

PAPER

[View Article Online](#)
[View Journal](#) | [View Issue](#)Cite this: *Nanoscale Adv.*, 2025, 7, 1671

X-ray photoelectron spectroscopy of metal oxide nanoparticles: chemical composition, oxidation state and functional group content†

Gregory P. Lopinski,^a Oltion Kodra,^b Filip Kunc,^a David C. Kennedy,^a Martin Couillard^b and Linda J. Johnston^a

Surface chemistry drives the interaction of a material with its surroundings, therefore it can be used to understand and influence the fate of nanomaterials when used as functional materials or when released to the environment. Here we have used X-ray photoelectron spectroscopy (XPS) to probe the chemical composition, oxidation state and functional group content of the near surface region of four families of commercially available metal oxide nanoparticles from several different suppliers. The analyzed nanoparticles varied in size and surface functionalization (unfunctionalized vs. amine, stearic acid, and PVP-coated samples). Survey and high-resolution scans have provided information on the atomic composition of the samples, including an estimate of the stoichiometry of the metal oxide, the presence of functional groups and the identification and quantification of any impurities on the surface. The presence of significant impurities for some samples and the variation from the expected oxidation state in other cases are relevant to studies of the environmental and health impacts of these materials as well as their use in applications. The functional group content measured by XPS shows a similar trend to earlier quantitative nuclear magnetic resonance (qNMR) data for aminated samples. This indicates that XPS can be a complementary probe of surface functional group content in cases where the functional group contains a unique element not otherwise present on the nanoparticles.

Received 15th November 2024
Accepted 3rd December 2024

DOI: 10.1039/d4na00943f

rsc.li/nanoscale-advances

1 Introduction

The importance of thorough characterization of the physical chemical properties of nanomaterials is increasingly recognized as an essential element for quality control during production, reproducible use in a variety of applications and investigations of potential environmental health and safety concerns.¹ Despite the availability of validated methods that have been tested in round robin comparisons for many properties, there are still gaps in availability of standard protocols for assessing some measurands.^{2,3} For example, surface chemistry is widely recognized as a key parameter for assessment of nanomaterials since it is often the main factor that controls the interaction of the material with its environment, both for applications where the nanomaterial is part of a complex assembly with nano-enabled properties and in scenarios where release of the material results in localization in environmental samples or ingestion by organisms.^{4,5} Nanomaterial surfaces are frequently modified by

adsorption or by covalent attachment of functional groups (FGs) in order to control the colloidal stability of the material, to ensure compatibility of the material with its surroundings for various applications and to allow for targeting of the material for biological applications. In addition, the surface composition may be different from the “bulk” due to the presence of surface impurities or changes in metal oxidation state at the surface.⁶ Thorough characterization is particularly important in the case of commercially available nanomaterials as data provided by the suppliers is not always complete and/or reliable.

Detailed studies of surface chemistry are complicated by a number of factors.⁵ These include the wide range of compositions of the nanomaterial and possible surface modifications and impurities, the compatibility of methods with different material compositions, and the fact that some methods infer surface properties from bulk measurements while others interrogate a thin surface layer. Bulk methods include thermogravimetric analysis (TGA), which is most useful when combined with FT-IR or mass spectrometry of evolved gases to provide structural information, quantitative NMR (qNMR) in either solution or the solid state and ICP-MS of dissolved materials.^{7–11} Both qNMR and ICP-MS have the advantages of providing accurate and traceable measurements, information on the chemical identity of the surfaces or FGs and potentially also information on possible impurities.

^aMetrology Research Centre, National Research Council Canada, Ottawa, ON, K1A 0R6, Canada. E-mail: gregory.lopinski@nrc-cnrc.gc.ca^bClean Energy Innovation Research Centre, National Research Council Canada, Ottawa, ON, K1A 0R6, Canada† Electronic supplementary information (ESI) available. See DOI: <https://doi.org/10.1039/d4na00943f>

However they typically involve optimized extraction of surface groups or dissolution of the sample and are not surface specific, providing compositional information for the entire sample.^{11,12} Methods with more surface sensitivity include X-ray photoelectron spectroscopy (XPS) which provides atomic composition of the near surface region and avoids time-consuming sample preparation steps, as well as optical probe methods which give estimates of probe-accessible surface groups and conductometry which can be used to quantify acidic or basic FGs for some materials.^{4,8,9,13–15}

Previous work from our group has used a combination of qNMR, XPS, TGA and optical probe methods to quantify surface coatings and FGs on a range of commercial silica and zinc oxide nanoparticles (NPs), providing a detailed evaluation of the relative merits and the range of applicability of the various methods.^{11,16,17} More recently we have carried out detailed physical chemical characterization of representative nanoparticles of four families of metal oxides prior to investigating their potential deleterious biological effects using assays to evaluate cytotoxicity and the role of reactive oxygen species.^{13,18,19} That work measured particle size using electron microscopy and dynamic light scattering, surface charge, specific surface area and, where relevant, qNMR measurement of surface coating or FG content. We have now completed a detailed XPS study of the same four families of commercial metal oxide nanomaterials from different suppliers and with varying sizes and surface chemistries. XPS is a valuable method for assessing the atomic composition of planar surfaces and is being increasingly applied to study the surface of nanomaterials.^{4,20–22} The XPS signal comes from the near surface region, with the probing depth limited by the mean free path of the photoelectrons (~5 nm in the current work). Other advantages of this method are that it requires minimal sample preparation and is compatible with most material compositions. XPS can provide information on the metal oxidation state in the vicinity of the surface, which may differ considerably from the bulk, as well as the presence of impurities that are often related to the synthesis method or to the presence of adventitious contamination. XPS can also be used to quantify FG content, particularly for organic coatings or FGs that contain elements such as sulfur, nitrogen or halogens.

In this paper we report XPS results that examine the chemical composition and metal oxidation state for a total of 35 different CeO₂, NiO, Fe₂O₃ and Mn₂O₃ nanomaterials that have been studied in earlier work. We also examine FG content for the 16 of these samples that have been modified to add a stearic acid or polymer coating or covalently modified with silane chemistry to attach an aminoalkyl functionality and correlate the results to those previously obtained by qNMR for some of the same samples. The types of metal oxide nanoparticles investigated here are currently being explored for a diverse range of applications including gas sensing, biomedicine, wastewater treatment and energy generation/storage.^{23–27} The surface chemical characterization of these nanoparticles provided by XPS contributes new information that is expected to help assess their suitability for these and other applications.

2 Materials and methods

2.1 Materials

Four series of bare and surface modified NiO, CeO₂, Fe₂O₃ and Mn₂O₃ NPs were purchased from several commercial suppliers. A total of 33 materials, some unfunctionalized and some modified with amine, stearic acid or polyvinylpyrrolidone surface functional groups or coatings were purchased from eight suppliers (US Research Nanomaterials, mkNano, Sigma, Skyspring, American Elements, Lanxess, Nano and Amorphous Materials and Nanografi). Two additional functionalized samples were prepared in-house. In the tables, figures and discussion to follow, samples will be referred to by a sample code (*i.e.* Ni-01). Surface modifications are often described by abbreviations (SA = stearic acid, PVP = polyvinylpyrrolidone, NH₂ = amine functionality resulting by modification with APTES (aminopropyl triethoxy silane) or another method). The set of materials and sample codes are summarized in Table 1 along with nominal particle sizes specified by the suppliers and equivalent diameters based on TEM measurements of the as-received particles. While the nominal sizes ranged from 10 to 100 nm, the TEM measurements indicated significant differences with the nominal values, typically showing smaller diameter values. For example, in the case of NiO, all the particles are in the 10–20 nm range rather than the 20–90 nm range indicated by the suppliers. The CeO₂ particles range between 10 and 64 nm, whereas the Fe₂O₃ and Mn₂O₃ particles all exhibit average diameters in the 35–60 nm range. In general, the TEM measurements on these particles also indicate that the particles have rather broad size distributions as evidenced by the large standard deviations (~25–40% of the average diameter).

2.2 X-ray photoelectron spectroscopy

XPS was carried out using an Axis Ultra DLD spectrometer (Kratos Analytical, Manchester, UK) with monochromated Al K α X-rays. The angle between the source and the spectrometer is 60°. As-received dry metal oxide nanopowders were mounted directly on the sample holder with adhesive tape and transferred into the XPS system. For measurements the samples were oriented normal to the spectrometer resulting in a take-off angle of 90°. Three distinct areas (300 × 700 μ m) were measured on each sample. Survey spectra were first obtained in order to estimate the relative atomic composition of the sample and detect any impurities that may be present. High resolution (HR) spectra were subsequently acquired in regions corresponding to the strongest core level transitions for the major elements present on these samples. These HR scans were used for more accurate quantification of the major elements present with the detailed structure of the peaks also providing additional information on the chemical state of the elements. The spectrometer pass energy was set at 80 eV for the survey spectra and 20 eV for the HR scans. A charge neutralizer was employed to compensate for sample charging. The energy scale for all spectra was referenced by calculating the offset required to shift the main peak in the C 1s region to 284.8 eV and applying the same shift to the other spectra taken at the same point.



Table 1 NiO, CeO₂, Fe₂O₃ and Mn₂O₃ NPs used in this study, along with the supplier, nominal size and surface modification as provided by the supplier. The average size (equivalent circular diameter and standard deviation) as measured by TEM^{13,18,19} are also shown. TEM sizes for the functionalized particles are not provided in this table but generally have similar sizes to the corresponding bare particle from the same supplier and are noted in Tables 2–5. Supplier abbreviations: USRN is US Research Nanomaterials; NAM is Nano and Amorphous Materials. Samples with NRC in the sample code were functionalized in-house at NRC

Oxide/supplier	Size (nm)		Functionalization			
	Nominal	TEM	None	PVP	SA	NH ₂
NiO						
Skyspring	50	15 ± 5	Ni-01			
mkNano	90	19 ± 8	Ni-02			
Sigma	<50	12 ± 4	Ni-03			
USRN	18	21 ± 9	Ni-04	Ni-05 Ni-04 NRC	Ni-06	Ni-07
USRN	15–35	19 ± 6	Ni-08			
CeO₂						
USRN	50	27 ± 11	Ce-01	Ce-10		
USRN	10–30	14 ± 5	Ce-02			
USRN	10	10 ± 2	Ce-03	Ce-04	Ce-05 Ce-03 NRC	Ce-09
NAM	50–105	20 ± 11	Ce-06			
mkNano	70	58 ± 25	Ce-07			
mkNano	35	64 ± 28	Ce-08			
Fe₂O₃						
Sigma	<50	35 ± 14	Fe-01			
USRN	30	34 ± 16	Fe-02	Fe-03	Fe-06	Fe-04 Fe-05
Lanxess	90	57 ± 20	Fe-07			
Mn₂O₃						
USRN	30	53 ± 17	Mn-01	Mn-02	Mn-03	Mn-04
USRN	<100	47 ± 13	Mn-05			
mkNano	50	41 ± 17	Mn-06			
Am. Element	<100	37 ± 14	Mn-07			
Nanografi	28	37 ± 15	Mn-08			

Although the drawbacks of referencing spectra to the C 1s peak in this manner have been discussed,^{28,29} a recent review of a large experimental data set has shown that this method, when judiciously applied, can yield consistent results to an accuracy of ±0.2 eV.³⁰ Data analysis was carried out with the CasaXPS software (Casa Software, Teignmouth, UK). The atomic compositions were obtained using the area under each peak after subtracting Shirley or linear backgrounds and applying Kratos relative sensitivity factors. Decomposition of the HR spectra into various components was carried out using mixed Gaussian–Lorentzian (GL30) lineshapes.

Each sample was measured at three distinct points for both survey and HR scans, reporting both the average atomic composition and the associated standard deviation. This standard deviation generally reflects the homogeneity of the sample, with several other factors making larger contributions to the total uncertainty associated with quantitative determination of sample composition using XPS data.³¹ Measurement of the integrated peak intensities, background subtraction, transmission function corrections and use of relative sensitivity factors all contribute to the uncertainty budget. An interlab

comparison using XPS to measure the composition of core-shell nanoparticles found agreement between multiple labs to within ~10%.³²

3 Results and discussion

XPS spectra were measured for all the samples summarized in Table 1. These same samples have been examined in previous studies that were directed at investigating several families of metal oxide nanomaterials for which extensive physical chemical characterization of their properties was obtained for correlation with biological assays that were aimed at understanding potential cytotoxicity, generation of reactive oxygen species and activation of apoptotic or immune response pathways.^{13,18,19} The selection of multiple samples of each metal oxide was intended to probe similarities and differences related to supplier, size and surface chemistry and to provide some of the data necessary for development of a regulatory framework for nanomaterial use within Canada. The physical chemical characterization consisted of size and specific surface area measurements by TEM and BET, respectively, dispersion and measurement of size and charge in



solution by dynamic light scattering and zeta potential and characterization of surface functional groups by qNMR and TGA. It is important to note that detailed characterization is essential to ensure that one understands the material properties and can make comparisons with literature data for biological assays. It is also common to purchase commercial nanomaterials for use in biological assays without measuring their physical chemical properties which is essential to determine whether comparable materials are examined in various studies. As seen in Table 1, the TEM measurements indicate that the nominal values for size provided by the supplier were frequently different from values measured in our experiments. This generally had the effect of narrowing the range of sizes that were examined. It should be noted that, apart from the Mn_2O_3 samples, the agreement of our in-house measurements with the supplier data was reasonably good for the functionalized samples (and a bare sample of the same size) purchased from USRN. The size of the nanoparticles is potentially relevant to the quantitative interpretation of the XPS measurements. However, as the diameter of even the smallest (~ 10 nm diameter) particles studied here are larger than the effective probing depth ($3\times$ the inelastic mean free path of the photoelectrons), curvature and other size effects are not expected to play a significant role.²⁰

Although XPS was not employed in the initial studies it became obvious that a consideration of the impurity content in the materials and the metal oxidation state were potentially important factors. XPS is also capable of assessing the surface functional groups directly on as received materials in powder form. Our previous work had indicated that qNMR is superior to TGA for measurement of total functional group content, providing adequate sensitivity, the ability to conclusively identify functional groups and the potential for traceable measurements with metrological rigor.³³ TGA is generally only a semi-quantitative method and, unless combined with FTIR or mass spec measurements, often fails to elucidate the surface composition. However, qNMR does require removal of groups from the surface prior to analysis. The capability of XPS to measure dry powder samples directly, while providing

quantitative information on the presence of impurities, metal oxidation state and surface composition, makes this technique an attractive addition to the surface chemistry tool box.

The XPS results are discussed in three different sections below. First, we discuss information obtained regarding the chemical composition for each of the four types of metal oxide nanomaterials. Then, we summarize what has been learned about the metal oxidation state in these materials. Finally, we compare XPS assessment of the functional group content on these particles with previous results obtained by qNMR.

3.1 Chemical composition

3.1.1 NiO nanomaterials. Survey scans were initially run for all samples in order to identify and quantify the elements present. A representative example is shown in Fig. 1 for unmodified NiO NPs obtained from four different suppliers. The strongest peaks can be assigned to Ni 2p and O 1s with additional features due to other Ni related photoemission and Auger transitions clearly visible. Detailed examination of the region between 500 and 150 eV reveals a number of additional peaks. The C 1s peak is observed on all bare nanoparticle samples and can be attributed to a combination of residual hydrocarbons left over from the synthesis procedure as well as adventitious contamination from air exposure. In addition, a number of other peaks can be assigned to elements (*i.e.*, Cl, Bi and Br) not expected to be present on these samples. This figure clearly illustrates that the identity and amount of these impurities vary, even for unfunctionalized samples, an observation which suggests that different synthesis routes used by the four suppliers lead to different types and variable levels of impurities.

The atomic composition of all the NiO NPs derived from the XPS survey scans is summarized in Table 2. The TEM measured average diameters indicate that all the particles were in the 10–20 nm size range, with standard deviations ranging from 25 to 70% of the average. As expected, for the bare particles, Ni and O are the primary elements present making up 83 to 93 atomic%

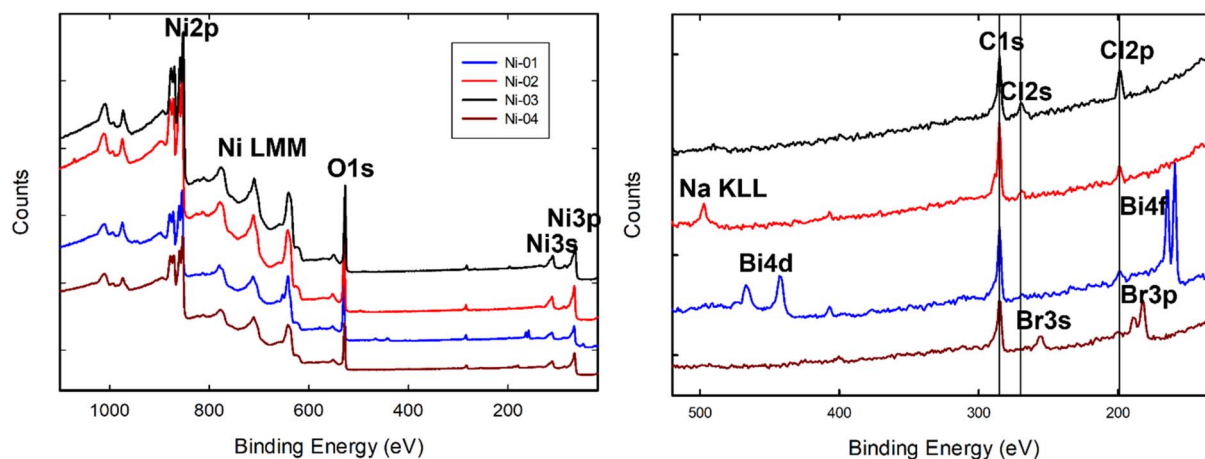


Fig. 1 Survey spectra for commercial samples of unmodified NiO nanoparticles from four different suppliers. The energy region between 520 and 130 eV is magnified at right to show the peaks assigned to various impurities present on the different samples.



Table 2 Atomic composition (%) of various NiO NPs based on intensities of the indicated core level emission peaks measured in XPS survey scans. The bold number is the average of 3 measurements on the sample while the number below this is the standard deviation. Blank entries indicate no peak was observed above the background

	Diameter	Ni 2p	O 1s	C 1s	N 1s	Si 2p	Br 3p	Cl 2p	Mn 3p	Bi 4f	Na 1s	Ni/O	C/Ni
Ni-01 Skyspring, bare	15 ± 5 nm	39.8 0.6	43.2 0.8	9.3 0.6				0.3 0.1	6.7 0.2	0.6 0.04		0.92 0.03	0.24 0.02
Ni-02 mkNano, bare	19 ± 8 nm	52.2 1.0	40.0 0.5	6.7 0.5				0.4 0.04			0.7 0.1	1.30 0.04	0.13 0.01
Ni-03 Sigma, bare	12 ± 4 nm	54.5 0.7	39.2 0.4	5.7 0.4				0.7 0.04				1.39 0.03	0.10 0.01
Ni-04 USRN, bare	21 ± 9 nm	49.9 0.7	37.9 0.2	9.5 0.6	0.3 0.1		2.2 0.2	0.2 0.04				1.32 0.02	0.19 0.02
Ni-04-NRC USRN, PVP@NRC	nd	30.2 0.8	33.0 1.4	31.5 1.5	3.4 0.4		1.7 0.1					0.92 0.04	1.04 0.07
Ni-05 USRN, PVP	17 ± 12 nm	48.8 1.0	38.0 0.5	10.7 0.6	0.5 0.04		2.0 0.04					1.28 0.04	0.22 0.02
Ni-06 USRN, SA	14 ± 6 nm	15.6 1.1	22.0 0.2	61.4 0.9			1.1 0.05					0.71 0.06	3.96 0.32
Ni-07 USRN, APTES	21 ± 14 nm	20.5 1.7	35.4 0.5	30.8 1.2	6.1 0.1		1.9 0.1					0.58 0.05	1.52 0.20
Ni-08 USRN, bare	19 ± 6 nm	50.8 0.3	42.1 0.4	6.7 0.3		5.3 0.6		0.4 0.05				1.21 0.01	0.13 0.01

of the sample as probed by XPS. The standard deviation of the three measurements at different spots on each sample is typically less than 1%, indicating that the samples are fairly homogeneous. As discussed above, the uncertainties in atomic compositions extracted from measured XPS intensities are considerably greater and typically on the order of 10%. A significant amount of carbon is observed on all the samples, with the unmodified samples exhibiting carbon content of between 5 and 10% (C/Ni ratios of 0.1–0.24) due to adventitious contamination from handling in air and/or residual hydrocarbons left over from the nanoparticle synthesis. In addition to C, the other impurities observed in Fig. 1 are quantified in Table 2. Ni-01 shows a substantial amount of Mn (6.7%) and a smaller amount of Bi (0.6%), both of which have been detected by ICP-MS measurements of the bulk composition for this sample.¹⁹ Ni-04 and the related modified samples all show the presence of Br, while all five unmodified samples show small amounts (<0.7%) of Cl. A small amount of Na was detected on Ni-02.

On three of the four functionalized samples, the carbon atomic fraction increases significantly to 30–60%, with corresponding C/Ni ratios from 1 to 4. The increase in C/Ni ratio is expected for the modified samples due to the carbon associated with the APTES, SA and PVP coatings as well as a decreased Ni 2p signal due to attenuation by the coatings. In contrast, the Ni-05 PVP modified particles exhibit a carbon content (and C/Ni ratio) similar to that of the bare USRN sample of the same size (Ni-04) and only a small increase in nitrogen content, both of which indicate only a minimal PVP coating. On the other hand, the in-house PVP-modified sample (Ni-04-NRC, prepared from the bare USRN Ni-04 particles) exhibits three times more carbon than the USRN PVP sample (Ni-05) and a seven-fold increase in nitrogen content, consistent with a substantial PVP coating. The APTES modified sample shows substantially larger N 1s and Si 2p signals than the corresponding bare

sample, as expected for amino-silane modification. The SA-coated NPs (Ni-06) exhibit a large increase in the C 1s fraction as well as a decrease in the Ni 2p signal, consistent with a substantial amount of functionalization. We note that the TEM size measurements for all the functionalized particles show no significant size increase due to the coating which is not surprising given the large distribution of diameters coupled with the fact that the functionalization layers are only expected to be 1–2 nm thick.

High resolution (HR) scans of the Ni 2p, O 1s, C 1s and N 1s regions were also recorded for all the NiO samples, with an additional scan of the Si 2p region for the APTES functionalized sample Ni-07. Atomic compositions derived from this data are summarized in Table S1† and are in reasonable agreement with those obtained from the survey scans with a few systematic differences. In comparing the atomic compositions from HR and survey data it is important to note that the contributions of impurities are ignored in the HR data although this is typically a small effect in most samples, as the total impurity content is usually less than a few percent. In general, the higher density of points in the HR scans (points every 0.1 eV as compared to every 1 eV in the survey scans) results in a better estimate of the background and a more accurate determination of the peak area. Comparing the data in Tables 2 and S1,† the HR data systematically yields lower relative values for the Ni 2p signal suggesting that the Shirley background fits to the survey scans typically lead to overestimation of the Ni fraction. The HR data allow the background to be fit more accurately, with the bare particles exhibiting Ni/O ratios of 0.74 for Ni-01 and ranging from 1 to 1.16 for the other four bare NiO particles. The lower value for Ni-01 is likely associated with the significant Mn contamination (likely in the form of MnO_x based on the energy of the Mn core levels) on this sample. Apart from Ni-01 the Ni/O ratios are in reasonable agreement with the expected value of 1



based on the stoichiometry of NiO, considering the uncertainties in extracting relative compositions from XPS data. The HR data also improves the ability to quantify the N 1s signal. Two of the bare samples, Ni-03 and Ni-04 show N 1s levels just above the detection limit (0.1–0.2%) whereas the Ni-05 PVP sample shows a significantly larger, but still small (0.8%) signal. As observed in the survey scan data, the HR data for the NRC-prepared PVP sample (Ni-04-NRC) shows considerably larger N and C signals which are consistent with a good level of PVP modification ($4.6\times$ that of Ni-05). Ni-07, an APTES modified sample, showed an even larger N signal as well as a significant Si 2p signal. The measured Si/N ratio from the HR data is 1, as expected for an APTES functionalized sample which should have one Si atom for every N atom.

Examination of high-resolution spectra provides additional insight into the chemical composition of the samples as seen in Fig. 2, showing the C 1s and O 1s regions for four different unmodified NiO particles. As these particles have not been intentionally modified the features in the C 1s region are attributed to residual hydrocarbons from the synthesis process or resulting from exposure to the lab air. Interestingly the shape of the C 1s spectrum is similar for the four particles, and can be fit with three peaks at 284.8, 285.8 and 288.3 eV which are attributed to C–C, C–O and O=C–O species respectively.³⁴ Fractions of these three species differ somewhat in the four samples but the C–C always dominates (65–80%), with smaller amounts of C–O (5–20%) and O=C–O (10–15%) species. This implies that this adventitious contamination contributes an oxygen signal of ~ 25 –35% of the carbon fraction present on the particles. As the carbon fractions on the unmodified particles are $< 10\%$ this means the oxygen fractions associated with the contamination are $< 3\%$, much smaller than the total oxygen fraction observed on these samples (38–43%). The O 1s region, also shown in Fig. 2, is rather similar for all the unmodified samples exhibiting two peaks. Based on previous work for NiO

nanoparticles, the stronger peak is attributed to the lattice oxygen within the metal oxide particles while the weaker feature can be assigned to hydroxyl groups on the surface.³⁵ While for samples Ni-01 and Ni-04 the lattice oxygen peak is observed at 529.5 eV, as previously reported for NiO, this peak is observed to shift to 529.2 eV on Ni-02 and Ni-03. Blume *et al.*³⁶ have identified an additional O 1s peak at 528.9 eV which they assign to oxygen vacancies in NiO_{1-x} so this shift to lower binding energy may be due to an increase in oxygen vacancies in these two samples.

An interesting observation from both the survey scan and HR data (Tables 2 and S1†) is that the Ni/O ratio decreases for modified NPs. There appears to be an inverse correlation between the Ni/O ratio and the carbon content. Fig. 3 illustrates this correlation by plotting the Ni 2p and O 1s signals as

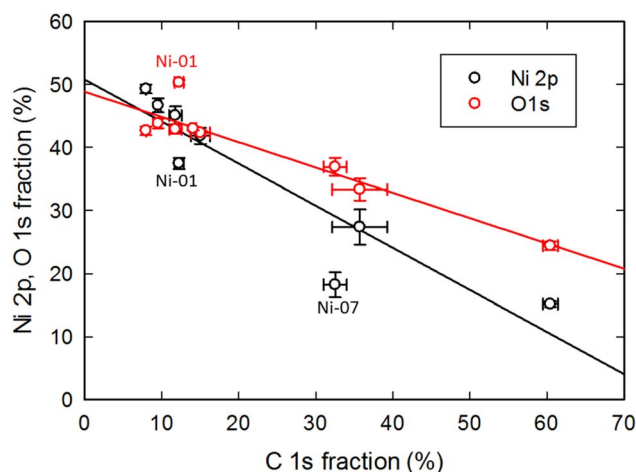


Fig. 3 Correlation of Ni 2p and O 1s signal intensities with the C 1s signal (based on the HR data) for the eight NiO samples. Outlier points are labelled and discussed in the text.

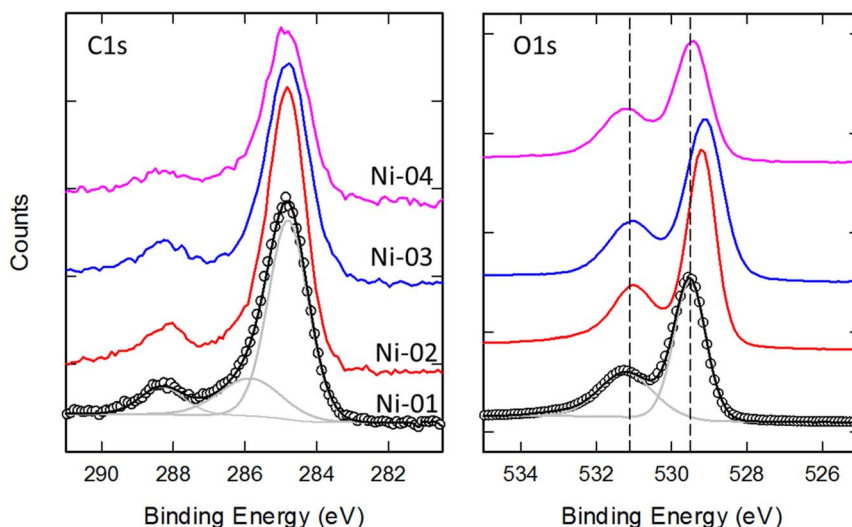


Fig. 2 High resolution spectra of the C 1s and O 1s regions for four unmodified NiO nanoparticle samples. Fits to the Ni-01 spectra are shown by the solid grey (components) and black (sum) lines.



a function of the C 1 s signal for the eight NiO samples. For samples with a low amount of carbon, the Ni and O fractions are approximately similar, as expected for NiO, but as the carbon fraction increases the Ni 2p signal is seen to decline faster than that of the O 1 s, leading to a decrease in the measured Ni/O ratio. This is consistent with the majority of the carbon residing on the surface of the particles and attenuating both the Ni and O signals from the particles, with the faster decrease of the Ni signal arising from the shorter inelastic mean free path of the lower kinetic energy Ni 2p photoelectrons as compared with the higher kinetic energy O 1s electrons. The data points indicated in the figure that are furthest from the linear trend line are cases in which there is a significant quantity of elements other than carbon (Mn in the case of Ni-01 and N and Si for Ni-07) on the surface of the particles that acts to attenuate the Ni and O photoelectrons.

3.1.2 Fe₂O₃ nanomaterials. Atomic composition, based on survey scans, for the bare and surface-modified Fe₂O₃ NPs is summarized in Table 3 (selected survey spectra are shown in Fig. S1†). The two unmodified samples Fe-01 and Fe-02 show a higher carbon fraction (17%) than observed for the bare NiO NPs. In terms of other impurities Fe-01 exhibits only a trace amount of Cl whereas Fe-02 and most of the corresponding functionalized USRN NPs all show Mg, Na, Cl and Ca impurities. Apart from Ca, the surface impurities observed here are not reported in the bulk purity assessment provided by USRN (measurement method not specified). Similar to the commercial NiO-PVP sample, Fe-03 shows no increase in either C 1s or N 1s compared to the corresponding bare sample of the same size. The USRN NH₂ functionalized samples (Fe-04, Fe-05) show the expected increase in nitrogen, although the amount of N is ~3× higher for the APTES-modified sample (Fe-05). For Fe-05 the silicon and nitrogen fractions are equal (within uncertainties) as expected. On these Fe₂O₃ samples the Si 2s peak is used for quantification of silicon as the Fe 3s peak interferes with the Si 2p peak which is typically employed. Fe-04 exhibits no silicon peaks, indicating that the amine functionalization on this sample was obtained using a non-silane route. This is consistent

with the supplier specification of both NH₂ and COOH functional groups, suggesting amino acid modification. The SA modified sample (Fe-06) shows a large increase in the C 1s signal, accompanied by attenuation of the Fe 2p as expected for this modification. Fe-07 from Lanxess is sold as a pigment (Colortherm, Red 110 M) and exhibits a larger O fraction and a reduced Fe fraction as well as a substantial Si 2s signal. Apart from carbon (12.4%) this sample exhibits no other detectable impurities. The significant Si signal on this sample suggests these NPs are either derived from a silicon containing iron oxide such as fayalite (Fe₂SiO₄) or consist of iron oxide particles modified with a silica coating. Data from HR scans allows us to discriminate between these two possibilities as discussed below.

High resolution spectra of all Fe₂O₃ samples were also recorded and the resulting atomic composition data summarized in Table S2.† There is good agreement between the atomic compositions obtained from survey and HR data with the differences smaller than observed for the NiO samples. For the PVP modified sample (Fe-03), the improved S/N in the HR scans allows a small peak in N 1s region to be detected, corresponding to 0.3 ± 0.1% atomic composition. This level is barely above the noise and indicative of only a small amount of PVP being detected. The measured Fe/O ratio is slightly higher from the HR scans (~0.8 for the unmodified particles), but still close to the expected value of 0.66 for Fe₂O₃. As observed for the NiO particles, this ratio decreases for modified particles, attributed to greater attenuation of the Fe 2p signal compared to the O 1s due to the surface modification, as discussed for the NiO samples. This decrease in Fe/O ratio is particularly significant for the SA modified (Fe-06) and Lanxess (Fe-07) samples.

Additional insight into the chemical composition of some of these Fe₂O₃ particles is obtained by looking at the HR spectra of the O 1s region shown in Fig. 4. For Fe-01, the O 1s peak can be fit with two components a sharp peak centered at 529.9 eV and a broader weaker peak at 531.4 eV, consistent with previous observations for Fe₂O₃.³⁷ As for NiO, the two peaks are assigned to lattice oxygen in the bulk of the particles and hydroxyl groups on the surface, respectively. For Fe-02, the basic peak structure is

Table 3 Atomic composition (%) of various Fe₂O₃ NPs based on intensities of the indicated core level emission peaks measured in XPS survey scans. The bold number is the average of 3 measurements on the sample with the number below the standard deviation. Blank entries indicate no peak was observed above the background

	Diameter	Fe 2p	O 1s	C 1s	N 1s	Si 2s	Cl 2p	Na 1s	Mg KLL	Ca 2p	Fe/O	C/Fe
Fe-01 Sigma, bare	35 ± 14 nm	35.9 0.7	46.6 1.0	17.3 0.7			0.3 0.04				0.77 0.03	0.48 0.02
Fe-02 USRN, bare	34 ± 16 nm	32.8 0.4	47.2 0.1	17.3 0.4			0.4 0.1	1.4 0.02	0.6 0.1	0.4 0.01	0.69 0.01	0.53 0.02
Fe-03 USRN, PVP	28 ± 8 nm	31.2 0.4	47.4 0.4	19.3 0.7			0.4 0.1	0.7 0.1	0.6 0.02	0.3 0.02	0.66 0.01	0.62 0.03
Fe-04 USRN, NH ₂	27 ± 6 nm	33.6 0.6	49.7 0.7	13.6 1.2	0.7 0.1		0.4 0.2	1.0 0.1	0.6 0.1	0.3 0.02	0.68 0.004	0.39 0.04
Fe-05 USRN, APTES	26 ± 6 nm	28.6 0.5	47.8 0.6	16.7 0.6	2.3 0.3	2.4 0.2	0.4 0.1	0.8 0.04	0.6 0.1	0.4 0.1	0.60 0.02	0.58 0.02
Fe-06 USRN, SA	24 ± 5 nm	11.1 0.1	31.0 0.8	57.2 0.9					0.6 0.03	0.2 0.05	0.36 0.01	5.17 0.14
Fe-07 Lanxess	57 ± 20 nm	19.4 0.7	57.8 0.8	12.4 1.3		10.3 0.6					0.34 0.01	0.68 0.08



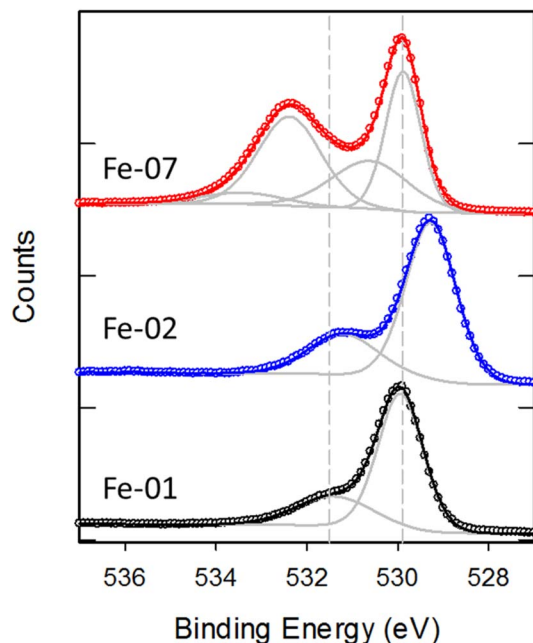


Fig. 4 High resolution spectra of the O 1s region for three different Fe_2O_3 nanoparticle samples. Fits are shown by the solid grey lines with the overall fit shown by the solid black, blue and red lines.

the same but the lattice oxygen peak is shifted to 529.4 eV. Fe-07 exhibits different structure in the O 1s region that can be fit with four peaks at 529.9, 530.5, 532.4 and 533.6 eV. The two higher binding energy features were not observed on the other Fe oxide particles and coincide with the expected energy of the O 1s in SiO_2 (532.4 eV) and for adsorbed water (533–534 eV). The Si 2p peak in this sample is observed at 103.2 as in SiO_2 . This strongly

suggests the Fe-07 particles are coated with a silica shell, which also accounts for the strong attenuation of the Fe signal observed on this sample. The other possibility for the presence of Si in these particles is that they consist of fayalite (Fe_2SiO_4). However, the Si 2p and O 1s binding energies in fayalite have been reported as 102 eV and 531.1 eV, respectively³⁸ inconsistent with the peak positions observed here. TEM imaging of Fe-07 (see Fig. S2†) does not show any definitive evidence for the presence of a silica shell on these particles, but the presence of a thin (~ 1 nm) shell cannot be ruled out. An EDX scan of the particles (also shown in Fig. S2†) shows the presence of Fe, O and C but no Si. Although EDX has good lateral spatial resolution, it probes the entire depth of the particles, effectively probing the “bulk” composition. Therefore, if the particles consisted of fayalite, the significant amount of Si ($\sim 14\%$) expected based on the stoichiometry would be easily detected. However, if the observed Si signal comes from a thin ~ 1 nm shell surrounding the particles it is likely too weak to be detected in EDX. Therefore, the electron microscopy data for Fe-07 is also consistent with the presence of a nm thick silica shell coating a Fe_2O_3 core.

HR spectra of the C 1s and O 1s regions are also expected to provide insight into the chemical composition for the other modified Fe_2O_3 particles. While changes associated with the different modifications are observed in the C 1s spectra (as shown in Fig. S3†), the presence of adventitious carbon contamination observed on all the particles complicates the interpretation of these changes. In contrast the O 1s region for all the modified samples is very similar to that of the bare samples as the strong oxygen signal from the Fe_2O_3 particles dominates the spectra.

3.1.3 CeO_2 nanomaterials. The atomic compositions obtained from survey scans for CeO_2 NPs are shown in Table 4.

Table 4 Atomic composition (%) of CeO_2 NPs from XPS survey scans. The bold number is the average of 3 measurements on the sample with the number below the standard deviation. Blank entries indicate no peak was observed above the background

	Diameter	Ce 3d	O 1s	C 1s	N 1s	Si 2s	Cl 2p	F 1s	P 2s	Na 1s	Ce/O	C/Ce
Ce-01 USRN, bare	27 ± 11 nm	30.1 1.8	55.8 0.6	12.2 1.4			1.9 0.2				0.54 0.04	0.41 0.07
Ce-02 USRN, bare	14 ± 5 nm	29.4 0.8	49.3 1.1	18.9 1.6			2.5 0.04				0.60 0.02	0.64 0.07
Ce-03 USRN, bare	10 ± 2 nm	36.3 0.2	55.0 0.3	6.2 0.3			0.2 0.1	2.3 0.3			0.66 0.003	0.17 0.01
Ce-03-NRC USRN, SA@NRC	nd	10.2 0.3	24.6 0.2	65.2 0.5							0.41 0.01	6.38 0.22
Ce-04 USRN, PVP	10 ± 2 nm	33.2 0.6	54.3 0.7	9.6 0.7			0.3 0.03	2.6 0.1			0.61 0.02	0.29 0.02
Ce-05 USRN, SA	nd	20.3 0.6	42.9 1.2	36.6 1.7			0.3 0.1				0.47 0.01	1.81 0.14
Ce-06 NAM, bare	20 ± 11 nm	22.4 0.3	53.5 0.4	12.6 0.6					9.1 0.3	2.4 0.3	0.42 0.01	0.56 0.02
Ce-07 mkNano, bare	58 ± 25 nm	26.2 0.9	51.3 0.5	20.9 0.9			0.2 0.1			1.4 0.3	0.51 0.02	0.80 0.06
Ce-08 mkNano, bare	64 ± 28 nm	32.1 0.4	55.7 0.4	8.3 0.3				2.5 0.2		1.4 0.3	0.58 0.02	0.26 0.01
Ce-09 USRN, APTES	nd	15.5 0.4	43.3 0.9	25.0 1.3	6.9 0.5	8.3 0.7	1.0 0.2				0.36 0.01	1.61 0.09
Ce-10 USRN, PVP	nd	29.8 1.1	55.3 1.7	13.5 2.3			1.5 0.2				0.54 0.02	0.45 0.09



The carbon content of the bare particles is highly variable, ranging from a low of 6.3% (Ce-03) to a high of 20.9% (Ce-07), although we note that the Ce 4s peak (at 289 eV in CeO₂)³⁹ interferes with the C 1s peak, increasing the uncertainty in carbon quantification using survey scan data. This is particularly problematic for samples containing a substantial fraction of O–C=O species which also give rise to a C 1s feature in this range.³⁴ As discussed above for NiO particles, most of the unmodified metal oxide samples studied here show a feature in this region due to adventitious carbon contamination, although it is typically less than 15% of the total carbon signal. All of the CeO₂ samples (apart from the SA modified Ce-05) also contained halogen impurities, eight with Cl and three with F. Interestingly, the NAM sample (Ce-06) stands out with a large amount of P (>9%) along with some Na (~2%) impurities. The P 2p peak is observed at ~133 eV, consistent with the P being bound to oxygen. The carbon content is seen to increase for all the modified samples, with particularly large increases observed for the SA coated particles (Ce-05 and Ce-03 NRC). Significant Si and N fractions were observed for the APTES modified sample, Ce-09. As for the other PVP-coated metal oxide NPs, N was not detected in the survey scans of either of the two USRN PVP-coated samples (Ce-04 or Ce-10).

Atomic compositions derived from the HR scans for the CeO₂ oxide samples (summarized in Table S3†) are in general agreement with the survey data, although there are significant differences on some samples, primarily due to the better ability to separate C 1s and Ce 4s contributions in the HR scans. The C/Ce ratios are generally higher in the HR data, except for Ce-03-NRC, indicating that the attempts to exclude the contributions of the Ce 4s peak to the C 1s region in the survey scans led to underestimating the C fraction. The Ce/O ratios are slightly lower based on the HR data, ranging from 0.39–0.53 for all but one of the unmodified samples, in agreement with the expected value of 0.5 for CeO₂. Ce-06 exhibits a lower Ce/O ratio of 0.31, probably as a result of the phosphate contamination. On the modified samples the Ce/O ratio is observed to decrease to

0.28–0.4, consistent with the Ce 3d electrons suffering greater attenuation due to a shorter mean free path relative to O 1s electrons, as discussed above for Ni and Fe oxides. As for the Ni and Fe oxides the Ce 3d and O 1s fractions decrease linearly with the C 1s signal (see Fig. S4†). Interestingly, HR scans of the N 1s region show a small level of nitrogen signal for Ce-04 and Ce-10 that was not detected in the survey scans, consistent with a small amount of PVP functionalization.

3.1.4 Mn₂O₃ nanomaterials. Atomic composition data from survey scans for the Mn₂O₃ NPs are summarized in Table 5. All the unmodified samples exhibit a large amount of carbon contamination (28–36%) with only small amounts of an additional Cl impurity (<0.4%) detected on most samples. Mn-08 exhibits the lowest amount of carbon and no detectable Cl. The absence of metal impurities on these samples is generally consistent with recent ICP MS studies.¹³ While those studies did show a small (0.4% by mass) amount of Cu for Mn-08, no Cu was detected in the current work, although the expected level is close to the detection limit of our XPS measurements. The large carbon content observed on Mn-01, the unmodified USRN sample, limits the ability to draw firm conclusions regarding the surface modifications of these particles based on the magnitude of the C 1s signal alone, although the SA-coated sample (Mn-03) does show a substantial increase in the carbon fraction. Note that the large decrease in the metal to oxygen ratio seen for the other SA modified metal oxides is neither expected nor observed for this sample since Mn 2p and O 1s electrons have similar kinetic energies (640 and 530 eV, respectively) and hence suffer similar amounts of attenuation upon passing through the SA coating. The APTES functionalized sample (Mn-04) shows the expected N 1s and Si 2p signals but at a smaller level than for the other metal oxide NPs. In contrast, no N signal is observed for the PVP modified sample (Mn-02), similar to the case of the other USRN PVP modified oxide NPs.

Atomic compositions based on HR data (Table S4†) are in reasonable agreement with that from the survey scans. The Mn and O contents are slightly lower than from the survey data

Table 5 Atomic composition (%) of Mn₂O₃ nanoparticles from XPS survey scans. The bold number is the average of 3 measurements on the sample with the number below the standard deviation. Blank entries indicate no peak was observed above the background

	Diameter	Mn 2p	O 1s	C 1s	Si 2p	N 1s	Cl 2p	Mn/O	C/Mn
Mn-01 USRN, bare	53 ± 17 nm	22.0 0.3	42.0 0.4	35.8 0.7			0.2 0.1	0.52 0.01	1.63 0.05
Mn-02 USRN, PVP	54 ± 17 nm	21.7 0.1	41.8 0.3	36.3 0.2			0.2 0.1	0.52 0.01	1.67 0.00
Mn-03 USRN, SA	56 ± 27 nm	14.7 0.4	26.8 0.3	58.5 0.6				0.55 0.01	3.98 0.15
Mn-04 USRN, APTES	49 ± 21 nm	20.2 1.1	43.2 0.9	34.3 2.0	1.1 0.0	1.0 0.1	0.3 0.04	0.47 0.02	1.71 0.19
Mn-05 USRN, bare	47 ± 13 nm	23.3 0.5	43.1 0.8	33.5 1.2			0.2 0.1	0.54 0.01	1.44 0.08
Mn-06 mKNano, bare	41 ± 17 nm	21.5 0.4	42.1 0.3	36.0 0.7			0.4 0.04	0.51 0.01	1.68 0.06
Mn-07 Am. Elem., bare	37 ± 14 nm	20.3 2.2	40.4 1.8	39.0 4.0			0.3 0.05	0.50 0.03	1.95 0.39
Mn-08 Nanografi, bare	37 ± 15 nm	25.9 1.0	46.0 0.2	28.1 1.1				0.56 0.02	1.09 0.09



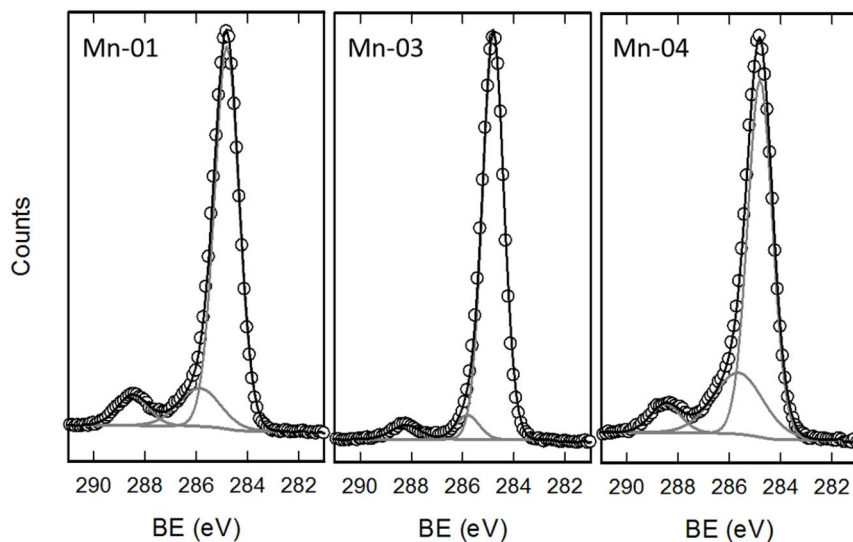


Fig. 5 High resolution spectra of the C 1s region for three Mn_2O_3 samples. Fits are shown by the solid grey (components) and black (sum) lines.

while the C fractions are somewhat higher. The Mn/O ratios are between 0.44 and 0.53, all smaller than the expected value of 0.66 for Mn_2O_3 . This is likely due to the presence of additional oxygen associated with the high level of adventitious carbon contamination observed on these samples. The unmodified sample with the lowest level of carbon (Mn-08) exhibits the highest Mn/O ratio, consistent with this explanation.

Fig. 5 shows high resolution scans of the C 1s region for the unmodified USRN sample (Mn-01) along with two modified samples of the same size from the same supplier. The C 1s spectrum for Mn-01 is similar to that observed on the other unmodified metal oxide NPs. As for the NiO samples discussed above, the majority of the C 1s signal ($\sim 78\%$) is assigned to aliphatic carbon, while the features at 286 eV and 288.5 eV arise from C–O (14%) and O–C=O (8%) species respectively. For Mn-01 which exhibits a carbon content of $\sim 40\%$ this suggests that $\sim 12\%$ of the $\sim 39\%$ oxygen fraction on this sample originates from the hydrocarbon contamination layer and not from the metal oxide particles. Reducing the oxygen fraction by removing the contribution from this contamination, yields a revised Mn/O ratio of ~ 0.7 , much closer to the expected value for Mn_2O_3 . The C 1s region for Mn-03 and Mn-04 samples in Fig. 5 shows changes that can be attributed to the modifications. For the SA modified particles (Mn-03) the C–C feature is a greater fraction of the total signal (89%) with smaller amounts in the C–O (7%) and O–C=O (4%) components, consistent with the presence of a long chain hydrocarbon. Note that for stearic acid the ratio of O–C=O to C–C species should be 0.058, quite close to the 0.05 which is observed here. For the APTES modified sample (Mn-04) the feature at 286 eV is more prominent (24%) than in the other two samples. This feature can be attributed to both C–N (from the terminal amine) and C–O (from unhydrolyzed ethoxy groups) on APTES. This shows that even in samples with substantial amounts of adventitious carbon contamination, it is possible to observe differences due to surface modifications in HR spectra of the C 1s region.

3.2 Oxidation state

The metal oxidation state plays an important role in the reactivity of metal oxide nanoparticles and their use in a variety of applications. While the oxidation state is generally determined by the metal/oxygen ratio, the stoichiometry (and hence the oxidation state) in the near surface region can differ from that in the bulk. While XPS data facilitates determination of the metal/oxygen ratio in the near surface region (the first ~ 5 nm of material), the measured values can differ from the expected bulk stoichiometry due to attenuation effects of contaminants and functional layers, as well as additional oxygen contributions from these layers. Together with the inherent uncertainty of $\sim 10\text{--}15\%$ for chemical compositions determined by XPS this means that the composition data alone is often insufficient to distinguish between closely related compositions (*i.e.* Mn_2O_3 with Mn/O = 0.66 as compared to MnO_2 with Mn/O = 0.5). However, XPS can also provide direct information on the metal oxidation state in these particles through analysis of the detailed structure of selected metal core level emission features. While some studies attempt to assign and quantify oxidation states by associating each state with a single peak at a specific binding energy this approach has been shown to be invalid for all of the transition metals investigated here.^{37,40,41} Instead, in addition to spin–orbit splitting, the full extended multiplet, shake-up and plasmon loss structures for each oxidation state must be taken into account, recognizing that each oxidation state typically gives rise to multiple features in the experimental spectrum.

For the NiO particles, high resolution scans of the Ni 2p region were examined in order to investigate potential variations in oxidation state. The Ni 2p emission envelope for the unmodified NiO particles is shown in Fig. 6 to exhibit a rather complex structure with multiple features visible. The envelope can be split into two parts associated with the two spin–orbit components. As seen in this figure the $2p_{3/2}$ segment can be fit with five peaks, following the approach of Biesinger *et al.*⁴² with the fitting parameters obtained here similar to parameters used



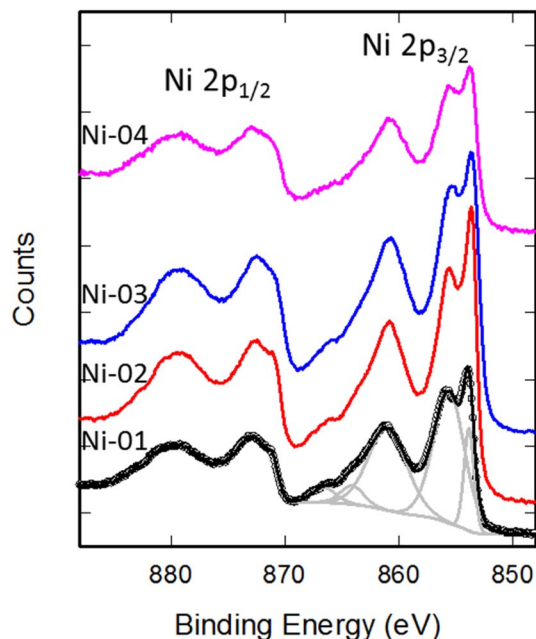


Fig. 6 High resolution spectra of Ni 2p region for four unmodified NiO samples. Fits for Ni-01 are shown by the solid grey (components) and black (sum) lines.

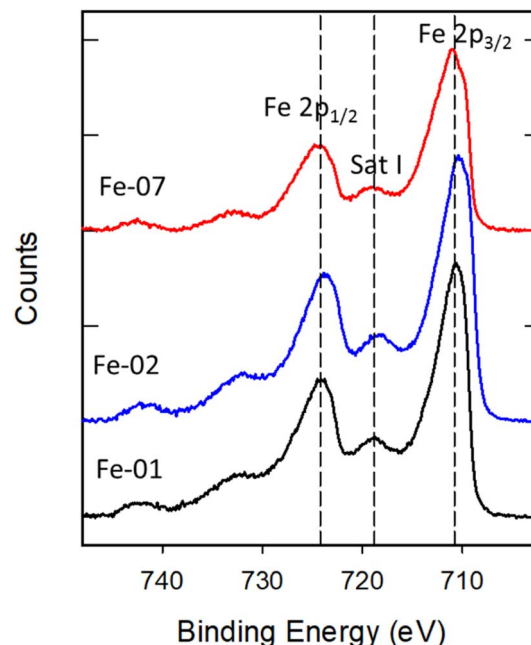


Fig. 7 High resolution spectra of Fe 2p region after Shirley background subtraction for three Fe_2O_3 nanomaterials.

for bulk NiO standard samples (see Table S5[†]). This is consistent with the nickel being primarily in the +2 oxidation state. While the fit is only shown for Ni-01, the other unmodified NiO samples exhibit the same structure, indicating the same oxidation state. No significant changes in this structure (see Fig. S5[†]) or required fitting parameters are observed for the modified particles, indicating that the various modifications do not appear to change the oxidation state.

For the iron oxide nanoparticles the two unmodified (Fe-01 and Fe-02) and the silica coated Fe-07, all exhibit Fe 2p envelopes with a similar structure as shown in Fig. 7, with two main spin-orbit ($2p_{1/2}$ and $2p_{3/2}$) peaks along with three additional less intense satellite features. Fitting the Fe $2p_{3/2}$ peak with multiplet components and comparing with similar fits to appropriate reference samples from the literature³⁷ indicates these spectra are consistent with the expected stoichiometry of Fe_2O_3 in which the iron is present as Fe^{3+} (see Table S6[†]). The presence of the satellite feature at 719 eV (Sat I) is also indicative of Fe^{3+} . Although the three spectra are quite similar, the $2p_{1/2}$ and $2p_{3/2}$ peaks and the Sat I feature for Fe-02 exhibit a slight shift to lower binding energy, suggesting the possible presence of a small amount of Fe^{2+} in this sample. Note that if Fe-07 contained fayalite the Fe would primarily be in the +2 oxidation state, inconsistent with spectrum and the fits for this sample, providing further evidence that this sample consists of silica coated Fe_2O_3 particles, as previously concluded.

For CeO_2 particles, the Ce 3d envelope is quite complex with six peaks clearly resolved and additional features appearing as shoulders as seen in Fig. 8 which depicts Ce 3d spectra for two samples of unmodified NPs. The spectrum observed for Ce-01 is characteristic of samples which are primarily Ce^{4+} ,^{43,44} as

expected for CeO_2 . All of the Ce samples studied here exhibit similar spectra to that of Ce-01 (upper figure), with the exception of Ce-06 (lower figure) in which the relative intensities of the various features are quite different. For cerium oxide samples, the presence of relatively distinct, non-overlapping features facilitates fitting XPS spectra in order to determine the Ce oxidation state as reviewed recently.⁴³ For samples containing a mixture of Ce^{4+} and Ce^{3+} , spectra can be fit with 10 peaks with six peaks from Ce^{4+} and four peaks corresponding to Ce^{3+} . It is necessary to constrain the fitting parameters to reflect the physical origin and relationships between the various features. Adopting this approach to the current data, makes it possible to quantify the Ce^{4+} and Ce^{3+} fractions in these samples. As indicated by the fits shown in Fig. 8, for Ce-01 the Ce^{4+} fraction (red) is dominant while in Ce-06 the Ce^{3+} fraction (blue) increases. Quantifying the fractions based on these fits provides estimates of Ce^{4+} content of 81% for Ce-01 and 42% for Ce-06, with the remaining fraction assumed to be Ce^{3+} . Ce^{4+} fractions for the other CeO_2 samples are given in Table S4[†] with most of them ranging between 70 and 80% and Ce-09 (APTES modified) at 62%. Although the standard deviations on these fractions from the three different measurements on each sample is ~2%, the absolute uncertainty in this quantitative oxidation state determination is likely considerably larger. Nonetheless, it is clear that the Ce-06 sample stands apart as having a larger Ce^{3+} content. This was also the sample that exhibited a significant fraction of P (see Table 4) which appeared to be in the form of phosphate. The increase in Ce^{3+} content on this sample may be explained by the presence of cerium phosphate (CeO_4P) which contains Ce in the +3 oxidation state. It is interesting to note that an earlier study of the biological effects of CeO_2 nanoparticles on aquatic organisms



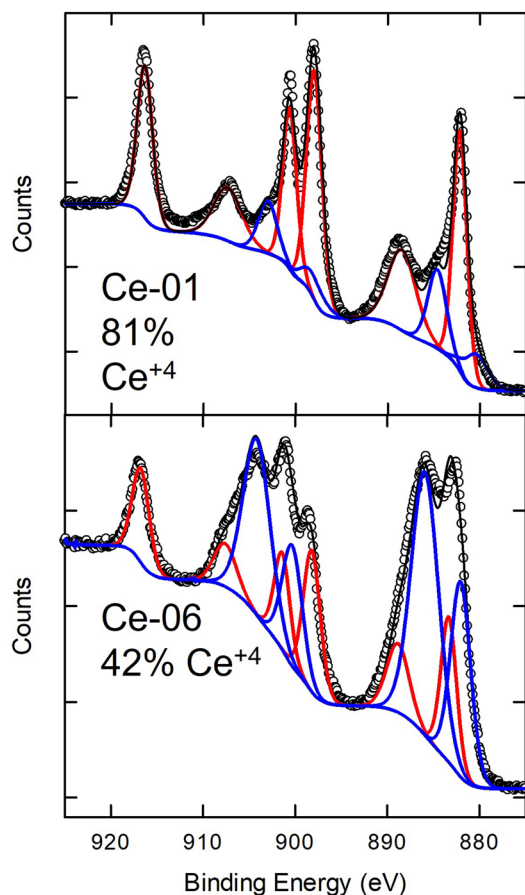


Fig. 8 High resolution spectra of Ce 3d region for two different CeO₂ nanoparticles. The hollow circles are the experimental data points while the solid black line represents the total fit to the data which is a sum of the contributions of two different oxidation states, represented by the red (+4) and blue (+3) lines.

had observed that the presence of Ce³⁺ surface states was the main driver of toxicity.⁴⁵ By contrast, our examination of the cytotoxicity of the same NPs examined here did not find an increased toxicity for Ce-06.¹⁸

Compared with Ni, Fe and Ce, the Mn2p region exhibits less structure as seen in Fig. 9a. There are two peaks due to spin-orbit splitting but the width of the peaks (considerably larger than the spectrometer resolution) as well as the asymmetric peak shape suggests the presence of multiple unresolved features within each of these peaks. All the unmodified Mn oxide samples show this similar 2p structure, which is consistent with expectations for Mn³⁺. Mn-08 shows a subtle change in this structure with a sharper peak present on the lower binding energy side of the 2p_{3/2} envelope. By comparing the spectra with reference spectra for Mn oxides in the literature^{37,46} this feature suggests the presence of some Mn⁴⁺. Multiplet splitting of the Mn 3s is often used for oxidation state determination.⁴⁶ HR spectra of the Mn 3s region are shown in Fig. 9b. All samples apart from Mn-08 exhibit Mn 3s splittings in the range 5.07–5.16, consistent with a Mn valence slightly larger than 3 (~3.2). For Mn-08 this splitting is reduced to 4.64 eV which implies a higher valence (~3.7). The O 1s region, shown in Fig. 9c also shows a slight downshift of ~0.3 eV in the lattice oxygen peak. A similar downshift has been observed previously for MnO₂ samples³⁷ so this observation is consistent with the Mn-08 sample containing a significant fraction of Mn⁴⁺ as suggested by the Mn 2p and Mn 3s spectra.

3.3 Correlation of surface functional group content with qNMR data

As discussed above, XPS has provided information regarding the surface functional group content for the modified metal oxide NPs studied here. In previous work, thermogravimetric analysis (TGA) and qNMR were used to quantify the amount of

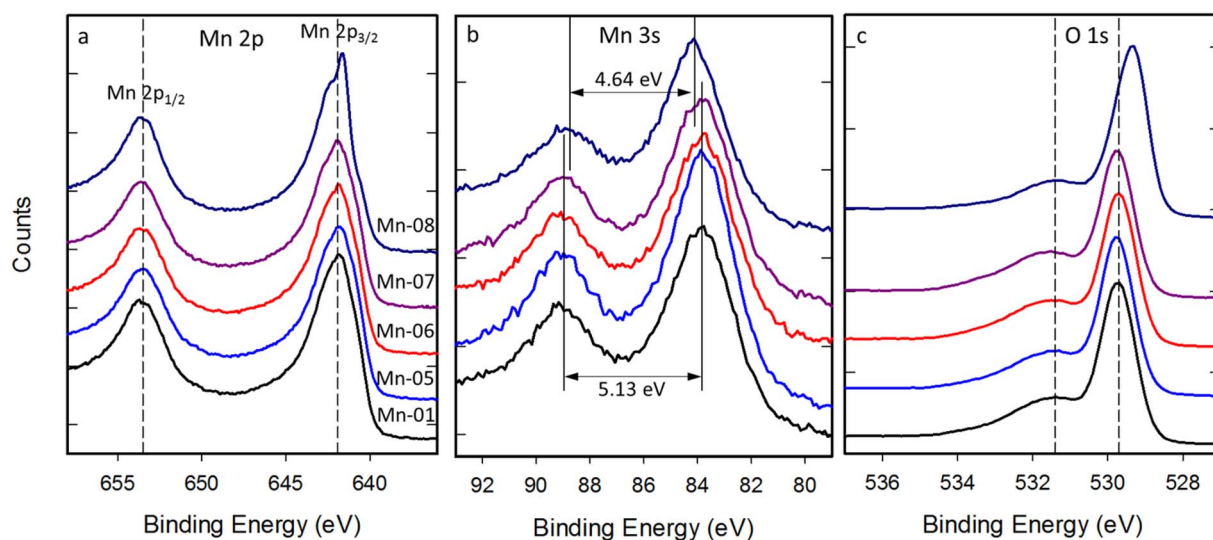
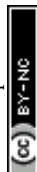


Fig. 9 High resolution spectra of (a) Mn 2p, (b) Mn 3s and (c) O 1s regions for five unmodified Mn₂O₃ nanoparticles.



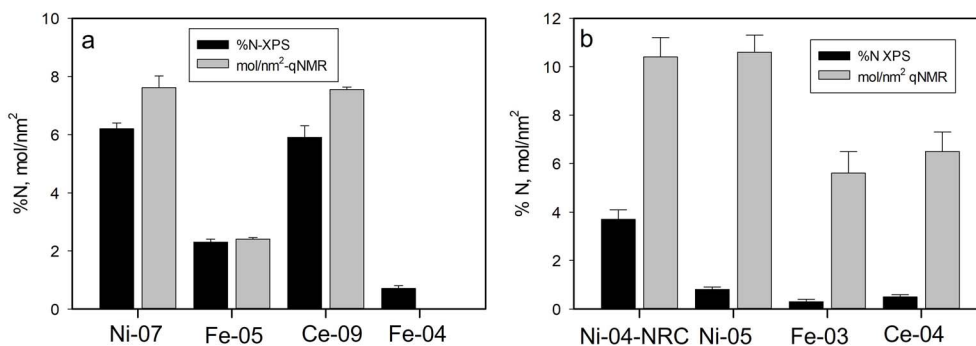


Fig. 10 Comparison of %N measured by XPS with molecules per nm² from qNMR for (a) aminated and (b) PVP modified metal oxide NPs. The qNMR signal for Fe-04 was weak and not quantified.

functionalization on the Ni, Fe and Ce oxide samples investigated here, with general agreement found between those two methods.³³ Here we compare the current XPS data with the previous qNMR results which were determined to be more reliable than the TGA data. XPS measures the atomic composition of the particles directly with the effective probing depth determined by the escape depth of the photoelectrons as discussed above. Photoelectrons from atoms on the surface are seen at full intensity while those emitted from deeper in the sample are subject to scattering and hence their response is attenuated. In contrast, qNMR typically measures the concentration of the functional group in solution after release from the particles, with the concentration being expressed in $\mu\text{mol g}^{-1}$. To facilitate comparison with XPS data, the functional group concentration determined by qNMR can be converted to a coverage of molecules on the surface of the particles, based on the size and density of the particles. This surface coverage is expected to be correlated with XPS signals of these functional groups as demonstrated previously for aminated silica nanoparticles.¹⁷ This correlation is only expected in cases where the functional groups are all attached to the surface of solid (*i.e.* non-porous) particles and hence detectable by XPS with minimal attenuation and also for which these groups contain a unique chemical element that is not also present in the particles or as adventitious contamination. Unfortunately, this precludes the use of two common elements found in surface functional groups, carbon and oxygen, for quantitative analysis of functional group content as these are typically present as adventitious contamination for samples exposed to air. However, for surface modifications that contain nitrogen, as in the case of the aminated and PVP modified particles, we expect the N content to be correlated with the surface coverage of functional groups determined by qNMR.

The relationship between XPS and qNMR data for the aminated metal oxide particles is shown in Fig. 10a. APTES modified samples (Ni-07, Ce-09 and Fe-05) all showed significant N 1s signals in the XPS measurements, corresponding to fractional atomic compositions of N of 2–6%. From the figure we can see a good correlation between the XPS %N and the molecules per nm² based on the qNMR, suggesting that XPS can be used as a quantitative measure of the

surface coverage of amine functional groups on the metal oxide nanoparticles. We note that the non-silane amine functionalized sample studied here (Fe-04) was investigated previously by qNMR and gave only a weak signal consistent with the presence of glycine that was not analyzed quantitatively.⁴⁷ This is consistent with the XPS observations that indicate this sample exhibits approximately one third the nitrogen content seen for the APTES functionalized Fe₂O₃ sample of the same size (Fe-05).

The PVP modified particles also exhibit a nitrogen signal which can be used for XPS quantification. As seen in Fig. 10b, the XPS and qNMR data for these particles do not follow the same trend. While Ni-04-NRC, PVP modified in-house from the unmodified USRN NPs, showed substantial N content, the three commercial PVP modified NPs all exhibited much lower N fractions, suggesting a low level of PVP modification. In contrast, the qNMR data for Ce-04, Ni-05 and Fe-03 all indicated significant amounts of PVP, comparable to or exceeding the levels of amine functionalization measured on the APTES samples.³³ Comparing N signals for the two PVP modified NiO samples, we see that the commercial Ni-05 sample exhibits only 20% of the %N content observed for the in-house sample, even though the qNMR results indicate these two samples exhibit the same level of functionalization. It is interesting to note that for the commercial samples, the XPS %N results suggest that Ni-05 exhibits the largest amount of PVP followed by Ce-04 and then Fe-03, the same trend indicated by the qNMR results. While the reason for the much lower N signal in the commercial samples is unclear, the differences suggest that the commercial PVP modification was likely prepared by a different method than used for the in-house modification. One possibility is that the commercial metal oxide samples were prepared in the presence of PVP⁴⁸ rather than by coating synthesized particles. The former approach would likely lead to less PVP on the surface of the particles and hence a reduced N signal. TEM images of the in-house PVP modified NiO (Ni-04-NRC) particles did not show significant differences compared with the corresponding unmodified particles (Ni-04) (images provided in Fig. S6†). This is not surprising since the thin (~1 nm) coating consisting of light elements (C, O and N) is not expected to generate significant contrast to be clearly distinguished in the images.



4. Conclusions

In this study we have used XPS to examine the chemical composition, oxidation state and surface functional group content of commercially available metal oxide nanoparticle samples from a number of different suppliers. Survey and high-resolution scans have provided information on the atomic composition of the samples, including estimates of the stoichiometry of the metal oxide, the presence of functional groups for modified samples as well as the identification and quantification of any impurities present on the samples. Surprisingly, many of the samples contained substantial amounts of impurities, considerably larger than expected from bulk measurements of impurity concentrations, suggesting these impurities are present at the surface of the particles. The presence of these contaminants may influence their performance in certain applications, and should be taken into account when considering the toxicity and environmental impact of these nanomaterials. The metal oxidation state in these particles is important for some applications and detailed analysis of the metal core level photoelectron spectra was used to provide information regarding the metal oxidation state in the near surface region. In general, the metal oxide particles studied here exhibited the expected oxidation state based on their specified stoichiometry, although a few exceptions were noted. The surface modifications generally did not appear to change the oxidation state. In terms of assessing functional group content, XPS can be complementary to other probes of chemical content such as qNMR. However, XPS is only suitable for functional groups on the surface of the nanoparticles and which contain unique elements not already present on the samples. Good correlation was observed between the nitrogen content observed by XPS and the functional group content obtained by qNMR for amine functionalized metal oxide nanoparticles.

Data availability

Data supporting this article have been included as part of the ESI.†

Conflicts of interest

The authors declare no conflicts of interest.

Acknowledgements

We wish to acknowledge helpful discussions on qNMR with Andreas Brinkman.

References

- 1 D. Comandella, S. Gottardo, I. M. Rio-Echevarria and H. Rauscher, *Nanoscale*, 2020, **12**, 4695–4708.
- 2 J. Labuda, J. Barek, Z. Gajdosechova, H. Goenaga-Infante, L. J. Johnston, Z. Mester and S. Shtykov, *Pure Appl. Chem.*, 2023, **95**, 133–163.
- 3 F. Loosli, K. Rasmussen, H. Rauscher, R. K. Cross, N. Bossa, W. Peijnenburg, J. Arts, M. Matzke, C. Svendsen, D. Spurgeon, P. A. Clausen, E. Ruggiero, W. Wohlleben and F. van der Kammer, *NanoImpact*, 2022, **25**, 100375.
- 4 J. Radnik, X. Knigge, E. Andresen, U. Resch-Genger, D. J. H. Cant, A. G. Shard and C. A. Clifford, *Anal. Bioanal. Chem.*, 2022, **414**, 4331–4345.
- 5 D. Geißler, N. Nirmalananthan-Budau, L. Scholtz, I. Tavernaro and U. Resch-Genger, *Microchim. Acta*, 2021, **188**, 321.
- 6 D. R. Baer, M. H. Engelhard, G. E. Johnson, J. Laskin, J. Lai, K. Mueller, P. Munusamy, S. Thevuthasan, H. Wang, N. Washton, A. Elder, B. L. Baisch, A. Karakoti, S. V. N. T. Kuchibhatla and D. Moon, *J. Vac. Sci. Technol., A*, 2013, **31**, 050820.
- 7 P. A. Clausen, V. Kofoed-Sørensen, A. W. Nørgaard, N. M. Sahlgren and K. A. Jensen, *Materials*, 2019, **12**, 3657.
- 8 A. Hennig, H. Borchering, C. Jaeger, S. Hatami, C. Würth, A. Hoffmann, K. Hoffmann, T. Thiele, U. Schedler and U. Resch-Genger, *J. Am. Chem. Soc.*, 2012, **134**, 8268–8276.
- 9 I.-L. Hsiao, S. Fritsch-Decker, A. Leidner, M. Al-Rawi, V. Hug, S. Diabaté, S. L. Grage, M. Meffert, T. Stoeger, D. Gerthsen, A. S. Ulrich, C. M. Niemeyer and C. Weiss, *Small*, 2019, **15**, 1805400.
- 10 N. Kong, J. Zhou, J. Park, S. Xie, O. Ramström and M. Yan, *Anal. Chem.*, 2015, **87**, 9451–9458.
- 11 F. Kunc, V. Balhara, A. Brinkmann, Y. Sun, D. M. Leek and L. J. Johnston, *Anal. Chem.*, 2018, **90**, 13322–13330.
- 12 A. Hennig, P. M. Dietrich, F. Hemmann, T. Thiele, H. Borchering, A. Hoffmann, U. Schedler, C. Jäger, U. Resch-Genger and W. E. S. Unger, *Analyst*, 2015, **140**, 1804–1808.
- 13 L. J. Johnston, X. Du, A. Zborowski and D. C. Kennedy, *Nanomaterials*, 2024, **14**, 198.
- 14 H. J. H. Kang, R. F. Ali, M. T. Y. Paul, M. J. Radford, I. Andreu, A. W. H. Lee and B. D. Gates, *Chem. Commun.*, 2019, 10452–10455.
- 15 N. Nirmalananthan-Budau, B. Rühle, D. Geißler, M. Moser, C. Kläber, A. Schäfer and U. Resch-Genger, *Sci. Rep.*, 2019, **9**, 17577.
- 16 F. Kunc, O. Kodra, A. Brinkmann, G. P. Lopinski and L. J. Johnston, *Nanomaterials*, 2020, **10**, 678.
- 17 Y. Sun, F. Kunc, V. Balhara, B. Coleman, O. Kodra, M. Raza, M. Chen, A. Brinkmann, G. P. Lopinski and L. J. Johnston, *Nanoscale Adv.*, 2019, **1**, 1598–1607.
- 18 M. Bushell, F. Kunc, X. Du, A. Zborowski, L. J. Johnston and D. Kennedy, *International Journal of Translational Medicine*, 2022, **2**, 522–536.
- 19 F. Kunc, M. Bushell, L. J. Johnston and D. Kennedy, *Nanomaterials*, 2022, **12**, 1822.
- 20 D. R. Baer, *J. Vac. Sci. Technol., A*, 2020, **38**, 031201.
- 21 D. R. Baer, D. J. Gaspar, P. Nachimuthu, S. D. Techane and D. G. Castner, *Anal. Bioanal. Chem.*, 2010, **396**, 983–1002.
- 22 A. Müller, T. Heinrich, S. Tougaard, W. S. M. Werner, M. Hronek, V. Kunz, J. Radnik, J. M. Stockmann, V.-D. Hodoroba, S. Benemann, N. Nirmalananthan-



- Budau, D. Geißler, K. Sparnacci and W. E. S. Unger, *J. Phys. Chem. C*, 2019, **123**, 29765–29775.
- 23 M. S. Chavali and M. P. Nikolova, *SN Appl. Sci.*, 2019, **1**, 607.
- 24 U. Farooq, T. Ahmad, F. Naaz and S. ul Islam, *Energy Fuels*, 2023, **37**, 1577–1632.
- 25 T. Naseem and T. Durrani, *Environ. Chem. Ecotoxicol.*, 2021, **3**, 59–75.
- 26 M. P. Nikolova and M. S. Chavali, *Biomimetics*, 2020, **5**, 27.
- 27 Y. Yoon, P. L. Truong, D. Lee and S. H. Ko, *ACS Nanosci. Au*, 2022, **2**, 64–92.
- 28 G. Greczynski and L. Hultman, *ChemPhysChem*, 2017, **18**, 1507–1512.
- 29 G. Greczynski and L. Hultman, *Prog. Mater. Sci.*, 2020, **107**, 100591.
- 30 M. C. Biesinger, *Appl. Surf. Sci.*, 2022, **597**, 153681.
- 31 C. R. Brundle and B. V. Crist, *J. Vac. Sci. Technol., A*, 2020, **38**, 041001.
- 32 N. A. Belsey, D. J. H. Cant, C. Minelli, J. R. Araujo, B. Bock, P. Brüner, D. G. Castner, G. Ceccone, J. D. P. Counsell, P. M. Dietrich, M. H. Engelhard, S. Fearn, C. E. Galhardo, H. Kalbe, J. W. Kim, L. Lartundo-Rojas, H. S. Luftman, T. S. Nunney, J. Pseiner, E. F. Smith, V. Spampinato, J. M. Sturm, A. G. Thomas, J. P. W. Treacy, L. Veith, M. Wagstaffe, H. Wang, M. Wang, Y.-C. Wang, W. Werner, L. Yang and A. G. Shard, *J. Phys. Chem. C*, 2016, **120**, 24070–24079.
- 33 F. Kunc, M. Gallernault, O. Kodra, A. Brinkmann, G. P. Lopinski and L. J. Johnston, *Anal. Bioanal. Chem.*, 2022, **414**, 4409–4425.
- 34 T. R. Gengenbach, G. H. Major, M. R. Linford and C. D. Easton, *J. Vac. Sci. Technol., A*, 2021, **39**, 013204.
- 35 M. A. Peck and M. A. Langell, *Chem. Mater.*, 2012, **24**, 4483–4490.
- 36 R. Blume, W. Calvet, A. Ghafari, T. Mayer, A. Knop-Gericke and R. Schlögl, *ChemPhysChem*, 2023, **24**, e2023002.
- 37 M. C. Biesinger, B. P. Payne, A. P. Grosvenor, L. W. M. Lau, A. R. Gerson and R. S. C. Smart, *Appl. Surf. Sci.*, 2011, **257**, 2717–2730.
- 38 P. Guo and C. Wang, *RSC Adv.*, 2017, **7**, 4437–4443.
- 39 K. I. Maslakov, Y. A. Teterin, A. J. Popel, A. Y. Teterin, K. E. Ivanov, S. N. Kalmykov, V. G. Petrov, P. K. Petrov and I. Farnan, *Appl. Surf. Sci.*, 2018, **448**, 154–162.
- 40 M. C. Biesinger, B. P. Payne, B. R. Hart, A. P. Grosvenor, N. S. McIntyre, L. W. Lau and R. S. Smart, *J. Phys.: Conf. Ser.*, 2008, **100**, 012025.
- 41 A. P. Grosvenor, B. A. Kobe, M. C. Biesinger and N. S. McIntyre, *Surf. Interface Anal.*, 2004, **36**, 1564–1574.
- 42 M. C. Biesinger, B. P. Payne, L. W. M. Lau, A. Gerson and R. S. C. Smart, *Surf. Interface Anal.*, 2009, **41**, 324–332.
- 43 E. Paparazzo, *J. Phys.: Condens. Matter*, 2018, **30**, 343003.
- 44 C. M. Sims, R. A. Maier, A. C. Johnston-Peck, J. M. Gorham, V. A. Hackley and B. C. Nelson, *Nanotechnology*, 2019, **30**, 085703.
- 45 G. Pulido-Reyes, I. Rodea-Palomares, S. Das, T. S. Sakthivel, F. Leganes, R. Rosal, S. Seal and F. Fernández-Piñas, *Sci. Rep.*, 2015, **5**, 15613.
- 46 E. S. Ilton, J. E. Post, P. J. Heaney, F. T. Ling and S. N. Kerisit, *Appl. Surf. Sci.*, 2016, 475–485.
- 47 F. Kunc and L. J. Johnston, unpublished results.
- 48 K. M. Koczur, S. Mourdikoudis, L. Polavarapu and S. E. Skrabalak, *Dalton Trans.*, 2015, **44**, 17883–17905.

