

Cite this: *J. Mater. Chem. B*, 2022, 10, 9202

Nano-assemblies of phosphonium-functionalized diblock copolymers with fabulous antibacterial properties and relationships of structure-activity†

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As a novel antimicrobial material, quaternary phosphonium salts (QPSs) have been drawing close attention because of their excellent antimicrobial capacity with high activity and low bacterial survivability. Polymeric QPSs (PQPSs) also exhibit selectivity and long-term stability, however the polymerization of QPSs is severely challenged by low controllability and narrow selectivity of cation type. In this study, high-conversion RAFT polymerization is employed to prepare innovative phosphonium-functionalized diblock copolymers (PFDCs) with desired molecular weights and particle sizes. The excellent antibacterial activity of the PFDCs achieves lowest MIC values of 40 and 60 $\mu\text{g mL}^{-1}$ (i.e., 1.4 and 2.2 $\mu\text{mol L}^{-1}$) against *E. coli* and *S. aureus*, respectively. Mixing with an ink, dye, and latex coating does not weaken the antibacterial activity of the PFDCs, which inhibited 99.9% *E. coli*, showing broad applicability in different media. The effects of the cation type, synthesis medium, crosslinking content, and particle size on the morphology and antibacterial activity are studied. In summary, the RAFT polymerization of QPSs through the versatile design of ionic liquid monomers and the polymerization-induced self-assembly (PISA) method for constructing nano-assemblies with various micromorphology and particle size provides an exceedingly efficient way to build up multifunctional and multi-morphological polymeric nano-objects that open up vast possibilities in the fields of antibiotics, drug delivery, templated synthesis, and catalysis.

Received 20th August 2022,
Accepted 17th October 2022

DOI: 10.1039/d2tb01778d

rsc.li/materials-b

1. Introduction

Evolving and increasing numbers of superbugs (e.g., MRSA, MDRSP, VRE, MRAB, etc.), which have adapted to antibiotics owing to their abuse during infection treatments, are becoming one of the major risks that severely threaten human health and even the environment.¹ The World Health Organization (WHO) reported that over 700 thousand people are killed by antibiotic-resistant bacteria every year, including infants and children.² Fading disinfection or dysfunction of traditional antibiotics has significantly weakened the therapy of infections, especially against some vital superinfections.³ As a result, reinforcing the antimicrobial activity of antibiotics is now imperative to curb the havoc wreaked by superbugs. From the perspective of chemistry, creating specific cations with high polarity, e.g.,

quaternary ammonium ions, imidazole ions, and haloamides, is attractive and practicable due to the high antibacterial activity of these large cations.^{4–6} Polymeric antibiotics designed with the introduction of high polar cation ions are now gaining more and more attention due to their low toxicity, improved biodistribution, pharmacokinetics, biofilm penetration capacity, and bacterial selectivity, revealing great prospects as a new generation antibiotic.^{7,8} Compared to conventional quaternary cationic polymers, PQPSs have more effective antimicrobial activity thanks to the larger phosphorus atom and lower electronegativity that induce stronger polarizability.⁹ Moreover, the high chemical and environmental stability of quaternary phosphonium salts allow them to be applied under various conditions and in different applications, such as the design of ionic liquids and nonviral gene delivery.^{10,11}

However, the synthesis of PQPSs is restricted by its low controllability, high price, and limited variety, causing their development to lag behind that of quaternary ammonium salts, imidazole salts, and pyrrolidine salts. The ionic liquids are novel organic solvents with a low melting point that have already been widely used in the preparation of functional polymers.¹² High designability and variety of the chemical

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† Electronic supplementary information (ESI) available. See DOI: <https://doi.org/10.1039/d2tb01778d>

structure of both cations and anions can enable the synthesis of many polyelectrolytes, such as polycations, polyanions, polyzwitterions, and polyampholytes.² Bringing quaternary phosphonium ions into an ionic liquid system significantly extends the variety of the cation type, which can be further designed as a highly selective antibiotic.¹³ On the other hand, employing reversible addition-fragmentation chain transfer (RAFT) polymerization in the preparation of poly(ionic liquids) has also been reported, as well as the advantages of high reactivity, high conversion rate, narrow molecular weight distribution, and good repeatability.¹⁴ RAFT polymerization-induced self-assembly (PISA), which realizes the precise manipulation of both molecular weight and particle morphology and size,¹⁵ is now commonly used to obtain multifunctional polymeric nanoparticles, *e.g.* micelles and vesicles,¹⁶ and it is also one of the optimal methods for the expandable design of PQPSs.

Our work aimed to prepare PQPSs through highly efficient RAFT polymerization with a tunable particle size and excellent antibacterial ability. To enhance the positivity of the quaternary phosphonium ion, the phenyl group with strong electron dispersion capacity was designed as the side group and introduced into the ionic liquids. Subsequently, the quaternary phosphonium ionic liquids were easily polymerized with a chain transfer agent to obtain a RAFT agent for further copolymerization with other monomers. Styrene (St) was used for the polymerization of the second block to control the molecular weight and particle size. Altering the hydrophilic quaternary phosphonium ion and hydrophobic styrene monomer provides high adaptability of the copolymers in various dispersions and application fields. The phosphonium-functionalized diblock copolymers (PFDCs) were further studied to evaluate their antibacterial activity, and the effects of the chemical structure of the cations and the molecular weight of the particles were studied to comprehend the possible antibacterial mechanism. The PFDCs with surprising antibacterial activity prepared through simple RAFT polymerization show great potential for future practical applications.

2. Experimental section

2.1 Materials and reagents

Azobisisobutyronitrile (AIBN, 98%, Shanghai Macklin Biochemical Co., Ltd, China) was recrystallized from methanol twice and stored in the fridge before use. Ionic liquid (IL) monomers of a quaternary phosphonium salt (QPS) (4-vinylbenzyl tributylphosphonium chloride (QPSBu₃IL)) and 4-vinylbenzyl triphenyl-phosphonium chloride (QPSPh₃IL) were prepared according to Scheme 1. Styrene (St, CP), ethylene glycol dimethacrylate (EGDMA, 98%), methyltrioctylammonium chloride (TOMAC, 97%), and 1-dodecanethiol (98%) were purchased from Shanghai Aladdin Biochemical Technology Co., Ltd, China. Carbon disulfide (CS₂, 99%, Shanghai Saen Chemical Technology Co., Ltd) and Luria-Bertani (LB) broth medium (Qingdao Hope Bio-Technology Co., Ltd) were obtained commercially. *Escherichia coli* (*E. coli*, Gram-negative)

and *Staphylococcus aureus* (*S. aureus*, Gram-positive) were kindly provided by Prof. Shiquan Niu (College of life sciences, Northwest Normal University, China) and incubated in nutrient agar (Qingdao Hi-Tech Industrial Park Haibo Biological Co., Ltd, biological-reagent grade). Deuterium oxide (D₂O), Deuterated dimethyl sulfoxide (DMSO-d₆), and deuterated chloroform (CDCl₃) were of analytic grade and used as received without further purification. All other commercial reagents such as 2,6-di-*tert*-butyl-4-methylphenol (BHT), ether, acetone, 1,4-dioxane, methanol, isopropyl alcohol, *n*-hexane, trichloromethane, concentrated hydrochloric acid (HCl, 36–38%, w/w) and sodium hydroxide were of analytical grade and used as received unless otherwise noted. A dialysis bag (MWCO 8000–14 000 g mol⁻¹) was pretreated with 2% sodium bicarbonate solution and 1.0 mmol L⁻¹ ethylenediamine tetraacetic acid (EDTA) solution, then washed with distilled water and stored in the fridge. All reactions were carried out under a nitrogen atmosphere with magnetic stirring unless otherwise noted. Distilled water was used throughout the experiment.

2.2 Synthesis

2.2.1 Synthesis of 2-(dodecylthiocarbonothioylthio)-2-methylpropionic acid chain transfer agent (DCMAT). As illustrated in Schemes 1(a), 1-dodecanethiol (20.21 g, 0.10 mol), acetone (58.01 g, 1.01 mol), and TOMAC (1.01 g, 2.50 mmol) were mixed and magnetically stirred in a precooled (0 °C) jacketed reactor under a nitrogen atmosphere. 50% (w/w) sodium hydroxide solution (9.01 g, 0.11 mol) was added dropwise in 10 min. The stirring was continued for another 20 min, then a mixture of CS₂ (7.60 g, 0.10 mol) and acetone (10.00 g, 0.17 mol) was sequentially added within 20 min, and the color turned red gradually. After 10 minutes, chloroform (17.80 g, 0.15 mol) was added at once, followed by the dropwise addition of 50% (w/w) sodium hydroxide solution (40.00 g, 0.50 mol) in 20 min. The reaction was stirred overnight under a nitrogen atmosphere. To stop the reaction, 200 mL of water and subsequently 80 mL of concentrated HCl were added to adjust the pH value of the solution to lower than 4. A yellow precipitate was generated during the acidification. The precipitate was harvested through vacuum filtration and then dissolved in 2-propanol, while the undissolved parts were discarded. The mixture was then pre-dried by rotary evaporation to eliminate most of the 2-propanol. Afterward, the pre-dried solid was dissolved in hot hexane to obtain a light-red saturated solution, followed by eliminating the undissolved parts through vacuum evaporation, then cooled in a refrigerator to obtain yellow crystallized DCMAT. The yield of DCMAT was 12.21 g (33.0%, m.p. 60–63 °C). The chemical structure was characterized using NMR and FTIR as follows: ¹H NMR spectrum (DMSO-d₆, δ ppm from TMS, Fig. S1a, ESI[†]): 0.89 (t, 3H), 1.37–1.47 (m, 20H), 1.69 (s, 6H), 3.28 (t, 2H) and 12.96 (s, 1H); ¹³C NMR spectrum (CDCl₃, δ ppm from TMS, Fig. S1b, ESI[†]): 14.13, 22.68, 25.17, 28.95, 29.10, 29.34, 29.43, 29.54, 29.61, 31.90, 37.04, 55.51, 178.86 and 220.75; FTIR (KBr, cm⁻¹, Fig. S1c, ESI[†]): 1716, 1282, 1170 and 1068. The molecular weight of DCMAT (C₁₇H₃₃O₂S₃) was characterized by ESI-MS (*m/z* calculated: 365.16, found: 365.1).



Scheme 1 Schematic illustration of the synthesis of the CTA agent, IL monomers, and PFDCs (a), and the general assembly pattern for the micromorphology (b). (i) 2-(dodecylthiocarbonothioylthio)-2-methylpropionic acid chain transfer agent (DCMAT); (ii) quaternary phosphonium-based IL monomers (QPSPh₃IL and QPSBu₃IL); (iii) macromolecular chain transfer agent (macro-CTA, Ph₆₀ and Bu₆₀); (iv) phosphonium-functionalized diblock copolymers (PFDCs, Ph₆₀St_y and Bu₆₀St_y); (v) PEGDMA crosslinked Bu₆₀/Ph₆₀ diblock copolymers (BuE_m and PhE_m).

2.2.2 Synthesis of quaternary phosphonium-based IL monomers (QPSPh₃IL and QPSBu₃IL). QPSPh₃IL was prepared according to a reported method,¹⁷ as shown in Scheme 1(a). 4-Vinylbenzyl chloride (9.16 g, 60 mmol) and triphenylphosphine (18.88 g, 72 mmol) were added into a dried flask containing 100 mL of acetone, and the mixture was magnetically stirred at 60 °C for 48 h under a nitrogen atmosphere. After the reaction, the mixture cooled down to room temperature, then 150 mL of ether was added dropwise, which gradually generated a white precipitate. The precipitate was sequentially filtered, washed with ether, and vacuum-dried at room temperature for 24 h to obtain a pure product. The yield rate was 85.0% (m.p. 121.9 °C). The chemical shifts were measured by NMR as follows: ¹H NMR spectrum (D₂O, δ ppm from TMS, Fig. S2a, ESI⁺): 7.63–7.41 (m, 3H), 7.40–7.30 (m, 12H), 6.90 (d, 2H), 6.68 (d, 2H), 6.41–6.34 (m, 1H), 5.49 (d, 1H), 4.98 (d, 1H) and 4.43 (d, 2H). ¹³C NMR spectrum (D₂O, δ ppm from TMS, Fig. S2b, ESI⁺): 137.7, 135.7, 134.1, 131.4, 130.1, 126.6, 117.6, 116.7, 115.4 and 77.9. The molecular weight of QPSPh₃IL (C₂₇H₂₄P) was characterized by ESI-MS (*m/z* calculated: 379.46, found: 379.20).

QPSBu₃IL was prepared according to a reported method,¹⁸ as shown in Scheme 1(a). 4-Vinylbenzyl chloride (8.06 g, 53 mol) and tributylphosphine (10.52 g, 53 mol) were added into a dried flask containing 100 mL of acetone, and the mixture was magnetically stirred at 60 °C for 48 h under a nitrogen atmosphere. After the reaction, the mixture cooled down to room temperature, then 150 mL of diethyl ether was added dropwise, which gradually generated a white precipitate. The precipitate was sequentially filtered, washed with diethyl ether, and vacuum-dried at room temperature for 24 h to obtain the pure product. The yield rate was 78.6% (m.p. 127.3 °C). The chemical shifts were measured by NMR as follows: ¹H NMR (D₂O, δ ppm from TMS, Fig. S3a, ESI⁺): 0.68 (t, 9H), 1.12–1.36 (m, 6H), 1.88 (s, 6H), 3.39 (t, 6H), 5.11 (d, 2H), 5.65 (d, 1H), 6.45–6.63 (m, 2H) and 7.0–7.4 (m, 4H). ¹³C NMR spectrum (D₂O, δ ppm from TMS, Fig. S3b, ESI⁺): 12.8, 17.3, 17.8, 22.7, 23.5, 25.5, 26.1, 115.3, 127.2, 128.1, 130.3, 136.0 and 137.4. The molecular weight of QPSBu₃IL (C₂₁H₃₆P) was characterized by ESI-MS (*m/z* calculated: 319.49, found: 319.31).

2.2.3 RAFT polymerization for the synthesis of macro-CTA. The macromolecular chain transfer agent (macro-CTA),

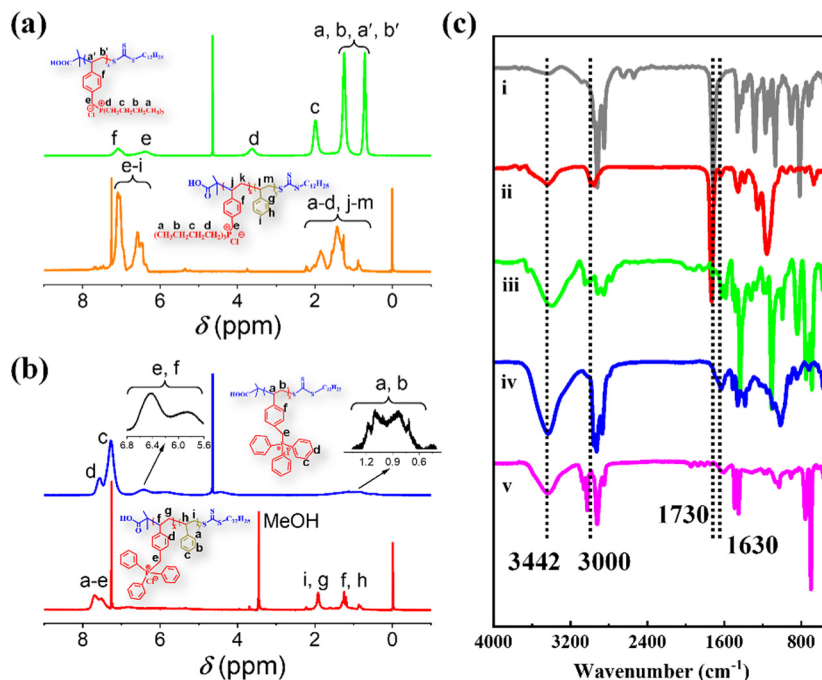


Fig. 1 (a) and (b) ¹H NMR spectra of Bu₆₀ (green), Bu₆₀St₁₂₀-M (orange), Ph₆₀ (blue) and Ph₆₀St₁₂₀-M (red). (c) FTIR spectra of DCMAT (ii), Ph₆₀ (iii), Ph₆₀St₁₂₀-M (iii), Bu₆₀ (iv) and Bu₆₀St₁₂₀-M (v).

poly(tributyl-(4-vinylbenzyl)-phosphonium chloride) (Bu₆₀), was synthesized by RAFT solution polymerization and is shown in Scheme 1(a). Briefly, DCMAT (91 mg, 2.5×10^{-4} mol), QPSBu₃IL monomer (5.3241 g, 0.015 mol), and anhydrous methanol (20 g) were added into a 100 mL Schlenk flask to achieve a target degree of polymerization (DP) of 60. Subsequently, the AIBN initiator (8.2 mg, 5×10^{-5} mol, $n_{\text{CTA}}/n_{\text{AIBN}} = 5.0$) was added to the mixture, followed by repeated vacuuming and filling with nitrogen for 30 min to eliminate dioxygen. The Schlenk flask was then tightly sealed and immersed in an oil bath at 70 °C. The final monomer concentration was fixed at 21% (w/w). The reaction proceeded for 12 h and was quenched by exposing the mixture to air and plunging the tube into liquid nitrogen. After thawing, the polymer solution was transferred into a dialysis bag and dialyzed against abundant DI water for 24 h. A yellow precipitate was generated during dialysis, then freeze-dried to obtain the macro-CTA agent (5.1986 g, 96% in conversion rate as determined by ¹H NMR). The product was coded Bu₆₀. Aqueous APC analysis (vs. poly (ethylene oxide) standards) indicated M_n and M_w/M_n values of 21 466 g mol⁻¹ and 1.14, respectively (Table S1, ESI[†]). The chemical shifts of Bu₆₀ were measured by NMR as follows: ¹H NMR spectrum (D₂O, broad peak δ ppm from TMS, Fig. 1): 7.09 (Ar-H, *ortho* to CH₂-P), 6.39 (Ar-CH₂-P), 3.63 (P-CH₂-(CH₂)₂CH₃), 2.00 (P-CH₂-CH₂-CH₂-CH₃ and backbone CH), 1.24 (P-(CH₂)₂-CH₂-CH₃ and backbone CH₂), and 0.71 (P-(CH₂)₃-CH₃ and backbone CH₃). ³¹P{¹H} NMR spectrum (D₂O, δ ppm, Fig. S4, ESI[†]): 25.4.

Another macro-CTA, poly(triphenyl-(4-vinylbenzyl)-phosphonium chloride) chain transfer agent (Ph₆₀), was also synthesized by RAFT solution polymerization. Briefly, DCMAT (91 mg, 2.5×10^{-4} mol), QPSPh₃IL monomer (6.2337 g, 0.015 mol), and anhydrous methanol

(20 g) were added into a 100 mL Schlenk flask to achieve a target degree of polymerization (DP) of 60. Subsequently, AIBN initiator (8.2 mg, 5×10^{-5} mol, $n_{\text{CTA}}/n_{\text{AIBN}} = 5.0$) was added to the mixture, followed by repeated vacuuming and filling with nitrogen for 30 min to eliminate dioxygen. The flask was then tightly sealed and immersed in an oil bath at 70 °C. The final monomer concentration was fixed at 24% (w/w). The reaction proceeded for 12 h and was quenched by exposing the mixture to air and plunging the tube into liquid nitrogen. After thawing, the polymer solution was transferred into a dialysis bag and dialyzed against abundant DI water for 24 hours. A light-yellow precipitate was generated during dialysis, then freeze-dried to obtain the macro-CTA agent (5.9452 g, 94% in conversion rate as determined by ¹H NMR). The product was coded Ph₆₀. Aqueous APC analysis (vs. poly (ethylene oxide) standards) indicated M_n and M_w/M_n values of 24 667 g mol⁻¹ and 1.13, respectively (Table S1, ESI[†]). The chemical shifts of Ph₆₀ were measured by NMR as follows: ¹H NMR spectrum (D₂O, broad peak δ ppm from TMS, Fig. 1): 7.57 (P-Ar-H, *para*), 7.30 (P-Ar-H, *ortho*), 6.49 (H-Ar-CH₂-P), 5.90 (H-Ar-CH₂-P), 1.2 (P-(CH₂)₃-CH₃ and backbone CH and CH₂), and 0.6 (P-(CH₂)₃-CH₃ and backbone CH₃). ³¹P{¹H} NMR (D₂O, δ ppm, Fig. S4, ESI[†]): 32.8.

2.2.4 RAFT polymerization for the synthesis of phosphonium-functionalized diblock copolymers (PFDCs). A series of poly[(tributyl-(4-vinylbenzyl)-phosphonium chloride-styrene] (Bu₆₀St_y) diblock copolymers was synthesized by RAFT polymerization. For brevity, the example synthesis of Bu₆₀St₁₂₀ is given as follows. Briefly, AIBN (0.82 mg, 5.0×10^{-6} mol, $n_{\text{Bu}_60}/n_{\text{AIBN}} = 2.0$), Bu₆₀ (169.8 mg, 1.0×10^{-5} mol) and methanol (1670.5 mg) were added into a Schlenk flask. After dissolution, styrene (St, 125 mg, 1.2×10^{-3} mol, $n_{\text{St}}/n_{\text{Bu}_60} = 120$) was added. The final content of St and Bu₆₀ was fixed at 15% (w/w). The flask was deoxygenated five

Table 1 Particle properties of non-crosslinked PFDCs with different substrate ratios measured by DLS

PFDCs					PFDCs				
Synthesis and measured in methanol					Synthesis and measured in 1,4-dioxane				
Block copolymer ^a	$n_{St}:n_{m_C}:n_I$	D_h^b (nm)	PDI ^c	ζ -potential (mV)	Block copolymer ^a	$n_{St}:n_{m_C}:n_I$	D_h (nm)	PDI	ζ -potential (mV)
Bu ₆₀ St ₃₀ -M	30:1:0.5	120	0.187	51.2	Bu ₆₀ St ₃₀ -D	30:1:0.5	126	0.154	49.8
Bu ₆₀ St ₁₂₀ -M	120:1:0.5	245	0.202	42.6	Bu ₆₀ St ₁₂₀ -D	120:1:0.5	244	0.150	43.2
Bu ₆₀ St ₂₀₀ -M	200:1:0.5	571	0.230	34.6	Bu ₆₀ St ₂₀₀ -D	200:1:0.5	464	0.121	33.3
Bu ₆₀ St ₇₀₀ -M	700:1:0.5	1255	0.208	28.7	Bu ₆₀ St ₇₀₀ -D	700:1:0.5	997	0.166	28.5
Ph ₆₀ St ₃₀ -M	30:1:0.5	40	0.289	55.8	Ph ₆₀ St ₃₀ -D	30:1:0.5	31	0.107	52.0
Ph ₆₀ St ₁₂₀ -M	120:1:0.5	185	0.275	48.2	Ph ₆₀ St ₁₂₀ -D	120:1:0.5	124	0.111	46.1
Ph ₆₀ St ₂₀₀ -M	200:1:0.5	427	0.202	44.2	Ph ₆₀ St ₂₀₀ -D	200:1:0.5	348	0.293	40.5
Ph ₆₀ St ₇₀₀ -M	700:1:0.5	1033	0.393	42.6	Ph ₆₀ St ₇₀₀ -D	700:1:0.5	1063	0.213	38.7

^a D: 1,4-dioxane; M: methanol; m_C : macro-CTA; I: AIBN. ^b Average hydrodynamic diameter of the resultant nanoparticles. ^c Polydispersity index obtained from the accessory software.

times by consecutive freezing-vacuuming-inflating with a nitrogen-thawing cycle. After the final cycle, the flask was thawed to room temperature and placed in an oil bath at 70 °C. The precursor solution was stirred for 16 h to ensure complete conversion of the St monomer. The reaction was then quenched by immersing the flask in liquid nitrogen. After thawing, the polymer solution was transferred into a dialysis bag and dialyzed against abundant DI water for 24 hours. The precipitate was generated during dialysis, then freeze-dried to obtain the product. The product was coded Bu₆₀St₁₂₀-M, where M represents methanol solvent. Other diblock copolymers of Bu₆₀St_y with different St monomer contents were prepared according to a similar protocol (Table 1). The conversion rate of St monomer for each Bu₆₀St_y copolymer was determined by using the ¹H NMR spectrum recorded in CDCl₃ and the data are summarized in Table S1 (ESI[†]). For comparison, a copolymer prepared by using 1,4-dioxane as solvent was also synthesized, and coded Bu₆₀St₁₂₀-D where D represents 1,4-dioxane solvent.

A series of poly[triphenyl-(4-vinylbenzyl)-phosphonium chloride-styrene] (Ph₆₀St_y) diblock copolymers, was also synthesized by RAFT polymerization. For brevity, the example synthesis of Ph₆₀St₁₂₀ is given as follows. Briefly, AIBN (0.82 mg, 5.0×10^{-6} mol, $n_{Ph_{60}}/n_{AIBN} = 2.0$), Ph₆₀ (202.1 mg, 1.0×10^{-5} mol) and methanol (1853.6 mg) were added into a Schlenk flask. After dissolution, styrene (St, 125 mg, 1.2×10^{-3} mol, $n_{St}/n_{Ph_{60}} = 120$) was added. The final content of St and Ph₆₀ was fixed at 15% (w/w). The flask was deoxygenated five times by consecutive freezing-vacuuming-inflating with a nitrogen-thawing cycle. After the final cycle, the flask was thawed to room temperature and placed in an oil bath at 70 °C. The precursor solution was stirred for 16 h to ensure complete conversion of the St monomer. The reaction was then quenched by immersing the Schlenk flask in liquid nitrogen. After thawing, the polymer solution was mixed with diethyl ether, which generated a precipitate. The precipitate was washed with diethyl ether three times and freeze-dried to obtain the product. The product was coded Ph₆₀St₁₂₀-M, where M represents methanol solvent. Other diblock copolymers of Ph₆₀St_y with different St monomer contents were prepared according to a similar protocol (Table 1). The conversion rate of St monomer for each Ph₆₀St_y copolymer was determined by using the ¹H NMR spectrum recorded in

CDCl₃ and the data are summarized in Table S1 (ESI[†]). For comparison, a copolymer prepared by using 1,4-dioxane as solvent was also synthesized, which was coded Ph₆₀St₁₂₀-D where D represents 1,4-dioxane solvent.

2.2.5 RAFT polymerization for the synthesis of PEGDMA crosslinked Bu₆₀/Ph₆₀ diblock copolymer. The synthesis was accomplished through one-step RAFT solution copolymerization by adding Bu₆₀ (or Ph₆₀), EGDMA, and AIBN (initiator) into methanol. For example, Bu₆₀ (216.6 mg, 10 μmol), EGDMA (59.5 mg, 30 μmol), AIBN (4 mg, 0.025 mmol), and anhydrous methanol (30 mL) were added to a 100 mL Schlenk flask. The Schlenk flask was deoxygenated by three cycles of consecutive freezing-vacuuming-inflating with a nitrogen-thawing cycle. After the final cycle, the Schlenk flask was thawed to room temperature and placed in an oil bath at 70 °C. The polymerization proceeded for 20 h and was quenched by immersing the Schlenk flask in liquid nitrogen. After thawing, the polymer solution was transferred into a dialysis bag and dialyzed against abundant DI water for 24 hours. The precipitate was generated during dialysis, then freeze-dried to obtain the product. The product was coded BuE₃₀, where Bu and E represent Bu₆₀ and EGDMA, respectively, and the number 30 is the initial dose in micromoles of EGDMA. Other PEGDMA crosslinked copolymers were similarly prepared and numbered, as shown in Table S2 (ESI[†]).

2.3 Characterization

¹H, ¹³C, and ³¹P NMR spectra were recorded on a Bruker AM spectrometer (400 MHz) at 300 K with tetramethylsilane (TMS) as the internal standard. All samples were dissolved in CDCl₃ to prepare a 2% (w/w) solution. Peak multiplicity was abbreviated as follows: singlet (s), doublet (d), triplet (t), quartet (q), and multiplet (m).

Electrospray ionization mass spectrometry (ESI-MS, Agilent 7890/5975, USA) was used to identify the molecular weight of the monomer and CTA samples.

Fourier-Transform infrared (FTIR) spectra were measured on a DIGILAB FTS3000 spectrophotometer by using the KBr method with wavenumber ranging from 4000 cm⁻¹ to 400 cm⁻¹.

The average molecular weight of the PFDCs (1 mg mL⁻¹ in HPLC THF solvent) was recorded using ACQUITY advanced polymer chromatography (APC) (Waters, USA) equipped with an ACQUITY APC XT 200 column (2.5 μm, 4.6 × 150 mm) and refractive index detector.

The surface compositions and chemical states of elements were determined by using ESCALAB Xi+ X-ray photoelectron spectroscopy (XPS) (Thermo Fisher Scientific Inc.) equipped with a monochromatic Al Kα X-ray source (1486.6 eV) with a spot size of 200–900 μm and an energy resolution of < 0.43 eV.

The morphology of the samples was observed using scanning electron microscopy (SEM, JEOL JSM 6701F) at an accelerating voltage of 5 kV.

The particle size and zeta potential (ζ) of the samples in the as-prepared solutions (1 mg mL⁻¹ in methanol or 1,4-dioxane) were measured at 25 °C on a Malvern Panalytical Zetasizer Nano ZS90 instrument equipped with a 4 mW He–Ne laser operating at a wavelength of 633 nm and an avalanche photodiode (APD) detector. Each sample was measured three times, and the ζ values were calculated according to the Smolouchowski–Helmholtz equation.

The contact angle (CA) of the samples was recorded on an SL200B goniometer (KINO Scientific Inc., USA). Briefly, dried sample powders were firstly ground and integrated by a tablet machine to obtain the membrane sample. The membrane sample was loaded on a slide and mounted on the platform. A drop of DI water (4 μL) was added from the top with an injection device. The images of the liquid on the sample were collected by a computer, and the contact angle was calculated automatically using the software.

2.4 Evaluation of antibacterial activity

The antibacterial activity of the PFDCs against *Escherichia coli* (*E. coli*, Grams-negative) and *Staphylococcus aureus* (*S. aureus*, Grams-positive) was evaluated in this work. Both strains were stored at 4 °C before the experiment. To activate the bacteria, a bacterial suspension (in LB broth) was incubated at 37 °C for 24 h. A bacterial suspension of 10⁷ CFU mL⁻¹ was prepared before the test. A nutrient agar hydrogel was used as the culture medium. The antibacterial activity of the PFDCs was assessed by using three different methods, colony formation assay, minimum inhibitory concentration (MIC), and SEM imaging.

2.4.1 Colony formation assay. This method was used to evaluate the bactericidal efficacy of the PFDCs. A suitable amount of activated *E. coli* or *S. aureus* was mixed with 10 mL of sterilized water until the absorbance of the suspension reached 1.0 (at 600 nm), then the suspension was incubated in a 37 °C water bath for 12 h with a shaking speed of 200 rpm. Then, 1 mL of the cultured suspension was serially diluted with sterilized water. 40.00 μg mL⁻¹ PFDCs was added to the diluted suspension at room temperature. The mixed solution was then spread on the agar (with LB broth) and the plates were incubated at 37 °C for a period of time. The control group was without any PFDC treatment. The number of bacterial colonies on the agar plates (90 mm in diameter) was counted

and the antibacterial rate (A.R., %) was calculated according to eqn (1):¹⁹

$$\text{Antibacterial rate(\%)} = \frac{A - B}{A} \times 100 \quad (1)$$

where *A* and *B* are the mean numbers of bacterial colonies for the control and PFDC-treated samples, respectively. All experiments were conducted in triplicate.

2.4.2 Evaluation of minimum inhibitory concentration (MIC). From the colony formation assay, the Ph₆₀St₃₀-M sample showed the best bactericidal efficacy, and, as a result, the MIC value for this sample was further measured. Different concentrations of Ph₆₀St₃₀-M (0, 5, 10, 30, 40, 60, and 200 μg mL⁻¹) were added into 5 mL of activated *E. coli* and *S. aureus* bacterial suspensions (10⁷ CFU mL⁻¹). The suspensions were incubated in a 37 °C water bath for 24 h under shaking. After that, 100 μL of the incubated suspension was evenly spread on the agar (with LB broth), and the plate was cultured at 37 °C for 24 h. After 24 h, each agar plate was recorded by a digital camera. The MIC value was defined as the minimum concentration of Ph₆₀St₃₀-M that inhibited the visible growth of the microbes.

2.4.3 SEM imaging. SEM was utilized to observe the bacterial morphology after the Ph₆₀St₃₀-M sample was treated. In this work, *E. coli* was selected as a representative to evaluate the effect of the Ph₆₀St₃₀-M sample. The *E. coli* was allowed to grow to a concentration of 10⁷ CFU mL⁻¹, then harvested by centrifugation and resuspended in saline. The bacteria were incubated again under agitation for 4 hours at 37 °C at 200 rpm. After that, the suspension was centrifuged at 10 000 rpm for 10 min to harvest the bacteria. The bacteria were further dehydrated by 25%, 75%, 90%, and 100% (v/v) aqueous ethanol solutions every 15 min. The dehydrated bacteria were gently spread on conductive tape and then observed using SEM.

3. Results and discussion

3.1 Synthesis of phosphonium-functionalized diblock copolymers (PFDCs)

To prepare a series of PFDCs with long-acting and broad-spectrum antibacterial activity, RAFT polymerization-induced self-assembly (RAFT PISA) was employed to realize their efficient synthesis and micromorphology manipulations. As illustrated in Scheme 1(a), the requisite chain transfer agent (CTA) and quaternary phosphonium ionic liquid monomers were firstly synthesized.^{17,18} DCMAT (CTA) could be obtained through a designed nucleophilic reaction with phase transfer catalysis (PTC) according to R. Shea's work.²⁰ Briefly, in the presence of a base, 1-dodecanethiol reacted with CS₂, followed by nucleophilic reactions with chloroform and acetone, and yellow crystalline DCMAT was obtained after subsequent acidification. The characterization using NMR and FTIR spectra of DCMAT (Fig. S1, ESI†) demonstrated the successful synthesis of the desired product. On the other hand, the ionic liquid monomers of the QPS, 4-vinylbenzyl-tributylphosphonium chloride (QPSBu₃IL), and 4-vinylbenzyl-triphenylphosphonium chloride (QPSPh₃IL) were prepared using the substitution

reaction between 4-vinylbenzyl chloride and tributylphosphine or triphenylphosphine. Fig. S2 and S3 (ESI[†]) show the NMR spectra with peak resolution to identify the chemical structure of QPSBu₃IL and QPSPh₃IL. Further, by mixing QPSBu₃IL or QPSPh₃IL, the DCMAT CTA, and AIBN (initiator) in the desired molar ratio, the active macro-CTA with a shamrock shape (regarding the electron cloud distribution), namely Bu₆₀ or Ph₆₀, was synthesized through RAFT solution polymerization.

In the subsequent polymerization, the RAFT and PISA process was considered to regulate and control the micromorphology of the particles of PFDCs. Constructing an insoluble second block during RAFT polymerization from the reaction system is an effective method to realize PISA and produce nano-assemblies, which simplifies the synthesis process, shortens the equilibration time, and avoids extremely dilute conditions.^{21,22} Styrene is a typical rigid monomer that dissolves in methanol but precipitates after polymerization. Therefore, by mixing the Bu₆₀ or Ph₆₀ macro-CTA, styrene monomer, and AIBN (initiator) with methanol solvent, PFDC nano-assemblies could be achieved *via* the RAFT and PISA process at a relatively high polymer dosing concentration (15%, w/w), as shown in Scheme 1(b). Another solvent, 1,4-dioxane, was also employed to study the effect of solvent on the micromorphology of the nano-assemblies. The sample codes and their conversion rate and molecular weight are summarized in Table S1 (ESI[†]). A conversion rate beyond 96% revealed the high efficiency of the RAFT polymerization for PFDCs. Also, the experimental molecular weight was close to the theoretical calculation, indicating that the RAFT and PISA process in this work was a highly predictable polymerization method. The synthesis of PFDCs was also accompanied by the micromorphology formation by PISA.

A relatively high density of quaternary phosphonium cations allowed substantial intramolecular interactions that induced further self-assembly. Based on the principle of polymer physics, the shamrock cations are sequenced in the *trans* or *gauche* steric arrangement along the polymer chain due to steric hindrance and electrostatic repulsion and leading to a staggered distribution (Scheme 1(b)). The flexibility of the polymer chain and hydrophilic-hydrophobic interactions between polymer and solvent allowed the planar curling and twisting of PSt segments through the conjugations between adjacent phenyl groups on the inner side, and further lateral stacking on the outer side. Such a chain arrangement formed a spherical morphology with minimal configurational entropy and acted as the core encapsulated by Bu₆₀ or Ph₆₀ segments due to their higher affinity to solvent (methanol). A more interesting morphology was obtained through the altering of synthesis factors, which will be discussed in the next section.

Additional crosslinking along with RAFT polymerization by the introduction of EGDMA was also investigated to study the effect of crosslinking on the micromorphology and antibacterial activity of the nano-assemblies. After the styrene monomer was replaced by EGDMA for RAFT polymerization with Bu₆₀ or Ph₆₀, PEGDMA crosslinked PFDCs were obtained. Double vinyl groups allowed not only the polymerization of EGDMA itself

but also copolymerization with different macro-CTAs, leading to the so-called “crosslinking polymerization”.²³ The introduction of PEGDMA led to the formation of a crosslinked network between macro-CTA and a ribbon-like structure, which then twisted due to the chain flexibility and connected end-to-end through conjugation between quaternary phosphonium ions and the phenylene groups, giving a circle sheet or Mobius loop structure, which will be discussed further in the next section.

The PEGDMA crosslinked PFDCs coded RE_m are listed in Table S2 (ESI[†]), where R represents Bu or Ph, and m represents the initial dose in micromoles of EGDMA. The successful synthesis of PEGDMA crosslinked PFDCs manifested the favorable universality of the RAFT PISA method that might inspire more types and morphologies of PQPS in view of the required functionalities.

3.2 Chemical structure and micromorphology of PFDCs

Chemical structure characterization of the PFDCs was conducted on the representative copolymers Ph₆₀St₁₂₀-M and Bu₆₀St₁₂₀-M to exemplify each step during polymerization, and the corresponding macro-CTA (Bu₆₀ and Ph₆₀) was characterized by using ¹H NMR and FTIR spectra, as shown in Fig. 1. The NMR peaks were also identified according to common knowledge of organic chemistry, as labeled in Fig. 1(a) and (b). Fig. S4 (ESI[†]) shows the ³¹P NMR spectra of Ph₆₀, Bu₆₀, Ph₆₀St₁₂₀-M, and Bu₆₀St₁₂₀-M, wherein the sharp signal for each sample confirmed the presence of phosphorus atoms in the polymer backbone. The higher chemical shift of Ph₆₀ (32.83 ppm) than that of Bu₆₀ (25.41 ppm) demonstrated that the triphenyl-phosphonium cation had lower electron density than that of the tributyl-phosphonium cation because of the stronger electron delocalization capacity provided by the phenyl groups. Copolymerization with St monomer did not sharply change the chemical shift in the ³¹P NMR spectra, which confirmed the diblock copolymerization.

FTIR spectra were used to confirm the existence of the fundamental functional groups on the polymer molecules, as illustrated in Fig. 1(c). The characteristic peaks at 3442, 1730, 1180 and 1028 cm⁻¹ appeared in the case of both macro-CTAs and the PFDCs, which were attributed to O–H in carboxyl groups, C=O in carboxyl groups, and thiocarbonyl (C=S) groups, respectively, which belonged to DCMAT, one of the starting materials.²⁴ The peaks at 1109 and 755 cm⁻¹, 3000 and 3021 cm⁻¹, and 1630 cm⁻¹, which were assigned to the vibration of P–C, C–H aromatic stretching, and aromatic double bond stretching, respectively, were only observed in the spectra of the macro-CTAs and PFDCs, suggesting successful copolymerization.²⁵ On the other hand, the chemical composition of the PEGDMA crosslinked PFDCs was characterized using XPS, as shown in Fig. S5 (ESI[†]). The survey spectrum clearly displayed the existence of C, O, P, S, and Cl elements for the PhE₂₀, BuE₂₅, and BuE₃₀ samples as representatives. The peaks at 284.1, 531.7, 133.4, 163.64, and 196.9 eV were assigned to C1s, O1s, P2p, S2p, and Cl2p, respectively.²⁶ This also confirmed that the PEGDMA crosslinked PFDCs were successfully synthesized.

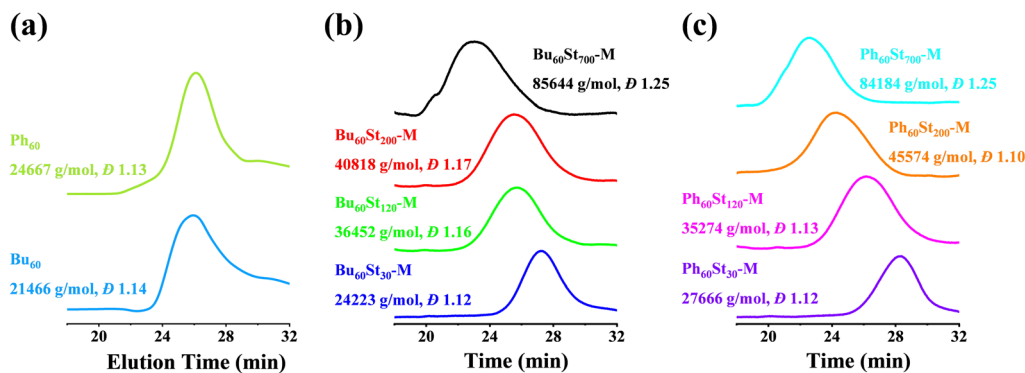


Fig. 2 APC traces of the macro-CTAs and PFDCs.

The molecular weights of the macro-CTAs and PFDCs were measured using advanced polymer chromatography (APC), and the trace spectra are shown in Fig. 2 and the conversion rate and molecular weight are summarized in Table S1 (ESI[†]). The macro-CTAs Bu₆₀ and Ph₆₀ with the theoretical molecular weight ($M_{n,th}$) of 21 662 and 25 259 g mol⁻¹ showed a similar retention time at around 26 min, while the experimental molecular weight ($M_{n,APC}$) showed smaller values of 21 466 and 24 667 g mol⁻¹, respectively, as shown in Fig. 2(a). All PFDCs were designed with a target degree of polymerization (DP) for the St monomer, and the corresponding theoretical molecular weight was calculated and is listed in Table S1 (ESI[†]). The retention time of the representative PFDCs ranged from 22 to 28 min (Fig. 2(b) and (c)), increasing in the order of descending molecular weight. The APC result showed that the experimental molecular weight of the obtained PFDCs was in good agreement with the theoretical value with an error less than 10%, suggesting the accurate controllability of the RAFT polymerization. Also, the high conversion rate of over 95% of the RAFT polymerization suggested the favorable possibility for industrial production of the PFDCs. Synthesis in different solvents with different relative polarities (methanol: 0.762; 1,4-dioxane: 0.164) had no significant influence on either molecular weight or conversion rate (Table S1, ESI[†]), suggesting a weak solvent dependence of the RAFT polymerization of PFDCs. The single peak of the APC curve and the polydispersity value measured by the APC method (\bar{D}) ranged from 1.10 to 1.25, indicating a high uniformity of the synthesized PFDCs obtained from the living radical polymerization.²⁷ The APC results confirmed that the RAFT polymerization designed in this work was highly applicable to industrial production.

Contact angles (CAs) of the PFDC membranes were further measured to classify the hydrophilicity and hydrophobicity, as shown in Fig. 3. The macro-CTAs Bu₆₀ and Ph₆₀ were soluble in water possibly due to the strong polarization effect of the quaternary phosphonium ions. Undoubtedly, copolymerization with hydrophobic styrene strikingly enhanced the hydrophobicity of the PFDCs, which led to an increase in contact angle. Meanwhile, a bigger contact angle was observed with the increase of St content in both Bu₆₀St_y-M and Ph₆₀St_y-M groups. It was reported that the CA of PSt was around 85° in thin film

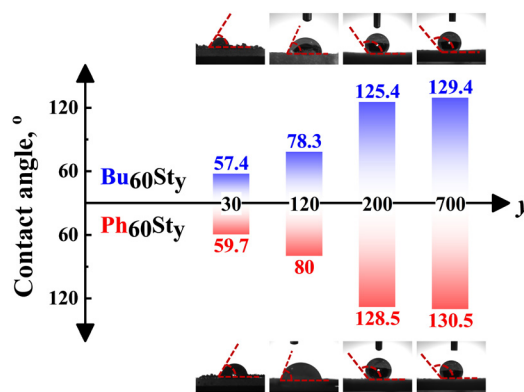


Fig. 3 The contact angle (CA) of Bu₆₀St_y-M and Ph₆₀St_y-M in membrane form. The Bu₆₀ and Ph₆₀ macro-CTAs were soluble in water.

form,²⁸ which increased to 147° in foam form²⁹ and 155° with a honeycomb microstructure,³⁰ suggesting a nonnegligible effect of surface morphology on the hydrophobicity. The CA of the PFDCs with high St contents showed a bigger value (125–130°) than the case of the thin films; as a result, it was reasonable to deduce that a special morphology of the PFDCs was produced during polymerization. However, interestingly, the CA of Ph₆₀St_y-M was similar to that of Bu₆₀St_y-M even at a relatively low St content. The CA of several PEGDMA crosslinked PFDCs was also measured, as shown in Fig. S6 (ESI[†]). PEGDMA crosslinked PFDCs had a low CA, suggesting favorable hydrophilicity due to the low DP of the EGDMA monomer, while the CA increased with EGDMA content because of the insolubility of PEGDMA in water. It should be noted that the CA of PhE₂₀ was bigger than that of BuE₂₀, suggesting lower hydrophilicity.

As described above, when the copolymerized component was St, the CA tended to be the same, while it was markedly different in the case of EGDMA. The paradoxical phenomenon occurring between St and EGDMA monomers regarding the trends of contact angle possibly derived from the conjugation effect between the first macro-CTA block and the second block. In the case of PSt, an ultrawide conjugation effect along the whole polymer chain among phenyl groups averaged the charges and gave a similar surface with nearly equal electricity, leading to a similar contact angle regardless of the macro-CTA



Fig. 4 SEM images of PFDCs. Insets show the principal 3D schematic model for the nano-assemblies. M and D represent the methanol and 1,4-dioxane synthesis media, respectively.

type. However, crosslinking polymerization by EGDMA only provided micromorphology tunability instead of averaging the polymer charges. The intermolecular conjugation effect of phenyl groups of the PhE_m copolymer might induce a micellar shape with a hydrophilic inner (π - π stacking) but hydrophobic outer, resulting in a more hydrophobic surface and a higher CA compared to the BuE_m copolymer, whose *n*-butyl side groups had weak interactions between each other and induced less special micromorphology or specific effects on the hydrophobicity.³¹ In summary, the hydrophilicity of the PFDCs could be controlled by quantitatively adjusting the ratio of two monomers, which will be useful for different practical applications.

Scanning electronic microscopy (SEM) was further conducted to examine the micromorphology of the PFDCs, as shown in Fig. 4. All PFDCs assembled into nanoparticles with a diameter ranging from 50 to 1600 nm related to the St content. With an increase in molar ratio of St monomer, the particle size of the nano-assemblies also increased. However, a dramatic decrease was observed from macro-CTA to PFDCs with relatively low contents of St (e.g. $y = 30$ and 120), which was induced by the π - π stacking between phenyl groups on cations and the PSt second block, which reduced the size of the particles. The large size of the macro-CTA might have originated from the adjacent quaternary phosphonium cations causing both strong electrostatic repulsion and steric hindrance that tremendously hampered the polymer chain

assembly and expanded the size of the particles. The much smaller particle size of Ph₆₀St₃₀ synthesized in both methanol and 1,4-dioxane compared to that of Bu₆₀St₃₀ and Ph₆₀ macro-CTA suggested that π - π stacking also occurred between phenyl groups on the cations and the PSt block, leading to a stronger intramolecular interaction that contracted the particle size of the nano-assemblies a lot more. Crosslinking polymerization with EGDMA increased the particle size of the Ph₆₀-type PFDC but decreased it in the case of the Bu₆₀-type PFDC, which might be due to the synergy of polymer chain elongation and intramolecular interactions (or repulsions) between PEGDMA and quaternary phosphonium cations.

The major morphology of the nano-assemblies was a dispersed sphere, while in some cases, a special morphology in the shape of a ball-bearing ring, cuboid, disk-bowl (Chinese shoe-shape gold ingot), circle sheet, wrinkled sphere, or twisted strip loop (*Möbius* loop) was observed (Fig. 4). Such special morphologies appeared randomly regardless of either St content or crosslinking, while their size was mostly located in a narrow range from 300 to 450 nm, suggesting a size-related effect on the morphology. However, the changes in the special morphology did not follow a specific trend, making it difficult to identify the formation mechanism. One thing that could be certain is that the stacking and packing of macromolecules began with a positively charged nucleus and expanded from point to planar, and finally to a three-dimensional structure. Excess polymer segments are attached to the surface of the spherical particles



Fig. 5 St content dependence of particle size (D_h) (a) and ζ -potential (b) for PFDCs measured by DLS. (c) $M_{n,APC}$ dependence of particle size (D_h) for the PFDCs.

and assembled as either cones or smaller spheres. A proper explanation for the special morphology is that the intramolecular interactions between phenyl groups and electrostatic repulsion among quaternary phosphonium cations induced certain polymer chain arrangements then various special morphologies,^{32–34} and even different contact angles as described above.

To obtain the particle information of the PFDC nano-assemblies, DLS was conducted to measure the hydrodynamic diameter (D_h), polydispersity index (PDI), and ζ -potential, as summarized in Table 1 and Table S2 (ESI[†]). Fig. 5 shows the tendency of D_h and PDI with the molar ratio of St monomer. The D_h value measured by DLS strongly agreed with the SEM results. A proximate linear increase of D_h with the molar ratio of St in both $Bu_{60}St_y$ and $Ph_{60}St_y$ PFDCs manifested the tunability of particle size through controlling DP (Fig. 5(a) and (c)). While in the case of the PEGDMA crosslinked PFDCs, the D_h value seemed to decrease in a logarithmic manner (Fig. S7a, ESI[†]), probably due to a different polymer chain rearrangement process during polymerization. The ζ -potential of all PFDCs was higher than +20 mV, indicating that they were positively charged particles. The ζ -potential decreased linearly with increasing molar ratio of St in the range from 0 to 200 and tended to be flat after that for the $Bu_{60}St_y$ and $Ph_{60}St_y$ PFDCs (Fig. 5(b)), suggesting lower dispersion stability. The ζ -potential slightly increased in the case of the PEGDMA crosslinked PFDCs (Fig. S7b, ESI[†]), which was attributed to a morphology stability increase through crosslinking. A higher ζ -potential of Ph_{60} -type PFDCs than that of Bu_{60} -type PFDCs suggested a stronger stabilization effect induced by the interactions between phenyl groups of quaternary phosphonium ions and the second block along the polymer chain, which was also consistent with the smaller particle size shown in Fig. 4. A sudden change in D_h and ζ -potential from macro-CTA to PFDCs was also observed from DLS results, demonstrating that copolymerization with St or EGDMA was able to stabilize the dispersion of PFDC nano-assemblies.

The nanoparticle size and dispersion condition of the PFDCs could be directly visualized using the Tyndall effect in either methanol or 1,4-dioxane, as shown in Fig. S8 and S9 (ESI[†]). Homogenous suspensions of PFDCs with a distinct single light going through appeared as a white turbid emulsion with the increase of copolymerized St monomer (Fig. S8, ESI[†])

and crosslinking (Fig. S9, ESI[†]), also suggesting an increase of particle size but lower dispersion stability of the PFDCs. Selection of the type of suspension regarding the polymer molecular weight and particle size is essential in practical production. That means that there is a strong dependence of the molecular weight on either particle size or suspension condition. This could lead to the versatile production of phosphonium-functionalized diblock copolymers, and widen their application.

3.3 Antibacterial activities of PFDCs

The introduction of quaternary phosphonium cations brought about a high antibacterial activity for the PFDCs against *E. coli* and *S. aureus*, which was evaluated by using colony formation assay, as shown in Fig. 6 and Fig. S10 (ESI[†]). Semi-qualitative measurement by colony counting was used to characterize the antibacterial activities and confirm the effect of the molar ratio of macro-CTA and St monomer. The PFDCs showed significant antibacterial activities because of the quaternary phosphonium cations, with antibacterial rates beyond 95.7% and even 99.9% in most of the cases against *E. coli*. It is well known that lysis caused by cations originates from the electrostatic interactions between them and the negatively charged cell envelope of bacteria, which leads to a breakdown of cell envelope structure and leakage of organoids and ultimately death.^{35–37} In addition, with the increased ratio of St monomer, the A.R. value decreased significantly in both cases of *E. coli* and *S. aureus*, indicating a strong content dependence of antibacterial activity for the quaternary phosphonium cations (Fig. 6(b) and (c)). Notably, polymerization in either methanol or 1,4-dioxane did not alter the antibacterial activity of the PFDCs (Fig. 6(a)), making them highly adaptable in different production processes and applications. Interestingly, the PFDCs with different macro-CTAs (Bu_{60} or Ph_{60}) showed similar antibacterial activity against the same bacteria, suggesting that the type of site groups on the phosphorus atom had no effect on the antibacterial activity. Some researchers have reported that the length of the site group can affect the antibacterial activity,³⁸ but in our case, the geometrical length of the *n*-butyl and phenyl groups under ideal conditions were similar and had a very slight effect on antibacterial activity if this theory was applicable (Fig. 6(e)). Steric hindrance of the side groups might also affect the antibacterial activity,³⁹ but this is not discussed here. The



Fig. 6 (a) Photographs of the culture plates of *E. coli* and *S. aureus* after exposure to different PFDCs at an initial dose of 40 $\mu\text{g mL}^{-1}$. The A.R. value is the antibacterial rate according to the colony counting measurements. (b) and (c) The molar ratio of macro-CTA dependence of A.R. value for Bu₆₀-type (b) and Ph₆₀-type (c) PFDCs. (d) Particle size (D_h) dependence of A.R. value for Bu₆₀St_y and Ph₆₀St_y PFDCs. The light-yellow region is in the D_h range and highlights the special micromorphologies. (e) Calculation of the length of *n*-butyl and phenyl groups using a minimal energy model (calculated by ChemBio 3D Ultra). Red, blue, and yellow colors represent the phenyl group, *n*-butyl group, and phosphorus atom, respectively. (f) Schematic diagram of the particles with different sizes and positive charge density in contact with *E. coli*. The positive charge was supplied by the quaternary phosphonium cation, which had a trident shape according to the schematic diagram (e).

PEGDMA crosslinked PFDCs all showed excellent antibacterial activity against *E. coli* and *S. aureus*, as shown in Fig. S10 (ESI[†]), suggesting that the antibacterial capacity was endowed by the introduction of quaternary phosphonium cations instead of the nature of crosslinking.

As well as the content of macro-CTA, the particle size of the PFDCs showed a strong effect on antibacterial activity, as shown in Fig. 6(d). Generally, a bigger particle size led to a smaller A.R. value (*i.e.*, antibacterial activity), and this tendency was more striking against *S. aureus* as the A.R. value decreased a lot more with the increase of D_h . A feasible explanation for this is that the total number of positive charges was constant on the surface of the PFDCs, while a large particle size led to a decrease in the positive charge density, leading to fewer cations in contact with the bacteria. This also explains why the A.R. value of the macro-CTA was a bit lower than that of either Ph₆₀St₃₀ or Bu₆₀St₃₀. On the other hand, *E. coli* has a long cylinder shape with a larger contact area and was able to contact more spherical PFDC particles compared to *S. aureus*, which had a circular shape. The two factors described here both affected the antibacterial activity, as illustrated in Fig. 6(f).

Fig. 6(d) highlights the D_h region that produced the special morphologies observed using SEM, and it seems that the special morphologies had no specific antibacterial activity against *E. coli*, while in the case of *S. aureus*, the ball-bearing ring shaped PFDC had a better antibacterial activity, which might be due to its higher topological adaptability and contact area to contact with the spherical *S. aureus*, as shown in Fig. S11 (ESI[†]).

The results of the colony formation assay showed that the Ph₆₀St₃₀-M sample had the highest antibacterial activity against *E. coli* and *S. aureus*, whereby the A.R. values reached 99.9% and 99.7%, respectively. Therefore, the Ph₆₀St₃₀-M sample was further applied to evaluate the minimal inhibition concentration (MIC), as shown in Fig. 7. By measuring the optical density (OD) at 600 nm, the concentration dependence was examined to quantify the antibacterial activity against *E. coli* and *S. aureus* (10^7 CFU mL⁻¹) for the Ph₆₀St₃₀-M sample. At a dose of 40.00 ($\mu\text{g mL}^{-1}$), *E. coli* growth was completely inhibited while in the case of *S. aureus*, a higher dose of 60 ($\mu\text{g mL}^{-1}$) was required for full inhibition (Fig. 7), which was consistent with the colony formation assay results. In other words, the MIC for

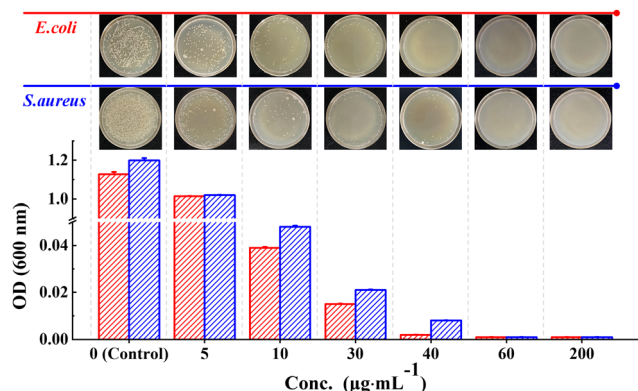


Fig. 7 Concentration dependence of the OD value for the P₆₀St₃₀-M sample. The images show the visible inhibition effect at corresponding concentrations against *E. coli* and *S. aureus*.

E. coli and *S. aureus* was 40 (1.4 $\mu\text{mol L}^{-1}$) $\mu\text{g mL}^{-1}$ and 60 $\mu\text{g mL}^{-1}$ (2.2 $\mu\text{mol L}^{-1}$), respectively, comparable to some commercial antibiotics and better than the reported polymeric antibiotics with a single active site (Table 2). Therefore, the synthesized copolymers showed excellent antibacterial activity.

The time-related antibacterial effectiveness of the Ph₆₀St₃₀-M sample was also evaluated against *E. coli* to measure the long-acting antibacterial capacity, as shown in Fig. 8(a). A 10^7 CFU mL^{-1} *E. coli* suspension mixed with 40.00 $\mu\text{g mL}^{-1}$ Ph₆₀St₃₀-M and sufficient LB broth was incubated for 10 days to observe the growth condition of the bacteria. Unsurprisingly, only one small colony of bacteria grew within the whole experiment period compared to the control group, suggesting a long-acting antibacterial capacity of the PFDCs. SEM was also used to examine the cell membrane integrity of *E. coli* after Ph₆₀St₃₀-M treatment, as shown in Fig. 8(b) and (c). The *E. coli* maintained its rod-like shape in the absence of Ph₆₀St₃₀-M, while the outer membrane was damaged and cells exhibited a hollow morphology after being treated, indicating leakage of organoids that was responsible for the death of the bacteria.

Antibacterial materials in various forms and composites are crucial and frequently used in our daily lives. As demonstrated above, PFDCs in suspension forms like inks, dyes, or paints are one of the most common applications in practical usage. As illustrated in Fig. 8(d), Ph₆₀St₃₀-M samples mixed with blue ink

or Rhodamine B (dye), or coated with commercial white latex paint were incubated with *E. coli* (10^7 CFU mL^{-1}). Without the presence of the PFDC, *E. coli* grew very well with either the ink, dye, or paint. After the introduction of 40.00 $\mu\text{g mL}^{-1}$ Ph₆₀St₃₀-M, an A.R. value of 99.9% for all mixing plates suggested that almost no bacteria grew in the plate compared to the control group, suggesting that the antibacterial activity was not affected after mixing with an ink, dye, or paint. The results suggested that the synthesized PFDCs had great potential as highly efficient antibacterial materials in many practical applications. They could also exhibit adaptability in different solvents by altering the ratio of hydrophilic and hydrophobic blocks in the copolymers.

4. Conclusion

Here, we demonstrated a RAFT polymerization method for the highly efficient synthesis of phosphonium-functionalized diblock copolymers with tunable particle size and excellent antibacterial activity. The conversion rate for the RAFT polymerization of PFDCs was beyond 95% synthesized in either methanol (strong polarity) or 1,4-dioxane (weak polarity) solvent, showing favorable producibility. In addition, polymerization-induced self-assembly (PISA) of PFDCs produced nano-assemblies of PFDCs whose particle size could be controlled by altering the initial dosing of St or EGDMA monomer. The quaternary phosphonium cations endowed the PFDCs with excellent antibacterial activity with a MIC of 40 and 60 $\mu\text{g mL}^{-1}$ (1.4 and 2.2 $\mu\text{mol L}^{-1}$) against *E. coli* and *S. aureus*, respectively, and the antibacterial mechanism was proposed regarding the size and charge density aspects. Interestingly, some special morphologies were generated during the synthesis of the PFDCs within a specific diameter range, which might have surprising antibacterial activity for specialized applications. Tuning the content of the monomer made it possible to modify the hydrophilicity and hydrophobicity of the PFDCs, which broadened their application. Combining the high practicability and functionality of PFDCs provides great prospects for their application as highly efficient and broad-spectrum antibiotics.

Table 2 Comparison of MIC between PFDCs and other reported antibacterial materials

Sample	MIC		Ref.
	<i>E. Coli</i>	<i>S. aureus</i>	
Ampicillin	2.4 $\mu\text{mol L}^{-1}$	2 $\mu\text{g mL}^{-1}$	40 and 41
Tetracycline	4.7 $\mu\text{mol L}^{-1}$	64 $\mu\text{g mL}^{-1}$	40 and 41
Chitosan oligomer	80 $\mu\text{g mL}^{-1}$	80 $\mu\text{g mL}^{-1}$	42
Polyhexamethylene guanidine hydrochloride-based disinfectant	50 $\mu\text{g mL}^{-1}$	400 $\mu\text{g mL}^{-1}$	43
Cationic random methacrylamide polymers	2.7 $\mu\text{mol L}^{-1}$	3.8 $\mu\text{mol L}^{-1}$	44
N-Alkyl imidazolium-based poly(ionic liquids)	1.2 $\mu\text{mol L}^{-1}$	2.5 $\mu\text{mol L}^{-1}$	38
Imidazolium-type poly(ionic liquids)	9 $\mu\text{mol L}^{-1}$	12 $\mu\text{mol L}^{-1}$	45
Quaternary phosphonium salt carbon sphere	150 $\mu\text{g mL}^{-1}$	100 $\mu\text{g mL}^{-1}$	46
Quaternary ammonium salt-based crosslinked micelles	298.4 $\mu\text{mol L}^{-1}$	69.6 $\mu\text{mol L}^{-1}$	47
Phosphonium-based polymeric biocides	2.7 $\mu\text{mol L}^{-1}$	2.7 $\mu\text{mol L}^{-1}$	13
PFDCs	1.4 $\mu\text{mol L}^{-1}$	2.2 $\mu\text{mol L}^{-1}$	This work

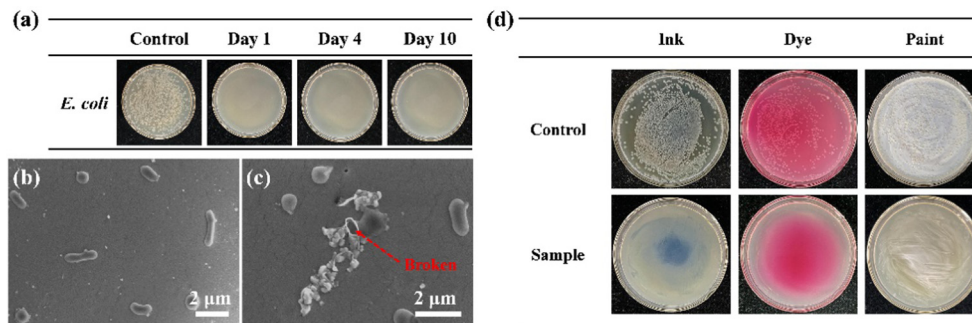


Fig. 8 (a) Photographs of the culture plates of *E. coli* after exposure to $\text{Ph}_{60}\text{St}_{30}\text{-M}$ for different numbers of days (10^7 CFU mL^{-1} , 37°C). (b) and (c) SEM images of *E. coli* before (b) and after (c) being treated with $\text{Ph}_{60}\text{St}_{30}\text{-M}$. (d) Photographs of the culture plates of *E. coli* after exposure to the modified ink, dye, and paint, respectively.

Author contributions

Peng Cao: methodology, investigation, data curation, writing – original draft, formal analysis. Xue Bai: investigation, validation. Yufeng He: resources. Pengfei Song: resources. Rongmin Wang: resources, supervision, project administration, funding acquisition, conceptualization. Junchao Huang: writing – review & editing, data curation, visualization, formal analysis.

Conflicts of interest

The authors declared no conflict of interest.

Acknowledgements

This research was funded by the National Natural Science Foundation of China (21865030).

References

- 1 F. D. Lowy, *Nat. Med.*, 2007, **13**, 1418–1420.
- 2 N. Nikfarjam, M. Ghomi, T. Agarwal, M. Hassanpour, E. Sharifi, D. Khorsandi, M. Ali Khan, F. Rossi, A. Rossetti, E. Nazarzadeh Zare, N. Rabiee, D. Afshar, M. Vosough, T. Kumar Maiti, V. Mattoli, E. Lichtfouse, F. R. Tay and P. Makvandi, *Adv. Funct. Mater.*, 2021, **31**, 2104148.
- 3 J. N. Pendleton and B. F. Gilmore, *Int. J. Antimicrob. Agents*, 2015, **46**, 131–139.
- 4 M. Wang, J. Shi, H. Mao, Z. Sun, S. Guo, J. Guo and F. Yan, *Biomacromolecules*, 2019, **20**, 3161–3170.
- 5 M. D. T. Torres, S. Voskian, P. Brown, A. Liu, T. K. Lu, T. A. Hatton and C. de la Fuente-Nunez, *ACS Nano*, 2021, **15**, 966–978.
- 6 B. Wang, T. Li, W. Guo, R. Wang, Y. Li, X. Zhu, P. Song and Y. He, *Int. J. Biol. Macromol.*, 2021, **174**, 198–206.
- 7 A. M. Carmona-Ribeiro and L. D. De Melo Carrasco, *Int. J. Mol. Sci.*, 2013, **14**, 9906–9946.
- 8 M.-H. Xiong, Y. Bao, X.-Z. Yang, Y.-H. Zhu and J. Wang, *Adv. Drug Delivery Rev.*, 2014, **78**, 63–76.
- 9 H. Zhou, D. Tang, X. Kang, H. Yuan, Y. Yu, X. Xiong, N. Wu, F. Chen, X. Wang, H. Xiao and D. Zhou, *Adv. Sci.*, 2022, **9**, 2200732.
- 10 Y. Xiong, J. Liu, Y. Wang, H. Wang and R. Wang, *Angew. Chem., Int. Ed.*, 2012, **51**, 9114–9118.
- 11 V. Loczenski Rose, F. Mastrotto and G. Mantovani, *Polym. Chem.*, 2017, **8**, 353–360.
- 12 W. Qian, J. Texter and F. Yan, *Chem. Soc. Rev.*, 2017, **46**, 1124–1159.
- 13 T. J. Cuthbert, B. Hisey, T. D. Harrison, J. F. Trant, E. R. Gillies and P. J. Ragona, *Angew. Chem., Int. Ed.*, 2018, **57**, 12707–12710.
- 14 B. Zhang, X. Yan, P. Alcouffe, A. Charlot, E. Fleury and J. Bernard, *ACS Macro Lett.*, 2015, **4**, 1008–1011.
- 15 Q. Zhang, R. Zeng, Y. Zhang, Y. Chen, L. Zhang and J. Tan, *Macromolecules*, 2020, **53**, 8982–8991.
- 16 J. Cornel Erik, J. Jiang, S. Chen and J. Du, *CCS Chem.*, 2020, **3**, 2104–2125.
- 17 Y. Xiong, H. Wang, R. Wang, Y. Yan, B. Zheng and Y. Wang, *Chem. Commun.*, 2010, **46**, 3399–3401.
- 18 Y. Xiong, Y. Wang, H. Wang and R. Wang, *Polym. Chem.*, 2011, **2**, 2306–2315.
- 19 S. Chen, S. Chen, S. Jiang, M. Xiong, J. Luo, J. Tang and Z. Ge, *ACS Appl. Mater. Interfaces*, 2011, **3**, 1154–1162.
- 20 J. T. Lai, D. Filla and R. Shea, *Macromolecules*, 2002, **35**, 6754–6756.
- 21 M. Semsarilar, V. Ladmiraal, A. Blanazs and S. P. Armes, *Langmuir*, 2013, **29**, 7416–7424.
- 22 C. Gonzato, M. Semsarilar, E. R. Jones, F. Li, G. J. P. Krooshof, P. Wyman, O. O. Mykhaylyk, R. Tuinier and S. P. Armes, *J. Am. Chem. Soc.*, 2014, **136**, 11100–11106.
- 23 M. Chen, J.-W. Li, W.-J. Zhang, C.-Y. Hong and C.-Y. Pan, *Macromolecules*, 2019, **52**, 1140–1149.
- 24 A. J. D. Magenau, N. Martinez-Castro, D. A. Savin and R. F. Storey, *Macromolecules*, 2009, **42**, 8044–8051.
- 25 B. Zeytuncu, E. Çakmakçı and M. V. Kahraman, *Polym. Compos.*, 2015, **36**, 946–954.
- 26 R. Bai, J. Kang, O. Simalou, W. Liu, H. Ren, T. Gao, Y. Gao, W. Chen, A. Dong and R. Jia, *ACS Biomater. Sci. Eng.*, 2018, **4**, 2193–2202.
- 27 C.-M. Liao, C.-C. Hsu, F.-S. Wang, B. B. Wayland and C.-H. Peng, *Polym. Chem.*, 2013, **4**, 3098–3104.
- 28 Y. Li, J. Q. Pham, K. P. Johnston and P. F. Green, *Langmuir*, 2007, **23**, 9785–9793.
- 29 N. Zhang, S. Zhong, T. Chen, Y. Zhou and W. Jiang, *RSC Adv.*, 2017, **7**, 22946–22953.
- 30 P. Escalé, W. Van Camp, F. Du Prez, L. Rubatat, L. Billon and M. Save, *Polym. Chem.*, 2013, **4**, 4710–4717.

- 31 S. Hosseini, H. Savaloni and M. Gholipour Shahraki, *Appl. Surf. Sci.*, 2019, **485**, 536–546.
- 32 P. Xu, L. Gao, C. Cai, J. Lin, L. Wang and X. Tian, *Angew. Chem., Int. Ed.*, 2020, **59**, 14281–14285.
- 33 H. Zhu, X. Wang, Y. Cui, J. Cai, F. Tian, J. Wang and H. Qiu, *Macromolecules*, 2019, **52**, 3479–3485.
- 34 H. Qiu, A. M. Oliver, J. Gwyther, J. Cai, R. L. Harniman, D. W. Hayward and I. Manners, *Macromolecules*, 2019, **52**, 113–120.
- 35 A. Bhatnagar, W. Hogland, M. Marques and M. Sillanpää, *Chem. Eng. J.*, 2013, **219**, 499–511.
- 36 B. Wang, F. Wang, Y. Kong, Z. Wu, R.-M. Wang, P. Song and Y. He, *Colloids Surf., A*, 2018, **549**, 122–129.
- 37 T. Zhang, J. Guo, Y. Ding, H. Mao and F. Yan, *Sci. China: Chem.*, 2019, **62**, 95–104.
- 38 C. Fang, L. Kong, Q. Ge, W. Zhang, X. Zhou, L. Zhang and X. Wang, *Polym. Chem.*, 2019, **10**, 209–218.
- 39 T. R. Stratton, B. M. Applegate and J. P. Youngblood, *Bio-macromolecules*, 2011, **12**, 50–56.
- 40 C. L. Martini, C. C. Lange, M. A. V. P. Brito, J. B. Ribeiro, L. C. Mendonça and E. K. Vaz, *J. Dairy Res.*, 2017, **84**, 202–205.
- 41 V. Hoerr, G. E. Duggan, L. Zbytnuik, K. K. H. Poon, C. Große, U. Neugebauer, K. Methling, B. Löffler and H. J. Vogel, *BMC Microbiol.*, 2016, **16**, 82.
- 42 A. Shanmugam, K. Kathiresan and L. Nayak, *Biotechnol. Rep.*, 2016, **9**, 25–30.
- 43 M. K. Oulé, R. Azinwi, A.-M. Bernier, T. Kablan, A.-M. Maupertuis, S. Mauler, R. K. Nevry, K. Dembélé, L. Forbes and L. Diop, *J. Med. Microbiol.*, 2008, **57**, 1523–1528.
- 44 A. Tyagi and A. Mishra, *ACS Omega*, 2021, **6**, 34724–34735.
- 45 Z. Zheng, Q. Xu, J. Guo, J. Qin, H. Mao, B. Wang and F. Yan, *ACS Appl. Mater. Interfaces*, 2016, **8**, 12684–12692.
- 46 Y. Yang, Q. Shi, J. Feng, X. Shu and J. Feng, *RSC Adv.*, 2014, **4**, 50708–50712.
- 47 F. Liu, D. He, Y. Yu, L. Cheng and S. Zhang, *Bioconjugate Chem.*, 2019, **30**, 541–546.