



Biomaterials for mRNA delivery

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Messenger RNA (mRNA) has recently emerged with remarkable potential as an effective alternative to DNA-based therapies because of several unique advantages. mRNA does not require nuclear entry for transfection activity and has a negligible chance of integrating into the host genome which excludes the possibility of potentially detrimental genomic alternations. Chemical modification of mRNA has further enhanced its stability and decreased its activation of innate immune responses. Additionally, mRNA has been found to have rapid expression and predictable kinetics. Nevertheless, the ubiquitous application of mRNA remains challenging given its unfavorable attributes, such as large size, negative charge and susceptibility to enzymatic degradation. Further refinement of mRNA delivery modalities is therefore essential for its development as a therapeutic tool. This review provides an exclusive overview of current state-of-the-art biomaterials and nanotechnology platforms for mRNA delivery, and discusses future prospects to bring these exciting technologies into clinical practice.

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Introduction

Messenger RNA (mRNA), a natural biomolecule, is a transient entity that mediates the translation of genetic information from genes encoded in DNA to proteins located throughout the cell. The physical and temporal qualities of mRNA have allowed its use as a safe genetic material for gene-based therapy which does not require genomic integration.^{1,2} It is suitable for this use because of its potential to avoid nuclear localization and also because it allows rapid protein expression even in non-dividing and hard-to-transfect cells (*e.g.* dendritic cells and macrophages). These properties make mRNA an attractive molecule for immunotherapy.³ Another advantage of mRNA as a genetic element is its predictable, consistent protein expression kinetics, especially compared to DNA transfection which follows random onset time courses.^{4–7} However, relative to the technologies developed for DNA delivery, mRNA delivery strategies still require much more improvement and testing. Fortunately some of the carriers and biomaterials established for DNA and small interfering RNA (siRNA) have also been exhibiting a promising foundation for the development of mRNA delivery technologies.

mRNA has been investigated for more than half a century, but its widespread applications in medical research and in the

development of novel therapeutic modalities have been limited due to its perceived instability, susceptibility to degradation, insufficient translatability and immunostimulatory effects.^{8–10} Such challenges have partially been resolved thanks to an improved understanding of the structure of mRNA and its relationship to mRNA stability, as well as the development of a variety of chemical modification methods.^{10–15} These breakthroughs have subsequently facilitated the synthesis of mRNA with many different structural modifications (*e.g.*, anti-reverse cap analogues (ARCA), 3'-globin UTR and poly-A tail), which still possess functional activity for immunotherapy and gene-based therapy. With these advances in stability and functionality, the use of mRNA as a therapeutic tool is now becoming a reality.¹⁴

Nevertheless, like other nucleic acids (*e.g.*, DNA and siRNA), naked mRNA cannot readily cross the cell membrane on its own and thus requires delivery systems to enhance its cell permeation.¹⁶ Viral vectors have been used as mRNA carriers but may suffer from their potential immunologic side effects and toxicity as well as the vector-size limitations.^{1,17} Non-viral strategies such as electroporation, gene gun and sonoporation have been more thoroughly investigated as mRNA delivery systems.^{16,18,19} However, the *ex vivo* manipulation of cells with mRNA transfection using such approaches, while feasible, is highly laborious, expensive and overall ill-suited for extensive applications.^{20–22}

Biomaterials represent an important step forward from the aforementioned non-viral strategies and have demonstrated promising potential for the delivery of various biomacromolecules such as DNA and siRNA.^{1,23} Compared to viral vectors and *ex vivo* technologies mentioned above, biomaterials are more biocompatible and diversified, and can be easily formu-

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lated for effective *in vivo* delivery and controlled release of therapeutics. Recently, biomaterials have also attracted considerable attention for mRNA delivery.^{5,24,25} For example, protamine has demonstrated remarkable abilities to ameliorate the transfection capabilities of mRNA and several protamine-mRNA complexes are now under clinical trials in cancer patients.^{26,27} Moreover, biomaterials-based nanoparticle platforms have in recent years been gradually applied to mRNA vaccine development and mRNA-based gene therapy.^{1,28,29}

In this review, we summarize the strategies for chemical modification of mRNA, provide an overview of the currently available biomaterials and nanotechnology platforms for mRNA delivery and a critical analysis of how these mRNA delivery systems may be further improved to potentiate their therapeutic utility and discuss the challenges and opportunities in this exciting field.

mRNA modification for clinical translation

Owing primarily to its instability, mRNA was originally considered to be unsuitable for use as a therapeutic molecule, despite the fact that research on mRNA delivery into cells was pioneered over three decades ago.^{30,31} Recent advances in molecular and structural biology, along with substantial pro-

gress in the understanding of mRNA biology and its degradation mechanism, have promoted the development of various chemical modification methods to improve mRNA stability and translation capacity.^{8,32} Below, we briefly outline the various modifications of mRNA's structure, including different capping techniques and elements that can be included to improve stability.

In eukaryotic cells, mature mRNA is generally comprised of five distinct portions (Fig. 1a): (i) a cap structure, (ii) a 5' untranslated region (5' UTR), (iii) an open reading frame (ORF), (iv) a 3' untranslated region (3' UTR) and (v) a poly(A) tail (a tail of 100–250 adenosine residues).^{33,34} With recent progress in molecular biology techniques, *in vitro* transcription is commonly utilized to produce functional mRNA using a bacteriophage promoter.⁵ During *in vitro* mRNA transcription it has been observed that almost half of the caps are oriented in reverse which makes them unrecognizable to the cap-binding protein.^{35,36} Anti-reverse cap analogs (ARCAs) were developed to solve this problem. These are modifications in which OCH₃ is used to replace or remove natural 3' OH cap groups to avoid inappropriate cap orientation (Fig. 1b and c).¹¹ It was found, moreover, that modification in the C2' position, as well as the C3' position, can also prevent inappropriate cap orientation.¹² Additional modifications of ARCA structure have been reported for the



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therapy and subunit antigens for vaccine therapy to prevent infectious diseases. He achieved several research awards for outstanding research performances include "Brain Korea 21 (BK21) Award" in 2012, the "Best Young Scientist Award" in 2013 from Seoul National University and some others in various scientific meetings. He has authored about 25 peer-reviewed articles and 3 of his research works have been issued/pending national and international patents. Currently, he is a Postdoctoral Fellow under Prof. Omid Farokhzad and Prof. Jinjun Shi at Harvard Medical School and Brigham & Women's Hospital (HMS/BWH), USA since Feb 2014, and working on developing mRNA nanotherapeutics for cancer immunotherapy & gene therapy, immunosenescence and asthma nanotherapeutics. Dr. Islam is a member of National Postdoctoral Association of USA and an Association of Young Biotechnologist of Bangladesh. He is also serving as a Co-Chair in the Communication Executive Committee of Postdoctoral Leadership Council of BWH at HMS.



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purpose of improving the efficiency and stability of mRNA.⁵ For example, tetraphosphate ARCAs are reported to improve translation efficiency relative to that of other cap analogs,¹² as well as phosphorothioate ARCAs, which confer hydrolysis resistance to mRNA, thus increasing translational stability.¹³

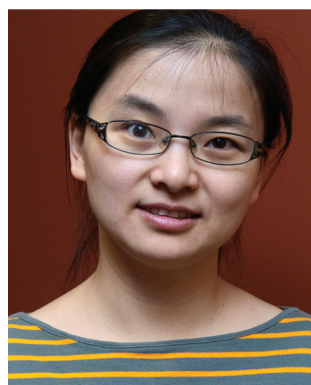
Polyadenylation, the addition of a poly adenine (A) tail to the 3' end of mRNA, is catalyzed by poly(A) polymerase and is part of the process that produces mature mRNA in eukaryote cells. The poly(A) tail acts as the binding site for poly(A)-binding protein (PABP) which is exported with mRNA from nucleus to cytoplasm where it further binds to and recruit proteins that facilitate translation, such as translation initiation factor 4F (eIF4F).³⁷ The poly(A) tail makes the mRNA molecule more stable as shortening or removal of the poly(A) tail accelerates mRNA degradation *via* enhanced exonucleotide digestion.³⁸ *E. coli* poly(A) polymerase (E-PAP) I has been optimized to add a poly(A) tail of at least 150 adenines to the 3' terminal of *in vitro* transcribed mRNA. In some reports, instead of enzymatically attached poly(A) tail, *in vitro* transcribed RNA generated from plasmid templates consists of coding sequencing and a poly(A) tail with 30 to 120 nucleotides in length.^{39–42} The additional adenine residues confer stability to the *in vitro* transcribed capped mRNA and may increase its translational efficiency.⁴³ Where direct comparisons have been made, longer poly(A) tails >100 adenines have generally been found to be preferable to smaller ones.^{40,41,44}

The widely studied adenylate–uridylylate rice elements (AREs) are important mRNA decay signals in the 3' untranslated regions (3' UTRs) of most eukaryotic mRNAs. mRNAs which contain AREs demonstrate reduced stability, perhaps due to

the removal of the poly(A) tail. Stability is increased, however, when AREs are replaced with the 3' UTR of a stable mRNA species such as β -globin mRNA.⁴⁵ Iron responsive elements (IREs) represent an additional type of 3' UTR which affect mRNA stability depending on their precise location in mRNA structure. In this case, the mRNA half-life increases when IREs occupy the 3' UTR, whereas when that are located at the 5' position, mRNA translational ability is improved.⁴⁶

The multiple means of modulating structural elements of the mRNA, including the 5' cap, 5'- and 3'-UTRs, the coding region, and the poly(A) tail, improve the intracellular stability and translational efficiency of *in vitro* transcribed mRNA. Although the degree of improvement for modulation of *in vitro* transcribed mRNA appears to depend on the means of modification, the cell type and cell differentiation state, reports range from modest levels of improvement to 2–3 orders of magnitude,^{12,40–42,47} which leads to the significantly higher protein expression and prolonged persistence of the protein from a range of a few minutes to longer than 1 week.^{40,47,48} Moreover, the incorporation of chemically modified nucleotides, such as substitution of cytidine triphosphate and uridine triphosphate with naturally occurring 5-methylcytidine and pseudouridine (ψ) triphosphate, respectively, into mRNA further suppresses the immune-stimulating property of *in vitro* transcribed mRNA.¹⁰

It is also noteworthy to clearly distinguish the different immune-stimulating effects that mRNA can have. First, there is an inherent response of the body to foreign, non-self mRNA. This is the type of response that the body uses to prevent threats such as RNA viruses, through the endosomal recognition.⁴⁹ These RNA molecules can be encountered through



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Dr Harshal R. Zope was born in Bhusawal, India on 26th Jan 1986. He pursued his interest in research by joining Institute of Biotechnology and Bioinformatics, University of Pune, India. In Pune University, he obtained a 5 years integrated M.Sc. degree in Biotechnology (2008) as well as M. Tech degree in Biotechnology (2009). Next, he pursued his interest by joining Leiden University for his PhD research under the supervision of Dr Alexander Kros (Feb 2010). During his PhD, he worked on peptide based functional biomaterials for various applications such as understanding membrane fusion, drug delivery and vaccine development. Currently, Harshal Zope is a Postdoctoral fellow at the Laboratory of Nanomedicine, Harvard Medical School/Brigham and Women's Hospital (HMS/BWH), Boston, USA. In the group of Prof. Omid Farokhzad and Prof. Jinjun Shi, he is developing a novel nanotechnology platform for HIV vaccine and cancer therapy.

markers for cancer progression and chemoresistance and dissecting the underlying molecular mechanisms.

immune sensors such as Toll-like receptors (TLRs).⁵⁰ Some of the aforementioned mRNA modifications are specifically designed to reduce this innate immune response to minimize inherent host response. Second, the delivered therapeutic mRNA molecules can induce the desired immune response through stimulating appropriate immune cells and presentation of the specific antigen. This can also be referred to as the immune-stimulating effect of the mRNA, but in this case is a designed antigen-specific effect as opposed to an inherent reaction to foreign materials. It is also important to mention that the innate immunity is not preferable for mRNA-based gene therapy applications, where we only need to transfer mRNA into the cells and facilitate the expression of the protein of interest for therapeutic purpose. The results of these actions will be henceforth discussed with a variety of mRNA molecules and delivery methods.^{10,51–53}

Naked mRNA, in different modified forms, has already been tested in clinical settings. The direct intradermal injections of naked mRNA were applied as a phase I/II non-randomized clinical trial in patients with stage IV renal cell cancer.⁵⁴ In this study, granulocyte-macrophage colony stimulating factor (GM-CSF) was used as an adjuvant with mRNA encoding several tumor-associated antigens including mucin 1, carcinoembryonic, human epidermal growth factor receptor 2, telomerase, survivin and melanoma-associated antigen 1. Results revealed that vaccinations were well tolerated with no severe side effects and induced clinical responses.⁵⁴ Recently, CureVac has developed self-adjuvanted two-component mRNA vaccines comprised of both free mRNA and protamine-com-

plexed mRNA for the treatment of castration-resistant prostate cancer and stage IIIB/IV non-small cell lung cancer. They have demonstrated excellent safety as well as high levels of cellular immunity with T and B cell responses in several clinical trials.^{55–59} Moderna is another company that has been investigating mRNA as a vaccine and therapeutic technology, with two ventures Onkaido Therapeutics and Valera actively exploring the discovery and development of mRNA-based treatments in oncology and infectious diseases, respectively. Some representative studies using modified naked mRNA are summarized in Table 1.

Whereas naked modified mRNA shows feasibility in their application as vaccines, several challenging issues remain elusive, such as enzymatic degradation, rapid elimination by renal excretion or by the mononuclear phagocyte system (MPS), as well as poor cellular uptake and endosomal escape, in particular for systemic mRNA delivery. Hence, to achieve more efficient mRNA delivery, a great number of biomaterials and nanoparticle platforms have been developed to protect mRNA from nuclease degradation and facilitate its cytosolic delivery for improved protein expression (Fig. 2 and 3). In the remainder of this review, we focus on recent advances in the delivery of mRNA to cells and tissues by means of contemporary biomaterials and nanotechnologies, including protamine complexes, lipid nanoparticles, polymeric nanoparticles, lipid-polymer hybrid nanoparticles, and gold nanoparticles.

Protamine–mRNA complexes

Protamine is a natural cationic protein, which gives it an excellent ability to complex nucleic acid including mRNA (Fig. 3a)



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Dr Bruce Zetter is the Charles Nowiszwski Professor of Cancer Biology at Boston Children's Hospital and Harvard Medical School. He is highly regarded nationally and internationally as a leader in the research of tumor progression, cancer diagnosis, cancer metastasis, and tumor angiogenesis. His current research interests are focused on tumor metastasis, novel cancer treatments and on tests that predict future outcomes for



Jinjun Shi

Dr Shi has developed many multifunctional nanoparticle systems for the delivery of chemotherapeutics, proteins, vaccines, and nucleic acids, one of which has led to the first-in-human clinical trial of a synthetic nanoparticle vaccine for smoking cessation and relapse prevention. He has authored more than 30 papers, and holds over 25 issued/pending national and international patents. He has also received many awards, such as the NIH K99/R00 Career Development Award, the AACR Scholar-in-Training Award, the Movember-PCF Challenge Award, and the PCF Young Investigator Award.

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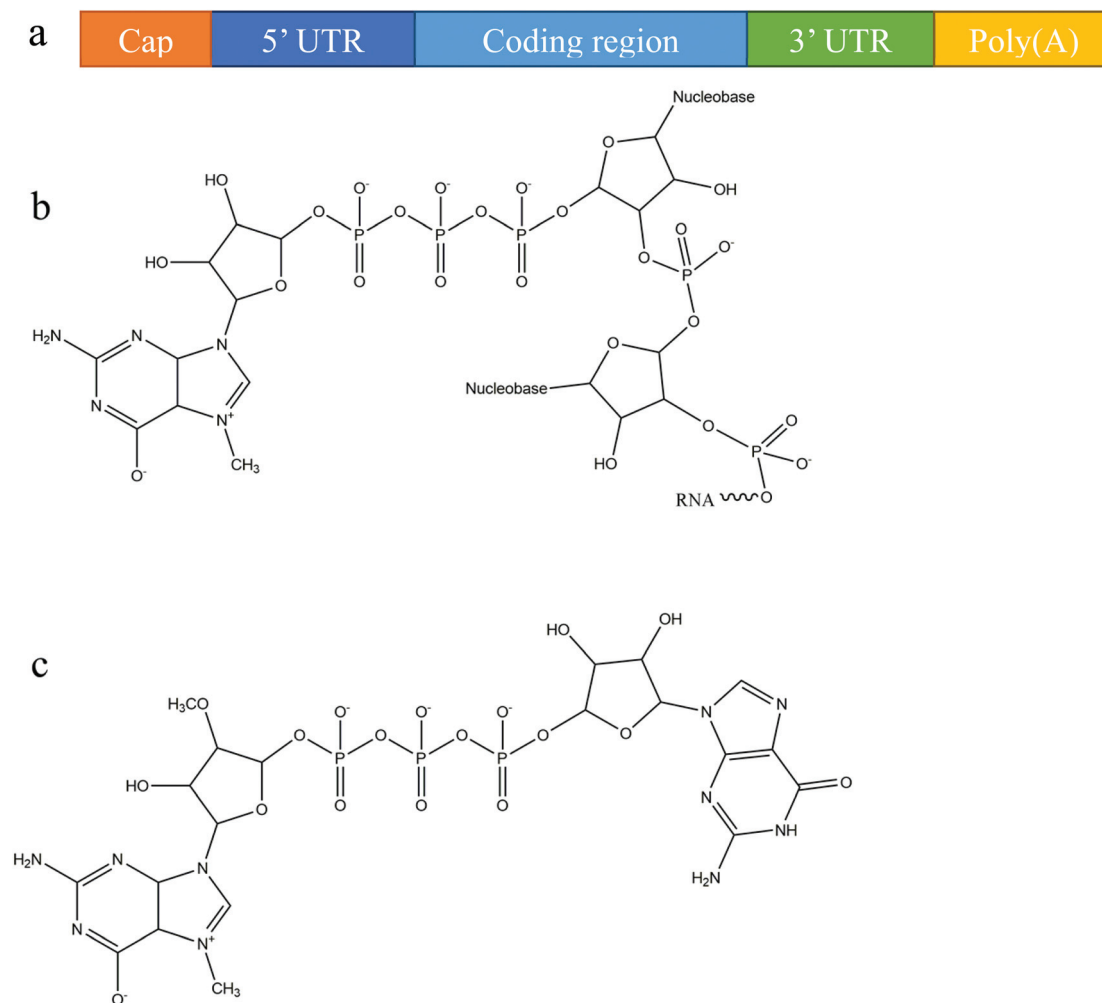


Fig. 1 mRNA structural features: (a) a basic structure of a eukaryotic mRNA with five distinct components, (b) the cap structure of the 5' end of eukaryotic mRNA, and (c) the structure of anti-reverse cap analogues (ARCA): 3'-O-Me-m7G(5')ppp(5')G.

Table 1 Naked mRNA and its activity in animal models and patients

Category	Stage	mRNA	Activity and efficiency	Ref.
Naked mRNA	C57BL/6 mice	Luciferase and human CEA	After intramuscular injection mice demonstrated anti-CEA antibody response	119
	CAF ₁ (H-2 ^a) mice	S1509a	Intradermal injection induced immunity in tumors	120
	Mice	Chloramphenicol acetyltransferase and luciferase	After intramuscular injection CAT activity was detected and, in a separate experiment, a dose-response effect with luciferase was determined	121
Naked mRNA with GM-CSF	Genetically engineered mice (C57BL6/CFW background)	Human VEGF-A	After intramyocardial injection there was improved heart function and long-term survival because of the directed differentiation of endogenous heart progenitors	122
	Patients with stage IIIB/IV non-small cell lung cancer	MAGE-C1, MAGE-C2, NY-ESO-1, BIRC5, 5T4	No dose limiting toxicity and good safety profile, along with significant induction of T and B cell responses in 65% of patients.	59
Naked mRNA with GM-CSF	Patients with stage IV renal cell cancer	MUC1, CEA, Her-2/neu, telomerase, survivin and MAGE-A1	The intradermal mRNA injections resulted in clinical response without any severe side effects and induced CD4 ⁺ and CD8 ⁺ T cell responses for the specific antigens.	54
	Patients with stage III and IV metastatic melanoma	Autologous mRNA	Intradermal injection; safe and feasible; increase in antitumor humoral immune response in some patients;	24

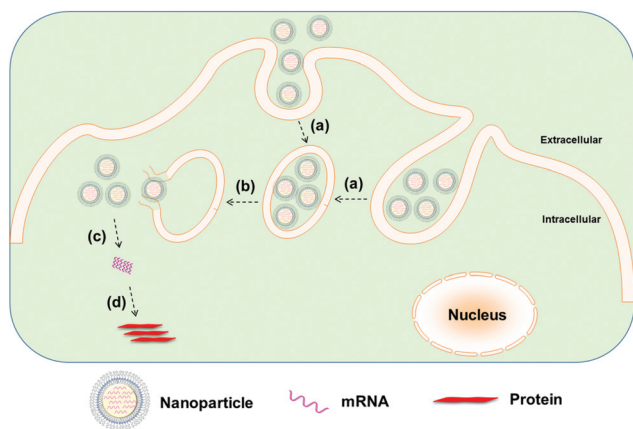


Fig. 2 Nanoparticle-mediated delivery of mRNA: (a) cellular internalization of nanoparticles into endosome, (b) endosomal escape, (c) release of mRNA from nanoparticles, and (d) mRNA translation to protein without genomic integration.

and provides increased uptake of mRNA and transfection capabilities.⁶⁰ It has been shown that protamine can effectively complex with mRNA and these complexes can act as a danger signal that activates murine cells through a MyD88-dependent pathway involving TLR7 and TLR8.⁶⁰ Hoerr *et al.* demonstrated that these complexes degrade within 2 h when incubated in serum, which limits their abilities for survival in the circulation *in vivo*.⁵³ Nevertheless, they later showed that even partially degraded mRNA–protamine complexes can still exhibit immunostimulatory activity for over 100 h.⁶⁰ Moreover, it was found that protamine–mRNA complexes strongly activated a variety of white blood cells (such as granulocytes, B cells and NK cells) and overall significantly stimulated immune response compared to that of protamine–DNA systems.^{53,60} In a clinical study by Weide *et al.* the efficacy of this system was demonstrated by testing intradermally administered protamine-complexed mRNAs coding for Melan-A (a melanoma antigen), Tyrosinase (an enzyme which catalyzes the pro-

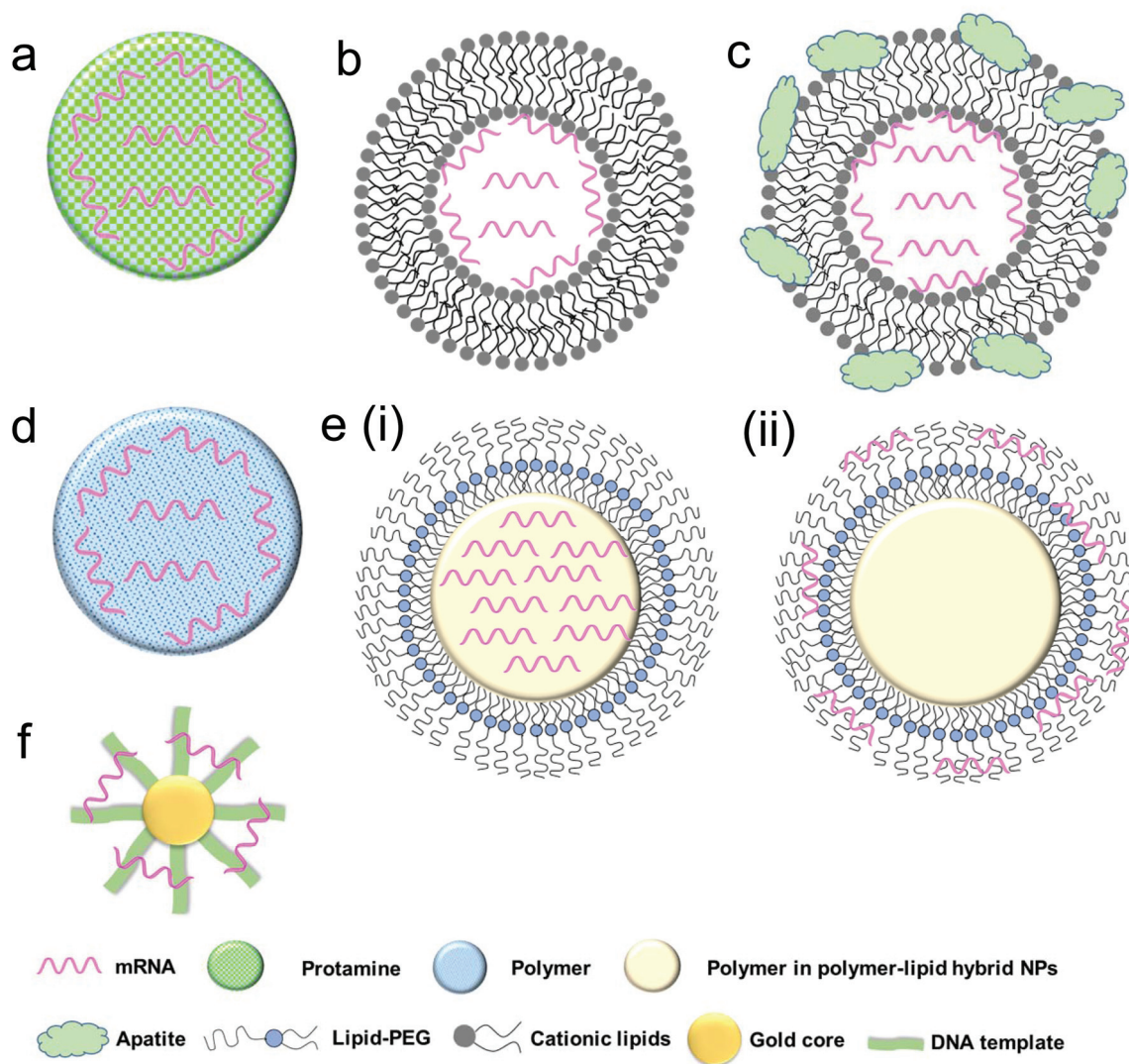


Fig. 3 Schematic representation of various biomaterial-based systems for mRNA delivery: (a) protamine–mRNA complex; (b) lipid nanoparticle; (c) lipid nanoparticle with inorganic compounds (e.g. apatite); (d) cationic polymeric nanoparticle; (e) lipid–polymer hybrid nanoparticles including (i) mRNA–polymer complex core surrounded by a lipid shell and (ii) polymer core surrounded by a lipid shell with mRNA absorbed onto the surface; and (f) gold nanoparticle.

duction of melanin), gp100 (a protein involved in melanosome maturation), Mage-A1 (a melanoma antigen), Mage-A3 (a melanoma antigen) and Survivin (a protein involved in the regulation of apoptosis) metastatic melanoma patients.²⁶ No side effects greater than grade II (mild to moderate) were observed and overall there was a complete clinical response, including significant effect on the frequency of immunosuppressive cells and increase of antigen-specific T cells.²⁶

Recently, CureVac has also explored the use of protamine-complexed mRNA. It was found in an *in vivo* study that a two-component mRNA vaccine composed of both free and protamine-complexed mRNA resulted in successful antigen expression and immune stimulation, which were mediated by Toll-like receptor 7.⁵¹ A balanced adaptive immunity with both cellular (T cell-mediated) and humoral immunity was achieved which not only showed prophylactic activity but also exhibited therapeutic efficacy against tumor. CureVac has also put forward a clinical trial for patients with castrate-resistant prostate cancer using mRNA-encoding for prostate-specific antigen (PSA), prostate-specific membrane antigen (PSMA), prostate stem cell antigen (PSCA) and six transmembrane epithelial antigen of the prostate 1 (STEAP1).²⁷ After intradermal vaccination, 80% of the subjects demonstrated immune response to the delivered mRNA antigen and 60% against multiple antigens, which ultimately correlated with longer survival.²⁷ The currently investigated protamine-complexed mRNA systems and their therapeutic efficacy are summarized in Table 2.

Lipid nanoparticles

The field of lipid nanoparticles for nucleic acid delivery is relatively mature compared to other nanotechnologies. As such, quite a variety of different lipid formulations have been tested for mRNA delivery. Cationic lipids are used ubiquitously owing to their favourable electrostatic interactions with negatively charged mRNA to form nanoparticles (Fig. 3b). This field was pioneered in 1989 with a study of the use of DOTMA (*N*-[1-(2,3-dioleoyloxy)propyl]-*N,N,N*-trimethylammonium chloride) to transfect human, rat, mouse, xenopus (frog) and drosophila cells with luciferase mRNA.³¹ The clinical development of such cationic lipid tools has however been hampered by their toxicity.³¹ Despite these concerns, DOTMA remains a commonly used material, along with DOTAP (1,2-dioleoyloxy-3-trimethylammonium propane chloride).⁶¹ An important note about these cationic lipids is that while positively charged lipid nanoparticles can be effective *in vitro*, *in vivo* results are less promising because cationic liposomes can be quickly eliminated by the mononuclear phagocyte system (MPS).

The coating of poly(ethylene) glycol (PEG) layer on lipid carriers has been widely used for the delivery of nucleic acid payloads, such as siRNA⁶² and DNA,⁶³ to improve the formulation process, reduce aggregation and increase the blood circulation time.^{64–67} However, the surface PEGylation has also been shown to decrease cellular uptake, an effect which may be minimized by optimizing PEG size and content.^{63,68,69} PEG modification has also been applied to nanoparticle-based

mRNA delivery, particularly to polymer nanoparticles⁷⁰ and lipid-polymer hybrid nanoparticles,^{71,72} both of which will be discussed in later sections. The application of PEGylation to lipid-based mRNA delivery systems to improve their *in vivo* efficacy remains elusive. Another strategy is the incorporation of a helper lipid such as 1,2-dioleoyl-*sn*-glycero-3-phosphoethanolamine (DOPE) to reduce aggregation of the lipid systems (as well as to improve endosomal escape).^{73–76}

To further enhance mRNA transfection potency in both mitotic and non-mitotic cells, Akaike and co-workers developed a different approach by coating inorganic carbonate apatite nanoparticles on liposomal carriers (Fig. 3c). It was evident that these inorganic additives helped to increase mRNA uptake through effective endocytosis. They also demonstrated that decorating mRNA-containing DOTAP-apatite particles with RGD, which is known for its ability to bind integrins,⁷⁷ enhanced cytoplasmic expression of delivered mRNA. This method is versatile and holds a great potential due to its abilities for targeting and efficient delivery of mRNA.^{61,78–80}

More recently, a novel lipid/protamine/mRNA nanoparticle platform was proposed and comprehensively explored for systemic delivery to tumors.¹⁰ In this study DOTAP liposomes were used to encapsulate mRNA-protamine complexes and then coated with DSPE-PEG and DSPE-PEG-anisamide.¹⁰ These particles demonstrated stability against degradation in serum, high *in vitro* transfection ability in NCI-H460 cells, low cytotoxicity (even at a concentration 50 times higher than the dose they found to induce effective transfection), accumulation in tumor site and anticancer action *in vivo*.¹⁰ Together this approach presents a very promising hybrid system for the systemic delivery of mRNA. Currently, Tekmira Pharmaceuticals is also investigating lipid nanoparticles for systemic mRNA delivery to the liver.

Until now, lipid nanoparticles have been widely utilized, particularly to introduce mRNA to immune cells for vaccine purposes. The detailed overview of currently available lipid-based nanoparticles for mRNA delivery was presented in several recent reports,^{3,16,25,81} and a summary of these advances is presented in Table 2. As the lipid nanoparticles represent a more thoroughly explored system, their further development and optimization may open up the foundation for the creation of more effective mRNA delivery systems.

Polymeric nanoparticles

Polymeric nanoparticles have emerged as effective delivery vehicles for a variety of payloads, such as DNA,⁸² siRNA,⁸³ mRNA,²⁸ proteins,⁸⁴ chemotherapeutic agents⁸⁵ and others. To date, cationic polymers have been used primarily because of their favourable electrostatic interactions with negatively charged nucleic acids (Fig. 3d) and cell membranes. As compared with other delivery vehicles such as liposomes, multiple parameters must be considered in their design. Pack *et al.* enumerated some of these in their recent report in which they emphasized optimization with respect to synthesis of mRNA carriers, transfection and endosomal escape capabilities.⁸⁶

Table 2 Biomaterials for mRNA delivery

Category	Platform	Stage	mRNA	Activity and efficiency	Ref.
Protamine–mRNA complex	Protamine complexed β -gal mRNA	HeLa-K ^b cells injected into B6 (H2 ^b) mice and BALB/c mice	β -Gal and GFP	Successful CTL response, dependent on injection site	53
		Human PBMC	β -Gal, EGFP or CMV pp65	Complexes triggered strong IL-6 and TNF- α release, activation of innate immune system and other APCs	60
		Murine BM-DC	β -Gal or CMV pp65	Stimulated mouse BM-DC: release of IL-6 and IL-12 and up-regulation of CD86	123
	Protamine complexed Melan-A, Tyrosinase, gp100, Mage-A1, Mage-A3, and Survivin mRNA	Patients with metastatic melanoma	Melan-A, Tyrosinase, gp100, Mage-A1, Mage-A3, and Survivin	Increased frequency of immunosuppressive and vaccine-directed T cells	26
		BALB/c and C57BL/6 mice	OVA (GgOVA), control (Ec β -gal sh), PSMA (HsPSMA) and STEAP (HsSTEAP)	Demonstrated antitumor effects after intravenous injection; activated the adaptive and innate immune systems	51
	CureVac® (CV9201)	Patients with stage IIIB/IV non-small cell lung cancer	MAGE-C1, MAGE-C2, NY-ESO-1, survivin and 5T4	Ability to inhibit established tumors	58
	CureVac® (CV9202)	Patients with stage IV non-small cell lung cancer	NY-ESO-1, MAGEC1, MAGEC2, 5 T4, Survivin, and MUC1	This is a phase I/IIa clinical trial	57
	CureVac® (CV9202) in combination with radiotherapy	Patients with stage IV non-small cell lung cancer	NY-ESO-1, MAGEC1, MAGEC2, 5 T4, Survivin, and MUC1	mRNA vaccines potentially induced immune response to antigens expressed in tumors	57
		Patients with castrate-resistant prostate cancer	PSA, PSCA, PSMA and STEAP1	19 patients have been recruited thus far, recruitment finished by the end of 2014	27,55
	CureVac® (CV9103)			Intradermal injections; maximum tolerated dose not defined; activation of TLR7	
CureVac® (CV9104)	Patients with metastatic castrate-refractory prostate cancer	mRNA-encoding six antigens (this is an advancement of CV9103 encoding two more antigens)	Antigen-specific T-cells were detected in 79% of patients and 58% of the immunological responders reacted against multiple antigens; Increased antigen-specific T-cells and antigen-unspecific B cells in 74% of patients	27,124	
Lipid nanoparticle	DOTAP liposomes covered with apatite nanoparticles	HeLa	Luciferase	Together with ARCA (over cap) had over 100-fold improvement compare to DOTAP, percentage not determined	78
		HeLa	Luciferase	9–14 better than mRNA liposome alone, percentage not determined	79
		NIH 3T3 HeLa	Luciferase	DOTAP-apatite outperformed DOTAP alone and Lipofectamine 2000, percentage not determined	80
	Fibronectin complexed to DOTAP liposomes covered with apatite nanoparticles	HUVEC	Luciferase	Fn-DOTAP-apatite more than 50-times better than DOTAP, percentage not determined	61
		HeLa	Luciferase		
	DOTAP/DOPE/DSPE-PEG-2000-biotin	Primary murine bone marrow-derived DC from C57BL/6 mice	TriMix mRNA encoding CD40-ligand, TLR4 and CD70	19%	19
	DOTAP/DOPE/DSPE-PEG-2000-biotin lipoplex loaded microbubbles	DC primary cultures from the bone marrow of C57BL/6 mice	Luciferase	24%	21
DOTMA	NIH 3T3	Luciferase	Percentage not determined, at least 20–30% RNase resistant	31	

Table 2 (Contd.)

Category	Platform	Stage	mRNA	Activity and efficiency	Ref.
	Lipofectamine 2000 and TransIT	Neurospheres from subventricular zone of adult C57BL/6 mice	EGFP	40–50%	125
	MLRI/DOPE and TransFast	CHO NIH 3T3	GFP and luciferase	>50% >45%	126
	Novel cationic lipids: X2, S1, S2, S3, 2X3 and 2D3 with DOPE (helper lipid)	DC cells cultured from the bone marrow of C57BL/6 mice	EGFP and B-16	Up to 47% of DC progenitors	127
	DOTAP/cholesterol liposome with DSPE-PEG and DSPE-PEG-AA, encapsulating protamine/mRNA cores	NCI-H460 xenograft	Herpes simplex virus 1-thymidine kinase (HSV1-tk)	Up to 57% of immature DCs 68–78%	10
	DOTAP/DOPE	HeLa	Luciferase, GFP and CXCR4	~80%	92
	Stemfect	JAWS II DC2.4 Human primary DCs Murine primary DCs	Luciferase and GFP Luciferase and GFP Luciferase and GFP Luciferase and GFP	80% >97% >50% >60%	128
Polymeric nanoparticle	Linear PEI	BEAS-2B	Luciferase	5-Fold higher than Lipo 2000	70
	Poly(DMAEMA-co-OEGMA)	BEAS-2B	Luciferase	Same level as Lipo 2000	
	Poly(DMAEMA) with PEG	BEAS-2B	Luciferase	3-Fold improvement after PEGylation	
	Branched PEI (25 kDa)	PC3	GFP	~30%	93
	Linear PEI	HeLa	Luciferase, GFP and CXCR4	40%	92
	Branched PEI (2 kDa) conjugated to melittin, with chloroquine	HeLa	GFP	52.2%	91
	Triblock copolymer (comprising DMAEMA, PEGMA, DEAEMA and BMA)	HUVEC DC2.4 RAW264.7	EGFP and OVA	71.6% 50% 77%	28
Polymer-lipid hybrid nanoparticle	Poly-(β -amino ester) core, phospholipid bilayer shell	DC2.4	Luciferase and GFP	30%	106
	Mannosylated histidylated lipopolyplexes	DC2.4	EGFP and MART-1	60%	71
	DMDHP	CHO	Luciferase and GFP	70–80%	129
Gold nanoparticle	DNA oligonucleotide-conjugated gold nanoparticle	HeLa HepG2	BAX	Fluorescent signal uniformly detected, percentage not determined.	29

Abbreviations: (*N,N*-[bis(2-hydroxyethyl)]-*N*-[2,3-bis(tetradecanoyloxy) propyl] ammonium chloride (DMDHP); 1,2-dioleoyl-3-trimethylammonium-propane (DOTAP); 1,2-dioleoyl-*sn*-glycero-3-phosphoethanolamine (DOPE); 1,2-distearoyl-*sn*-glycero-3-phosphoethanolamine-*N*-[biotinyl(polyethylene glycol)-2000] (ammonium salt); 1,2-distearoyl-phosphatidylethanolamine-polyethylene glycol (DSPE-PEG); 1,2-distearoyl-phosphatidylethanolamine-polyethylene glycol-anisamide (DSPE-PEG-AA); Antigen-presenting cell (APC); Anti-Reverse Cap Analog (ARCA); Bone-marrow-derived dendritic cell (BM-DC); Carcinoembryonic antigen (CEA); Cluster of differentiation 70 (CD70); Cluster of differentiation 86 (CD86); C-X-C chemokine receptor type 4 (CXCR-4); Cytomegalovirus (CMV); Cytotoxic T cell (CTL); Dendritic cell (DC); Diethylaminoethyl methacrylate (DEAEMA) and butyl methacrylate (BMA); Diethylaminoethyl methacrylate (DMAEMA); Enhanced green fluorescent protein (EGFP); Granulocyte-macrophage colony stimulating factor (GM-CSF); Green fluorescent protein (GFP); Human bronchial epithelia cell line (BEAS-2B); Human epidermal growth factor receptor 2 (Her-2/neu); Interleukin 12 (IL-12); Interleukin 6 (IL-6); Melanoma-associated antigen 1 (MAGE-A1); Melanoma-associated antigen family C1 (MAGE-C1); Melanoma-associated antigen family C2 (MAGE-C2); Mucin 1 (MUC1); New York esophageal squamous cell carcinoma 1 (NY-ESO-1); *N*-[1-(2,3-dioleoyloxy)propyl]-*N,N,N*-trimethylammonium chloride (DOTMA); Oligo(ethylene glycol) methyl ether methacrylate (OEGMA); Ovalbumin (OVA); Peripheral blood mononuclear cell (PBMC); Poly(ethylene glycol) methyl ether methacrylate (PEGMA); Polyethylenimine (PEI); Prostate-specific antigen (PSA); Prostate-specific membrane antigen (PSMA); Prostate stem cell antigen (PSCA); Six transmembrane epithelial antigen of the prostate 1 (STEAP1); Toll-like receptor (TLR); Tumor necrosis factor alpha (TNF- α); Tumor-associated antigen (TAA); Vascular endothelial growth factor-A (VEGF-A); β -Galactosidase (β -gal); Trophoblast glycoprotein (5T4).

Additionally, polymers to be used for pharmaceutical purposes should be biocompatible, non-toxic and non-immunogenic.

Generally, polymeric nanoparticles are prepared by nanoprecipitation^{87,88} or emulsion techniques.^{89,90} When properly optimized, these polymeric carriers may serve as a highly functional delivery vehicle for mRNA. The surfaces can be easily modified with other materials such as ligands to confer targeting abilities and other polymers to afford properties such as pH-responsive release for tuning the release profile.⁹¹ The particles themselves must overcome extra- and intracellular barriers to ensure that the mRNA is available for protein expression (Fig. 2). Additionally, specific to polymeric particles, the charge and size of the polymers used must be carefully considered since the binding strength between the mRNA and polymer is a key factor in mRNA expression efficiency.⁹¹

Among various polymeric carriers for gene delivery, polyethylenimine (PEI) (Fig. 4a) is a classic material and is always used as a standard control in nucleic acid transfection. Rejman *et al.* explored the use of linear PEI polyplexes in HeLa cells and found moderate mRNA transfection efficiency of approximately 40% using C-X-C chemokine receptor (CXCR) type 4. This was exceeded by a cationic lipid formulation (DOTAP/DOPE) (Fig. 4b and c).⁹² However, concerns regarding PEI include non-degradability and potential toxicity (depending on the molecular weight and type of PEI). Recently, a few different studies have investigated the efficiency of modified PEI as an mRNA delivery system with high transfection efficiency and reduced toxicity. Read *et al.* explored the use of reducible polycations with histidine and polylysine residues (HIS-RPCs) and, with their optimised formulations, found

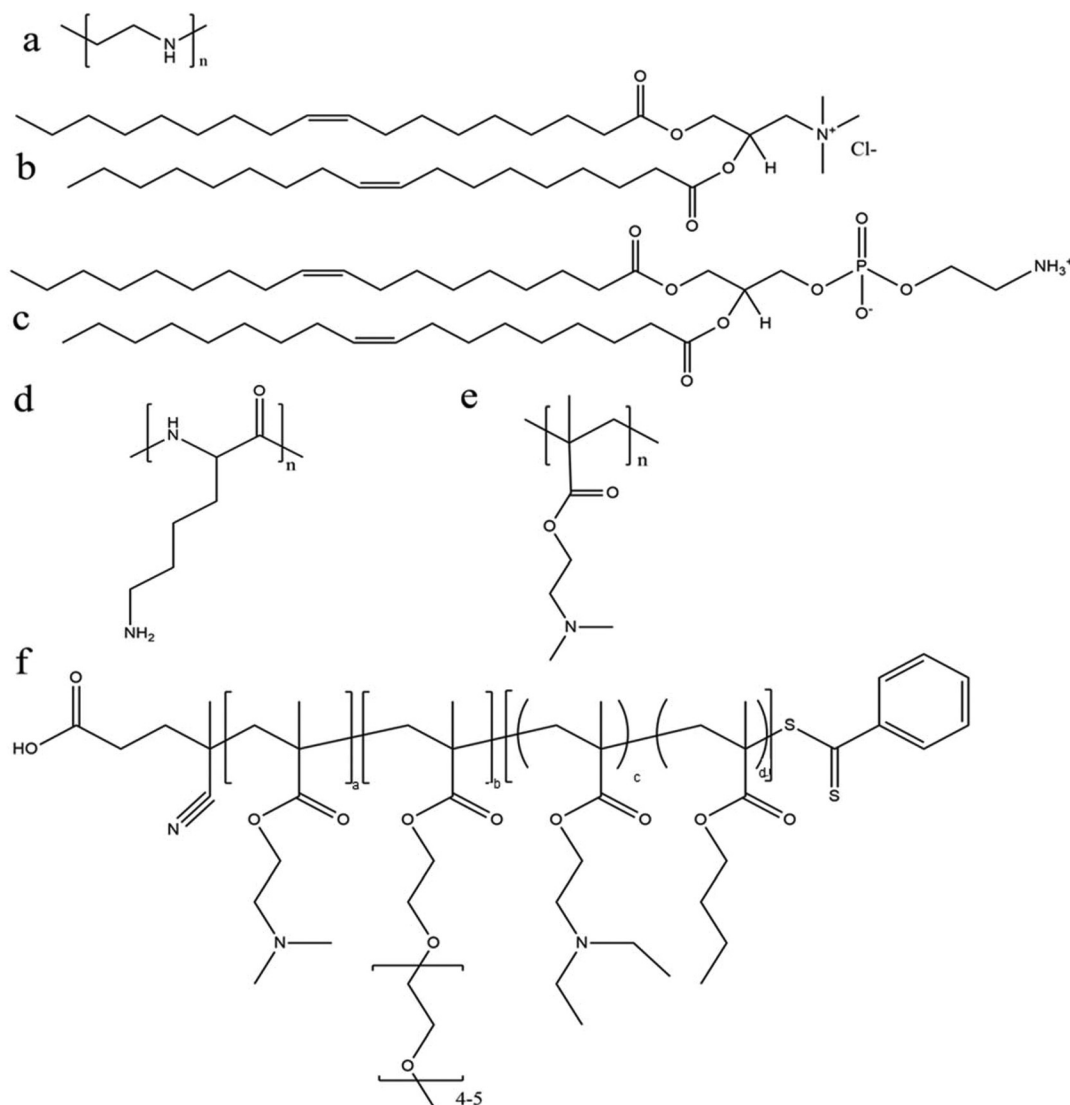


Fig. 4 Chemical structure of some representative polymers and lipids used for mRNA delivery: (a) polyethylenimine (PEI); (b) 1,2-dioleoyl-3-trimethylammonium-propane (chloride salt) (DOTAP); (c) 1,2-dioleoyl-*sn*-glycero-3-phosphoethanolamine (DOPE); (d) poly(L-lysine) (PLL); (e) poly(2-(dimethylamino)ethyl methacrylate) (p(DMAEMA)); and (f) poly(DMAEMA-PEGMA-DEAEMA-co-BMA).

transfection efficiencies of over 90% in human prostate cancer cells (PC3).⁹³ In contrast, they found that nanoparticles made of unmodified branched PEI (25 kDa) were only able to achieve efficiencies of about 30%.⁹³ Bettinger *et al.* also investigated the transfection activity of PEI (branched but with low molecular weight of 2 kDa) along with another well-known cationic polymer, poly(L-lysine) (Fig. 4d). In this study, they included additive compounds to increase transfection capability of PEI and, here, they found that the polymer in the presence of chloroquine (an agent to aid endosomolysis) and in conjugation to melittin (a membrane-active peptide) significantly increased the transfection efficiency.⁹¹ With these additives, transfection efficiencies of 52.2% and 71.6% were achieved in HeLa and HUVEC cells, respectively, and were greater than with the cationic lipid (DOTAP) alone.⁹¹ The effect of molecular weight of PEI was also investigated with the conclusion that overall higher MW polymers had better endosomolytic activity but bound mRNA too tightly for it to be released.⁹¹ These studies and their conflicting results also bring forward the important note that the efficacy and utility of a polymer candidate can be markedly improved through the use of additives and tuning of the MW and size of the particle.

The effect of PEGylation on the polymer–mRNA binding efficacy and mRNA transfection abilities was also investigated. It was demonstrated that adding PEG side chains to poly(*N,N*-diethylaminoethyl methacrylate) (p(DMAEMA)) (Fig. 4e) increased its ability to complex mRNA and tendency to form monodispersed particles, as compared to their unmodified variants as well as linear PEI (used as a positive control).⁷⁰ Nonetheless, linear PEI displayed the highest mRNA transfection capabilities and the PEGylated mRNA nanoparticle exhibited significantly higher transfection capability over its non-PEGylated counterpart.⁷⁰ Moreover, through testing with an influenza-peptide 7 (an endolysosomal release peptide), it was found that the PEGylated copolymer further supported the endosomal release of their mRNA payload and improved transfection efficiency,⁷⁰ suggesting further investigation into this polymer chemistry may be fruitful.

With rapid advances in polymer chemistry, a vast variety of other cationic polymers have been synthesized and applied to mRNA delivery. Recently a novel triblock polymer has been developed (Fig. 4f) using: (i) diethylaminoethyl methacrylate (DMAEMA) to promote mRNA condensation, (ii) poly(ethylene glycol) methyl ether methacrylate (PEGMA) to enhance stability and biocompatibility and (iii) a copolymer of diethylaminoethyl methacrylate (DEAEMA) and butyl methacrylate (BMA) to facilitate cytosolic entry.²⁸ The triblock polymer nanoparticles were ranged in size from 86 to 216 nm after complexation with mRNA and achieved transfection efficiency of up to 77% in RAW264.7 macrophage cells and 50% in DC2.4 dendritic cells, compared with approximately 30% in both cell lines where the commercial transfection agent Lipofectamine 2000 was used.²⁸ A further study was performed using a number of differing formulations made of the similar three polymers and found that the optimal arrangement was DMAEMA-PEGMA-DEAEMA-*co*-BMA, which provided an ideal

balance between stability and charge shielding.²⁸ With this arrangement, the copolymers showed decreased cytotoxicity *in vitro* with the decreasing of molecular weights for the second (PEGMA) block.²⁸

The use of polymeric nanoparticles for mRNA delivery is still in its infancy but has shown great potential. The field of polymeric mRNA delivery began with the use of diethylaminoethyl-dextran³⁰ and has now evolved to rival the abilities of many well-established lipid systems. Various novel polymeric carriers as mRNA delivery systems are summarized in Table 2. The future of this field largely depends on the further discovery and optimization of more efficient polymers to improve mRNA transfection efficiency, as well as safety and biocompatibility.

Lipid–polymer hybrid nanoparticles

The lipid–polymer hybrid nanoparticle platform has several potential advantages when compared with either lipid- or polymer-based nanoparticles for the delivery of therapeutic compounds including small molecules, peptides, proteins and oligonucleotides, and has gained significant attention in last decade.^{94–98} Structurally, the core of the nanoparticle is made up of a polymeric material coated with lipids and/or lipid–PEGs which can enhance the nanoparticle stability and pharmacokinetics, and confer surface tunable properties.^{94,99–101} Alternatively, the properties of the core can be tuned in order to respond to external stimuli which can help to facilitate endosomal escape.^{71,102,103} We have also recently reported the development of several such systems which exhibited sustained delivery of siRNA and enhanced gene silencing.^{104,105} Lipid–polymer hybrid nanoparticle systems have also been employed as delivery vehicles for mRNA.^{71,72,102}

There are two structural strategies which have been employed to design lipid–polymer hybrid nanoparticles for mRNA delivery. In the first method, negatively charged mRNA is complexed with the cationic polymer and then the surface can be decorated with lipids or lipid–PEG conjugates (Fig. 3e (i)).^{10,106} In the second method, mRNA is adsorbed to the surface of the cationic nanoparticle core (Fig. 3e (ii)).¹⁰⁷ Using both methods, mRNA delivery can be achieved; however release profiles may vary and it has been hypothesized by Su *et al.*¹⁰² that surface adsorbed mRNA has a faster release profile. They also showed that adsorbed mRNA has increased stability compared to naked mRNA which suggests that even if the mRNA is on the surface of nanoparticle, degradation can still be reduced due to the complexation.

To increase endosomal mRNA delivery, various pH-responsive polymers have been used. Upon endocytosis, changes in pH cause a conformational change in the polymer which results in osmotic shock followed by endosome disruption. For example, a PEGylated derivative of histidylated polylysine and L-histidine-(*N,N*-di-*n*-hexadecylamine)ethylamide liposomes (termed as ‘histidylated lipopolyplexes’) was prepared for an mRNA based cancer vaccine in which polylysine was used to complex mRNA, and histidylation contributed in charge switching at endosomal

pH condition due to the imidazole group with pI of 6. Additionally, they found that mannosylation of this delivery system significantly enabled targeted mRNA delivery to dendritic cells through interaction with the mannose receptor.⁷¹ The PEGylated histidylated mRNA lipopolyplexes were also efficient in inducing an anti-B16 specific cellular immune response and conferring protection against B16F10 melanoma in mice.⁷² In another report, poly- β -amino ester (PBAE), developed by Su *et al.*, also used pH-responsiveness to disrupt endosomal membrane.¹⁰⁶ The PBAE polymer was coated with lipids containing DOTAP which was used to adsorb mRNA on the particle as described earlier. Both of the pH-responsive systems have great potential for developing an effective delivery system for mRNA.¹⁰²

A summary of the advances on lipid-polymer hybrid nanoparticles is presented in Table 2. As of now it is difficult to postulate which is the most efficient method for mRNA delivery since there is a lack of sufficient studies to compare different mRNA delivery systems and determine the optimum one. Evidently, there is a compelling need to investigate various delivery options and establish quick optimization methods to screen a large number of mRNA delivery systems at once. This would accelerate the developmental progress of this highly promising biomedical research field.

Gold nanoparticles

Another type of nanoparticle delivery system for mRNA is gold nanoparticle-DNA oligonucleotide (AuNP-DNA) conjugates (Fig. 3f).²⁹ This system has demonstrated significant transfection activity in HeLa (cervical carcinoma) and HepG2 (hepatocyte carcinoma) cells (detected using confocal fluorescent microscopy). The nanoparticles were tested *in vivo* using direct injection into xenograft tumors and the results showed that the (AuNP-DNA conjugate)-mRNA nanoparticles were able to induce the production of biologically functionalized Bcl-2-associated X (BAX) protein,²⁹ suggesting a promising start for this mRNA nanoparticle formulation. While gold nanoparticles have not been extensively studied for mRNA delivery, their promising results with other nucleic acids (*e.g.*, siRNA and DNA) indicate that they have significant potential for the delivery of mRNA.

Indeed, the gold nanoparticle-oligonucleotide system has been more widely studied with DNA and siRNA.^{108–110} This system has demonstrated significant cellular uptake in a number of cell lines.^{109,111–113} Recently, the mechanism behind the uptake of gold oligonucleotide particles was further elucidated. It has been shown that their uptake is highest in serum-free conditions and the membrane proteins responsible for the uptake of these particles are the scavenger receptors.¹¹¹ Other delivery systems based upon gold nanoparticles include gold nanoparticles passivated with BSA-SV40 large T antigen conjugates,¹¹⁴ layer-by-layer gold nanoparticles with polymers for the delivery of siRNA^{115,116} and plasmid DNA delivery using carbon dot-gold nanoparticles conjugated with PEI.¹¹⁷ These exciting techniques and technologies all may have the potential to be applied to mRNA delivery,

demonstrating that the field of gold nanoparticle-mediated mRNA transport is rich with new possibilities.

Conclusion

mRNA-based vaccine and gene therapy technologies are innovative, promising and rapidly emerging strategies in biomedical research for the purpose of treating acquired and congenital diseases. Although naked mRNA has prospective clinical efficacy, this field faces various challenges such as the delivery issues, targeting administration and short-term gene expression, all of which are critical drawbacks and could be the major limitations to translate this field to widespread clinical application. The rapid advancements in biomaterials and nanotechnology could significantly help overcome these obstacles. This review has receptively presented the progress of cutting-edge biomaterials and nanotechnology platforms to improve the efficacy of mRNA delivery. While further improvements are necessary, it is encouraging to see that some of the mRNA-based treatment strategies^{24,26,118} have already entered the clinical trial stage within a short time following their development. More efforts are still required to find novel biomaterials/nanoparticles and optimized formulations which can provide high transfection activity and biocompatibility with minimal carrier-specific toxicity/immunostimulation, high selectivity and specificity, and effective systemic *in vivo* delivery and prolonged protein expression (particularly for gene therapy). With the advancement and proper design of biomaterials, it is expected that mRNA technology will be of high interest in clinical applications for years to come.

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References

- 1 H. Yin, R. L. Kanasty, A. A. Eltoukhy, A. J. Vegas, J. R. Dorkin and D. G. Anderson, *Nat. Rev. Genet.*, 2014, **15**, 541–555.
- 2 E. S. Quabius and G. Krupp, *Nat. Biotechnol.*, 2015, **32**, 229–235.
- 3 R. P. Deering, S. Kommareddy, J. B. Ulmer, L. A. Brito and A. J. Geall, *Expert Opin Drug Deliv.*, 2014, **11**, 885–899.
- 4 J. Lee, D. Boczkowski and S. Nair, *Methods Mol. Biol.*, 2013, **969**, 111–125.
- 5 A. Yamamoto, M. Kormann, J. Rosenecker and C. Rudolph, *Eur. J. Pharm. Biopharm.*, 2009, **71**, 484–489.

- 6 C. Leonhardt, G. Schwake, T. R. Stogbauer, S. Rappl, J. T. Kuhr, T. S. Ligon and J. O. Radler, *Nanomedicine*, 2014, **10**, 679–688.
- 7 T. S. Ligon, C. Leonhardt and J. O. Radler, *PLoS One*, 2014, **9**, e107148.
- 8 X. Wu and G. Brewer, *Gene*, 2012, **500**, 10–21.
- 9 E. Grudzien-Nogalska, J. Kowalska, W. Su, A. N. Kuhn, S. V. Slepencov, E. Darzynkiewicz, U. Sahin, J. Jemielity and R. E. Rhoads, *Methods Mol. Biol.*, 2013, **969**, 55–72.
- 10 Y. Wang, H. H. Su, Y. Yang, Y. Hu, L. Zhang, P. Blancafort and L. Huang, *Mol. Ther.*, 2013, **21**, 358–367.
- 11 J. Stepinski, C. Waddell, R. Stolarski, E. Darzynkiewicz and R. E. Rhoads, *RNA*, 2001, **7**, 1486–1495.
- 12 J. Jemielity, T. Fowler, J. Zuberek, J. Stepinski, M. Lewdorowicz, A. Niedzwiecka, R. Stolarski, E. Darzynkiewicz and R. E. Rhoads, *RNA*, 2003, **9**, 1108–1122.
- 13 E. Grudzien-Nogalska, J. Jemielity, J. Kowalska, E. Darzynkiewicz and R. E. Rhoads, *RNA*, 2007, **13**, 1745–1755.
- 14 J. Peng, E. L. Murray and D. R. Schoenberg, *Methods Mol. Biol.*, 2008, **419**, 215–230.
- 15 M. S. Kormann, G. Hasenpusch, M. K. Aneja, G. Nica, A. W. Flemmer, S. Herber-Jonat, M. Huppmann, L. E. Mays, M. Illenyi, A. Schams, M. Griese, I. Bittmann, R. Handgretinger, D. Hartl, J. Rosenecker and C. Rudolph, *Nat. Biotechnol.*, 2011, **29**, 154–157.
- 16 G. Tavernier, O. Andries, J. Demeester, N. N. Sanders, S. C. De Smedt and J. Rejman, *J. Controlled Release*, 2011, **150**, 238–247.
- 17 C. E. Thomas, A. Ehrhardt and M. A. Kay, *Nat. Rev. Genet.*, 2003, **4**, 346–358.
- 18 M. D. Mattozzi, M. J. Voges, P. A. Silver and J. C. Way, *J. Vis. Exp.*, 2014, **86**, e51234.
- 19 H. Dewitte, S. Van Lint, C. Heirman, K. Thielemans, S. C. De Smedt, K. Breckpot and I. Lentacker, *J. Controlled Release*, 2014, **194**, 28–36.
- 20 S. Van Meirvenne, L. Straetman, C. Heirman, M. Dullaers, C. De Greef, V. Van Tendeloo and K. Thielemans, *Cancer Gene Ther.*, 2002, **9**, 787–797.
- 21 M. L. De Temmerman, H. Dewitte, R. E. Vandenbroucke, B. Lucas, C. Libert, J. Demeester, S. C. De Smedt, I. Lentacker and J. Rejman, *Biomaterials*, 2011, **32**, 9128–9135.
- 22 H. Dewitte, S. Van Lint, C. Heirman, K. Thielemans, S. C. De Smedt, K. Breckpot and I. Lentacker, *J. Controlled Release*, 2014, **194c**, 28–36.
- 23 R. Kanasty, J. R. Dorkin, A. Vegas and D. Anderson, *Nat. Mater.*, 2013, **12**, 967–977.
- 24 B. Weide, J. P. Carralot, A. Reese, B. Scheel, T. K. Eigentler, I. Hoerr, H. G. Rammensee, C. Garbe and S. Pascolo, *J. Immunother.*, 2008, **31**, 180–188.
- 25 P. Midoux and C. Pichon, *Expert Rev. Vaccines*, 2015, **14**, 221–234.
- 26 B. Weide, S. Pascolo, B. Scheel, E. Derhovanessian, A. Pflugfelder, T. K. Eigentler, G. Pawelec, I. Hoerr, H. G. Rammensee and C. Garbe, *J. Immunother.*, 2009, **32**, 498–507.
- 27 K. J. Kallen, U. Gnad-Vogt, B. Scheel, G. Rippin and A. Stenzl, *J. Immunother. Cancer*, 2013, **1**, 1–1.
- 28 C. Cheng, A. J. Convertine, P. S. Stayton and J. D. Bryers, *Biomaterials*, 2012, **33**, 6868–6876.
- 29 J. H. Yeom, S. M. Ryou, M. Won, M. Park, J. Bae and K. Lee, *PLoS One*, 2013, **8**, e75369.
- 30 G. Koch, *Curr. Top. Microbiol. Immunol.*, 1973, **62**, 89–138.
- 31 R. W. Malone, P. L. Felgner and I. M. Verma, *Proc. Natl. Acad. Sci. U. S. A.*, 1989, **86**, 6077–6081.
- 32 M. Strenkowska, J. Kowalska, M. Lukaszewicz, J. Zuberek, W. Su, R. E. Rhoads, E. Darzynkiewicz and J. Jemielity, *New J. Chem.*, 2010, **34**, 993–1007.
- 33 A. K. Banerjee, *Microbiol. Rev.*, 1980, **44**, 175–205.
- 34 R. J. Jackson, *Cell*, 1993, **74**, 9–14.
- 35 A. E. Pasquinelli, J. E. Dahlberg and E. Lund, *RNA*, 1995, **1**, 957–967.
- 36 A. Cai, M. Jankowska-Anyszka, A. Centers, L. Chlebicka, J. Stepinski, R. Stolarski, E. Darzynkiewicz and R. E. Rhoads, *Biochemistry*, 1999, **38**, 8538–8547.
- 37 L. Weill, E. Belloc, F. A. Bava and R. Mendez, *Nat. Struct. Mol. Biol.*, 2012, **19**, 577–585.
- 38 D. Schwartz, C. J. Decker and R. Parker, *RNA*, 2003, **9**, 239–251.
- 39 V. F. Van Tendeloo, P. Ponsaerts, F. Lardon, G. Nijs, M. Lenjou, C. Van Broeckhoven, D. R. Van Bockstaele and Z. N. Berneman, *Blood*, 2001, **98**, 49–56.
- 40 S. Holtkamp, S. Kreiter, A. Selmi, P. Simon, M. Koslowski, C. Huber, O. Tureci and U. Sahin, *Blood*, 2006, **108**, 4009–4017.
- 41 M. Mockey, C. Goncalves, F. P. Dupuy, F. M. Lemoine, C. Pichon and P. Midoux, *Biochem. Biophys. Res. Commun.*, 2006, **340**, 1062–1068.
- 42 U. Sahin, K. Kariko and O. Tureci, *Nat. Rev. Drug Discovery*, 2014, **13**, 759–780.
- 43 G. J. Cao and N. Sarkar, *Proc. Natl. Acad. Sci. U. S. A.*, 1992, **89**, 10380–10384.
- 44 A. C. Beckel-Mitchener, A. Miera, R. Keller and N. I. Perrone-Bizzozero, *J. Biol. Chem.*, 2002, **277**, 27996–28002.
- 45 G. Shaw and R. Kamen, *Cell*, 1986, **46**, 659–667.
- 46 R. D. Klausner, T. A. Rouault and J. B. Harford, *Cell*, 1993, **72**, 19–28.
- 47 K. J. Kallen and A. Thess, *Therapeutic Advances in Vaccines*, 2014, vol. 2, pp. 10–31.
- 48 K. Kariko, A. Kuo and E. Barnathan, *Gene Ther.*, 1999, **6**, 1092–1100.
- 49 S. S. Diebold, T. Kaisho, H. Hemmi, S. Akira and C. Reis e Sousa, *Science*, 2004, **303**, 1529–1531.
- 50 S. R. Nallagatla, R. Toroney and P. C. Bevilacqua, *RNA Biol.*, 2008, **5**, 140–144.
- 51 M. Fotin-Mleczek, K. M. Duchardt, C. Lorenz, R. Pfeiffer, S. Ojkic-Zrna, J. Probst and K. J. Kallen, *J. Immunother.*, 2011, **34**, 1–15.

- 52 M. Fotin-Mleczek, K. Zanzinger, R. Heidenreich, C. Lorenz, A. Thess, K. M. Duchardt and K. J. Kallen, *J. Gene Med.*, 2012, **14**, 428–439.
- 53 I. Hoerr, R. Obst, H. G. Rammensee and G. Jung, *Eur. J. Immunol.*, 2000, **30**, 1–7.
- 54 S. M. Rittig, M. Haentschel, K. J. Weimer, A. Heine, M. R. Muller, W. Brugger, M. S. Horger, O. Maksimovic, A. Stenzl, I. Hoerr, H. G. Rammensee, T. A. Holderried, L. Kanz, S. Pascolo and P. Brossart, *Mol. Ther.*, 2011, **19**, 990–999.
- 55 H. Kubler, T. Maurer, A. Stenzl, S. Feyerabend, U. Steiner, M. Schostak, W. Schultze-Seemann, F. Vom Dorp, L. Pilla and G. Viatali, *J. Clin. Oncol.*, 2011, **29**, 4535.
- 56 S. Rausch, C. Schwentner, A. Stenzl and J. Bedke, *Hum. Vaccin. Immunother.*, 2014, **10**, 3146–3152.
- 57 M. Sebastian, A. Papachristofilou, C. Weiss, M. Fruh, R. Cathomas, W. Hilbe, T. Wehler, G. Rippin, S. D. Koch, B. Scheel, M. Fotin-Mleczek, R. Heidenreich, K. J. Kallen, U. Gnad-Vogt and A. Zippelius, *BMC Cancer*, 2014, **14**, 748.
- 58 ClinicalTrials.gov, Trial of an RNActive®-Derived Cancer Vaccine in Stage IIIB/IV Non Small Cell Lung Cancer (NSCLC), <https://clinicaltrials.gov/ct2/show/NCT00923312?term=cv9201&rank=1>, (accessed 06 April, 2015).
- 59 M. Sebastian, L. V. Boehmer, A. Zippelius, F. Mayer, M. Reck, D. Atanackovic, M. Thomas, F. Schneller, S. W. Jan, E. Goekkurt, H. Bernhard, A. Groeschel, B. Scheel, S. D. Koch, T. Lander, G. Rippin, V. Wiegand, U. S. Gnad-Vogt, K. J. Kallen and A. Knuth, *J. Clin. Oncol.*, 2012, **2012**, 30, Abstract: 2573.
- 60 B. Scheel, R. Teufel, J. Probst, J. P. Carralot, J. Geginat, M. Radsak, D. Jarrossay, H. Wagner, G. Jung, H. G. Rammensee, I. Hoerr and S. Pascolo, *Eur. J. Immunol.*, 2005, **35**, 1557–1566.
- 61 F. T. Zohra, Y. Maitani and T. Akaike, *Biol. Pharm. Bull.*, 2012, **35**, 111–115.
- 62 Y. Bao, Y. Jin, P. Chivukula, J. Zhang, Y. Liu, J. Liu, J. P. Clamme, R. I. Mahato, D. Ng, W. Ying, Y. Wang and L. Yu, *Pharm. Res.*, 2013, **30**, 342–351.
- 63 P. Harvie, F. M. Wong and M. B. Bally, *J. Pharm. Sci.*, 2000, **89**, 652–663.
- 64 J. Heyes, K. Hall, V. Taylor, R. Lenz and I. MacLachlan, *J. Controlled Release*, 2006, **112**, 280–290.
- 65 M. C. Woodle, *Adv. Drug Delivery Rev.*, 1998, **32**, 139–152.
- 66 V. P. Torchilin, *Nat. Rev. Drug Discovery*, 2005, **4**, 145–160.
- 67 M. L. Immordino, F. Dosio and L. Cattel, *Int. J. Nanomedicine*, 2006, **1**, 297–315.
- 68 H. Hatakeyama, H. Akita, K. Kogure, M. Oishi, Y. Nagasaki, Y. Kihira, M. Ueno, H. Kobayashi, H. Kikuchi and H. Harashima, *Gene Ther.*, 2007, **14**, 68–77.
- 69 L. Y. Song, Q. F. Ahkong, Q. Rong, Z. Wang, S. Ansell, M. J. Hope and B. Mui, *Biochim. Biophys. Acta*, 2002, **1558**, 1–13.
- 70 S. Uzgun, G. Nica, C. Pfeifer, M. Bosinco, K. Michaelis, J. F. Lutz, M. Schneider, J. Rosenecker and C. Rudolph, *Pharm. Res.*, 2011, **28**, 2223–2232.
- 71 F. Perche, T. Benvegna, M. Berchel, L. Lebegue, C. Pichon, P. A. Jaffres and P. Midoux, *Nanomedicine*, 2011, **7**, 445–453.
- 72 M. Mockey, E. Bourseau, V. Chandrashekar, A. Chaudhuri, S. Lafosse, E. Le Cam, V. F. Quesniaux, B. Ryffel, C. Pichon and P. Midoux, *Cancer Gene Ther.*, 2007, **14**, 802–814.
- 73 D. Hirsch-Lerner, M. Zhang, H. Eliyahu, M. E. Ferrari, C. J. Wheeler and Y. Barenholz, *Biochim. Biophys. Acta*, 2005, **1714**, 71–84.
- 74 L. Wasungu, M. C. Stuart, M. Scarzello, J. B. Engberts and D. Hoekstra, *Biochim. Biophys. Acta*, 2006, **1758**, 1677–1684.
- 75 H. Farhood, N. Serbina and L. Huang, *Biochim. Biophys. Acta*, 1995, **1235**, 289–295.
- 76 L. Wasungu and D. Hoekstra, *J. Controlled Release*, 2006, **116**, 255–264.
- 77 S. E. D'Souza, M. H. Ginsberg and E. F. Plow, *Trends Biochem. Sci.*, 1991, **16**, 246–250.
- 78 F. T. Zohra, E. H. Chowdhury, S. Tada, T. Hoshiba and T. Akaike, *Biochem. Biophys. Res. Commun.*, 2007, **358**, 373–378.
- 79 F. T. Zohra, E. H. Chowdhury, M. Nagaoka and T. Akaike, *Anal. Biochem.*, 2005, **345**, 164–166.
- 80 F. T. Zohra, E. H. Chowdhury and T. Akaike, *Biomaterials*, 2009, **30**, 4006–4013.
- 81 K. K. Phua, S. K. Nair and K. W. Leong, *Nanoscale*, 2014, **6**, 7715–7729.
- 82 J. U. Menon, P. Ravikumar, A. Pise, D. Gyawali, C. C. Hsia and K. T. Nguyen, *Acta Biomater.*, 2014, **10**, 2643–2652.
- 83 H. Gul-Uludag, J. Valencia-Serna, C. Kucharski, L. A. Marquez-Curtis, X. Jiang, L. Larratt, A. Janowska-Wieczorek and H. Uludag, *Leuk. Res.*, 2014, **38**, 1299–1308.
- 84 M. C. Chen, K. Sonaje, K. J. Chen and H. W. Sung, *Biomaterials*, 2011, **32**, 9826–9838.
- 85 A. A. Ranade, P. P. Bapsy, S. Nag, D. Raghunadharao, V. Raina, S. H. Advani, S. Patil, A. Maru, V. P. Gangadharan, C. Goswami, J. S. Sekhon, K. Sambasivaiah, P. Parikh, A. Bakshi and R. Mohapatra, *Asia Pac. J. Clin. Oncol.*, 2013, **9**, 176–181.
- 86 D. W. Pack, A. S. Hoffman, S. Pun and P. S. Stayton, *Nat. Rev. Drug Discovery*, 2005, **4**, 581–593.
- 87 L. Zhang, J. M. Chan, F. X. Gu, J. W. Rhee, A. Z. Wang, A. F. Radovic-Moreno, F. Alexis, R. Langer and O. C. Farokhzad, *ACS Nano*, 2008, **2**, 1696–1702.
- 88 J. Wu, N. Kamaly, J. Shi, L. Zhao, Z. Xiao, G. Hollett, R. John, S. Ray, X. Xu, X. Zhang, P. W. Kantoff and O. C. Farokhzad, *Angew. Chem., Int. Ed.*, 2014, **53**, 8975–8979.
- 89 M. G. Nava-Arzaluz, E. Pinon-Segundo, A. Ganem-Rondero and D. Lechuga-Ballesteros, *Recent Pat. Drug Delivery Formulation*, 2012, **6**, 209–223.
- 90 E. Pinon-Segundo, M. G. Nava-Arzaluz and D. Lechuga-Ballesteros, *Recent Pat. Drug Delivery Formulation*, 2012, **6**, 224–235.
- 91 T. Bettinger, R. C. Carlisle, M. L. Read, M. Ogris and L. W. Seymour, *Nucleic Acids Res.*, 2001, **29**, 3882–3891.
- 92 J. Rejman, G. Tavernier, N. Bavarsad, J. Demeester and S. C. De Smedt, *J. Controlled Release*, 2010, **147**, 385–391.

- 93 M. L. Read, S. Singh, Z. Ahmed, M. Stevenson, S. S. Briggs, D. Oupicky, L. B. Barrett, R. Spice, M. Kendall, M. Berry, J. A. Preece, A. Logan and L. W. Seymour, *Nucleic Acids Res.*, 2005, **33**, e86.
- 94 L. Zhang, J. M. Chan, F. X. Gu, J.-W. Rhee, A. Z. Wang, A. F. Radovic-Moreno, F. Alexis, R. Langer and O. C. Farokhzad, *ACS Nano*, 2008, **2**, 1696–1702.
- 95 S. Krishnamurthy, R. Vaiyapuri, L. Zhang and J. M. Chan, *Biomater. Sci.*, 2015, **3**, 923–936.
- 96 C. Salvador-Morales, L. Zhang, R. Langer and O. C. Farokhzad, *Biomaterials*, 2009, **30**, 2231–2240.
- 97 B. Mandal, H. Bhattacharjee, N. Mittal, H. Sah, P. Balabathula, L. A. Thoma and G. C. Wood, *Nanomedicine*, 2013, **9**, 474–491.
- 98 S. D. Kong, M. Sartor, C. M. Hu, W. Zhang, L. Zhang and S. Jin, *Acta Biomater.*, 2013, **9**, 5447–5452.
- 99 C. Salvador-Morales, L. Zhang, R. Langer and O. C. Farokhzad, *Biomaterials*, 2009, **30**, 2231–2240.
- 100 D. E. Owens III and N. A. Peppas, *Int. J. Pharm.*, 2006, **307**, 93–102.
- 101 B. T. Luk, R. H. Fang and L. Zhang, *Theranostics*, 2012, **2**, 1117–1126.
- 102 X. Su, J. Fricke, D. G. Kavanagh and D. J. Irvine, *Mol. Pharm.*, 2011, **8**, 774–787.
- 103 C. Clawson, L. Ton, S. Aryal, V. Fu, S. Esener and L. Zhang, *Langmuir*, 2011, **27**, 10556–10561.
- 104 J. Shi, Z. Xiao, A. R. Votruba, C. Vilos and O. C. Farokhzad, *Angew. Chem., Int. Ed.*, 2011, **123**, 7165–7169.
- 105 J. Shi, Y. Xu, X. Xu, X. Zhu, E. Pridgen, J. Wu, A. R. Votruba, A. Swami, B. R. Zetter and O. C. Farokhzad, *Nanomedicine*, 2014, **10**, 897–900.
- 106 X. Su, J. Fricke, D. G. Kavanagh and D. J. Irvine, *Mol. Pharm.*, 2011, **8**, 774–787.
- 107 D. M. Anderson, L. L. Hall, A. R. Ayyalapu, V. R. Irion, M. H. Nantz and J. G. Hecker, *Hum. Gene Ther.*, 2003, **14**, 191–202.
- 108 S. M. Ryou, S. Kim, H. H. Jang, J. H. Kim, J. H. Yeom, M. S. Eom, J. Bae, M. S. Han and K. Lee, *Biochem. Biophys. Res. Commun.*, 2010, **398**, 542–546.
- 109 N. L. Rosi, D. A. Giljohann, C. S. Thaxton, A. K. Lytton-Jean, M. S. Han and C. A. Mirkin, *Science*, 2006, **312**, 1027–1030.
- 110 J. H. Kim, J. H. Yeom, J. J. Ko, M. S. Han, K. Lee, S. Y. Na and J. Bae, *J. Biotechnol.*, 2011, **155**, 287–292.
- 111 P. C. Patel, D. A. Giljohann, W. L. Daniel, D. Zheng, A. E. Prigodich and C. A. Mirkin, *Bioconjugate Chem.*, 2010, **21**, 2250–2256.
- 112 D. A. Giljohann, D. S. Seferos, W. L. Daniel, M. D. Massich, P. C. Patel and C. A. Mirkin, *Angew. Chem., Int. Ed.*, 2010, **49**, 3280–3294.
- 113 D. A. Giljohann, D. S. Seferos, P. C. Patel, J. E. Millstone, N. L. Rosi and C. A. Mirkin, *Nano Lett.*, 2007, **7**, 3818–3821.
- 114 J. A. Ryan, K. W. Overton, M. E. Speight, C. N. Oldenburg, L. Loo, W. Robarge, S. Franzen and D. L. Feldheim, *Anal. Chem.*, 2007, **79**, 9150–9159.
- 115 A. Elbakry, A. Zaky, R. Liebl, R. Rachel, A. Goepferich and M. Breunig, *Nano Lett.*, 2009, **9**, 2059–2064.
- 116 E. Zhao, Z. Zhao, J. Wang, C. Yang, C. Chen, L. Gao, Q. Feng, W. Hou, M. Gao and Q. Zhang, *Nanoscale*, 2012, **4**, 5102–5109.
- 117 J. Kim, J. Park, H. Kim, K. Singha and W. J. Kim, *Biomaterials*, 2013, **34**, 7168–7180.
- 118 H. Kübler, A. Stenzl, W. Schultze-Seemann, F. vom Dorp, L. Pilla, C. Hampel, D. Jocham and K. Miller, *Eur. J. Cancer*, 2011, **47**, S498–S499.
- 119 R. M. Conry, A. F. LoBuglio, M. Wright, L. Sumerel, M. J. Pike, F. Johannig, R. Benjamin, D. Lu and D. T. Curiel, *Cancer Res.*, 1995, **55**, 1397–1400.
- 120 R. D. Granstein, W. Ding and H. Ozawa, *J. Invest. Dermatol.*, 2000, **114**, 632–636.
- 121 J. A. Wolff, R. W. Malone, P. Williams, W. Chong, G. Acsadi, A. Jani and P. L. Felgner, *Science*, 1990, **247**, 1465–1468.
- 122 L. Zangi, K. O. Lui, A. von Gise, Q. Ma, W. Ebina, L. M. Ptaszek, D. Spater, H. Xu, M. Tabebordbar, R. Gorbatov, B. Sena, M. Nahrendorf, D. M. Briscoe, R. A. Li, A. J. Wagers, D. J. Rossi, W. T. Pu and K. R. Chien, *Nat. Biotechnol.*, 2013, **31**, 898–907.
- 123 B. Scheel, S. Braedel, J. Probst, J. P. Carralot, H. Wagner, H. Schild, G. Jung, H. G. Rammensee and S. Pascolo, *Eur. J. Immunol.*, 2004, **34**, 537–547.
- 124 ClinicalTrials.gov, Trial of RNActive®-Derived Prostate Cancer Vaccine in Metastatic Castrate-refractory Prostate Cancer, <https://www.clinicaltrials.gov/ct2/show/NCT01817738?term=curevac&rank=2>, (accessed 12 May, 2015).
- 125 S. McLenachan, D. Zhang, A. B. Palomo, M. J. Edel and F. K. Chen, *PLoS One*, 2013, **8**, e83596.
- 126 S. Zou, K. Scarfo, M. H. Nantz and J. G. Hecker, *Int. J. Pharm.*, 2010, **389**, 232–243.
- 127 O. O. Markov, N. L. Mironova, M. A. Maslov, I. A. Petukhov, N. G. Morozova, V. V. Vlassov and M. A. Zenkova, *J. Controlled Release*, 2012, **160**, 200–210.
- 128 K. K. Phua, K. W. Leong and S. K. Nair, *J. Controlled Release*, 2013, **166**, 227–233.
- 129 D. M. Anderson, L. L. Hall, A. R. Ayyalapu, V. R. Irion, M. H. Nantz and J. G. Hecker, *Hum. Gene Ther.*, 2003, **14**, 191–202.