



Cite this: *J. Anal. At. Spectrom.*, 2022, **37**, 1203

Size determination of nanoparticles by ICP-ToF-MS using isotope dilution in microdroplets

Marcus von der Au, Sebastian Faßbender,  Michail Ioannis Chronakis, 
 Jochen Vogl  and Björn Meermann *

Within this work, the combination of a microdroplet generator and an ICP-ToF-MS for nanoparticle analysis is presented. For the size determination of platinum nanoparticles an on-line isotope dilution analysis approach was developed. The $^{194}\text{Pt}/^{195}\text{Pt}$ isotopic ratio was used for the characterization of the particles, while the $^{182}\text{W}/^{183}\text{W}$ isotopic ratio was monitored simultaneously for mass bias correction. The on-line ID-MDG-sp-ICP-ToF-MS approach was deployed for the size determination of three platinum nanoparticle samples (50 nm, 63 nm, 70 nm); for validation, complementary size characterization techniques (sp-ICP-ToF-MS and TEM) were used. The robustness of this technique was evidenced, by using sodium chloride concentrations up to 100 mg L^{-1} as a matrix component. Our new on-line ID MDG-sp-ICP-ToF-MS approach is a promising tool for the fast and reliable determination of nanoparticles' size in severe matrix concentrations, e.g., environmental samples.

Received 28th February 2022
 Accepted 9th May 2022

DOI: 10.1039/d2ja00072e

rsc.li/jaas

Introduction

The incorporation of nanoparticles in a variety of products, as well as the release of such particles in the environment, has dramatically increased during the past years.¹ Therefore, small particles in the nanometer range have become a major concern in terms of ecotoxicology and human health.^{2,3} Furthermore, nanoparticles find their way into regulations and thus, uniform definitions.⁴ Hence, the presence of nanoparticles (NP) and their composition must be monitored, and rapid and reliable techniques are essential for this purpose. A detector which allows for an extremely sensitive elemental detection of single particles in short transient signals is ICP-MS in single particle mode, sp-ICP-MS.⁵ The utilization of this easy-to-use technique, which is mainly based on the appropriate dilution of a nanoparticle suspension, increased rapidly after its introduction in 1993 by Nomizu *et al.* and developed further by Degueldre *et al.* in 2003.^{6–8} Although the collection of data is simple, the quantification as well as deviation of nanoparticles' size information is difficult.⁹ One of the key aspects, which makes this technique difficult to use, is the determination of the transport efficiency (TE). Since most approaches are using liquid standards to quantify the metal content of the nanoparticles, the TE of the sample introduction system being used must be determined. For this purpose, adequately characterized standards are needed.⁹ The microdroplet generator (MDG), as a sample introduction system, is an upcoming technique that eliminates

the need for these standards.^{10–16} This technique allows for calibration of the system by introducing a liquid in the form of monodisperse microdroplets resulting in the same type of signal as nanoparticles.¹⁴ By producing monodisperse droplets, the volume of the droplets and therefore the absolute mass of introduced calibrants are known and the system can be calibrated, since a TE for a droplet of 100% can be achieved.¹⁴

The frequency of the generation of microdroplets can be adjusted in the range of a few Hz up to the kHz range. Since the ion cloud of the droplet is within the time range of the typical nanoparticle's ion cloud, fast ion counting, and data acquisition is necessary. With classic ICP-MS (scanning) mass analyzers, the number of isotopes which can be determined in a single event is limited. With the latest development of the ICP-ToF-MS, this limitation can be overcome and simultaneous information about nearly the whole periodic table can be acquired in short transient signals/single nanoparticles.¹⁷ Due to the (quasi) simultaneous detection of all elements/all isotopes at the same time also isotope dilution analysis (IDA) is possible within single particle events.¹⁸ With the monodisperse droplets generated by a microdroplet generator, the absolute mass of the isotopic standard introduced into the plasma is known. Therefore, all values needed for the on-line ID approach are accessible. Furthermore, by using the ToF mass analyzer more than one isotope system can be investigated. Also, the possibility to investigate and characterize more than one element in nanoparticles allows for the differentiation between natural and engineered nanoparticles.¹⁹

The suitability of on-line ID for nanoparticles has already been reported by Telgmann *et al.*²⁰ and Sötebier *et al.*²¹ for silver nanoparticles in recent years. Since for these studies scanning

Division 1.1 – Inorganic Trace Analysis, Federal Institute for Materials Research and Testing (BAM), Richard-Willstätter-Straße 11, 12489 Berlin, Germany. E-mail: bjoern.meermann@bam.de



mass analyzers were used, it was not possible to determine both silver isotopes ($^{107}\text{Ag}/^{109}\text{Ag}$) in single particles. Furthermore, this limitation does not allow for a simultaneous mass bias correction. By using a mass bias correction, the correctness of the data may be improved, which may affect the result and therefore the determined diameter. In addition, both studies rely on the correct determination of the TE and the stability of this value during the measurements. This lowers the precision and reproducibility of these approaches.

Within our study, the use of the MDG as sample introduction system is being proposed for the direct mass quantification within single nanoparticles *via* on-line ID. With this approach, the TE is not needed, and a direct calibration *via* liquid standards is enabled. Furthermore, by using an ICP-ToF-MS detector, the isotope ratio can be determined for each individual particle; for calibration as well as mass bias correction a second isotope system is applicable.

Experimental

For the on-line ID MDG-sp-ICP-ToF-MS measurements, a combination of an ICP-ToF-MS system (2R, TOFWERK, Switzerland) and a microdroplet generator introduction, control and autosampler system (TOFWERK) equipped with a 50 μm nozzle microdroplet generator (microdrop Technologies

were used for the mass bias correction; a Cs standard solution was used as internal standard for droplet monitoring (all standard solutions: 1000 mg L^{-1} Certipur, Merck KGaA, Germany). During measurements the three elements (*i.e.*, Cs, W, Pb) were diluted to a mass concentration of 10 $\mu\text{g L}^{-1}$ each. For the isotope spike solution, the reference material ERM-AE141 ($w(\text{Pt}) = 19.90 \text{ mg kg}$; $n(^{194}\text{Pt})/n(\text{Pt}) = 0.9143$; $n(^{195}\text{Pt})/n(\text{Pt}) = 0.0676$; BAM) was used. The data treatment, calculations and fits were done in Origin 2021 Pro (OriginLab Corporation, USA). All ICP-ToF-MS measurements were carried out in triplicates.

Results and discussion

Isotope dilution is one of the most precise methods available for mass fraction determination. However, the exact knowledge of the spike's mass fraction as well as the isotope ratio of the spike and the absolute amount of spike solution introduced into the system is required to provide correct results. For the determination of the mass fraction of interest in single particles with a microdroplet generator (MDG) as sample introduction system, the following equation (eqn (1)) was derived and applied. Since the volume of a nanoparticle (about 10^{-8} pL) is negligible (compared to the volume of the droplets, which is about 10^2 pL), it is excluded from the calculations:

$$m_{\text{sample}} = \frac{M_{\text{Pt},x} \times m_{\text{spike, stock}} \times m_{\text{spike, sample}} \times w_{\text{spike, stock}} \times V_{\text{droplet}} \times \left(R_{^{194}\text{Pt}/^{194}\text{Pt, measured}} \times A_{^{195}\text{Pt}, y} - A_{^{194}\text{Pt}, y} \right)}{M_{\text{Pt},y} \times m_{\text{total, stock}} \times V_{\text{total, sample}} \times \left(A_{^{194}\text{Pt}, x} \times R_{^{194}\text{Pt}/^{195}\text{Pt, measured}} - A_{^{195}\text{Pt}, x} \right)} \quad (1)$$

GmbH, Germany) was used. The sp-ICP-ToF-MS analysis was carried out with the standard sample introduction system that consists of a concentric nebulizer and a cyclonic spray chamber. The sp-ICP-ToF-MS parameters were tuned daily to optimize the signal intensity. The on-line ID MDG-sp-ICP-ToF-MS approach was optimized daily to ensure the introduction of all generated droplets into the plasma and a high sensitivity. The helium flow rate used for the MDG was kept at 0.5 L min^{-1} and the droplet frequency at 50 Hz. Also, the RF power for all experiments was set at 1550 W.

The platinum and gold nanoparticles used were obtained from nanoComposix (50 nm and 70 nm platinum NPs and 100 nm Gold NPs, USA). Further TEM characterized platinum particles were provided by division 6.5 (Federal Institute for Materials Research and Testing (BAM), Germany). For the analysis with the sp-ICP-ToF-MS and the MDG-sp-ICP-ToF-MS approaches, all samples were prepared in precleaned PP-vials (DigiTUBEs 15 mL and 50 mL, SCP Science Corp., USA) leached with diluted sub-boiled nitric acid ($\phi = 1.3\%$) in ultra-pure water for at least one week. The particles were diluted according to the different methods to achieve a sufficient number of investigated particles. For external calibration, gold and platinum standard solutions were used. For the on-line ID-MDG-sp-ICP-ToF-MS approach, Pb and W standard solutions

Within eqn (1) M is the natural (M_x) and spiked (M_y) molar mass of the analyte, $m_{\text{spike, stock}}$ is the mass of the spike solution used (ERM-AE-141) to prepare the stock solution, $m_{\text{total, stock}}$ is the total mass of the stock solution prepared, $m_{\text{spike, sample}}$ is the mass of the stock solution added to the sample and m_{sample} the obtained sample mass; $w_{\text{spike, stock}}$ is the mass fraction of the platinum in the spike solution (ERM-AE-141); V_{droplet} is the volume of the generated droplet and $V_{\text{total, sample}}$ the volume of the final sample solution; R is the ratio of the isotopic system used ("measured" refers to the uncorrected ratio and xy refers to the corrected ratio) and A (x refers to the sample and y refers to the spike) the abundance of the isotopes taken for calculation. By using a reference material, only the $^{194}\text{Pt}/^{195}\text{Pt}$ isotope ratio in the sample and V_{droplet} must be determined, which makes the calculation fast and easy. The values regarding the isotopic abundances and atomic mass are provided by the CIAAW²² (for the natural platinum isotopic composition) and the reference material certificate (for the platinum isotopic composition and the concentration of the spike solution). The masses and volumes used in the calculation are determined prior to the analysis either volumetrically or gravimetrically.

V_{droplet} is automatically calculated *via* the diameter and provided by the MDG-System software. To ensure the correct droplet volume, the build-in microscope and the software used



were size calibrated by means of a 50 μm pin provided by TOFWERK. Since V_{droplet} [pL] is calculated by the software, also the final volume $V_{\text{total,sample}}$ was measured volumetrically to avoid conversion errors and to keep the equation consistent. The advantage of the ICP-ToF-MS lies in the quasi-simultaneous detection of (nearly) all elements as well as isotopes, all time in short transient signals – thus, all information needed for the on-line ID MDG-sp-ICP-ToF-MS approach is obtained within one single run.

To cope with potential mass bias effects a second isotopic system was monitored simultaneously for correction. For platinum, two close systems with a similar isotopic pattern were tested for this purpose. Since the chance that Pb in the sample is higher, the $^{182}\text{W}/^{183}\text{W}$ (natural abundance 26.3%/30.6%) isotope ratio was used, which has a similar isotopic pattern and mass in comparison to the $^{194}\text{Pt}/^{195}\text{Pt}$ (natural abundance 32.9%/33.5%) isotope ratio. For the platinum $^{194}\text{Pt}/^{195}\text{Pt}$ isotope ratio correction with the $^{182}\text{W}/^{183}\text{W}$ isotopic ratio, the following equations (eqn (2) and (3)) were used. In these equations f is the mass bias correction factor.

$$f = \frac{R_{^{182}\text{W}/^{183}\text{W,measured}} \times A_{^{183}\text{W}}}{A_{^{182}\text{W}} \times \ln\left(\frac{M_{^{182}\text{W}}}{M_{^{183}\text{W}}}\right)} \quad (2)$$

$$R_{^{194}\text{Pt}/^{195}\text{Pt,xy}} = 10^{\frac{R_{\text{Pt}}}{\left(\frac{M_{^{194}\text{Pt,natural}}}{M_{^{195}\text{Pt,natural}}}\right)^f}} \quad (3)$$

Taking eqn (3) into account, a modified version of eqn (1) (eqn (4)) can be derived, taking the correction factor into account. Eqn (4) was applied for all further calculations upon on-line MDG-sp-ICP-ToF-MS analyses:

$$m_{\text{sample}} = \frac{M_{\text{Pt,x}} \times m_{\text{spike,stock}} \times m_{\text{spike,sample}} \times w_{\text{spike,stock}} \times V_{\text{droplet}} \times \left(R_{^{194}\text{Pt}/^{195}\text{Pt,xy}} \times A_{^{195}\text{Pt,y}} - A_{^{194}\text{Pt,y}}\right)}{M_{\text{Pt,y}} \times m_{\text{total,stock}} \times V_{\text{total,sample}} \times \left(A_{^{194}\text{Pt,x}} \times R_{^{194}\text{Pt}/^{195}\text{Pt,xy}} - A_{^{195}\text{Pt,x}}\right)} \quad (4)$$

The nanoparticles' diameter (d_{NP}) can then be obtained using m_{sample} and the density of platinum particles (ρ_{Pt}), supposing a spherical shape for the particles (verified by TEM) and assuming that the particle consists of Pt only (100%) (eqn (5)).

$$d_{\text{NP}} = 2 \times \left(\frac{3 \times m_{\text{sample}}}{\rho_{\text{Pt}} \times 4 \times \pi}\right)^{\frac{1}{3}} \quad (5)$$

With the high number of droplets measured using the MDG-sp-ICP-ToF-MS approach, a Gaussian distribution was observed for all particles/droplets. Since the fluctuation from droplet to droplet was too high for the determination of the values needed for m_{sample} calculation based on a droplet-to-droplet basis the center of Gaussian fits were applied. Histograms were

calculated *via* Origin 2021 Pro. The droplet volume (Fig. 1(a)) and the $^{182}\text{W}/^{183}\text{W}$ isotopic ratio (Fig. 1(b)) are not affected by the added isotope spike and therefore for these two a mono-modal distribution is expected. In contrast, for the $^{194}\text{Pt}/^{195}\text{Pt}$ ratio, a bimodal distribution should occur – (i) the isotope ratio of the spike solution within a single droplet (Fig. 1c(i)) and (ii) the mixed isotope ratio of the spike solution and a Pt-nanoparticle (Fig. 1c(ii)).

For the on-line isotope dilution analysis, the shifts in the isotopic ratios are crucial. Therefore, at first, different spike concentrations were used to identify an appropriate working range for the on-line ID-MDG-sp-ICP-ToF-MS approach. To derive an appropriate isotope ratio of the spike solution, a threshold was set: once a distinction of droplets from the MDG with and without Pt-particles included using the Origin software package is not possible anymore, an upper spike concentration limit is obtained. Thus, the optimal spike concentration/spike isotope ratio lies below this threshold. For isotopic spike optimization two different commercially available platinum nanoparticle suspensions (nominal size 50 nm and 70 nm) were investigated: five different spike isotope ratios for the 50 nm and four different spike isotope ratios for the 70 nm platinum nanoparticles were tested. The results obtained by the on-line ID-MDG-sp-ICP-ToF-MS approach in comparison to classical sp-ICP-ToF-MS are shown in Fig. 2. To determine the mass fraction with the classical sp-ICP-ToF-MS approach, the calibration strategy by Pace *et al.*²³ was applied.

Fig. 2 illustrates the correlation of the Pt nanoparticle's diameter and the applied spike concentration as well as the resulting isotope ratio of the blend. For both nanoparticle sizes a threshold for the Pt isotopic ratio of the blend of 6 was observed. With lower values an underestimation of the diameter occurs. This is mainly due to “underspiking” and the resulting error magnification, since the spike concentration and spike isotope ratio do not sufficiently change the “natural” Pt

isotope ratio of the particles.^{24,25} Once the $^{194}\text{Pt}/^{195}\text{Pt}$ isotope ratio reaches an appropriate value (approx. $^{194}\text{Pt}/^{195}\text{Pt} = 6$), a reproducible diameter of $42 \text{ nm} \pm 1 \text{ nm}$ and $62 \text{ nm} \pm 1 \text{ nm}$ is achieved. Due to the high discrepancy between the given size according to the given and the calculated size of the nanoparticles, a “classical” sp-ICP-ToF-MS approach was also applied to verify the obtained values. This approach provided identical values for the particles' diameter of $42 \text{ nm} \pm 1 \text{ nm}$ and $62 \text{ nm} \pm 1 \text{ nm}$ respectively. Also, the “classical” sp-ICP-ToF-MS approach delivered the same particle diameter, lower than the values provided by the manufacturer. Thus, it was concluded that the nanoparticles' size had changed, most probably during the storage time. To verify this assumption, a third nanoparticle sample was investigated. For this purpose, a synthesized and well characterized platinum nanoparticle sample was used. For





Fig. 1 Volume distribution of the droplets of the MDG (a); isotope ratio distribution of the tungsten ($^{182}\text{W}/^{183}\text{W}$) (b) and platinum ($^{194}\text{Pt}/^{195}\text{Pt}$) without (c, (i)) and with (c, (ii)) platinum nanoparticles (50 nm) present in the droplets.



Fig. 2 Calculated diameters for the platinum nanoparticles (nominal diameter 50 nm and 70 nm). The squares and circles indicate the results obtained by on-line ID-MDG-sp-ICP-ToF-MS for different $^{194}\text{Pt}/^{195}\text{Pt}$ isotopic ratios and the dashed lines indicate the results obtained by classical sp-ICP-ToF-MS. The error bars represent the standard deviation of three independent measurements.

these particles, a mean diameter of 63 nm was determined using TEM. For the on-line ID-MDG-sp-ICP-ToF-MS measurements, a Pt isotope ratio ($^{194}\text{Pt}/^{195}\text{Pt}$) above 6 was adjusted; the on-line ID-MDG-sp-ICP-ToF-MS approach delivered a diameter of $64 \text{ nm} \pm 1 \text{ nm}$. Thus, a perfect agreement between TEM and on-line ID-MDG-sp-ICP-ToF-MS results could be shown. The results are within the same range as the “classical” sp-ICP-MS approach, but with less effort and steps and therefore this method is less prone to errors. Also, the stability, identity, and size of nanoparticles, which are applied for calibration of the transport efficiency, are difficult to check and control, since usually another technique and/or instrument (e.g., TEM) is needed. Whereas the concentration and isotope ratio of the spike solution can be determined relatively easily.

To test the matrix robustness of the proposed method sodium chloride was added to samples containing the spike solution and platinum nanoparticles with the nominal diameter of 50 nm. A mixed $^{194}\text{Pt}/^{195}\text{Pt}$ isotopic ratio of the natural

platinum nanoparticles and the platinum spike of a ratio of about 10 was adjusted in accordance with the previous findings. The calculated diameter for the samples were $42 \text{ nm} \pm 1 \text{ nm}$ for the sample without added sodium chloride and $42 \text{ nm} \pm 1 \text{ nm}$ for the test solutions with added sodium chloride at three different concentration levels: 1 mg L^{-1} , 10 mg L^{-1} and 100 mg L^{-1} . The determined diameter of the Pt nanoparticles was constant even at the highest sodium chloride concentration. However, the formation of sodium chloride crystals at the tip of the nozzle of the MDG was observed, therefore higher sodium chloride concentrations were not tested. The obtained results illustrate the matrix robustness of the proposed method within the tested range. As the results suggest, the on-line ID-MDG-sp-ICP-ToF-MS approach is suitable to characterize the size of nanoparticles also in heavy (environmental) matrices, with little to no sample preparation.

Conclusion

The combination of on-line ID, MDG and sp-ICP-ToF-MS is fit for purpose for nanoparticle size characterization. With higher reproducibility and better characterization capabilities of the droplets produced by the MDG the precise determination of the nanoparticles' size is enabled by on-line ID. With on-line ID for the size determination of nanoparticles, TE as well as calibration curve determination becomes redundant. Our new approach is highly reproducible and robust against matrix effects – the latter is of particular interest for environmental samples, *i.e.*, diluted sea water. In addition, the ToF mass analyzer has no limit in the number of simultaneously detectable isotopes/elements, thus, more than one type of nanoparticles can be characterized simultaneously – provided that the element under investigation comprises at least two interference free and stable isotopes. In addition, this work lays the foundation for the characterization of multi-element NPs as well as for uncertainty calculations, which would allow this technique as complementary tool for the certification of candidate materials.

Conflicts of interest

The authors declare no competing financial interest.



Acknowledgements

The authors want to thank Andreas F. Thünemann and Tina Rybak (Federal Institute for Materials Research and Testing (BAM), Division 6.5 – Synthesis and Scattering of Nanostructured Materials) for providing the platinum nanoparticles. Furthermore, the authors want to thank the German Federal Ministry for Economic Affairs and Climate Action (BMWK) for the financial support.

References

- 1 Q. Bai, Y. Yin, Y. Liu, H. Jiang, M. Wu, W. Wang, Z. Tan, J. Liu, M. H. Moon and B. Xing, *Appl. Spectrosc. Rev.*, 2021, 1–22.
- 2 R. D. Handy and B. J. Shaw, *Health Risk Soc.*, 2007, 9, 125–144.
- 3 R. D. Handy, F. Von der Kammer, J. R. Lead, M. Hassellöv, R. Owen and M. Crane, *Ecotoxicology*, 2008, 17, 287–314.
- 4 European Commission, *Commission Recommendation of 18 October 2011 on the Definition of Nanomaterial (2011/696/EU)*, 2011.
- 5 S. Fernández-Trujillo, M. Jiménez-Moreno, Á. Ríos and R. d. C. R. Martín-Doimeadios, *J. Anal. At. Spectrom.*, 2021, 36, 528–534.
- 6 M. D. Montano, J. W. Olesik, A. G. Barber, K. Challis and J. F. Ranville, *Anal. Bioanal. Chem.*, 2016, 408, 5053–5074.
- 7 C. Degueldre and P. Y. Favarger, *Colloids Surf., A*, 2003, 217, 137–142.
- 8 T. Nomizu, S. Kaneco, T. Tanaka, T. Yamamoto and H. Kawaguchi, *Anal. Sci.*, 1993, 9, 843–846.
- 9 F. Laborda, A. C. Gimenez-Ingalaturre and E. Bolea, in *Comprehensive Analytical Chemistry*, Elsevier, 2021, vol. 93, pp. 35–67.
- 10 S. Gschwind, L. Flamigni, J. Koch, O. Borovinskaya, S. Groh, K. Niemax and D. Günther, *J. Anal. At. Spectrom.*, 2011, 26, 1166–1174.
- 11 K. Mehrabi, R. Kaegi, D. Günther and A. Gundlach-Graham, *Environ. Sci.: Nano*, 2021, 8, 1211–1225.
- 12 L. Hendriks, A. Gundlach-Graham and D. Günther, *J. Anal. At. Spectrom.*, 2019, 34, 1900–1909.
- 13 L. Hendriks, A. Gundlach-Graham and D. Günther, *Chimia*, 2018, 72, 221–226.
- 14 O. Borovinskaya, S. Gschwind, B. Hattendorf, M. Tanner and D. Günther, *Anal. Chem.*, 2014, 86, 8142–8148.
- 15 J. W. Olesik and S. E. Hobbs, *Anal. Chem.*, 1994, 66, 3371–3378.
- 16 M. P. Dziewatkoski, L. B. Daniels and J. W. Olesik, *Anal. Chem.*, 1996, 68, 1101–1109.
- 17 M. von der Au, O. Borovinskaya, L. Flamigni, K. Kuhlmeier, C. Büchel and B. Meermann, *Algal Res.*, 2020, 49, 101964.
- 18 S. Faßbender, M. von der Au, M. Koenig, J. Pelzer, C. Piechotta, J. Vogl and B. Meermann, *Anal. Bioanal. Chem.*, 2021, 413, 5279–5289.
- 19 A. Praetorius, A. Gundlach-Graham, E. Goldberg, W. Fabienke, J. Navratilova, A. Gondikas, R. Kaegi, D. Günther, T. Hofmann and F. von der Kammer, *Environ. Sci.: Nano*, 2017, 4, 307–314.
- 20 L. Telgmann, C. Metcalfe and H. Hintelmann, *J. Anal. At. Spectrom.*, 2014, 29, 1265–1272.
- 21 C. A. Sötebier, D. J. Kutscher, L. Rottmann, N. Jakubowski, U. Panne and J. Bettmer, *J. Anal. At. Spectrom.*, 2016, 31, 2045–2052.
- 22 IUPAC Commission on Isotopic Abundances and Atomic Weights, (accessed February 15th 2022), <https://ciaaw.org>.
- 23 H. E. Pace, N. J. Rogers, C. Jarolimek, V. A. Coleman, C. P. Higgins and J. F. Ranville, *Anal. Chem.*, 2011, 83, 9361–9369.
- 24 J. D. Fassett and P. J. Paulsen, *Anal. Chem.*, 1989, 61, 643A–[649A].
- 25 J. Vogl, *J. Anal. At. Spectrom.*, 2007, 22, 475–492.

