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## Rapid biocompatible macrocyclization of peptides with decafluoro-diphenylsulfone†

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In this manuscript, we describe modification of Cys-residues in peptides and proteins in aqueous solvents *via* aromatic nucleophilic substitution ( $S_NAr$ ) with perfluoroarenes (fAr). Biocompatibility of this reaction makes it attractive for derivatization of proteins and peptide libraries comprised of 20 natural amino acids. Measurement of the reaction rates for fAr derivatives by  $^{19}F$  NMR with a model thiol donor ( $\beta$ -mercaptoethanol) in aqueous buffers identified decafluoro-diphenylsulfone (DFS) as the most reactive  $S_NAr$  electrophile. Reaction of DFS with thiol nucleophiles is >100 000 faster than analogous reaction of perfluorobenzene; this increase in reactivity enables application of DFS at low concentrations in aqueous solutions compatible with biomolecules and protein complexes irreversibly degraded by organic solvents (*e.g.*, bacteriophages). DFS forms macrocycles when reacted with peptides of the general structure  $X_n$ -Cys- $X_m$ -Cys- $X_l$ , where X is any amino acid and  $m = 1$ –15. It formed cyclic peptides with 6 peptide hormones—oxytocin, urotensin II, salmon calcitonin, melanin-concentrating hormone, somatostatin-14, and atrial natriuretic factor (1–28) as well as peptides displayed on M13 phage. Rates up to  $180 M^{-1} s^{-1}$  make this reaction one of the fastest Cys-modifications to-date. Long-term stability of macrocycles derived from DFS and their stability toward oxidation further supports DFS as a promising method for modification of peptide-based ligands, cyclization of genetically-encoded peptide libraries, and discovery of bioactive macrocyclic peptides.

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## Introduction

Rapid, site-specific modifications of polypeptides are critical for chemical modification of proteins, synthesis of antibody–drug conjugates, and generation of genetically-encoded libraries with unnatural moieties. Biocompatible site-specific cross-linking of two side-chain residues in polypeptides is a rapidly growing field because such modification can increase stability and efficacy of peptide-based therapeutics (for recent reviews see<sup>1–6</sup>). While such cross-linking or macrocyclization can be performed on amino acid side-chains that contain unnatural reactive groups,<sup>7–9</sup> modification of naturally-occurring residues is particularly attractive because it can be used for modification of readily-available peptides/proteins and phage display libraries comprised of 20 natural amino acids.<sup>10–16</sup>

In this article, we report rapid, biocompatible modification of Cys residues *via* nucleophilic aromatic substitution ( $S_NAr$ ) reaction with perfluoroarenes (fAr). We overcome the deficiencies of previously known  $S_NAr$ -reagents, such as 1-chloro-

2,4-dinitrobenzene (CDNB),<sup>17</sup> which are susceptible to background reactivity with nucleophilic residues in proteins, or perfluorobenzene<sup>18</sup> and perfluorobiphenyl<sup>18,19</sup> which suffer from low reactivity and poor solubility in water. The latter reagents are effective in organic solvents, such as DMF, but are not suitable for modification of biological entities that are irreversibly deactivated by high concentrations of organic solvents (*e.g.*, proteins, bacteriophage, *etc.*). While glutathione transferase can be used to catalyze  $S_NAr$ -reactions in water,<sup>20</sup> the enzyme acts only on peptides that contain unnatural N-terminal- $\alpha$ -Glu.<sup>20,21</sup> Hence, improved fAr cross-linking agents provide the ability to access diverse chemical modalities through native polypeptide sequences.

## Results and discussion

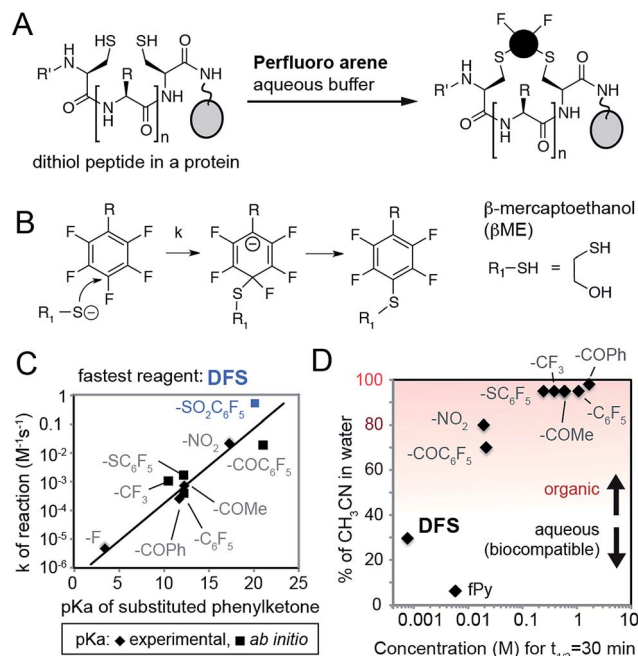
To identify  $S_NAr$  derivatives for aqueous bioconjugations, we explored the rate of the reaction of a model thiol donor,  $\beta$ -mercaptoethanol ( $\beta$ ME), and twelve fAr reagents using  $^{19}F$  NMR (see ESI Fig. S1–S3,† examples of  $^{19}F$  NMR traces) in aqueous Tris-buffer pH = 8.5 supplemented with  $CH_3CN$  as co-solvent (Fig. 1A–C). Reactivity of most perfluoroarenes was hampered by both poor solubility and poor reactivity (Fig. 1D); many reactions required 80% or more  $CH_3CN$  as co-solvent. Similar to studies in organic solvents,<sup>22</sup> the reactivity in aqueous

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**Fig. 1** (A) Design of reagents for perfluoroarene macrocyclization by cross-linking two Cys-residues in a protein in aqueous conditions. (B) Mechanism of  $S_NAr$  reaction. (C) Rate constants ( $k$ ) of the reaction between substituted fAr and  $\beta$ ME, as measured by  $^{19}F$  NMR, increase with the increasing electronegativity of R group. As a measure of electronegativity, we used  $pK_a$  in a series of phenylketones for which many values are known<sup>23</sup> or can be calculated (ESI Fig. S4†). (D) To find the  $S_NAr$  reagents for rapid modification in water, we plotted molar concentration of fAr necessary to reach 50% conversion in 30 minutes (calculated as  $\ln(2)/(1800 \times k)$ , where  $k$  is the rate constant) and % of organic co-solvent necessary to dissolve the fAr at this concentration. Most fAr are poorly reactive and/or too insoluble in aqueous solvents; only **DFS** and perfluoropyridine (fPy) can be used in conditions that require low amount of organic co-solvent.

conditions correlates with the electronegativity of the substituent on the aromatic ring (Fig. 1C) yielding the fastest reactivity for perfluoronitrobenzene ( $R = NO_2$ ), decafluorobenzophenone ( $R = -COC_6F_5$ ), or decafluoro-diphenylsulfone ( $R = -SO_2C_6F_5$ , **DFS**) reagents, or heteroaromatic derivatives such as perfluoropyridine.

We then validated the reactivity and specificity of the most reactive substrate (**DFS**) using model peptides  $H_2N-SWCXC-CONH_2$ , where  $X = Asp, Ala, Ser, \text{ or } Arg$  that contained two types of nucleophiles: N-terminal amine and thiol (Fig. 2A). We were pleased to find that **DFS** reacted remarkably fast with Cys side-chains (Fig. 2B and ESI Fig. S5 and S6†) in conditions that required as little as 10% of  $CH_3CN$  co-solvent (Fig. 1) or as little as 5% DMF co-solvent (ESI Fig. S7†). As  $S_NAr$  reaction alters the aromatic  $\pi$ -system,<sup>17,24</sup> we measured rate constants by measuring changes in absorption at 300 nm (ESI Fig. S5 and S6, S8–S11†) and validated these rates by  $^{19}F$  NMR (ESI Fig. S10 and S11†) and LCMS (ESI Fig. S7†). Conveniently, the reaction was accelerated in solvents with high water content, although the rate decreased in water without any  $CH_3CN$  due to limited solubility of **DFS** (ESI Fig. S12†). At pH 8.5 and water content of 70–80%, the rate constant was  $100\text{--}180\text{ M}^{-1}\text{ s}^{-1}$  for positively



**Fig. 2** (A)  $S_NAr$  reaction of **DFS** with peptides is sequence and solvent-dependent. (B) Rates in all conditions were measured by monitoring absorbance at 320 nm, and identities of the products were confirmed by NMR and Liquid Chromatography Mass-Spectrometry (LCMS, ESI Fig. S7 and S12–S15†). (C) The rate of the reaction correlates with acidity of the thiol;  $pK_a$  were estimated by measuring the absorbance of peptides at 240 nm as described in ESI Fig. S18† and were in accordance with previously reported values for CXC peptides.<sup>25</sup>

charged peptide sequence **SWCRC** and  $50\text{--}80\text{ M}^{-1}\text{ s}^{-1}$  for neutral analog **SWCAC** (Fig. 2B).

The rate of the reaction with **DFS** was sequence dependent (Fig. 2B and C and ESI Fig. S5†). Positively-charged Arg residue next to Cys accelerated the reaction, negatively charged Asp suppressed it. This observation can be explained by higher basicity of thiol in **CRC** vs. **CDC** (Fig. 2C). Additionally, the positive charge on the Arg side-chain can stabilize the intermediate carbanion (Fig. 1B). We anticipate that analogous strong dependence of the rate constant on the  $pK_a$  of thiol (Fig. 2C) exist in other  $S_NAr$  reactions between peptides and perfluoroarenes<sup>18,19</sup> because  $S_NAr$  reactions with thiols exhibit large Brønsted coefficients.<sup>20</sup> Substrate-dependence was the most pronounced when solvation was poor (e.g., high  $CH_3CN$  content); increased solvation in solutions with high aqueous content masked the effect (Fig. 2B and ESI Fig. S6†). The pH-dependence of the rate plateaued at pH 8–9 confirming that thiolate is the nucleophile (ESI Fig. S19 and S20†). In acidic pH, the rate dropped exponentially (i.e., decrease of  $\log(k)$  was linear with pH). Reaction with perfluoropyridine exhibited similar solvent-dependent and substrate-dependent profiles (ESI Fig. S8, S9, and S21†), however, the rates were 10–100 times slower.

LCMS confirmed the identity of the products during the reaction and 24–48 hours after its completion (ESI Fig. S12–S15†). LCMS confirmed that the reaction was selective for Cys in the presence of free N-terminus and large excess of **DFS**.



Side-reactivity was observed only after prolonged exposure to **DFS**. Specifically, peptides SWCAC or SWCRC incubated with large excess of **DFS** (10 eq., 2 mM) for 48 hours contained ~20% of the product with two **DFS** moieties. Less than 5% of this by-product was visible after 48 hours in the presence of 0.2 mM **DFS**. In contrast, Cys arylation under analogous conditions was complete in 5–10 minutes (ESI Fig. S6†). We observed this by-product in three different peptide sequences (ESI Fig. S13–S15†) and proposed it to be a product of  $S_NAr$  reaction between **DFS** and N-terminus. While the side-reaction of N-terminal amine is possible, it is  $>10^4$  times slower than Cys-arylation. This data still do not completely rule out the possibility that the cyclization could also happen between the thiol and the terminal amine. To show that cyclization occurs exclusively on Cys residues, we treated peptide with **DFS** and then with thiol-specific reagent, biotin-PEG<sub>2</sub>-iodoacetamide (**BIA**). While **BIA** reacted readily with thiols in starting material, we detected no reaction between **BIA** and octafluoro-diphenylsulfone (**OFS**)-peptide conjugate (Fig. S17†).

To confirm that **DFS** is useful for modification of complex peptides, we modified peptide CYIQNCP LG (oxytocin, OT). Reaction yielded a single product with expected mass and apparent rate constant of  $37.2 M^{-1} s^{-1}$  as measured by LCMS (Fig. 3, ESI Fig. S16†). To stop the reaction before completion, we quenched it with acid and observed mono-arylated intermediates of oxytocin M1 and M2 in a transient open-chain state (Fig. 3A and B). Both intermediates disappeared over the course of the reaction (Fig. 3C). To confirm that **DFS** can form cyclic peptides regardless of their ring size, we tested reaction between **DFS** and five other peptide hormones of general structure  $X_N$ -Cys- $X_M$ -Cys- $X_L$ , where X is any amino acid other than Cys and  $M$ —the number of amino acids flanked by two Cys—ranged from 4 to 15. Urotensin II (UT,  $M = 4$ ), salmon calcitonin (CT,  $M = 5$ ), melanin-concentrating hormone (MCH,  $M = 8$ ), somatostatin-14 (ST,  $M = 10$ ), and atrial natriuretic factor (1–28) (ANF,  $M = 15$ ) formed cyclic peptides in reaction with **DFS** regardless of their ring size. We used HPLC to estimate the rate of cyclization (Fig. 3C, S22–S23†) by quenching the reaction with acid in the presence of 1 mM **DFS**. After 30 seconds, we detected 10–90% of the final cyclized product and <5–60% of starting material remaining. After 10 min, only urotensin II, which displayed the slowest kinetics, contained ~5% of the unreacted starting material. Other peptides were completely converted to final cyclic products.

We then confirmed reaction of **DFS** with thiols on intact M13 bacteriophage, which is composed of 5 types of proteins and a single-stranded DNA genome. The phage virion is an ideal model for bioconjugation because (i) it can be engineered to display any peptide sequence, (ii) modification can be detected in individual phage particles,<sup>14</sup> (iii) biocompatibility of the modification can be readily quantified by measuring the fraction of M13 particles that can replicate in bacteria; reagents that damage the proteins or DNA disrupt this replication<sup>14</sup> (ESI Fig. S24†). We validated the modification on M13 phage using a biotin capture assay developed previously by our group (Fig. 4).<sup>14,26</sup> Briefly, a phage displaying a disulfide-peptide ACPARSPLEC was reduced with TCEP and the resulting thiols

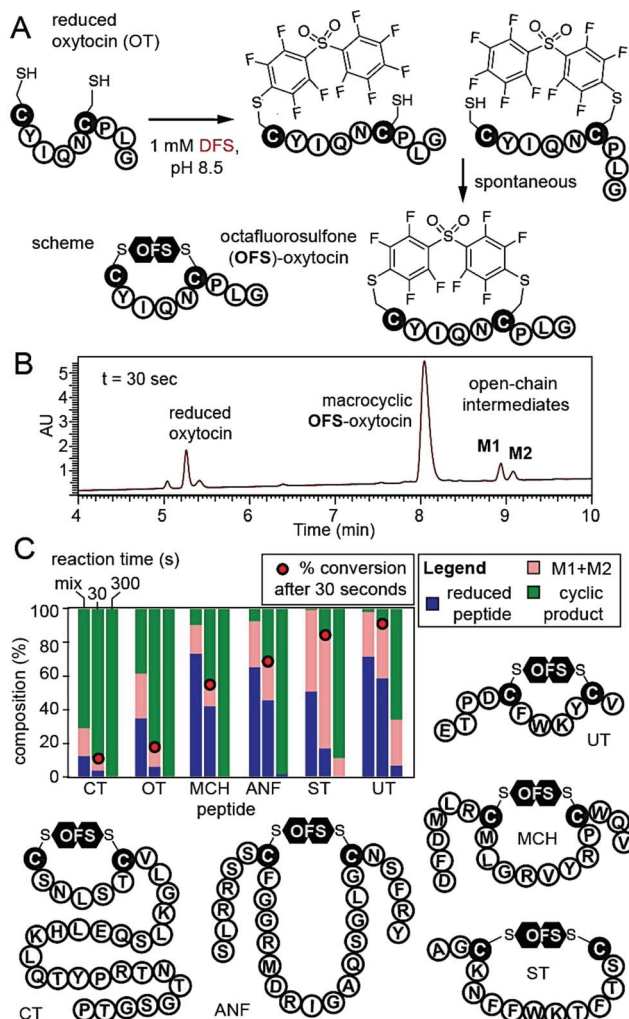


Fig. 3 (A and B) Cyclization of reduced oxytocin (OT) by **DFS**. Quenching the reaction by 2% TFA at 30 s revealed ~90% consumption of starting material, 80% of product, and 12% of open-chain intermediates M1 and M2. (C) Stacked bar representation of the composition of reaction mixture determined by HPLC shows that both M1 and M2 of OT were completely converted to product by 5 min. Analogous studies with five other peptide hormones in the presence of 1 mM **DFS**. Fraction of the reactant, sum of M1 + M2 intermediates, and products was determined by integration of the HPLC traces at 215 nm (see ESI Fig. S16, S22, and S23† for raw traces).

were alkylated using **BIA** (Fig. 4B and C). The reaction yielded ~95% of the biotinylated phage, as determined by titrating the phage before and after exposure to streptavidin-coated beads (Fig. 4D). We then used a “pulse-chase” approach<sup>13–16,26</sup> to quantify reaction of phage with **DFS**. “Pulsing” the reduced phage with 0.5 mM **DFS** in 30% CH<sub>3</sub>CN-Tris buffer (pH 8.5) for 30 min consumed ~75% phage-displayed thiols. “Chase” of the reaction with **BIA** yielded only 20% of biotinylated phage. In contrast, the phage remained reduced and susceptible to alkylation by **BIA** when it was “pulsed” with buffer that contained no **DFS**. Thus, disappearance of thiol indicates the reaction with **DFS** and is not due to side reactions (e.g., oxidation). The same “pulse-chase” quantification confirmed that peptide ACPARSPLEC displayed on phage reacts with

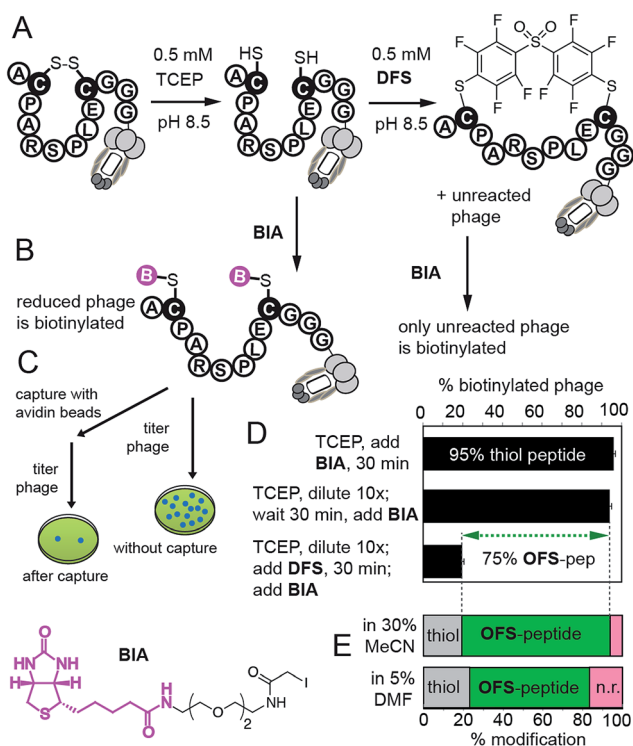


perfluoropyridine at much slower rate than **DFS** (Fig. S25†). We further validated the “pulse-chase” method on purified peptide SWCDYRC and confirmed that the peptide “pulsed” by **DFS** for 30 minutes forms a product that does not react with **BIA** (Fig. S17†).

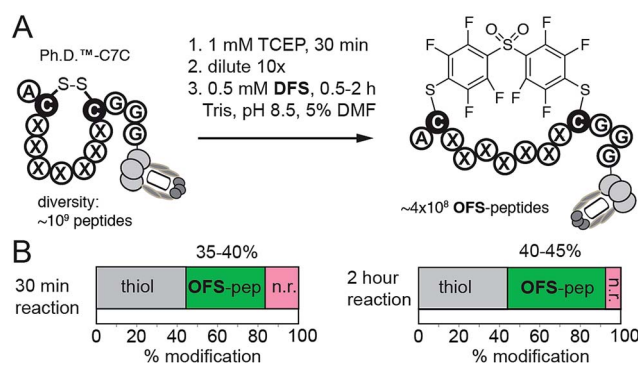
While M13 phage is stable in solutions that contain 30% CH<sub>3</sub>CN, in many applications such as protein modifications, such a high amount of acetonitrile might not be tolerated. To this end, **DFS**-mediated cyclization can be performed in aqueous solutions containing only 5% DMF as co-solvent. We observed that solubility of **DFS** in Tris buffer (pH 8.5) containing 5% DMF (v : v) is approximately 500 μM. This concentration is sufficient to perform rapid modifications of synthetic peptides, phage-displayed peptides, and phage-displayed libraries of peptides. For example, reaction between 100 μM **DFS** and 100 μM peptide SWCDYRC required approximately 100 seconds to reach 50% conversion and 15 min to reach completion (ESI Fig. S7†). LCMS monitoring of this reaction estimated the 2<sup>nd</sup> order rate constant to be 100–130 M<sup>-1</sup> s<sup>-1</sup> (ESI Fig. S7†). We then used these conditions to modify a phage clone that contained peptide ACPARSPLEC. The modification efficiency by

0.5 mM **DFS** in 5% DMF for 30 minutes was ~60%, (Fig. 4E). Finally, these conditions can also be used to modify commercially available libraries of peptides displayed on phage with 7 random amino acids in between the cysteine residues (New England Biolabs (Ph.D.<sup>TM</sup>-C7C, Fig. 5)). A decrease in chemical modification efficiency of libraries to 40%, when compared to modification of an individual clone (Fig. 4E), is not surprising because library contains 10<sup>9</sup> peptide sequences with diverse reactivity. This decrease in modification is also consistent with our previous reports.<sup>13,15</sup> **DFS** thus can conveniently convert readily available libraries of peptide-disulfides to genetically-encoded macrocyclic libraries that contain unique perfluoroarene cross-linker. We have employed these libraries in selection of peptide-macrocycle ligands for protein targets but results of these investigations extend beyond the scope of this manuscript and will be described in our subsequent report.

Cross-linking of two Cys residues is one of the most common modifications of synthetic peptides and recombinant proteins. To demonstrate advantages of **DFS** over well-established cysteine cross-linkers, we selected *α,α'*-dibromo-*meta*-xylene (**DBMX**) reagent as a reference (ESI Fig. S26†). Reagents structurally and electronically similar to **DBMX** are commonly used to stabilize secondary structure of peptides and synthesize genetically-encoded macrocycle libraries.<sup>27–29</sup> The major advantage of **OFS**-peptide conjugates (perfluoroaryl thioethers) is their higher stability to oxidation when compared to conjugates made with **DBMX** (benzyl thioethers). For example, we routinely use NaIO<sub>4</sub> to convert N-terminal Ser to N-terminal aldehyde.<sup>14,30,31</sup> Treatment with NaIO<sub>4</sub> readily converted SWCDYRC-**OFS** conjugate to *Ald*-WCDYRC-**OFS**, where “*Ald*” denotes aldehyde group (Fig. 6), and we observed no oxidation of aryl thioethers. As an example of application, we introduced N-terminal glycosylation<sup>14,30,31</sup> in SWCDYRC-**OFS** conjugate (Fig. 6). One-pot two-step reaction quantitatively converted peptide-**OFS** conjugate to *Ald*-peptide-**OFS** and then to glycopeptide-**OFS** conjugate. Such transformation would be



**Fig. 4** (A) Quantification strategy for the reaction between **DFS** and M13 phage displaying a peptide ACPARSPLEC.<sup>13</sup> (B) Reaction of reduced phage with biotin-iodoacetamide (**BIA**) biotinylates the phage. (C) Capture assay measures the yield of biotinylation. (D) Capture assay determined that 95% of phage is reduced (reactive with **BIA**). Diluting the reducing agent 10-fold does not change the number of reduced thiols, but exposing the reduced phage to 0.5 mM **DFS** in 30% MeCN/Tris buffer for 30 min consumes reactive thiols yielding 80% **OFS**-modified phage. Error bars represent standard deviations from two experiments. (E) Capture can be summarized as stack bar: bottom bar shows that ~60% of phage is modified by 0.5 mM solution of **DFS** in aqueous Tris buffer containing 5% DMF as co-solvent.



**Fig. 5** (A) An aqueous solution of **DFS** containing 5% DMF as co-solvent can conveniently convert commercially available phage displayed library of 10<sup>9</sup> disulfide heptapeptides—Ph.D.<sup>TM</sup>-C7C from New England Biolabs—to a library of **OFS**-macrocyclic peptides. (B) Biotin capture, analogous to that described in Fig. 4C–E, quantified the efficiency of this modification. After 30 minutes to 2 hours reaction with **DFS**, 35–45% of the library is modified with **DFS** yielding ~350–450 million of diverse phage-displayed **OFS**-macrocycles. Data is averaged from two independent experiments.



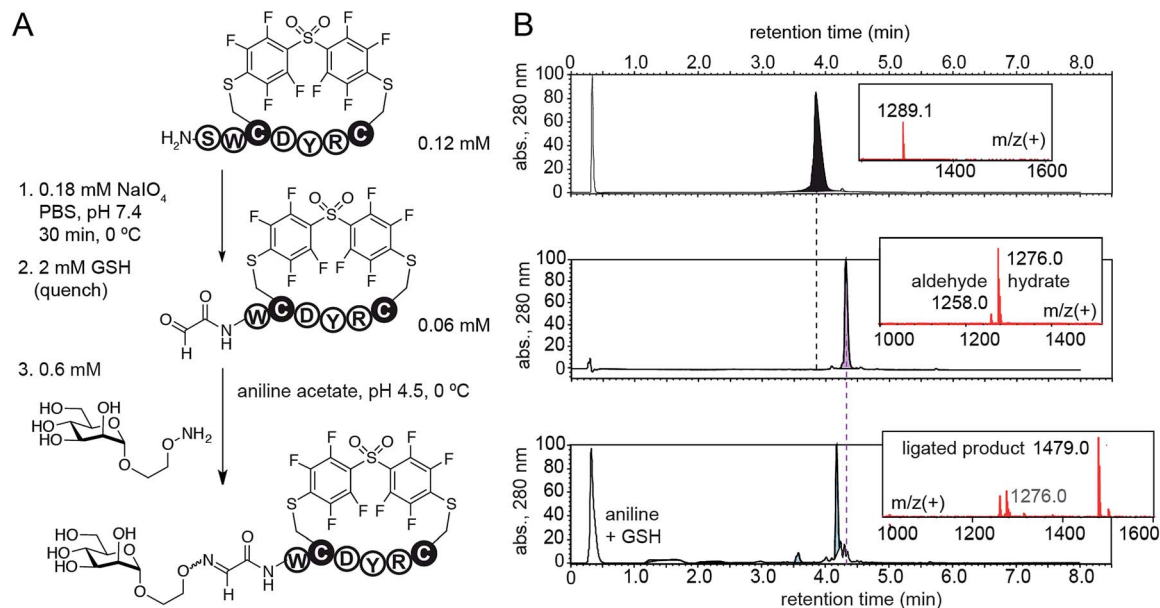


Fig. 6 Robust oxidation of SWCDYRC-OFS conjugate to peptide-aldehyde and subsequent one-pot conversion to a cyclic glycopeptide.

challenging to achieve in conjugates that contain thioether bonds derived from benzyl or haloacetyl cross-linkers (ESI Fig. S27†). Treatment of SWCDYRC-DMMX conjugate by NaIO<sub>4</sub> led to concomitant oxidation of serine and benzyl-thioether (ESI Fig. S27D†), whereas treatment of SWCDYRC-OFS in identical conditions produced no detectable traces of oxidized product (ESI Fig. S27C†).

To compare the stability of OFS-thioethers towards oxidation with stability of other conjugates, we exposed them to conditions recently used to evaluate stability of diverse thioethers (1 mM H<sub>5</sub>IO<sub>6</sub> at 37 °C).<sup>32</sup> Consistent with the published results,<sup>32</sup> we observed that H<sub>5</sub>IO<sub>6</sub> rapidly oxidized benzyl-thioether bonds and converted SWCDYRC-DMMX conjugate to four oxidized products in 10 minutes (ESI Fig. S27F†). In contrast, half-life of oxidation for thioethers in SWCDYRC-OFS by H<sub>5</sub>IO<sub>6</sub> was 16 hours (ESI Fig. S27E†). This value exceeds the reported oxidation half-lives for tolyl- or 4-cyanophenyl-thioether (1 and 6 h, see ESI Fig. S4† in ref. 32). As the rate of oxidation of aryl thioether scales with electronegativity of the aryl substituent,<sup>32</sup> it is not surprising that the peptide-OFS macrocycles exhibit high resistance to oxidation. We believe that fast reactivity of DFS combined with exceptional oxidative stability of resulting OFS-thioethers will make it possible to conduct robust multi-step modification of peptides, which combines Cys cross-linking and N-terminal ligations to functionalize natural proteins and diversify genetically-encoded libraries.

## Conclusions

The DFS framework combines fast reactivity up to 180 M<sup>-1</sup> s<sup>-1</sup>, specificity to Cys-residues, biocompatibility with complex multiprotein complexes (here, bacteriophage), and maximum performance at neutral to mildly-basic pH. Its reactivity significantly exceeds the rates of substitution reactions of Cys with

CDNB ( $k = 1 \text{ M}^{-1} \text{ s}^{-1}$ ),<sup>17</sup> haloacetyl cross-linkers ( $k = 10\text{--}20 \text{ M}^{-1} \text{ s}^{-1}$  at pH 8.5),<sup>13,33</sup> bis-bromobenzyl cross-linkers ( $k = 30 \text{ M}^{-1} \text{ s}^{-1}$  at pH 8.8),<sup>34</sup> bis-methyl-maleimide cross-linkers ( $k = 1\text{--}3 \text{ M}^{-1} \text{ s}^{-1}$ ),<sup>35</sup> and allenamides<sup>36</sup> ( $k = 16\text{--}30 \text{ M}^{-1} \text{ s}^{-1}$ ).<sup>16</sup> Recent summaries from the Bode<sup>37</sup> and Prescher<sup>38</sup> groups highlight that in modern bio-orthogonal reactions, very few reach rates of  $>100 \text{ M}^{-1} \text{ s}^{-1}$  as exhibited by DFS. To date only two Cys-reactive reagents, dichlorotetrazene<sup>39</sup> and maleimide (rates up to  $800 \text{ M}^{-1} \text{ s}^{-1}$ ),<sup>37</sup> can rival DFS in reactivity, but the products are susceptible to long-term hydrolytic instability.<sup>39–42</sup> In contrast, DFS adducts of oxytocin are stable for over 2 weeks at pH = 7.4 (ESI Fig. S28†). We note that the size of the DFS is compatible to the size of many previously reported thiol crosslinkers such as bisbromoxylene,<sup>27–29</sup> decafluorobenzene,<sup>18,19</sup> benzophenone,<sup>32</sup> bismaleimide, chloroacetamide-azobenzene,<sup>13</sup> and others. Introduction of DFS into proteins or peptides will be context dependent: it can provide favourable stabilization of secondary structure but it can also interfere with biological function. In contrast to many known Cys cross-linkers, DFS produces conjugates that are stable to oxidation; this stability will enable development of multi-step transformations that convert peptides made of natural amino acids to functionality-rich macrocyclic derivatives. We anticipate that this reaction will serve as a platform for development of new bioconjugation and macrocyclization strategies as well as post-translational diversification of genetically-encoded libraries.

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## References

- 1 A. Bhat, L. R. Roberts and J. J. Dwyer, *Eur. J. Med. Chem.*, 2015, **94**, 471–479.
- 2 C. P. R. Hackenberger and D. Schwarzer, *Angew. Chem., Int. Ed.*, 2008, **47**, 10030–10074.
- 3 N. Tsomaia, *Eur. J. Med. Chem.*, 2015, **94**, 459–470.
- 4 C. J. White and A. K. Yudin, *Nat. Chem.*, 2011, **3**, 509–524.
- 5 V. Azzarito, K. Long, N. S. Murphy and A. J. Wilson, *Nat. Chem.*, 2013, **5**, 161–173.
- 6 M. Pelay-Gimeno, A. Glas, O. Koch and T. N. Grossmann, *Angew. Chem., Int. Ed.*, 2015, **54**, 8896–8927.
- 7 H. E. Blackwell and R. H. Grubbs, *Angew. Chem., Int. Ed.*, 1998, **37**, 3281–3284.
- 8 C. E. Schafmeister, J. Po and G. L. Verdine, *J. Am. Chem. Soc.*, 2000, **122**, 5891–5892.
- 9 L. D. Walensky, A. L. Kung, I. Escher, T. J. Malia, S. Barbuto, R. D. Wright, G. Wagner, G. L. Verdine and S. J. Korsmeyer, *Science*, 2004, **305**, 1466–1470.
- 10 A. Angelini and C. Heinis, *Curr. Opin. Chem. Biol.*, 2011, **15**, 355–361.
- 11 E. Baslé, N. Joubert and M. Pucheault, *Chem. Biol.*, 2010, **17**, 213–227.
- 12 J. M. Smith, J. R. Frost and R. Fasan, *J. Org. Chem.*, 2013, **78**, 3525–3531.
- 13 M. R. Jafari, L. Deng, P. I. Kitov, S. Ng, W. L. Matochko, K. F. Tjhung, A. Zeberoff, A. Elias, J. S. Klassen and R. Derda, *ACS Chem. Biol.*, 2014, **9**, 443–450.
- 14 S. Ng, M. R. Jafari, W. L. Matochko and R. Derda, *ACS Chem. Biol.*, 2012, **7**, 1482–1487.
- 15 S. Ng and R. Derda, *Org. Biomol. Chem.*, 2016, DOI: 10.1039/c5ob02646f.
- 16 M. R. Jafari, J. Lakusta, R. J. Lundgren and R. Derda, *Bioconjugate Chem.*, 2016, DOI: 10.1021/acs.bioconjchem.6b00026.
- 17 A. M. Gold, *Biochemistry*, 1968, **7**, 2106–2115.
- 18 A. M. Spokoyny, Y. Zou, J. J. Ling, H. Yu, Y.-S. Lin and B. L. Pentelute, *J. Am. Chem. Soc.*, 2013, **135**, 5946–5949.
- 19 Y. Zou, A. M. Spokoyny, C. Zhang, M. D. Simon, H. Yu, Y.-S. Lin and B. L. Pentelute, *Org. Biomol. Chem.*, 2014, **12**, 566–573.
- 20 B. S. Nieslanik and W. M. Atkins, *J. Am. Chem. Soc.*, 1998, **120**, 6651–6660.
- 21 C. Zhang, A. M. Spokoyny, Y. Zou, M. D. Simon and B. L. Pentelute, *Angew. Chem., Int. Ed.*, 2013, **52**, 14001–14005.
- 22 R. D. Chambers, J. A. Cunningham and D. J. Spring, *J. Chem. Soc. C*, 1968, 1560–1565.
- 23 F. G. Bordwell, *Acc. Chem. Res.*, 1988, **21**, 456–463.
- 24 L. Ekladius and H. K. King, *Biochem. J.*, 1957, **65**, 128–131.
- 25 L. D. Lavis, T. J. Rutkoski and R. T. Raines, *Anal. Chem.*, 2007, **79**, 6775–6782.
- 26 P. I. Kitov, D. F. Vinals, S. Ng, K. F. Tjhung and R. Derda, *J. Am. Chem. Soc.*, 2014, **136**, 8149–8152.
- 27 N. K. Bashiruddin and H. Suga, *Curr. Opin. Chem. Biol.*, 2015, **24**, 131–138.
- 28 Y. V. Guillen Schlippe, M. C. T. Hartman, K. Josephson and J. W. Szostak, *J. Am. Chem. Soc.*, 2012, **134**, 10469–10477.
- 29 P. Timmerman, J. Beld, W. C. Puijk and R. H. Meloen, *ChemBioChem*, 2005, **6**, 821–824.
- 30 S. Ng, E. Lin, P. I. Kitov, K. F. Tjhung, O. O. Gerlits, L. Deng, B. Kasper, A. Sood, B. M. Paschal, P. Zhang, C.-C. Ling, J. S. Klassen, C. J. Noren, L. K. Mahal, R. J. Woods, L. Coates and R. Derda, *J. Am. Chem. Soc.*, 2015, **137**, 5248–5251.
- 31 K. F. Tjhung, P. I. Kitov, S. Ng, E. N. Kitova, L. Deng, J. S. Klassen and R. Derda, *J. Am. Chem. Soc.*, 2016, **138**, 32–35.
- 32 E. V. Vinogradova, C. Zhang, A. M. Spokoyny, B. L. Pentelute and S. L. Buchwald, *Nature*, 2015, **526**, 687–691.
- 33 R. L. Lundblad, in *Chemical Reagents For Protein Modification*, Taylor and Francis Group, LLC, New York, 2014, ch. 7, pp. 217–337.
- 34 L. E. J. Smeenk, N. Dailly, H. Hiemstra, J. H. van Maarseveen and P. Timmerman, *Org. Lett.*, 2012, **14**, 1194–1197.
- 35 Y. Chen, C. M. Clouthier, K. Tsao, M. Strmiskova, H. Lachance and J. W. Keillor, *Angew. Chem., Int. Ed.*, 2014, **53**, 13785–13788.
- 36 A. Abbas, B. Xing and T.-P. Loh, *Angew. Chem., Int. Ed.*, 2014, **53**, 7491–7494.
- 37 F. Saito, H. Noda and J. W. Bode, *ACS Chem. Biol.*, 2015, **10**, 1026–1033.
- 38 D. M. Patterson, L. A. Nazarova and J. A. Prescher, *ACS Chem. Biol.*, 2014, **9**, 592–605.
- 39 S. P. Brown and A. B. Smith III, *J. Am. Chem. Soc.*, 2015, **137**, 4034–4037.
- 40 S. C. Alley, D. R. Benjamin, S. C. Jeffrey, N. M. Okeley, D. L. Meyer, R. J. Sanderson and P. D. Senter, *Bioconjugate Chem.*, 2008, **19**, 759–765.
- 41 A. D. Baldwin and K. L. Kiick, *Bioconjugate Chem.*, 2011, **22**, 1946–1953.
- 42 N. Toda, S. Asano and C. F. Barbas, *Angew. Chem., Int. Ed.*, 2013, **52**, 12592–12596.

