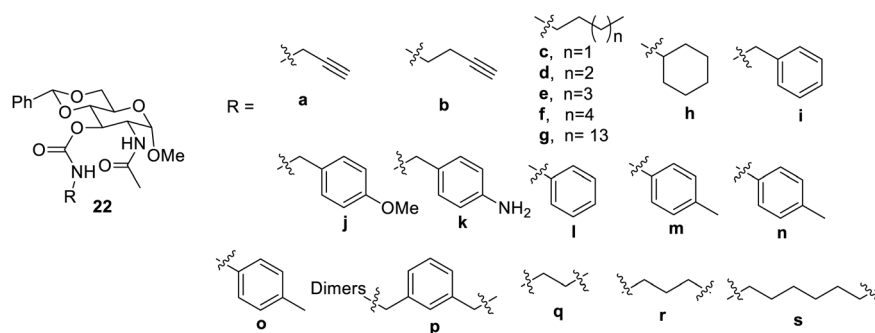


Fig. 14 C-3 carbamates of *N*-acetyl-D-glucosamines.



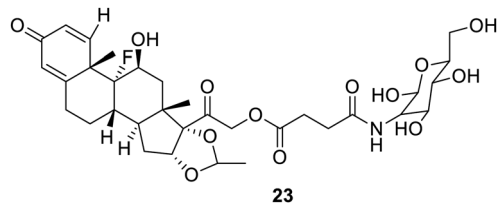


Fig. 15 Triamcinaolone acetamide substituted with succinate and glucosamines.

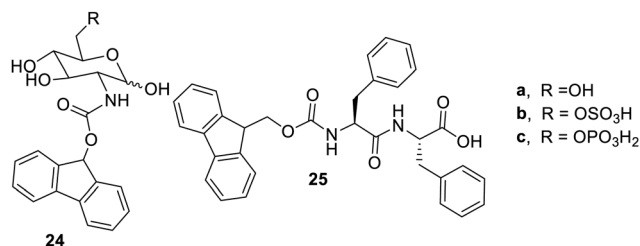


Fig. 16 Fmoc-glucosamine derivatives as new materials.

not form gel but their mixture makes gel easily. The first type of the material is used when no change in the properties of the gel is required. The second one gives rise to the material whose properties depend upon the assembly of the components at the molecular level.⁵⁷

The gelation properties of Fmoc-Glucosamine derivatives⁵⁸ were studied for the two-component system in cell culture. The compounds **24a** and **25** as shown in Fig. 16 were soluble in water with low solubility. The solubility of compound **24a** was enhanced by compounds **24b** and **24c**. The compound **25** co-assembled with **24b** and **24c** to undergo gelation.⁵⁹ The advanced research has mentioned that binary component gels have reduced toxicity and may be used as biomimetics. Compound **24c** (*N*-fluorenylmethoxycarbonylglucosamine-6-phosphate) is a gelator precursor and dissociates alkaline phosphate and yields gelator **24a**.⁶⁰ The tumor cells in breast cancer and osteosarcoma have excess of alkaline phosphate and compounds **24a** and **24c** due to their sugar moiety binding GLUT1 (glucose transporter 1), and they stop glycolysis in the cell. The cleaved precursor **24c** *via* ALP, the resulting molecule undergoes self-assembly and disturbs the cell function and promoting cell death to the tumor cells. This biomaterial may become effective for the treatment of cancer.

2.3. D-Mannose, D-arabinose and D-galactose derivatives as LMOGs

Low-molecular weight organogelators (LMOGs) based on mannose **26** have also been reported which have self-healing and phase selective properties in gelation,⁶¹ as shown in Fig. 17. They have been found to gel a large number of organic solvents, oils, fuels, alcohols, aromatic solvents, *etc.* Arabinose glycosides⁶² **27a–c** were synthesized and studied for their gelation in aforesaid solvents. Their gelation action in the fuels was reported to be very effective for cleaning oil spills. An important

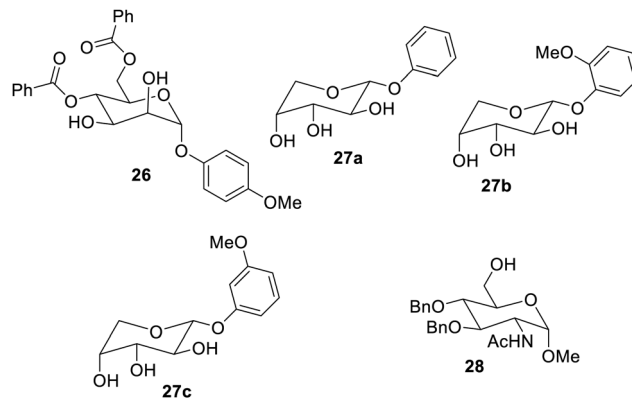


Fig. 17 D-Mannose, D-arabinose and D-glucose derivatives as gelators.

multifunctional gelator **28** based on glucose has also been reported, which has the ability to form organogels and hydrogels. Gelator **28** shows the shortest gelation time (5 s) for *N*-acetyl-glucosamine-based gelators, and it was also proved to be efficient not only for organic solvents but also for oils such diesel and petrol. Organogels of the same gelator have been proven to be useful for the removal of waterborne synthetic dyes, whereas their hydrogels were explored as growth media for the fabrication of gold nanoparticles⁶³ (Fig. 17).

Galactose derivatives have also been reported as new materials on the basis of their gelation. The combination of squaramides and galactosyl moieties was used to synthesize⁶⁴ low-molecular weight gels, as shown in Fig. 18. It was found that the symmetrical triazole derivatives **29a** and **29b** were weak gelators, giving gels only in ethyl alcohol and ethyl acetate. However, the amphiphilic derivative **30a** showed very good gelation properties in a large number of solvents with lower gelation concentrations. The amphiphilic derivative **30b** after amputation of the acetyl group under basic conditions behaved as a supergelator in aqueous ethyl alcohol (1 : 1) with a minimum gelation concentration of 0.1% w/v. This research gave a new material, which shows potential for novel applications.

2.4. Glyconamide derivatives by modification at the anomeric position

Various linear derivatives have been reported as gelators, synthesized by C-1 anomeric functionalization. The amide and ester derivatives of glyconic acid and glyconolactone have been reported as new gelation materials.

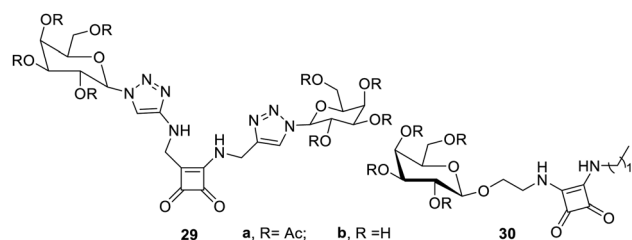


Fig. 18 Novel glycosyl squaramide gelators.



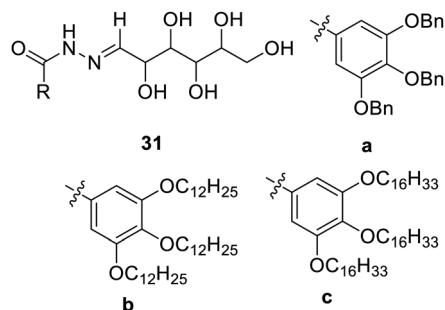


Fig. 19 Poly aryl ether glucose-based low-molecular weight gelators.

Poly alkyl ether and poly aryl ether glucose-cored LMGs have been reported.⁶⁵ Strong gel formation has been reported by poly aryl ether glucose cored low-molecular weight gelators **31a–c**. The fibrous networking of the gel makes it a good novel material for the controlled release of the drug (Fig. 19).

The reaction between galactonolactone and amines such as hexyl, heptyl, and octyl amines yielded galactonamides⁶⁶ **32a**, which have been reported to be effective gelators in various solvents (Fig. 20). The minimum gelation concentration for these was 1%, 0.45% and 0.5% for hexyl, heptyl and octyl derivatives respectively. These gels have been successfully employed for the growth of neuronal cells. *N*-Heptyl galactonamide (**32b**) in DMSO has been reported to form an injectable gel in water bath.^{67,68}

The gluconamide amphiphiles were extracted from cashew nut. The derivatives⁶⁹ **33a** and **33b** were able to form gels easily. The former derivative formed gels in water and the latter in the organic solvent at a concentration of 1% w/v and 0.8% w/v respectively. The derivatives were found to contain a fibrous network characteristic of the supramolecular gel and possess notable antibacterial properties (Fig. 21).

D-Gluconic gelators⁷⁰ have also been reported, which are similar to gluconamide-based gelators. Such gelators contain 3,4-chlorobenzylidene acetal-protecting groups, and are shown in Fig. 22. Among the synthesized gelator, compound **34c** was most prominent and found to form gels in a large number of organic solvents and aqueous mixtures. The gel of compound **34c** in *n*-butyl acetate has exceptional extraction properties for pump oil at room temperature and has good mechanical strength. The gel of compound **34c** in chloroform was sensitive to anions. These gelators were found to form moldable clear gels, which may be employed as valuable materials in the optics.

The interaction of the terminal amino group on *D*-gluconic acetal derivatives with aliphatic acid leading to the formation of

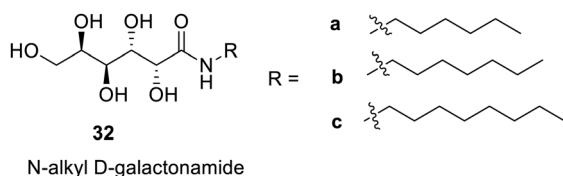


Fig. 20 Galactonamide-based gelators.

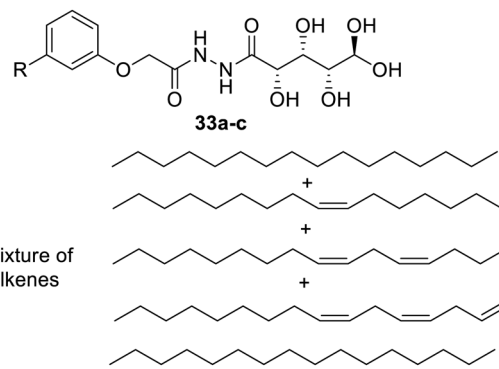


Fig. 21 Gluconamide derivatives as gelators.

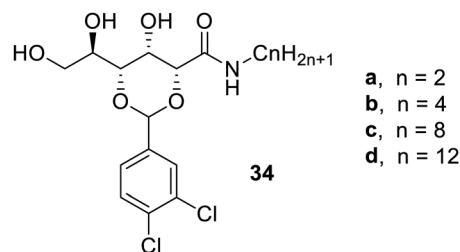


Fig. 22 *D*-Gluconic acetal-based gelators.

two-component gels was also studied.⁷¹ It was found in the study that the gelation property was dependent on the aliphatic chain length **35b–d** in comparison to **35a**. The compound **35c** was reported as a super gelator in chlorobenzene and *o*-xylene with a concentration of 0.1% w/w. The compounds **35b–d** in comparison to **35a** were able to give a co-gel with *o*-xylene, dioxane, and chlorobenzene (Fig. 23). The compound **35c** formed strong, self-healing co-gels with stearic acid, myristic acid, and capric acid in chlorobenzene. This co-gel was clear and may be used in the optical apparatus. The compounds **36a–b** were good gelators in various organic solvents such as nitrobenzene, chlorobenzene, benzene, and toluene at a minimal concentration of 0.06–0.19% w/w.⁷²

2.5. Alditol derivatives from reduced sugar alcohols

According to a study,⁷³ 1,3-dibenzylidene-*D*-sorbitol and 2,4-dibenzylidene-*D*-sorbitol (DBS) can produce gels in polyethylene glycol (PEG) that is impacted by silica due to the disruption of the hydrogen connection between DBS and PEG. The inclusion

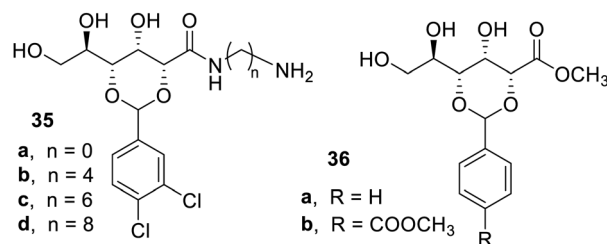


Fig. 23 *D*-Gluconic acetal-based gelators.





Fig. 24 Sorbitol monoacetal and diacetal as gelators.



Fig. 25 D-Mannitol derivative for gelation in ionic liquids.

of silica had an impact on the temperature and time of gel formation. A spherulite structure with lamellar packing was discovered through structural analysis. This study brought forward the concept of modification of the self-assembly in solvents by the addition of silica, which led to the development of versatile hybrid materials.

Another research developed mono- and di-benzylidene-protected sorbitol low-molecular weight gelators,⁷⁴ as shown in Fig. 24. Compound **37b** was not a hydrogelator but was able to form gels in most of the solvents and in aqueous ethyl alcohol. Compound **38a** was the best hydrogelator, while the compounds **38b** and **38c** did not show any gelation properties. These multicomponent gels may be employed in the tunable soft materials, which have numerous applications.

Carbohydrate-based LMGs, as shown in Fig. 25, have been found to form gels in ionic liquids.⁷⁵ Ammonium phosphonium, pyridinium and imidazolium-based ionic liquid gels were obtained in this research. It was found that a higher amount of the gel was required in the case of bulky cations if the ionic liquid. The gelator **39** was able to form gels with most of the ionic liquids. Ionic liquid gels were found to be thermally more stable than the ionic liquid itself. Reversible ionic liquid gels may play numerous roles in the electrochemistry.

Xylitol-based⁷⁶ organogelators have also been reported as shown in Fig. 26, which yield gel in crude oil and most of the organic solvents. Compound **40a** was found to be a powerful



Fig. 26 Xylitol-based organogelators.

gelator for crude oil and organic solvents. The gelation of the crude oil and refinery effluent was observed to be very fast within minutes. Thus, such gelators proved themselves as new materials, which may be used in the refinery process to minimize the waste.

2.6. Glycosyl triazole derivatives as molecular gelators

Triazoles and other heterocycles are also used as functional groups in low-molecular weight gelators. The hydrogen bond acceptors and donors of triazoles make it simple to create them by the click reaction, and they also readily establish π - π interactions. Such properties of the triazole make it a valuable low-molecular weight gelator.^{77,78}

2.6.1. C-1 triazole glycoside derivatives. A series of self-assembling sugar triazoles were synthesized from D-maltose, D-glucose and D-glucosamine triazole derivatives, by cycloaddition of sugar azides and alkynes. These sugar triazoles have also been found to yield new materials for gelation,^{31-33,40} as shown in Fig. 27. Several analogs to the triazole moiety at anomeric position were synthesized by long-chain alcohol derivative **41c**, which was a gelator in aqueous solutions, polar solutions and a mixture of both solutions. The compound **41a** formed gels in water and DMSO in the ratio 1 : 1 and 1 : 2. The compounds **41c** and **41e** were also found to be good gelators. The derivatives without polar groups **41b** and **41d** were able to yield gels in isopropanol and in a mixture of ethyl alcohol and water in a ratio of 1 : 2.

The dimers of these sugar triazoles **42a-b** were found to be more promising gelators than the monomers (Fig. 28). The compound **42a** formed gels in ethanol and toluene as well as the aqueous mixture of DMSO and ethanol.^{31,38}

Some effective low-molecular weight peracetylated D-glucosamine triazole gelators were obtained from D-glucosamines⁴⁰ (Fig. 29). The compounds **43a-d** and **43a-b** were hydrogelators



Fig. 27 Gelators based on glucosyl triazole moieties.



Fig. 28 Dimers of sugar triazoles as gelators.





Fig. 29 Triazole derivatives of D-glucosamine gelators.



Fig. 30 Benzylidene acetal-protected triazole glycoside gelators.

and yielded gels in aqueous DMSO and alcohol. The derivatives **43a–b** were reported to be gelators in toluene. The dimers of these **44b** and **44c** are still more effective gelators and represent effective hydrogelators.

The influence of the functional group on making the molecule an effective gelator was also studied.⁷⁹ For this protected sugar, triazoles with 4,6-*O*-benzylidene acetal **45** and **46** were formed from azides. The compounds **45b** and **45c** were found to possess effective gelation properties for engine oil. The compound **45d** yielded gel in aqueous DMSO. The *N*-acetyl D-glucosamine triazole derivatives **46a** and **46b** yielded gel in aqueous DMSO and ethanol as shown in Fig. 30.

D-Galactose substituted with benzylidene acetal-protected triazoles was found to be an effective gelator,⁸⁰ as shown in Fig. 31. It was observed that **47a–c** were gelators for oils, while the aromatic compounds yielded gels in organic solvents. The

gelator containing an anthracene ring was found to be fluorescent, and shows potential for use in medical diagnosis and optical applications.

Triazolylarabinosides⁸¹ have also been reported as organogelators, as shown in Fig. 32. The derivatives **48a** and **48g** acted as supergelators in kerosene, diesel and petrol, and hence, they may be employed as new materials for the solidification of crude oil and its various products, which are used for the purification and recovery of oils.

2.6.2. Triazole at other position than anomeric position in carbohydrate. LMGs have also been obtained from peracetyl glucosamine triazoles.³² Triazoles at 2-acyl positions are shown in Fig. 33; however, triazoles may also be linked at other positions. The modification of structure made the gels stronger, which is due to stronger hydrogen bonding and π - π bonding. The gelators produced **49a–r** have shown good



Fig. 31 Benzylidene acetal-protected triazoles as gelators.





Fig. 32 Triazolyrarabinoside as a gelator.

gelation in aqueous ethanol and DMSO. Compound **49a** was the best hydrogelator along with **49f,g**, **49i,m** and **49o,q**. The reported gelators may be treated as new soft materials synthesized from carbohydrates.

The *N*-acetyl-glucosamines having triazoles linked at C-6 positions (Fig. 34) have also been synthesized and reported to be good gelators in oils and organic solvents.⁸² The compounds **50a,b** and **50f** were found to exhibit phase-selective gelation in crude oil, which became incapable of gelation in powdered form using ethyl acetate as a carrier.

2.6.3. Nucleoside and glycosyl triazole hybrids. Uracil-based gelators have also been synthesized.⁸³ The compound **51** showed microstructures, which confirm the existence of gel-like structures in 2% of ethyl alcohol and methyl alcohol. The gel formed here exhibited thixotropy (sol-gel conversion) on treatment with stress. Such uracil-based gelators add new materials as injectable biomaterials, as shown in Fig. 35.

Bolaamphiphiles which were generated with nucleosides were reported to form hydrogels at a lower gelation concentration.⁸⁴ The compound **52** was found to remain intact for almost less than a month in mouse without any toxicity and immunogenicity. Therefore, this compound is reported to be a new biomaterial, which may find many applications in the live tissues, as shown Fig. 36.

The disulfide linkage in the bolaamphiphiles **53**⁸⁵ was also explored for the sustained release of proteins containing thiols. The thixotropic nature of the resulting gels made them suitable for injecting into the body. The disulfide group enables the thiol-disulfide exchange, so these hydrogels may be employed for their functionalization with thiol-containing proteins for their sustained release in the tissues (Fig. 37).

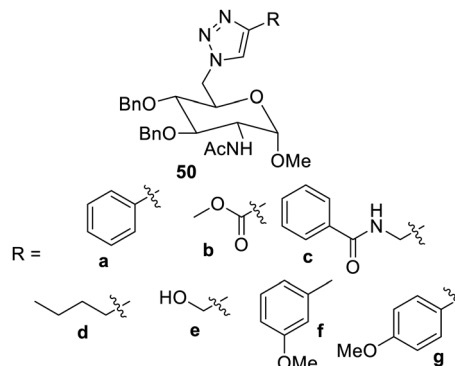
Fig. 34 *N*-Acetyl-glucosamines having triazoles linked at C-6 positions.

Fig. 35 Uracil gelator amphiphiles.

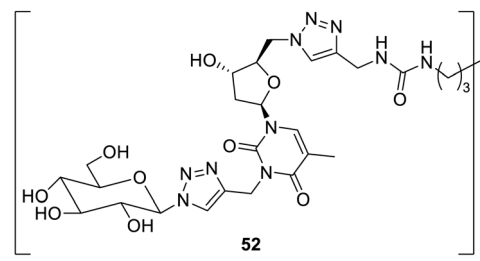


Fig. 36 Urea based amphiphiles as gelators.

2.7. Disaccharide derivatives and glycoclusters

2.7.1. Disaccharides. To explore the disaccharide gelators, various peracetyl triazole derivatives of lactose and maltose

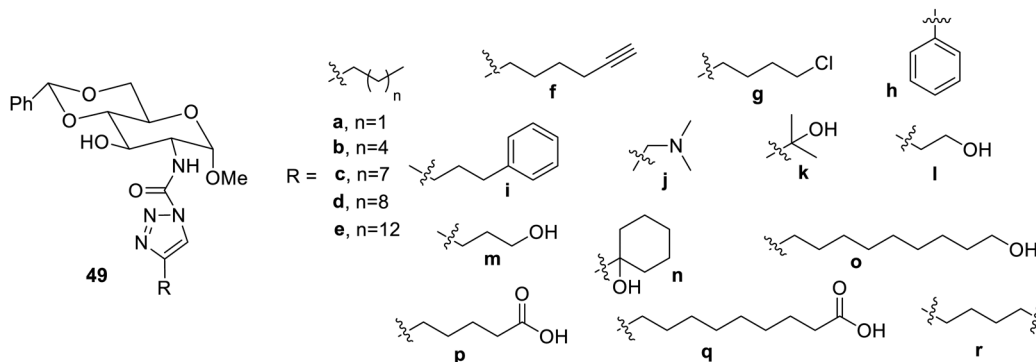


Fig. 33 Triazole gelators with repeating chains.



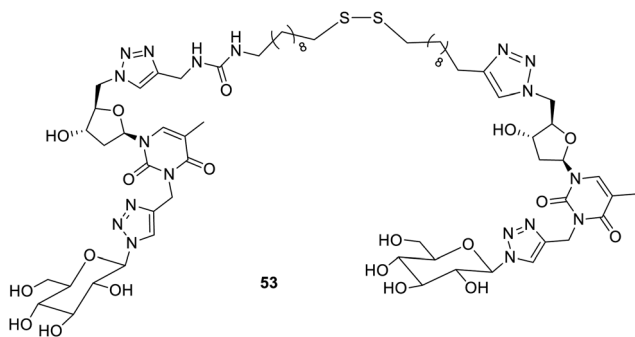


Fig. 37 Disulfide linkage in the bolaamphiphiles containing urea.



Fig. 38 Maltosyl triazole derivatives as gelators.

were studied.³³ It was found that maltose yielded better gelators in various solvents. The compound 55 at a concentration of 2.0–5.0 mg L⁻¹ was the most effective gelator in aqueous DMSO and ethyl alcohol, as shown in Fig. 38.

Thiolactose-based amphiphiles have also been reported as low-molecular weight gelators, which yielded colloidal solutions in water, as shown in Fig. 39.⁸⁶ The compound 56a yielded hydrogels easily at lower concentrations. The compound 56b showed chain inversion. The binding nature of these amphiphiles to the peanut agglutinin lectin lead to the conclusion that these structures may be helpful in designing nanostructures to study sugar-specific ligands.

2.7.2. Branched glycoclusters. Glycoconjugates are dendritic compounds that lie in between small molecules and polymers, which may be tailored to meet the specific physical and mechanical requirements of the materials.^{87,88} Glycoconjugates incorporated with catalysts may be employed in the synthesis. The dendrimers have uniformity in the molecular weight, and hence, they are preferred over polymers for introduction of required functional groups. The dendritic gelators made up of LMWG subunits are potential new materials which are stable,

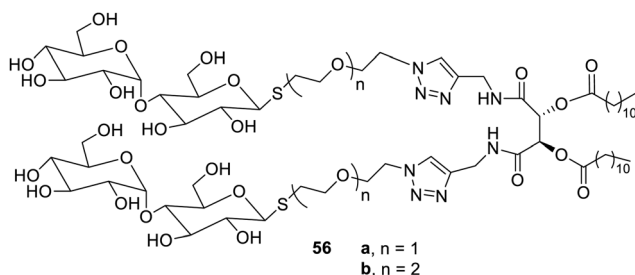


Fig. 39 Thiolactose-based amphiphilic gelators.

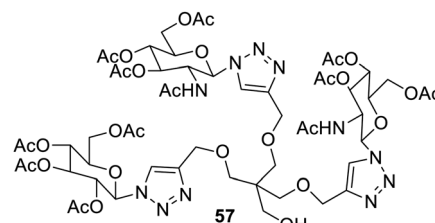


Fig. 40 Branched glycoclusters as gelators.

flexible, with better functionality than LMWG. The trimeric glycoconjugate 57, as shown in Fig. 40, was reported to be an effective hydrogelator, which may be employed for the sustained release of Vitamin B12, riboflavin and naproxen sodium.

The dendritic glycoclusters have been synthesized⁸⁸ and the compounds with few branches were appreciable hydrogelators. Hexameric and trimeric gelators were combined to produce co-metallogels, which can be used in supramolecular catalysts that can be recycled and used repeatedly. Such materials may reduce the use of metal ions in the catalysis for the synthetic processes.

2.8. Nucleotide and nucleoside-based gelators

Nucleotide and nucleoside are the self-assembled and may be employed for supramolecular studies. They are basically ribose sugar having pyrimidine or purine forming intermolecular attraction *via* hydrogen bonding, electrostatic interactions, and π - π interaction.⁸⁹ Out of the four nucleobases, guanidine is the most desired molecule for such gelator systems, as it has biochemically notable extended two-dimension ribbon with G-4 wires. Supramolecular substances based on nucleic acid are significant for biotechnical and biomedical applications.⁹⁰

2.8.1. Deoxy ribose-based gelator. Cytosine-based gelator⁹¹ 58 is shown in Fig. 41. *N*⁴-Octanoyl-2'-deoxycytidine 58 was one such hydrogelator, which exhibited fluorescence in an aqueous ethanol solution due to π - π interaction and protected chromophore quenching, leading to enhanced absorption and fluorescence. These self-assembling gelators may find application in drug delivery systems.

Further research has been done on guanosine-based⁹² gelators. Among these gelators, the 8-aza-20-deoxyisoguanosine derivative 59 (Fig. 42) was discovered to be fluorescent under UV light, and exhibits thixotropy, where the gel form was restored as soon as the radiation was turned off. The gelation



Fig. 41 Cytosine-based gelators.



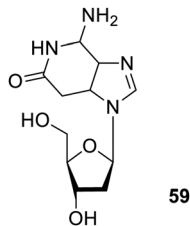


Fig. 42 Guanosine nucleoside as a gelator.

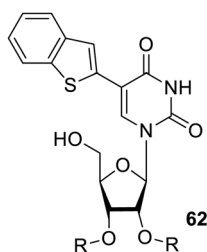


Fig. 43 Cytosine and guanosine derivative-based lipid nucleotides.

process was temperature and pH dependent, which make it a potential molecule in the nanomedicine.

Nucleotide-lipids⁹³ having pyrimidine and purine bases have also been reported, as shown in Fig. 43, which were able to yield hydrogels. Cytosine and guanosine derivatives **60** and **61** are the main reported compounds, as shown in Fig. 44. Among them, cytosine hydrogels were stabilized by NaCl but guanosine hydrogels stabilized even in the absence of NaCl. It was established that the supramolecular structure of such hydrogels was dependent on nucleobase employed along with other conditions. These compounds **60** and **61** were further studied for the slow release of diazepam from the matrix of the gel, and it confirmed the potential of this gelator for controlled delivery of drugs.

2.8.2. From ribose derivatives. Guanosine-based hydrogels⁹⁴ formed by adding 4% (w/v) guanosine and pyridine-4-boric acid in 0.14 M KCl (aqueous) were found to be stable for quite long time and the sol-gel transition may



- a, R = octanosyl
b, R = myristoyl
c, R = palmitoyl
d, R = oleyl

Fig. 44 Benzothiophene-based nucleolipid gelators.

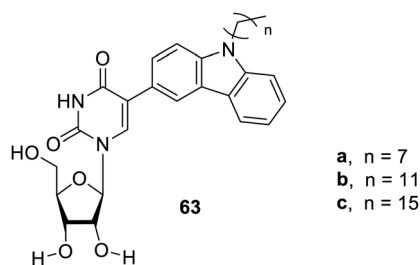
be maneuverer by adjusting the components. They exhibited imbibition in aqueous solvents and have shown appreciable conductivity. The gel formed may be used in electrochemical biosensing.

Guanosine monophosphate⁹⁵ crosslinked with Ca^{2+} and Fe^{3+} was synthesized, which exhibited pH-dependent zero-order doxorubicin release, which is a well-known chemotherapeutic agent. In the acidic buffer, the release of the drug was at fast rate. This hydrogel was reported to be stable at a pH of 7.4 for three weeks, and there was release of just 3% of the drug. This establishes the metallo gel as a potential candidate for drug delivery.

2.8.3. Fluorescence functions of nucleotide. 5-Benzothiophene⁹⁶ on modification gave a series of compound **62**, as shown in Fig. 44. The compounds **62b,c** yielded gels in DMSO at low temperatures. The compounds in DMSO were fluorescent under UV radiation. It was noted that the gelator **62c** was found to be undergo sol-gel transition on changing the temperature and sonicating. The gel was found to be unstable toward Hg^{2+} among several other metal ions added to the gel. The gel was also found to be stable in various anions such as I^- , Cl^- , Br^- , and HSO_4^- . This gelator shows potential to be used as an optical material, which is responsive towards various stimuli.

Carbazole⁹⁷ gelators **63a-c** (Fig. 45) have also been reported to possess numerous properties such as electrical conduction, charge carrier, and luminescence. These are presently employed as chemo sensors for fluorescence imaging and several other related biomedical applications. The exceptional self-assembling property of these gelators is due to the hydrogen bonding. These gelators may forecast numerable morphologies in various solvents, which need to be explored for their future applications.

It was reported that the enzymatic self-assembly triggered adenosine-based gelators,⁹⁸ as shown in Fig. 46. Treatment of a mixture of hydrogelator naphthalene-Phe-Phe-Lys **64** and adenosine with phosphoryl chloride yielded a phosphorylated form of hydrogelator **65**. The dephosphorylated form **64** has fibrous structures, while the phosphorylated form does not have any fibrous structure and the self-assembly here was due to the enzyme. This work represents the enzymatic action for the production of hydrogels using adenosine nucleoside.



- a, n = 7
b, n = 11
c, n = 15

Fig. 45 Carbazole nucleosides as gelators.



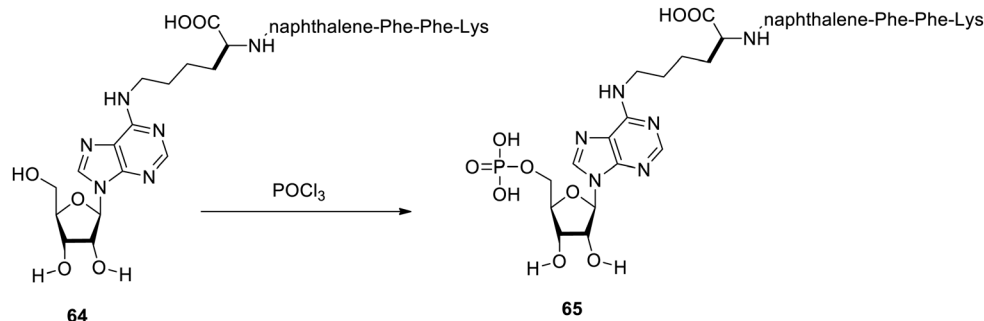


Fig. 46 Adenosine monophosphate-based gelators.

3. Stimulus-responsive gels

Smart materials are in great demand in the diagnosis and treatment of diseases. One such substance that uses chemicals to stimulate biomolecules is a stimulus-responsive gel.⁹⁹ The stimulation may be physical, chemical or biological. The enzymes which are biological stimuli have advantages for the peptide bond generation, and the other product is generally water. The physical stimulation is caused by electric field, magnetic field and temperature or radiation. The chemical stimulation includes pH changes and specific ion-presence. The sol-gel transition is reversible and may be employed for the stimulation. Stimulus-responsive hydrogels are biocompatible and are used in the immunotherapy as non-immunogenic vaccines. Many more applications of the stimulus-responsive gels need to be explored for biomedical applications.

3.1. pH-Responsive gelators

The gelators have been designed to work on change in pH as external stimuli. The pH stimulation can be of two types. The first one are those which work on protonation and deprotonation,¹⁰⁰ while the second one are those which work on dissociation of the gelator functional groups on changing the pH of the medium. The first type sol-gel transformation work on the principle of production of positive and negative charges on the system by protonation and deprotonation of the gelator respectively.¹⁰¹ The second type of the gelators have labile pH-dependent functional groups,¹⁰² which break away as the pH is changed. Nano particles based on cytidine-5'-monophosphate (5'-CMP)-mediated akaganeite (β -FeOOH) were used to give hydrogels,¹⁰³ in the pH range of 5.5–8.5 and the temperature range of 30–40 °C. The hydrogel gave a colloidal solution at a lower pH. The porous nature of the hydrogel was reported due to the conformation change from C2' endo to C3' endo. This change enhanced the non-covalent forces. The presence of the ribose sugar made this gelator non-cytotoxic, injectable, and self-healing. Therefore, it may be used for the loading and release of the drug at the desired pH in the organic tissues, thereby making it a potential biomedical material.

Carbohydrate-based pH-responsive D-glucosamine gelators have also been studied, as given in Fig. 47. The protecting group of benzylidene acetal in **66** was changed to more labile

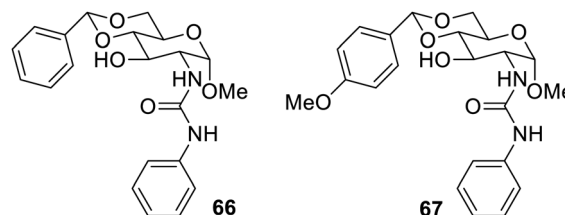


Fig. 47 Carbohydrate-based pH-responsive D-glucosamine gelators.

and pH-responsive *p*-methoxybenzylidene acetal in **67**. The gelator **67** dissolved completely in sulfuric acid in 48 hours. This enables the gelator **67** suitable for the drug release in acidic media.

3.2. Photosensitive gelators

Light has also been used to stimulate the gelation action of several gelators. Light or radiation-controlled transformation of gels may enhance the potential of gelators in many biomedical applications.¹⁰⁴ Carbohydrate moieties with photo-responsive azobenzene functional groups have been reported as photo-sensitive gelators.^{105,106}

3.3. Photo-isomerizable gelators

Besides sol-gel transformations, there are several other changes which occur in response to the light falling on the gelators. Bolaamphiphilic glycolipid LMGs **68** having *N*-alkyl-2-anilino-3-chloromaleimide have shown additional thermochromism,¹⁰⁷ as shown in Fig. 48. This material upon increase in temperature became transparent on the alteration of the congregation without affecting the assembly of gels. The bathochromic shift was observed under UV-visible radiation. This research has given an overview of the application of gelators responsible for glycol-related enzymes.

Supramolecular gelators responsive to multiple stimuli have also been reported. Glucose-based multi-stimuli-responsive supramolecular gelators¹⁰⁸ **69–72** containing a photosensitive aryl azo moiety and a metal and pH-responsive hydroxyl group have been synthesized, as shown in Fig. 49. Upon exposure to light, the sugar azobenzene derivatives collapsed on treatment with UV radiation for 30 min. The exposure to the visible radiation changed it to the solution in 2–4 days. The gel was





Fig. 48 Gelators having *N*-alkyl-2-anilino-3-chloromaleimide.

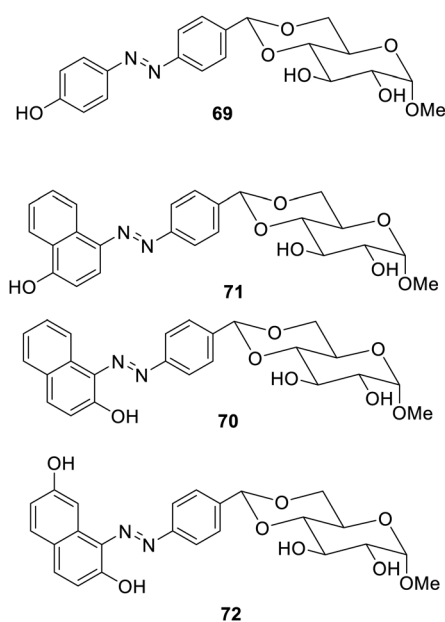


Fig. 49 Glucose-based multi-stimuli-responsive supramolecular gelators.

found to become a solution at a pH higher than 12.4 and lower than 1.6.

Another azobenzene carbohydrate macrocyclic low-molecular weight gelator has been reported which shows *cis-trans* isomerism on stimulation with the radiation, as shown in Fig. 50.¹⁰⁹ The azo moiety predominates in *cis* form 73a with radiation at a wavelength 365 nm, while *trans* form 73b dominates at a wavelength of 435 nm. Half-life of the *Z* form 73b is 51 days. These gelators may be employed as new photo responsive, biocompatible materials.

Stilbene has been introduced in the gelator to make it photosensitive. Glyconucleoside bolaamphiphilic low-molecular weight gelators¹¹⁰ have been reported as photo-sensitive gelation. The compounds 74 and 75 were found to form gels in aqueous ethanol (Fig. 51). The *E-Z* isomerism in stilbene upon UV radiation causes sol-gel transformation. The UV irradiation with a 312 nm deformed supramolecular structure lead to only *Z*-isomers in dominance. These photo-responsive materials may be considered to replace the

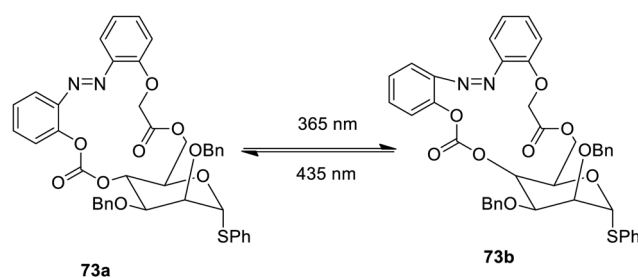


Fig. 50 *cis-trans* isomerism in azobenzene carbohydrate.

polymer-based radiation stimulation employed in drug delivery.

3.3.1. Photoreactive gelators containing diacetylene. Poly-diacetylene exhibits colour changes upon interaction with environmental changes and biomolecules. They follow non-linear optics and the colour changes make them suitable for making new materials, which are photo-responsive. The diacetylene group when incorporated on the carbohydrates yield significant multi-stimulant gelators, which may be employed for making sensors. Diacetylene containing glycol lipids 76–79 have been studied for their gelation in aqueous and organic solvents. Novel materials with required properties may be formed by the crosslinking of the polymeric groups. Blue-Red transition was shown by the diacetylene compounds³⁰ on changing the temperature. Polymerized diacetylene gelators are shown in Fig. 52.

The compounds 79f and 79g polymerized on exposure to the UV-radiation and may be used as new materials for photo-stimulating applications.

Urea and amide functional groups may also be used for synthesizing the gelators,⁴² as shown in Fig. 53. These compounds yielded gels in numerous solvents and their aqueous mixtures. Compound 80c gave gel in toluene, ethanol and isopropyl alcohol easily with low gelation concentration followed by compound 81b. These molecules were reported to be sensitive towards UV radiation proposing to be new materials in this field.

Gelators based on β-D-glucopyranoside diacetylene have been reported as shown in Fig. 54.¹¹¹ Compound 82 yielded gels at low concentrations in non-polar solvents and gels. The diyne groups existed in close proximity in gels facilitating the



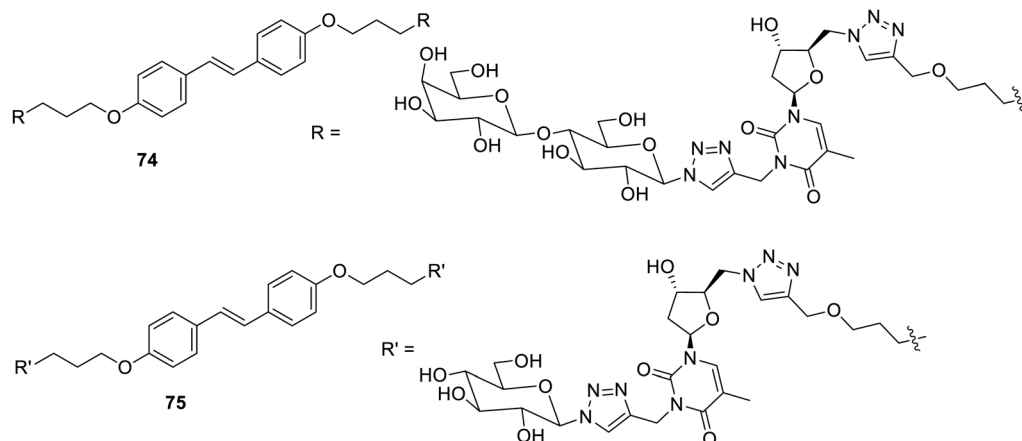


Fig. 51 Glyconucleoside bolaamphiphilic low-molecular weight gelators with stilbene.

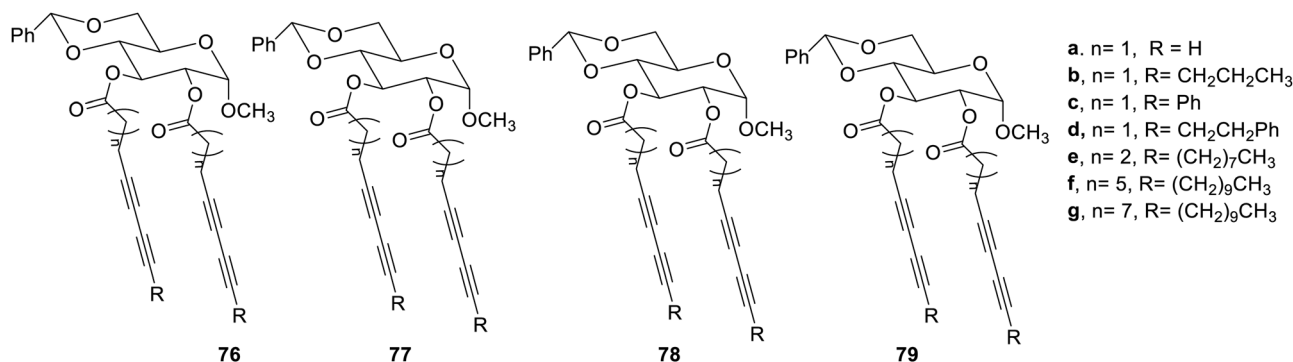


Fig. 52 Diacetylene glycolipids from D-glucose.

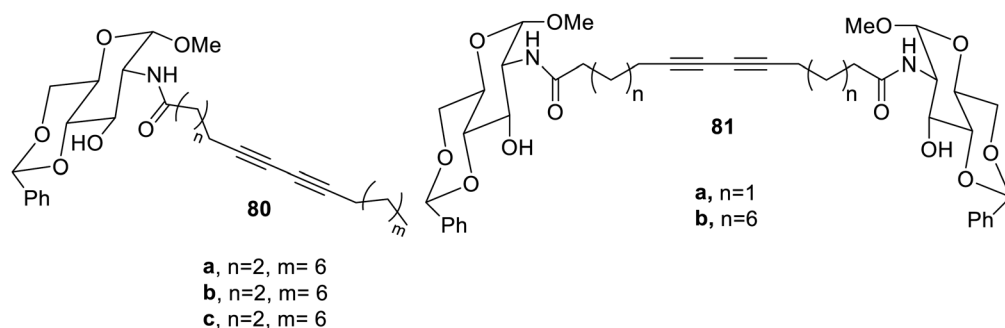


Fig. 53 Diacetylene with urea and amide derivatives as gelators.

polymerization on exposure to the UV radiation (300 nm) for two days. The resulting polymer **83** has exceptional attraction for galactose-binding lectin, which was much more than that of monosaccharides. Such materials pose themselves as biomaterials with exceptional binding with lectin.

3.4. Enzyme-responsive gelators and biomedical applications

The gelation process has also been induced by the enzymes¹¹² which are catalysts for the synthesis in biochemical processes.

The enzymes may be employed for the drug delivery or delivery of the specific biomolecules at the specific site. If the gelator has the enzymatic moiety, then on degradation of the enzyme in a specific environment, the gelator will also degrade leading to the release of desired drugs or the biomolecule.¹¹³ Enzymatic stimulation is preferred than other stimulations, as they are harmless, more specific, and selective in their action. Carbohydrate-based materials as gelators whose gelation is induced by the enzymes are lesser





Fig. 54 β -D-Glucopyranoside diacetylene as a photo-stimulating gelator.

in number than those of peptide and other polymer-based gelators.

The enzyme β -galactosidase has been exploited frequently for the stimulating supramolecular gelators. Glyconucleo bolaamphiphiles (GNBA) have been employed for the generation of stimuli-responsive materials.¹¹⁴ Gelator precursors **84** and **85** based on the lactose were generated and used to attach nucleotides with carbohydrates (Fig. 55). The compound **84** gave a viscous solution in PBS. While the compound **85** yielded hydrogels. Both the compounds **84** and **85** yielded hydrogels **86a** and **86b** upon treatment with enzyme β -galactosidase, and it was found that the supramolecules formed after enzymatic reactions had good mechanical strength. This study concluded that materials which are stimulated by enzyme β -galactosidase may be employed to alter the internal cell functioning, which may be utilized in therapeutics.

Supramolecular 4-nitro-2,1,3-benzoxadiazole attached with diphenylalanine and tyrosine through β -galactose.¹¹⁵ The solubility of the precursor **87** (Fig. 56) was enhanced by the sugar attached to tyrosine, which made the cleavage by β -galactosidase yielding self-assembly **88**. The cleavage of enzyme took place at 37 °C in saline solutions buffered with phosphates. The gelator precursors on incubation with HeLa cancer cells resulted in the self-assembly of the compound, and this

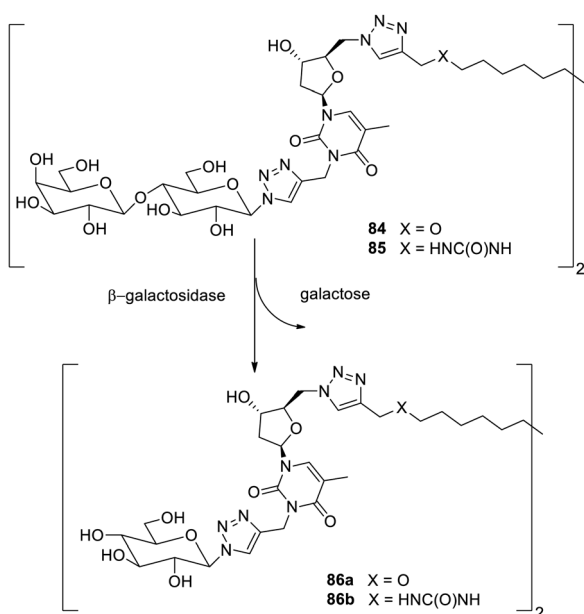


Fig. 55 Glyconucleo-bolaamphiphiles as gelators stimulated by enzymes.

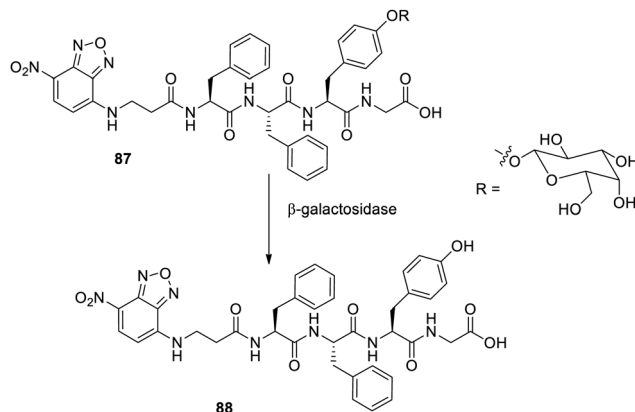


Fig. 56 Gelator precursor for β -galactosidase.

material may be employed for detecting the type and extent of these cells. The results of the research also confirmed toxicity on cancer cells, which was due to the self-assembly inside the cell occurring upon cleavage of galactose in the presence of β -galactosidase.

The gelator¹¹⁶ **89** based on glucosamine peptide yielded hydrogels, which were biocompatible (Fig. 57). The derivatives of such gelators when combined with the drug olsalazine (anti-inflammatory) yielded compound **90**. In an acidic medium, the molecule undergoes self-assembly yielding nanofibrous structures, which release the drug mesalazine on treatment with azoreductase. Another material involving glycopeptide was found to have potential applications in the functioning of cells.¹¹⁷

Enzyme stimulation has also been reported involving enzyme histidine triad nucleotide-binding protein 1. Here nucleoside phosphoramidate having bases such as pyrimidine and purine was employed as the gelator and yielded the hydrogel in the presence of enzymes.¹¹⁸

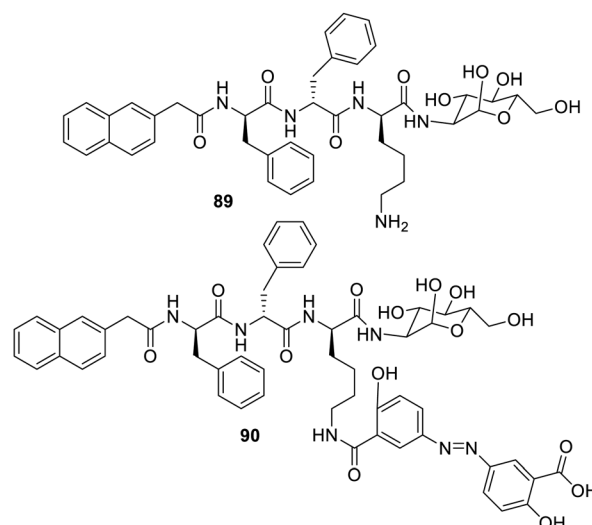


Fig. 57 Glucosamine peptide gelators.





Fig. 58 Glycolipid containing unsaturated tail.

Glycolipids with a *N*-methyl-D-glucamine group were synthesized¹¹⁹ (Fig. 58). The glycolipid had a polar head with an unsaturated chain. It was found in the research that the gelators **91a** and **92b** with small unsaturated chains yielded hydrogels, while the gelator with longer unsaturated tails gave organogelation, which was induced by water *via* agitation. Hydrogel formation was enhanced upon increasing the polar head group size. The gelation of these hydrogels was found to engulf the bioactive compound riboflavin and curcumin, immaterial of their hydrophobic or hydrophilic nature. The engulfing property was further enhanced on increasing the size of the polar headgroup. The release of the bioactive molecule was carried out using trypsin which is an amidase. The bioactive molecule was released on the hydrolysis of glycolipid. It was also found that the hydrogels also exhibited potential for the removal of copper and chromium from the edible oils, which was further enhanced upon increasing the unsaturation of the chain.

Alkaline phosphatase is the most prominent enzyme used for stimulation in carbohydrate-based gels. In one of the reported research studies,¹²⁰ peptide-containing phosphate was cleaved by alkaline phosphate. The gelator here contained a naphthyl group for the self-assembly and galactose head-group to interact with the lectin on *Pseudomonas aeruginosa* to screen for the antibacterial property of the gelator. The dephosphorylation and production of the fibrous structure occurred, which rendered galactose on the outer boundary of the self-assembled fibre, making its interaction with the lectin on the bacterial cell wall. Here the enzyme stimulated the self-assembly and formation of a gel, which was proved to have bactericidal effects.

Similar molecules have a naphthyl group on peptide with a phosphate group and attaching alkaline phosphatase for enzyme stimulation and carbohydrate for binding to the lectin. Phe-Phe-Ser-Tyr(H₂PO₃) and D-mannose composing gelators based on glycopeptide were found to be bactericidal with healing effects on wounds.¹²¹ The mannose here interacted with lectin to cause protein agglutination, which causes adhesion of the bacteria on *Escherichia coli*. The other tested bacteria were *Staphylococcus aureus*, but in this case, the protein of the bacterial cell wall does not interact with mannose.

4. New carbohydrate-based gelators

Glucose- and maltose-based Amphiphilic β -C-glycosylbarbiturate hydrogelators **93a-d** and **94a-d** were synthesized by a H₂O₂-mediated green method, as shown in Fig. 59. These hydrogelators produce glycol nanostructures that self-assemble due to intermolecular interactions and packing factors. Supramolecular gelation of water at neutral pH can be obtained using the synthesized gelators. It was interesting to note that amyloglucosidase from *Aspergillus niger* was used to hydrolyze a precursor made of maltose into a hydrogelator made of glucose.¹²²

Amphiphilic *N*-glycosyl naphthalimides¹²³ (glucose, galactose and xylose derived) **95d**, **96d**, and **97d** have also been synthesized by a green method, which furnished the gel in both hydrophilic and hydrophobic solvents due to their supramolecular self-assembly. These gels possess molecular level interactions such as H-bonding, p-p stacking and van der Waals forces. Compound **95d** (Fig. 60) shows hydrogelation in DMSO-H₂O (40% v/v) with a critical gelation concentration (CGC) of 0.5% (wt/v) and organogelation in CHCl₃, cyclohexane and xylene with a CGC of 0.3, 0.8 and 0.6% (wt/v).

Further, synthesis of C-2 carbamates of *para*-phenylethylidene acetal-protected D-glucosamine derivatives¹²⁴ was also reported along with their gelation properties. The study also reported isopropyl carbamate as a highly efficient hydrogelator, which also exhibited the ability to form gel electrolytes. Gelators **98** and **99** possessed ability to form metalogels with various metal ions, and copper and cobalt xerogels showed good catalytic properties, as shown in Fig. 61. Gelator **99** was reported to form a highly stable gel with naproxen sodium and facilitated controlled release.

The research group further progressed to study the effect of diacetylene functional groups introduced at the anomeric position by using *N*-acetyl-D-glucosamine derivatives on gelating properties. The diacetylenic group was attached at the anomeric position using α -glycosidic linkage separated by one, two and three methylene groups from anomeric oxygen. The glycosides formed red and purple coloured gels, which were prone to photopolymerization. The compound **100**, a hydroxyl-modified derivative, could form hydrogels at a low concentration of 1 wt%. The derivative **101** which was modified using carboxylic acid formed transparent gels and has the ability to immobilize basic solutions, which encourages its use as a pH-responsive gelator. The synthesized gelators, as



Fig. 59 Amphiphilic β -C-glycosylbarbiturates as gelators.



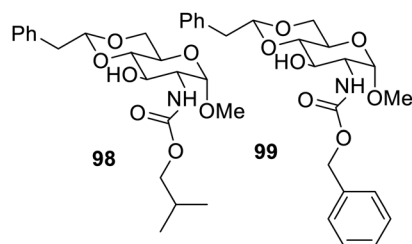
Fig. 60 *N*-Glycosyl naphthalimide derivatives as gelators.

Fig. 61 Carbamate derivatives as new gelators.

shown in Fig. 62, could be used as stimulus-responsive materials, which are highly advantageous in biomedical applications.¹²⁵

In another work, a highly environment-friendly and biologically safe cross-linker, oxidized sucrose (OS) was used to form carboxymethyl chitosan hydrogel (CMCG) **102** (Fig. 63) of a polysaccharide, carboxymethyl chitosan (CMC).¹²⁶ The structural characterization revealed that the reaction proceeded smoothly even in the absence of catalysts *via* cross-linking of aldehyde groups in OS and primary amine groups in CMC. This study focuses on the utilization of oxidised sucrose as a natural cross-linking agent for selective linking of amino sugars to facilitate the formation of polysaccharide gels, which hold enormous potential for biomedical applications.

Recent years have also witnessed the synthesis and utilization of gelators for various applications. The ability of *N*-linear saturated fatty acyl-GABA present in the human brain synthesized using amide-bonded γ -aminobutyric acid (GABA) and linear saturated fatty acids of varying lengths to gelate organic solvents has been reported by Komba *et al.* Along with this, ester derivatives of GABA¹²⁷ and 1,5-anhydro-D-glucitol (1,5-AG) **103a-d** or D-glucopyranose (Glc) **104a-d** (Fig. 64) also delivered



Fig. 62 Diacetylene-containing glycolipids as gelators.



Fig. 63 Carboxymethyl chitosan hydrogel.



Fig. 64 Ester derivatives of GABA and (1,5-AG) and (Glc).

gelating properties, which demonstrated high gel hardness and transparency of *N*-tetradecanoic acyl-GABA bonded to 1,5-AG *via* ester linkage (C14GABA-AG).

Another interesting study reports the effect of halide substitution on gelation properties by demonstrating the synthesis of *para*-chlorobenzylidene acetal-protected D-glucosamine amide derivatives. The self-assembling characteristics of these amides were examined in a variety of solvents, organics as well as mixtures, which revealed the property of these derivatives to form gels at low minimum gelation concentrations (MGCs) even below 0.1 wt%. The exploration resulted in the formation of a highly stable gel in water and EtOH/H₂O by a gelator¹²⁸ benzamide **105** (Fig. 65) at an extremely low concentration of 0.04 wt%. The gel was further employed for entrapment as well as controlled release of chloramphenicol and naproxen as well as for dye removal for toluidine blue aqueous solutions.

Alkyl-substituted sugar-aminobenzoate-based organogelator **106** has also been synthesized at low costs by a simple and effective method, as shown in Fig. 65. It can remove water-soluble dyes from their concentrated aqueous solutions. Organogelator **106** shows high gelling ability towards a wide range of solvents along with the exceptional detection ability towards



Fig. 65 D-Glucosamine amide and alkyl derivatives as gelators.



copper metal ions with a state change from gel to solution¹²⁹ Gel-sol transformation takes place due to high stability of [Cu(G13)] complex formation.

5. Conclusions

This review presented an overview of carbohydrate-based gelators as new materials and their potential applications in various emerging fields such as drug delivery, environmental remediation, antibacterial agents, tissue engineering, thixotropy and wound healing. Carbohydrates are natural materials which are compatible for cells and tissues of living beings, and hence, they are extensively researched and useful for the generation of soft materials. The carbohydrate-based LMGs that fall into this category with numerous potential applications in almost all fields including biomedical applications are discussed in the review. Ribose, arabinose, galactose, mannose, glucosamine, and glucose have been studied for the development of LMGs. The role of the C-1 oxidized and reduced sugar has also been reviewed. The functionalization of carbohydrates for specific stimulation has also been discussed with several examples, which enrich the literature for the development of novel hydro or organogelators for specific requirement in the future. This review compiles stimulus-responsive gelators that are sensitive to acids, bases, enzymes, photoradiation and ions. Thus, the carbohydrate-based gelator as a soft material is an emerging field, which may be studied in collaboration with material scientists to develop new materials for future use.

Conflicts of interest

The authors declare that there is no conflict of interest regarding this manuscript.

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References

- 1 S. Mitchell, R. Qin, N. Zheng and J. Pérez-Ramírez, *Nat. Nanotechnol.*, 2021, **16**, 129–139.
- 2 J. Zhou, J. Li, X. Du and B. Xu, *Biomaterials*, 2017, **129**, 1–27.
- 3 H. Shigemitsu and I. Hamachi, *Acc. Chem. Res.*, 2017, **50**, 740–750.
- 4 J. Wang, X. Miao, Q. Fengzhao, C. Ren, Z. Yang and L. Wang, *RSC Adv.*, 2013, **3**, 16739–16746.
- 5 T. Kato, Y. Hirai, S. Nakaso and M. Moriyama, *Chem. Soc. Rev.*, 2007, **36**, 1857–1867.
- 6 S. S. Babu, V. K. Praveen and A. Ajayaghosh, *Chem. Rev.*, 2014, **114**, 1973–2129.
- 7 A. Ajayaghosh, V. K. Praveen and C. Vijayakumar, *Chem. Soc. Rev.*, 2008, **37**, 109–122.
- 8 J. Morris, J. Bietsch, K. Bashaw and G. Wang, *Gels*, 2021, **7**, 1–61.
- 9 B. O. Okesola and D. K. Smith, *Chem. Soc. Rev.*, 2016, **45**, 4226–4251.
- 10 A. M. Vibhute and K. M. Sureshan, *ChemSusChem*, 2020, **13**, 5343–5360.
- 11 J. Zhang and C. Y. Su, *Coord. Chem. Rev.*, 2013, **257**, 1373–1408.
- 12 A. Y. Y. Tam and V. W. W. Yam, *Chem. Soc. Rev.*, 2013, **42**, 1540–1567.
- 13 Y. Zheng, G. Li and Y. Zhang, *ChemNanoMat*, 2016, **2**, 364–375.
- 14 M. Häring and D. D. Díaz, *Chem. Commun.*, 2016, **52**, 13068–13081.
- 15 F. Versluis, J. H. van Esch and R. Eelkema, *Adv. Mater.*, 2016, **28**, 4576–4592.
- 16 R. Tian, J. Chen and R. Niu, *Nanoscale*, 2014, **6**, 3474–3482.
- 17 R. Dong, Y. Pang, Y. Su and X. Zhu, *Biomater. Sci.*, 2015, **3**, 937–954.
- 18 B. Escuder, F. Rodríguez-Llansola and J. F. Miravet, *New J. Chem.*, 2010, **34**, 1044–1054.
- 19 F. Rodríguez-Llansola, B. Escuder and J. F. Miravet, *J. Am. Chem. Soc.*, 2009, **131**, 11478–11484.
- 20 A. Döring, W. Birnbaum and D. Kuckling, *Chem. Soc. Rev.*, 2013, **42**, 7391–7420.
- 21 N. Singh, K. Zhang, C. A. Angulo-Pachón, E. Mendes, J. H. Van Esch and B. Escuder, *Chem. Sci.*, 2016, **7**, 5568–5572.
- 22 N. Basu, A. Chakraborty and R. Ghosh, *Gels*, 2018, **4**(2), 52.
- 23 R. Kubota, S. Liu, H. Shigemitsu, K. Nakamura, W. Tanaka, M. Ikeda and I. Hamachi, *Bioconjugate Chem.*, 2018, **29**, 2058–2067.
- 24 M. Liu, G. Ouyang, D. Niu and Y. Sang, *Org. Chem. Front.*, 2018, **5**, 2885–2900.
- 25 J. Hoque, N. Sangaj and S. Varghese, *Macromol. Biosci.*, 2019, **19**, 1–16.
- 26 H. Wang, Z. Feng and B. Xu, *Theranostics*, 2019, **9**, 3213–3222.
- 27 S. Datta and S. Bhattacharya, *Chem. Soc. Rev.*, 2015, **44**, 5596–5637.
- 28 K. Soundarajan, R. Periyasamy and T. Mohan Das, *RSC Adv.*, 2016, **6**, 81838–81846.
- 29 G. Wang, S. Cheuk, K. Williams, V. Sharma, L. Dakessian and Z. Thornton, *Carbohydr. Res.*, 2006, **341**, 705–716.
- 30 X. Nie and G. Wang, *J. Org. Chem.*, 2006, **71**, 4734–4741.
- 31 H. P. R. Mangunuru, J. R. Yerabolu, D. Liu and G. Wang, *Tetrahedron Lett.*, 2015, **56**, 82–85.
- 32 G. Wang, A. Chen, H. P. R. Mangunuru and J. R. Yerabolu, *RSC Adv.*, 2017, **7**, 40887–40895.
- 33 I. S. Okafor and G. Wang, *Carbohydr. Res.*, 2017, **451**, 81–94.
- 34 A. Chen, S. B. Adhikari, K. Mays and G. Wang, *Langmuir*, 2017, **33**, 8076–8089.



- 35 G. Wang, H. Yang, S. Cheuk and S. Coleman, *Beilstein J. Org. Chem.*, 2011, **7**, 234–242.
- 36 S. Cheuk, E. D. Stevens and G. Wang, *Carbohydr. Res.*, 2009, **344**, 417–425.
- 37 N. Goyal, S. Cheuk and G. Wang, *Tetrahedron*, 2010, **66**, 5962–5971.
- 38 G. Wang, S. Cheuk, H. Yang, N. Goyal, P. V. Narasimha Reddy and B. Hopkinson, *Langmuir*, 2009, **25**, 8696–8705.
- 39 N. Goyal, H. P. R. Mangunuru, B. Parikh, S. Shrestha and G. Wang, *Beilstein J. Org. Chem.*, 2014, **10**, 3111–3121.
- 40 H. P. R. Mangunuru, J. R. Yerabolu and G. Wang, *Tetrahedron Lett.*, 2015, **56**, 3361–3364.
- 41 H. P. R. Mangunuru, H. Yang and G. Wang, *Chem. Commun.*, 2013, **49**, 4489–4491.
- 42 G. Wang, N. Goyal, H. P. R. Mangunuru, H. Yang, S. Cheuk and P. V. N. Reddy, *J. Org. Chem.*, 2015, **80**, 733–743.
- 43 K. Soundarajan, M. Rajasekar and T. M. Das, *Mater. Sci. Eng., C*, 2018, **93**, 776–781.
- 44 A. D. Ludwig, A. Saint-Jalmes, C. Mériadec, F. Artzner, O. Tasseau, F. Berrée and L. Lemiègre, *Chem. – Eur. J.*, 2020, **26**, 13927–13934.
- 45 C. Mahendar, M. K. Dixit, Y. Kumar and M. Dubey, *J. Mater. Chem. C*, 2020, **8**, 11008–11012.
- 46 K. Lalitha, K. Gayathri, Y. S. Prasad, R. Saritha, A. Thamizhanban, C. U. Maheswari, V. Sridharan and S. Nagarajan, *Gels*, 2018, **4**(1), 1.
- 47 F. Ono, K. Ichimaru, O. Hirata, S. Shinkai and H. Watanabe, *Chem. Lett.*, 2020, **49**, 156–159.
- 48 J. Brinksma, B. L. Feringa, R. M. Kellogg, R. Vreeker and J. Van Esch, *Langmuir*, 2000, **16**, 9249–9255.
- 49 N. N. Adarsh, D. K. Kumar and P. Dastidar, *Tetrahedron*, 2007, **63**, 7386–7396.
- 50 M. Suzuki, M. Yumoto, H. Shirai and K. Hanabusa, *Tetrahedron*, 2008, **64**, 10395–10400.
- 51 A. Pal, Y. K. Ghosh and S. Bhattacharya, *Tetrahedron*, 2007, **63**, 7334–7348.
- 52 J. Morris, P. Kozlowski and G. Wang, *Langmuir*, 2019, **35**, 14639–14650.
- 53 A. Chen, L. P. Samankumara, C. Garcia, K. Bashaw and G. Wang, *New J. Chem.*, 2019, **43**, 7950–7961.
- 54 D. Wang, A. Chen, J. Morris and G. Wang, *RSC Adv.*, 2020, **10**, 40068–40083.
- 55 T. Xiong, X. Li, Y. Zhou, Q. Song, R. Zhang, L. Lei and X. Li, *Acta Biomater.*, 2018, **73**, 275–284.
- 56 E. R. Draper and D. J. Adams, *Chem*, 2017, **3**, 390–410.
- 57 L. E. Buerkle and S. J. Rowan, *Chem. Soc. Rev.*, 2012, **41**, 6089–6102.
- 58 L. S. Birchall, S. Roy, V. Jayawarna, M. Hughes, E. Irvine, G. T. Okorogheye, N. Saudi, E. de Santis, T. Tuttle, A. A. Edwards and R. V. Ulijn, *Chem. Sci.*, 2011, **2**, 1349–1355.
- 59 A. Brito, Y. M. Abul-Haija, D. S. Da Costa, R. Novoa-Carballal, R. L. Reis, R. V. Ulijn, R. A. Pires and I. Pashkuleva, *Chem. Sci.*, 2019, **10**, 2385–2390.
- 60 A. Brito, P. M. R. Pereira, R. L. Reis, R. V. Ulijn, J. S. Lewis, R. A. Pires and I. Pashkuleva, *Nanoscale*, 2020, **12**, 19088–19092.
- 61 K. B. Pal and B. Mukhopadhyay, *ChemistrySelect*, 2017, **2**, 967–974.
- 62 N. P. Pathak, Rajkamal and S. Yadav, *Chem. Commun.*, 2020, **56**, 2999–3002.
- 63 C. Narayana, R. K. Upadhyay, R. Chaturvedi and R. Sagar, *New J. Chem.*, 2017, **41**, 2261–2267.
- 64 J. Ramos, S. Arufe, H. Martin, D. Rooney, R. B. P. Elmes, A. Erxleben, R. Moreira and T. Velasco-Torrijos, *Soft Matter*, 2020, **16**, 7916–7926.
- 65 R. Kannan, V. Muthuvijayan and E. Prasad, *New J. Chem.*, 2017, **41**, 7453–7462.
- 66 A. Chalard, L. Vaysse, P. Joseph, L. Malaquin, S. Souleille, B. Lonetti, J. C. Sol, I. Loubinoux and J. Fitremann, *ACS Appl. Mater. Interfaces*, 2018, **10**, 17004–17017.
- 67 A. Chalard, P. Joseph, S. Souleille, B. Lonetti, N. Saffon-Merceron, I. Loubinoux, L. Vaysse, L. Malaquin and J. Fitremann, *Nanoscale*, 2019, **11**, 15043–15056.
- 68 A. Chalard, M. Mauduit, S. Souleille, P. Joseph, L. Malaquin and J. Fitremann, *Addit. Manuf.*, 2020, **33**, 101162.
- 69 Y. Siva Prasad, S. Manikandan, K. Lalitha, M. Sandeep, R. Vara Prasad, R. Arun Kumar, C. S. Srinandan, C. Uma Maheswari, V. Sridharan and S. Nagarajan, *Nano Sel.*, 2020, **1**, 510–524.
- 70 X. Guan, K. Fan, T. Gao, A. Ma, B. Zhang and J. Song, *Chem. Commun.*, 2016, **52**, 962–965.
- 71 J. Liu, J. Li, P. Lin, N. Zhang, X. Han, B. Zhang and J. Song, *Chem. Commun.*, 2016, **52**, 13975–13978.
- 72 K. Fan, X. Wang, X. Wang, H. Yang, G. Han, L. Zhou and S. Fang, *RSC Adv.*, 2020, **10**, 37080–37085.
- 73 W. C. Lai and P. H. Huang, *Soft Matter*, 2017, **13**, 3107–3115.
- 74 G. C. Dizon, G. Atkinson, S. P. Argent, L. T. Santu and D. B. Amabilino, *Soft Matter*, 2020, **16**, 4640–4654.
- 75 P. McNeice, Y. Zhao, J. Wang, G. F. Donnelly and P. C. Marr, *Green Chem.*, 2017, **19**, 4690–4697.
- 76 C. S. Kesava Raju, B. Pramanik, R. Ravishankar, P. V. Chalapathi Rao and G. Sriganesh, *RSC Adv.*, 2017, **7**, 37175–37180.
- 77 J. E. Moses and A. D. Moorhouse, *Chem. Soc. Rev.*, 2007, **36**, 1249–1262.
- 78 H. C. Kolb, M. G. Finn and K. B. Sharpless, *Angew. Chem., Int. Ed.*, 2001, **40**, 2004–2021.
- 79 A. Chen, I. S. Okafor, C. Garcia and G. Wang, *Carbohydr. Res.*, 2018, **461**, 60–75.
- 80 B. P. Krishnan and K. M. Sureshan, *Chem. – Asian J.*, 2018, **13**, 187–193.
- 81 Rajkamal, N. P. Pathak, D. Chatterjee, A. Paul and S. Yadav, *RSC Adv.*, 2016, **6**, 92225–92234.
- 82 C. Narayana, P. Kumari, G. Tiwari and R. Sagar, *Langmuir*, 2019, **35**, 16803–16812.
- 83 M. A. Ramin, J. Baillet, S. Benizri, L. Latxague and P. Barthélémy, *New J. Chem.*, 2016, **40**, 9903–9906.
- 84 M. A. Ramin, L. Latxague, K. R. Sindhu, O. Chassande and P. Barthélémy, *Biomaterials*, 2017, **145**, 72–80.
- 85 N. D. Bansode, K. R. Sindhu, C. Morel, M. Rémy, J. Verget, C. Boiziau and P. Barthélémy, *Biomater. Sci.*, 2020, **8**, 3186–3192.



- 86 M. E. Cano, P. H. Di Chenna, D. Lesur, A. Wolosiuk, J. Kovensky and M. L. Uhrig, *New J. Chem.*, 2017, **41**, 14754–14765.
- 87 A. Chen, D. Wang, J. Bietsch and G. Wang, *Org. Biomol. Chem.*, 2019, **17**, 6043–6056.
- 88 G. Wang, D. Wang, J. Bietsch, A. Chen and P. Sharma, *J. Org. Chem.*, 2020, **85**, 16136–16156.
- 89 G. M. Peters and J. T. Davis, *Chem. Soc. Rev.*, 2016, **45**, 3188–3206.
- 90 J. Baillet, V. Desvergnés, A. Hamoud, L. Latxague and P. Barthélémy, *Adv. Mater.*, 2018, **30**, 1–24.
- 91 M. G. F. Angelerou, P. W. J. M. Frederix, M. Wallace, B. Yang, A. Rodger, D. J. Adams, M. Marlow and M. Zelzer, *Langmuir*, 2018, **34**, 6912–6921.
- 92 H. Zhao, D. Jiang, A. H. Schäfer and F. Seela, *Chem-PlusChem*, 2017, **82**, 778–784.
- 93 B. Alies, M. A. Ouelhazi, A. Patwa, J. Verget, L. Navailles, V. Desvergnés and P. Barthélémy, *Org. Biomol. Chem.*, 2018, **16**, 4888–4894.
- 94 J. Li, H. Wei, Y. Peng, L. Geng, L. Zhu, X. Y. Cao, C. Sen Liu and H. Pang, *Chem. Commun.*, 2019, **55**, 7922–7925.
- 95 N. Thakur, B. Sharma, S. Bishnoi, S. Jain, D. Nayak and T. K. Sarma, *ACS Appl. Bio Mater.*, 2019, **2**, 3300–3311.
- 96 A. Nuthanakanti and S. G. Srivatsan, *Nanoscale*, 2016, **8**, 3607–3619.
- 97 X. Jia, J. Zhao, S. Xu, F. Zhang, J. Sun and R. Lu, *Eur. J. Org. Chem.*, 2018, 1910–1915.
- 98 X. Du, J. Li, Y. Gao, Y. Kuang and B. Xu, *Chem. Commun.*, 2012, **48**, 2098–2100.
- 99 M. Ikeda, *Polym. J.*, 2019, **51**, 371–380.
- 100 Y. Qiu and K. Park, *Adv. Drug Delivery Rev.*, 2012, **64**, 49–60.
- 101 A. G. Cheetham, R. W. Chakraborty, W. Ma and H. Cui, *Chem. Soc. Rev.*, 2017, **46**, 6638–6663.
- 102 W. Gao, J. M. Chan and O. C. Farokhzad, *Mol. Pharm.*, 2010, **7**, 1913–1920.
- 103 A. Kumar and Priyanka, *New J. Chem.*, 2019, **43**, 14997–15013.
- 104 E. R. Draper and D. J. Adams, *Chem. Commun.*, 2016, **52**, 8196–8206.
- 105 M. J. Clemente, R. M. Tejedor, P. Romero, J. Fitremann and L. Oriol, *RSC Adv.*, 2012, **2**, 11419–11431.
- 106 R. Rajaganesh, A. Gopal, T. Mohan Das and A. Ajayaghosh, *Org. Lett.*, 2012, **14**, 748–751.
- 107 R. Oosumi, M. Ikeda, A. Ito, M. Izumi and R. Ochi, *Soft Matter*, 2020, **16**, 7274–7278.
- 108 Z. Khayat and H. Zali-Boeini, *Dyes Pigm.*, 2018, **159**, 337–344.
- 109 C. Lin, S. Maisonneuve, R. Métivier and J. Xie, *Chem. – Eur. J.*, 2017, **23**, 14996–15001.
- 110 J. Baillet, A. Gaubert, D. M. Bassani, J. Verget, L. Latxague and P. Barthélémy, *Chem. Commun.*, 2020, **56**, 3397–3400.
- 111 B. P. Krishnan, S. Raghu, S. Mukherjee and K. M. Sureshan, *Chem. Commun.*, 2016, **52**, 14089–14092.
- 112 J. Gao, J. Zhan and Z. Yang, *Adv. Mater.*, 2020, **32**, 1–13.
- 113 J. Mu, J. Lin, P. Huang and X. Chen, *Chem. Soc. Rev.*, 2018, **47**, 5554–5573.
- 114 J. Baillet, A. Gaubert, J. Verget, L. Latxague and P. Barthélémy, *Soft Matter*, 2020, **16**, 7648–7651.
- 115 T. Xu, Y. Cai, X. Zhong, L. Zhang, D. Zheng, Z. Gao, X. Pan, F. Wang, M. Chen and Z. Yang, *Chem. Commun.*, 2019, **55**, 7175–7178.
- 116 J. Zhou, M. O’Keeffe, G. Liao, F. Zhao, C. Terhorst and B. Xu, *Tetrahedron*, 2016, **72**, 6078–6083.
- 117 J. Zhou, X. Du, X. Chen and B. Xu, *Biochemistry*, 2018, **57**, 4867–4879.
- 118 H. T. West, C. M. Csizmar and C. R. Wagner, *Biomacromolecules*, 2018, **19**, 2650–2656.
- 119 K. P. C. Sekhar, D. K. Swain, S. A. Holey, S. Bojja and R. R. Nayak, *Langmuir*, 2020, **36**, 3080–3088.
- 120 S. Liu, H. Li, J. Zhang, X. Tian and X. Li, *RSC Adv.*, 2020, **10**, 33642–33650.
- 121 J. Li, S. Liang, Y. Yan, X. Tian and X. Li, *Macromol. Biosci.*, 2019, **19**, 1–10.
- 122 S. Yao, R. Brahmi, A. Bouschon, J. Chen and S. Halila, *Green Chem.*, 2023, **25**, 330–335.
- 123 A. K. Rachamalla, V. P. Rebaka, T. Banoo, R. Pawar, M. Faizan, K. Lalitha and S. Nagarajan, *Green Chem.*, 2022, **24**, 2451–2463.
- 124 P. Sharma and G. Wang, *Gels*, 2022, **8**, 191, DOI: [10.3390/gels8030191](https://doi.org/10.3390/gels8030191).
- 125 G. Wang, D. Wang, A. Chen, I. S. Okafor and L. P. Samankumara, *ACS Omega*, 2022, **7**, 11330–11342.
- 126 H. Kono, J. Noda and H. Wakamori, *Molecules*, 2022, **27**, 6137, DOI: [10.3390/molecules27186137](https://doi.org/10.3390/molecules27186137).
- 127 S. Komba and R. Iwaura, *ACS Omega*, 2021, **6**, 20912–20923.
- 128 J. Bietsch, M. Olson and G. Wang, *Gels*, 2021, **7**, 21–23.
- 129 P. V. Bhavya, K. Soundarajan, J. G. Malecki and T. Mohan Das, *ACS Omega*, 2022, **7**, 39310–39324.

