



Cite this: *Nanoscale*, 2022, **14**, 13915

Dye-sensitized lanthanide containing nanoparticles for luminescence based applications

Clémence Cheignon, * Ali A. Kassir, † Lohona K. Soro † and
Loïc J. Charbonnière *

Due to their exceptional luminescent properties, lanthanide (Ln) complexes represent a unique palette of probes in the spectroscopic toolkit. Their extremely weak brightness due to forbidden Ln electronic transitions can be overcome by indirect dye-sensitization from the antenna effect brought by organic ligands. Despite the improvement brought by the antenna effect, (bio)analytical applications with discrete Ln complexes as luminescent markers still suffers from low sensitivity as they are limited by the complex brightness. Thus, there is a need to develop nano-objects that cumulate the spectroscopic properties of multiple Ln ions. This review firstly gives a brief introduction of the spectral properties of lanthanides both in complexes and in nanoparticles (NPs). Then, the research progress of the design of Ln-doped inorganic NPs with capping antennas, Ln-complex encapsulated NPs and Ln-complex surface functionalized NPs is presented along with a summary of the various photosensitizing ligands and of the spectroscopic properties (excited-state lifetime, brightness, quantum yield). The review also emphasizes the problems and limitations encountered over the years and the solutions provided to address them. Finally, a comparison of the advantages and drawbacks of the three types of NP is provided as well as a conclusion about the remaining challenges both in the design of brighter NPs and in the luminescence based applications.

Received 25th May 2022,
Accepted 29th August 2022

DOI: 10.1039/d1nr06464a

rsc.li/nanoscale

1. Introduction

Because of their exceptional electronic properties,¹ lanthanide (Ln) ions are very particular ones in the spectroscopic toolbox. They display line like emission spectra which are specific signatures for each luminescent ion of the series, and their excited state lifetimes are three to six orders of magnitude longer than organic compounds or luminescent d-block coordination compounds, allowing for a very sensitive time-resolved (TR) detection. However, the same reasons providing them with these exceptional properties, *i.e.* the forbidden character of the electronic transitions by Laporte's and also often spin selection rules, confer on them very poor absorption properties. Molar absorption coefficients of lanthanide ions are very weak, rarely exceeding unity, apart for the exceptional case of Yb complexes.² Fortunately, during the second world war, the discovery of Weissman³ that lanthanide ions could be indirectly photosensitized by coordinated aromatic ligands (also called antennas), changed the game. Since then, a

plethora of researchers tried to improve the luminescence properties of Ln complexes, by coordinating them with adapted antenna which collect photons and transfer them to the Ln cation. It is worth noting that a proper selection of the antenna is required to ensure that it coordinates with the Ln ion, it absorbs light at the desired excitation wavelength and that its excited state is close enough to the excited state of the Ln to allow energy transfer.⁴ Thanks to the antenna effect, the brightness of the complexes (*B*), defined as the product of the molar absorption coefficient by the luminescence quantum yield,⁵ have begun to become acceptable in comparison with organic fluorophores.⁶

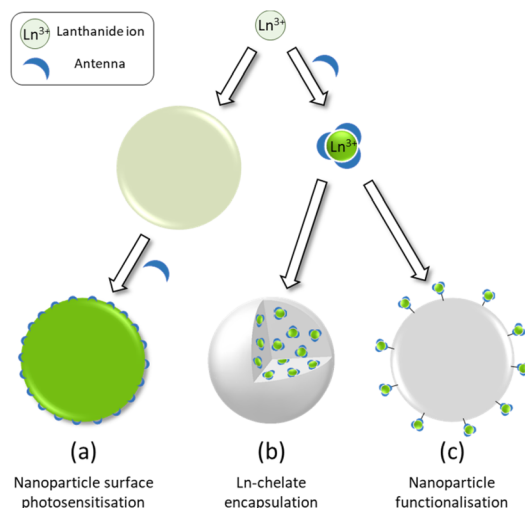
Embedding the Ln ions into synthetic pre-organized macrocyclic structures,⁷ further led to kinetically and thermodynamically stable complexes which can find applications in luminescence labeling.^{8–10} However, even for the best of these labels and despite the advantages of TR detection, these labels are still far from the best fluorophores, such as luminescent proteins or semiconducting nanocrystals.¹¹

An obvious further step in the development of brighter Ln based luminescent marker was then to try to gather numerous Ln ions into nanoscopic structures. The aim of this review is to highlight the three main approaches towards Ln based nanoscopic scaffolds that are illustrated in Scheme 1. These comprise the development of

Equipe de Synthèse Pour l'Analyse (SynPA), Institut Pluridisciplinaire Hubert Curien (IPHC), UMR 7178 CNRS/Université de Strasbourg, ECPM, Bâtiment R1N0, 25 rue Becquerel, 67087 Strasbourg, Cedex 2, France. E-mail: ccheignon@unistra.fr, l.charbonn@unistra.fr

†These authors contributed equally to this work.





Scheme 1 Schematic representation of the three strategies for the design of dye-sensitized lanthanide containing nanoparticles.

surface capped Ln nanoparticles (Ln-NPs) using antenna ligands (Scheme 1a), the incorporation of discrete molecular complexes into nanoscopic structures (Scheme 1b) and the preparation of nanoscopic scaffolds surface functionalized by Ln complexes (Scheme 1c). Before going into the details of each approach, we will first deal with some of the basic spectroscopic properties of luminescent Ln ion to compare them at the level of molecular complexes or at the nanoscopic scale.

2. Basic spectral properties of Ln ion in complexes and their comparison with Ln doped NPs

2.1. Absorption properties

The absorption A of a compound in solution is defined as its propensity to absorb photons at a certain wavelength. A is proportional to the molar absorption coefficient ϵ_λ (in $\text{mol}^{-1} \text{L cm}^{-1}$) and is determined using the Beer-Lambert law:

$$A = \log\left(\frac{I_{\lambda 0}}{I_\lambda}\right) = \epsilon_\lambda \cdot l \cdot c \quad (1)$$

In which $I_{\lambda 0}$ and I_λ are respectively the intensities of the incoming and outgoing light source at the wavelength λ , l is the length of the optical path (in cm) and c the concentration of the absorbing species in the sample (in mol L^{-1}). Alternatively, the absorption may be related to σ the absorption cross section (in cm^2) which represents the effective area that a photon needs to cross in order to be absorbed. While chemists generally prefer ϵ , physicists and spectroscopists prefer σ but both are related by the following relationship:

$$\sigma = 0.382 \times 10^{-20} \times \epsilon \quad (2)$$

Considering the absorption properties of a Ln complex, it is important to clearly differentiate the absorption due to the ligand coordinated to the metal and the intrinsic absorption of the Ln ion itself. For the later, f-f electronic transitions are forbidden by Laporte's rule ($\Delta L = \pm 1$) and for most of them by the spin multiplicity rule too ($\Delta S = 0$). For a full detail of the selection rules in Ln compounds, the reader is invited



Clémence Cheignon

Clémence Cheignon obtained her PhD in analytical chemistry at the University Paul Sabatier (Toulouse, France) in 2016 under the supervision of Dr Fabrice Collin and Dr Christelle Hureau in Prof. Peter Faller's group, where she also carried out post-doctoral research. After one year as a Temporary Lecturer and Research assistant at the Institut des Biomolécules Max Mousseron (Montpellier, France)

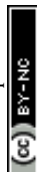
in the group of Prof. Christine Enjalbal, she moved to Strasbourg (France) and obtained the position of assistant professor at the Engineer School of chemistry (ECPM) of Strasbourg University (France) in 2019. She is conducting research in the SynPA group led by Dr Loïc Charbonnière at IPHC, her current research is focused on the design and characterization of luminescent lanthanide nanoparticles and their application as probes in (bio)analytical applications.



Ali A. Kassir

Ali Kassir obtained his BSc degree in chemistry from Lebanese University in 2017. He completed his master's in analytical sciences at Strasbourg University in 2019. Currently, he is a PhD student in the SynPA team under the direction of Dr Loïc Charbonnière and the co-supervision of Dr Clémence Cheignon, and working on the development of new ELISA detection methods based on ultraviolet right lanthanide nanoparticles.

His main interests are centered on nanomaterials and bioanalytical applications.



to consider specialized literature.¹ As a result, the absorption coefficients of f-f transitions in Ln complexes are very weak ($<1 \text{ mol}^{-1} \text{ L cm}^{-1}$) and are rarely determined, except for some cases of Yb complexes,^{2,12} the corresponding $^2\text{F}_{5/2} \leftarrow ^2\text{F}_{7/2}$ electronic transition being unique and allowed by the spin selection rule. In contrast, the electronic transition centered on the ligands are generally associated to allowed $^1\pi\pi^*$ transitions with large absorption coefficients ($\epsilon > 1000 \text{ mol}^{-1} \text{ L cm}^{-1}$). Additionally, the incorporation of multiple aromatic antenna coordinated to the Ln allows to cumulate their absorption properties.¹³

When Ln ions are embedded into nanoparticles, as the 4f orbitals have a small radial extension,¹⁴ and considering that these orbitals are protected from the surroundings by filled 5s and 5p orbitals, they poorly participate into the bonding with the surrounding ligands or coordinating ions and are only weakly influenced by the environment. As a result, the absorption coefficients of the f-f transitions are corresponding to the sum of the Ln atoms in the NPs and they do not benefit of any confinement effect, as it is the case for semi-conducting nanoparticles.¹⁵ Similarly, the absorption associated to the ligands coordinated to Ln-NPs are generally the result of the sum of the absorption of each ligand. In some instances, these absorptions have been determined using spectrophotometric titrations allowing to reach very large absorption coefficients for the whole NPs ($\epsilon > 10^7 \text{ mol}^{-1} \text{ L cm}^{-1}$),¹⁶ making such NPs of relevance for labeling and bio-analytical applications.

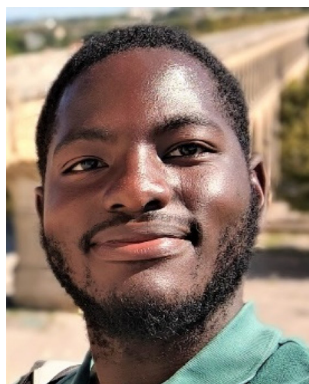
2.2. Luminescence properties

The luminescence quantum yield (ϕ) is defined as the ratio of emitted photons over absorbed photons. When an atom or a molecule has absorbed a photon, it reaches an excited state and can decay back to the ground state by radiative processes, resulting in luminescence, with a rate constant k_r , and a radiative lifetime τ_r ($k_r = 1/\tau_r$) or by numerous other non-radiative processes, with rate constants k_{nr} . The luminescence is then related to these rate constants by:

$$\phi = \frac{k_r}{k_r + \sum k_{nr}} = \frac{\tau_{\text{obs}}}{\tau_r} \quad (3)$$

When one determines the excited state lifetime of the molecule by spectroscopic means, the observed decay, τ_{obs} , corresponds to the inverse of the sum of all the radiative and non-radiative phenomena leading to the right part of eqn (3). Considering eqn (3), it is obvious that the smaller the rates of non-radiative processes, the larger the luminescence quantum yield.

For discrete luminescent lanthanide complexes, the most important non-radiative phenomena are generally due to energy transfer to the overtones of high energy oscillators such as OH, NH and CH bonds of the organic ligands or of closely placed molecules of solvents.¹⁷ These energy losses have been deeply studied by different authors for visible lanthanide emitters such as Eu^{17,18} and Tb,¹⁷ but also for near infrared (NIR) ones such as Yb¹⁷ and Nd¹⁹ and dual NIR-visible Ln emitters



Lohona K. SORO

Lohona SORO obtained his master's degree in analytical chemistry in 2019 at the University of Strasbourg. He is now a PhD student under the supervision of Dr Loïc Charbonnière in the SynPA team. His research mainly focuses on optimizing the upconversion efficiency of lanthanide-based upconverting devices in solution.



Loïc J. Charbonnière

Loïc Charbonnière got a degree of engineer in chemistry from the chemistry school of Strasbourg (France) in 1991 and a master in chemistry from the University of Strasbourg in 1993. He earned the grade of Dr in 1996 for his thesis on the synthesis of dinuclear triple stranded helicates under the supervision of Prof. Alan Williams in Geneva (Switzerland). After a first post-doctoral period in Lausanne (Switzerland, 1997) with Prof.

Jean-Claude Bünzli and a second one in Strasbourg (1998) with Dr Françoise Arnaud-Neu, he obtained the position of associate researcher at the French National Research Centre (CNRS) in 1998 in the team of Dr Raymond Ziessel. In 2011, he created a team dedicated to the synthesis of new chemical tools for analytical application, renamed the SynPA team in 2018. In 2011, he also earned the grade of Research Director at the CNRS. His main scientific interests lie in organic synthesis and coordination chemistry, with a strong liking for lanthanides and their exceptional properties and all their potential uses at the border of different disciplines.



Dy and Sm.²⁰ The particular interest of these studies relies on the possibility to access to the hydration number q , *i.e.* the number of water molecules directly bonded to the first coordination sphere of the Ln cation. By determining the luminescence lifetime of the complexes in light water $\tau_{\text{H}_2\text{O}}$ and in heavy water $\tau_{\text{D}_2\text{O}}$, q can be determined using the general formula:

$$q = A \left(\frac{1}{\tau_{\text{H}_2\text{O}}} - \frac{1}{\tau_{\text{D}_2\text{O}}} - B \right) - C \quad (4)$$

In which A , B and C are constants depending on the studied Ln.^{17–20} The impact of these non-radiative processes can be highly detrimental, especially for NIR emitters in water as the energy levels of the Ln excited states are close to the first and second overtones of the OH oscillators of water at *ca.* 7000 and 10 500 cm^{-1} , corresponding to *ca.* 1430 and 952 nm respectively.¹⁷ Although less important in general, the oscillators associated to C–H bonds present in the framework of the antenna ligands can also have a strong impact on the luminescence, justifying the importance of their deuteration to avoid the corresponding losses.²¹

In the case of Ln-NPs, two kinds of Ln environments have to be taken into account. Ln atoms in the very external layers can also be influenced by OH, NH and CH oscillators of the surrounding medium and their luminescence can be quenched by the same mechanisms as for discrete Ln complexes. In contrast, Ln atoms embedded in the core of the NPs are not influenced by the surrounding medium. However, they are still prone to non-radiative deactivation through impurities of the matrix with high-frequency vibrations, such as OH-groups²² or energy transfer processes assisted by phonons, quasiparticles having the energy of a vibration of the lattice of the crystalline core. However, phonons of the lattice are generally of much lower energy than OH, NH or CH oscillators. For example, when the OH stretching vibration of water is observed at 3500 cm^{-1} , the highest phonon energy observed in common solid matrices do not exceed 1200 cm^{-1} .²³ The non-radiative rate constant, k_{nr} , can then be related to the energy of the phonon, ΔE_{p} , and to the energy difference between the ground and excited states of the lanthanide ΔE_{Ln} by the following relation:²⁴

$$k_{\text{nr}} = A \cdot e^{-\left(\frac{B \times \Delta E_{\text{Ln}}}{\Delta E_{\text{p}}}\right)} \quad (5)$$

In which A and B are constants depending on the solid matrix. From eqn (5), one can realize that the smaller the energy of the phonon, the lower the rate constant for non-radiative decays. In some instances, the non-radiative decays can be so small that the observed luminescence lifetimes of the Ln atoms are close to the radiative lifetimes, affording an almost quantitative lanthanide centered luminescence quantum yield (eqn (3)), as it is for example observed for some Eu doped GdF_3 NPs²⁵ for which a luminescence lifetime of 10.5 ms is observed when the radiative lifetime of Eu is 9.7 ms for the aqua ion and up to 11 ms in other media.²⁶

Considering the position of the Ln atom in the NP (outer, intermediate or inner layers), it is in some instances possible to differentiate the different environment of the Ln atoms. In the case of Eu doped LaF_3 NPs,²⁷ it was possible to extract up to three different lifetimes for the Eu emission with values of 7.04 (48%), 1.85 (38%) and 0.4 (14%) ms. These lifetimes, in agreement with the respective populations indicated in brackets, were respectively attributed to core Eu atoms, Eu atoms close to the surface and Eu atoms at the surface of the NPs, which are severely quenched by water molecules and citrate capping ligands.

The decreased luminescence properties of surface atoms might be an important issue, especially if the doping in luminescent Ln atoms becomes high. In that case, energy migration can occur within the inner Ln atoms, which might be transferred up to the surface atoms and is then partially quenched. Once again, this is particularly true for NIR Ln emitters, providing the NPs with poor luminescence efficiency. In the case of NP with upconversion (UC) properties (see below) or downconversion and downshifting in the NIR domain,²⁸ in which NIR photons have to be accumulated, such a surface quenching effect is particularly deleterious. To overcome this problem, core/shell structures have been designed to protect the Ln emitters by an outer shell, thereby displacing the active Ln atoms from the surface of the NP. For example, NPs based on NaYF_4 doped with a mixture of Nd^{3+} , Yb^{3+} and Er^{3+} have seen their UC efficiency increased by a factor 50 when a supplementary shell of NaYF_4 is added at the surface of the NPs.²⁹ The core/shell approach can even be extended to multiple shells in order to localize finally the Ln absorbers or sensitizers and the energy acceptors with a controlled directionality of the energy transfer processes.^{30,31}

It has to be kept in mind that, if the increase of the doping ratio of active luminescent Ln species is important to increase the NPs luminescence, increasing too much this ratio can have deleterious effects and can lead to the phenomenon of concentration quenching for which larger doping ratios lead to decreased luminescence efficiencies.^{32,33}

The position of the emitting Ln cations into the NPs is also of large importance for energy transfer applications with organic dyes at the surface, both for ligand to Ln energy transfer (dye-sensitization) or for Ln to dye at the surface, such as for Förster resonance energy transfer (FRET) applications. In the case of dye-sensitization, although the exact mechanisms are prone to case by case debates between the Dexter and Förster mechanisms,^{34,35} a close contact between the sensitizing ligand and the Ln atoms at the first layers of the surface is mandatory for an efficient sensitization of the Ln atoms. In the second case of energy transfer from the Ln atoms to dyes at the vicinity of the surface, the mechanism generally assumed is that of a dipole–dipole Förster type energy transfer. Within the frame of the theoretical treatment of energy transfer with the Förster's formalism,³⁶ the energy transfer efficiency is dependent on the donor–acceptor distance r to the inversed sixth power, thereby decreasing rapidly when the donor–acceptor distance increases. Thus Ln atoms close to the



surface of the Ln NP are more efficient than those in the core of the NPs.³⁷ Ultimately, the distance between the dye anchored at the surface of Ln-NPs can be chemically or biochemically modulated to improve the ET, for example by playing with the size of biomolecules such as full antibodies, or some of their fragments of smaller size.³⁸

As a last point, NPs have a large advantage over Ln complexes in the possibility of mixing different Ln atoms, providing the NPs with multiplexing capabilities. One might argue that the synthesis of controlled heteropolynuclear complexes is also feasible,^{39,40} but generally at the expense of important synthetic efforts (see for example ref. 41 and 42). For Ln-NPs, a simple mixing of the adequate concentrations at the first step of the synthesis allows to obtain a large panel of NPs with spectroscopically unique signatures with applications in barcoding,⁴³ multiplexing, *i.e.* multiple analysis with a same sample⁴⁴ or multimodal analytical devices.⁴⁵ On the other hand, the larger size and composition of NPs compared to Ln complexes can bring significant issues regarding toxicity,^{46,47} possible unwanted size dependent pharmacokinetic and bio-distribution properties,⁴⁸ or simple stability troubles such as leaching in biological media.^{49,50}

With regards to these different considerations on Ln luminescence, the next chapters aim at reviewing the main works reporting on the three principal strategies developed so far for improving Ln luminescence with the help of nanoscopic scaffolds.

3. Ln ion doped NPs with capping photosensitizers

In general, the efficiency of Ln doped NPs luminescence depends on the structure, local site symmetry, and phonon energy of the host materials.⁵¹ Low phonon energies of host materials are favorable for Ln doping to achieve intense luminescence, as it allows for low multiphonon relaxation rates and minimal nonradiative energy losses. Fluorides, owing to the low phonon energy ($\approx 350\text{ cm}^{-1}$) and high chemical stability, are considered to be the most efficient for many applications.⁵² Crystal structure also significantly affects the Ln luminescence, as low-symmetry is more desirable than high-symmetry due to the higher 4f–4f transition probabilities. For example, Krämer *et al.* reported that the $\text{NaYF}_4:\text{Yb}^{3+}/\text{Er}^{3+}$ UC efficiency of green emission is approximately 10 times stronger in hexagonal phase compared to cubic phase,⁵³ when Quintanilla *et al.* revealed that the photoluminescence quantum yield of α -phase $\text{NaGdF}_4:\text{Er}^{3+},\text{Yb}^{3+}$ surpasses that of β - NaGdF_4 for sizes below 20 nm, which can be related to distortion of the crystal lattice when the UC NPs become smaller.⁵⁴

Consequently, with typical bare NPs, it is essential to carefully select the host material with appropriate parameters of interest for achieving optimal Ln luminescence in a given application.

In the last decade, new challenges have emerged for the engineering and synthesis of ultrabright lanthanide-based NPs (Ln-NPs). One creative approach involves the coordination of fluorescent organic ligands (antennas) at the NP surface to sensitize the incorporated Ln^{3+} , allowing light harvesting over broader wavelength ranges with greatly enhanced NPs absorption cross section. These ligands also contribute in the shielding of Ln emission from high-energy oscillators in the chemical matrix. Thus, a very efficient sensitization of Ln-NPs can be obtained through surface coverage with an adequate capping ligand and the surface-modified NPs display huge improvements of their photophysical properties and their brightness in particular.

In this case, an appropriate selection of the capping antenna becomes very important for the spectroscopic performance of the Ln-NPs as the main pathway of reaching Ln related excited state is through the energy transfer from the antenna and not from its direct excitation anymore, subsequently the nature of the host material seems to have less impact on the final luminescence performance of dye-sensitized Ln-NPs. This is illustrated by the diversity of suitable matrices doped with Ln^{3+} that are reported to be effective for dye-sensitization, such as oxides, fluorides, phosphates, vanadates, silicates, and hydroxides, as presented below.

This organic/inorganic dual nanomaterial received increasing attention thanks to the possibility of combining the low cost and versatility of organic molecules with the chemical and physical properties of the inorganic materials, giving a great utility for many applications, such as luminescent sensors, optical fibers, lasers, amplifiers, solar cells, electroluminescent devices, and various time-correlated luminescence applications. In this section, we will review the different spectral conversion mechanisms illustrated in Fig. 1 and investigated in dye-sensitized Ln-NPs: (1) downshifting, where one high-energy photon is transformed into one lower energy photon; (2) upconversion, where two or more low energy photons are converted into one high-energy photon; and (3) quantum cutting, also called downconversion, in which one high-energy photon is converted into several lower energy photons. A comparison of the photoluminescence properties of the various NPs (when studied) is summarized at the end of this section in Table 1, and the various dye-sensitizing ligands reported in

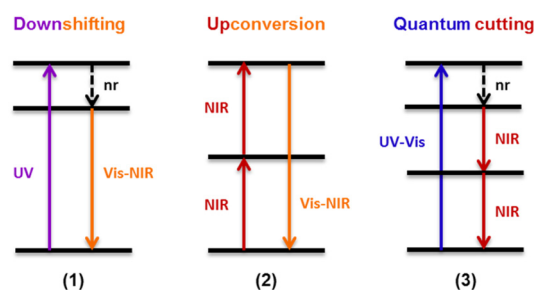


Fig. 1 Schematic representation of different spectral conversion mechanisms possible with Ln^{3+} -doped NPs. nr = non-radiative.



Table 1 Comparison of the photoluminescence properties of the Ln doped NPs with capping dye-sensitizers

Matrix	Ln	Antenna	Diameter (nm)	λ excitation (nm)	Lifetime of emitting Ln (μ s)	Enhancement factor ^a / B^b (L mol ⁻¹ cm ⁻¹)	Quantum yield (%)	Ref.
LaF ₃	Eu	1	232 \pm 65 ^c	305	125	135	1.85 \pm 0.28	55
LaF ₃	Tb	2	25–40	282		44		58
LaF ₃	Eu	5	3–4	278		100		61
LaF ₃	Eu	6	3.5	275	510 ^{31%} 1990 ^{69%}			62
LaF ₃	Tb	6	3	319	2296 ^{55%} 8305 ^{45%}			63
LaF ₃	Tb	7	3.32	312	641 ^{37%} 2604 ^{63%}			64
		8		320	1713 ^{34%} 3453 ^{66%}			
LaF ₃	Tb	9	5.2	265		100		65
LaF ₃	Eu	10	25	337		100	70	66
		12						
LaF ₃	Tb	14	20–25	307	1580 ^{48%} 3760 ^{52%}	$B = 2.1 \times 10^6$	13	16
		15		337	1110 ^{47%} 2590 ^{53%}	$B = 2.2 \times 10^6$	29	
LaF ₃	Eu	16	10	350		180	61.6	67 and 68
LaF ₃	Eu	2	25–40	286		88		69 and 70
		16		378				
LaF ₃	Eu	17	16	377		20		71
		18						
LaF ₃	Eu	18	4.4 \pm 1.2	457	80 ^{53%} 320 ^{31%}		13.25	73
					1650 ^{16%}			
LaF ₃	Tb	20	20–25	337	660 ^{18%} 2070 ^{82%}		38 \pm 1	75
		21						
LaF ₃	Tb	22	20–25	335	726		8–15	76
CaF ₂	Tb	11	11 \pm 4	311	2740			80
NaYF ₄	Tb	24	60–110	320	2350 [—] 570 [—]	330	19	81
	Eu				1860 [—] 7200 [—]		14	
LiYF ₄	Eu	25	90 \times 40	365	2057 ^{53%} 9135 ^{47%}		31	82
NaGdF ₄	Eu	26		317		10 000	3.3 \pm 0.6	83,84
NaGdF ₄	Eu	2–18	33.9	277			27.6	85
Y ₂ O ₃	Eu	27	6.4 \pm 1.5	270			19	86
Y ₂ O ₃	Tb	28	100	290	1250	100		87
YPO ₄	Eu	16	23.2 \pm	350		4700	26 \pm 2	91
			8.8					
YVO ₄	Eu	16	10–15	369		14		92
LaPO ₄	Eu	16	7	350	850	33	22	94
SiO ₂	Eu	29	207 \pm 13	343	83 ^{7%} 283 ^{31%}	190	49	97
					1085 ^{62%}			
NaYF ₄	Nd	30	5.3 \pm 0.6	340	4.1 ^{20%} 68 ^{80%}			101
	Yb		6.0 \pm 0.6		1.1 ^{15%} 3.7 ^{63%}			
					12.6 ^{22%} (in DMSO)			
NaYF ₄	Yb	31	5.2–6.3	460	3 ^{6%} 16.5 ^{24%}	300		102
					111 ^{70%}			
CaF ₂	Yb/Nd	33	4.4 \pm 0.1	467	17	2100		103
NaYF ₄	Yb/Er	34	16	806		3300	0.12	109
NaGdF ₄	Yb/Er	34	12	806		33 000	5.3	110
NaYF ₄	Yb/Er	35	20	783		80		111
		36		820		70		
		37		808		200		
		38		845		100		
NaYF ₄ :Yb/Er@NaYF ₄ :Yb	Yb/Er	34	35	806		1000		112
NaLuF ₄ :Yb/Er@NaLuF ₄ :Yb,Pr	Yb/Er	36	18	820		800		113
NaYbF ₄ :Tm@NaYF ₄ :Nd	Yb/Tm	37	54	808			4.8	114
NaYF ₄ :Yb/Er@NaYF ₄ :Nd/Yb	Yb/Er	41	33	800			9.2	116
NaYF ₄ :Yb/X@NaYbF ₄ @NaYF ₄ :Nd (X = Er, Ho, Tm or Pr)	Yb/X	41	52	800			13	117
NaGdF ₄ :Yb/Er@NaGdF ₄ :Yb@NaNdF ₄ :Yb	Yb/Er	37	56.4	808		7		119
NaYF ₄ :Yb,Er,Nd	Yb/Er	34	50	806				120
NaYF ₄ :Yb,Er@NaYF ₄ :Yb,Nd	Yb/Er	35	16	783		34	1	121
NaYF ₄	Tb/Yb	6	2.71	272	1370 ^{68%} 3192 ^{32%}			123
NaYF ₄	Pr/Yb	42	22 \times 55	397	21.7	30		124
NaYF ₄	Tb/Yb	43	30	405		2260		125
NaGdF ₄ :Nd/Yb@NaGdF ₄ :Nd	Nd/Yb	26	10	355	626 \pm 48			126

^a Enhancement factor is the ratio of the dye-sensitized NPs luminescence to that of bare NPs. ^b B = Brightness. ^c Hydrodynamic diameter measured by DLS.



the literature for downshifting and for both upconversion and downconversion are illustrated in Fig. 2 and Fig. 6 respectively.

3.1. Downshifting NPs

Most of the studies published in the field of Ln-NPs sensitized by organic antennas have been focused on the downshifting mechanism, by shifting UV-blue light to more advantageous and better exploited visible and NIR emission.

a UV to visible conversion. Eu^{3+} and Tb^{3+} are the most reported Ln in such spectral conversion. These respectively red and green emitters may find interesting applications in numerous fields based on fluorescence detection.

Charbonnière *et al.* were first to report such type of spectral conversion.⁵⁵ In a pioneering work of 2008, they described sensitization of 5%Eu-doped LaF_3 -AEP (AEP for aminoethyl-phosphate) NPs by partial AEP exchange with 6-carboxy-5'-methyl-2,2'-bipyridine (**1**) in water. The NPs excitation spectrum reflected the characteristic absorption bands of the ligand with a maximum at 305 nm, and the ligand to Eu^{3+} energy transfer produced a 135-fold enhanced emission. This general strategy was then expanded for different combinations of chromophores, Ln, and inorganic host material.

Later, Kokuoz *et al.* architecturally developed a new type of "core-shell nanostructure" where specific emitting Ln are constrained by an undoped layer, forming a shell protecting those in the core.⁵⁶ They demonstrated that benzoic acid (**2**) and phthalic acid (**3**) could efficiently sensitize Eu^{3+} and Tb^{3+} doped LaF_3 core-shell NPs, showing that an extra undoped LaF_3 shell did not inhibit the sensitization of core Eu^{3+} , proving that energy-transfer can penetrate to sites which are not directly ligand-bound, referring to a dipole-dipole mechanism.

2 was then frequently investigated as Chen *et al.* confirmed its ability to sensitize Eu-doped LaF_3 NPs,⁵⁷ while Yang *et al.* proved that it can also sensitize Tb-doped LaF_3 NPs.⁵⁸ Taking advantage of these previous works, Wang *et al.* used **2** in a $\text{Eu}^{3+}/\text{Tb}^{3+}$ co-doped LaF_3 system in order to sensitize both Ln^{3+} , realizing multicolor Ln-NPs by a single wavelength excitation with tunable emission spectra through controlling $\text{Eu}^{3+}/\text{Tb}^{3+}$ molar ratios.⁵⁹

Then, Kokuoz *et al.* covered Eu-doped LaF_3 NPs with 3,4-formylphenylbenzoic acid (**4**), and demonstrated that through a balance between ligand and lanthanide emission, these NPs exhibit color tunability from red to greenish blue as a function of selected excitation wavelengths ranging from 250 to 400 nm.⁶⁰

In a similar system, Cross *et al.* reported a strong sensitization of 5% Eu-doped LaF_3 -citrate NPs upon citrate exchange by dipicolinate ligands (**5**), increasing the emission intensity by a factor of 100.⁶¹

In 2014, Li *et al.* demonstrated that 1,2,4,5-benzenetetracarboxylic acid (**6**), due to its high symmetry and various coordination with Ln^{3+} , can efficiently bind to the surface of Eu-doped LaF_3 NPs and strongly sensitize and protect Eu^{3+} luminescence,⁶² while Li *et al.* highlighted its ability to sensitize Tb as well, in Tb-doped LaF_3 .⁶³ In a later study, they also

introduced salicylate (**7**) and 5-sulfosalicylate (**8**) as efficient sensitizers of Tb-doped LaF_3 .⁶⁴ Different benzoic acid derivatives were then exploited as Ghosh and Luwang described the sensitization of Tb^{3+} doped LaF_3 NPs by *p*-amino-benzoic acid (**9**), reporting at least 100-fold enhancement in luminescence intensity.⁶⁵ They also applied their model for the quantification of nitro explosive compounds in aqueous medium whereas Khudoleeva *et al.* aimed for bioimaging applications by performing surface modification of Eu-doped LaF_3 with terephthalate (**10**) or 2,6-naphthalenedicarboxylate (**12**), increasing luminescence intensity by two orders of magnitude.⁶⁶

Then, in a wide study, Goetz *et al.* reported the sensitization of 10% Tb-doped LaF_3 using eleven different ligands derived from dipicolinic acid **5** and 2-hydroxyisophthalic acid (**13**) with varying coordination and photosensitizing abilities.¹⁶ The two most effective photosensitizing ligands were 5-cyano,2-hydroxyisophthalic acid (**14**) and the bis(2-hydroxybenzoic acid) (**15**) as they provided very promising brightness values up to $2.2 \times 10^6 \text{ L mol}^{-1} \text{ cm}^{-1}$ which exceed those of QDs and semiconducting nanopolymers. The NPs were used for highly sensitive time-resolved luminescence applications, like imaging in HeLa cells by fluorescence microscopy.

Beyond carboxylic acid ligands, Janssens *et al.* introduced β -diketonates as efficient sensitizers for Eu^{3+} , as they showed the possibility to sensitize 5%Eu-doped LaF_3 -OA (OA = oleic acid) NPs by partial surface coverage with thenoyltrifluoroacetone (**16**) and the resultant luminescence intensity was enhanced 180 times.⁶⁷ They also succeeded to press these modified NPs at high pressure, creating transparent solid discs of high density.⁶⁸ **16** was shown to be as efficient as **2** at photosensitizing Eu-doped LaF_3 NPs, as Wang *et al.* observed a 40-fold luminescence intensity enhancement for both ligands.⁶⁹ In a recent work, this team reported the coordination of Eu-doped LaF_3 with both ligands simultaneously. This mixed hybrid possesses a broadband excitation spectrum (200~400 nm) which perfectly covers the entire ultraviolet (UV) spectral range and can be extremely beneficial to the enhancement of the conversion efficiency of silicon solar cells.⁷⁰

Advanced studies aimed to find different sensitizers for Eu^{3+} in such type of NPs. Safronikhin *et al.* proved that dibenzoylmethane (**17**) and 1,10-phenanthroline (**18**) are able to sensitize EuF_3 NPs,⁷¹ while Irfanullah *et al.* showed the ability of **18** and 9-oxidophenalenone (**19**) to sensitize water dispersed 5%Eu-doped LaF_3 NPs.^{72,73} The latest ligand ensured great protection of Eu^{3+} from non-radiative deactivation through high-energy vibrations of **19**, as proved by the relatively long lifetime of Eu^{3+} emission up to 0.41 ms. In a recent study, Adusumalli *et al.* tested $\text{LaF}_3:\text{Eu}^{3+}$ and $\text{SrF}_2:\text{Eu}^{3+}$ NPs photosensitized by different ligands (**5**, **6**, **10** and **13**) for their haemocompatibility.⁷⁴ Therefore, flow cytometry was used to analyse the possible NPs binding to the red blood cell membrane. They showed that these nanomaterials are non-cytotoxic compounds *in vitro* and can be further investigated for biomedical *in vivo* applications. Many studies claimed that these Ln-NPs are particularly appealing for bioanalytical applications, but no proof of concept was provided until Charpentier *et al.*



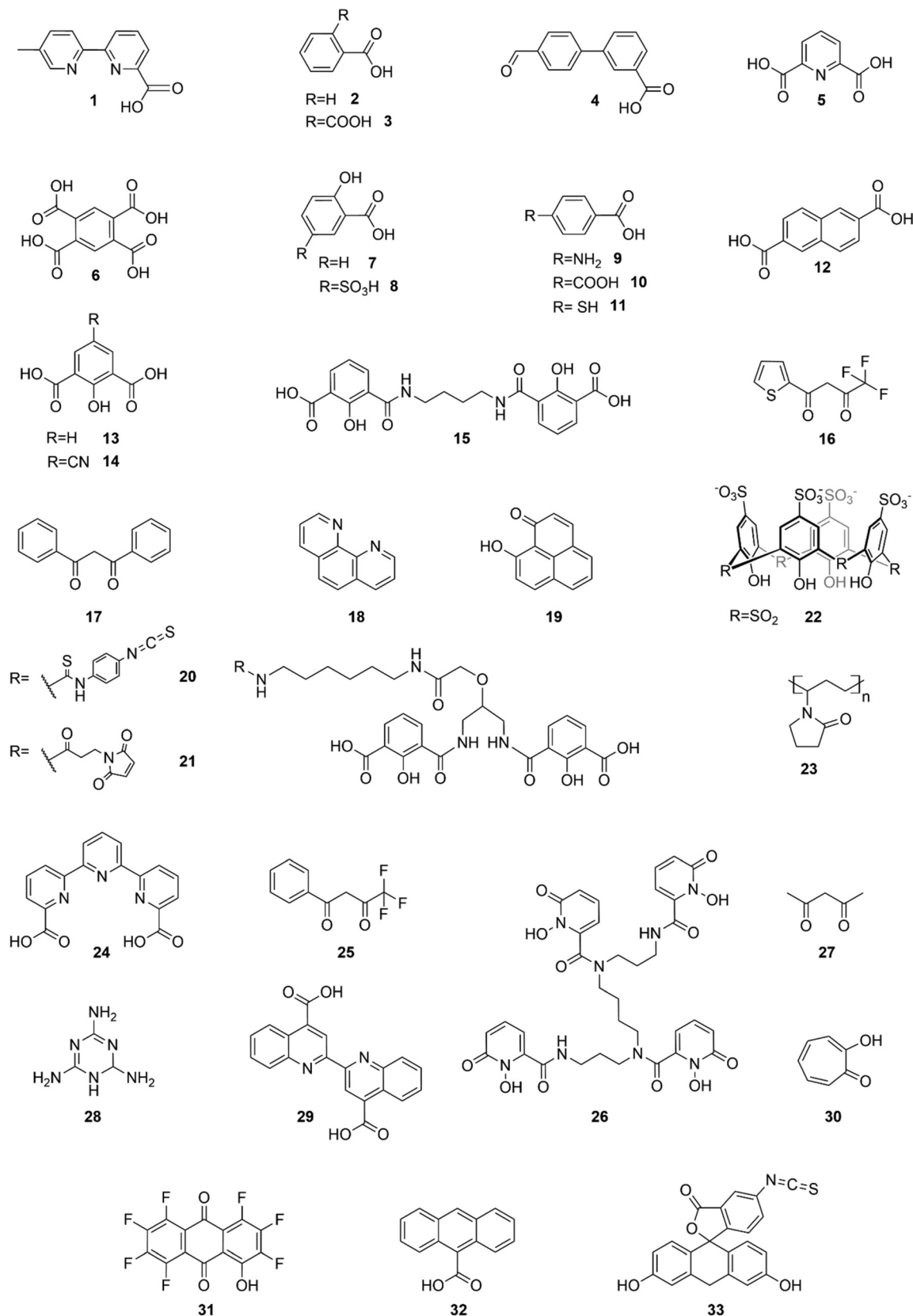


Fig. 2 Dye-sensitizing ligands reported in the literature for surface sensitization of downshifting NPs.



recently developed novel ligands which can be simultaneously used for both sensitization of Ln-NPs and bioconjugation.⁷⁵ They designed two linked hydroxy-isophthalic acid based units, where the linker was modified to introduce an activated function that react either with amine (**20**) or thiol (**21**) functions, making them linkable to many biomolecules of interest. These ligands showed important sensitization of 10%Tb-doped LaF₃ leading to exceptional NP brightness up to $1.8 \times 10^6 \text{ L mol}^{-1} \text{ cm}^{-1}$, while their bioapplicability was demonstrated in two prototypical approaches for bioimaging and bio-sensing. In further studies, Cheignon *et al.* used the same Ln-NPs for a surface coordination by *p*-sulfonato-sulfoxocalix[4] arene (**22**), which showed strong photosensitization of Tb emission upon ligand excitation.⁷⁶ Such design allows to exploit the calixarene cavity to a potential inclusion of aromatic compounds. An interaction with Rhodamine 6G was proved by FRET experiments that revealed a strong association to the surface of Tb-NPs, as shown in Fig. 3.

Besides LaF₃, alkali metal fluoride matrices have also been investigated as suitable Ln³⁺ host material. Wang *et al.* reported the sensitization properties of **2** in CaF₂ doped with either Eu³⁺ or Tb³⁺, showing a drastic increase in emission intensity compared with corresponding Eu and Tb molecular complexes.⁷⁷ However, those NPs could not be dispersed properly in water or organic solvents, which limited their application. Then, Song *et al.* synthesized Tb-doped CaF₂-OA which showed good dispersibility in chloroform and toluene.⁷⁸ Fluorescence measurements showed that even the surface coating OA, when excited, could slightly sensitize Tb³⁺, while in

a later report, they performed poly(*N*-vinyl-2-pyrrolidone) (**23**) coverage of their NPs doped with Eu, Tb, or Tb/Ce, using each pyrrolidone moiety of the polymer as an individual antenna coordinated to the NP surface.⁷⁹ **23** exhibited efficient sensitization of Tb or Eu-doped NPs, while for Tb/Ce co-doped system, simultaneous energy transfer from Ce³⁺ and the ligand was observed, evidencing a synergistic enhancing effect between a co-dopant and an organic antenna for the first time.

In 2019, Adusumalli *et al.* demonstrated that 4-mercapto-benzoic acid (**11**) is an efficient sensitizer for the Tb-doped CaF₂ NPs.⁸⁰ The prepared capped NPs were used to develop an approach for selective detection of nitroaromatic pollutants in water.

Other fluoride matrices were investigated as host material for dye-sensitized Ln-NPs. Eu or Tb-doped NaYF₄ nanocrystals have been prepared by Gauthier *et al.* and four N-heterocyclic organic ligands were tested to promote sensitization of Eu³⁺ or Tb³⁺ luminescence.⁸¹ The terpyridine derivative **24** showed the best performance for both Ln, presenting up to a 330-fold enhancement in emission intensity. Then, Samanta *et al.* proposed Eu-doped LiYF₄ NPs coating by 4,4,4-trifluoro-1-phenyl-1,3-butanedione (**25**) which provided efficient sensitization.⁸² They also proved the applicability of this model in Si solar cell efficiency enhancement.

On another hand, Agbo *et al.* detailed the photophysics of Eu-doped NaGdF₄ NPs and studied their sensitization by a hydroxypyridone derivative (**26**), responsible for a 10⁴-fold increase in luminescence intensity.⁸³ They also applied their model as a tool of choice to overcome the constraints of UV solar spectrum/semiconductor bandgap mismatch.⁸⁴ In a similar system, Song *et al.* used **2** and 1,10-phenanthroline (**18**) for surface modification in order to enhance the luminescence performance of Ln-doped NaGdF₄ (Ln = Tb, Eu, Dy) NPs.⁸⁵ Interestingly, the overlap in excitation bands for both Gd³⁺ ions and ligands ensured simultaneous energy transfer of Gd³⁺ → Ln³⁺ and ligands → Ln³⁺ under a single wavelength excitation.

Away from fluoride-based matrices, Y₂O₃ is one of the most studied metal-oxide matrices for Ln-NPs. First study of Eu-doped Y₂O₃ sensitization by acetylacetonate (**27**) was published by Dai *et al.* showing that the excitation of ligands leads to strongly enhanced white light emission arising from efficient intramolecular energy transfer to Eu³⁺ as well as Y₂O₃ oxygen vacancies.⁸⁶ Those optical properties allow for applications in UV LED pumped solid-state lighting. Then, Stagi *et al.* proved the sensitization effectiveness of Tb-doped Y₂O₃ by 2,4,6-triamino-s-triazine (**28**) presenting a 10²-fold luminescence enhancement.⁸⁷

The photosensitizing ability of the well-known β-diketonate (**16**) has been investigated with various types of matrices. Ji *et al.* and Chen *et al.* used it to sensitize Eu-doped Y₂O₃ NPs, showing a greatly enhanced luminescence intensity,^{88,89} while Balderas *et al.* employed the sensitized NPs to produce transparent poly(methyl methacrylate) (PMMA) luminescent films with an extended and tunable excitation wavelength range from 200 to 550 nm.⁹⁰

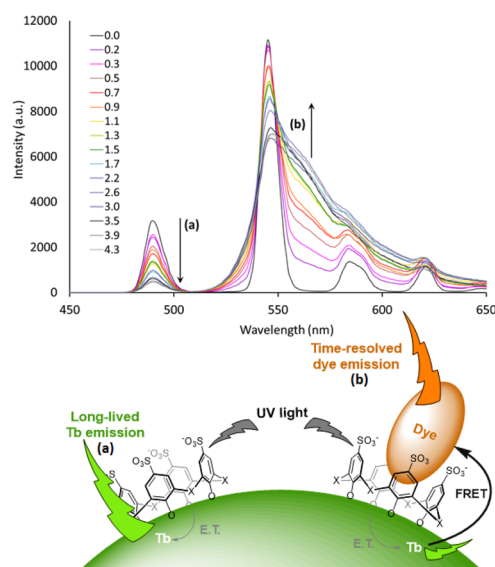


Fig. 3 Time-resolved emission spectra of Tb NP capped with **22** upon addition of different equivalents of rhodamine 6G. The intensity decrease of the Tb emission and increase of rhodamine emission are marked with arrows a and b respectively. Inset: representation of the proposed supramolecular assembly. NP photosensitization by surface capping ligands and host-guest interaction with the aromatic charged dye. Adapted with permission from ref. 76. Copyright 2021 Wiley-VCH Verlag GmbH & Co. KGaA.



Chen *et al.* showed that **16** could also sensitize Eu-doped $\text{YPO}_4(\text{OA})$ NPs with a resulting ~ 4700 -fold brighter emission,⁹¹ while only a very low sensitization occurred for Eu-doped YVO_4 NPs as demonstrated by Tang *et al.*⁹²

Moreover, the sensitization of Eu-doped LaOF NPs by **16**, prepared by He *et al.* by annealing Eu-doped LaF_3 NPs, resulted in higher luminescence enhancement compared with sensitized Eu-doped LaF_3 NPs.⁹³ Thus, Vats *et al.* succeeded to increase the luminescence emission of Eu-doped LaPO_4 NPs through **16** sensitization.⁹⁴ Interestingly, photoluminescence and lifetime measurements clearly showed that sensitization takes place only at the surface of LaPO_4 NPs, while the core Eu^{3+} remains unsensitized.

Embedding Ln^{3+} into a host material such as alkali halides, semiconductors, and metal oxides has been widely investigated. Gonçalves *et al.* introduced luminescent Eu-doped SnO_2 NPs where Ln^{3+} were found to be essentially incorporated into the cassiterite structure, substituting Sn^{4+} , while for high concentration they are also located at the particles surface.⁹⁵ Capping with another β -diketonate **25** presented high light output under UV excitation in water, due to an efficient sensitization and protection of Eu^{3+} emission. In 2015, Eu-doped ZnO nanowall structures have been achieved by an electrochemical deposition method by Kang *et al.* where Eu^{3+} were uniformly distributed in the ZnO-Zn(OH)_2 core/shell structure.⁹⁶ Surface modification by 1,10-phenanthroline (**18**) generated an additional sharp Eu^{3+} emission while the energy transfer from ZnO to Eu^{3+} appears to be extremely weak without ligand capping. The results led to propose a unique cascade energy transfer model between ZnO, **18** and Eu^{3+} .

Lately, Artizzu *et al.* suggested a concept model system based on purely silica-based core/shell NPs where Eu^{3+} ions are confined into a thin silica layer and are efficiently photosensitized through 2,2'-biquinoline-4,4'-carboxylate (**29**) covalently grafted on the surface of the outer shell.⁹⁷ A remarkable intensity enhancement of Eu-based NPs luminescence by 190-fold was reported, showing that silica matrices are suitable and a highly performing host alternative to commonly investigated nanocrystals for the development of Ln-based luminescent materials.

b UV-Vis to NIR conversion. Other lanthanides such as Yb^{3+} , Nd^{3+} and Dy^{3+} attracted particular interest due to their near infrared (NIR) emission appealing for several applications such as biosensing and biomedical imaging.⁹⁸ Additionally, they are efficient tools for solar cell technologies thanks to their NIR emission lines above the Si bandgap in silicon-based devices.^{99,100}

The first report on the use of organic ligands to sensitize NIR luminescence of Ln-NPs was published by Zhang *et al.* in 2007. In this pioneering work, a strategy was established to both protect and sensitize the NIR luminescence of 20%Nd or Yb-doped NaYF_4 NPs by direct coordination of tropolonate (**30**) ligands to the NPs surface, as proven by longer luminescence lifetimes than for the corresponding molecular complex $[\text{Ln}(\text{Trop})_4]^-$.¹⁰¹ In a similar NaYF_4 system, Lu *et al.* reported the sensitization of Yb-doped NaYF_4 NPs through the excitation of

capping 2-hydroxy-perfluoro-anthraquinone (**31**). The overall Yb^{3+} NIR emission intensity is increased by a factor of 300.¹⁰²

In 2016, Utochnikova *et al.* developed a photoluminescence study on sensitization of YbF_3 and EuF_3 , with 2,6-naphthalenedicarboxylate (**12**) or 9-anthracenate (**32**). Both ligands enhanced EuF_3 luminescence intensity up to 100 times, while in the case of the NIR emitting YbF_3 system, successful sensitization was achieved only with **32**.¹⁰⁴ In a later study, they reported the luminescence properties of rarely studied Dy-based NPs, DyF_3 and Dy-doped LaF_3 , both sensitized by terephthalic acid (**10**) with enhanced Dy^{3+} NIR emission observed upon surface modification.¹⁰⁵

Recently, Liu *et al.* presented a novel type of dye-sensitized $\text{CaF}_2:\text{Yb}^{3+}/\text{CaF}_2:\text{Nd}^{3+}$ core/shell NP, with fluorescein isothiocyanate FITC ligands (**33**) capping the surface of NPs and acting as efficient visible light harvester (Fig. 4a).¹⁰³ The strong spectral overlap between **33** emission and Nd^{3+} absorption (Fig. 4b) as well as between Nd^{3+} emission and Yb^{3+} absorption (Fig. 4c), makes Nd^{3+} a suitable “energy bridge” to realize effective multistep sequential dye $\rightarrow \text{Nd}^{3+} \rightarrow \text{Yb}^{3+}$ energy transfer. This ultraefficient cascade mechanism resulted in a remarkable enhancement of about 2100 times of the Yb^{3+} luminescence intensity, which is the highest figure of merit reported in literature so far for NIR-emitting analogous systems.

3.2. Dye-sensitized upconversion NPs

In the last decade, lanthanide-doped upconversion NPs (UCNPs) have attracted worldwide attention due to their capability of generating shorter-wavelength photons under infrared

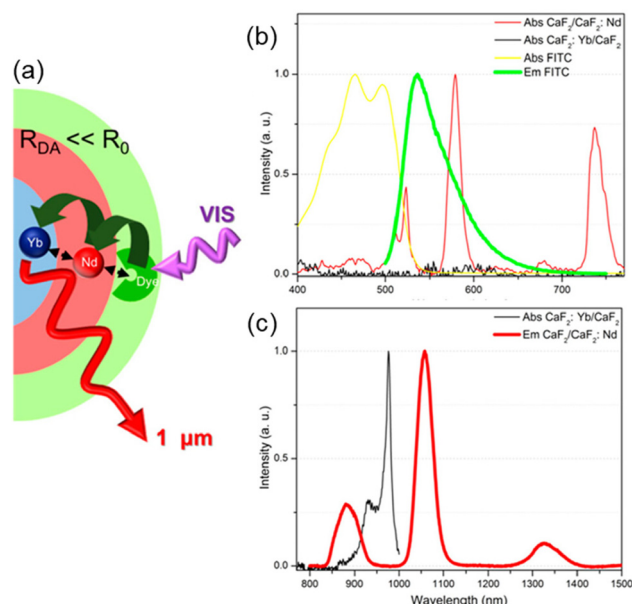


Fig. 4 (a) Architectural design of $\text{CaF}_2:\text{Yb}^{3+}/\text{CaF}_2:\text{Nd}^{3+}$ core/shell NP dye-sensitized by **33** and (b–c) spectral overlaps between donor–acceptor emission–absorption bands for multistep sequential energy transfer. Adapted with permission from ref. 103. Copyright 2019, American Chemical Society.



light excitation, with promising applications in biomedical imaging, photodynamic therapy, solar cells, and display technologies.¹⁰⁶

Generally, upconversion NPs consist of a couple of lanthanide dopants (typically $\text{Yb}^{3+}/\text{Ho}^{3+}$, $\text{Yb}^{3+}/\text{Er}^{3+}$, or $\text{Yb}^{3+}/\text{Tm}^{3+}$) in an organic host material. Yb^{3+} (or alternatively Nd^{3+}) ions are usually doped to function as sensitizer ions, absorbing the 980 nm (or 808 nm for Nd) laser irradiation and then successively transfer their excitation energy to nearby co-doping activators (Ho^{3+} , Er^{3+} , Tm^{3+} ...), finally leading to the emission of upconverted light (UCL).¹⁰⁸ However, the weak and narrow absorption bands of lanthanide ions pose a fundamental limit of UCNPs to withhold their brightness, creating a long-standing hurdle for the field. Dye-sensitization emerged once again to address this performance-limiting problem.

In 2012, Zou *et al.* reported the first attempt to enhance the brightness of $\text{NaYF}_4:\text{Yb}^{3+}/\text{Er}^{3+}$ UCNPs through organic carboxylated cyanine dye IR-806 (**34**) sensitization.¹⁰⁹ FRET occurs from the excited **34** to the Yb^{3+} absorption centers on the surface of NPs, which further sensitized Er^{3+} to produce UCL. The overall upconversion is dramatically enhanced by a factor of 3300. In a similar system, Garfield *et al.* tested the same typical $\text{Yb}^{3+}/\text{Er}^{3+}$ couple and **34** in a NaGdF_4 inorganic host lattice, and reported a 33 000 times increase in brightness and a 100-fold increase in efficiency over uncapped UCNPs.¹¹⁰ Later, this approach was extended and applied to a set of $\text{NaYF}_4:\text{Yb}^{3+}/\text{X}^{3+}$ ($\text{X} = \text{Er}$, Tm or Ho) NPs and many NIR dyes with distinctive absorption ranges for more efficient absorption and a tunable excitation band in a wide spectral range. In their study, Wu *et al.* used a series of NIR dyes and showed their ability to sensitize $\text{NaYF}_4:20\%\text{Yb}^{3+}$, $2\%\text{Er}^{3+}$ UCNPs.¹¹¹ These included commercially available dyes IR-783 (**35**) and IR-820 (**36**) and their respective carboxylate derivatives IR-808 (**37**) and IR-845 (**38**) which were synthesized.

Thanks to the modular nature of NIR dyes, Lee *et al.* suggested a simultaneous use of multiple types of dyes on the surface which can dramatically widen the photon absorption window of $\text{NaYF}_4:20\%\text{Yb}^{3+}$, $0.5\%\text{Tm}^{3+}$ UCNPs, to the entire visible and NIR range.¹⁰⁷ Three types of dye sensitizers, **34**, BODIPY-FL (**39**) and Cy3.5 (**40**), were chosen to perform sensitization. The sufficient spectral overlap between the emission spectrum of one sensitizer and the absorption spectrum of another enabled a cascade FRET sensitization of the UCNPs, while empowering collective absorption spectra of the three ligands as shown in Fig. 5. In 2016, Wu *et al.* introduced core/shell nanostructured $\text{NaYF}_4:\text{Yb}^{3+}/\text{Er}^{3+}@\text{NaYF}_4:\text{Yb}^{3+}$ UCNPs where Yb^{3+} sensitizer was directly doped in the shell.¹¹² Through a sensitization by **34**, and Ln^{3+} extra protection provided by the core/shell design, they succeeded to amplify upconversion efficiency, and aimed for their application in controlling neuronal activity and bioimaging.

Similar results were also observed by Yin *et al.* using **36** sensitized $\text{NaLuF}_4:\text{Yb}^{3+}/\text{Er}^{3+}@\text{NaLuF}_4:\text{Yb}^{3+},\text{Pr}^{3+}$ core/shell UCNPs, with a 800 times UCL enhancement.¹¹³

Despite recent progress, several drawbacks persist as the limitation of spectral overlap between the emission of NIR

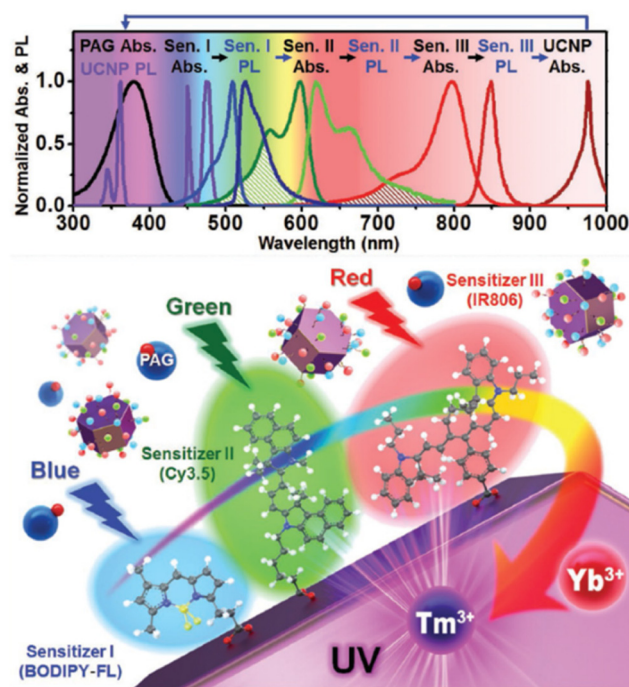


Fig. 5 Design of multi-dye-sensitized UCNPs for wide-range photo-absorption and upconversion. Reprinted with permission from ref. 107. Copyright 2016 Wiley-VCH Verlag GmbH & Co. KGaA.

dyes and the absorption of the typically used sensitizer Yb^{3+} ions, restraining energy transfer efficiency from the dye to the UCNPs. Then, Chen *et al.* proposed further advances involving the concept of multistep cascade energy transfer in which **37** is used to sensitize Nd^{3+} doped in the shell layer, whereas Yb^{3+} and Tm^{3+} are a second sensitizer and activator respectively, in the core of an epitaxially designed core/shell ($\text{NaYbF}_4:\text{Tm}^{3+}0.5\%$) $@\text{NaYF}_4:\text{Nd}^{3+}$ UCNPs.¹¹⁴ They demonstrated that a multistep cascade energy transfer dye $\rightarrow \text{Nd}^{3+} \rightarrow \text{Yb}^{3+}$ is about 1.5 times more efficient than the direct energy transfer from **37** to Yb^{3+} yielding an upconversion efficiency nearly ~ 100 times higher than typically reported for Ln-UCNPs.

Later, Wei *et al.* showed that optimal doping concentration of Nd^{3+} in such UCNPs system could be increased from 2 to 20 mol% resulting in additional 10 times higher upconversion brightness.¹¹⁵

In additional studies, Chen *et al.* proved that the achieved UCL intensities from the sensitization of $\text{NaYF}_4:\text{Yb}^{3+}/\text{Er}^{3+}@\text{NaYF}_4:\text{Nd}^{3+}/\text{Yb}^{3+}$ core/shell NPs by indocyanine green organic dye (**41**) were 2–3 times higher than that from sensitized NPs incorporating only Yb^{3+} or Nd^{3+} in the shell layer.¹¹⁶ This was attributed to the strong overlap of the emission spectrum of this ligand with the absorption peaks of Nd^{3+} and Yb^{3+} exhibiting a synergistic effect from the multidimensional energy transfer involving multiple pathways: **41** \rightarrow shell $\text{Yb}^{3+} \rightarrow$ core Yb^{3+} ; **41** \rightarrow shell $\text{Nd}^{3+} \rightarrow$ core Yb^{3+} ; **41** \rightarrow shell $\text{Nd}^{3+} \rightarrow$ shell $\text{Yb}^{3+} \rightarrow$ core Yb^{3+} . Shao *et al.* reported a higher luminescence enhancement ($\times 28$) with $\text{NaYF}_4:\text{Yb}^{3+}/\text{Er}^{3+}@\text{NaYF}_4:\text{Yb}^{3+}/\text{Nd}^{3+}$ core/shell NPs sensitized by **34**¹¹⁷ and then extended the core/shell concept by



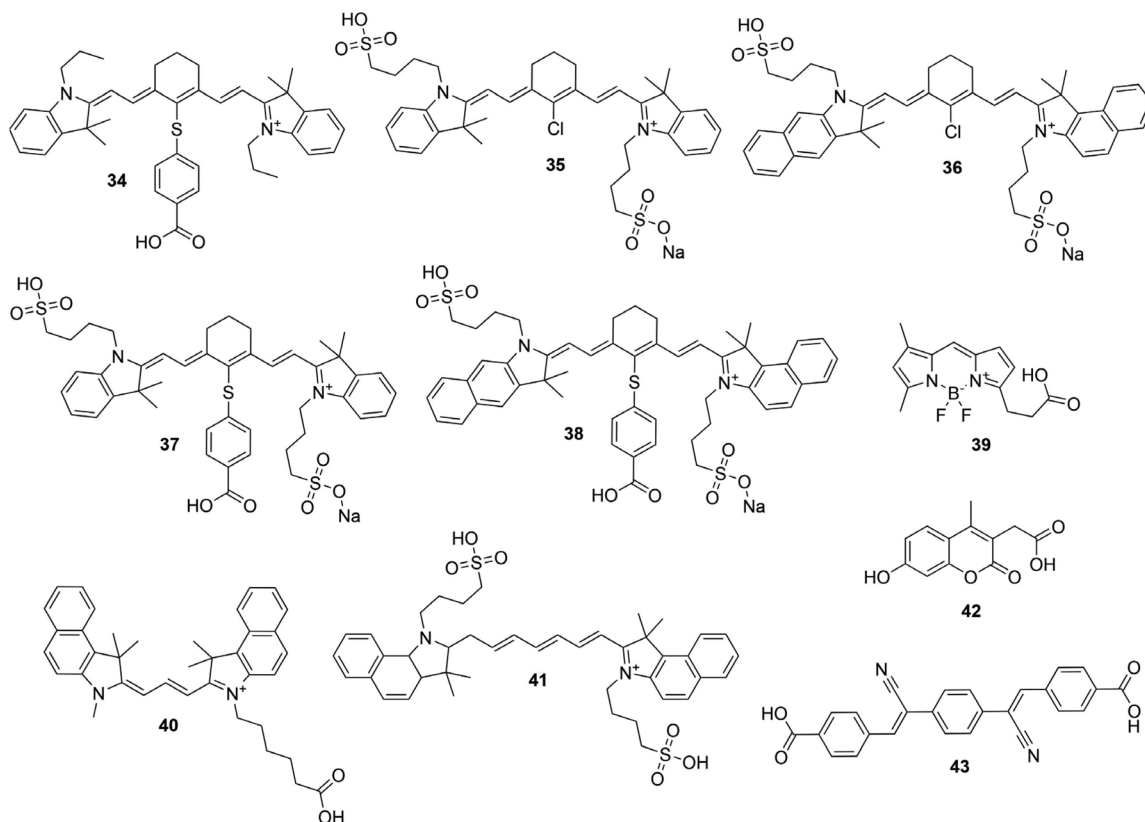


Fig. 6 Reported photosensitizing ligands for upconversion and downconversion related NPs.

presenting $\text{NaYF}_4\text{:Yb}^{3+}/\text{X}^{3+}@\text{NaYbF}_4@\text{NaYF}_4\text{:Nd}^{3+}$ core/shell/shell nano-structure ($\text{X} = \text{Er}, \text{Ho}, \text{Tm}$ or Pr) sensitized by **41**.¹¹⁸ Thanks to a cascade energy transfer pathway: **41** \rightarrow Nd^{3+} (outer shell) \rightarrow Yb^{3+} (inner shell) \rightarrow $\text{Yb}^{3+}/\text{X}^{3+}$ (core), the brightness was 4-times increased with a broad excitable spectral range which facilitates their use in bioapplications.

Finally, by combining cascade and multidimensional energy transfer pathways, Xu *et al.* demonstrated that the UCL intensity of $\text{NaGdF}_4\text{:Yb}^{3+}/\text{Er}^{3+}@\text{NaGdF}_4\text{:Yb}^{3+}@\text{NaNdF}_4\text{:Yb}^{3+}$ NPs could be enhanced over 7-fold by sensitization with **37**.¹¹⁹ The advantage of this core/shell/shell design with Nd^{3+} and Yb^{3+} co-doped in the outer shell is that nearly 100% Nd^{3+} allow efficient extraction of the excited-dye energy, while the Yb -containing shell layer minimizes possible back energy transfer from the core to the surrounding environment, thus maximizing the UCL efficiency.

Such dye-sensitized UCNPs have attracted worldwide attention due to their promising applications. Among recent works, Wei *et al.* found that the temperature increases significantly through energy conversion of **34** dye sensitized $\text{NaYF}_4\text{:Yb}^{3+}, \text{Er}^{3+}, \text{Nd}^{3+}$ UCNPs. When applied to a tumor, NPs are found successfully distributed to tumor cells, which can be killed efficiently based on the photothermal effect of the NPs.¹²⁰ Therefore, they are promising to be used for effective thermal therapy of tumors with real-time temperature monitoring.

Dye-sensitized UCNPs found interest in biophotonic applications like biological imaging, multimodal imaging and photodynamic therapy. In addition, the use of these UCNPs in solar cells have achieved rapid progresses. The highest power conversion efficiency was reported by Bi *et al.* at 21.1% using **35** dye sensitized core/shell $\text{NaYF}_4\text{:Yb}^{3+}, \text{Er}^{3+}@\text{NaYF}_4\text{:Yb}^{3+}, \text{Nd}^{3+}$ UCNPs and coupled with plasmonic Au nanorods film.¹²¹ This work indicates that insulating dye-sensitized UCNPs are of great significance for the future of solar cells devices.

3.3. Dye-sensitized quantum cutting NPs

Another conversion mechanism known as quantum cutting or downconversion (DC, not to be confused with the conventional downshifting), allows to transform the energy of one absorbed photon into two or more emitted low-energy photons, with quantum efficiency potentially larger than 100% (Fig. 1). This phenomenon has been demonstrated in various $\text{Ln}^{3+}\text{--Yb}^{3+}$ co-doped couples ($\text{Ln} = \text{Tb}, \text{Tm}, \text{Pr}, \text{Ho}$ or Dy), where Yb^{3+} functions as an ideal acceptor with its unique NIR emission band at around 1000 nm, specifically important for photovoltaic applications.¹²²

While the low absorption cross section of Ln hampers practical application, organic-inorganic hybrids are an alternative option to sensitize NIR quantum cutting in Ln-NPs. This approach has been described for the first time in 2013 by Li *et al.* with a combination of a UV-absorbing sensitizer **6** and



$\text{Tb}^{3+}/\text{Yb}^{3+}$ co-doped NaYF_4 NPs.¹²³ Upon absorption of **6**, the energy is non-radiatively transferred to coordinated Tb^{3+} , and by cooperative energy transition through downconversion, Tb^{3+} transfers the energy to Yb^{3+} , which in turn undergoes a multiphoton relaxation and subsequent emission in the near infrared region. As a downconversion luminescent converter, this kind of material meant to be useful in Si-based solar cells to reduce thermalization loss and enhance conversion efficiency of solar cells *via* spectral modification. But this type of spectral conversion mechanism remains rarely studied. In 2018, Wang et al. reported another Ln combination for an efficient dye-sensitized downconversion using a strong UV/blue absorbing Coumarin dye (**42**) to sensitize quantum cutting of $\text{Pr}^{3+}/\text{Yb}^{3+}$ couple in NaYF_4 NPs.¹²⁴ Photoluminescence and lifetime measurements, demonstrated a FRET from Coumarin to Pr^{3+} followed by downconversion, resulting in Yb^{3+} NIR emission with ~ 30 times enhancement with respect to the bare NPs.

The efficiency of downconversion emission in these NPs is limited by the concentration quenching effect due to non-radiative recombination paths set by intramolecular motions mechanisms, common to most aromatic hydrocarbons and their derivatives.¹¹⁵ Shao *et al.* introduced the concept of an ultimate photosensitization by aggregation-induced enhanced emission luminophores (AIEE LP) dyes to overcome this limitation.¹²⁵ This concept illustrated in Fig. 7 was demonstrated by completely covering the surface of $\text{Yb}^{3+}/\text{Tb}^{3+}$ co-doped NaYF_4 NPs with a dicyanostilbene derivative AIEE dye (**43**), which is designed for efficient attachment to the NPs at high density to maximize absorbance while passivating the surface. The energy transfer resulted in a 2260-fold enhancement of multiphoton downconversion by quantum cutting relative to

the dye-free NPs. In a prototypical application, this quantum cutting of UV photons to NIR photons matching with Si solar cells bandgap produced a 4% increase in efficiency under concentrated solar illumination.

Recently, Agbo *et al.* demonstrated the role that core/shell structures can play in this type of spectral conversion. They used **26** as a UV photosensitizer of $\text{NaGd}_{1-x}\text{Nd}_x\text{Yb}_y\text{F}_4/\text{NaGd}_{1-x}\text{Nd}_x\text{F}_4$ core/shell NPs,¹²⁶ enabling an efficient, step-wise ligand-donor-acceptor ($\text{26-Nd}^{3+}\text{-Yb}^{3+}$) energy transfer that mitigates the possibility of efficiency losses arising from the direct sensitization of Yb^{3+} by **26**. Using a detailed power dependence study, they described a spectral transformation through partial utilization of decay channels that permit the production of two NIR photons per UV photon absorbed. These results are important to challenge the energetic mismatches between solar illumination and the spectral response of commercial photovoltaic cells. However, dye-sensitized quantum cutting, with external quantum efficiency larger than 100%, is yet to be reported.

To summarize, Ln incorporation into inorganic matrix covered with capping antennas offers a very high protection from environment related deactivation, as highlighted by long excited-state lifetimes exhibited by numerous NPs (see Table 1). Various types of organic matrices can host Ln and the doping level can be tuned as desired. However, despite these important and promising merits, this type of nano-objects still has some drawbacks, such as the stabilization of the organic dyes on the surface of NPs, as the coordination of the ligands to metallic ions on the surface suggested an electrostatic, non-stable bonding nature. Moreover, the NP (bio)functionalization for analytical applications is a tricky point as significant organic synthesis work is needed in order to obtain appropriate antennas with an active function. The photostability of the organic dyes over time on the surface of NPs could also be a point to be considered as they are not protected. To overcome these points, organic/inorganic systems could be isolated from the environment (by encapsulation inside polymeric or silica coatings) to prevent dissociations or photochemical reactions and facilitate the functionalization.

4. Ln-Complexes encapsulated into NPs

A second strategy for cumulating the properties of Ln complexes is their encapsulation inside NPs. Incorporation of several Ln complexes into one NP has two advantages: (i) it allows to drastically increase the brightness of the entity compared to the single complex, by cumulating the absorption properties of chelates, and (ii) the complexes are protected from their environment and thus suffer less from deactivation.

In order to optimize the synthesis and obtain NPs with the best luminescence properties, it is important to have an estimation of the number of Ln-complexes (or in some cases the weight percentage (wt%) relative to the polymer) in a NP. While the complex encapsulation is often assumed to be quan-

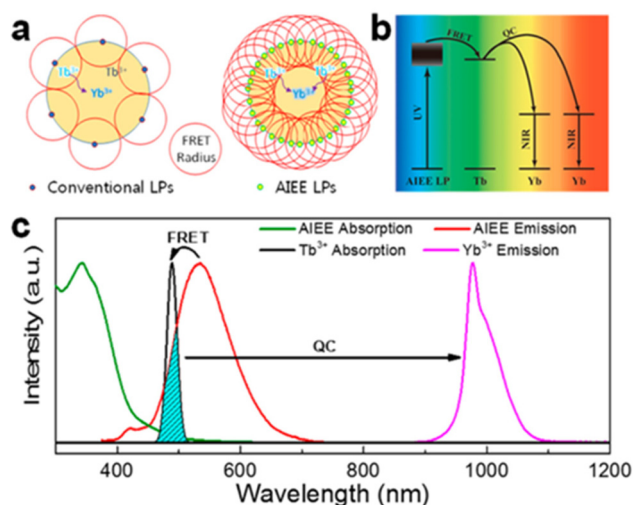


Fig. 7 (a) Schematic illustration of the difference between conventional emission and AIEE in the sensitization of quantum cutting. (b) Energy-transfer pathway from the AIEE LPs to NPs. (c) Normalized absorption and photoluminescence spectra of the AIEE dye and NPs. Reprinted with permission from ref. 125. Copyright (2018) American Chemical Society.



titative (the initial and final wt% are considered to be equal), an estimation may be done by assuming that no quenching occurs in the NPs and thus by directly comparing the photoluminescence intensities of NPs with complexes in the solution at the same entity concentration.¹²⁷ Proper quantifications of the number of complexes into a NP can be realized by comparing the luminescence intensity of released complexes after NP dissociation with Ln-complex standards,¹²⁸ or by Ln quantification by Inductively Coupled Plasma Atomic Emission Spectroscopy of the remaining supernatant or after NP digestion.¹²⁹

In this part of the review, we will introduce the different examples of Ln-complex doped NPs described in the literature over the years. The photosensitizing ligands encountered in the literature are reported in Fig. 8. The main reports are related to encapsulation in either polymer or silica NPs. The photoluminescence properties of the various NPs (when studied) are summarized at the end of this section in Table 2.

4.1. Polymer NPs

Polymeric NPs are a well-suited matrix for the encapsulation of Ln complexes. Extensively studied as biomaterials for biomedical applications^{130,131} in particular for drug delivery¹³² because of their remarkable stability in biological environment and their biocompatibility, they are also used as matrix for dye-encapsulated NPs.¹³³ The techniques of preparation of the dye-loaded polymer NPs are beyond the scope of this review but are well addressed in other publications.^{133,134} Very briefly, they can be obtained by either polymerization of monomer units (such as emulsion, micro-emulsion...) or by preparation from preformed polymers (e.g. self-assembly, nano-precipitation), in the presence of the dye to be encapsulated.

The first attempt of Tb-chelate and Eu-chelate doped polymer NPs synthesis in the literature was reported by Tamaki *et al.* in 2002.¹³⁵ Sub-micron PEG-coated polystyrene (PSt) particles (diameter of *circa* 200 nm) were encapsulating β -diketonate (**44** or **45**) ternary Tb³⁺ or Eu³⁺ complexes coordinated to an acryl derivative of phenanthroline (**46** or **47**). The acryl function enabled to covalently bind the complexes to the host matrix thanks to its involvement in the copolymerization process and to avoid a possible leakage from the particles over time. At the same period, [Eu(**48**)₃] doped PSt NPs were also commercialized and have been extensively used by Härmä and colleagues over the years for improving bioanalytical methods (immunoassays,^{136–139} cell imaging,¹⁴⁰ protein quantification¹⁴¹ and protein aggregation^{142,143}). The 107 nm NPs in particular were found to contain 31 000 chelates and to exhibit an Eu-based luminescence emission lifetime of *circa* 720 μ s.¹²⁸ Härmä's group was also the first to describe the synthesis and characterization of Sm(III) and Dy(III) chelates doped polymer based NPs with respectively β -diketonate derivative **49** and dipicolinic acid derivatives (**51**, **52** or **53**) as photosensitizing ligands, as well as their Eu(III) and Tb(III) counterparts.¹⁴⁴ NPs of around 45 nm were obtained, exhibiting photoluminescence emission in the visible (from green to red depending on the Ln ion) with lifetimes of 618, 695 and 89 μ s

for Eu, Tb and Sm doped polymeric NPs respectively, Dy-related lifetime being too short to be detected here. Their potential application as fluorescent label was highlighted with a sandwich immunoassay after surface functionalization by streptavidin.

Tamaki *et al.* studied the impact of the use of PSt and PMMA as polymer matrix for encapsulation of [Eu(**16**)₃(**18**)] and [Ln(**25**)₃(**18**)] (Ln = Eu, Tb, Sm) complexes by precipitation.¹⁴⁵ Only [Ln(**25**)₃(**18**)] with PSt matrix resulted in relatively uniform and luminescent NPs, probably thanks to the interaction between the phenyl moieties of **25** and styrene, but even there, very large particles in the 1–2 μ m range were obtained. This highlights the complexity of the NP preparation and complex encapsulation steps for preparing luminescent NPs with size uniformity in the nm range.

Yuan Wang and colleagues designed poly(methylmethacrylate-*co*-methacrylic acid) NPs embedding *circa* 3500 [Eu(**16**)₃(**54**)] complexes per NP, for application as cell imaging probe.¹⁴⁶ 52 nm water-dispersible nanospheres were obtained by co-precipitation, exhibiting two-photon-sensitized luminescence properties: Eu ion emits red light through two-photon excitation of **54** at 832 nm *via* a singlet energy-transfer pathway predominantly. With high two-photon excitation action cross sections ($\delta \times \Phi$ estimated at 1.2×10^5 GM), they are promising probes for two-photon excitation microscopy, as demonstrated by the NP bioconjugation and use as bionanoprobe for imaging of live cancer cells.

In 2012, Desbiens *et al.* demonstrated that the doping level of Ln-complex in the PSt NPs has a strong influence on the monomer conversion degree (conversion of monomeric carbon-carbon double bonds into polymeric carbon-carbon single bonds) and the Ln content in the NPs.¹²⁹ In this study, PSt NPs were synthesized by mini-emulsion polymerization in the presence of various concentrations of [Eu(**16**)₃(**18**)] complex (from 2 to 7 wt% relatively to styrene) and the results indicated that only a maximal doping level of 2% by weight in final particles could be achieved. At higher doping levels, neither the monomer conversion degree nor the final Eu content in the NPs are reproducible anymore, probably due to the limited solubility of the complex in styrene. This assumption has been supported by the study of Aikawa *et al.*,¹⁴⁷ where two complexes [Eu(**16**)₃(TOPO)₂] (TOPO = Tri-Octyl Phosphine Oxide, O=P(*n*-octyl)₃) and [Eu(**16**)₃(H₂O)₂], were incorporated into PSt NPs. The styrene conversion was remarkably reduced with the latter complex compared with the former one, as it was not dissolved in styrene. Thanks to the high solubility of [Eu(**16**)₃(TOPO)₂] in styrene, a maximal loading ratio of 15 wt% Eu-chelate was achieved, but the NPs started to aggregate at upper Eu content.

At the same period, new types of lanthanide doped polymer based NPs have emerged. Tan *et al.* described the use of lanthanide coordination polymer nanoparticle composed of adenine and dipicolinic acid (**5**) coordinated to Tb(III), as probe for Hg²⁺ detection.¹⁴⁸ Photoinduced energy transfer between adenine and **5** caused by hydrogen bonds between the two prevents Tb photosensitization by **5**, but the presence



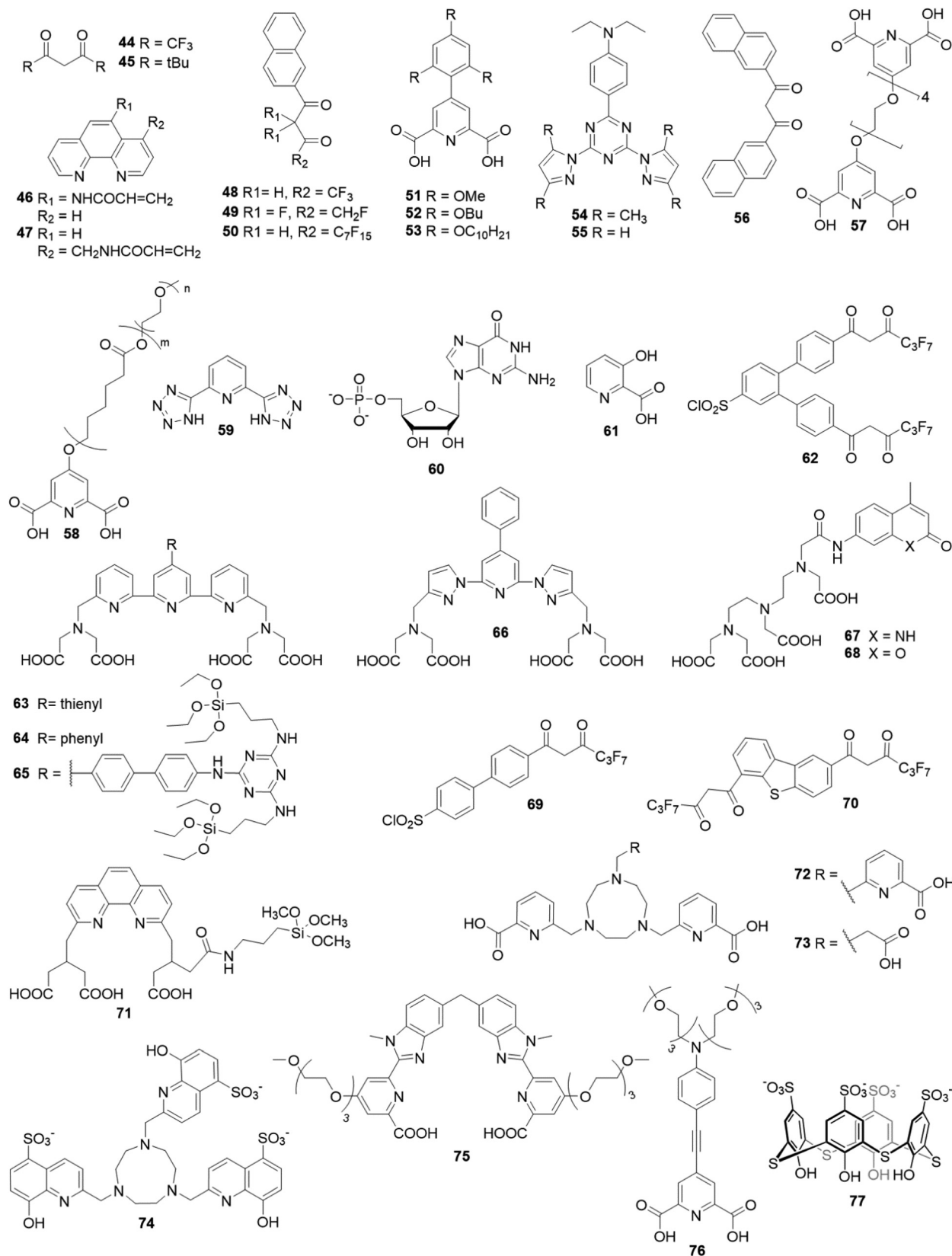


Fig. 8 Photosensitizing ligands present in the Ln complexes encapsulated in the NPs.



Table 2 Comparison of the photoluminescence properties of the Ln-complex encapsulated NPs

Matrix	Ln	Antenna	Complexes per NP	Diameter (nm)	ϵ/B (L mol ⁻¹ cm ⁻¹)	QY (%)	Lifetime (μ s)	λ excitation (nm)	Ref.
Polymer	Eu	48		107			720		128
	Eu	49; 53	1400	45			618	340	144
	Tb		1000				695	320	
	Sm		250				89	341	
	Eu	16; 54	3500	52	$\epsilon = 6.5 \times 10^4$ $B = 7.0 \times 10^7$	31	586	412 (2PE @ 832)	146
	Eu	16; 18	2 wt%	52 to 94			720	343	129
	Eu	18; 50	60 wt%	15		31.5	509	342	149
	Eu	59	2 wt%	376 ^a		60	2920	300	154
	Eu	16; 18	40 wt% (5000)	34	$B = 4 \times 10^7$	26	1300	350	155
	Eu	16; 55	156	15		14	756	410	157
Silica	Eu	62		37 \pm 3			384	336	170
	Eu	63		50 \pm 5		1.1	770	336	172
	Tb	66	~1500	42 \pm 3			1520	320	127
	Tb	66		45 \pm 3		10	2000	324	173
	Tb	67		50 \pm 3			1500	328	175
	Eu	68		55 \pm 3			600	328	176
	Eu	53; 68		10		21	397	335	177
	Eu	53; 61		36 \pm 3		66	390	406	178
	Eu ₂	54; 69		42 \pm 3		31	346	345–390	179
	Tb	71		40 \pm 5			1950	280	180
	Eu						1200		
	Eu	61		45 \pm 5			450	380	181
	Tb	72		50			2060	279	182
	Eu						1230		
	Yb	74				0.11	4.17	375	
	Nd–Yb	72; 74				0.095 (Yb) 0.018 (Nd)	0.058 (Nd)	272	
						0.13 (Yb)	4.01 (Yb)	375	
	Tb	64		56 \pm 4			1200	335	183
	Eu						1280		
	Eu ₂	75	2380	90 \pm 10		28	2400	330	185
	Eu	56	1000	10	$\epsilon = 5 \times 10^7$ $B = 3 \times 10^6$	6	243	350	196
	Eu	56	2 wt%	20–30			250	400	186
							(25 °C)		
	Eu	76	5 wt%	70 \pm 20 ^a			750 \pm 20	370 (2PE@740)	187
	Tb	77		22 \pm 3		3		330	191
	Tb–Yb	77		59 \pm 4			1116	330	192
	Tb–Gd						1247		
Zirconia	Tb	66		33 \pm 4		8.9	2000	324	174

^a Hydrodynamic diameter measured by DLS. 2PE = two-photon excitation.

of mercuric ion leads to an enhancement of Tb luminescence, originating from the coordination of the mercuric cation to adenine. The detection has been shown to be specific to Hg (no luminescence enhancement for other metals ions) with a detection limit of 0.2 nM.

Chiu and colleagues designed Eu-doped polymer dots with a matrix able to photosensitize the Eu-chelate *via* FRET.¹⁴⁹ High amounts (up to 80 wt%) of two different Europium-(β -diketonate) complexes ([Eu(**50**)₃(**18**)] and [Eu(**56**)₃(TOPO)₂]) were embedded in a fluorescent poly(9-vinylcarbazole) (PVK) matrix, giving rise to Eu-related red emission luminescence through excitation of PVK at 342 nm. A maximal QY of 33.5% was obtained with 60 wt% [Eu(**50**)₃(**18**)], the latter decreasing for higher doping ratios, possibly due to concentration quenching. The polymer dots were employed as probe for living cell imaging. They also proposed the preparation of

polymer dots containing [Eu(**16**)₃(**18**)] complex covalently linked to the matrix, with the aim of avoiding complex leakage.¹⁵⁰

Complex coacervate core micelles (nanometric colloidal complexes produced by the co-assembly of ionic-neutral block copolymers with oppositely charged species¹⁵¹) containing Eu and Gd ions were proposed by Wang *et al.* for potential applications both in luminescence spectroscopy and magnetic relaxation measurements.¹⁵² Both Ln ions are chelated by a dipicolinate derivative (**57**), an anionic coordination polymer that electrostatically interacts with a cationic neutral diblock polymer to form micelles in aqueous solution. Micelle-like NPs were also proposed by Thévenaz *et al.* by coordinating Eu (III) or Tb(III) with a dipicolinate moiety functionalized with diblock copolymers based on poly(ethylene glycol) (PEG) and poly(ϵ -caprolactone) (PCL) segments (**58**).¹⁵³ Solvent displace-



ment allowed to form green or red emitting NPs composed of a PCL core (solid sphere < 47 nm in diameter) containing Ln-complexes and a PEG corona (vesicle >47 nm).

In 2013, Wartenberg *et al.* reported the use of pyridine-bis-tetrazolate ligand (**59**) as sole photosensitizer of Tb, Eu, Dy and Sm in Vis-NIR luminescent polymeric NPs containing the four $[\text{Ln}(\mathbf{59})_3]^{3-}$ complexes.¹⁵⁴ The authors highlighted the importance of using a cationic surfactant to incorporate efficiently the anionic complexes into PMMA matrix and obtain luminescent NPs. Once again, the doping level is a limiting parameter, as a maximum of 2 wt% was reached without destabilizing the NPs, similar to the doping level of $[\text{Eu}(\mathbf{16})_3(\mathbf{18})]$ in Pst NPs seen previously.¹²⁹ 6 years later, Cardoso Dos Santos *et al.* have overcome this issue as they succeeded to encapsulate up to 40 wt% of $[\text{Eu}(\mathbf{16})_3(\mathbf{18})]$ complex into NPs (corresponding to 5000 complexes per NP) composed of PMMA copolymers bearing sulfonate (PMMA-SO₃H; 1 or 3 mol%) or carboxylate (PMMA-COOH; 10 mol%) groups (Fig. 9).¹⁵⁵ Two emission lifetimes were measured, a short one (around 0.5 ms, similar to the free complex in acetonitrile) attributed to the complex at the surface of the NP and a long one (up to 1.3 ms) attributed to the complexes protected in the core of the NP, the lifetime exceeding that of Eu complex in solid state (see Fig. 9a for decay profiles). With a quantum yield up to 26% (Fig. 9b) and a remarkable brightness exceeding $10^7 \text{ L mol}^{-1} \text{ cm}^{-1}$, the authors have reported a very promising candidate for single-particle and live-cell imaging.

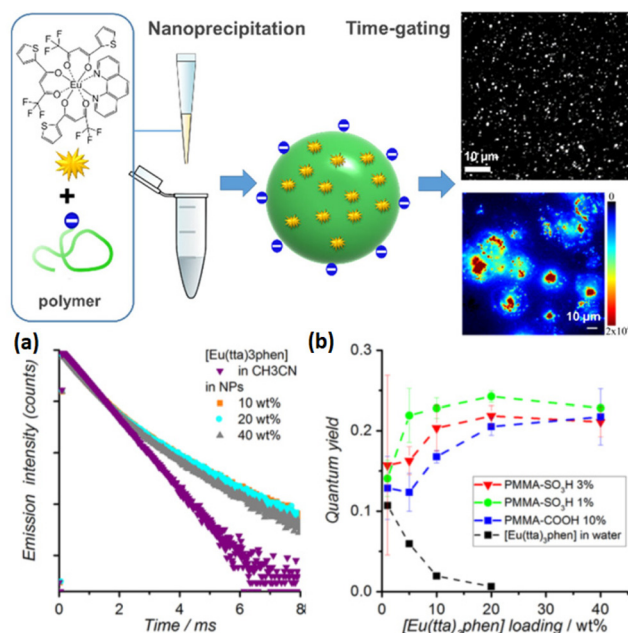


Fig. 9 Top: schematic representation of the preparation of Eu-complex doped polymer NPs for single particle and live-cell imaging. Bottom: (a) time-resolved emission decay profiles of Eu ($\lambda_{\text{exc}} = 350 \text{ nm}$, $\lambda_{\text{em}} = 620 \text{ nm}$) measured for solutions of $[\text{Eu}(\mathbf{16})_3(\mathbf{18})]$ in acetonitrile and of PMMA-COOH 10% NPs loaded with different doping rates of $[\text{Eu}(\mathbf{16})_3(\mathbf{18})]$ and (b) photoluminescence quantum yield as a function of Eu-chelate doping level. Adapted with permission from ref. 155. Copyright (2019) American Chemical Society.

Various analytical and bioanalytical applications of Ln-complex doped polymeric NPs have been made in the last years. A ratiometric nanothermometer for intracellular temperature measurement in real time was reported by Takei *et al.*¹⁵⁶ Thermosensitive $[\text{Eu}(\mathbf{16})_3]$ complex and thermoinsensitive rhodamine 101 were incorporated into PMMA NPs, and the ratio of both emission intensities is related to the temperature of the NP environment. Wang *et al.* developed another two-photon excitation polymeric nanoprobe encapsulating $[\text{Eu}(\mathbf{16})_3(\mathbf{55})]$ complex¹⁵⁷ very similar to the previous one ($[\text{Eu}(\mathbf{16})_3(\mathbf{54})]$)¹⁴⁶ with a lower two-photon excitation cross section (estimated at $3.7 \times 10^3 \text{ GM}$ for excitation at 800 nm). Functionalized with a tumor-targeting agent, the NPs have been used as nanocarrier and luminescent probe for imaging. Gao *et al.* designed a coordination polymer NP composed of Tb and Eu ions with **60** as Tb photosensitizer, for the ratiometric detection of the anthrax biomarker dipicolinic acid (DPA, **5**).¹⁵⁸ The presence of DPA induces both an enhancement of Tb and a quenching of Eu emissions by coordinating and photosensitizing Tb ions in addition to preventing Tb to Eu energy transfer. Finally, immunochromatographic assays have also been developed with $[\text{Eu}(\mathbf{17})_3(\mathbf{18})]$ doped PSt NPs for detection of procalcitonin,¹⁵⁹ an indicator of bloodstream infections and sepsis, and even more recently for the trendy detection of anti-SARS-CoV-2 IgG.¹⁶⁰

4.2. Silica NPs

Silica NPs are commonly employed as biomaterial because of their water solubility, high stability, limited toxicity and good biodegradability. They are also easily produced with a tunable size and their surface functionalization is simple and versatile.¹⁶¹ Besides their application as nanocarriers for drug delivery,^{161,162} silica NPs have also emerged as a matrix of choice for dye encapsulation as it is transparent to visible light and not involved in energy/electron transfer processes.¹⁶³ The preparation of dye-doped silica NPs is well described in the literature^{164–166} and will not be addressed here. Dye-Doped silica NPs have been reported as luminescent probe for numerous applications (chemosensor, bioprobe in imaging or immunoassay, (bio)marker quantification...),^{163,165} but one major limitation of these short-lived emitting nanoprobes is the media autofluorescence that generates a high background signal and thus decreases the method sensitivity. To overcome this issue, long-lived emission of Ln-complex is a very promising alternative.

The first attempt of Eu-chelate doped silica NPs synthesis have been proposed by Trindade and colleagues in 2003.¹⁶⁷ SiO₂ NPs doped with complexes of Eu³⁺ ion coordinated by three 3-hydroxypicolinic acids (**61**) were prepared by a sol-gel method adapted from the Stöber process,¹⁶⁸ leading to quite large NPs of 130 nm in diameter with an Eu emission lifetime of 0.5 ms. However, the experimental results strongly suggest that the Eu³⁺ complexes would be located at the surface of the NP, Eu³⁺ being also coordinated by silanol groups. Using a similar method based on Stöber process, Zhao *et al.* succeeded to encapsulate $[\text{Eu}(\mathbf{17})_3(\mathbf{18})]$ complexes into Silica particles, but

here again, with a very large size (300 nm).¹⁶⁹ These studies highlight the difficulty of efficiently and homogeneously encapsulating complexes into NPs.

Jingli Yuan's group designed 20 to 50 nm silica NPs doped with various chelates (Eu³⁺ with **62**^{170,171} or **63**,¹⁷² Tb with **66**^{127,173}) by water-in-oil microemulsion. Encapsulation was realized either with free Ln-complex^{127,171–173} or by copolymerization of Ln-complex bound to 3-aminopropyl(triethoxy)silane (APTES).¹⁷⁰ With the latter, Ln complexes are covalently bound to silicon atom, thus more protected from dye leaking and less sensitive to the different handling steps (washing, surface modification, bioconjugation...). The encapsulation of Ln-complex is beneficial as less photobleaching is observed, however emission lifetimes and quantum yields are much lower than those of the free complexes once encapsulated in the NPs. These results could originate from the exciting light absorbance by the matrix and/or from a concentration quenching due to a high concentration of Ln-complexes in the NPs. These NPs, functionalized with amino groups, were then conjugated with a biomarker (streptavidin or antibody) prior to be used as luminescent probes in fluoroimmunoassays.

Encapsulation of Tb chelated by **66** in zirconia NPs as an alternative to silica NPs has also been proposed in parallel.¹⁷⁴ Doped zirconia NPs have been shown to be more stable than their silica counterpart¹²⁷ in high pH aqueous media and to have a slightly longer Tb emission lifetime (2.0 vs. 1.5 ms), however the limit of detection (LOD) of the immunoassay realized with ZrO₂ NPs is much higher. As the NP conjugation is non-covalent (Lewis acid–base conjugation), dissociation probably occurs at low concentrations or during washing steps, explaining the high LOD compared with covalently-conjugated silica NPs.

Chen *et al.* proposed the synthesis of silica NPs embedding Tb³⁺ coordinated to a polyaminocarboxylate derivative with a Carbostyryl **124** as photosensitizer [Tb(**67**)]¹⁷⁵ and its Eu counterpart [Eu(**68**)] with a coumarin **120**.¹⁷⁶ These 50 nm large NPs exhibit emission lifetime of 1.5 ms and 0.6 ms for Tb and Eu respectively, upon excitation of the antenna ($\lambda_{\text{exc}} = 328$ nm), similar to the ones in free complexes. Each NP is reported to be as bright as about 340 free Tb complexes and 1300 free Eu complexes respectively, highlighting a potential 100 to 1000-fold increase of sensitivity for biomarker detection and quantification. DNA sandwich hybridization assays were successfully realized with the detection-oligonucleotide labelled luminescent NPs and capture-oligonucleotide linked to a magnetic bead to detect and quantify the targeted DNA, with a detection 100-fold (for Tb) and 50-fold (for Eu) more sensitive than the Fluorescein isothiocyanate (FITC) technique.

From 2008, Jingli Yuan's group focused on the design of red-emitting Eu-doped silica NPs with excitation in UV-Vis range (from 200 to 450 nm) with [Eu(**54**)–(**69**)₃] complex¹⁷⁷ and excitation in visible light (406 nm) with [Eu(**54**)–(**62**)] complex.¹⁷⁸ Both **62** and **69** were linked to APTES to be covalently bound to the matrix. The best properties were obtained with [Eu(**54**)–(**62**)] complex, the related NPs exhibiting a Eu emission lifetime of 0.39 ms and a QY of 66%. Other

UV-Vis excited NPs were proposed with a dinuclear [Eu₂(**70**)₃(**55**)₂] complex (excitation up to 475 nm) with a similar lifetime but a QY of 31%.¹⁷⁹ These three different NPs were utilized as luminescent probe for the detection of environmental pathogens by time-gated luminescence imaging.

In the late 2000s, several studies also emerged with NPs containing two different Ln ions. The first report was made by Zhang *et al.* with the preparation of NPs encapsulating different ratios of Tb/Eu ions both chelated and photosensitized by (**71**) and exhibiting longer emission lifetime than the related free complexes (1.2 ms vs. 0.42 ms for Eu, 1.95 ms vs. 0.64 ms for Tb).¹⁸⁰ Six years after their first attempt,¹⁶⁷ Trindade and colleagues proposed a different strategy of [Ln(**61**)₃] complexes (Ln = Tb, Eu or Tb-Eu) encapsulation based on the formation of a core of complexes and a shell of amorphous silica as a protective layer, by reverse microemulsion.¹⁸¹ This new method is more adapted, as the photoluminescence results confirmed the presence of Ln-complexes in the NPs and the size of NPs is more suitable for bioassays (45 nm vs. 127 nm previously¹⁶⁷). In 2010, Samuel *et al.* proposed two types of dual-mode lanthanide doped luminescent silica NPs with a tunable size between 10 and 100 nm.¹⁸² Visible and NIR emitting nanospheres containing Tb(**72**)–Eu(**72**) and Nd(**72**)–Yb(**74**) respectively exhibited emission lifetimes similar to their corresponding free complexes and the emission of the NIR emitting NPs could be tuned by varying the excitation wavelength, suggesting a potential application as multiluminescent bar codes. Jiang *et al.* designed NPs with emission colors from green to red by co-binding different molar ratios of Eu(**64**) and Tb(**64**).¹⁸³ Wartenberg *et al.* designed their own Tb–Eu co-doped silica NPs with **72** or **73** as antennas,¹⁸⁴ and proposed a new method to study the efficiency of the incorporation process by using ¹⁵²Eu chelates for radioactivity measurements.

The effect of Ln-complex encapsulation on the ligand triplet-state energy level was questioned by Eliseeva *et al.*, and [Eu₂(**75**)₃] binuclear helicates incorporation into silica NPs was shown to have no impact on it.¹⁸⁵

The beginning of the 2010s' was also marked by the breakthrough of new kind of probes in this field. To the best of our knowledge, the first thermometer composed of Ln-complex encapsulated in silica matrix was presented by Wolfbeis and colleagues in 2010. They first tried to encapsulate a high content of [Eu(**56**)₃(TOPO)₂] complex into 2-bis(trimethoxysilyl) decane but the resulting luminescence intensity was reduced by concentration quenching.¹⁸⁶ To overcome this issue, an hybrid NP was prepared by using PMMA polymeric material as co-matrix to compensate the decrease of Eu complex concentration. This hybrid nanoprobe was employed for sensing and imaging of physiological temperatures (from 25 to 45 °C with a precision of ±0.3 °C) thanks to the dependence of both photoluminescence intensity and emission lifetime to temperature upon visible–light excitation.¹⁸⁶ One year later, Philippot *et al.* presented the first Ln-complex doped silica NPs with two-photon excitation.¹⁸⁷ The encapsulation of [Eu(**76**)₃]^{3–} complex was beneficial as the NPs are luminescent



even in aqueous solution whereas the free complex is not, because of ligand substitution by water molecules. The 5 wt% doped NPs exhibited typical Eu red emission upon excitation into 76 at 370 nm, and they are also promising probes for two-photon microscopy imaging thanks to the two-photon absorption ability of 76 in the NIR (740 nm).

The last five years have been marked by a decrease of the number of publications compared with the beginning of the century. The published studies are more focused on the analytical/bioanalytical applications, such as the detection of Cardiac Troponin I by [Eu(65)] doped NPs¹⁸⁹ or anthrax biomarker (dipicolinic acid (5), named DPA) detection by a time-resolved ratiometric fluorescent probe.¹⁸⁸ In this study, the surface of silica NPs encapsulating Tb(III) ions coordinated to DPA (Tb/DPA@SiO₂ in Fig. 10a) was functionalized with a guanosine monophosphate bound to Eu(III) (Eu/GMP). In the presence of DPA in solution, Eu ions are chelated by the latter through the replacement of a water molecule in the Eu/GMP complex. Upon DPA excitation, both Tb(III) and Eu(III) ions are dye-sensitized, thus emitting green and red light respectively. The green emitting light related to Tb emission serves as a stable reference, and the DPA in solution is detected and quantified by measuring the ratio of Eu over Tb emission intensities as shown by the emission spectra for various DPA concentrations (Fig. 10b).

This time-resolved ratiometric fluorescent assay has revealed a good selectivity and sensitivity (LOD of 7.3 nM)

towards DPA detection in biological samples. Moreover, a paper-based visual sensor (Fig. 10c) was developed for fast detection by naked eyes or with a smartphone under a UV lamp, but with a low sensitivity that has to be improved (LOD of 1 μM). A similar method was also developed for mercury detection.¹⁹⁰

Finally, in 2019, *p*-sulfonatothiocalix[4]arene (77) was used as (i) coordinating photosensitizer of Tb ion in green emitting doped silica NPs for immunoassay application¹⁹¹ and as (ii) photosensitizer of Tb ion and chelator of Yb ion in dual visible-NIR luminescent silica NPs.¹⁹² In the case of Tb–Yb doped NPs, besides the Tb-related visible emission, a Tb(III) → Yb(III) energy transfer occurs and leads to NIR emission through UV excitation of 77 (320 nm). The internalization into cells of these dual visible–NIR emitting NPs highlighted their possible use as probes in cellular imaging.

To sum up, encapsulation of Ln-complexes into NPs is a good strategy to cumulate the spectroscopic performances of the luminescent complexes and protect them from quenching. However, to control the size of NPs and to obtain an appropriate loaded amount of complex in the NPs without destabilizing the polymerization reaction and complex leakage is a real challenge, as highlighted by the literature. The question of the quantity of Ln-complex to be encapsulated to have the brightest NPs has been studied for years, and a subtle equilibrium has to be respected to encapsulate the highest number of complexes to cumulate their properties without suffering from concentration quenching. The use of bulky groups or bulky counterions is also a good strategy to control the organization of complexes in the NPs and decrease the risk of concentration quenching.¹³³ Complex leakage over time can also be avoided by using complexes linked to a monomeric unit to be covalently linked to the matrix.

Finally, these NPs are well suited for (bio)analytical applications as (i) the host matrix (polymer or silica) is biocompatible and (ii) reactive groups (such as amino or carboxylate groups) can be introduced at the surface of NPs during their synthesis or with an extra polymerization step, allowing for (bio)functionalization.

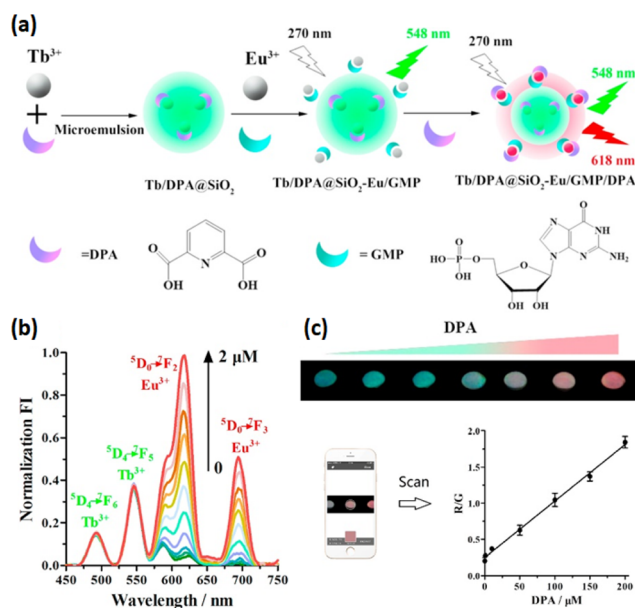


Fig. 10 Anthrax biomarker (DPA) detection via a dual-lanthanide based silica NPs. (a) Representation of the luminescent probe. (b) Emission spectra of Tb/DPA@SiO₂-Eu/GMP probe to different concentrations of DPA upon DPA excitation. (c) Fluorescence color image of the designed paper-based visual sensor over various concentrations of DPA solution under 254 nm UV light excitation and standard curve obtained with a smartphone. Reprinted from ref. 188, Copyright (2017), with permission from Elsevier.

5. NPs surface functionalized by Ln-complexes

The functionalization of NPs with lanthanide complexes gained interest over the years. The combination of their properties led to new systems very promising for applications such as medical imaging or luminescent probes. In this part of the review, we will mainly overview the different systems presented in the literature. The photosensitizing ligands used to elaborate those systems are illustrated in Fig. 11 and 15.

5.1. Functionalized quantum dots

Quantum dots (QDs) are used for almost three decades for their size dependent optical properties with high absorption cross sections¹⁹³ and narrow emission bands.^{194,195}



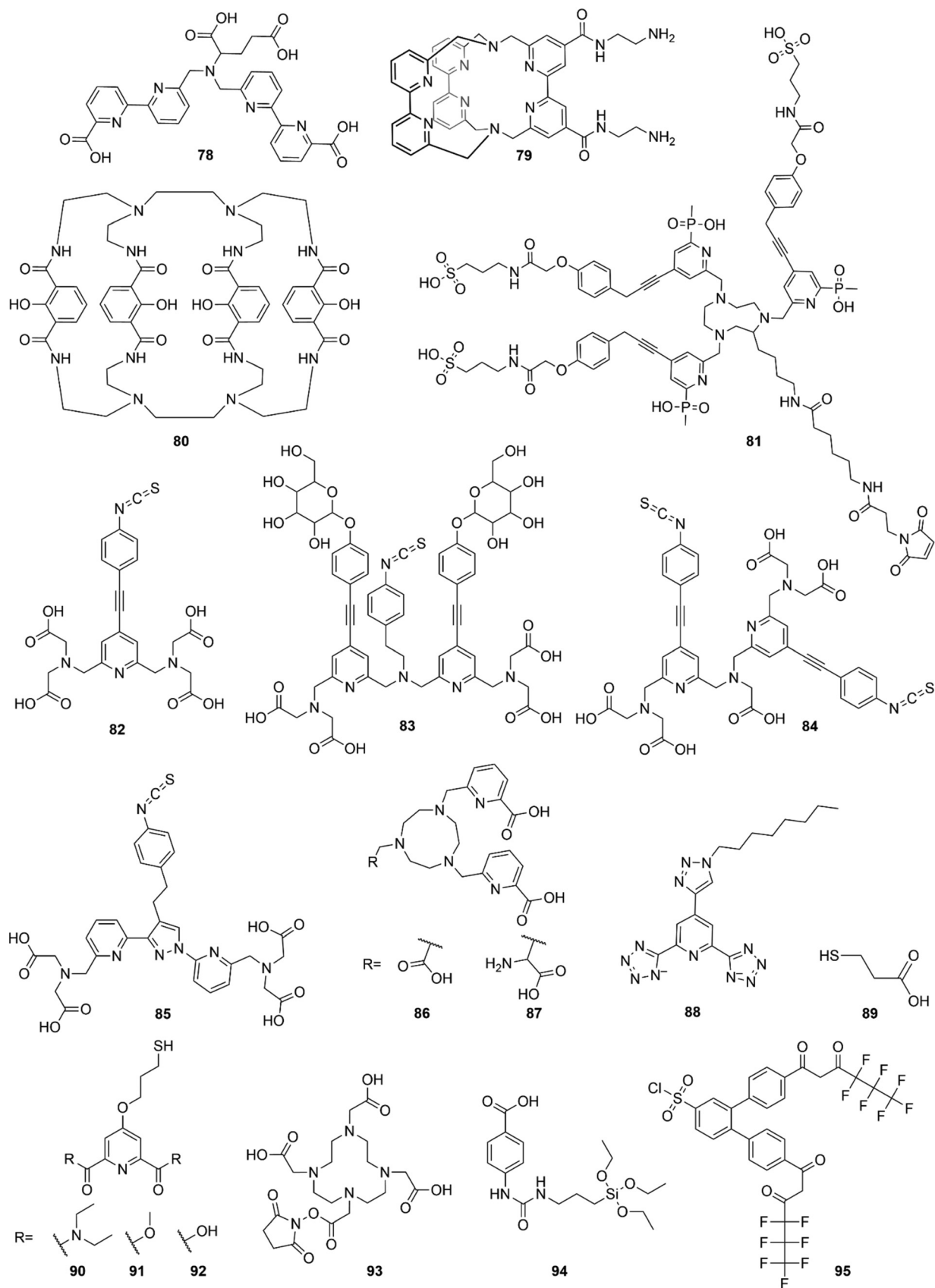


Fig. 11 Photosensitizing ligands used to form the Ln complexes and functionalize the NPs, part 1.



Table 3 Comparison of the main properties of the coated QDs

Ln	Antenna	Number of coated complexes	Lifetime ^a (μs)	FRET Distance (nm)	FRET efficiency (%)	λ _{exc} (nm)	Ref.
Tb	78	5 to 7	170 and 560	7.2–7.6	63	308	203
Tb	78	6		8.4		315	204
Tb	78	5.6		10	up to 70	308	205
Eu	78; 79			9.6			
Tb	80	6		5 to 11		340	209
Tb	80			6 to 10		339	208
Tb	80			10.1 and 7.5		340 and 400	213
Tb	80			6.3 to 8.1	up to 94	339 and 400	214
Tb	80		70 to 170	6 to 7.5	up to 97	308 and 340	211
Tb	80			5.5 to 11.5			212
Tb	80			5.4 to 11.1		340	217
Tb	80			10		337	218
Tb	80			14		340	216
Tb	80		740 and 1820	10.3 and 12.2		330–340	43
Eu	81		610 and 1090	9.2 and 11.1			
Tb	80			10		337	219
Tb	80			1 to 10			215
Tb	86	75–80				275–320	223
Eu							
Eu	89; 90; 91; 92		Up to 1430			267–305	225
Tb			Up to 1470				
Eu, Tb, Yb	93		0.005 to 0.009			220 to 440	226

^a Ln³⁺ excited-state lifetimes after functionalization of the QDs.

QDs offer an improvement in term of stability toward photobleaching in comparison to organic fluorophores^{197–199} and can be used as an energy donor with dyes but also interact with luminescent donors as fluorescent proteins,²⁰⁰ luminescent polymers,²⁰¹ or lanthanide complexes in the case of the FRET process. Despite those positive points, the FRET process is particularly limited by donor–acceptor distance as its efficiency depends on it.²⁰² This part of the review will be mainly focused on FRET with QDs as acceptors and lanthanides as donors. The photoluminescence properties of those systems are summarized in Table 3.

Löhmansröben and Hildebrandt firstly worked on this subject by introducing a system composed of CdSe/ZnS QDs as acceptor functionalized with [Tb(78)] chelates as donor into a FRET process based on a Time-Resolved Fluoroimmunoassay format (TR-FIA).²⁰³ This assay relies on biotin–streptavidin recognition process where Tb-chelate labelled streptavidin interacts with biotins coated at the surface of the QDs. The use of a donor with a long-lived excited state such as Tb (1.5 ms) is a great advantage for energy transfer to the QD, leading to a FRET efficiency of 63 ± 4%, and allowed the use of time-resolved acquisition thanks to the increased decay time of the QDs (few hundreds of μs) enhancing the sensitivity of the detection procedure. These results were confirmed in a more detailed analysis of the decay time of the FRET process.²⁰⁴ Eu complexes were also successfully employed as energy donor in FRET experiments with QDs, however the use of [Tb(78)] chelates showed better results than [Eu(78)] and [Eu(79)] complexes, with a LOD under 2 pM and generally a better FRET efficiency (up to 70%).²⁰⁵

Those pioneer reports illustrating the Ln-complex functionalization of QDs and the related FRET process have paved the way for many researchers, in particular Hildebrandt

and his collaborators whose work has largely contributed to the scientific progress of the field. Various studies were realized with a commercial [Tb(80)] complex (named Lumi4-Tb) labelled with streptavidin and biotin tagged QDs for the development of new ultrasensitive assays with multiplexed detection. This method is based on the FRET interaction between different QDs emitting at different wavelengths and [Tb(80)] present in the same sample. Their system is expected to contribute to applications in biology and medicine for example, as it provides a LOD in the sub-picomolar level.²⁰⁶

A new assay for multiplexed ultrasensitive detection allowing the measurement of manifold clinical parameters simultaneously within one sample (Fig. 12, left panel) was reported,²⁰⁷ with a LOD lower than 1 pM, offering a better sensitivity than a well-established homogeneous immunoassay FRET pair (40 to 240-fold higher). A spectroscopic ruler²⁰⁸ was also designed, which permitted to correlate the efficiency of

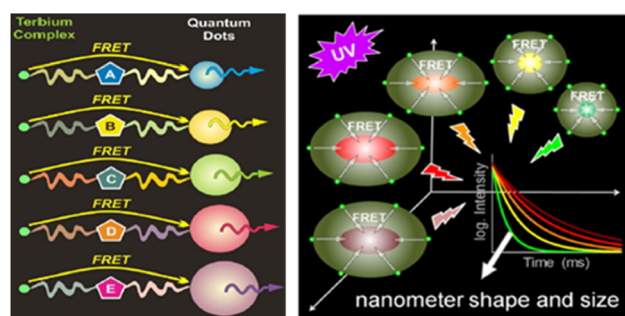


Fig. 12 Principle of multiplexed Tb-complex to QDs FRET applied for ultra-sensitive diagnostics (left) and spectroscopic ruler measurements (right). Reprinted with permission from ref. 207 and 208. Copyright (2010), Wiley-VCH Verlag GmbH & Co. KGaA.



the energy transfer with the donor–acceptor distance, and consequently to provide information about the size and shape of the QD (as illustrated in Fig. 12, right panel).²⁰² Such tool was realized by optimizing the energy transfer between QDs emitting at different wavelengths and [Tb(80)] complexes without the help of a FRET enhancer.^{209,210} A good energy transfer with distance over 10 nm and 11 nm were obtained without and with FRET enhancer respectively, making possible applications in life sciences and nanotechnology.²⁰⁸ The spectroscopic ruler measurement can also be set by others methods,^{211,212} such as QDs coated with [Tb(80)] functionalized peptides or maltose-binding proteins, allowing to increase the precision in shape and size determination compared to other analytical methods like TEM, DLS and HPLC.²¹²

In 2012, they demonstrated the ability of QDs to be, in the same system, both an acceptor and a donor for FRET relays (Fig. 13).²¹³ QDs were functionalized with [Tb(80)] and a dye with the help of peptides. In such a system, excited Tb-complex transfers the energy to the QD which then transfers it to the dye. This process has been used for protease activity and DNA hybridization by using trypsin and chymotrypsin labelled with the Tb-complex and the dye, respectively. Moreover, despite the presence of only one QD, this system was also shown to be employable for multiplexed protease sensing with time-gated FRET relay by conjugating the QDs with dye/Tb-complex labelled peptides (Fig. 13).²¹⁴ The system was studied with the use of trypsin which gave a very high FRET efficiency (around 94%).

Recently, Xu, Qiu and Hildebrandt combined nucleic acid amplification and NPs for microRNA (miRNA) biosensing.²¹⁵ [Tb(80)]-to-QD related FRET was combined with rolling circle amplification (RCA) for a specific miR-21 assay. A subpicomolar LOD, a multiplexing capability and FRET with donor–acceptor distances of about 1 to 10 nm were obtained. This work also highlighted that the development of such systems requires many modification and optimization steps and is more complex than what can be found in the literature.

In addition to the biotin–streptavidin recognition system, other methods for the QDs functionalization by Ln-complexes were investigated by Hildebrandt and co-workers.^{216–218} The team proposed a system where [Tb(80)] and a dye were linked to the QDs using DNA components, as shown in Fig. 14.²¹⁶ The dye was used as a bridge for energy transfer from the Tb-complex to the QDs, allowing to overcome the limitations of

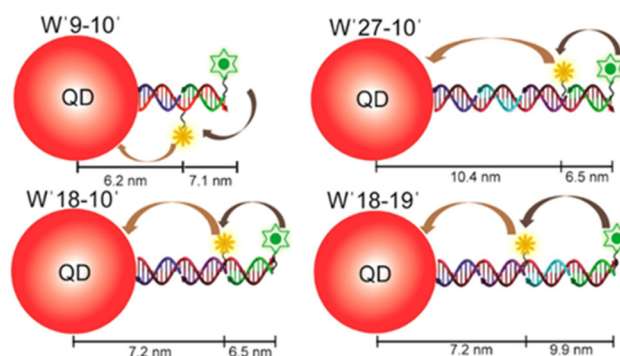


Fig. 14 Representation of the four [Tb-complex](green)–Dye(yellow)–QD-DNA systems with the different distances in between fluorophores. Reprinted with permission from ref. 216. Copyright (2017) American Chemical Society.

energy transfer due to the donor–acceptor distance and the long lifetime of the QDs, by reaching a FRET distance of *ca.* 14 nm but with a lower FRET efficiency (between 10 and 20%) compared to the previously mentioned systems.

A system using peptides functionalized with [Tb(80)] complex on one side and a hexahistidine tag on the other side was also proposed. They were assembled to a zinc-rich surface of dihydrolipoic acid capped QDs. Both this system and another based on biotin–streptavidin recognition were used to evaluate the influence of plasma on photophysical properties by comparing them in buffer and plasma.²¹⁷ Biotin–streptavidin binding system was shown to be the most suitable for clinical assays as it reached a low picomolar range of LOD when zinc–histidine interaction related LOD was 100 times higher. A few years later, they proposed another system made of QDs with a SiO₂ shell of different thickness (6 and 12 nm) on which two maleimide functionalized [Tb(80)] and [Eu(81)] complexes were attached forming one single nanoparticle,⁴³ allowing for the detection of different cells in a single view thanks to four specific photoluminescence decays given by Tb and Eu complexes. Thanks to a Red/Green/Blue (RGB) coding by the NPs, the living cells can be labelled and distinguished individually in mixtures, and the coding range can be extended with the different QDs colors, increasing the possibilities of fluorescence encoding. More recently, they succeeded to accomplish sensitive quantification of biological targets and long-distance conformational analysis with a sub-nanometer resolution for DNA and peptides.²¹⁹ Meanwhile, the surface bio-functionalisation of the NPs was extended to antibodies and their fragments, to offer applications into the field of biological diagnosis applied to the detection of prostate specific antigens.^{220,221}

In parallel to the work realised by Hildebrandt and co-workers, different research groups reported the use of Ln-complex functionalized QDs. In 2007, Weller and co-workers worked on competitive binding assays with CdTe QDs coated with streptavidin and [Eu(82)], [Eu(83)], [Eu(84)] and [Tb(85)] complexes coupled to biotin or estradiol.²²² A few years later, Mazzanti and co-workers covalently attached [Tb

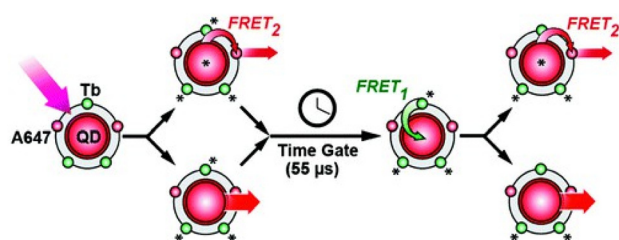


Fig. 13 Principle of the time-gated Tb to QD to Dye FRET relay. Reprinted from ref. 214. Copyright (2012) American Chemical Society.



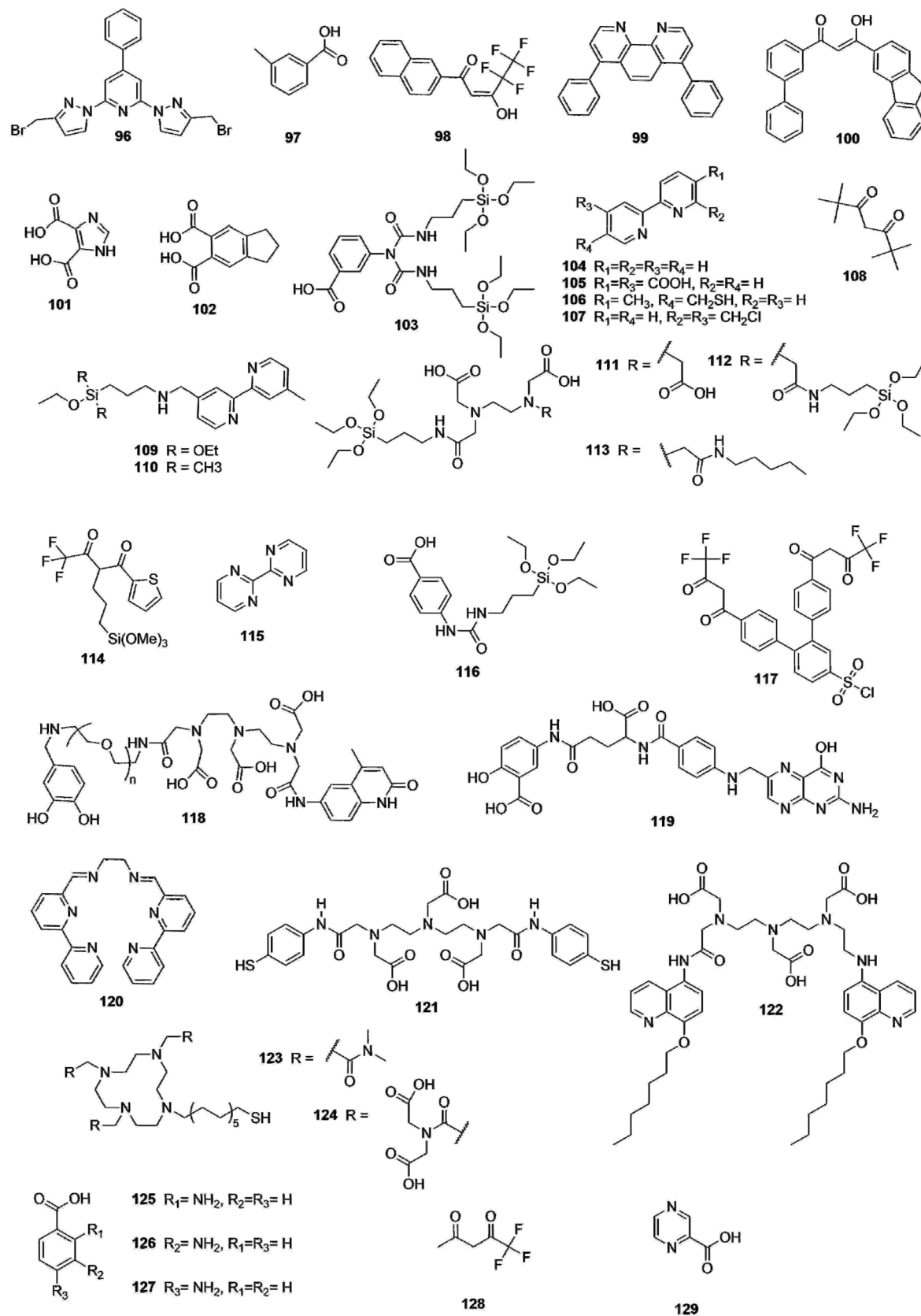


Fig. 15 Ligands used to form the Ln complexes and functionalize the NPs, part 2.

(86)] and [Eu(86)] complexes to QDs.²²³ Their main interest was the binding of a Gd-chelate for MRI probe but their study on luminescent Tb/Eu-complexes attached to QDs opened the possibility to produce a bimodal probe made with Tb/Eu complexes and the QDs for their luminescent aspect and Gd-chelates for their magnetic aspect. They also proposed a new series of bi-luminescent hybrid materials with **87** and **88** complexing Ln(III) (Ln = Eu, Tb, Gd and Yb).²²⁴ The attachment of the complexes was allowed by the functionalization of the ligands with thiol functions. In this case, the FRET process was made with the QDs emitting in a range of 525–540 nm as a donor and the Ln complexes as an acceptor. Bettencourt-Dias and co-workers aimed to synthesize surface-modified QDs by capping them with different ligands (**89**, **90**, **91** and **92**) complexing both Tb(III) and Eu(III).²²⁵ QDs were capped with the ligands and then mixed with the desired amount of Ln(III) in a one-pot synthesis method. Overall emission efficiencies, up to 79% and 36% for [Tb(**90**)] and [Eu(**90**)] respectively, are obtained making those frameworks suitable for imaging and sensing applications. Zhu and co-workers worked on the functionalization of Carbon Quantum Dots (CDots) with Ln complexes.²²⁶ CDots were chosen because they are free of heavy metals and display better biocompatibility and lower toxicity than QDs.²²⁷ The complexes were made with Ln(III) ions (Eu, Tb, Yb) and ligand **93** and were covalently linked to the CDots thanks to branched polyethylenimine forming an amine bond. This system has been characterized and could be used for multicolour imaging as they exhibited high fluorescence quantum yield (10 to 15%).

5.2. Functionalized silica NPs

As presented in section 4.2, the use of silica NPs is widely used for the encapsulation of Ln-complexes. However, it is also possible to directly coat the surface of the silica NPs with the Ln-complexes to combine the properties of the lanthanide and the nanoscale silica chemistry,^{228,229} as demonstrated by the unexpected silica NP functionalization by [Ln(**61**)₃] obtained by Trindade and colleagues in 2003 (instead of complex encapsulation, see Section 4.2 for more details).¹⁶⁷ To perform such systems, two methods are mainly used: (i) the organic modification of silica NPs surface using either amino, dipyridyl groups²²⁸ or β -diketonate²³⁰ ligands prior to Ln ion or chelate complexation and (ii) the grafting of organosilylated Ln(III) complexes on the surface of the silica NPs. The photoluminescence properties of the functionalized silica NPs are summarized in Table 4.

Using the former method, Wang and co-workers were the first to propose a facile strategy to design Ln-complex coated silica NPs,²³¹ one year after Trindade's report. The coating was firstly made with the ligand **94** and Tb(III) was added to form the complexes at the silica NPs surface. As compared to the free complex, the use of silica NPs increased the lifetime of the Tb(III), (1.3 to 2.2 ms) considering that the structure of the resulting nanocomposite is relatively rigid and limits the non-radiative transitions due to the vibrations of the ligands. This

Table 4 Comparison of the main properties of the silica NPs

Ln	Antenna	QY (%)	Lifetime ^a (ms)	Enhancement factor ^b	λ_{exc} (nm)	Ref.
Tb	94		2.2		306	231
Eu	95		0.398			232
Tb	96		2.224			
Eu	17		0.9 to 1.07		245 and 395	233
Tb	26		0.58 to 1.03		295 and 345	
Eu	97		0.31		324	234
Tb			0.76		290	
Eu	98; 99	43	1.05		385	235
Eu	18; 100	30	0.786		440	236
Eu	101		0.37 and 0.89		285	237
Tb	102		0.86 and 1.45		309	
Dy	18		0.017	3.4	313	238
	104		0.019	2.7	306	
Eu	103		0.57 and 0.7	2.02 and 2.14	293 and 294	239
Eu, Tb	16, 17		0.45 to 0.52			240
Eu	114		0.26		365	241
	16; 108		0.65			242

^a Ln³⁺ lifetimes after functionalization of the NPs. ^b Luminescence enhancement factor: comparison between the free Ln³⁺-complex intensity and the functionalized NPs intensity.

successful approach opened the gate to new systems combining silica NPs and Ln-complex.

In 2007, Ye Xu and Qingge Li covalently coated the surface of the silica NPs with [Eu(**95**)] and [Tb(**96**)] complexes in order to design a luminescent probe for a time-resolved immuno-fluorometric assay (TR-FIA) for detection of hepatitis B with a LOD 20- to 30-fold lower than ELISA.²³² Moreover, their Eu³⁺/Sm³⁺ dual-labelling related TR-FIA was shown to be less sensitive than the individual Eu(III) TR-FIA, due to the low fluorescence yield and short emission lifetime of Sm. The next year, Bing Yan and Bing Zhou designed another system where [Eu(**17**)] and [Tb(**26**)] complexes were covalently attached to mesoporous MCM-41 silica NPs.²³³ The ligands were first functionalized with tri-alkoxysilyl group allowing the bonding to the silica NPs before complexing the lanthanide. This was the first report presenting modified β -diketonates derivatives bonded mesoporous host. A similar system has also been proposed with **97** as ligand complexing either Eu³⁺ or Tb³⁺.²³⁴ In 2009, the team of Reddy designed a hybrid material with the MCM-41 silica NPs.²³⁵ They covalently linked a [Eu(**98**)₃(**99**)] to the silica NPs by grafting **98** to a coupling agent by a co-condensation route. This system exhibited an overall quantum yield of 43% and an intrinsic quantum yield reaching 81% for an energy transfer efficiency of 53%. The ligand replacement by **18** and **100** resulted into lower overall and intrinsic quantum yields (30% and 53%) with a quite similar energy transfer efficiency (57%).²³⁶ Huo and co-workers immobilized



[Eu(**101**)₃(H₂O)₂] and [Tb(**102**)₃(H₂O)₂] complexes onto mesoporous silica NPs with the use of a silane compound to obtain new luminescent hybrid nanomaterials.²³⁷ The nanomaterial containing Tb exhibited higher lifetimes (864 and 1450 μs) than the one containing Eu (372 and 893 μs). The absence of emission from the ligand in the emission spectrum of [Tb(**102**)₃(H₂O)₂] indicated a more efficient energy transfer from the ligand to the Tb(III). Li and co-workers proposed silica NPs functionalization using [Dy(**103**)(**18**)₂(ClO₄)₃(H₂O)₂] and [Dy(**103**)(**104**)₂(ClO₄)₃(H₂O)₂].²³⁸ **103** was used as a silane coupling agent to link Dy to the silica NPs, enhancing the fluorescence emission of the lanthanide by 2.71 and 3.41 times for [Dy(**103**)(**18**)₂(ClO₄)₃(H₂O)₂] and [Dy(**103**)(**104**)₂(ClO₄)₃(H₂O)₂], respectively. The grafting of Eu complex instead of Dy on the same NPs, using a silane coupling linked technique to graft the complex to the silica NPs surface, led to a less important increase (approximately 2 times improvement).²³⁹ In 2018, Qian and co-workers presented the self-assembly of nanocomposites made with Ln³⁺ complexes and silica NPs.²⁴⁰ The complexes were designed with β-diketone ligands as **16** and **17** and were coordinated to the NPs by the intermediary of **107** which was used to functionalize the silica NPs and to sensitize the Ln ions. Their first method presented good results of forming Ln(III) complexes coated silica NPs but it is limited by the heterogeneous surface and a low grafting ratio. To overcome this issue, a second method consisting in grafting of organosilylated Ln(III) complexes on the surface of the silica NPs was developed. Menu and co-workers were the first to propose a system with this method, by using dipyrindine derivatives with an alkoxy silane group to form [Eu(**108**)₃(**109**)] and [Eu(**108**)₃(**110**)] complexes prior to attach them on the surface of the silica NPs.²²⁸ By comparison with free complexes, the grafted complexes on the silica NPs were found to be more efficient when excited in the UV (260 nm) or in the near UV (340 nm). Two new probes for *E. coli* bacteria bioimaging were also proposed, based on the functionalization of silica NPs with [Eu(**114**)₃] and [Eu(**16**)₃(**108**)],^{236,237} before proposing a new tool for biological analysis with a Vis-NIR-emitting probe.²³⁹ A [Yb(**16**)₃(**115**)] complex was linked to [Ru(**104**)(**109**)], resulting in a [Yb(**16**)₃(**115**)Ru(**104**)(**109**)]Cl₂ complex that was covalently bonded to the silica NPs by the intermediary of **109**. This system confers the possibility of having emission in the visible and in the NIR by excitation into the singlet Metal-Ligand Charge Transfer transition of the Ru(II). An energy transfer of 73% is obtained from Ru(II) to Yb(III). Both good luminescence and biological properties were demonstrated, essential qualities for a potential biomarker.

Taylor and Lin presented a method for the detection of dipicolinic acid (DPA, **5**) using different EDTA based ligands for Tb(III) coordination.²⁴³ [Tb(**111**)(H₂O)₃], [Tb(**112**)(H₂O)₃] and [Tb(**113**)(H₂O)₃] complexes were individually covalently attached to silica NPs and in the presence of DPA, Tb(III) emission was observed thanks to the replacement of water molecules by DPA. The selectivity of the method has also been tested and presented high selectivity for DPA.

5.3. Functionalized metal NPs

This section will be dedicated to the various types of metallic NPs functionalized with Ln complexes, namely iron, silver and gold NPs. A comparison of the photoluminescence properties of the systems (when applicable) will be presented in Table 5.

a Functionalized iron NPs (Fe-NPs). In the last decade, magnetic NPs has become an important tool in various fields such as data storage, magnetic fluids and bioapplications, thanks to their unique magnetic properties (superparamagnetic, high magnetic susceptibility).^{244–246} In this context, superparamagnetic iron oxide (SPIO) NPs such as magnetite (Fe₃O₄) and maghemite (Fe₂O₃) are the most used as they are biocompatible.²⁴⁷ However, they are relatively unstable as they tend to aggregate and are easily oxidized, thus losing their magnetic properties.²⁴⁸ To overcome these limitations, Fe-NPs are often functionalized with organic compounds, silica shell and polymers to stabilize them and maintain their physical characteristics.^{248–250} The stabilized Fe-NPs could then be functionalized with Ln(III)-complexes, opening the possibility to make bifunctional NPs combining magnetic and optical properties, which will be the main focus of this part. This bifunctional material can be particularly useful in biological applications where magnetic separation and optical detection are needed. In 2007, Hur and colleagues prepared a nanomaterial by functionalizing Fe₃O₄-silica NPs with ligand **105** complexing either Tb(III) or Eu(III),²⁵¹ while Shi and his team covalently grafted [Tb(**116**)₃] complexes onto Fe₃O₄-silica NPs to obtain a bifunctional material with a quantum yield of 5.6% and a saturation magnetization (SM) of 7.44 emu g^{−1},²⁵² SM corresponding to the maximum of magnetization reached in a sufficiently large magnetic field.²⁵³

Jingli Yuan's team designed NPs with magnetic and optical properties by combining Fe₃O₄ NPs and [Eu(**117**)] complex.²⁵⁴ The NPs were firstly coated with poly(vinylpyrrolidone) (PVP) to stabilize the Fe-NPs before [Eu(**117**)] copolymerization with free 3-aminopropyltriethoxysilane (APS), tetraethyl orthosilicate and Fe-NPs which led to a covalent bond between Eu-complexes and the NPs thanks to the presence of APS. With a S/N ratio 4 times higher when using time-resolved fluorescence imaging than steady-state fluorescence imaging and a SM of 2.5 emu g^{−1} which is enough for bio-separation,²⁵⁵ it has been proposed as a probe for biological labelling, detection and cell separation. A few years after, Yang and his team coated two [Tb(**118**)(**119**)] complexes on the surface of Fe₃O₄ NPs.²⁵⁶ Functionalized NPs were used as luminescent probe for targeted HeLa cells imaging thanks to the affinity between **119** and the folate receptor of the cells, and would be able to provide contrast in MRI. Another luminomagnetic system was proposed by Ni *et al.*, composed of Fe₃O₄ NPs functionalized with smaller NPs containing [Eu(**17**)₃(**18**)]Cl₃ complexes (Fig. 16), both NPs owning a protective silica shell.²⁵⁷ These functionalized Fe-NPs with an average size of 570 nm can be used in optical imaging as well as for effective magnetic separation. In 2014, Malta and co-workers chose a different path to functionalize the Fe-NPs by using calixarene-Ln(III) com-



Table 5 Comparison of the main properties of the metal NPs

NP composition	Ln	Antenna	Functionalized NP diameter (nm)	QY (%)	Lifetime ^a (ms)	Enhancement factor ^b	λ_{exc} (nm)	Ref.
Fe-NPs	Tb	116	120 to 160	5.6	0.9		307	252
	Eu	117			0.35		334	254
	Tb	118		0.096	1.15		325	256
		118; 119						
	Eu	16	26	14	0.23		372	258
	Eu	16; 98			0.35		365	259
	Eu	25		0.37	0.35 (solid) and 0.64 (suspension)		270 and 310	260
	Eu	120			1.41 ^{28%} and 0.66 ^{72%}		332	261
	Tb				0.43 ^{63%} and 1.66 ^{37%}		338	
	Eu	16		29	0.28		340 and 370	262
Au-NPs	Tb	27		13	0.2		330	
	Eu	106		0.96	0.36		320	265
	Tb			0.38	0.73			
	Eu	121			8×10^{-4}		266	266
	Eu	122			0.29		340	268
	Eu	48	230		0.19 and 0.67		395	269
	Eu	48	120		0.16 ^{14%} 0.67 ^{90%}		330	270
	Eu	124			0.54		279	271
	Tb	80		12.2	0.63			
Ag-NPs					4 to 6.3		371	272
	Eu	16	22	21	0.37		343	274
	Eu	16; 104			0.62	21	343	275
	Sm	2; 18; 104				3.8	427, 441, 463	276
	Dy					5.9		
	Eu	2; 129				2 to 6		277
	Tb			6.5 and 23		3 to 26		

^a Ln³⁺ lifetime after functionalization of the NPs. Percentages are the relative distribution of the lifetimes obtained. ^b Luminescence enhancement factor between the free complex and the functionalized NPs.

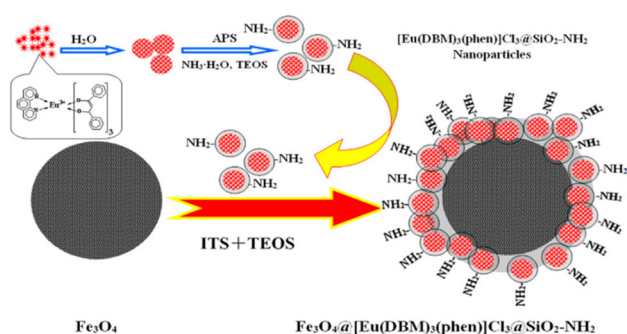


Fig. 16 Preparation of a functionalized Fe-NPs with Ln-complexes containing NPs Reprinted from ref. 257, Copyright (2013), with permission from Elsevier.

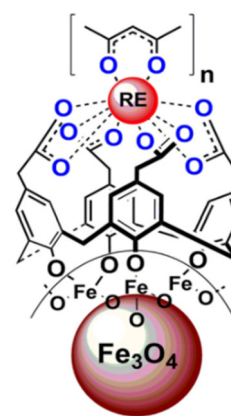


Fig. 17 Structure of Fe₃O₄@calix-Ln(β-diketonate). Reprinted with permission from ref. 258. Copyright (2014) American Chemical Society.

plexes.²⁵⁸ Thanks to the one-pot method, [Eu(16)] and [Tb(27)] complexes were successfully attached to the Fe-NPs with a calixarene as intermediate (Fig. 17) for the stabilization of the NPs. The efficiency obtained with this system was lower than the complex *i.e.* 14% and 29% for the Fe₃O₄@calix-Eu(16) and the free [Eu(16)₃(H₂O)₂] complex, respectively. Two years later, Ribeiro and his team chose to work with other SPIO NPs,

Fe₂O₃, for designing their bifunctional magnetic luminescent particles composed of [Eu(16)(98)] complexes grafted on the surface of the silica shell of the NPs.²⁵⁹

By comparison with the original complex, the luminescence lifetime decreased from 0.60 ms to 0.35 ms which might be



due to some defects on the SPIO NPs, and the SM decreased from 60 emu g⁻¹ to 0.12 emu g⁻¹. In 2019, Carlos Gerales and his team proposed a system with maghemite covered by a silica shell and impregnated with [Eu(25)₃(H₂O)₂], acting as a luminescent probe,²⁶⁰ exhibiting enhancement of the quantum efficiency from 0.19 (free complex) to 0.37 for the NP. [Gd(25)₃((H₂O)₂)] was also added, affording a bimodal system able to act as a MRI contrast agent in addition to its luminescent role. The following year, Stefan Lis and his team proposed a new system with the expectation of improving the bifunctionality (magnetism and luminescence), that Fe-NPs and Ln-complexes can provide.²⁶¹ They prepared Fe₃O₄@silica core-shell NPs in which [Ln(120)] (Ln = Tb or Eu) are linked to the NPs by the intermediary of terephthalic acid. Unfortunately, a decrease of both luminescence intensity and lifetime of the system is observed in comparison to the respective Ln-complexes at R.T and at 77 K, explained by the low content of chain complexes and by the high absorption of magnetic cores. Recently, new luminescent-magnetic architectures were developed by covering magnetite NPs with biocompatible chitosan (CS) and glutathione (GSH) functionalized with either [Eu(16)₃(H₂O)₂] or [Tb(27)₃(H₂O)₃], leading to Fe₃O₄-CS-Eu/Tb and Fe₃O₄-GSH-Eu/Tb, with promising bioapplications.²⁶² Quantum emission efficiencies of 29% and 13% and lifetimes of 0.28 ms and 0.2 ms were obtained for the Fe₃O₄-CS-Eu and Fe₃O₄-GSH-Eu NPs, respectively. The values of the former are close to the free Eu-complex (29% and 0.26 ms) and a decrease is observed with those of the latter. These results can be related to the number of water molecules found in the first coordination sphere of the Eu atoms which are of 2.5 for Fe₃O₄-CS-Eu and 5 for Fe₃O₄-GSH-Eu.

b Functionalized gold NPs (Au-NPs). The biocompatibility of Au-NPs along with the simplicity of surface coating with organic and biological molecules make these NPs a nanostructure of choice for functionalization and for biological applications.^{263,264} No specific activity related to Au-NPs properties is described, Au-NPs are mostly used as biocompatible nanocarriers for the Ln complexes.

The first examples were reported in 2006, when Thomas and his team coated the surface of the Au-NPs with [Ln(106)₃] (Ln = Eu(III) or Tb(III)).²⁶⁵ The phosphorescent nanomaterial was proposed as sensor for biologically important cations such as Ni(II), Ca(II), Mg(II) and others. Different metal ions were added to solutions of functionalized NPs, which may cause a replacement of Ln(III) by the metal ion depending on its affinity for the ligand, leading to a decrease of Ln photoluminescence intensity. The presence of transition metal ions has been shown to induce a higher decrease of the luminescence intensity compared to alkaline earth metals. In parallel, Pikramenou's team designed a nanomaterial composed of Au-NPs coated with [Eu(121)] complexes.²⁶⁶ The ligand conferred a strong affinity with the surface of Au-NPs thanks to the presence of thiol functions, known to be poorly coordinating towards Ln atoms.²⁶⁷ A luminescent pH probe for human platelets was then designed, composed of Au-NPs coated by [Eu(122)] complexes and by peptides.²⁶⁸ The Eu related lumi-

nescent properties of the NPs make them suitable optical probes for imaging in cells whereas the peptides allow the NPs to cross the cell membrane. By the intermediary of the peptide, the nano-objects associate with the cell membrane at a pH of 7.4, and upon reduction of the pH (≤6.5), the NPs enter the cell membrane. This pH-mediated internalization is triggered by the pH-dependent peptide translocation across the membrane. [Eu(122)] complex was chosen for its neutrality, its good stability, and its inertness towards the peptide. The NPs luminescence was monitored in platelets and displayed luminescence lifetimes of 288 μs and 277 μs in spread and fixed platelets respectively, which are close to the lifetimes of the NPs in solution (290 μs) and confirmed the presence of the NPs in the cells. In 2008, Gunnlaugsson and co-workers presented their own Ln-complex functionalized Au-NPs as a probe for luminescent sensing of biological substrates.²⁶⁹ [Eu(123)] functionalized Au-NPs were prepared from DMAP-Au-NPs by exchange of DMAP (4-(dimethylamino)-pyridine) with Eu-chelate. The NPs exhibited no luminescence when the ligand was excited but after the addition of ligand **48**, this antenna sensitized the Eu excited state, leading to the observation of red emission. The interaction of this system with Bovine Serum Albumin (BSA) was also studied, resulting in the quenching of the Eu(III) luminescence, explained by the strong interaction between BSA and ligand **48**.²⁷⁰ With the same sensitizer, [Eu(124)] coated Au-NPs were also proposed for analysis of the structure of microdamaged bovine bones.²⁷¹ In 2018, Werts and co-workers used the system of [Tb(80)] labelled streptavidin (mentioned in QDs part) to coat the surface of biotinylated Au-NPs for nanosurface energy transfer,²⁷² an energy-transfer mechanism measuring (bio)molecular interactions at longer distances than FRET.²⁷³

c Functionalized silver NPs (Ag-NPs). The functionalization of metal NPs with Ln(III)-complexes was also proposed using Ag-NPs with expectations of improving the optical properties of the Ln(III) complexes thanks to the fact that absorption frequency of Ln complexes is close to the surface plasmon resonance of Ag-NPs.²⁷⁴ Zhang and co-workers were the first to present a surface coating of Ag-NPs by [Eu(16)₃(H₂O)₂] in 2005,²⁷⁴ with a system prepared by mixing AgNO₃, NaBH₄ as a reducer and the complexes in solution.

Ag colloidal NPs exhibited a luminescence as intense as the free Eu-complex with similar lifetimes and quantum yield (0.386 ms and 23.1% for the complex and 0.368 ms and 21% for the Ag colloidal NPs) due to the low energy transfer between the Eu-complex and the metal, leading to a weak quenching effect, caused by the aggregation of the complexes at the surface of the Ag-NPs which induced a red-shift of the absorption band (J-aggregation).^{278,279} The ligand **104** was then employed in addition to **16** as chelator in Ln complexes to substitute the water molecules.²⁷⁵ In addition of [Eu(16)₃104], [Tb(16)₃104] was used to stabilize the nanocomposite as it helps reducing the quenching effect. An enhancement of 21% for the luminescence emission intensity of the [Eu(16)₃104] complex was then obtained compared to the one of the complex itself. Following their steps, Chu and



co-workers studied the effect of the presence of the silica shell between Ln complexes and Ag-NP surface on Ln luminescence.²⁷⁶ Three different core-shell Ag@SiO₂ NPs were prepared with varying silica shell thicknesses (10, 25 and 80 nm) and with Ln complexes attached at the surface of the NPs. Sm(III) and Dy(III) complexes were chosen with **2**, **18** and **104** as photosensitizing ligands. All the compounds exhibited improved photoluminescence properties than the free Ln complexes with a 5.9-fold and 3.8-fold enhancement of emission intensity for the Dy(III) and Sm(III) complexes coated Ag-NPs, respectively. The impact of the position of substituents in aromatic ligands on the luminescence of Sm(III) and Dy(III) complexes functionalized Ag@SiO₂ NPs was also studied with **125** (–NH₂ in *ortho* position), **126** (–NH₂ in *meta* position), and **127** (–NH₂ in *para* position) as chelators.²⁸⁰ A stronger emission was observed with amino group in *meta* position (ligand **126**) as the positive conjugation effect of amino group is weaker.

In 2014, Yan and colleagues proposed a new approach to obtain a white emitting soft material using [Eu(**16**)₄], [Sm(**17**)₄] or [Eu(**128**)₄] complexes linked to Ag-NPs thanks to a thiol-functionalized ionic liquid (SHIL).²⁸¹ By changing the number of complexes bound to Ag-NPs, the luminescence colour of the soft hybrid material was adjusted thanks to the band gap differences between the Ln(III) complexes and the NPs. Among the different systems, only Ag-SHIL-[Eu(**128**)₄]₃, Ag-SHIL-[Sm(**17**)₄]₃ and Ag-SHIL-[Eu(**16**)₄]₃ systems displayed white luminescence.

To the best of our knowledge, the last report was made by Zhao's team in 2020,²⁷⁷ where the different factors impacting the metal-enhanced luminescence of NPs were investigated. For this purpose, Ag-NPs were synthesized with different thickness of the silica shell upon which [Eu(**2**)₃], [Eu(**129**)₃], [Tb(**2**)₃] and [Tb(**129**)₃] complexes were separately adsorbed. Excitation wavelength has been considered as a key factor with a luminescent enhancement from 1.55 to 14.97 times when changing the excitation wavelength from 255 to 380 nm between the nanocomposite and the free complex. The silica shell thickness was also considered as a key factor as the distance between the complex and the core of the NPs influences the luminescence of Ln. The best quantum yield (23%) was obtained with a thickness of 24 nm compared to the free complex (3.1%) and a shell with nearly the double of the thickness (6.5%).

In this section, the functionalization of different NPs by Ln(III) complexes have been overviewed. Those new nanomaterials were useful for multiple applications such as imaging and sensing, and led to increased luminescence properties (lifetimes, emission intensity) of the complexes compared to the free ones, particularly in the case of the silica NPs, Au-NPs and Ag-NPs. Limitations related to the FRET distance were overcome with these types of NPs by reaching a distance up to 14 nm with the use of a dye as a bridge.²¹⁶ However, this improvement comes with a limited FRET efficiency (less than 20%). Bifunctional nano-objects were also designed with Fe-NPs, making possible to combine magnetic and luminescence properties. However, to the best of our knowledge, no Gd based NPs with Ln-complex functionalized surface that could

combine photoluminescence and MRI properties has been described up to date. Such a luminescence/MRI bimodal probe would be of great interest,²⁸² especially with NIR-light absorbing and emitting Ln complexes.

6. Outlook and conclusion

Bringing the antenna effect of photosensitizing ligands to the level of Ln-NPs has created a new playground for chemists and physicists with extremely bright nano-objects. The complementary advantages of long-lived excited states and spectral signatures of each Ln ions further afford highly sensitive luminescence detection and the capacity of multiplexed detection with numerous (bio)analytical applications.

A direct comparison of the performances between the different types of NPs and even in the same category is quite difficult as few studies present a full spectroscopic characterization (excited-state lifetime, quantum yield and brightness). However, it can be highlighted that each type of NPs has its advantages and limitations that make them the most suitable depending on the application. For example, by comparing the higher Tb lifetimes of each category of NPs, Ln-doped NPs with surface capping antennas exhibit the highest Tb lifetimes (up to 8.3 ms – quite close to the radiative lifetime²⁸³ – vs. 2.06 ms for Ln-complex encapsulated NPs and 1.66 ms for Ln-complex surface functionalized NPs). Embedding the atoms in the core of the NPs clearly affords a better Ln protection from deactivation caused by the environment, the latter being still important for the encapsulated or grafted complexes. However, (bio)functionalization remains a limiting factor for analytical applications as numerous/complex steps of organic synthesis are required to form functionalized antennas, when surface with active function can be more easily obtained for polymer or silica NPs. Finally, surface decoration of NPs with Ln complexes has a unique advantage of offering bifunctional detections by the combination of luminescent properties of Ln complexes with the specific properties of the NPs (such as surface plasmon resonance for Ag-NPs, magnetism for Fe-NPs) or to present multiplexed optical detection and FRET possibilities with QDs. However, in this case, Ln complexes are not protected from the environment and consequently are more impacted by solvent related deactivations like free complexes, potentially leading to lower spectroscopic performances compared with the first two types of NPs.

As seen in the last three sections, most of the light harvesting range of dye-sensitized NPs is in the UV-Vis zone, which might provide biocompatibility issues. The development of NIR-excited Ln containing NPs would be of particular importance to comply with the biological window and allow non-invasive study of biological specimens. Several strategies, already illustrated in few studies but demanding deeper development, can be considered: the use of NIR-light absorbing antennas in combination with NIR absorbing Ln (such as Yb or Er) either for upconversion or downshifting, or two-photon excitation antennas excited in the NIR range.



Considering the relative youth of the field, it is surmised that new systems will be developed to better tune the up and down energy transfer processes, with surface ligands playing the rule of energy bridges between the core NPs and the external surroundings,²⁸⁴ or with new spectroscopic properties arising from the interaction of the NPs with chemical entities at the surface as observed for the triplet excitons enhancement occurring for some organic dye interacting with Ln-NPs.²⁸⁵

The maturation stage of these new tools will probably soon open them the doors of commercial applications. In particular, the bio-analytical fields are already tackling the use of Ln-NPs for *in vitro* applications such as lateral flow immunoassays,²⁸⁶ ELISA like detection,²⁸⁷ or luminescence microscopy.²⁸⁸ However, the high sensitivity obtained with Ln-NPs is based on the development of dedicated instruments, working with a time-delayed detection mode, that will require dedicated time-resolved readers to unravel the best of the sensitivity of these nano-objects. Additionally, if few reports studied their long term chemical or colloidal stabilities,^{46,47} and, if applications have already been found in *in vitro* and *in cellulo* diagnostics, numerous questions remain to be answered concerning the biocompatibility and toxicity of these nano-objects before any eventual long term *in vivo* applications could be safely envisaged.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

This work was funded by the French National Research Agency (ANR) through the contract number ANR-11-LABX-0058_NIE within the investissement d'Avenir program ANR-10-IDEX-0002-02 and through the project ANR-21-CE19-0016-01. The French Ministère de l'enseignement Supérieur, de la Recherche et des Technologies is gratefully acknowledged for financial support (LKS).

References

- 1 J.-C. G. Bünzli, *Coord. Chem. Rev.*, 2015, **293**, 19–47.
- 2 C. Doffek and M. Seitz, *Angew. Chem., Int. Ed.*, 2015, **54**, 9719–9721.
- 3 S. I. Weissman, *J. Chem. Phys.*, 1942, **10**, 214–217.
- 4 J.-C. G. Bünzli and C. Piguet, *Chem. Soc. Rev.*, 2005, **34**, 1048–1077.
- 5 S. J. Butler, M. Delbianco, L. Lamarque, B. K. McMahon, E. R. Neil, R. Pal, D. Parker, J. W. Walton and J. M. Zwieter, *Dalton Trans.*, 2015, **44**, 4791–4803.
- 6 L. D. Lavis and R. T. Raines, *ACS Chem. Biol.*, 2008, **3**, 142–155.
- 7 N. Sabbatini, S. Perathoner, V. Balzani, B. Alpha and J.-M. Lehn, in *Supramolecular Photochemistry*, ed. V. Balzani, Springer Netherlands, Dordrecht, 1987, pp. 187–206.
- 8 S. J. Butler, L. Lamarque, R. Pal and D. Parker, *Chem. Sci.*, 2014, **5**, 1750–1756.
- 9 N. Weibel, L. J. Charbonnière, M. Guardigli, A. Roda and R. Ziessel, *J. Am. Chem. Soc.*, 2004, **126**, 4888–4896.
- 10 J. Xu, T. M. Corneillie, E. G. Moore, G.-L. Law, N. G. Butlin and K. N. Raymond, *J. Am. Chem. Soc.*, 2011, **133**, 19900–19910.
- 11 P. Reiss, M. Protière and L. Li, *Small*, 2009, **5**, 154–168.
- 12 N. Souri, P. Tian, C. Platas-Iglesias, K.-L. Wong, A. Nonat and L. J. Charbonnière, *J. Am. Chem. Soc.*, 2017, **139**, 1456–1459.
- 13 L. Charbonnière, S. Mameri, P. Kadjane, C. Platas-Iglesias and R. Ziessel, *Inorg. Chem.*, 2008, **47**, 3748–3762.
- 14 A. J. Freeman and R. E. Watson, *Phys. Rev.*, 1962, **127**, 2058–2075.
- 15 I. L. Medintz, H. T. Uyeda, E. R. Goldman and H. Mattoussi, *Nat. Mater.*, 2005, **4**, 435–446.
- 16 J. Goetz, A. Nonat, A. Diallo, M. Sy, I. Sera, A. Lecointre, C. Lefevre, C. F. Chan, K.-L. Wong and L. J. Charbonnière, *ChemPlusChem*, 2016, **81**, 526–534.
- 17 A. Beeby, I. M. Clarkson, R. S. Dickins, S. Faulkner, D. Parker, L. Royle, A. S. de Sousa, J. A. G. Williams and M. Woods, *J. Chem. Soc., Perkin Trans. 2*, 1999, 493–504.
- 18 R. M. Supkowski and W. DeW. Horrocks, *Inorg. Chim. Acta*, 2002, **340**, 44–48.
- 19 T. Kimura and Y. Kato, *J. Alloys Compd.*, 1998, **271**–273, 867–871.
- 20 T. Kimura and Y. Kato, *J. Alloys Compd.*, 1995, **225**, 284–287.
- 21 C. Doffek, N. Alzakhem, C. Bischof, J. Wahsner, T. Güden-Silber, J. Lügger, C. Platas-Iglesias and M. Seitz, *J. Am. Chem. Soc.*, 2012, **134**, 16413–16423.
- 22 C. Homann, L. Krukewitt, F. Frenzel, B. Grauel, C. Würth, U. Resch-Genger and M. Haase, *Angew. Chem., Int. Ed.*, 2018, **57**, 8765–8769.
- 23 M. Haase and H. Schäfer, *Angew. Chem., Int. Ed.*, 2011, **50**, 5808–5829.
- 24 R. Englman and J. Jortner, *Mol. Phys.*, 1970, **18**, 145–164.
- 25 Q. Ju, Y. Liu, D. Tu, H. Zhu, R. Li and X. Chen, *Chem. – Eur. J.*, 2011, **17**, 8549–8554.
- 26 J.-C. G. Bünzli, *Chem. Rev.*, 2010, **110**, 2729–2755.
- 27 M. Irfanullah, N. Bhardwaj and A. Chowdhury, *Dalton Trans.*, 2016, **45**, 12483–12495.
- 28 Y. Zhong and H. Dai, *Nano Res.*, 2020, **13**, 1281–1294.
- 29 A. Lay, C. Siefe, S. Fischer, R. D. Mehlenbacher, F. Ke, W. L. Mao, A. P. Alivisatos, M. B. Goodman and J. A. Dionne, *Nano Lett.*, 2018, **18**, 4454–4459.
- 30 Z. Zhuo, Y. Liu, D. Liu, P. Huang, F. Jiang, X. Chen and M. Hong, *Chem. Sci.*, 2017, **8**, 5050–5056.
- 31 X. Cheng, D. Tu, W. Zheng and X. Chen, *Chem. Commun.*, 2020, **56**, 15118–15132.
- 32 F. Auzel, G. Baldacchini, L. Laversenne and G. Boulon, *Opt. Mater.*, 2003, **24**, 103–109.



- 33 Z. Liu, Q. Meng, H. Liu, C. Yao, Q. Meng, W. Liu and W. Wang, *Opt. Mater.*, 2013, **36**, 384–389.
- 34 P. A. Tanner, L. Zhou, C. Duan and K.-L. Wong, *Chem. Soc. Rev.*, 2018, **47**, 5234–5265.
- 35 M. W. Mara, D. S. Tatum, A.-M. March, G. Doumy, E. G. Moore and K. N. Raymond, *J. Am. Chem. Soc.*, 2019, **141**, 11071–11081.
- 36 N. Hildebrandt and I. L. Medintz, *FRET - Förster Resonance Energy Transfer: From Theory to Applications*, Wiley-VCH Verlag GmbH & Co. KGaA, First edition., 2014.
- 37 S. Bhuckory, E. Hemmer, Y.-T. Wu, A. Yahia-Ammar, F. Vetrone and N. Hildebrandt, *Eur. J. Inorg. Chem.*, 2017, **2017**, 5186–5195.
- 38 K. D. Wegner, Z. Jin, S. Lindén, T. L. Jennings and N. Hildebrandt, *ACS Nano*, 2013, **7**, 7411–7419.
- 39 E. Debroye and T. N. Parac-Vogt, *Chem. Soc. Rev.*, 2014, **43**, 8178–8192.
- 40 A. M. Nonat and L. J. Charbonnière, *Coord. Chem. Rev.*, 2020, **409**, 213192.
- 41 S. Faulkner and S. J. A. Pope, *J. Am. Chem. Soc.*, 2003, **125**, 10526–10527.
- 42 M. Sy, D. Esteban-Gómez, C. Platas-Iglesias, A. Rodríguez-Rodríguez, R. Tripier and L. J. Charbonnière, *Eur. J. Inorg. Chem.*, 2017, **2017**, 2122–2129.
- 43 C. Chen, L. Ao, Y. T. Wu, V. Cifliku, M. Cardoso Dos Santos, E. Bourrier, M. Delbianco, D. Parker, J. M. Zwier, L. Huang and N. Hildebrandt, *Angew. Chem., Int. Ed.*, 2018, **57**, 13686–13690.
- 44 X. Liu, W. Wu, D. Cui, X. Chen and W. Li, *Adv. Mater.*, 2021, **33**, 2004734.
- 45 H. Xing, W. Bu, S. Zhang, X. Zheng, M. Li, F. Chen, Q. He, L. Zhou, W. Peng, Y. Hua and J. Shi, *Biomaterials*, 2012, **33**, 1079–1089.
- 46 E. Fröhlich, *J. Nanobiotechnol.*, 2017, **15**, 84.
- 47 A. Gnach, T. Lipinski, A. Bednarkiewicz, J. Rybka and J. A. Capobianco, *Chem. Soc. Rev.*, 2015, **44**, 1561–1584.
- 48 V. Mittelheisser, P. Coliat, E. Moeglin, L. Goepf, J. G. Goetz, L. J. Charbonnière, X. Pivot and A. Detappe, *Adv. Mater.*, 2022, e2110305.
- 49 O. Dukhno, F. Przybilla, V. Muhr, M. Buchner, T. Hirsch and Y. Mély, *Nanoscale*, 2018, **10**, 15904–15910.
- 50 S. Lahtinen, A. Lyytikäinen, H. Pääkkilä, E. Hömppi, N. Perälä, M. Lastusaari and T. Soukka, *J. Phys. Chem. C*, 2017, **121**, 656–665.
- 51 W. Zheng, P. Huang, D. Tu, E. Ma, H. Zhu and X. Chen, *Chem. Soc. Rev.*, 2015, **44**, 1379–1415.
- 52 S. Heer, K. Kömpe, H.-U. Güdel and M. Haase, *Adv. Mater.*, 2004, **16**, 2102–2105.
- 53 K. W. Krämer, D. Biner, G. Frei, H. U. Güdel, M. P. Hehlen and S. R. Lüthi, *Chem. Mater.*, 2004, **16**, 1244–1251.
- 54 M. Quintanilla, E. Hemmer, J. Marques-Hueso, S. Rohani, G. Lucchini, M. Wang, R. R. Zamani, V. Roddatis, A. Speghini, B. S. Richards and F. Vetrone, *Nanoscale*, 2022, **14**, 1492–1504.
- 55 L. J. Charbonnière, J.-L. Rehspringer, R. Ziessel and Y. Zimmermann, *New J. Chem.*, 2008, **32**, 1055–1059.
- 56 B. Kokuz, C. Kucera, J. R. DiMaio and J. Ballato, *Opt. Mater.*, 2009, **31**, 1327–1330.
- 57 N. Chen, Y. He and G. Du, *Mater. Sci. Semicond. Process.*, 2014, **24**, 62–67.
- 58 Y. Yang, B. Liu, L. Tang and N. Chen, *Mater. Sci. Semicond. Process.*, 2015, **30**, 513–517.
- 59 J. Wang, W. Wang, L. Zhang, H. Yao and Z. Li, *Mater. Lett.*, 2013, **106**, 281–283.
- 60 B. Kokuz, J. Dimaio, C. Kucera, J. Evanoff David and J. Ballato, *J. Am. Chem. Soc.*, 2008, **130**, 12222–12223.
- 61 A. M. Cross, P. S. May, F. C. J. M. van Veggel and M. T. Berry, *J. Phys. Chem. C*, 2010, **114**, 14740–14747.
- 62 S. W. Li, H. J. Ren and S. G. Ju, *J. Nanosci. Nanotechnol.*, 2014, **14**, 3677–3682.
- 63 S. Li, X. Zhang, Z. Hou, Z. Cheng, P. Ma and J. Lin, *Nanoscale*, 2012, **4**, 5619–5626.
- 64 S. Li, X. Li, Y. Jiang, Z. Hou, Z. Cheng, P. Ma, C. Li and J. Lin, *RSC Adv.*, 2014, **4**, 55100–55107.
- 65 D. Ghosh and M. N. Luwang, *RSC Adv.*, 2015, **5**, 10468–10478.
- 66 V. Yu. Khudoleeva, V. V. Utochnikova, A. S. Kalyakina, I. M. Deygen, A. A. Shiryaev, L. Marciniak, V. A. Lebedev, I. V. Roslyakov, A. V. Garshev, L. S. Lepnev, U. Schepers, S. Bräse and N. P. Kuzmina, *Dyes Pigm.*, 2017, **143**, 348–355.
- 67 S. Janssens, G. V. M. Williams and D. Clarke, *J. Appl. Phys.*, 2011, **109**, 023506.
- 68 S. Janssens, G. Williams, D. Clarke and M. Sharp, *Basic Solid State Phys.*, 2011, **248**, 439–443.
- 69 X. Wang, Y. Yang, N. Chen, B. Liu and G. Liu, *J. Nanosci. Nanotechnol.*, 2016, **16**, 3729–3734.
- 70 X. Wang, Y. Yang, G. Du, N. Chen and A. Zhang, *J. Nanosci. Nanotechnol.*, 2018, **18**, 1876–1881.
- 71 A. Safronikhin, H. Ehrlich, N. Kuzmina and G. Lisichkin, *Appl. Surf. Sci.*, 2014, **307**, 482–488.
- 72 M. Irfanullah, N. Bhardwaj and A. Chowdhury, *Dalton Trans.*, 2016, **45**, 12483–12495.
- 73 M. Irfanullah, D. K. Sharma, R. Chulliyil and A. Chowdhury, *Dalton Trans.*, 2015, **44**, 3082–3091.
- 74 V. N. K. B. Adusumalli, L. Mrówczyńska, D. Kwiatek, Ł. Piosik, A. Lesicki and S. Lis, *ChemMedChem*, 2021, **16**, 1640–1650.
- 75 C. Charpentier, V. Cifliku, J. Goetz, A. Nonat, C. Cheignon, M. Cardoso Dos Santos, L. Francés-Soriano, K.-L. Wong, L. J. Charbonnière and N. Hildebrandt, *Chem. – Eur. J.*, 2020, **26**, 14602–14611.
- 76 C. Cheignon, M. Heurté, R. C. Knighton, A. A. Kassir, A. Lecointre, A. Nonat, A. Boos, C. Christine, Z. Asfari and L. J. Charbonnière, *Eur. J. Inorg. Chem.*, 2021, **2021**, 3761–3770.
- 77 J. Wang, Z. Wang, X. Li, S. Wang, H. Mao and Z. Li, *Appl. Surf. Sci.*, 2011, **257**, 7145–7149.
- 78 L. Song and L. Xue, *Appl. Surf. Sci.*, 2012, **258**, 3497–3501.
- 79 L. Song, J. Gao and J. Li, *J. Lumin.*, 2014, **151**, 18–21.



- 80 V. N. K. B. Adusumalli, H. V. S. R. M. Koppiseti, N. Madhukar and V. Mahalingam, *Microchim. Acta*, 2019, **186**, 389.
- 81 N. Gauthier, O. Racourt, D. Imbert and M. Mazzanti, *J. Nanopart. Res.*, 2013, **15**, 1723.
- 82 T. Samanta, S. K. Jana, A. E. Praveen and V. Mahalingam, *Dalton Trans.*, 2017, **46**, 9646–9653.
- 83 P. Agbo, T. Xu, M. Sturzbecher-Hoehne and R. J. Abergel, *ACS Photonics*, 2016, **3**, 547–552.
- 84 P. Agbo and R. J. Abergel, *Inorg. Chem.*, 2016, **55**, 9973–9980.
- 85 Y. Song, B. Shao, Y. Feng, W. Lü, G. Liu and H. You, *Dalton Trans.*, 2016, **45**, 9468–9476.
- 86 Q. Dai, M. E. Foley, C. J. Breshike, A. Lita and G. F. Strouse, *J. Am. Chem. Soc.*, 2011, **133**, 15475–15486.
- 87 L. Stagi, D. Chiriu, A. Ardu, C. Cannas, C. M. Carbonaro and P. C. Ricci, *J. Appl. Phys.*, 2015, **118**, 125502.
- 88 L. Ji, N. Chen, G. Du, M. Yan and W. Shi, *Ceram. Int.*, 2014, **40**, 3117–3122.
- 89 N. Chen, L. Ji and G. Du, *J. Lumin.*, 2014, **153**, 259–263.
- 90 U. Balderas, S. Carmona, L. Mariscal, I. Martínez and C. Falcony, *Chem. Phys.*, 2018, **511**, 1–6.
- 91 J. Chen, Q. Meng, P. S. May, M. T. Berry and C. Lin, *J. Phys. Chem. C*, 2013, **117**, 5953–5962.
- 92 L. Tang, W. Gui, K. Ding, N. Chen and G. Du, *J. Alloys Compd.*, 2014, **590**, 277–282.
- 93 Y. He, N. Chen and G. Du, *J. Am. Ceram. Soc.*, 2014, **97**, 1931–1936.
- 94 B. G. Vats, M. Shafeeq, P. Singhal and S. Neogy, *ChemistrySelect*, 2018, **3**, 4930–4938.
- 95 R. R. Gonçalves, Y. Messaddeq, M. A. Aegerter and S. J. L. Ribeiro, *J. Nanosci. Nanotechnol.*, 2011, **11**, 2433–2439.
- 96 J.-S. Kang, Y.-K. Jeong, J.-G. Kang, L. Zhao, Y. Sohn, D. Pradhan and K. T. Leung, *J. Phys. Chem. C*, 2015, **119**, 2142–2147.
- 97 F. Artizzu, D. Loche, D. Mara, L. Malfatti, A. Serpe, R. V. Deun and M. F. Casula, *J. Mater. Chem. C*, 2018, **6**, 7479–7486.
- 98 J. Huang and K. Pu, *Chem. Sci.*, 2021, **12**, 3379–3392.
- 99 J.-C. G. Bünzli and S. V. Eliseeva, *J. Rare Earths*, 2010, **28**, 824–842.
- 100 T. Fix, A. Nonat, D. Imbert, S. Di Pietro, M. Mazzanti, A. Slaoui and L. J. Charbonnière, *Prog. Photovolt. Res. Appl.*, 2016, **24**, 1251–1260.
- 101 J. Zhang, C. M. Shade, D. A. Chengelis and S. Petoud, *J. Am. Chem. Soc.*, 2007, **129**, 14834–14835.
- 102 H. Lu, Y. Peng, H. Ye, X. Cui, J. Hu, H. Gu, A. N. Khlobystov, M. A. Green, P. J. Blower, P. B. Wyatt, W. P. Gillin and I. Hernández, *Sci. Rep.*, 2017, **7**, 1–10.
- 103 J. Liu, A. M. Kaczmarek, F. Artizzu and R. Van Deun, *ACS Photonics*, 2019, **6**, 659–666.
- 104 V. V. Utochnikova, A. S. Kalyakina, L. S. Lepnev and N. P. Kuzmina, *J. Lumin.*, 2016, **170**, 633–640.
- 105 V. Y. Khudoleeva, V. V. Utochnikova, A. S. Goloveshkin, L. Marciniak, A. V. Knotko, L. S. Lepnev and N. P. Kuzmina, *Dyes Pigm.*, 2019, **160**, 890–897.
- 106 S. Wilhelm, *ACS Nano*, 2017, **11**, 10644–10653.
- 107 J. Lee, B. Yoo, H. Lee, G. D. Cha, H.-S. Lee, Y. Cho, S. Y. Kim, H. Seo, W. Lee, D. Son, M. Kang, H. M. Kim, Y. I. Park, T. Hyeon and D.-H. Kim, *Adv. Mater.*, 2017, **29**, 1603169.
- 108 K. L. Reddy, R. Balaji, A. Kumar and V. Krishnan, *Small*, 2018, **14**, 1801304.
- 109 W. Zou, C. Visser, J. A. Maduro, M. S. Pshenichnikov and J. C. Hummelen, *Nat. Photonics*, 2012, **6**, 560–564.
- 110 D. J. Garfield, N. J. Borys, S. M. Hamed, N. A. Torquato, C. A. Tajon, B. Tian, B. Shevitski, E. S. Barnard, Y. D. Suh, S. Aloni, J. B. Neaton, E. M. Chan, B. E. Cohen and P. J. Schuck, *Nat. Photonics*, 2018, **12**, 402–407.
- 111 X. Wu, H. Lee, O. Bilsel, Y. Zhang, Z. Li, T. Chen, Y. Liu, C. Duan, J. Shen, A. Punjabi and G. Han, *Nanoscale*, 2015, **7**, 18424–18428.
- 112 X. Wu, Y. Zhang, K. Takle, O. Bilsel, Z. Li, H. Lee, Z. Zhang, D. Li, W. Fan, C. Duan, E. M. Chan, C. Lois, Y. Xiang and G. Han, *ACS Nano*, 2016, **10**, 1060–1066.
- 113 D. Yin, Y. Liu, J. Tang, F. Zhao, Z. Chen, T. Zhang, X. Zhang, N. Chang, C. Wu, D. Chen and M. Wu, *Dalton Trans.*, 2016, **45**, 13392–13398.
- 114 G. Chen, J. Damasco, H. Qiu, W. Shao, T. Y. Ohulchanskyy, R. R. Valiev, X. Wu, G. Han, Y. Wang, C. Yang, H. Ågren and P. N. Prasad, *Nano Lett.*, 2015, **15**, 7400–7407.
- 115 W. Wei, G. Chen, A. Baev, G. S. He, W. Shao, J. Damasco and P. N. Prasad, *J. Am. Chem. Soc.*, 2016, **138**, 15130–15133.
- 116 G. Chen, W. Shao, R. R. Valiev, T. Y. Ohulchanskyy, G. S. He, H. Ågren and P. N. Prasad, *Adv. Opt. Mater.*, 2016, **4**, 1760–1766.
- 117 Q. Shao, X. Li, P. Hua, G. Zhang, Y. Dong and J. Jiang, *J. Colloid Interface Sci.*, 2017, **486**, 121–127.
- 118 W. Shao, G. Chen, A. Kuzmin, H. L. Kutscher, A. Pliss, T. Y. Ohulchanskyy and P. N. Prasad, *J. Am. Chem. Soc.*, 2016, **138**, 16192–16195.
- 119 J. Xu, M. Sun, Y. Kuang, H. Bi, B. Liu, D. Yang, R. Lv, S. Gai, F. He and P. Yang, *Dalton Trans.*, 2017, **46**, 1495–1501.
- 120 Y. Wei, S. Liu, C. Pan, Z. Yang, Y. Liu, J. Yong and L. Quan, *Int. J. Nanomed.*, 2020, **15**, 1409–1420.
- 121 W. Bi, Y. Wu, C. Chen, D. Zhou, Z. Song, D. Li, G. Chen, Q. Dai, Y. Zhu and H. Song, *ACS Appl. Mater. Interfaces*, 2020, **12**, 24737–24746.
- 122 X. Huang, S. Han, W. Huang and X. Liu, *Chem. Soc. Rev.*, 2012, **42**, 173–201.
- 123 S. Li, Z. Hou, Z. Cheng, H. Lian, P. Ma, C. Li and J. Lin, *RSC Adv.*, 2013, **3**, 5491–5497.
- 124 Z. Wang and A. Meijerink, *J. Phys. Chem. Lett.*, 2018, **9**, 1522–1526.
- 125 W. Shao, C.-K. Lim, Q. Li, M. T. Swihart and P. N. Prasad, *Nano Lett.*, 2018, **18**, 4922–4926.
- 126 P. Agbo, J. S. Kanady and R. J. Abergel, *Front. Chem.*, 2020, **8**, 579942.



- 127 Z. Ye, M. Tan, G. Wang and J. Yuan, *Anal. Chem.*, 2004, **76**, 513–518.
- 128 H. Härmä, T. Soukka and T. Lövgren, *Clin. Chem.*, 2001, **47**, 561–568.
- 129 J. Desbiens, B. Bergeron, M. Patry and A. M. Ritcey, *J. Colloid Interface Sci.*, 2012, **376**, 12–19.
- 130 F. Danhier, E. Ansorena, J. M. Silva, R. Coco, A. Le Breton and V. Préat, *J. Controlled Release*, 2012, **161**, 505–522.
- 131 A.-I. Moreno-Vega, T. Gómez-Quintero, R.-E. Nuñez-Anita, L.-S. Acosta-Torres and V. Castaño, *J. Nanotechnol.*, 2012, **2012**, 936041.
- 132 K. M. El-Say and H. S. El-Sawy, *Int. J. Pharm.*, 2017, **528**, 675–691.
- 133 A. Reisch and A. S. Klymchenko, *Small*, 2016, **12**, 1968–1992.
- 134 K. Li and B. Liu, *Chem. Soc. Rev.*, 2014, **43**, 6570–6597.
- 135 K. Tamaki and M. Shimomura, *Int. J. Nanosci.*, 2002, **01**, 533–537.
- 136 T. Soukka, H. Härmä, J. Paukkunen and T. Lövgren, *Anal. Chem.*, 2001, **73**, 2254–2260.
- 137 T. Soukka, J. Paukkunen, H. Härmä, S. Lönnberg, H. Lindroos and T. Lövgren, *Clin. Chem.*, 2001, **47**, 1269–1278.
- 138 T. Soukka, K. Anttonen, H. Härmä, A.-M. Pelkkikangas, P. Huhtinen and T. Lövgren, *Clin. Chim. Acta*, 2003, **328**, 45–58.
- 139 L. Kokko, K. Sandberg, T. Lövgren and T. Soukka, *Anal. Chim. Acta*, 2004, **503**, 155–162.
- 140 V. Väisänen, H. Härmä, H. Lilja and A. Bjartell, *Luminescence*, 2000, **15**, 389–397.
- 141 A. Valanne, J. Suojanen, J. Peltonen, T. Soukka, P. Hänninen and H. Härmä, *Analyst*, 2009, **134**, 980–986.
- 142 S. Pihlasalo, T. Deguchi, M. Virtamo, J. Jacobino, K. Chary, F. R. López-Picón, G. Brunhofer-Bolzer, R. Huttunen, A. Fallarero, P. Vuorela and H. Härmä, *Anal. Chem.*, 2017, **89**, 2398–2404.
- 143 S. Pihlasalo, J. Kirjavainen, P. Hänninen and H. Härmä, *Anal. Chem.*, 2011, **83**, 1163–1166.
- 144 P. Huhtinen, M. Kivelä, O. Kuronen, V. Hagren, H. Takalo, H. Tenhu, T. Lövgren and H. Härmä, *Anal. Chem.*, 2005, **77**, 2643–2648.
- 145 K. Tamaki, H. Yabu, T. Isoshima, M. Hara and M. Shimomura, *Colloids Surf., A*, 2006, **284–285**, 355–358.
- 146 G. Shao, R. Han, Y. Ma, M. Tang, F. Xue, Y. Sha and Y. Wang, *Chem. – Eur. J.*, 2010, **16**, 8647–8651.
- 147 T. Aikawa, A. Mizuno, M. Kohri, T. Taniguchi, K. Kishikawa and T. Nakahira, *Colloids Surf., B*, 2016, **145**, 152–159.
- 148 H. Tan, B. Liu and Y. Chen, *ACS Nano*, 2012, **6**, 10505–10511.
- 149 W. Sun, J. Yu, R. Deng, Y. Rong, B. Fujimoto, C. Wu, H. Zhang and D. T. Chiu, *Angew. Chem., Int. Ed.*, 2013, **52**, 11294–11297.
- 150 Q. Li, J. Zhang, W. Sun, J. Yu, C. Wu, W. Qin and D. T. Chiu, *Langmuir*, 2014, **30**, 8607–8614.
- 151 I. K. Voets, A. de Keizer and M. A. Cohen Stuart, *Adv. Colloid Interface Sci.*, 2009, **147–148**, 300–318.
- 152 J. Wang, A. H. Velders, E. Gianolio, S. Aime, F. J. Vergeldt, H. V. As, Y. Yan, M. Drechsler, A. de Keizer, M. A. C. Stuart and J. van der Gucht, *Chem. Commun.*, 2013, **49**, 3736–3738.
- 153 D. C. Thévenaz, C. A. Monnier, S. Balog and G. L. Fiore, *Biomacromolecules*, 2014, **15**, 3994–4001.
- 154 N. Wartenberg, O. Raccurt, D. Imbert, M. Mazzanti and E. Bourgeat-Lami, *J. Mater. Chem. C*, 2013, **1**, 2061–2068.
- 155 M. Cardoso Dos Santos, A. Runser, H. Bartenlian, A. M. Nonat, L. J. Charbonnière, A. S. Klymchenko, N. Hildebrandt and A. Reisch, *Chem. Mater.*, 2019, **31**, 4034–4041.
- 156 Y. Takei, S. Arai, A. Murata, M. Takabayashi, K. Oyama, S. Ishiwata, S. Takeoka and M. Suzuki, *ACS Nano*, 2014, **8**, 198–206.
- 157 W. Yang, L.-M. Fu, X. Wen, Y. Liu, Y. Tian, Y.-C. Liu, R.-C. Han, Z.-Y. Gao, T.-E. Wang, Y.-L. Sha, Y.-Q. Jiang, Y. Wang and J.-P. Zhang, *Biomaterials*, 2016, **100**, 152–161.
- 158 N. Gao, Y. Zhang, P. Huang, Z. Xiang, F.-Y. Wu and L. Mao, *Anal. Chem.*, 2018, **90**, 7004–7011.
- 159 T. Liao, F. Yuan, C. Shi, C.-X. He and Z. Li, *RSC Adv.*, 2016, **6**, 103463–103470.
- 160 Z. Chen, Z. Zhang, X. Zhai, Y. Li, L. Lin, H. Zhao, L. Bian, P. Li, L. Yu, Y. Wu and G. Lin, *Anal. Chem.*, 2020, **92**, 7226–7231.
- 161 L. B. Capeletti, L. M. D. Loiola, A. S. Picco, M. da Silva Liberato and M. B. Cardoso, in *Smart Nanoparticles for Biomedicine*, ed. G. Ciofani, Elsevier, 2018, pp. 115–129.
- 162 A. Bitar, N. M. Ahmad, H. Fessi and A. Elaissari, *Drug Discovery Today*, 2012, **17**, 1147–1154.
- 163 S. Bonacchi, D. Genovese, R. Juris, M. Montalti, L. Prodi, E. Rampazzo and N. Zaccheroni, *Angew. Chem., Int. Ed.*, 2011, **50**, 4056–4066.
- 164 G. Yao, L. Wang, Y. Wu, J. Smith, J. Xu, W. Zhao, E. Lee and W. Tan, *Anal. Bioanal. Chem.*, 2006, **385**, 518–524.
- 165 V. Gubala, G. Giovannini, F. Kunc, M. P. Monopoli and C. J. Moore, *Cancer Nanotechnol.*, 2020, **11**, 1.
- 166 M. Montalti, L. Prodi, E. Rampazzo and N. Zaccheroni, *Chem. Soc. Rev.*, 2014, **43**, 4243–4268.
- 167 P. C. R. Soares-Santos, H. I. S. Nogueira, V. Félix, M. G. B. Drew, R. A. Sá Ferreira, L. D. Carlos and T. Trindade, *Chem. Mater.*, 2003, **15**, 100–108.
- 168 W. Stöber, A. Fink and E. Bohn, *J. Colloid Interface Sci.*, 1968, **26**, 62–69.
- 169 D. Zhao, W. Qin, C. Wu, G. Qin, J. Zhang and S. Lü, *Chem. Phys. Lett.*, 2004, **388**, 400–405.
- 170 X. Hai, M. Tan, G. Wang, Z. Ye, J. Yuan and K. Matsumoto, *Anal. Sci.*, 2004, **20**, 245–246.
- 171 Z. Ye, M. Tan, G. Wang and J. Yuan, *J. Mater. Chem.*, 2004, **14**, 851.
- 172 M. Tan, Z. Ye, G. Wang and J. Yuan, *Chem. Mater.*, 2004, **16**, 2494–2498.
- 173 Z. Ye, M. Tan, G. Wang and J. Yuan, *Talanta*, 2005, **65**, 206–210.



- 174 Z. Ye, M. Tan, G. Wang and J. Yuan, *J. Fluoresc.*, 2005, **15**, 499–505.
- 175 Y. Chen, Y. Chi, H. Wen and Z. Lu, *Anal. Chem.*, 2007, **79**, 960–965.
- 176 Y. Chen and Z. Lu, *Anal. Chim. Acta*, 2007, **587**, 180–186.
- 177 J. Wu, G. Wang, D. Jin, J. Yuan, Y. Guan and J. Piper, *Chem. Commun.*, 2008, 365–367.
- 178 J. Wu, Z. Ye, G. Wang, D. Jin, J. Yuan, Y. Guan and J. Piper, *J. Mater. Chem.*, 2009, **19**, 1258–1264.
- 179 L. Tian, Z. Dai, L. Zhang, R. Zhang, Z. Ye, J. Wu, D. Jin and J. Yuan, *Nanoscale*, 2012, **4**, 3551–3557.
- 180 H. Zhang, Y. Xu, W. Yang and Q. Li, *Chem. Mater.*, 2007, **19**, 5875–5881.
- 181 K. O. Iwu, P. C. R. Soares-Santos, H. I. S. Nogueira, L. D. Carlos and T. Trindade, *J. Phys. Chem. C*, 2009, **113**, 7567–7573.
- 182 J. Samuel, G. Tallec, P. Cherns, W. L. Ling, O. Raccurt, O. Poncelet, D. Imbert and M. Mazzanti, *Chem. Commun.*, 2010, **46**, 2647–2649.
- 183 H. Jiang, G. Wang, W. Zhang, X. Liu, Z. Ye, D. Jin, J. Yuan and Z. Liu, *J. Fluoresc.*, 2010, **20**, 321–328.
- 184 N. Wartenberg, O. Raccurt, E. Bourgeat-Lami, D. Imbert and M. Mazzanti, *Eur. J. Inorg. Chem.*, 2013, **2013**, 1493–1498.
- 185 S. V. Eliseeva, B. Song, C. D. B. Vandevyver, A.-S. Chauvin, J. B. Wacker and J.-C. G. Bünzli, *New J. Chem.*, 2010, **34**, 2915–2921.
- 186 H. Peng, M. I. J. Stich, J. Yu, L. Sun, L. H. Fischer and O. S. Wolfbeis, *Adv. Mater.*, 2010, **22**, 716–719.
- 187 C. Philippot, A. Bourdolle, O. Maury, F. Dubois, B. Boury, S. Brustlein, S. Brasselet, C. Andraud and A. Ibanez, *J. Mater. Chem.*, 2011, **21**, 18613–18622.
- 188 Q.-X. Wang, S.-F. Xue, Z.-H. Chen, S.-H. Ma, S. Zhang, G. Shi and M. Zhang, *Biosens. Bioelectron.*, 2017, **94**, 388–393.
- 189 K. R. Kim, Y. D. Han, H. J. Chun, K. W. Lee, D.-K. Hong, K.-N. Lee and H. C. Yoon, *Biosensors*, 2017, **7**, 48.
- 190 H. Tan, Q. Li, C. Ma, Y. Song, F. Xu, S. Chen and L. Wang, *Biosens. Bioelectron.*, 2015, **63**, 566–571.
- 191 E. Fanizza, N. Depalo, S. Fedorenko, R. M. Iacobazzi, A. Mukhametshina, R. Zairov, A. Salatino, F. Vischio, A. Panniello, V. Laquintana, M. L. Curri, A. Mustafina, N. Denora and M. Striccoli, *Int. J. Mol. Sci.*, 2019, **20**, 3139.
- 192 S. Fedorenko, D. Gilmanova, A. Mukhametshina, I. Nizameev, K. Kholin, B. Akhmadeev, A. Voloshina, A. Sapunova, S. Kuznetsova, A. Daminova, S. Katsyuba, R. Zairov and A. Mustafina, *J. Mater. Sci.*, 2019, **54**, 9140–9154.
- 193 A. P. Alivisatos, *Science*, 1996, **271**, 933–937.
- 194 X. Peng, M. C. Schlamp, A. V. Kadavanich and A. P. Alivisatos, *J. Am. Chem. Soc.*, 1997, **119**, 7019–7029.
- 195 P. Reiss, J. Bleuse and A. Pron, *Nano Lett.*, 2002, **2**, 781–784.
- 196 H. Peng, C. Wu, Y. Jiang, S. Huang and J. McNeill, *Langmuir*, 2007, **23**, 1591–1595.
- 197 A. R. Kortan, R. Hull, R. L. Opila, M. G. Bawendi, M. L. Steigerwald, P. J. Carroll and L. E. Brus, *J. Am. Chem. Soc.*, 1990, **112**, 1327–1332.
- 198 X. Wu, H. Liu, J. Liu, K. N. Haley, J. A. Treadway, J. P. Larson, N. Ge, F. Peale and M. P. Bruchez, *Nat. Biotechnol.*, 2003, **21**, 41–46.
- 199 J. K. Jaiswal, H. Mattoussi, J. M. Mauro and S. M. Simon, *Nat. Biotechnol.*, 2003, **21**, 47–51.
- 200 N. N. Mamedova, N. A. Kotov, A. L. Rogach and J. Studer, *Nano Lett.*, 2001, **1**, 281–286.
- 201 I. Potapova, R. Mruk, C. Hübner, R. Zentel, T. Basché and A. Mews, *Angew. Chem., Int. Ed.*, 2005, **44**, 2437–2440.
- 202 L. Stryer and R. P. Haugland, *Proc. Natl. Acad. Sci. U. S. A.*, 1967, **58**, 719–726.
- 203 N. Hildebrandt, L. J. Charbonnière, M. Beck, R. F. Ziessel and H. G. Löhmannsröben, *Angew. Chem., Int. Ed.*, 2005, **44**, 7612–7615.
- 204 N. Hildebrandt, L. J. Charbonnière and H.-G. Löhmannsröben, *J. Biomed. Biotechnol.*, 2007, **2007**, 1–6.
- 205 L. J. Charbonnière, N. Hildebrandt, R. F. Ziessel and H. G. Löhmannsröben, *J. Am. Chem. Soc.*, 2006, **128**, 12800–12809.
- 206 N. Hildebrandt, K. D. Wegner and W. R. Algar, *Coord. Chem. Rev.*, 2014, **273274**, 125–138.
- 207 D. Geissler, L. J. Charbonnière, R. F. Ziessel, N. G. Butlin, H. G. Löhmannsröben and N. Hildebrandt, *Angew. Chem., Int. Ed.*, 2010, **49**, 1396–1401.
- 208 F. Morgner, D. Geißler, S. Stufner, N. G. Butlin, H. G. Löhmannsröben and N. Hildebrandt, *Angew. Chem., Int. Ed.*, 2010, **49**, 7570–7574.
- 209 D. Geissler, N. G. Butlin, D. Hill, H.-G. Löhmannsröben and N. Hildebrandt, in *Optics InfoBase Conference Papers*, ed. I. Georgakoudi, J. Popp and K. Svanberg, 2009, vol. 7368, p. 73680P.
- 210 I. Gryczynski, J. Malicka, Z. Gryczynski, J. R. Lakowicz and C. D. Geddes, *J. Fluoresc.*, 2002, **12**, 131–133.
- 211 D. Geissler, H.-G. Löhmannsröben, L. J. Charbonnière, R. F. Ziessel, N. G. Butlin, I. L. Medintz, H. Mattoussi and N. Hildebrandt, *Proc. SPIE 7575, Colloidal Quantum Dots for Biomedical Applications V*, 2010, 75750E.
- 212 K. D. Wegner, F. Morgner, E. Oh, R. Goswami, K. Susumu, M. H. Stewart, I. L. Medintz and N. Hildebrandt, *Chem. Mater.*, 2014, **26**, 4299–4312.
- 213 W. R. Algar, A. P. Malanoski, K. Susumu, M. H. Stewart, N. Hildebrandt and I. L. Medintz, *Anal. Chem.*, 2012, **84**, 10136–10146.
- 214 W. R. Algar, D. Wegner, A. L. Huston, J. B. Blanco-Canosa, M. H. Stewart, A. Armstrong, P. E. Dawson, N. Hildebrandt and I. L. Medintz, *J. Am. Chem. Soc.*, 2012, **134**, 1876–1891.
- 215 J. Xu, X. Qiu and N. Hildebrandt, *Nano Lett.*, 2021, **21**, 4802–4808.
- 216 S. A. Díaz, G. Lasarte Aragonés, S. Buckhout-White, X. Qiu, E. Oh, K. Susumu, J. S. Melinger, A. L. Huston, N. Hildebrandt and I. L. Medintz, *J. Phys. Chem. Lett.*, 2017, **8**, 2182–2188.
- 217 F. Morgner, S. Stufner, D. Geißler, I. L. Medintz, W. R. Algar, K. Susumu, M. H. Stewart, J. B. Blanco-



- Canosa, P. E. Dawson and N. Hildebrandt, *Sensors*, 2011, **11**, 9667–9684.
- 218 T. Hallaj, M. Amjadi, X. Qiu, K. Susumu, I. L. Medintz and N. Hildebrandt, *Nanoscale*, 2020, **12**, 13719–13730.
- 219 C. Léger, A. Yahia-Ammar, K. Susumu, I. L. Medintz, A. Urvoas, M. Valerio-Lepiniec, P. Minard and N. Hildebrandt, *ACS Nano*, 2020, **14**, 5956–5967.
- 220 L. Mattera, S. Bhuckory, K. D. Wegner, X. Qiu, F. Agnese, C. Lincheneau, T. Senden, D. Djurado, L. J. Charbonnière, N. Hildebrandt and P. Reiss, *Nanoscale*, 2016, **8**, 11275–11283.
- 221 S. Bhuckory, L. Mattera, K. D. Wegner, X. Qiu, Y.-T. Wu, L. J. Charbonnière, P. Reiss and N. Hildebrandt, *Chem. Commun.*, 2016, **52**, 14423–14425.
- 222 H. Härmä, T. Soukka, A. Shavel, N. Gaponik and H. Weller, *Anal. Chim. Acta*, 2007, **604**, 177–183.
- 223 G. J. Stasiuk, S. Tamang, D. Imbert, C. Poillot, M. Giardiello, C. Tisseyre, E. L. Barbier, P. H. Fries, M. De Waard, P. Reiss and M. Mazzanti, *ACS Nano*, 2011, **5**, 8193–8201.
- 224 J. K. Molloy, C. Lincheneau, M. M. Karimdjy, F. Agnese, L. Mattera, C. Gateau, P. Reiss, D. Imbert and M. Mazzanti, *Chem. Commun.*, 2016, **52**, 4577–4580.
- 225 R. A. Tigaa, G. J. Lucas and A. De Bettencourt-Dias, *Inorg. Chem.*, 2017, **56**, 3260–3268.
- 226 F. Wu, L. Yue, L. Yang, K. Wang, G. Liu, X. Luo and X. Zhu, *Colloids Surf., B*, 2019, **175**, 272–280.
- 227 H. Su, Y. Liao, F. Wu, X. Sun, H. Liu, K. Wang and X. Zhu, *Colloids Surf., B*, 2018, **170**, 194–200.
- 228 S. Cousinié, M. Gressier, C. Reber, J. Dexpert-Ghys and M. J. Menu, *Langmuir*, 2008, **24**, 6208–6214.
- 229 L. J. Charbonnière, N. Weibel, C. Estournes, C. Leuvrey and R. Ziessel, *New J. Chem.*, 2004, **28**, 777–781.
- 230 E. DeOliveira, C. R. Neri, O. A. Serra and A. G. S. Prado, *Chem. Mater.*, 2007, **19**, 5437–5442.
- 231 L. Lu, F. Liu, G. Sun, H. Zhang, S. Xi and H. Wang, *J. Mater. Chem.*, 2004, **14**, 2760–2762.
- 232 Y. Xu and Q. Li, *Clin. Chem.*, 2007, **53**, 1503–1510.
- 233 B. Yan and B. Zhou, *J. Photochem. Photobiol., A*, 2008, **195**, 314–322.
- 234 Y. Li and B. Yan, *J. Solid State Chem.*, 2008, **181**, 1032–1039.
- 235 D. B. Ambili Raj, S. Biju and M. L. P. Reddy, *J. Mater. Chem.*, 2009, **19**, 7976–7983.
- 236 V. Divya, S. Biju, R. L. Varma and M. L. P. Reddy, *J. Mater. Chem.*, 2010, **20**, 5220–5227.
- 237 D. Zhang, X. Wang, Z. A. Qiao, D. Tang, Y. Liu and Q. Huo, *J. Phys. Chem. C*, 2010, **114**, 12505–12510.
- 238 W. X. Li, Y. S. Zheng, X. F. Cao, J. Bai, Z. F. Fu, J. R. Bao and Y. L. Li, *J. Lumin.*, 2016, **178**, 470–478.
- 239 Z. F. Fu, W. X. Li, J. Bai, J. R. Bao, X. F. Cao and Y. S. Zheng, *Luminescence*, 2017, **32**, 327–333.
- 240 Y. Y. Ma, H. X. Huang, M. Chen and D. J. Qian, *J. Lumin.*, 2018, **203**, 277–285.
- 241 A. P. Duarte, M. Gressier, M. J. Menu, J. Dexpert-Ghys, J. M. A. Caiut and S. J. L. Ribeiro, *J. Phys. Chem. C*, 2012, **116**, 505–515.
- 242 A. P. Duarte, L. Mauline, M. Gressier, J. Dexpert-Ghys, C. Roques, J. M. A. Caiut, E. Deffune, D. C. G. Maia, I. Z. Carlos, A. A. P. Ferreira, S. J. L. Ribeiro and M. J. Menu, *Langmuir*, 2013, **29**, 5878–5888.
- 243 K. M. L. Taylor and W. Lin, *J. Mater. Chem.*, 2009, **19**, 6418–6422.
- 244 S. Mornet, S. Vasseur, F. Grasset, P. Veverka, G. Goglio, A. Demourgues, J. Portier, E. Pollert and E. Duguet, *Prog. Solid State Chem.*, 2006, **34**, 237–247.
- 245 Y. Jun, J. Choi and J. Cheon, *Chem. Commun.*, 2007, 1203–1214.
- 246 D. Patel, J. Y. Moon, Y. Chang, T. J. Kim and G. H. Lee, *Colloids Surf., A*, 2008, **313–314**, 91–94.
- 247 N. Lewinski, V. Colvin and R. Drezek, *Small*, 2008, **4**, 26–49.
- 248 W. Wu, Q. He and C. Jiang, *Nanoscale Res. Lett.*, 2008, **3**, 397–415.
- 249 M. Ma, Y. Zhang, W. Yu, H. Y. Shen, H. Q. Zhang and N. Gu, *Colloids Surf., A*, 2003, **212**, 219–226.
- 250 X. Shen, X. Fang, Y. Zhou and H. Liang, *Chem. Lett.*, 2004, **33**, 1468–1469.
- 251 J. Choi, J. C. Kim, Y. B. Lee, I. S. Kim, Y. K. Park and N. H. Hur, *Chem. Commun.*, 2007, 1644.
- 252 S. Y. Yu, H. J. Zhang, J. B. Yu, C. Wang, L. N. Sun and W. D. Shi, *Langmuir*, 2007, **23**, 7836–7840.
- 253 H. M. Lu, W. T. Zheng and Q. Jiang, *J. Phys. D: Appl. Phys.*, 2007, **40**, 320–325.
- 254 J. Wu, Z. Ye, G. Wang and J. Yuan, *Talanta*, 2007, **72**, 1693–1697.
- 255 H. Lu, G. Yi, S. Zhao, D. Chen, L. Guo and J. Cheng, *J. Mater. Chem.*, 2004, **14**, 1336.
- 256 B. Wang, J. Hai, Q. Wang, T. Li and Z. Yang, *Angew. Chem., Int. Ed.*, 2011, **50**, 3063–3066.
- 257 Y. Liu, G. Cheng, Z. Wang, J. Zhang, D. Sun, G. Hong and J. Ni, *Mater. Lett.*, 2013, **97**, 187–190.
- 258 L. U. Khan, H. F. Brito, J. Hölsä, K. R. Pirota, D. Muraca, M. C. F. C. Felinto, E. E. S. Teotonio and O. L. Malta, *Inorg. Chem.*, 2014, **53**, 12902–12910.
- 259 R. R. Silva, A. P. Duarte, R. M. Sábio, J. M. A. Caiut, M. Gressier, M. J. Menu, A. Franco and S. J. L. Ribeiro, *ChemistrySelect*, 2016, **1**, 5923–5928.
- 260 S. L. C. Pinho, J. Sereno, A. J. Abrunhosa, M.-H. Delville, J. Rocha, L. D. Carlos and C. F. G. C. Geraldès, *Inorg. Chem.*, 2019, **58**, 16618–16628.
- 261 S. Goderski, S. Kanno, K. Yoshihara, H. Komiya, K. Goto, T. Tanaka, S. Kawaguchi, A. Ishii, J. Shimoyama, M. Hasegawa and S. Lis, *ACS Omega*, 2020, **5**, 32930–32938.
- 262 A. de Espindola, C. S. Chagas, E. Barbosa, C. E. Castro, F. L. A. Fonseca, P. S. Haddad and C. Molina, *J. Magn. Magn. Mater.*, 2022, **545**, 168751.
- 263 N. L. Rosi, D. A. Giljohann, C. S. Thaxton, A. K. R. Lytton-Jean, M. S. Han and C. A. Mirkin, *Science*, 2006, **312**, 1027–1030.
- 264 Y. Zhao, Y. Tian, Y. Cui, W. Liu, W. Ma and X. Jiang, *J. Am. Chem. Soc.*, 2010, **132**, 12349–12356.



- 265 B. I. Ipe, K. Yoosaf and K. G. Thomas, *J. Am. Chem. Soc.*, 2006, **128**, 1907–1913.
- 266 D. J. Lewis, T. M. Day, J. V. MacPherson and Z. Pikramenou, *Chem. Commun.*, 2006, 1433–1435.
- 267 L. Prodi, M. Montalti, N. Zaccheroni, G. Pickaert, L. Charbonnière and R. Ziessel, *New J. Chem.*, 2003, **27**, 134–139.
- 268 A. Davies, D. J. Lewis, S. P. Watson, S. G. Thomas and Z. Pikramenou, *Proc. Natl. Acad. Sci. U. S. A.*, 2012, **109**, 1862–1867.
- 269 J. Massue, S. J. Quinn and T. Gunnlaugsson, *J. Am. Chem. Soc.*, 2008, **130**, 6900–6901.
- 270 S. Comby and T. Gunnlaugsson, *ACS Nano*, 2011, **5**, 7184–7197.
- 271 E. M. Surender, S. Comby, B. L. Cavanagh, O. Brennan, T. C. Lee and T. Gunnlaugsson, *Chem*, 2016, **1**, 438–455.
- 272 C. Chen, C. Midelet, S. Bhuckory, N. Hildebrandt and M. H. V. Werts, *J. Phys. Chem. C*, 2018, **122**, 17566–17574.
- 273 A. Samanta, Y. Zhou, S. Zou, H. Yan and Y. Liu, *Nano Lett.*, 2014, **14**, 5052–5057.
- 274 Y. Sun, H. Jiu, D. Zhang, J. Gao, B. Guo and Q. Zhang, *Chem. Phys. Lett.*, 2005, **410**, 204–208.
- 275 S. Wu, Y. Sun, X. Wang, W. Wu, X. Tian, Q. Yan, Y. Luo and Q. Zhang, *J. Photochem. Photobiol., A*, 2007, **191**, 97–103.
- 276 J. Kang, Y. Li, Y. Chen, A. Wang, B. Yue, Y. Qu, Y. Zhao and H. Chu, *Mater. Res. Bull.*, 2015, **71**, 116–121.
- 277 Y. Zhao, A. Wang, J. Kang, H. Chu, H. Zhang and Y. Zhao, *J. Photochem. Photobiol., A*, 2020, **400**, 112678.
- 278 E. E. Jelley, *Nature*, 1936, **138**, 1009–1010.
- 279 H. Fukumoto and Y. Yonezawa, *Thin Solid Films*, 1998, **327–329**, 748–751.
- 280 J. Kang, Y. Zhao, H. Chu and Y. Zhao, *J. Photochem. Photobiol., A*, 2018, **365**, 119–124.
- 281 Y. Mei and B. Yan, *Inorg. Chem. Commun.*, 2014, **40**, 39–42.
- 282 W. Mnasri, M. Parvizian and S. Ammar-Merah, *Nanomaterials*, 2021, **11**, 354.
- 283 W. T. Carnall, in *Handbook on the Physics and Chemistry of Rare Earths*, Elsevier, 1979, vol. 3, pp. 171–208.
- 284 C. Lin, Z. Xia, L. Zhang, X. Chen, Q. Sun, M. Lu, Z. Yuan, X. Xie and L. Huang, *ACS Appl. Mater. Interfaces*, 2020, **12**, 31783–31792.
- 285 S. Han, R. Deng, Q. Gu, L. Ni, U. Huynh, J. Zhang, Z. Yi, B. Zhao, H. Tamura, A. Pershin, H. Xu, Z. Huang, S. Ahmad, M. Abdi-Jalebi, A. Sadhanala, M. L. Tang, A. Bakulin, D. Beljonne, X. Liu and A. Rao, *Nature*, 2020, **587**, 594–599.
- 286 F. Mousseau, C. F. Tarrisse, S. Simon, T. Gacoin, A. Alexandrou and C. I. Bouzigues, *Nanoscale*, 2021, **13**, 14814–14824.
- 287 S. Zhou, W. Zheng, Z. Chen, D. Tu, Y. Liu, E. Ma, R. Li, H. Zhu, M. Huang and X. Chen, *Angew. Chem.*, 2014, **126**, 12706–12710.
- 288 L. Francés-Soriano, N. Hildebrandt and L. J. Charbonnière, in *Reference Module in Chemistry, Molecular Sciences and Chemical Engineering*, Elsevier, 2022. <https://doi.org/10.1016/B978-0-12-823144-9.00095-9>.

