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Post COVID-19 complications and follow up biomarkers

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Millions of people were infected by the coronavirus disease (COVID-19) epidemic, which left a huge burden on the care of post COVID-19 survivors around the globe. The self-reported COVID-19 symptoms were experienced by an estimated 1.3 million people in the United Kingdom (2% of the population), and these symptoms persisted for about 4 weeks from the beginning of the infection. The symptoms most frequently reported were exhaustion, shortness of breath, muscular discomfort, joint pain, headache, cough, chest pain, cognitive impairment, memory loss, anxiety, sleep difficulties, diarrhea, and a decreased sense of smell and taste in post-COVID-19 affected people. The post COVID-19 complications were frequently related to the respiratory, cardiac, nervous, psychological and musculoskeletal systems. The lungs, liver, kidneys, heart, brain and other organs had been impaired by hypoxia and inflammation in post COVID-19 individuals. The upregulation of substance "P" (SP) and various cytokines such as tumor necrosis factor-alpha (TNF- α), interleukin 6 (IL-6), interleukin 10 (IL-10), interleukin 1 beta (IL-1 β), angiotensin-converting enzyme 2 (ACE2) and chemokine C-C motif ligand 3 (CCL3) has muddled respiratory, cardiac, neuropsychiatric, dermatological, endocrine, musculoskeletal, gastrointestinal, renal and genitourinary complications in post COVID-19 people. To prevent these complications from worsening, it was therefore important to study how these biomarkers were upregulated and block their receptors.

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1. Introduction and background

The COVID-19 viral infection was first reported in Wuhan, China, in December 2019 and its prevalence caused respiratory distress syndrome worldwide.¹⁻³ It was estimated that 5 million people were facing COVID-19 across the globe.⁴ It was reported on March 5, 2022, that there were 446 511 318 cases globally recorded with 6 004 421 deaths (WHO, 2022).⁵ In Pakistan, there were 1 516 150 cases reported and the number of deaths increased to 30 287 (NHSRC, 2022).⁶ The COVID-19 pandemic infected millions of people around the world, putting a financial burden on the treatment of COVID-19 survivors. An estimated 1.3 million people living in their houses in the United Kingdom, which was 2% of the population, experienced self-reported COVID-19 symptoms and these symptoms persisted for ≥ 4 weeks after the onset of the COVID-19 infection.⁷

According to the World Health Organization (WHO), post COVID-19 is defined as symptoms and complications that

appeared after COVID-19 recovery, continued for ≥ 12 weeks and had a worsening impact on the patients' health and quality of life. The most common symptoms are fatigue, shortness of breath and neural dysfunction that may worsen over time.⁸

The post COVID-19 survivors continued to develop physical, mental and psychological symptoms and complications. It was reported that fatigue, shortness of breath (SOB), myalgia, arthralgia, headache, cough, chest pain, anxiety, memory loss and sleep disorders were present in post COVID-19 people. Post-COVID-19 individuals experienced impaired quality of life, and mental health and employment issues.⁹

Currently, there is limited literature available that reports possible risk factors, pathophysiology and biomarker expression in post COVID-19. This review focuses on the post COVID-19 symptoms, biomarkers, complications of respiratory, cardiovascular, hematological, neuropsychiatric, dermatological, diabetes mellitus, renal, gastrointestinal and genitourinary diseases. Moreover, we will discuss potential biomarkers that may escalate health impairments in COVID-19 (Fig. 1).

2. Post COVID-19 symptoms

Post COVID-19 caused systemic inflammation and hypoxia which could damage the kidneys, liver, heart, brain and other body organs.^{10,11} It was reported that dyspnea and reduced

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Long-Term Complications of Post COVID-19

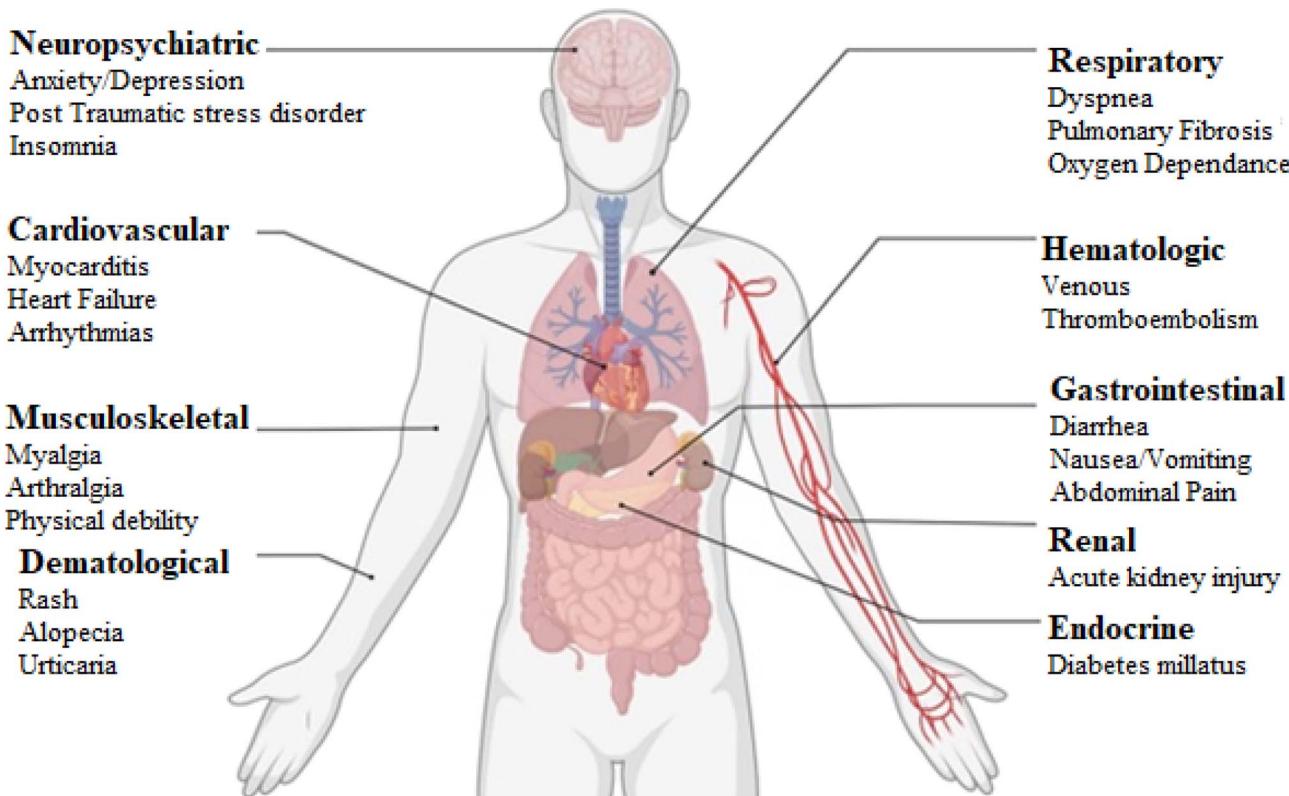


Fig. 1 Schematic representation of COVID-19 terms sequelae observed following the COVID-19 infection. This figure has been reproduced from ref. 30 with permission from American Journal of Physiology-Cell Physiology, copyright 2021.

exercise tolerance developed in 10–40% of the hospitalized patients within 2–4 months and shortness of breath developed in 65% of patients after discharge from the intensive care unit (ICU). It was documented that 10–35% of COVID-19 patients did not require hospitalization after post-COVID-19 symptoms.¹²

Hannah *et al.* (2020) studied responses from 3762 participants with confirmed COVID-19 from 56 countries and the duration of COVID-19 was at least 28 days. It was reported that 96% of patients experienced the symptoms of fatigue, dry cough, shortness of breath, headaches, muscle ache, chest tightness and sore throat apart from the loss of taste and smell beyond 90 days. The post COVID-19 symptoms that continued after 6 months were fatigue, post-exertional malaise and cognitive dysfunction. Most of the people recovered approximately after seven months from COVID-19, but not all the symptoms were relieved.¹³ The common clinical systemic complications of post COVID-19 were interstitial lung disease (organized pneumonia and pulmonary fibrosis), pulmonary embolism, chronic cough, cavity lesions, lower airway pulmonary hypertension lung fibrosis, arterial thrombosis, venous thromboembolism, cardiac thrombosis, strokes, inflammation, dermatological complications and overall neuropsychiatric dysfunctions¹⁴ A meta-analysis of studies involving $n = 17\,794$ patients revealed that troponin and aspartate levels increased with the worsening of COVID-19 conditions. Furthermore, this

study described that comorbidities of cerebrovascular diseases, chronic heart diseases, obstructive pulmonary disease, hypertension, chronic kidney disease, metastatic neoplasm and diabetes mellitus increased the risk of mortality and severity in post COVID-19 patients.¹⁵

Post COVID-19 affected survivors at all levels of age including adults, children and those who were not hospitalized, but possible risk factors were reported including female sex, early dyspnea and prior psychiatric disorders.²⁸ Older age, severe pneumonia, long ICU duration, mechanical ventilation duration, smoking and chronic alcoholism history were predictors for lung fibrosis development.²⁹ These symptoms may be controlled by overcoming blockage in the expression pathways of systemic biomarkers which contributed to escalate post COVID-19 worsening.

3. Post COVID-19 pathological complications

3.1 Respiratory complications

The most common pulmonary symptom reported following COVID-19 is dyspnea which persisted in 22.9 to 53% of patients 2 months after COVID-19.^{31,32} One of the most serious complications of the post-acute period is the development of



pulmonary fibrosis.³³ The post-COVID-19 survivors have the potential to develop coughs or shortness of breath and if symptoms worsen, they can develop extensive fibrosis of the lungs.³⁴ Oxygen dependency was reported in up to 6.6% of survivors. Diffuse alveolar damage, cytokines recruiting lymphocytes, macrophages and neutrophils which recruit fibroblasts result in pulmonary fibrosis.³⁵ Therefore, the risk of developing fibrotic lung changes increased in post COVID-19 survivors.³⁶ After COVID-19 recovery, most of the lungs would heal without complications but minimal fibrosis could develop which can progress to an advanced phase of fibrosis.³⁷

In our opinion, strategies should be developed to control the development of pulmonary fibrosis by looking at which main inflammatory agents contributed to the production of cytokine storms at the alveolar level in order to speed up the healing process and get rid of pulmonary fibrosis (Table 1).

3.2 Cardiovascular complications

The COVID-19 infection caused systematic inflammation, which further caused cardiovascular complications, morbidity and mortality. It was reported that troponin levels increased by 7–17% with the severity of the COVID-19 infection.³⁸ Chest pain

was reported in 21% of patients 60 days after discharge from the hospital. The susceptibility to cardiovascular complications such as ischemic and non-ischemic heart disease, cerebrovascular disorders, pericarditis, heart failure, myocarditis, shocks, dysrhythmia and thromboembolic complications increased.^{39,40} The most serious cardiovascular complication in COVID-19 patients was myocarditis, and deaths were reported in 7% of the patients with myocarditis.⁴¹ Eman and Toraih (2020) concluded that an increase in cardiac enzymes could be used as a biomarker for the identification of those patients who are at a higher risk of suffering from post-COVID-19 complications.¹⁵

It seems that post-COVID-19 cardiovascular complications occurred due to inflammatory changes caused by the production of cytokines and substance P. So, it may be possible to use systemic anti-inflammatory drugs to defeat the causative agents.

3.3 Hematological complications

It was reported that COVID-19 also invaded vascular endothelial cells and increases pro-inflammatory cytokine release and vascular endothelial injury which resulted in impaired fibrinolysis, thrombosis and hematological complications.⁴²

Table 1 Clinical symptoms of COVID-19 survivors

Symptoms	% Of symptoms observed												
	Ref.	16	17	18	19	20	21	22	23	24	25	26	27
Fatigue	28	53	52	—	39	—	98	63	—	—	—	—	31
Fever	—	—	—	—	—	—	75	—	—	—	—	—	—
Diaphoresis	24	—	—	—	—	—	—	—	—	—	—	—	—
Post-activity tachypnea	21	—	—	—	—	—	—	—	—	—	—	—	—
Palpitation	—	—	—	11	—	—	—	—	11	—	—	19	—
Chest pain	12	22	—	13	—	5	73	—	—	—	—	10	—
Psychosocial distress	23	—	—	—	—	17	—	—	—	39	—	—	—
Anxiety	7	—	—	—	—	—	—	—	7	—	30	16	—
Depression	—	—	—	—	—	—	—	23	—	33	—	—	—
Malaise	—	—	—	—	—	—	—	—	—	—	—	—	33
Concentration impairment	—	—	—	—	—	—	—	—	—	—	—	—	26
Headache	—	—	—	—	—	—	83	—	6	—	—	—	—
Weakness	—	—	—	—	—	—	—	—	—	—	—	—	41
Myalgia	—	—	—	—	22	6	88	—	9	—	—	—	—
Arthralgia	8	27	—	16	—	6	78	9	9	—	—	—	—
Inability to walk	—	—	—	—	—	—	40	—	—	—	—	—	—
Cough	7	—	—	—	11	—	73	7	6	—	—	—	—
Dyspnea	—	43	—	8	39	6	87	26	15	—	—	—	25
Wheezing	—	—	—	—	—	—	48	—	—	—	—	—	—
Abdominal pain	—	—	—	—	5	—	—	—	—	—	—	—	—
Diarrhea	—	—	—	—	—	—	59	—	—	—	—	—	—
Anorexia	—	—	—	—	—	—	—	—	8	—	—	—	—
Rhinorrhea	—	—	—	—	—	—	34	—	—	—	—	—	—
Ageusia	—	—	—	—	—	—	—	—	7	10	—	—	—
Sore throat	—	—	—	—	—	—	71	—	—	—	—	—	—
Anosmia	—	—	—	—	11	5	—	26	12	—	—	—	—
Sleep disorders	17	—	—	23	24	—	—	—	—	—	—	17	—
Dizziness	—	—	—	—	—	—	—	6	5	—	—	—	—
Hair loss	29	—	—	—	—	—	—	22	—	—	24	—	—
Reduced quality of life	—	—	—	—	—	—	—	—	—	—	—	26	—
Number of symptoms observed	10/30	4/30	1/30	5/30	7/30	6/30	13/30	12/30	8/30	3/30	5/30	6/30	—



Endothelial pathway activation increased the risk of thrombotic events in post COVID-19 survivors. The microvascular dysfunction enhanced the expression of tissue factors in response to the inflammatory cytokines as well as the effects of hypoxia that contributed to hematological complications.^{43,44} So, this revealed to us that certain cytokines caused vascular endothelial injuries which may be reduced by inhibiting the production of cytokines by using medicines.

3.4 Neuropsychiatric complications

COVID-19 infections invade the neural system and are involved in hypoxic brain injury, metabolic imbalances and oxidative stress which lead to worse neurogenic symptoms or complications in survivors.⁴⁵ The two-month monitoring revealed that loss of smell was 13.11% and taste was 11% post-traumatic stress disorder 28%, depression 31%, anxiety 42% and insomnia 40% in post COVID-19. Sun *et al.* (2021) reported that plasma cytokines IL-4 and IL-6 increased in post COVID-19 recovering individuals from 1 to 3 months. The peripheral and neuro-inflammation in post COVID-19 infection increased neurological complications. Individuals recovering from COVID-19 experienced concealed neural and neurological symptoms.⁴⁶ It was reported that depression was 31%, anxiety was 42%, obsessive compulsive disorders were 20% and insomnia was 40%. A record of $n = 402$ adults with psychiatric symptoms after surviving COVID-19 showed that increased inflammation was related to worsening depression in COVID-19 survivors, and inflammation based on peripheral lymphocytes, neutrophils and platelets can be used as biomarkers for psychiatric disorders.⁴⁷

Over 30% of post COVID-19 survivors complained of altered memory status.⁴⁸ The neurological symptoms such as headache, fatigue, brain fog, dizziness, memory loss, confusion, dysautonomia and difficulty focusing, are associated with post COVID-19 infections.^{49–51} Therefore, early screening and detailed rehabilitation strategies may be necessary to prevent and manage post-COVID-19 complications and their worsening.

3.5 Dermatological complications

The literature reported that dermatological complications developed in COVID-19 due to deposition in dermal capillaries or interstitials.⁵² Dermal complications such as maculopapular rash, vesicular rash microvascular vasculitis, urticaria, chilblain and petechiae are associated with COVID-19.^{53,54} A meta-analysis reported that the most common complication reported was maculopapular exanthema in 36.1% of patients with COVID-19. The papulovesicular rash (34.7%), painful red acral purple papules (15.3%) and urticaria (9.7%) were reported in the hands and feet in post-COVID-19 patients.⁵⁵ It seems that there are certain other biological components which are related to post-COVID-19 and worsening of dermatology. So we should look further to see which more components are involved and deposited in dermal capillaries and interstitials to intensify complications. Proper medications should be taken to cure these dermatological ailments.

3.6 Diabetes mellitus complications

COVID-19 causes increased oxidative stress and the release of pro-inflammatory cytokines in diabetic patients as compared to healthy individuals. COVID-19 damaged β cells and is related to an insulin deficiency which led to hyperglycemia.⁵⁶ The post COVID-19 patients with diabetes mellitus types I and II developed hyperglycemia, loss of acute compensation for diabetes and diabetic ketoacidosis.⁵⁷ The data showed that the prevalence of diabetes was 15.4% in post COVID-19 patients. Therefore, it seems that oxidative stress and cytokine production after post COVID-19 and other complex interrelated processes contributed to hyperglycemia and had an impact on β Langerhans cells and the appearance of glucose uptake receptors on the cell surface.^{58,59} Psychotherapy and medicines that may minimize the secretions of cytokines may be helpful.

3.7 Renal complications

The renal function was reported to be abnormal in 35% of patients at the time of discharge from hospitals, and in those patients 30% required dialysis. During follow-up in the same study, 36% of patients with residual kidney disease had recovered at the time of discharge but 14% of those who had recovered before discharge had recurrent kidney disease. Direct COVID-19 viral damage, systemic hypoxia, abnormal coagulation and effects of inflammatory cytokines were the contributing factors to acute kidney injury.^{60,61} Renal abnormalities occurred in most patients with COVID-19 pneumonia. Renal complications in COVID-19 were associated with higher mortality.⁶² Kidney dialysis is required in this situation.

3.8 Gastrointestinal complications

Diarrhea was among the top 10 most common symptoms and its prevalence was reported to be 6%. Other COVID-19 symptoms were nausea, vomiting, abdominal pain and loss of appetite. In post COVID-19 patients with acute liver injury, abnormal liver function may persist but improve over time.⁶³ Assessment of dietary intake and personalized dietary recommendations are the best strategies to recover.

3.9 Musculoskeletal complications

Arthritis was associated with some kind of viral infection. COVID-19 caused myalgia and arthralgia without true inflammatory arthritis in most patients. Post COVID-19 was less related to rheumatological diseases in patients.^{64,65} Fatigue, myalgia and arthralgia were most commonly reported in post COVID-19 survivors.^{66,67} It was documented that musculoskeletal complications arise due to high pro-inflammatory cytokines that are related to post COVID-19.^{68,69} Elevated levels of the pro-inflammatory factor interleukin-6 may be related to myalgia and joint pain.⁷⁰ Drugs such as siltuximab, sarilumab, and tocilizumab which have antiinflammatory effects, and inhibit the production of IL-6, may be used on the prescription of a doctor for recovery in post COVID-19 patients.



3.10 Genitourinary complications

It was shown by the literature survey that there was a significant reduction in dihydrotestosterone, testosterone, and orchitis in several patients. Viral testicular inflammation was reported in up to 19% of patients. In female patients, luteinizing hormone levels were elevated and preterm delivery took place with no vertical transmission.⁷¹ COVID-19 is associated with severe physiological changes in the testes and impairment of spermatogenesis, hormonal imbalances secondary to hypogonadism and sexual dysfunction.⁷² Hypogonadism correlates with unregulated pro-inflammatory cytokines TNF- α , IL-1 β and IL-6.⁷³ A recent study found that 30% of COVID-19 patients still had persistent symptoms or complications after nine months and the majority of non-hospitalized patients (85%) had mild symptoms.⁷⁴ So, it seemed that production of TNF- α , IL-1 β and IL-6 resulted in increased post genitourinary complications and may be stopped by blocking the pathways that cause the production of these inflammatory components. Psychological support and proper nutrition, including micro and macronutrients, are required for recovery, which may upgrade the sexual functioning of patients.

4. Biomarkers of COVID-19

Humans have approximately 20 000–25 000 protein-encoding genes.⁷⁵ There could be over 1 000 000 proteoforms including splice variants and essential post-translational modifications.⁷⁶ There were 29 predicted proteins in the COVID-19 virus with four main structural proteins (s, e, m, and n).⁷⁷ The post translation proteins (PTMs) could provide additional signatures to detect COVID-19 and its effects. The consideration of PTMs such as glycosylation and lipidation could provide more opportunities for the detection of viral proteins. A concentration on COVID-19 protein-derived glyco-conjugate peptides may enable the specificity and sensitivity of detection of the viral protein to be improved.⁷⁸

Early biomarker identification improved the management of COVID-19 patients and the appropriate allocation of healthcare resources during the pandemic. Anne *et al.* (2020) systematically reviewed routine laboratory tests of 45 studies for severe COVID-19 disease and described that an increased level of lymphocytes, neutrophil count, C reactive protein (CRP), lactate dehydrogenase, D-dimer, and aspartate aminotransferase and a low level of hemoglobin was observed in severe COVID-19 patients.⁷⁹ A significant association was presented between lymphopenia, thrombocytopenia, and elevated levels of procalcitonin, CRP, lactate dehydrogenase (LDH), and D-dimer and COVID-19 severity.⁸⁰ Protein markers of neuronal dysfunction such as amyloid beta, neurofilament light, neurogranin, total tau and p-T181-tau increased in COVID-19 compared to historic controls.⁸¹ Raymond's (2020) meta-analysis on $n = 967$ patients from six studies concluded that N terminal pro B type natriuretic peptide (NTP-BNP) was associated with increased mortality in COVID-19.⁸² Li *et al.* (2020) systematically reviewed 4189 patients with COVID-19 from 28 studies, which revealed that higher troponin, creatinine kinase myoglobin binding

myoglobin and NT-BNP were associated with higher mortality in COVID-19.⁸³ NTP-BNP can be used as a biomarker in patients without chronic heart failure, but has a poor prognosis in patients with sepsis and acute respiratory distress syndrome (ARDS).⁸⁴

Jing *et al.* (2020) conducted a systematic review regarding the blood coagulation biomarkers in patients with severe COVID-19 by searching databases from PubMed, Embase, Web of Science and the Cochrane Library. A record of 13 studies with $n = 1341$ adult patients showed that low platelets, higher D dimers and higher fibrinogen were associated with severity in patients with COVID-19. No correlation was manifested between activated partial thromboplastin time (APTT) or prothrombin time (PT) and the severity of COVID-19.⁸⁵ Jiaxi Xu and Eric Lazartigues (2022) studied ACE2 in COVID-19 patients with severe neurological symptoms and the possibility that COVID-19 can infect and injure the central nervous system in humans. It was determined that the expression of ACE2 in human neurons is related to COVID-19 by using human pluripotent stem cell-derived neurons and the possibility of a patient's ability to respire and worsen respiratory failure.⁸⁶ Interleukin 4 (IL-4) is a cytokine that is a potent regulator of immunity and is secreted primarily by mast cells, T helper 2 cells (Th2), eosinophils and basophils. Interleukin 4 (IL-4) is related to abnormal learning, memory, neurogenesis and neurological disorders. Sachin (2012) conducted extensive research on the role of IL-4 in immunity. The study evidence indicated that it has a critical role in the central nervous system (CNS) for memory and learning functions.⁸⁷

The data suggested that a systematic and longitudinal assessment of cytokines is required for the COVID-19 disease. A cytokine storm profile associated with severe COVID-19 disease was characterized by increased interleukin 2 (IL-2), interleukin 7 (IL-7), granulocyte-colony stimulating factor, interferon- γ inducible protein 10, monocyte chemoattractant protein 1, macrophage inflammatory protein 1 alpha and tumor necrosis factor alpha.⁸⁸ Elevated concentrations of immunological interleukin markers were consistent with the "cytokine storm" hypothesis, whereas elevated concentrations of D dimer were related to inappropriate blood coagulation as a possible contributor to pathogenesis.⁸⁹ Wenjie *et al.* (2020) reviewed published articles according to preferred reporting items for systematic reviews and meta-analyses (PRISMA) guidelines and evaluated the risk factors associated with mortality in COVID-19. Data ($n = 4659$) retrieved from 14 studies showed that hypertension, coronary heart disease and diabetes were likely more associated with mortality in COVID-19 patients. The greater levels of troponin, CRP, Interleukin-6 (IL-6), D dimer, creatinine and alanine transaminase and low level of albumin were presented in the mortality group. The increased acute response was related to cardiac inflammation and renal, liver and hematologic organ damage in COVID-19.⁹⁰ Yujun *et al.* (2020) provided evidence that interleukin 6 (IL-6) induced the exhaustion of lymphocytes in COVID-19 and that pro-inflammatory IL-6 levels increased in severely ill COVID-19 patients as compared to moderately ill patients.⁹¹ Sun *et al.* (2021) studied neurological sequelae in COVID-19 from 1 to 3



Table 2 Predictive immune biomarkers of COVID-19

Type	Family	Biomarkers	Amino acid	Ref.
Tachykinin peptides	Neuropeptide family	SP	11	109 and 110
Cytokines ¹¹¹	Interleukin family	IL-6	212	112 and 113
		IL-7	177	114
		1L-1 β	269	115
		1L-18	193	114
		TNF- α	269	116 and 117
	Interferon family	IFN-I	166	118
	Chemokine family	CCR2	374	119
Anaphylatoxins	Glycoproteins	MCP1/CCL2	25	120–122
Metalloenzymes	Zinc metalloenzyme	C3a, C5a	77, 74	123 and 124
Mucine	Glycosylated proteins	ACE2	805	86, 125–127
Brain natriuretic peptide	N-terminal prohormone of brain natriuretic peptide	MUC-1 & SAC	72	128
Protein	Apolipoproteins family	NT-proBNP	76	82
	Pentameric protein	SAA1	122	129 and 130
Cells	—	CRP	187	131 and 132
		T cells		133–137
		NK cells		138 and 139
		Eosinophils, basophils, and mast cell		140 and 141
Immunoglobulin protein	Antibodies	Anti-SARS-CoV-2 N/SIgM/IgG ab		46 and 142

months and found that cytokines (IL-4 and IL-6), antibody levels and neuronal enriched extracellular vesicle protein cargo were increased in COVID-19 patients.⁸¹ Piotr Lorkiewicz and Napoleon Waszkiewicz (2021) searched PubMed, Scopus and Google Scholar scientific literature databases to evaluate the development of post-COVID depression. They found that increased levels of IL-6, interleukin 1 β (IL-1 β), tumor necrosis factor alpha (TNF- α), interferon-gamma (IFN- γ), interleukin 10 (IL-10), interleukin 2 (IL-2), soluble interleukin 2 receptor (SIL-2R), CRP, monocyte chemoattractant protein 1 (MCP-1), and serum amyloid A1 (SAA1), and a declining level of brain derived neurotrophic factor (BDNF) and tryptophan were associated with the development of depression in post-COVID-19.⁹²

The substance “P” (SP) is a neuropeptide related to the tachykinin peptide family. The SP primary structure consists of 11 amino acids (RKPQQFFGLM) that have a C terminal α amidated methionine residue. SP is released in the central and peripheral nervous systems and distributed in the human brain (Kast *et al.*, 2015).⁹³ Neurokinin 1 is a receptor for SP which is distributed in the gastrointestinal tract, lung, urinary tract, muscles, peripheral tissues, immune cells and thyroid in mammals.^{94,95} SP is involved in the regulation of the cardiovascular system, dilatation of the vascular system, neural response, gastric motility, inflammation, pain, depression and respiratory mechanisms.^{96–98} SP is released from nerves, macrophages, eosinophils, lymphocytes and dendrites, and binds to NK1. The mast cells produced histamines, chemokines and cytokines in response to the SP stimulus. SP inhibited transforming growth factor beta 1 (TGF- β 1), released macrophages and acted as an immune suppressor. Like interleukin 4, interferon gamma (IFN- γ) and TNF- α activated upregulation of SP-NK1.^{99–102} An elevated level of SP is involved in inflammation of the respiratory, gastrointestinal and musculoskeletal systems.¹⁰³ SP acts as a neuromodulator functioning in an

autocrine, endocrine, and paracrine manner. It is also released from non-neuronal cells like immune cells.¹⁰⁴ SP is associated with inflammation in asthma and chronic obstructive lung diseases. Viral infections like respiratory syncytial virus (RSV) caused upregulation of SP expression and were linked with inflammation in rat lungs.¹⁰⁵

The COVID-19 virus stimulates the trigeminal ganglion (TG) *via* trigeminal nerves. The trigeminal nerve has three branches named ophthalmic, maxillary and mandibular nerves and noxious stimulus-activated nociceptor afferents that have cell bodies present in the TG. As a result of nerve injury, neuropeptides such as SP are secreted which are involved in inflammation (Goto *et al.*, 2017).¹⁰⁶

Upregulation of SP impairment of mucociliary transport helps to secrete mucus or secretion from the airway. Nuclear factor kappa B (NF- κ B) pathways were reported to be involved in the SP overexpression in airways causing inflammation and inhibition of Interlukin10 (Sponchiado *et al.*, 2020).¹⁰⁷ Cytokine storms were aroused in the COVID-19 infection and contributed to airway inflammation (Riffat, 2021).¹⁰⁸

The cytokine storm syndrome is reported in critically ill COVID-19 patients and presents with high inflammatory mediators, systemic inflammation and multiple organ injury or failure.¹⁴³ Pro-inflammatory cytokines such as TNF- α , IL-6, IL-10, IL-1 β and chemokines increase in COVID-19 patients.¹⁴⁴ There are certain biomarkers such as TNF- α , IL-6, IL-10, IL-1 β chemokines (CCL2 and CCR2) and neuropeptides (substance P) which are produced systemically in post COVID-19 patients and are responsible for deregulation of organ functions (Table 2).

Substance P (SP) expression increased with the onset of viral infections. COVID-19 caused the induction of SP which has a counterpart in neurogenic inflammation in the lung. Stress-induced effects on intracerebral SP levels were mainly obtained from brain tissue measurements. Substance P



counterparts escalate the cytokine storm in COVID-19 patients. Therefore, it may be a good biomarker to follow up with post COVID-19 patients.

The minimum detectable limit of detection for TNF- α is 10 pg mL $^{-1}$. TNF- α levels were higher in the SARS-CoV-2 (+) symptomatic group (16 pg mL $^{-1}$) compared with the SARS-CoV-2 (-) symptomatic individuals (7.6 pg mL $^{-1}$). TNF- α and IL-6 are counterparts of the cytokine storm.¹⁴⁵ Since previous findings suggested the involvement of TNF- α -induced inflammation and the manifestation of long term COVID symptoms,¹⁴⁶ as a biomarker for the prognosis of severe COVID-19, targeting TNF- α using antibodies or inhibitors is a good biomarker.

The lower limit for the detection of IL-6 is 1.5 pg mL $^{-1}$, whereas the upper limit for detection is 5000 pg mL $^{-1}$ without any prior dilution.¹⁴⁷ But it may vary due to different ELISA kits. Some studies reported that it may be increased in sepsis (>1000 pg mL $^{-1}$ at COVID-19 onset).¹⁴⁸ In addition, patients with such complicated forms of COVID-19 had nearly threefold higher serum IL-6 levels than those with non-complicated disease.¹⁴⁹ IL-6 is a suitable biomarker to follow up with post COVID-19 patients. Monoclonal antibodies may be effective in reducing IL 6 expression in COVID 19.¹⁵⁰

COVID-19 patients had higher IL-18 levels compared to healthy subjects (103 [210] pg mL $^{-1}$ vs. 310 [502] pg mL $^{-1}$, $p = 0.006$). Levels of circulating IL-18 were considered increased in different diseases in relation to the disease characteristics and severity.¹⁵¹ High levels of IL-18 have been found in the blood of patients with many different types of allergic diseases. Serum IL-18 concentrations have a significant correlation with COVID-19 severity.¹⁵²

In normal healthy adults, an IL-1 β concentration in the serum ranges from 0.5 to 12 pg mL $^{-1}$. Interleukin-1 β (IL-1 β) is a member of the interleukin 1 family of cytokines that are instrumental in inflammatory responses. IL-1 β is an important, robust pro-inflammatory cytokine involved in the body's immune response against infection and injury. In the cytokine storm, the levels of several cytokines such as IL-1 β , IL-10 and TNF- α increased which correlate with disease severity.¹⁵³ But the range of minimum and maximum values was also narrower on days 3 and 6 than on day 0. Studies on longitudinal cytokine profiles in patients across all severities of COVID-19 are still limited.¹⁵⁴

IFN-I has important roles in protecting the lungs from the spread of respiratory viruses and reducing inflammation in the lung. Integrated immune analysis, including immune cell analysis, whole-blood transcriptomics and cytokine quantification in COVID-19 patients impaired the IFN-I response as a result of low IFN-I levels. The assay range is approximately 0.4–298 pg mL $^{-1}$. The reference range for a healthy population is less than 2.0.¹⁵⁵

Chemokine receptor CCR2 expression causes high monocyte numbers, which contribute to inflammation. During COVID-19 infection, CCR2 stimulates monocytes to enter the lung parenchyma and causes monocyte driven cytokine storm inflammation.¹⁵⁶ The critical point in this therapeutic approach is to pay attention to other mechanisms involved in migration of

monocytes and macrophages to the lung and which on worsening, cause fibrotic changes.

Monocyte chemoattractant protein-1 (MCP-1/CCL2) is one of the key chemokines that regulate the migration and infiltration of monocytes/macrophages.¹⁵⁷ The circulating levels of CCL2 were nearly doubled (264 vs. 134 pg mL $^{-1}$) in the severe group compared to the non-severe group of COVID-19.¹⁵⁸

Airway mucus from patients with COVID-19 had a higher level of MUC5AC, which may be detected only when secretions are taken by bronchoscopy.¹²⁸ Serum amyloid A (SAA) is the most prominent acute phase reactant as its serum levels in the acute phase response demonstrate the most notable increase. In healthy individuals, SAA is present at a blood concentration below 3 mg L $^{-1}$. Its level rises during the COVID-19 infection.¹⁵⁹ There is less published data on SAA as a biomarker of COVID-19 severity, and future longitudinal studies should be conducted to assess the clinical relevance of SAA with the severity of COVID-19 and follow-up biomarkers.

CRP might be related to the production of inflammatory cytokines, which fight against microbes, but when the immune system becomes hyperactive, it can damage lung tissue.¹⁶⁰ CRP is a valuable early marker but CRP levels in patients with COVID-19 who may progress from non-severe to severe cases need to be further studied in large-scale multicenter studies.

Basophils play an important role in promoting antibody response and mast cells may take part in the pro-coagulative status typical of COVID-19 patients. The limitations of this biomarker are the number and timing of serum sampling that affect the results in long COVID-19.¹⁶¹

The detection limitations of biomarkers for COVID-19 are a BSL-3 laboratory, well trained technicians, state of the art calibrated instruments, and high quality standardized kits that have high sensitivity and specificity that may detect threshold value for a biomarker.¹⁶² In the longitudinal studies, participants should be involved in the study from the very beginning to the end with their consent for COVID-19 biomarkers. Phlebotomists should be well trained for sample drawing and samples should be handled and stored properly and carefully as prescribed by the kit manufacturer's protocol.¹⁶³ Some patients who are PCR negative for COVID-19 have biomarkers for COVID-19. These results are false positives and they should be reported clinically in order to retain validity of data. Standardized techniques should be used for biomarker detection. The balance between assay sensitivity and specificity must be weighed to reduce the risk of false positives. The limitations of biomarkers are number and timing of serum sampling that affect result in long COVID-19. The assay should be performed within the given time limit mentioned in kit protocol.¹⁶⁴ The aforementioned conditions should be fulfilled properly to avoid detect limitations of biomarkers for COVID-19. These may be possible detection limitations for COVID-19 biomarkers and need further to be explored.

5. Outlook

Although great progress has been made in post COVID-19 complications and biomarkers, there are still several



problems to be addressed. The expression pathways for developing complications with respect to biomarkers should be investigated by using inhibitors to reduce and stop the cytokine storm in post-COVID-19 patients. A release of the substance P counterpart for worsening symptoms and complications in post-COVID-19 patients should be sorted out to lessen and cure COVID-19. It is suggested that patients with COVID-19 who experience symptoms should manage by blocking mast cells and their mediators, which could be useful for cure and needed to be investigated. Currently, the major constraint to controlling the symptoms and complications of COVID-19 patients is a lack of drugs that inhibit the production pathways for biomarkers. Neurological and musculoskeletal complications remained for long periods, and other complications were refractory. Drugs should be developed so that the care of these patients can be optimized to cure COVID-19 patients and give them a better, healthier life. Finally, expression pathways of biomarkers related to post-COVID-19 complications have not been investigated. We must take more serious steps for patients and be more concerned about limiting the expression pathways for the generation of these biomarkers, and there should be nationwide surveillance programmes to improve patients' health and quality of life.

6. Conclusion

Patients' education will be useful in identifying the post COVID-19 syndrome, which is defined by lingering symptoms and complications that occur after 4 weeks. After COVID-19, the expression of SP and several cytokines (TNF α , IL-6, IL-10, IL-1, ACE2 and CCL3) result in complications involving the respiratory system, heart, brain, endocrine system, skin, musculoskeletal system, gastrointestinal tract, kidneys and genitourinary system. Pulmonary fibrosis development is the worst complication because it affects the quality of life. We should rule out and block the SP expression pathway as well as that of other cytokines, which will help to overcome post COVID-19 complications.

Conflicts of interest

There are no conflicts to declare.

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References

- 1 J. She, J. Jiang, L. Ye, L. Hu, C. Bai and Y. Song, *Clin. Transl. Med.*, 2020, **9**, 1–7.
- 2 W. Wu, A. Wang and M. Liu, *Lancet*, 2020, **395**, 497–506.
- 3 D. McGonagle, K. Sharif, A. O'Regan and C. Bridgewood, *Autoimmun. Rev.*, 2020, **19**, 102537.
- 4 D. M. Altmann and R. J. Boyton, *BMJ*, 2021, **372**.
- 5 W. H. Organization, *World Health Organization 2020 guidelines on physical activity and sedentary behaviour*, 2020.
- 6 S. Shareef, S. Akhtar, N. Tufail and F. Ahmad, *J. Univ. Med. Dent. Coll.*, 2022, **13**, 299–303.
- 7 E. Rezel-Potts, A. Douiri, X. Sun, P. J. Chowienczyk, A. M. Shah and M. C. Gulliford, *PLoS Med.*, 2022, **19**, e1004052.
- 8 E. Wynberg, H. D. van Willigen, M. Dijkstra, A. Boyd, N. A. Kootstra, J. G. van den Aardweg, M. J. van Gils, A. Matser, M. R. de Wit and T. Leenstra, *Clin. Infect. Dis.*, 2022, **75**, e482–e490.
- 9 M. Chutiyami, U. M. Bello, D. Salihu, D. Ndwiga, M. A. Kolo, R. Maharaj, K. Naidoo, L. Devar, P. Pratitha and P. Kannan, *Int. J. Nurs. Stud.*, 2022, **104211**.
- 10 A. Gupta, M. V. Madhavan, K. Sehgal, N. Nair, S. Mahajan, T. S. Sehrawat, B. Bikdeli, N. Ahluwalia, J. C. Ausiello and E. Y. Wan, *Nat. Med.*, 2020, **26**, 1017–1032.
- 11 Y. Cao, A. Hirosi and S. Montgomery, *BMJ Open*, 2020, **10**, e043560.
- 12 H. C. Maltezou, A. Pavli and A. Tsakris, *Vaccines*, 2021, **9**, 497.
- 13 H. E. Davis, G. S. Assaf, L. McCorkell, H. Wei, R. J. Low, Y. Re'em, S. Redfield, J. P. Austin and A. Akrami, *eClinicalMedicine*, 2021, **38**, 101019.
- 14 S. SeyedAlinaghi, A. M. Afsahi, M. MohsseniPour, F. Behnezhad, M. A. Salehi, A. Barzegary, P. Mirzapour, E. Mehraeen and O. Dadras, *Arch. Acad. Emerg. Med.*, 2021, **9**.
- 15 E. A. Toraih, R. M. Elshazli, M. H. Hussein, A. Elgaml, M. Amin, M. El-Mowafy, M. El-Mesery, A. Ellythy, J. Duchesne and M. T. Killackey, *J. Med. Virol.*, 2020, **92**, 2473–2488.
- 16 Q. Xiong, M. Xu, J. Li, Y. Liu, J. Zhang, Y. Xu and W. Dong, *Clin. Microbiol. Infect.*, 2021, **27**, 89–95.
- 17 A. Carfi, R. Bernabei, F. Landi and C.-P.-A. C. S. G. Gemelli Against, *JAMA*, 2020, **324**, 603–605.
- 18 H. Fogarty, L. Townsend, H. Morrin, A. Ahmad, C. Comerford, E. Karampini, H. Englert, M. Byrne, C. Bergin, J. M. O'Sullivan, I. Martin-Loeches, P. Nadarajan, C. Bannan, P. W. Mallon, G. F. Curley, R. J. S. Preston, A. M. Rehill, D. McGonagle, C. Ni Cheallaigh, R. I. Baker, T. Renné, S. E. Ward and J. S. O'Donnell, *J. Thromb. Haemostasis*, 2021, **19**, 2546–2553.
- 19 C. Carvalho-Schneider, E. Laurent, A. Lemaignen, E. Beaufils, C. Bourbao-Tournois, S. Laribi, T. Flament, N. Ferreira-Maldent, F. Bruyère, K. Stefic, C. Gaudy-Graffin, L. Grammatico-Guillon and L. Bernard, *Clin. Microbiol. Infect.*, 2021, **27**, 258–263.
- 20 D. T. Arnold, F. W. Hamilton, A. Milne, A. J. Morley, J. Viner, M. Attwood, A. Noel, S. Gunning, J. Hattrick, S. Hamilton, K. T. Elvers, C. Hyams, A. Bibby, E. Moran, H. I. Adamali, J. W. Dodd, N. A. Maskell and S. L. Barratt, *Thorax*, 2021, **76**, 399–401.



21 M. Bellan, D. Soddu, P. E. Balbo, A. Baricich, P. Zeppegno, G. C. Avanzi, G. Baldon, G. Bartolomei, M. Battaglia, S. Battistini, V. Binda, M. Borg, V. Cantaluppi, L. M. Castello, E. Clivati, C. Cisari, M. Costanzo, A. Croce, D. Cuneo, C. De Benedittis, S. De Vecchi, A. Feggi, M. Gai, E. Gambaro, E. Gattoni, C. Gramaglia, L. Grisafi, C. Guerriero, E. Hayden, A. Jona, M. Invernizzi, L. Lorenzini, L. Loretì, M. Martelli, P. Marzullo, E. Matino, A. Panero, E. Parachini, F. Patrucco, G. Patti, A. Pirovano, P. Prosperini, R. Quaglino, C. Rigamonti, P. P. Sainaghi, C. Vecchi, E. Zecca and M. Pirisi, *JAMA Netw. Open*, 2021, **4**, e2036142.

22 A. Dennis, M. Wamil, J. Alberts, J. Oben, D. J. Cuthbertson, D. Wootton, M. Crooks, M. Gabbay, M. Brady, L. Hishmeh, E. Attree, M. Heightman, R. Banerjee and A. Banerjee, *BMJ Open*, 2021, **11**, e048391.

23 C. Huang, L. Huang, Y. Wang, X. Li, L. Ren, X. Gu, L. Kang, L. Guo, M. Liu, X. Zhou, J. Luo, Z. Huang, S. Tu, Y. Zhao, L. Chen, D. Xu, Y. Li, C. Li, L. Peng, Y. Li, W. Xie, D. Cui, L. Shang, G. Fan, J. Xu, G. Wang, Y. Wang, J. Zhong, C. Wang, J. Wang, D. Zhang and B. Cao, *Lancet*, 2021, **397**, 220–232.

24 K. Stavem, W. Ghanima, M. K. Olsen, H. M. Gilboe and G. Einvik, *Thorax*, 2021, **76**, 405–407.

25 H. Ahmed, K. Patel, D. C. Greenwood, S. Halpin, P. Lewthwaite, A. Salawu, L. Eyre, A. Breen, R. O'Connor, A. Jones and M. Sivan, *J. Rehabil. Med.*, 2020, **52**, jrm00063.

26 A. Parker, E. H. Louw, U. Lalla, C. F. N. Koegelenberg, B. W. Allwood, H. Rabie, S. I. Sibeko, J. J. Taljaard and S. Lahri, *S. Afr. Med. J.*, 2020, **110**, 957–958.

27 M. Michelen, L. Manoharan, N. Elkheir, V. Cheng, A. Dagens, C. Hastie, M. O'Hara, J. Suett, D. Dahmash, P. Bugaeva, I. Rigby, D. Munblit, E. Harriss, A. Burls, C. Foote, J. Scott, G. Carson, P. Olliaro, L. Sigfrid and C. Stavropoulou, *BMJ Glob Health*, 2021, **6**.

28 S. J. Yong, *Infect. Dis.*, 2021, **53**, 737–754.

29 A. Combrink, N. Irwin, G. Laudin, K. Naidoo, S. Plagerson and A. Mathee, *S. Afr. Med. J.*, 2010, **100**, 297–299.

30 A. D. Desai, M. Lavelle, B. C. Boursiquot and E. Y. Wan, *Am. J. Physiol. Cell Physiol.*, 2022, **322**, C1–C11.

31 V. Chopra, S. A. Flanders, M. O'Malley, A. N. Malani and H. C. Prescott, *Ann. Intern. Med.*, 2021, **174**, 576–578.

32 S. Mandal, J. Barnett, S. E. Brill, J. S. Brown, E. K. Denneny, S. S. Hare, M. Heightman, T. E. Hillman, J. Jacob and H. C. Jarvis, *Thorax*, 2021, **76**, 396–398.

33 H. F. Schwensen, L. K. Borreschmidt, M. Storgaard, S. Redsted, S. Christensen and L. B. Madsen, *J. Clin. Pathol.*, 2021, **74**, 400–402.

34 Z. L. Yang, C. Chen, L. Huang, S. C. Zhou, Y. N. Hu, L. M. Xia and Y. Li, *Front. Med.*, 2020, **7**, 605088.

35 L. T. McDonald, *Am. J. Physiol. Lung Cell. Mol. Physiol.*, 2021, **320**, L257–L265.

36 A.-R. Koczulla, *N. Engl. J. Med.*, 2014, **371**, 781.

37 P. M. George, S. L. Barratt, R. Condliffe, S. R. Desai, A. Devaraj, I. Forrest, M. A. Gibbons, N. Hart, R. G. Jenkins and D. F. McAuley, *Thorax*, 2020, **75**, 1009–1016.

38 F. Zhou, *Lancet*, 2020, **395**, 1054–1062.

39 J. H. Tanne, *BMJ*, 2022, 376–378.

40 F. Farshidfar, N. Koleini and H. Ardehali, *JCI Insight*, 2021, **6**.

41 Q. Ruan, K. Yang, W. Wang, L. Jiang and J. Song, *Intensive Care Med.*, 2020, **46**, 846–848.

42 R. Escher, N. Breakey and B. Lämmle, *Thromb. Res.*, 2020, **190**, 62.

43 T. Hadid, Z. Kafri and A. Al-Katib, *Blood Rev.*, 2021, **47**, 100761.

44 M. G. Lazzaroni, S. Piantoni, S. Masneri, E. Garrafa, G. Martini, A. Tincani, L. Andreoli and F. Franceschini, *Blood Rev.*, 2021, **46**, 100745.

45 M. Pennisi, G. Lanza, L. Falzone, F. Fisicaro, R. Ferri and R. Bella, *Int. J. Mol. Sci.*, 2020, **21**, 5475.

46 B. Sun, Y. Feng, X. Mo, P. Zheng, Q. Wang, P. Li, P. Peng, X. Liu, Z. Chen and H. Huang, *Emerging Microbes Infect.*, 2020, **9**, 940–948.

47 M. Mazza and R. Lorenzo, Anxiety and depression in COVID-19 survivors: role of inflammatory and clinical predictors, *Brain Behav. Immun.*, 2020, **89**, 594–600.

48 E. Garrigues, P. Janvier, Y. Kherabi, A. Le Bot, A. Hamon, H. Gouze, L. Doucet, S. Berkani, E. Oliosi and E. Mallart, *J. Infect.*, 2020, **81**, e4–e6.

49 A. S. Nordvig, K. T. Fong, J. Z. Willey, K. T. Thakur, A. K. Boehme, W. S. Vargas, C. J. Smith and M. S. Elkind, *Neurol. Clin. Pract.*, 2021, **11**, e135–e146.

50 M.-I. Stefanou, L. Palaiodimou, E. Bakola, N. Smyrnis, M. Papadopoulou, G. P. Paraskevas, E. Rizos, E. Boutati, N. Grigoriadis and C. Krogias, *Ther. Adv. Chronic Dis.*, 2022, **13**, 20406223221076890.

51 S. Shanbehzadeh, M. Tavahomi, N. Zanjari, I. Ebrahimi-Takamjani and S. Amiri-Arimi, *J. Psychosom. Res.*, 2021, **147**, 110525.

52 C. Magro, J. J. Mulvey, D. Berlin, G. Nuovo, S. Salvatore, J. Harp, A. Baxter-Stoltzfus and J. Laurence, *Transl. Res.*, 2020, **220**, 1–13.

53 S. Recalcati, *J. Eur. Acad. Dermatol. Venereol.*, 2020, **34**(5), e212–e213.

54 C. Galván Casas, A. Catala, G. Carretero Hernández, P. Rodríguez-Jiménez, D. Fernández-Nieto, A. Rodríguez-Villa Lario, I. Navarro Fernández, R. Ruiz-Villaverde, D. Falkenhain-López and M. Llamas Velasco, *Br. J. Dermatol.*, 2020, **183**, 71–77.

55 M. Sachdeva, R. Gianotti, M. Shah, L. Bradanini, D. Tosi, S. Veraldi, M. Ziv, E. Leshem and R. P. Dodiuk-Gad, *J. Dermatol. Sci.*, 2020, **98**, 75–81.

56 R. Unnikrishnan and A. Misra, *Nutr. Diabetes*, 2021, **11**, 21.

57 A. K. Singh and K. Khunti, *Annu. Rev. Med.*, 2022, **73**, 129–147.

58 A. Abdi, M. Jalilian, P. A. Sarbarzeh and Z. Vlaisavljevic, *Diabetes Res. Clin. Pract.*, 2020, **166**, 108347.

59 M. Faghir-Gangi, H. Moameri, N. Abdolmohamadi and S. Nematollahi, *Clin. Diabetol.*, 2020, **9**, 271–278.

60 L. Chan, K. Chaudhary, A. Saha, K. Chauhan, A. Vaid and I. Paranjpe, *J. Am. Soc. Nephrol.*, 2021, **32**, 151–160.



61 Y. Peleg, S. Kudose, V. D'Agati, E. Siddall, S. Ahmad, T. Nickolas, S. Kisselev, A. Gharavi and P. Canetta, *Kid. Int. Rep.*, 2020, **5**, 940–945.

62 G. Pei, *J. Am. Soc. Nephrol.*, 2020, **31**, 1157–1165.

63 Y.-W. An, S. Song, W.-X. Li, Y.-X. Chen, X.-P. Hu, J. Zhao, Z.-W. Li, G.-Y. Jiang, C. Wang and J.-C. Wang, *Int. J. Med. Sci.*, 2021, **18**, 176.

64 M. Marks and J. L. Marks, *Clin. Med.*, 2016, **16**, 129.

65 H. Zacharias, S. Dubey, G. Koduri and D. D'Cruz, *Autoimmun. Rev.*, 2021, **20**, 102883.

66 E. M. Amenta, A. Spallone, M. C. Rodriguez-Barradas, H. M. El Sahly, R. L. Atmar and P. A. Kulkarni, Postacute COVID-19: an overview and approach to classification, in *Open forum infectious diseases*, Oxford University Press, US, 2020, vol. 7(12), p. ofaa509.

67 L. Liang, B. Yang, N. Jiang, W. Fu, X. He, Y. Zhou, W.-L. Ma and X. Wang, *J. Korean Med. Sci.*, 2020, **35**, 418.

68 F. Salamanna, F. Veronesi, L. Martini, M. P. Landini and M. Fini, *Front. Med.*, 2021, 392.

69 P. K. Dos Santos, E. Sigoli, L. J. Braganca and A. S. Cornachione, *Front. Physiol.*, 2022, 510.

70 K. Nakamura, K. Saito, Y. Hara, T. Aoyagi, K. Kitakawa, Y. Abe, H. Takemura, F. Ikeda, M. Kaku and K. Kanemitsu, *BMC Infect. Dis.*, 2018, **18**, 1–5.

71 M. A. Khalili, K. Leisegang, A. Majzoub, R. Finelli, M. K. P. Selvam, R. Henkel, M. Mojgan and A. Agarwal, *World J. Men's Health*, 2020, **38**, 506.

72 B. Ebner, Y. Volz, J.-N. Mumm, C. G. Stief and G. Magistro, *Nat. Rev. Urol.*, 2022, **19**, 344–356.

73 L. M. Barton, E. J. Duval, E. Stroberg, S. Ghosh and S. Mukhopadhyay, *Am. J. Clin. Pathol.*, 2020, **153**, 725–733.

74 J. K. Logue, N. M. Franko, D. J. McCulloch, D. McDonald, A. Magedson, C. R. Wolf and H. Y. Chu, *JAMA Netw. Open*, 2021, **4**, e210830.

75 H. G. Sequencing, *Nature*, 2004, **431**, 931–945.

76 R. Aebersold, J. N. Agar, I. J. Amster, M. S. Baker, C. R. Bertozzi, E. S. Boja, C. E. Costello, B. F. Cravatt, C. Fenselau and B. A. Garcia, *Nat. Chem. Biol.*, 2018, **14**, 206–214.

77 F. Wu, S. Zhao, B. Yu, Y. Chen, W. Wang and Z. Song, *Nature*, 2020, **579**, 265–269.

78 Y. Watanabe, J. D. Allen, D. Wrapp, J. S. McLellan and M. Crispin, *bioRxiv*, 2020, preprint, DOI: [10.1101/2020.03.26.010322](https://doi.org/10.1101/2020.03.26.010322).

79 A. Alnor, M. B. Sandberg, C. Gils and P. J. Vinholt, *J. Appl. Lab. Med.*, 2020, **5**, 1038–1049.

80 P. Malik, U. Patel and D. Mehta, *BMJ Evid. Based Med.*, 2020, **26**, 107–108.

81 B. Sun, N. Tang, M. J. Peluso, N. S. Iyer, L. Torres, J. L. Donatelli, S. E. Munter, C. C. Nixon, R. L. Rutishauser and I. Rodriguez-Barraquer, *Cells*, 2021, **10**, 386.

82 R. Pranata, I. Huang, A. A. Lukito and S. B. Raharjo, *Postgrad. Med. J.*, 2020, **96**, 387–391.

83 J.-W. Li, T.-W. Han, M. Woodward, C. S. Anderson, H. Zhou, Y.-D. Chen and B. Neal, *Prog. Cardiovasc. Dis.*, 2020, **63**, 518–524.

84 M. Brueckmann, G. Huhle, S. Lang, K. K. Haase, T. Bertsch, C. Weiß, J. J. Kaden, C. Putensen, M. Borggrefe and U. Hoffmann, *Circulation*, 2005, **112**, 527–534.

85 J. Lin, H. Yan, H. Chen, C. He, C. Lin, H. He, S. Zhang, S. Shi and K. Lin, *J. Med. Virol.*, 2021, **93**, 934–944.

86 J. Xu and E. Lazartigues, *Cell. Mol. Neurobiol.*, 2022, **42**, 305–309.

87 P. Sachin, S. Gadani, J. Cronk, G. Norris and J. Kipnis, *J. Immunol.*, 2012, **189**, 4213–4219.

88 C. Huang and L. Ren, *Lancet*, 2021, **397**, 220–232.

89 A. D. Whetton, G. W. Preston, S. Abubeker and N. Geifman, *J. Proteome Res.*, 2020, **19**, 4219–4232.

90 W. Tian, W. Jiang, J. Yao, C. J. Nicholson, R. H. Li, H. H. Sigurslid, L. Wooster, J. I. Rotter, X. Guo and R. Malhotra, *J. Med. Virol.*, 2020, **92**, 1875–1883.

91 Y. Tang, J. Liu, D. Zhang, Z. Xu, J. Ji and C. Wen, *Front. Immunol.*, 2020, **11**, 1708.

92 P. Lorkiewicz and N. Waszkiewicz, *J. Clin. Med.*, 2021, **10**, 4142.

93 R. Kast, H. Schirok, S. Figueroa-Pérez, J. Mittendorf, M. Gnoth, H. Apeler, J. Lenz, J. Franz, A. Knorr and J. Hütter, *Br. J. Pharmacol.*, 2007, **152**, 1070–1080.

94 K. Ebner, S. B. Sartori and N. Singewald, *Curr. Pharm. Des.*, 2009, **15**, 1647–1674.

95 M. Muñoz and R. Coveñas, *Amino Acids*, 2014, **46**, 1727–1750.

96 M. Samsam, R. Coveñas, R. Ahangari, J. Yajeya, J. Narváez and G. Tramu, *PAIN®*, 2000, **84**, 389–395.

97 R. Bang, G. Sass, A. K. Kiemer, A. M. Vollmar, W. L. Neuhuber and G. Tiegs, *J. Pharmacol. Exp. Ther.*, 2003, **305**, 31–39.

98 M. Munoz and R. Covenas, *Peptides*, 2013, **48**, 1–9.

99 M. B. Johnson, A. D. Young and I. Marriott, *Front. Cell. Neurosci.*, 2017, **10**, 296.

100 I. Marriott and K. L. Bost, *Neuroimmune Circuits, Drugs of Abuse, and Infectious Diseases*, 2001, 247–254.

101 M. Covas, L. Pinto and R. Victorino, *Clin. Exp. Immunol.*, 1994, **96**, 384–388.

102 I. Marriott and K. L. Bost, *J. Immunol.*, 2000, **165**, 182–191.

103 T. M. O'Connor, J. O'Connell, D. I. O'Brien, T. Goode, C. P. Bredin and F. Shanahan, *J. Cell. Physiol.*, 2004, **201**, 167–180.

104 I. Chernova, J.-P. Lai, H. Li, L. Schwartz, F. Tuluc, H. M. Korchak, S. D. Douglas and L. E. Kilpatrick, *J. Leucocyte Biol.*, 2009, **85**, 154–164.

105 G. Piedimonte, M. M. Rodriguez, K. A. King, S. McLean and X. Jiang, *Am. J. Physiol. Lung Cell. Mol. Physiol.*, 1999, **277**, L831–L840.

106 T. Goto, H. Iwai, E. Kuramoto and A. Yamanaka, *Jpn. Dent. Sci. Rev.*, 2017, **53**, 117–124.

107 M. Sponchiado, Y. S. Liao, K. R. Atanasova, E. N. Collins, V. Schurmann, L. Bravo and L. R. Reznikov, *Physiol. Rep.*, 2021, **9**, e14749.

108 R. Mehbob, F. J. Ahmad, A. Qayyum, M. A. Rana, S. A. Gilani, M. A. Tariq and J. Akram, *MedRxiv*, 2020, preprint, DOI: [10.1101/2020.08.01.20166678](https://doi.org/10.1101/2020.08.01.20166678).



109 R. Mehboob, M. Kurdi, A. Bamaga, N. Aldardeir, H. Nasief, L. H. Moshref, T. Alsinani, A. O. Rayes and R. H. Jabbad, *Front. Med.*, 2021, **8**, 727593.

110 S. Smieszek, *Eur. Respir. J.*, 2021, **58**(55), OA4114.

111 M. Muñoz and R. J. P. Coveñas, 2013, **48**, 1–9.

112 J. Liu, X. Zheng, Q. Tong, W. Li, B. Wang, K. Sutter, M. Trilling, M. Lu, U. Dittmer and D. Yang, *J. Med. Virol.*, 2020, **92**, 491–494.

113 Z. Feng, B. Diao, R. Wang, G. Wang, C. Wang, Y. Tan, L. Liu, C. Wang, Y. Liu and Y. Liu, *MedRxiv*, 2020, preprint, DOI: [10.1101/2020.03.27.20045427](https://doi.org/10.1101/2020.03.27.20045427).

114 Y. Jamilloux, T. Henry, A. Belot, S. Viel, M. Fauter, T. El Jammal, T. Walzer, B. François and P. Sève, *Autoimmun. Rev.*, 2020, **19**, 102567.

115 E. Toniato, R. Ross and S. Kritas, *J. Biol. Regul. Homeost Agents*, 2020, **34**, 11–16.

116 E. Kamphuis, T. Junt, Z. Waibler, R. Forster and U. Kalinke, *Blood*, 2006, **108**, 3253–3261.

117 A. Ivagnès, M. Messaoudene, G. Stoll, B. Routy, A. Fluckiger, T. Yamazaki, K. Iribarren, C. P. Duong, L. Fend and A. Caignard, *Oncotarget*, 2018, **7**, e1386826.

118 S. Xia, Y. Tao, L. Cui, Y. Yu and S. Xu, *J. Immunol. Res.*, 2019, **2019**, 5370706.

119 F. Leuschner, G. Courties, P. Dutta, L. J. Mortensen, R. Gorbatov, B. Sena, T. I. Novobrantseva, A. Borodovsky, K. Fitzgerald and V. Koteliansky, *Eur. Heart J.*, 2015, **36**, 1478–1488.

120 M. S. Abers, O. M. Delmonte, E. E. Ricotta, J. Fintzi, D. L. Fink, A. A. A. de Jesus, K. A. Zaremba, S. Alehashemi, V. Oikonomou and J. V. Desai, *JCI Insight*, 2021, **6**, 144455.

121 M. Catanzaro, F. Fagiani, M. Racchi, E. Corsini, S. Govoni and C. Lanni, *Signal Transduction Targeted Ther.*, 2020, **5**, 84.

122 Y. Zhao, L. Qin, P. Zhang, K. Li, L. Liang, J. Sun, B. Xu, Y. Dai, X. Li and C. Zhang, *JCI Insight*, 2020, **5**, 139834.

123 L. Gralinski, T. Sheahan, T. Morrison, V. Menachery, K. Jensen, S. Leist, A. Whitmore, M. Heise and R. Baric, *mBio*, 2018, **9**, 10–128.

124 T. Gao, M. Hu, X. Zhang, H. Li, L. Zhu, H. Liu, Q. Dong, Z. Zhang, Z. Wang and Y. Hu, *MedRxiv*, 2020, preprint, DOI: [10.1101/2020.03.29.20041962](https://doi.org/10.1101/2020.03.29.20041962).

125 R. D. Lopes, A. V. S. Macedo, R. J. Moll-Bernardes, A. Feldman, G. D. A. S. Arruda, A. S. de Souza, D. C. de Albuquerque, L. Mazza, M. F. Santos and N. Z. Salvador, *Am. Heart J.*, 2020, **226**, 49–59.

126 W. Tai, L. He, X. Zhang, J. Pu, D. Voronin, S. Jiang, Y. Zhou and L. Du, *Cell. Mol. Immunol.*, 2020, **17**, 613–620.

127 H. Kai and M. Kai, *Hypertens. Res.*, 2020, **43**, 648–654.

128 Z. Xu, L. Shi, Y. Wang, J. Zhang, L. Huang, C. Zhang, S. Liu, P. Zhao, H. Liu and L. Zhu, *Lancet Respir. Med.*, 2020, **8**, 420–422.

129 M. Pieri, M. Ciotti, M. Nuccetelli, M. A. Perrone, M. T. Calio, M. S. Lia, M. Minieri and S. Bernardini, *Int. Immunopharmacol.*, 2021, **95**, 107512.

130 A. Zinelli, P. Palogiannis, C. Carru and A. A. Mangoni, *Int. J. Infect. Dis.*, 2021, **105**, 668–674.

131 D. Azoulay, M. Shehadeh, S. Chepa, E. Shaoul, M. Baroum, N. A. Horowitz and E. Kaykov, *J. Infect.*, 2020, **81**, e79–e81.

132 A. Bhargava, E. A. Fukushima, M. Levine, W. Zhao, F. Tanveer, S. M. Szpunar and L. Saravolatz, *Clin. Infect. Dis.*, 2020, **71**, 1962–1968.

133 F. Wang, J. Nie, H. Wang, Q. Zhao, Y. Xiong, L. Deng, S. Song, Z. Ma, P. Mo and Y. Zhang, *J. Infect. Dis.*, 2020, **221**, 1762–1769.

134 C. Guo, B. Li, H. Ma, X. Wang, P. Cai, Q. Yu, L. Zhu, L. Jin, C. Jiang and J. Fang, *bioRxiv*, 2020, **22**(5), bbab085, DOI: [10.1101/864488](https://doi.org/10.1101/864488).

135 C. Qin, L. Zhou, Z. Hu, S. Zhang, S. Yang and Y. Tao, *Clin. Infect. Dis.*, 2020, **71**, 762–768.

136 L. Lei, H. Qian, X. Yang, X. Zhang, D. Zhang, T. Dai, R. Guo, L. Shi, Y. Cheng and B. Zhang, *J. Cell. Mol. Med.*, 2020, **24**, 11603–11606.

137 D. Pinto, Y.-J. Park, M. Beltramello, A. C. Walls, M. A. Tortorici, S. Bianchi, S. Jaconi, K. Culap, F. Zatta and A. De Marco, *bioRxiv*, 2020, preprint, DOI: [10.1101/2020.04.07.023903](https://doi.org/10.1101/2020.04.07.023903).

138 O. J. McElvaney, N. L. McEvoy, O. F. McElvaney, T. P. Carroll, M. P. Murphy, D. M. Dunlea, O. Ní Choileáin, J. Clarke, E. O'Connor and G. Hogan, *Am. J. Respir. Crit. Med.*, 2020, **202**, 812–821.

139 D. Pinto, Y.-J. Park, M. Beltramello, A. C. Walls, M. A. Tortorici, S. Bianchi, S. Jaconi, K. Culap, F. Zatta and A. De Marco, Structural and functional analysis of a potent sarbecovirus neutralizing antibody, *bioRxiv*, 2020, preprint, DOI: [10.1101/2020.04.07.023903](https://doi.org/10.1101/2020.04.07.023903).

140 L. M. Buja, D. Wolf, B. Zhao, B. Akkanti, M. Do, L. Lelenwa, N. Do, G. Ottaviani, M. Tarek and D. O. Trujillo, *Cardiovasc. Pathol.*, 2019, **48**, 107233.

141 F. Yasui, C. Kai, M. Kitabatake, S. Inoue, M. Yoneda, S. Yokochi, R. Kase, S. Sekiguchi, K. Morita and T. Hishima, *J. Immunol.*, 2008, **181**, 6337–6348.

142 E. Adams, M. Ainsworth, R. Anand, M. I. Andersson, K. Auckland, J. K. Baillie, E. Barnes, S. Beer, J. I. Bell and T. Berry, *MedRxiv*, 2020, preprint, DOI: [10.1101/2020.04.15.20066407](https://doi.org/10.1101/2020.04.15.20066407).

143 W. Zhang, Y. Zhao, F. Zhang, Q. Wang, T. Li, Z. Liu, J. Wang, Y. Qin, X. Zhang and X. Yan, *Clin. Immunol.*, 2020, **214**, 108393.

144 B. Silva Andrade, S. Siqueira, W. R. de Assis Soares, F. de Souza Rangel, N. O. Santos, A. dos Santos Freitas, P. Ribeiro da Silveira, S. Tiwari, K. J. Alzahrani and A. Goes-Neto, *Viruses*, 2021, **13**, 700.

145 M. J. Pereson, M. N. Badano, N. Aloisi, R. Chuit, M. E. de Bracco and P. Bare, *Microbiol. Spectrum*, 2022, **10**, e01411–e01421.

146 M. J. Peluso, S. Lu, A. F. Tang, M. S. Durstenfeld, H.-e. Ho, S. A. Goldberg, C. A. Forman, S. E. Munter, R. Hoh and V. Tai, *J. Infect. Dis.*, 2021, **224**, 1839–1848.

147 D. Talwar, S. Kumar, S. Acharya, N. Raisinghani, S. Madaan, V. Hulkoti, A. Akhilesh, S. Khanna, D. Shah and S. Nimkar, *Ind. J. Crit. Care Med.*, 2022, **26**, 39.

148 D. Zhao, F. Yao, L. Wang, L. Zheng, Y. Gao, J. Ye, F. Guo, H. Zhao and R. Gao, *Clin. Infect. Dis.*, 2020, **71**, 756–761.



149 B. K. Patterson, J. Guevara-Coto, R. Yogendra, E. B. Francisco, E. Long, A. Pise, H. Rodrigues, P. Parikh, J. Mora and R. A. Mora-Rodríguez, *Front. Immunol.*, 2021, **12**, 2520.

150 F. Zeng, Y. Huang, Y. Guo, M. Yin, X. Chen, L. Xiao and G. Deng, *Int. J. Infect. Dis.*, 2020, **96**, 467–474.

151 H. Satış, H. S. Özger, P. A. Yıldız, K. Hızel, Ö. Gulbahar, G. Erbaş, G. Aygencel, O. G. Tunçcan, M. A. Öztürk and M. Dizbay, *Cytokine*, 2021, **137**, 155302.

152 S. A. Ihim, S. D. Abubakar, Z. Zian, T. Sasaki, M. Saffarioun, S. Maleknia and G. Azizi, *Front. Immunol.*, 2022, **13**, 919973.

153 F. Coperchini, L. Chiovato, G. Ricci, L. Croce, F. Magri and M. Rotondi, *Cytokine Growth Factor Rev.*, 2021, **58**, 82–91.

154 Z.-S. Xu, T. Shu, L. Kang, D. Wu, X. Zhou, B.-W. Liao, X.-L. Sun, X. Zhou and Y.-Y. Wang, *Signal Transduction Targeted Ther.*, 2020, **5**, 100.

155 J. Hadjadj, N. Yatim, L. Barnabei, A. Corneau, J. Boussier, N. Smith, H. Péré, B. Charbit, V. Bondet and C. Chenevier-Gobeaux, *Science*, 2020, **369**, 718–724.

156 R. A. Grant, L. Morales-Nebreda, N. S. Markov, S. Swaminathan, M. Querrey, E. R. Guzman, D. A. Abbott, H. K. Donnelly, A. Donayre and I. A. Goldberg, *Nature*, 2021, **590**, 635–641.

157 S. Lee, G. Y. Yoon, S. J. Lee, Y.-C. Kwon, H. W. Moon, Y.-J. Kim, H. Kim, W. Lee, G. U. Jeong and C. Kim, *Microbiol. Spectr.*, 2022, **10**, e02322–e02371.

158 C. Huang, Y. Wang, X. Li, L. Ren, J. Zhao, Y. Hu, L. Zhang, G. Fan, J. Xu and X. Gu, *Lancet*, 2020, **395**, 497–506.

159 I. Sorić Hosman, I. Kos and L. Lamot, *Front. Immunol.*, 2021, **11**, 631299.

160 S. A. B. Garcia and N. Guzman, in *StatPearls [Internet]*, StatPearls Publishing, 2022.

161 A. P. Rathore and A. L. St John, *Curr. Opin. Immunol.*, 2020, **66**, 74–81.

162 G. K. Jena, C. N. Patra and J. Sruti, in *Coronavirus Drug Discovery*, Elsevier, 2022, pp. 37–48.

163 A. Niraula, B. Gelal and M. Lamsal, *J. Nepal Med. Assoc.*, 2020, **58**, 1107.

164 A. N. Masterson, B. B. Muhoberac, A. Gopinadhan, D. J. Wilde, F. T. Deiss, C. C. John and R. Sardar, *Anal. Chem.*, 2021, **93**, 8754–8763.

