



Cite this: *Green Chem.*, 2022, **24**, 4174

Influence of storage temperature on the stability of heat treated phycocyanin- λ -carrageenan complexes in liquid formulations†

Stephan Buecker,^a Lutz Grossmann,^{a,b} Myriam Loeffler,^c Elena Leeb^d and Jochen Weiss^{id}*^a

The protein chromophore complex present in cyanobacteria like *Arthrospira platensis* can be utilized as a natural food colorant, but the appearance is prone to changes under environmental conditions that alter the protein-chromophore interactions. The study investigates the color stability of phycocyanin stabilized by λ -carrageenan during storage at 4 and 25 °C under different heating and pH conditions. Phycocyanin solutions and combinations with λ -carrageenan (1 : 4) were prepared at a pH of 2.5–6.0, with and without thermal treatment at 70 °C and 90 °C. Visual appearance, absorption spectra, zeta potential, and the diameter of the structures formed were observed over a period of 28 days. Considerable color losses were determined for pure phycocyanin at a pH of 2.5 to 4.5 when the samples were stored at 25 °C, which was improved by the formation of soluble complexes with λ -carrageenan in associative conditions. Interestingly, soluble complexes at a pH < pI were found to be more resistant at high treatment temperatures with a higher storage stability than co-soluble polymers at a pH > pI, but soluble structures formed at a pH of 2.5 had a significantly larger diameter (871 ± 185 nm) that increased in size during storage (2025 ± 1012 nm after 14 d at 25 °C) than at a pH of 3.0 (200 ± 5 nm) when heated to 90 °C. These results were related to changes in protein structure and mass transport phenomena and might be of high importance for low-viscous food formulations.

Received 1st March 2022,
Accepted 14th April 2022

DOI: 10.1039/d2gc00809b

rsc.li/greenchem

1 Introduction

The use of coloring foods in food formulations instead of synthetic colors is compelling. Their plant origin and exclusively physical processing make them not only a natural but also a renewable and vegan source that is very attractive to consumers.^{1–4} When the Southampton Study declared, in 2007, that six synthetic dyes may increase hyperactivity in children, a radical drop in their commercial use was noticed.⁵ Nonetheless, colorants are typically added to food formulations to make them more appealing to consumers, to account for natural color losses during production and differences in raw material

quality, or color fading over time. A convincing alternative to synthetic dyes is the use of food colorants that are derived from color-intensive food-grade raw materials including vegetables, fruits, or cyanobacteria. These foods are exclusively processed by physical processing methods and H₂O extraction to obtain a concentrated extract that is rich in the desired pigment.⁶ Their origin and minimal processing make them not only a natural but also a renewable and vegan source that is very attractive to consumers.^{1–4} However, these extracts are often less stable than their synthetic counterparts, which presents a challenge for producing stable foods with a long shelf life, thus minimizing food losses.

Up to now, phycocyanin (PC) present in eukaryote algae and cyanobacteria such as *Arthrospira platensis* (also known as *Spirulina*) is the only authorized food coloring in the European Union with a blue appearance, which is a major challenge to replace synthetic blue dyes in many food formulations.⁷ PC consists of the more blueish green-appearing allophycocyanin (aPC) and the royal blue-appearing C-phycocyanin (CPC). Both aPC and CPC are alike in their tertiary structure, but differ in amino acid sequence, number of chromophores (phycocyanobilins), and molecular weight.⁸ Each holds an α - and β -subunit with globular folding. In aPC, both subunits bind one chromo-

^aDepartment of Food Material Science, Institute of Food Science and Biotechnology, University of Hohenheim, Garbenstrasse 21, 70599 Stuttgart, Germany.

E-mail: j.weiss@uni-hohenheim.de; Tel: +49 711 459 24415

^bDepartment of Food Science, University of Massachusetts Amherst, Amherst, MA 01003, USA

^cDepartment of Microbial and Molecular Systems, Leuven Food Science and Nutrition Research Centre, KU Leuven - Ghent Technology Campus, Gebroeders de Smetstraat 1, 9000 Ghent, Belgium

^dGNT Europa GmbH, 52072 AachenKackertstrasse 22, Germany

† Electronic supplementary information (ESI) available. See DOI: <https://doi.org/10.1039/d2gc00809b>

phore, whereas in CPC, the β -subunit attaches two chromophores. The chromophores are covalently bound to the apoprotein by thioether bonds.^{8–10} In solution, PC is present as a complex mixture of monomers, trimers, and hexamers. Monomers are the predominant form at acidic pH values. They assemble into hexamers around their isoelectric point (pI) of a pH of 4.6.¹¹ A further increase of the pH leads to dissociation into trimers.⁸

A major drawback of utilizing chromophores that are stabilized by proteins is the sensitivity of protein structures to denaturation. This influences the blue appearance of PC-containing products.¹² However, many food formulations undergo thermal, pH, and ionic changes during production and storage. For example, a lot of beverages have a pH of 3.0, which is challenging in terms of PC storage stability due to an increased number of unstable PC monomers.^{8,13} Additionally, pasteurization takes place at <100 °C and is one of the most common treatments to preserve foods, but alters the structure of proteins.¹² It is already known that adverse temperatures and pH levels affect PC stability and, thus, color intensity.¹⁴

Previous studies have shown that PC can be stabilized by complexation with different carrageenans, whereas λ - and ι -carrageenan proved to efficiently stabilize PC against thermal- and acidic-degradation, such as precipitation and color loss.^{12,15,16} Carrageenans consist of repeating disaccharide units of 3-linked β -D-galactopyranose and 4-linked α -galactopyranose or 3,6-anhydro- α -galactopyranose, have a high molecular weight (200–800 kDa) and are commonly found in red algae (*Rhodophyta*). With three sulfate groups per disaccharide unit, λ -carrageenan carries a strong negative charge, which seemed to be beneficial for the complexation with PC as the complexes are primarily formed by electrostatic interactions.^{15,17,18} Additionally, hydrophobic interactions and hydrogen bonds affect the complex structure.¹⁸

While carrageenan was found to be promising to stabilize PC against heat treatments, far too little attention has been paid to the influence of storage time and storage temperature on these complexes. However, this is of significant importance, since color stability throughout the shelf life is of high relevance in food formulations. For this reason, this study aims to elucidate the effect of pH and temperature treatment on the color stability of PC and its mixtures with λ -carrageenan during 28 days of storage at different storage conditions.

2 Materials and methods

2.1 Materials

An *Arthrospira platensis* protein concentrate rich in PC (henceforth referred to as PC powder) obtained by spray drying was kindly supplied by GNT International B.V. (Mierlo, Netherlands) and was chosen to enhance the pertinence of this study for the food and beverage industry. The λ -carrageenan (λ C) powder was obtained from TIC Gums Inc. Both source materials were chosen to be comparable to pre-

vious studies. (White Marsh, USA). Hydrochloric acid (HCl), sodium hydroxide (NaOH), and sodium azide (NaN₃) were purchased from Carl Roth GmbH & Co. KG (Karlsruhe, Germany).

2.2 Methods

2.2.1 Solution preparation. For the preparation of the mixed/individual biopolymer solutions, stock solutions of 0.1% (w/w) PC powder and/or 0.15% (w/w) λ C was/were dissolved in double-distilled (DI) water and hydrated for 180 minutes. The solutions containing both biopolymers were formed by first dissolving λ C at 500 rounds per minute. Afterwards PC powder was added in a PC: λ C ratio of 1:4 while being stirred for another 5 minutes.¹⁸ To exclude turbidity effects of microbial growth, 0.05% (w/w) NaN₃ was added to the samples. Hereinafter, the 0.1% (w/w) PC powder solutions, which yield in a final PC concentration of 0.0369% (w/w), are referred to as 0.04% (w/w) PC solutions.

2.2.2 Formation of mixtures and electrostatic complexes. PC: λ C solutions with an initial pH of 6.7 were adjusted to a pH of 6.0, 5.0, 4.5, 4.0, 3.5, 3.0, and 2.5 (± 0.02) using either HCl or NaOH (0.1–3.0 M) and solutions were kept overnight at 4 °C. Before the heat treatment, the pH values of all samples were checked and readjusted if necessary. The pH maintained stable throughout the observed storage period.

2.2.3 Heat treatment. Heat treatments were carried out in a forced convective water bath (Lauda Dr R. Wobser GmbH & Co. KG, Germany). 45 g of each liquid sample was placed in 50 mL conical PP tubes (VWR International GmbH, Pennsylvania, USA), and the sample solutions were heated to the appropriate core temperatures. To reach a core temperature of 70 and 90 °C, 15 and 30 minutes were required, respectively. After a holding time of 1 min at the respective core temperature, samples were immediately cooled down to 25 °C using a second water bath at 25 °C.

2.2.4 Storage tests. The heat-treated samples were stored at either 4 °C (common refrigerator temperature) or 25 °C in a climate cabinet (HCP50, Memmert GmbH & Co. KG, Schwabach, Germany) over a period of 28 days.

2.3 Analyses

In order to investigate the influence of storage time and different storage temperatures on color shifts and intensity, pure PC and PC: λ C (1:4) mixtures were analyzed during the storage period on day 0, 3, 6, 14, and 28.

2.3.1 Protein & PC content. The protein content of the samples was determined according to the method provided by Dumas using a Dumatherm device (C. Gerhardt GmbH & Co. KG, Koenigswinter, Germany) and a nitrogen-to-protein conversion factor of 6.27, which was suggested for *Arthrospira platensis*.¹⁹ Afterward, the total color active PC concentration was calculated according to Yoshikawa Belay *et al.* (2008) using the following equations:²⁰

$$C_{\text{C-phycocyanin}} = 0.162A_{620} - 0.098A_{650} \text{ (g L}^{-1}\text{)} \quad (1)$$

$$C_{\text{allophycocyanin}} = 0.180A_{650} - 0.042A_{620} \text{ (g L}^{-1}\text{)} \quad (2)$$

with A_{620} and A_{650} corresponding to the absorption of aPC and CPC at 620 and 650 nm.

2.3.2 Zeta potential & particle size. To investigate the microelectrophoretic behavior and size of the complexes formed, measurements were performed using a Zetasizer Nano ZS (Malvern Instruments, Worcestershire, UK). Here, depending on the velocity and the direction of the moving complexes in an electric field, the net charge of the complexes can be determined using the Smoluchowski approximation. The particle size was determined by dynamic light scattering at a laser wavelength of 633 nm and the backscattered light was detected at a measurement angle of 173° . No interferences with the samples were observed due to the absorption shift of PC to 650–670 nm upon complexation and below pH 5.0. The results were consistent and showed good measurement signals, which agreed with previous findings.¹⁸ During the zeta potential and size determination, the temperature was kept constant at 25 °C and the measurement was carried out in triplicate.

2.3.3 Transmittance and absorption. Transmittance and absorption of the samples were detected using an Ultrospec 2100 *pro* (Biochrom GmbH, Berlin, Germany) spectrophotometer. Samples were transferred to semimicro UV cuvettes (BRAND GmbH & Co. KG, Wertheim, Germany) and the device was blanked with DI water. The transmittance was measured at 780 nm and the absorption spectrum was recorded from 240 nm to 750 nm, respectively. A turbidity baseline correction was performed by measuring the absorption at 750 nm to account for scattering and absorption of colloidal particles. Results are reported as relative values to unheated 0.04% (w/w) PC solutions at a pH of 6.0, which show the highest color intensity.

2.3.4 Visual appearance. Pictures of the samples were taken inside a photo box under steady light conditions. 2.5 mL of each sample were transferred into macro cuvettes (BRAND GmbH & Co. KG, Wertheim, Germany), which were always placed on the same fixed positions. The pictures were taken after a period of 30 min to account for potential PC precipitation effects.

Average colors of the solutions were extracted by GIMP version 2.10.24 (available at: <https://www.gimp.org/>).

2.4 Modeling and calculations

2.4.1 Protein modeling. The structure of a CPC hexamer stack was adapted from 'RCSB Protein Data bank' (available at: <https://www.rcsb.org/structure/1HA7>). Amino-acid sequences of group C–X were deleted in the text file. Further modification of the protein structure was carried out by the freeware Swiss-PdbViewer version 4.1.0 (Swiss Institute of Bioinformatics, Lausanne, Switzerland) (available at: https://spdbv.vital-it.ch/download_prerelease.html). The hydrophobic patches (green), as well as the positive (blue) and negative (red) surface charges, were visualized for the α -subunit and the α - β -heterodimer (monomer).

2.4.2 Calculation of the theoretical pI of PC. The webtool 'Compute pI/Mw' by the Swiss Institute of Bioinformatics (SIB)

(available at: https://web.expasy.org/compute_pi/) was used for the calculation of the theoretical pI of PC, which is based on the combined theoretical pIs of CPC and aPC, whereas CPC was considered to contribute 85% and aPC 15.15% to the pI of PC. The ratio of aPC and CPC was calculated according to eqn (1) and (2).

2.5 Statistics

Each experiment was conducted twice and all measurements were repeated at least two times. Means and standard deviations were determined with Excel (Microsoft Inc., Redmond, USA). SPSS Statistics 27 (IBM Inc., Armonk, USA) was used to statistically evaluate significant differences between results gained from PC concentration calculations and size measurements using a one-way ANOVA followed by a *post-hoc* Tukey test ($\alpha = 0.05$).

3 Results and discussion

In a previous study, it was shown that pure PC was unstable around its pI due to decreased electrostatic repulsion among PC and that heat treatment has a profound impact on the color stability of pure PC.^{14,18} Initially, the protein and PC content in the powder were analyzed, and based on these results, the storage stability was investigated.

3.1 Protein content

The protein content of the PC powder was determined as $89.83 \pm 0.25\%$, which is considerably higher than that of untreated biomass with a dry matter protein content of typically 46–63%.²¹ This is attributed to the production of PC powder, which involves cell and cell wall disruption, separation of insoluble compounds, and filtration steps, such as ion-exchange filtration.^{22,23,23} Additionally, the protein concentration of cyanobacteria can be optimized by modifying the growth conditions, *e.g.*, luminosity, temperature, stirring speed, dissolved solids, pH, water quality, and macro as well as micronutrients.²⁴ According to the manufacturer, the protein content was 88.75% (Appendix Table 4) and was determined *via* the Kjeldahl method. This is about 1.2% less than our result, but corresponds to the protein values of the Dumas method, which are usually up to 1.4% higher.²⁵

According to the calculations from eqn (1) and (2), the amount of color active PC was 36.9%. Thus, around 41% of the protein in PC powder can be classified as color active PC. The CPC concentration was determined to be 31.3% (w/w) and the aPC content 5.6% (w/w). This is in alignment with the literature, which proposed that aPC is a minor component of *Arthrospira platensis*.²⁶

3.2 Color fading of pure PC solutions

The color stability of PC-containing solutions during storage is a key quality parameter that is important to ensure a consistent color intensity and so minimize food waste. Therefore, the color stability of unheated and heated (70/90 °C) pure PC and

mixed biopolymer solutions (PC : λ C; 1 : 4) was investigated at a pH of 2.5–6.0 over a period of 28 days at 4 °C and 25 °C. These two temperatures were selected to reflect typical storage

conditions at room temperature or under refrigeration conditions. Measurements were conducted on days 0, 3, 6, 14, and 28, while only stable (precipitated solutions were excluded) solutions were considered in the analysis.

Fig. 1 shows the unheated and heat-treated 0.04% (w/w) PC solutions at day 0 and selected samples (pH 2.5, 3.0, and 6.0) after 28 days of storage at 4 and 25 °C. These pH values were chosen because solutions at pH values of 3.5, 4.0, 4.5 precipitated. Further, the pH values of 2.5, as well as 3.0, reflect the pH values of most beverage formulations. A pH of 6.0 was selected as $\text{pH} > \text{pI}$.¹³

PC was found to precipitate at around a pH of 4.0 on day 0, which can be related to the pI of PC ($\text{pH} \sim 4.6$).^{8,11} The apparent pIs of the solutions were calculated from the zeta potential measurements (Appendix Table 3) to be 4.05, 4.28, and 4.37 for unheated solutions, and solutions heated to 70 or 90 °C, respectively. The change of pI might be attributed to protein unfolding upon thermal denaturation.¹²

Unheated PC solutions with a pH of 2.5, 3.0, 5.0, or 6.0 (pH 4 data not shown) were found to be visually stable against precipitation for the entire 28 days of storage, independent of the storage temperature used, whereas precipitates could also be detected in heated PC solutions having a pH of 5.0 (Fig. 2). This could be related to the aforementioned pI shift at higher temperatures leading to a reduced electrostatic repulsion of PC

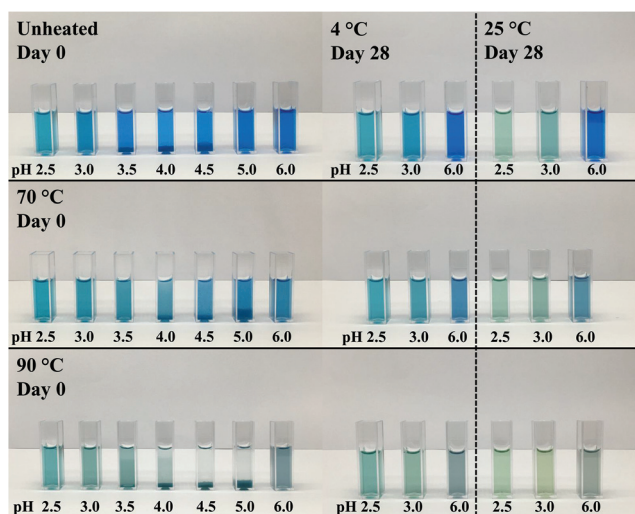


Fig. 1 Visual color appearance of 0.04% (w/w) PC solutions unheated and subjected to thermal treatments at 70 and 90 °C at pH 2.5–6.0 at day 0 and after 28 days of storage at 4 and 25 °C.

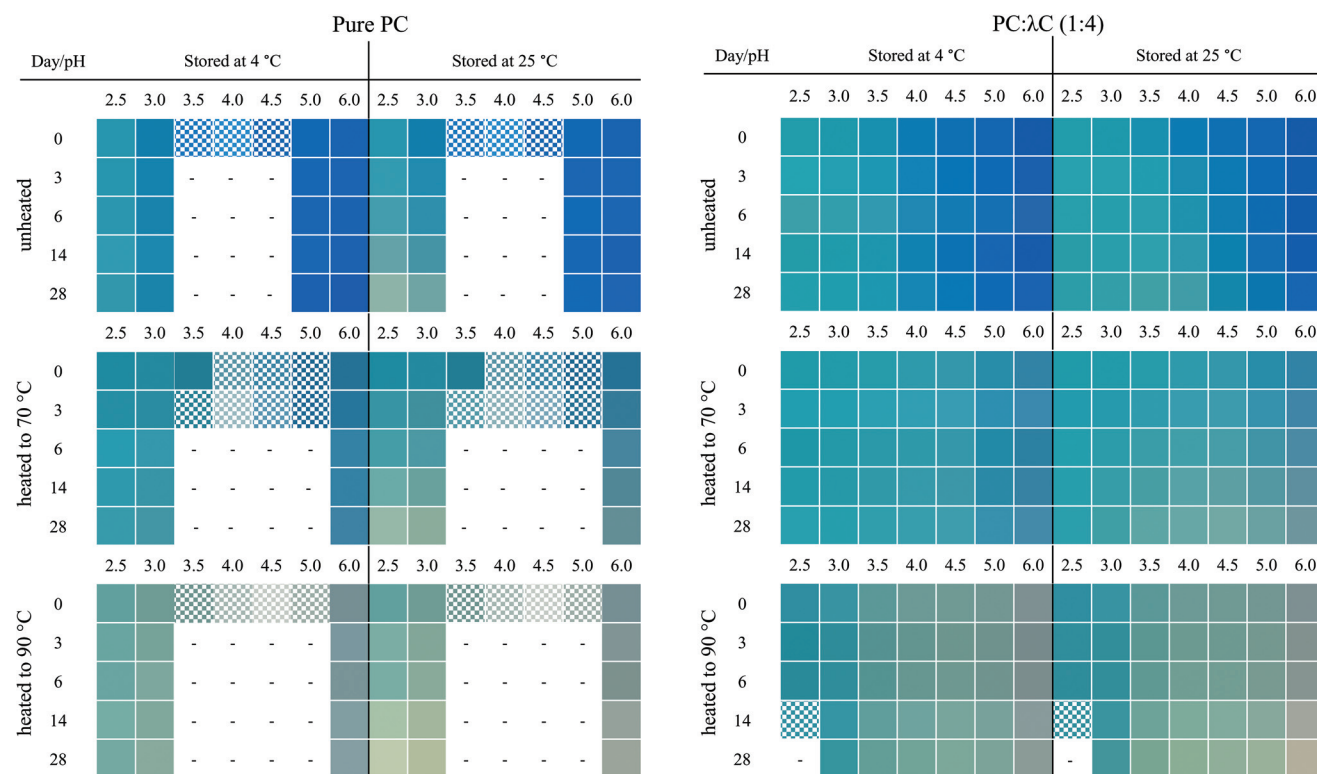


Fig. 2 Colors of 0.04% (w/w) PC solutions and PC : λ C (1 : 4) solutions at day 0, 3, 6, 14 and 28 over the pH range of 2.5–6.0. Solutions were prepared unheated, heated to 70 and 90 °C and stored at either 4 or 25 °C. Chequered fields illustrate the supernatant color of precipitated samples. The samples in empty fields were excluded because of precipitation. Average color of the samples was extracted from pictures by GIMP version 2.10.24.

at pH 5.0. Small changes in the protein structure by thermal treatments were already reported to cause a significant color loss which is also in alignment with our findings.^{10,14}

In contrast to the stability of the solution to precipitation, differences in color stability were found depending on the storage temperatures used. All color losses at 4 and 25 °C were discussed in turn. The color of PC solutions that were not heat treated or heated to a core temperature of 70 °C was found to be visually stable when stored at 4 °C over the course of 28 days (Fig. 1). Minor color losses at 4 °C were reported by Kannaujiya & Sinha (2016).²⁷ Solutions heated to 90 °C, with subsequent storage at 4 °C, appeared to undergo a color shift from blue to a dark grey at a pH of 6.0 and to greenish turquoise at a pH of 2.5 and 3.0, respectively. A green color can be related to a further dissociation of PC into monomers over time.^{8,27} Recently it was shown that the planarity of the chromophore significantly impacts the color appearance.²⁸ The grey color was described as color fading and might be related to aggregation of PC particles.⁶

The color shift and fading were more pronounced at 25 °C storage temperature (Fig. 1). For instance, at a pH of 2.5 and 3.0, the color shift towards a greenish turquoise was observed for both the unheated PC solutions and the solutions heated to 70 °C. A loss of color intensity could be noticed for almost all solutions stored at 25 °C for 28 days. These observations can be related to the extreme temperature sensitivity of PC.²⁷ At a pH of 6.0, the color of the unheated solutions remained stable at 25 °C, with only small visual changes. This could be due to PC stacks being organized as hexamers, the predominant PC structure at a pH of 6.0, which was reported to be more stable than monomeric PC.¹⁴

3.2.1 PC content. From the absorption measurements, the a color-active PC amount was calculated according to

Yoshikawa, Belay (2008).²⁰ The PC amounts of the 0.04% (w/w) PC solutions are presented in Table 1.

A pronounced PC degradation was facilitated by heating the solutions to either 70 or 90 °C. At a pH of 3.0, the initial PC concentration of unheated solutions and solutions heated to 70 °C was significantly lower than at a pH of 6.0. A possible explanation for this might be that, for acidic pH values, PC is primarily present as a monomer that has a lower absorption than PC hexamers.⁸ Furthermore, PC was reported to be more temperature resistant when assembled as hexamer.^{14,29} However, in PC solutions heated to 90 °C, the initial PC concentration on day 0 was equal at a pH of 3.0 and 6.0, but the overall PC concentration was lower compared to untreated solutions and solutions heated to 70 °C. This can be attributed to the 6th order degradation kinetics that phycocyanin follows.³⁰ Thus, denaturation of highly concentrated PC occurs much faster. Since the initial concentration of the PC solution at pH 3.0 was lower, both values converge with time.

There was a tendency that PC degradation at the same storage temperature was stronger at pH 3.0 than at pH 6.0, which could be due to increased stability of PC hexamers.¹⁴ This was especially prevalent at 25 °C storage temperature. Solutions stored at 4 °C having a pH of 6.0 showed a higher PC stability, while the strongest decrease in PC concentration was observed for untreated solutions prepared at a pH of 3.0. This is in alignment with the findings of Kannaujiya & Sinha (2016) and can be related to the temperature sensitivity of the PC protein structure.²⁷ Unheated solutions exhibited the greatest PC color loss over time (Table 1). These solutions had the highest initial concentration, whereas the lowest PC concentration was observed in solutions heated to 90 °C. These relationships may partly be explained by the fact that PC denaturation slows down significantly at low concentrations.³⁰

Table 1 Total PC concentrations (mg mL⁻¹) and relative PC concentrations with respect to unheated pH 6.0 samples of 0.04% (w/w) PC solutions at a pH of 3.0 and 6.0. The solutions were prepared unheated and heated to 70 or 90 °C and stored for a period of 28 days at either 4 or 25 °C

Heat treatment	Day	<i>c</i> _{PC} in mg mL ⁻¹ at pH 3		<i>c</i> _{PC} in mg mL ⁻¹ at pH 6	
		4 °C	25 °C	4 °C	25 °C
None	0	0.24 ± 0.01 ^{AA} (63.2%)		0.38 ± 0.01 ^{AB} (100%)	
	3	0.22 ± 0.01 ^b (57.9%)	0.19 ± 0.01 ^b (50.0%)	0.38 ± 0.01 ^{ab} (100%)	0.38 ± 0.01 ^a (100%)
	6	0.21 ± 0.01 ^b (55.3%)	0.17 ± 0.01 ^c (44.7%)	0.38 ± 0.01 ^{ab} (100%)	0.37 ± 0.01 ^b (97.4%)
	14	0.20 ± 0.01 ^c (52.6%)	0.13 ± 0.01 ^d (34.2%)	0.37 ± 0.01 ^{ab} (97.4%)	0.36 ± 0.01 ^c (94.7%)
	28	0.19 ± 0.01 ^c (50.0%)	0.09 ± 0.01 ^e (23.7%)	0.37 ± 0.01 ^b (97.4%)	0.35 ± 0.01 ^d (92.1%)
70 °C	0	0.14 ± 0.01 ^{AA} (36.8%)		0.15 ± 0.01 ^{AB} (39.5%)	
	3	0.14 ± 0.01 ^a (36.8%)	0.13 ± 0.01 ^b (34.2%)	0.15 ± 0.01 ^a (39.5%)	0.15 ± 0.02 ^a (39.5%)
	6	0.14 ± 0.01 ^{ab} (36.8%)	0.11 ± 0.01 ^c (28.9%)	0.15 ± 0.01 ^a (39.5%)	0.14 ± 0.02 ^a (36.8%)
	14	0.13 ± 0.01 ^b (34.2%)	0.09 ± 0.01 ^d (23.7%)	0.15 ± 0.01 ^{ab} (39.5%)	0.13 ± 0.02 ^a (34.2%)
	28	0.13 ± 0.01 ^c (34.2%)	0.07 ± 0.01 ^e (18.4%)	0.14 ± 0.01 ^b (36.8%)	0.11 ± 0.02 ^a (28.9%)
90 °C	0	0.07 ± 0.01 ^{AA} (18.4%)		0.07 ± 0.02 ^{AA} (18.4%)	
	3	0.07 ± 0.01 ^a (18.4%)	0.06 ± 0.01 ^b (15.8%)	0.07 ± 0.01 ^a (18.4%)	0.07 ± 0.01 ^a (18.4%)
	6	0.07 ± 0.01 ^a (18.4%)	0.06 ± 0.01 ^c (15.8%)	0.07 ± 0.01 ^a (18.4%)	0.07 ± 0.01 ^a (18.4%)
	14	0.06 ± 0.01 ^a (15.8%)	0.04 ± 0.01 ^d (10.5%)	0.07 ± 0.01 ^a (18.4%)	0.06 ± 0.01 ^a (15.8%)
	28	0.06 ± 0.01 ^a (15.8%)	0.03 ± 0.01 ^e (7.9%)	0.07 ± 0.01 ^a (18.4%)	0.05 ± 0.01 ^a (13.2%)

Indices with small letters indicate significant differences among solutions prepared with the same heat treatment in one column ($\alpha < 0.05$). Indices with capital letters indicate a difference between a pH of 3.0 and 6.0 within the same row determined by *t*-test ($\alpha < 0.05$).

3.3 Color fading of PC : λ C complex solutions

As pointed out in the previous chapter, pure PC solutions precipitated at a pH of 3.5–5.0 and were unstable against thermal treatments. Further, a color shift from blue, at a pH of 6.0, to turquoise, at a pH of 2.5, was observed. It was related to a change in the PC monomer, trimer, and hexamer concentration and an increasing chromophore planarity.²⁸ Together, the results showed that PC was less stable under acidic pH values or high storage temperatures (25 °C). This study aimed to improve the storage stability of PC by complexation with λ C. In the upcoming section, the present findings are compared to the appearance and absorption measurements of PC : λ C (1 : 4) solutions. The PC concentration in the complexes was identical to the pure PC solutions and all measurements were performed equally.

3.3.1 Visual appearance. To analyze the stability of PC : λ C mixtures, the solutions were again visually analyzed. Fig. 2 illustrates the average colors of the pictures from 0.04% (w/w) PC solutions and PC : λ C (1 : 4) solutions unheated and heated (70 or 90 °C) on day 0, 3, 6, 14, 28 stored at either 4 °C or 25 °C.

First, the effects of pH were evaluated. All biopolymer complex solutions were stable against precipitation on day 0 (Fig. 2 and Appendix Fig. 7), which was not the case for PC solutions, especially not at a pH of 4 (close to the pI) (Fig. 1). Moreover, all solutions, except for solutions prepared at a pH of 2.5 and heated to 90 °C, were stable against precipitation over the storage period of 28 days. The improvement against pH-dependent precipitation around the isoelectric point can be related to an increase in bulk viscosity, steric interactions among polysaccharide residues, and electrostatic repulsion, which was described for carrageenan-whey protein complexes.³¹

The effects of storage on color were described below. Unheated solutions showed almost no color loss over time, independent of the pH used. The unheated PC : λ C solutions stored at a pH of 4.0 showed a shift towards turquoise when stored at 25 °C, while the same solutions stored at 4 °C had a more bluish color after 28 days of storage. This indicates an altering protein structure with a simultaneous modification of the phycocyanobilin structure which was accelerated by the increased storage temperature.³² Structural changes could be due to hydrophobic interactions between PC proteins that evolve over time.³³

Next, the heating effects were specified. The color loss caused by thermal denaturation appeared to be stronger at higher pH values, whereas an intense turquoise color was maintained at a pH of 2.5 and 3.0, especially when the solutions were treated at 90 °C. A possible explanation for this might be that PC is stabilized by λ C at acidic pH values below the pI of ~pH 4.6 by electrostatic complexation between the anionic λ C and the positively charged PC, as previously reported. These interactions do not occur, or to a far lesser extent, at pH values above the pI, as for solutions prepared at a pH of 6.

From here on, the various effects of storage time and temperature, heat treatment and pH were considered in combi-

nation. At a pH of 2.5, PC : λ C solutions that were heated to 90 °C precipitated (Appendix Fig. 8), which could be due to the formation of large and dense complexes. These structures are fostered by the aggregation of PC by hydrophobic interactions and simultaneous complexation of PC and λ C by electrostatic interactions.^{12,33} However, the sedimentation occurred slowly and is hence not visible in Fig. 2.

Over the observed storage time, a temperature of 4 °C had only a minor influence on the appearance of those solutions that have been heated to 70 °C and adjusted to a pH of 2.5–4.5. The intensity of the blue color at a pH of 5.0–6.0 was slightly less compared to the unheated solutions. As suggested previously, this is due to electrostatic interactions at lower pH values. At a pH of 4.5, which is above the pI, λ C might attach to some positively charged patches on the PC surface.^{18,34}

However, increasing the storage temperature from 4 to 25 °C had a remarkable effect on the color appearance and intensity of the heated solutions. Mixed biopolymer solutions heated to 70 °C showed a color shift from turquoise towards a greyish green at a pH of 3.5–4.5, and to a light grey at a pH of 5.0 to 6.0. The shift towards green can be related to PC dissociation causing a color shift from 620 to 346–360 nm, while the grey color might be associated with protein aggregation causing diffuse light scattering.^{6,8} The effects were even more pronounced for PC : λ C solutions that have been heated to 90 °C and could already be observed under cold storage conditions. One explanation could be that, due to the strong thermal treatment, hydrophobic patches which were previously hidden inside the PC structure became exposed and resulted in PC aggregation over time.^{12,33} However, independent of the storage time and temperature, all the mixed biopolymer solutions were found to maintain a turquoise color at a pH 2.5 of and 3.0.

3.3.2 Absorption. The color stability and appearance are directly related to the quaternary structure of PC. CPC forms a trimer at a pH of 6.0 ($A_{\max} \sim 620$ nm), a hexamer stack at a pH of 4.0 ($A_{\max} \sim 620$ nm), and a monomer at a pH of 3.0 ($A_{\max} \sim 614$ nm), whereas aPC forms trimers ($A_{\max} \sim 650$ nm) over a broad range of pHs.^{29,32} Monomers were reported to have a lower absorption maximum and to be less stable than trimers and hexamers.^{8,14,29,35} Thus, aPC appears more intense at acidic pH values.

The absorption of PC : λ C solutions (1 : 4) was measured to quantify the color shift and reveal changes in the spectrum with pH change, heat treatment, as well as storage time and temperature. The absorption of solutions is directly linked to the visual appearance of the solutions and is subsequently presented for unheated (Fig. 3) and heated (Fig. 4) solutions at a pH of 3.0 and a pH of 6.0 that have been stored for 0 and 28 days at either 4 °C (A) or 25 °C (B), respectively. Solutions with a pH of 3.0 show a high stability of PC : λ C solutions against precipitation and color degradation (see above) even at high treatment temperatures. A pH of 6.0 served as a standard reference for solutions with a pH > pI. The latter solutions showed poorer color stabilization.

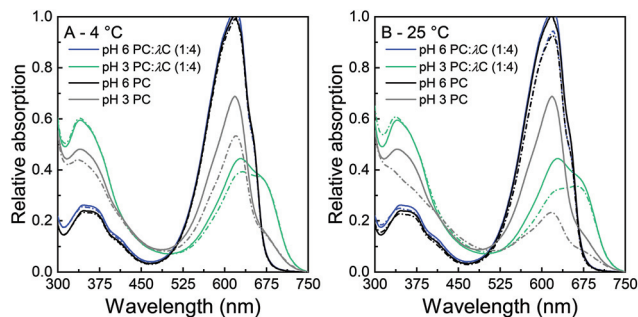


Fig. 3 Relative absorption of unheated PC:λC (1:4) solutions and 0.04% (w/w) PC solutions having a pH of 3.0 or 6.0 at 300–750 nm. Solutions were stored for 28 days at A: 4 °C or B: 25 °C. Solid lines illustrate samples on day 0. Dotted lines illustrate samples on day 28.

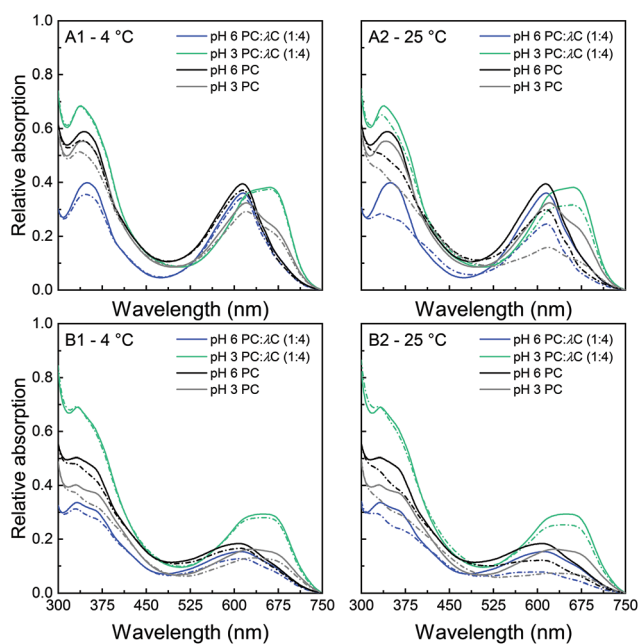


Fig. 4 Relative absorption of PC:λC (1:4) solutions at 300–750 nm and 0.04% (w/w) PC solutions having a pH of 3.0 or 6.0 at 300–750 nm. A: Solutions heated to 70 °C and B: solutions heated to 90 °C. Solutions were stored for 28 days at A1/B1: 4 °C or A2/B2: 25 °C, respectively. Solid lines illustrate samples on day 0. Dotted lines illustrate samples on day 28.

Unheated solutions (Fig. 3) had a lower absorption at a pH of 3.0 as compared to a pH of 6.0 which can be attributed to PC hexamers dissociating to monomers at low pH values.⁸ During storage, solutions that were stored at 25 °C had a greater decrease in absorption intensity compared to those stored at 4 °C, but overall losses were smaller compared to pure unheated PC solutions. Similar to the prevention of the pI-dependent formation of PC aggregates without the presence of λC, this effect might be related to an increased viscosity, steric interactions among polysaccharide residues, and electrostatic repulsion.^{18,31}

The second major difference observed in the absorption of PC:λC solutions is the peak at 660 nm. Individual PC solutions had a maximal absorption at 620 nm while decreasing the pH in the biopolymer mixture led to an increasing shift towards the red region. It is well known that anionic surfactants and urea can cause protein dissociation.⁹ In addition the separation of the PC subunits by urea was reported to cause an absorption shift from 620 to 662 nm.^{9,36} Thus, the previous results are likely to be related to a rearrangement or dissociation of the α- and β-subunits due to the complexation with λC coming with planarity changes of the phycocyanobilin.¹⁸

Three findings from the absorption measurements are of particular interest. First, the pH 3.0 PC:λC solutions had a higher absorption at 620 nm as compared to solutions prepared at a pH of 6.0. Second, A_{620} decreased significantly over time at 4 °C for PC:λC solutions adjusted to a pH of 6.0 and heated to 90 °C. At a 25 °C storage temperature, all PC:λC solutions having a pH of 6.0 significantly decreased in A_{620} over time, whereas PC:λC solutions at a pH of 3.0, heated to 90 °C, remained stable over the period of 28 days. Third, PC:λC solutions adjusted to a pH of 3.0 were found to be more heat resistant than solutions adjusted to a pH of 6.0. The latter finding is contrary to the results we found for pure heated PC solutions.

The results pinpoint the stability improvement against storage-related color fading in acidic pH values due to electrostatic interactions of λC and PC, especially after thermal treatments (Fig. 4). Thus, the color might additionally be stabilized by hydrophobic interactions which build up during the heat treatment.^{12,37}

3.3.3 Complex size. In the previous sections, we described enhanced color stability of PC by electrostatic complexation with λC. A stable turquoise color was initially determined for solutions at a pH of 3.0 and 2.5. However, after 14 days of storage, the latter solutions that were heat-treated at 90 °C precipitated (Fig. 2 and Appendix Fig. 8). To get a better understanding of the formed complexes (unheated/heated) and the observed changes at different storage temperatures (4 and 25 °C) over the course of 28 days, a DLS size analysis was performed and the results summarized in Table 2.

On day 0, the unheated biopolymer mixtures had a similar diameter of about 900 nm at both pH values. With increasing storage time, the complex diameter decreased. The electrostatic attraction of both biopolymers might slowly cause association of the biopolymers, resulting in an increase in density. According to Veis, Aranyi (1960) the structural rearrangement could be driven by an electrostatic entropy gain caused by detached counter ions.³⁸

The rearrangement of the complexes was accelerated by a higher storage temperature, which could be due to an increased Brownian motion and a decreased solution viscosity at higher temperatures, both influencing the movement of biopolymers in solutions. Also, hydrophobic interactions might develop at higher temperatures.³³ Moreover, unheated PC:λC complexes at a pH of 2.5 were found to be smaller than the

respective solutions at a pH of 3.0, which could be a result of an increased electrostatic attraction at lower pH values. This is in agreement with the transmittance measurements. At acidic pH values, the transmittance of complex solutions decreased, which was shown to be a result of strong protein-polysaccharide associations³⁹ (Appendix Fig. 8).

Heating the complexes to 70 °C led to smaller complexes compared to the unheated mixtures, which could either be a result of an accelerated chemical reaction between the biopolymers or hydrophobic interactions among PC due to the

thermal treatment favoring association interactions.^{12,40} Further, positively charged amino acid residues could be exposed due to structural rearrangements which promote electrostatic interactions.⁴¹ The complexes were again similar in diameter at a pH of 2.5 and 3.0 when heated to 70 °C on day 0. With prolonged storage, it was evident that complexes formed at a pH of 2.5 possessed smaller diameters than at a pH of 3.0. Independent of the pH, a storage temperature of 25 °C led again to smaller complexes as compared to cold storage. The rearrangement might be related to carrageenans' backbone flexibility and increased hydrophobic interactions at higher temperatures.^{31,37,42}

Heating the solutions to 90 °C during preparation caused an entirely different effect. At a pH of 2.5, the complexes had a significantly larger diameter than at a pH of 3.0 and the solutions became very inhomogeneous in size, which might be related to aggregation. In principle, secondary aggregation of protein-carbohydrate complexes is reduced by steric interactions and electrostatic repulsion. However, the PC:λC solutions showed a significantly decreased magnitude in zeta potential from a pH of 6.0 to 2.5, when the solutions were heated to 90 °C (Appendix Table 3). This could either be related to charge neutralization or dissociation of λC which has a pK_a of around 2.0.⁴³ Thus, complex-complex association might be a result of decreased electrostatic repulsion. Further, the increasing aggregation of PC might be related to hydrophobic interactions. As the solutions precipitated after 14 days, an ongoing rearrangement and aggregation of the complexes over time led to dense and big particles that tend to precipitate. Zeeb *et al.* (2018) reported that biopolymer complexes of whey protein isolate and pectin showed an Ostwald ripening-like dependence on the size after thermal treatments due to increasing hydrophobic and decreasing electrostatic interactions.⁴⁴ To the contrary, stable solutions could be produced at a pH of 3.0, which might be linked to the higher magnitude of the zeta potential and thereby stronger electrostatic repulsion (Appendix Table 3).

Table 2 Z-Average diameter of PC:λC (1:4) complex solutions unheated and heated to 70 or 90 °C at pH 2.5 and 3.0 and stored over 28 days at either 4 or 25 °C. PC:λC (1:4) complex solutions contained 0.15% (w/w) λC and 0.04% (w/w) PC

Heat treatment	Day	<i>d</i> in nm at pH 2.5		<i>d</i> in nm at pH 3.0	
		4 °C	25 °C	4 °C	25 °C
None	0	890 ± 155 ^{aA}		907 ± 14 ^{aA}	
	3	868 ± 126 ^a	526 ± 31 ^b	901 ± 9 ^a	698 ± 80 ^b
	6	770 ± 82 ^a	476 ± 31 ^b	835 ± 23 ^{ab}	562 ± 41 ^{bc}
	14	715 ± 60 ^a	360 ± 17 ^b	780 ± 52 ^{bc}	420 ± 25 ^{cd}
	28	547 ± 12 ^a	316 ± 7 ^b	694 ± 11 ^c	354 ± 10 ^d
70 °C	0	484 ± 57 ^{aA}		646 ± 59 ^{aA}	
	3	464 ± 62 ^a	416 ± 29 ^{ab}	614 ± 79 ^a	554 ± 10 ^{ab}
	6	436 ± 92 ^a	386 ± 5 ^{abc}	545 ± 90 ^a	509 ± 50 ^{ab}
	14	448 ± 57 ^a	336 ± 20 ^{bc}	603 ± 62 ^a	410 ± 3 ^{bc}
	28	424 ± 26 ^a	291 ± 4 ^c	532 ± 8 ^a	360 ± 29 ^c
90 °C	0	871 ± 185 ^{aA}		200 ± 5 ^{aB}	
	3	1355 ± 960 ^a	1661 ± 938 ^a	202 ± 10 ^a	202 ± 11 ^a
	6	1305 ± 950 ^a	1734 ± 904 ^a	200 ± 9 ^a	202 ± 7 ^a
	14	1382 ± 1068 ^a	2025 ± 1012 ^a	199 ± 12 ^a	202 ± 14 ^a
	28	^a	^a	201 ± 9 ^a	210 ± 5 ^a

Indices with small letters indicate differences within one column of solutions prepared with the same heat treatment ($\alpha = 0.05$). Indices with capital letters indicate a difference between a pH of 2.5 and 3.0 within one row calculated from a *t*-test ($\alpha = 0.05$). ^aWere excluded because of precipitation.

Table 3 Zeta potential (mV) of 0.04% (w/w) PC solutions and PC:λC (1:4) complex solutions unheated and subjected to heat treatments at 70 or 90 °C at a pH of 2.5–6.0

Heat treatment	pH						
	2.5	3.0	3.5	4.0	4.5	5.0	6.0
0.04% PC solutions							
None	23.1 ± 0.4	24.0 ± 0.4	16.7 ± 0.1	1.7 ± 1.2	-16.4 ± 3.0	-20.1 ± 0.4	-12.4 ± 0.1
70 °C	22.9 ± 0.9	22.9 ± 1.3	21.0 ± 0.7	14.1 ± 1.2	-10.9 ± 2.2	-25.3 ± 2.3	-26.6 ± 0.4
90 °C	22.3 ± 1.6	23.3 ± 0.3	20.7 ± 1.0	15.9 ± 3.0	-5.4 ± 4.4	-21.7 ± 3.4	-24.3 ± 2.4
PC:λC (1:4) complex solutions							
None	-59.5 ± 2.3	-58.3 ± 1.0	-57.1 ± 1.3	-56.9 ± 1.8	-59.2 ± 0.1	-57.8 ± 1.0	-56.3 ± 1.8
70 °C	-56.7 ± 0.7	-54.6 ± 1.9	-55.3 ± 1.4	-58.1 ± 0.8	-56.0 ± 1.4	-57.2 ± 1.9	-55.8 ± 0.1
90 °C	-36.7 ± 1.4	-45.6 ± 0.1	-50.5 ± 1.2	-55.0 ± 0.6	-56.3 ± 1.2	-55.6 ± 1.3	-57.2 ± 0.8

3.4 Mechanistic insights

Summarizing the results of previous studies and the present investigations, a hypothetical scheme is proposed in this study, which is summarized in Fig. 5.

Phycocyanin carries differently charged patches on its surface (Fig. 5A). At a pH of 6.0, PC trimers and carrageenan coexist in solution but do not attract each other (Fig. 5B). Lowering the pH to 3.0 causes the surface charge of PC to become more positive leading to the association of the poly-

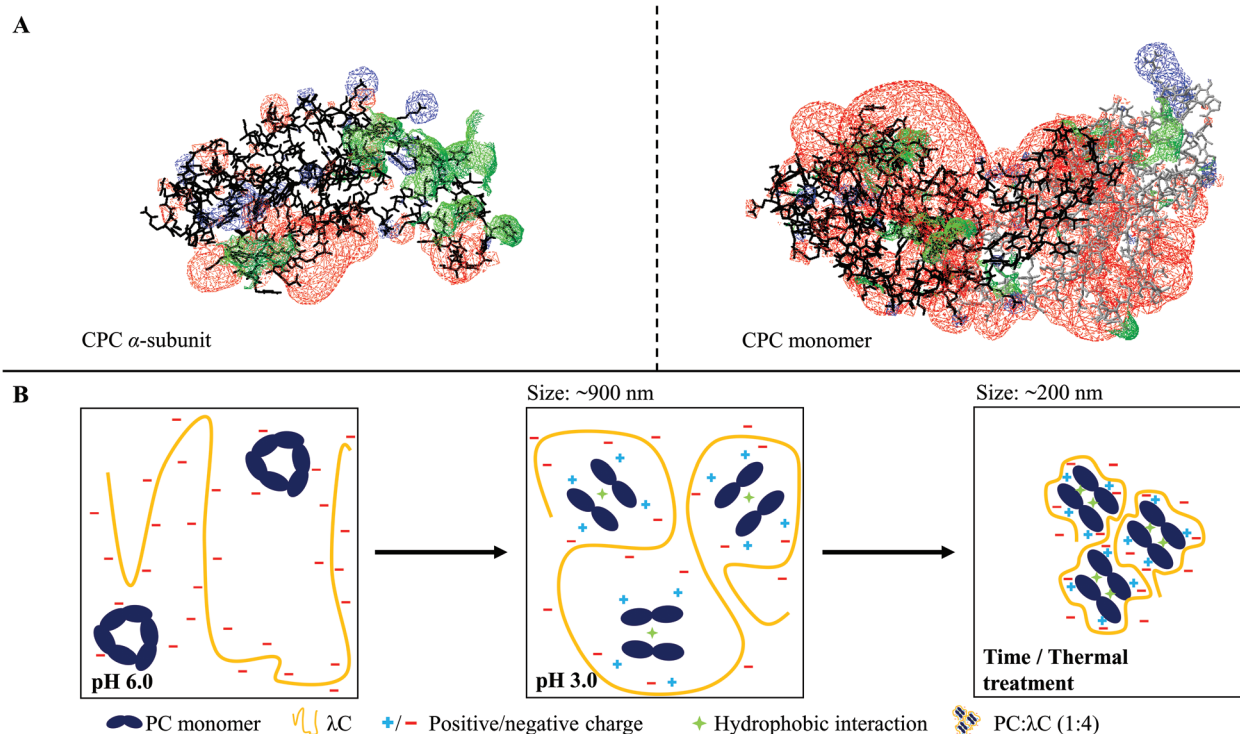


Fig. 5 A: Structure of a C-phycocyanin taken from Uniprot at pH 6.0 (α -subunit and monomer). Black amino acids belong to the α - and grey structures to the β -subunit. Positively charged surfaces are blue, negative charges are red, and hydrophobic groups are green. (<https://www.rcsb.org/structure/1HA7>) B: Mechanistic drawing of the PC : λ C (1 : 4) complex formation influenced by pH, thermal treatment, and time.

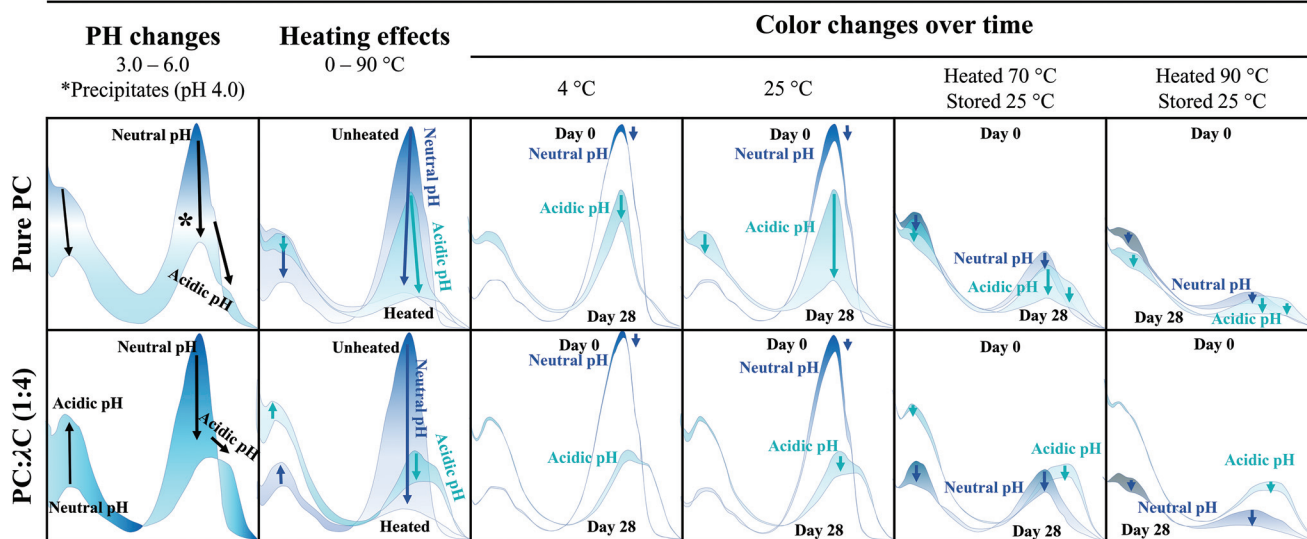


Fig. 6 Schematic illustration of the absorption spectrum (300–750 nm) from 0.04% (w/w) PC solutions and PC : λ C (1 : 4) solutions influenced by pH changes from 2.5–6.0, thermal treatments up to 90 °C and 28 days of storage at 4 °C and 25 °C.

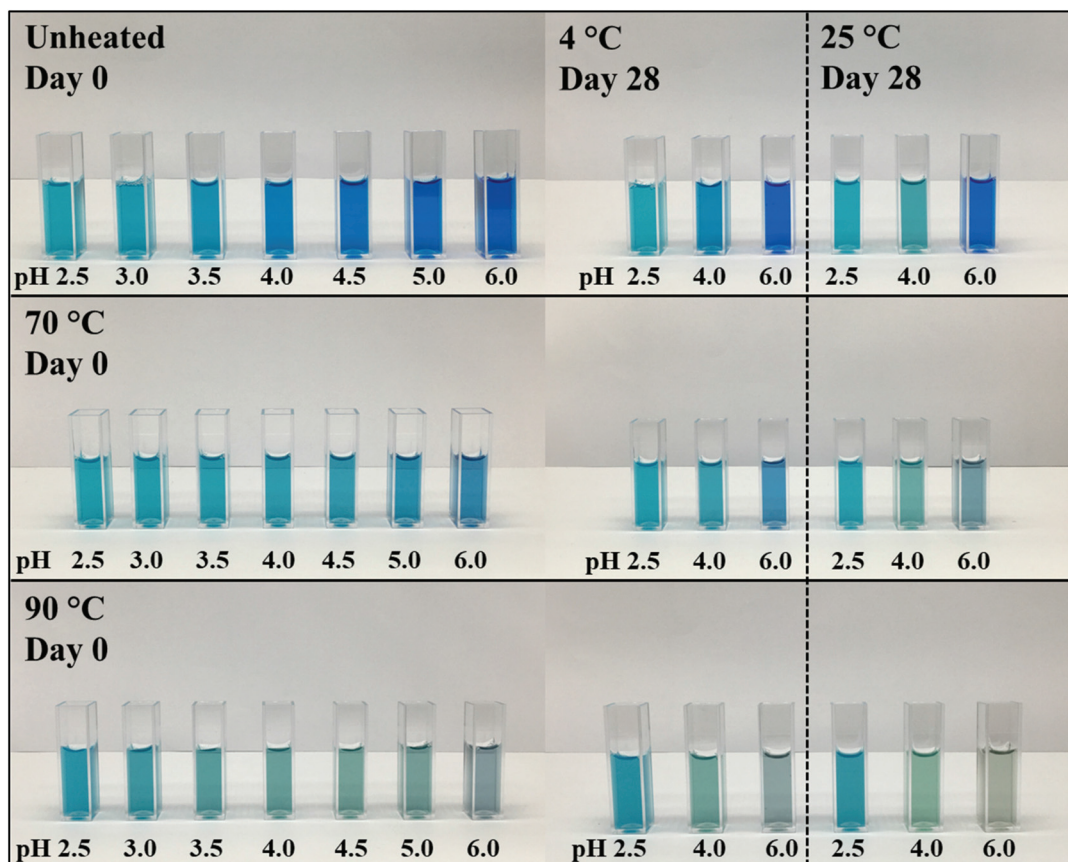


Fig. 7 Visual color appearance of PC : λC (1 : 4) solutions (~0.04% (w/w) PC) unheated and subjected to thermal treatments at 70 and 90 °C at a pH of 2.5–6.0 and stored over a period of 28 days at either 4 or 25 °C.

mers, which form stable soluble complexes. The PC structure shifts to monomers and the absorption shift towards red, which might indicate a further dissociation into α - and

β -subunits caused by the complex formation (Fig. 5B). Ongoing interactions and rearrangements of hydrophobic nature cause smaller or denser PC:λC complexes over time, which is favored by higher storage temperatures (Fig. 5B). Moreover, heat treatments during sample preparation lead to smaller or denser complexes as a result of increased hydrophobic interactions between PC trapped within the complex and accelerated electrostatic complexation.

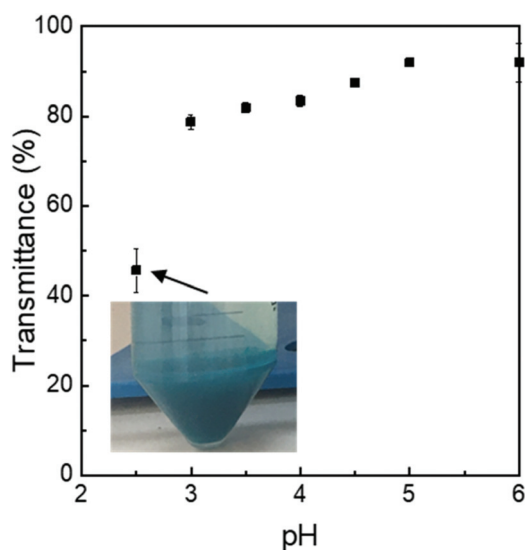


Fig. 8 Relative transmittance of PC:λC (1:4) complexes (~0.04% (w/w) PC) prepared at 90 °C and stored for 14 days at 25 °C plotted over pH.

4 Conclusion

Fig. 6 compiles our key findings on the effects of storage time and temperature on heated and unheated PC solutions and PC:λC complex solutions in a schematic illustration.

The color intensity of pure PC decreased over time especially at low pH values (pH 2.5 to 4.5), high heating (90 °C), and storage (25 °C) temperatures. However, electrostatic complexation with λC considerably increased the color stability at pH < pI and facilitated a stabilization in color over the 28-day storage period especially at a pH of 3.0 even when heated to 90 °C and stored at 25 °C. These results might be of great interest for low-viscous food applications such as smoothies or soft drinks that are stored either at refrigerated or room temperature and formulated with natural colorants.

Table 4 Nutritional values of the *Arthrospira platensis* extract and λ -carrageenan according to the manufacturers

	Arthrospira platensis extract Value (unit)	λ -Carrageenan
Energy	364 (kcal per 100 g)	272 (kcal per 100 g)
Protein	87.5 (g per 100 g)	1.0 (g per 100 g)
Carbohydrates	1.2 (g per 100 g)	67.00 (g per 100 g)
Sugar	1.17 (g per 100 g)	0.00 (g per 100 g)
Fat	0.60 (g per 100 g)	0.00 (g per 100 g)
Saturated fatty acids	0.36 (g per 100 g)	0.00 (g per 100 g)
Dietary fiber	0.5 (g per 100 g)	67 (g per 100 g)
Common salt	1.51 (g per 100 g)	17.2 (g per 100 g)

Conflicts of interest

There are no conflicts to declare.

Appendix

See Fig. 7 and 8 and Tables 3 and 4.

Acknowledgements

This project was partly funded by GNT Europa GmbH.

References

- D. L. Zink, *Emerging Infect. Dis.*, 1997, **3**, 467–469.
- C. Spence, *Int. J. Gastron. Food Sci.*, 2019, **17**, 1–10.
- L. Jespersen, L. D. Strmdahl, K. Olsen and L. H. Skibsted, *Eur. Food Res. Technol.*, 2005, **220**, 261–266.
- T. T. L. Cheung, A. F. Junghans, G. B. Dijksterhuis, F. Kroese, P. Johansson, L. Hall and D. T. D. de Ridder, *Appetite*, 2016, **106**, 2–12.
- B. Weiss, *Environ. Health Perspect.*, 2008, **116**, 240–241.
- Handbook on natural pigments in food and beverages. Industrial applications for improving food color*, ed. R. Carle and R. M. Schweiggert, Woodhead Publishing, Duxford, UK, 2016, number 295.
- European Parliament and European Council Regulation (EC) No 1333/2008 in *OJEU* L 354/16, 2008, pp. 1–18.
- D. S. Berns and R. MacColl, *Chem. Rev.*, 1989, **89**, 807–825.
- K. L. Thoren, K. B. Connell, T. E. Robinson, D. D. Shellhamer, M. S. Tammaro and Y. M. Gindt, *Biochemistry*, 2006, **45**, 12050–12059.
- N. Adir, R. Vainer and N. Lerner, *Biochim. Biophys. Acta, Bioenerg.*, 2002, **1556**, 168–174.
- Z. Zhang, S. Cho, Y. Dadmohammadi, Y. Li and A. Abbaspourrad, *Food Hydrocoll.*, 2021, **110**, 106055.
- G. O. Phillips and P. A. Williams, *Handbook of food proteins*, Woodhead Publishing, Cambridge UK, Philadelphia, 2011, no. 222.
- A. Reddy, D. F. Norris, S. S. Momeni, B. Waldo and J. D. Ruby, *J. Am. Dent. Assoc., JADA*, 2016, **147**, 255–263.
- R. Chaiklahan, N. Chirasuwan and B. Bunnag, *Process Biochem.*, 2012, **47**, 659–664.
- Y. Li, Z. Zhang and A. Abbaspourrad, *Food Hydrocoll.*, 2021, **119**, 106852.
- J. L. MacDonald and E. Leeb, EP3692804A1, 2020-08-12.
- W. R. Blakemore, *Reference Module in Food Science*, Elsevier, 2016.
- S. Buecker, L. Grossmann, M. Loeffler, E. Leeb and J. Weiss, *Food Chem.*, 2022, **380**, 132157.
- (a) C. Safi, M. Charton, O. Pignolet, F. Silvestre, C. Vaca-Garcia and P.-Y. Pontalier, *J. Appl. Phycol.*, 2013, **25**, 523–529; (b) R. Matissek, M. Fischer and G. Steiner, *Lebensmittelanalytik*, Springer Berlin Heidelberg, Berlin, Heidelberg, 2018.
- N. Yoshikawa and A. Belay, *J. AOAC Int.*, 2008, **91**, 524–529.
- E. W. Becker, *Biotechnol. Adv.*, 2007, **25**, 207–210.
- (a) S. T. Silveira, L. K. d. M. Quines, C. A. V. Burkert and S. J. Kalil, *Bioprocess Biosyst. Eng.*, 2008, **31**, 477–482; (b) CN101942014B.
- 秦松, 刘冰, 闫鸣艳, 林甜甜, 衣悦涛, CN101942014B, 2010-08-23.
- (a) S. Benelhadj, A. Gharsallaoui, P. Degraeve, H. Attia and D. Ghorbel, *Food Chem.*, 2016, **194**, 1056–1063; (b) A. Vonshak, *Spirulina platensis (Arthrospira). Physiology, cell-biology, and biotechnology*, Taylor & Francis, London, Bristol PA, 1997.
- M. Thompson, L. Owen, K. Wilkinson, R. Wood and A. Damant, *Analyst*, 2002, **127**, 1666–1668.
- (a) H.-N. Su, B.-B. Xie, X.-L. Chen, J.-X. Wang, X.-Y. Zhang, B.-C. Zhou and Y.-Z. Zhang, *J. Appl. Phycol.*, 2010, **22**, 65–70; (b) M. Moranças, J.-L. Mouget and J. Dumay, *Microalgae in Health and Disease Prevention*, Elsevier, 2018, pp. 145–175.
- V. K. Kannaujiya and R. P. Sinha, *J. Appl. Phycol.*, 2016, **28**, 1063–1070.
- N. Soulier and D. A. Bryant, *Photosynth. Res.*, 2021, **147**, 11–26.
- L. Böcker, T. Hostettler, M. Diener, S. Eder, T. Demuth, J. Adamcik, K. Reineke, E. Leeb, L. Nyström and A. Mathys, *Food Chem.*, 2020, **316**, 126374.
- L. Böcker, S. Ortmann, J. Surber, E. Leeb, K. Reineke and A. Mathys, *Innovative Food Sci. Emerging Technol.*, 2019, **52**, 116–121.
- G. O. Phillips and P. A. Williams, *Handbook of Hydrocolloids*, Woodhead Pub, Cambridge, 2nd edn, 2009.
- R. MacColl, *J. Struct. Biol.*, 1998, **124**, 311–334.
- X. Li, L. Li, Y. Ma, R. Wang, Y. Gu and L. Day, *Food Biosci.*, 2020, **34**, 100530.
- C. K. W. Koo, C. Chung, J.-T. R. Fu, A. Sher, P. Rousset and D. J. McClements, *Food Res. Int.*, 2019, **123**, 779–789.
- (a) R. MacColl, L. E. Eisele and A. Menikh, *Biopolymers*, 2003, **72**, 352–365; (b) O. Kao, D. S. Berns and R. MacColl, *Eur. J. Biochem.*, 1971, **19**, 595–599.

- 36 R. MacColl and D. Guard-Friar, *Phycobiliproteins*, CRC Press, Boca Raton Fla., 1987.
- 37 C. Tanford, *The hydrophobic effect. Formation of micelles and biological membranes*, Wiley, New York, 2nd edn, 1980.
- 38 A. Veis and C. Aranyi, *J. Phys. Chem.*, 1960, **64**, 1203–1210.
- 39 (a) F. Weinbreck, R. de Vries, P. Schrooyen and C. G. de Kruif, *Biomacromolecules*, 2003, **4**, 293–303; (b) S. Kim, J. Huang, Y. Lee, S. Dutta, H. Y. Yoo, Y. M. Jung, Y. Jho, H. Zeng and D. S. Hwang, *Proc. Natl. Acad. Sci. U. S. A.*, 2016, **113**, E847–E853.
- 40 G. Job and R. Rüffler, *Physical Chemistry from a Different Angle*, Springer International Publishing, Cham, 2016.
- 41 G. Saelensminde, Ø. Halskau and I. Jonassen, *Extremophiles*, 2009, **13**, 11–20.
- 42 (a) X. Lu, S. Xie, L. Wang, H. Xie, Q. Lei and W. Fang, *Chem. Phys.*, 2020, **538**, 110910; (b) G. Berth, J. Vukovic and M. D. Lechner, *J. Appl. Polym. Sci.*, 2008, **110**, 3508–3524.
- 43 N. Al-Zebari, S. M. Best and R. E. Cameron, *J. Phys. Mater.*, 2019, **2**, 15003.
- 44 B. Zeeb, L. Mi-Yeon, M. Gibis and J. Weiss, *Food Hydrocoll.*, 2018, **74**, 53–61.