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# A bioinspired oxoiron(IV) motif supported on a N<sub>2</sub>S<sub>2</sub> macrocyclic ligand<sup>+</sup>

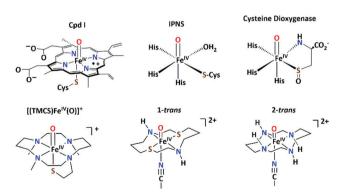
Jennifer Deutscher, <sup>(b)</sup><sup>a</sup> Philipp Gerschel,<sup>b</sup> Katrin Warm, <sup>(b)</sup><sup>a</sup> Uwe Kuhlmann,<sup>c</sup> Stefan Mebs, <sup>(b)</sup><sup>d</sup> Michael Haumann, <sup>(b)</sup><sup>d</sup> Holger Dau, <sup>(b)</sup><sup>d</sup> Peter Hildebrandt, <sup>(b)</sup><sup>c</sup> Ulf-Peter Apfel <sup>(b)</sup>\*<sup>be</sup> and Kallol Ray <sup>(b)</sup>\*<sup>a</sup>

A mononuclear oxoiron(IV) complex 1-*trans* bearing two equatorial sulfur ligations is synthesized and characterized as an active-site model of the elusive sulfur-ligated Fe<sup>IV</sup>=O intermediates in nonheme iron oxygenases. The introduction of sulfur ligands weakens the Fe=O bond and enhances the oxidative reactivity of the Fe<sup>IV</sup>=O unit with a diminished deuterium kinetic isotope effect, thereby providing a compelling rationale for nature's use of the *cis*-thiolate ligated oxoiron(IV) motif in key metabolic transformations.

Sulfur-ligated oxoiron(iv) centers are proposed as key oxidants in the catalytic cycles of various heme and non-heme iron oxygenases (Scheme 1).<sup>1–3</sup> Iron(IV)–oxo porphyrin  $\pi$ -cation radical (Cpd I) intermediates containing a thiolate ligand trans to the oxo group have been isolated and spectroscopically characterized in a number of heme enzymes.<sup>4-6</sup> The increased basicity of the oxoiron(IV) core caused by the strong electron donation from the trans-sulfur ligand, is discussed as a strategy to perform hydrogen atom abstraction by Cpd I at a lower redox potential without performing oxidative destruction of the surrounding enzyme environment.<sup>5,7-10</sup> However, similar knowledge on the effect of cis-sulfur ligands on the reactivity of oxoiron(IV) cores in nonheme enzymes is lacking. Notably, identification of cis thiolateligated oxoiron(w) species remained elusive in biology, although they are suggested to be reactive intermediates for a wide range of chemical transformations, including sulfur-oxygenation,

hydrogen-atom abstraction, and C–S bond formation reactions in non-heme enzymes.

For over 40 years, small-molecule complexes synthesized as active-site models of the high-valent intermediates in heme and non-heme oxygenases have advanced our understanding of the catalytic cycles.<sup>3,11–20</sup> Despite these efforts, the synthesis of an oxoiron(IV) porphyrin complex with a thiolate ligand has not yet been achieved. Furthermore,  $[(TMCS)Fe^{IV}(O)]^+$  (TMCS = 1-mercaptoethyl-4,8,11-trimethyl-1,4,8,11-tetraazacyclotetra-decane; (Scheme 1)), represents the only synthetic complex<sup>21</sup> thus far to model the RS-Fe<sup>IV</sup>=O unit associated with the active oxidants of cytochrome P450<sup>2,6</sup> and chloroperoxidase.<sup>4,5,10</sup> Similarly, a recently reported  $[(Me_3TACN)Fe^{IV}(O)(S_2Si(CH_3)_2)]$  (Me<sub>3</sub>TACN = 1,4,7-trimethyl-1,4,7-triazacyclononane)<sup>13</sup> complex represents the only model complex for the postulated oxoiron(IV) core containing a sulfur ligation cis to the oxo group in non-heme oxygenases.<sup>22,23</sup> However, the thermal instability of the compound has prevented any reactivity studies. Herein we report the synthesis and characterization of the  $S = 1 \text{ Fe}^{\text{IV}} = \text{O complex } [(\text{dithiacyclam})\text{Fe}^{\text{IV}}(\text{O})(\text{CH}_3\text{CN})]^{2+1}$ (1-trans, dithiacyclam<sup>24,25</sup> = 1,8-dithia-4,11-diazacyclotetradecane), which contains two thioether sulfur coordination sites poised cis to the oxo group. A comparative study between 1-trans,



<sup>a</sup> Institut für Chemie Humboldt-Universität zu Berlin, Brook-Taylor-Str. 2, 12489, Berlin, Germany. E-mail: kallol.ray@hu-berlin.de

<sup>b</sup> Anorganische Chemie 1 Ruhr-Universität Bochum, Universitätsstraße 150, 44780, Bochum, Germany

Scheme 1 Top: Proposed structures of the thiolate-ligated oxoiron(IV) reactive intermediates in biology; bottom: structures of [(TMCS)Fe<sup>IV</sup>(O)]<sup>+</sup>, 1-trans and 2-trans.

<sup>&</sup>lt;sup>c</sup> Institut für Chemie Technische, Universität Berlin, Fakultät II Straße des 17, Juni 135, 10623, Berlin, Germany

<sup>&</sup>lt;sup>d</sup> Institut für Physik Freie, Universität Berlin, Arnimallee 14, 14195 Berlin, Germany

<sup>&</sup>lt;sup>e</sup> Department of Electrosynthesis, Fraunhofer UMSICHT, Osterfelder Str. 3,

<sup>46047</sup> Oberhausen, Germany

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containing a N<sub>2</sub>S<sub>2</sub> macrocyclic ligand, and [Fe<sup>IV</sup>(O)(cyclam)(CH<sub>3</sub>CN)]<sup>2+</sup> (2-*trans*; cyclam = 1,4,8,11-tetraazacyclotetradecane; (Scheme 1)),<sup>26</sup> based on the popular N<sub>4</sub>-donor cyclam ligand, provides some insight how *cis*-sulfur coordination influences the reactivity and spectroscopic properties of the oxoiron(rv) unit.

Combining the tetradentate dithiacyclam ligand with  $Fe(OTf)_2(CH_3CN)_2$  in acetonitrile yielded the iron(II) complex in two isomeric forms [Fe<sup>II</sup>(dithiacvclam)(CH<sub>3</sub>CN)<sub>2</sub>](OTf)<sub>2</sub> (**1a**-*trans*) and [Fe<sup>II</sup>(dithiacyclam)(OTf)<sub>2</sub>] (**1a**-*cis*). Crystals suitable for X-ray diffraction analysis were grown by vapour diffusion of diethyl ether into an acetonitrile solution of the iron(II) complex at -15 °C for **1a**-*cis* or at -40 °C for **1a**-*trans*. The X-ray structure of 1a-trans displays a six-coordinate geometry with axially bound CH<sub>3</sub>CN ligands (Fig. 1; Table S2, ESI<sup>†</sup>). The N<sub>2</sub>S<sub>2</sub> donor atoms of dithiacyclam occupy the equatorial coordination sites and show average Fe-S and Fe-N distances of 2.252(6) Å and 1.985(18) Å, respectively. In contrast, in 1a-cis the sulfur donor atoms occupy the axial coordination sites and show average Fe-S distances of 2.466(14) Å (Table S1, ESI<sup>+</sup>); the nitrogen atoms of the dithiacyclam and the oxygen atoms of the triflate (OTf) anions occupy equatorial positions with average Fe-N and Fe-O distances of 2.2085(4) Å and 2.143(3) Å, respectively. The zero-field Mößbauer spectrum of 1a-cis in acetone (Fig. S1, ESI<sup>+</sup>) at 15 K reveals a single quadrupole doublet with an isomer shift of  $\delta = 1.10 \text{ mm s}^{-1}$  and a large quadrupole splitting of  $(\Delta E_{\rm O})$  = 3.26 mm s<sup>-1</sup>, demonstrating that the iron( $\pi$ ) center remains in the high-spin configuration (S = 2). In addition, the <sup>1</sup>H- and <sup>19</sup>F-NMR spectra (Fig. S2, ESI<sup>†</sup>) of 1a-cis in  $d_6$ -acetone at -85 °C display paramagnetically shifted peaks, indicative of the coordination of both OTf anions and further supporting the high-spin iron(II) assignment. In CH<sub>2</sub>Cl<sub>2</sub>/CH<sub>3</sub>CN solution, CH<sub>3</sub>CN gradually replaces the bound triflates of 1a-cis to form **1a**-*trans* with an S = 0 Fe<sup>II</sup> ground-state, as evident from <sup>1</sup>H-NMR (at -85 °C), which reveals peaks between 0 and 4 ppm (Fig. S3, ESI,<sup> $\dagger$ </sup> left), and <sup>19</sup>F-NMR (at -85 °C; Fig. S3, ESI,<sup> $\dagger$ </sup> right), which shows a singlet at -79.0 ppm corresponding to free OTf anions. Freezing the solution leads to the partial re-binding of OTf anion to Fe<sup>II</sup>; zero-field Mößbauer measurement (Fig. S4, ESI<sup>†</sup>) at 15 K shows a major quadrupole doublet with a new high-spin Fe<sup>II</sup> signal ( $\delta$  = 1.15 mm s<sup>-1</sup> and  $\Delta E_Q$  = 2.31 mm s<sup>-1</sup>; 73%) with a significantly reduced  $\Delta E_Q$  relative to **1a-cis**, presumably corresponding to the *trans*-[(dithiacyclam)Fe<sup>II</sup>(OTf)(CH<sub>3</sub>CN)]<sup>+</sup> complex. An additional doublet with  $\delta$  = 0.52 mm  $s^{-1}$  and

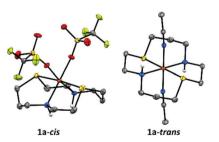


Fig. 1 Molecular structures of **1a**-*cis* and **1a**-*trans* obtained by XRD. Atoms are displayed as thermal ellipsoids at 50% probability level; triflate counter ions for **1a**-*trans* are omitted for clarity. Atom types: Fe: orange; N: blue; C: grey; O: red; S: yellow; F: green; H: white.

 $\Delta E_{\rm Q} = 0.26 \text{ mm s}^{-1}$  corresponds to the low-spin  $S = 0 \text{ Fe}^{\rm II}$  center in **1a**-*trans* (27%). Thus, CH<sub>3</sub>CN-binding favours the *trans* configuration and the coordination of both CH<sub>3</sub>CN is necessary for stabilizing the low-spin Fe<sup>II</sup> state in **1a**.

A solution of **1a**-*trans* in a  $CH_2Cl_2/CH_3CN$  solvent mixture (95:5) at -85 °C with 4 equiv. of 2-(tert-butylsulfonyl)iodosylbenzene  $({}^{t}BuSO_{2}C_{6}H_{4}IO, {}^{s}PhIO)^{27}$  led to the formation of a pale green intermediate **1-***trans* ( $t_{1/2}$  = 10000 s at -65 °C) with absorption maxima at 596 nm ( $\varepsilon_{max}$  = 226 M<sup>-1</sup> cm<sup>-1</sup>) and 815 nm ( $\varepsilon_{max}$  = 549  $M^{-1}$  cm<sup>-1</sup>), which are typical of S = 1 oxoiron(iv) cores (Fig. 2; left).11 The characteristic near-infrared band in 1-trans (a) 815 nm is significantly red-shifted relative to that in 2-trans,<sup>26</sup> which is consistent with a weakened equatorial field in 1-trans. Notably, in the absence of CH<sub>3</sub>CN, **1-***trans* was not generated.<sup>28</sup> An electron spray ionization mass spectrum (Fig. S5, ESI<sup>+</sup>) of **1-***trans* exhibited a signal at m/z = 794.96, consistent with its formulation as  $\{[(dithiacyclam)Fe^{IV}(O)(OTf)](^{S}PhIO)\}^{+}$  (m/z calc =794.97), which is shifted by 4 units to m/z = 798.97, when sPhI<sup>18</sup>O was used to generate 1-trans. <sup>19</sup>F-NMR (at -85 °C in a 95:5 mixture of CD<sub>2</sub>Cl<sub>2</sub> and CD<sub>3</sub>CN) shows a singlet at -77.0 ppm corresponding to the free OTf anions in 1-trans (Fig. S6, ESI<sup>†</sup>). The zero-field Mößbauer spectrum of 1-trans in frozen acetone/ CH<sub>2</sub>Cl<sub>2</sub>/CH<sub>3</sub>CN solution and recorded at 15 K exhibits a doublet representing about 84% of the iron with  $\Delta E_{\rm O}$  = 1.21 mm s<sup>-1</sup> and  $\delta = 0.13 \text{ mm s}^{-1}$  corresponding to the presence of an Fe<sup>IV</sup> center (Fig. 2; right); the remaining 16% of the signals with  $\Delta E_{\rm O}$  = 1.57 mm s<sup>-1</sup> and  $\delta$  = 0.55 mm s<sup>-1</sup> correspond to a high-spin Fe<sup>III</sup> product, arising from the decay of 1-trans.

The Fe K-edge X-ray absorption (Fig. S7 and Table S4, ESI<sup>†</sup>) spectrum of **1-***trans* reveals a K-edge energy of 7122.7 eV, which is lower relative to **2-***trans* (7123.9 eV). Furthermore, the pre-edge transition in **1-***trans* is less intense, which may reflect a less covalent Fe=O bond in **1-***trans* relative to **2-***trans*. The resonance Raman (rR) spectrum of **1-***trans* exhibits a  $\nu$ (Fe=O) stretching mode at 793 cm<sup>-1</sup> (Fig. 2, inset), which is red-shifted by 49 cm<sup>-1</sup> relative to that in **2-***trans* ( $\nu$ (Fe=O) = 842 cm<sup>-1</sup>),<sup>26</sup> thereby demonstrating an elongation of the Fe=O bond by ~0.02 Å in **1-***trans* relative to **2-***trans*.<sup>29</sup> However, within the error of the

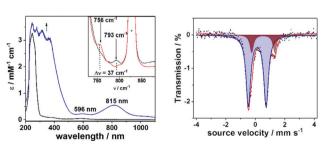
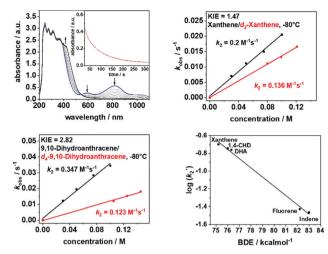


Fig. 2 Left: UV-Vis-spectrum of **1a-trans** (black) and **1-trans** (blue) in a 95:5 mixture of CH<sub>2</sub>Cl<sub>2</sub> and CH<sub>3</sub>CN at -90 °C; the inset shows the resonance Raman spectra of <sup>16</sup>O- (black) and <sup>18</sup>O-labelled (red) **1-trans** (4 mM) upon 413 nm irradiation at -90 °C. Solvent signal is indicated by an asterisk. Right: Zero-field Mößbauer spectrum (black) of a frozen sample of **1-trans** in a solvent mixture of acetone/CH<sub>2</sub>Cl<sub>2</sub>/CH<sub>3</sub>CN (10:0.95:0.05) and simulation (red) with  $\delta$  = 0.13 mm s<sup>-1</sup> and  $\Delta E_{\Omega}$  = 1.21 mm s<sup>-1</sup> for the main species (blue, 84%). The minor species with  $\delta$  = 0.55 mm s<sup>-1</sup> and  $\Delta E_{\Omega}$  = 1.57 mm s<sup>-1</sup> corresponds to the decay of **1-trans** (brown, 16%).



**Fig. 3** Top left: Changes in the UV-Vis-spectra of a 1 mM solution of **1-***trans* in CH<sub>2</sub>Cl<sub>2</sub>/CH<sub>3</sub>CN (95:5) at -80 °C upon addition of 100 eq xanthene; the inset shows the time trace for the decay of the 815 nm band and its pseudo-first order fit; top right: plots of the pseudo-first order rate constants  $k_{obs}$  vs. the substrate concentrations for xanthene and  $d_2$ -xanthene in order to determine the kinetic isotopic effect (KIE) for the reaction of **1-***trans* with xanthene; bottom left: plots of the pseudo-first order 9,10-dihydroanthracene (DHA) and  $d_4$ -DHA in order to determine the kinetic isotopic effect (KIE) for the logarithms of the second-order rate constants  $k_2'$  vs. the C-H BDEs of the substrates with **1-***trans* in CH<sub>2</sub>Cl<sub>2</sub>/CH<sub>3</sub>CN (95:5).

extended X-ray absorption fine structure (EXAFS) analysis, the Fe=O distances in 1-trans and 2-trans are not discernible; in both cases a distance of 1.67  $\pm$  0.02 Å has been obtained.<sup>26</sup> The DFT optimized geometry of 1-trans in the S = 1 state slightly underestimates the Fe-O bond (calculated (a) 1.655 Å) from the EXAFS data (Tables S5 and S6, ESI<sup>+</sup>), and as a result the calculated  $\nu$ (Fe–O) is overestimated. This is a common problem that is encountered in the oxoiron(iv) chemistry.<sup>8,13</sup> However, a geometry scan of the Fe-O bond length (Table S6, ESI<sup>+</sup>) reveals a relatively flat surface potential, with the structures exhibiting Fe-O bond lengths of 1.66–1.70 Å being within 1 kcal mol<sup>-1</sup> in energy from the lowest energy structure. In particular, a constrained optimization with a fixed Fe-O distance (a) 1.68 Å for 1-trans gives a calculated  $\nu$ (Fe–O) of 794 cm<sup>-1</sup>, in good agreement with the experiment. Similarly, a constrained geometry with an Fe-O distance of 1.66 Å can account for the experimental  $\nu$ (Fe–O) of 842 cm<sup>-1</sup> for 2-*trans*. In summary, a 49 cm<sup>-1</sup> red-shift in the  $\nu$ (Fe–O) of 1-*trans* relative to 2-*trans*, translates to an Fe=O elongation of only 0.02 Å (based on both constrained DFT optimization and Badger's rule<sup>29</sup>), which is not clearly discernible within the error of the EXAFS analysis, but is reflected in a less-intense pre-edge transition at the Fe K-edge.

The introduction of the equatorial sulfur ligands also exhibits a significant effect on the reactivity of the oxoiron(iv) unit (Fig. 3 and Fig.s S8-S12, ESI<sup>+</sup>). In reactions with substrates containing C-H bonds like xanthene, 1,4-cyclohexadiene (CHD), dihydroanthracene (DHA), fluorene and indene, 1-trans reacts at least 3-4 orders of magnitude faster than 2-trans (Table 1). Furthermore, low kinetic isotope effects (KIEs) of 1.47 and 2.82 were determined in the reaction of 1-trans with xanthene (Fig. 3, top right) and DHA (Fig. 3, bottom left), respectively, which are in contrast to the previously reported value of 20.0 for reaction of xanthene with 2-trans.<sup>26</sup> Nevertheless, when the logarithms of the second order rate constants  $(k_2)$  were plotted vs. the BDE C-H values of the substrates, the linear correlation previously reported for 2-trans were found to be also valid for 1-trans (Fig. 3, bottom right). Thus, although proton-transfer is involved in the ratedetermining step of the oxidation of C-H bonds by 1-trans, the large tunneling contribution in hydrogen atom abstraction (HAA), which is observed in 2-trans and in most high-valent metal-oxo mediated HAA reactions,<sup>11,12,30</sup> is not applicable for 1-trans. In particular, in a previous study the axial thiolate ligand of [Fe<sup>IV</sup>(O)(TMCS)]<sup>+</sup> has been suggested to play a unique role in facilitating tunnelling, thereby resulting in a large KIE of 80 for DHA oxidation.<sup>31</sup> A contrasting effect is now demonstrated for the equatorial sulphur ligation, which reduces the tunnelling contribution to a minimum. The effect of N versus S donors in an otherwise identical ligand environment is also reflected in the higher oxygen atom transfer (OAT) ability of 1-trans relative to 2-trans (Table 1).

In summary, a minor 0.02 Å elongation of the Fe=O bond upon introduction of the equatorial sulfur ligands is shown to have a dramatic influence on the spectroscopic (Table S7, ESI<sup>†</sup>) and oxidative reactivity properties of the Fe<sup>IV</sup>=O unit. Notably, **1**-*trans*, similar to the previously reported<sup>13</sup> [(Me<sub>3</sub>TACN)-Fe<sup>IV</sup>(O)(S<sub>2</sub>Si(CH<sub>3</sub>)<sub>2</sub>)] complex features a very low  $\nu$ (Fe–O), which establishes a general trend of the activation of the Fe–O bond in oxoiron(IV) complexes involving *cis*-sulphur ligands. The enhanced reactivity of **1**-*trans* relative to **2**-*trans*, can presumably

Table 1      Reactivity comparison of 1-trans and 2-trans				
Substrate	$BDE^{a}$ [kcal mol <sup>-1</sup> ]	$k_2 \left[ \mathrm{M}^{-1} \mathrm{s}^{-1} \right]$ 1-trans	$k_2[M^{-1}s^{-1}]$ 2-trans	Product $(yield)^d$
Xanthene	75.2	$205^{b}$	$1.1 imes 10^{-1}$	Xanthone (36%)
1,4-CHD	76	$370^{b}$	$9.7 imes10^{-2}$	
DHA	76.3	$355^{b}$	$4.9 imes10^{-2}$	Anthracene (48%)
Fluorene	82.2	$18.2^{c}$	$\textbf{7.1} \times \textbf{10}^{-3}$	Fluorenone (11%)
Indene	83	17.6 <sup>c</sup>	$5.8\times10^{-3}$	Indenone
Thioanisol	_	87 <sup>c</sup>	_	
PPh <sub>3</sub>	_	n.d	5.9	$OPPh_3$ (28%)

<sup>*a*</sup> Values taken from ref. 32. <sup>*b*</sup> Values calculated for 20 °C from experimental values determined at -80 °C using van't Hoff equation. <sup>*c*</sup> Values calculated for 15 °C from experimental values determined at -65 °C using van't Hoff equation. <sup>*d*</sup> Specified yields correspond to the reactivity of **1-***trans*. n.d: the reaction was too fast for kinetic studies.

be attributed to the positive shift in the redox potential upon sulphur ligation, as evident from cyclic voltammetry experiments, which shows a 170 mV positive shift in the  $\mbox{Fe}^{2+/3+}$ potential (Fig. S13, ESI<sup>+</sup>) in 1a-trans relative to 2a-trans. In addition, a change in mechanism in the C-H bond oxidation reactions from HAA to proton coupled electron transfer (PCET)<sup>33</sup> is evident from the reduction of KIE from a value of 20 in 2-trans to 1.47 in **1-trans** (using xanthene as a substrate). This drastic downshift in KIE is unique for sulfur substitution and is not observed in the oxygen substituted oxoiron(IV) center.<sup>34</sup> Understanding this lowering of KIE will require further experimental and computational work. However the significant effect of the equatorial sulfur ligation on the physical and chemical properties of oxoiron(iv) cores may provide a compelling rationale for nature's use of the *cis*-thiolate ligated oxoiron(IV) motif in key metabolic transformations that involve the activation of strong C-H bonds.

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### Conflicts of interest

There are no conflicts to declare.

#### Notes and references

- 1 M. Costas, M. P. Mehn, M. P. Jensen and L. Que, *Chem. Rev.*, 2004, 104, 939–986.
- 2 B. Meunier, S. P. de Visser and S. Shaik, *Chem. Rev.*, 2004, **104**, 3947–3980.
- 3 J. B. Gordon and D. P. Goldberg, *Reference Module in Chemistry, Molecular Sciences and Chemical Engineering*, Elsevier, 2020, DOI: 10.1016/B978-0-12-409547-2.14906-6.
- 4 M. T. Green, J. H. Dawson and H. B. Gray, Science, 2004, 304, 1653-1656.
- 5 K. L. Stone, R. K. Behan and M. T. Green, Proc. Natl. Acad. Sci. U. S. A., 2005, 102, 16563–16565.
- 6 J. Rittle and M. T. Green, Science, 2010, 330, 933-937.
- 7 F. Ogliaro, S. P. de Visser and S. Shaik, J. Inorg. Biochem., 2002, 91, 554-567.
- 8 M. T. Green, J. Am. Chem. Soc., 2006, 128, 1902-1906.

- 9 R. K. Behan, L. M. Hoffart, K. L. Stone, C. Krebs and M. T. Green, J. Am. Chem. Soc., 2006, 128, 11471–11474.
- 10 K. L. Stone, R. K. Behan and M. T. Green, *Proc. Natl. Acad. Sci. U. S. A.*, 2006, **103**, 12307–12310.
- 11 X. Engelmann, I. Monte-Pérez and K. Ray, Angew. Chem., Int. Ed., 2016, 55, 7632–7649.
- 12 R. A. Baglia, J. P. T. Zaragoza and D. P. Goldberg, *Chem. Rev.*, 2017, 117, 13320–13352.
- 13 J. B. Gordon, A. C. Vilbert, I. M. DiMucci, S. N. MacMillan, K. M. Lancaster, P. Moënne-Loccoz and D. P. Goldberg, J. Am. Chem. Soc., 2019, 141, 17533–17547.
- 14 L. Vicens, G. Olivo and M. Costas, ACS Catal., 2020, 10, 8611-8631.
- 15 J. Chen, Z. Jiang, S. Fukuzumi, W. Nam and B. Wang, *Coord. Chem. Rev.*, 2020, 421, 213443.
- 16 T. Devi, Y.-M. Lee, W. Nam and S. Fukuzumi, *Coord. Chem. Rev.*, 2020, 410, 213219.
- 17 S. Fukuzumi, K.-B. Cho, Y.-M. Lee, S. Hong and W. Nam, *Chem. Soc. Rev.*, 2020, **49**, 8988–9027.
- 18 S. Hong, Y.-M. Lee, K. Ray and W. Nam, Coord. Chem. Rev., 2017, 334, 25-42.
- 19 W. Nam, Y.-M. Lee and S. Fukuzumi, Acc. Chem. Res., 2018, 51, 2014–2022.
- 20 M. Sankaralingam, Y.-M. Lee, W. Nam and S. Fukuzumi, *Coord. Chem. Rev.*, 2018, 365, 41–59.
- 21 M. R. Bukowski, K. D. Koehntop, A. Stubna, E. L. Bominaar, J. A. Halfen, E. Münck, W. Nam and L. Que, *Science*, 2005, **310**, 1000–1002.
- 22 D. Kumar, W. Thiel and S. P. de Visser, J. Am. Chem. Soc., 2011, 133, 3869–3882.
- 23 S. Aluri and S. P. de Visser, J. Am. Chem. Soc., 2007, 129, 14846-14847.
- 24 P. Gerschel, B. Battistella, D. Siegmund, K. Ray and U.-P. Apfel, *Organometallics*, 2020, **39**, 1497–1510.
- 25 P. Gerschel, K. Warm, E. Farquhar, U. Englert, M. Reback, D. Siegmund, K. Ray and U.-P. Apfel, *Dalton Trans.*, 2019, **48**, 5923–5932.
- 26 D. Kass, T. Corona, K. Warm, B. Braun-Cula, U. Kuhlmann, E. Bill, S. Mebs, M. Swart, H. Dau and M. Haumann, *J. Am. Chem. Soc.*, 2020, 142, 5924–5928.
- 27 D. Macikenas, E. Skrzypczak-Jankun and J. D. Protasiewicz, J. Am. Chem. Soc., 1999, 121, 7164–7165.
- 28 Since we start with a pure solution of 1a-*trans*, the sole product that is formed is 1-*trans* (see also ref. 33). Oxidation of a solution of 1a-*cis* leads to only iron(m)products; so we presume that 1-*cis* is too unstable to be trapped in high-yields.
- 29 Acoording to Badger's rule (R. M. Badger, *J. Chem. Phys.*, 1935, 3, 710–714), which is previously shown to be valid for oxoiron(rv) complexes (see ref. 8).
- 30 (a) D. Mandal and S. Shaik, J. Am. Chem. Soc., 2016, 138, 2094-2097;
  (b) D. Mandal, D. Mallick and S. Shaik, Acc. Chem. Res., 2018, 51, 107-117;
  (c) E. J. Klinker, S. Shaik, H. Hirao and L. Que Jr., Angew. Chem., Int. Ed., 2009, 48, 1291-1295.
- 31 J. E. M. N. Klein, D. Mandal, W.-M. Ching, D. Mallick, L. Que, Jr. and S. Shaik, *J. Am. Chem. Soc.*, 2017, **139**, 18705–18713.
- 32 Y.-R. Luo, Comprehensive Handbook of Chemical Bond Energies, CRC Press, 2007, DOI: 10.1201/9781420007282.
- 33 J. M. Meyer, Acc. Chem. Res., 2010, 44, 36-46.
- 34 I. Monte-Pérez, X. Engelmann, Y.-M. Lee, M. Yoo, E. Kumaran, E. R. Farquhar, E. Bill, J. England, W. Nam, M. Swart and K. Ray, *Angew. Chem., Int. Ed.*, 2017, 56, 14384–14388.