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ESIPT-based fluorescence probe for the rapid detection of hypochlorite (HOCl/CIO[−])†

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ESIPT-based fluorescence probes are emerging as an attractive tool for the detection of biologically relevant analytes owing to their unique photophysical properties. In this work, we have developed an ESIPT-based fluorescence probe (TCBT-OMe) for the detection of HOCl/CIO[−] through the attachment of a bioorthogonal dimethylthiocarbamate linker. TCBT-OMe was shown to rapidly detect HOCl/CIO[−] (<10 s) at biologically relevant concentrations (LoD = 0.16 nM) and have an excellent selectivity towards others ROS/RNS and amino acids. Therefore, TCBT-OMe was tested in live cells and was successfully shown to be able to detect endogenous and exogenous HOCl/CIO[−] in HeLa cells. Additionally, TCBT-OMe acts as a dual input logic gate for Hg²⁺ and H₂O₂. Interestingly, Hg²⁺ alone gradually causes a fluorescence response but requires >30 min to produce a fluorescence response. Test strips containing TCBT-OMe were prepared and were demonstrated as an effective way to detect HOCl/CIO[−] in water. Furthermore, TCBT-OMe was shown to detect exogenously added HOCl/CIO[−] in three different water samples with little interference thus demonstrating the effectiveness as a method for the detection of HOCl/CIO[−] in drinking water samples.

Hypochlorous acid (HOCl) is a biologically important reactive oxygen species (ROS), which partially dissociates to form its hypochlorite anion (CIO[−]) under physiological conditions. In biological systems, myeloperoxidase, an enzyme found in leukocytes produces HOCl/CIO[−] by catalysing the reaction between Cl + H₂O₂ → HOCl.¹ This vital ROS is used in immune defence systems due to its microbicidal properties.¹ Unfortunately, excessive

production of HOCl/CIO[−] can lead to the damage of a range of biological targets such as amino acids, proteins, carbohydrates and lipids.^{2,3} As a consequence, HOCl/CIO[−] has been associated with a number of diseases causing cell and tissue damage.⁴

In addition to its role in biological systems, HOCl/CIO[−] is produced by the chlorination of water (Cl₂ + H₂O → HOCl), which is the most common method for the treatment of water especially in public swimming pools.⁵ NaOCl (Bleach) is also extensively used as a disinfectant for both domestic and industrial purposes. Unfortunately, over-exposure to HOCl/CIO[−], results in swimming pool-associated asthma, irritation to the oesophagus, throat and spontaneous vomiting (http://www.who.int/water_sanitation_health/dwq/chlorine.pdf).⁶ Additionally, there is an increased risk of bladder cancer associated with chlorinated by-products produced from chlorinated water.^{7,8} Therefore, given the potential health hazard towards animals and humans, the development of an effective method for HOCl/CIO[−] detection is required.

Within our research group, we are interested in developing reaction-based fluorescence sensors for the detection of biologically important analytes.^{9–13} Small-molecule fluorescence probes are a particular attractive tool owing to their high sensitivity, selectivity and high spatial and temporal resolution.¹⁴ In particular, we are interested in using Excited State Intramolecular Proton Transfer (ESIPT)-based fluorescence probes due to their excellent photophysical properties, which include intense luminescence, photostability and a large Stokes shift.^{15,16} Previously, we reported an ESIPT-based fluorescence probe for the detection of peroxynitrite (ONOO[−]) through the use of a benzyl boronic ester protecting group (Scheme 1).¹⁵ This protecting group blocked the ESIPT process and therefore a low fluorescence intensity was observed. The addition of ONOO[−], resulted in the fluorophore's deprotection and an increase in fluorescence intensity was observed.

In this work, we believed a methoxy-hydroxybenzothiazole (HBT-OMe) fluorophore would provide an effective ESIPT fluorescence probe for the detection of HOCl/CIO[−] (see ESI,† S1).^{17,18}

To obtain TCBT-OMe we first prepared HBT-OMe by the addition of a 2 : 1 H₂O₂–(30% in H₂O)/HCl solution to 2-aminothiophenol

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Scheme 1 (a) Our previously reported ESIPT probe for the detection of ONOO^- . (b) This work – a thiocarbamate linker-based ESIPT **TCBT-OMe** for the detection of HOCl/CLO^- .

and *O*-vanillin in EtOH. This reaction proceeded quickly and smoothly, in a good yield (68%). With **HBT-OMe** in hand, four equivalents of dimethylthiocarbamoyl chloride was then added slowly to a solution of **HBT-OMe** in DCM. DIPEA was subsequently added dropwise to the reaction, which produced **TCBT-OMe** in excellent yield (72%).

We then evaluated the UV-Vis of **TCBT-OMe** with the addition of HOCl/CLO^- (10 μM), which resulted in the formation of a UV absorption peak at ~ 310 nm (see ESI,† Fig. S1). Bhattacharyya *et al.* have reported that the fluorescence emission of the ESIPT process can be effected by intermolecular hydrogen bonding.^{19,20} Therefore, evaluation of ESIPT-based fluorescence probes are commonly carried out in the presence of the surfactant cetyl trimethylammonium bromide (CTAB, 1 mM) or by using a large ratio of organic solvent.^{19,21–23} It is believed that the formation of a micellar environment creates a hydrophobic pocket that aids the ESIPT process. Therefore, we evaluated the ability of **TCBT-OMe** to detect HOCl/CLO^- by fluorescence in the presence of CTAB, 1 mM. As shown in Fig. 1a, **TCBT-OMe** was found to be very sensitive towards HOCl/CLO^- reacting with micromolar concentrations to produce a large increase in fluorescence (~ 42 fold – Fig. S3, ESI†). **TCBT-OMe** was shown to rapidly react with HOCl/CLO^- producing a fluorescence response within less than 10 s (see ESI,† Fig. S4) and have a very low Limit of Detection (LoD) of 0.16 nM (see ESI,† Fig. S5). HOCl/CLO^- (35 μM) was added to **TCBT-OMe** at different pH values and a bell-shaped curve was observed. The largest fluorescence response was seen at the pK_a of $\text{HOCl/CLO}^- = 7.53$ (Fig. S5, ESI†) suggestive of general acid–base catalysis being in operation. (see ESI,† Scheme S1 for proposed mechanism).

We then evaluated the selectivity of **TCBT-OMe** towards other reactive oxygen/nitrogen species (ROS/RNS) and amino acids (Fig. 1b). Remarkably, **TCBT-OMe** had an excellent selectivity towards HOCl/CLO^- therefore permitting its use as a fluorescence probe for the detection of HOCl/CLO^- in live cells. As shown in Fig. 2, **TCBT-OMe** was successfully used to visualise endogenously stimulated HOCl/CLO^- in HeLa cells using phorbol 12-myristate 13-acetate (**PMA**, which is a ROS stimulant that induces the production of HOCl/CLO^-). Separately, HeLa cells were

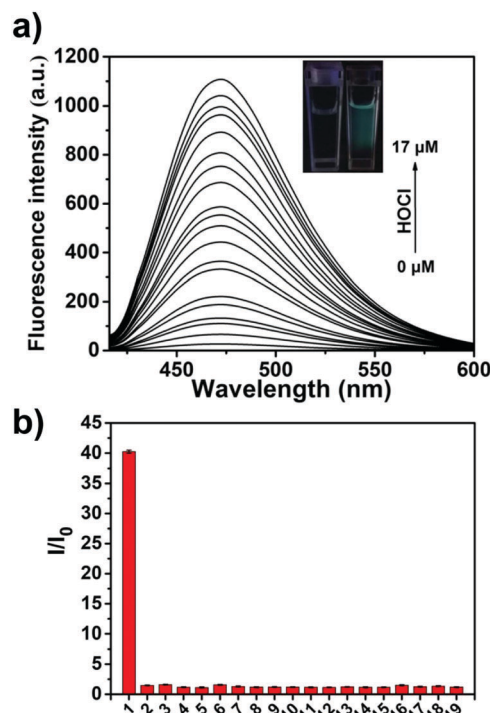


Fig. 1 (a) Fluorescence spectra of **TCBT-OMe** (5 μM) with increasing additions of HOCl/CLO^- (from 0 to 17 μM) in PBS buffer (pH 7.4, containing 1% DMSO, 1 mM CTAB). Measurements were taken after 1 min. $\lambda_{\text{ex}} = 310$ nm. Slit widths: ex = 6 nm em = 4 nm. (b) Selectivity bar chart of **TCBT-OMe** in PBS pH 7.4, containing 1% DMSO, 1 mM CTAB with HClO (15 μM) and other interfering reagents (ROS/RNS and various amino acids). 1, HClO ; 2, blank; 3, ONOO^- ; 4, H_2O_2 ; 5, ROO^\bullet ; 6, $\bullet\text{OH}$; 7, $\bullet\text{O}_2^-$; 8, $^1\text{O}_2$; 9, NO; 10, glycine; 11, asparagine; 12, cysteine; 13, homocysteine; 14, glutathione; 15, arginine; 16, histidine; 17, serine; 18, glycine; 19, threonine. Note: the concentration of **TCBT-OMe** and each interfering species are 5 μM and 100 μM respectively, 30 min wait before measurement in buffer solution. $\lambda_{\text{ex}} = 310$ nm/ $\lambda_{\text{em}} = 472$ nm error bars represent s.d. Measurements were taken after 30 min. $\lambda_{\text{ex}} = 310$ nm. Slit widths: ex = 6 nm, em = 4 nm.

also pretreated with 4-aminobenzoic acid hydrazide (**ABAH**, which is a specific inhibitor of MPO which suppressed the generation of HOCl) and as expected only weak fluorescence was observed. **TCBT-OMe** was also able to detect HOCl/CLO^- added exogenously to the HeLa cells.

The dimethylthiocarbamate linker of **TCBT-OMe** has previously been used in the construction of dual input molecular logic gate²⁴ for the detection of Hg^{2+} 'AND' H_2O_2 (see ESI† Scheme S2 for proposed mechanism).^{25,26} Therefore, we evaluated the ability of **TCBT-OMe** to perform molecular logic with the input of Hg^{2+} and H_2O_2 . The presence of solely H_2O_2 (120 μM) led to a small increase in fluorescence intensity (dashed line), however, with subsequent additions of Hg^{2+} (0–9 μM) a large fluorescence response was observed (Fig. 3a). To demonstrate that both analytes are required, Hg^{2+} was added first, followed by the addition of H_2O_2 (0–180 μM). As shown in Fig. 3b, the subsequent addition of H_2O_2 rapidly led to an increase in fluorescence intensity. **TCBT-OMe** was shown to be selective towards Hg^{2+} over other metal cations in the presence of H_2O_2 (see ESI,† Fig. S9). Interestingly, Hg^{2+} alone resulted in a slow increase in fluorescence intensity (see ESI,† Fig. S10). This is believed to be



Fig. 2 (a) From top to bottom: HeLa cells were pretreated with **TCBT-OMe** (40 μM) for 30 min; HeLa cells pretreated with **TCBT-OMe** (40 μM) were then left for 30 min after preincubation with PMA (1.2 $\mu\text{g mL}^{-1}$) for 90 min; HeLa cells pretreated with **TCBT-OMe** (40 μM) were then left for 30 min after preincubation with 250 μM ABAH for 70 min; HeLa cells loaded with **TCBT-OMe** (40 μM) for 30 min followed by the exogenous addition of 8 μM NaOCl for 5 min. Scale bar: 25 μm λ_{ex} = 420 nm/ λ_{em} = 420–590 nm. (b) The histogram shows the semi-quantitative calculation of averaged fluorescence intensity (FI) of each fluorescence panel in the displayed images by ImageJ software.

due to the instability of the dimethylcarbonate formed from the reaction of **TCBT-OMe** with Hg^{2+} .

Despite this interesting dual responsive reactivity of **TCBT-OMe**, this 'AND' logic requires minutes to fully react, whereas HOCl/ClO^- reacts with **TCBT-OMe** within seconds. Therefore, due to the significantly greater reactivity of **TCBT-OMe** towards HOCl/ClO^- over Hg^{2+} , we believed we could use it as an effective method for the detection of HOCl/ClO^- in drinking water sources.

We produced test strips by simply soaking a commercially available test strip in water containing **TCBT-OMe** (0.8 mM). After drying, test strips impregnated with **TCBT-OMe** were placed in water containing HClO/ClO^- (0–200 μM). As shown in Fig. 4, there is a clear colour/intensity difference in the test strips that have been dipped into water containing various concentrations of HClO/ClO^- .

In addition to detecting HClO/ClO^- in water, **TCBT-OMe** was added into three different water samples containing 1 mM CTAB (Sample A, tap water from University of Bath; Sample B, water from the Avon River (Bath); Sample C, water from Roman spa in Bath). Interestingly, little interference was observed for the exogenous addition of HClO/ClO^- to each water sample (>95% recovery) – see ESI,† Table S1.

In summary, we have developed an ESIPT-based fluorescence **TCBT-OMe** for the detection of HClO/ClO^- . **TCBT-OMe** was shown to have a very high sensitivity and selectivity towards HClO/ClO^- fully reacting within 10 s and having a LoD of 0.16 μM . Significantly, **TCBT-OMe** was able to detect endogenous and

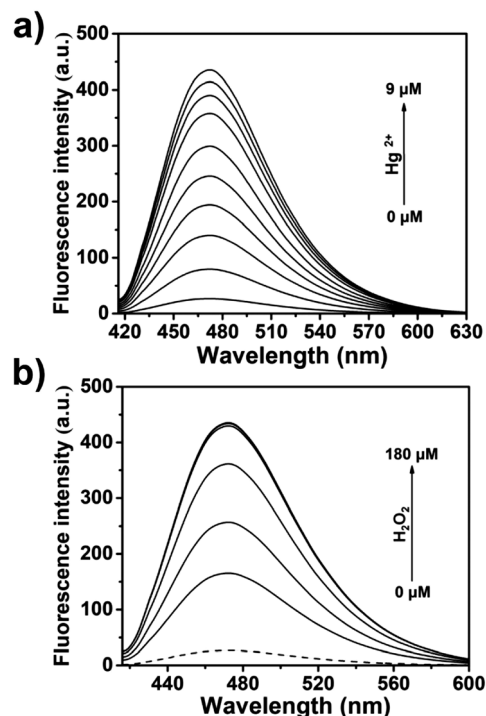


Fig. 3 (a) Fluorescence spectra of **TCBT-OMe** (5 μM) in the presence of H_2O_2 (120 μM) – (dashed line represent probe and H_2O_2) with increasing concentrations of Hg^{2+} (0–9 μM) in buffer solution pH 7.4, 1% DMSO, 1 mM CTAB 14 min wait between measurement. λ_{ex} = 310 nm. Slit widths: ex = 6 nm em = 4 nm. (b) Fluorescence spectra of **TCBT-OMe** (5 μM) in the presence of Hg^{2+} (9 μM) – (dashed line represents probe and Hg^{2+}) with increasing concentrations of H_2O_2 (final concentration: 0, 20, 40, 80, 100, 120, 140 μM and 180 μM) in PBS pH 7.4, containing 1% DMSO, 1 mM CTAB. 14 min wait between measurement in buffer solution. λ_{ex} = 310 nm. Slit widths: ex = 6 nm em = 4 nm.



Fig. 4 Photograph showing the colour changes of **TCBT-OMe** impregnated test strips after addition to water samples containing different concentrations of HClO/ClO^- under UV light (365 nm).

exogenous HClO/ClO^- in HeLa cells. Additionally, **TCBT-OMe** was shown as a dual input logic gate with Hg^{2+} and H_2O_2 as inputs. Interestingly, Hg^{2+} alone gradually produced a fluorescence response but required >30 min to produce a significant fluorescence response. Test strips containing **TCBT-OMe** were developed and shown to be an effective way to detect HClO/ClO^- in water. Furthermore, **TCBT-OMe** was shown to detect exogenously added HClO/ClO^- in three different water samples with little interference demonstrating its effectiveness as a method to detect HClO/ClO^- in drinking water samples.

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Conflicts of interest

No conflicts of interest.

Notes and references

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