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Polymers from sugars and CS₂: synthesis and ring-opening polymerisation of sulfur-containing monomers derived from 2-deoxy-D-ribose and D-xylose†

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Thionocarbonate (–O–CS–O–) and xanthate (–S–CS–O–) cyclic monomers were synthesised by cyclothiocarbonation of 2-deoxy-D-ribose- and D-xylose-derived diols with carbon disulfide, then polymerised using organocatalytic ring-opening methods. Regular polymer linkages were obtained, with the sugar backbone influencing the regioselectivity of monomer opening. Thermal analysis revealed lower glass transition temperatures compared to carbonate analogues and a low onset of thermal degradation.

Natural monosaccharides are a pool of readily available and functional building blocks that are cheap, non-toxic, stereochemically diverse, and offer a renewable alternative to petroleum-based resources for the synthesis of monomers and polymers.¹ Sugars have been used in polymer synthesis for example as pendant groups² or incorporated into main polymer chains *via* step-growth methods from aldoric esters and alditols.^{1b} Driven by the versatility and accurate control offered by ring-opening polymerisation methods (ROP),³ combined with the ability of sugars to be functionalised to adjust the properties of the resulting polymers, recent efforts have been devoted to the synthesis and subsequent ROP of sugar-based cyclic monomers.^{1a} These include lactams,⁴ phosphoesters⁵ and cyclic carbonates.⁶ Some of our own work in this field has involved using CO₂ to produce sugar-based cyclic carbonates without the need for phosgene derivatives.^{6a–c}

Substitution of some oxygen atoms with sulfur in polymer backbones can result in enhanced physical (*e.g.* increased crystallinity), thermal, mechanical, electrical and optical properties, as well as advanced characteristics such as adhesion to metals, biological and chemical resistance, and biocompatibility.⁷ Therefore, we set out to utilise carbon disulfide (CS₂), the sulfur analogue of CO₂, to make novel sugar-based materials. Sulfur-containing analogues of sugar-based cyclic carbonates have been reported,⁸ but these have been synthesised using CSCI₂ or Im₂CS (Im = imidazole) reagents, and investigated mainly for their tendency to undergo O–S rearrangements.^{6i,8b–e,9} No polymerisation studies have been reported. Furthermore, while being used in the viscose process, CS₂ has only been explored in polymer synthesis as a monomer for homopolymerisation or copolymerisation with epoxides and oxetanes.^{7b,10} Herein, we report the synthesis and polymerisation of novel cyclic xanthate and thionocarbonate monomers from sugar diols and carbon disulfide.

First, the synthesis of a 6-membered cyclic thionocarbonate *trans*-fused to a sugar furanose ring was targeted. Our hypothesis was that CS₂, conversely to CO₂,^{6b,c} would allow the cyclo-carbonation of the *trans* 1,3-diol motif of ribofuranose sugars, because of the longer C–S bond (155.3 pm) in CS₂ compared to C–O (116.3 pm) in CO₂. The resulting monomer would also be highly strained, and therefore prone to ROP. Using an analogous procedure to the one reported previously in our group for the synthesis of cyclic carbonates from diols and CO₂,¹¹ CS₂ was added to a solution of 1-O-methyl-2-deoxy-D-ribofuranose in acetonitrile, in the presence of 1,8-diazabicyclo[5.4.0]-undec-7-ene (DBU). The reaction mixture changed rapidly from colourless to yellow, and ¹H NMR analysis revealed insertion of CS₂ into the sugar hydroxy groups (Fig. S1 and S2 in the ESI†). Furthermore, the C₃ and C₅ atoms of the sugar moiety remained deshielded (signals around 75 ppm), so no O–S rearrangement is believed to take place at this stage. Subsequent addition of mesyl chloride in the presence of triethylamine then led to the formation of cyclic xanthate **1**, which was isolated by column chromatography in a 10% yield

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†Electronic supplementary information (ESI) available: Experimental procedures, NMR spectra of monomers and polymers. Single-crystal X-ray diffraction data for **1**, **2** and **3**. Images of SEC traces, TGA-MS, MALDI-ToF MS and DSC traces. CCDC 1583233–1583235. For ESI and crystallographic data in CIF or other electronic format see DOI: 10.1039/c8py00119g





Scheme 1 Synthesis of monomers **1–3** (see ESI† for detailed procedures).

(Scheme 1). Integration of the anomeric protons in the ^1H NMR spectrum confirmed a 1 : 1 mixture of the two anomers (Fig. S3†). To the best of our knowledge, **1** is the first cyclic xanthate (and in fact the first cyclic carbonate analogue), *trans*-fused to a sugar furanose ring. The nature of **1** was determined by NMR and FTIR spectroscopies, as well as electrospray ionisation mass spectrometry and elemental analysis (see ESI†). In particular, the $^{13}\text{C}\{^1\text{H}\}$ NMR signal for the C=S bond is observed around 208 ppm, characteristic of the C(S)SO environment in cyclic xanthates.^{9e} Because of an adjacent, more electropositive sulfur atom, the C₅ signal in **1** (Fig. S4†) also appears at significantly lower chemical shift (37–38 ppm) compared to the related cyclic carbonates (67 ppm).^{6b} The $^3J_{\text{H3H4}}$ coupling constants (8.7 and 8.8 Hz) are also larger than for the *cis*-configured cyclic monomers we reported previously (4.9 and 5.7 Hz),^{6b} and consistent with a *trans*-fused cyclic monomer. The structure was further corroborated by X-ray diffraction of a co-crystal of both anomers, obtained by recrystallization from hexanes (Fig. S40†). For the α anomer, the furanose ring adopts a 4-*exo*-3-*endo* twist conformation ($^3\text{T}_4$), whereas for the β anomer, the furanose ring has a 3-*endo* (^3E) conformation. From previous mechanistic understanding of the analogous reaction with CO_2 ,¹¹ **1** is not the expected thionocarbonate product, which would form from insertion of CS_2 into a hydroxy group, mesylation of the resulting xanthate, then cyclisation *via* a nucleophilic addition–elimination pathway. As cyclo-thiocarbonation attempts using Im_2CS proved unsuccessful, we suspect that the *trans*-configuration of the diol prevents cyclisation, or that the product is highly unstable. Formation of **1** could however be explained by a putative minor pathway (hence the low yields obtained): insertion of CS_2 into the secondary hydroxy group, followed by mesylation of the 5-OH, then cyclisation *via* intramolecular nucleophilic substitution. **1** could also result from various S–O rearrangements.

Using 1,2-*O*-isopropylidene- α -D-xylofuranose as a substrate was then considered, as in this case the *cis*-configuration of

the 1,3-diol motif should facilitate cyclisation and give the expected thionocarbonate. Using the same procedure, two products were isolated after purification by column chromatography: xanthate **2** and thionocarbonate **3**, in 15% and 48% yield respectively (Scheme 1). **2** displayed a xanthate signal in the $^{13}\text{C}\{^1\text{H}\}$ NMR signal at 208.6 ppm, while a signal at 187.4 ppm was observed for **3**, consistent with a thionocarbonate species (Fig. S15 and S24† respectively).^{9e} Confirmation of the structure of **2** and **3** by NMR and FTIR spectroscopies, as well as electrospray ionisation mass spectrometry and elemental analysis, was further corroborated by single-crystal X-ray diffraction (Fig. S41 and S42†). As expected, using a *cis* 1,3-diol motif yielded thionocarbonate **3**, likely *via* an addition–elimination mechanism for the ring-closing step.¹¹ However, **2** was still formed in small quantity. The possibility that **2** could result from an alternative $\text{S}_{\text{N}}2$ -type mechanism was therefore verified experimentally. 1,2-*O*-isopropylidene- α -D-xylofuranose was tosylated at the 5-position into **4** (Scheme S5†), which, upon addition of CS_2 and DBU, should only lead to the substitution of the tosyl group by the xanthate salt (Scheme S6†). Formation of **2** was indeed observed with approximately 50% conversion after 30 minutes (Fig. S39†). While **3** has been previously synthesised using Im_2CS ,^{8c} to the best of our knowledge **2** is the first cyclic xanthate *cis*-fused to a sugar furanose ring. These two compounds are the sulfur analogues of the carbonate monomer reported by Gross and coworkers.⁶ⁱ

Ring-opening polymerisation of monomers **1–3** were next studied at room temperature in dichloromethane, using 1,5,7-triazabicyclo[4.4.0]dec-5-ene (TBD) organocatalyst, 4-methylbenzyl alcohol initiator, and 1 mol L^{-1} initial monomer concentration (Table 1). TBD is one of the most active organo-

Table 1 Ring-opening polymerisation of **1–3**^a

Entry	M	[M]:[C]:[I] ^b	Time (h)	Conv. ^c (%)	M_n (calc) ^d	M_n (SEC) ^e	D^f
1	1	25 : 1 : 1	0.25	>99	5.3	5.1	1.6
2	1	50 : 1 : 1	0.25	>99	10.4	5.2	1.8
3	1	100 : 1 : 1	0.25	>99	20.7	11.3	2.2
4	2	25 : 1 : 1	0.25	86	5.5	6.0	1.5
5	2	50 : 1 : 1	0.25	87	10.9	5.3	2.2
6	2	50 : 1 : 1	2	>99	12.5	12.6	1.5
7	2	100 : 1 : 1	0.25	86	21.5	15.7	1.7
8	3	25 : 1 : 1	0.25	42	2.6	5.2	1.5
9	3	50 : 1 : 1	0.25	40	4.8	7.4	1.3
10	3	50 : 1 : 1	2	66	7.8	6.7	1.2
11	3	50 ^g : 1 : 1	0.25	52	6.2	7.5	1.5
12	3	50 ^g : 1 : 1	0.5	61	7.2	10.0	1.3
13	3	50 ^g : 1 : 1	3	62	7.3	10.3	1.3
14	3	100 ^g : 1 : 1	0.25	44	10.3	10.6	1.5

^a Polymerisations carried out at room temperature in anhydrous CH_2Cl_2 solvent with initial $[\text{M}]_0 = 1 \text{ mol L}^{-1}$ (M = monomer), unless stated otherwise; all entries correspond to separate experiments. ^b C is the catalyst, TBD, and I is the initiator, 4-MeBnOH. ^c Conversion measured by ^1H NMR determined by relative integration of the anomeric proton in the ^1H NMR spectrum in CDCl_3 . ^d In 10^3 g mol^{-1} , calculated as $M_n(I) + (M_n(\text{monomer}) \times [\text{monomer}]_0 / [\text{I}]_0 \times \text{conv}/100\%)$. ^e In 10^3 g mol^{-1} , estimated by SEC (RI detector) *versus* polystyrene standards with THF eluent. ^f $D = M_n/M_w$. ^g $[\text{M}]_0 = 1.58 \text{ mol L}^{-1}$.



catalysts for the ROP of cyclic carbonates,¹² which usually gives good polymerisation control and limits the amount of cyclic species formed by direct nucleophilic initiation, as with diazabicyclo[5.4.0]undec-7-ene (DBU).¹³ Early trials with DBU gave indeed poor control and decreased activity. Monomer conversion was determined by ¹H NMR spectroscopy. Conformational changes brought about by the release of ring strain upon opening led for all monomers to a downfield shift of H-3 and coalescing of the signals assigned to H-5, as well as a general broadening of the resonances (Fig. S43, S54 and S65† for 1–3, respectively). Polymerisation of 1 proceeded rapidly, reaching >99% conversion in less than 15 min at various monomer: initiator: catalyst feed ratios (Table 1, entries 1–3). Polymerisation of 2 was slightly slower, consistent with the less strained nature of this xanthate compared to 1 (Table 1, entries 5–8), with around 86% monomer conversion after 15 min and >99% after 2 h for a monomer: initiator: catalyst ratio of 50:1:1 (Table 1, entries 5 and 6). Polymerisation of 3 (entries 8–14 in Table 1) proceeded even slower and reached a monomer conversion plateau (66% after 2 hours), indicating a concentration dependent equilibrium polymerisation. A slightly higher initial concentration of 3 (1.58 mol L⁻¹) did not lead to a significant increase in monomer conversion (Table 1, entries 8 and 11). Overall, as expected from the *trans* configuration on its fused furanose ring and the resulting high strain of its xanthate ring, monomer 1 is more reactive towards ROP than monomers 2 and 3, which feature a *cis*-fused furanose ring and are less strained cyclic monomers. Xanthate 2 also appears more reactive than analogous thionocarbonate 3, and we suggest that this is because C–S bonds are easier to break than C–O bonds.

Size exclusion chromatography (SEC) in THF confirmed the polymeric nature of the products, and was used to estimate number-average molecular weights (*M_n*) and dispersities (*D*) versus polystyrene standards. Polymers of up to 11 300 g mol⁻¹ (*D* 2.2) and 15 700 g mol⁻¹ (*D* 1.7) could be achieved from 1 and 2, respectively, but limited agreement between predicted and SEC *M_n*, as well as broad and varying distributions, were observed. The ROP of 3 was more controlled, with better agreement between theoretical molecular weights and those determined by SEC, as well as narrower dispersities. Polymers of up to 10 600 g mol⁻¹ (*D* 1.5) could be obtained from 3. However, for most polymers, inconsistencies and discrepancies between theoretical *M_n*, SEC *M_n* and those estimated by NMR (via the relative integration of the 4-methylbenzyl alcohol end-group, which only accounts for linear polymers) highlight the limited control of the ROP of 1–3 under these organocatalytic conditions. This suggests the formation of cyclic species by backbiting or sensitivity to traces of chain-transfer agent like adventitious moisture (leading to smaller *M_n* than expected), as well as *trans* chain exchange phenomena, as sometimes seen in the ROP of cyclic carbonates, including in the polymerisation of sugar-based monomers.^{6b,c,i}

The FTIR spectra of all polymers were characterised by several strong absorption bands in the 1290–1020 cm⁻¹ region and none at 1757 cm⁻¹, indicative of C=S^{8d,14} but no C=O

bonds (Fig S53, S64 and S75†). Analysis by ¹³C{¹H} NMR also supported the absence of O–S rearrangement during polymerisation as no carbonyl resonance was observed. ¹³C{¹H} NMR was used to investigate further the microstructure of polymers (Fig. 1). For poly(3), three distinct thionocarbonate environments (differing by 0.3–1 ppm) were detected around 193.8 ppm (compared to 187.4 ppm for the monomer), and assigned to tail–tail (or head–head), head–tail, and head–head (or tail–tail) thionocarbonate linkages (Fig. S66†). Their 1 : 2 : 1 ratio suggests random cleavage of the thioacyl–oxygen bond at either side of the thionocarbonate carbonyl and subsequent nonselective propagation of the chain to yield regiorandom polymers. For poly(1), one distinct xanthate resonance was observed at 213.0 ppm (compared to the monomer signals at 208.1 and 207.8 for both anomers) (Fig. S44†). Thus, the thio-carbonyl region suggests a preference for regioregular opening of 1 (likely to liberate a more acidic primary thiol) and subsequent selective propagation of the chain to yield a poly (xanthate). In stark contrast, the polymer resulting from xanthate 2 displayed mainly two distinct thiocarbonyl resonances of similar intensities. Based on the literature,^{9b,d} the resonance at 222.5 ppm is assigned to a trithiocarbonate environment (C(S)₂), and the one at 193.1 ppm is assigned to a thionocarbonate (C(S)O₂) (Fig. S55†). This suggests an alternating opening of the monomer at either side of the xanthate thiocarbonyl, and subsequent selective propagation of the chain to yield regioregular polymers with alternating C(S)₂ and C(S)O₂ linkages. The origin of this regioregularity is so far unknown.

Analysis of the polymers by Matrix-assisted laser desorption/ionisation time-of-flight (MALDI-ToF) mass spectrometry was conducted to confirm the microstructures deduced by NMR, but proved extremely challenging. However, for poly(3) a major cyclic polymer series, with no end-groups and an integer number of sugar thionocarbonate repeat units (*m/z* ~232.25) was observed, likely due to backbiting of the polymer chain. A minor linear polymer series with 4-MePhCH₂O and OH end groups was also present (Fig. S71†). Poly(1) yielded poor data, although two different polymer series (cyclic and

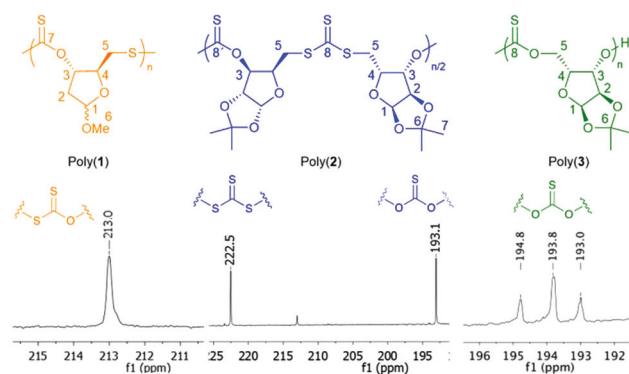


Fig. 1 Ring-opening polymerisation of monomers 1–3 and linkages observed by ¹³C NMR for the respective polymers.



linear) with sugar xanthate repeat units ($m/z \sim 206.27$) were detected, in agreement with NMR analysis (Fig. S49†). However, results from poly(2) (Fig. S60†) disagreed with the major alternating sequence inferred by NMR, revealing poly(xanthate) series (cyclic and linear) with repeating unit $m/z \sim 248.31$, which could come from the selective ionisation of side-products. Such poly(xanthate) traces can actually be detected by NMR in the ^{13}C NMR spectrum (trace signal at 213.0 ppm in Fig. 1).

Thermal properties of the polymers were evaluated by thermogravimetric analysis (TGA) and differential scanning calorimetry (DSC) of representative samples. Analysis of poly(1) with M_n 11 300 g mol^{-1} (Table 1, entry 3) showed two degradation steps with onsets at $\sim 120^\circ\text{C}$ and $\sim 182^\circ\text{C}$, with associated mass losses of 58 and 10% respectively (Fig. S50 and S51†). A glass transition temperature (T_g) of 55°C was measured (Fig. S52†). Although direct comparison with the analogous polycarbonate is not possible for synthetic reasons, poly(1) displays thermal properties which are very similar to the reported values for a related polycarbonate with a *cis*-fused motif ($T_g \sim 58^\circ\text{C}$ and T_d (onset) $\sim 125^\circ\text{C}$ for M_n 25 600 g mol^{-1}).^{6b} Xylose-derived polymers were slightly more thermally stable. Poly(2) with M_n 12 500 g mol^{-1} (Table 1, entry 6) featured two thermal degradation steps (Fig. S61 and S62†), with onsets at $\sim 77^\circ\text{C}$ and $\sim 135^\circ\text{C}$, corresponding to mass losses of 6% and 72%, respectively. TGA analysis of poly(3) with M_n 10 600 g mol^{-1} (Table 1, entry 14) showed three degradation steps, with onsets at $\sim 77^\circ\text{C}$ (10% mass loss), $\sim 161^\circ\text{C}$ then $\sim 298^\circ\text{C}$ (combined 79% mass loss) (Fig. S72 and S73†). Similar T_g values were measured for poly(2) ($\sim 46^\circ\text{C}$, Fig. S63†) and poly(3) ($\sim 48^\circ\text{C}$, Fig. S74†). These T_g values are significantly lower than the value for the analogous xylose carbonate, reported by Gross and coworkers (T_g 128°C), for which no thermal degradation was reported.⁶ⁱ

This decrease in glass transition temperatures when replacing O by S is consistent with the literature, in particular for aromatic polycarbonates.^{7i,15} For aliphatic polymers, data is rare, but poly(trimethylene monothiocarbonate) (PTMMTC) has been shown to have a T_g between -17 and -25°C ,^{7o} similar to the one reported for poly(trimethylene carbonate) (PTMC) of -25°C .^{6b} However, Darensbourg, Zhang and coworkers further noted that when S/O rearrangements occurred (yielding for example thionocarbonate linkages), T_g fell to -41°C . PTMMTC is also semi-crystalline (up to 71% crystallinity while PTMC is amorphous) and displays better thermal stability with onset of degradation occurring at 228.5°C vs. 197°C for PTMC. Another example by Endo and coworkers is the comparison done between a norbornene-substituted poly(trimethylene carbonate) and the analogous polythionocarbonate, for which the sulfur aliphatic polymer shows a decrease in T_g (82°C vs. 108°C) but an increase in the temperature of degradation onset ($T_{d10\%}$ 258°C vs. 207°C).¹⁶ A possible explanation for the decrease in T_g is the difference in the bond lengths of C–O (1.43 Å) and C–S (1.815 Å) as well as the van der Waals radii of O (1.52 Å) and S (1.85 Å) atoms, which increase the free volume in the polymer. Asymmetrical link-

ages have also been invoked to explain the enhanced intermolecular interactions between poly(thiocarbonate) chains, leading to crystallinity and higher thermal stability.

Conclusions

In summary, in our attempt to use CS_2 in the direct cyclothiocarbonation of the *trans* 1,3-diol motif of a ribofuranose, we discovered that cyclic xanthate structures could be accessed. We thus isolated the first two examples of 6-membered cyclic xanthate monomers, *trans* and *cis*-fused to sugar furanose rings, derived from natural sugars 2-deoxy-D-ribose and D-xylose, respectively. Polymers from these two monomers and from a xylose-derived thionocarbonate, also made using CS_2 , were obtained by ROP under mild reaction conditions, with organocatalyst TBD and 4-MeBnOH alcohol initiator. MALDI-ToF MS revealed both linear and cyclic topologies, in agreement with the limited control over the polymerisation observed. $^{13}\text{C}\{^1\text{H}\}$ NMR analysis suggests that no O–S rearrangement occurs during polymerisation, and that both the nature of the monomer and that of the sugar used influence the regioselectivity of ring-opening. While the deoxyribose xanthate monomer yields a poly(xanthate) species, the xylose cyclic xanthate produces a polymer with alternating trithiocarbonate and thionocarbonate linkages, and the xylose-derived poly(thionocarbonate) shows regiorandom linkages. Thermal analysis revealed lower glass transition temperatures compared to carbonate analogues and a low onset of thermal degradation. Building from this communication, current efforts are focusing on controlling the polymerisation of these novel sulfur-containing monomers and extending our methodology to other sugar feedstocks, in order to accurately investigate the effect of replacing O by S in the linkages of sugar-based polycarbonates, and gain a better understanding of the structure–property relationships of the resulting polymers. Because of their sugar backbone and the high refractive index and Abbe's number of sulphur analogues of polycarbonates,¹⁷ these materials could be used in optical and biomedical applications where degradability and biocompatibility is required.

Conflicts of interest

There are no conflicts to declare.

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Notes and references

- (a) G. L. Gregory, E. M. Lopez-Vidal and A. Buchard, *Chem. Commun.*, 2017, **53**, 2198–2217; (b) J. A. Galbis, M. d. G. García-Martín, M. V. de Paz and E. Galbis, *Chem. Rev.*, 2016, **116**, 1600–1636.
- (a) A. J. Varma, J. F. Kennedy and P. Galgali, *Carbohydr. Polym.*, 2004, **56**, 429–445; (b) V. Ladmiral, E. Melia and D. M. Haddleton, *Eur. Polym. J.*, 2004, **40**, 431–449; (c) R. Narain, D. Jhurry and G. Wulff, *Eur. Polym. J.*, 2002, **38**, 273–280; (d) M. Okada, *Prog. Polym. Sci.*, 2001, **26**, 67–104; (e) J. A. Galbis and M. d. G. García-Martín, in *Carbohydrates in Sustainable Development II*, ed. A. P. Rauter, P. Vogel and Y. Queneau, Springer Berlin, Heidelberg, 2010, pp. 147–176; (f) X. Feng, A. J. East, W. B. Hammond, Y. Zhang and M. Jaffe, *Polym. Adv. Technol.*, 2011, **22**, 139–150; (g) J. A. Galbis and M. d. G. García-Martín, in *Monomers, Polymers and Composites from Renewable Resources*, ed. A. Gandini, Elsevier, Amsterdam, 2008, pp. 89–114.
- T. Endo, in *Handbook of Ring-Opening Polymerization*, Wiley-VCH Verlag GmbH & Co. KGaA, 2009, pp. 53–63.
- (a) E. L. Dane and M. W. Grinstaff, *J. Am. Chem. Soc.*, 2012, **134**, 16255–16264; (b) E. L. Dane, S. L. Chin and M. W. Grinstaff, *ACS Macro Lett.*, 2013, **2**, 887–890; (c) C. Ghobril, B. Heinrich, E. L. Dane and M. W. Grinstaff, *ACS Macro Lett.*, 2014, **3**, 359–363; (d) S. E. Stidham, S. L. Chin, E. L. Dane and M. W. Grinstaff, *J. Am. Chem. Soc.*, 2014, **136**, 9544–9547; (e) S. L. Chin, Q. Lu, E. L. Dane, L. Dominguez, C. J. McKnight, J. E. Straub and M. W. Grinstaff, *J. Am. Chem. Soc.*, 2016, **138**, 6532–6540; (f) R. Xiao, E. L. Dane, J. Zeng, C. J. McKnight and M. W. Grinstaff, *J. Am. Chem. Soc.*, 2017, **139**, 14217–14223; (g) M. Bueno, J. A. Galbis, M. G. García-Martín, M. V. De Paz, F. Zamora and S. Muñoz-Guerra, *J. Polym. Sci., Part A: Polym. Chem.*, 1995, **33**, 299–305; (h) M. d. G. García-Martín, M. V. de Paz Báñez and J. A. Galbis, *J. Carbohydr. Chem.*, 2000, **19**, 805–815; (i) M. d. G. García-Martín, M. V. de Paz Báñez and J. A. G. Pérez, *Carbohydr. Res.*, 1993, **240**, 301–305.
- Y.-Y. T. Tsao and K. L. Wooley, *J. Am. Chem. Soc.*, 2017, **139**, 5467–5473.
- (a) G. L. Gregory, L. M. Jenisch, B. Charles, G. Kociok-Köhn and A. Buchard, *Macromolecules*, 2016, **49**, 7165–7169; (b) G. L. Gregory, G. Kociok-Köhn and A. Buchard, *Polym. Chem.*, 2017, **8**, 2093–2104; (c) G. L. Gregory, E. M. Hierons, G. Kociok-Köhn, R. I. Sharma and A. Buchard, *Polym. Chem.*, 2017, **8**, 1714–1721; (d) K. Tezuka, K. Koda, H. Katagiri and O. Haba, *Polym. Bull.*, 2015, **72**, 615–626; (e) K. Mikami, A. T. Lonacker, T. P. Gustafson, N. F. Zinnel, P.-J. Pai, D. H. Russell and K. L. Wooley, *J. Am. Chem. Soc.*, 2013, **135**, 6826–6829; (f) O. Haba, H. Tomizuka and T. Endo, *Macromolecules*, 2005, **38**, 3562–3563; (g) M. Azechi, K. Matsumoto and T. Endo, *J. Polym. Sci., Part A: Polym. Chem.*, 2013, **51**, 1651–1655; (h) R. Kumar, W. Gao and R. A. Gross, *Macromolecules*, 2002, **35**, 6835–6844; (i) Y. Shen, X. Chen and R. A. Gross, *Macromolecules*, 1999, **32**, 2799–2802.
- (a) A. Kultys, *Sulfur-Containing Polymers in Encyclopedia of Polymer Science and Technology*, John Wiley and Sons, Inc., 2010; (b) M. Luo, Y. Li, Y.-Y. Zhang and X.-H. Zhang, *Polymer*, 2016, **82**, 406–431; (c) E. Marianucci, C. Berti, F. Pilati, P. Manaresi, M. Guaita and O. Chiantore, *Polymer*, 1994, **35**, 1564–1566; (d) J. C. Seferis, in *Polymer handbook*, ed. J. Brandrup, E. H. Immergut and E. A. Grulke, Wiley, New York, 4th edn, 1999, pp. VI-571–VI-582; (e) R. K. Sathir and K. F. Schoch, *Chem. Mater.*, 1996, **8**, 1281–1286; (f) B. Ochiai and T. Endo, *Prog. Polym. Sci.*, 2005, **30**, 183–215; (g) W. Kuran, *Prog. Polym. Sci.*, 1998, **23**, 919–992; (h) G. Montaudo, C. Puglisi, C. Berti, E. Marianucci and F. Pilati, *J. Polym. Sci., Part A: Polym. Chem.*, 1989, **27**, 2277–2290; (i) M. F. Rey-Stolle, F. Fernández-Martín, L. H. Tagle and I. Hernández-Fuentes, *Polym. Int.*, 1998, **46**, 93–98; (j) H. R. Kricheldorf and G. Schwarz, *J. Macromol. Sci., Pure Appl. Chem.*, 2007, **44**, 625–649; (k) J.-L. Yang, H.-L. Wu, Y. Li, X.-H. Zhang and D. J. Darensbourg, *Angew. Chem., Int. Ed.*, 2017, **56**, 5774–5779; (l) C. G. Overberger and J. Weise, *J. Polym. Sci., Part B: Polym. Lett.*, 1964, **2**, 329–331; (m) T. J. Bannin and M. K. Kiesewetter, *Macromolecules*, 2015, **48**, 5481–5486; (n) C. G. Overberger and J. K. Weise, *J. Am. Chem. Soc.*, 1968, **90**, 3533–3537; (o) H.-L. Wu, J.-L. Yang, M. Luo, R.-Y. Wang, J.-T. Xu, B.-Y. Du, X.-H. Zhang and D. J. Darensbourg, *Macromolecules*, 2016, **49**, 8863–8868; (p) M. Luo, X.-H. Zhang and D. J. Darensbourg, *Acc. Chem. Res.*, 2016, **49**, 2209–2219.
- (a) M. Benazza, R. Kanso and G. Demailly, *Carbohydr. Res.*, 2010, **345**, 346–351; (b) Y. Tsuda, S. Noguchi and H. Niino, *Chem. Pharm. Bull.*, 2001, **49**, 1210–1213; (c) M. Adiyaman, S. P. Khanapure, S. W. Hwang and J. Rokach, *Tetrahedron Lett.*, 1995, **36**, 7367–7370; (d) D. Trimnell, W. N. Doane, C. R. Russell and C. E. Rist, *Carbohydr. Res.*, 1971, **17**, 319–326; (e) D. Trimnell, W. M. Doane and C. R. Russell, *Carbohydr. Res.*, 1972, **22**, 351–359.
- (a) X.-H. Zhang, F. Liu, X.-K. Sun, S. Chen, B.-Y. Du, G.-R. Qi and K. M. Wan, *Macromolecules*, 2008, **41**, 1587–1590; (b) D. J. Darensbourg, J. R. Andreatta, M. J. Jungman and J. H. Reibenspies, *Dalton Trans.*, 2009, 8891–8899; (c) D. J. Darensbourg, S. J. Wilson and A. D. Yeung, *Macromolecules*, 2013, **46**, 8102–8110; (d) J. Diebler, H. Komber, L. Häußler, A. Lederer and T. Werner, *Macromolecules*, 2016, **49**, 4723–4731; (e) M. Luo, X.-H. Zhang and D. J. Darensbourg, *Macromolecules*, 2015, **48**, 5526–5532.
- M. Luo, X.-H. Zhang and D. J. Darensbourg, *Polym. Chem.*, 2015, **6**, 6955–6958.
- G. L. Gregory, M. Ulmann and A. Buchard, *RSC Adv.*, 2015, **5**, 39404–39408.



- 12 (a) F. Nederberg, B. G. G. Lohmeijer, F. Leibfarth, R. C. Pratt, J. Choi, A. P. Dove, R. M. Waymouth and J. L. Hedrick, *Biomacromolecules*, 2007, **8**, 153–160; (b) M. K. Kiesewetter, E. J. Shin, J. L. Hedrick and R. M. Waymouth, *Macromolecules*, 2010, **43**, 2093–2107.
- 13 D. Pati, X. Feng, N. Hadjichristidis and Y. Gnanou, *Macromolecules*, 2017, **50**, 1362–1370.
- 14 (a) R. M. Silverstein, F. X. Webster, D. J. Kiemle and D. L. Bryce, in *Spectrometric Identification of Organic Compounds*, John Wiley and Sons, New York, 8th edn, 2014, ch. 2; (b) B. Tamami and A. R. Kiasar, *J. Chem. Res., Synop.*, 1998, 454–455.
- 15 M. Yazdani-Pedram, E. Soto, L. H. Tagle, F. R. Diaz, L. Gargallo and D. Radić, *Thermochim. Acta*, 1986, **105**, 149–160.
- 16 K. Kakimoto, N. Nemoto, F. Sanda and T. Endo, *Chem. Lett.*, 2002, **31**, 156–157.
- 17 M. Luo, X.-H. Zhang, B.-Y. Du, Q. Wang and Z.-Q. Fan, *Polym. Chem.*, 2015, **6**, 4978–4983.

