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# Surface tension models for binary aqueous solutions: a review and intercomparison†

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The liquid–air surface tension of aqueous solutions is a fundamental quantity in multi-phase thermodynamics and fluid dynamics and thus relevant in many scientific and engineering fields. Various models have been proposed for its quantitative description. This Perspective gives an overview of the most popular models and their ability to reproduce experimental data of ten binary aqueous solutions of electrolytes and organic molecules chosen to be representative of different solute types. In addition, we propose a new model which reproduces sigmoidal curve shapes (Sigmoid model) to empirically fit experimental surface tension data. The surface tension of weakly surface-active substances is well reproduced by all models. In contrast, only few models successfully model the surface tension of aqueous solutions with strongly surface-active substances. For substances with a solubility limit, usually no experimental data is available for the surface tension of supersaturated solutions and the pure liquid solute. We discuss ways in which these can be estimated and emphasize the need for further research. The newly developed Sigmoid model best reproduces the surface tension of all tested solutions and can be recommended as a model for a broad range of binary mixtures and over the entire concentration range.

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## 1. Introduction

The surface tension of aqueous solutions is a key physical property in many fields, influencing *e.g.* the efficiency of distillation trays,<sup>1</sup> the biological functioning of lung surfactants,<sup>2</sup> the size distribution of aerosol produced by medical nebulizers,<sup>3</sup> and the number of aerosol particles activated to cloud droplets in the atmosphere.<sup>4</sup> It depends on the type and concentration of the solutes, temperature, and the curvature of the surface, the latter becoming relevant only in droplets with radii of curvature approaching the molecular scale.<sup>5</sup> Various techniques allow to measure the surface tension of large drops or bulk solutions. Instruments are also emerging to measure the surface tension of small droplets, such as atomic force microscopy<sup>6</sup> and holographic optical tweezers.<sup>7</sup> Most measurements of the surface tension of aqueous solutions reported in the literature, however, are bulk measurements and restricted

to aqueous solutions with one solute only (binary solutions). Data of aqueous solutions with more than one solute (multi-component solutions) are scarce. Mathematical models of surface tension have been developed for different purposes, be it simply to convert discrete measurement data points into an analytical expression, or to predict the surface tension for unknown systems. The approach and the complexity of surface tension models in the literature varies widely, from models based on molecular dynamics,<sup>8–10</sup> machine learning models,<sup>11–14</sup> to empirical and thermodynamic models, each type of model being justified by its ability to fulfill a specific goal (*e.g.*, accuracy, simplicity, or robustness). Given the importance of surface tension across various disciplines, it is important to develop mathematical tools that can accurately describe how surface tension varies as a function of solution composition.

In many models for bulk aqueous solutions, surface tension is expressed as a function of composition at a constant temperature, using the surface tension of the pure components as input parameters. They differ with respect to their physical basis and derivation, their number of fitting parameters, and their ability to fit differently-shaped surface tension curves as a function of solute concentration. Some are mathematically equivalent, although having been derived in different ways. Moreover, some can be more easily extended to multi-component solutions than others. Often, a certain model is favoured for

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particular substances or in a specific field, such as liquid metal alloys, which is the major application area for the Butler equation,<sup>15</sup> while atmospheric sciences rely often on the Szyszkowski–Langmuir model for the description of surface tension of liquid aerosol particles at high relative humidity.<sup>16–42</sup> However, a systematic, interdisciplinary comparison of the performance of these models is lacking. In this study, we fill this gap by analyzing the performance of common surface tension models for bulk binary aqueous solutions.

The effect a solute has on the surface tension of water varies greatly with the type of solute. Electrolytes are known to increase the surface tension of the solution. Other substances, such as simple alcohols, have a lower pure component surface tension than water and thus lower the surface tension of the solution, with a more or less pronounced deviation from a linear mixing rule. Substances with a strong tendency to partition to the surface drastically lower the surface tension of a solution even when they are present in only very small amounts. The term “surfactants”, which stands for surface-active-agents, usually refers to this latter kind of substance (strong tendency to partition), while those substances with a weaker tendency to partition can be named “weak surfactants”.

In this study, we investigate whether a model can be found to apply to all of these solute types, as such a model would have several advantages: First, describing all substances with a single model allows a compressed representation of experimental data, requiring only fit parameters to be reported. Second, if the parameters of this model have physical meaning, the substances can be compared to each other by comparison of their parameters (*e.g.*, their extent to which they partition to the surface). Lastly, such a model has the potential to be extended to a multi-component model for mixtures of different types of surface active species. Since experimental data for multi-component solutions are scarce, but experimental data for binary solutions are broadly available, a predictive model for multi-component solutions could be developed that is based on fitting to binary experimental data. In atmospheric sciences, electrolytes, weakly surface active organics and strong surfactants can be mixed together in one aerosol particle. Therefore, the ability of the binary model to fit all these substance types individually is a prerequisite for a model to predict the surface tension of the mixture.

In aiming for a universal surface tension model, a challenge is posed by substances whose pure liquid surface tension values are unknown because they crystallize in bulk volumes at the temperature where the surface tension is required (*e.g.* electrolytes). Small droplets containing such substances can easily supersaturate and remain liquid at concentrations where larger volumes would crystallize. Hence, for such systems a model is required that can predict surface tension at concentrations where measured values are lacking. Here, we test the ability of the models to predict the surface tension of crystallizing substances, and discuss different approaches to deal with these substances.

In the following, we first summarize the theoretical background of the most common surface tension models for

aqueous bulk solutions and analyze their mathematical form. Second, we analyze the applicability and performance of comparable surface tension models on a set of ten test substances. In our results and discussion, we report difficulties and constraints of the different models, the best fit parameters for the tested substances as well as knowledge gaps suggesting needs for further research.

## 2. Overview of literature surface tension models

In the literature, various semi-empirical surface tension models for aqueous solutions can be found. Here, we summarize the main assumptions behind popular models, some characteristics of their equations and examples of their applications. A summary of these models is given in Table 1. All models are a function of either the mole fraction  $x_i$ , the activity  $a_i$ , or the molarity  $c_i$  of the solute  $i$ , with

$$x_i + x_w = 1, \quad (1)$$

$$c_i = \rho \frac{x_i}{x_i \tilde{M}_i + x_w \tilde{M}_w}, \quad (2)$$

where  $x_w$  is the mole fraction of water,  $\rho$  is the density of the solution, and  $\tilde{M}_i$  and  $\tilde{M}_w$  are the molar mass of the solute and of water, respectively. In this study, we define  $a_i = \gamma_i x_i$  for organic substances, where  $\gamma_i$  is the activity coefficient of the solute, and  $a_i = \gamma_i m_i$  for electrolytes, where  $m_i$  is the molality of the solute. In principle, the enrichment of solute in the surface leads to its depletion in the bulk phase and therefore, the composition of the bulk phase  $x_i^{\text{bulk}}$  needs to be distinguished from the total composition  $x_i^{\text{total}}$ . This effect, however, only becomes relevant in small volumes, where the number of solute molecules at the surface  $N_i^{\text{surf}}$  accounts for a non-negligible fraction of the total number of solute molecules  $N_i^{\text{total}}$ . In large volumes, due to the lower surface-to-volume ratio,  $N_i^{\text{surf}} \ll N_i^{\text{total}} \approx N_i^{\text{bulk}}$ . In the following, therefore, we simply define  $x_i := x_i^{\text{bulk}} \approx x_i^{\text{total}}$ .

### 2.1 Gibbsian surface tension models

Gibbs's work on the thermodynamics of surfaces in 1876 was a milestone in the science of surfaces and set the basis for many surface tension models thereafter. For bulk systems, the main idea in Gibbsian surface tension models is the concept of the Gibbs dividing surface—a theoretical phase of zero thickness, lying between the bulk phase and the gas phase. While for a real surface, the concentration of a substance follows a certain profile across the surface, in the Gibbs approach, this concentration profile is modelled by a step function, and the amount of substance that is under or over accounted by this approach is compensated for in the Gibbs dividing surface (surface excess).<sup>74–76</sup> Different approaches exist about where to place the Gibbs dividing surface. Commonly, it is defined as the equimolar surface with respect to water, *i.e.* where the surface excess of water equals zero. Another approach defines the Gibbs dividing surface as the special equimolar surface where the sum of the surface excesses of all substances weighted by



**Table 1** Summary of common semi-empirical surface tension models in the literature, as described in Section 2. Models marked with an asterisk are further evaluated in the next section

| Name                          | Ref.      | Equation for binary mixtures                                                                                                                                                                           | Fit parameters                               | Multi-component solutions              | Used in         |
|-------------------------------|-----------|--------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|----------------------------------------------|----------------------------------------|-----------------|
| Li & Lu model*                | 43        | $\sigma = \sigma_w + RT \Gamma^{\max} \ln \frac{1}{1 + a_i K}$                                                                                                                                         | $\Gamma^{\max}, K$                           | Two equations proposed <sup>43</sup>   | 44–46           |
| Szyszkowski–Langmuir equation | 47 and 48 | $\sigma = \sigma_w - RT \alpha \ln(1 + c_i/\beta)$                                                                                                                                                     | $\alpha, \beta$                              | See <sup>21,25,27,49</sup>             | 16–42           |
| Statistical model*            | 50–52     | $\sigma = \sigma_w + \frac{kT}{rS_w} \ln \frac{1 - Ka_i}{1 - Ka_i(1 - C)}$                                                                                                                             | $r, K, C$                                    | See Boyer <i>et al.</i> <sup>53</sup>  | 54              |
| Butler equation*              | 55–59     | $\begin{cases} \sigma = \sigma_w + \frac{RT}{A_w} \ln \left( \frac{a_w^{\text{surf}}}{a_w} \right) \\ \sigma = \sigma_i + \frac{RT}{A_i} \ln \left( \frac{a_i^{\text{surf}}}{a_i} \right) \end{cases}$ | —                                            | Inherent                               | 44,45 and 60–64 |
| Tamura model*                 | 59 and 65 | $\sigma^{1/4} = \psi_w^{\text{surf}} \sigma_w^{1/4} + (1 - \psi_w^{\text{surf}}) \sigma_i^{1/4}$                                                                                                       | —                                            | Not inherent                           | 44 and 45       |
| Eberhart model*               | 66        | $\sigma = \frac{x_w \sigma_w + S x_i \sigma_i}{x_w + S x_i}$                                                                                                                                           | $S$                                          | Not inherent                           | 44 and 45       |
| Shereshefsky model            | 67        | $\sigma = \sigma_w - \frac{(\sigma_w - \sigma_i) x_i e^{\frac{\Delta F_s}{RT}}}{1 - x_i \left( e^{\frac{\Delta F_s}{RT}} - 1 \right)}$                                                                 | $\Delta F_s$                                 | Not inherent                           | 68              |
| Connors–Wright model*         | 69        | $\sigma = \sigma_w - \left( 1 + \frac{b x_w}{1 - a x_w} \right) x_i (\sigma_w - \sigma_i)$                                                                                                             | $a, b$                                       | See Shardt <i>et al.</i> <sup>70</sup> | 68 and 70       |
| Chunxi model                  | 71        | $\sigma = x_i \sigma_i + x_w \sigma_w - \frac{x_i x_w RT}{x_w + x_i A_{wi}} \left( \frac{\partial A_{wi}}{\partial A} \right) \left( 1 - \frac{1}{A_{wi}} \right)$                                     | $A_{wi}, \frac{\partial A_{wi}}{\partial A}$ | See Chunxi <i>et al.</i> <sup>71</sup> | 32,72 and 73    |

their molar volume equals zero, which is equivalent to setting the volume of the surface phase to zero.<sup>75</sup> The location of the Gibbs dividing surface does not affect the magnitude of the surface tension for bulk systems, but it plays a role for liquid volumes of finite size, *i.e.* for small droplets with a diameter < 10 nm.<sup>75</sup>

Another fundamental ingredient for Gibbsian surface tension models is the Gibbs adsorption equation, which relates the composition of the surface to the surface tension. It is usually written as

$$\sum_{i=1}^n \Gamma_i^s d\mu_i + A d\sigma = 0, \quad (3)$$

where  $\Gamma_i^s$  is the molar surface excess and  $\mu_i$  the chemical potential of substance  $i$ ,  $A$  is the surface area and  $\sigma$  the surface tension. A derivation of the Gibbs adsorption equation was given by Elliot.<sup>76</sup>

To relate the surface composition to the bulk composition, a Langmuir adsorption isotherm is often used. Langmuir adsorption was originally derived for the adsorption of an adsorbent from the gas phase to a solid substrate. It assumes (i) that the adsorbent behaves like an ideal gas with a certain partial pressure, (ii) that the adsorption rate  $K$  equals the desorption rate in equilibrium and (iii) that the solid substrate has a maximum number of binding sites  $\Gamma^{\max}$ .<sup>77</sup> Analogously, to model the partitioning of a solute  $i$  from a liquid bulk phase to a liquid–gas interface, the partial pressure is replaced by the activity of the solute in the bulk  $a_i$ , leading to:

$$\Gamma_i^s = \Gamma^{\max} \frac{a_i K}{1 + a_i K}, \quad (4)$$

where  $\Gamma^{\max}$  and  $K$  can be obtained by fitting to experimental data. These assumptions together with placing the Gibbs dividing surface at the equimolar surface with respect to water, leads to the model described by Li and Lu<sup>43</sup> (**Li & Lu model**) which for a binary water–solute mixture reads as

$$\sigma = \sigma_w + RT \Gamma^{\max} \ln \frac{1}{1 + a_i K}, \quad (5)$$

where  $\Gamma^{\max}$  and  $K$  are fit parameters. Li and Lu<sup>43</sup> applied the model to aqueous electrolyte solutions using Pitzer activity coefficients and proposed two equations for calculating the surface tension of mixed electrolyte aqueous solutions without introducing more free parameters. Topping *et al.*<sup>44</sup> and Booth *et al.*<sup>45</sup> applied this model to various single and multiple solute mixtures of organic and inorganic compounds typical for the atmosphere and compared its performance with the one of the Butler equation and the model by Tamura *et al.*<sup>65</sup> (see below).

Sorjamaa *et al.*<sup>75</sup> proposed a similar model consisting of a set of equations to calculate the surface tension of binary and multi-component solutions with the following differences from the Li & Lu model: (i) the Gibbs dividing surface is placed at the “special equimolar surface”, (ii) the activity coefficients of the solutes are calculated from Henrys law constants, and (iii) ideality is assumed for water.

Lastly, a famous model following the Gibbs–Langmuir approach is known as the **Szyszkowski–Langmuir equation**, sometimes also called “Szyszkowski equation of state”<sup>24</sup> or “Szyszkowski–Langmuir adsorption isotherm”.<sup>16</sup> Originally formulated as an entirely empirical model by Szyszkowski<sup>47</sup>



based on experiments with aqueous solutions containing fatty acids, this model was later interpreted thermodynamically by Langmuir.<sup>48</sup> It can be written as

$$\sigma = \sigma_w - RT\alpha \ln(1 + c_i/\beta), \quad (6)$$

where  $\alpha$  and  $\beta$  are fit parameters and  $c_i$  is the molar concentration of the solute  $i$  in the bulk. Note that different notations exist *e.g.* with  $\beta c_i$  instead of  $c_i/\beta$  which leads to different fit parameters.<sup>16</sup> Eqn (6) can be derived from the Li & Lu model (eqn (5)) by assuming a dilute solution ( $a_i \approx c_i$ ) and hence replacing  $a_i$  with  $c_i$  leads to  $\Gamma^{\max} = \alpha$  and  $K = \beta^{-1}$ . Thus, the Szyszkowski–Langmuir equation is only accurate for dilute solutions with decreasing accuracy as concentration increases (see Section S3 in ESI†). The Szyszkowski–Langmuir equation originally has no formulation for multi-component solutions, but some equations have been suggested for how to extend the Szyszkowski–Langmuir equation for multi-component solutions.<sup>21,25,27,49</sup> In atmospheric sciences the Szyszkowski–Langmuir equation is a popular model that has been used for various substances (*e.g.* dicarboxylic acids, humic substances, sodium dodecyl sulfate).<sup>16–42</sup>

## 2.2 Statistical mechanics model

Aiming for a model that works equally well for non-electrolyte and electrolyte solutes over the full range of concentrations, Wexler and Dutcher<sup>50</sup> derived a surface tension model from a Gibbs free energy expression using statistical mechanics (**Statistical model**). The sorption of solute molecules to the liquid–vapor interface is modelled by representing the surface as a monolayer and the bulk as a multilayer. The model is formulated for binary aqueous mixtures as

$$\sigma = \sigma_w + \frac{kT}{rS_w} \ln \frac{1 - Ka_i}{1 - Ka_i(1 - C)}, \quad (7)$$

where  $r$  is the number of water molecules displaced from the surface by each solute molecule,  $C$  and  $K$  are parameters related to the energy of the multilayer and the monolayer, respectively, and  $S_w$  is the area occupied by a water molecule. The fit parameter  $r$  is positive for surfactants and negative if the solute is depleted from the surface. According to the derivation of the model,  $K$  and  $C$  are defined as  $K = \exp(\varepsilon_{SB}/kT)$  and  $C = \exp((\varepsilon_{SS} - \varepsilon_{SB})/kT)$ , where  $\varepsilon_{SB}$  and  $\varepsilon_{SS}$  are the energies of each solute molecule in the bulk and surface, respectively, and  $k$  is the Boltzmann's constant.<sup>51</sup> Therefore,  $K$  and  $C$  can only be positive. The parameter  $K$  also has to fulfill  $Ka_i = 1 - N_{SS}/N_s$ ,<sup>50</sup> where  $N_s$  is the total number of solute molecules and  $N_{SS}$  is the number of the solute molecules in the surface. From examining this equation at  $a_i = 1$ , it can be deduced that  $K$  has to be within  $[0,1]$ , since the right side of the equation is within  $[0,1]$ . When  $\sigma_w$  and  $S_w$  are known, the model has three free parameters, namely  $r$ ,  $K$  and  $C$ . However, if  $\sigma_i$  is known as well,  $C$  can be eliminated as a free parameter by replacing it with

$$C = 1 - \left[ 1 - (1 - K) \exp\left(\frac{rS_w(\sigma_w - \sigma_i)}{kT}\right) \right] / K. \quad (8)$$

For substances with strong partitioning to the surface,  $K$  becomes very small and  $C$  very large. For this case, the model can be reduced to a simpler form with only the two free parameters  $r$  and  $K$ :

$$\sigma = \sigma_w - \frac{kT}{rS_w} \ln(1 + Ka_i), \quad (9)$$

which is mathematically equivalent to the Li & Lu model (eqn (5)) with  $RT\Gamma^{\max} = kT/(rS_w)$ . If in this case  $\sigma_i$  is known,  $K$  can be expressed as a function of  $\sigma_i$  and  $r$  leading to a one parameter model.

In Boyer<sup>51</sup> the predictive capabilities of the model were further improved by relating the model parameters to solute molecular properties, thereby reducing the number of free parameters. To this end, a number of alcohols, polyols and electrolytes have been tested. For the electrolytes, the root mean squared error (RMSE) of the fully predictive surface tension model is between 0.4 and 3.6 mN m<sup>−1</sup>. For the organics, a one-parameter version of the model leads to equally good RMSE values between 0.3 and 3.5 mN m<sup>−1</sup>. Results from their fully predictive model for alcohols are only shown for methanol, 1,2-ethanediol and 1,3-butanediol. For these three alcohols, their predictive model provides excellent agreement with the models that have one or two fitted parameters.

Boyer and Dutcher<sup>78</sup> extended the single-solute model to a multiple-solute version, in order to model the surface tension of aqueous organic acid solutions including partial dissociation of the acid. For the seven di- and tricarboxylic acids that were examined, this “ternary model” shows a very similar fitting capability as the one-parameter-model neglecting dissociation (“binary model”), with differences in the RMSE of the models < 0.02 mN m<sup>−1</sup>. This suggests that the inclusion of the dissociation of the acids does not improve the model considerably; however, the ternary model laid a foundation for the development of a general model for multi-component solutions as followed up by Boyer *et al.*<sup>53</sup> Miles *et al.*<sup>54</sup> have applied the multi-component model to picoliter NaCl–glutaric acid–water solution droplets of varying composition reaching good agreement with surface tension measurements obtained using a microdroplet dispenser and a holographic optical tweezer. A summary of all these model developments can be found in Boyer and Dutcher.<sup>52</sup>

Recently, Liu and Dutcher<sup>79</sup> extended the surface tension model from Wexler and Dutcher<sup>50</sup> to considering multiple surface layers to model the concentration depth profile at the surface. They compared model fits to experimental data for several organic and inorganic substances with good agreement, but the results are not compared to the simpler model version with a monolayer assumption for the surface. Thus it remains unclear if the higher complexity of the model actually leads to improved surface tension predictions.

In all mentioned publications from this research group, activity coefficients were calculated with their own activity coefficients model, which is based on the statistical mechanics of multilayer sorption.<sup>80–82</sup>



### 2.3 The Butler equation

In 1932, Butler,<sup>55</sup> derived a set of equations to calculate the surface tension of bulk mixtures of any number of substances, which is commonly written as

$$\sigma = \sigma_i \frac{A_i^0}{A_i} + \frac{RT}{A_i} \ln \left( \frac{a_i^{\text{surf}}}{a_i} \right) \quad i = 1, 2, \dots, n, \quad (10)$$

where  $n$  is the total number of substances and  $a_i^{\text{surf}}$  and  $a_i$  are the activities of solute  $i$  in the surface phase and the bulk, respectively. Usually, the molar surface areas of the pure substance  $A_i^0$  and of the substance in the mixture  $A_i$  are set equal, which further simplifies eqn (10). With predictive values for  $\sigma_i$ ,  $A_i$  and activity coefficients as a function of composition, the so-called **Butler equation** needs no fitting to experiments. It has to be noted that the equation cannot be solved when the activity of a substance reaches zero in the bulk (division by zero) or in the surface (undefined logarithm of zero) which is e.g. the case for the pure substances where the mole fractions are  $x_i = 0$  or  $x_i = 1$ . For a given bulk composition, the surface mole fraction of each substance  $x_i^{\text{surf}}$  (including solutes and solvents) and the surface tension ( $n + 1$  unknowns) can be calculated by solving the system of  $n + 1$  equations formed by eqn (10) and

$$\sum_{i=1}^n x_i^{\text{surf}} = 1. \quad (11)$$

The performance of the Butler equation is determined by the choice of  $\sigma_i$ ,  $A_i$  and the activity coefficients. Therefore, for optimal performance, experimental values for  $\sigma_i$  should be used, if available. To determine  $A_i$  and the activity coefficients, many different equations or methods have been proposed. For aqueous mixtures, Poling *et al.*<sup>59</sup> recommend the method of Suarez *et al.*,<sup>57</sup> where activity coefficients are obtained with a version of the “Universal Quasichemical Functional Group Activity Coefficients” (UNIFAC) model<sup>83</sup> and molar surface areas are calculated with the equation from Goldsack and White:<sup>84</sup>

$$A_i = 1.021 \times 10^8 V_c^{6/15} V_b^{4/15}, \quad (12)$$

where  $V_c$  is the critical molar volume in  $\text{cm}^3 \text{mol}^{-1}$ ,  $V_b$  is the bulk liquid molar volume in  $\text{cm}^3 \text{mol}^{-1}$  of substance  $i$  and  $A_i$  is in  $\text{cm}^2 \text{mol}^{-1}$ . Suarez *et al.*<sup>57</sup> also estimated  $A_i$  from fitting to experimental surface tension data of binary solutions for 20 different components. Such fitted values for  $A_i$  led to much better surface tension predictions. Poling *et al.*<sup>59</sup> gives an example for the correct application of the Suarez method for a propanol–water mixture. Alternatively,  $A_i$  is often calculated as

$$A_i = V_i^{2/3} N_A^{1/3}, \quad (13)$$

where  $V_i$  is the molar volume of substance  $i$  and  $N_A$  is Avogadro's constant, following the suggestion from Sprow and Prausnitz<sup>56</sup> to assume a nearly spherical molecule.

The Butler equation has been used for a variety of compounds, including liquid metal alloys,<sup>85,86</sup> electrolyte solutions<sup>60,61</sup> and atmospherically-relevant mixtures. Cai and Griffin<sup>62</sup> calculated the surface tension of secondary organic aerosol particles applying

the procedure described by Poling *et al.*<sup>59</sup> This calculation was carried out in the context of evaluating for which compositions and particle sizes of secondary organic aerosol particles the Kelvin effect has to be considered for the gas–liquid partitioning of semi-volatile substances in aerosol chamber experiments. A comparison of the modelled surface tension values to experimental data was not done by Cai and Griffin,<sup>62</sup> due to lack of experimental data.

In contrast, Topping *et al.*<sup>44</sup> and Booth *et al.*<sup>45</sup> systematically evaluated the performance of the predictive Butler equation (referred to as the model by Sprow & Prausnitz or Suarez) by comparing to experimental data for various single and multiple solute mixtures of atmospherically-relevant organic and inorganic compounds. They pointed out that for many atmospherically-relevant compounds  $\sigma_i$  is unknown, since the pure compounds tend to crystallize in the bulk at atmospheric conditions, hence making surface tension measurements in the bulk impossible. Therefore, they applied models to estimate  $\sigma_i$ , which resulted in poor performance compared to a surface tension model with fit parameters such as the Li & Lu model.

Werner *et al.*<sup>63,64</sup> applied a Butler equation-based method of Li *et al.*<sup>60</sup> to aqueous succinic acid solutions with<sup>64</sup> and without<sup>63</sup> inorganic salts and compared their results to experiments. Here, the original system of equations was reduced to one equation only, namely that for water. Further, they assumed, that  $m_{\text{succinic acid}}^{\text{surf}} = g \cdot m_{\text{succinic acid}}^{\text{bulk}}$  where  $m$  is the molality and  $g$  is the surface enrichment factor, the latter of which they obtained from X-ray photoelectron spectroscopy measurements and from molecular dynamics simulations. The modelled surface tensions could reproduce the trend of the measurements. It was also shown that inorganic salts further increase the surface enrichment of succinic acid.

### 2.4 Parachor model

A semi-empirical model for the surface tension of a single organic solute in water was described by Tamura *et al.*<sup>65</sup> (**Tamura model**). It is based on the Macleod–Sugden correlation, which relates  $\sigma_i$  to liquid and vapor densities *via* a temperature independent parameter called the “parachor”. The final model equation is:<sup>59</sup>

$$\sigma^{1/4} = \psi_w^{\text{surf}} \sigma_w^{1/4} + (1 - \psi_w^{\text{surf}}) \sigma_i^{1/4}, \quad (14)$$

where  $\psi_w^{\text{surf}}$  is the superficial volume fraction of water in the surface layer. It is calculated from<sup>44,59</sup>

$$\log_{10} \frac{\psi_w^{\text{surf}}}{1 - \psi_w^{\text{surf}}} = \log_{10} \left[ \frac{(x_w V_w)^q}{x_i V_i} (x_w V_w + x_i V_i)^{1-q} \right] + 44.1 \frac{q}{T} \left[ \frac{\sigma_i V_i^{2/3}}{q} - \sigma_w V_w^{2/3} \right], \quad (15)$$

where  $T$  is the temperature,  $q$  is a parameter that depends on the length and type of organic substance and  $V_i$  and  $V_w$  are the molar volume of substance  $i$  and water, respectively. This model, like the Butler equation, cannot be solved for  $x_i = 0$  or  $x_i = 1$  due to the logarithmic function.



Topping *et al.*<sup>44</sup> and Booth *et al.*<sup>45</sup> tested the Tamura model as a fully predictive model for aqueous solutions of various organic and inorganic substances and a number of aqueous multi-component solutions. Since the Tamura model has no version for multi-component solutions, an additive approach was used for these, such that the deviations from  $\sigma_w$  caused by the individual solutes are simply added to give the total deviation of  $\sigma$ . The comparison to experimental data revealed large discrepancies of the model predictions, mostly due to the high uncertainty in  $\sigma_i$  values.

## 2.5 Simple thermodynamic models

Based on the assumption that the surface tension of a binary liquid mixture is an average of the pure substance surface tensions weighted by the surface concentrations, Eberhart<sup>66</sup> derived a simple equation for the mixture surface tension, which for a single-solute–water mixture reads as

$$\sigma = \frac{x_w \sigma_w + S x_i \sigma_i}{x_w + S x_i}, \quad (16)$$

where  $S$  is a dimensionless parameter describing the surface layer enrichment in the solute (**Eberhart model**). Based on the same assumption, but with a different derivation, Shereshefsky<sup>67</sup> found the equation

$$\sigma = \sigma_w - \frac{(\sigma_w - \sigma_i) x_i e^{\frac{\Delta F_s}{RT}}}{1 - x_i \left( e^{\frac{\Delta F_s}{RT}} - 1 \right)}, \quad (17)$$

where  $\Delta F_s$  describes the free energy change of replacing water in the surface region by a mole of solute (**Shereshefsky model**). This model is mathematically equivalent to the Eberhart model, with  $S = e^{\frac{\Delta F_s}{RT}}$ .

A simple model with two free parameters was derived by Connors and Wright<sup>69</sup> (**Connors–Wright model**), based on the same assumption of  $\sigma$  being an average weighted by  $x_i^{\text{surf}}$  but with additional assumptions about the state of the surface layer and by applying a Langmuir adsorption isotherm to describe the partitioning between bulk and the surface. For a single-solute–water mixture, it is written as

$$\sigma = \sigma_w - \left( 1 + \frac{b x_w}{1 - a x_w} \right) x_i (\sigma_w - \sigma_i), \quad (18)$$

where  $a$  and  $b$  are fit parameters. For  $a = b$ , the model reduces to the Shereshefsky/Eberhart model, with

$$a = b = 1 - 1/e^{\frac{\Delta F_s}{RT}} = 1 - 1/S. \quad (19)$$

The parameters  $a$  and  $b$  influence the slope of the surface tension–solute mole fraction curve at low and high  $x_i$ , respectively (see Fig. 1). If  $b = 0$ , the equation reduces to a simple linear mixing rule.

Another two-parameter model was derived by Chunxi *et al.*<sup>71</sup> (**Chunxi model**) based on the Wilson equation<sup>87</sup> as

$$\sigma = x_i \sigma_i + x_w \sigma_w - \frac{x_i x_w RT}{x_w + x_i A_{wi}} \left( \frac{\partial A_{wi}}{\partial A} \right) \left( 1 - \frac{1}{A_{wi}} \right), \quad (20)$$

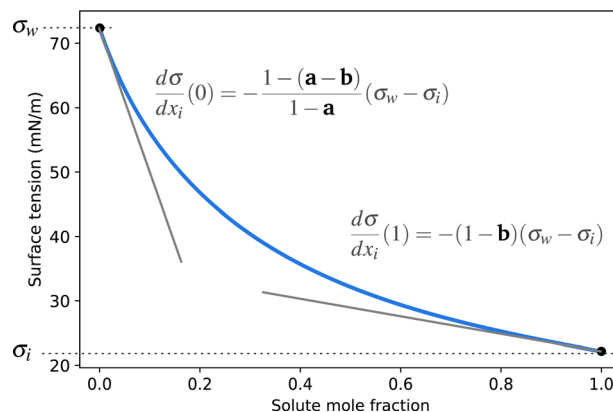


Fig. 1 Illustration of the Connors–Wright function (blue solid line) and the influence of the parameters  $a$  and  $b$  on its slopes.

where  $A_{wi}$  and  $(\partial A_{wi}/\partial A)$  are fit parameters. Even though the derivation of the Chunxi and the Connors–Wright model differ substantially, the final forms of the models are mathematically equivalent, as shown by Shardt and Elliott.<sup>68</sup>

Shardt and Elliott<sup>68</sup> extended the Shereshefsky, Chunxi (referred to as Li *et al.*) and the Connors–Wright model to predict  $\sigma$  as a function of composition and temperature and compared their performance for 15 organic substances in a temperature range from 293 to 323 K. Overall, the Connors–Wright and the Chunxi model performed slightly better than the Shereshefsky model because of their additional fit parameter. Shardt and Elliott<sup>68</sup> recommend to use the Connors–Wright model, since the parameters in the Chunxi model are temperature-dependent while the ones in the Connors–Wright model are not. Shardt *et al.*<sup>70</sup> further extended the Connors–Wright model to multi-component solutions and nonaqueous systems that may contain a supercritical component.

In the field of atmospheric sciences, Hyvarinen *et al.*<sup>72</sup> applied the Chunxi model for aqueous solutions of several dicarboxylic acids and *cis*-pinonic acid. The model was fitted to their own surface tension data at varying composition and temperature. The surface tensions of the supercooled pure acids were estimated with the method of MacLeod–Sugden and their temperature dependence was fitted to a linear equation. Vanhanen *et al.*<sup>73</sup> applied the multi-component version of the Chunxi model to ternary solutions of NaCl, succinic acid and water. Finally, Prisle *et al.*<sup>32</sup> fitted the Chunxi model to experimental  $\sigma$  data of sodium laurate solutions in a sensitivity study to predict the surface partitioning of fatty acid sodium salts during cloud droplet activation.

## 3. Methodology of model comparison

In order to assess the performance of surface tension models proposed in the literature, we compared their ability to fit experimental surface tension data. All models were tested with the same set of test substances, irrespective of their original purpose. To evaluate the surface tension model performance independently of the choice of activity coefficient models, we



used the same activity coefficient model for all models and all substances. Experimental data as well as modelling was focused on ambient temperature in this study ( $20\text{ }^{\circ}\text{C} < T < 25\text{ }^{\circ}\text{C}$ ). Note that models with temperature independent fit parameters allow the application of the fit parameters to any temperature, since the temperature dependence can be represented by  $\sigma_w(T)$  and  $\sigma_i(T)$ , as shown by Shardt and Elliott<sup>68</sup> for the Connors–Wright model. Therefore, many findings of this study should be valid at other temperatures, too. All models were fitted with the `curve_fit()` function in the module `optimize` of the Python package `SciPy`.<sup>88</sup>

### 3.1 Tested models

We evaluated the performance of the models marked with an asterisk in Table 1 as well as one additional model proposed by us (see below). To provide a level playing field, only models were selected that are a function of solute mole fraction or solute activity in the bulk and thus applicable to the entire concentration range. Therefore, the Szyszkowski–Langmuir equation was not analyzed, being only valid for dilute solutions as outlined above. We preferred the Eberhart model over the mathematically equivalent Shereshefsky model, since its fit parameters are dimensionless and therefore temperature independent. For the same reason, we preferred the Connors–Wright model over the Chunxi model. This leaves us with seven models to be tested. Also, all models were expressed in a form to be a direct function of  $\sigma_w$  and  $\sigma_i$ . This implied the reformulation of some models, as described in the following.

The **Li & Lu model** was tested in two versions: the original model using  $a_i$  (non-ideal) and a simplified version, where ideality is assumed by replacing  $a_i$  with  $x_i$  (ideal). Although the Li & Lu model is not a direct function of  $\sigma_i$ , a value for  $\sigma_i$  can be derived by examining the model at  $x_i = 1$  and replacing the parameter  $K$ :

$$\sigma = \sigma_w + RT\Gamma^{\max} \ln \frac{1}{1 + \left( \exp\left(\frac{\sigma_w - \sigma_i}{RT\Gamma^{\max}}\right) - 1 \right) a_i} \quad (21)$$

If  $\sigma_i$  is known, this form of the Li & Lu model has only one fit parameter ( $\Gamma^{\max}$ ).

The **Statistical model** exists in different versions with zero to three fit parameters. Fully predictive or 1-parameter versions were developed only for alcohols and electrolytes so far and thus will not be examined here. The simplified version for very small  $K$  and high  $C$  values (eqn (9)), which applies to surface tension lowering substances (surfactants) is equivalent to the Li & Lu model and is thus covered by this model. Hence, in the following, with “Statistical model” we refer to the full model (eqn (7)). For a better comparison to the other models, here too, we rewrite the model as a function of  $\sigma_i$ , by replacing the parameter  $C$  (eqn (8)). Thus, if  $\sigma_i$  is known, the Statistical model has the two fit parameters  $r$  and  $K$ . The original model, based on its derivation, only allows  $K$  within  $[0,1]$ . In this study, however, we found that for surfactants, restricting  $K$  within  $[0,1]$  leads to fits very similar to the Li & Lu model, while

allowing  $K$  to be negative improved the fits. This is clearly an unphysical usage of the original Statistical model turning it into a mixing rule that lacks thermodynamic background. Due to its improved performance we nevertheless include it in this study and label it Statistical unconstrained or abbreviated Stat. uncon. in the following. We also test the Statistical unconstrained model in an ideal and a non-ideal version.

To reduce the number of tested model versions, we performed a preliminary comparison between three versions of the **Butler equation**:

- Calculating  $A_i$  with eqn (12) (Goldsack and White<sup>84</sup>)
- Calculating  $A_i$  with eqn (13) (Sprow and Prausnitz<sup>56</sup>)
- Making  $A_i$  a fit parameter.

In agreement with Suarez *et al.*,<sup>57</sup> fitting  $A_i$  turned out to be clearly superior to the fully predictive models (see Section 4 in ESI†). Therefore, we selected this option for the model comparison of the Butler equation in an ideal and a non-ideal version.

In the **Tamura model**, the parameter  $q$  represents the number of segments in a solute molecule, *e.g.* the carbon number for fatty acids, and should be limited to whole numbers. Tamura *et al.*<sup>65</sup> suggest values for  $q$  for certain substances, but not for all substances that we intend to model and hence, we use it as a fit parameter. We also allow non-integer numbers for  $q$  here, due to better fit performance. The molar volumes of the solutes were calculated as  $V_i = \bar{M}/\rho_i$  with the molecular weight  $\bar{M}$  and the subcooled liquid density  $\rho_i$ . For organics,  $\rho_i$  was calculated using UManSysProp<sup>89</sup> (method: Girolami, critical properties method: Nannoolal), and for NaCl,  $\rho_i = 2.09\text{ g cm}^{-3}$  was used.<sup>90</sup> The molar volume of water was set to  $V_w = 18\text{ cm}^3\text{ mol}^{-1}$ .

The **Eberhart model** and the **Connors–Wright model** were used as shown in eqn (16) and (18), respectively.

Inspired by the “S”-shape of the surface tension curves on the logarithmic  $x$ -axis scale, we proposed a parametrized sigmoid function (**Sigmoid model**) in addition to the other models. We used the logistic function as a basis:

$$f(z) = \frac{1}{1 + e^{-z}}, \quad (22)$$

which is a scaled version of the hyperbolic tangent, both part of the group of sigmoid functions. We define  $-z = \ln(x_i)$  and introduce four parameters that allow to shift and scale the function, which leads to

$$\sigma(x_i) = a + b \frac{c}{c + x_i^d} \quad (23)$$

Evaluating this function at  $\sigma(x_i = 0)$  and  $\sigma(x_i = 1)$  allows for replacing the parameters  $a$  and  $b$  and introducing  $\sigma_w$  and  $\sigma_i$ , which results in

$$\sigma(x_i) = \sigma_w - (\sigma_w - \sigma_i)(c + 1) \frac{x_i^d}{c + x_i^d} \quad (24)$$

The parameters  $c$  and  $d$  in this equation both influence the position of and the slope at the inflection point  $x_{\text{infl}}$  of the sigmoid function. To differentiate between the position and



slope and access them *via* two independent parameters, we replace  $c$  by a new parameter  $p = \log_{10}(c)/d$ , which results in our final model equation

$$\sigma(x_i) = \sigma_w - (\sigma_w - \sigma_i)(10^{pd} + 1)^{-\frac{x_i^d}{10^{pd} + x_i^d}} \quad (25)$$

Now, the parameter  $p$  determines the position of the inflection point as  $p = \log_{10}(x_{\text{infl}})$ , which allows for a direct comparison of different substances with this parameter. The parameter  $d$  influences the slope at the inflection point. From this slope, the tangent line at the inflection point can be calculated, enabling an estimate of the critical micelle concentration (CMC) as illustrated in Fig. 2. The resulting equation for the estimated CMC is

$$\log_{10}(x_{\text{CMC}}) = \frac{2}{d \ln(10)} + p, \quad (26)$$

and hence, the parameter  $d$  determines the distance of the estimated CMC from the inflection point *via*

$$\log_{10}(x_{\text{CMC}}) - \log_{10}(x_{\text{infl}}) = \frac{2}{d \ln(10)}. \quad (27)$$

### 3.2 Tested substances

Ten different compounds for which surface tension as a function of aqueous solution concentration has been measured were selected in a way to cover a broad range of different curve types. Three types of model substances were chosen: substances that crystallize in bulk volumes and therefore lack experimental surface tension values of the pure solute (NaCl, sucrose, levoglucosan, and glutaric acid), highly miscible solutes with low surface tension (1,2-ethanediol and methanol), and finally, surfactants of different strength (1,6-hexanediol, butyric acid, nonanoic acid, and Triton X-100). Detailed information on the different datasets is given in Table 2. Surface tension curves as a function of mole fraction are shown in Section 1 in the ESI† If the temperature of the measurement is not given in the reference,  $T = 298.15$  K was assumed. For conversion of mole concentrations ( $\text{mol L}^{-1}$ ) or molalities ( $\text{mol kg}^{-1}$ ) to mole fractions, for water and

NaCl, densities of  $\rho_w = 1000 \text{ kg m}^{-3}$  and  $\rho_{\text{NaCl}} = 2090 \text{ kg m}^{-3}$  were used,<sup>90</sup> respectively, and for the organic substances,  $\rho_i$  was calculated with UManSysProp<sup>89</sup> (method: Girolami, critical properties method: Nannoolal).

### 3.3 Calculation of activity coefficients

For the calculation of activity coefficients the “Aerosol Inorganic–Organic Mixtures Functional groups Activity Coefficients” (AIOMFAC) model was used.<sup>110,111</sup> This model combines the Pitzer model<sup>112</sup> and the UNIFAC model<sup>83</sup> and was designed with a focus on substances that are relevant in atmospheric science. It is available at <http://www.aiomfac.caltech.edu/index.html>.

For organic solutions, using the AIOMFAC activity coefficients is equivalent to using UNIFAC. For alcohol groups, the new parametrization by Marcolli and Peter<sup>113</sup> was used. For electrolytes (*e.g.* NaCl), AIOMFAC is based on the Pitzer model. The activity coefficients are ion specific and molality based with the 1-molal solution as the reference state. Hence, for the calculation of  $a_{\text{NaCl}}$ , the mole fraction  $x_{\text{NaCl}}$  was converted to molality and multiplied by the mean activity coefficient  $\gamma_{\text{NaCl}} = \gamma_{\text{Na}^+} = \gamma_{\text{Cl}^-}$ .<sup>110</sup> The so-derived  $a_{\text{NaCl}}$  is not bound to  $[0,1]$ . As a result, small modifications of the Li & Lu model and the Statistical model were necessary to fulfill the condition of  $\sigma_i = \sigma(x_i = 1)$ , while the Butler equation did not require any modifications (see Section 5 in ESI†). Due to the combination of the UNIFAC and the Pitzer model in AIOMFAC, the results in this study are valid, too, when activities are calculated with the UNIFAC model for organic solutes and the Pitzer model for inorganic solutes.

With AIOMFAC-based equilibrium calculations, liquid–liquid phase separation (LLPS) can also be predicted.<sup>114,115</sup> For the solutions examined in this study, LLPS was predicted for strong surfactants, *i.e.* nonanoic acid and Triton X-100, for a certain concentration range. Substances with a strong amphiphilic structure are known to form structures of high surfactant content—known as micelles—above a certain concentration in the solution which is known as the critical micelle concentration (CMC). Since micelle formation can be regarded as a form of phase separation, we assumed that the onset of LLPS in AIOMFAC-based equilibrium calculations corresponds to the formation of micelles and thus LLPS was explicitly allowed when calculating the activity coefficients. Using activity coefficients of the phase separated system also led to much better fit performance than applying activities calculated for a one-phase system (*i.e.* suppressing LLPS). In the case of LLPS, it was assumed that the surfactant-rich phase determines the surface tension of the gas–liquid interface, based on its lower surface tension. As interfacial energies between the two liquid phases are of minor relevance in large droplets and bulk systems, they were neglected. Thus, the surface tension was calculated for the composition of the solute rich (outer) phase, which for nonanoic acid and Triton X-100 was the almost pure surfactant. The result of this approach is presented and discussed in Subsection 4.3.

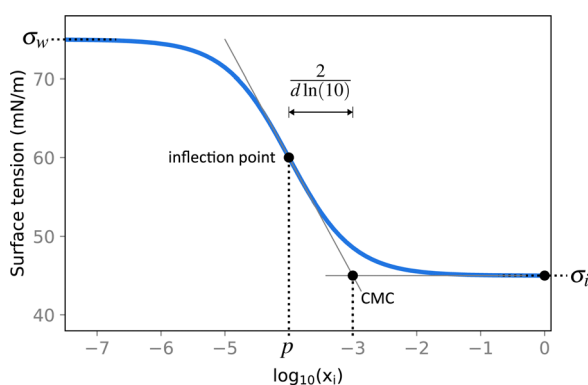
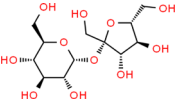
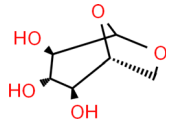
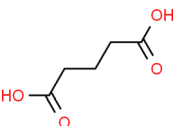
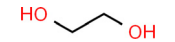
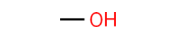
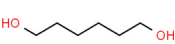
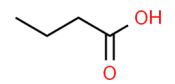
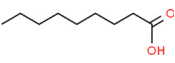
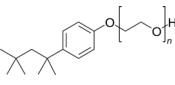


Fig. 2 Illustration of the Sigmoid model (blue solid line, eqn (25)) with parameters  $\sigma_w = 75 \text{ mN m}^{-1}$ ,  $\sigma_i = 45 \text{ mN m}^{-1}$ ,  $p = -4$ , and  $d = 0.869$ . The critical micelle concentration “CMC” is estimated by intersecting the tangent line at the inflection point with a horizontal line at  $\sigma(x_i \rightarrow \infty)$ .



**Table 2** Tested substances, experimental surface tension data sources and the surface tension of pure water  $\sigma_w$  that was used.  $\sigma_w$  was calculated with IAPWS-IF97<sup>91</sup> for the average of the minimum and maximum temperature of the experimental data sets. Under uncertainty, either the standard deviation  $S$  or the accuracy  $A$  as reported in the literature is given

| Name           | Structure                                                                           | Experimental surface tension data |          |                                           |                                                        | $\sigma_w$<br>(mN m <sup>-1</sup> ) |
|----------------|-------------------------------------------------------------------------------------|-----------------------------------|----------|-------------------------------------------|--------------------------------------------------------|-------------------------------------|
|                |                                                                                     | Type of measurement               | $T$ (°C) | Ref.                                      | Uncertainty                                            |                                     |
| NaCl           | $\text{Na}^+ \text{Cl}^-$                                                           | Wilhelmy plate                    | 25 °C    | Aumann <i>et al.</i> <sup>17</sup>        | —                                                      | 72.36                               |
|                |                                                                                     | Holographic optical tweezer       | —        | Boyer <i>et al.</i> <sup>53</sup>         | $S < 0.03 \text{ mN m}^{-1}$                           |                                     |
|                |                                                                                     | Sessile bubble tensiometer        | 23       | Ozdemir <i>et al.</i> <sup>92</sup>       | $S < 0.09 \text{ mN m}^{-1}$                           |                                     |
|                |                                                                                     | Nouy ring tensiometer             | 20       | Tuckermann <sup>42</sup>                  | —                                                      |                                     |
|                |                                                                                     | Capillary rise method             | 25.2     | Vanhanen <i>et al.</i> <sup>73</sup>      | —                                                      |                                     |
| Sucrose        |    | Wilhelmy plate                    | 25       | Aumann <i>et al.</i> <sup>17</sup>        | $S = 0.5\text{--}1\%$                                  | 71.97                               |
|                |                                                                                     | Various                           | 25       | Washburn <i>et al.</i> <sup>93</sup>      | $A < 0.5 \text{ mN m}^{-1}$                            |                                     |
| Levogluconan   |    | Wilhelmy plate                    | 25       | Aumann <i>et al.</i> <sup>17</sup>        | $S = 0.5\text{--}1\%$                                  | 72.36                               |
|                |                                                                                     | Pendant drop                      | —        | Topping <i>et al.</i> <sup>44</sup>       | —                                                      |                                     |
|                |                                                                                     | Wilhelmy plate                    | 20       | Tuckermann and Cammenga <sup>94</sup>     | —                                                      |                                     |
| Glutaric acid  |    | Pendant drop                      | —        | Topping <i>et al.</i> <sup>44</sup>       | —                                                      | 72.36                               |
|                |                                                                                     | Wilhelmy plate                    | 20       | Lee and Hildemann <sup>95</sup>           | $S < 0.3 \text{ mN m}^{-1}$                            |                                     |
|                |                                                                                     | Pendant drop                      | 21       | Booth <i>et al.</i> <sup>45</sup>         | —                                                      |                                     |
|                |                                                                                     | Wilhelmy plate                    | 25       | Aumann <i>et al.</i> <sup>17</sup>        | $S = 0.5\text{--}1\%$                                  |                                     |
|                |                                                                                     | Wilhelmy plate                    | 24.85    | Boyer and Dutcher <sup>78</sup>           | $S < 0.03 \text{ mN m}^{-1}$                           |                                     |
|                |                                                                                     | Holographic optical tweezer       | —        | Boyer <i>et al.</i> <sup>53</sup>         | $A = 1 \text{ mN m}^{-1}$                              |                                     |
|                |                                                                                     | Holographic optical tweezer       | —        | Bzdek <i>et al.</i> <sup>96</sup>         | —                                                      |                                     |
| 1,2-ethanediol |   | Wilhelmy plate                    | —        | Bzdek <i>et al.</i> <sup>96</sup>         | —                                                      | 71.97                               |
|                |                                                                                     | Pendant drop                      | Ambient  | Varga <i>et al.</i> <sup>97</sup>         | —                                                      |                                     |
|                |                                                                                     | Bubble pressure tensiometer       | 25       | Messow <i>et al.</i> <sup>98</sup>        | —                                                      |                                     |
|                |                                                                                     | Nouy ring tensiometer             | 25       | Basařova <i>et al.</i> <sup>99</sup>      | $S < 0.2 \text{ mN m}^{-1}$ ,                          |                                     |
|                |                                                                                     | Wilhelmy plate                    | 19.85    | Gliński <i>et al.</i> <sup>100</sup>      | $A < 0.15 \text{ mN m}^{-1}$                           |                                     |
| Methanol       |  | Drop volume tensiometer           | —        | Maximino <sup>101</sup>                   | $A = 0.4 \text{ mN m}^{-1}$                            | 72.37                               |
|                |                                                                                     | Bubble pressure tensiometer       | 25       | Semenov and Pokrovskaya <sup>102</sup>    | $S < 1\%$                                              |                                     |
|                |                                                                                     | —                                 | —        | —                                         | —                                                      |                                     |
| 1,6-hexanediol |  | Capillary rise method             | 25       | Romero <i>et al.</i> <sup>103</sup>       | $S < 0.3 \text{ mN m}^{-1}$                            | 71.97                               |
| Butyric acid   |  | Drop volume tensiometer           | 25       | Granados <i>et al.</i> <sup>104</sup>     | $S < 0.03 \text{ mN m}^{-1}$                           | 71.98                               |
|                |                                                                                     | Pendant drop                      | —        | Suárez and Romero <sup>105</sup>          | $S = 0.01 \text{ mN m}^{-1}$                           |                                     |
|                |                                                                                     | Capillary rise method             | 24.85    | Donaldson and Anderson <sup>106</sup>     | —                                                      |                                     |
| Nonanoic acid  |  | Nouy ring tensiometer             | 21.85    | Lunkenheimer <i>et al.</i> <sup>107</sup> | —                                                      | 72.46                               |
|                |                                                                                     | Bubble pressure tensiometer       | —        | Badban <i>et al.</i> <sup>108</sup>       | —                                                      |                                     |
| Triton X-100   |  | Nouy ring tensiometer             | 19.85    | Zdziennicka <i>et al.</i> <sup>109</sup>  | $A = 0.3\text{--}0.7\%$<br>$S < 0.2 \text{ mN m}^{-1}$ | 72.76                               |

## 4 Results and discussion

The resulting fit parameters for all substances and all models are shown in Table 3. In this table, for each fit the pure solute surface tension  $\sigma_i$ , which is determined either by fitting, taken from the literature, or set to the experimental value is also listed, since the other fit parameters are specific for this  $\sigma_i$  value. The corresponding RMSE values are summarized in Table 4. The results are presented and discussed by dividing the substances into the three groups: moderately surface active

substances (1,2-ethanediol, methanol), substances with unknown  $\sigma_i$  (NaCl, levoglucosan, sucrose, glutaric acid) and intermediate to strong surface active substances (1,6-hexanediol, butyric acid, nonanoic acid, Triton X-100).

### 4.1 Moderately surface active substances

As pure liquids, 1,2-ethanediol and methanol both have a surface tension  $\sigma_i$  that is substantially lower than that of water. Consequently, in an aqueous solution with these compounds,



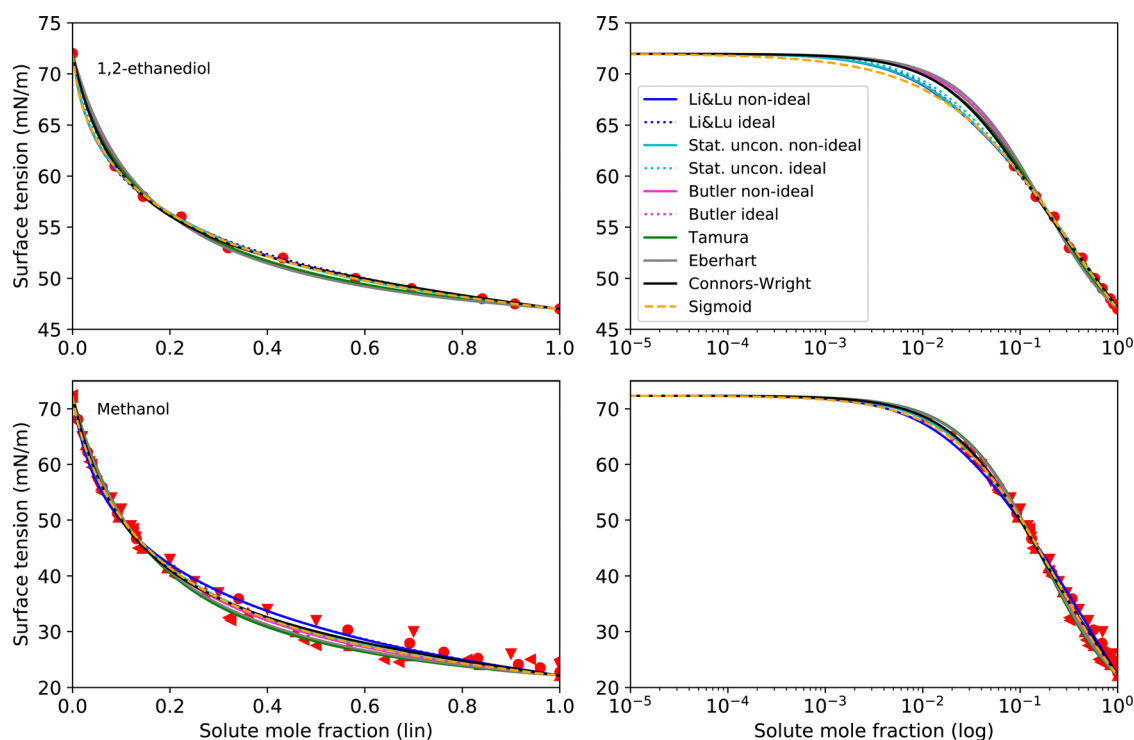


**Table 3** Fit parameters including the pure solute surface tension  $\sigma_i$  for all tested substances and models.  $\sigma_i$  was either taken as the lowest datapoint at the highest concentration measured (same value for all models), from extrapolation of the temperature function reported by Janz<sup>116</sup> (denoted by the superscript 'Janz'), or it was made a fit parameter (denoted by the superscript 'Fit'). The name of some substances was abbreviated. For full names see Table 2

|                                                    | Li & Lu                          |                                             | Stat. uncon.                     |           | Butler                                       |            | Tamura                                 |                                        | Eberhart                        |                                   | Connors–Wright                   |          | Sigmoid                         |        |
|----------------------------------------------------|----------------------------------|---------------------------------------------|----------------------------------|-----------|----------------------------------------------|------------|----------------------------------------|----------------------------------------|---------------------------------|-----------------------------------|----------------------------------|----------|---------------------------------|--------|
|                                                    | non-ideal                        | ideal                                       | $r$                              | $K$       | $r$                                          | $K$        | $A_i$ ( $\text{m}^2 \text{mol}^{-1}$ ) | $A_i$ ( $\text{m}^2 \text{mol}^{-1}$ ) | $q$                             | $S$                               | $a$                              | $b$      | $p$                             | $d$    |
| NaCl<br>$\sigma_i$ (mN $\text{m}^{-1}$ )           | $-3.784$<br>$1821^{\text{Fit}}$  | $-1.76 \times 10^7$<br>$156.4^{\text{Fit}}$ | $-0.013$<br>$97.42^{\text{Fit}}$ | $-0.084$  | $9.25 \times 10^{-5}$<br>$4475^{\text{Fit}}$ | $0.983$    | $6363$<br>$169.73^{\text{Janz}}$       | $7024$<br>$169.73^{\text{Janz}}$       | $3.50$<br>$211.06^{\text{Fit}}$ | $0.848$<br>$169.73^{\text{Janz}}$ | $0.934$<br>$175.28^{\text{Fit}}$ | $-0.031$ | $1.08$<br>$187.88^{\text{Fit}}$ | $1.16$ |
| Sucrose<br>$\sigma_i$ (mN $\text{m}^{-1}$ )        | $-0.588$<br>$85.45^{\text{Fit}}$ | $-2.357$<br>$87.54^{\text{Fit}}$            | $-28.25$<br>$99.54^{\text{Fit}}$ | $0.99994$ | $-34.00$<br>$98.48^{\text{Fit}}$             | $0.999999$ | $63637$<br>$81.80^{\text{Fit}}$        | $57027$<br>$82.90^{\text{Fit}}$        | $6.24$<br>$80.08^{\text{Fit}}$  | $6.613$<br>$83.58^{\text{Fit}}$   | $0.993$<br>$121.13^{\text{Fit}}$ | $0.0113$ | $1.67$<br>$105.25^{\text{Fit}}$ | $0.82$ |
| Levoglucozan<br>$\sigma_i$ (mN $\text{m}^{-1}$ )   | $3.769$<br>$45.67^{\text{Fit}}$  | $0.417$<br>$66.97^{\text{Fit}}$             | $0.066$<br>$56.37^{\text{Fit}}$  | $-8.865$  | $0.0973$<br>$69.29^{\text{Fit}}$             | $-58.90$   | $306433$<br>$69.08^{\text{Fit}}$       | $335696$<br>$69.11^{\text{Fit}}$       | $4.34$<br>$66.86^{\text{Fit}}$  | $60.13$<br>$69.29^{\text{Fit}}$   | $0.983$<br>$69.34^{\text{Fit}}$  | $1.0$    | $-2.03$<br>$70.04^{\text{Fit}}$ | $1.84$ |
| Glutaric<br>$\sigma_i$ (mN $\text{m}^{-1}$ )       | $2.304$<br>$41.35^{\text{Fit}}$  | $1.602$<br>$46.25^{\text{Fit}}$             | $6.707$<br>$44.74^{\text{Fit}}$  | $-1.256$  | $9.567$<br>$49.18^{\text{Fit}}$              | $-1.750$   | $217162$<br>$51.72^{\text{Fit}}$       | $240178$<br>$51.71^{\text{Fit}}$       | $3.94$<br>$48.23^{\text{Fit}}$  | $73.02$<br>$53.56^{\text{Fit}}$   | $0.991$<br>$31.85^{\text{Fit}}$  | $0.377$  | $-1.57$<br>$50.26^{\text{Fit}}$ | $0.67$ |
| Ethanediol<br>$\sigma_i = 47.00 \text{ mN m}^{-1}$ | $2.259$                          | $2.421$                                     | $7.189$                          | $-0.0436$ | $6.034$                                      | $-0.245$   | $44924$                                | $44624$                                | $1.96$                          | $6.894$                           | $0.907$                          | $0.747$  | $-0.51$                         | $0.66$ |
| Methanol<br>$\sigma_i = 22.14 \text{ mN m}^{-1}$   | $5.672$                          | $5.259$                                     | $1.880$                          | $-0.898$  | $2.199$                                      | $-0.800$   | $38529$                                | $38938$                                | $0.946$                         | $6.776$                           | $0.890$                          | $0.762$  | $-0.65$                         | $0.83$ |
| Hexanediol<br>$\sigma_i = 42.60 \text{ mN m}^{-1}$ | $1.257$                          | $0.800$                                     | $0.00056$                        | $-49.08$  | $3.034$                                      | $-115.8$   | $401369$                               | $525173$                               | $7.38$                          | $335.0$                           | $0.99698$                        | $1.0$    | $-2.53$                         | $0.94$ |
| Butyric<br>$\sigma_i = 26.51 \text{ mN m}^{-1}$    | $2.702$                          | $1.480$                                     | $1.5 \times 10^{-7}$             | $-19.23$  | $3.4 \times 10^{-5}$                         | $-214.1$   | $217095$                               | $332118$                               | $4.35$                          | $215.1$                           | $0.99528$                        | $1.0$    | $-2.30$                         | $1.21$ |
| Nonanoic<br>$\sigma_i = 33.01 \text{ mN m}^{-1}$   | $1.770$                          | $0.673$                                     | $4.7 \times 10^{-5}$             | $-116.5$  | $5.4 \times 10^{-5}$                         | $-264239$  | $512197$                               | $1139169$                              | $10.6$                          | $264242$                          | $0.999996$                       | $1.0$    | $-5.38$                         | $1.69$ |
| TritonX100<br>$\sigma_i = 33.80 \text{ mN m}^{-1}$ | $0.874$                          | $0.4503$                                    | $9.7 \times 10^{-6}$             | $-99.04$  | $3.593$                                      | $-615468$  | $2.7 \times 10^8$                      | $1968337$                              | $18.0$                          | $3153538$                         | $0.9999997$                      | $1.0$    | $-6.52$                         | $0.87$ |

**Table 4** Root mean squared errors (RMSE) of the surface tension fits reported in Table 3 in  $\text{mN m}^{-1}$ . The cells are colored with a linear gradient from white ( $1 \text{ mN m}^{-1}$ ) to red ( $10 \text{ mN m}^{-1}$ )

|                | Li&Lu<br>non-ideal | Li&Lu<br>ideal | Stat.uncon.<br>non-ideal | Stat.uncon.<br>ideal | Butler<br>non-ideal | Butler<br>ideal | Tamura | Eberhart | Connors-<br>Wright | Sigmoid |
|----------------|--------------------|----------------|--------------------------|----------------------|---------------------|-----------------|--------|----------|--------------------|---------|
| NaCl           | 0.73               | 0.79           | 0.73                     | 0.73                 | 0.77                | 0.78            | 0.72   | 0.78     | 0.72               | 0.72    |
| Sucrose        | 0.12               | 0.10           | 0.12                     | 0.37                 | 0.11                | 0.10            | 0.12   | 0.10     | 0.10               | 0.09    |
| Levogluconan   | 0.59               | 0.59           | 0.59                     | 0.59                 | 0.59                | 0.59            | 0.59   | 0.59     | 0.59               | 0.58    |
| Glutaric acid  | 1.38               | 1.52           | 1.35                     | 1.35                 | 2.37                | 2.41            | 2.06   | 2.89     | 1.39               | 1.35    |
| 1,2-ethanediol | 0.22               | 0.25           | 0.22                     | 0.21                 | 0.47                | 0.47            | 0.38   | 0.57     | 0.24               | 0.22    |
| Methanol       | 1.59               | 1.58           | 1.38                     | 1.37                 | 1.41                | 1.41            | 1.74   | 1.61     | 1.36               | 1.38    |
| 1,6-hexanediol | 3.66               | 3.97           | 1.26                     | 0.63                 | 1.76                | 2.35            | 4.11   | 0.71     | 0.70               | 0.66    |
| Butyric acid   | 5.02               | 6.42           | 2.40                     | 1.61                 | 2.37                | 4.44            | 5.26   | 1.61     | 1.59               | 1.31    |
| Nonanoic acid  | 8.68               | 12.11          | 3.97                     | 4.04                 | 8.09                | 11.72           | 11.56  | 4.04     | 4.04               | 1.86    |
| Triton X-100   | 18.57              | 11.29          | 17.26                    | 0.90                 | 19.59               | 10.12           | 11.75  | 1.32     | 1.32               | 1.02    |
| average        | 2.44               | 3.04           | 1.34                     | 1.18                 | 1.99                | 3.44            | 3.83   | 1.42     | 1.20               | 0.92    |



**Fig. 3** Surface tension fits (lines) and experimental data (red symbols) for 1,2-ethanediol and methanol on linear (left) and logarithmic scales (right). Different symbols of the experimental data refer to different sources (see Table 2 and Fig. S2 in ESI†).

the surface tension is lower than  $\sigma_w$ , but the deviation from a linear mixing rule is not very large. Thus, the preference of these substances to partition to the surface of the solution is only moderate. The continuously decreasing surface tension with increasing solute mole fraction of these moderately surface active substances is reproduced well by all models (see Fig. 3) resulting in low RMSE values ( $<2 \text{ mN m}^{-1}$ ). The higher RMSE values for methanol than for 1,2-ethanediol are the result of the larger scatter of its experimental data. The ideal and non-ideal versions of the models using activity coefficients lead to very similar RMSE values, since 1,2-ethanediol and methanol mix almost ideally with water. This result suggests that the benefit of activity coefficients is not clear for these substances, and the additional effort of calculating them is not justified.

## 4.2 Substances with unknown $\sigma_i$

For NaCl, sucrose, levoglucosan, and glutaric acid, the surface tension of the pure liquid substance  $\sigma_i$  is not known, because these substances crystallize in the bulk at room temperature. Yet, the surface tension models we examine in this study are, directly or indirectly, a function of  $\sigma_i$ . While for bulk systems, the modelling can be restricted to concentrations below the solubility limit, for small droplets like aerosol particles, modelling beyond the solubility limit is required as these particles can strongly supersaturate. Smooth water uptake and loss of levitated levoglucosan particles during humidity cycles suggests that they do not crystallize at all and remain liquid at room temperature.<sup>117</sup> Therefore,  $\sigma_i$  of levoglucosan should be considered a real quantity, but one that has not yet been measured.



In contrast, glutaric acid and NaCl have been observed to eventually crystallize even in micrometer-sized droplets<sup>118,119</sup> and sucrose transforms into a glass<sup>120</sup> at high concentrations. For these three substances,  $\sigma_i$  can be considered a hypothetical quantity and thus, priority should be laid on accurately modelling the range of concentrations that can be realized. Moreover, in semi-empirical modelling, other fit parameters may compensate for differences in the choice of  $\sigma_i$  thereby allowing good fit performance independent of the exact choice of  $\sigma_i$ . Due to this dependence of the fit parameters on the choice of  $\sigma_i$ , especially in cases where  $\sigma_i$  is not well constrained, its value employed in the surface tension parameterization needs to be reported together with the other fit parameters to ensure reproducibility in modelling surface tensions by other groups. Note that Topping *et al.*<sup>44</sup> found a high sensitivity in the choice of  $\sigma_i$  when they used predictive models, because in this case, the model has no fit parameters to ensure the reproduction of the data.

The different options to obtain a value for  $\sigma_i$  include:

- (i) Extrapolate from concentration-dependent measurements with a thermodynamic model (*i.e.* by fitting  $\sigma_i$ ).
- (ii) Resort to measurements of  $\sigma_i$  at a higher temperature where the substance is liquid and extrapolate to the desired temperature.
- (iii) Use prediction tools like the Macleod–Sugden correlation or the corresponding states correlation (see Poling *et al.*<sup>59</sup>) based on physical properties like density, boiling point, and critical molar volume.
- (iv) Use group contribution approaches and artificial neural network (ANN) models fed with experimental data from similar substances (derivation from other substances).

Making  $\sigma_i$  a fit parameter (option i) leads to the best RMSE values, due to having one additional adjustable parameter. Therefore, here we present surface tension fits with all tested models and for all tested substances, that use  $\sigma_i$  as a fit parameter with two exceptions: For the Eberhart model and the Butler equation, fitting NaCl with unknown  $\sigma_i$  was unsuccessful due to numerical issues (too many free parameters). In this case,  $\sigma_i$  was taken from an extrapolation of the temperature function by Janz<sup>116</sup> (option ii), which is based on molten salt measurements. Extrapolation of this function gives a value of  $\sigma_{\text{NaCl}} = 169.73 \text{ mN m}^{-1}$  at  $T = 25^\circ\text{C}$ , which is frequently used for surface tension parameterizations of NaCl solutions.<sup>18,26,73,121,122</sup> The results following this approach for NaCl, sucrose, levoglucosan and glutaric acid are shown in Fig. 4 and the corresponding parameters in Table 3.

As can be seen from Fig. 4, fitting  $\sigma_i$  with the various models results in a large range of  $\sigma_i$  values. Therefore, when the surface tension of solutions in the supersaturated concentration range is of interest, a better constraint of  $\sigma_i$  is desirable. For NaCl, an alternative to fitting  $\sigma_i$  is given by temperature extrapolation following Janz,<sup>116</sup> as mentioned above.

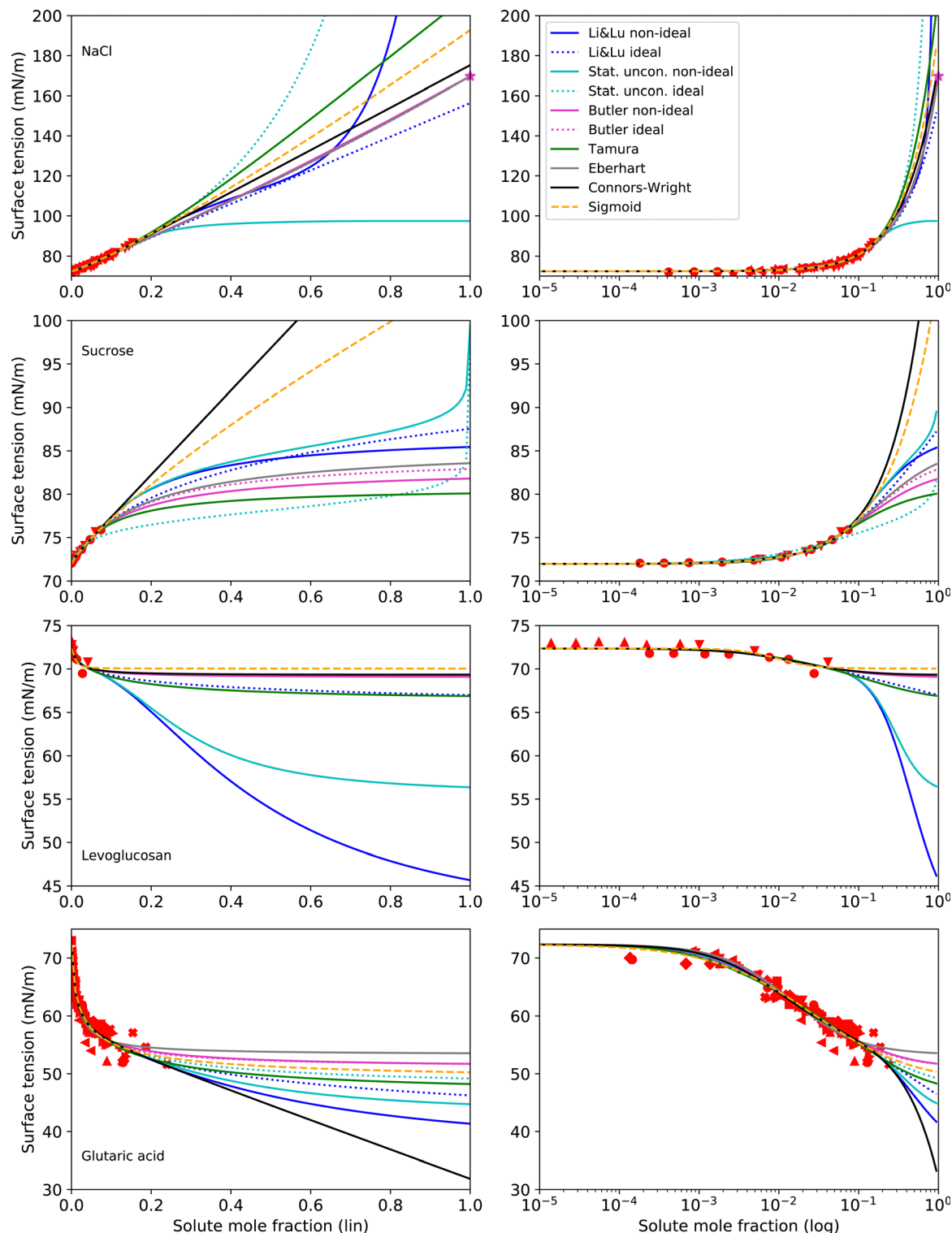
For sucrose and levoglucosan, no data is available for an extrapolation from higher temperature (ii), and sucrose even decomposes before melting. Prediction tools (iii) also fail because physical properties like boiling point and critical molar

volume data cannot be determined. In this case, only group contribution approaches or ANN models can be used (iv), either to predict the lacking quantities for option (iii) or to directly predict  $\sigma_i$ . However, no model to directly predict  $\sigma_i$  was available in the literature for sugars and polyols, evidencing a lack of research in this field. For levoglucosan, a value of  $\sigma_{\text{levoglucosan}} = 22.71 \text{ mN m}^{-1}$  calculated with Macleod–Sugden's method and Yens–Wood densities could be found,<sup>44</sup> which seems too low in comparison with molecules of similar structures like sugars.

For glutaric acid, several predictions of its pure liquid surface tension  $\sigma_{\text{glutaric}}$  can be found in the literature. Mulero *et al.*<sup>123</sup> carefully screened surface tension data of pure organic acids and suggest functions for the temperature dependence for the NIST REFPROP program.<sup>124</sup> The data thereby is mostly based on predictions with the Macleod–Sugden method (iii) at higher temperature, where the acids are liquid. An extrapolation to room temperature (ii) results in  $\sigma_{\text{glutaric}} = 43.22 \text{ mN m}^{-1}$  at  $T = 25^\circ\text{C}$ . Using the Macleod–Sugden method combined with the predictive Yens–Wood method for the calculation of sub-cooled liquid densities, Topping *et al.*<sup>44</sup> reported a value of  $\sigma_{\text{glutaric}} = 38.88 \text{ mN m}^{-1}$  at  $T = 25^\circ\text{C}$ . The same authors report a value of  $\sigma_{\text{glutaric}} = 56.1 \text{ mN m}^{-1}$  predicted by ACDlabs Chemschetch 5.0. This spread highlights the uncertainties related to  $\sigma_i$  for organic acids and the need for further research.

Since for sucrose, levoglucosan and glutaric acid, no good prediction of  $\sigma_i$  could be obtained by options (ii) to (iv), the question arises if option (i) can be used to estimate  $\sigma_i$ . The large spread in fitted  $\sigma_i$  values in Fig. 4 suggests that some models are less physically constrained and simply follow the trend of the data, while others seem to be more robust and less prone to predict unphysical  $\sigma_i$  values. Using the data from methanol and butyric acid, we tested the capability of the models to extrapolate to high concentrations by fitting the models to only a part of the experimental data (Section 6 in the ESI†). Comparing the thereby predicted  $\sigma_i$  with the actual  $\sigma_i$  of these two substances showed that the Butler equation seems to be a model that is relatively robust in extrapolating the surface tension even from very limited experimental data. However, the Butler equation seems to be strongly dependent on the activity coefficient model that is being used and only precise substance-specific activity coefficients would allow to actually examine the predictive capabilities of the Butler equation. From the extrapolation test and the fits for sucrose and levoglucosan in Fig. 4, it can also be seen that if the data are limited to too narrow of a concentration range, the uncertainty in the predicted  $\sigma_i$  values becomes very large for all models. A solution to this problem could be to take measurements of the surface tension at a higher temperature or to change to a solvent that allows for a higher solubility such that experimental data over a wider concentration range can be obtained, and thus providing a better prediction of  $\sigma_i$ . From the current study, we cannot yet recommend a final methodology to obtain precise surface tension fits for the supersaturated concentration range of crystallizing substances, but we suggest to test the aforementioned approaches in future studies.





**Fig. 4** Surface tension fits (lines) and experimental data (red symbols) for substances with unknown  $\sigma_i$  on linear (left) and logarithmic (right) x-axis scales. All models use  $\sigma_i$  as a fit parameter except for NaCl, where the Eberhart model and the Butler equation use  $\sigma_i = 169.73 \text{ mN m}^{-1}$  (purple star), as suggested by a temperature extrapolation from molten salt measurements.<sup>116</sup> No values for  $\sigma_i$  of sucrose could be found in the literature. For levoglucosan, a prediction with the Macleod–Sugden correlation and Yens–Wood densities suggests a value of  $\sigma_i = 22.71 \text{ mN m}^{-1}$ .<sup>44</sup> Values for  $\sigma_i$  of glutaric acid reported in the literature include  $43.22 \text{ mN m}^{-1}$  (NIST, temperature extrapolation),<sup>123</sup>  $38.88 \text{ mN m}^{-1}$  (Macleod–Sugden and Yens–Wood) and  $56.1 \text{ mN m}^{-1}$  (ACDlabs Chemschetch 5.0).<sup>44</sup> Different symbols of the experimental data refer to different sources (see Table 2 and Fig. S2 in ESI†). Note that for NaCl and glutaric acid, the curve fits of the non-ideal and ideal versions of the Butler equation completely overlay. For levoglucosan, the curve fits of the models Stat. uncon. ideal, Butler ideal, Eberhart and Connors–Wright completely overlay.



### 4.3 Intermediate to strong surfactants

The surface tension curves for aqueous solutions of 1,6-hexanediol, butyric acid, nonanoic acid and Triton X-100 are characterized by a strong decrease in surface tension in a narrow, low concentration range and a plateauing of the surface tension at higher concentrations, as is characteristic for substances with strong surface partitioning. The concentration at which the surface tension curves change from a negative slope to a plateau at low surface tension marks the concentration at which the surface is fully covered with solute molecules and where for strongly amphiphilic molecules the formation of micelles in the bulk starts (CMC). While for 1,6-hexanediol and butyric acid, the CMC lies at  $x_{\text{CMC}} \approx 10^{-2}$  to  $10^{-1}$ , for nonanoic acid and Triton X-100 it is at a substantially lower concentration ( $x_{\text{CMC}} \approx 1 \times 10^{-5}$ ), which is why we categorize the former as “intermediate surfactants” and the latter as “strong surfactants”.

As can be seen in Fig. 5, the Li & Lu models, the Butler equation (ideal) and the Tamura model were not able to reproduce the steep surface tension decrease at low concentration for intermediate and even less so for strong surfactants. In contrast, with the Stat. uncon. ideal, Eberhart, Connors–Wright and Sigmoid models, the shape of the experimental data could be reproduced closely for both the intermediate and strong surfactants.

The performance of the models using activity coefficients (Butler, Stat. uncon., Li & Lu, all non-ideal) was found to be highly dependent on the activity coefficients and LLPS prediction by AIOMFAC. As can be seen in the fits for nonanoic acid in Fig. 5, the LLPS from  $x_i = 2.8 \times 10^{-4}$  to  $x_i = 0.613$  (grey-shaded area) is responsible for the abrupt changes in  $\sigma$  at the left and right end of the LLPS region and the plateau in between for these models. The Butler equation (non-ideal) has an additional kink at  $x_i \approx 10^{-7}$ . To understand this distinctive curve shape, the reader is reminded that in the Butler theory, the liquid phase is divided into a bulk phase and a surface phase. At the kink at  $x_i \approx 10^{-7}$ , the composition of the surface phase  $x_i^{\text{surf}}$ , which is restricted—same as the bulk phase—to compositions that do not phase separate, changes abruptly from surfactant-poor ( $x_i^{\text{surf}} = 2.8 \times 10^{-4}$ ) to surfactant-rich ( $x_i^{\text{surf}} = 0.613$ ). In the subsequent decrease in  $\sigma$ , the surface phase further enriches in solute, until, at  $x_i = 0.613$ , the bulk eventually phase separates. At this point, the surfactant-rich (bulk) phase determines the surface tension. The resulting step-like surface tension fit of the non-ideal Butler equation nicely reproduces the steep  $\sigma$  decrease and the abrupt plateauing of the nonanoic acid data, but only a close agreement of the actual CMC and the predicted LLPS onset would ensure a good fit. In the case of nonanoic acid, the  $x_i$  of LLPS onset predicted by AIOMFAC is slightly too high. Since AIOMFAC is a group contribution model, the activity and LLPS predictions for a single substance cannot be expected to match the data exactly.

While for nonanoic acid, AIOMFAC predicts the LLPS onset at a higher mole fraction than the actual CMC, for Triton X-100, the opposite is the case. For Triton X-100, AIOMFAC predicts

LLPS from  $x_i = 2 \times 10^{-7}$  to  $x_i = 0.192$ , while the CMC is found at  $x_{\text{CMC}} \approx 10^{-5}$ . As a result, no proper fits could be made with the models using non-ideality. A solution to this problem could be to introduce an additional fit parameter that serves to correct the AIOMFAC activity coefficients for the substance of concern in order to match the onset of LLPS with the CMC.

While non-ideal and ideal model versions achieved similar RMSE values for 1,2-ethanediol and methanol, the non-ideal versions of the Li & Lu model and the Butler equation indeed improved the fit performance for intermediate and strong surfactants, since for these substances non-ideality is much more pronounced. This is not the case for the Statistical unconstrained model, probably due to the loss of its physical basis, since we used the parameters in an unphysical way (*i.e.* allowing  $K$  to be negative). The reader is reminded that a physically correct application of the Statistical model for surface active substances leads to the same results as the Li & Lu model.

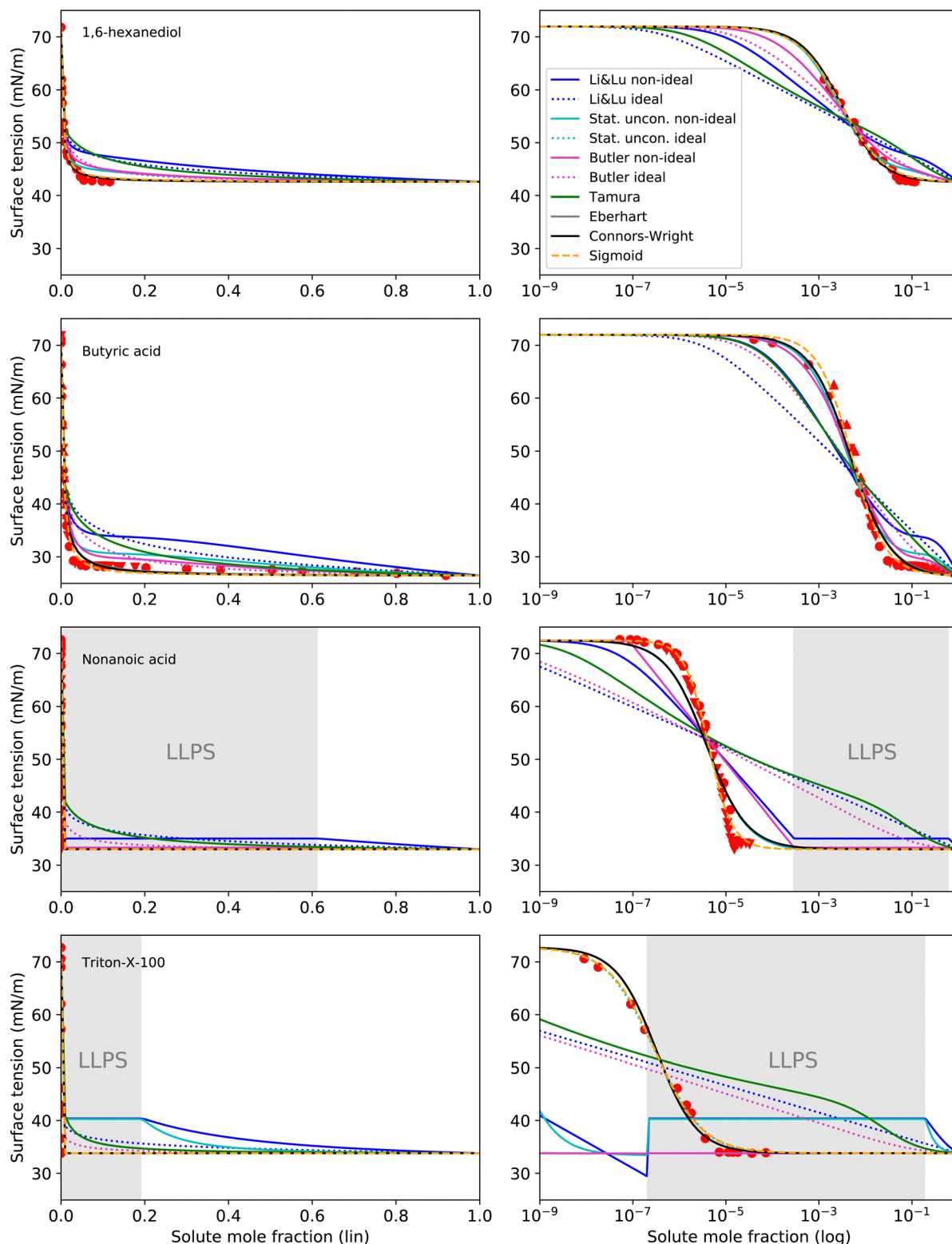
### 4.4 Strengths and weaknesses of the models

Finally, we discuss the strengths and weaknesses of the models with regard to programming and computational effort, meaning and precision of fit parameters, predictive capabilities, application to multi-component solutions, extrapolation, and derivation of the surface composition.

All models tested in this paper are mathematically rather simple equations and thus not very computationally expensive and numerically very stable. The time to write the code for fitting the models and to debug it was mostly short and the resulting code not very long. The Butler equation is the only model that raised some numerical difficulties and took a considerable amount of time to be coded, debugged and executed for the fitting to experimental data. The reasons are that (i) a system of equations needs to be solved instead of an analytical expression, (ii) the calculation of activity coefficients is required, and (iii) activity values too close to zero must be detected by the code with an appropriate threshold to avoid errors from the logarithmic function trying to solve for zero.

The Eberhart, Connors–Wright, Stat. uncon. ideal and Sigmoid models all fit the surface tension data of all studied substances accurately. The versatility of these models ensures that a single framework of fit parameters can be chosen to report the effect of different solutes on surface tension. All solutes can then be compared quantitatively to each other within that framework to lend insight into, for example, which are more surface active. To facilitate a comparison, the fit parameters should be easy to interpret and convenient to report. The fit parameters of the Eberhart and Connors–Wright models require mathematical manipulation to describe their influence on the shape of the surface tension curve (*e.g.*, Fig. 1). Additionally, strong surfactants like Triton-X-100 bring these models to the limit of their fitting capacity as manifested by the many significant digits that must be reported for the fit parameters to describe the data accurately (*e.g.* for the Connors–Wright model,  $a = 0.9999997$ ). Alternatively, the Sigmoid model has tangible fit parameters (the concentration at the inflection point and the CMC), and only a few significant digits are





**Fig. 5** Surface tension fits (lines) and experimental data (red symbols) for intermediate to strong surface active substances on linear (left) and logarithmic (right) x-axis scale. Liquid-liquid phase separation (LLPS) was predicted by AIOMFAC for nonanoic acid and Triton X-100 in the grey shaded concentration range. Different symbols of the experimental data refer to different sources (see Table 2 and Fig. S2 in ESI†). Note that the curve fits of the models Stat. uncon. ideal, Eberhart and Connors-Wright partly or completely overlay.

required to ensure high precision. Combined with its high accuracy, the Sigmoid model is easily interpretable and convenient,

making it an elegant tool for the numerical reporting and comparison of solutes surface activeness.



Compared to the Szyszkowski–Langmuir equation, which is often used in atmospheric sciences, the Sigmoid model provides a smooth curve over the whole concentration range, while the Szyszkowski–Langmuir equation needs to be combined with a constant value at concentrations above the CMC, which can be challenging for numerical solvers.

The Sigmoid model, being a purely empirical model, requires experimental data, *i.e.* it has no predictive capabilities at the current state of development. Possible relationships between molecular properties and the fit parameters have yet to be researched. Such relationships have been found for the Statistical model for alcohols, polyols and electrolytes. Similarly, the Tamura model suggests predictive model parameters for certain solute groups. Only the Butler equation provides a fully predictive surface tension model through thermodynamic derivation that is valid for all types of solutes. However, it requires activity coefficients and molar surface areas of all substances in the mixture. Due to the difficulties in obtaining precise values for these, the predictive capabilities of the Butler equation are often unsatisfactory, and a measurement of the surface tension is a more reliable option compared to a prediction based on molecular properties.

The ability of the models to extrapolate to concentrations without experimental data was discussed in Section 4.2 and a preliminary extrapolation test suggests that the Butler equation is the most robust model to extrapolate (ESI† Section 6), possibly due to a strong thermodynamic background. Yet, more research is required to reach a more definitive conclusion.

For processes like heterogeneous surface reactions or bulk depletion in tiny droplets, the surface composition  $x_i^{\text{surf}}$  corresponding to a certain bulk composition is required additionally to surface tension. In the Butler equation,  $a_i^{\text{surf}}$  is a direct model output and allows to back-calculate  $x_i^{\text{surf}}$ . The Statistical model allows a derivation of  $x_i^{\text{surf}}$  as a function of  $a_i$  and the fit parameters based on eqn (8) in Wexler and Dutcher.<sup>50</sup> Also for the Li & Lu model, a derivation of the number of moles in the surface and the bulk has been suggested.<sup>46</sup> The Eberhart and the Connors–Wright models assume that a simple linear mixing rule

$$\sigma = \sigma_i x_i^{\text{surf}} + \sigma_w x_w^{\text{surf}} \quad (28)$$

holds in the derivation of the models. Based on this equation and the final model equation, the surface composition can be derived for the Eberhart model as

$$x_i^{\text{surf}} = \frac{S}{S - 1 + 1/x_i} \quad (29)$$

and for the Connors–Wright model as

$$x_i^{\text{surf}} = x_i \left[ 1 + \frac{b(1 - x_i)}{1 - a(1 - x_i)} \right]. \quad (30)$$

Eqn (28) is also used in combination with Szyszkowski–Langmuir fits in the “Monolayer model” by Malila and Prisle<sup>122</sup> to calculate the bulk depletion in atmospheric aerosol particles. The empirical Tamura and Sigmoid models have no definition of  $x_i^{\text{surf}}$  from a thermodynamic standpoint. Yet, assuming the

validity of eqn (28) also for the Sigmoid model, an equation for  $x_i^{\text{surf}}$  can be derived by setting eqn (28) equal to eqn (25), resulting in

$$x_i^{\text{surf}} = (10^{pd} + 1) \frac{x_i^d}{10^{pd} + x_i^d}. \quad (31)$$

A model with a thermodynamic background brings the advantage that an equation for solutions with more than one solute can be derived. As such, the Butler equation is inherently multi-component compatible, and the Statistical, Li & Lu and Connors–Wright models each have multi-component counterparts (Table 1). For the Tamura, Eberhart, Shereshefsky and Sigmoid models, no multi-component versions of the models have been suggested yet.

## 5. Summary and conclusions

In this study, we reviewed empirical, semi-empirical and thermodynamic surface tension models for aqueous solutions. Furthermore, we derived a new empirical model that is based on the logistic function (Sigmoid model). We compared the ability of six frequently-used models as well as the proposed Sigmoid model to fit experimental surface tension data of binary aqueous mixtures. Our study revealed the following findings:

- The Sigmoid model was found to have the best overall fit performance of all models, with an average RMSE of  $0.92 \text{ mN m}^{-1}$  and individual RMSE values  $< 2 \text{ mN m}^{-1}$ . For all tested substances, the experimental data could be excellently reproduced (see Fig. 6). We therefore recommend this model to describe experimental surface tension data with an analytical equation.

- From the models proposed in the literature, the Connors–Wright model was found to be the most versatile in fitting different curve shapes. In contrast to the empirical Sigmoid model, it has a simple thermodynamic basis and a formulation for multi-component solutions. Its capability to predict the surface tension of multi-component solutions for a wide variety of compounds, however, remains to be tested.

- For weakly surface active substances like 1,2-ethanediol or methanol, all seven models achieved good results with RMSE values  $< 2 \text{ mN m}^{-1}$ . The Li & Lu model, the Tamura model and the Butler equation (ideal) were found not to be able to fit the surface tension of surfactants. The Statistical model is equivalent to the Li & Lu model for intermediate and strong surfactants when restricting the fit parameters to values within the physically meaningful limits. An unphysical usage of the model by allowing negative values for  $K$  leads to a model with good fit performance, especially when assuming ideality (“Stat. uncon. ideal”) although lacking a thermodynamic basis.

- For substances that are solid at room temperature, the pure liquid surface tension  $\sigma_i$  is unknown. We discussed various options to estimate its value, including to make  $\sigma_i$  a fit parameter, which means estimating its value from an extrapolation of the experimental data using one of the tested



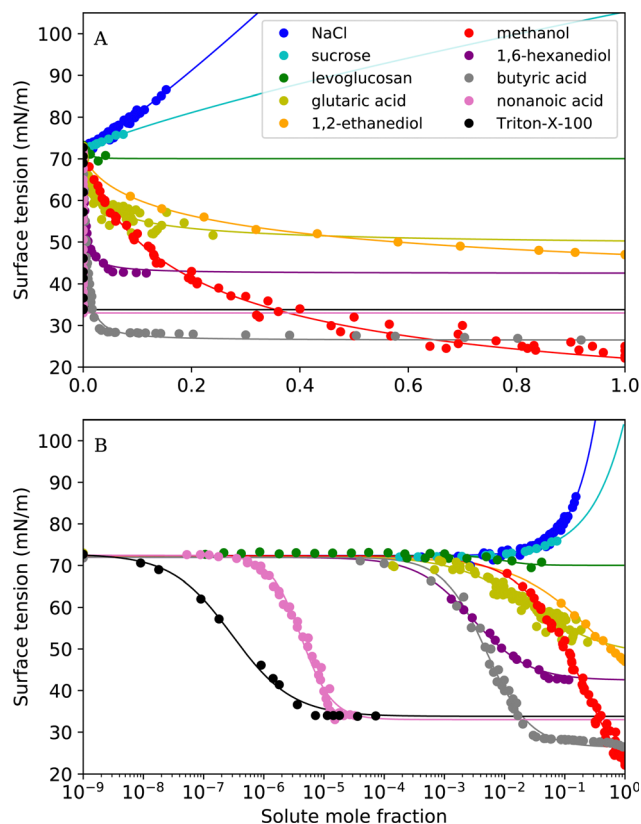


Fig. 6 Surface tension fits with the Sigmoid model for all tested substances (A) on a linear x-axis scale (B) on a logarithmic x-axis scale.

surface tension models. Depending on the model, the values obtained for  $\sigma_i$  by such extrapolation are more or less subject to physical constraints. According to our analysis, the Butler equation seems to be a robust model for extrapolating to unknown concentrations as long as accurate, substance-specific activity coefficients can be provided. Overall, more research about the various methods of obtaining  $\sigma_i$  is clearly needed, especially for sugars.

- AIOMFAC has proven a useful tool to calculate activity coefficients for a broad range of substances including organic and inorganic substances. For strong surfactants, AIOMFAC predicts a liquid–liquid phase separation into one surfactant-rich and a water-rich phase within a wide concentration range which can be interpreted as the formation of micelles. This prediction of micelle formation (in the form of a liquid–liquid phase separation) is crucial for surface tension fits for strong surfactants but it is not very accurate in AIOMFAC, since AIOMFAC is a group contribution method. A solution to this problem could be a tuning of the activity coefficients to the specific substance. This could be the subject of future work.

To conclude, the Sigmoid model was found to excellently reproduce the surface tension of binary aqueous solutions over the whole concentration range and for a broad range of solutes. This allows modelling the surface tension of aqueous solutions in a universal and simple way, which can be helpful in many fields. Further research should be directed to extending the

Sigmoid model to capture multi-component solutions and to obtaining better estimates of the pure liquid surface tension  $\sigma_i$  of crystallizing substances.

## Data availability statement

Data for this paper, including the experimental surface tension data, activity coefficients calculated with AIOMFAC, fitted curves, and fit parameters are available at <https://doi.org/10.5281/zenodo.7548346>.

## Author contributions

J. K. reviewed the literature, analyzed the models, visualized the results and wrote the original draft. N. S. and C. M. directly supervised this work and contributed to the methodology and validation and C. M. acquired the funding. M. H., C. F. and B. N. provided the surface tension data. T. P. contributed with resources and supervising. All authors contributed to conceptualization and writing (review & editing), and all authors have approved the final version of the paper.

## Conflicts of interest

There are no conflicts to declare.

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## References

- 1 S. Syeda, A. Afacan and K. Chuang, *Chem. Eng. Res. Des.*, 2004, **82**, 762–769.
- 2 J. Pérez-Gil, *Biochim. Biophys. Acta, Biomembr.*, 2008, **1778**, 1676–1695.
- 3 O. Mc Callion, K. Taylor, M. Thomas and A. Taylor, *Int. J. Pharm.*, 1996, **129**, 123–136.
- 4 K. A. Wokosin, E. L. Schell and J. A. Faust, *Environ. Sci.: Atmos.*, 2022, **2**, 775–828.
- 5 R. C. Tolman, *J. Chem. Phys.*, 1949, **17**, 333–337.
- 6 H. D. Lee and A. V. Tivanski, *Annu. Rev. Phys. Chem.*, 2021, **72**, 235–252.
- 7 R. Power and J. Reid, *Rep. Prog. Phys.*, 2014, **77**, 074601.



- 8 B. Minofar, P. Jungwirth, M. R. Das, W. Kunz and S. Mahiuddin, *J. Phys. Chem. C*, 2007, **111**, 8242–8247.
- 9 S. Mahiuddin, B. Minofar, J. M. Borah, M. R. Das and P. Jungwirth, *Chem. Phys. Lett.*, 2008, **462**, 217–221.
- 10 J. C. Neyt, A. Wender, V. Lachet, A. Ghoufi and P. Malfreyt, *J. Chem. Phys.*, 2013, **139**, 024701.
- 11 N. S. Mousavi, B. Vaferi and A. Romero-Martínez, *Ind. Eng. Chem. Res.*, 2021, **60**, 10354–10364.
- 12 Z. Nieto, V. M. K. Kottedda, A. Rodriguez, S. S. Kumar, V. Kumar and A. Bronson, Proceedings of the 5th Joint US-European Fluids Engineering Division Summer Meeting, ASME, 2018, vol. 3, p. V003T21A003.
- 13 M. Pierantozzi, A. Mulero and I. Cachadiña, *Molecules*, 2021, **26**, 1636.
- 14 T. Soori, S. M. Rassoulinejad-Mousavi, L. Zhang, A. Rokoni and Y. Sun, *Fluid Phase Equilib.*, 2021, **538**, 113012.
- 15 I. Egry, E. Ricci, R. Novakovic and S. Ozawa, *Adv. Colloid Interface Sci.*, 2010, **159**, 198–212.
- 16 A. Asa-Awuku, A. P. Sullivan, C. J. Hennigan, R. J. Weber and A. Nenes, *Atmos. Chem. Phys.*, 2008, **8**, 799–812.
- 17 E. Aumann, L. Hildemann and A. Tabazadeh, *Atmos. Environ.*, 2010, **44**, 329–337.
- 18 B. R. Bzdek, J. P. Reid, J. Malila and N. L. Prisle, *Proc. Natl. Acad. Sci.*, 2020, **117**, 8335–8343.
- 19 M. Facchini, M. Mircea, F. Sandro and R. Charlson, *Nature*, 1999, **401**, 257–259.
- 20 M. R. Giordano, D. Z. Short, S. Hosseini, W. Lichtenberg and A. A. Asa-Awuku, *Environ. Sci. Technol.*, 2013, **47**, 10980–10986.
- 21 S. Henning, T. Rosenørn, B. D'Anna, A. A. Gola, B. Svenningsson and M. Bilde, *Atmos. Chem. Phys.*, 2005, **5**, 575–582.
- 22 H. Kokkola, R. Sorjamaa, A. Peräniemi, T. Raatikainen and A. Laaksonen, *Geophys. Res. Lett.*, 2006, **33**, L10816.
- 23 T. B. Kristensen, N. L. Prisle and M. Bilde, *Atmos. Res.*, 2014, **137**, 167–175.
- 24 Z. Li, A. L. Williams and M. Rood, *J. Atmos. Sci.*, 1998, **55**, 1859–1866.
- 25 J. J. Lin, J. Malila and N. L. Prisle, *Environ. Sci.: Processes Impacts*, 2018, **20**, 1611–1629.
- 26 J. J. Lin, T. B. Kristensen, S. M. Calderón, J. Malila and N. L. Prisle, *Environ. Sci.: Processes Impacts*, 2020, **22**, 271–284.
- 27 A. Z. Mazurek, S. J. Pogorzelski and A. D. Kogut, *Atmos. Environ.*, 2006, **40**, 4076–4087.
- 28 R. McGraw and J. Wang, *J. Chem. Phys.*, 2021, **154**, 024707.
- 29 H. S. Morris, V. H. Grassian and A. V. Tivanski, *Chem. Sci.*, 2015, **6**, 3242–3247.
- 30 B. Noziere, C. Baduel and J.-L. Jaffrezo, *Nat. Commun.*, 2014, **5**, 3335.
- 31 S. S. Petters and M. D. Petters, *J. Geophys. Res.: Atmos.*, 2016, **121**, 1878–1895.
- 32 N. Prisle, T. Raatikainen, R. Sorjamaa, B. Svenningsson, A. Laaksonen and M. Bilde, *Tellus B*, 2008, **60**, 416–431.
- 33 N. L. Prisle, T. Raatikainen, A. Laaksonen and M. Bilde, *Atmos. Chem. Phys.*, 2010, **10**, 5663–5683.
- 34 N. L. Prisle, M. Dal Maso and H. Kokkola, *Atmos. Chem. Phys.*, 2011, **11**, 4073–4083.
- 35 N. L. Prisle, A. Asmi, D. Topping, A.-I. Partanen, S. Romakkaniemi, M. Dal Maso, M. Kulmala, A. Laaksonen, K. E. J. Lehtinen, G. McFiggans and H. Kokkola, *Geophys. Res. Lett.*, 2012, **39**, L05802.
- 36 N. L. Prisle and B. Molgaard, *Atmospheric Chemistry and Physics Discussions*, 2018, **2018**, 1–23.
- 37 N. L. Prisle, *Atmos. Chem. Phys.*, 2021, **21**, 16387–16411.
- 38 T. Raatikainen and A. Laaksonen, *Geosci. Model Dev.*, 2011, **4**, 107–116.
- 39 S. Romakkaniemi, H. Kokkola, J. N. Smith, N. L. Prisle, A. N. Schwier, V. F. McNeill and A. Laaksonen, *Geophys. Res. Lett.*, 2011, **38**, L03807.
- 40 A. Schwier, D. Mitroo and V. F. McNeill, *Atmos. Environ.*, 2012, **54**, 490–495.
- 41 R. Sorjamaa and A. Laaksonen, *J. Aerosol Sci.*, 2006, **37**, 1730–1736.
- 42 R. Tuckermann, *Atmos. Environ.*, 2007, **41**, 6265–6275.
- 43 Z. Li and B. C.-Y. Lu, *Chem. Eng. Sci.*, 2001, **56**, 2879–2888.
- 44 D. O. Topping, G. B. McFiggans, G. Kiss, Z. Varga, M. C. Facchini, S. Decesari and M. Mircea, *Atmos. Chem. Phys.*, 2007, **7**, 2371–2398.
- 45 A. M. Booth, D. O. Topping, G. McFiggans and C. J. Percival, *Phys. Chem. Chem. Phys.*, 2009, **11**, 8021–8028.
- 46 D. Topping, *Geosci. Model Dev.*, 2010, **3**, 635–642.
- 47 B. von Szyszkowski, *Z. Phys. Chem.*, 1908, **64U**, 385–414.
- 48 I. Langmuir, *J. Am. Chem. Soc.*, 1917, **39**, 1848–1906.
- 49 A. N. Schwier, G. A. Viglione, Z. Li and V. Faye McNeill, *Atmos. Chem. Phys.*, 2013, **13**, 10721–10732.
- 50 A. S. Wexler and C. S. Dutcher, *J. Phys. Chem. Lett.*, 2013, **4**, 1723–1726.
- 51 H. Boyer, A. Wexler and C. Dutcher, *J. Phys. Chem. Lett.*, 2015, **6**, 3384–3389.
- 52 H. C. Boyer and C. S. Dutcher, *J. Phys. Chem. A*, 2017, **121**, 4733–4742.
- 53 H. C. Boyer, B. R. Bzdek, J. P. Reid and C. S. Dutcher, *J. Phys. Chem. A*, 2017, **121**, 198–205.
- 54 R. Miles, M. Glerum, H. Boyer, J. Walker, C. Dutcher and B. Bzdek, *J. Phys. Chem. A*, 2019, **123**, 3021–3029.
- 55 J. A. V. Butler, *Proc. R. Soc. London, Ser. A*, 1932, **135**, 348–375.
- 56 F. B. Sprow and J. M. Prausnitz, *Trans. Faraday Soc.*, 1966, **62**, 1105–1111.
- 57 J. T. Suarez, C. Torres-Marchal and P. Rasmussen, *Chem. Eng. Sci.*, 1989, **44**, 782–785.
- 58 M. S. C. S. Santos and J. C. R. Reis, *ACS Omega*, 2021, **6**, 21571–21578.
- 59 B. E. Poling, J. M. Prausnitz and J. P. O'Connell, *Properties of Gases and Liquids*, McGraw-Hill Education, New York, 5th edn, 2001, ch. 12.
- 60 Z.-B. Li, Y.-G. Li and J.-F. Lu, *Ind. Eng. Chem. Res.*, 1999, **38**, 1133–1139.
- 61 Y.-F. Hu and H. Lee, *J. Colloid Interface Sci.*, 2004, **269**, 442–448.
- 62 X. Cai and R. J. Griffin, *J. Atmos. Chem.*, 2005, **50**, 139–158.



- 63 J. Werner, J. Julin, M. Dalirian, N. L. Prisle, G. Öhrwall, I. Persson, O. Björneholm and I. Riipinen, *Phys. Chem. Chem. Phys.*, 2014, **16**, 21486–21495.
- 64 J. Werner, M. Dalirian, M.-M. Walz, V. Ekholm, U. Wideqvist, S. Lowe, G. Öhrwall, I. Persson, I. Riipinen and O. Björneholm, *Environ. Sci. Technol.*, 2016, **50**, 7434–7442.
- 65 M. Tamura, M. Kurata and H. Odani, *Bull. Chem. Soc. Jpn.*, 1955, **28**, 83–88.
- 66 J. G. Eberhart, *J. Phys. Chem.*, 1966, **70**, 1183–1186.
- 67 J. Shereshefsky, *J. Colloid Interface Sci.*, 1967, **24**, 317–322.
- 68 N. Shardt and J. A. W. Elliott, *Langmuir*, 2017, **33**, 11077–11085.
- 69 K. A. Connors and J. L. Wright, *Anal. Chem.*, 1989, **61**, 194–198.
- 70 N. Shardt, Y. Wang, Z. Jin and J. A. Elliott, *Chem. Eng. Sci.*, 2021, **230**, 116095.
- 71 L. Chunxi, W. Wenchuan and W. Zihao, *Fluid Phase Equilib.*, 2000, **175**, 185–196.
- 72 A.-P. Hyvärinen, H. Lihavainen, A. Gaman, L. Vairila, H. Ojala, M. Kulmala and Y. Viisanen, *J. Chem. Eng. Data*, 2006, **51**, 255–260.
- 73 J. Vanhanen, A.-P. Hyvärinen, T. Anttila, T. Raatikainen, Y. Viisanen and H. Lihavainen, *Atmos. Chem. Phys.*, 2008, **8**, 4595–4604.
- 74 R. Defay, I. Prigogine, A. Bellemans and D. Everett, *Surface Tension and Adsorption*, Wiley, 1966.
- 75 R. Sorjamaa, B. Svenningsson, T. Raatikainen, S. Henning, M. Bilde and A. Laaksonen, *Atmos. Chem. Phys.*, 2004, **4**, 2107–2117.
- 76 J. A. W. Elliott, *J. Phys. Chem. B*, 2020, **124**, 10859–10878.
- 77 I. Langmuir, *J. Am. Chem. Soc.*, 1918, **40**, 1361–1403.
- 78 H. C. Boyer and C. S. Dutcher, *J. Phys. Chem. A*, 2016, **120**, 4368–4375.
- 79 S. Liu and C. S. Dutcher, *J. Phys. Chem. A*, 2021, **125**, 1577–1588.
- 80 C. S. Dutcher, X. Ge, A. S. Wexler and S. L. Clegg, *J. Phys. Chem. C*, 2011, **115**, 16474–16487.
- 81 C. S. Dutcher, X. Ge, A. S. Wexler and S. L. Clegg, *J. Phys. Chem. C*, 2012, **116**, 1850–1864.
- 82 C. S. Dutcher, X. Ge, A. S. Wexler and S. L. Clegg, *J. Phys. Chem. A*, 2013, **117**, 3198–3213.
- 83 A. Fredenslund, R. L. Jones and J. M. Prausnitz, *AIChE J.*, 1975, **21**, 1086–1099.
- 84 D. E. Goldsack and B. R. White, *Can. J. Chem.*, 1983, **61**, 1725–1729.
- 85 M. Vermot des Roches, A. E. Gheribi and P. Chartrand, *Calphad*, 2019, **65**, 326–339.
- 86 A. Dobosz, R. Novakovic and T. Gancarz, *J. Mol. Liq.*, 2021, **343**, 117646.
- 87 G. M. Wilson, *J. Am. Chem. Soc.*, 1964, **86**, 127–130.
- 88 P. Virtanen, R. Gommers, T. E. Oliphant, M. Haberland, T. Reddy, D. Cournapeau, E. Burovski, P. Peterson, W. Weckesser, J. Bright, S. J. van der Walt, M. Brett, J. Wilson, K. J. Millman, N. Mayorov, A. R. J. Nelson, E. Jones, R. Kern, E. Larson, C. J. Carey, Í. Polat, Y. Feng, E. W. Moore, J. VanderPlas, D. Laxalde, J. Perktold, R. Cimrman, I. Henriksen, E. A. Quintero, C. R. Harris, A. M. Archibald, A. H. Ribeiro, F. Pedregosa, P. van Mulbregt and SciPy 1.0 Contributors, *Nat. Methods*, 2020, **17**, 261–272.
- 89 D. Topping, M. Barley, M. K. Bane, N. Higham, B. Aumont, N. Dingle and G. McFiggans, *Geosci. Model Dev.*, 2016, **9**, 899–914.
- 90 S. L. Clegg and A. S. Wexler, *J. Phys. Chem. A*, 2011, **115**, 3393–3460.
- 91 W. Wagner and H.-J. Kretzschmar, *International Steam Tables: Properties of Water and Steam Based on the Industrial Formulation IAPWS-IF97*, Springer Berlin Heidelberg, Berlin, Heidelberg, 2008, pp. 7–150.
- 92 O. Ozdemir, S. I. Karakashev, A. V. Nguyen and J. D. Miller, *Miner. Eng.*, 2009, **22**, 263–271.
- 93 E. Washburn, C. West and C. Hull, *International Critical Tables of Numerical Data, Physics, Chemistry and Technology*, National Research Council, 1928, vol. 4.
- 94 R. Tuckermann and H. K. Cammenga, *Atmos. Environ.*, 2004, **38**, 6135–6138.
- 95 J. Y. Lee and L. M. Hildemann, *Atmos. Environ.*, 2014, **89**, 260–267.
- 96 B. R. Bzdek, R. M. Power, S. H. Simpson, J. P. Reid and C. P. Royall, *Chem. Sci.*, 2016, **7**, 274–285.
- 97 Z. Varga, G. Kiss and H.-C. Hansson, *Atmos. Chem. Phys.*, 2007, **7**, 4601–4611.
- 98 U. Messow, P. Braeuer, A. Schmidt, C. Bilke-Krause, K. Quitzsch and U. Zilles, *Adsorption*, 1998, **4**, 257–267.
- 99 P. Basařová, T. Váchová and L. Bartovská, *Colloids Surf., A*, 2016, **489**, 200–206.
- 100 J. Gliński, G. Chavepeyer, J.-K. Platten and P. Smet, *J. Chem. Phys.*, 1998, **109**, 5050–5053.
- 101 R. B. Maximino, *Phys. Chem. Liq.*, 2009, **47**, 475–486.
- 102 I. A. Semenov and M. A. Pokrovskaya, *Theor. Found. Chem. Eng.*, 2014, **48**, 90–95.
- 103 C. Romero, M. S. Páez, J. A. Miranda, D. J. Hernández and L. E. Oviedo, *Fluid Phase Equilib.*, 2007, **258**, 67–72.
- 104 K. Granados, J. Gracia-Fadrique, A. Amigo and R. Bravo, *J. Chem. Eng. Data*, 2006, **51**, 1356–1360.
- 105 F. Suárez and C. M. Romero, *J. Chem. Eng. Data*, 2011, **56**, 1778–1786.
- 106 D. J. Donaldson and D. Anderson, *J. Phys. Chem. A*, 1999, **103**, 871–876.
- 107 K. Lunkenheimer, W. Barzyk, R. Hirte and R. Rudert, *Langmuir*, 2003, **19**, 6140–6150.
- 108 S. Badban, A. E. Hyde and C. M. Phan, *ACS Omega*, 2017, **2**, 5565–5573.
- 109 A. Zdziennicka, K. Szymczyk, J. Krawczyk and B. Jańczuk, *Fluid Phase Equilib.*, 2012, **318**, 25–33.
- 110 A. Zuend, C. Marcolli, B. P. Luo and T. Peter, *Atmos. Chem. Phys.*, 2008, **8**, 4559–4593.
- 111 A. Zuend, C. Marcolli, A. M. Booth, D. M. Lienhard, V. Soonsin, U. K. Krieger, D. O. Topping, G. McFiggans, T. Peter and J. H. Seinfeld, *Atmos. Chem. Phys.*, 2011, **11**, 9155–9206.



- 112 K. S. Pitzer, *Activity Coefficients in Electrolyte Solutions*, CRC Press, 2nd edn, 1991.
- 113 C. Marcolli and T. Peter, *Atmos. Chem. Phys.*, 2005, **5**, 1545–1555.
- 114 A. Zuend, C. Marcolli, T. Peter and J. H. Seinfeld, *Atmos. Chem. Phys.*, 2010, **10**, 7795–7820.
- 115 A. Zuend and J. H. Seinfeld, *Fluid Phase Equilib.*, 2013, **337**, 201–213.
- 116 G. J. Janz, *J. Phys. Chem. Ref. Data*, 1980, **9**, 791–830.
- 117 D. M. Lienhard, D. L. Bones, A. Zuend, U. K. Krieger, J. P. Reid and T. Peter, *J. Phys. Chem. A*, 2012, **116**, 9954–9968.
- 118 A. A. Zardini, S. Sjogren, C. Marcolli, U. K. Krieger, M. Gysel, E. Weingartner, U. Baltensperger and T. Peter, *Atmos. Chem. Phys.*, 2008, **8**, 5589–5601.
- 119 C. Peng, M. N. Chan and C. K. Chan, *Environ. Sci. Technol.*, 2001, **35**, 4495–4501.
- 120 B. Zobrist, V. Soonsin, B. P. Luo, U. K. Krieger, C. Marcolli, T. Peter and T. Koop, *Phys. Chem. Chem. Phys.*, 2011, **13**, 3514–3526.
- 121 C. S. Dutcher, A. S. Wexler and S. L. Clegg, *J. Phys. Chem. A*, 2010, **114**, 12216–12230.
- 122 J. Malila and N. L. Prisle, *J. Adv. Model. Earth Syst.*, 2018, **10**, 3233–3251.
- 123 A. Mulero, I. Cachadiña and E. L. Sanjuán, *J. Phys. Chem. Ref. Data*, 2016, **45**, 033105.
- 124 E. Lemmon, M. Huber and M. McLinden, NIST Standard Reference Database 23: Reference Fluid Thermodynamic and Transport Properties-REFPROP, Version 9.1, 2013, [https://tsapps.nist.gov/publication/get\\_pdf.cfm?pub\\_id=912382](https://tsapps.nist.gov/publication/get_pdf.cfm?pub_id=912382).

