

Cite this: *Nanoscale Adv.*, 2021, 3, 2710

Ligand exchange reactions on thiolate-protected gold nanoclusters

Yanan Wang  and Thomas Bürgi *

As a versatile post-synthesis modification method, ligand exchange reaction exhibits great potential to extend the space of accessible nanoclusters. In this review, we summarized this process for thiolate-protected gold nanoclusters. In order to better understand this reaction we will first provide the necessary background on the synthesis and structure of various gold clusters, such as Au₂₅(SR)₁₈, Au₃₈(SR)₂₄, and Au₁₀₂(SR)₄₄. The previous investigations illustrated that ligand exchange is enabled by the chemical properties and flexible gold–sulfur interface of nanoclusters. It is generally believed that ligand exchange follows a S_N2-like mechanism, which is supported both by experiments and calculations. More interesting, several studies show that ligand exchange takes place at preferred sites, *i.e.* thiolate groups –SR, on the ligand shell of nanoclusters. With the help of ligand exchange reactions many functionalities could be imparted to gold nanoclusters including the introduced of chirality to achiral nanoclusters, size transformation and phase transfer of nanoclusters, and the addition of fluorescence or biological labels. Ligand exchange was also used to amplify the enantiomeric excess of an intrinsically chiral cluster. Ligand exchange reaction accelerates the prosperity of the nanocluster field, and also extends the diversity of precise nanoclusters.

Received 8th March 2021

Accepted 2nd April 2021

DOI: 10.1039/d1na00178g

rsc.li/nanoscale-advances

1. Introduction

During the past decade, the interest in monolayer-protected metal clusters has drastically increased. The progress in the field has been reported in review articles focussing for example on precise synthesis,^{1–5} metal doping,^{6,7} and applications of gold nanoclusters.^{8,9} In general, the chemical properties of

nanomaterials largely depend on their surface properties. As a flexible surface chemistry method, ligand exchange reaction (LER) is therefore an important tool for the development of nanomaterials. For instance, Dinkel and co-workers summarized the findings obtained by *in situ* spectroscopic measurements to probe the ligand exchange at the surface of colloidal gold and silver nanoparticles. Specifically, the results obtained by the inherently surface sensitive technique second-harmonic scattering (SHS) was discussed.¹⁰ In addition, Xu and co-workers concentrated on exchange reactions on metal–organic

Department of Physical Chemistry, University of Geneva, 30 Quai Ernest-Ansermet, 1211 Geneva 4, Switzerland. E-mail: thomas.buergi@unige.ch



Yanan Wang received her Bachelor's degree in Applied Chemistry and Master's degree in bioengineering from China University of Petroleum (East China) in 2014 and 2017, respectively. Since 2017, she has been working in the group of Prof. Thomas Bürgi as a PhD candidate in the Department of Physical Chemistry of the University of Geneva. Currently, her research mainly focuses on

the synthesis and characterization of chiral gold nanoclusters and their potential applications.



Thomas Bürgi studied chemistry and obtained his PhD (1995) at the University of Berne (Switzerland). After a postdoc at MIT, he did his habilitation at ETH, Zürich. He became assistant professor at the University of Neuchâtel (Switzerland, 2005) and full professor at the University of Heidelberg (2008). In 2010 he moved to the University of Geneva, where he is professor of physical chemistry.

His research focuses on fundamental aspects and applications of chiral metal clusters, plasmon-based metamaterials and the development of *in situ* spectroscopy.



frameworks (MOFs).¹¹ More focussing on metal clusters, Zhu and co-worker published a review on transformations of precise nanoclusters induced by ligand exchange.¹² They focused on the size conversion of gold and silver nanoclusters during ligand exchange reaction. However, a comprehensive summary focusing on the ligand exchange reaction of thiolate-protected gold clusters is still rare.¹³

Recently, many new findings have emerged on the mechanism and peculiarities of the ligand exchange reactions on thiolate-protected clusters, including ligand exchange between clusters,¹⁴ ligand exchange including chiral thiols,¹⁵ and ligand exchange induced amplification of enantiomeric excess.¹⁶ These new findings are of great interest and further expanded the knowledge in the field of the ligand exchange reaction on thiolate-protected gold nanoclusters. Considering the recent findings in the field and its implications for applications an attempt to summarize to current knowledge seems appropriate.

The ligand exchange reaction is an important post-synthesis method for the modification and further functionalization of gold nanoclusters. In this review, we specifically address the ligand exchange reactions (LERs) on thiolate-protected gold nanoclusters. The main points to be addressed are illustrated in Scheme 1. We mainly focus on atomically precise gold nanoclusters with known crystal structure, such as Au₂₅(SR)₁₈, Au₃₈(SR)₂₄, and Au₁₀₂(SR)₄₄, which allows to obtain molecular level insight into the process. The clusters contain diverse surface motifs, *e.g.* monomeric and dimeric Au–sulfur staples and the distinct surface thiolates show different reactivity towards ligand exchange. It is also emerging from recent experiments and calculations that the surface region of these clusters, *i.e.* the Au–sulfur interface, is far from being static. Furthermore, not only free thiols in solution can exchange with thiols on the cluster, but thiols can exchange among the clusters without addition of free thiols. In addition, LERs also takes place between free thiols and supported clusters. Ligand

exchange can lead to important modifications of the cluster properties. For example, LERs can induce chirality on achiral nanoclusters and also lead to the enhancement of enantiomeric excess of chiral clusters. LERs can promote size transfer of clusters, and phase transfer between organic and aqueous phase. In general, ligand exchange reaction is an important process that largely extends the diversity of precise nanoclusters and their potential for applications. In the following, after an introduction to monolayer-protected gold clusters, we summarize the current knowledge on LERs with special focus on the more recent findings.

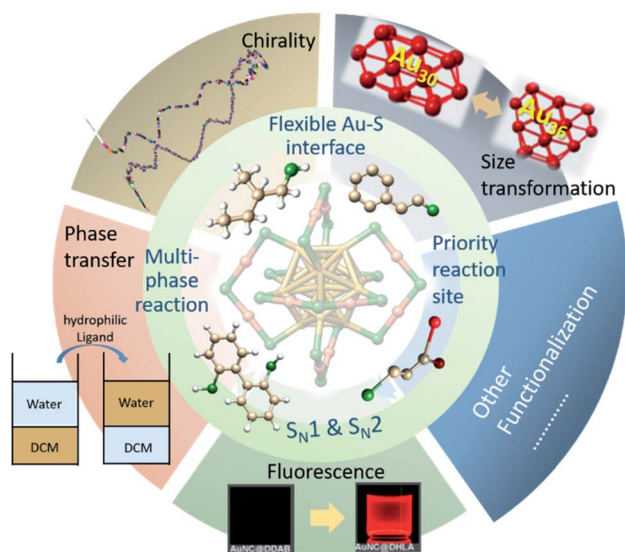
2. Precise monolayer-protected gold nanoclusters

2.1 Synthesis of precise gold nanoclusters

Metal nanoclusters are composed of tens to hundreds of atoms and bridge the gap between single atoms or molecules and nanoparticles.^{5,17} Different from the plasmon excitation in nanoparticles, which is a collective excitation of conduction electrons, metal nanoclusters are characterized by strong quantum confinement effect and molecular-like properties.^{2,18} Nowadays, metal nanoclusters include several sub-classes, such as thiolate-protected silver nanoclusters, phosphine-protected gold nanoclusters, thiolate-protected gold nanoclusters and so on. Among them, thiolate-protected gold nanoclusters are the most studied due to their relatively long history and unique properties.^{3,18,19} In the 1990s, triggered by the work of Brust and Schiffrin, several groups started the research on gold nanoclusters and obtained a series of ultrasmall species.²⁰ Typically in these syntheses mixtures of different cluster sizes were obtained. At this size every atom counts and minor changes in composition can have large effects on electronic and optical properties of the nanoclusters. Therefore, the precise control of the composition and purity is critically important.³

In order to obtain pure nanoclusters, Whetten and co-workers tried to separate cluster mixtures by polyacrylamide gel electrophoresis (PAGE)²¹ and solvent-extraction,^{22,23} however, it is challenging to obtain pure cluster samples in this way. Murray and co-workers prepared the Au₂₅ cluster by a two-phase protocol^{24,25} or the conversion of Au₁₁ to Au₂₅,²⁶ but mixtures of clusters of different size were obtained. After purification by solvent-extraction and other methods, clusters of different size were obtained with very low yield. Later, Tsukuda and co-workers applied PAGE,²⁷ recycling size exclusion chromatograph²⁸ and solvent-extraction²⁹ to isolate the precise nanoclusters in a time-consuming strategy. After about ten years of struggling to obtain pure cluster samples in high yield, there was a strong need for efficient methods towards synthesis of precise gold nanoclusters.

The first breakthrough was reported for Au₂₅, a prototypical gold cluster. In 2008 Zhu and co-workers reported a high yield synthesis of Au₂₅(SCH₂CH₂Ph)₁₈ by kinetic control with a two-phase method.³⁰ Later on, by using THF as solvent, Wu and co-workers achieved a big improvement of the synthesis of Au₂₅ nanoclusters using a one-pot method.³¹ Importantly, a “size-



Scheme 1 Schematic diagram of ligand exchange reactions on thiolate-protected gold nanoclusters.



focusing” process was identified in the growth of Au₂₅ nanoclusters, which accounts for the formation of atomically monodisperse Au₂₅ nanoclusters from the one-pot reaction.

Similar to the success in the synthesis of monodisperse Au₂₅, a facile solution synthetic method for obtaining monodisperse Au₃₈(S-C₁₂H₂₅)₂₄ was developed by exploring a two phase ligand exchange process in which glutathione-capped Au_n clusters are utilized as the starting material.³² Here a LER was carried out at elevated temperature (e.g. 80 °C) to obtain high purity Au₃₈(S-C₁₂H₂₅)₂₄ clusters, still at relatively low yield. After that, Qian and co-workers modified the protocol by replacing methanol by acetone in the first step (synthesis of Au_n(SG)_m) and by using an excess of phenylethylthiol (PhC₂H₄SH) as the etching ligand for the ligand exchange reaction.³³ Using these modifications they could increase the synthesis yield of Au₃₈ to 25% (Au atom).³³ This work illustrated that the formation of Au₃₈(SC₂H₄Ph)₂₄ resulted from the conversion from larger clusters in the thiol etching induced growth process in a remarkable “size-focusing” process. The above results revealed that size-focusing is a very efficient methodology for synthesizing atomically precise gold nanoclusters. The underlying principle can be illustrated as shown in Fig. 1.² Initially several nanoclusters with different sizes and vastly different stability coexist in the reaction mixture. The most stable nanocluster within the initially controlled size distribution will be sieved out.² After the success in obtaining Au₂₅ and Au₃₈ nanoclusters using the size focusing method, a series of Au_n(SR)_m nanoclusters were synthesized in high yield and on a large scale, using this methodology, such as Au₂₃,³⁴ Au₆₄,³⁵ Au₉₉,³⁶ Au₁₄₄ (ref. 37) and Au₃₃₃.³⁸ As the size-focusing involves core-etching from larger clusters to smaller clusters, the size (distribution) of the initial thiolate-protected gold cluster is critical. In addition, the high yield product may still coexist with some by-products, making additional purification steps such as size exclusion chromatography (SEC) necessary. Knoppe and co-workers used SEC to separate Au₃₈ and Au₄₀ clusters, which differ by only two gold atoms.³⁹ This example demonstrates the power of this separation technique.

The development of synthesis methods resulted in several monodisperse nanoclusters including Au₂₅(SR)₁₈, Au₃₈(SR)₂₄,

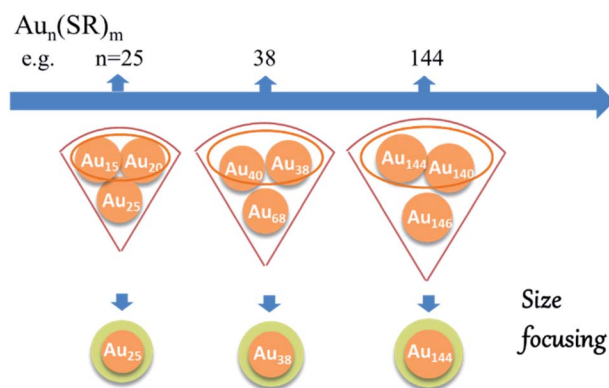


Fig. 1 Schematic of the “size-focusing” process. Adapted with permission from ref. 2. Copyright 2010 American Chemical Society.

Au₉₉(SR)₄₂,³⁶ and Au₁₄₄(SR)₆₀.³⁷ Such clusters have already become excellent candidates for applications in catalysis,^{36,40,41} as optical active materials,^{42,43} as chemical sensors,^{44,45} and for biological applications.^{46,47}

The availability of numerous atomically precise thiolate-protected gold clusters allowed one to study the evolution of properties as a function of size. Compared with the bulk metal, which has continuous energy band structure, ultrasmall nanoclusters possess size-dependent electronic structure, a HOMO–LUMO gap and molecule-like properties. The HOMO–LUMO gap of Au₂₅(SR)₁₈ was found to be 1.3 eV,³⁰ which is bigger than the value of 1.0 eV found for Au₃₈(SR)₂₄.³³ This implies that the larger clusters have a smaller energy gap, as expected. Having many additional clusters at hand, the relationship between HOMO–LUMO gap and size of the nanoclusters became clearer. There is a general trend of shrinking gap with increasing size. Plots of the HOMO–LUMO gap E_g against $n^{-1/3}$ or against $\ln(n)$ (n : number of gold atoms) revealed linear relationship and two groups of clusters.¹⁷ The two sets of clusters have difference structure and atomic packing, which will be addressed in the next subsection.

2.2 Structure of precise monolayer-protected gold nanoclusters

In order to understand the fundamental science of nanoclusters, it is necessary to determine their structures. X-ray single-crystal diffraction is the most straightforward and important technique for the structure determination of nanoclusters. Among the most important milestones in the research history of structural characterization of nanoclusters goes back to 2007, when Kornberg and co-workers published the first total X-ray structure of a thiolate-protected gold cluster, containing 102 gold atoms and 44 *p*-mercaptobenzoic acid (*p*-MBA) ligands.⁴⁸ The structure of Au₁₀₂(*p*-MBA)₄₄ was unexpected and did not match the view emerging from the “cluster of clusters” growth mechanism proposed for cluster chemistry.⁴⁹ As shown in Fig. 2A, the gold core has a decahedron structure and is surrounded by a gold thiolate layer characterized by “staple” motifs. In these structures, one gold atom is bound to two sulfur atoms. Similar structures were previously predicted theoretically as gold-thiolate rings.⁵⁰ The Au₁₀₂(*p*-MBA)₄₄ cluster contains 19 short and two long motif SR(AuSR)_x ($x = 1, 2$). In addition, the cluster is chiral because of the geometry of the equatorial atoms (Fig. 2B). The stability of the cluster structure has several origins. The cluster has 58 free electrons and thus has a closed electronic shell within a superatom model.⁵¹ The thiolate monolayer contributes as well to the stability of the cluster and within this layer additional stability stems from the π - π stacking between phenyl rings of the *p*-MBA molecules.

Owing to the development of precise synthesis of nanoclusters and X-ray single-crystal diffraction, structure determination of gold nanoclusters ushered in a period of vigorous development after the first breakthrough of Au₁₀₂(*p*-MBA)₄₄.⁴⁸ The experimental crystal structure of ultrasmall gold nanocluster Au₂₅(SR)₁₈ was reported separately by Jin’s and Murray’s groups.^{52,53} Grönbeck and co-workers also predicted the same



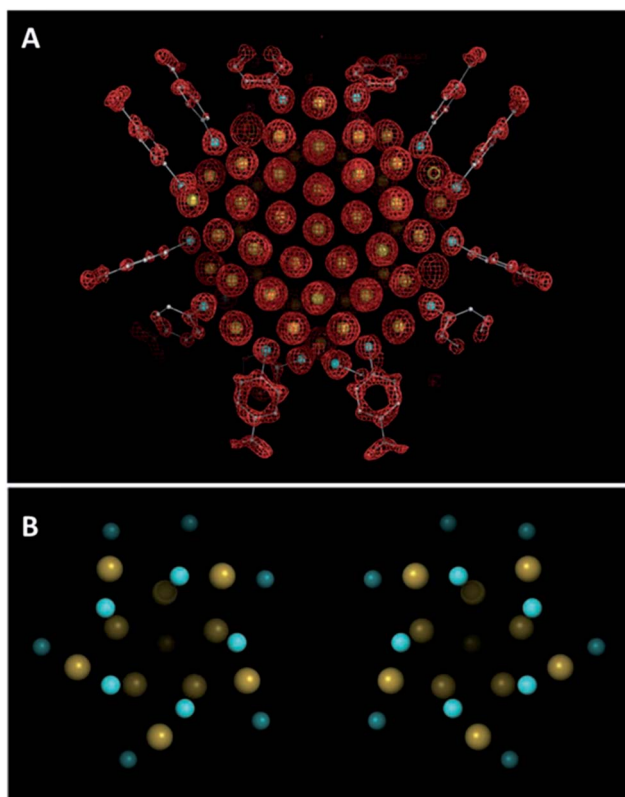


Fig. 2 X-ray crystal structure of the $\text{Au}_{102}(\text{p-MBA})_{44}$ crystal. (A) Electron density map and atomic structure (Au atoms depicted as yellow spheres, and *p*-MBA shown as framework and with small spheres [S in cyan, C in gray, and O in red]). (B) Top view of the two enantiomeric clusters. Colour scheme as in (A), only the sulfur atoms of *p*-MBA are shown. Adapted with permission from ref. 48. Copyright 2007 American Association for the Advancement of Science.

$\text{Au}_{25}(\text{SR})_{18}$ structure by DFT calculations.⁵⁴ X-ray crystallographic analysis shows that the Au_{25} cluster has an Au_{13} icosahedral core (Fig. 3) surrounded by a monolayer containing 12 gold atoms and 18 thiolates (Fig. 3). The crystal structure reveals three types of gold atoms ((i) one central gold atom (ii) 12 gold atoms as the vertices of the icosahedron, and (iii) 12 gold atoms forming staple) and two types of SR groups (terminal SR groups which link to the gold core, and central SR groups which link to

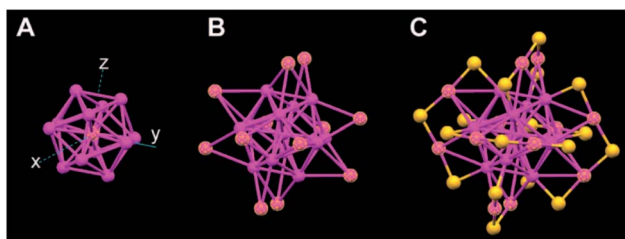


Fig. 3 Crystal structure of the $\text{Au}_{25}(\text{SR})_{18}$ cluster. (A) The icosahedral Au_{13} core; (B) the Au_{13} core plus the exterior 12 Au atoms; (C) whole Au_{25} cluster protected by 18 thiolate ligands (just S is shown for clarity, magenta, Au; yellow, S). Adapted with permission from ref. 52. Copyright 2008 American Chemical Society.

two staple gold atoms). The high symmetry structure shown above was obtained from the anion $\text{Au}_{25}(\text{SR})_{18}^-$, which is an 8 electron superatom with S^2P^6 electronic structure, but a similar structure was found for neutral $\text{Au}_{25}(\text{SR})_{18}$.⁵²

Compared with $\text{Au}_{25}(\text{SR})_{18}$, the crystal structure of $\text{Au}_{38}(\text{SR})_{24}$ is slightly more complex.⁵⁵ As illustrated in Fig. 4A, the $\text{Au}_{38}(\text{SR})_{24}$ cluster has a face-fused biicosahedral Au_{23} core is covered by six dimeric ($-\text{SR}-\text{Au}-\text{SR}-\text{Au}-\text{SR}-$) and three monomeric staples ($-\text{SR}-\text{Au}-\text{SR}-$). A similar structure has been predicted by Zeng⁵⁶ just slightly differing in the arrangement of the dimeric staples on the icosahedral Au_{13} unit. The monomeric staples were found for $\text{Au}_{38}(\text{SR})_{24}$ but not for $\text{Au}_{25}(\text{SR})_{18}$ which is just protected by six dimeric staples. This observation led to the prediction that the smaller size clusters need longer staples for the protection considering the large curvature for small gold core.⁵⁷

Similar to the case of $\text{Au}_{102}(\text{SR})_{44}$ and $\text{Au}_{38}(\text{SR})_{24}$, the structure of $\text{Au}_{28}(\text{TBBT})_{20}$ (where TBBT = 4-*tert*-butylbenzenethiolate) is also chiral (Fig. 4B).^{48,55,58} The cluster has an Au_{20} chiral kernel protected by four dimeric “staples” and eight bridging thiolates, and the chirality of the structure arises due to the rotational arrangement of the four dimeric staples and the arrangement of eight bridging thiolates. Later, Jin’s group

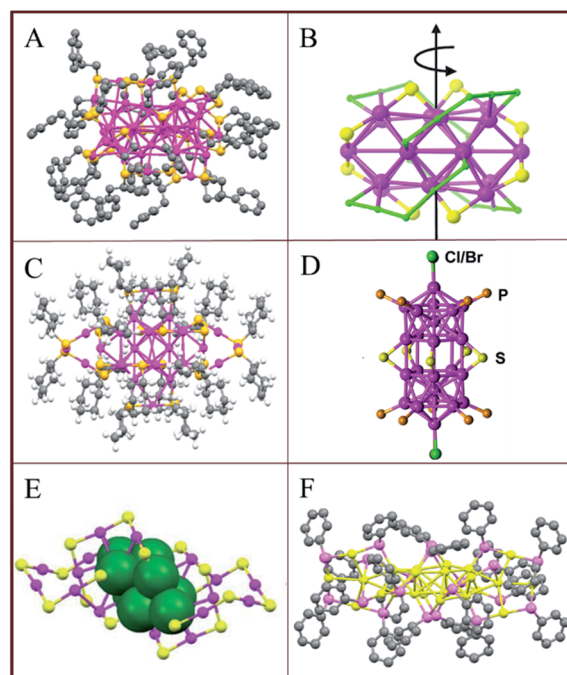


Fig. 4 Crystal structures of (A) $\text{Au}_{38}(\text{PET})_{24}$, adapted with permission from ref. 55. Copyright 2010 American Chemical Society. (B) $\text{Au}_{28}(-\text{TBBT})_{20}$, adapted with permission from ref. 58. Copyright 2013 American Chemical Society. (C) $\text{Au}_{28}(\text{S}-c-\text{C}_6\text{H}_{11})_{20}$, adapted with permission from ref. 59. Copyright 2016 American Chemical Society. (D) $[\text{Au}_{24}(\text{PPh}_3)_{10}(\text{SC}_2\text{H}_4\text{Ph})_5\text{X}_2]^+$, adapted with permission from ref. 60. Copyright 2012 American Chemical Society. (E) $\text{Au}_{24}(\text{SCH}_2\text{Ph}-t\text{Bu})_{20}$, adapted with permission from ref. 61. Copyright 2014 Royal Society of Chemistry. (F) $\text{Au}_{24}(\text{SeC}_6\text{H}_5)_{20}$, adapted with permission from ref. 62. Copyright 2014 American Chemical Society. The ligands are partially omitted for clarity.



reported an isomer of $\text{Au}_{28}(\text{TBBT})_{20}$ obtained after the ligand exchange with excess $\text{SH}-c\text{-C}_6\text{H}_{11}$ ligand. The obtained $\text{Au}_{28}(\text{S}-c\text{-C}_6\text{H}_{11})_{20}$ has two trimeric staples arranged on the surface as shown in Fig. 4C.⁵⁹ The above results revealed that, for a given number of gold atoms, the structure of a cluster may be modified by the ligand. This was also observed for Au_{24} clusters.^{60–62} $\text{Au}_{24}\text{L}_{20}$ clusters have different structures depending on the ligands (L): (i) Mixed ligand shell composed of phosphine and thiolate (Fig. 4D),⁶⁰ (ii) thiolate (Fig. 4E)⁶¹ and (iii) selenolate-capped Au_{24} clusters (Fig. 4F).⁶² This confirms that the structure of gold clusters is affected by the protecting ligand on the surface.

Apart from the clusters we mentioned above, the crystal structure of some other clusters have also been reported including $\text{Au}_{18}(\text{SR})_{14}$,⁶³ $\text{Au}_{20}(\text{TBBT})_{16}$,⁶⁴ $\text{Au}_{21}(\text{S}-t\text{Bu})_{15}$,⁶⁵ $\text{Au}_{36}(\text{SR})_{24}$,⁶⁶ $\text{Au}_{92}(\text{SR})_{44}$,⁶⁷ $\text{Au}_{130}(p\text{-MBT})_{50}$,⁶⁸ $\text{Au}_{144}(\text{SCH}_2\text{Ph})_{60}$.⁶⁹ Among those clusters, $\text{Au}_{144}(\text{SR})_{60}$ has attracted a lot of attention. The structure of $\text{Au}_{144}(\text{SCH}_2\text{Ph})_{60}$ was predicted by Olga Lopez-Acevedo and co-workers in 2009,⁷⁰ but the experimental structure was published almost one decade later, in 2018, by Zhikun Wu and co-workers.⁶⁹ The $\text{Au}_{144}(\text{SCH}_2\text{Ph})_{60}$ has a Au_{12} icosahedral inner core covered by two Au shells resulting in a Au_{114} core. 30 monomeric staple motifs are distributed on the surface of the Au_{114} core in a highly ordered pattern. Because of the arrangement of the 30 monomeric staples, the $\text{Au}_{144}(\text{SCH}_2\text{Ph})_{60}$ structure is chiral. With more and more crystal structures of gold clusters determined, the influence of size and structure on the HOMO–LUMO gap can be established. Atom packing plays an important role for E_g , as mentioned above, and the FCC (face-centered cubic) structures have much higher E_g energies than the icosahedral ones of comparable sizes.¹⁷

2.3 Flexibility of the gold–sulfur interface of gold nanoclusters

The formation of a gold–sulfur bond is the driving force for the anchoring of thiol ligands on either gold surfaces, nanoparticles or clusters,⁷¹ leading to the formation of self-assembled monolayers (SAM).⁷² From the available structural investigations it emerges that the staple-type binding motif is the preferred structure element of the gold–sulfur interface, as first evidenced by the crystal structure determination of $\text{Au}_{102}(\text{SR})_{44}$.⁴⁸ After analysis of various nanocluster structures such as $\text{Au}_{25}(\text{SR})_{18}$, $\text{Au}_{38}(\text{SR})_{24}$ and $\text{Au}_{28}(\text{SR})_{20}$, the bridging motif (Fig. 5A), monomeric staple (Fig. 5B) and dimeric staple motif (Fig. 5C) are the most common surface structure elements. Actually the tetrameric staple motif also has been

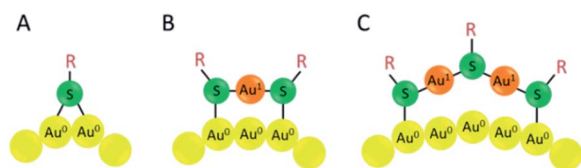


Fig. 5 Schematic drawing of the bridging thiolate (A), monomeric Au–SR–Au staple motif (B) and dimeric Au–SR–Au–SR–Au staple motif. Yellow: Au (0) atom; orange: Au(I) atom; green: sulphur atom.

evidenced for $\text{Au}_{24}(\text{SCH}_2\text{Ph}-t\text{Bu})_{20}$.⁶¹ Also other staple motifs are expected in order to adapt to the different curvatures of nanoclusters.

The properties of gold–sulfur interface and its dynamics determine the further functionalization of thiolate-protected clusters and their applications. However, due to the experimental difficulties in probing the dynamic properties of the interface, this field was barren for a long time. Since the structure of the $\text{Au}_{38}(\text{SR})_{24}$ cluster been solved it became a good candidate for studying the gold–sulfur interface.⁵⁵ The cluster is intrinsically chiral due to the chiral arrangement of the dimeric staples at the poles of the cluster. Dolamic and co-workers reported the separation of the enantiomers of $\text{Au}_{38}(\text{SCH}_2\text{CH}_2\text{Ph})_{24}$ (ref. 73) by chiral high-performance liquid chromatography. Using enantiopure $\text{Au}_{38}(\text{SR})_{24}$ clusters Knoppe and co-workers showed that the cluster can racemize (Fig. 6, top).⁷⁴ The reaction was started with A- $\text{Au}_{38}(\text{SCH}_2\text{CH}_2\text{Ph})_{24}$ and C- $\text{Au}_{38}(\text{SCH}_2\text{CH}_2\text{Ph})_{24}$ (A: anticlockwise, C: clockwise), respectively, and the evolution of the clusters was followed by circular dichroism spectroscopy. From this investigation, the racemization of the enantiopure $\text{Au}_{38}(\text{SR})_{24}$ clusters takes place at modest temperatures without significant decomposition. Furthermore, from the temperature-dependent kinetic data, the activation barrier for the rearrangement within the thiolate–gold interface of the $\text{Au}_{38}(\text{SR})_{24}$ clusters is about 28 kcal mol^{-1} , which is lower than the gold–sulfur bond energy (about 50 kcal mol^{-1}). The relatively low activation barrier indicated that the racemization process of $\text{Au}_{38}(\text{SR})_{24}$ proceeds without complete Au–S bond

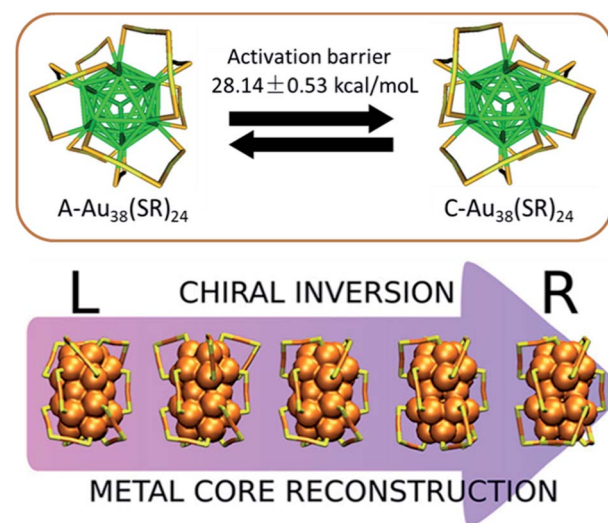


Fig. 6 Schematic drawing of the racemization reaction of $\text{Au}_{38}(\text{SR})_{24}$ cluster. (Top) In this scheme (view along the long axis of the cluster), the left-handed cluster (left, A- $\text{Au}_{38}(\text{SR})_{24}$) is converted into the right-handed enantiomer (right, C- $\text{Au}_{38}(\text{SR})_{24}$) and vice versa. The two enantiomers are separated by an activation barrier of ca. 28 kcal mol^{-1} as determined from experiments. Adapted with permission from ref. 74. Copyright 2012 American Chemical Society. (Bottom) Density functional theory computational simulation of the chiral inversion of $\text{Au}_{38}(\text{SR})_{24}$ clusters through rotation of the three gold core atoms at the poles of the cluster. Adapted with permission from ref. 75. Copyright 2019 American Chemical Society.



breaking and evidenced the flexibility of the gold-thiolate interface. In addition, using density functional theory (DFT) computations, Häkkinen and co-workers proposed a mechanism for this inversion of the Au–S framework of $\text{Au}_{38}(\text{SR})_{24}$ as shown in Fig. 6 (bottom).⁷⁵ In their model, the racemization proceeds *via* a rotational reconstruction of the metal core without any Au–S bonds being broken.

The flexibility of the gold–sulfur interface is affected by many factors, such as Pd doping, which is normally used for increasing the stability of the cluster and enhance its reactivity in catalytic and ligand exchange reactions. Barrabés and co-workers reported that the racemization process takes place at significantly lower temperature after doping the $\text{Au}_{38}(\text{SR})_{24}$ cluster with two Pd atoms compared with the parent cluster.⁷⁶ This case indicated that the Pd doping of the cluster renders the Au–S interface more flexible. In contrast, after introducing of a rigid dithiolate, 1,1'-binaphthyl-2,2'-dithiol (BINAS), into the ligand shell of the $\text{Au}_{38}(\text{SR})_{24}$ cluster, the racemization drastically slows down.⁷⁷ For example, the racemization of $\text{Au}_{38}(2\text{-PET})_{22}(\text{BINAS})_1$ at 70 °C is about 27 times slower compared to the parent cluster, meaning that the introduction of the dithiol reduces the flexibility of the Au–S interface. Interestingly, the vibrational spectrum of the Au–S interface is also drastically influenced by introducing one BINAS dithiol into the ligand shell of the cluster, possibly related to the fact that the dithiol bridges two staples.⁷⁸ Similarly, the racemization of the $\text{Au}_{40}(\text{SR})_{24}$ cluster was reported to take place at higher temperature compared with $\text{Au}_{38}(\text{SR})_{24}$ but with a similar activation barrier, further confirming the flexibility of gold–sulfur interface.⁷⁹

The racemization described above is not the only dynamic process observed for the $\text{Au}_{38}(\text{SR})_{24}$ cluster. In fact, ligands can migrate on the cluster surface between different symmetry unique sites. It has been reported that HPLC can be used to separate and isolate one specific regioisomer after ligand exchange between $\text{Au}_{38}(2\text{-PET})_{24}$ clusters and enantiopure chiral [2.2]-paracyclophane-4-thiol 1 (PCP-4-SH).⁸⁰ The isolated species is stable at room temperature, however, the adsorbed thiolate migrates between the different symmetry-unique sites at 80 °C as followed by HPLC.⁸⁰ The mechanism underlying this observation is not yet clear: it might involve the exchange of two ligands between different sites on one cluster or the exchange of ligands between clusters (see below).

The chemical properties and flexibility of the gold–sulfur interface, as discussed above, form the basis for the post-synthesis modification and further functionalization of the gold nanoclusters. Among them, ligand exchange reaction is extraordinarily sparkly and we will now focus on this topic in the following sections.

3. Ligand exchange reactions

3.1 Mechanism of ligand exchange reactions

Depending on the flexibility and the reactivity of the gold–sulfur interface, the thiolates within the ligand shell can be displaced by other thiols in the solution, which is named place-exchange reactions, or more commonly ligand exchange reactions (LERs).

For thiolate-protected clusters this reaction was first reported by Murry and co-workers.^{81–85} LERs were studied by ¹H NMR spectroscopy, thermal analysis and IR spectroscopy.^{81,82,84,85} Results of these experiments revealed some factors which may affect the process. The chemical reactivity is mostly independent of the nanoparticle or cluster size,⁸⁴ but the stability of the sample is a factor determining the fate of the cluster after the ligand exchange reactions.⁸⁶ The rate and extent of ligand exchange reactions was found to increase with increasing positive electronic charge on the Au cluster core,⁸² but it decreases with increasing size of the entering ligand and the carbon chain length of the protecting monolayer.⁸⁵ The ligand exchange reactions discussed in the following sections have been summarized in Table 1.

Evidence from related investigations suggested an associative pathway for the mechanism of ligand exchange reactions, which has been corroborated by various experimental and computational studies.^{14,87,88} For example, Heinecke and co-workers reported the first crystal structure of $\text{Au}_{102}(p\text{-MBA})_{40}(p\text{-BBT})_4$ (*p*-BBT = parabromobenzene thiol), which is formed by the ligand exchange reaction of $\text{Au}_{102}(p\text{-MBA})_{44}$ with *p*-BBT as the incoming ligand.⁸⁸ Available crystal structures of $\text{Au}_{102}(p\text{-MBA})_{44}$ and $\text{Au}_{102}(p\text{-MBA})_{40}(p\text{-BBT})_4$ were underpinned by a computational DFT study on energetics, reaction intermediates and pertinent transition states during the ligand exchange reactions, revealing microscopic details of this process. The associative ligand exchange mechanism, as emerging from a computational study, is illustrated in Fig. 7, where methane thiol is the incoming ligand. For associative ligand exchange, the more solvent exposed sulfur sites on $\text{Au}_{102}(p\text{-MBA})_{44}$ are more reactive and “nucleophilic attacked” by the incoming thiol(ate), creating an intermediate that has both incoming and outgoing ligands simultaneously bound to the accessible gold atom (Fig. 7b). Then the intermediate structure changed to the well-known “hemiring” unit with one Au replaced by an H atom (Fig. 7c). Depending on the orientation of the residue, the observed bond length between the hydrogen of the incoming ligand and the sulfur of the outgoing ligand become shorter and finally the sulfur of the outgoing ligand is released from the gold core (Fig. 7d). To complete exchange, the hydrogen atom of the adsorbed methane thiol is transferred to the outgoing ligand, which is then released to the solution (Fig. 7e). The intermediate states b (gold bound to three sulfur atoms) and d (breaking of S–H bond) have maximum energy during the exchange process.

A similar process may take place when labelling proteins with nanoclusters (as shown in Fig. 8). The initial reaction of this modification is a ligand exchange reactions of the anti-influenza N9 neuraminidase NC10 antibody against a glutathione ligand on a gold cluster as studied by means of *ab initio* QM/MM calculations.⁸⁷ Compared to the reaction with free thiol, the example displayed here concerns a large protein and may have two substitution modes as illustrated in Fig. 8. In the calculations ligand exchange of the side and apex glutathione ligands were considered, and showed that the intermediate from the side ligand is more stable. This investigation indicates that the essential features of the ligand exchange reaction are



Table 1 Summary of the examples of ligand exchange reactions mentioned in each section. The order in the table follows the order of discussion in the text

Section	Topic	Reaction	Reference
3.1	Ligand exchange with free ligand	$\text{Au}_{102}(\text{p-MBA})_{44} + \text{parabromobenzene thiol (p-BBT)} \rightarrow \text{Au}_{102}(\text{p-MBA})_{40}(\text{p-BBT})_4$	88
		Anti-influenza N9 neuraminidase NC10 antibody + $\text{Au}_{25}(\text{SG})_{18} \rightarrow \text{ScFv-Au}_{25}(\text{SG})_{18}$ -complex	87
		$[\text{Au}_{25}(\text{SC}_2\text{H}_4\text{Ph})_{18}]^- + \text{HSCH}_2\text{Ph-}^t\text{Bu} \rightarrow \text{Au}_{24}(\text{SCH}_2\text{Ph-}^t\text{Bu})_{20}$	86
	Intercluster ligand exchange	$\text{Au}_{24}\text{Cd}(\text{SC}_2\text{H}_4\text{Ph})_{18} + \text{HSCH}_2\text{Ph-}^t\text{Bu} \rightarrow \text{Au}_{24}\text{Cd}(\text{SCH}_2\text{Ph-}^t\text{Bu})_{18} + \text{Au}_{24}\text{Cd}(\text{SCH}_2\text{Ph-}^t\text{Bu})_{17}(\text{SC}_2\text{H}_4\text{Ph})_1$	
		$[\text{Au}_{25}(\text{SC}_{10}\text{H}_{21})_{18}]^- + [\text{Au}_{25}(\text{SC}_{12}\text{H}_{25})_{18}]^- \rightarrow \text{Au}_{25}(\text{SC}_{12}\text{H}_{25})_{18-x}(\text{SC}_{10}\text{H}_{21})_x$	89
		$\text{Au}_{25}(\text{SBut})_{18} + \text{Au}_{25}(2\text{-PET})_{18} \rightarrow \text{Au}_{25}(\text{SBut})_{18-x}(2\text{-PET})_x$	14
		$\text{Au}_{38}(2\text{-PET})_{24-2x}(\text{R-BINAS})_x + \text{Au}_{25}(2\text{-PET})_{18} \rightarrow \text{Au}_{25}(2\text{-PET})_{16}(\text{R-BINAS})_1 + \text{Au}_{38}(2\text{-PET})_{24-2x}(\text{R-BINAS})_x$	16
3.2	Site-selective ligand exchange	$\text{Au}_{24}\text{Pd}(\text{SC}_2\text{H}_4\text{Ph})_{18} + \text{C}_{12}\text{H}_{25}\text{SH} \rightarrow \text{Au}_{24}\text{Pd}(\text{SC}_2\text{H}_4\text{Ph})_{18-x}(\text{SC}_{12}\text{H}_{25})_x$	92
		$[\text{Au}_{25}(\text{SC}_2\text{H}_4\text{Ph})_{18}]^- + \text{HSePh/HTePh} \rightarrow [\text{Au}_{25}(\text{SC}_2\text{H}_4\text{Ph})_{18-x}(\text{SePh/TePh})_x]$	99
		$\text{Au}_{24}\text{Cd}(\text{SCH}_2\text{Ph})_{18} + \text{SCH}_2\text{Ph-}^t\text{Bu} \rightarrow \text{Au}_{24}\text{Cd}(\text{SCH}_2\text{Ph-}^t\text{Bu})_{17}(\text{SCH}_2\text{Ph})_1$	86
		$\text{PdAu}_{24}(2\text{-PET})_{18} + \text{S-BINAS} \rightarrow \text{PdAu}_{24}(2\text{-PET})_{16}(\text{S-BINAS})_1$	108
3.3		$\text{Au}_{11}(\text{PPh}_3)_7\text{Cl}_3$ (deposited on Al_2O_3) + L-glutathione(GSH) $\rightarrow \text{Au}_{11}(\text{PPh}_3)_7(\text{GS})_x\text{Cl}_{3-x}$	109
4.1	Ligand exchange induced chirality	$[\text{Au}_{25}(\text{PET})_{18}]^- [[\text{TOA}^+] + \text{R/S-BINAS} \rightarrow [\text{Au}_{25}(\text{PET})_{18-2x}(\text{R/S-BINAS})_x]^- [[\text{TOA}^+]$	107
		$[\text{Au}_{38}(\text{SR})_{24}]/[\text{Au}_{40}(\text{SR})_{24}] + \text{R/S-BINAS} \rightarrow [\text{Au}_{38}(\text{SR})_{24-2x}(\text{R/S-BINAS})_x]/[\text{Au}_{40}(\text{SR})_{24-2x}(\text{R/S-BINAS})_x]$	106
		$\text{A-Au}_{38}(2\text{-PET})_{24} + \text{C-Au}_{38}(2\text{-PET})_{22}(\text{R-BINAS})_1 \rightarrow \text{C-Au}_{38}(2\text{-PET})_{24} + \text{A-Au}_{38}(2\text{-PET})_{22}(\text{R-BINAS})_1$	16
4.2	Ligand exchange induced size transformation	$\text{Au}_{28}(\text{SPh-}^t\text{Bu})_{20} + \text{HS-c-C}_6\text{H}_{11} \leftrightarrow \text{Au}_{28}(\text{S-c-C}_6\text{H}_{11})_{20} + \text{HSPH-}^t\text{Bu}$	59
		$\text{Au}_{25}(\text{PET})_{18} + 4\text{-tert-butylbenzenethiol (TBBTH)} \rightarrow \text{Au}_{28}(\text{TBBT})_{20}$	58
		$\text{Au}_{11}(\text{PPh}_3)_7\text{Cl}_3 + \text{L-glutathione (GSH)} \rightarrow \text{Au}_{25}(\text{GS})_{18}$	26 and 110
		$\text{Au}_{144}(\text{SC}_2\text{H}_4\text{Ph})_{60} + \text{HSPH} \rightarrow \text{Au}_{99}(\text{SPh})_{42}$	128
		$\text{Au}_{38}(\text{SC}_2\text{H}_4\text{Ph})_{24} + \text{HSPH-}^t\text{Bu} \rightarrow \text{Au}_{36}(\text{SPh-}^t\text{Bu})_{24}$	120
		$\text{Au}_{23}(\text{S-c-C}_6\text{H}_{11})_{16} + \text{HTBBT} \rightarrow \text{Au}_{36}(\text{TBBT})_{24}$	127
		$\text{Au}_{329}(\text{PET})_{84} + \text{HSPH-}^t\text{Bu} \rightarrow \text{Au}_{279}(\text{SPh-}^t\text{Bu})_{84}$	129
		$\text{Au}_{36}(\text{SPhX})_{24}$ (where X = -H or - ^tBu) + $\text{HS-}^t\text{Bu} \rightarrow \text{Au}_{30}(\text{S-}^t\text{Bu})_{18}\text{Au}_{30}(\text{S-}^t\text{Bu})_{18} + \text{HSPHX}$ (where X = -H or - ^tBu) $\rightarrow \text{Au}_{36}(\text{SPhX})_{24}$	12 and 125
		$\text{Au}_{44}(\text{TBBT})_{28} + 2,4\text{-dimethylbenzenethiol (2,4-DMBT)} \leftrightarrow \text{Au}_{44}(2,4\text{-DMBT})_{26} + \text{TBBTH}$	130 and 131
		$\text{Au}_{43}(\text{S-c-C}_6\text{H}_{11})_{25} + 2,4\text{-DMBT} \leftrightarrow \text{Au}_{44}(2,4\text{-DMBT})_{26} + \text{HS-c-C}_6\text{H}_{11}$	
$\text{Au}_{43}(\text{S-c-C}_6\text{H}_{11})_{25} + \text{TBBTH} \rightarrow \text{Au}_{44}(\text{TBBT})_{28} + \text{HS-c-C}_6\text{H}_{11}$			
$\text{Au}_{25}(\text{SBut})_{18} + \text{R/S-phenylpropane-1-thiol (R/S-PPT)} \rightarrow \text{Au}_{28}(\text{R/S-PPT})_{21}$	15		
4.3	Ligand exchange induced phase transfer	$\text{Au}_{24}\text{Ag}_{20}(2\text{-Spy})_4(\text{PA})_{20}\text{Cl}_2 + \text{mercaptosuccinic acid (MSA)} \rightarrow \text{Au}_{24}\text{Ag}_{20}\text{-MSA}$	138
		$[\text{Ag}_{141}\text{Br}_{12}(\text{S-Adm})_{40}]^{3+} + \text{Phenylacetylene (PAH)} \rightarrow [\text{Ag}_{141}\text{Br}_{12-n}\text{Cl}_n(\text{S-Adm})_{38}\text{PA}_2]^{3+}$	139
		$\text{Au}_x\text{Ag}_{44-x}(\text{PhC}\equiv\text{C})_m(\text{Ph}_3\text{P})_n\text{Cl}_p + \text{Tiopronin} \rightarrow \text{Au}_x\text{Ag}_{44-x}(\text{SR})_w\text{Cl}_z$	140
4.4	Addition of fluorescence properties	$\text{AuNC@DDAB} + \text{dihydrolipoic acid (DHLA)} \rightarrow \text{AuNC@DHLA}$	141
		$\text{AuNAC@Ag (NAC = N-acetylcysteine)} + \text{L-glutathione (GSH)} \rightarrow \text{AuNAC@AgSG}$	147

independent of the nature of the thiolate ligands and the presence of the protein. But the presence of positive residues at the protein C-terminal tail is critical for forming attractive intermolecular interactions with the carboxylate groups of the SG ligands, facilitating the adsorption of the protein cysteine on the gold cluster surface.

Because of the fast dynamics and complexity of the ligand exchange it is difficult to disclose the microscopic details of the $\text{S}_{\text{N}}2$ -like process by experimental alone. However, with the assistance of computational method such as DFT calculations,

some light could be shed on the $\text{S}_{\text{N}}2$ -like mechanism of initial ligand exchange. Interestingly, recently Wu and co-workers proposed an unimolecular nucleophilic substitution ($\text{S}_{\text{N}}1$)-like mechanism (in addition to a $\text{S}_{\text{N}}2$ -like process),⁸⁶ based on single crystal structure analysis of ligand exchanged clusters. The crystal structures showed that part of the sulfur atoms retained their configuration upon ligand exchange, which seems not possible for a $\text{S}_{\text{N}}2$ -like process. However, this argument only holds if the configuration at the sulfur atom is retained in solution, which was not addressed.



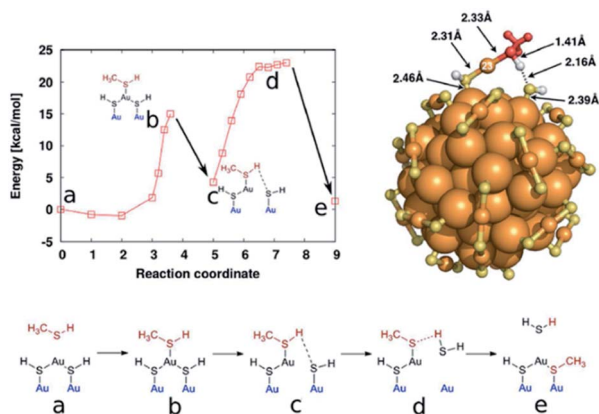


Fig. 7 Scheme of ligand exchange process with methane thiol and $\text{Au}_{102}(\text{SR})_{44}$ clusters. Top-left panel: energy behavior during exchange process (corresponding configuration were depicted as a sketch in the bottom panel); top-right panel: hemiring-like intermediate *c*. Configurations close to *b* and *d* have been confirmed to be at the local energy maximum by structural relaxations to the intermediate and final states. Adapted with permission from ref. 88. Copyright 2012 American Chemical Society.

Apart from the possibility for exchange of thiols *via* free ligand, as discussed above, there is a second mechanism for thiol exchange, which takes place through intercluster collisions. In 2013, Yoshiki Niihori and co-workers observed the intercluster LERs between $\text{Au}_{25}(\text{SC}_{10}\text{H}_{21})_{18}$ and $\text{Au}_{25}(\text{SC}_{12}\text{H}_{25})_{18}$ when they studied the influence of Pd atom doping of thiolate-protected Au_{25} nanoclusters on ligand exchange reactivity.⁸⁹ In that case the authors assumed that this exchange resulted from the detachment of ligand or gold–ligand species from the cluster.⁸⁹ Later, Salassa and co-workers carried out the intercluster LERs between $\text{Au}_{25}(\text{SBut})_{18}$ and $\text{Au}_{25}(\text{2-PET})_{18}$, and experimentally showed that the process is fast. As shown in Fig. 9A, 15 min after mixing the two clusters at room

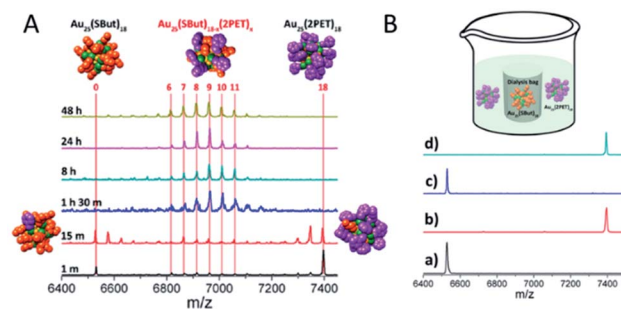


Fig. 9 (A) Positive-ion MALDI mass spectra of $\text{Au}_{25}(\text{SBut})_{18}$ and $\text{Au}_{25}(\text{2-PET})_{18}$ mixture collected at different time after mixing the two clusters; (B) experiment where $\text{Au}_{25}(\text{SBut})_{18}$ was put inside a dialysis membrane whereas $\text{Au}_{25}(\text{2-PET})_{18}$ was added outside the dialysis membrane. Positive-ion MALDI mass spectra of the solution inside/outside the dialysis membrane at $t = 0$ (a/b) and after 5 days (c/d). Adapted with permission from ref. 14. Copyright 2017 American Chemical Society.

temperature peaks from clusters with mixed ligand layer already appear in the mass spectrum.¹⁴ In addition, the interclusters ligand exchange seems to take place without release of thiol or thiol–gold complex into the solution, as illustrated in Fig. 9B. In this experiment $\text{Au}_{25}(\text{SBut})_{18}$ was put inside a dialysis membrane and thus the two clusters were physically separated, whereas low molecular weight species could still penetrate and pass the membrane. The MALDI mass spectra showed that the clusters did not undergo any exchange of thiols.¹⁴

Recently, we confirmed that bidentate thiol ligands can also exchange between clusters by observing the mass peak belonging to $\text{Au}_{25}(\text{2-PET})_{16}(\text{R-BINAS})_1$ ($\text{BINAS} = 1,1'$ -binaphthyl-2,2'-dithiol) after mixing $\text{Au}_{38}(\text{2-PET})_{24-2x}(\text{R-BINAS})_x$ (x corresponds to the number of BINAS ligands, which was about 2 on average in this experiment) and $\text{Au}_{25}(\text{2-PET})_{18}$ clusters.¹⁶ Compared to the exchange of monothiols higher temperatures are needed for the process to take place, 70 °C in this case.

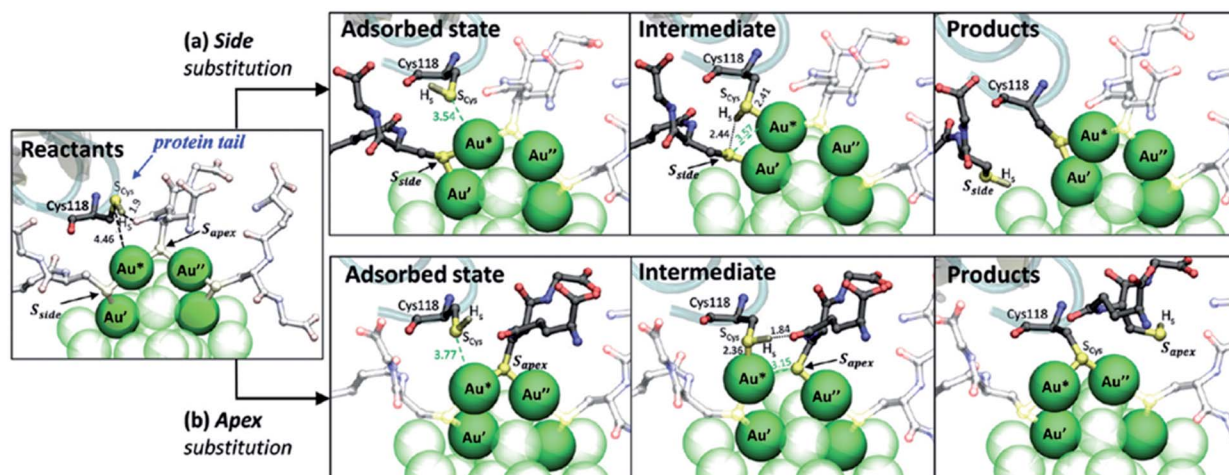


Fig. 8 Mechanism of the ligand exchange reaction in the $\text{scFv}\cdots\text{Au}_{25}(\text{SG})_{18}^-$ complex computed by QM/MM calculations. (a) Substitution at the side SG. (b) Substitution at the apex SG. For clarity, only three SG ligands of the dimeric staples are shown, and other SG groups and gold atoms far from the reaction centre are shown colorless or semitransparent. Part of the secondary structure of the protein is displayed, together with the nucleophilic cysteine. Water molecules are not shown. Adapted with permission from ref. 87. Copyright 2017 Royal Society of Chemistry.



However, compared with the free thiol exchange, the mechanism of intercluster ligand exchange is still rather unexplored and further investigations are needed.

Over the past decades, since the first report of ligand exchange reactions for gold nanoclusters, this reaction has been widely used to introduce new ligands to the parent nanoclusters,^{77,88,90–92} and to add functionalities or chemical properties to clusters as a post-modification method.^{87,93} This methodology strongly extends the possibilities to modify surface properties of gold nanoclusters, normally without changing the metal core.⁹³ Understanding the mechanism of the ligand exchange reaction may help us engineer the surface chemistry of nanomaterials in order to build multifunctional nano-platforms.

3.2 Site-selective ligand exchange reactions

Several reports show that during the ligand exchange reaction, many sites on the ligand shell exchange slowly or not at all, whereas others are comparatively reactive.^{85,94,95} These observations revealed that the ligand shell is heterogeneous and offers a diversity of ligand binding sites and the exchange reaction occurs preferentially at selected ones.⁸³ Before going into detail, it is essential to discuss the diversity sulfur groups in the ligand shell which represent the reactive site.

Since the dynamic exchange process were mostly investigated with $\text{Au}_{25}(\text{SR})_{18}$ and $\text{Au}_{38}(\text{SR})_{24}$,^{80,96} which are easily prepared and relatively stable, we focus some more on their crystal structure as illustrated in Fig. 10. The $\text{Au}_{25}(\text{SR})_{18}$ cluster (Fig. 10 left structure) has an icosahedral Au_{13} core surrounded by six SR-Au-SR-Au-SR staple units as we mentioned before.^{77,89,92,97–99} Because of the symmetry of $\text{Au}_{25}(\text{SR})_{18}$, six dimeric staples share the same chemical environment, but the SR groups in one staple can be divided into central –SR group, which is bound to the two gold atoms in the staple, and terminal –SR groups, which are linked to the gold core.¹⁰⁰ In contrast, $\text{Au}_{38}(\text{SR})_{24}$ consists of a biicosahedral Au_{23} core

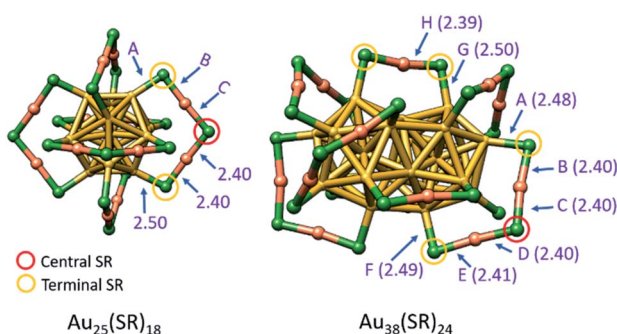


Fig. 10 Crystal structures of $\text{Au}_{25}(\text{SR})_{18}$ and $\text{Au}_{38}(\text{SR})_{24}$ clusters. Some terminal –SR (orange circle) and central –SR (red circle) are marked in the scheme. $\text{Au}_{25}(\text{SR})_{18}$ nanocluster has three possible ligand substitution sites marked as A, B, C, and $\text{Au}_{38}(\text{SR})_{24}$ has eight possible ligand substitution sites marked as A–H. Bond distances also are given in Å. Color code: Au core = gold, Au staple = orange, S = green. Adapted with permission from ref. 100 and ref. 102. Copyright 2015 & 2016 American Chemical Society.

(Fig. 10 right structure) and is covered not only by six dimeric units but also by three monomeric units. The structure of $\text{Au}_{38}(\text{SR})_{24}$ is elongated, with the three monomeric staples at the equator and the six dimeric staples at the two poles.¹⁰¹ The dimeric staples can be divided into central –SR group and terminal –SR groups. However, the two terminal –SR groups in the dimeric staples of $\text{Au}_{38}(\text{SR})_{24}$ have different chemical environment.^{80,102} Overall the 24 thiolate ligands are divided into four groups of symmetry unique sites with different chemical environment, which can give rise to preferential exchange sites during LERs.^{88,92}

From recent experimental and computational studies, it became clear that LERs between clusters and free monothioles in solution start preferentially at the terminal SR groups of $\text{Au}_{25}(\text{SR})_{18}$ and $\text{Au}_{38}(\text{SR})_{24}$ clusters,^{92,100,102} via an associative $\text{S}_{\text{N}}2$ -like mechanism.⁸⁸ This tendency was experimentally demonstrated by Yoshiki Niihori and co-workers.⁹² In this case the isomer distributions of the product after ligand exchange between $\text{Au}_{24}\text{Pd}(\text{SC}_2\text{H}_4\text{Ph})_{18}$ and $\text{C}_{12}\text{H}_{25}\text{SH}$ were determined by high resolution high-performance liquid chromatography. The quantitative evaluation of the expected coordination isomers and the products obtained by the reactions, showed that the exchange reaction starts at the thiolate which is bound to the core site. This also holds for exchange with other chalcogenate ligands such as HSePh or -HTePh.⁹⁹ Niihori and co-workers also found that Pd doping of the $\text{Au}_{25}(\text{SR})_{18}$ cluster drastically increased the ligand exchange rate. Pd doping reduces the number of valence electrons of the metal core, which facilitates the attack by the incoming ligand. Furthermore Pd doping induces the distortion of the cluster geometry.⁸⁹

In addition, the site preference of LERs can be investigated by computational methods. Aikens and co-workers employed density functional theory (DFT) to examine the ligand exchange on model $\text{Au}_{25}(\text{SH})_{18}^-$ and $\text{Au}_{38}(\text{SH})_{24}$ clusters with an incoming methanethiol.^{100,102} They calculated the intermediates and transition states, and predicted the barrier heights and reaction energies for this ligand exchange process. In Fig. 10, the different possible ligand substitution sites are marked on the crystal structure of $\text{Au}_{25}(\text{SR})_{18}$ (Fig. 10 left) and $\text{Au}_{38}(\text{SR})_{24}$ (Fig. 10 right) nanoclusters. The former cluster offers three different sites, whereas the latter has eight. The energies of the intermediates, transition state and products corresponding to ligand exchange at different sites determined by DFT calculation are summarized at Table 2. The results for Au_{25} indicated that the most favourable ligand exchange process is at site B, and the site C has highest energy barrier, which resulted in ligand exchange at the central SR group. This tendency was consistent with the one for $\text{Au}_{38}(\text{SR})_{24}$ as is evident from the calculated energies listed at Table 2. LERs at site C had higher energy barriers compared with other sites, meaning that the reaction between the staple gold atom and the sulfur atom of the central-SR units will proceed at a slower rate. Here, we should mention that each sulfur atom is participating in two bonds, for example Au_{25} A&B, Au_{38} C&D, and Au_{38} E&F, which represent different reactive sites, however, those two sites will lead to the same product. Above predictions from computational studies on $\text{Au}_{25}(\text{SH})_{18}^-$ and $\text{Au}_{38}(\text{SR})_{24}$ indicated that the



Table 2 Summary of energies (in eV) for ligand exchange extracted from ref. 100 and ^{102a}

	TS1	Int	TS2	Products
Au ₂₅ A	0.56	0.19	0.76	-0.01
Au ₂₅ B	0.62	0.29	0.65	0.05
Au ₂₅ C	0.78	0.35	1.15	-0.04
Au ₃₈ A	0.68	0.22	0.80	-0.06
Au ₃₈ B	0.61	0.29	0.76	-0.07
Au ₃₈ C	0.85	0.21	0.85	-0.05
Au ₃₈ D	0.59	0.34	0.89	-0.11
Au ₃₈ E	0.59	0.21	0.80	-0.04
Au ₃₈ F	0.75	0.06	0.56	-0.04
Au ₃₈ G	0.54	0.21	0.65	-0.05
Au ₃₈ H	0.91	0.15	0.77	-0.05

^a Energies are relative to the original cluster and free ligand.

exchange is preferred at the terminal SR groups, which is congruous with the experimental results.^{88,92} However, this might not always be the case as shown by Zhu and co-workers. After the ligand exchange reaction between Au₂₄Cd(SCH₂Ph)₁₈ and SCH₂Ph-^tBu Au₂₄Cd(SCH₂Ph-^tBu)₁₇(SCH₂Ph)₁ has been obtained.⁸⁶ The integrated area of the peaks extracted from ¹H NMR spectra of this species illustrated that the remaining parent ligand SCH₂Ph was equally distributed among the terminal and central sulfur atoms, in contrast with the results discussed above. This result may be caused by several reasons, such as steric hindrance within the ligand shell or the inevitable migration of the ligand on the cluster surface at 80 °C. In addition, Pengo and co-workers also showed by NMR spectroscopy that the distribution of incoming ligand on the central and terminal sites depends on the properties of the ligand.⁹⁸

Apart from the monothiolate ligands, nanoclusters are also reactive towards dithiol ligands. Amala Dass and co-workers systematic studied LERs with aliphatic dithiol ligands of various chain length, HS-(CH₂)_n-SH.¹⁰⁴ They documented that C₃ and C₄ prefer interstaple coupling, and C₅ and C₆ are good candidates for intrastaple binding, whereas the length of C₂ ligand is not enough for the bidentate binding. Aromatic dithiols have also been used for the LERs. However, they are sterically more demanding and more rigid compared to aliphatic thiols. One example is 1,1'-binaphthalene-2,2'-dithiol (BINAS), which is a chiral rigid dithiol ligand (Fig. 11 left), used to introduce chirality to achiral clusters.^{77,105} Several studies provide insight into the LERs on BINAS with Au₂₅ and Au₃₈ clusters,^{77,103,105-108} and from the experimental and computational results it emerges that the bidentate ligand connects two neighboring staples by interstaple coupling.^{103,105,108} Sels and co-workers reported the isolation of different exchange products and isomers of clusters containing one or two BINAS adsorbed PdAu₂₄(2-PET)₁₈ clusters.¹⁰⁸ The investigated structure of the PdAu₂₄(SR)₁₆(S-BINAS)₁ cluster is shown in Fig. 11, which indicates the two different sulfur environments of the cluster and marks some bond distances.¹⁰³ As displayed in Fig. 11, there are several possible binding sites for BINAS at the ligand shell, illustrated with curved lines. However, considering the distances between the two S-atoms in undistorted BINAS, 4.1 Å,



Fig. 11 3D sketch of S-BINAS (left) and MAu₂₄(SR)₁₆(S-BINAS)₁ (right), where M = Pt, Pd. Other ligands apart from BINAS are omitted for clarity. The two types of sulfur environments are indicated with S_{core} (also called terminal) and S_{apex} (also called central). Color code: Au = yellow, S = green, C = beige, H = white, Pt and Pd = blue. Adapted with permission from ref. 103. Copyright 2017 American Chemical Society.

the interstaple binding site between a S_{apex} and a S_{core} sulfur atom has the appropriate distance, 4.05 Å. Based on X-ray absorption spectroscopy (XAS), the authors also showed that the BINAS interstaple binding mode does not perturb the cluster structure.¹⁰³

Compared with the monothiolate ligand exchange it may be easier to isolate isomers of clusters with mixed ligand shell when using dithiolate ligands. A better understanding of the preferred reactive sites of the LERs will enable the rational design of mixed ligand shell clusters.

3.3 Comparison of ligand exchange reactions of clusters in solution and on supports

LERs is a process that takes advantage of the dynamic nature of the thiol-gold surface,⁷² and in most cases it is performed with clusters in solution. As a further development, Truttmann and co-workers recently reported the first successful ligand exchange reaction on supported (immobilized) Au₁₁ nanoclusters.¹⁰⁹ In this case, Au₁₁(PPh₃)₇Cl₃ clusters were deposited on Al₂O₃, and then the dropcasts sample was exposed to a solution of thiol ligand. In such a procedure cluster-ligand combinations are possible that are not accessible through normal LERs due to solvent incompatibility. Indeed, in the study mentioned above the hydrophilic thiol ligand, L-glutathione, and the hydrophobic 2-PET (2-phenylethanethiol) ligand were used. According to previous reports, when performing ligand exchange with Au₁₁(PPh₃)₇Cl₃ and GSH in solution, the Au₁₁ clusters grow to form Au₂₅.^{26,110} Here, in contrast, no cluster growth has been observed as indicated from MALDI-MS after ligand exchange reaction on supported clusters. As evidenced by IR (PM-IRRAS, ATR-IR) and NMR spectroscopy, partial ligand exchange took place and the halide ligands of the parent nanocluster were primarily exchanged with thiolates.

In contrast to the ligand exchange in solution, the reaction on the supported cluster did not induce a change of the core size of the nanoclusters. However, due to the hindered



accessibility of the ligand on the support side of the cluster, the number of exchanged ligands was relatively low compared to typical solution phase ligand exchange reactions. In addition, similar to the functionalization in solution, fluorescence could be introduced to supported clusters by reacting them with fluorescein probe, which also provides new insight for use of the clusters in biological applications.¹⁰⁹

4. Functionalization of gold nanoclusters induced by LERs

4.1 Chirality induced by LERs

Chirality is a geometric property of objects, which widely exists in nature from molecules over proteins^{111,112} to even larger structures. During the past decade, with the use of X-ray single-crystal diffraction, it has emerged that chirality is a ubiquitous property for gold nanoclusters. Because of the potential applications in sensing, catalysis, molecular recognition and so on, chiral gold nanoclusters have attracted a lot of interest.^{113–115}

The reported chiral gold nanoclusters can be categorized into three types: (i) chiral Au–S framework with achiral ligands, (ii) achiral Au–S framework with chiral ligands,^{114,115} and (iii) a combination of the two (chiral Au–S framework and chiral ligand). The type I chiral nanoclusters are sometimes also called intrinsically chiral nanoclusters, and some examples are Au₂₀(SR)₁₆, Au₂₈(SR)₂₀, Au₃₈(SR)₂₄, Au₁₀₂(SR)₄₄ and Au₁₃₃(SR)₅₂, in which all the different R groups are achiral. The chirality of type II nanoclusters is due to the ligands SR. Fundamentally, such chiral clusters can be prepared by direct synthesis using a chiral thiol.¹¹⁶ However, resulting from the solubility and steric effect of different ligands, direct synthesis of nanoclusters with some ligands was unsuccessful.⁹³ Alternatively, ligand exchange is a good method to incorporate chirality or other functionality onto the gold nanoclusters.

By using ligand exchange on achiral Au_m(SR)_n with chiral thiols (SR*), a series of chiral Au_m(SR)_{n-x}(SR*)_x clusters with chiroptical activity have become available.^{15,107,117,119} For example, Si and co-workers reported the ligand exchange on [Au₂₅(PET)₁₈]⁻[TOA⁺] (in the following named as Au₂₅) with R/S-



Fig. 12 (A) CD spectra of Au₂₅ clusters before exchange with R/S-BINAS (a), after exchange with R-BINAS (b), and after exchange with S-BINAS (c); (B) UV-vis absorption spectra of the corresponding samples. Adapted with permission from ref. 107. Copyright 2009 American Chemical Society.

BINAS.¹⁰⁷ As illustrated in Fig. 12, the CD (left panel) and UV-Vis absorption (right panel) spectra of Au₂₅ clusters were recorded before (a) and after exchange with R/S-BINAS (b/c). As expected, the Au₂₅ cluster is optically inactive before ligand exchange. Once the cluster exchange with BINAS, intense bands at 305, 352, and 390 nm are observed in the CD spectra with opposite sign for clusters covered by the two enantiomers of BINAS. The CD signals refer to the chiral ligand but the spectra are different compared with the one of free BINAS. Furthermore, Fig. 12 reveals that the absorption spectrum of the cluster became less defined after exchange with BINAS. Apart from Au₂₅ clusters, similar exchange with R/S-BINAS also takes place on Au₃₈ and Au₄₀ clusters.¹⁰⁶ In addition, as we mentioned before, the introduction of BINAS ligand on Au₃₈ clusters stabilized the structure against inversion (enantiomerization), probably due to the reduced flexibility of the gold–sulfur interface.⁷⁷ Also, different from the LERs with monothiols, exchange with dithiol ligand reduces the number of isomers. Due to the reduction of the reaction rate after the first exchange, it is also relatively easy to obtain species with just one ligand exchanged.^{77,105}

Compared with the direct synthesis, the ligand exchange of nanoclusters may lead to a heterogeneous system with a distribution of the number of exchanged ligands when the exchange is incomplete. In order to obtain completely exchanged nanoclusters, large excess of free incoming ligand with respect to the clusters and/or repetition of the exchange reaction are necessary, with purification of the intermediate cluster after each step.

The optical activity is usually considerably stronger for the type I chiral clusters, *i.e.* clusters with intrinsic chirality in their Au–S framework, compared to type II chiral clusters, *i.e.* achiral Au–S framework with chiral ligands. The absorption spectrum of a given cluster is roughly ligand independent. In contrast,

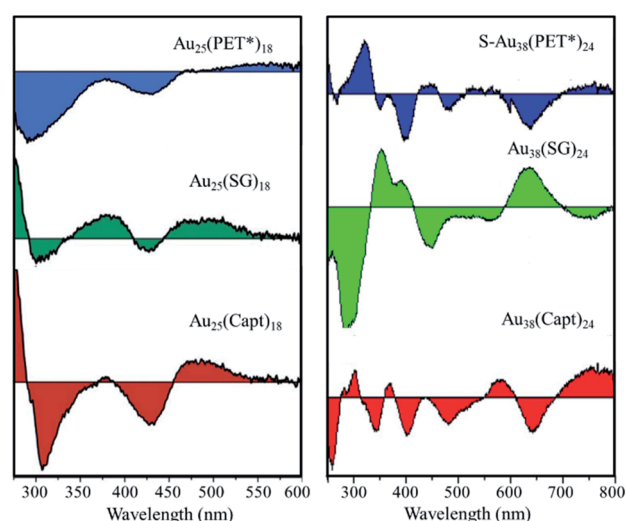


Fig. 13 Effect of the chiral ligand type on the CD spectra of Au₂₅(SR*)₁₈ and Au₃₈(SR*)₂₄. PET* = S-phenylpropane-1-thiol, SG = glutathione, Capt = captopril. Left panel: adapted with permission from ref. 117. Copyright 2012 Royal Society of Chemistry. Right panel: adapted with permission from ref. 118. Copyright 2014 Wiley-VCH.



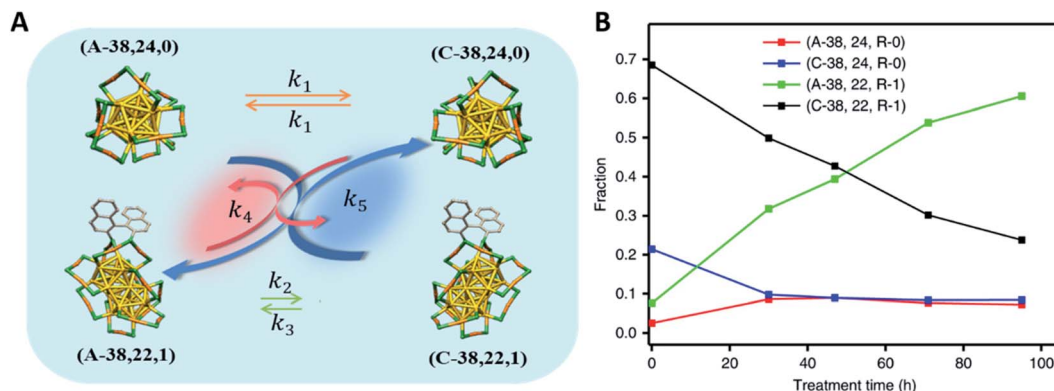


Fig. 14 Amplification of the enantiomeric excess after introducing chiral R-BINAS to the Au₃₈ nanoclusters. (A) Scheme for the dynamic inversion system, (B) evolution of different clusters species under 70 °C as a function of time. The nomenclature of the four different species assigned as follow: (A-38, 24, R-0) corresponded to A-Au₃₈(2-PET)₂₄; Adapted with permission from ref. 16. Copyright 2020 Nature Research.

when the chirality of the nanoclusters is due to the ligand, the chiroptical properties will be ligand-dependent,¹⁰⁷ which offer various possibilities to influence the optical response. Fig. 13 shows circular dichroism (CD) spectra of Au₂₅ and Au₃₈ clusters stabilized with three different chiral R* groups. All these chiral Au₂₅(SR*)₁₈ nanoclusters reveal the typical UV/Vis absorption spectrum of Au₂₅, but their CD signals show differences depending on the R* groups (Fig. 13 left panel).¹¹⁷ A similar phenomenon was also observed for Au₃₈(SR*)₂₄ nanoclusters with different chiral thiolate ligands (Fig. 13 right panel).¹¹⁸ This ligand-dependence provides a possibility to engineer the optical characteristics of a given gold nanocluster.

In addition, the chiral ligand R-BINAS was also used to amplify the enantiomeric excess of Au₃₈(SR)₂₄ at 70 °C.¹⁶ In a mixture containing the two enantiomers of the Au₃₈ cluster and the corresponding clusters containing one R-BINAS in their ligand shell (diastereomers), the fraction of anticlockwise (A) clusters increases with time at the expense of clockwise (C) clusters (Fig. 14B). For the experiment shown in Fig. 14B the initial sample was prepared with high fraction of C-Au₃₈. The sample was then heated to 70 °C and the HPL chromatograms were recorded as a function of time. The fractions of different species were determined from the HPL chromatograph and plotted as function as time (Fig. 14B), showing the increase of A-Au₃₈. The proposed mechanism of the process is shown in Fig. 14A. This dynamic inversion is due to the diastereoselective intercluster exchange of R-BINAS between chiral clusters and the fast racemization of Au₃₈(2-PET)₂₄. This example shows that the dynamic nature of these clusters can be used as a benefit. Overall, the ligand exchange reaction with chiral ligands will continue to add new impetus to the field of chiral nanomaterials and its applications.

4.2 LERs induced size transformation of clusters

LERs is a process that takes advantage of the dynamic nature of the thiol-gold interface,⁷² and in most cases it does not cause any change in size or structure of the cluster, only replacing one capping ligand by another one.⁹⁴ During the past decade, due to the development of precise synthesis methods and structure

determination by X-ray single-crystal diffraction numerous new nanoclusters were reported. LERs became an important new methodology for controlling the size and structure of nanoclusters, and the process was named ligand-exchange-induced size/structure transformation (LEIST for short), which was first proposed by the Jin's group.¹²⁰

Up to now, most of the reported LEIST work focuses on PPh₃-stabilized^{121,122} and thiolate-protected Au nanoclusters.^{123–125} However, the driving forces for LEIST is quite different for these two kinds of Au nanoclusters.¹² First, for LEIST of PPh₃-stabilized Au nanoclusters, exchanging the phosphine ligands by thiolate ligands changes the surface bonding completely (from Au–P to Au–S). The different coordination modes, Au–P vs. Au–S, lead to different surface motifs and drives the structural transformation of the cluster.^{12,121} Second, for LEIST of thiolate-protected Au nanoclusters, the new thiolates would not significantly alter the gold–ligand coordination; however, due to the distinctly different physical–chemical properties (*e.g.*, size, rigidity, bulkiness, interactions *etc.*) of different thiolate ligands, the original structures may not be the most stable for the ligand-exchanged nanoclusters, and thus the structural transformation takes place.¹² In the following we mainly focus on the LEIST on thiolate-protected Au nanoclusters. Depending on the size or structural change of the thiolate-protected Au nanoclusters resulting from the LEIST, this process been classified into three groups.

(I) Transformation between structural isomers without size change. For example, Au₂₈(SR)₂₀ nanocluster reversibly changes its structure upon ligand exchange between R = *c*-C₆H₁₁ and R = Ph^tBu at elevated temperatures (*e.g.*, 80 °C) as shown in Fig. 15.⁵⁹ The structures of these two nanoclusters are remarkable different. Au₂₈(SPh^tBu)₂₀ contains a FCC Au₂₀ kernel capped by four Au₂(SR)₃ staple motifs and eight bridging thiolates, whereas Au₂₈(S-*c*-C₆H₁₁)₂₀ adopts a more loose structure with a FCC Au₂₀ kernel plus two Au₁(SR)₂, two Au₃(SR)₄, and eight bridging SR staple-like structures. DFT calculations revealed that the origin of reversible isomerization lay in the thiolate ligand's carbon tail structure, which was found to dictate the



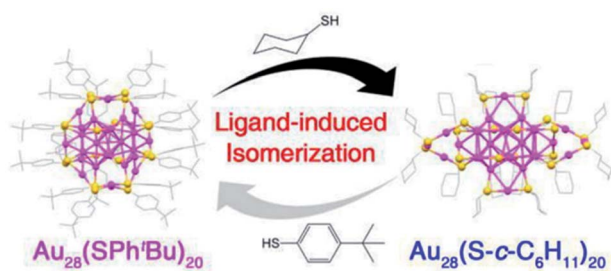


Fig. 15 Ligand-exchange induced transformation between $\text{Au}_{28}(\text{SPh}^t\text{Bu})_{20}$ and $\text{Au}_{28}(\text{S}-c\text{-C}_6\text{H}_{11})_{20}$ nanoclusters. Color codes: purple sphere, Au; yellow sphere, S; gray sphere, C. For clarity, all H atoms are omitted. Adapted with permission from ref. 59. Copyright 2016 American Chemistry Society.

isomer's stability.⁵⁹ In LEIST the isomerization of clusters is rare.

(II) Transformations from a smaller to a larger nanocluster.^{26,58,61,110,125–127} An interesting example is the transformation of $\text{Au}_{25}(\text{PET})_{18}$ into $\text{Au}_{28}(\text{TBBT})_{20}$.⁵⁸ $\text{Au}_{25}(\text{PET})_{18}$ is probably the most studied thiolate-protected nanocluster, due to its prototypical character. At 80 °C and in large excess of TBBT the $\text{Au}_{25}(\text{PET})_{18}$ cluster is transformed into $\text{Au}_{28}(\text{TBBT})_{20}$. It is worth to mention that the resulting $\text{Au}_{28}(\text{TBBT})_{20}$ cluster is chiral, and the origin of chirality is primarily rooted in the rotating arrangement of the four dimeric staples as well as the arrangement of the bridging thiolates as we mentioned before. Moreover, the pair of enantiomers of $\text{Au}_{28}(\text{TBBT})_{20}$ can be separated by chiral-HPLC.⁵⁸ Another example is the transformation of Au_{11} to Au_{25} upon ligand exchange with GSH.^{26,110}

(III) Transformations from a larger to a smaller nanocluster.^{12,64,120,123–125,128,129} For instance, highly stable

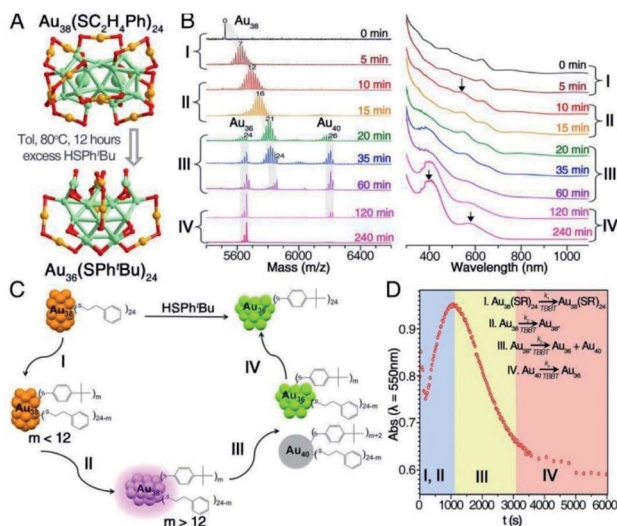


Fig. 16 (A) Ligand-exchange induced transformation from $\text{Au}_{38}(\text{SC}_2\text{H}_4\text{Ph})_{24}$ to $\text{Au}_{36}(\text{SPh}^t\text{Bu})_{24}$. (B) Time-dependent ESI-MS and UV-vis spectra of the transformation. (C) Reaction pathway for the transformation. (D) Kinetics (monitored by absorbance at 550 nm) for the transformation. Adapted with permission from ref. 120. Copyright 2013 American Chemical Society.

$\text{Au}_{144}(\text{SC}_2\text{H}_4\text{Ph})_{60}$ reacted with thiophenol, HSPH, to form a different 99 atom cluster species $\text{Au}_{99}(\text{SPh})_{42}$.¹²⁸ In addition, Zeng and co-worker reported that, by LEIST, the very stable and widely investigated $\text{Au}_{38}(\text{SC}_2\text{H}_4\text{Ph})_{24}$ cluster was transformed to $\text{Au}_{36}(\text{SPh}^t\text{Bu})_{24}$ as shown in Fig. 16A.¹²⁰ To unravel details of the intriguing one-size-to-another size transformation, Zeng carried out time-dependent mass spectrometry and optical spectroscopy analyses (Fig. 16B) and found a remarkable disproportionation in the transformation of rod-like biicosahedral $\text{Au}_{38}(\text{SC}_2\text{H}_4\text{Ph})_{24}$ to tetrahedral $\text{Au}_{36}(\text{SPh}^t\text{Bu})_{24}$. From the evolution of the mass spectra and corresponding UV curves, the reaction pathway can be roughly divided into four stages (Fig. 16C): (i) (0–5 min) ligand exchange reaction occurs between PET and TBBT; (ii) (10–15 min) ligand exchange reaction continues leading to TBBT-triggered structural distortion of Au_{38} with an optical feature at 550 nm (Fig. 16D); (iii) (20–60 min) critical stage for the disproportionation of Au_{38} to Au_{36} and Au_{40} , and (iv) size conversion of Au_{40} to Au_{36} evidenced by the decrease and complete disappearance of the Au_{40} peak in time-dependent mass spectra. Recently, other LEIST, such as transformations from $\text{Au}_{23}(\text{S}-c\text{-C}_6\text{H}_{11})_{16}$ to $\text{Au}_{36}(\text{TBBT})_{24}$ and from $\text{Au}_{329}(\text{PET})_{84}$ to $\text{Au}_{279}(\text{SPh}^t\text{Bu})_{84}$ also revealed similar pathways but without disproportionation. In these cases the conversion through ligand exchange followed by the size focusing ultimately lead to size growth.^{127,129}

Apart from the irreversible transformation mentioned above, Amala Dass and co-workers presented the first reversible interconversion between two nanomolecules $\text{Au}_{36}(\text{SPhX})_{24}$, (where $\text{X} = -\text{H}$ or ^tBu) and $\text{Au}_{30}(\text{S}^t\text{Bu})_{18}$ as illustrated in Fig. 17.^{12,125} In this case, the gold core converted between bicuboctahedral Au_{20} and 4-fused cuboctahedron Au_{24} , and the staple arrangement of these two cluster surfacers was also different. More interesting, the reversible conversion easily takes place under the same thermochemical conditions with different thiol ligands. Later, Wu and co-workers achieved the interconversion among $\text{Au}_{44}(\text{TBBT})_{28}$, $\text{Au}_{44}(2,4\text{-DMBT})_{26}$, and $\text{Au}_{43}(\text{S}-c\text{-C}_6\text{H}_{11})_{25}$ nanoclusters by the ligand exchange,^{130,131} and they also investigated the thermostability of these nanoclusters. As monitored by time-dependent optical absorption, the $\text{Au}_{44}(2,4\text{-DMBT})_{26}$ cluster was less thermostable than $\text{Au}_{44}(\text{TBBT})_{28}$, and much more stable than $\text{Au}_{43}(\text{S}-c\text{-C}_6\text{H}_{11})_{25}$ at 80 °C.¹³⁰ The interconversion not only offer many possibilities to obtain new nanoclusters but also leads to valuable insight into the inherent influence of the ligand on the composition and atomic structure of thiolate-protected gold clusters.

Focussing on the experimental conditions of the LEIST, mentioned above, it seems that the conditions necessary for LEIST to take place are large excess of incoming ligand, normally more than 100 times compared to the endogenous ligand, and elevated temperatures.¹²⁴ In addition, it was argued that the incoming thiol must be significantly different from the endogenous thiol for the transformation to happen.⁶¹ However, we reported a new transformation of $\text{Au}_{25}(\text{SR})_{18}$ into $\text{Au}_{28}(\text{SR})_{21}$ by LER with a chiral ligand (R- or S-phenylpropane-1-thiol) at mild conditions, *i.e.* at room temperature and with low thiol excess (with incoming to outgoing ligand molar ratio 2 : 1).¹⁵ In this case, the van der Waals interactions within the ligand shell and





Fig. 17 Schematic of the conditions for the interconversion between $\text{Au}_{30}(\text{S}^t\text{Bu})_{18}$ and $\text{Au}_{36}(\text{SPhX})_{24}$. Reaction take place under the same thermochemical conditions with different thiol ligands (thiophenol and *tert*-butyl). Left panel: crystal structure of $\text{Au}_{30}(\text{S}^t\text{Bu})_{18}$, right panel: crystal structure of $\text{Au}_{36}(\text{SPhX})_{24}$. Color code: Au core atom: red, Au staple atom: green, sulphur: yellow, carbon: gray. Hydrogen atoms has been eliminated for clarity. Adapted with permission from ref. 125. Copyright 2017 American Chemistry Society.

the “bulkiness” of the methyl are the main factors driving this process. In summary, LEIST provides new and exciting avenues to explore novel atom-precise nanoclusters and opens a new strategy to investigate the size growth of nanoclusters.

4.3 LERs induced organic/aqueous phase transfer of clusters

There are many types of ligands that can be used to stabilize nanoparticles or nanoclusters, as shown in Fig. 18.¹³² Depending on the solubility of the surface ligand, one can distinguish hydrophobic and hydrophilic clusters. However, for some (biological) applications, the hydrophobic clusters need to be dissolved in aqueous solutions. Also hydrophilic clusters sometimes need to be made accessible to reactions in organic

solvent. In this situation, phase transfer will be the first option to relocate the clusters to the desired phase.

In general, for phase transfer processes, reagents such as tetraoctylammonium bromide,^{133,134} (–)-1*R*,2*S*-*N*-dodecyl-*N*-methylephedrinium bromide¹³⁵ and others have been successfully used. These chemical reagents form an additional molecular layer on the clusters and thus change their surface properties. This strategy was mainly used for the phase transfer from aqueous to organic phase.^{133–137} In addition to this strategy, ligand exchange has also been utilized as an important strategy for the phase transfer. For instance, when hydrophobic $\text{Au}_{11}(\text{PPh}_3)_7\text{Cl}_3$ reacted with GSH ligand in solution, the Au_{11} clusters grew to form water-soluble Au_{25} nanoclusters,^{26,110} which is a good evidence for ligand exchange. In addition, Zheng and co-workers reported the crystal structure of intermetallic nanocluster $\text{Au}_{24}\text{Ag}_{20}(\text{2-SPy})_4(\text{PA})_{20}\text{Cl}_2$ and also investigated the phase transfer process after performing ligand exchange with mercaptosuccinic acid (MSA),¹³⁸ leading to transfer of the $\text{Au}_{24}\text{Ag}_{20}$ cluster from DCM to water phase. Other reported cases of phase transfer induced by ligand exchange reactions, include ligands such as (phenylacetylene) PAH¹³⁹ and Tiopronin.¹⁴⁰ The latter ligand also introduced cancer therapy functions to the cluster.

It emerges from the cases mentioned above, that the phase transfer from aqueous to organic phase relies on phase transfer reagents or ligand modification, and in many cases the core size of the cluster is maintained. However, the inverse process, transfer from organic to aqueous phase, mostly relies on the ligand exchange reactions. Furthermore, the phosphine-stabilized clusters are more prone to be transferred to another phase after reaction with thiol ligands. For the choice of the ligand used for the phase transfer, three considerations have to be made: (i) the affinity to the metal core of new ligand compared with the original one; (ii) the solubility of the new ligand in the target solvent, and (iii) the capability of the ligand to maintain the core size of the cluster after phase transfer.

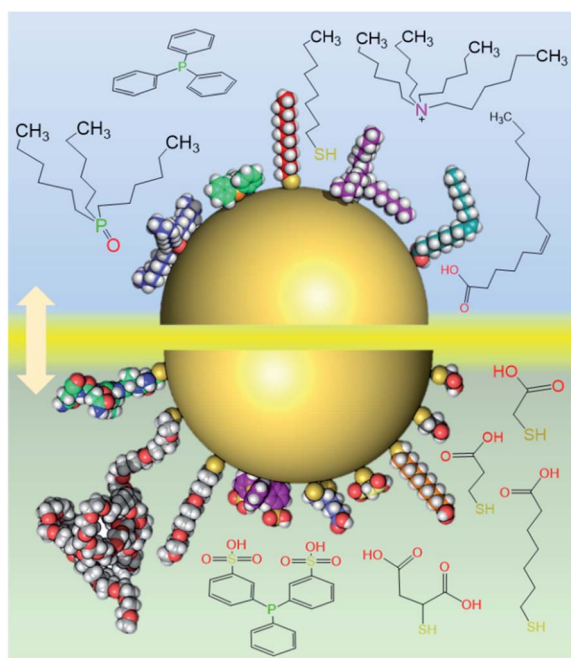


Fig. 18 A nanoparticle (nanocluster) stabilized with different hydrophobic (top panel) and hydrophilic (bottom panel) ligand molecules (skeletal drawings or space filling models were used to represent the ligands). Adapted with permission from ref. 132. Copyright 2010 Royal Society.

4.4 Florescence induced by LERs

Fluorescent gold nanoclusters have been widely used for biological applications such as cell identification, to study interaction, differentiation, and tracking.^{109,142–145} Compared with the



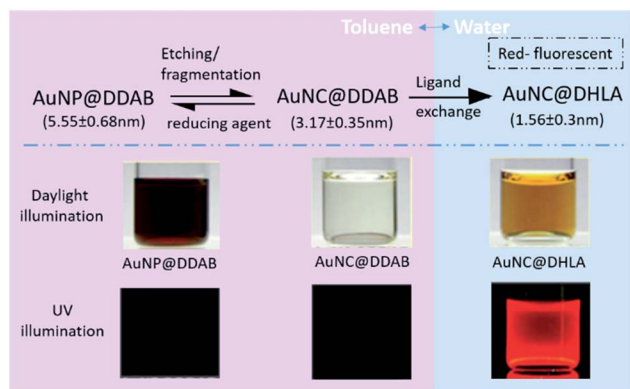


Fig. 19 Strategy to fabricate hydrophilic fluorescent Au nanoclusters. Size distribution of three different Au nanoclusters extracted from 100 particles. Pictures of particle solutions under daylight and UV excitation illustrated respectively. Adapted with permission from ref. 141. Copyright 2009 American Chemical Society.

more traditional QDs, fluorescent gold nanoclusters have decent quantum yield, excellent biocompatibility, good photostability, and lower cytotoxicity. The clusters had negligible influence on the cell viability at the considered dose.¹⁴⁶ However, because of the synthesis method and the solubility of ligands, the diversity of the fluorescent gold nanoclusters has been limited.

LER is a very efficient strategy for preparing fluorescent gold nanoclusters. As one of the most intuitive cases, the non-fluorescent gold nanoclusters (AuNC@DDAB) can be converted to brightly red emitting nanoclusters (AuNC@DHLA) through an elegant ligand exchange reaction with dihydrolipoic acid (DHLA) as shown in Fig. 19.¹⁴¹ In this system, the DDAB-stabilized gold nanoparticles (AuNP@DDAB) are etched by the addition of Au precursors (HAuCl₄ or AuCl₃) to smaller nanoclusters (AuNC@DDAB), and the organic soluble and hydrophobic AuNC@DDAB become water soluble upon ligand exchange with dihydrolipoic acid (AuNC@DHLA). The AuNP@DDAB solution shows red color due to surface plasmon absorption, which is absent to AuNC@DDAB and AuNC@DHLA. More interesting, after ligand exchange with DHLA, the AuNC@DHLA solution shows strong red photoluminescence as shown in the Fig. 19. Here, the phase transfer and addition of fluorophore has been achieved together by the LERs.

In addition, Xu *et al.* reported ligand exchange by GSH and *N*-acetylcysteine (NAC) on AuNAC@Ag. The products showed maximum 20-fold fluorescence enhancement.¹⁴⁷ During this ligand exchange process, silver ions and GSH have synergistic effects and the PL enhancement was found to be proportional to the concentration of GSH. This fluorescence enhancement was also used for selective imaging of intracellular glutathione.¹⁴⁷ Besides dissolved clusters, the fluorescence can also be introduced to supported nanoclusters by using ligand exchange on the immobilized sample, as we mentioned above.¹⁰⁹

5. Conclusions

In conclusion, ligand exchange reaction plays an important role in the field of nanoclusters. As one of the post-synthesis

methods for the modification and functionalization of the nanoclusters, the process is enabled by the flexibility of the gold-sulfur interface. Experimental and computational investigations show that LERs between clusters and free monothiolates in solution starts preferably at the terminal SR groups of the staple motifs, which are linked to the gold core, *via* an associative S_N2-like mechanism. The process takes place in solution phase but also on immobilized Au nanoclusters. Understanding of the preferred reactive sites of the process will help us engineer precise clusters with mixed ligand shell. Compared with the monothiolate ligand exchange, dithiolate ligands may offer an easier way to obtain precise clusters with mixed ligand shell. Intercluster ligand exchange is also an important property of thiolate-protected metal clusters and has to be considered whenever clusters with different ligands are in the same solution. Despite the considerable effort made, the microscopic details of intercluster ligand exchange remain obscure.

Ligand exchange reactions are in general easy to perform and offer a great potential to introduce or amplify properties of the clusters, such as introduction of chirality to achiral nanoclusters and the amplification of optical activity and enantiomeric excess, size transformation of the cluster, phase transfer, and the addition of fluorescent groups. Thus, ligand exchange reactions will continue to play an important role in the future and a better understanding of the process will further increase the potential of this method.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

TB acknowledges the generous support of the Swiss National Science Foundation (grant 200020_192232) and the University of Geneva. YW thanks the China Scholarship Council fellowship (201706450070).

References

- 1 R. Jin, *Nanoscale*, 2010, **2**, 343–362.
- 2 R. Jin, H. Qian, Z. Wu, Y. Zhu, M. Zhu, A. Mohanty and N. Garg, *J. Phys. Chem. Lett.*, 2010, **1**, 2903–2910.
- 3 H. Qian, M. Zhu, Z. Wu and R. Jin, *Acc. Chem. Res.*, 2012, **45**, 1470–1479.
- 4 R. Jin, C. Zeng, M. Zhou and Y. Chen, *Chem. Rev.*, 2016, **116**, 10346–10413.
- 5 I. Chakraborty and T. Pradeep, *Chem. Rev.*, 2017, **117**, 8208–8271.
- 6 A. Ghosh, O. F. Mohammed and O. M. Bakr, *Acc. Chem. Res.*, 2018, **51**, 3094–3103.
- 7 X. Kang, Y. Li, M. Zhu and R. Jin, *Chem. Soc. Rev.*, 2020, **49**, 6443–6514.
- 8 X.-R. Song, N. Goswami, H.-H. Yang and J. Xie, *Analyst*, 2016, **141**, 3126–3140.
- 9 S. Maity, D. Bain and A. Patra, *Nanoscale*, 2019, **11**, 22685–22723.



- 10 R. Dinkel, W. Peukert and B. Braunschweig, *J. Phys.: Condens. Matter*, 2017, **29**, 133002.
- 11 M.-M. Xu, Q. Chen, L.-H. Xie and J.-R. Li, *Coord. Chem. Rev.*, 2020, **421**, 213421.
- 12 X. Kang and M. Zhu, *Chem. Mater.*, 2019, **31**, 9939–9969.
- 13 Y. Niihori, S. Hossain, B. Kumar, L. V. Nair, W. Kurashige and Y. Negishi, *APL Mater.*, 2017, **5**, 053201.
- 14 G. Salassa, A. Sels, F. Mancin and T. Bürgi, *ACS Nano*, 2017, **11**, 12609–12614.
- 15 Y. Wang, B. Nieto-Ortega and T. Bürgi, *Chem. Commun.*, 2019, **55**, 14914–14917.
- 16 Y. Wang, B. Nieto-Ortega and T. Bürgi, *Nat. Commun.*, 2020, **11**, 4562.
- 17 R. Jin, *Nanoscale*, 2015, **7**, 1549–1565.
- 18 T. Tsukuda, *Bull. Chem. Soc. Jpn.*, 2012, **85**, 151–168.
- 19 C. M. Aikens, *Acc. Chem. Res.*, 2018, **51**, 3065–3073.
- 20 M. Brust, M. Walker, D. Bethell, D. J. Schiffrin and R. Whyman, *J. Chem. Soc., Chem. Commun.*, 1994, 801–802, DOI: 10.1039/C39940000801.
- 21 T. G. Schaaff and R. L. Whetten, *J. Phys. Chem. B*, 2000, **104**, 2630–2641.
- 22 R. L. Whetten, J. T. Khoury, M. M. Alvarez, S. Murthy, I. Vezmar, Z. L. Wang, P. W. Stephens, C. L. Cleveland, W. D. Luedtke and U. Landman, *Adv. Mater.*, 1996, **8**, 428–433.
- 23 T. G. Schaaff, M. N. Shafiqullin, J. T. Khoury, I. Vezmar, R. L. Whetten, W. G. Cullen, P. N. First, C. Gutiérrez-Wing, J. Ascensio and M. J. Jose-Yacamán, *J. Phys. Chem. B*, 1997, **101**, 7885–7891.
- 24 R. L. Donkers, D. Lee and R. W. Murray, *Langmuir*, 2004, **20**, 1945–1952.
- 25 J. B. Tracy, M. C. Crowe, J. F. Parker, O. Hampe, C. A. Fields-Zinna, A. Dass and R. W. Murray, *J. Am. Chem. Soc.*, 2007, **129**, 16209–16215.
- 26 Y. Shichibu, Y. Negishi, T. Tsukuda and T. Teranishi, *J. Am. Chem. Soc.*, 2005, **127**, 13464–13465.
- 27 Y. Negishi, K. Nobusada and T. Tsukuda, *J. Am. Chem. Soc.*, 2005, **127**, 5261–5270.
- 28 H. Tsunoyama, P. Nickut, Y. Negishi, K. Al-Shamery, Y. Matsumoto and T. Tsukuda, *J. Phys. Chem. C*, 2007, **111**, 4153–4158.
- 29 N. K. Chaki, Y. Negishi, H. Tsunoyama, Y. Shichibu and T. Tsukuda, *J. Am. Chem. Soc.*, 2008, **130**, 8608–8610.
- 30 M. Zhu, E. Lanni, N. Garg, M. E. Bier and R. Jin, *J. Am. Chem. Soc.*, 2008, **130**, 1138–1139.
- 31 Z. Wu, J. Suhan and R. Jin, *J. Mater. Chem.*, 2009, **19**, 622–626.
- 32 H. Qian, M. Zhu, U. N. Andersen and R. Jin, *J. Phys. Chem. A*, 2009, **113**, 4281–4284.
- 33 H. Qian, Y. Zhu and R. Jin, *ACS Nano*, 2009, **3**, 3795–3803.
- 34 A. Das, T. Li, K. Nobusada, C. Zeng, N. L. Rosi and R. Jin, *J. Am. Chem. Soc.*, 2013, **135**, 18264–18267.
- 35 C. Zeng, Y. Chen, G. Li and R. Jin, *Chem. Mater.*, 2014, **26**, 2635–2641.
- 36 G. Li, C. Zeng and R. Jin, *J. Am. Chem. Soc.*, 2014, **136**, 3673–3679.
- 37 H. Qian and R. Jin, *Nano Lett.*, 2009, **9**, 4083–4087.
- 38 H. Qian, Y. Zhu and R. Jin, *Proc. Natl. Acad. Sci. U. S. A.*, 2012, **109**, 696–700.
- 39 S. Knoppe, J. Boudon, I. Dolamic, A. Dass and T. Bürgi, *Anal. Chem.*, 2011, **83**, 5056–5061.
- 40 G. Li and R. Jin, *Acc. Chem. Res.*, 2013, **46**, 1749–1758.
- 41 T. Kawawaki and Y. Negishi, *Nanomaterials*, 2020, **10**, 238.
- 42 M. Zhou, T. Higaki, Y. Li, C. Zeng, Q. Li, M. Y. Sfeir and R. Jin, *J. Am. Chem. Soc.*, 2019, **141**, 19754–19764.
- 43 A. Sels, R. Azoulay, W. J. Buma, M. A. J. Koenis, V. P. Nicu and T. Bürgi, *J. Phys. Chem. C*, 2019, **123**, 22586–22594.
- 44 Z. Wu, M. Wang, J. Yang, X. Zheng, W. Cai, G. Meng, H. Qian, H. Wang and R. Jin, *Small*, 2012, **8**, 2028–2035.
- 45 T. Shu, J. Wang, L. Su and X. Zhang, *Anal. Chem.*, 2016, **88**, 11193–11198.
- 46 S. K. Katla, J. Zhang, E. Castro, R. A. Bernal and X. Li, *ACS Appl. Mater. Interfaces*, 2018, **10**, 75–82.
- 47 D. Yang, G. Yang, S. Gai, F. He, G. An, Y. Dai, R. Lv and P. Yang, *Nanoscale*, 2015, **7**, 19568–19578.
- 48 P. D. Jadzinsky, G. Calero, C. J. Ackerson, D. A. Bushnell and R. D. Kornberg, *Science*, 2007, **318**, 430–433.
- 49 B. K. Teo and H. Zhang, *Coord. Chem. Rev.*, 1995, **143**, 611–636.
- 50 H. Häkkinen, M. Walter and H. Grönbeck, *J. Phys. Chem. B*, 2006, **110**, 9927–9931.
- 51 M. Walter, J. Akola, O. Lopez-Acevedo, P. D. Jadzinsky, G. Calero, C. J. Ackerson, R. L. Whetten, H. Grönbeck and H. Häkkinen, *Proc. Natl. Acad. Sci. U. S. A.*, 2008, **105**, 9157–9162.
- 52 M. Zhu, C. M. Aikens, F. J. Hollander, G. C. Schatz and R. Jin, *J. Am. Chem. Soc.*, 2008, **130**, 5883–5885.
- 53 M. W. Heaven, A. Dass, P. S. White, K. M. Holt and R. W. Murray, *J. Am. Chem. Soc.*, 2008, **130**, 3754–3755.
- 54 J. Akola, M. Walter, R. L. Whetten, H. Häkkinen and H. Grönbeck, *J. Am. Chem. Soc.*, 2008, **130**, 3756–3757.
- 55 H. Qian, W. T. Eckenhoff, Y. Zhu, T. Pintauer and R. Jin, *J. Am. Chem. Soc.*, 2010, **132**, 8280–8281.
- 56 Y. Pei, Y. Gao and X. C. Zeng, *J. Am. Chem. Soc.*, 2008, **130**, 7830–7832.
- 57 D.-e. Jiang, W. Chen, R. L. Whetten and Z. Chen, *J. Phys. Chem. C*, 2009, **113**, 16983–16987.
- 58 C. Zeng, T. Li, A. Das, N. L. Rosi and R. Jin, *J. Am. Chem. Soc.*, 2013, **135**, 10011–10013.
- 59 Y. Chen, C. Liu, Q. Tang, C. Zeng, T. Higaki, A. Das, D.-e. Jiang, N. L. Rosi and R. Jin, *J. Am. Chem. Soc.*, 2016, **138**, 1482–1485.
- 60 A. Das, T. Li, K. Nobusada, Q. Zeng, N. L. Rosi and R. Jin, *J. Am. Chem. Soc.*, 2012, **134**, 20286–20289.
- 61 A. Das, T. Li, G. Li, K. Nobusada, C. Zeng, N. L. Rosi and R. Jin, *Nanoscale*, 2014, **6**, 6458–6462.
- 62 Y. Song, S. Wang, J. Zhang, X. Kang, S. Chen, P. Li, H. Sheng and M. Zhu, *J. Am. Chem. Soc.*, 2014, **136**, 2963–2965.
- 63 S. Chen, S. Wang, J. Zhong, Y. Song, J. Zhang, H. Sheng, Y. Pei and M. Zhu, *Angew. Chem., Int. Ed.*, 2015, **54**, 3145–3149.
- 64 C. Zeng, C. Liu, Y. Chen, N. L. Rosi and R. Jin, *J. Am. Chem. Soc.*, 2014, **136**, 11922–11925.



- 65 L. Xiong, S. Yang, X. Sun, J. Chai, B. Rao, L. Yi, M. Zhu and Y. Pei, *J. Phys. Chem. C*, 2018, **122**, 14898–14907.
- 66 C. Zeng, H. Qian, T. Li, G. Li, N. L. Rosi, B. Yoon, R. N. Barnett, R. L. Whetten, U. Landman and R. Jin, *Angew. Chem., Int. Ed.*, 2012, **51**, 13114–13118.
- 67 C. Zeng, C. Liu, Y. Chen, N. L. Rosi and R. Jin, *J. Am. Chem. Soc.*, 2016, **138**, 8710–8713.
- 68 Y. Chen, C. Zeng, C. Liu, K. Kirschbaum, C. Gayathri, R. R. Gil, N. L. Rosi and R. Jin, *J. Am. Chem. Soc.*, 2015, **137**, 10076–10079.
- 69 N. Yan, N. Xia, L. Liao, M. Zhu, F. Jin, R. Jin and Z. Wu, *Sci. Adv.*, 2018, **4**, eaat7259.
- 70 O. Lopez-Acevedo, J. Akola, R. L. Whetten, H. Grönbeck and H. Häkkinen, *J. Phys. Chem. C*, 2009, **113**, 5035–5038.
- 71 E. Pensa, E. Cortés, G. Corthey, P. Carro, C. Vericat, M. H. Fonticelli, G. Benítez, A. A. Rubert and R. C. Salvarezza, *Acc. Chem. Res.*, 2012, **45**, 1183–1192.
- 72 T. Bürgi, *Nanoscale*, 2015, **7**, 15553–15567.
- 73 I. Dolamic, S. Knoppe, A. Dass and T. Bürgi, *Nat. Commun.*, 2012, **3**, 798.
- 74 S. Knoppe, I. Dolamic and T. Bürgi, *J. Am. Chem. Soc.*, 2012, **134**, 13114–13120.
- 75 S. Malola and H. Häkkinen, *J. Am. Chem. Soc.*, 2019, **141**, 6006–6012.
- 76 N. Barrabés, B. Zhang and T. Bürgi, *J. Am. Chem. Soc.*, 2014, **136**, 14361–14364.
- 77 S. Knoppe and T. Bürgi, *Phys. Chem. Chem. Phys.*, 2013, **15**, 15816–15820.
- 78 B. Varnholt, P. Oulevey, S. Lubber, C. Kumara, A. Dass and T. Bürgi, *J. Phys. Chem. C*, 2014, **118**, 9604–9611.
- 79 B. Varnholt, I. Dolamic, S. Knoppe and T. Bürgi, *Nanoscale*, 2013, **5**, 9568–9571.
- 80 L. Beqa, D. Deschamps, S. Perrio, A.-C. Gaumont, S. Knoppe and T. Bürgi, *J. Phys. Chem. C*, 2013, **117**, 21619–21625.
- 81 A. C. Templeton, M. J. Hostetler, C. T. Kraft and R. W. Murray, *J. Am. Chem. Soc.*, 1998, **120**, 1906–1911.
- 82 Y. Song and R. W. Murray, *J. Am. Chem. Soc.*, 2002, **124**, 7096–7102.
- 83 R. L. Donkers, Y. Song and R. W. Murray, *Langmuir*, 2004, **20**, 4703–4707.
- 84 R. Guo, Y. Song, G. Wang and R. W. Murray, *J. Am. Chem. Soc.*, 2005, **127**, 2752–2757.
- 85 M. J. Hostetler, A. C. Templeton and R. W. Murray, *Langmuir*, 1999, **15**, 3782–3789.
- 86 N. Yan, N. Xia and Z. Wu, *Small*, 2020, 2000609.
- 87 V. Rojas-Cervellera, L. Raich, J. Akola and C. Rovira, *Nanoscale*, 2017, **9**, 3121–3127.
- 88 C. L. Heinecke, T. W. Ni, S. Malola, V. Mäkinen, O. A. Wong, H. Häkkinen and C. J. Ackerson, *J. Am. Chem. Soc.*, 2012, **134**, 13316–13322.
- 89 Y. Niihori, W. Kurashige, M. Matsuzaki and Y. Negishi, *Nanoscale*, 2013, **5**, 508–512.
- 90 M. Klunker, M. Mondeshki, M. N. Tahir and W. Tremel, *Langmuir*, 2018, **34**, 1700–1710.
- 91 T. W. Ni, M. A. Tofanelli, B. D. Phillips and C. J. Ackerson, *Inorg. Chem.*, 2014, **53**, 6500–6502.
- 92 Y. Niihori, Y. Kikuchi, A. Kato, M. Matsuzaki and Y. Negishi, *ACS Nano*, 2015, **9**, 9347–9356.
- 93 E. S. Shibu, M. A. H. Muhammed, T. Tsukuda and T. Pradeep, *J. Phys. Chem. C*, 2008, **112**, 12168–12176.
- 94 G. H. Woehrle, L. O. Brown and J. E. Hutchison, *J. Am. Chem. Soc.*, 2005, **127**, 2172–2183.
- 95 L. O. Brown and J. E. Hutchison, *J. Am. Chem. Soc.*, 1997, **119**, 12384–12385.
- 96 X. Kang, H. Chong and M. Zhu, *Nanoscale*, 2018, **10**, 10758–10834.
- 97 C. A. Fields-Zinna, J. F. Parker and R. W. Murray, *J. Am. Chem. Soc.*, 2010, **132**, 17193–17198.
- 98 P. Pengo, C. Bazzo, M. Boccalon and L. Pasquato, *Chem. Commun.*, 2015, **51**, 3204–3207.
- 99 S. Hossain, W. Kurashige, S. Wakayama, B. Kumar, L. V. Nair, Y. Niihori and Y. Negishi, *J. Phys. Chem. C*, 2016, **120**, 25861–25869.
- 100 A. Fernando and C. M. Aikens, *J. Phys. Chem. C*, 2015, **119**, 20179–20187.
- 101 S. Knoppe, R. Azoulay, A. Dass and T. Bürgi, *J. Am. Chem. Soc.*, 2012, **134**, 20302–20305.
- 102 A. Fernando and C. M. Aikens, *J. Phys. Chem. C*, 2016, **120**, 14948–14961.
- 103 A. Sels, G. Salassa, S. Pollitt, C. Guglieri, G. Rupprechter, N. Barrabés and T. Bürgi, *J. Phys. Chem. C*, 2017, **121**, 10919–10926.
- 104 V. R. Jupally, R. Kota, E. V. Dornshuld, D. L. Mattern, G. S. Tschumper, D.-e. Jiang and A. Dass, *J. Am. Chem. Soc.*, 2011, **133**, 20258–20266.
- 105 B. Molina, A. Sánchez-Castillo, S. Knoppe, I. L. Garzón, T. Bürgi and A. Tlahuice-Flores, *Nanoscale*, 2013, **5**, 10956–10962.
- 106 S. Knoppe, A. C. Dharmaratne, E. Schreiner, A. Dass and T. Bürgi, *J. Am. Chem. Soc.*, 2010, **132**, 16783–16789.
- 107 S. Si, C. Gautier, J. Boudon, R. Taras, S. Gladiali and T. Bürgi, *J. Phys. Chem. C*, 2009, **113**, 12966–12969.
- 108 A. Sels, N. Barrabés, S. Knoppe and T. Bürgi, *Nanoscale*, 2016, **8**, 11130–11135.
- 109 V. Truttman, C. Herzig, I. Illes, A. Limbeck, E. Pittenauer, M. Stöger-Pollach, G. Allmaier, T. Bürgi, N. Barrabés and G. Rupprechter, *Nanoscale*, 2020, **12**, 12809–12816.
- 110 L. C. McKenzie, T. O. Zaikova and J. E. Hutchison, *J. Am. Chem. Soc.*, 2014, **136**, 13426–13435.
- 111 S. Mason, *Trends Pharmacol. Sci.*, 1986, **7**, 20–23.
- 112 J. L. Bada, *Nature*, 1995, **374**, 594–595.
- 113 C. Gautier and T. Bürgi, *ChemPhysChem*, 2009, **10**, 483–492.
- 114 H. Yao, *Prog. Nat. Sci.: Mater. Int.*, 2016, **26**, 428–439.
- 115 C. Zeng and R. Jin, *Chem. - Asian J.*, 2017, **12**, 1839–1850.
- 116 M. Zhu, H. Qian, X. Meng, S. Jin, Z. Wu and R. Jin, *Nano Lett.*, 2011, **11**, 3963–3969.
- 117 S. Kumar and R. Jin, *Nanoscale*, 2012, **4**, 4222–4227.
- 118 Q. Xu, S. Kumar, S. Jin, H. Qian, M. Zhu and R. Jin, *Small*, 2014, **10**, 1008–1014.
- 119 S. Knoppe, N. Kothalawala, V. R. Jupally, A. Dass and T. Bürgi, *Chem. Commun.*, 2012, **48**, 4630–4632.
- 120 C. Zeng, C. Liu, Y. Pei and R. Jin, *ACS Nano*, 2013, **7**, 6138–6145.



- 121 M.-B. Li, S.-K. Tian, Z. Wu and R. Jin, *Chem. Mater.*, 2016, **28**, 1022–1025.
- 122 S. Wang, H. Abroshan, C. Liu, T.-Y. Luo, M. Zhu, H. J. Kim, N. L. Rosi and R. Jin, *Nat. Commun.*, 2017, **8**, 848.
- 123 P. R. Nimmala, S. Theivendran, G. Barcaro, L. Sementa, C. Kumara, V. R. Jupally, E. Apra, M. Stener, A. Fortunelli and A. Dass, *J. Phys. Chem. Lett.*, 2015, **6**, 2134–2139.
- 124 C. Zeng, Y. Chen, A. Das and R. Jin, *J. Phys. Chem. Lett.*, 2015, **6**, 2976–2986.
- 125 A. Dass, T. C. Jones, S. Theivendran, L. Sementa and A. Fortunelli, *J. Phys. Chem. C*, 2017, **121**, 14914–14919.
- 126 Z. Gan, J. Chen, J. Wang, C. Wang, M.-B. Li, C. Yao, S. Zhuang, A. Xu, L. Li and Z. Wu, *Nat. Commun.*, 2017, **8**, 14739.
- 127 M. P. Maman, A. S. Nair, H. Cheraparambil, B. Pathak and S. Mandal, *J. Phys. Chem. Lett.*, 2020, **11**, 1781–1788.
- 128 P. R. Nimmala and A. Dass, *J. Am. Chem. Soc.*, 2014, **136**, 17016–17023.
- 129 S. K. Eswaramoorthy, N. A. Sakthivel and A. Dass, *J. Phys. Chem. C*, 2019, **123**, 9634–9639.
- 130 H. Dong, L. Liao and Z. Wu, *J. Phys. Chem. Lett.*, 2017, **8**, 5338–5343.
- 131 L. Liao, S. Zhuang, C. Yao, N. Yan, J. Chen, C. Wang, N. Xia, X. Liu, M.-B. Li, L. Li, X. Bao and Z. Wu, *J. Am. Chem. Soc.*, 2016, **138**, 10425–10428.
- 132 R. A. Sperling and W. J. Parak, *Philos. Trans. R. Soc., A*, 2010, **368**, 1333–1383.
- 133 M. A. Habeeb Muhammed and T. Pradeep, *J. Cluster Sci.*, 2009, **20**, 365–373.
- 134 J. W. Padelford, T. Wang and G. Wang, *ChemElectroChem*, 2016, **3**, 1201–1205.
- 135 S. Knoppe, O. A. Wong, S. Malola, H. Häkkinen, T. Bürgi, T. Verbiest and C. J. Ackerson, *J. Am. Chem. Soc.*, 2014, **136**, 4129–4132.
- 136 H. Yao and M. Iwatsu, *Langmuir*, 2016, **32**, 3284–3293.
- 137 H. Yao and S. Tsubota, *Chem. Phys.*, 2017, **493**, 149–156.
- 138 Y. Wang, H. Su, C. Xu, G. Li, L. Gell, S. Lin, Z. Tang, H. Häkkinen and N. Zheng, *J. Am. Chem. Soc.*, 2015, **137**, 4324–4327.
- 139 L. Ren, P. Yuan, H. Su, S. Malola, S. Lin, Z. Tang, B. K. Teo, H. Häkkinen, L. Zheng and N. Zheng, *J. Am. Chem. Soc.*, 2017, **139**, 13288–13291.
- 140 X. Zan, Q. Li, Y. Pan, D. J. Morris, P. Zhang, P. Li, H. Yu and M. Zhu, *ACS Appl. Nano Mater.*, 2018, **1**, 6773–6781.
- 141 C.-A. J. Lin, T.-Y. Yang, C.-H. Lee, S. H. Huang, R. A. Sperling, M. Zanella, J. K. Li, J.-L. Shen, H.-H. Wang, H.-I. Yeh, W. J. Parak and W. H. Chang, *ACS Nano*, 2009, **3**, 395–401.
- 142 Y. Bai, T. Shu, L. Su and X. Zhang, *Crystals*, 2020, **10**, 357.
- 143 Y. Guo, H. T. N. N. Amunyela, Y. Cheng, Y. Xie, H. Yu, W. Yao, H.-W. Li and H. Qian, *Food Chem.*, 2021, **335**, 127657.
- 144 Y. Zheng, L. Lai, W. Liu, H. Jiang and X. Wang, *Adv. Colloid Interface Sci.*, 2017, **242**, 1–16.
- 145 L. Shang, F. Stockmar, N. Azadfar and G. U. Nienhaus, *Angew. Chem., Int. Ed.*, 2013, **52**, 11154–11157.
- 146 Y. Wang, L. Hu, L. Li and J.-J. Zhu, *Journal of Analysis and Testing*, 2017, **1**, 13.
- 147 X. Hu, Y. Zheng, J. Zhou, D. Fang, H. Jiang and X. Wang, *Chem. Mater.*, 2018, **30**, 1947–1955.

