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On the way to potential antifungal compounds: synthesis and *in vitro* activity of 2-benzofuranylacetic acid amides†

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Crop losses caused by microbial infections are a significant global issue, especially in tropical regions. The development of novel antimicrobial agents, particularly antifungal agents, has been explored from various perspectives, including chemical synthesis. However, conventional approaches typically involve synthesizing new and potent compounds on a small scale (a few milligrams), making the scale-up of the reaction a major challenge. In this manuscript, we present a method for the synthesis of new and active (against *Fusarium oxysporum*) benzofuranyl acetic acid amides. Our strategy allows us to synthesize the key precursor on the gram scale, enabling the production of sufficient quantities of other active compounds within short timeframes for conducting biological studies. All the reactions used in this manuscript are recognized by their industrial application.

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Introduction

Microorganisms' resistance to current treatments poses a significant challenge for humans, not only from a medical perspective but also in various other fields such as veterinary, economy, biochemistry, and agriculture.^{1,2} The issue of crop losses due to microbial pathogenesis is particularly severe in tropical countries, but it affects the global community as a whole.^{3,4}

Fusarium oxysporum (FOX) is a highly infectious pathogen that poses a significant threat to global agricultural production. It is responsible for causing vascular wilt⁵ and contaminating crops such as wheat, barley, corn, bananas, and cotton, resulting in substantial worldwide losses estimated at approximately 80%.^{6–8} This invasive plant pathogen also contributes to the presence of mycotoxins in food, which not only jeopardizes food safety⁹ but also endangers the health of both humans and animals, making them susceptible to fungal infections.

In this context, remarkable progress is being made in the development of new bioactive compounds against phytopathogens such as FOX on a weekly basis.^{10–18} However, many of

these advancements come with a significant drawback. Most active compounds are derived from natural sources, resulting in limited quantities that make it challenging to conduct thorough testing and field evaluations. While chemical synthesis has been regarded as a viable alternative for obtaining larger quantities of active compounds, recent developments often involve the use of expensive or complex materials, or techniques. Additionally, even if the active compounds are successfully synthesized, the amounts obtained are often insufficient for conducting field trials. Consequently, when a highly promising compound is discovered, the chemical industry faces new challenges in scaling up production, requiring the re-optimization of reaction conditions at each step. Therefore, it is crucial to provide potentially scalable protocols when synthesizing new compounds.

In light of these considerations, our research focused on the synthesis of benzofuranyl acetic acid amides using easily scalable processes. This investigation was inspired by the numerous active compounds that possess a benzofuran ring, analogue heterocycles and at least one amide bond,^{19–23} as illustrated in Fig. 1.

The synthesis of benzofuranyl acetic acid amides is quite uncommon. Only a few reports can be found, which are based on different methods such as palladium-catalyzed amino-carbonylative cyclization,²⁴ Pd-catalyzed alkyne cyclization,²⁵ gold-catalyzed domino cyclization,²⁶ and the coupling of benzofurane boronic acids with bromoketones.²⁷

Alternatively, a plausible approach is the synthesis of benzofuranyl acetic acids and their esters, which provides a few more options. These options include cyclizations induced by

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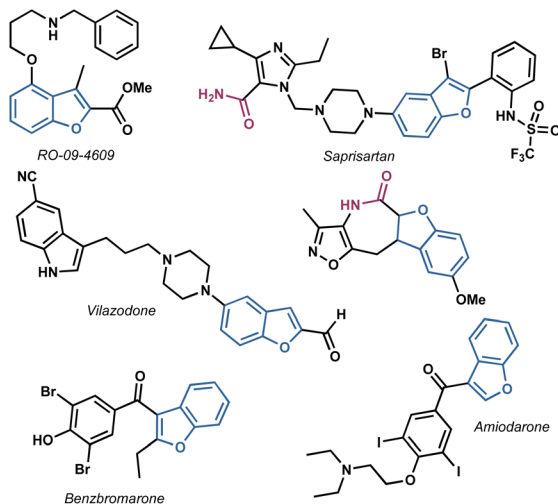


Fig. 1 Structures of some active benzofuran-based compounds.

radicals^{28,29} or transition metals,^{30–32} functionalization of benzofuran derivatives,^{33–36} or formal cycloadditions.^{37–39}

Regarding the antifungal potential of benzofuran derivatives, the available literature is limited. Certain authors have assessed the activity against *FOX* of those derivatives and observed inhibition halos ranging from 28 to 32 mm, demonstrating better activity compared to the positive control nystatin, which exhibited a 23 mm halo.⁴⁰ Moreover, Xie *et al.*⁴¹ have reported the *in vivo* and *in vitro* biological activity of some molecules containing a benzofuran ring against other phytopathogenic fungi such as *Colletotrichum iagenarium* and *Erysiphe graminis*. They obtained data comparable to the positive control *enoxastrobin*, showing inhibition up to 100%.

It is worth noting that while numerous active compounds containing the benzofuran ring have been described and show promising bioactivities, only a few reports suggest that the mentioned antifungal activity can indeed be attributed to the benzofuran ring.^{42–44}

Considering the privileged position of the amide bond in bioactive molecules and the scarcity of reports where a benzofuran served as the structural core of antifungal compounds, we identified benzofuranyl acetic acid amides as our target molecules (Fig. 2). Our hypothesis was that the amide bond would confer stability against hydrolysis (compared to esters), increase the molecules' polarity, thereby enhancing their solubility in water. Additionally, the introduction of polar functional groups in the amide aromatic ring would aid in this solubility enhancement. However, the ultimate activity would predominantly rely on the benzofuran core. The lack of reliable synthetic

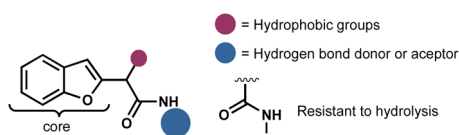


Fig. 2 Schematic structure of target molecules.

protocols for such molecules emphasizes the need to explore new synthetic strategies to develop them as promising agents for phytopathogenic control.

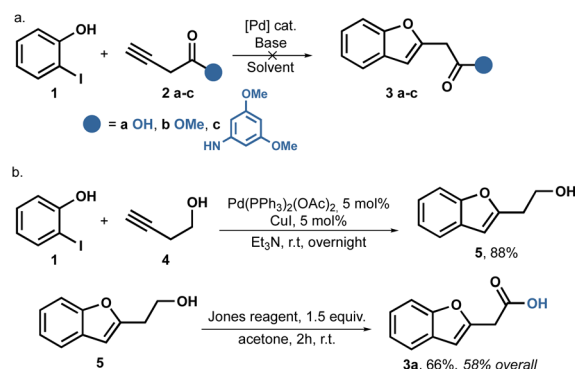
In light of these possibilities, we have conceived a method for synthesizing benzofuranyl acetic acid amides and the benzofuran core using either Sonogashira coupling-cyclization or rearrangements of methyl ketones. In this study, we present the results of employing these strategies and evaluate the *in vitro* activities of the obtained compounds against *Fusarium oxysporum*.

Results and discussion

We initiated our study by following the Sonogashira coupling method as described previously.^{38,45,46} It is well-known that when alkynes are coupled with *o*-iodophenols using the Sonogashira reaction, *o*-alkynyl alcohols are produced.^{47,48} These alcohols are *in situ* activated by the metal catalyst, leading to cyclizations and the formation of benzofurans. Based on this knowledge, we aimed to synthesize the desired amides using a one or two step process starting with 3-butynoic acid **2a**, its methyl ester **2b**, and the propynyl amide **2c**. Unfortunately, all our attempts to carry out this reaction were unsuccessful. We utilized $\text{Pd}(\text{PPh}_3)_2\text{Cl}_2$ as the catalyst, CuI as the cocatalyst, and Et_3N in stoichiometric amounts. However, in all cases, we were able to recover the starting phenol unchanged. We hypothesize that the allene is formed *in situ*, which is somehow less reactive (Scheme 1a).

Despite attempting to change the base to Na_2CO_3 , Cs_2CO_3 , or the catalyst for $\text{Pd}(\text{AcO})_2$, we did not achieve better results. Fortunately, when we switched to $\text{Pd}(\text{PPh}_3)_2(\text{OAc})_2$ as the catalyst, we observed some conversion only when using the alcohol **4** as the substrate; no reaction was observed with the acid **2a**, ester **2b**, or amide **2c**.

Consequently, we shifted our focus to a multi-step strategy and employed butynol **4** as the coupling partner. To our delight, the reaction proceeded smoothly, when using Et_3N as the solvent and successfully achieved full conversion, ultimately isolating the product **5** with a yield of 88%. Furthermore, a Jones oxidation allowed us to obtain benzofuranyl acetic acid



Scheme 1 Sonogashira coupling to benzofuranyl acetic acid **3a**.



3a in two steps, achieving an overall yield of 58%, as shown in Scheme 1b.

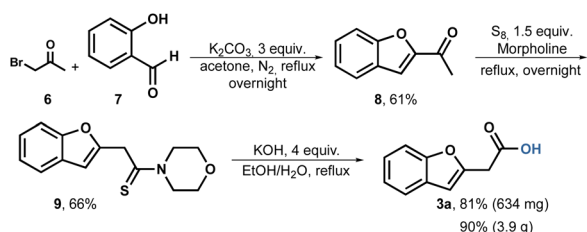
Despite the successful outcome of this sequence, we encountered difficulties when attempting to scale up the reaction. Increasing the reaction scale resulted in lower yields, and we were only able to replicate the yields up to 4 mmol scale.

Based on these findings, we conducted a Willgerdt–Kindler rearrangement.⁴⁹ The reaction sequence begins by forming methyl ketone **8** through the combination of commercially available bromoacetone **6** and salicylic aldehyde **7**. Under basic conditions, the reagents are heated, resulting in the production of methyl ketone **8** in a scale of up to 5.7 grams and a yield of 61%. For the Willgerdt–Kindler rearrangement, we employed sulfur as the reagent and morpholine as the base and solvent. To our satisfaction, the thioamide **9** was obtained at a scale of 4 grams with a yield of 66% after crystallization from ethanol. Subsequently, the hydrolysis of compound **9** proceeded smoothly using aqueous ethanol, achieving an 81% yield in the milligram scale and 90% in gram scale after acid-base extractions (Scheme 2).

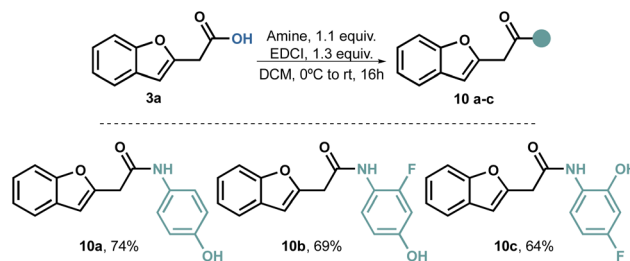
Up to this point, we have successfully employed two methods to achieve the synthesis of benzofuranyl acetic acid **3a**, yielding satisfactory overall results on a multi-gram scale. Subsequently, our focus shifted towards obtaining *N*-aryl amides.

We selected aminophenols as the reacting amines, with the hope of enhancing the water solubility of the resulting products. Since the primary goal of this research was to establish a dependable method for obtaining potential antifungal agents suitable for field trials, water solubility played a crucial role. Therefore, incorporating hydroxy groups into the aromatic ring could aid in achieving this essential feature.

Our attempts to perform a transamidation of thioamide^{50–52} **9** with aromatic amines proved unsuccessful. Consequently, we explored alternative traditional amidation conditions,^{53,54} including the direct amidation of the carboxylic acid;⁵⁵ however, the use of coupling reagents proved to be relatively straightforward. Initially, we conducted the amidations using dicyclohexylcarbodiimide (DCC) as the coupling reagent. While the reaction itself proceeded well, we encountered several challenges in purifying the product. Regardless of the solvent used, separating the product from the cyclohexyl urea by-product proved difficult. To overcome this issue, we switched to EDCI, as the urea by-product is water-soluble. With this modification, we successfully obtained compounds **10a–c** in acceptable to good yields (Scheme 3).



Scheme 2 Willgerdt–Kindler rearrangement to benzofuranyl acetic acid **3a**.



Scheme 3 Amidation of **3a** with hydroxyanilines.

One noteworthy aspect of this amidation process is its reliability, as we found no preconceived limitations. Consequently, the same protocol can be applied to various types of amines.

With the aim of establishing a dependable approach for acquiring amides with a benzofuranyl acetic core that possess structural diversity, our objective was to introduce alkyl groups in α positions.

It is widely acknowledged that the alkylation of primary and secondary amides poses a challenge primarily due to the deprotonation occurring at the more acidic nitrogen. Therefore, we revisited the deprotonation process of methyl ester **11**, which had been obtained by esterifying **3a** with excellent yield.

Extensive optimization was required for the deprotonation–alkylation process. Initially, we used *n*-BuLi as the deprotonating agent, but it resulted in virtually no conversion. Upon switching to LDA, we managed to isolate 35% of product **12a**. To improve the yield, we introduced TMEDA as an additive, which increased the yield to 51%. However, it was only when we used HMPA as a additive that we achieved full conversion and were able to isolate the desired product **12a** in a 75% yield. Through this process, we established that the combination of LDA and HMPA proved to be the optimal choice for achieving alkylation of **11** with alkyl iodides. Consequently, we extended the method to EtI, obtaining **12b** with a 73% yield.

The objective of introducing these alkyl groups was to evaluate their potential impact on the antifungal properties of the target compound. As we did not observe any significant effects in this case (*vide infra*), we did not carry out additional alkylations. Nonetheless, it is plausible to assume that this reaction may be limited to primary unhindered alkyl iodides. Subsequent hydrolysis under traditional conditions yielded alkylated acids **13a** and **13b** in excellent yields.

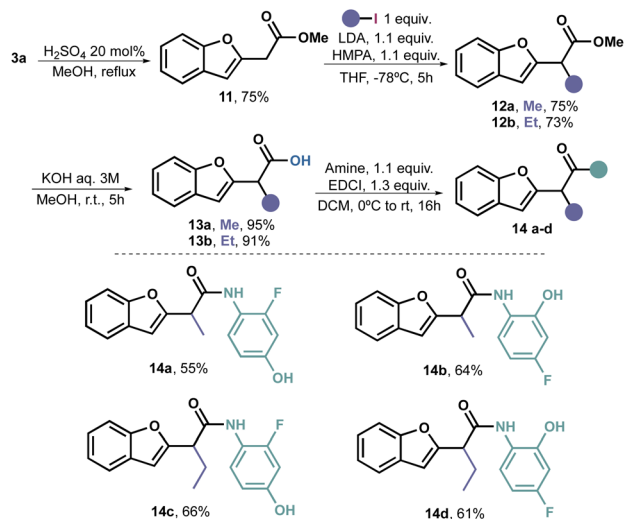
To complete the amidation, we followed the previously described procedure, resulting in the synthesis of four additional amides (**14a–d**) with acceptable to good yields (Scheme 4).

Armed with these results, we decided to conduct biological activity tests using *F. oxysporum* for this purpose (Table 1).

FOX is a pathogenic fungus that affects crops such as bananas, tomatoes, tropical fruits, and flowers, among others.⁷ Therefore, it has been used as a model system for testing antifungal activities for many years.

Initially, we tested the thioamide **9** as an analogue of the target molecules, but it showed no activity. Among compounds **10a–c**, only **10b** exhibited some inhibition on *F. oxysporum* growth (IC₅₀ = 0.42 mM). This suggests that the substitution pattern of





Scheme 4 Synthesis of α -alkyl benzofuranyl acetic acid amides **14**.

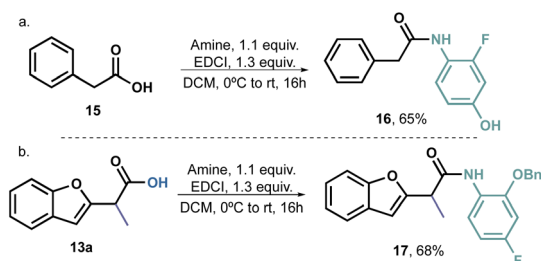
Table 1 IC_{50} values against *Fusarium oxysporum* through mycelial growth inhibition

Compound	IC_{50}^a (mM)
9	NA ^b
10a	NA
10b	0.42 ± 0.16
10c	NA
14a	1.27 ± 0.15
14b	10.99 ± 3.38
14c	0.70 ± 0.16
14d	1.12 ± 0.18
16	ND ^c
17	NA
Epoxiconazole (μ M)	0.06 ± 0.02

^a Data expressed as mean $\pm$ standard deviation using three replicates.

^b NA = not activity. ^c ND = not determined.

2-fluoro-4-hydroxy in the amine allows for some activity. This hypothesis was confirmed by the results obtained with compounds **14a** and **14c** ($IC_{50} = 1.27$ and 0.70 mM, respectively). Additionally, when comparing compounds **14a** with **14c** and **14b** with **14d**, there appears to be an apparent effect of the size of the alkyl group.



Scheme 5 Amidation of **15** and **13a** to **16** and **17** respectively.

Furthermore, we decided to prepare an analogue amide from phenylacetic acid **16** (Scheme 5a) and an *o*-benzyloxy derivative **17** (Scheme 5b). Compound **16** exhibited an unusual growth of fungus, rendering it impossible to determine the IC_{50} . However, this behaviour was solely observed at high concentrations, thus indicating that benzofurans have indeed potential antifungal activity and confirming our working hypothesis. On the other hand, the decision to use the benzyl group is supported by literature reports, which often highlight the presence of large hydrophobic groups in the structure of antifungal agents. Therefore, our aim was to determine whether the position 2 of the aromatic ring played a significant role in the compounds' activity. Since compound **17** did not exhibit any activity, it is reasonable to assume that small hydrogen bond acceptors would be more promising as substituents for this position.

Although the activities of compounds **10b** ($IC_{50} = 0.42$ mM) and **14c** are not comparable to the positive control or other previously described potent antifungal compounds, their inhibition suggests that modifying the alkyl chain or functionalizing the benzofuran ring may lead to the discovery of new and potent inhibitors of *F. oxysporum*. The synthetic method described in this paper can be utilized for both types of functionalization and modifications. It should be relatively easy to functionalize the benzofuran ring at least in three different stages, and alkylating with longer chains is plausible.

It is worth noting that when evaluated for antibacterial activity against *Agrobacterium tumefaciens* and *Pectobacterium carotovorum*, the compounds showed no activity.

Conclusions

In conclusion, we have described two distinct methods for synthesizing benzofuranyl acetic acid using commercially available materials. Both methods are applicable on a 2 mmol scale, with one of them being suitable for multigram quantities. Additionally, we have presented the alpha alkylation of these substrates and subsequent amidation under mild reaction conditions. In total, we have synthesized eight new benzofuranyl acetic acid amides and provided preliminary results on their biological activity against *Fusarium oxysporum*. Our ongoing research focuses on further functionalizing the molecules described in this study and testing their biological activity against other fungal strains. The results will be presented in due course.

Experimental section

General information

All the solvents were used as received from the supplier without any modifications. All reagents were used as received from commercial suppliers. Reactions progress was monitored by thin layer chromatography (TLC) performed on aluminium plates coated with silica gel F254 with 0.2 mm thickness TLC plates were visualized using ultraviolet (UV) light at 254 nm or stained with *p*-anisaldehyde, vanillin, ninhydrin, or phosphomolybdic acid solutions. Flash column chromatography was performed using silica gel 60 (230–400 mesh). Neat infrared



spectra were recorded using a Thermo Nicolet-Nexus (FT-IR) with PIKE MIRacle ATR cell. Wavenumbers (ν_{max}) are reported in cm^{-1} . High-resolution mass spectrometry was recorded using an Agilent 5973 (70 eV) spectrometer using electrospray ionization (ESI). GC-MS was recorded in a Thermo Scientific Trace 1300. NMR spectra were recorded using a BRUKER Advance III HD Ascend 400 spectrometer. Chemical shifts are given in parts per million (ppm, δ), referenced to TMS (^1H and ^{13}C) and trifluoroacetic acid (^{19}F) ($\delta = -75.39$), the solvent peak of CDCl_3 defined at $\delta = 7.26$ ppm (^1H NMR) and $\delta = 77.16$ (^{13}C NMR), $\text{DMSO}-d_6$ defined at $\delta = 2.5$ ppm (^1H NMR) and $\delta = 39.70$ (^{13}C NMR). Coupling constants are quoted in Hz (J). ^1H NMR splitting patterns were designated as singlet (s), doublet (d), triplet (t), quartet (q), (td) triplet of doublets and multiplet (m). Splitting patterns that could not be interpreted or easily visualized were designated as multiplet (m) or broad (br). Melting points were measured on solids after chromatography using a 1101D-MEL-TEMP melting point apparatus.

Experimental procedures and characterization

2-(Benzofuran-2-yl)ethan-1-ol (5). To a two-neck round bottom flask equipped with a magnetic stirrer was added $\text{Pd}(\text{PPh}_3)_2(\text{OAc})_2$ (0.3 mmol, 0.224 g) followed by CuI (0.3 mmol, 0.06 g) and 2-iodophenol (6.0 mmol, 1.33 g). The flask was then evacuated three times using a Schlenk line. Then, at rt, dry and deoxygenated Et_3N (12.0 mL, 0.5 M) was added followed by but-3-yn-1-ol (6.6 mmol, 0.5 mL) under nitrogen atmosphere. The reaction mixture was then stirred for overnight at room temperature. The mixture was concentrated, diluted with EtOAc (10 mL). The EtOAc layer was washed with water and brine, dried over anhydrous Na_2SO_4 , filtered and concentrated. The residue was purified by flash chromatography (DCM-100%) to afford 2-(benzofuran-2-yl) ethan-1-ol (5) as a colorless oil (0.858 g, 88%). ^1H NMR (400 MHz, CDCl_3) δ : 7.51 (d, $J = 8.5$ Hz, 1H), 7.43 (d, $J = 8.8$ Hz, 1H), 7.30–7.13 (m, 2H), 6.51 (s, 1H), 3.99 (t, $J = 6.2$ Hz, 2H), 3.05 (t, $J = 6.4$ Hz, 2H), 1.71 (bs, 1H). ^{13}C NMR (100 MHz, CDCl_3) δ : 156.1, 154.9, 128.8, 123.7, 122.8, 120.6, 111.0, 103.8, 60.9, 32.2. FT-IR (neat) $\nu(\text{cm}^{-1})$: 3332, 1600, 1454, 1249, 1045, 740, 428. HRMS (ESI): calc. For $\text{C}_{10}\text{H}_{10}\text{NaO}_2$ [$\text{M} + \text{Na}$] $^+$ 185.0578; found: 185.0576.

2-(Benzofuran-2-yl)acetic acid (3a). (Method A): to a stirring solution of 2-(benzofuran-2-yl)ethan-1-ol 5 (2.32 mmol, 0.375 g) in acetone (23 mL), was added Jones reagent (3.48 mmol, 2.7 mL). The reaction mixture was stirred for 2 hours. Methanol and water were added, the mixture was extracted with EtOAc (3 $\times$ 15 mL), the organic phases were reunited, dried over anhydrous Na_2SO_4 and concentrated under reduced pressure and the crude product was purified by flash chromatography (DCM-EtOAc 80 : 20) to afford 2-(benzofuran-2-yl) acetic acid (3a) as a brown solid (0.272 g, 66%). (Method B): 2-(benzofuran-2-yl)-1-morpholinoethane-1-thione 9 (21.8 mmol, 5.705 g) were dissolved in 30.0 mL of ethanol and subsequently KOH (87.2 mmol, 4.89 g) solution in 20.0 mL of a mixture 1:1 ethanol- H_2O was added; the resulting mixture was heated to reflux, and the reaction was followed by TLC. The crude was concentrated, and the basic aqueous solution washed three

times with EtOAc to remove organic impurities and morpholine; the aqueous phase was acidified with HCl, the mixture extracted with EtOAc (3 $\times$ 20 mL). The organic phases were combined, dried over anhydrous Na_2SO_4 and concentrated under reduced pressure to produce pure 2-(benzofuran-2-yl) acetic acid (3a) as a brown solid (3.9 g, 90%). ^1H NMR (400 MHz, CDCl_3) δ : 7.53 (dd, $J = 7.6$, 0.8 Hz, 1H), 7.46–7.43 (m, 1H), 7.28–7.16 (m, 2H), 6.66 (d, $J = 0.6$ Hz, 1H), 3.88 (s, 2H). ^{13}C NMR (100 MHz, CDCl_3) δ : 174., 155.1, 149.8, 128.5, 124.3, 123.0, 121.0, 111.3, 105.7, 34.4. FT-IR (neat) $\nu(\text{cm}^{-1})$: 3105, 2607, 1716, 1608, 1454, 1384, 1253, 1103, 959, 813, 740, 659. HRMS (ESI): calc. for $\text{C}_{10}\text{H}_8\text{NaO}_3$ [$\text{M} + \text{Na}$] $^+$ 199.0371; found: 199.0374, mp: 96–99 $^\circ\text{C}$.

1-(Benzofuran-2-yl)ethan-1-one (8). To a solution of 1-bromopropan-2-one (58.5 mmol, 8.00 g) in acetone were added potassium carbonate (175.5 mmol, 23.40 g) and 2-hydroxybenzaldehyde (64.4 mmol, 7.9 g) under N_2 . The resulting mixture was stirred at reflux overnight. The crude mixture was poured into crash ice, and the solid filtered and washed with water. Dry crude product was purified by flash chromatography (pentane-EtOAc 95 : 5) to afford 1-(benzofuran-2-yl)ethan-1-one (8) as a white solid (5.72 g, 61%), alternatively the product may be recrystallized from 95% ethanol obtaining 54% yield. ^1H NMR (400 MHz, CDCl_3) δ : 7.72 (d, $J = 7.9$ Hz, 1H), 7.59 (d, $J = 8.4$ Hz, 1H), 7.53–7.44 (m, 2H), 7.32 (t, $J = 7.5$ Hz, 1H), 2.62 (s, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ : 188.8, 155.7, 152.7, 128.4, 127.1, 124.0, 123.4, 113.2, 112.5, 26.6. FT-IR (neat) $\nu(\text{cm}^{-1})$: 3120, 3086, 1670, 1612, 1554, 1292, 1172, 1076, 972, 748, 636. HRMS (ESI): calc. for $\text{C}_{10}\text{H}_9\text{O}_2$ [$\text{M} + \text{H}$] $^+$ 161.0597; found 161.0602, mp: 67–71 $^\circ\text{C}$.

2-(Benzofuran-2-yl)-1-morpholinoethane-1-thione (9). A mixture of 1-(benzofuran-2-yl)ethan-1-one 8 (25.3 mmol, 4.06 g) and sulfur (38.0 mmol, 1.217 g) in 3.27 mL morpholine was refluxed overnight with agitation. After cooling the dark viscous mixture, 10 mL of water and extracted with EtOAc (3 $\times$ 15 mL). The combined organic layers were washed with brine, dried over anhydrous Na_2SO_4 , and concentrated under reduced pressure. The crude product was recrystallized from 95% to afford pure thioamide 9 in 66% yield (4.36 g), alternatively 9 may be purified by flash chromatography (DCM-CHX 90 : 10) to obtain 2-(benzofuran-2-yl)-1-morpholinoethane-1-thione (9) as a brown solid (4.03 g, 61%). ^1H NMR (400 MHz, CDCl_3) δ : 7.57 (d, $J = 7.2$ Hz, 1H), 7.46 (d, $J = 7.9$ Hz, 1H), 7.35–7.18 (m, 2H), 6.69 (s, 1H), 4.51 (s, 2H), 4.42 (t, $J = 5.0$ Hz, 2H), 3.93 (d, $J = 4.9$ Hz, 2H), 3.84 (t, $J = 5.0$ Hz, 2H), 3.74 (t, $J = 4.9$ Hz, 2H). ^{13}C NMR (100 MHz, CDCl_3) δ : 196.3, 154.7, 152.4, 128.3, 124.1, 123.0, 120.9, 111.0, 104.6, 66.4, 51.2, 50.3, 43.9. FT-IR (neat) $\nu(\text{cm}^{-1})$: 3086, 2858, 1608, 1481, 1273, 1111, 1026, 956, 887, 817, 759, 686. HRMS (ESI): calc. for $\text{C}_{14}\text{H}_{16}\text{NO}_2\text{S}$ [$\text{M} + \text{H}$] $^+$, 262.0902; found, 262.0903, mp: 156–160 $^\circ\text{C}$.

General procedure for amidation

To a stirred solution of the corresponding acid (1.0 equiv.), and aniline derivative (1.1 equiv.), in DCM at 0 $^\circ\text{C}$; was added EDCI (1-ethyl-3-(3-dimethylaminopropyl)carbodiimide 1.3 equiv.), the mixture is stirred and left under stirring until reaching r.t.



gradually and then stirred for 16 h at r.t. The mixture was poured into 10% HCl solution and extracted with EtOAc (3 × 20 mL); and the combined organic extracts were combined, washed with brine, dried over anhydrous Na₂SO₄, filtered and concentrated under reduced pressure. The crude mixture was purified using flash chromatography to afford the pure product.

2-(Benzofuran-2-yl)-*N*-(4-hydroxyphenyl)acetamide (10a).

According to the general procedure, to a stirred solution of 2-(benzofuran-2-yl)acetic acid **3a** (0.57 mmol, 0.10 g), 4-aminophenol (0.62 mmol, 0.068 g), 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide (0.74 mmol, 0.1415 g) and DCM (3.5 mL); were used to obtain 2-(benzofuran-2-yl)-*N*-(4-hydroxyphenyl)acetamide (**10a**) as a yellow solid (0.113 g, 74.4%). Purified with DCM 100%. ¹H NMR (400 MHz, DMSO-*d*₆) δ: 10.04 (s, 1H), 9.23 (s, 1H), 7.55 (dd, *J* = 25.4, 7.7 Hz, 2H), 7.38 (d, *J* = 8.9 Hz, 2H), 7.32–7.17 (m, 2H), 6.92–6.61 (m, 3H), 3.86 (s, 2H). ¹³C NMR (100 MHz, DMSO-*d*₆) δ: 165.9, 154.6, 153.9, 153.8, 131.1, 128.9, 124.2, 123.2, 121.4, 121.1, 115.6, 111.2, 105.1, 36.9. FT-IR (neat) ν(cm^{−1}): 3529, 1659, 1539, 1454, 1346, 1296, 1234, 1161, 960, 825, 752. HRMS (ESI): calc. for C₁₆H₁₃NO₃ (M + H⁺), 268.0974; found, 268.0969, mp: 191–194 °C.

(Benzofuran-2-yl)-*N*-(2-fluoro-4-hydroxyphenyl)acetamide (10b).

According to the general procedure, to a stirred solution of 2-(benzofuran-2-yl)acetic acid **3a** (0.57 mmol, 0.10 g), 4-amino-3-fluorophenol (0.62 mmol, 0.079 g), 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide (0.74 mmol, 0.1415 g) and DCM (3.5 mL); were used to obtain 2-(benzofuran-2-yl)-*N*-(2-fluoro-4-hydroxyphenyl)acetamide (**10b**) as a light yellow solid (0.112 g, 69.2%). Purified with DCM 100%. ¹H NMR (400 MHz, DMSO-*d*₆) δ: 9.90–9.74 (m, 2H), 7.56 (dd, *J* = 24.2, 8.2 Hz, 2H), 7.46 (t, *J* = 9.0 Hz, 1H), 7.31–7.18 (m, 2H), 6.76 (s, 1H), 6.69–6.55 (m, 3H), 3.94 (s, 2H). ¹³C NMR (100 MHz, DMSO-*d*₆) δ: 166.6, 155.9 (d, *J* = 11.0 Hz), 155.4 (d, *J* = 244.8 Hz), 154.4, 153.4, 128.6, 126.6 (d, *J* = 3.5 Hz), 123.9, 122.9, 120.9, 117.0 (d, *J* = 12.7 Hz), 111.1 (d, *J* = 2.9 Hz), 111.0, 104.9, 103.0 (d, *J* = 22.0 Hz), 36.0. ¹⁹F-NMR (374 MHz, DMSO-*d*₆) δ: −122.3. FT-IR (neat) ν(cm^{−1}): 3224, 3047, 1616, 1546, 1450, 1365, 1157, 1103, 956, 817, 752. HRMS (ESI): calc. for C₁₆H₁₃FO₃ (M + H⁺), 286.0869; found, 286.0871, mp: 170–174 °C.

2-(Benzofuran-2-yl)-*N*-(4-fluoro-2-hydroxyphenyl)acetamide (10c).

According to the general procedure, to a stirred solution of 2-(benzofuran-2-yl)acetic acid **3a** (0.57 mmol, 0.10 g), 2-amino-5-fluorophenol (0.62 mmol, 0.079 g), 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide (0.74 mmol, 0.1415 g) and DCM (3.5 mL); were used to obtain 2-(benzofuran-2-yl)-*N*-(4-fluoro-2-hydroxyphenyl)acetamide (**10c**) as a light brown solid (0.103 g, 63.6%). Purified with CHX-DCM 50 : 50/DCM 100%. ¹H NMR (400 MHz, DMSO-*d*₆) δ: 10.38 (s, 1H), 9.46 (s, 1H), 7.78 (dd, *J* = 8.9, 6.4 Hz, 1H), 7.55 (dd, *J* = 25.9, 7.5 Hz, 2H), 7.37–7.10 (m, 2H), 6.77 (s, 1H), 6.67 (dd, *J* = 10.3, 2.8 Hz, 1H), 6.60 (td, *J* = 8.7, 2.8 Hz, 1H), 4.01 (s, 2H). ¹³C NMR (100 MHz, DMSO-*d*₆) δ: 166.7, 159.1 (d, *J* = 240.4 Hz), 154.4, 153.5, 149.7 (d, *J* = 11.1 Hz), 128.6, 123.9, 123.7 (d, *J* = 9.9 Hz), 122.9, 122.8 (d, *J* = 2.9 Hz), 120.9, 111.0, 105.2 (d, *J* = 21.9 Hz), 104.9, 102.7 (d, *J* = 24.9 Hz), 36.3. ¹⁹F-NMR (374 MHz, DMSO-*d*₆) δ: −117.4. FT-IR (neat) ν(cm^{−1}): 3371, 2897, 2750, 1654, 1527, 1423, 1238, 1157, 1103, 979, 933,

759. HRMS (ESI): calc. for C₁₆H₁₃FO₃ (M + H⁺), 286.0869; found, 286.0869, mp: 165–168 °C.

Methyl 2-(benzofuran-2-yl)acetate (11). 2-Benzofuranyl acetic acid **3a** (4.54 mmol, 0.800 g) in methanol (5 mL) was added, then H₂SO₄ (0.91 mmol, 253 μL) was added; the mixture was refluxed for 12 h using a CaCl₂ trap. The reaction was monitored by TLC. After cooling the mixture, H₂O (5 mL) was added, and it was extracted with EtOAc (3 × 10 mL). The organic phase was washed with NaHCO₃, dried over anhydrous Na₂SO₃ and concentrated under reduced pressure and the crude product was purified by flash chromatography (pentane 100%) to afford methyl 2-(benzofuran-2-yl)acetate (**11**) as a yellow liquid (0.648 g, 75%). ¹H NMR (400 MHz, CDCl₃) δ: 7.53 (d, *J* = 7.3 Hz, 1H), 7.45 (d, *J* = 8.2 Hz, 1H), 7.30–7.16 (m, 2H), 6.63 (s, 1H), 3.85 (s, 2H), 3.76 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ: 169.5, 155.1, 150.7, 128.6, 124.1, 122.9, 120.9, 111.2, 105.3, 52.6, 34.6. FT-IR (neat) ν(cm^{−1}): 2940, 1739, 1454, 1250, 1007, 957, 741. HRMS (ESI): calc. For C₁₁H₁₁O₃ [M + H]⁺ 191.0707; found: 191.0703.

Methyl 2-(benzofuran-2-yl)propanoate (12a). To a freshly prepared solution of LDA (5.4 mmol in THF 5.4 mL), a solution methyl 2-(benzofuran-2-yl)acetate **11** (5.2 mmol, 1.0 g) in dry THF (15.4 mL) was added dropwise at −78 °C and HMPA (5.4 mmol, 0.97 g) and stirred for 1 h at this temperature. CH₃I (5.2 mmol, 2.6 mL) was added dropwise, and the mixture was stirred at −78 °C for 4 h. After the reaction was complete, saturated aqueous NH₄Cl was added and extracted with EtOAc (3 × 10 mL). The combined organic layers were dried over anhydrous MgSO₄. After filtration, concentration under reduced pressure, and purification by flash chromatography (pentane-EtOAc, 90 : 10) to afford methyl 2-(benzofuran-2-yl)propanoate (**12a**) as a colorless liquid (0.80 g, 75%). ¹H NMR (400 MHz, CDCl₃) δ: 7.52 (d, *J* = 7.7 Hz, 1H), 7.44 (d, *J* = 8.0 Hz, 1H), 7.22 (dt, *J* = 19.6, 7.4 Hz, 2H), 6.57 (s, 1H), 3.96 (q, *J* = 7.3 Hz, 1H), 3.72 (s, 3H), 1.62 (d, *J* = 7.3 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ: 172.6, 156.3, 154.8, 128.4, 124.0, 122.8, 120.8, 111.2, 103.2, 52.5, 39.9, 15.8. FT-IR (neat) ν(cm^{−1}): 2989, 1743, 1610, 1455, 1380, 1201, 1095, 945, 740. HRMS (ESI): calc. for C₁₂H₁₂O₃Na [M + Na]⁺: 227.0679; found: 227.0675.

Methyl 2-(benzofuran-2-yl)butanoate (12b). To a freshly prepared solution of LDA (2.8 mmol in THF 2.3 mL), a solution methyl 2-(benzofuran-2-yl)acetate **11** (2.6 mmol, 0.5 g) in dry THF (7.7 mL) was added dropwise at −78 °C and HMPA (2.8 mmol, 0.49 g) and stirred for 1 h at this temperature. Ethyl iodide (2.6 mmol, 0.21 mL) was added dropwise, and the mixture was stirred at −78 °C for 4 h. After the reaction was complete, saturated aqueous NH₄Cl was added and extracted with EtOAc (3 × 10 mL). The combined organic layers were dried over anhydrous MgSO₄. After filtration, concentration under reduced pressure, and purification by flash chromatography (pentane 100) to afford methyl 2-(benzofuran-2-yl)butanoate (**12b**) as a colorless liquid (0.42 g, 73%). ¹H NMR (400 MHz, CDCl₃) δ: 7.52 (d, *J* = 7.2 Hz, 1H), 7.45 (d, *J* = 8.0 Hz, 1H), 7.31–7.15 (m, 2H), 6.60 (s, 1H), 3.74 (d, *J* = 10.4 Hz, 4H), 2.23–1.97 (m, 2H), 0.99 (t, *J* = 7.4 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ: 172.1, 155.1, 154.6, 128.4, 123.9, 122.7, 120.8, 111.2, 103.9, 76.7, 52.3, 47.3, 24.2, 12.0. FT-IR (neat) ν(cm^{−1}): 2970,

1735, 1454, 1253, 1002, 798, 748. HRMS (ESI): calc. for $C_{13}H_{15}O_3$ $[M + H]^+$ 219.1016; found: 219.1014.

2-(Benzofuran-2-yl)propanoic acid (13a). To a solution of methyl 2-(benzofuran-2-yl) propanoate **12a** (2.6 mmol, 0.53 g) in methanol 26 mL was added a solution of aqueous KOH 3 M (4.3 mL). The resulting mixture was stirred for 3 h at room temperature. After evaporation of MeOH, the mixture was acidified with HCl 2 M, then saturated with NaCl and extracted with DCM. The organic layer was washed with brine and dried over anhydrous Na_2SO_4 . After filtration, concentration under reduced pressure to afford 2-(benzofuran-2-yl)propanoic acid (**13a**) as a white solid (0.47 g, 95%). 1H NMR (400 MHz, $CDCl_3$) δ : 7.52 (d, J = 7.5 Hz, 1H), 7.45 (d, J = 8.0 Hz, 1H), 7.28–7.16 (m, 2H), 6.62 (s, 1H), 3.99 (q, J = 7.2 Hz, 1H), 1.64 (d, J = 7.2 Hz, 3H). ^{13}C NMR (100 MHz, $CDCl_3$) δ : 178.0, 155.5, 154.8, 128.0, 124.1, 122.7, 120.7, 111.0, 103.5, 39.7, 15.5. FT-IR (neat) ν (cm^{-1}): 3200, 1693, 1604, 1454, 1215, 1068, 937, 802, 740, 671. HRMS (ESI): calc. for $C_{22}H_{20}NaO_6$ $[2M + Na]^+$, 403.3858; found, 403.3864, mp: 98–102 °C.

2-(Benzofuran-2-yl)butanoic acid (13b). To a solution of methyl 2-(benzofuran-2-yl)butanoate **12b** (1.8 mmol, 0.38 g) in methanol (26 mL) was added a solution of aqueous KOH 3 M (2.9 mL). The resulting mixture was stirred for 5 h at room temperature. After evaporation of MeOH, the mixture was acidified with HCl 2 M, then saturated with NaCl and extracted with DCM. The organic layer was washed with brine and dried over anhydrous Na_2SO_4 . After filtration, concentration under reduced pressure to afford 2-(benzofuran-2-yl)butanoic acid (**13b**) as a yellow solid (0.33 g, 91%). 1H NMR (400 MHz, $CDCl_3$) δ : 7.53 (d, J = 7.6 Hz, 1H), 7.46 (d, J = 8.1 Hz, 1H), 7.35–7.14 (m, 2H), 6.65 (s, 1H), 3.78 (t, J = 7.5 Hz, 1H), 2.40–1.86 (m, 2H), 1.02 (t, J = 7.4 Hz, 3H). ^{13}C NMR (100 MHz, $CDCl_3$) δ : 177.5, 154.8, 154.2, 128.3, 124.1, 122.8, 120.9, 111.2, 104.4, 47.2, 24.0, 11.9. FT-IR (neat) ν (cm^{-1}): 2974, 1697, 1600, 1412, 1254, 1006, 871, 752. HRMS (ESI): calc. for $C_{12}H_{13}O_3$ $[M + H]^+$ 205.0858; found, 205.0856, mp: 75–78 °C.

2-(Benzofuran-2-yl)-N-(2-fluoro-4-hydroxyphenyl)propanamide (14a). According to the general procedure, to a stirred solution of 2-(benzofuran-2-yl)propanoic acid **13a** (0.64 mmol, 0.122 g), 4-amino-3-fluorophenol (0.67 mmol, 0.08 g), 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide (0.83 mmol, 0.159 g) and DCM (3.0 mL); were used to obtain 2-(benzofuran-2-yl)-N-(2-fluoro-4-hydroxyphenyl)propanamide (**14a**) as a white solid (0.105 g, 55%). Purified with (DCM 100%). 1H NMR (400 MHz, $DMSO-d_6$) δ : 9.77 (d, J = 18.9 Hz, 2H), 7.58 (d, J = 7.2 Hz, 1H), 7.52 (d, J = 7.8 Hz, 1H), 7.43 (t, J = 8.9 Hz, 1H), 7.28–7.11 (m, 2H), 6.74 (s, 1H), 6.59 (dd, J = 20.2, 11.5 Hz, 2H), 4.34–3.98 (m, 1H), 1.52 (d, J = 7.0 Hz, 3H). ^{13}C NMR (101 MHz, $DMSO-d_6$) δ : 170.4, 158.6, 157.0, 156.2 (d, J = 10.8 Hz), 154.5 (d, J = 6.7 Hz), 128.7, 127.0 (d, J = 3.4 Hz), 124.2, 123.2, 121.2, 117.2 (d, J = 12.7 Hz), 111.4 (d, J = 2.6 Hz), 103.3, 16.2. ^{19}F -NMR (374 MHz, $DMSO-d_6$) δ = –122.2. FT-IR (neat) ν (cm^{-1}): 3350, 2952, 1664, 1535, 1428, 1280, 1151, 955, 832, 750, 650. HRMS (ESI): calc. for $C_{17}H_{15}FNO_3$ $[M + H]^+$, 300.1036; found, 300.1029, mp: 134–140 °C.

2-(Benzofuran-2-yl)-N-(4-fluoro-2-hydroxyphenyl)propanamide (14b). According to the general procedure, to

a stirred solution of 2-(benzofuran-2-yl)propanoic acid **13a** (0.51 mmol, 0.097 g), 2-amino-5-fluorophenol (0.53 mmol, 0.067 g), 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide (0.66 mmol, 0.127 g) and DCM (2.6 mL); were used to obtain 2-(benzofuran-2-yl)-N-(4-fluoro-2-hydroxyphenyl)propanamide (**14b**) as a white solid (0.117 g, 64%). Purified with (DCM-CHX 90 : 10). 1H NMR (400 MHz, $CDCl_3$) δ : 8.80 (s, 1H), 7.74 (s, 1H), 7.58 (d, J = 7.6 Hz, 1H), 7.50 (d, J = 8.1 Hz, 1H), 7.35–7.18 (m, 2H), 6.85 (dd, J = 8.7, 5.7 Hz, 1H), 6.72 (d, J = 10.5 Hz, 2H), 6.59–6.49 (m, 1H), 4.06 (q, J = 7.3 Hz, 1H), 1.75 (d, J = 7.2 Hz, 3H). ^{13}C NMR (100 MHz $CDCl_3$) δ : 171.7, 161.7 (d, J = 245.2 Hz), 155.3, 154.9, 150.4 (d, J = 12.3 Hz), 127.9, 124.8, 123.3, 123.2 (d, J = 10.3 Hz), 121.6 (d, J = 3.1 Hz), 121.2, 111.2, 107.3 (d, J = 23.1 Hz), 107.0 (d, J = 24.6 Hz), 104.6, 41.5, 15.8. ^{19}F -NMR (374 MHz, $CDCl_3$) δ : –114.2. FT-IR (neat) ν (cm^{-1}): 3379, 2931, 1647, 1539, 1431, 1280, 1149, 968, 840, 752, 655, 459. HRMS (ESI): calc. for $C_{17}H_{15}FNO_3$ $(M + H)^+$, 300.1036; found, 300.1040, mp: 138–142 °C.

2-(Benzofuran-2-yl)-N-(2-fluoro-4-hydroxyphenyl)butanamide (14c). According to the general procedure, to a stirred solution of 2-(benzofuran-2-yl)butanoic acid **13b** (0.35 mmol, 0.071 g), 4-amino-3-fluorophenol (0.42 mmol, 0.053 g), 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide (0.45 mmol, 0.087 g) and DCM (2.2 mL); were used to obtain 2-(benzofuran-2-yl)-N-(2-fluoro-4-hydroxyphenyl)butanamide (**14c**) as a white solid (0.072 g, 66%). Purified with DCM 100%. 1H NMR (400 MHz, $DMSO-d_6$) δ : 7.79 (t, J = 8.8 Hz, 1H), 7.63–7.53 (m, 2H), 7.48 (d, J = 8.0 Hz, 1H), 7.33–7.15 (m, 2H), 6.70 (s, 1H), 6.61–6.49 (m, 2H), 6.13 (s, 1H), 3.80–3.67 (m, 1H), 2.34–2.21 (m, 1H), 2.18–2.07 (m, 1H), 1.05 (t, J = 7.4 Hz, 3H). ^{13}C NMR (100 MHz, $DMSO-d_6$) δ : 169.8, 155.5, 155.0 (d, J = 6.9 Hz), 154.0 (d, J = 10.2 Hz), 153.1, 128.3, 124.5, 124.3 (d, J = 2.1 Hz), 123.2, 121.1, 118.3 (d, J = 13.3 Hz), 111.4 (d, J = 3.3 Hz), 111.3, 105.0, 103.4 (d, J = 22.6 Hz), 50.1, 24.5, 12.3. ^{19}F -NMR (374 MHz, $DMSO-d_6$) δ = –121.9. FT-IR (neat) ν (cm^{-1}): 3603, 3364, 2967, 1647, 1512, 1454, 1250, 1107, 802, 744. HRMS (ESI): calc. for $C_{18}H_{17}FNO_3$ $[M + H]^+$ 314.1186; found, 314.1187; mp: 124–130 °C.

2-(Benzofuran-2-yl)-N-(4-fluoro-2-hydroxyphenyl)butanamide (14d). According to the general procedure, to a stirred solution of 2-(benzofuran-2-yl)butanoic acid **13b** (0.35 mmol, 0.071 g), 2-amino-5-fluorophenol (0.42 mmol, 0.053 g), 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide (0.45 mmol, 0.087 g) and DCM (2.2 mL); were used to obtain 2-(benzofuran-2-yl)-N-(4-fluoro-2-hydroxyphenyl)butanamide (**14d**) as a brown oil (0.066 g, 61%). Purified with CHX-DCM 80 : 20. 1H NMR (400 MHz, $CDCl_3$) δ : 8.79 (s, 1H), 7.87 (s, 1H), 7.56 (d, J = 7.8 Hz, 1H), 7.48 (d, J = 8.2 Hz, 1H), 7.36–7.15 (m, 2H), 6.91 (d, J = 5.3 Hz, 1H), 6.88–6.59 (m, 2H), 6.61–6.43 (m, 1H), 4.09–3.70 (m, 1H), 2.33–2.21 (m, 1H), 2.20–2.06 (m, 1H), 1.05 (t, J = 7.4 Hz, 3H). ^{13}C NMR (100 MHz $CDCl_3$) δ : 171.4, 161.6 (d, J = 246.2 Hz), 155.0, 154.2, 150.3 (d, J = 9.9 Hz), 128.0, 124.7, 123.3, 123.2 (d, J = 10.3 Hz), 121.7 (d, J = 3.2 Hz), 121.2, 111.3, 107.4 (d, J = 23.0 Hz), 106.8 (d, J = 22.7 Hz), 105.4, 49.4, 24.5, 12.2. ^{19}F -NMR (374 MHz, $CDCl_3$) δ = –114.3. FT-IR (neat) ν (cm^{-1}) = 2970, 1654, 1608, 1454, 1254, 1153, 976, 802. HRMS (ESI): calc. for $C_{18}H_{17}FNO_3$ $[M + H]^+$ 314.1186; found, 314.1181.

N-(2-fluoro-4-hydroxyphenyl)-2-phenylacetamide (16). According to the general procedure, to a stirred solution of 2-



phenylacetic acid **15** (0.51 mmol, 0.069 g), 4-amino-3-fluorophenol (0.56 mmol, 0.071 g), 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide (0.66 mmol, 0.127 g) and DCM (3.2 mL); were used to obtain *N*-(2-fluoro-4-hydroxyphenyl)-2-phenylacetamide (**16**) as a white solid (0.082 g, 65%). Purified with DCM 100%. ¹H NMR (400 MHz, DMSO-*d*₆) δ: 9.71 (d, *J* = 43.0 Hz, 2H), 7.44–7.21 (m, 6H), 6.94–6.32 (m, 2H), 3.63 (s, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ: 169.5, 155.8 (d, *J* = 11.0 Hz), 155.5 (d, *J* = 244.7 Hz), 136.4, 129.3, 128.5, 126.7, 126.6 (d, *J* = 4.3 Hz), 117.2 (d, *J* = 12.6 Hz), 111.1 (d, *J* = 2.9 Hz), 102.9 (d, *J* = 22.1 Hz), 42.6. ¹⁹F-NMR (374 MHz, DMSO-*d*₆) δ = −122.3. FT-IR (neat) ν(cm^{−1}): 3159, 1616, 1539, 1458, 1315, 1230, 1153, 964, 848, 763. HRMS (ESI): calc. for C₁₄H₁₃FNO₂ [M + H]⁺ 246.0930; found, 246.0925, mp: 169–173 °C.

2-(Benzofuran-2-yl)-N-(2-(benzyloxy)-4-fluorophenyl)propanamide (17). According to the general procedure, to a stirred solution of 2-(benzofuran-2-yl)propanoic acid **3a** (0.47 mmol, 0.09 g), 2-(benzyloxy)-4-fluoroaniline (0.49 mmol, 0.11 g), 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide (0.61 mmol, 0.117 g) and DCM (2.4 mL); were used to obtain 2-(benzofuran-2-yl)-*N*-(2-(benzyloxy)-4-fluorophenyl) propanamide (**17**) as a white solid (120 mg, 68%). Purified with pentane-EtOAc 90 : 10. ¹H NMR (400 MHz, CDCl₃) δ: 8.32 (t, *J* = 7.5 Hz, 1H), 8.16 (s, 1H), 7.69–7.02 (m, 9H), 6.73–6.51 (m, 3H), 4.97 (s, 2H), 4.15–3.71 (m, 1H), 1.68 (d, *J* = 6.6 Hz, 3H). ¹³C NMR (100 MHz CDCl₃) δ: 169.1, 159.0 (d, *J* = 242.9 Hz), 156.5, 154.9, 148.1 (d, *J* = 9.9 Hz), 135.6, 128.8, 128.2, 127.6, 124.0 (d, *J* = 3.3 Hz), 120.7 (d, *J* = 9.2 Hz), 111.1, 120.9, 123.0, 124.3, 128.5, 107.2 (d, *J* = 21.7 Hz), 103.8, 100.1 (d, *J* = 27.1 Hz), 71.0, 42.5, 15.7. ¹⁹F-NMR (374 MHz, CDCl₃) δ: −115.8. FT-IR (neat) ν(cm^{−1}): 3271, 3032, 1874, 1654, 1535, 1284, 1222, 972, 825, 732. HRMS (ESI): calc. for C₂₄H₂₁FNO₃ (M + H)⁺, 390.1505; found, 390.1499, mp: 118–122 °C.

Author contributions

C. M.-M. investigation, methodology, validation, writing-review and editing and visualisation; P. B.-M. investigation biological activity, validation, writing-review and editing and visualisation; L M. P writing-review and editing and funding acquisition and D. G.-S. conceptualisation, project administration, resources, writing-original draft, writing-review and editing, funding acquisition, supervision, and visualisation.

Conflicts of interest

There are no conflicts to declare.

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Notes and references

- 1 L. Serwecińska, *Water*, 2020, **12**, 3313.
- 2 C. Manyi-Loh, S. Mamphweli, E. Meyer and A. Okoh, *Molecules*, 2018, **23**, 795.
- 3 J. B. Ristaino, P. K. Anderson, D. P. Bebbler, K. A. Brauman, N. J. Cuniffe, N. V. Fedoroff, C. Finegold, K. A. Garrett, C. A. Gilligan, C. M. Jones, M. D. Martin, G. K. MacDonald, P. Neenan, A. Records, D. G. Schmale, L. Tateosian and Q. Wei, *Proc. Natl. Acad. Sci. U. S. A.*, 2021, **118**, e2022239118.
- 4 M. Tudi, H. Daniel Ruan, L. Wang, J. Lyu, R. Sadler, D. Connell, C. Chu and D. T. Phung, *Int. J. Environ. Res. Public Health*, 2021, **18**, 1112.
- 5 T. R. Gordon, *Annu. Rev. Phytopathol.*, 2017, **55**, 23–39.
- 6 F. Shabani and L. Kumar, *PLoS One*, 2013, **8**, e83404.
- 7 L. Burgess and W. Bryden, *Microbiol. Aust.*, 2012, **33**, 22–25.
- 8 D. H. M. Alkhalifah, E. Damra, M. B. Melhem and W. N. Hozzein, *Microorganisms*, 2023, **11**, 468.
- 9 E. Sandulachi, A. Ghendov-Mosan, D. Cojocari and R. Sturza, *Adv. Microbiol.*, 2021, **11**, 541–553.
- 10 G. B. Eshun, F. J. Osonga, T. Erdogan, A. Gölcü and O. A. Sadik, *RSC Adv.*, 2023, **13**, 21781–21792.
- 11 A. Akhmedov, R. Gamirov, Y. Panina, E. Sokolova, Y. Leonteva, E. Tarasova, R. Potekhina, I. Fitsev, D. Shurpik and I. Stoikov, *Org. Biomol. Chem.*, 2023, **21**, 4863–4873.
- 12 L. Tai, S.-X. Sun, K. Yue, J.-Q. Chai, S.-T. Hou, G.-Y. Dai, C.-L. Yang and M. Chen, *New J. Chem.*, 2023, **47**, 12850–12860.
- 13 I. I. Ozturk, K. Turk, A. M. Grześkiewicz, M. Kubicki, C. N. Banti and S. K. Hadjikakou, *New J. Chem.*, 2023, **47**, 12779–12789.
- 14 L. Du, S. Haldar, J. B. King, A. O. Mattes, S. Srivastava, K. L. Wendt, J. You, C. Cunningham and R. H. Cichewicz, *J. Nat. Prod.*, 2023, DOI: [10.1021/acs.jnatprod.3c00382](https://doi.org/10.1021/acs.jnatprod.3c00382).
- 15 S. Y. Jeong, A. Alishir, S. Zhang, Y. Zhang, S. Choi, C. Pang, H. Y. Bae, W. H. Jung and K. H. Kim, *J. Nat. Prod.*, 2023, DOI: [10.1021/acs.jnatprod.3c00193](https://doi.org/10.1021/acs.jnatprod.3c00193).
- 16 X.-y. Zhi, Y. Zhang, Y.-f. Li, Y. Liu, W.-p. Niu, Y. Li, C.-r. Zhang, H. Cao, X.-j. Hao and C. Yang, *J. Agric. Food Chem.*, 2023, **71**, 11680–11691.
- 17 C. Yang, S. Sun, W. Li, Y. Mao, Q. Wang, Y. Duan, R. Csuk and S. Li, *J. Agric. Food Chem.*, 2023, **71**, 11341–11349.
- 18 M. Noman, Azizullah, T. Ahmed, Y. Gao, H. Wang, X. Xiong, J. Wang, J. Lou, D. Li and F. Song, *J. Agric. Food Chem.*, 2023, **71**, 11667–11679.
- 19 A. A. Abbas and K. M. Dawood, *RSC Adv.*, 2023, **13**, 11096–11120.
- 20 H. X. Ngo, S. Garneau-Tsodikova and K. D. Green, *MedChemComm*, 2016, **7**, 1285–1306.

- 21 Y. Liu, Y. Wang, G. Dong, Y. Zhang, S. Wu, Z. Miao, J. Yao, W. Zhang and C. Sheng, *MedChemComm*, 2013, **4**, 1551–1561.
- 22 M. Masubuchi, H. Ebiike, K.-i. Kawasaki, S. Sogabe, K. Morikami, Y. Shiratori, S. Tsujii, T. Fujii, K. Sakata, M. Hayase, H. Shindoh, Y. Aoki, T. Ohtsuka and N. Shimma, *Bioorg. Med. Chem.*, 2003, **11**, 4463–4478.
- 23 X. Cao, M. Wang, Z. Li and X. Xu, *J. Agric. Food Chem.*, 2023, **71**, 8757–8768.
- 24 B. Gabriele, R. Mancuso, G. Salerno and M. Costa, *J. Org. Chem.*, 2007, **72**, 9278–9282.
- 25 N. Thirupathi, M. Hari Babu, V. Dwivedi, R. Kant and M. Sridhar Reddy, *Org. Lett.*, 2014, **16**, 2908–2911.
- 26 Y. He, Z. Li, K. Robeyns, L. Van Meervelt and E. V. Van der Eycken, *Angew. Chem., Int. Ed.*, 2018, **57**, 272–276.
- 27 B. Li, T. Li, M. A. Aliyu, Z. H. Li and W. Tang, *Angew. Chem., Int. Ed.*, 2019, **58**, 11355–11359.
- 28 K.-O. Kim and J. Tae, *Synthesis*, 2005, **2005**, 387–390.
- 29 E. J. Enholm and G. Prasad, *Tetrahedron Lett.*, 1989, **30**, 4939–4942.
- 30 B. Gabriele, R. Mancuso, G. Salerno and M. Costa, *Adv. Synth. Catal.*, 2006, **348**, 1101–1109.
- 31 Y. Fang and C. Li, *J. Org. Chem.*, 2006, **71**, 6427–6431.
- 32 T. Huang, Y. Shao, S. Tang and J. Sun, *Org. Biomol. Chem.*, 2023, **21**, 5752–5756.
- 33 V. Ramella, Z. He, C. G. Daniliuc and A. Studer, *Eur. J. Org. Chem.*, 2016, **2016**, 2268–2273.
- 34 A. Nakatani, K. Hirano, T. Satoh and M. Miura, *Chem. –Eur. J.*, 2013, **19**, 7691–7695.
- 35 Y. Yamane, K. Yoshinaga, M. Sumimoto and T. Nishikata, *ACS Catal.*, 2019, **9**, 1757–1762.
- 36 W. Tian, B. Li, D. Tian and W. Tang, *Chin. Chem. Lett.*, 2022, **33**, 197–200.
- 37 M. Chakravarty and K. C. Kumara Swamy, *J. Org. Chem.*, 2006, **71**, 9128–9138.
- 38 P. K. Mandal and D. K. Chand, *Synthesis*, 2015, **47**, 1661–1668.
- 39 C. E. Castro, E. J. Gaughan and D. C. Owsley, *J. Org. Chem.*, 1966, **31**, 4071–4078.
- 40 P. Borrego-Muñoz, F. Ospina and D. Quiroga, *Molecules*, 2021, **26**, 3997.
- 41 Y.-Q. Xie, Y.-B. Huang, J.-S. Liu, L.-Y. Ye, L.-M. Che, S. Tu and C.-L. Liu, *Pest Manage. Sci.*, 2015, **71**, 404–414.
- 42 H. Xu, Z. Hou, Z. Liang, M.-B. Guo, X. Su and C. Guo, *Chin. J. Chem.*, 2019, **37**, 1245–1250.
- 43 B. Shankar, P. Jalapathi, M. Ramesh, A. K. Kumar, M. Ragavender and G. Bharath, *Russ. J. Gen. Chem.*, 2016, **86**, 1711–1721.
- 44 S.-S. Du, X.-F. Luo, J.-X. An, Z.-J. Zhang, S.-Y. Zhang, Y.-R. Wang, Y.-Y. Ding, W.-Q. Jiang, B.-Q. Zhang, Y. Ma, Y. Zhou, Y.-M. Hu and Y.-Q. Liu, *Pest Manage. Sci.*, 2023, **79**, 2748–2761.
- 45 M. Csékei, Z. Novák and A. Kotschy, *Tetrahedron*, 2008, **64**, 8992–8996.
- 46 M. J. Moure, R. SanMartin and E. Domínguez, *Adv. Synth. Catal.*, 2014, **356**, 2070–2080.
- 47 M. Nakamura, L. Ilies, S. Otsubo and E. Nakamura, *Angew. Chem., Int. Ed.*, 2006, **45**, 944–947.
- 48 M. Nakamura, L. Ilies, S. Otsubo and E. Nakamura, *Org. Lett.*, 2006, **8**, 2803–2805.
- 49 D. L. Priebbenow and C. Bolm, *Chem. Soc. Rev.*, 2013, **42**, 7870–7880.
- 50 A. Ojeda-Porras and D. Gamba-Sánchez, *Tetrahedron Lett.*, 2015, **56**, 4308–4311.
- 51 G. Li, Y. Xing, H. Zhao, J. Zhang, X. Hong and M. Szostak, *Angew. Chem., Int. Ed.*, 2022, **61**, e202200144.
- 52 D. R. Goud and U. Pathak, *Synthesis*, 2012, **44**, 3678–3682.
- 53 A. Ojeda-Porras and D. Gamba-Sánchez, *J. Org. Chem.*, 2016, **81**, 11548–11555.
- 54 R. M. de Figueiredo, J.-S. Suppo and J.-M. Campagne, *Chem. Rev.*, 2016, **116**, 12029–12122.
- 55 A. Ojeda-Porras, A. Hernández-Santana and D. Gamba-Sánchez, *Green Chem.*, 2015, **17**, 3157–3163.

