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Synthesis and evaluation of isothiazolo[4,5-*b*]pyridines as cyclin G-associated kinase (GAK) inhibitors†

Yulia Ivanova,^a Sander Spittaels,^a Ling-Jie Gao,^b Dominique Schols,^c Luc Van Meervelt,^d Mathy Froeyen,^b Wim Dehaen^a and Steven De Jonghe^{*c}

Isothiazolo[4,3-*b*]pyridines have been extensively explored as inhibitors of cyclin G-associated kinase (GAK). In order to expand the structure–activity relationship study and to discover other chemotypes that act as GAK inhibitors, the closely related isothiazolo[4,5-*b*]pyridine scaffold was explored. An easy and efficient synthetic procedure to access 3,5- and 3,6-dihalogenated isothiazolo[4,5-*b*]pyridines as key building blocks was developed. Regioselective functionalization with various substituents was performed. None of the newly synthesized isothiazolo[4,5-*b*]pyridines were active as GAK inhibitors. Molecular modeling was applied to rationalise their inactivity as GAK binders.

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Introduction

Cyclin G-associated kinase (GAK), also known as auxilin II, is a serine-threonine protein kinase that belongs to the Numb-associated kinase (NAK) family.¹ GAK plays a crucial role in clathrin-mediated endocytosis (CME) *via* phosphorylation of Thr144 and Thr156 of adaptor protein complexes 1 and 2 (AP1 and AP2).² This phosphorylation is necessary for the recruitment of clathrin and AP-2 to the plasma membrane and is essential for the formation of coated vesicles during CME.^{3–6} In a later stage of CME, GAK is involved in the uncoating of these vesicles, allowing the recycling of clathrin back to the cell surface and finalising the cycle.⁷

The GAK-regulated clathrin-mediated intracellular trafficking of cellular cargo proteins is a host process that is frequently hijacked by multiple viruses. It has been shown that GAK is an essential factor in the early and late stages of the lifecycle of various, unrelated viruses, such as the hepatitis C virus (HCV),⁸ the dengue virus (DENV),⁹ the Ebola virus

(EBOV),⁹ and more recently the severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2).¹⁰ A preclinical proof-of-concept was achieved using erlotinib which is a potent, albeit non-selective GAK inhibitor, in DENV and EBOV murine models leading to reduced viremia, morbidity, and mortality.⁹ Overall, these data validate GAK inhibition as a promising strategy for the development of broad-spectrum antiviral agents. Besides the interest in GAK as a host target for antiviral therapy, GAK is also involved in various other human pathologies, ranging from neurodegenerative disorders (*e.g.* Parkinson's disease)¹¹ and various cancers (such as osteosarcoma¹² and prostate cancer¹³).

In the past years, our group extensively explored isothiazolo[4,3-*b*]pyridine chemistry as a central scaffold for the discovery of potent and selective GAK inhibitors as broad-spectrum antiviral agents. Previous structure–activity relationship (SAR) studies were mainly focused on modifications of the substituents at positions 3, 5 and 6 of the isothiazolo[4,3-*b*]pyridine core (Fig. 1).^{14–20} The first isothiazolo[4,3-*b*]pyridine-based GAK inhibitor (compound 1) possessed a substituted phenyl moiety at position 6 and a morpholine at position 3 and displayed high GAK affinity (IC₅₀ = 51 nM). Later on, it was shown that the insertion of a hydrogen bond acceptor moiety on the substituent at position 3 (*e.g.* compound 2) led to an improved potency (IC₅₀ = 14 nM). Switching the aryl substituent on the pyridine core from position 6 to position 5 resulted in compound 3 with an equally high GAK affinity (IC₅₀ = 24 nM). Moreover, these isothiazolo[4,3-*b*]pyridine based GAK inhibitors exhibited *in vitro* antiviral activity against HCV, DENV, and EBOV, in the low micromolar range, while lacking cytotoxicity.

Besides modification of the substitution pattern, a scaffold hopping strategy has also been applied, while keeping the sub-

^aKU Leuven, Department of Chemistry, Sustainable Chemistry for Metals and Molecules, Celestijnenlaan 200F, B-3001 Leuven, Belgium

^bKU Leuven, Department of Pharmaceutical and Pharmacological Sciences, Rega Institute for Medical Research, Laboratory of Medicinal Chemistry, Herestraat 49, box 1030, 3000 Leuven, Belgium

^cKU Leuven, Department of Microbiology, Immunology and Transplantation, Rega Institute for Medical Research, Laboratory of Virology and Chemotherapy, Herestraat 49, box 1043, 3000 Leuven, Belgium. E-mail: steven.dejonghe@kuleuven.be

^dKU Leuven, Department of Chemistry, Biomolecular Architecture, Celestijnenlaan 200F, B-3001 Leuven, Belgium

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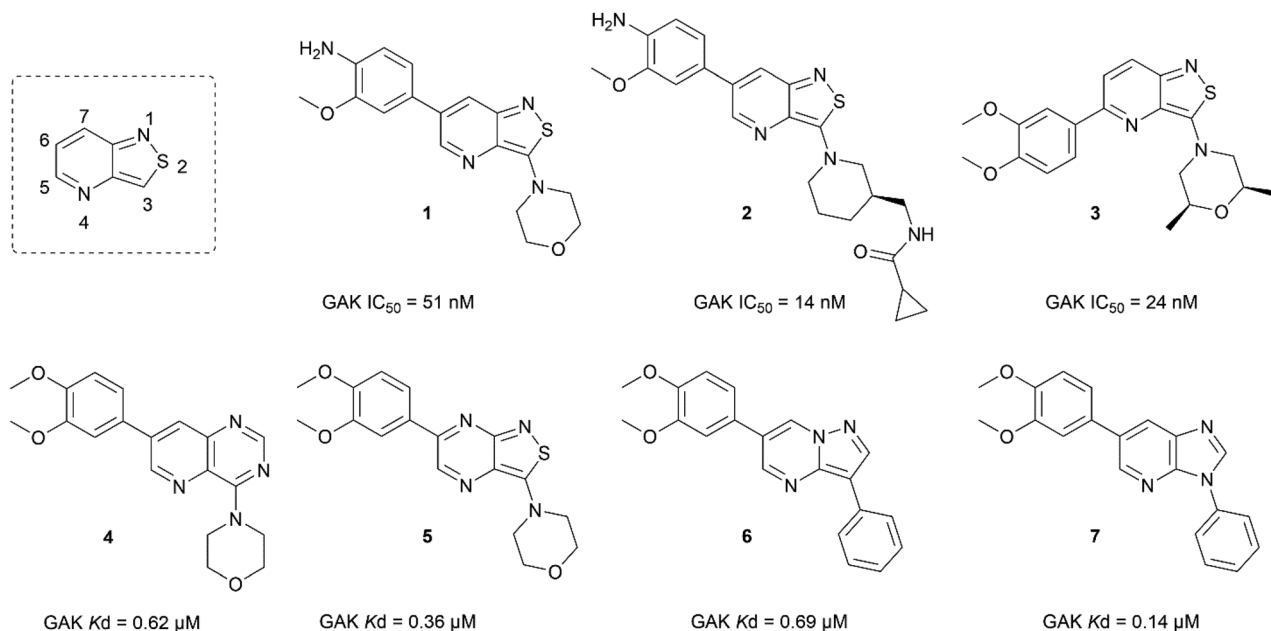


Fig. 1 Previously reported GAK inhibitors.

stitution pattern unchanged.¹⁷ A series of 13 novel scaffolds (including 6–5 and 6–6 bicyclic scaffolds) were prepared, from which 4 core structures displayed GAK affinity with dissociation constants (K_d values) below 1 μM (Fig. 1), including the quinazoline analogue **4**, the isothiazolo[3,4-*b*]pyrazine derivative **5**, the pyrazolo[1,5-*a*]pyrimidine **6** and imidazo[4,5-*b*]pyridine **7**.

Despite this scaffold hopping approach demonstrating the potential of altering the 5-membered cycle, derivatives of the closely related isothiazolo[4,5-*b*]pyridine core have not been studied as potential GAK inhibitors. This structure results from a simple swap of the nitrogen and sulfur yielding a scaffold which is of similar size and shape to the parent isothiazolo[4,3-*b*]pyridine lead molecule (Fig. 2). In addition, a SciFinderTM search revealed that isothiazolo[4,5-*b*]pyridines are not very well studied in organic and medicinal chemistry, with only 889 compounds known based on this skeleton (accessed

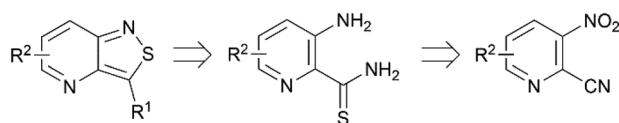
on May 2024). Hence, although the isothiazolo[4,5-*b*]pyridine scaffold is not completely novel, it is definitively less explored compared to other privileged scaffolds in medicinal chemistry, such as purines (661K compounds known, SciFinderTM, accessed on May 2024) or pyrrolo[2,3-*b*]pyridines (358K compounds known, SciFinderTM, accessed on May 2024). Therefore, in this study, a synthetic approach towards isothiazolo[4,5-*b*]pyridines is developed. A variety of groups was introduced onto the scaffold, including the substitution pattern of the known isothiazolo[4,3-*b*]pyridine based GAK inhibitors, since this allows for a side-by-side comparison of both series and for an evaluation of the importance of the central scaffold for GAK affinity.

Results and discussion

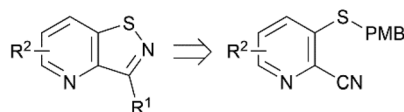
Chemistry

Potent GAK inhibitors based on a isothiazolo[4,3-*b*]pyridine scaffold bear an aryl residue (*e.g.* 3,4-dimethoxyphenyl) on the pyridine ring, whereas position 3 of the isothiazolo[4,3-*b*]pyridine scaffold tolerates more structural variety for GAK affinity. In contrast to isothiazolo[4,3-*b*]pyridines, which can be accessed from a 3-aminopyridine-2-carbothioamide analogue (derived from a suitable 3-nitropicolonitrile derivative) by treatment with an oxidising agent to construct the isothiazole moiety, isothiazolo[4,5-*b*]pyridines require an appropriate 3-mercaptopycolonitrile analogue that will then undergo oxidative ring closure to form the isothiazole ring (Fig. 2). Compound **12** was, thus, considered a key intermediate since it easily allows for structural modifications at position 3. Therefore, the 3,4-dimethoxyphenyl functionality was introduced as the first step (Scheme 1). A Suzuki coupling between

Previous work: isothiazolo[4,3-*b*]pyridine



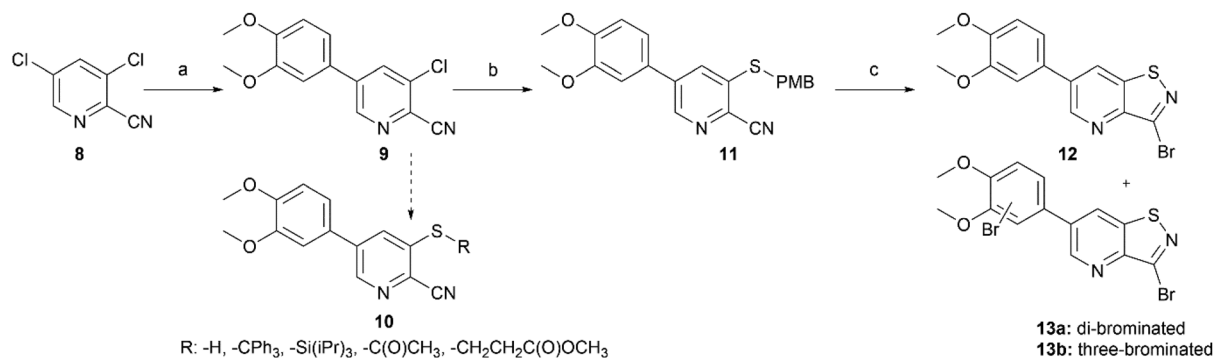
This work: isothiazolo[4,5-*b*]pyridine



R^1 : N-, S-, O-substituents, substituted phenyl; R^2 : substituted phenyl, alkynyl; PMB: *p*-methoxybenzyl

Fig. 2 Medicinal chemistry strategy.





Scheme 1 Initial synthetic approach towards isothiazolo[4,5-*b*]pyridines. Reagents and conditions: (a) 3,4-dimethoxyphenylboronic acid, Pd(PPh₃)₄, K₂CO₃, THF/H₂O, 100 °C, 72%; (b) 4-methoxybenzylthiol, NaH, DMF, rt, 81%; (c) Br₂, EtOAc, rt or reflux.

the commercially available 3,5-dichloropicolinonitrile **8** and 3,4-dimethoxyphenylboronic acid, catalysed by Pd(PPh₃)₄, using K₂CO₃ as the base in a THF/H₂O solvent mixture proceeds preferentially at position 5, resulting in compound **9**, along with some minor amounts of the disubstituted side-product. The correct regiochemistry of compound **9** was elucidated *via* a 2D NMR NOESY experiment (see ESI†). The presence of clear NOE correlations between the proton at position 6 of the pyridine ring and both protons at the *ortho* position of the dimethoxyphenyl moiety confirmed that the 3,4-dimethoxyphenyl group was attached to position 5 of the pyridine scaffold.

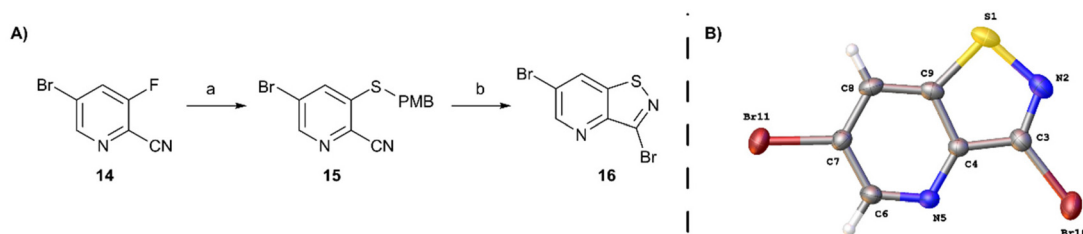
For the introduction of sulfur onto the pyridine scaffold, various reagents were explored to obtain the compound with the general structure **10**. Applying sodium sulfide or protected thiol reagents (such as triphenylmethanethiol, triisopropylsilanethiol and potassium thioacetate) only afforded messy reaction mixtures containing mainly side products. Although the reaction of pyridine **9** with sodium methanethiolate afforded the desired derivative, the subsequent cleavage of the methyl group under acidic conditions (HCl or HBr) was troublesome. Treatment of **9** with methyl 3-mercaptopropionate and sodium methoxide yielded the thioether, however, the desired compound was not isolated pure due to a low-yielding reaction. Gratifyingly, the reaction of **9** with *p*-methoxybenzylthiol at room temperature yielded (*p*-methoxybenzyl)thio-picolinonitrile **11** in good yield. The assembly of fused isothiazoles and isothiazolones by oxidative S–N bond formation is a known process.²¹ The synthesis of fused isothiazoles from the free thiol and neighbouring nitrile in the bromine atmosphere has been reported in several patents, however in rather unsatisfactory yields below 50%.^{22–24} In addition, these procedures start from an isolated free thiol, that is generated *via* acidic cleavage of a *S*-protected precursor, adding one more step to the synthetic sequence.²⁵ Since *p*-methoxybenzyl (PMB) deprotection can be effected under oxidative conditions,²⁶ we reasoned that both reactions (*i.e.* PMB cleavage and isothiazole formation) could be combined to deliver the isothiazolo[4,5-*b*]pyridine **12** by treatment of picolinonitrile **11** with bromine in a single step. The benefit of this procedure is the formation of

the free thiol *in situ* and its immediate further conversion, avoiding dimer formation. However, bromination of the electron-rich 3,4-dimethoxyphenyl moiety hampered the isolation of the desired product **12**. Mass spectral analysis of the crude reaction mixtures indicated that besides the desired compound **12**, di- and three-brominated side products (**13a** and **13b**, respectively) were formed when performing the reaction at reflux temperature, whereas the formation of a mono-brominated compound was observed at room temperature. To minimise bromination of the 3,4-dimethoxyphenyl moiety, various reaction circumstances (such as reducing the number of equivalents of bromine, decreasing the reaction temperature, or the slow and dropwise addition of bromine) were attempted. Unfortunately, none of these conditions was able to avoid over-bromination, preventing the isolation of compound **12** as a pure substance.

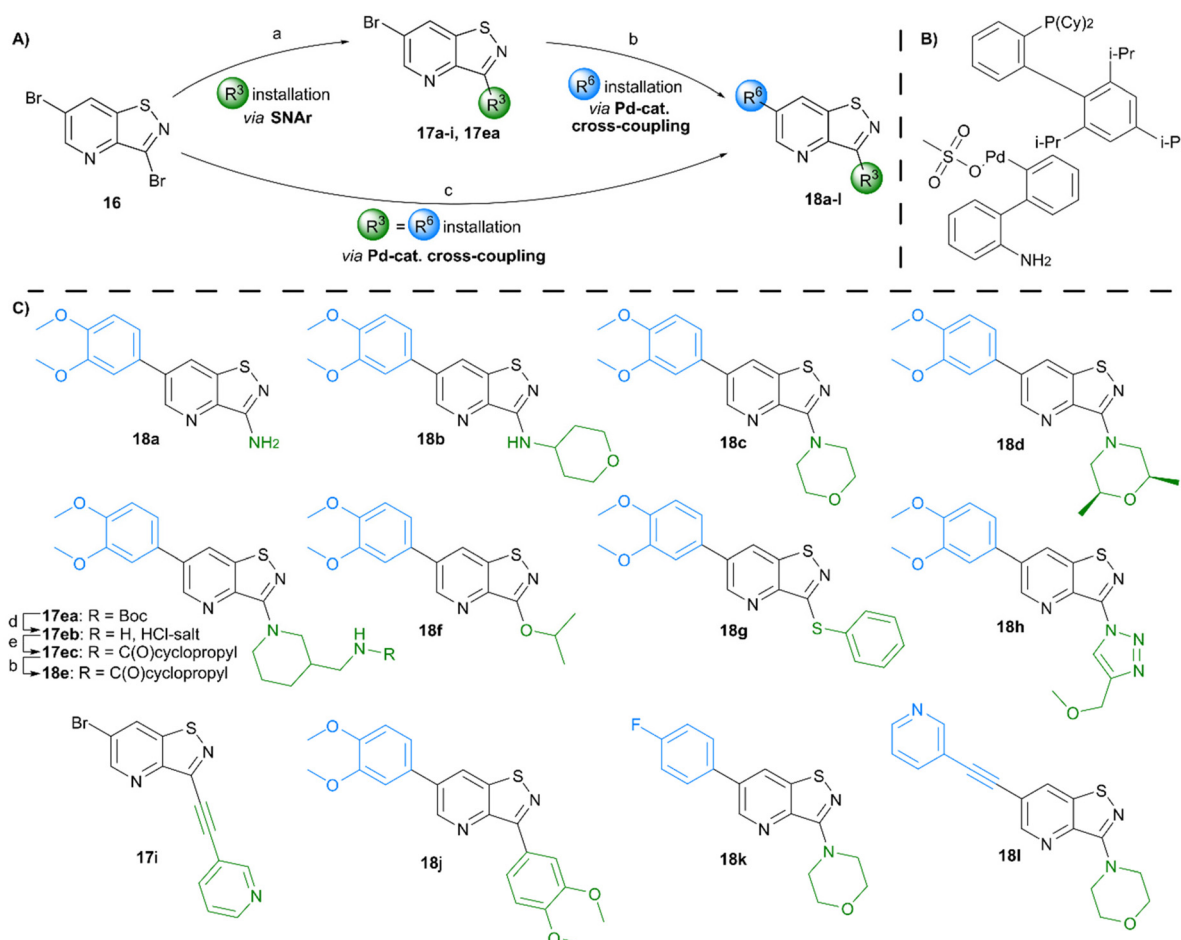
In order to avoid bromination of the dimethoxyphenyl residue, an alternative strategy was developed (Scheme 2). Commercially available 5-bromo-3-fluoropicolinonitrile **14** was chosen as the starting material to ensure selectivity during the nucleophilic aromatic substitution. Treatment of **14** with *p*-methoxybenzylthiol using K₂CO₃ as base afforded compound **15** in good yield. The following oxidative cyclisation with bromine furnished the 3,6-dibromo-isothiazolo[4,5-*b*]pyridine **16** in a single step in high yield, offering convenient access to the previously not reported key intermediate. The structure of **16** was confirmed by single-crystal X-ray crystallography (CCDC 2357539† and Scheme 2B).

Various groups were regioselectively introduced at position 3 of the isothiazolo[4,5-*b*]pyridine scaffold (Scheme 3). A nucleophilic aromatic substitution of 3,6-dibromoisothiazolo[4,5-*b*]pyridine **16** with different *N*-nucleophiles yielded compounds **17a–d** and **17ea**. The 3-amino congener **17a** was obtained in a lower 62% yield due to its insoluble nature, hampering its purification. Reactions resulting in secondary (**17b**), and tertiary (**17c–d** and **17ea**) amines proceeded smoothly with yields of 80% and 92–97%, respectively. Acidic cleavage of the Boc-protecting group of **17ea** resulted in the intermediate **17eb**, which was then followed by coupling with cyclopropanecarbonyl chloride, affording compound **17ec** in good yield.





Scheme 2 Two-step procedure towards 3,6-dibromo-isothiazolo[4,5-*b*]pyridine. Reagents and conditions: (A) (a) *p*-methoxybenzylthiol, K_2CO_3 , DMA, 0 °C, 85%; (b) Br_2 , EtOAc, 80 °C, 88%. (B) The molecular structure of **16**, showing the atom labeling and displacement ellipsoids drawn at the 30% probability level.



Scheme 3 Synthesis of 3,6-disubstituted isothiazolo[4,5-*b*]pyridines. Reagents and conditions: (A) (a) for **17a–ea**: *N*-nucleophile, EtOH or DMF, rt to 78 °C, 62–97%; for **17f**: *i*-PrOH, NaH, DMF, 50 °C, 18%; for **17g**: thiophenol, DMF, 50 °C, 53%; for **17h**: (i) NaN_3 , DMF, 0 °C, (ii) methyl propargyl ether, $CuSO_4 \cdot 5H_2O$, sodium ascorbate, DMSO/ H_2O , rt, 28%; for **17i**: 3-ethynylpyridine, LiHMDS, THF, 0 °C, 80%; (b) for Suzuki coupling: 3,4-dimethoxyphenylboronic acid or 4-fluorophenylboronic acid, $Pd(PPh_3)_4$ or XPhos $Pd\ G3$, K_2CO_3 or Cs_2CO_3 , dioxane/ H_2O , 80 °C or 100 °C, 51–87%; for Sonogashira coupling: 3-ethynylpyridine, $Pd(PPh_3)_2Cl_2$, Et_3N , DMF, 100 °C, 71%; (c) 3,4-dimethoxyphenylboronic acid, XPhos $Pd\ G3$, Cs_2CO_3 , dioxane/ H_2O , 80 °C, 48%; (d) HCl/dioxane 4 M, 0 °C to rt, quant.; (e) cyclopropanecarbonyl chloride, pyridine, DCM, 0 °C to rt, 82%. (B) Structure of XPhos $Pd\ G3$ catalyst. (C) Reaction scope.

The scope was extended to oxygen- and sulfur-containing nucleophiles to obtain corresponding compounds **17f** and **17g**. To access the 3-isopropoxy analogue, sodium hydride was used as a base, while the reactions with *S*- and *N*-nucleophiles proceeded without any additional base. Interestingly, position 3

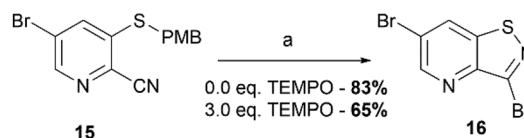
was also susceptible to substitution with sodium azide, resulting in the formation of the 3-azido intermediate, which was directly used in a Cu(I)-catalyzed azide–alkyne cycloaddition affording the 1,4-disubstituted triazole analogue **17h** in an overall yield of 28%. Using 3-ethynylpyridine and lithium bis



(trimethylsilyl)amide (LiHMDS) as strong base allowed to isolate compound **17i**. Position 3 was also susceptible to carbon–carbon bond formation. A Suzuki reaction allowed to install a 3,4-dimethoxyphenyl substituent simultaneously at positions 3 and 6, yielding compound **18j**. To furnish the fully decorated scaffold, Pd-catalysed reactions were performed on intermediates **17a–i**. A Suzuki procedure, utilizing tetrakis(triphenylphosphine)palladium, 3,4-dimethoxyphenylboronic acid and potassium carbonate, was used to prepare the final compounds **18a–h** in fair to excellent yields (51–87%). Reaction with (4-fluorophenyl)boronic acid proceeded equally well, producing compound **18k** in 85%. When a Suzuki coupling was applied to 3-(pyridin-3-ylethynyl)isothiazolo[4,5-*b*]pyridine **17i**, this only resulted in the degradation of the triple bond, most likely due to Pd complexation, and no desired compound was isolated. The bromine at position 6 was also susceptible to the Sonogashira coupling, affording compound **18l** in 71% yield. The correct regiochemistry was elucidated *via* a 2D NMR NOESY experiment, taking compound **18c** as a model substrate (see ESI†).

For the synthesis of 3,5-disubstituted isothiazolo[4,3-*b*]pyridines, an alternative synthetic scheme was established (Scheme 4). Commercially available 6-chloro-3-nitropicolonitrile **19** was reacted with *p*-methoxybenzylthiol at -45°C . This lower temperature allows for the selective substitution of the nitro group at position 3, leaving the chlorine at position 6 intact. The subsequent oxidative cyclisation was performed at 0°C to avoid halogen exchange, yielding the 3-bromo-5-chloroisothiazolo[4,5-*b*]pyridine **21** in good yield. The reaction of key intermediate **21** with morpholine and *cis*-2,6-dimethylmorpholine gave intermediates **22a** and **22b**, in yields of 57 and 71%, respectively. The lower yields, in comparison with the synthesis of **17c–d** can be rationalised by the reduced selectivity, since position 5 of key intermediate **21** is activated by the presence of the neighbouring nitrogen. The final step consisted of a Suzuki coupling, which was performed using XPhos Pd G3 as a third-generation catalyst and cesium carbonate as the base to activate the less reactive chlorine functional-ity, furnishing the final compounds **23a–b** in good yields.

In order to shed a light on its mechanism, the oxidative cyclization reaction was run in the presence of 2,2,6,6-tetramethylpiperidine 1-oxyl (TEMPO), a free radical trapping reagent (Scheme 5). Adding three equivalents of TEMPO to the



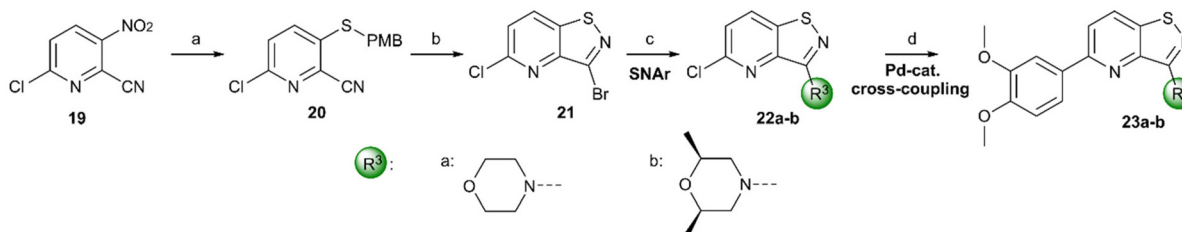
Scheme 5 Free radical trapping experiment. Reagents and conditions: (a) Br_2 (2.0 eq.), EtOAc, 80°C , 4 h.

reaction mixture allowed to isolate 3,6-dibromo-isothiazolo[4,5-*b*]pyridine **16** in 65% yield. This suggests that this reaction does not follow a radical pathway, but has rather an ionic nature. The lower yield in the control reaction with TEMPO is due to the purification technique (precipitate filtration and further recrystallization), which is dependent on the scale and solubilizing potential of the initial reaction mixture.

The proposed ionic mechanism is depicted in Scheme 6. Intermediate **A** is formed by reaction of compound **15** with bromine. The subsequent elimination of cation **E**, similarly to the previously described mechanism of PMB deprotection,²⁶ yields species **B**, containing a S–Br bond. Activation of the nitrile group *via* protonation by a catalytic amount of hydrogen bromide present in bromine, leads to intermediate **C**, which undergoes bromide attack to form the imine species **D**. An intramolecular cyclization, with concomitant elimination of HBr, furnishes the final product **16**. The previously formed cation **E** is attacked by bromide forming PMB bromide **F**. Its electron-rich character makes intermediate **F** susceptible for overbromination affording compounds **G**, explaining the requirement of more than one equivalent of bromine. Moreover, upon the formation of compounds **G**, hydrogen bromide is released which acts as a catalyst in the synthesis of compound **16**.

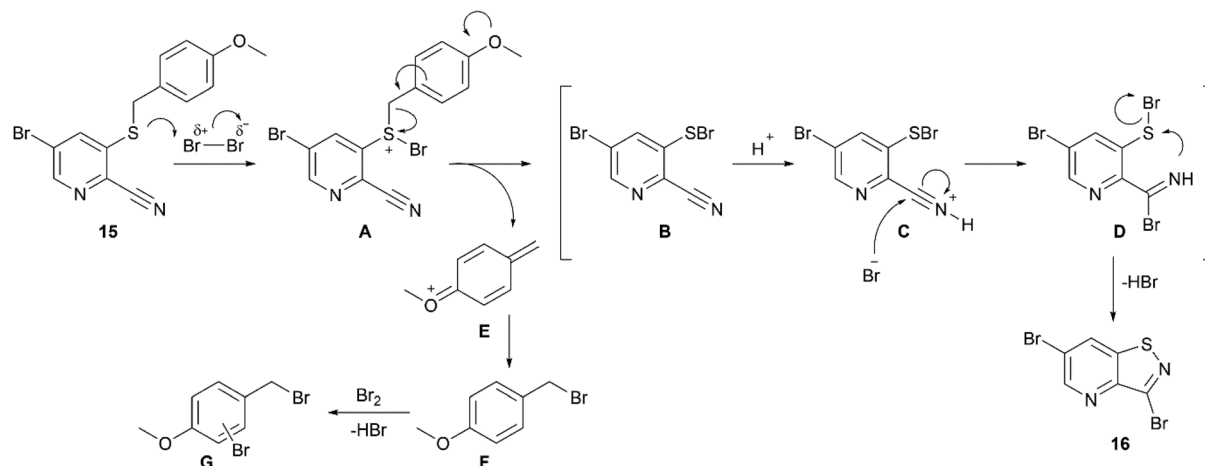
GAK binding affinity evaluation

To assess GAK binding affinity of a selection of the isothiazolo[4,5-*b*]pyridines (**18c–f** and **23a–b**) the commercially available LanthaScreen™ Europium kinase binding assay was employed (Table 1).²⁷ In this competitive assay, the ATP binding site is exposed to a Alexa Fluor 647-labeled conjugate or tracer and the potential GAK ligands. A high time-resolved fluorescence resonance energy transfer (TR-FRET) signal resulting from the antibody and tracer binding, indicates the occupation of the



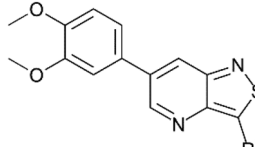
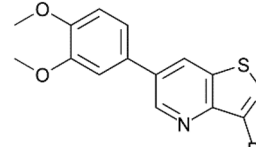
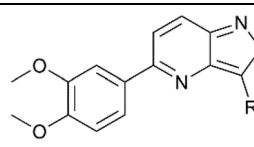
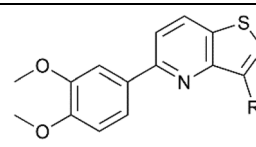
Scheme 4 Synthesis of 3,5-disubstituted isothiazolo[4,5-*b*]pyridines. Reagents and conditions: (a) *p*-methoxybenzylthiol, K_2CO_3 , DMF, -45°C , 63%; (b) Br_2 , EtOAc, 0°C , 75%; (c) $(\text{R}^3)_2\text{NH}$, EtOH, reflux, 57–71%; (d) 3,4-dimethoxyphenylboronic acid, XPhos Pd G3, Cs_2CO_3 , dioxane/ H_2O , 80°C , 89% – quant. PMB – *p*-methoxybenzyl.





Scheme 6 Proposed mechanism of cyclization.

Table 1 GAK binding data of isothiazolo[4,5-*b*]pyridines, compared to reported isothiazolo[4,3-*b*]pyridines

	Isothiazolo[4,3- <i>b</i>]pyridines ^a	Isothiazolo[4,5- <i>b</i>]pyridines ^a
	 24 IC₅₀ = 0.442 μM	 18c IC₅₀ > 10 μM
	25 IC₅₀ = 0.136 μM^c	18d IC₅₀ > 10 μM
	26 IC₅₀ = 0.014 μM^d	18e IC₅₀ > 10 μM
	27 IC₅₀ = 0.458 μM	18f IC₅₀ > 10 μM
	Isothiazolo[4,3- <i>b</i>]pyridines ^a	Isothiazolo[4,5- <i>b</i>]pyridines ^a
	 28 IC₅₀ = 0.124 μM^c	 23a IC₅₀ > 3.33 μM^b
	3 IC₅₀ = 0.024 μM^c	23b IC₅₀ > 3.33 μM^b

^a IC₅₀ = half-maximal inhibitory concentration. Values are the mean of two independent experiments. ^b At higher concentrations, these compounds exhibited intrinsic fluorescence against the acceptor wavelength, interfering with the assay readout. ^c Data from ref. 19. ^d Data from ref. 20.



ATP site. When a compound is endowed with high GAK binding affinity, it displaces the tracer leading to the reduction in the TR-FRET signal. The purity of the analysed samples was above 95% (ESI, Table S1†).

The goal of this research was to evaluate the importance of the central isothiazolo[4,3-*b*]pyridine scaffold for GAK affinity, by the synthesis of a series of isothiazolo[4,5-*b*]pyridines. It is well-known from previous research that an electron-rich aryl group (such as a dimethoxyphenyl, trimethoxyphenyl or amino-methoxyphenyl) at position 5 or 6 of the central core is optimal for GAK affinity. On the other hand, prior SAR studies revealed that position 3 of the isothiazolo[4,3-*b*]pyridine scaffold tolerates more structural variety, such as various *N*- and *O*-nucleophiles. Hence, within the newly synthesized isothiazolo[4,5-*b*]pyridines, 6-(3,4-dimethoxyphenyl) (compounds **18c-f**), as well as 5-(3,4-dimethoxyphenyl) derivatives (compounds **23a-b**) were selected for GAK affinity evaluation due to their high structural similarity to the previously reported potent inhibitors.

In order to study the influence of swapping the sulfur and nitrogen atoms within the isothiazole moiety, a side-by-side comparison of the GAK binding affinity of both scaffolds was performed, keeping the substitution pattern intact. Therefore, the previously reported isothiazolo[4,3-*b*]pyridines **3** and **24-28** (Fig. 3) were included for comparison purposes. Isothiazolo[4,3-*b*]pyridines **24** and **27** were previously shown to be endowed with potent GAK affinity in the KINOMEScan™ platform of DiscoverX. To confirm their GAK binding in the Lanthascreen™ assay (which was used to evaluate GAK affinity of the newly synthesized isothiazolo[4,5-*b*]pyridines) com-

pounds **24** and **27** were resynthesized according to known procedures.^{14,15} Biochemical evaluation in the Lanthascreen™ binding assay revealed that these compounds behaved as potent GAK ligands (Table 1), as expected based on the DiscoverX data. A good correlation between both assay formats was observed, which is in agreement with previous literature findings.²⁸

As can be clearly derived from the data in Table 1, all isothiazolo[4,5-*b*]pyridines completely lack GAK affinity, whereas the parent isothiazolo[4,3-*b*]pyridines do show GAK affinity. Hence, swapping the sulfur and nitrogen atoms within the isothiazole moiety had a detrimental effect on GAK inhibition.

Molecular modeling

To rationalise the lack of GAK affinity of the various isothiazolo[4,5-*b*]pyridines, a close inspection of the experimentally determined binding mode of isothiazolo[4,3-*b*]pyridine **1** to GAK (PDB entry 4Y8D¹⁵) was performed (Fig. 4, top panel). Besides van der Waals interactions of the morpholinyl and 4-amino-3-methoxyphenyl moieties, four interactions of the isothiazolo[4,3-*b*]pyridine scaffold are observed.

(i) A classical hydrogen bond between the nitrogen of the isothiazole moiety and the backbone NH of Cys126 (N/N distance 3.0 Å); this bond is well-known as a stabilizing interaction.

(ii) A non-conventional hydrogen bond between cys126.SG acting as a hydrogen bond donor and the isothiazole nitrogen (SG/N distance 3.6 Å); this type of hydrogen bond is an important interaction.²⁹

(iii) A non-conventional hydrogen bond between the sulfur of the isothiazole moiety and the hydroxyl group of Thr123 (S/O distance 3.4 Å); since sulfur is a rather poor H-bond acceptor, this H-bond is rather weak.

(iv) A chalcogen bond (ChB) interaction between the sulfur of the isothiazole and the backbone oxygen of Glu124 (S/O distance 3.2 Å, C-S...O angle is 153.2°).³⁰ The ability of sulfur to act as a ChB donor is favoured because of the electron-withdrawing capacity of the nitrogen, covalently attached to the sulfur.

Overall, the combination of these interactions contributes to the excellent GAK binding affinity of compound **1**.

The corresponding isothiazolo[4,5-*b*]pyridine (compound **29**) was then superimposed (by adjusting the flexible dihedral angles) on the isothiazolo[4,3-*b*]pyridine **1** (which displays potent GAK affinity). A different interaction pattern of compound **29** with GAK was observed, when compared to GAK inhibitor **1** (Fig. 4, bottom).

(i) The sulfur atom of the isothiazole moiety is engaged in a hydrogen bonding with the backbone nitrogen of Cys126 (S/N distance 2.7 Å). This interaction is rather weak as sulfur functions as hydrogen bond acceptor.

(ii) The thiol group of Cys126 (as H-bond donor) makes a weak hydrogen bond with the isothiazole sulfur as H-bond acceptor (S/S distance 3.3 Å).

(iii) The isothiazole sulfur has a ChB interaction with Glu124.O (distance S/O 3.1 Å; angle O-S-C 150.94°).

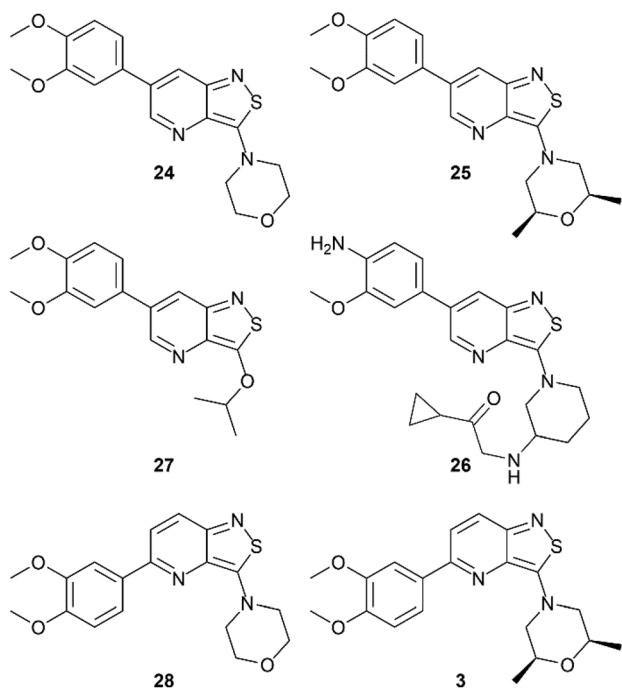


Fig. 3 Known isothiazolo[4,3-*b*]pyridines included in current SAR study.



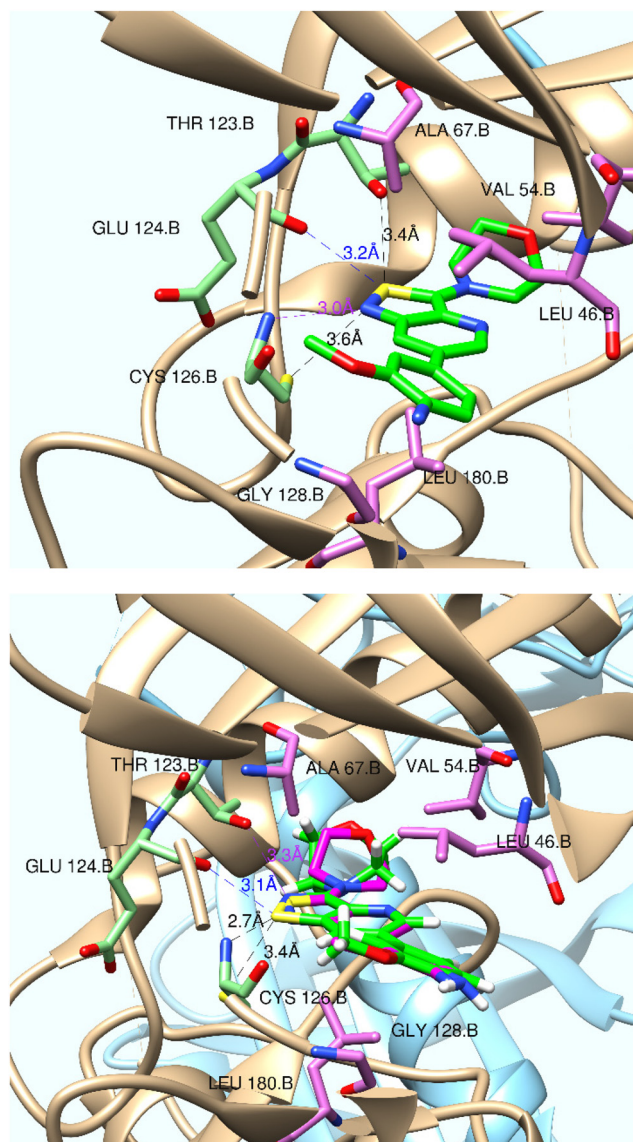


Fig. 4 Top: interactions between isothiazolo[4,3-*b*]pyridine **1** (green) and GAK in PDB 4Y8D (resolution 2.1 Å). Bottom: isothiazolo[4,5-*b*]pyridine **29** (green), superimposed on compound **1** (magenta). Figures were created using UCSF Chimera.³¹

(iv) The nitrogen of the isothiazole can hydrogen bond to Thr123.OG/HOG. However, the N/O distance of 3.4 Å makes this classical hydrogen bond rather weak.

It is clear that the superimposition model with the isothiazolo[4,5-*b*]pyridine **29** still shows important interactions with GAK. Only the classical hydrogen bond might be weaker based on the distance criterion. However, a comparison of the interactions based on this superimposition model does not offer a satisfying explanation for the complete lack of GAK affinity of isothiazolo[4,5-*b*]pyridines.

The dipole moment is a widely used parameter to explain bioactivities of molecules. In order to investigate if differences in the polarity of a molecule are related to different GAK inhibition, the dipole moment of both scaffolds was calculated

Table 2 Calculated dipole moments using open MOPAC22 at the PM6 level,³² having the same conformation as the ligand in PDB 4Y8D

Cmpd	Structure	Dipole moment (Debye)
1		0.785
29		4.647

(Table 2). The dipole moment of the novel isothiazolo[4,5-*b*]pyridine **29** is drastically increased, when compared to the original isothiazolo[4,3-*b*]pyridine analogue **1** (Table 2).

To further investigate how dissimilar the isothiazolo[4,3-*b*]pyridine **1** and the isothiazolo[4,5-*b*]pyridine **29** are in terms of electrostatics, the molecular electrostatic potential surfaces

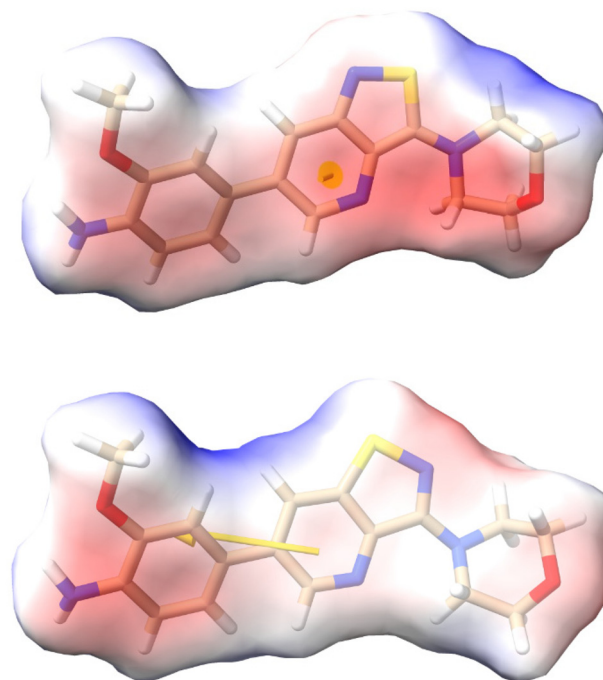


Fig. 5 Electrostatic potential calculated by APBS around the 2 scaffolds (having a conformation similar to compound **1** in PDB 4Y8D). Top: isothiazolo[4,3-*b*]pyridine **1**. Bottom: isothiazolo[4,5-*b*]pyridine **29**. The yellow arrow is the dipole vector starting at the centre of mass origin. The dipole points from – to + charges. Colors: red = negative potential; blue = positive potential; white = neutral. Figure created in ChimeraX.³³



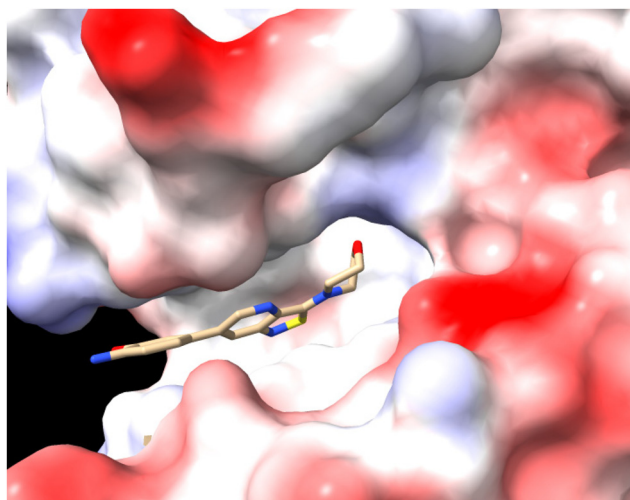


Fig. 6 The GAK ATP binding site in PDB 4Y8D, as coloured by Coulomb potential (red = negative signal, white = neutral, blue = positive potential).

around both scaffolds (having a similar conformation as compound **1** in 4Y8D) was calculated. The blue (positive) potential around the sulfur of the isothiazolo[4,3-*b*]pyridine **1** (Fig. 5, top) matches the red negative potential in the GAK binding pocket (Fig. 6). This electrostatic complementarity between GAK and isothiazolo[4,3-*b*]pyridine **1** is beneficial for GAK affinity.

In contrast, for the isothiazolo[4,5-*b*]pyridine **29** (Fig. 5, bottom), a more negative potential (red) is observed at the nitrogen of the isothiazole moiety, which is incompatible with the red negative potential at this position in the ATP binding site of GAK (Fig. 6), which explains the inactivity of isothiazolo[4,5-*b*]pyridines as GAK binders.

Conclusion

To probe the importance of the isothiazolo[4,3-*b*]pyridine scaffold for GAK binding, the closely related isothiazolo[4,5-*b*]pyridines (with a substitution pattern that is known to be beneficial for GAK affinity) were prepared. Surprisingly, the isothiazolo[4,5-*b*]pyridines were completely devoid of GAK affinity. Molecular modeling allowed to rationalise the lack of GAK binding due to a number of weaker hydrogen interactions, as well as of a less favourable electrostatic complementarity.

Despite the negative GAK data, the chemistry presented in this work is still of high value. A straightforward and high-yielding two-step synthesis of the dihalo-isothiazolo[4,5-*b*]pyridine building blocks **16** and **21** was established. These key intermediates can be used in various synthetic transformations (such as nucleophilic aromatic substitutions and palladium-catalyzed cross-coupling reactions), allowing regio-selective functionalisation of the isothiazolo[4,5-*b*]pyridine scaffold. It allows the preparation of isothiazolo[4,5-*b*]pyridine

based compound libraries with broad structural diversity that can be used in various drug discovery screening programs.

Experimental section

Chemistry

General information. Chemicals purchased from commercial sources (BLDPharm, J&K Scientific, Merck, Acros Organics, TCI Chemicals, or Fluorochem) were used without further purification unless otherwise stated. Dry reaction solvents were received from commercial sources. TLC analysis was performed using silica gel on TLC aluminum sheets bought from Sigma-Aldrich (pore diameter of 60 Å, fluorescent indicator at 254 nm). Product visualization performed by UV irradiation at 254 nm. Flash column chromatography was performed with a CombiFlash EZ prep apparatus using BGB Scorpius Silica 60 Å Irregular – 12 g cartridges. Reactions requiring dry conditions were performed in the flame-dried screw-capped reaction tubes. If heating was required, the reactions were performed in screw-capped reaction tubes placed in an aluminum heating block on a hot plate with magnetic stirring. ^1H and ^{13}C nuclear magnetic resonance (NMR) spectroscopy were performed on a Bruker Avance III HD 400 spectrometer (400 MHz for ^1H NMR and 101 MHz for ^{13}C NMR) or on a Bruker Avance II+ 600 spectrometer (600 MHz for ^1H NMR and 151 MHz for ^{13}C NMR) at ambient conditions. Chemical shifts (δ) are reported in parts per million (ppm) and referenced for all ^1H NMR spectra to tetramethylsilane (TMS, 0 ppm), or the residual signals of d_1 -chloroform (CDCl_3 , 7.26 ppm) or dimethyl sulfoxide- d_6 ($\text{DMSO}-d_6$, 2.50 ppm). All ^{13}C NMR spectra were referenced to the residual signals of CDCl_3 (77.16 ppm) or $\text{DMSO}-d_6$ (39.52 ppm). High resolution mass spectra (HRMS) were acquired on a quadrupole orthogonal acceleration time-of-flight mass spectrometer (Synapt G2 HDMS, Waters, Milford, MA) equipped with an electrospray ionization source. Samples were dissolved in 50% (v/v) acetonitrile, infused at $5\ \mu\text{L}\ \text{min}^{-1}$ and spectra were obtained in positive (or negative) ionization mode with a resolution of 15 000 (FWHM) using leucine enkephalin as lock mass.

3-Chloro-5-(3,4-dimethoxyphenyl)pyridine-2-carbonitrile (9). To a screw-capped reaction tube equipped with a magnetic stirring bar commercially available (from BLDPharm) 3,5-dichloropyridine-2-carbonitrile **8** (1.73 g, 10.0 mmol, 1.0 eq.), (3,4-dimethoxyphenyl)boronic acid (2.00 g, 11.0 mmol, 1.1 eq.) and potassium carbonate (2.76 g, 20.0 mmol, 2.0 eq.) were added and dissolved in the mixture of THF (40 ml) and water (10 ml). The reaction mixture was degassed with argon, and $\text{Pd}(\text{PPh}_3)_4$ (0.578 g, 0.5 mmol, 0.05 eq.) was added. The mixture was degassed again and was allowed to stir overnight at 100 °C. After disappearance of the starting material, the mixture was diluted with water (5 ml) and extracted three times with ethyl acetate. The combined organic layers were dried over anhydrous magnesium sulfate, filtered and concentrated *in vacuo*. The crude residue was purified by silica gel flash column chromatography (eluting with a mixture of hexanes/EtOAc in a



ratio of 8/2). The title compound was obtained as a yellow solid (72% yield, 1.97 g).

MP 164–167 °C **¹H NMR** (400 MHz, CDCl₃) δ (ppm): 8.79 (d, J = 2.0, 1H), 7.97 (d, J = 2.0, 1H), 7.19 (dd, J = 8.3, 2.1, 1H), 7.06 (d, J = 2.1, 1H), 7.01 (d, J = 8.3, 1H), 3.98 (s, 3H), 3.96 (s, 3H). **¹³C NMR** (101 MHz, CDCl₃) δ (ppm): 151.0, 150.1, 147.0, 140.9, 136.0, 134.6, 130.6, 127.2, 120.6, 115.1, 112.0, 110.2, 56.3, 56.2. **HRMS** (ESI⁺): m/z calculated for C₁₄H₁₁ClN₂O₂ [M + H]⁺: 275.05817; found: 275.0585.

5-(3,4-Dimethoxyphenyl)-3-((4-methoxybenzyl)thio)picolinonitrile (11). To a flame-dried screw-capped reaction tube equipped with a magnetic stirring bar 3-chloro-5-(3,4-dimethoxyphenyl)pyridine-2-carbonitrile **9** (824.0 mg, 3.0 mmol, 1.0 eq.) was added. The reaction tube was evacuated and backfilled with argon three times. Reagent was dissolved in anhydrous dimethylformamide (4 ml) and 4-methoxy- α -toluenethiol (0.46 ml, 3.3 mmol, 1.1 eq.) was added. The resulting mixture was cooled down to 0 °C and sodium hydride (79.2 mg, 3.3 mmol, 1.1 eq.) was added to the reaction mixture in small portions under the argon flow. The reaction was allowed to stir for 4 h at room temperature. The mixture was quenched with water (5 ml) and extracted three times with ethyl acetate. The combined organic layers were dried over anhydrous magnesium sulfate, filtered and concentrated *in vacuo*. The crude residue was purified by silica gel flash column chromatography (eluting with a mixture of hexanes/EtOAc in a ratio of 85/15) and further washed with diethyl ether providing the title compound as an orange solid (81% yield, 945.0 mg).

MP 136–137 °C **¹H NMR** (400 MHz, CDCl₃) δ (ppm): 8.66 (d, J = 2.0, 1H), 7.67 (d, J = 2.0, 1H), 7.26–7.23 (m, 2H), 7.05 (dd, J = 8.3, 2.1, 1H), 6.96 (d, J = 8.3, 1H), 6.88 (d, J = 2.1, 1H), 6.85–6.82 (m, J = 8.6, 2H), 4.23 (s, 2H), 3.94 (s, 3H), 3.93 (s, 3H), 3.77 (s, 3H). **¹³C NMR** 159.4, 150.6, 149.9, 146.7, 139.3, 138.6, 136.1, 132.7, 130.3, 128.2, 127.6, 120.3, 116.2, 114.3, 111.9, 110.1, 56.3, 56.2, 55.4, 38.3. **HRMS** (ESI⁺): m/z calculated for C₂₂H₂₀N₂O₃S [M + H]⁺: 393.12673; found: 393.1264.

5-Bromo-3-((4-methoxybenzyl)thio)picolinonitrile (15). To a flame-dried screw-capped reaction tube equipped with a magnetic stirring bar, commercially available (from BLDPharm) 5-bromo-3-fluoropicolinonitrile **14** (2.01 g, 10.0 mmol, 1.0 eq.) and potassium carbonate (2.07 g, 15.0 mmol, 1.5 eq.) were added. The reaction tube was evacuated and backfilled with argon three times. Reagents were dissolved in anhydrous dimethylacetamide (10 ml). The resulting mixture was cooled down to 0 °C and 4-methoxy- α -toluenethiol (1.46 ml, 10.5 mmol, 1.05 eq.) was added drop-wise to the reaction mixture. The reaction was allowed to stir 4 h at 0 °C. The mixture was quenched with water (10 ml) and extracted three times with ethyl acetate. The combined organic layers were dried over anhydrous magnesium sulfate, filtered and concentrated *in vacuo*. Recrystallization from methanol provided the title compound as white crystals (85% yield, 2.84 g).

MP 92–93 °C **¹H NMR** (600 MHz, CDCl₃) δ (ppm): 8.50 (d, J = 2.0 Hz, 1H), 7.77 (d, J = 2.0 Hz, 1H), 7.25–7.22 (m, 2H), 6.87–6.84 (m, 2H), 7.37–7.33 (m, 2H), 4.20 (s, 2H), 3.80 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ (ppm): 159.5, 148.9, 140.9, 139.4, 131.9, 130.2, 126.2, 124.4, 115.2, 114.3, 55.3, 37.8. **HRMS** (ESI⁺): m/z calculated for C₁₄H₁₁BrN₂OS [M + H]⁺: 334.9849; found: 334.9845.

3,6-Dibromoisothiazolo[4,5-*b*]pyridine (16). To a screw-capped reaction tube equipped with a magnetic stirring bar 5-bromo-3-((4-methoxybenzyl)thio)picolinonitrile **15** (168.0 mg, 0.5 mmol, 1.0 eq.) was added and dissolved in ethyl acetate (3 ml). Bromine (0.05 ml, 1.0 mmol, 2.0 eq.) was added to the reaction mixture. The reaction was allowed to stir 4 h at 80 °C. The formed precipitate was filtered off and further recrystallized from methanol providing the title compound as white needle-like crystals (88% yield, 442.0 mg).

MP 185–186 °C **¹H NMR** (400 MHz, DMSO-*d*₆) δ (ppm): 9.06 (d, J = 2.0 Hz, 1H), 8.97 (d, J = 2.0 Hz, 1H). **¹³C NMR** (101 MHz, DMSO-*d*₆) δ (ppm): 150.8, 146.6, 146.1, 137.9, 132.7, 120.6. **HRMS** (ESI⁺): m/z calculated for C₆H₂Br₂N₂S [M + H]⁺: 292.8379; found: 292.8379.

Nucleophilic substitution

General procedure A. To a screw-capped reaction tube equipped with a magnetic stirring bar 3,6-dibromoisothiazolo[4,5-*b*]pyridine **16** (1.0 eq.) or 3-bromo-5-chloroisothiazolo[4,5-*b*]pyridine **21** (1.0 eq.) were added and dissolved in ethanol (0.05 M). The nitrogen-containing nucleophile (3.0 eq.) was then added to the reaction mixture. The reaction was heated at reflux overnight. After disappearance of the starting material, volatiles were evaporated *in vacuo* and the crude residue was purified by silica gel flash column chromatography (eluting with a mixture of hexanes/MTBE in a ratio of 9/1).

6-Bromoisothiazolo[4,5-*b*]pyridin-3-amine (17a). To a screw-capped reaction tube equipped with a magnetic stirring bar ammonia solution in methanol (7 N) (0.49 ml, 3.4 mmol, 5.0 eq.) and dimethylformamide (1 ml) were added. To the resulting mixture the solution of dibromoisothiazolo[4,5-*b*]pyridine **16** (200.0 mg, 0.68 mmol, 1.0 eq.) in dimethylformamide (1 ml) was added dropwise. The reaction was allowed to stir 2 h at room temperature. The mixture was diluted with water (4 ml), the formed precipitate was filtered and further purified by silica gel flash column chromatography (eluting with a mixture of hexanes/EtOAc in a ratio of 9/1). The title compound was obtained as off-white solid (62% yield, 97.0 mg).

MP 109–112 °C **¹H NMR** (400 MHz, CDCl₃) δ (ppm): 8.42 (d, J = 2.1 Hz, 1H), 8.28 (d, J = 2.1 Hz, 1H), 2.87 (br s, 2H). **¹³C NMR** (101 MHz, CDCl₃) δ (ppm): 149.2, 148.0, 133.7, 125.5, 124.5, 114.7. **HRMS** (ESI⁺): m/z calculated for C₆H₄BrN₃S [M + H]⁺: 229.93825; found: 229.9390.

6-Bromo-*N*-(tetrahydro-2H-pyran-4-yl)isothiazolo[4,5-*b*]pyridin-3-amine (17b). To a screw-capped reaction tube equipped with a magnetic stirring bar 3,6-dibromoisothiazolo[4,5-*b*]pyridine **16** (147.0 mg, 0.5 mmol, 1.0 eq.) and 4-aminotetrahydropyran (151.7 mg, 1.5 mmol, 3.0 eq.) were added and dissolved in dimethylformamide (2 ml). The reaction was allowed to stir overnight at room temperature. The mixture was diluted with water (4 ml), the formed precipitate was filtered and further purified by silica gel flash column chromatography (eluting with a



mixture of hexanes/EtOAc in a ratio of 8/2). The title compound was obtained as off-white solid (80% yield, 125.0 mg).

MP 125–127 °C ^1H NMR (600 MHz, CDCl_3) δ (ppm): 8.43 (d, $J = 2.0$ Hz, 1H), 8.16 (d, $J = 2.0$ Hz, 1H), 4.01–3.98 (m, 2H), 3.38 (dt, $J = 2.0, 11.8$ Hz, 2H), 3.02–2.97 (m, 1H), 2.94 (d, $J = 5.1$ Hz, 1H), 1.99–1.96 (m, 2H), 1.57–1.50 (m, 2H). ^{13}C NMR (151 MHz, CDCl_3) δ (ppm): 148.5, 148.3, 134.1, 125.4, 124.9, 114.7, 66.6, 57.4, 33.8. **HRMS** (ESI^+): m/z calculated for $\text{C}_{11}\text{H}_{12}\text{BrN}_3\text{OS}$ [$\text{M} + \text{H}$] $^+$: 313.99577; found: 313.9964.

4-(6-Bromoisothiazolo[4,5-*b*]pyridin-3-yl)morpholine (17c). This compound was prepared according to the general procedure A using 3,6-dibromoisothiazolo[4,5-*b*]pyridine **16** (294.0 mg, 1.0 mmol), morpholine (0.26 ml, 3.0 mmol), and ethanol (20 ml). The title compound was obtained as off-white solid (92% yield, 919.0 mg).

MP 154–156 °C ^1H NMR (400 MHz, CDCl_3) δ (ppm): 8.63 (d, $J = 2.0$ Hz, 1H), 8.28 (d, $J = 2.0$ Hz, 1H), 4.02–4.00 (m, 4H), 3.91–3.89 (m, 4H). ^{13}C NMR (101 MHz, CDCl_3) δ (ppm): 160.3, 147.9, 147.3, 142.0, 130.6, 119.6, 66.9, 47.9. **HRMS** (ESI^+): m/z calculated for $\text{C}_{10}\text{H}_{10}\text{BrN}_3\text{OS}$ [$\text{M} + \text{H}$] $^+$: 299.9801; found: 299.9804.

(2*S*,6*R*)-4-(6-Bromoisothiazolo[4,5-*b*]pyridin-3-yl)-2,6-dimethylmorpholine (17d). This compound was prepared according to the general procedure A using 3,6-dibromoisothiazolo[4,5-*b*]pyridine **16** (235.2 mg, 0.8 mmol), (2*S*,6*R*)-2,6-dimethylmorpholine (0.297 ml, 2.4 mmol), and ethanol (16 ml). The title compound was obtained as off-white solid (97% yield, 774.0 mg).

MP 94–96 °C ^1H NMR (400 MHz, CDCl_3) δ (ppm): 8.63 (d, $J = 2.0$ Hz, 1H), 8.25 (d, $J = 2.0$ Hz, 1H), 4.81–4.78 (m, 2H), 3.88–3.83 (m, 2H), 2.75 (dd, $J = 10.5, 12.9$ Hz, 2H), 1.27 (d, $J = 6.4$ Hz, 6H). ^{13}C NMR (101 MHz, CDCl_3) δ (ppm): 159.9, 147.8, 147.3, 142.0, 130.6, 119.5, 71.7, 52.9, 19.1. **HRMS** (ESI^+): m/z calculated for $\text{C}_{12}\text{H}_{14}\text{BrN}_3\text{OS}$ [$\text{M} + \text{H}$] $^+$: 328.0114; found: 328.0110.

tert-Butyl ((1-(6-bromoisothiazolo[4,5-*b*]pyridin-3-yl)piperidin-3-yl)methyl)carbamate (17ea). This compound was prepared according to the general procedure A using 3,6-dibromoisothiazolo[4,5-*b*]pyridine **16** (449.1 mg, 0.8 mmol), racemic 3-(Boc-aminomethyl)piperidine (0.297 ml, 2.4 mmol), and ethanol (16 ml). The title compound was obtained as off-white solid (97% yield, 774.0 mg).

MP 110–112 °C ^1H NMR (600 MHz, CDCl_3) δ (ppm): 8.65 (d, $J = 1.8$ Hz, 1H), 8.26 (d, $J = 1.8$ Hz, 1H), 5.73 (br s, 1H), 4.08–4.03 (m, 2H), 3.73–3.70 (m, 1H), 3.58–3.55 (m, 1H), 3.30–3.26 (m, 1H), 3.19–3.15 (m, 1H), 1.95 (br s, 1H), 1.86–1.78 (m, 2H), 1.70–1.66 (m, 1H), 1.48 (s, 9H), 1.46–1.43 (m, 1H). ^{13}C NMR (151 MHz, CDCl_3) δ (ppm): 160.9, 156.4, 147.8, 147.2, 141.9, 130.8, 119.4, 79.1, 51.3, 48.5, 42.4, 35.7, 28.6, 28.1, 23.4. **HRMS** (ESI^+): m/z calculated for $\text{C}_{17}\text{H}_{23}\text{BrN}_4\text{O}_2\text{S}$ [$\text{M} + \text{H}$] $^+$: 427.0798; found: 427.0794.

(1-(6-Bromoisothiazolo[4,5-*b*]pyridin-3-yl)piperidin-3-yl)methanaminium chloride (17eb). A slightly adapted procedure by Martinez-Gualda *et al.*²⁰ was used. To the round bottom reaction flask equipped with a magnetic stirring bar *tert*-butyl ((1-(6-bromoisothiazolo[4,5-*b*]pyridin-3-yl)piperidin-3-yl)methyl)

carbamate **17ea** (650.0 mg, 1.52 mmol, 1.0 eq.) was added and dissolved in 1,4-dioxane (30 ml). The resulting mixture was cooled down to 0 °C, hydrogen chloride solution 4.0 M in 1,4-dioxane (66.0 ml, 15.2 mmol, 10.0 eq.) was added drop-wise. The reaction was allowed to stir overnight at room temperature. After disappearance of the starting material, volatiles were evaporated *in vacuo* and the crude residue was triturated with dichloromethane to result in the title compound as a white solid (99% yield, 548.0 mg).

MP 209–210 °C ^1H NMR (400 MHz, $\text{DMSO}-d_6$) δ (ppm): 8.89 (d, $J = 2.2$ Hz, 1H), 8.77 (d, $J = 2.2$ Hz, 1H), 8.17 (br s, 3H), 4.70–4.67 (m, 1H), 4.47–4.43 (m, 1H), 3.20–3.13 (m, 1H), 3.02 (dd, $J = 9.9, 13.0$ Hz, 1H), 2.86–2.70 (m, 2H), 2.80–1.99 (m, 1H), 1.94–1.86 (m, 1H), 1.79–1.73 (m, 1H), 1.61–1.50 (m, 1H), 1.36–1.27 (m, 1H). ^{13}C NMR (101 MHz, $\text{DMSO}-d_6$) δ (ppm): 159.6, 147.5, 147.2, 141.3, 132.0, 118.9, 50.4, 47.5, 41.5, 33.5, 27.6, 23.6. **HRMS** (ESI^+): m/z calculated for $\text{C}_{12}\text{H}_{15}\text{BrN}_4\text{S}$ [$\text{M} + \text{H}$] $^+$: 327.0274; found: 327.0278.

***N*-((1-(6-Bromoisothiazolo[4,5-*b*]pyridin-3-yl)piperidin-3-yl)methyl)cyclopropanecarboxamide (17ec).** A slightly adapted procedure by Martinez-Gualda *et al.*²⁰ was used. To a flame-dried screw-capped reaction tube equipped with a magnetic stirring bar (1-(6-bromoisothiazolo[4,5-*b*]pyridin-3-yl)piperidin-3-yl)methanaminium chloride **17eb** (363.7 mg, 1.0 mmol, 1.0 eq.) was added. The reaction tube was evacuated and backfilled with argon three times. The reagent was dissolved in anhydrous dichloromethane (2 ml) and anhydrous pyridine (0.241 ml, 3.0 mmol, 3.0 eq.) was added. The resulting mixture was cooled down to 0 °C, cyclopropanecarbonyl chloride (0.109 ml, 1.2 mmol, 1.2 eq.) was added. The reaction was allowed to stir overnight at room temperature. After disappearance of the starting material, volatiles were evaporated *in vacuo* and the crude residue was purified by silica gel flash column chromatography (eluting with a mixture of DCM/EtOAc in a ratio of 9/1) providing the title compound as an off-white solid (82%, 323.0 mg).

MP 162–163 °C ^1H NMR (600 MHz, CDCl_3) δ (ppm): 8.62 (d, $J = 2.0$ Hz, 1H), 8.27 (d, $J = 2.0$ Hz, 1H), 6.51 (br s, 1H), 4.15–4.11 (m, 2H), 3.67–3.60 (m, 2H), 3.52–3.48 (m, 1H), 3.25–3.20 (m, 1H), 2.00–1.94 (m, 1H), 1.88–1.78 (m, 2H), 1.71–1.65 (m, 1H), 1.50–1.45 (m, 1H), 1.43–1.39 (m, 1H), 1.00–0.98 (m, 2H), 0.78–0.75 (m, 2H). ^{13}C NMR (151 MHz, CDCl_3) δ (ppm): 173.6, 160.8, 147.9, 147.1, 142.0, 130.9, 119.4, 51.2, 48.5, 41.6, 35.7, 28.3, 23.5, 15.1, 7.2. **HRMS** (ESI^+): m/z calculated for $\text{C}_{16}\text{H}_{19}\text{BrN}_4\text{OS}$ [$\text{M} + \text{H}$] $^+$: 395.0536; found: 395.0531.

6-Bromo-3-isopropoxyisothiazolo[4,5-*b*]pyridine (17f). To a flame-dried screw-capped reaction tube equipped with a magnetic stirring bar 3,6-dibromoisothiazolo[4,5-*b*]pyridine **16** (294.0 mg, 1.0 mmol, 1.0 eq.) was added. The reaction tube was evacuated and backfilled with argon three times. The reagent was dissolved in anhydrous dimethylformamide (2 ml) and anhydrous isopropanol (0.077 ml, 1.0 mmol, 1.0 eq.) was added. Sodium hydride (48.0 mg, 2.0 mmol, 2.0 eq.) was added to the reaction mixture in small portions under the argon flow. The reaction was allowed to stir overnight at 50 °C.



Volatiles were evaporated *in vacuo* and the crude residue was purified by silica gel flash column chromatography (eluting with a mixture of hexanes/MTBE in a ratio of 95/5) providing the title compound as light-brown solid (18% yield, 50.0 mg).

MP 69–70 °C **¹H NMR** (600 MHz, CDCl₃) δ (ppm): 8.75 (d, J = 1.9 Hz, 1H), 8.30 (d, J = 1.9 Hz, 1H), 5.45 (sept, J = 6.8 Hz, 1H), 1.53 (d, J = 6.8 Hz, 6H). **¹³C NMR** (151 MHz, CDCl₃) δ (ppm): 161.9, 149.4, 146.4, 139.5, 130.8, 120.6, 73.2, 22.1. **HRMS** (ESI⁺): m/z calculated for C₉H₉BrN₂OS [M + H]⁺: 272.9692; found: 272.9691.

6-Bromo-3-(phenylthio)isothiazolo[4,5-*b*]pyridine (17g). To a screw-capped reaction tube equipped with a magnetic stirring bar 3,6-dibromoisothiazolo[4,5-*b*]pyridine **16** (147.0 mg, 0.5 mmol, 1.0 eq.) was added and dissolved in dimethylformamide (2 ml). Thiophenol (0.06 ml, 0.55 mmol, 1.1 eq.) was then added to the reaction mixture. The reaction was allowed to stir overnight at 50 °C. The mixture was diluted with water and extracted three times with ethyl acetate. The combined organic layers were dried over anhydrous magnesium sulfate, filtered and concentrated *in vacuo*. The crude residue was purified by silica gel flash column chromatography (eluting with a mixture of hexanes/EtOAc in a ratio from 10/0 to 9/1). The title compound was obtained as off-white solid (53% yield, 86.4 mg).

MP 118–121 °C **¹H NMR** (600 MHz, CDCl₃) δ (ppm): 8.80 (d, J = 1.8 Hz, 1H), 8.40 (d, J = 1.8 Hz, 1H), 7.74–7.71 (m, 2H), 7.48–7.46 (m, 3H). **¹³C NMR** (101 MHz, CDCl₃) δ (ppm): 160.9, 149.6, 147.1, 146.5, 135.1, 130.6, 129.7, 129.6, 128.1, 120.6. **HRMS** (ESI⁺): m/z calculated for C₁₂H₇BrN₂S₂ [M + H]⁺: 322.93073; found: 322.9312.

6-Bromo-3-(4-(methoxymethyl)-1H-1,2,3-triazol-1-yl)isothiazolo[4,5-*b*]pyridine (17h). To a screw-capped reaction tube equipped with a magnetic stirring bar 3,6-dibromoisothiazolo[4,5-*b*]pyridine **16** (147.0 mg, 0.5 mmol, 1.0 eq.) was added and dissolved in dimethylformamide (2 ml). The resulting mixture was cooled down to 0 °C and sodium azide (48.8 mg, 0.75 mmol, 1.5 eq.) was added. The reaction was allowed to stir 2 h at 0 °C. The formed precipitate was filtered, washed with water, dried under vacuum, and incorporated in the following reaction without further purification. To a screw-capped reaction tube equipped with a magnetic stirring bar, the precipitate was added and dissolved in dimethylsulfoxide (1 ml). Water (1 ml), methyl propargyl ether (0.044 ml, 0.55 mmol, 1.1 eq.), sodium ascorbate (9.9 mg, 0.05 mmol, 0.1 eq.) and copper sulfate pentahydrate (12.5 mg, 0.05 mmol, 0.1 eq.) were then added to the reaction mixture. The reaction was allowed to stir overnight at room temperature. The mixture was diluted with water and extracted three times with dichloromethane. The combined organic layers were dried over anhydrous sodium sulfate, filtered and concentrated *in vacuo*. The crude residue was purified by silica gel flash column chromatography (eluting with a mixture of dichloromethane/EtOAc in a ratio of 1/1). The title compound was obtained as off-white solid (28% yield, 45.5 mg).

MP 192–193 °C **¹H NMR** (600 MHz, DMSO-*d*₆) δ (ppm): 9.17 (d, J = 1.8 Hz, 1H), 9.11 (s, 1H), 9.07 (d, J = 1.8 Hz, 1H), 4.64 (s, 2H), 3.36 (s, 3H). **¹³C NMR** (151 MHz, DMSO-*d*₆) δ (ppm):

150.9, 148.6, 144.2, 140.3, 132.9, 124.2, 120.3, 64.5, 57.5. **HRMS** (ESI⁺): m/z calculated for C₁₀H₈BrN₅OS [M + H]⁺: 325.97061; found: 325.9709.

6-Bromo-3-(pyridin-3-ylethynyl)isothiazolo[4,5-*b*]pyridine (17i). To a flame-dried screw-capped reaction tube equipped with a magnetic stirring bar 3,6-dibromoisothiazolo[4,5-*b*]pyridine **16** (294.0 mg, 1.0 mmol, 1.0 eq.) was added. To another flame-dried screw-capped reaction tube equipped with a magnetic stirring bar 3-ethynylpyridine (103.1 mg, 1.0 mmol, 1.0 eq.) was added. Both reaction tube were evacuated and backfilled with argon three times, to each tube anhydrous tetrahydrofuran (2 × 6 ml) was added. The resulting mixtures were cooled down to 0 °C. In the tube, containing alkyne, lithium hexamethyldisilazide solution in tetrahydrofuran (1.0 M, 1.0 ml, 1.0 mmol, 1.0 eq.) was added. The resulting mixture was transferred to the reaction tube, containing dibromoisothiazolo[4,5-*b*]pyridine **16**, *via* double-sided needle. This mixture was allowed to stir 2 h at 0 °C. After disappearance of the starting material, volatiles were evaporated *in vacuo* and the crude residue was purified by silica gel flash column chromatography (eluting with a mixture of hexanes/EtOAc in a ratio of 85/15). The title compound was obtained as white solid (80% yield, 252.0 mg).

MP 139–141 °C **¹H NMR** (400 MHz, CDCl₃) δ (ppm): 8.80 (dd, J = 0.9, 2.2 Hz, 1H), 8.65 (dd, J = 1.7, 4.9 Hz, 1H), 8.60 (d, J = 2.0 Hz, 1H), 8.25 (d, J = 2.0 Hz, 1H), 7.87–7.84 (ddd, J = 1.7, 2.2, 7.9 Hz, 1H), 7.37–7.33 (ddd, J = 0.9, 4.9, 7.9 Hz, 1H). **¹³C NMR** (101 MHz, CDCl₃) δ (ppm): 152.9, 150.2, 150.0, 139.3, 138.3, 137.2, 128.5, 125.7, 123.4, 118.8, 114.2, 98.4, 74.0. **HRMS** (ESI⁺): m/z calculated for C₁₃H₆BrN₃S [M + H]⁺: 315.95390; found: 315.9532.

Suzuki coupling

General procedure B. To the screw-capped reaction tube equipped with a magnetic stirring bar 3-substituted-6-bromoisothiazolo[4,5-*b*]pyridine (1.0 eq.), (3,4-dimethoxyphenyl)boronic acid (1.1 eq.) and potassium carbonate (2.0 eq.) were added and dissolved in the mixture of dioxane and water (4/1 ratio respectively, 0.1M). The reaction mixture was degassed with argon, and Pd(PPh₃)₄ (0.05 eq.) was added. The mixture was degassed again and was allowed to stir overnight at 100 °C. After disappearance of the starting material, volatiles were evaporated *in vacuo* and the crude residue was purified by silica gel flash column chromatography (eluting with a mixture of hexanes/EtOAc in a ratio of 8/2, unless mentioned otherwise).

6-(3,4-Dimethoxyphenyl)isothiazolo[4,5-*b*]pyridin-3-amine (18a). To a screw-capped reaction tube equipped with a magnetic stirring bar 6-bromoisothiazolo[4,5-*b*]pyridin-3-amine **17a** (50.0 mg, 0.21 mmol, 1.0 eq.), (3,4-dimethoxyphenyl)boronic acid (43.5 mg, 0.239 mmol, 1.1 eq.) and potassium carbonate (60.1 mg, 0.44 mmol, 2.0 eq.) were added and dissolved in a mixture of dioxane (1.8 ml) and water (0.4 ml). The reaction mixture was degassed with argon, and XPhos Pd G3 (18.4 mg, 0.02 mmol, 0.1 eq.) was added. The mixture was degassed again and was allowed to stir overnight at 100 °C.



After disappearance of the starting material, volatiles were evaporated *in vacuo* and the crude residue was purified by silica gel flash column chromatography (eluting with a mixture of hexanes/EtOAc in a ratio of 6/4). The title compound was obtained as off-white solid (64% yield, 40.1 mg).

MP 125–127 °C $^1\text{H NMR}$ (600 MHz, CDCl_3) δ (ppm): 8.83 (d, $J = 2.0$ Hz, 1H), 8.23 (d, $J = 2.0$ Hz, 1H), 7.22 (dd, $J = 1.8$, 8.0 Hz, 1H), 7.14 (d, $J = 1.8$ Hz, 1H), 7.02 (d, $J = 8.0$ Hz, 1H), 5.40 (br s, 2H), 3.98 (s, 3H), 3.96 (s, 3H). $^{13}\text{C NMR}$ (151 MHz, CDCl_3) δ (ppm): 158.7, 149.9, 149.8, 146.7, 144.9, 140.8, 135.6, 130.1, 125.9, 120.5, 111.9, 110.8, 56.3, 56.2. **HRMS** (ESI^+): m/z calculated for $\text{C}_{14}\text{H}_{13}\text{N}_3\text{O}_2\text{S}$ [$\text{M} + \text{H}$] $^+$: 288.08011; found: 288.0789.

6-(3,4-Dimethoxyphenyl)-*N*-(tetrahydro-2*H*-pyran-4-yl)isothiazolo[4,5-*b*]pyridin-3-amine (18b). This compound was prepared according to the general procedure B using 6-bromo-*N*-(tetrahydro-2*H*-pyran-4-yl)isothiazolo[4,5-*b*]pyridin-3-amine **17b** (60.0 mg, 0.19 mmol), (3,4-dimethoxyphenyl)boronic acid (38.2 mg, 0.21 mmol), potassium carbonate (52.8 mg, 0.38 mmol), $\text{Pd}(\text{PPh}_3)_4$ (11.0 mg, 0.01 mmol), dioxane (1.6 ml) and water (0.4 ml). The title compound was obtained as pale brown solid (69% yield, 49.2 mg).

MP 167–168 °C $^1\text{H NMR}$ (600 MHz, CDCl_3) δ (ppm): 8.78 (d, $J = 1.9$ Hz, 1H), 8.21 (d, $J = 1.9$ Hz, 1H), 7.21 (dd, $J = 1.9$, 8.2 Hz, 1H), 7.13 (d, $J = 1.9$ Hz, 1H), 7.01 (d, $J = 8.2$ Hz, 1H), 5.76 (d, $J = 7.9$ Hz, 2H), 4.23–4.16 (m, 1H), 4.07–4.04 (m, 2H), 3.98 (s, 3H), 3.96 (s, 3H), 3.60 (dt, $J = 1.9$, 11.5 Hz, 2H), 2.22–2.19 (m, 2H), 1.72–1.65 (m, 2H). $^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ (ppm): 157.6, 149.9, 149.8, 146.2, 144.6, 140.9, 135.9, 130.2, 126.0, 120.4, 111.9, 110.8, 67.0, 56.24, 56.20, 48.7, 33.6. **HRMS** (ESI^+): m/z calculated for $\text{C}_{19}\text{H}_{21}\text{N}_3\text{O}_3\text{S}$ [$\text{M} + \text{H}$] $^+$: 372.13762; found: 372.1377.

4-(6-(3,4-Dimethoxyphenyl)isothiazolo[4,5-*b*]pyridin-3-yl)morpholine (18c). This compound was prepared according to the general procedure B using 4-(6-bromoisothiazolo[4,5-*b*]pyridin-3-yl)morpholine **17c** (98.0 mg, 0.33 mmol), (3,4-dimethoxyphenyl)boronic acid (65.4 mg, 0.36 mmol), potassium carbonate (90.2 mg, 0.65 mmol), $\text{Pd}(\text{PPh}_3)_4$ (18.9 mg, 0.016 mmol), dioxane (2.6 ml) and water (0.7 ml). The title compound was obtained as off-white solid (87% yield, 101.2 mg).

MP 132–134 °C $^1\text{H NMR}$ (400 MHz, CDCl_3) δ (ppm): 8.82 (d, $J = 2.2$ Hz, 1H), 8.20 (d, $J = 2.2$ Hz, 1H), 7.21 (dd, $J = 2.0$, 8.2 Hz, 1H), 7.13 (d, $J = 2.0$, 1H), 7.00 (d, $J = 8.2$, 1H), 4.06–4.04 (m, 4H), 3.96 (s, 3H), 3.94 (s, 3H), 3.92–3.90 (m, 4H). $^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ (ppm): 160.7, 149.9, 149.7, 146.7, 145.6, 142.3, 134.4, 129.9, 125.6, 120.3, 111.9, 110.6, 66.9, 56.2, 56.1, 47.9. **HRMS** (ESI^+): m/z calculated for $\text{C}_{18}\text{H}_{19}\text{N}_3\text{O}_3\text{S}$ [$\text{M} + \text{H}$] $^+$: 358.1219; found: 358.1212.

(2*S*,6*R*)-4-(6-(3,4-Dimethoxyphenyl)isothiazolo[4,5-*b*]pyridin-3-yl)-2,6-dimethylmorpholine (18d). This compound was prepared according to the general procedure B using (2*S*,6*R*)-4-(6-bromoisothiazolo[4,5-*b*]pyridin-3-yl)-2,6-dimethylmorpholine **17d** (107.2 mg, 0.33 mmol), (3,4-dimethoxyphenyl)boronic acid (65.4 mg, 0.36 mmol), potassium carbonate (90.2 mg, 0.65 mmol), $\text{Pd}(\text{PPh}_3)_4$ (18.9 mg, 0.016 mmol), dioxane

(2.6 ml) and water (0.7 ml). The title compound was obtained as off-white solid (76% yield, 96.0 mg).

MP 151–152 °C $^1\text{H NMR}$ (600 MHz, CDCl_3) δ (ppm): 8.84 (d, $J = 1.9$ Hz, 1H), 8.20 (d, $J = 1.9$ Hz, 1H), 7.22 (dd, $J = 2.1$, 8.1 Hz, 1H), 7.14 (d, $J = 2.1$ Hz, 1H), 7.01 (d, $J = 8.1$ Hz, 1H), 4.88 (d, $J = 11.4$ Hz, 2H), 3.97 (s, 3H), 3.95 (s, 3H), 3.93–3.87 (m, 2H), 2.79 (dd, $J = 2.2$, 11.4 Hz, 2H), 1.29 (d, $J = 6.3$ Hz, 6H). $^{13}\text{C NMR}$ (151 MHz, CDCl_3) δ (ppm): 160.4, 149.9, 149.8, 146.7, 145.6, 142.4, 134.4, 129.9, 125.6, 120.3, 111.9, 110.7, 71.8, 56.2, 53.0, 19.1. **HRMS** (ESI^+): m/z calculated for $\text{C}_{20}\text{H}_{23}\text{N}_3\text{O}_3\text{S}$ [$\text{M} + \text{H}$] $^+$: 386.1533; found: 386.1527.

***N*-((1-(6-(3,4-Dimethoxyphenyl)isothiazolo[4,5-*b*]pyridin-3-yl)piperidin-3-yl)methyl)cyclopropanecarboxamide (18e)**. This compound was prepared according to the general procedure B using *N*-((1-(6-bromoisothiazolo[4,5-*b*]pyridin-3-yl)piperidin-3-yl)methyl) cyclopropanecarboxamide **17ec** (60.0 mg, 0.15 mmol), (3,4-dimethoxyphenyl)boronic acid (30.4 mg, 0.17 mmol), potassium carbonate (42.0 mg, 0.30 mmol), $\text{Pd}(\text{PPh}_3)_4$ (8.8 mg, 0.008 mmol), dioxane (1.2 ml) and water (0.3 ml). The title compound was purified by silica gel flash column chromatography (eluting with a mixture of hexanes/EtOAc in a ratio of 6/4) and was obtained as off-white solid (84% yield, 58.0 mg).

MP 166–167 °C $^1\text{H NMR}$ (600 MHz, CDCl_3) δ (ppm): 8.82 (d, $J = 2.0$ Hz, 1H), 8.22 (d, $J = 2.0$ Hz, 1H), 7.21 (dd, $J = 2.2$, 8.2 Hz, 1H), 7.13 (d, $J = 2.2$ Hz, 1H), 7.02 (d, $J = 8.2$ Hz, 1H), 6.82 (br s, 1H), 4.10–4.05 (m, 2H), 3.97 (s, 3H), 3.96 (s, 3H), 3.89–3.86 (m, 1H), 3.76–3.73 (m, 1H), 3.60–3.55 (m, 1H), 3.29–3.25 (m, 1H), 2.02–2.01 (m, 1H), 1.89–1.81 (m, 2H), 1.74–1.69 (m, 1H), 1.56–1.51 (m, 1H), 1.47–1.43 (m, 1H), 1.02–0.99 (m, 2H), 0.77–0.74 (m, 2H). $^{13}\text{C NMR}$ (151 MHz, CDCl_3) δ (ppm): 173.6, 161.3, 149.9, 149.8, 146.8, 145.5, 142.3, 134.6, 129.9, 126.0, 120.4, 111.9, 110.8, 56.24, 56.21, 51.3, 48.6, 41.3, 35.6, 28.3, 23.4, 15.2, 7.2. **HRMS** (ESI^+): m/z calculated for $\text{C}_{24}\text{H}_{28}\text{N}_4\text{O}_3\text{S}$ [$\text{M} + \text{H}$] $^+$: 453.1955; found: 453.1947.

6-(3,4-Dimethoxyphenyl)-3-isopropoxyisothiazolo[4,5-*b*]pyridine (18f). This compound was prepared according to the general procedure B using 6-bromo-3-isopropoxyisothiazolo[4,5-*b*]pyridine **17f** (77.0 mg, 0.28 mmol), (3,4-dimethoxyphenyl)boronic acid (56.4 mg, 0.31 mmol), potassium carbonate (77.9 mg, 0.56 mmol), $\text{Pd}(\text{PPh}_3)_4$ (16.3 mg, 0.014 mmol), dioxane (2.2 ml) and water (0.6 ml). The title compound was obtained as off-white solid (71% yield, 66.0 mg).

MP 154–155 °C $^1\text{H NMR}$ (400 MHz, CDCl_3) δ (ppm): 8.92 (d, $J = 2.0$ Hz, 1H), 8.22 (d, $J = 2.0$ Hz, 1H), 7.22 (dd, $J = 2.2$, 8.3 Hz, 1H), 7.13 (d, $J = 2.2$ Hz, 1H), 7.01 (d, $J = 8.3$ Hz, 1H), 5.49 (sept, $J = 6.3$ Hz, 1H), 3.97 (s, 3H), 3.95 (s, 3H), 1.54 (d, $J = 6.3$ Hz, 1H). $^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ (ppm): 162.2, 149.9, 149.8, 147.7, 145.5, 139.8, 135.8, 129.9, 125.8, 120.6, 111.9, 110.8, 72.8, 56.3, 56.2, 22.2. **HRMS** (ESI^+): m/z calculated for $\text{C}_{17}\text{H}_{18}\text{N}_2\text{O}_3\text{S}$ [$\text{M} + \text{H}$] $^+$: 331.1111; found: 331.1112.

6-(3,4-Dimethoxyphenyl)-3-(phenylthio)isothiazolo[4,5-*b*]pyridine (18g). This compound was prepared according to the general procedure B using 6-bromo-3-(phenylthio)isothiazolo[4,5-*b*]pyridine **17g** (76.0 mg, 0.24 mmol), (3,4-dimethoxyphenyl)boronic acid (47.1 mg, 0.26 mmol), potassium carbonate



(65.0 mg, 0.47 mmol), Pd(PPh₃)₄ (13.6 mg, 0.012 mmol), dioxane (1.9 ml) and water (0.5 ml). The title compound was obtained as pale brown solid (51% yield, 46.0 mg).

MP 161–163 °C ¹H NMR (600 MHz, CDCl₃) δ (ppm): 9.00 (d, *J* = 2.0 Hz, 1H), 8.32 (d, *J* = 2.0 Hz, 1H), 7.77–7.76 (m, 2H), 7.49–7.47 (m, 3H), 7.24 (dd, *J* = 1.9, 8.3 Hz, 1H), 7.15 (d, *J* = 1.9 Hz, 1H), 7.03 (d, *J* = 8.3 Hz, 1H), 3.99 (s, 3H), 3.96 (s, 3H). ¹³C NMR (151 MHz, CDCl₃) δ (ppm): 160.5, 150.1, 149.9, 147.9, 147.5, 145.5, 135.7, 135.0, 129.8, 129.53, 129.52, 128.6, 125.5, 120.6, 112.0, 110.8, 56.3, 56.2. **HRMS** (ESI⁺): *m/z* calculated for C₂₀H₁₆N₂O₂S₂ [*M* + *H*]⁺: 381.07259; found: 381.0721.

6-(3,4-Dimethoxyphenyl)-3-(4-(methoxymethyl)-1*H*-1,2,3-triazol-1-yl)isothiazolo[4,5-*b*]pyridine (18h). This compound was prepared according to the general procedure B using 6-bromo-3-(4-(methoxymethyl)-1*H*-1,2,3-triazol-1-yl)isothiazolo[4,5-*b*]pyridine **17h** (40.0 mg, 0.12 mmol), (3,4-dimethoxyphenyl)boronic acid (24.5 mg, 0.135 mmol), potassium carbonate (33.9 mg, 0.245 mmol), Pd(PPh₃)₄ (7.1 mg, 0.006 mmol), dioxane (1.0 ml) and water (0.3 ml). The title compound was obtained as off-white solid (62% yield, 29.1 mg).

MP 195–197 °C ¹H NMR (600 MHz, CDCl₃) δ (ppm): 9.24 (s, 1H), 9.13 (d, *J* = 2.0 Hz, 1H), 8.44 (d, *J* = 2.0 Hz, 1H), 7.29 (dd, *J* = 2.3, 8.4 Hz, 1H), 7.19 (d, *J* = 2.3 Hz, 1H), 7.06 (d, *J* = 8.4 Hz, 1H), 4.78 (s, 2H), 4.00 (s, 3H), 3.98 (s, 3H), 3.50 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ (ppm): 150.5, 150.0, 149.4, 148.8, 148.1, 145.3, 140.5, 136.2, 129.0, 125.5, 122.9, 120.8, 112.1, 110.8, 66.0, 58.6, 56.3, 56.2. **HRMS** (ESI⁺): *m/z* calculated for C₁₈H₁₇N₅O₃S [*M* + *H*]⁺: 384.11247; found: 384.1125.

3,6-Bis(3,4-dimethoxyphenyl)isothiazolo[4,5-*b*]pyridine (18j). To the screw-capped reaction tube equipped with a magnetic stirring bar 3,6-dibromoisothiazolo[4,5-*b*]pyridine **16** (73.5 mg, 0.25 mmol, 1.0 eq.), (3,4-dimethoxyphenyl)boronic acid (182.0 mg, 1.0 mmol, 4.0 eq.) and cesium carbonate (488.7 mg, 1.5 mmol, 6.0 eq.) were added and dissolved in the mixture of dioxane (5.3 ml) and water (1.1 ml). The reaction mixture was degassed with argon, and XPhos Pd G3 (42.3 mg, 0.05 mmol, 0.2 eq.) was added. The mixture was degassed again and was allowed to stir 4 h at 80 °C. After disappearance of the starting material, volatiles were evaporated *in vacuo* and the crude residue was purified by silica gel flash column chromatography (eluting with a mixture of hexanes/EtOAc in a ratio of 7/3). The title compound was obtained as pale brown solid (48% yield, 48.7 mg).

MP 123–124 °C ¹H NMR (600 MHz, CDCl₃) δ (ppm): 9.08 (d, *J* = 1.8 Hz, 1H), 8.41 (dd, *J* = 1.9, 8.4 Hz, 1H), 8.39 (d, *J* = 1.8 Hz, 1H), 8.16 (d, *J* = 1.9 Hz, 1H), 7.28 (dd, *J* = 2.3, 8.4 Hz, 1H), 7.20 (d, *J* = 2.3 Hz, 1H), 7.06–7.03 (m, 2H), 4.04 (s, 3H), 4.00 (s, 3H), 3.98 (s, 3H), 3.97 (s, 3H). ¹³C NMR (151 MHz, CDCl₃) δ (ppm): 161.7, 150.7, 149.9, 149.8, 149.1, 148.3, 147.9, 147.2, 134.2, 129.9, 127.4, 125.2, 121.9, 120.5, 111.9, 111.5, 110.9, 110.8, 56.24, 56.21, 56.16, 56.1. **HRMS** (ESI⁺): *m/z* calculated for C₂₂H₂₀N₂O₄S [*M* + *H*]⁺: 409.1216; found: 409.1217.

4-(6-(4-Fluorophenyl)isothiazolo[4,5-*b*]pyridin-3-yl)morpholine (18k). This compound was prepared according to the general procedure B using 4-(6-bromoisothiazolo[4,5-*b*]pyridin-3-yl)morpholine **17c** (50.0 mg, 0.17 mmol), (4-fluorophenyl)

boronic acid (25.6 mg, 0.18 mmol), potassium carbonate (46.0 mg, 0.33 mmol), Pd(PPh₃)₄ (9.6 mg, 0.008 mmol), dioxane (1.3 ml) and water (0.4 ml). The title compound was obtained as off-white solid (84% yield, 44.4 mg).

MP 139–141 °C ¹H NMR (600 MHz, CDCl₃) δ (ppm): 8.81 (d, *J* = 2.0 Hz, 1H), 8.21 (d, *J* = 2.0 Hz, 1H), 7.63–7.61 (m, 2H), 7.23–7.20 (m, 2H), 4.07–4.05 (m, 4H), 3.94–3.92 (m, 4H). ¹³C NMR (151 MHz, CDCl₃) δ (ppm): 163.4 (*J*_{C-F} = 249.3 Hz), 160.7, 146.7, 145.6, 142.7, 133.7, 133.4 (*J*_{C-F} = 3.3 Hz), 129.5 (*J*_{C-F} = 8.3 Hz), 126.1, 116.5 (*J*_{C-F} = 22.0 Hz), 67.0, 47.9. **HRMS** (ESI⁺): *m/z* calculated for C₁₆H₁₄FN₃OS [*M* + *H*]⁺: 316.09143; found: 316.0915.

4-(6-(Pyridin-3-ylethynyl)isothiazolo[4,5-*b*]pyridin-3-yl)morpholine (18l). To a flame-dried screw-capped reaction tube equipped with a magnetic stirring bar 6-(4-(6-bromoisothiazolo[4,5-*b*]pyridin-3-yl)morpholine **17c** (50.0 mg, 0.17 mmol, 1.0 eq.), 3-pyridinyl ethyn (18.9 mg, 0.18 mmol, 1.1 eq.) and bis(triphenylphosphine)palladium chloride (23.4 mg, 0.03 mmol, 0.2 eq.) were added. The reaction tube was evacuated and back-filled with argon three times. Reagents were dissolved in anhydrous dimethylformamide (2 ml). Triethylamine (0.046 ml, 0.03 mmol, 2.0 eq.) was added and the reaction mixture was allowed to stir overnight at 100 °C. The mixture was diluted with water and extracted three times with ethylacetate. The combined organic layers were dried over anhydrous sodium sulfate, filtered and concentrated *in vacuo*. The crude residue was purified by silica gel flash column chromatography (eluting with a mixture of dichloromethane/EtOAc in a ratio of 9/1). The title compound was obtained as white solid (71% yield, 38.0 mg).

MP 180–181 °C ¹H NMR (600 MHz, CDCl₃) δ (ppm): 8.83 (d, *J* = 1.2 Hz, 1H), 8.72 (d, *J* = 1.8 Hz, 1H), 8.62 (dd, *J* = 1.2, 4.9 Hz, 1H), 8.26 (d, *J* = 1.8 Hz, 1H), 7.88–7.86 (m, 1H), 7.36–7.33 (ddd, *J* = 0.6, 4.9, 7.8 Hz, 1H), 4.05–4.03 (m, 4H), 3.93–3.91 (m, 4H). ¹³C NMR (101 MHz, CDCl₃) δ (ppm): 160.7, 152.6, 149.6, 148.4, 145.8, 142.3, 138.8, 130.9, 123.3, 119.6, 117.5, 90.6, 89.2, 67.0, 47.9. **HRMS** (ESI⁺): *m/z* calculated for C₁₇H₁₄N₄OS [*M* + *H*]⁺: 323.09610; found: 323.0960.

6-Chloro-3-((4-methoxybenzyl)thio)picolinonitrile (20). To a flame-dried screw-capped reaction tube equipped with a magnetic stirring bar 6-chloro-3-nitropicolonitrile **19** (917.8 mg, 5.0 mmol, 1.0 eq.) and potassium carbonate (1.04 g, 7.5 mmol, 1.5 eq.) were added. The reaction tube was evacuated and back-filled with argon three times. The reagents were dissolved in anhydrous dimethylformamide (10 ml). The resulting mixture was cooled down to –45 °C, *p*-methoxybenzylthiol (0.73 ml, 5.25 mmol, 1.05 eq.) was added drop-wise over an hour. The reaction was allowed to stir one more hour at –45 °C. The mixture was quenched with water (10 ml) and extracted three times with ethyl acetate. The combined organic layers were dried over anhydrous magnesium sulfate, filtered and concentrated *in vacuo*. Recrystallization from methanol provided the title compound as white crystals (63% yield, 915.0 mg).

MP 118–119 °C ¹H NMR (600 MHz, CDCl₃) δ (ppm): 7.58 (d, *J* = 8.7 Hz, 1H), 7.35 (d, *J* = 8.6 Hz, 1H), 7.21–7.19 (m, 2H), 6.84–6.82 (m, 2H), 4.18 (s, 2H), 3.79 (s, 3H). ¹³C NMR



(151 MHz, CDCl₃) δ (ppm): 159.5, 149.9, 141.1, 138.2, 134.1, 130.2, 127.8, 126.9, 114.8, 114.4, 55.5, 38.4. **HRMS** (ESI⁺): m/z calculated for C₁₄H₁₁ClN₂OS [M + H]⁺: 291.0353; found: 291.0356.

3-Bromo-5-chloroisothiazolo[4,5-*b*]pyridine (21). To a screw-capped reaction tube equipped with a magnetic stirring bar 6-chloro-3-((4-methoxybenzyl)thio)picolinonitrile **20** (405.0 mg, 1.4 mmol, 1.0 eq.) was added and dissolved in ethyl acetate (14 ml). The resulting mixture was cooled down to 0 °C, bromine (0.128 ml, 2.5 mmol, 1.8 eq.) was added to the reaction mixture. The reaction was allowed to stir 6 h at 0 °C. After the reaction completion, volatiles were evaporated *in vacuo* and the crude residue was further recrystallized from methanol providing the title compound as off-white crystals (75% yield, 261.0 mg).

MP 123–124 °C **¹H NMR** (600 MHz, CDCl₃) δ (ppm): 8.26 (d, J = 8.5 Hz, 1H), 7.54 (d, J = 8.6 Hz, 1H). **¹³C NMR** (151 MHz, CDCl₃) δ (ppm): 151.7, 148.4, 144.2, 137.9, 130.8, 124.3. **HRMS** (ESI⁺): m/z calculated for C₆H₂BrClN₂S [M + H]⁺: 248.8884; found: 248.8917.

4-(5-Chloroisothiazolo[4,5-*b*]pyridin-3-yl)morpholine (22a). This compound was prepared according to the general procedure A using 3-bromo-5-chloroisothiazolo[4,5-*b*]pyridine **21** (124.8 mg, 0.5 mmol), morpholine (0.131 ml, 1.5 mmol), and ethanol (10 ml). The title compound was obtained as a pale orange solid (57% yield, 73.0 mg).

MP 144–145 °C **¹H NMR** (400 MHz, CDCl₃) δ (ppm): 8.07 (d, J = 8.6 Hz, 1H), 7.36 (d, J = 8.6 Hz, 1H), 4.01–3.98 (m, 4H), 3.92–3.89 (m, 4H). **¹³C NMR** (101 MHz, CDCl₃) δ (ppm): 159.9, 147.6, 145.3, 142.9, 130.9, 122.5, 66.9, 47.8. **HRMS** (ESI⁺): m/z calculated for C₁₀H₁₀ClN₃OS [M + H]⁺: 256.0306; found: 256.0312.

(2*S*,6*R*)-4-(5-Chloroisothiazolo[4,5-*b*]pyridin-3-yl)-2,6-dimethylmorpholine (22b). This compound was prepared according to the general procedure A using 3-bromo-5-chloroisothiazolo[4,5-*b*]pyridine **21** (124.8 mg, 0.5 mmol), (2*S*,6*R*)-2,6-dimethylmorpholine (0.186 ml, 1.5 mmol), and ethanol (10 ml). The title compound was obtained as a pale orange solid (71% yield, 101.2 mg).

MP 128–129 °C **¹H NMR** (600 MHz, CDCl₃) δ (ppm): 8.06 (d, J = 8.6 Hz, 1H), 7.35 (d, J = 8.6 Hz, 1H), 4.77 (d, J = 10.8 Hz, 2H), 3.89–3.84 (m, 2H), 2.77 (dd, J = 2.1, 10.5 Hz, 2H), 8.06 (d, J = 6.2 Hz, 6H). **¹³C NMR** (101 MHz, CDCl₃) δ (ppm): 159.6, 147.5, 145.3, 142.8, 130.9, 122.5, 71.7, 52.9, 19.1. **HRMS** (ESI⁺): m/z calculated for C₁₂H₁₄ClN₃OS [M + H]⁺: 284.0619; found: 284.0624.

Suzuki coupling

General procedure C. To a screw-capped reaction tube equipped with a magnetic stirring bar 3-substituted-5-chloroisothiazolo[4,5-*b*]pyridine (1.0 eq.), (3,4-dimethoxyphenyl)boronic acid (2.0 eq.) and cesium carbonate (3.0 eq.) were added and dissolved in a mixture of dioxane and water. The reaction mixture was degassed with argon, and XPhos Pd G3 (0.1 eq.) was added. The mixture was degassed again and was allowed to stir for 4 h at 80 °C. After disappearance of the starting material, volatiles were evaporated *in vacuo* and the crude

residue was purified by silica gel flash column chromatography (eluting with a mixture of hexanes/EtOAc in a ratio of 8/2).

4-(5-(3,4-Dimethoxyphenyl)isothiazolo[4,5-*b*]pyridin-3-yl)morpholine (23a). This compound was prepared according to the general procedure C using 4-(5-chloroisothiazolo[4,5-*b*]pyridin-3-yl)morpholine **22a** (60.0 mg, 0.235 mmol), (3,4-dimethoxyphenyl)boronic acid (85.4 mg, 0.47 mmol), caesium carbonate (229.3 mg, 0.704 mmol), XPhos Pd G3 (19.9 mg, 0.023 mmol), dioxane (3.8 ml) and water (0.9 ml). The title compound was obtained as a light brown solid (99% yield, 83.0 mg).

MP 188–189 °C **¹H NMR** (600 MHz, CDCl₃) δ (ppm): 8.15 (d, J = 8.7 Hz, 1H), 7.80 (d, J = 8.7 Hz, 1H), 7.69 (d, J = 2.0 Hz, 1H), 7.60 (dd, J = 2.0, 8.5 Hz, 1H), 6.99 (d, J = 8.5 Hz, 1H), 4.15–4.13 (m, 4H), 3.99 (s, 3H), 3.96 (s, 3H), 3.96–3.94 (m, 4H). **¹³C NMR** (151 MHz, CDCl₃) δ (ppm): 160.8, 153.9, 150.6, 149.5, 144.2, 143.7, 131.7, 129.3, 119.8, 118.4, 111.4, 109.9, 67.0, 56.2, 55.9, 48.1. **HRMS** (ESI⁺): m/z calculated for C₁₈H₁₉N₃O₃S [M + H]⁺: 358.1219; found: 358.1213.

(2*S*,6*R*)-4-(5-(3,4-Dimethoxyphenyl)isothiazolo[4,5-*b*]pyridin-3-yl)-2,6-dimethylmorpholine (23b). This compound was prepared according to the general procedure C using (2*S*,6*R*)-4-(5-chloroisothiazolo[4,5-*b*]pyridin-3-yl)-2,6-dimethylmorpholine **22b** (80.0 mg, 0.282 mmol), (3,4-dimethoxyphenyl)boronic acid (102.6 mg, 0.564 mmol), caesium carbonate (275.6 mg, 0.85 mmol), XPhos Pd G3 (23.9 mg, 0.028 mmol), dioxane (4.5 ml) and water (1.1 ml). The title compound was obtained as a light brown solid (89% yield, 96.7 mg).

MP 162–163 °C **¹H NMR** (600 MHz, CDCl₃) δ (ppm): 8.15 (d, J = 8.7 Hz, 1H), 7.81 (d, J = 8.7 Hz, 1H), 7.73 (d, J = 2.1 Hz, 1H), 7.59 (dd, J = 2.0, 8.3 Hz, 1H), 6.99 (d, J = 8.3 Hz, 1H), 5.04 (d, J = 12.2 Hz, 1H), 4.00 (s, 3H), 3.97 (s, 3H), 3.96–3.91 (m, 2H), 2.84 (dd, J = 2.2, 12.7 Hz, 2H), 1.30 (d, J = 6.3 Hz, 6H). **¹³C NMR** (151 MHz, CDCl₃) δ (ppm): 160.4, 153.8, 150.5, 149.5, 144.3, 143.6, 131.7, 129.3, 119.8, 118.3, 111.3, 109.8, 71.8, 56.2, 55.9, 53.0, 19.2. **HRMS** (ESI⁺): m/z calculated for C₂₀H₂₃N₃O₃S [M + H]⁺: 386.1533; found: 386.1526.

GAK LanthaScreen™ Eu binding assay. The final compounds were evaluated in the LanthaScreen™ binding assay, as provided on a fee-for-service basis by Thermo Fisher Scientific (SelectScreen™ Profiling Lab). Briefly, 10 titrations of test compound in DMSO are transferred to a 384-well plate. This was followed by the sequential addition of the kinase buffer (50 mM HEPES pH 7.5, 0.01% BRIJ-35, 10 mM MgCl and 1 mM EGTA), the 2× kinase antibody (Eu Anti GST) mixture and the 4× Tracer 222 solution. After shaking for 30 s and a 1 h incubation period at room temperature, the plate was read on a fluorescence plate reader. When the bound tracer in the active site was displaced by the test compound, fluorescence was not observed. The collected data were then compared to a 0% displacement control with pure DMSO and a 100% displacement control with staurosporine, a known inhibitor of GAK and plotted against the logarithmic concentration parameter. The IC₅₀ values were subsequently extracted.

Crystal structure determination of 16. X-ray intensity data for **16** were collected at 293(2) K on an Agilent SuperNova diffr-



actometer with Eos CCD detector using MoK α radiation. The images were processed (unit cell determination, intensity data integration, correction for Lorentz and polarization effects, and empirical absorption correction) using CrysAlisPRO.³⁴ Using Olex2,³⁵ the structure was solved with the ShelXT³⁶ structure solution program using Intrinsic Phasing and refined with the ShelXL³⁷ refinement package using full-matrix least-squares minimization on F^2 . All hydrogen atoms were placed in idealized positions and refined in the 'riding mode'. Non-hydrogen atoms were refined anisotropically and hydrogen atoms with isotropic temperature factors were fixed at 1.2 times U_{eq} of the parent atoms. The representation of molecular structures was done using the program Olex2.³⁵ Crystal data, data collection, and structure refinement details are summarized in Table S2 (ESI†). Crystallographic data for the structure reported herein have been deposited with the Cambridge Crystallographic Data Centre as supplementary publication number CCDC 2357539.†

Computational chemistry

X-ray cocrystal structure of GAK with compound 1. PDB entry 4Y8D was used for all modelling calculations.¹⁵

Superposition of compound 29. The conformational energy of compound 29 was optimized in vacuum conditions using Open MOPAC at the PM6 level.³² Compound 29 was then fitted onto compound 1 in the ATP binding site of GAK by adjusting the flexible dihedral angles.

Calculation of dipole moments. Open MOPAC was used to calculate the dipole moment of isothiazolo[4,3-*b*]pyridine 1 and isothiazolo[4,5-*b*]pyridine 29 at the PM6 while maintaining the binding conformations level.³²

Calculation of electrostatic potentials. The atoms were assigned charges and sizes from the amber gaff2 force field using pdb2pqr. Then the electrostatic potential around the molecules was calculated by APBS using default parameters.³⁸ The potentials were then visualized in chimera.^{31,33}

Author contributions

W. D. and S. D. J. supervised the project. W. D., D. S., S. D. J., and Y. I. conceptualised the idea. S. S. performed the investigation of the initially proposed synthetic method depicted in Scheme 1 in course of his Master's thesis. L. J. G. performed the synthesis of some reference isothiazolo[4,3-*b*]pyridines. Y. I. performed the rest of the experimental work. Y. I. wrote the first draft of the manuscript. Y. I. constructed the ESI. L. V. M. performed the crystal structure determination. M. F. performed molecular modeling and wrote the related part of manuscript and ESI. All authors discussed and agreed to the final version of the manuscript.

Data availability

Crystallographic data for compound 16 have been deposited with the Cambridge Crystallographic Data Centre as sup-

plementary publication number CCDC 2357539. Other data supporting this article have been included as part of the ESI.†

Conflicts of interest

There is no conflict of interest to declare.

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References

- 1 C. Huang, C. Ji and J. Wang, *Mol. Biol. Rep.*, 2023, **50**, 4645–4652.
- 2 C. X. Zhang, Å. E. Y. Engqvist-Goldstein, S. Carreno, D. J. Owen, E. Smythe and D. G. Drubin, *Traffic*, 2005, **6**, 1103–1113.
- 3 A. Young, *Semin. Cell Dev. Biol.*, 2007, **18**, 448–458.
- 4 H. T. McMahon and E. Boucrot, *Nat. Rev. Mol. Cell Biol.*, 2011, **12**, 517–533.
- 5 M. A. Edeling, C. Smith and D. Owen, *Nat. Rev. Mol. Cell Biol.*, 2006, **7**, 32–44.
- 6 D. Fessart and F. Delom, *Int. J. Cell Biol.*, 2011, **2011**, 14.
- 7 D. W. Lee, X. Zhao, F. Zhang, E. Eisenberg and L. E. Greene, *J. Cell Sci.*, 2005, **118**, 4311–4321.
- 8 G. Neveu, A. Ziv-Av, R. Barouch-Bentov, E. Berkerman, J. Mulholland and S. Einav, *J. Virol.*, 2015, **89**, 4387–4404.
- 9 E. Bekerman, G. Neveu, A. Shulla, J. Brannan, S. Y. Pu, S. Wang, F. Xiao, R. Barouch-Bentov, R. R. Bakken, R. Mateo, J. Govero, C. M. Nagamine, M. S. Diamond, S. De Jonghe, P. Herdewijn, J. M. Dye, G. Randall and S. Einav, *J. Clin. Invest.*, 2017, **127**, 1338–1352.
- 10 M. Karim, S. Saul, L. Ghita, M. K. Sahoo, C. Ye, N. Bhalla, C. W. Lo, J. Jin, J. G. Park, B. Martinez-Gualda, M. P. East, G. L. Johnson, B. A. Pinsky, L. Martinez-Sobrido, C. R. M. Asquith, A. Narayanan, S. De Jonghe and S. Einav, *Antiviral Res.*, 2022, **204**, 105367.
- 11 A. Beilina, I. N. Rudenko, A. Kaganovich, L. Civiero, H. Chau, S. K. Kalia, L. V. Kalia, E. Lobbestael, R. Chia,



- K. Ndukwe, J. Ding, M. A. Nalls, M. Olszewski, D. N. Hauser, R. Kumaran, A. M. Lozano, V. Baekelandt, L. E. Greene, J. M. Taymans, E. Greggio and M. R. Cookson, *Proc. Natl. Acad. Sci. U. S. A.*, 2014, **111**, 2626–2631.
- 12 M. Susa, E. Choy, X. Liu, J. Schwab, F. J. Hornicek, H. Mankin and Z. Duan, *Mol. Cancer Ther.*, 2010, **9**, 3342–3350.
- 13 M. R. Ray, L. A. Wafa, H. Cheng, R. Snoek, L. Fazli, M. Gleave and P. S. Rennie, *Int. J. Cancer*, 2006, **118**, 1108–1119.
- 14 J. Li, S. Kovackova, S. Pu, J. Rozenski, S. De Jonghe, S. Einav and P. Herdewijn, *MedChemComm*, 2015, **6**, 1666–1672.
- 15 S. Kovackova, L. Chang, E. Bekerman, G. Neveu, R. Barouch-Bentov, A. Chaikvad, C. Heroven, M. Šála, S. De Jonghe, S. Knapp, S. Einav and P. Herdewijn, *J. Med. Chem.*, 2015, **58**, 3393–3410.
- 16 S. Y. Pu, R. Wouters, S. Schor, J. Rozenski, R. Barouch-Bentov, L. I. Prugar, C. M. O'Brien, J. M. Brannan, J. M. Dye, P. Herdewijn, S. De Jonghe and S. Einav, *J. Med. Chem.*, 2018, **61**, 6178–6192.
- 17 R. Wouters, J. Tian, P. Herdewijn and S. De Jonghe, *ChemMedChem*, 2019, **14**, 237–254.
- 18 R. Wouters, S. Y. Pu, M. Froeyen, E. Lescrinier, S. Einav, P. Herdewijn and S. De Jonghe, *Eur. J. Med. Chem.*, 2019, **163**, 256–265.
- 19 B. Martinez-Gualda, S. Y. Pu, M. Froeyen, P. Herdewijn, S. Einav and S. De Jonghe, *Bioorg. Med. Chem.*, 2020, **28**, 115188.
- 20 B. Martinez-Gualda, S. Saul, M. Froeyen, D. Schols, P. Herdewijn, S. Einav and S. De Jonghe, *Eur. J. Med. Chem.*, 2021, **213**, 113158.
- 21 Y. Ivanova, M. Smoljo, S. De Jonghe and W. Dehaen, *ARKIVOC*, 2024, 202312146.
- 22 F. Jakob, J. Alen, S. Kruger, D. Friebe, S. Hennen and Gruenenthal GMBH, Substituted pyrrolidine amides IV, *World Intellectual Property Organization*, 2020254551, 2020.
- 23 R. T. Skerlj, E. M. J. Bourque, W. J. Greenlee, P. T. Lansbury and Lysosomal Therapeutics INC, Substituted imidazo[1,2-b]pyridazines, substituted imidazo[1,5-b]pyridazines, related compounds, and their use in the treatment of medical disorders, *World Intellectual Property Organization*, WO2017192930, 2017..
- 24 S. Wang, E. Kumpulainen, J. Pystynen, A. Pohjakallio, A. Haikarainen and Orion Corporation, 2-(1-Heteroaryl)piperazin-4-yl)methyl-1,4-benzodioxane derivatives as α_2c antagonists, *World Intellectual Property Organization*, WO2016193551, 2016..
- 25 G. Zhou, Y. Wei, W. Li, H. Yuan, K. C. Liu and Acerand Therapeutics (Shanghai) CO, Acerand Therapeutics (Hong Kong) limited, Pyrazole-containing derivative, pharmaceutically acceptable salt thereof, preparation method therefor and application thereof *World Intellectual Property Organization*, WO2023116865, 2023..
- 26 N. Rival, A. Albornoz Grados, L. Schiavo, F. Colobert and G. Hanquet, *Tetrahedron Lett.*, 2015, **56**, 6823–6826.
- 27 C. S. Lebakken, S. M. Riddle, U. Singh, W. J. Frazee, H. C. Eliason, Y. Gao, L. J. Reichling, B. D. Marks and K. W. Vogel, *SLAS Discovery*, 2009, **14**, 924–935.
- 28 S. Verdonck, S. Y. Pu, F. J. Sorrell, J. M. Elkins, M. Froeyen, L. J. Gao, L. I. Prugar, D. E. Dorosky, J. M. Brannan, R. Barouch-Bentov, S. Knapp, J. M. Dye, P. Herdewijn, S. Einav and S. De Jonghe, *J. Med. Chem.*, 2019, **62**, 5810–5831.
- 29 P. Zhou, F. Tian, F. Lv and Z. Shang, *Proteins: Struct., Funct., Bioinf.*, 2009, **76**, 151–163.
- 30 A. Pizzi, A. Daolio, R. Beccaria, N. Demitri, F. Viani and G. Resnati, *Chem. – Eur. J.*, 2023, **29**, e202300571.
- 31 E. F. Pettersen, T. D. Goddard, C. C. Huang, G. S. Couch, D. M. Greenblatt, E. C. Meng and T. E. Ferrin, *J. Comput. Chem.*, 2004, **25**, 1605–1612.
- 32 J. J. P. Stewart, *J. Mol. Model.*, 2007, **13**, 1173–1213.
- 33 E. C. Meng, T. D. Goddard, E. F. Pettersen, G. S. Couch, Z. J. Pearson, J. H. Morris and T. E. Ferrin, *Protein Sci.*, 2023, **32**, e4792.
- 34 *CrysAlis PRO*, Agilent Technologies UK Ltd, Yarnton, Oxfordshire, England, 2012.
- 35 O. V. Dolomanov, L. J. Bourhis, R. J. Gildea, J. A. Howard and H. Puschmann, *J. Appl. Crystallogr.*, 2009, **42**, 339–341.
- 36 G. M. Sheldrick, *Acta Crystallogr., Sect. A: Found. Adv.*, 2015, **71**, 3–8.
- 37 G. M. Sheldrick, *Acta Crystallogr., Sect. C: Struct. Chem.*, 2015, **71**, 3–8.
- 38 E. Jurrus, D. Engel, K. Star, K. Monson, J. Brandi, L. E. Felberg, D. H. Brookes, L. Wilson, J. Chen, K. Liles, M. Chun, P. Li, D. W. Gohara, T. Dolinsky, R. Konecny, D. R. Koes, J. E. Nielsen, T. Head-Gordon, W. Geng, R. Krasny, G. W. Wei, M. J. Holst, J. A. McCammon and N. A. Baker, *Protein Sci.*, 2018, **27**, 112–128.

