



Cite this: DOI: 10.1039/d5sc05667e

All publication charges for this article have been paid for by the Royal Society of Chemistry

Mimicking sMMOH chemistry: trapping the Sc^{3+} -bound nonheme $\text{Fe}^{\text{III}}\text{--O--O--Fe}^{\text{III}}$ adduct prior to its conversion into an $\text{Fe}^{\text{IV}}_2(\mu\text{--O})_2$ core

Patrick M. Crossland,^{††a} Bittu Chandra,^{†a} Saikat Banerjee,^{†a} Chase S. Abelson,^a Yisong Guo,^{†b} Marcel Swart^{†*cd} and Lawrence Que^{†*a}

Di-iron systems that activate O_2 to form high-valent, oxo-bridged Fe^{IV}_2 or $\text{Fe}^{\text{III}}\text{Fe}^{\text{IV}}$ products are of great interest to bio-inorganic chemists due to their relevance to the chemistry of soluble methane monooxygenase (sMMOH), which incorporates both atoms of O_2 gas into a diiron(IV) complex with an Fe_2O_2 diamond core. In this study, the $[\text{Fe}^{\text{III}}_2(\text{Me}_3\text{NTB})_2(\mu\text{--O})(\mu\text{--O})_2]^{2+}$ adduct (Me_3NTB = tris((1-methyl-1H-benzo[d]imidazole-2-yl)methyl)amine) reacts with two Sc^{3+} to break the O–O bond that in turn forms the target $\text{Fe}^{\text{IV}}(\mu\text{--O})_2\text{Fe}^{\text{IV}}$ product. This study provides the first evidence that a Lewis acid can interact directly with a diferric-peroxo complex to initiate O–O bond cleavage, as evidenced via vibrational and X-ray absorption spectroscopy.

Received 28th July 2025
Accepted 27th September 2025

DOI: 10.1039/d5sc05667e

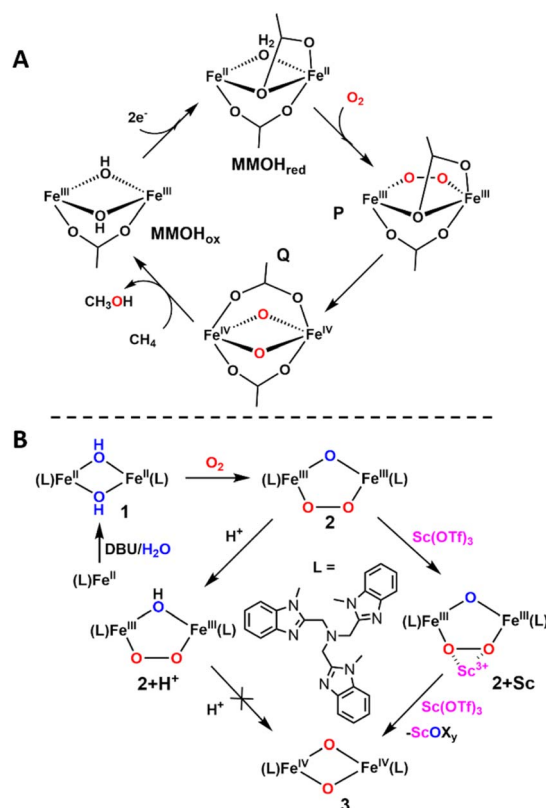
rsc.li/chemical-science

Introduction

In Nature, nonheme diiron enzymes have been found to activate dioxygen to perform a variety of reactions like the hydroxylation of methane and the eukaryotic initiation factor 5a, the oxidative desaturation of fatty acids, as well as the conversion of ribonucleotides to deoxyribonucleotides.¹ Of particular interest is the soluble methane monooxygenase hydroxylase enzyme (sMMOH), which hydroxylates the very strong C–H bond of methane ($\text{C–H BDFE} = 105 \text{ kcal mol}^{-1}$) to form methanol.² As shown in Scheme 1A, the catalytic cycle of sMMOH is initiated at its diferrous site, to which dioxygen binds to form intermediate **P**, which is best described as a diferric-peroxo species.³ **P** then converts into the diiron(IV) intermediate **Q** in a proton-dependent step^{3,4} that results in O–O bond cleavage and the incorporation of both atoms of O_2 into this key intermediate.⁵

Modeling this reaction has been a goal for synthetic inorganic chemists, and various diiron compounds have been used to mimic the proposed intermediates along the dioxygen activation pathway of sMMOH.^{6–17,26,27} Several diferric-peroxo intermediates have been identified that convert into higher-valent species by treatment with acid.^{15–17}

More recently, we have reported a biomimetic system based on a mononuclear $\text{Fe}^{\text{II}}(\text{Me}_3\text{NTB})$ precursor (Me_3NTB = tris((1-



Scheme 1 (A) Proposed catalytic cycle of sMMO and (B) generation of intermediates **2**, **2+H⁺**, **2+Sc**, and **3** in the biomimetic system.

^aDepartment of Chemistry, University of Minnesota, Minneapolis, MN 55455, USA. E-mail: larryque@umn.edu

^bDepartment of Chemistry, Carnegie Mellon University, Pittsburgh, PA 15213, USA. E-mail: ysguo@andrew.cmu.edu

^cIQCC and Department of Chemistry, University of Girona, 17003 Girona, Spain. E-mail: marcel.swart@udg.edu

^dICREA, Pg. Lluis Companys 23, 08010 Barcelona, Spain

[†] Current address: Dept. of Chemistry, University of Chicago, Chicago, IL 60637 (USA).

[‡] These authors contributed equally.

methyl-1*H*-benzo[*d*]imidazole-2-yl)-methyl)amine) that forms a diiron(II) derivative $[(\text{Me}_3\text{NTB})\text{Fe}^{\text{II}}(\mu\text{-OH})_2\text{Fe}^{\text{II}}(\text{Me}_3\text{NTB})]^{2+}$ (**1**) upon treatment with a suitable base, and is converted upon exposure to dioxygen into a $(\mu\text{-oxo})(\mu\text{-1,2-peroxo})$ diiron(III) species **2** $[(\text{Me}_3\text{NTB})\text{Fe}^{\text{III}}(\mu\text{-O})(\mu\text{-1,2-O}_2)\text{Fe}^{\text{III}}(\text{Me}_3\text{NTB})]^{2+}$. This diferric species in turn transforms into the diferryl diamond core complex **3** $[(\text{Me}_3\text{NTB})\text{Fe}^{\text{IV}}(\mu\text{-O})_2\text{Fe}^{\text{IV}}(\text{Me}_3\text{NTB})]^{4+}$ upon treatment with $\text{Sc}(\text{OTf})_3$.¹⁶ In this study, we report the characterization of a previously unidentified short-lived intermediate that forms immediately upon addition of Sc^{3+} to a solution of **2** in the course of its conversion into **3** (Scheme 1B). The fleeting **2+Sc** complex represents the first biomimetic example of a diferric-peroxo complex to be trapped with a bound Lewis acid prior to its conversion into a diiron(IV) complex having an $\text{Fe}^{\text{IV}}_2(\mu\text{-O})_2$ diamond core.

Results and discussion

Synthesis and UV-Vis characterization of **2+Sc** complex

The generation and characterization of $(\mu\text{-oxo})(\mu\text{-1,2-peroxo})$ diiron(III) complex (**2**) has been previously reported by our group (Scheme 1B).¹⁶ Treatment of a solution of **2** in MeCN at 233 K with 1 equiv. $\text{Sc}(\text{OTf})_3$ (or, Sc^{3+}) results in an immediate change in its UV-Vis features (Fig. 1 and S5), forming a species designated as **2+Sc** with a peak at *ca.* 350 nm, and a less intense visible band at *ca.* 490 nm. These peaks are blue-shifted relative to **2**, which showed bands at *ca.* 400 nm and 595 nm.¹⁶ An identical UV-Vis spectrum (Fig. S1) is generated directly upon the addition of 2 equiv. Sc^{3+} to **2**. However, over the course of an hour, this addition of a second equivalent of Sc^{3+} results in a red-shift to *ca.* 400 nm and *ca.* 600 nm for the less intense band (corresponding to **3**). Clearly, one equiv. Sc^{3+} is sufficient to generate the **2+Sc** adduct, but adding the second equivalent of Sc^{3+} to this solution is required to generate **3**. These observations suggest that the first Sc^{3+} ion binds to the O–O bond, and only after the addition of the second Sc^{3+} ion does O–O bond cleavage occurs.

Similar blue shifts (as observed for the change from **2** to **2+Sc**) have previously been observed in the UV-Vis features of mononuclear $\text{Fe}^{\text{III}}\text{-O}_2^{2-}$ complexes upon treatment with Lewis acids,^{18–20} as well as in di-copper-peroxo systems that bind exogenous Lewis acids.^{21–23} This is in contrast to the red-shift

typically found upon addition of protic acids to diferric-peroxo intermediates that result from protonation of the bridging oxo moiety.^{15,16} The opposite effects of protic and Lewis acids on the peroxo-based UV-Vis feature strongly suggest that the blue shift of the peroxo band of **2** upon Sc^{3+} addition comes from the interaction of the Sc^{3+} ion with the peroxo moiety (and not with the oxo bridge), resulting in a decrease in the basicity of the peroxo ligand that promotes the cleavage of the O–O bond.

Crucially, **2+Sc** persists for a few seconds at 233 K before undergoing a rearrangement over an induction period of *ca.* 10–15 minutes (Fig. S5), eventually converting into **3** (Fig. 1, right side). This time-lapse allows **2+Sc** to be trapped by rapid freeze-quenching for characterization by a variety of spectroscopic methods. While one equivalent of Sc^{3+} generates **2+Sc** (as observed by UV-Vis absorption, Fig. 1, left), the addition of the second equivalent allows us to follow the conversion of **2** to **3** by spectroscopic methods.

Resonance Raman spectroscopy

Resonance Raman spectroscopy has proven to be a useful probe for the features found in the visible spectra of **2** and **2+Sc** (Fig. 2). The O–O vibration for **2** is observed at 825 cm^{-1} and red-shifts by 9 cm^{-1} to 816 cm^{-1} upon addition of Sc^{3+} , reflecting the interaction of Sc^{3+} with the 1,2-peroxo bridge. To observe the corresponding O–O vibration of **2+Sc** requires irradiation with a 457 nm laser, due to the blue shift in the peroxo-to-iron charge transfer band upon Sc binding. No resonance-enhanced peaks are observed with the use of a 516 nm laser, as used to characterize **2**. The red-shift in the O–O vibration for **2+Sc** is about half as large as that found for Sc^{3+} binding to $[\text{Fe}^{\text{III}}(\text{TMC})(\eta^2\text{-O}_2)]^+$ from 826 to 807 cm^{-1} ,¹⁹ reflecting the higher basicity of the side-on bound peroxo moiety in the latter complex.

Interestingly, four (Fe–O)-based vibrations observed in **2** (at 715 , 527 , 518 , and 454 cm^{-1})¹⁶ disappear upon Sc^{3+} addition (Fig. S6), namely the $\nu_{\text{sym}}(\text{Fe-O}_2\text{-Fe})$, $\nu_{\text{asym}}(\text{Fe-O}_2\text{-Fe})$, $\nu_{\text{sym}}(\text{Fe-O-Fe})$, and $\nu_{\text{asym}}(\text{Fe-O-Fe})$ features, leaving the O–O stretch at 825 cm^{-1} (red-shifted to 816 cm^{-1}) to be the only vibration

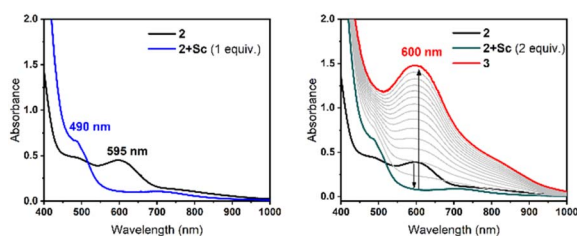


Fig. 1 UV-Vis spectral changes at -40°C upon treating 0.50 mM **2** in MeCN with 1 equiv. Sc^{3+} to generate **2+Sc** (left, blue line) and with 2 equiv. of Sc^{3+} (right, cyan line) *en route* to its conversion to **3** (spectral changes after adding 2 equiv. Sc^{3+} , right, spectra taken every 5 minutes).

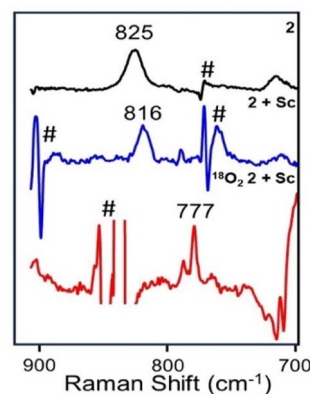


Fig. 2 Resonance Raman spectra at 77 K with 135° backscattering of **2** (561 nm excitation) and **2+Sc** (457 nm excitation). Black: $0.5\text{ mM }^{16}\text{O}_2\text{-2}$ in MeCN; Blue: $0.5\text{ mM }^{16}\text{O}_2\text{-2+Sc}$ in MeCN; Red: $0.5\text{ mM }^{18}\text{O}_2\text{-2+Sc}$ in CD_3CN . # indicates artifacts from solvent subtraction.



observed in its resonance Raman spectrum. This dramatic change shows Sc^{3+} binding to **2** to induce a structural alteration significantly distinct from protonation, leading to a distortion of the relatively planar $\text{Fe}^{\text{III}}(\mu\text{-O})(\mu\text{-1,2-O}_2)\text{Fe}^{\text{III}}$ moiety that disrupts the well-established vibrational patterns associated with such cores.^{6,7,15,16}

The interaction of Sc^{3+} with **2** differs substantially from that of adding a proton to the oxo bridge of **2**, for which Raman vibrations at 820 [$\nu(\text{O-O})$], 527 [$\nu_{\text{asym}}(\text{Fe-O}_2\text{-Fe})$], and 449 [$\nu_{\text{sym}}(\text{Fe-O}_2\text{-Fe})$] cm^{-1} are observed, the latter two being associated with deformations of the Fe-O-O-Fe unit in **2+H**.¹⁶

Note that our DFT calculations predict that the proton does not bind to the oxo bridge, but instead protonates one of the benzimidazole rings (Fig. 3, bottom). All attempts to optimize structures of **2** with a protonated oxo bridge lead to an Fe-Fe distance of 3.38 Å, or even an opened up [(L)(OH)Fe^{III}($\mu\text{-1,2-O}_2$)Fe^{III}(L)(MeCN)]³⁺ complex with an Fe...Fe distance of 4.39 Å. Instead, protonation of one of the benzimidazoles affords an Fe-Fe distance of 3.05 Å, leading to computed $\nu(\text{O-O})$, $\nu_{\text{asym}}(\text{Fe-O}_2\text{-Fe})$, and $\nu_{\text{sym}}(\text{Fe-O}_2\text{-Fe})$ vibrations that nearly match those of **2** itself.¹²

The **2+Sc** sample obtained using ¹⁸O₂ in CD₃CN shows three peaks at 785, 777, and 760 cm^{-1} in place of the sole feature at 816 cm^{-1} found in the corresponding ¹⁶O₂ experiment in CH₃CN. As the features at 760 cm^{-1} and 785 cm^{-1} can also be observed as shoulders to solvent-derived peaks in the ¹⁶O spectrum obtained in CH₃CN shown in Fig. 2, only the vibration at 777 cm^{-1} can be associated with the labeled O-O vibration of

2+Sc. In addition, there is a broad peak at 690 cm^{-1} that could arise from an Fe-O-Fe vibration,^{1,24} but this feature persists even in the spectrum of thermally-decayed **2+Sc** and thus must arise from some other byproduct.

Mössbauer and X-ray absorption spectroscopy

Mössbauer spectroscopy shows that the **2+Sc** adduct (Fig. 4) differs significantly from previously reported **2**.¹⁶ Complex **2** exhibits a single quadrupole doublet with $\delta = 0.49 \text{ mm s}^{-1}$ and $\Delta E_{\text{Q}} = 1.06 \text{ mm s}^{-1}$, indicating a twofold-symmetric anti-ferromagnetically coupled diferric center. In contrast, **2+Sc** exhibits a prominent pair of doublets with comparable intensities that can be associated with an antiferromagnetically coupled pair of high-spin diferric centers, with $\delta = 0.48 \text{ mm s}^{-1}$ and $\Delta E_{\text{Q}} = 1.60 \text{ mm s}^{-1}$ (for **2+Sc A**) and $\delta = 0.46 \text{ (A) mm s}^{-1}$ and $\Delta E_{\text{Q}} = 0.81 \text{ mm s}^{-1}$ for (**2+Sc B**), together representing 74% of the total Fe in the sample. There is also a third diferric type quadrupole doublet **2'** representing 18% of the total Fe, with parameters $\delta = 0.50 \text{ mm s}^{-1}$ and $\Delta E_{\text{Q}} = 1.14 \text{ mm s}^{-1}$ that are similar but distinct to those of **2**. However, unlike **2+Sc A** and **2+Sc B**, **2'** does not convert to **3**, as shown in Fig. 4C, which leads us to tentatively assign this minor di-ferric type species as a minor byproduct of little relevance to the chemistry of interest, and most likely no longer contains Sc.

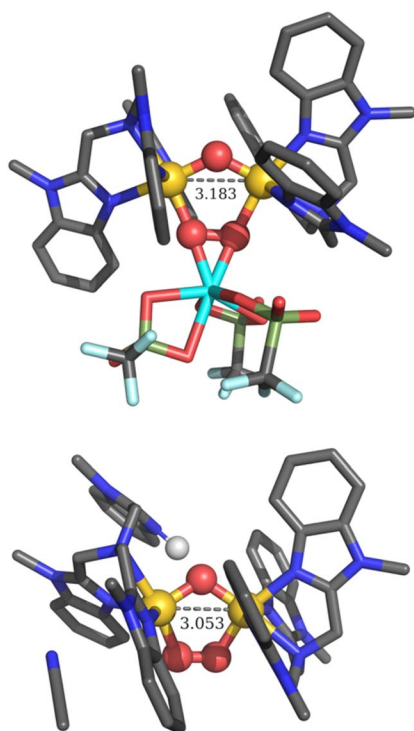


Fig. 3 Top: DFT optimized structure of **2+Sc**. Bottom: DFT computed structure (r2scan-3c, COSMO) for **2+H**, which exhibits the short Fe...Fe distance as previously inferred from EXAFS analysis.¹⁶

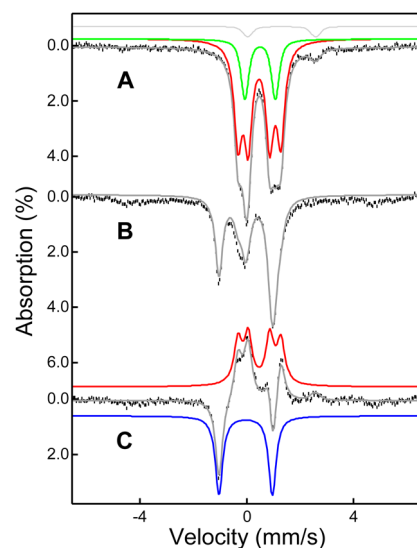


Fig. 4 4.2 K zero field Mössbauer spectra (vertical bars) of **2+Sc** (panel A) and upon conversion of **2+Sc** into **3** (panel B). Panel C shows the B – A difference spectrum highlighting the conversion of **2+Sc** to **3**. Panel A shows an overall simulation (grey solid line) with four species. The red solid line containing two distinct ferric doublets of equal intensity represents 74% of the iron in the sample and is assigned to the dominant **2+Sc** species with distinct Fe^{III} units. The green solid line represents 18% of the iron in the sample corresponding to a symmetric diferric species. The light grey line represents a minor ferrous species (~5%), likely to be the starting ferrous complex. Panel B shows an overall simulation (grey solid line) with 50% **3**, 25% **2+Sc** and 18% **2'**. Panel C shows the conversion of **2+Sc** (red solid line) into **3** (blue solid line).



Complex **2+Sc** shows a K-edge X-ray absorption near edge (XANES) feature at an energy that is 0.6 eV blue-shifted relative to that of **2** (Fig. S4), indicating a lower electron density on the ferric ions. Such a blue-shift may result from the decreased interaction between the O_2^{2-} ligand and Fe^{3+} ions due to competition with Sc^{3+} binding. Its pre-edge area decreases only slightly upon Sc^{3+} binding, from 18.2 units to 17.5 units (see Fig. S3 in SI), reflecting only a slight change to the symmetry around the iron center, in contrast to the larger effect of adding H^+ to **2**. Although **2+H⁺** has a K-edge energy of 7126.7 eV that is intermediate between those of **2** and **2+Sc**, its pre-edge area is significantly smaller due to the weakening of the Fe–oxo bonds upon protonation. In addition, the pre-edge region of **2+H⁺** is best fitted with two features, indicating a significant differentiation between the two (Brønsted/Lewis) acid complexes of **2**. Our DFT study reproduces this change in the number of features when going from **2** to **2+H** (Fig. S7). Clearly, **2** is affected quite differently by its interactions with H^+ and Sc^{3+} .

As expected, the structure for **2+Sc**, as derived from EXAFS analysis (Fig. 5), significantly differs from those of both the parent complex and the protonated complex **2+H⁺**. The observed Fe...Fe distance lengthens to 3.15 Å (vs. 3.07 and 3.09 Å for **2** and **2+H⁺**, respectively), and a Sc scatterer is found at 4.08 Å. When the latter is excluded from the fit, there is a prominent unassigned peak in the Fourier-transformed EXAFS spectrum, lending strong support for a Sc^{3+} -bound peroxo moiety. While it is possible that a scatterer at this distance could arise from a multiple scattering pathway in the imidazole of the ligand backbone, careful analysis of the EXAFS data precludes such a conclusion. The goodness of fit of the EXAFS data with a carbon shell at the same distance as the Sc shell (with no Sc included) is noticeably worse (Table S2, fit 6 compared to fit 5). Additionally, no such ligand-derived scatterers were observed in the initial report of **2**.¹⁶ DFT calculations support the results of this analysis, predicting an Fe...Fe distance of 3.183 Å and Fe...Sc distances of 4.10 Å (Fe1...Sc) and 4.01 Å (Fe2...Sc), in excellent agreement with experimental

results. The Sc...O distances in **2+Sc** are also slightly different, at 2.04 Å ($\text{O1}\cdots\text{Sc}$) vs. 2.08 Å ($\text{O2}\cdots\text{Sc}$).

Comparison with literature results

To the best of our knowledge, this study provides the first documented example of a Lewis acid adduct to a diferric-peroxo complex. To put this novel family of complexes into perspective, we compare their properties with those observed for other (μ -1,2-peroxo)diferric complexes in the literature.⁶ In previous work, our laboratory has noted a correlation between the diiron distance and the O–O vibrational frequency (Fig. 6). Within this context, the parent complex **2** has parameters that place it closest to the established trendline among the four related complexes derived from **2**. **2+H⁺** falls further away and **2+Sc** is even further. Clearly, an even more drastic structural change occurs upon Sc^{3+} binding, which is also reflected by the loss of all Fe–O vibrations (Fig. 2). This distinct behavior likely reflects the loss of the relatively planar [$\text{Fe}^{\text{III}}_2(\mu\text{-O})(\mu\text{-1,2-O}_2)$] framework found in **2** and emphasizes the unique role Sc^{3+} plays in promoting O–O bond cleavage.

The observed activation of the O–O bond by Sc^{3+} in this work stands in sharp contrast to the effect of adding a proton observed for other diferric-peroxo systems. For complexes supported by BnBQA (= *N*-benzyl-1-(quinolinyl)-*N*-(quinolinylmethyl)methanamine) and 6-Me₂-BPP (= bis(((6-methylpyridinyl)methyl)-glycine), protonation of their respective oxo bridges results in higher frequency O–O vibrations (854 vs. 928 cm^{-1} and 847 vs. 908 cm^{-1} , respectively).^{7,15} Protonation also does not lead to the formation of diiron(IV) species for the BnBQA and the 6Me₃TPA (= tris((6-methylpyridinyl)methyl)amine) complexes. Instead, both complexes simply decay upon addition of a protic acid to form $\text{Fe}^{\text{III}}\text{Fe}^{\text{IV}}$ products. In

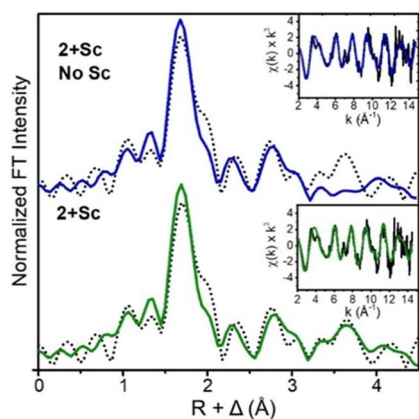


Fig. 5 Fourier transformed EXAFS and *k*-space EXAFS (insets) spectra of **2+Sc**, with the Sc scatterer at 4.08 Å included in the green colored spectral fits.

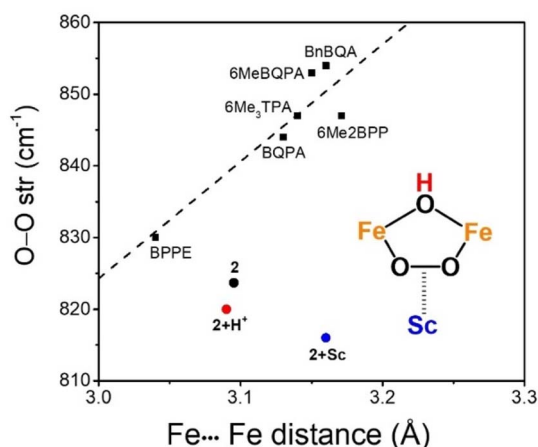


Fig. 6 Empirically derived correlation between O–O vibrational frequencies for nonheme (μ -oxo)diferric-peroxo intermediates and their diiron distances as deduced by EXAFS analysis. All points representing the oxo/peroxo complexes fall close to the dashed line. Compared to the other complexes shown, **2** falls furthest away from the dashed line, but even larger deviations are found for **2+H⁺** and **2+Sc**, reflecting more significant structural alterations relative to that of **2**.



contrast to what is observed upon protonation in the BnBQA and 6Me₃TPA systems, **2** converts into **3** upon addition of Sc³⁺, promoting O–O bond scission to generate the Fe^{IV}₂(μ-O)₂ diamond core of **3**.

This outcome parallels the behavior observed for mono-nuclear [Fe^{III}-η₂-O₂] complexes, which undergo O–O bond scission upon treatment with Lewis acids, leading to the subsequent formation of Fe^{IV} = O products with the help of an electron donor.^{18,19,25} This is not the case in the diferric system **2**, where the two required reducing equivalents needed to fully activate O₂ come from both irons. It was initially proposed, based on resonance Raman labeling experiments, that the Sc³⁺ ion scavenged the bridging oxo to allow for the dinuclear core to isomerize to a form suited for O–O cleavage. However, based on our current results, Sc³⁺ in fact plays a crucial bifunctional role by first priming the O–O bond for cleavage and subsequently scavenging the bridging oxo ligand.

The O–O bond in monoferric-peroxo complexes would appear to be more activated by binding Lewis acids than that of **2+Sc**. For example, in the case of [(TMC)Fe^{III}(η₂-O₂)]⁺ (TMC = 1,4,8,11-tetramethyl-1,4,8,11-tetraazacyclotetradecane), the O–O vibration shifts from 826 cm⁻¹ to 807 cm⁻¹ upon Sc³⁺ binding,¹⁹ which is almost twice as pronounced as the 10 cm⁻¹ red shift observed upon the conversion of **2** to **2+Sc**. The Fe...Sc³⁺ distance also differs significantly between mono- and diferric systems. EXAFS analysis of the [(TMC)Fe^{III}(μ-η₂:η₂-O₂)Sc]⁴⁺ adduct reveals an Fe...Sc³⁺ distance of 3.84 Å, which concurs with the 3.80 Å distance derived from DFT,¹⁸ but is significantly shorter than the 4.08 Å distance found for **2+Sc** by EXAFS analysis. Although this difference between mono- and diferric systems is not unexpected, it highlights the geometric constraints imparted by the dinuclear system.

Conclusions

The data presented in this paper represent the first documented evidence for Lewis-acid adduct formation to a 1,2-peroxodiferric species derived from the reaction of a synthetic diiron(II) complex with O₂ en route to its conversion into an Fe^{IV}₂(μ-O)₂ diamond core. This sequence of transformations mimics the steps involved in the conversion of the diiron(II) active site of soluble methane monooxygenase hydroxylase (sMMOH) upon exposure to O₂, namely the initial formation of the 1,2-peroxodiferric derivative **P** and its subsequent conversion into the diiron(IV) oxidant **Q**. Our results thus provide strong evidence for the formation of a Sc³⁺ adduct to the peroxo intermediate prior to inducing O–O bond scission to form a diiron(IV) complex with an Fe₂(μ-O)₂ diamond core.

Abbreviations

sMMOH	Soluble methane monooxygenase
BDfE	Bond dissociation free energy

Author contributions

Patrick M Crossland and Bittu Chandra contributed equally to this work. The manuscript was written through contributions of all authors. All authors have approved the final version of the manuscript.

Conflicts of interest

There are no conflicts to declare.

Data availability

The computational data have been uploaded to iochem-bd at <https://doi.org/10.19061/iochem-bd-4-90>, all the other data are available in the supplementary information (SI). Supplementary information: additional XAS data, experimental procedures, and methods. See DOI: <https://doi.org/10.1039/d5sc05667e>.

Acknowledgements

This work was supported by U. S. National Institutes of Health grants R35 GM-131721 to L. Q. (University of Minnesota) and R01 GM-125924 to Y. G. (Carnegie Mellon University), AEI/MCIN (grants PID2020-114548GB-I00 and PID2023-152415NB-I00 to M. S.), GenCat (grant 2021SGR00487) and a developer's license to M. S. by Software for Chemistry and Materials (SCM). XAS data were collected at the Stanford Synchrotron Radiation Lightsource Beamline 9-3. Use of the Stanford Synchrotron Radiation Lightsource, SLAC National Accelerator Laboratory, is supported by the U.S. Department of Energy, Office of Science, Office of Basic Energy Sciences under Contract No. DE-AC02-76SF00515. The SSRL Structural Molecular Biology Program is supported by the DOE Office of Biological and Environmental Research, and by the National Institutes of Health, National Institute of General Medical Sciences (P30GM133894). The contents of this publication are solely the responsibility of the authors and do not necessarily represent the official views of NIGMS or NIH. PC would like to acknowledge Dr Andrew J. Jasniowski for helpful discussions regarding XAS interpretation and Drs Jorge Martinez and Norman Zhao for useful discussions about data presentation.

Notes and references

- 1 A. J. Jasniowski and L. Que, Dioxygen Activation by Nonheme Diiron Enzymes: Diverse Dioxygen Adducts, High-Valent Intermediates, and Related Model Complexes, *Chem. Rev.*, 2018, **118**, 2554–2592.
- 2 R. Banerjee, J. C. Jones and J. D. Lipscomb, Soluble Methane Monooxygenase, *Annu. Rev. Biochem.*, 2019, **88**, 409–431.
- 3 S.-K. Lee and J. D. Lipscomb, Oxygen Activation Catalyzed by Methane Monooxygenase Hydroxylase Component: Proton Delivery during the O–O Bond Cleavage Steps, *Biochemistry*, 1999, **38**(14), 4423–4432.



- 4 C. E. Tinberg and S. J. Lippard, Revisiting the Mechanism of Dioxygen Activation in Soluble Methane Monooxygenase from *M. Capsulatus* (Bath): Evidence for a Multi-Step, Proton-Dependent Reaction Pathway, *Biochemistry*, 2009, **48**(51), 12145–12158.
- 5 R. Banerjee, Y. Proshlyakov, J. D. Lipscomb and D. A. Proshlyakov, Structure of the Key Species in the Enzymatic Oxidation of Methane to Methanol, *Nature*, 2015, **518**(7539), 431–434.
- 6 A. T. Fiedler, X. Shan, M. P. Mehn, J. Kaizer, S. Torelli, J. R. Frisch, M. Kodera and L. Que, Spectroscopic and Computational Studies of (μ -Oxo)(μ -1, 2-Peroxo)Diiron(III) Complexes of Relevance to Nonheme Diiron Oxygenase Intermediates, *J. Phys. Chem. A*, 2008, **112**(50), 13037–13044.
- 7 X. Zhang, H. Furutachi, S. Fujinami, S. Nagatomo, Y. Maeda, Y. Watanabe, T. Kitagawa and M. Suzuki, Structural and Spectroscopic Characterization of (μ -Hydroxo or μ -Oxo)(μ -Peroxo)Diiron(III) Complexes: Models for Peroxo Intermediates of Non-Heme Diiron Proteins, *J. Am. Chem. Soc.*, 2005, **127**(3), 826–827.
- 8 J. R. Frisch, V. V. Vu, M. Martinho, E. Münck and L. Que, Characterization of Two Distinct Adducts in the Reaction of a Nonheme Diiron(II) Complex with O₂, *Inorg. Chem.*, 2009, **48**(17), 8325–8336.
- 9 A. J. Skulan, M. A. Hanson, H. F. Hsu, L. Que and E. I. Solomon, Spectroscopic Study of [Fe₂O₂(5-Et₃-TPA)₂]³⁺: Nature of the Fe₂O₂ Diamond Core and Its Possible Relevance to High-Valent Binuclear Non-Heme Enzyme Intermediates, *J. Am. Chem. Soc.*, 2003, **125**, 7344–7356.
- 10 G. Xue, D. Wang, R. De Hont, A. T. Fiedler, X. Shan, E. Münck and L. Que, A Synthetic Precedent for the [FeIV₂(μ -O)₂] Diamond Core Proposed for Methane Monooxygenase Intermediate Q, *Proc. Natl. Acad. Sci. U. S. A.*, 2007, **104**, 20713–20718.
- 11 J. S. Pap, A. Draksharapu, M. Giorgi, W. R. Browne, J. Kaizer and G. Speier, Stabilisation of μ -Peroxido-Bridged Fe(III) Intermediates with Non-Symmetric Bidentate N-Donor Ligands, *Chem. Commun.*, 2014, **50**(11), 1326–1329.
- 12 T. Ookubo, H. Sugimoto, T. Nagayama, H. Masuda, T. Sato, K. Tanaka, Y. Maeda, H. Okawa, Y. Hayashi, A. Uehara and M. Suzuki, cis- μ -1,2-Peroxo Diiron Complex: Structure and Reversible Oxygenation, *J. Am. Chem. Soc.*, 1996, **118**(3), 701–702.
- 13 M. Kodera, Y. Taniike, M. Itoh, Y. Tanahashi, H. Shimakoshi, K. Kano, S. Hirota, S. Iijima, M. Ohba and H. Okawa, Synthesis, Characterization, and Activation of Thermally Stable μ -1,2- Peroxodiiron(III) Complex, *Inorg. Chem.*, 2001, **40**(19), 4821–4822.
- 14 S. V. Kryatov, S. Taktak, I. V. Korendovych, E. V. Rybak-Akimova, J. Kaizer, S. Torelli, X. Shan, S. Mandal, V. L. MacMurdo, A. Mairata I Payeras and L. Que, Dioxygen Binding to Complexes with FeII 2(μ -OH)₂ Cores: Steric Control of Activation Barriers and O₂- Adduct Formation, *Inorg. Chem.*, 2005, **44**(1), 85–99.
- 15 M. A. Cranswick, K. K. Meier, X. Shan, A. Stubna, J. Kaizer, M. P. Mehn, E. Mu and L. Que, Protonation of a Peroxodiiron(III) Complex and Conversion to a Diiron(III/IV) Intermediate: Implications for Proton-Assisted O-O Bond Cleavage in Nonheme Diiron Enzymes, *Inorg. Chem.*, 2012, (51), 10417–10426.
- 16 S. Banerjee, A. Draksharapu, P. M. Crossland, R. Fan, Y. Guo, M. Swart and L. Que, Sc³⁺-Promoted O-O Bond Cleavage of a (μ -1,2-Peroxo)Diiron(III) Species Formed from an Iron(II) Precursor and O₂ to Generate a Complex with an FeIV₂(μ -O)₂Core, *J. Am. Chem. Soc.*, 2020, **142**, 4285–4297.
- 17 V. L. MacMurdo, H. Zheng and L. Que, Model for the Cofactor Formation Reaction of *E. Coli* Ribonucleotide Reductase. From a Diiron(II) Precursor to an Fe(III)Fe(IV) Species via a Peroxo Intermediate, *Inorg. Chem.*, 2000, **39**(11), 2254–2255.
- 18 Y. M. Lee, S. Bang, Y. M. Kim, J. Cho, S. Hong, T. Nomura, T. Ogura, O. Troeppner, I. Ivanović-Burmazović, R. Sarangi, S. Fukuzumi and W. Nam, A Mononuclear Nonheme Iron(III)-Peroxo Complex Binding Redox-Inactive Metal Ions, *Chem. Sci.*, 2013, **4**(10), 3917–3923.
- 19 F. Li, K. M. Van Heuvelen, K. K. Meier, E. Münck and L. Que, Sc³⁺-Triggered Oxoiron(IV) Formation from O₂ and Its Non-Heme Iron(II) Precursor via a Sc³⁺-Peroxo-Fe³⁺ Intermediate, *J. Am. Chem. Soc.*, 2013, **135**(28), 10198–10201.
- 20 S. Bang, Y. M. Lee, S. Hong, K. B. Cho, Y. Nishida, M. S. Seo, R. Sarangi, S. Fukuzumi and W. Nam, Redox-Inactive Metal Ions Modulate the Reactivity and Oxygen Release of Mononuclear Non-Haem Iron(III)-Peroxo Complexes, *Nat. Chem.*, 2014, **6**(10), 934–940.
- 21 K. E. Dalle, T. Gruene, S. Dechert, S. Demeshko and F. Meyer, Weakly Coupled Biologically Relevant CuII₂(μ -H1:H1-O₂) Cis -Peroxo Adduct That Binds Side-on to Additional Metal Ions, *J. Am. Chem. Soc.*, 2014, **136**(20), 7428–7434.
- 22 A. Brinkmeier, K. E. Dalle, L. D'Amore, R. A. Schulz, S. Dechert, S. Demeshko, M. Swart and F. Meyer, Modulation of a μ -1,2-Peroxo Dicopper(II) Intermediate by Strong Interaction with Alkali Metal Ions, *J. Am. Chem. Soc.*, 2021, **143**(42), 17751–17760.
- 23 N. P. Vargo, J. B. Harland, B. W. Musselman, N. Lehnert, M. Z. Ertem and J. R. Robinson, Calcium-Ion Binding Mediates the Reversible Interconversion of Cis and Trans Peroxido Dicopper Cores, *Angew. Chem., Int. Ed.*, 2021, **60**(36), 19836–19842.
- 24 E. C. Wilkinson, Y. Dong, Y. Zang, H. Fujii, R. Faczkiewicz, G. Fraczkiewicz, R. S. Czernuszewicz and L. Que, Raman Signature of the Fe₂O₂ “diamond” Core, *J. Am. Chem. Soc.*, 1998, **120**, 955–962.
- 25 A. Yokoyama, J. E. Han, K. D. Karlin and W. Nam, An Isoelectronic NO Dioxygenase Reaction Using a Nonheme Iron(III)-Peroxo Complex and Nitrosonium Ion, *Chem. Commun.*, 2014, **50**(14), 1742–1744.
- 26 M. C. White, A. G. Doyle and E. N. Jacobsen, A Synthetically Useful, Self-Assembling MMO Mimic System for Catalytic Alkene Epoxidation with Aqueous H₂O₂, *J. Am. Chem. Soc.*, 2001, **123**, 7194–7195.
- 27 H. Guan, C.-H. Tung and L. Liu, Methane Monooxygenase Mimic Asymmetric Oxidant: Self-Assembling μ Hydroxo, Carboxylate-Bridged Diiron(III) Catalyzed Enantioselective Dehydrogenation, *J. Am. Chem. Soc.*, 2022, **144**, 5976–5984.

