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Tailoring the pore structure of iron oxide core@stellate mesoporous silica shell nanocomposites: effects on MRI and magnetic hyperthermia properties and applicability to anti-cancer therapies†

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Core-shell nanocomposites made of iron oxide core (IO NPs) coated with mesoporous silica (MS) shells are promising theranostic agents. While the core is being used as an efficient heating nanoagent under alternating magnetic field (AMF) and near infra-red (NIR) light and as a suitable contrast agent for magnetic resonance imaging (MRI), the MS shell is particularly relevant to ensure colloidal stability in a biological buffer and to transport a variety of therapeutics. However, a major challenge with such inorganic nanostructures is the design of adjustable silica structures, especially with tunable large pores which would be useful, for instance, for the delivery of large therapeutic biomolecule loading and further sustained release. Furthermore, the effect of tailoring a porous silica structure on the magneto- or photo-thermal dissipation still remains poorly investigated. In this work, we undertake an in-depth investigation of the growth of stellate mesoporous silica (STMS) shells around IO NPs cores and of their micro/mesoporous features respectively through time-lapse and *in situ* liquid phase transmission electron microscopy (LPTEM) and detailed nitrogen isotherm adsorption studies. We found here that the STMS shell features (thickness, pore size, surface area) can be finely tuned by simply controlling the sol-gel reaction time, affording a novel range of IO@STMS core@shell NPs. Finally, regarding the responses under alternating magnetic fields and NIR light which are evaluated as a function of the silica structure, IO@STMS NPs having a tunable silica shell structure are shown to be efficient as T_2 -weighted MRI agents and as heating agents for magneto- and photoinduced hyperthermia. Furthermore, such IO@STMS are found to display anti-cancer effects in pancreatic cancer cells under magnetic fields (both alternating and rotating).

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1. Introduction

Nowadays, a true challenge in the field of nanomedicine is the development of multifunctional materials that could be used

to perform imaging, drug delivery and other innovative therapies using a single platform, allowing the reduction of side effects and the improvement of diagnosis and therapeutic efficiency.^{1,2} Among the potential materials, core-shell iron

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oxide@silica nanoparticles (IO@silica NPs) are particularly interesting systems as the association of these two materials naturally combines several advantageous properties. Indeed, IO NPs are already commercially used for magnetic resonance imaging (MRI) as they are very good T_2 contrast agents.^{3–6} In addition, they are recognised as efficient heating agents under an alternating magnetic field (AMF) stimulus^{3,7–10} and their potential as heating agents by near-infrared (NIR) light irradiation is also emerging.^{11–14} Last but not least, IO NPs are known to have low toxicity and to be well internalised and degraded by cells,^{3,4} and given all these features, have great potential for theranostic applications. However, without a robust and efficient coating strategy, they suffer from rapid blood capillary agglomeration and elimination, reducing their efficiency.^{15,16} Among different coating possibilities (polymers, dendrimers, specific biomolecules, silica¹⁶), synthesising a porous silica (MS) shell around IO NPs presents some interesting features. Indeed, the size and shape of silica nanomaterials are tunable, the surface chemistry is versatile and very importantly, silica is recognised as a generally safe material by the FDA.^{17–19} Furthermore, MS NPs are reported to degrade *in vitro* and *in vivo* and the main dissolution product, silicic acid, is reported to be water soluble and non-toxic. Thus, the addition of an MS layer coating to IO NPs is very appealing for medical applications, especially for drug delivery.^{20–24}

Such MS shell coatings around IO NPs,^{20,21,25,26} but also other inorganic NPs,^{27–30} were usually synthesised using surfactant templating, with the most famous one being cetyltrimethylammonium bromide (CTAB). Control of the shell thickness is one of the crucial features of such a synthesis and it is mainly achieved by the amounts of reactants, as notably shown by Ye *et al.* with the molar ratio $[\text{CTAB}]/[\text{Fe}_3\text{O}_4]$.³¹ Recently, there has been tremendous interest in tuning the pore size, especially to tailor large pores around IO cores. Indeed, the use of CTAB leads to a pore size of ~ 3 nm, which is enough for the delivery of small drugs, but the delivery of larger molecules such as proteins, RNA or DNA has gained growing interest these last years.³²

Despite the fact that small pore MS coatings with adjustable shell thicknesses were already reported around superparamagnetic IO NPs (size in the range 10–25 nm), the tailoring of porous silica structures and tuned increases of pore size around such NPs remained quite limited. In some approaches, elegant pore size tuning was achieved using the swelling micelle method, which however limits the range of pore size at *ca.* 3–6 nm.³³ Very recently, interesting works have described the tunable design of MS shells having radially oriented pores around a magnetic core through so-called interfacial co-assembly in a bi-liquid phase where the addition of a water non-miscible apolar solvent was used as a way to expand/tune the pore size.³⁴ These works were nevertheless essentially focused on bigger controlled iron oxide-clusters (*ca.* up to hundreds of nms).^{35–38} For instance, by investigating different synthesis parameters (surfactant concentration, amount of organic solvent, or silica precursor, reaction temperature, reaction

time, *etc.*), Nemec and Kralj have developed versatile silica-shell morphologies around a wide range of magnetic inorganic cores, having hierarchical dual pore sizes from ~ 3 to ~ 40 nm, with centre-radial and raspberry-like pore geometries.³⁶ In another work, Fiedler *et al.*³⁸ have developed a powerful approach for synthesizing different silica shells of various thicknesses and porosities in the range *ca.* 5–10 nm that can be adjusted independently around IO cluster cores of various sizes, especially by changing the composition of the cyclohexane/TEOS phase.

As addressed in this present article, the pore increase can also be obtained by another method adapted from K. Zhang *et al.*³⁹ where CTA⁺, counterion tosylate (CTATos) is used to orient the silica structure towards stellate mesoporous silica STMS having a pore size of *ca.* 10–15 nm. Previously in our team, we synthesized and developed core-free STMS and core@shell IO@STMS (where IO NPs are made by thermal decomposition) for biological and environmental applications^{40–43} and the IO@STMS NPs were notably shown to be suitable systems for MRI, MHT and PHT.^{44–46} However, these IO@STMS core-shell nanostructures have been synthesised only at a given final core-shell size (*ca.* 120 nm) and to date, no work has reported in depth the growth mechanism of the STMS shell around IO NP cores or the possibilities to design IO@STMS NPs having a tailored shell structure and their resulting properties for nanomedicine applications (textural pore structure, colloidal stability, response under magnetic fields or NIR light...).

Hence to the best of our knowledge, there is no report that makes use of CTATos as a porogen surfactant combined with a controlled sol-gel reaction time applied around *ca.* 25 nm size iron oxide NPs (synthesized by thermal decomposition) to generate individual core@shell structures with tunable growth and pore structure (from 7 to 16 nm). Worthy of note is that in all the previous mentioned reports, no investigation of the silica pore structure effects on magneto- and photothermal dissipation and MRI properties was reported, and approaches based on the visualisation in real time through the *in situ* liquid phase TEM (LPTEM) method of the silica growth were not yet proposed.

In this work, we report the great control of the stellate mesoporous silica shell growth around these IO NPs, to afford a range of IO@STMS core-shell NPs designed with a tunable silica shell. First, with the aim of evidencing the silica shell growth with its reaction time around the IO NP core, we investigated the growth kinetic of STMS shells by time-lapse TEM imaging of the NPs taken out at different time-points of the synthesis and by performing *in situ* LPTEM imaging to have direct observation of the STMS shell growth. Then, using different synthesis times corresponding to different shell growths, denoted IO@STMS-*t* (*t* = 40, 60, 120 min, growth time), we deeply investigated the textural pore size properties (microporosity, mesoporosity) and the colloidal stability of these tunable core-shell NPs. We then evaluated the responses of the different IO@STMS-*t* NPs under external fields (magnetic field and NIR light) and the effects of pore structure or



shell thickness were discussed. Hence, their potential as T_2 contrast agents for MRI was evaluated by measurements of their relaxivities, and their potential as good heating agents for MHT and PHT was evaluated by specific absorption rate (SAR, $W\ g^{-1}$) measurements. Finally, the potential use of such IO@STMS- t NPs for anti-cancer applications was evaluated by investigating their cytotoxicity towards the pancreatic cancer cell line MIA Paca-2 in the presence and absence of magnetic field stimuli (alternating and rotating). The main concept of this work is represented in Scheme 1.

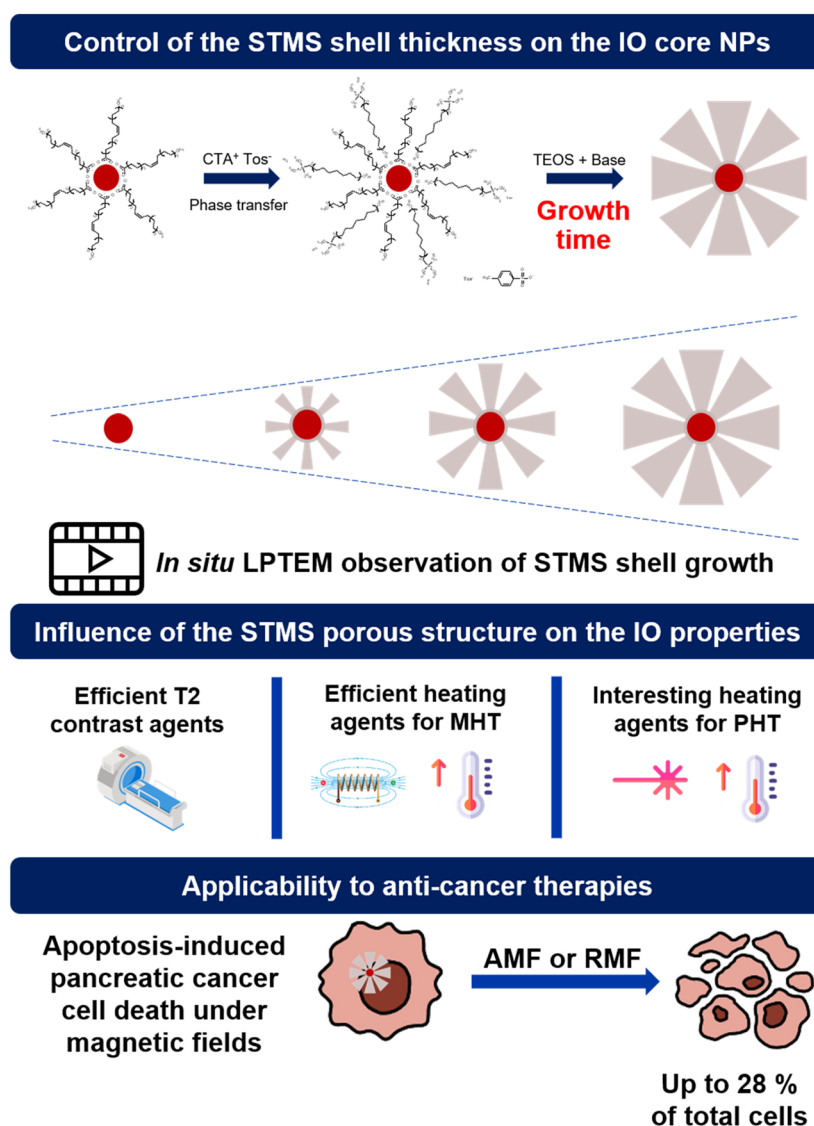
II. Materials and methods

II.1. Materials

All materials were used as provided. Anhydrous absolute ethanol (EtOH, CAS 64-17-5), chloroform (CHCl₃, CAS 67-66-3),

nitric acid 65% (HNO₃, CAS 7697-37-2), and acetone (C₃H₆O, CAS 67-64-1) were purchased from Carlo Erba Reagents. Dibenzylether (DBE, CAS C₁₄H₁₄O, CAS 103-50-4) and squalane (C₃₀H₆₂, CAS 111-01-3) were purchased from Acros Organics. Cetyltrimethylammonium *p*-toluene sulfonate (CTATos, CAS 138-32-9) and Trizma® base (AHMPD, CAS 77-86-1) were purchased from Sigma Life Science. Tetraethylorthosilicate (TEOS, CAS 78-10-4) was purchased from Aldrich chemistry, oleic acid (C₁₈H₃₄O₂, CAS 112-80-1) from Alfa Aesar, ferric chloride (FeCl₃, CAS 7705-08-0) from Sigma Aldrich, PBS from Sigma and sodium stearate (C₁₈H₃₅NaO₂, CAS 822-16-2) from TCI. Iron and indium (115In) plasma emission standards, 1 g L⁻¹, were purchased from AccuStandard.

For *in vitro* experiments, dimethylsulfoxide (DMSO, C₂H₆SO, CAS 67-68-5) and 3-(4,5-dimethylthiazolyl-2)-2,5-diphenyltetrazolium bromide (MTT, C₁₈H₁₆BrN₅S, CAS 298-93-1) were purchased from Sigma, and penicillin-streptomycin



Scheme 1 Representative scheme of the study.



(P4333) was purchased from Sigma-Aldrich. Dulbecco's Modified Eagle's Medium (DMEM, GlutaMAX™) and phosphate-buffered saline (PBS) were purchased from Life technologies. Foetal bovine serum (FBS) was purchased from Eurobio Scientific.

II.2. Synthesis of IO NPs

Thermal decomposition was used to synthesise oleic acid-stabilised iron oxide nanospheres with a mean diameter of around 20 nm following a recently reported procedure.⁴⁷ Briefly, iron stearate(III) was prepared by precipitation of sodium stearate and ferric chloride salts in an aqueous solution as described.⁴⁸ Then, 1.85 g (2 mmol) of the synthesised iron(III) stearate was mixed with 1.89 g of oleic acid (6.7 mmol) in a two-neck round-bottom flask in 19.5 mL (15.8 g) of squalane and 0.5 mL (0.53 g) of DBE. The mixture was heated at 120 °C and kept at this temperature for 60 min. The condenser was then connected to the flask and the solution was heated to 330 °C prior to being kept under reflux for 60 min under air. After cooling to room temperature, the viscous suspension was solubilised in 10 mL of chloroform. The NPs were precipitated by the addition of an excess of acetone and washed three times with chloroform and acetone (ratio 1 : 4, centrifugation 14 000g, 5 min). The NPs were then redispersed in chloroform and stored until further use.

II.3. Resuspension of IO NPs in deionised water

For some characterisations, oleic-acid-coated IO NPs were further coated with CTA⁺ surfactant with the aim of re-dispersing them in deionised water (dH₂O). To do so, 19.2 mg of CTATos was dissolved in 2 mL of dH₂O at 50 °C under stirring in a 5 mL glass vial. Then, the stirring was increased to 950 rpm and 488 µL of the IO solution at 4.1 mgFe mL⁻¹ in chloroform was added to the vial. The temperature was then increased to 65 °C and the solution kept under this vigorous stirring until full evaporation of the chloroform. The final solution was a 1 mgFe mL⁻¹ IO NP colloidal solution in dH₂O.

II.4. Synthesis of IO@STMS-*t* NPs

The protocol was used as described previously^{44–46} with some standardisations. Here, this procedure is described for IO NPs of 26.6 ± 2.1 nm. The amount and volume of IO NP solution are adapted with the IO NPs' diameter. In a 50 mL round bottom flask, 240 mg of CTATos was dissolved in 25 mL of dH₂O at 50 °C (oil bath) under stirring (300 rpm). Then, 27.6 mg of AHMPD pH buffer salt was added and dissolved. The stirring was then increased to high speed (950 rpm) prior to the addition of an adequate quantity of IO NPs in chloroform (31.91 mg, corresponding here to 7 mL). The oil bath temperature was increased to 65 °C to evaporate the chloroform. The mixture changed from hazy grey after addition of the IO NPs to limpid dark black after the evaporation of the chloroform. The mixture was left under stirring for ten additional minutes to be sure that all the chloroform was evaporated before increasing the temperature to 70 °C. Once the temperature of the oil bath was stabilised, the mixture was left

under stirring for 30 min to let its temperature stabilise too, prior to the addition of 1.5 mL of TEOS drop by drop for one min. The stirring was then reduced to 750 rpm and left for reaction.

For the kinetic study, a sample was collected at 5–10–20–30–40–50–60–90–120 min, quickly cooled with an ice bath and washed twice with EtOH (15 000g, 10 min). Without a kinetic study, the reaction was left for 40–60–90 and 120 min (series 1 and 2).

After reaction, the NPs were collected by centrifugation (12 000g, 20 min) and washed twice with 15 mL of EtOH (12 000g, 12 min). The CTATos was then extracted by dispersing the NPs in 20 mL of NH₄NO₃ (20 mg mL⁻¹ in EtOH) and heated at 70 °C under stirring. The extraction was then followed by two washings with 15 mL of dH₂O and two washings with 15 mL of EtOH (12 000g, 12 min). The extraction status was followed by the zeta potential in dH₂O as it was positive before extraction (~30 mV) and became negative when all the CTATos was extracted (~-20 mV). Here, a first extraction was performed for one night and a second for 1 h. The zeta potential was then stable, meaning that all the CTATos was removed. The NPs were then resuspended in EtOH prior to being used.

The particles were washed three times with dH₂O prior to being used for MRI, MHT or PHT properties evaluation. Particles were designated as IO@STMS-*t* with *t* being the STMS growth time.

II.5. *In vitro* biological experiments

II.5.1. Cell culture. The pancreatic cancer cell line MIA Paca-2 was cultured in complete DMEM medium containing 10% FBS and 100 IU mL⁻¹ penicillin/streptomycin under a humidified atmosphere of 95% air and 5% CO₂ at 37 °C. This DMEM/FBS/penicillin/streptomycin medium is designated as the “cell culture medium” for the following protocols.

II.5.2. Cytotoxicity. An average of 10⁴ cells per well were seeded in a 96-well plate, grown overnight, and incubated with increasing concentrations of IO@STMS-*t* NPs (from 0 to 100 µgFe mL⁻¹) in cell culture medium for 24, 48 or 72 h. The cells maintained in the incubation medium without NPs served as controls. Cell viability was then quantified using the MTT assay. The experiment was performed in quintuplicate.

Prior to use, the NPs (80 µL) were washed once with dH₂O prior to being resuspended in cell culture medium (centrifugation 12 000 rpm, 10 min).

II.5.3. Cellular uptake. An average of 3 × 10⁵ MIA Paca-2 cells were seeded onto 35 mm dishes in cell culture medium. The cells were incubated with nanoparticles at 0, 0.5 or 5 µgFe mL⁻¹ in cell culture medium for 72 h, at 37 °C under a 5% CO₂ atmosphere, washed twice with ice-cold PBS, and centrifuged (1500 rpm, 10 min). The amount of internalised IO NPs was determined through ICP-MS titration of Fe after acid digestion of the cell pellets in concentrated HNO₃ for 12 h. The results were expressed in pg of iron per cell and as the percentage of the initial mass of iron internalised. The experiment was performed in quadruplicate. Prior to being used, the



NPs (5 μL) were centrifuged and resuspended in dH_2O (centrifugation 12 000 rpm, 10 min).

II.5.4. Cytotoxicity under a magnetic stimulus: high-frequency alternating (AMF) or low-frequency rotating (RMF) magnetic field. An average of 25×10^3 MIA Paca-2 cells per compartment were seeded onto four-compartment CELLview™ dishes (Greiner Bio-One) and grown overnight in cell culture medium prior to being incubated with IO@STMS-*t* NPs at 0, 0.5 or 5 $\mu\text{gFe mL}^{-1}$ for 72 h at 37 °C. The cells were then washed with cell culture medium and exposed, or not, to an AMF ($f = 250$ kHz, $\mu_0 H = 20$ mT (16 kA m^{-1})) or an RMF ($f = 1$ Hz, $\mu_0 H = 40$ mT (32 kA m^{-1})) for 2 h. The temperature was strictly maintained at 37 °C and controlled using a thermal optical fibre probe (Reflex 4, Neoptix, Quebec City, QC, Canada) placed in the incubation medium during the magnetic field exposure. At the end of the experiment, the cells were placed under a humidified atmosphere with 5% CO_2 at 37 °C for further analyses. The cell death was then analysed by annexin V/propidium iodide labelling. The experiment was performed in quadruplicate. Prior to being used, the NPs (5 μL) were centrifuged and resuspended in dH_2O (centrifugation 12 000 rpm, 10 min).

II.5.5. Cytotoxicity under multiple magnetic stimuli: high-frequency alternating (AMF) or low-frequency rotating (RMF) magnetic field. An average of 10^4 MIA Paca-2 cells per compartment were seeded onto four-compartment CELLview™ dishes (Greiner Bio-One) and grown overnight in cell culture medium prior to being incubated or not with IO@STMS-40 NPs at 5 $\mu\text{gFe mL}^{-1}$ for 72 h at 37 °C. The cells were then washed twice with cell culture medium and exposed to an AMF ($f = 250$ kHz, $\mu_0 H = 20$ mT (16 kA m^{-1})) or an RMF ($f = 1$ Hz, $\mu_0 H = 40$ mT (32 kA m^{-1})) for 2 h every 24 h, for 3 days. The temperature was strictly maintained at 37 °C and controlled using a thermal optical fibre probe (Reflex 4, Neoptix, Quebec City, QC, Canada) placed in the incubation medium during the magnetic field exposure. The effects of magnetic field treatments were investigated on cell viability by counting the cell number using a cell counter (Beckman cell counter Z2) 24 h after the last exposure. Thus, cells were seeded on day 1 and incubated with particles from day 2 to day 4, and then sample 1 was exposed to nothing, AMF or RMF on day 5 and the cells were counted on day 6. Sample 2 was exposed to nothing, AMF or RMF on days 5 and 6 and the cells were counted on day 7. The same process was used for sample 3, which was exposed three times to nothing, AMF or RMF, and the cells were counted on day 8. For the sake of clarity, a scheme is presented in the Results and discussion section.

II.6. Characterisation methods

II.6.1. Transmission electron microscopy (TEM) and energy-dispersive X-ray analysis (EDX). The TEM imaging of IO and IO@STMS-*t* NPs was performed with a JEOL 2100 TEM instrument operating at 200 kV after deposition of the particles on carbon-coated copper grids. The JEOL Si(Li) EDX detector was used to determine the relative amount of Si

and Fe atoms. The open source software ImageJ was used to analyse the images and to determine the size distribution of the NPs.

In vitro samples were imaged using a Hitachi HU12A (Japan) TEM instrument operating at 75 kV. After their incubation with IO@STMS-*t* NPs ($t = 40, 60, 120$) at 5 $\mu\text{gFe mL}^{-1}$ for 72 h, the cells were washed with PBS and fixed with 4% glutaraldehyde in Sorensen buffer for 4 h at 4 °C. After the washes, the cells were post-fixed in 1% osmium tetroxide (osmium 2%, saccharose 0.25 mol L^{-1} , Sorensen buffer 0.05 mol L^{-1}) for 1 h at 20 °C, followed by washing with distilled water and uranyl acetate 2% for 12 h at 4 °C. After dehydration, 70 nm sections of cells embedded in EMBED 812 resin were stained with uranyl acetate and lead citrate. The samples could then be imaged.

II.6.2. *In situ* liquid phase TEM (LPTEM). *In situ* LPTEM was performed on a JEOL 2100F/Cs (S)TEM operating in continuous capture mode.

For these experiments, a Protochips liquid cell holder (Poseidon Select 510) was used for the *in situ* LPTEM analyses. The liquid cell holder contained a removable microchip composed of two Si_3N_4 membranes that isolated the liquid sample from the ultrahigh vacuum from the TEM column. The dimensions of the E-chip used in this work were 2×2 mm² or 6×4.5 mm². The thickness of the Si_3N_4 membranes was 50 nm, and the space between each membrane was 150 nm. The two microchips were washed in HPLC-grade acetone for ~2 min to remove a protective film and then washed with ethanol for 2 min to clean the Si_3N_4 membranes and they were submitted to plasma cleaning with an Ar/O₂ mixture for 30 s.

One of the microchips was placed on the *in situ* holder where a volume of 1 μL of the IO NPs was added. The second chip was then positioned on top to seal the liquid system (see Fig. S1† for a schematic representation). The sample holder containing the NPs was inserted into the microscope and TEOS was added by flow. Images were recorded using AXON Studio software. Images were exported from AXON studio and analysed with ImageJ.

II.6.3. X-ray diffraction (XRD). XRD patterns were collected with a Bruker D8 Discover diffractometer in Bragg Brentano geometry equipped with a monochromatic copper radiation source and an energy-resolved LYNXEYE XE-T detector in the 20–70° (2θ) range with a scan step of 0.03°. The measurements were performed at room temperature and high-purity silicon powder was used as an internal standard.

II.6.4. Dynamic light scattering (DLS) and zeta potential. DLS and zeta potential were performed on a Zetasizer Nano ZS (Malvern instruments). DLS measurements were recorded in triplicate at 25 °C and at a scattering angle of 173° using a 1 cm length plastic cell. The measurements were conducted with a concentration of 0.2 mg mL^{-1} of NPs in dH_2O .

Zeta potential measurements were recorded in triplicate at 25 °C using a DTS1070 folded capillary cell. The measurements were conducted in dH_2O at a concentration of NPs of 0.2 mg mL^{-1} or in PBS buffer (pH 7.4) when checking the extraction of the CTATos.



II.6.5. Nitrogen adsorption-desorption measurements. The textural properties of the prepared samples were studied by nitrogen adsorption-desorption measurements at $-196\text{ }^{\circ}\text{C}$. The nanoparticles were degassed under vacuum at ambient temperature (around $20\text{ }^{\circ}\text{C}$) for 3 h to desorb the moisture before analysis. The specific surface area was calculated by the Brunauer-Emmett-Teller (BET) method. Pore volume and pore size distribution were determined using the desorption branch by the Barrett-Joyner-Halenda (BJH) method, which is well suited for mesopore analysis. The Horvath-Kawazoe model was used for determining the pore-size distribution in a micro-pore analysis from a single adsorption.

II.6.6. Iron titration

II.6.6.1. By NMR ^1H -relaxometry. T_1 relaxation time measurements were used to quantify the amount of iron in the NPs for series 1 of IO@STMS-*t* NPs. To do so, 100 μL of IO@STMS-*t* NPs was collected and dried. Then, 323 μL of HNO_3 (65%) was used to completely dissolve the IO@STMS-*t* NPs. Some heating at $60\text{ }^{\circ}\text{C}$ could be used to help this digestion step. The sample was diluted precisely in a 10 mL calibrated flask and the T_1 relaxation was measured. The amount of iron was determined using a calibration curve established by measuring the longitudinal relaxivity r_1 of a standard solution of iron(III) nitrate at 2% of HNO_3 (calibration curve presented in Fig. S2†). The variation of the relaxation rates ($1/T_1$) as a function of $[\text{Fe}^{3+}]$ from 0 to 3.6 mmol L^{-1} was plotted and used for the calculations.

II.6.6.2. By inductively coupled plasma-mass spectrometry (ICP-MS). The iron content for series 2 of IO@STMS-*t* NPs and of the biological samples collected from the cellular uptake experiments was measured on an Agilent 8900 ICP-MS Triple Quad instrument. First, the cells were resuspended in 500 μL of dH_2O in order to be transferred to a glass vial for their digestion with 323 μL of HNO_3 (65%) for one night. The samples were then partially diluted in dH_2O , filtered with a $0.45\text{ }\mu\text{m}$ sterile PES syringe filter, and then diluted to 10 mL using a calibrated flask. The amount of acid was then reduced for the ICP-MS analysis by diluting 1.60 mL of sample to a final volume of 5 mL. Indium (10 ppb) was added as internal standard. Linear calibration functions were obtained (r^2 of ≥ 0.999) and are presented in Fig. S3†.

II.6.7. Relaxometry. A Bruker Minispec 60 working at a Larmor frequency of 60 MHz for protons (1.41 T) at $37\text{ }^{\circ}\text{C}$ was used to measure the longitudinal T_1 and transversal T_2 relaxation times of IO@STMS-*t* NPs. The longitudinal relaxivity r_1 and transverse relaxivity r_2 values were calculated according to the general equation of relaxivity given in eqn (1):

$$R_i = R_i^0 + r_i \times [\text{IO@STMS}] \quad (1)$$

where R_i is the respectively longitudinal ($i = 1$) or transverse ($i = 2$) relaxation rate ($R_i = 1/T_i$ in s^{-1}) in the presence of the NPs, R_i^0 the relaxation rate of the aqueous medium (in the absence of the NPs) and r_i the corresponding relaxivity value of the NPs (in $\text{s}^{-1} \text{mM}^{-1}$). To perform this experiment, the particles were diluted at 4–2–1–0.5–0.25 mmol Fe L^{-1} in dH_2O .

II.6.8. Magnetothermal measurements

II.6.8.1. By AC magnetometry. The heating efficiency of IO@STMS-*t* NPs was measured by AC magnetometry using the AC Hyster™ setup from NanoTech Solutions with pick-up coil technology.⁴⁹ To do so, 40 μL of aliquots of freshly sonicated suspensions of IO NPs in chloroform at 3.35 mgFe mL^{-1} or of IO@STMS-*t* NPs in EtOH at 0.5 mgFe mL^{-1} (thus 4.6 mgIO mL^{-1} and 0.69 mgIO mL^{-1} respectively) was introduced into 3 mm diameter 4 inches length NMR tubes (VWR, France). The magnetisation cycles $M(H)$ were then measured three times with a delay of 45 s between each measurement at a frequency f of 280 kHz and an amplitude H of 20 kA m^{-1} . Further measurements were performed some days later after sonication at 280 kHz or 217 kHz and 24 kA m^{-1} . The measured cycles were averaged and normalised by the exact weight of IO present in the tube to get the mass magnetisation in $\text{A m}^2 \text{kg}^{-1}$.

II.6.8.2. By calorimetry. The temperature profiles under an AMF stimulus were performed using a D5 series instrument equipped with a G2 multi-mode 1500 W driver (NanoScale Biomagnetics™, nB) and a CAL1 coil under MaNiaC™ software. Standard HPLC 1.5 mL vials well adapted for magnetothermal measurements were used and filled with 1 mL of IO-CTA⁺ or IO@STMS-*t* NPs at 0.5 mgFe mL^{-1} in dH_2O . An AMF with a frequency f of 303.50 kHz and an amplitude H of 300 G (24 kA m^{-1}) was applied and the temperature profiles were recorded for 5 min.

II.6.9. Photothermal measurements. The temperature profiles under NIR light irradiation were performed using an EA-PS 2042 10B power supply coupled with a 1064 nm wavelength laser beam generated by a CCMI apparatus from AeroDIODE™. The temperature was recorded using a temperature sensor. Briefly, 1 mL of IO-CTA⁺ or IO@STMS NPs at 0.5 mgFe mL^{-1} in dH_2O was added to a 1 cm path length plastic cell inserted in a closed CVH100 cuvette holder cell from Thorlabs. The laser was then applied with a power of 1.020 W and the temperature profiles recorded for 10 min.

II.6.10. Processing of the hyperthermia curve measurements to obtain the specific absorption rate (SAR) and the intrinsic loss power (ILP). The SAR is calculated using different equations depending on the method used to measure the hyperthermia properties of the NPs. Eqn (2) is used in the case of AC magnetometry:⁵⁰

$$\text{SAR} (\text{W g}^{-1}) = \frac{\mu_0}{m_{\text{IO}}} \times f \times \oint_{\text{cycle}} M_t(H_t) dH_t \quad (2)$$

where μ_0 is the magnetic permeability of a vacuum, m_{IO} is the mass of iron oxide in g (obtained as $m_{\text{IO}} = 1.38 \times m_{\text{Fe}}$) and $M_t(H_t)$ is the magnetization curve, while eqn (3) is used in the case of calorimetry (MHT and PHT):

$$\text{SAR} (\text{W g}^{-1}) = m_s \times \frac{C_s}{m_{\text{IO}}} \times \left[\frac{dT}{dt} \right]_{t=0} \quad (3)$$

where m_s and C_s are the mass in g and the heat capacity in $\text{J kg}^{-1} \text{K}^{-1}$ of the solvent respectively, m_{IO} is the mass of iron



oxide in g and $(dT/dt)t = 0$ is the derivative function of the temperature at $t = 0$. This term is determined by fitting the temperature curve with a second order polynomial function as described by Périgo *et al.*⁵⁰ The mass of iron oxide is determined as $m_{\text{IO}} = 1.38 \times m_{\text{Fe}}$ as calculated from the relative molar masses of Fe_3O_4 and Fe .

In order to compare the heating efficiency of NPs between different labs or studies, the ILP can be calculated following eqn (4):

$$\text{ILP}(\text{nH m}^2 \text{ kg}^{-1}) = \frac{\text{SAR} \times 10^9}{f \times H^2} \quad (4)$$

where the SAR is in W g^{-1} , f is the frequency of the magnetic field in kHz and H is the amplitude of the magnetic field in A m^{-1} .

II.6.11. Magnetic measurements (VSM). The magnetization curve of IO@STMS- t NPs was measured on a VSM magnetometer (PPMS, Quantum Design, USA) at room temperature. The samples were prepared by drying 5 μL of NP solution. VSM magnetization cycles were measured by applying a field from -3000 to 3000 mT with a sampling rate of approximately 10 mT s^{-1} .

II.6.12. MTT assay. First, 10 μL of MTT at 5 mg mL^{-1} was added to each well and the multi-well plate was then placed at 37°C for 2 h of incubation. After medium removal, 100 μL of DMSO was added and a new incubation of 1 h at 37°C was performed. The absorbance was then measured at 570 nm.

II.6.13. FITC-annexin V/propidium iodide labelling. Cell death was analysed by annexin V/propidium iodide (AnnV/PI) labelling 4 h after magnetic field exposure using a Cell Meter Annexin V apoptosis assay kit (AAT Bioquest, Sunnyvale, CA, USA) in accordance with the manufacturer's instructions. The counting of labeled cells was carried out through the analysis of confocal microscopy images (LSM 510, Zeiss) representing populations of ~ 2000 cells per experiment, using ImageJ software.

II.6.14. Statistical analysis of *in vitro* experiment results. Results are expressed as the mean $\pm$ SEM of at least three independent experiments. The statistical analysis was performed using a one-way ANOVA test and Tukey *post-hoc* test. Differences were considered significant when $p < 0.05$ and statistical significance was set as $*p < 0.05$, $**p < 0.01$, $***p < 0.001$, $****p < 0.0001$.

III. Results and discussion

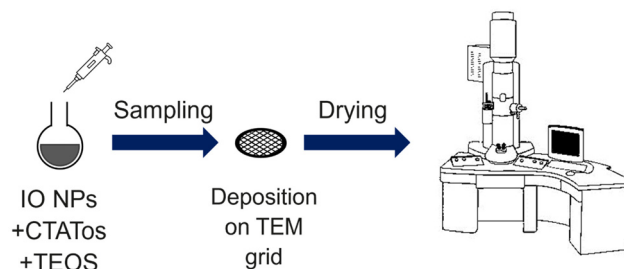
III.1. STMS growth on IO NPs through TEM imaging

III.1.1. Time-lapse TEM imaging. First, IO core NPs were synthesised by thermal decomposition and characterised as previously reported.^{47,48} As can be seen in Fig. S4,† the particles presented overall a spherical shape and a diameter of $ca. 26.6 \pm 2.1 \text{ nm}$. The XRD pattern showed the characteristic diffraction peaks of the inverse spinel structure, with an additional peak that could come from residual NaCl salts from the synthesis (marked with *). These spinel IO NPs can be assimilated

to oxidized magnetite ($\text{Fe}_{3-x}\text{O}_4$) as stated in several previous works achieved in our group.^{47,51,52} Then, the IO NPs were used for the synthesis of IO@STMS NPs following the surfactant phase-driven sol-gel reaction shown in Scheme 1 using an adapted and standardised protocol based on the one reported in previous works.^{44–46} Briefly, the oleic-acid stabilised IO NPs stored in chloroform were suspended in dH_2O thanks to a phase transfer helped by the CTATos surfactant under high-speed stirring and at 65°C . Controlled chloroform evaporation upon heating ensured phase transfer with the establishment of hydrophobic interactions between the surfactant molecules and the hydrophobic oleic acid bound to the IO NPs. The apolar tails of the surfactant and oleic acid interact by creating an interdigitated lipid-like bilayer. The positively charged external ammonium groups ensure electrostatic repulsion between the IO NPs in aqueous solution and overall a satisfying colloidal stability. Once the chloroform was evaporated and the CTATos-oleic-acid stabilised IO NPs were thus suspended in dH_2O , the TEOS precursor was added to the solution and condensed around the surfactant phase to form the STMS shell, displaying large porosities.

The very first step of our study was to evaluate the possibility to control the STMS shell growth around the IO NP core. To do so, we performed the “120 min” classical synthesis as described in the Materials and methods section (II.4.) and collected a sample at different time points. The reaction in the sample aliquot was stopped by immersion in an ice bath and the particles were then washed prior to being imaged by TEM, as represented in Scheme 2. The obtained time-lapse of the reaction with the corresponding particle size distribution is presented in Fig. 1. As can be seen, a few condensed silica spicules around the IO NPs can be observed at 5, 10, 20 and 30 min post-addition of TEOS, but a good silica layer with stellate morphology can be seen from 40 min. Its growth is then clearly observable at 50, 60, 90 and 120 min.

EDX measurements were also performed on the samples (Fig. 2A) and show a start of the silica growth between 20 min and 30 min post-addition of TEOS as the Si/Fe mass ratio increased from $ca. 0.06 \pm 0.03$ to $ca. 0.13 \pm 0.03$ and then to $ca. 1.03 \pm 0.21$ at 40 min post-addition of TEOS. Altogether, these analyses show that the STMS growth starts from 30 min post-addition of TEOS but that 40 min of reaction is necessary to



Scheme 2 Schematic representation of the procedure followed to perform the time-lapse TEM imaging.



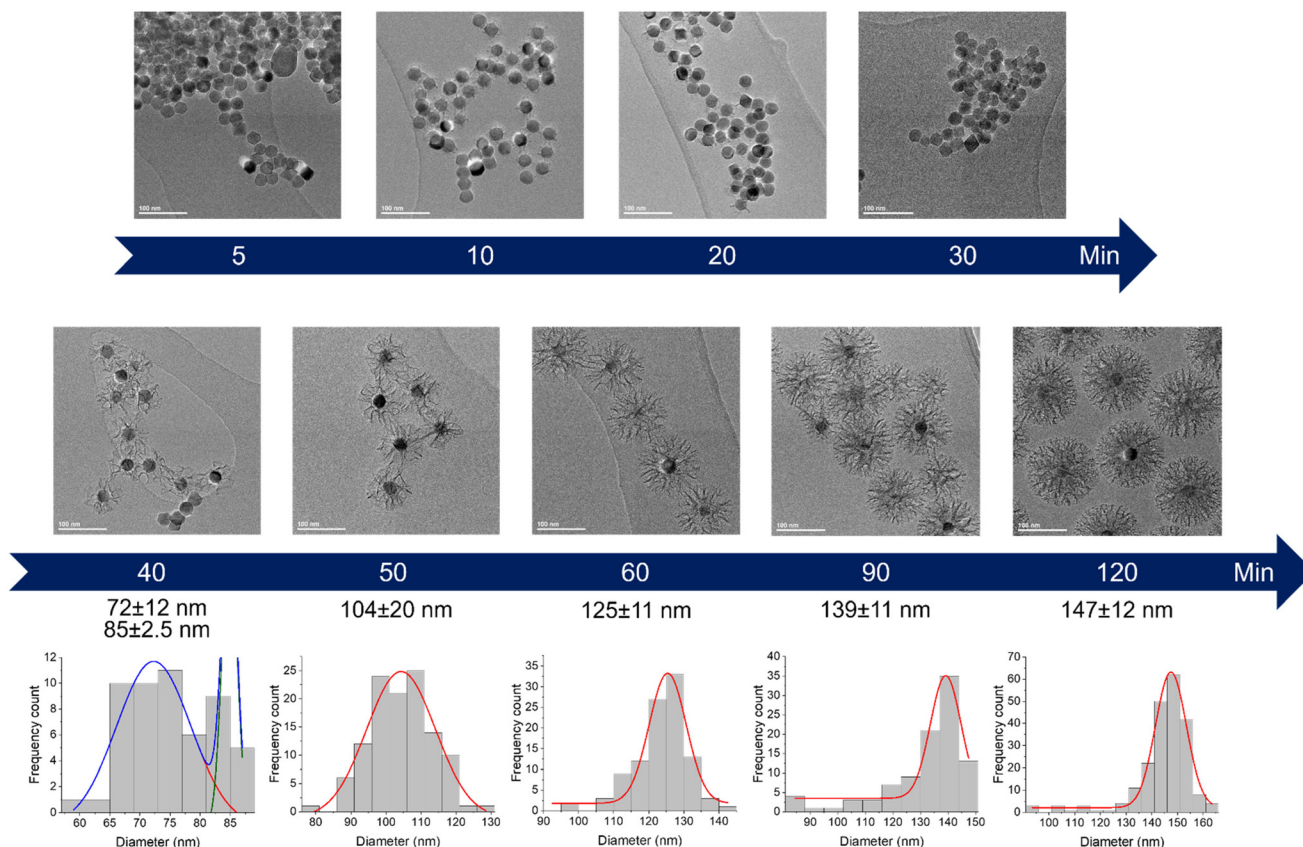


Fig. 1 Kinetic tracking of STMS growth on IO NPs by TEM with the corresponding IO@STMS NPs diameter distribution analysed by Gaussian fit.

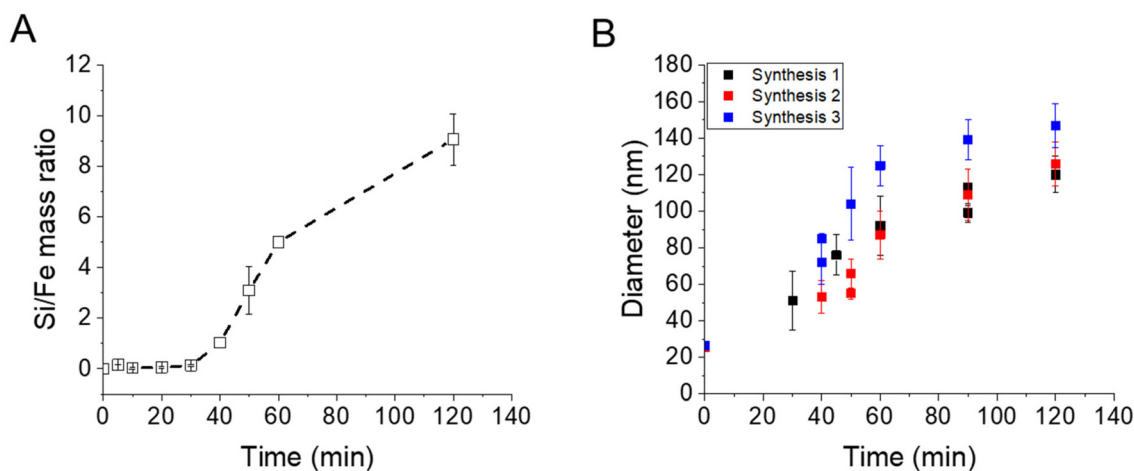
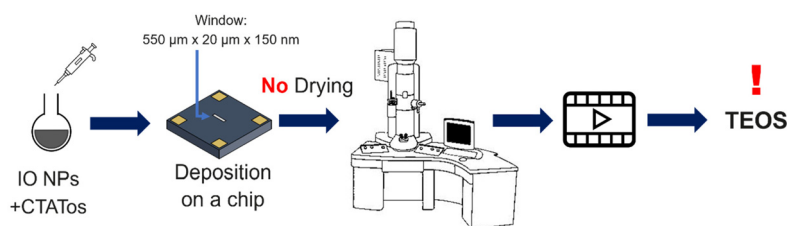


Fig. 2 (A) Kinetic tracking of STMS growth on IO NPs by Si/Fe mass ratio obtained by EDX. (B) Evolution of the IO@STMS NP final diameters measured by TEM on three different syntheses.

get a good STMS shell. The reproducibility of this timing was evaluated on two other syntheses performed on different IO NP batches. The evolution of the final IO@STMS NP diameters determined by TEM is shown in Fig. 2B and confirms that the kinetics of the reaction remains the same from batch to batch.

All these experiments show that we can modulate the STMS shell thickness around the IO NP core by simply playing on the synthesis time and by taking into consideration that a minimum of 40 min of reaction is necessary to get a minimal STMS shell.





Scheme 3 Schematic representation of the procedure followed to perform the first *in situ* LPTEM experiment.

III.1.2. *In situ* LPTEM imaging. The development and application of sealed environmental cells TEM (EC-TEM) facilitates the real-time monitoring of structural and chemical transformations in materials within reactive gas atmospheres at elevated temperatures, as well as (electro)-catalytic reactions in liquid environments. This innovative EC-TEM technology stands out as one of the most effective instruments for investigating local structure and material evolution in realistic environments, offering sub-nanometer resolution.^{53–55} Given its capabilities, EC-TEM serves as the ideal tool for tracking the growth processes occurring in the STMS growth on the IO NPs. The procedure followed for the first attempt is represented in Scheme 3. Basically, it was chosen to prepare the IO NP suspension in dH_2O , to place it in the chip, to find a region to observe by LPTEM and finally to inject the TEOS precursor. The obtained video is presented in ESI Video 1† and a time-lapse series was extracted and is presented in Fig. 3. It has to be noted that we worked in the liquid phase, rendering the TEM focalisation harder than in the dried phase. What can be specifically seen in these images is first the growing of a

shadow around the IO NPs and then the growing of “arms” around the particles, particularly well seen in the last pictures. Such “arm” structures can also be seen on other particles that were imaged in another region of the chip (Fig. S5†). These arms are the silica shell growing around the IO NPs and correspond quite well with the stellate structure of this shell.

The initial experiment provided an insightful glimpse into the silica growth on the IO NPs. However, we identified two key limitations: uncertainty regarding the timing of TEOS introduction into the cell, and concerns about the homogeneous dilution of TEOS on the cell. Thus, we performed a second experiment where we added the TEOS to the IO NP suspension before adding the solution to the chip for *in situ* LPTEM observation, as represented in Scheme 4. The resulting video is given in ESI Video 2,† and the corresponding time-lapse is given in Fig. 4. The growth of the STMS shell can clearly be seen by the increase in size of most of the particles visible in the selected region. The diameter of one particle was measured all along the duration of the video, which here corresponds precisely to the time post-addition of TEOS. As

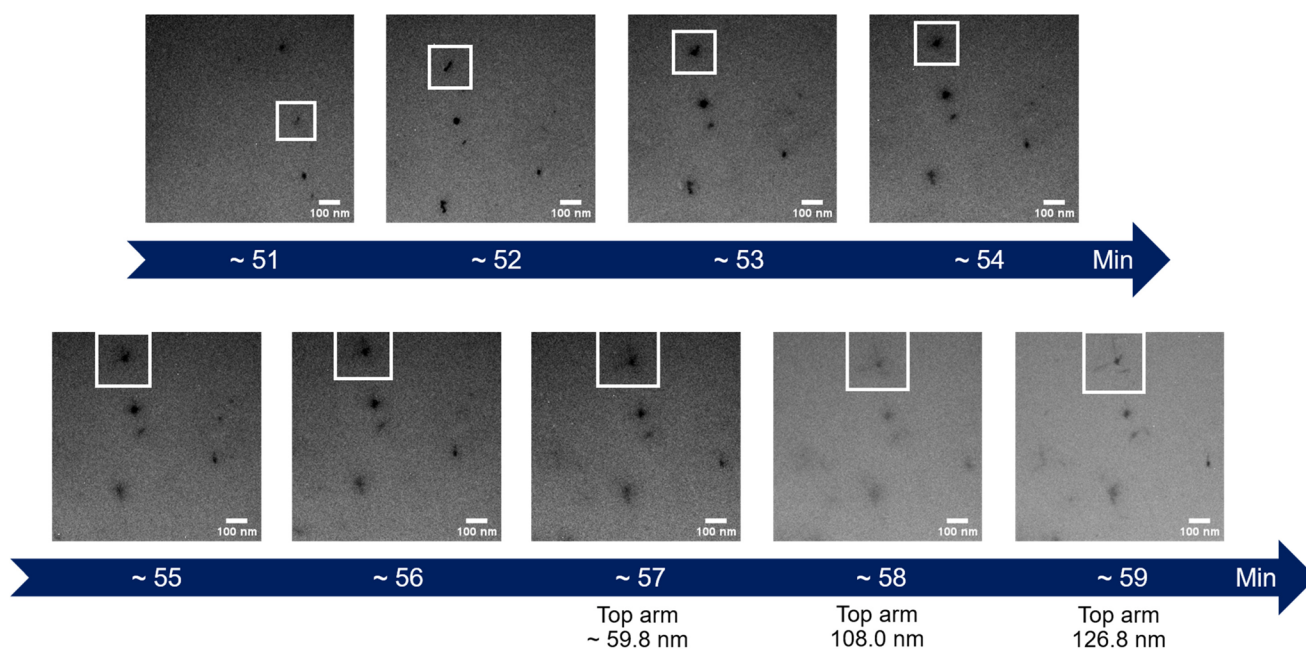
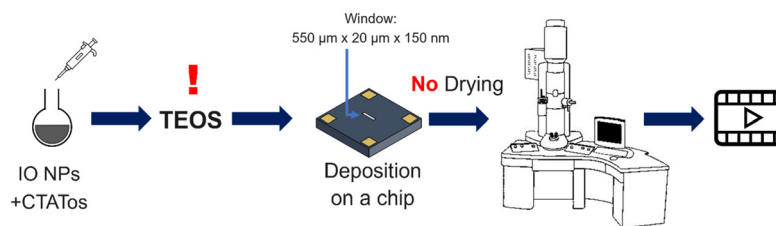


Fig. 3 Time-lapse images extracted from the *in situ* LPTEM video ESI Video 1.† The white box represents an interesting particle whose diameter is measured on the last images.





Scheme 4 Schematic representation of the procedure followed to perform the second *in situ* LPTEM experiment.

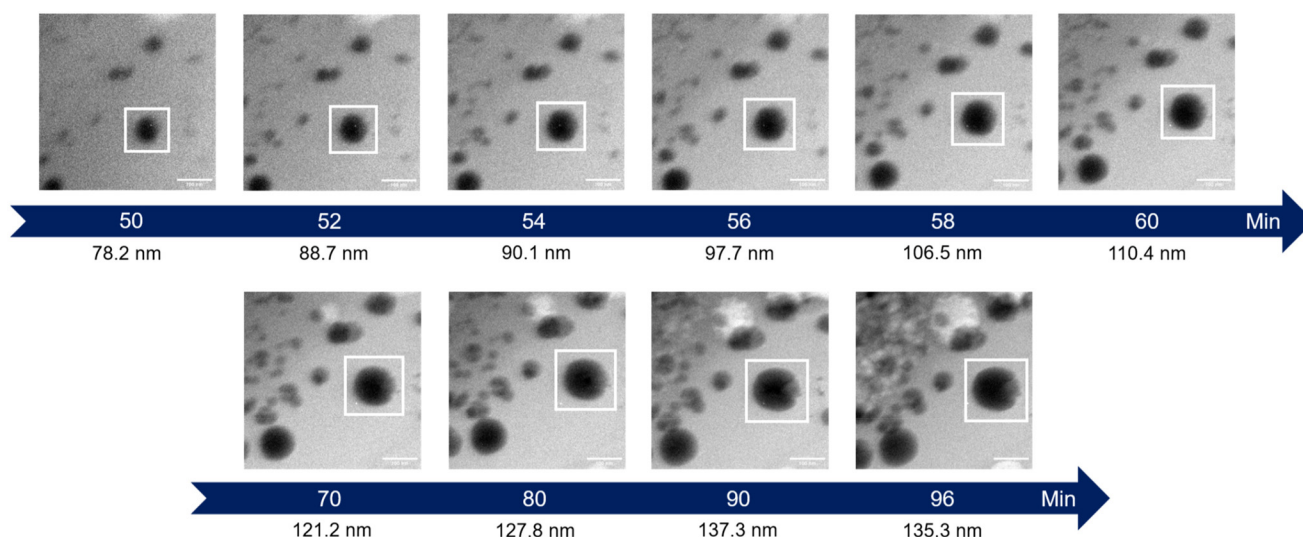


Fig. 4 Time-lapse images extracted from the *in situ* LPTEM video ESI Video 2.† The white box shows the particle whose diameter is indicated under the images.

evident from the extracted time-lapse series (Fig. 4) but also from the graph presented in Fig. 5, the growth was quite fast between 50 and 60 min post-addition of TEOS, with a growing speed estimated at $8.1 \pm 0.5 \text{ nm min}^{-1}$, but then it slowed down. This variation in growth kinetic compared with the experiment may stem from the confined conditions employed,

without any stirring and thus also under diffusion-limited conditions.

These two attempts of *in situ* LPTEM imaging allowed us to obtain two complementary videos, one showing the “arm” structure of the stellate silica around the IO NPs, and the second showing the homogeneous growth of the silica shell on the IO NPs core.

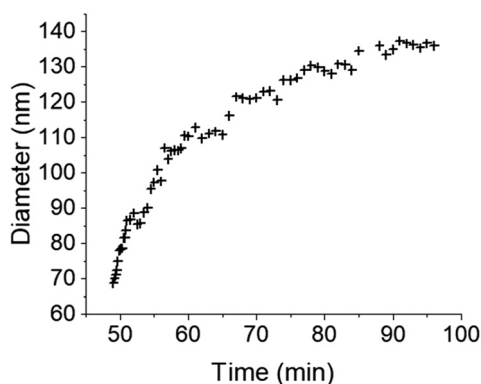


Fig. 5 Evolution of the IO@STMS NP final diameters measured on the *in situ* LPTEM images from ESI Video 2.†

III.2. Pore structure characterisation of tailored IO@STMS-*t* NPs

In a second step, we wanted to finely characterise the structural features of IO@STMS NPs having tuned shell thicknesses. For the sake of clarity, we denote the different batches as IO@STMS-*t* where *t* represents the growth time for the rest of the study, and chose the times according to the previous results: 40, 60 and 120 min. It is noteworthy that IO@STMS-*t* NPs usually come from the same IO NP batch, but sometimes also from another IO NP batch of very similar diameter. Such cases will be specified.

Fig. 6 shows the synthesised particle batches from the first series (denoted series 1 for the rest of the study). The IO@STMS-120 were the ones obtained by performing the kinetic study. The IO@STMS-40 and IO@STMS-60 were syn-



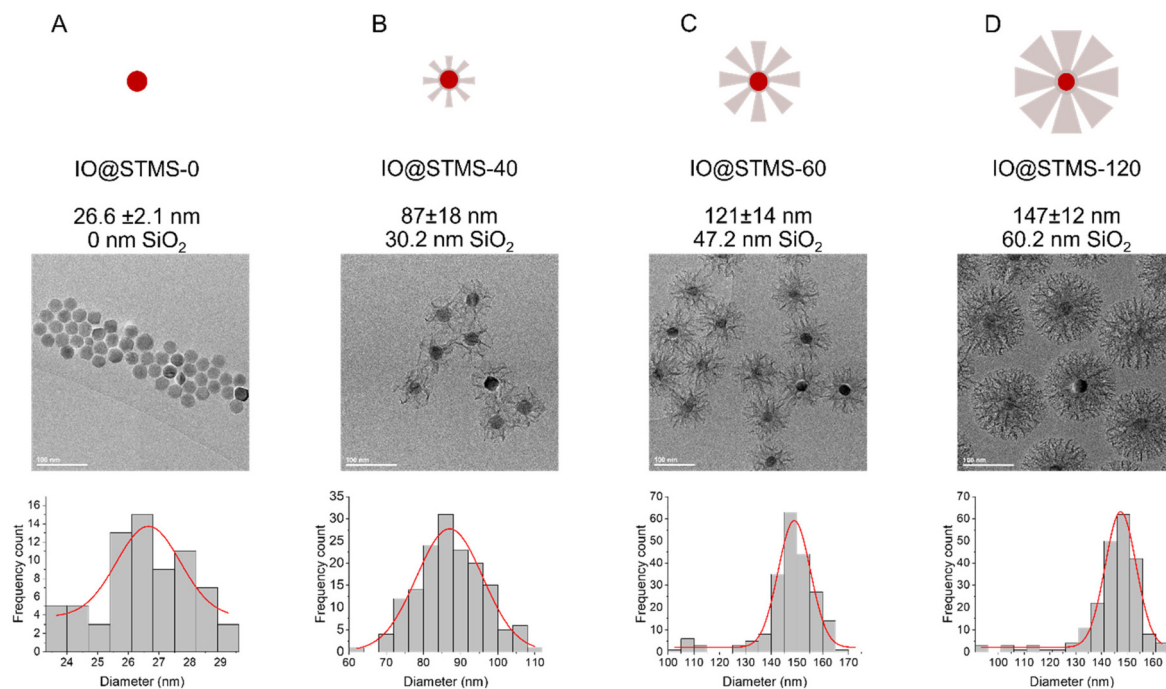


Fig. 6 Schematic representation, TEM and distribution of diameters for (A) IO@STMS-0, (B) IO@STMS-40, (C) IO@STMS-60 and (D) IO@STMS-120 obtained from series 1.

thesised in other experiments, whose respective diameters measured on TEM images correlate very well with the ones taken out during the kinetic study, as we got a diameter here of *ca.* 87 ± 18 nm *versus* *ca.* 85 ± 2.5 nm during the kinetic study for 40 min of reaction and *ca.* 121 ± 14 nm *versus* *ca.* 125 ± 11 nm for 60 min, showing again the good reproducibility of the synthesis and feasibility of the control of the STMS shell thickness.

We then characterised the porous texture/structure of the IO@STMS-*t* by nitrogen adsorption-desorption measurements in order to get precise information on the structural characteristics of the particles in terms of surface area, pore size, and pore volume (Fig. 7). We also added IO@STMS-90 to this porous texture study as it gave the same diameter as the IO@STMS-120 (*ca.* 149 ± 12 nm, see Fig. S6† for the TEM characterisation). In addition, for this characterisation, we had to synthesise a new batch of IO@STMS-40 due to the very low quantity obtained for the batch presented in Fig. 6 (see Fig. S7† for the TEM characterisation).

The nitrogen adsorption-desorption curves are presented in Fig. 7 and the BET surface (S_{BET}), pore volume (V) and representative pore size (D_p) values are regrouped in Table 1. Firstly, the global aspect of the adsorption-desorption isotherms (Fig. 7B, E, H and K) shows the dual micro-mesoporous character of our samples. Indeed, the shape of the curve at a relative pressure up to 0.6, with the small increase and then the plateau, corresponds more to a type I isotherm (according to the IUPAC classification), which is attributed to microporous materials (pore size below 2 nm); while the presence of the sharp increase at higher relative pressures (from

0.8) with the small hysteresis loop corresponds more to a type IV or V isotherm, which is attributed to mesoporous materials (pore size from 2 to 50 nm).⁵⁶ Unfortunately, the non-representative shape of the hysteresis loop does not allow determination of the pore shape. However, other characteristics could be extracted from the nitrogen adsorption-desorption measurements.

Secondly, the BET surface area could be determined and it is clear that the value decreases as the silica growth time increases, as it dropped from *ca.* $579 \text{ m}^2 \text{ g}^{-1}$ for IO@STMS-60 to *ca.* $466 \text{ m}^2 \text{ g}^{-1}$ and *ca.* $333 \text{ m}^2 \text{ g}^{-1}$ for IO@STMS-90 and IO@STMS-120 respectively. The global pore volume followed the same trends, as it dropped from *ca.* $1.59 \text{ cm}^3 \text{ g}^{-1}$ for IO@STMS-40 to *ca.* $1.16 \text{ cm}^3 \text{ g}^{-1}$ and *ca.* $0.67 \text{ cm}^3 \text{ g}^{-1}$ for IO@STMS-90 and IO@STMS-120 respectively. These first observations mean that increasing the reaction time allows the STMS shell first to grow rapidly in a radial way (from the NPs to the exterior), thus getting a higher silica shell thickness with large openings and thin walls, and then to condense inside the pores, thus getting a denser shell and thicker walls. One can object that the BET values and the global pore volume measured for the IO@STMS-40 are smaller (*ca.* $408 \text{ m}^2 \text{ g}^{-1}$ and *ca.* $1.03 \text{ cm}^3 \text{ g}^{-1}$ respectively) than the ones for IO@STMS-60 and thus do not follow the trend, but this is actually logical as the particles are still very small, especially for the batch used for this measurement. However, this kind of mechanism of STMS shell growth, that we represented in Scheme 5, is in great correlation with what is observed by TEM: we can clearly see large pores in the TEM picture “40 min” that we do not longer see in the TEM picture “120 min” in Fig. 1, and the



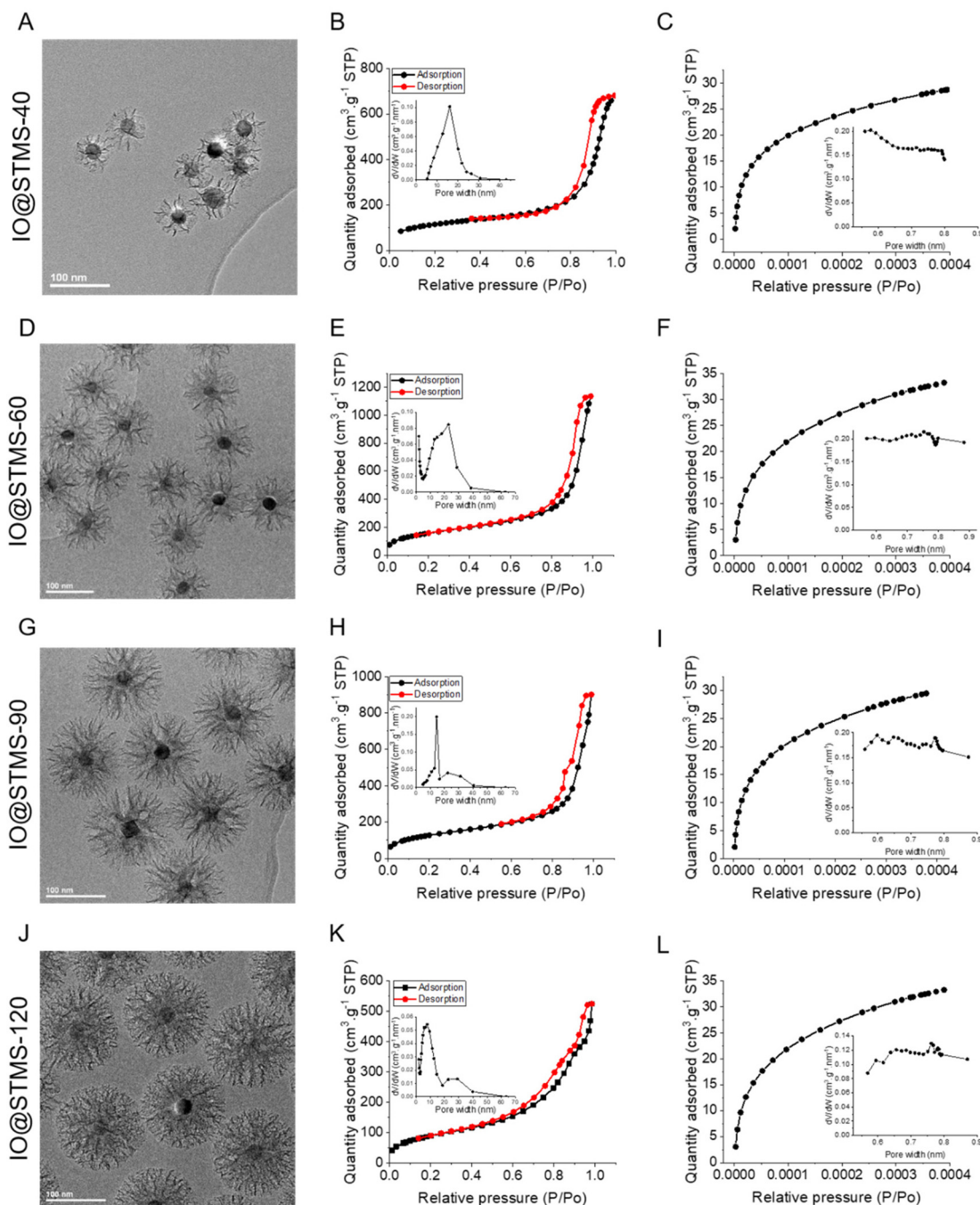


Fig. 7 TEM images (issued from Fig. 6, Fig. S6 and S7†), adsorption-desorption isotherms at high relative pressures including an inset with the BJH desorption pore volume plot, and adsorption isotherms at small relative pressures including an inset with the Horvath-Kawazoe differential pore volume plot for IO@STMS-40 (A–C), IO@STMS-60 (D–F), IO@STMS-90 (G–I) and IO@STMS-120 (J–L).

thin wall reminds us of the silica “arms” seen in the first *in situ* LPTM experiment (Fig. 3 and Fig. S5†). In addition, the comparison of IO@STMS-90 and IO@STMS-120 leads to the conclusion that even if the radial growth is done, the condensation reaction still occurs, reducing the global pore volume and resulting in a denser silica shell.

We nevertheless note that this global pore volume includes the volume of the micropores (V_{micro}), the volume of the meso-

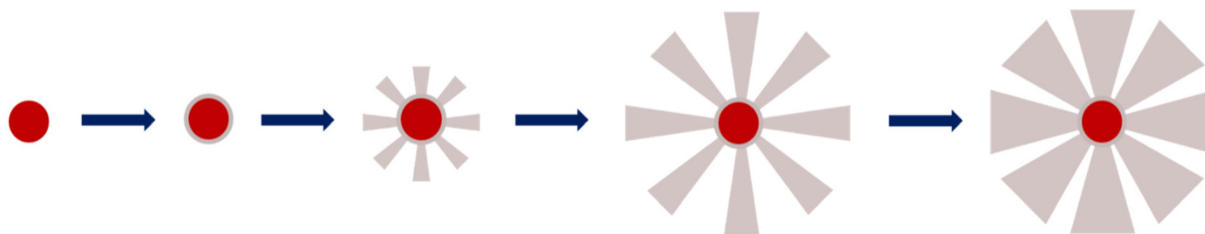
pores (V_{meso}) and the void volume corresponding to interparticle spaces. We thus used the Horvath-Kawazoe model (inset in Fig. 7C, F, I and L) and the BJH model (inset in Fig. 7B, E, H and K) to extract V_{micro} and V_{meso} and respectively their associated representative pore widths. The values reported in Table 1 confirm that we have a micro-mesoporous structure as V_{meso} does not corresponds to the total V_{pore} . The high values of V_{meso} compared with V_{micro} were expected as, even if the micro-



Table 1 Textural properties of the IO@STMS-*t* NPs (*t* > 0)

Sample	S_{BET} (m ² g ⁻¹)	V_{pores} (cm ³ g ⁻¹)	V_{meso} (cm ³ g ⁻¹)	V_{micro} (cm ³ g ⁻¹)	$D_{\text{p-micro}}$ (nm)	$D_{\text{p-meso}}$ (nm)
IO@STMS-40	408	1.03	0.92 (a)	0.041	0.5–0.8	16.1
IO@STMS-60	579	1.59	0.99 (b)	0.063	0.5–0.8	15.2
IO@STMS-90	466	1.16	0.70 (c)	0.056	0.5–0.8	14.5
IO@STMS-120	333	0.67	0.52 (c)	0.034	0.5–0.8	7.7

V_{meso} was determined by integrating the BJH pore volume plot between 5 and 24 nm (a) or between 5 and 18 nm (c). In the specific case of IO@STMS-60, the signal was decomposed to get the participation of the mesopores and the voids and the integration was performed according to the obtained peaks, thus being from 5 to 39 nm (b). V_{micro} was determined by integrating the Horvath–Kawazoe differential pore volume plot between 0.5 and 0.8 nm.

**Scheme 5** Schematic representation of the STMS shell growth mechanism around the IO NP core.

pores are present in a large amount as ultra-thin channels (around 0.5–0.8 nm diameter), they do not contribute importantly to the total pore volume and the mesopores still represent a largely higher volume. Furthermore, the Horvath–Kawazoe differential pore volume plot does not allow a determination of a representative micropore size. However, the decrease of V_{micro} with the increase of growth time is clear, and the quite low value of V_{micro} for IO@STMS-120 (*ca.* 0.034 cm³ g⁻¹) tends to show that the microporous component of the sample has been highly reduced by the silica condensation. The decrease of V_{meso} with silica growth time also seems logical given the previous conclusions. Regarding the pore size analysis, the evolution of the BJH desorption pore volume plot is quite interesting. Indeed, the plots show a bimodal distribution for IO@STMS-90 and IO@STMS-120, corresponding to a $D_{\text{p-meso}}$ of *ca.* 14.5 nm and *ca.* 7.7 nm respectively, with another small peak around 30 nm that is usually attributed to interparticle voids. However, the plots obtained for IO@STMS-40 and IO@STMS-60 do not show a bimodal distribution, meaning that the mesopores are still quite “open” up to 60 min of synthesis, *i.e.* large enough not to be very different in terms of interparticle voids on the plot. However, the analysis of these plots allowed the determination of a $D_{\text{p-meso}}$ of *ca.* 16.1 nm and *ca.* 15.2 nm for IO@STMS-40 and IO@STMS-60 respectively, which follows the global trends found with IO@STMS-90 and IO@STMS-120.

Altogether, these analyses confirm the micro–mesoporous character of the IO@STMS-*t* NPs and showed that the radial growth of the STMS shell is accompanied by a continuous silica condensation of the shell, even when the radial growth is stopped.

III.3. Colloidal stability of tailored IO@STMS-*t* NPs

Finally, the influence of the reaction time and thus of the STMS shell thickness on the colloidal stability of the particles was evaluated in EtOH (Fig. S8†) and in dH₂O (Fig. 8). The stability of the IO-CTA⁺ NPs in dH₂O (IO@STMS-0) was quite good after the phase transfer from chloroform to CTATos solution in dH₂O. The hydrodynamic diameter was determined to be *ca.* 45.9 ± 15.9 nm which indicates that some aggregates may be formed even with the surfactant covering. This is in correlation with the aggregation of the particles that we observed after a 24 h aging. Regarding the IO@STMS-*t* (with *t* > 0) NPs, the colloidal stability was overall good in both EtOH and dH₂O. The size graphs of intensity distribution show some very slight aggregation with the smaller STMS shell thickness and a progressive disappearance of these aggregates with the increase of the STMS shell thickness in both solvents. In addition, the size graphs of number distribution show that these aggregates were very few in the sample as only one peak is visible. Interestingly, the hydrodynamic diameter was found to be *ca.* 198 ± 2.4 nm (PDI *ca.* 0.24 ± 0.01), *ca.* 174 ± 0.3 nm (PDI *ca.* 0.19 ± 0.02) and *ca.* 177 ± 0.9 nm (PDI *ca.* 0.07 ± 0.01) in EtOH and *ca.* 183 ± 2.4 nm (PDI *ca.* 0.27 ± 0.02), *ca.* 163 ± 0.8 nm (PDI *ca.* 0.18 ± 0.02) and *ca.* 171 ± 1.1 nm (PDI *ca.* 0.07 ± 0.02) in dH₂O for IO@STMS-40, IO@STMS-60 and IO@STMS-120 respectively, indicating suitable colloidal stability.

The zeta potential of the particles was evaluated in dH₂O right after the colloidal stability study (Fig. S9†). The change from a positive value (*ca.* 45.4 ± 0.6 mV) for IO@STMS-0 to a negative value for IO@STMS-*t* (*ca.* -23.5 ± 0.2 mV, *ca.* -21.6 ±



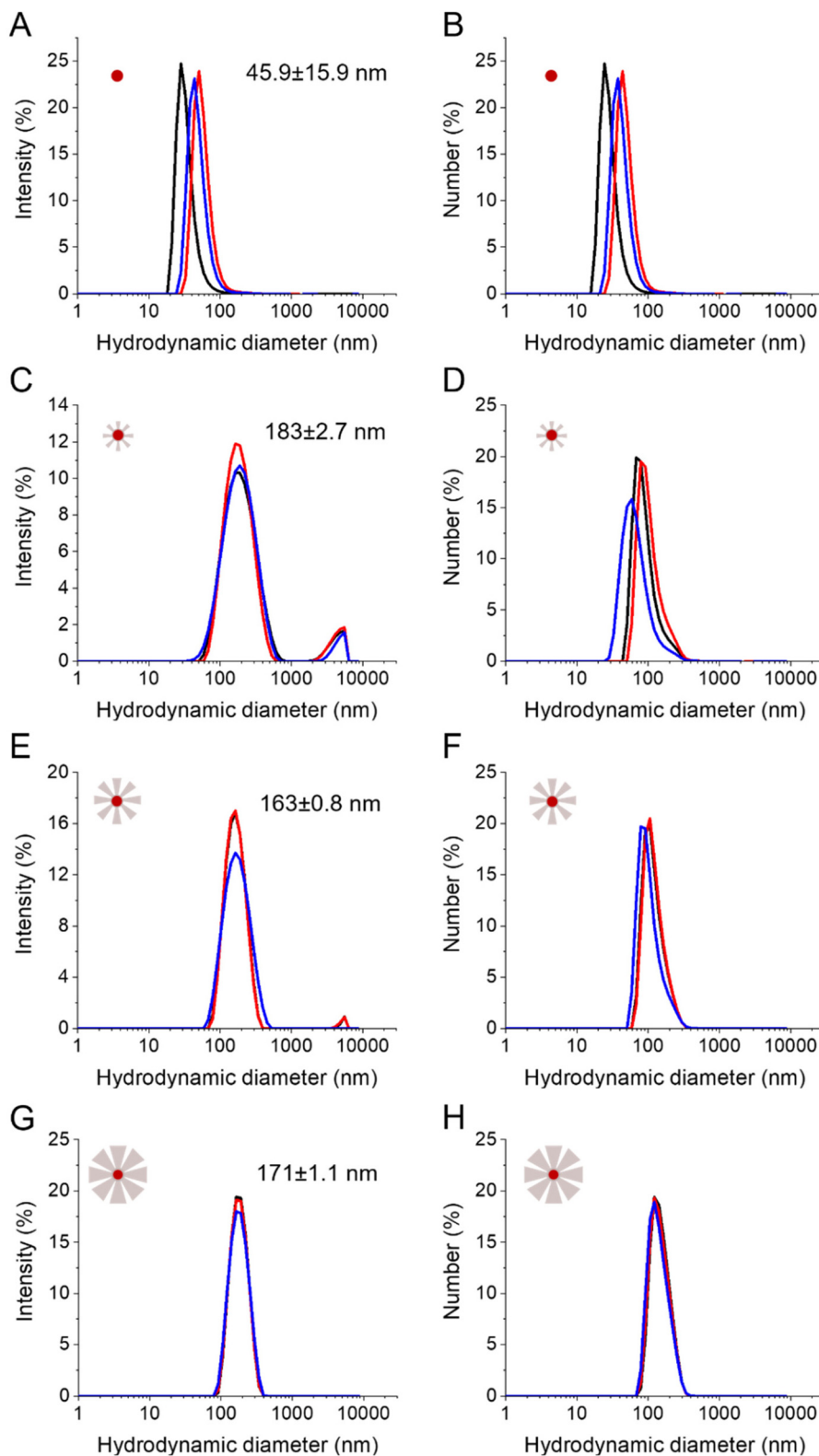


Fig. 8 Colloidal stability in dH₂O as shown from the distributions of intensity and of the number of hydrodynamic diameters respectively of IO@STMS-0 (A, B), IO@STMS-40 (C, D), IO@STMS-60 (E, F) and IO@STMS-120 (G, H).



0.2 mV and *ca.* -14.8 ± 0.8 mV for $t = 40, 60$ and 120 respectively) is due to the silica shell around the particles and was used, in practice, as a first indicator of the success of the silica synthesis before any TEM analysis. Given the colloidal stability, and more precisely the disappearance of the slight aggregates when the growth time increases, we were expecting an increase in absolute value of the zeta potential with the increase of growth time, as a higher absolute value would mean a higher electrostatic repulsion and thus a higher stability. However, the trend was the opposite, and this may be explained by the nature of the negative surface charge in silica NPs. Indeed, this negative surface charge comes from the silanolate groups that are coming from easily deprotonated Si-OH groups at the NPs' surface. When the growth time increases, the surface area decreases and Si-OH may cross-link more into Si-O-Si bridge groups, both effects contributing to reduce the negative zeta value.

Furthermore, we found that IO@STMS with a small STMS shell thickness have a similar/comparable hydrodynamic size to IO@STMS with a bigger shell (in the range 163–183 nm). We figure out that clusters composed of a couple to several NPs are formed for IO@STMS-40 (TEM core shell size of *ca.* 87 ± 18 nm) while the IO@STMS-120 may be dispersed individually. Factors such as the increase of hydrophilicity of the silica coating combined with the absence/reduction of magnetic dipolar interactions between iron oxide cores may explain this enhanced dispersion state for bigger shells. Conversely, an incomplete growth of silica that may favour H-bond interactions between the surface silanol groups of small STMS shells combined with potential magnetic dipolar interactions would explain their stabilisation into small clusters of several IO@STMS NPs. For instance, the difference of dipolar magnetic interactions existing between citrate-stabilised iron oxide NPs and silica-coated iron oxide was well evidenced by Kesse, Vichery and coworkers using ZFC/FC curves.⁵⁷

Overall, the DLS analyses show that the increase of STMS shell thickness improves the global colloidal stability of the IO@STMS-*t* NPs.

III.4. Influence of the STMS shell on the MRI, MHT and PHT properties of IO NPs

We showed in the previous sections that we were able to synthesise IO@STMS NPs with tunable silica shell thicknesses and pore structures. The next step of our study was to investigate the influence of the STMS shell thickness on the MRI, MHT and PHT properties of the IO NP core. From this section, it was particularly important for us to compare IO@STMS-*t* NPs with the same IO NP core, thus coming from the same series of syntheses. Series 1 presented in Fig. 6 allowed only one measurement for the following experiments. We thus synthesised a second series of IO@STMS-*t* NPs (denoted as series 2), whose characteristics are given in Fig. S10,† for which we could perform the measurements in triplicate and the *in vitro* studies with cancer cells presented in the last section of this article.

III.4.1. Magnetic resonance imaging (MRI). MRI is based on the nuclear magnetic resonance of the hydrogen atoms of

water molecules (or other tissues) under a static magnetic field (typically 1.5 T) and after stimulation with a radiofrequency field. Protons relax with time through two modes of relaxation and contrast agents such as magnetic core-shell NPs influence strongly these relaxation times with their concentration. These relaxation times are for longitudinal relaxation denoted T_1 and for transverse relaxation denoted T_2 , and their measurement gives access to the respective longitudinal and transverse relaxivities r_1 and r_2 that characterise contrast agents. Thus, we measured the longitudinal and transverse relaxation rates ($R_1 = 1/T_1$ and $R_2 = 1/T_2$ respectively) at different iron concentrations (Fig. 9A and B) for the different IO@STMS-*t* NPs. As can be seen in Fig. 9C, the longitudinal relaxivity r_1 is not impacted by the presence of a thin STMS shell, as it decreased only from *ca.* 8.7 ± 0.2 mM⁻¹ s⁻¹ for IO@STMS-0 to *ca.* 8.1 ± 1.3 mM⁻¹ s⁻¹ for IO@STMS-40 (STMS shell thickness of *ca.* 20.8 nm). However, this value decreased importantly when increasing the STMS shell thickness as it dropped to *ca.* 2.6 ± 0.1 mM⁻¹ s⁻¹ and *ca.* 1.2 ± 0.1 mM⁻¹ s⁻¹ for IO@STMS-60 and IO@STMS-120 respectively, which corresponds to a STMS shell thickness of *ca.* 30.8 nm and *ca.* 42.3 nm respectively. This tendency was also followed by the IO@STMS-*t* NPs obtained in the series 1, as can be seen in Fig. S11C and Table S1.† This phenomenon can be explained by the fact that the longitudinal relaxation is based on an inner coordination sphere mechanism where the proton relaxation occurs through direct contact probably through iron-OH₂ (metal-ligand bond) with the contrast agent. So, in our case, it is highly dependent on the access of water to the IO core NP. Thus, the increase of the STMS shell thickness, together with the linked decrease of the pore volume associated with silica shell densification, decreases this access to water and then the r_1 value.

Regarding the transverse relaxivity r_2 (Fig. 9D), the value is slightly increased in the presence of the STMS shell, with a very slight increase of the r_2 value when the shell thickness increases. Indeed, the r_2 went from *ca.* 356 ± 17 mM⁻¹ s⁻¹ for IO@STMS-0 to *ca.* 345 ± 9 mM⁻¹ s⁻¹, *ca.* 380 ± 5 mM⁻¹ s⁻¹ and *ca.* 409 ± 6 mM⁻¹ s⁻¹ for IO@STMS-40, IO@STMS-60 and IO@STMS-120 respectively. The IO@STMS-*t* NPs obtained from series 1 gave the same tendency, as shown in Fig. S11D and Table S1.† The first observation that can be drawn here is that the silica shell growth does not influence or hinder importantly these transverse relaxation modes of water protons. This is attributed to the long-range effect of this relaxation mode through dipolar interactions due to the high magnetic moment of the IO NPs. We can notice the slight r_2 increases from 0 to *t* min growth time which may be due to the colloidal stabilization brought by the STMS shell to the IO NPs and the increase of the hydrodynamic diameter, slowing down their Brownian motion and increasing the contact time between the water molecules and the IO NPs during the spin echoes. Importantly, the r_2 values are very high compared with commercial T_2 contrast agents (Combidex $r_2 = 65$ mM⁻¹ s⁻¹, Ferumoxytol $r_2 = 89$ mM⁻¹ s⁻¹, Resovist $r_2 = 189$ mM⁻¹ s⁻¹)⁵⁸ showing that these IO@STMS-*t* NPs are very good T_2 contrast agents.^{46,59,60}



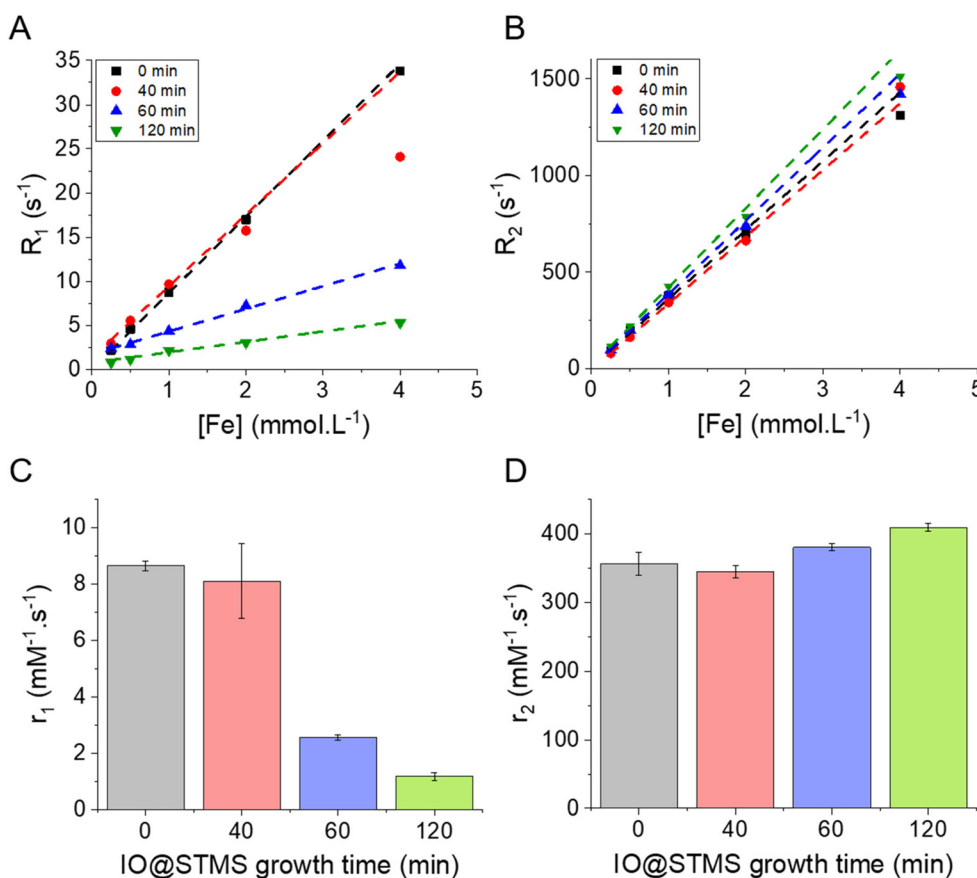


Fig. 9 (A) and (B) respectively the longitudinal $R_1 = 1/T_1$ (s⁻¹) and transverse $R_2 = 1/T_2$ (s⁻¹) relaxation rates as a function of the concentration of iron in the different IO@STMS-*t* NPs solutions. (C) and (D) respectively the evolution of the longitudinal and transverse relaxivities r_1 and r_2 as a function of the IO@STMS growth time. These results were obtained with the IO@STMS-*t* obtained from series 2.

III.4.2. Magnetic hyperthermia (MHT). As long as the frequency and amplitude respect the so-called Brezovich criterion,⁵⁰ the application of AMF is non-invasive, non-toxic and does not have a tissue penetration depth limit. More importantly, magnetic hyperthermia (MHT) allows the local increase of body temperature from 37 °C to 40–43 °C, which is enough to kill cancer cells as they are more sensitive to temperature than healthy cells.³ This ability to induce MHT is already used in preclinical and clinical trials,^{7–9} with a notable trial conducted in Berlin (Germany) by MagForce with NanoTherm™ technology where cancer cell apoptosis was shown to be induced by MHT. The use of IO NPs as bimodal agents for MRI and MHT has also been studied and reported.^{44,46,61}

The release of heat from IO NPs under AMF stimulation is usually described as arising from two phenomena: Néel relaxation, which corresponds to the reorientation of the magnetic moment inside the particle, and Brown relaxation, which corresponds to the reorientation of the whole particle in the medium. The heating process depends on several parameters such as the magnetocrystalline anisotropy, the NP volume and the environment viscosity. Usually, the heating capacity of particles is expressed with the specific absorption rate (SAR), which is most often determined by a calorimetric experiment

through eqn (3), given in Materials and methods (II.6.10.). Another way to determine the SAR consists of measuring the dynamic magnetization $M(t)$, which exhibits a phase shift compared with the instantaneous AMF vector $H(t)$, creating AC hysteresis loops, even for superparamagnetic IO NPs. In this AC magnetization curve $M_t(H_t)$, the surface area is equal to the heat dissipated during one period of the AMF $T = 2\pi/f$, hence the SAR:⁵⁰

$$\text{SAR (W g}^{-1}\text{)} = \frac{\mu_0}{m_{\text{IO}}} \times f \times \oint_{\text{cycle}} M_t(H_t) dH_t \quad (2)$$

where μ_0 is the magnetic permeability of a vacuum and m_{IO} is the mass of iron oxide in g (obtained as $m_{\text{IO}} = 1.38 \times m_{\text{Fe}}$).

The magnetothermal properties of the particles obtained from series 2 were evaluated by AC magnetometry using different conditions of frequency/magnetic fields f/H : 280 kHz/20 kA m⁻¹, 280 kHz/24 kA m⁻¹ and 217 kHz/24 kA m⁻¹ for IO@STMS-0 (3.35 mgFe mL⁻¹) and for IO@STMS-*t*, with *t* = 40, 60, 120 min (0.5 mgFe mL⁻¹). Representative hysteresis cycles are presented in Fig. 10A–C, where it can be noted that the ones obtained for IO@STMS-0 present a “square shape” rather



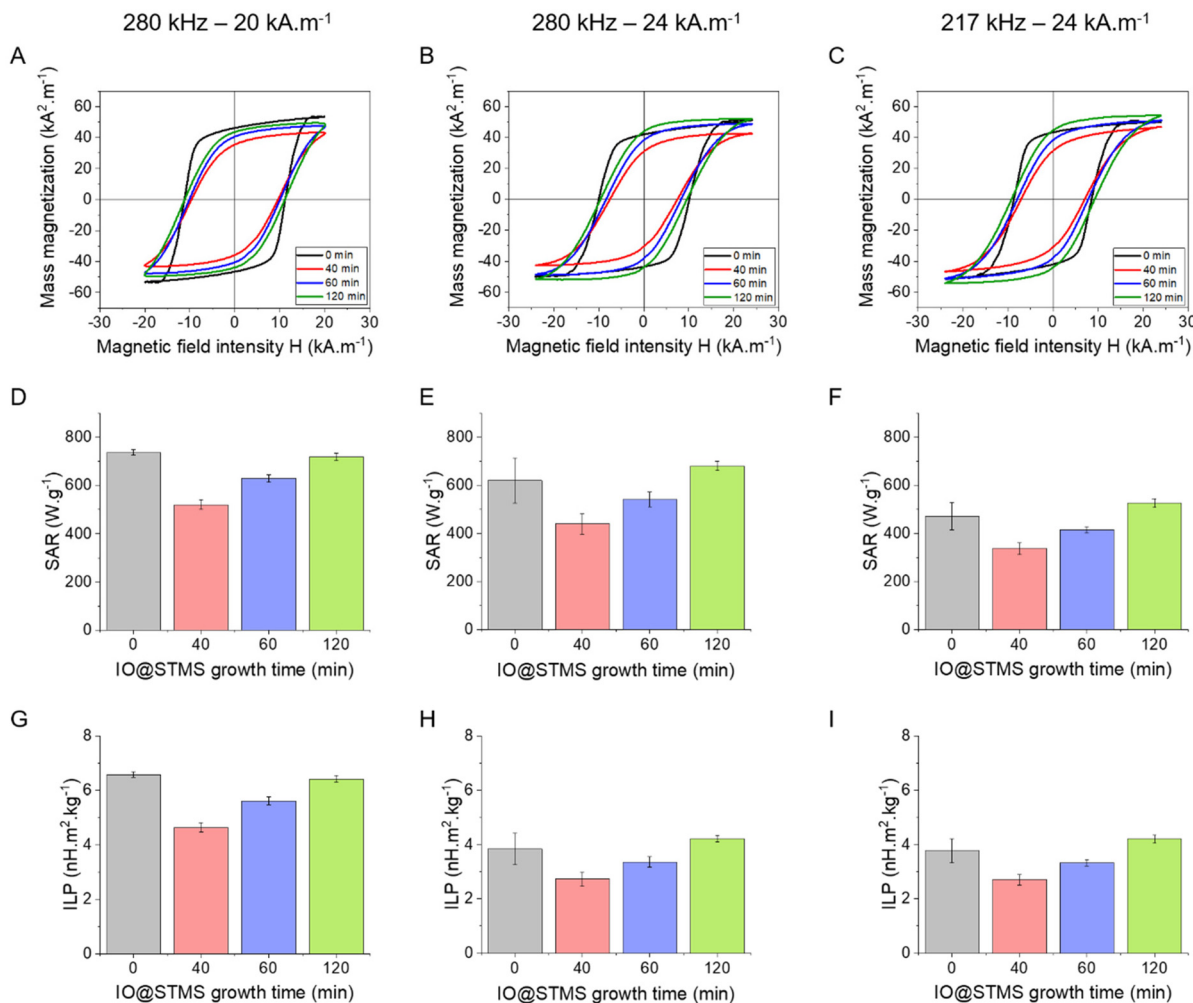


Fig. 10 (A)–(C) Representative hysteresis cycles for the different IO@STMS-*t* NPs (IO@STMS-0 and for IO@STMS-*t* (*t* = 40, 60, 120 min)) obtained from series 2 at different frequency–amplitude conditions. (D)–(F) Corresponding SAR values. (G)–(I). Corresponding ILP values.

than opened sigmoids. In previous work by Mille *et al.*,⁶² this phenomenon of a large opening of the hysteresis loops was ascribed to the organisation of the particles as chains of NPs during the magnetic field application. The chaining effect was also investigated by Martinez-Boubeta, Serantes and coworkers.^{63,64} The authors showed experimentally and using modelling that the formation of chain-like structures was explained by more favourable dipolar interaction energy as compared with thermal energy. Computation of hysteresis cycle evolution was achieved as a function of the number of particles within a chain and indicated that hysteresis cycles had more squared shapes and that the area of hysteresis loop increased (and thus SAR values) with the length of the chain.

In our case, macroscopic chains were indeed observed after the AC magnetometry measurement, and could be redispersed by sonication after the measurement. Such chains were also observed on some regions when performing *in situ* LPTM imaging (Fig. S12†). This chaining is likely explained by the large diameter of the IO NP cores ($d_{\text{core}} = 26.6$ nm), which

given the measured saturation magnetization of these Fe_{3-x}O₄ NPs achieved in Fig. S13† (56.5 emu g⁻¹ hence $M_s = 2.83 \times 10^5$ A m⁻¹ volume magnetization) leads to a dipolar parameter $\gamma = \frac{\mu_0 M_s^2 \pi^2 d_{\text{core}}^3}{36 k_B T} \sim 125$, which explains why the bare IO NPs (without silica) make chains. When the silica layer grows, the dipolar parameter decreases to much lower values $\gamma = \frac{\mu_0 M_s^2 \pi^2 d_{\text{core}}^6}{36 k_B T \cdot (d_{\text{core}} + 2\delta_{\text{shell}})^3} \sim 5.0$ for IO@STMS-40 ($\delta_{\text{shell}} = 30.2$ nm), ~ 2.1 for IO@STMS-60 ($\delta_{\text{shell}} = 47.2$ nm) and ~ 1.4 for IO@STMS-120 ($\delta_{\text{shell}} = 60.2$ nm).

Regarding the evolution of the saturation magnetization M_s (referred to iron oxide mass) as a function of the silica shell growth presented in Fig. S13,† it is worthy of note that M_s tends to decrease when the silica shell is reduced. The M_s of these samples was measured several months after their synthesis and we hypothesize that when a large silica shell is formed, the oxidized magnetite (Fe_{3-x}O₄) is preserved from



oxidation with time, while by decreasing the silica shell, oxidation in maghemite ($\gamma\text{-Fe}_2\text{O}_3$) is favoured, explaining this lowering of M_s .

Looking at the SAR values presented in Fig. 10D (280 kHz and 20 kA m^{-1}) the presence of the STMS shell on the IO NP core tends first to decrease the SAR, as it dropped from $ca. 736 \pm 11\text{ W g}^{-1}$ for IO@STMS-0 to $ca. 520 \pm 20\text{ W g}^{-1}$ for IO@STMS-40. The SAR values then increased with the increase of STMS growth time as it was measured at a value of $ca. 629 \pm 16\text{ W g}^{-1}$ and $ca. 718 \pm 15\text{ W g}^{-1}$ for IO@STMS-60 and IO@STMS-120 respectively under the same magnetic field. The same tendency is visible for the two other magnetic fields (Fig. 10E and F) and the global tendency could also be found for the particles obtained in series 1, whose results are presented in Fig. S14 and Table S2.† The global reduction of SAR with the addition of the silica shell is in agreement with previous studies performed in our team⁶⁵ and could correspond to the disappearance of the observed magnetic chains of iron oxide@CTA⁺ (IO@STMS-0) when a silica coating is achieved. With a small silica shell, the core-shell reorganised in small clusters which might have a limited Brown relaxation. Then, the increase of the SAR with the increase of the silica shell could come from the better colloidal stability brought by this higher shell thickness, which restores the Brown relaxation mechanism to the heat dissipation.

Unfortunately, the SAR values are difficult to compare from one article to another as they are very dependent on extrinsic parameters such as the frequency f and the amplitude H of the applied magnetic field.⁶⁶ Thus, we calculated the intrinsic loss power (ILP) following eqn (4) given in Materials and methods (II.6.10.), which assumes a linear dependency of SAR with f and quadratic dependency with H , as predicted by the linear response theory for superparamagnetic NPs.⁶⁷

Such calculations gave us an ILP ranging from $ca. 4.64 \pm 0.18$ to $ca. 6.58 \pm 0.10\text{ nH m}^2\text{ kg}^{-1}$ for series 2 at 280 kHz and 20 kA m^{-1} (Fig. 10G) and then from $ca. 2.7 \pm 0.27$ to $4.21 \pm 0.12\text{ nH m}^2\text{ kg}^{-1}$ when the amplitude is increased to 24 kA m^{-1} (Fig. 10H and I). The moderate decrease of the SAR and ILP values at 24 kA m^{-1} can come from the fact that the measurements were performed on the same samples and some days after the ones performed at 20 kA m^{-1} . We suppose here that the chaining effect happened in the first place and was beneficial for the SAR and thus the ILP, while the aging of the samples led to aggregates that could not be completely broken with sonication and thus reduced the ILPs. What can be noted is that these reduced values correlate well with the one obtained for series 1 (from $ca. 2.11\text{ nH m}^2\text{ kg}^{-1}$ (IO@STMS-40) to $ca. 4.08\text{ nH m}^2\text{ kg}^{-1}$ (IO@STMS-0) (Fig. S14C and Table S2†)). In addition, such values are quite good compared with the ILP of $1.18\text{ nH m}^2\text{ kg}^{-1}$ measured for NanoThermTM⁶⁸ and compared with the ILPs reported in the literature,¹⁰ which is very encouraging for anti-cancer applications using MHT therapies.

III.4.3. NIR-light photoinduced hyperthermia (PHT). Even if NIR light irradiation suffers from a tissue penetration of only 3–4 cm,^{69,70} this technique can be simply implemented and at

low cost. The potential of IO NPs as heating agents under such a stimulus has recently emerged with a first publication using them for photoinduced hyperthermia (PHT) from Yu *et al.* in 2011.¹⁴ The dual use of MHT and PHT is also studied. For example, Espinosa *et al.* reported that the hyperthermia induced by the administration of both stimuli to cubic IO NPs led to complete tumour regression in an *in vivo* model, explaining that the use of PHT restores MHT efficiency after cell internalisation, while the efficiency of MHT alone is reduced compared with expectations due to particle confinement.⁷¹

Unlike the case of gold and silver NPs, for which it is known that PHT is due to plasmonic resonance under laser irradiation, the mechanism in the case of IO NPs and the parameters influencing the photothermal response are not clearly understood yet. For example, Sadat *et al.* suggested that the heat generation is due to electronic transitions inside the IO lattice from the valence band to the conduction band.⁷² Furthermore, even if the mechanism and the parameters are still under debate, it seems that main parameters influencing PHT performances are not the size or shape but more precisely the quality of the crystallographic structure and the presence of defects (vacancies, dislocations, *etc.*) in the crystal structure. Indeed, in a previous work by Bertuit, Abou-Hassan and co-workers addressing the design of iron oxide nanoflowers having varying levels of defects, nanoflowers having excessive defects in terms of oxygen vacancies were found to decrease the photothermal effects. This was explained by the electron trapping inside the structure decreasing the electron-heat conversion.⁷³

Given all this information, it seemed interesting to us to test the potential of our NPs for PHT therapies. The PHT properties of our IO@STMS- t NPs were thus evaluated using NIR light irradiation at 1064 nm. The NIR laser was applied to the different core-shell NP suspensions at a fixed iron concentration (0.5 mgFe mL^{-1}) and the temperature profiles were plotted and converted into SAR values using the mass of iron oxide as a reference (Fig. 11). Regarding the temperature profile, the final ΔT was found to be quite similar for the IO@STMS- t NPs from series 2, with a slight increase of values between $t = 0$ ($ca. 23.1 \pm 0.4\text{ }^\circ\text{C}$) and $t = 40, 60$ or 120 ($ca. 25.2 \pm 0.2\text{ }^\circ\text{C}$, $ca. 23.9 \pm 0.8\text{ }^\circ\text{C}$ and $ca. 23.8 \pm 0.1\text{ }^\circ\text{C}$ respectively) while a slight decrease of the values can be seen for the IO@STMS- t NPs from series 1 ($ca. 21.9\text{ }^\circ\text{C}$ versus $ca. 19.1\text{ }^\circ\text{C}$, $ca. 19.8\text{ }^\circ\text{C}$ and $ca. 19.4\text{ }^\circ\text{C}$, Fig. S15†). Taking these results in their totality, they mean that the thermal transfer is overall not affected by the large pore silica shell whatever the thickness.

The final SAR values were determined to be $ca. 831 \pm 66\text{ W g}^{-1}$ for IO@STMS-0 and $ca. 1191 \pm 63\text{ W g}^{-1}$, $ca. 1023 \pm 93\text{ W g}^{-1}$ and $ca. 1123 \pm 71\text{ W g}^{-1}$ for IO@STMS-40, IO@STMS-60 and IO@STMS-120 respectively. Regarding the values obtained from series 1 (Fig. S15 and Table S3†), the SAR value decreased slightly with the addition of the STMS shell and then with the growing of the STMS shell thickness (from $ca. 909\text{ W g}^{-1}$ to $ca. 712\text{ W g}^{-1}$). Taking again these results in their totality, it is difficult to really give a tendency for the impact of the STMS



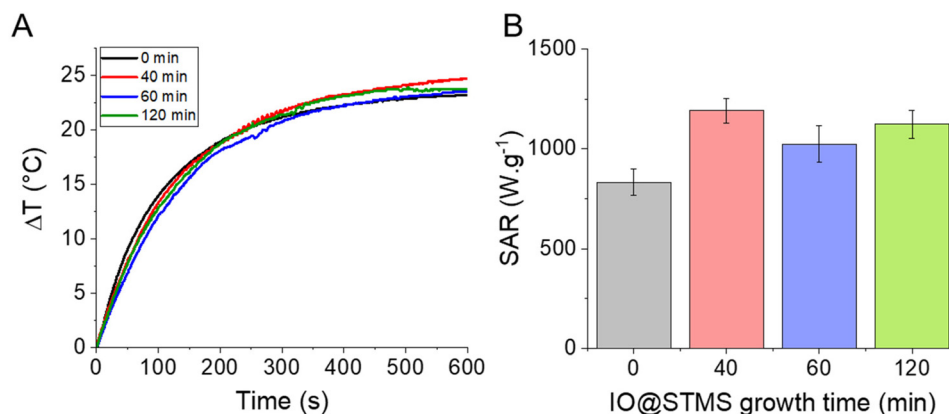


Fig. 11 (A) Temperature profiles as a function of time for the different IO@STMS-*t* NPs at 0.5 mgFe mL⁻¹ under NIR light irradiation. (B) Corresponding SAR values. These results were obtained with the IO@STMS-*t* obtained from series 2.

shell on the PHT properties of the IO NP core as the values are quite similar whatever the particle series. Unfortunately, a standardisation of the SAR like the calculation of the ILP for magnetothermal measurements is still unavailable for photo-thermal measurements, making the comparison of our values with the literature difficult. However, the high SAR values and the absence of a clear impact of the silica shell on these SAR values shows the promising aspect of the use of such NPs for PHT therapies.

III.5. Biological features: evaluation of NPs' applicability to anti-cancer therapies

In the next sections, our goal was to evaluate the potential of our nanoparticles for anti-cancer therapies using magnetic fields as external stimuli. Thus, the pancreatic cancer cell line MIA Paca-2 was chosen, as a model, to perform the *in vitro* experiments. In addition, as the STMS shell was added to the IO NP core to counteract their rapid blood capillary agglomeration and elimination, IO NPs without an STMS shell were not used for this study.

III.5.1. Cytotoxicity. The cytotoxicity of the IO@STMS-*t* NPs was first evaluated. To do so, MIA Paca-2 cells were incubated for 24, 48 or 72 h with an increasing concentration of nano-

particles, based on the concentration of iron. This choice was made as we still wanted to assess the potential impact of the STMS shell on the properties of the IO NP core, and not only to determine if our NPs were good candidates for anti-cancer therapies. The results presented in Fig. 12 clearly show the higher cytotoxicity of IO@STMS-120 toward cells from early time points, compared with IO@STMS-40 and IO@STMS-60. The cytotoxicity of these IO@STMS-*t* (*t* = 40 and 60) was still similar, with always more than 90% survival after 24 h of incubation up to 100 µgFe mL⁻¹. However, IO@STMS-60 presented more cytotoxicity than IO@STMS-40 at 24 h and 48 h of incubation. Altogether, these results show that a concentration up to 0.78 µgFe mL⁻¹ of iron can be used for all IO@STMS-*t* NPs up to 72 h of incubation, as the final cell viability was *ca.* 90.37 ± 4.80% and *ca.* 90.50 ± 2.17% for IO@STMS-40 and IO@STMS-60 respectively, which is still very good, and *ca.* 82.15 ± 2.99% for IO@STMS-120, which is still acceptable. Thus, the non-cytotoxic concentration of 0.5 µgFe mL⁻¹ was chosen for the following experiments for all nanoparticles. In addition, in order to potentially increase the efficiency of anti-cancer therapy using these nanoparticles and as the cytotoxicity of IO@STMS-40 and IO@STMS-60 is lower than IO@STMS-120 and is still acceptable at 5 µgFe mL⁻¹, we

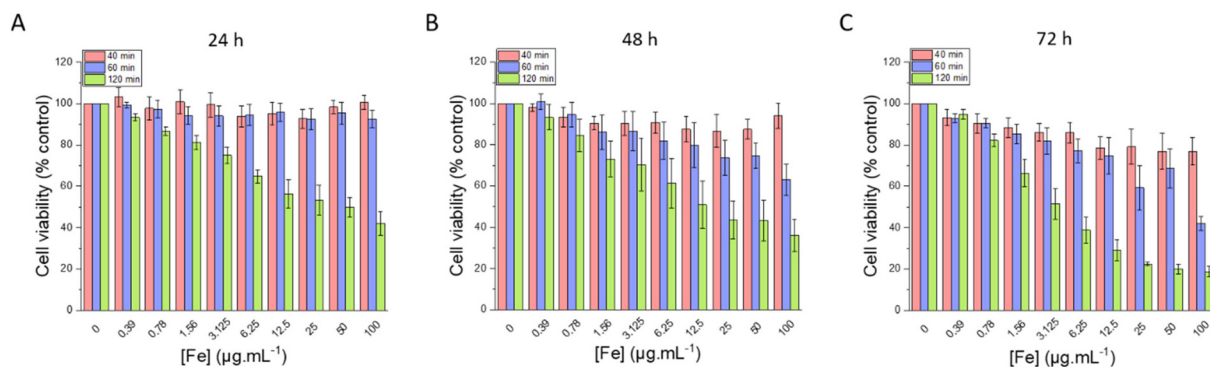


Fig. 12 Cell viability evaluated after (A) 24 h, (B) 48 h and (C) 72 h of incubation with different amounts of IO@STMS-*t* NPs, with *t* = 40, 60 or 120.



decided to test also this concentration with these two nanoparticle batches.

Regarding the chosen concentrations, to give some elements of comparison, in a previous work from our team,⁴⁶ we previously evidenced a hyperthermia induced-cytotoxic effect on HeLa cancer cells (*ca.* 65% metabolic activity reduction *vs.* control) under AMF using human serum albumin-coated IO@STMS-120 at a concentration of 25 $\mu\text{gIO@STMS mL}^{-1}$ that corresponds to 1.25 $\mu\text{gFe mL}^{-1}$. Field conditions providing cancer cell viability reduction were at 100 kHz and 357 Gauss (28.4 kA m^{-1}) whose product is below the safety limit of $H \times f = 5 \times 10^9 \text{ A m}^{-1} \text{ s}^{-1}$, commonly accepted for localized hyperthermia.⁷⁴ Similarly, a concentration of 16 $\mu\text{gFe mL}^{-1}$ of IO NPs coated with PEG was shown to induce the death of different cancer cell lines (pancreatic, gastric cancer) by intra-lysosomal magnetic hyperthermia (AMF: 40 mT, 275 kHz).⁷⁵

III.5.2. Cellular uptake. We then investigated the cellular uptake of the IO@STMS-*t* NPs. The cells were incubated for 72 h with IO@STMS-*t* NPs at a final concentration of 0.5 $\mu\text{gFe mL}^{-1}$ or 5 $\mu\text{gFe mL}^{-1}$. As shown in Fig. 13A, the cellular uptake was $\sim 0.04 \text{ pgFe per cell}$ and increased to $\sim 0.4 \text{ pgFe per cell}$ when the cells were incubated respectively with 0.5 $\mu\text{gFe mL}^{-1}$ or 5 $\mu\text{gFe mL}^{-1}$ of IO@STMS-*t* NPs, indicating that the uptake proportionally increased, by 10-fold, with the concentration of IO@STMS-*t* NPs. These cellular uptakes corresponded respectively to *ca.* $13 \pm 5\%$, $18 \pm 3\%$ and $21 \pm 3\%$ of the initial iron mass incubated of IO@STMS-40, IO@STMS-60 and IO@STMS-120 at 0.5 $\mu\text{gFe mL}^{-1}$ with the cells, suggesting a slight increase of cellular uptake with the increase of STMS shell thickness (Fig. 13B). However, our statistical test showed no significant difference between these values. In addition, similar percentages of the initial iron mass of IO@STMS-40 and IO@STMS-60, respectively *ca.* $14 \pm 4\%$ and *ca.* $14 \pm 2\%$, were taken up by cells incubated with 5 $\mu\text{gFe mL}^{-1}$. Thus, we

cannot conclude that the STMS shell has an impact on the cellular uptake of the NPs, as the values are quite close.

TEM was also performed on the samples incubated at 5 $\mu\text{gFe mL}^{-1}$ for 72 h in order to investigate the particles' localization inside the cells. As can be seen in Fig. 14, the particles are localized in the lysosomes, meaning that we would have endo-lysosomal hyperthermia under AMF.

Another interesting result obtained from this TEM intracellular imaging is that the silica shell appears to be importantly intracellularly degraded as only clustered IO NPs can be seen on the images presented in Fig. 14. However, it is important to note that silica shells could still be observed in very few imaged areas of IO@STMS-60 and IO@STMS-120. Indeed, as can be seen in Fig. S16A,† some IO NPs are not all clustered in the case of IO@STMS-60 and present a kind of corona that might be the silica shell degrading, being more transparent to the electron beam, leading thus to this poor contrast. However, the silica shell is clearly observable in the case of IO@STMS-120 as shown in Fig. S16B.†

The intracellular degradation of mesoporous silica NPs and also of iron oxide clusters@mesoporous silica NPs was previously reported to occur from some days to several weeks depending on the different parameters influencing the silica dissolution: NP concentration (silica limit solubility is reported in the range 140–160 $\mu\text{g mL}^{-1}$ under standard physiological conditions), NP aggregation state, pore size, degree of silica condensation, *etc.*^{37,76}

In our case, TEM intracellular imaging suggests that the degradation of the silica shell in the lysosomes is occurring rapidly (in three days) and is heterogeneous, which can come from the fact that the particle concentration is quite low, providing suitable low intra-lysosomal confinement and thus favouring silica shell dissolution.

III.5.3. Cytotoxicity under magnetic stimuli. The applicability to anti-cancer therapies using external stimuli was then

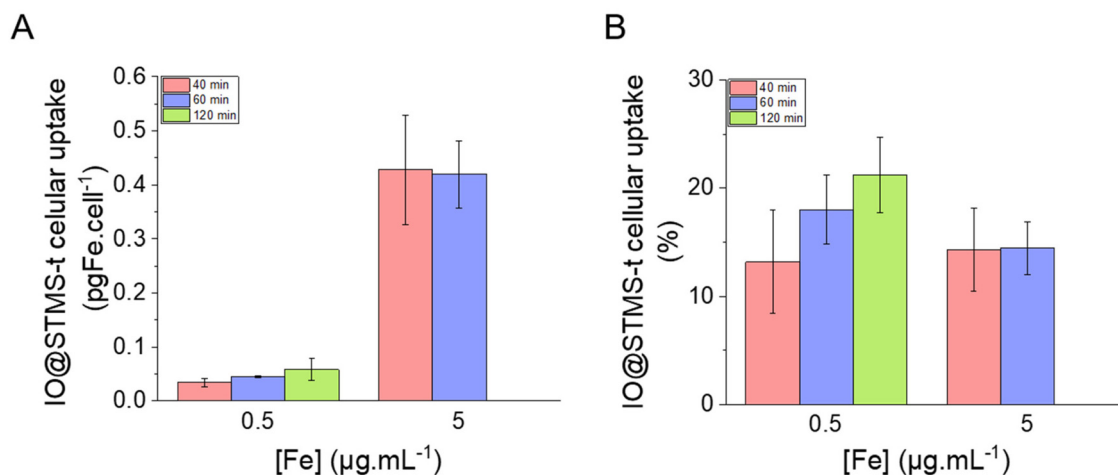
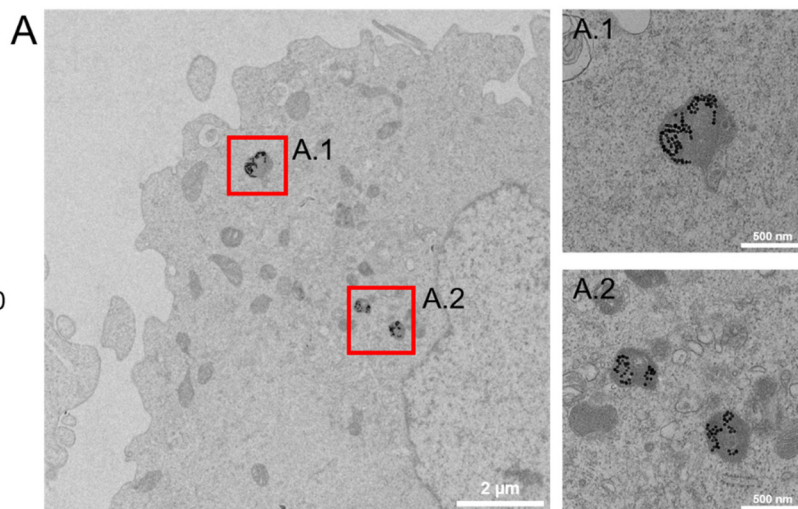


Fig. 13 IO@STMS-*t* NP cellular uptake in (A) pgFe per cell and in (B) percentage of incubated iron. Statistical analysis gave no significant difference between the groups.

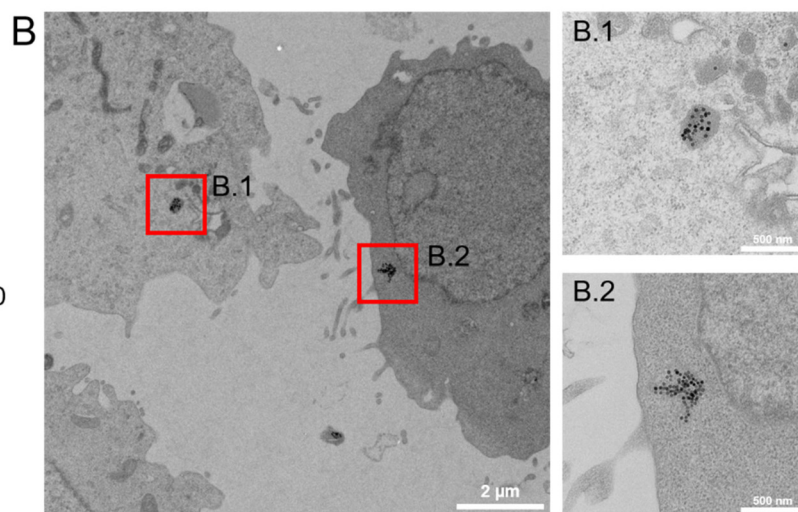




IO@STMS-40




IO@STMS-60




IO@STMS-120

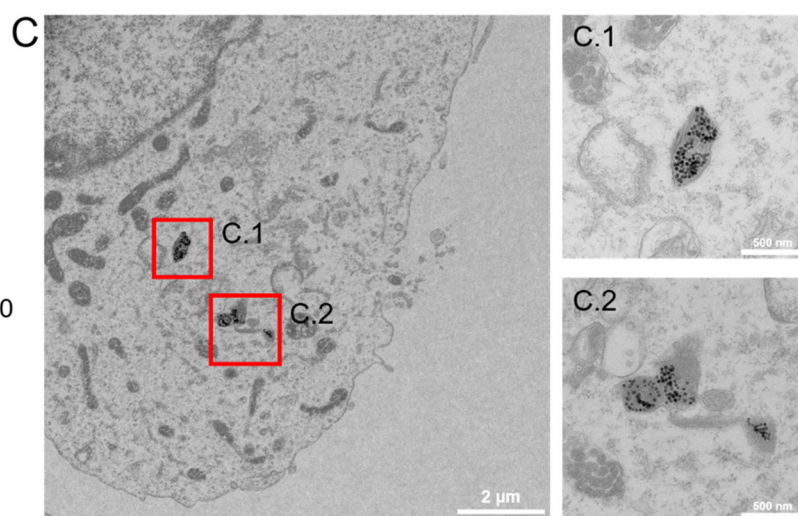


Fig. 14 TEM images of cells incubated at $5 \mu\text{gFe mL}^{-1}$ for 72 h with (A) IO@STMS-40, (B) IO@STMS-60 and (C) IO@STMS-120 and respective magnifications of lysosomes in A.1, A.2; B.1, B.2 and C.1, C.2.



studied. The cells were incubated or not with the IO@STMS-*t* NPs to allow their internalization, and then exposed or not to an AMF with a frequency f of 250 kHz and an amplitude $\mu_0 \cdot H$

of 20 mT (16 kA m^{-1}) for 2 h. The cells that were not incubated with NPs and not subjected to AMF served as a control to evaluate the efficiency of the treatment. Cell death was then

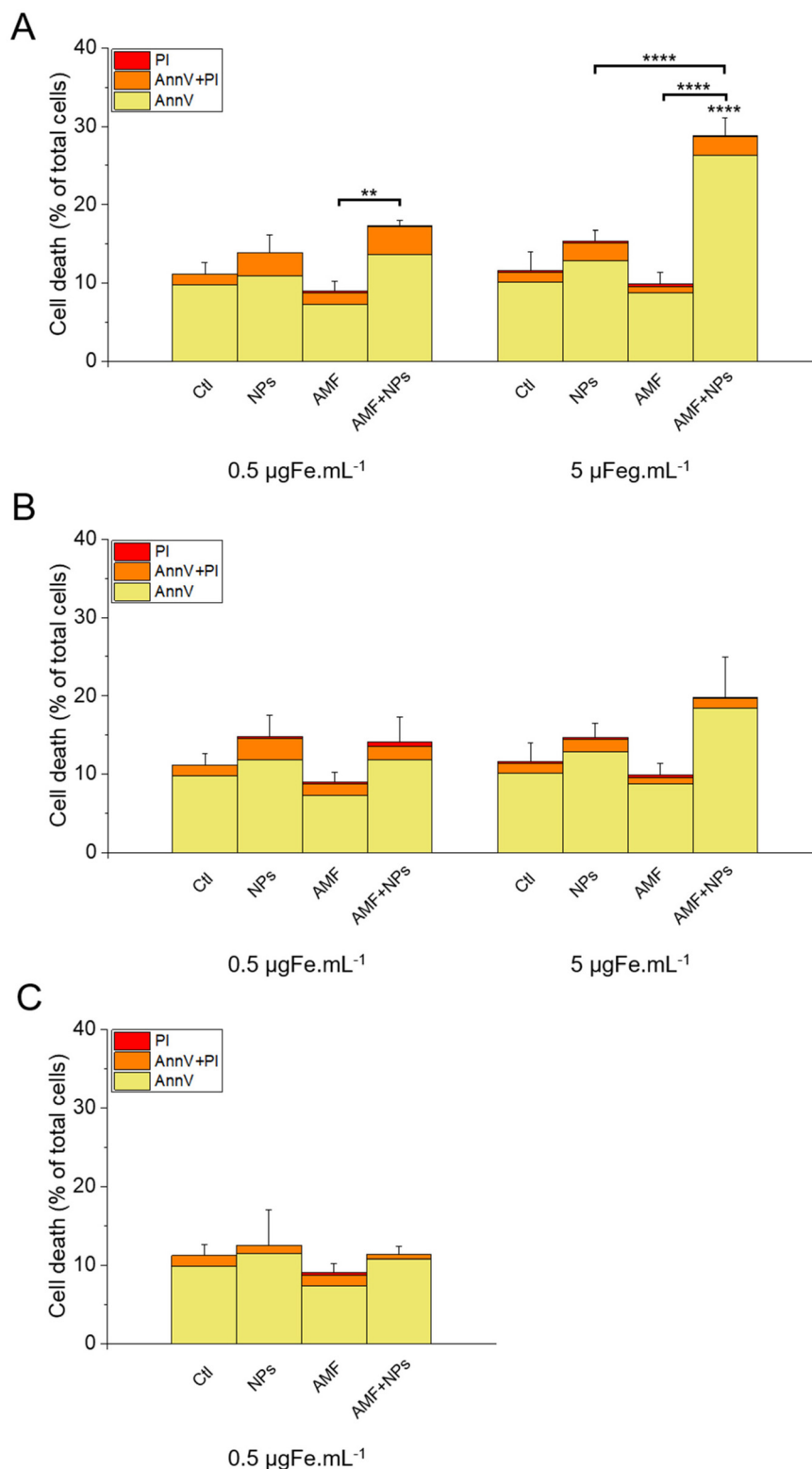


Fig. 15 Cell death evaluated in the presence or absence of IO@STMS-*t* NPs with or without an AMF stimulus for (A) IO@STMS-40, (B) IO@STMS-60 and (C) IO@STMS-120. Error bars and statistical analysis are on the total percentage.



evaluated using annexin V (AnnV) and propidium iodide (PI) labelling. This labelling was chosen in order to give a first insight on the mechanism of the cell death. Indeed, AnnV is a marker of early apoptosis, as it binds to certain phospholipids that normally face the interior of the cell (in contact with the cytoplasm) but face the exterior of the cells (in contact with the environment) in the early stages of apoptosis. Regarding PI, this molecule is a marker of necrosis, as it binds to DNA and can enter the cells only if the membrane becomes more permeable. The case where both molecules label the same cell corresponds more to late apoptotic cell death. However, it has to be kept in mind that this technique gives insight on the cell death mechanism, but that more specific tests should be performed to discriminate, for example, primary necrotic cell death from late apoptotic cell death.⁷⁷ In the case of anti-cancer therapies, a cell death mechanism by apoptosis is highly researched as it corresponds to a programmed suicide of the cell in response to the treatment without the release of detrimental molecules for other cells, while cell death by necrosis is an accidental cell death leading to the release of such molecules.

The results of this labelling are presented in Fig. 15. As expected, the incubation of the cells with NPs only did not significantly increase cell death compared with the control, as well as the treatment with AMF only. Interestingly, AMF exposure increased the mortality of MIA Paca-2 cancer cells incubated with IO@STMS-40, especially at 5 $\mu\text{gFe mL}^{-1}$ (Fig. 15A). Indeed, cell death was increased only from *ca.* 11.2 $\pm$ 1.5%, *ca.* 13.6 $\pm$ 2.2% and *ca.* 9.1 $\pm$ 1.2% for the control, NPs

only and AMF only respectively to *ca.* 17.3 $\pm$ 0.6% for AMF+NPs when the cells were incubated at 0.5 $\mu\text{gFe mL}^{-1}$ while it went up to *ca.* 28.8 $\pm$ 2.3% when the cells were incubated at 5 $\mu\text{gFe mL}^{-1}$, which was determined to be significantly higher than the % of dead cells measured for the other treatments. Moreover, the dead cells are mainly labelled by AnnV, indicating an apoptosis-related cell death pathway induced by IO@STMS-40 upon MHT treatment. In contrast, IO@STMS-60 and IO@STMS-120 did not decrease the viability of the MIA Paca-2 cancer cells upon MHT treatment (Fig. 15B and C).

Regarding these magnetothermal effects under AMF, we can only speculate that the improved cancer cell killing with the IO@STMS-40 sample as compared with the other samples is due to local heating effects at the surface of the IO NPs through the silica large-pore shell. Even if all of the AMF treatments on IO@STMS-*t* are made in macroscopically athermic conditions, we can hypothesize that the local temperature is higher when the silica shell is thinner and more opened. For instance, several teams showed a huge reduction in cancer cell viability using targeted magnetic nanoparticles without the need for a perceptible temperature rise.^{75,78,79} Furthermore, the total absence of a silica shell observed by the above intracellular TEM imaging in IO@STMS-40 as compared with IO@STMS-60 may also play a role in local related effects at the iron oxide surface (magnetothermal effect, ROS production, etc.) and thus might contribute to improving cytotoxicity.

Thus, these results tend to show that even if IO@STMS-40 has lower SAR values, at a slightly lower or

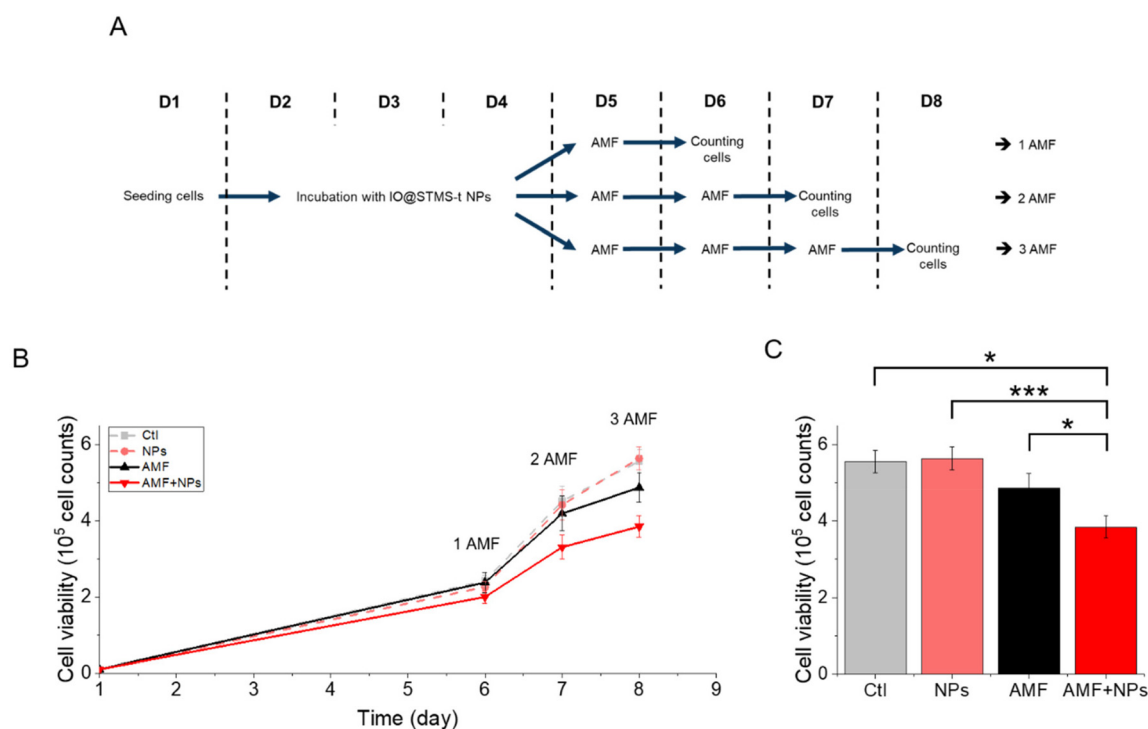


Fig. 16 (A) Schematic representation of the applied protocol for AMF. (B) Proliferation of MIA Paca-2 cancer cells along the treatment. Statistical analysis was done on the (C) proliferation at day 8.



similar amount of internalized iron, they are still the more promising nanoparticles for anti-cancer therapies using AMF.

IO@STMS-40 NPs were thus used for a complementary experiment analysing cell proliferation. The cells were incubated or not with these NPs at $5 \mu\text{gFe mL}^{-1}$ for 72 h and

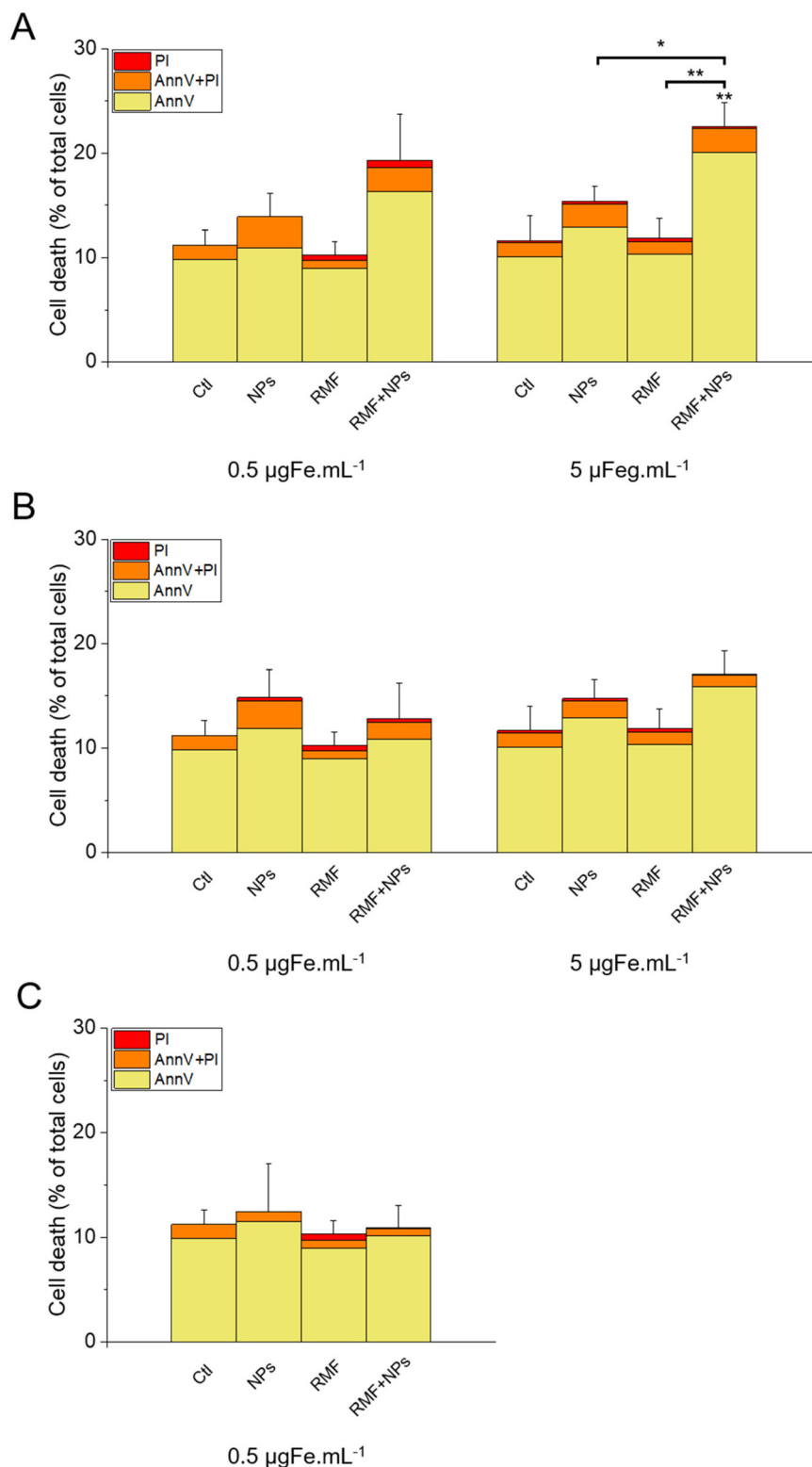


Fig. 17 Cell death evaluated in the presence or absence of IO@STMS-*t* NPs with or without an RMF stimulus for (A) IO@STMS-40, (B) IO@STMS-60 and (C) IO@STMS-120. Error bars and statistical analysis are on the total percentage.



exposed respectively one, two or three times to AMF (or not). A schematic representation of the protocol is given in Fig. 16A to better perceive it. The results, presented in Fig. 16B, show that cell proliferation is affected over time under exposure to AMF, and even more under exposure to AMF after being incubated with the NPs. Indeed, the number of cells dropped from $5.6 \times 10^5 \pm 3.0 \times 10^4$ without NPs or AMF, to $4.9 \times 10^5 \pm 3.9 \times 10^4$ after three exposures to AMF and to $3.8 \times 10^5 \pm 2.8 \times 10^4$ after incubation with NPs and three exposures to AMF, which has been determined to be significantly different, as shown in Fig. 16C. These results show again that IO@STMS-40 are promising NPs for anti-cancer therapies using AMF.

The application of an AMF to evaluate the potential of IO@STMS-*t* NPs for anti-cancer therapies makes sense as these particles are studied for magnetic hyperthermia treatment. However, it is also possible to use RMF in order to see if the NPs can induce cell death by mechanical forces and not only by the local increase of temperature. Similar experiments were then performed upon RMF application, with a frequency *f* of 1 Hz and an amplitude $\mu_0 * H$ of 40 mT (32 kA m^{-1}). As can be seen in Fig. 17, only $5 \mu\text{gFe mL}^{-1}$ of IO@STMS-40 increased significantly cell death: *ca.* $22.6 \pm 2.3\%$ comparatively to *ca.* $11.2 \pm 1.5\%$ of the control cells. Moreover, dead cells are also mainly labelled by AnnV, indicating an apoptosis-related cell death pathway induced by IO@STMS-40 motion upon RMF application. Regarding these RMF results, we could also hypothesize that NP chaining is more probable with a thinner silica shell and may induce more efficient torque

effects for IO@STMS-40 as compared with IO@STMS-60. However, these nanoparticles seemed less efficient under RMF compared with AMF: *ca.* $22.6 \pm 2.3\%$ versus *ca.* $28.8 \pm 2.3\%$ of cell death respectively after $5 \mu\text{gFe mL}^{-1}$ of IO@STMS-40 incubation.

Regarding the multiple application of RMF with or without IO@STMS-40 NPs, the results presented in Fig. 18 show a significant impact of the multiple exposure to RMF and to RMF with NPs. Indeed, the number of cells dropped from $5.6 \times 10^5 \pm 3.0 \times 10^4$ without NPs or RMF, to $4.8 \times 10^5 \pm 3.1 \times 10^4$ after three exposures to RMF and to $3.8 \times 10^5 \pm 2.3 \times 10^4$ after incubation with NPs and three exposures to RMF, showing one more time the great potential of IO@STMS-40 NPs for anti-cancer therapy under RMF.

Altogether, these results show that IO@STMS-40 are the best candidates between the IO@STMS-*t* NPs for anti-cancer therapies, whether using an AMF or RMF stimulus. The cell death may represent a low value ($\sim 23\text{--}29\%$ of total cells), which can be disappointing regarding the quite high SAR and ILP values determined in the previous section. However, multiple exposures to AMF or RMF significantly reduced the cell proliferation, showing its potential for such application.

In addition, we have to keep in mind that the IO@STMS-*t* NPs were used here without any polymer coating. Such a coating may enhance their internalization and decrease cytotoxicity which would allow an increase in the concentration of nanoparticles, and thus should increase their anti-cancer efficiency.

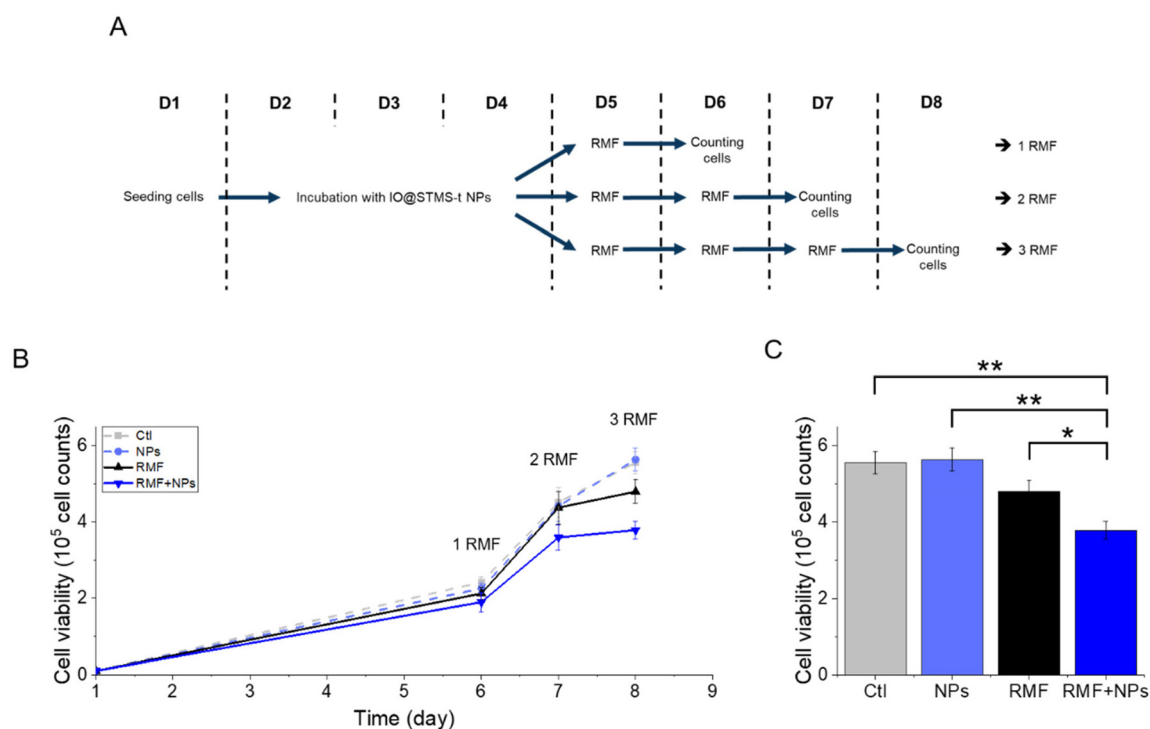


Fig. 18 (A) Schematic representation of the applied protocol for RMF. (B) Proliferation of MIA Paca-2 cancer cells along the treatment. Statistical analysis was done on the (C) proliferation at day 8 after three exposures to RMF or no exposure.



As a perspective, there are various strategies of polymer coatings around mesoporous silica that may be used to enhance cancer cell uptake. Among them, we can cite polycation surface modifications like the non-covalently adsorbed polyethyleneimine coatings ensuring a strong interaction with the negatively charged cell membrane, and yielding a high cellular uptake.⁸⁰ We can cite also the covalently bound polyarginine acting as a cancer cell-penetrating cationic polypeptide.⁸¹ Another surface modification strategy is the covalent conjugation of hyaluronic acid, a negatively charged polyelectrolyte, which ensures efficient cell internalization through targeting the CD44 receptor overexpressed in cancer cells.⁸²

IV. Conclusion

In this work, we have addressed the fine tuning of the STMS shell growth around an IO NP core by simply playing on the sol-gel time of the synthesis. The control of the STMS shell growth was evidenced as well by time-lapse TEM imaging and by *in situ* LPTM, which were found to be very complementary. The evolution of the pore structure was also deeply investigated by nitrogen adsorption-desorption measurements (BET, BJH and Horvath-Kawazoe methods) and we could show that increasing the shell thickness resulted in a decrease of the specific surface area, pore volume and pore size. This new synthesis approach based on a very simple procedure allows the preparation of a range of new IO@STMS core-shell NPs with tunable shell thickness and pore structure. Furthermore, all of these IO@STMS-*t* (with *t* being the growth time) were shown to have overall a good colloidal stability.

The impact of the STMS shell thickness on the MRI and hyperthermia properties of the IO NPs core was then studied. First, regarding MRI applications, we showed that STMS shell growth had no detrimental impact on the T_2 -weighted MRI relaxivities, making these IO@STMS-*t* NPs very good T_2 contrast agents with relaxivity values in the range of 320–377 mM⁻¹ s⁻¹. Regarding the evolution of the T_1 -weighted MRI relaxivities, which decreased strongly with silica shell thickness, these allow confirmation of the densification of the silica walls with the increase of growth time, limiting water accessibility. Regarding the hyperthermia properties of the IO@STMS-*t* NPs, we could see a slight decrease of the SAR value determined under an AMF stimulus and no clear impact on the SAR value determined under an NIR light stimulus, indicating that the stellate porous silica shell provides a low insulating property with silica shell growth. Overall, it does not affect the heat dissipation from the core to the surrounding media, making all these core-shell IO@STMS-*t* NPs efficient heating agents under AMF or NIR light irradiation stimulation.

Finally, several *in vitro* experiments using the pancreatic cancer cell line MIA Paca-2 were performed in order to evaluate the potential use of these IO@STMS-*t* NPs for anti-cancer applications. We were able to show that ~14% of the incubated particles were internalised by the cells and that the application of AMF or RMF improved the anti-cancer efficiency of IO@STMS-40.

This study showed the great interest from adding the STMS shell around the IO NP core as it notably stabilises the particles and does not hinder their interesting properties, namely MRI, MHT and PHT. In addition, the *in vitro* experiments showed the interest for using such core-shell NPs to improve anti-cancer treatment by using external magnetic fields. Such systems could then be envisioned for bimodal anti-cancer applications, with the combination of local hyperthermia with the thermally-induced controlled delivery of therapeutic molecules of a higher size than synthesised ones, such as protein, DNA or RNA. However, as such biomolecules are quite sensitive to thermal denaturation, evaluation of the local temperature at the surface of the IO@STMS NPs would also be future studies to investigate.

Data availability

Data for this article are available at Recherche.Data.Gouv repository at the <https://doi.org/10.57745/SVCWSI>.

Conflicts of interest

There are no conflicts of interest to declare.

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